



1 The Stratified Microbial Carbon Pump: Thermal Stratification Enhances Refractory
2 Dissolved Organic Carbon Production and Stabilizes Carbon in Alkaline Karst Waters
3 through Keystone Microbial Interaction Networks

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26

27 **Abstract:** Recalcitrant dissolved organic carbon (RDOC) represents a persistent fraction of the
28 carbon pool and contributes substantially to regional carbon budgets in terrestrial aquatic
29 ecosystems. Given the substantial carbon sequestration potential of karst waters, elucidating the
30 dynamics and persistence of RDOC is essential for understanding carbon retention in these
31 geologically distinctive environments. Karst reservoirs represent important yet highly complex
32 components of regional carbon cycling. Here, RDOC accumulation and its underlying mechanisms
33 were examined in the Dalongdong (DLD) Reservoir, a dissolved-carbon-rich karst water body, over
34 three contrasting thermal phases: incubating thermal stratification (ITS), obvious thermal
35 stratification (OTS), and mixing (MX). Our results revealed that RDOC dynamics were
36 predominantly regulated by microbial processes rather than benthic carbon inputs. The
37 establishment of thermal stratification generated pronounced physicochemical gradients that
38 facilitated vertical niche partitioning among keystone taxa, thereby regulating DOM bioavailability
39 through taxon-specific metabolic pathways. Bacterial network analysis further indicated that
40 facilitative interactions favored the generation of labile carbon, whereas competitive interactions
41 under environmental stress promoted RDOC accumulation. Notably, the distinctive geochemical
42 conditions of the karst system facilitated Ca–P co-precipitation, resulting in persistent phosphorus



43 limitation and an elevated C:P ratio in the water column. This nutrient imbalance reduced microbial
44 carbon use efficiency and suppressed extracellular enzyme activities, consequently favoring the
45 conversion of autochthonous labile carbon into more persistent RDOC. Collectively, these findings
46 suggest that the synergistic coupling of the biological carbon pump (BCP) and microbial carbon
47 pump (MCP), reinforced by geochemical phosphorus sequestration, represents an important
48 mechanism underlying long-term carbon retention in alkaline, calcium-rich aquatic systems. Our
49 findings further demonstrate that karst-specific geochemical conditions interact with thermal
50 stratification to regulate microbial processes governing carbon persistence. In particular, MCP
51 efficiency appears to be modulated by carbonate-weathering-derived dissolved inorganic carbon
52 (DIC), providing mechanistic insights into the enhanced carbon sequestration capacity of
53 geologically distinctive inland waters. Protecting these highly efficient carbon-sequestering
54 ecosystems may therefore contribute to atmospheric CO₂ removal and provide an additional
55 pathway for advancing global climate mitigation.

56

57 **Keywords:** Refractory dissolved organic carbon, Thermal stratification, Bacteria interactions,
58 Carbon Sink, Microbial Carbon Pump

59

60 1. Introduction

61 Carbonate rock weathering transfers atmospheric and soil-derived CO₂ into DIC, which is
62 subsequently transported into karst aquatic systems, including rivers, lakes, and reservoirs (**Liu et**
63 **al., 2011**). Within karst systems, carbonate dissolution consumes CO₂ from CO₂-rich water while



64 simultaneously producing mobile DIC according to the reaction $\text{CaCO}_3 + \text{CO}_2 + \text{H}_2\text{O} \rightarrow \text{Ca}^{2+} +$
65 2HCO_3^- (Liu et al., 2018). This CO_2 consumption and subsequent DIC transport constitute the
66 fundamental mechanism underlying the karst carbon sink. In contrast to non-karst landscapes,
67 carbonate weathering provides a distinctive and potentially sustained pathway for the transfer of
68 inorganic carbon through terrestrial aquatic systems. Global carbon budget assessments suggest an
69 unresolved uncertainty of approximately $0.4\text{--}1.2 \text{ Pg C yr}^{-1}$ in terrestrial carbon sinks, with the karst
70 carbon sink potentially accounting for an important fraction of this so-called “missing carbon
71 sink”(Harris et al., 2021; Houghton and Nassikas, 2017; Friedlingstein et al., 2021;
72 Friedlingstein et al., 2022; Friedlingstein et al., 2023). China contains the largest extent of karst
73 landscapes globally, covering approximately 3.46 million km^2 , and these extensive karst regions
74 therefore represent substantial potential for carbon sequestration (Chen et al., 2023).

75 Carbon sequestration in karst systems is governed not only by DIC formation and transport
76 but also by biological processes. The BCP, a concept originally developed in marine
77 biogeochemical research, describes the biological uptake of inorganic carbon through
78 photosynthesis and its subsequent incorporation into organic carbon (Volk and Hoffert, 1985). In
79 karst aquatic environments, aquatic photosynthesis enables the abundant DIC pool to be
80 incorporated into biologically produced organic carbon (Liu et al., 2018). However, the
81 functioning of the BCP in karst waters may differ fundamentally from that in non-karst aquatic
82 systems, where BCP activity is often constrained by the availability of inorganic carbon. Karst
83 waters are typically characterized by high DIC concentrations, elevated pH, and relatively low
84 concentrations of dissolved CO_2 , suggesting that conventional carbon-limitation paradigms may



85 not adequately explain BCP dynamics in these environments. Under such hydrochemical
86 conditions, the fertilizing effect of aqueous CO₂ may stimulate autotrophic organic carbon (AOC)
87 production, thereby strengthening the potential for carbon sequestration (**Shao et al., 2025; Yang**
88 **et al., 2016**).

89 In contrast to the BCP, the MCP represents a complementary biogeochemical pathway through
90 which carbon is retained in aquatic environments via microbial metabolic processes. Through
91 microbial transformation, labile dissolved organic carbon (LDOC) and semi-labile dissolved
92 organic carbon (SLDOC) can be converted into RDOC, which is considerably more resistant to
93 biological degradation. Owing to its persistence, RDOC may remain in aquatic environments for
94 hundreds to thousands of years, thereby contributing to the formation of long-lived and relatively
95 stable carbon reservoirs (**Jiao et al., 2024**). The MCP framework provides a mechanistic basis for
96 understanding how microorganisms, including bacteria, archaea, and zooplankton, transform
97 bioavailable dissolved organic carbon into more persistent forms through metabolic activity (**Jiao**
98 **et al., 2024**). The resulting RDOC commonly exhibits humic-like fluorescent characteristics and
99 possesses molecular structures that are highly resistant to degradation, facilitating its prolonged
100 retention in aquatic systems and contributing to the establishment of stable carbon pools (**Jiao et**
101 **al., 2024; Jiao et al., 2010**). Increasing attention has recently been paid to MCP-mediated carbon
102 transformation in karst aquatic environments, including lakes, rivers, and reservoirs. For example,
103 He et al. reported that microbially derived organic carbon (MDOC) represented the predominant
104 accumulated fraction of RDOC in the Lijiang River, while *Sporichthyaceae* and *Novosphingobium*
105 played important roles in organic carbon formation and stabilization before and after the rainy



106 season (**He et al., 2022**). In the Honghua Reservoir, Liuzhou, aerobic anoxygenic phototrophic
107 bacteria (AAPB) were found to enhance the stability of the karst carbon sink by reducing the
108 degradation rate of organic carbon and extending its residence time (**He et al., 2022**). Shi et al.
109 further suggested that microbial community composition and taxon–taxon relationships play
110 substantial roles in determining RDOC production, while keystone taxa such as *Fluviicola* and
111 *Polynucleobacter* may indirectly contribute to its formation by modulating interactions within the
112 microbial community (**Shi et al., 2025**). Taken together, available evidence indicates that carbon
113 persistence in karst aquatic environments is closely governed by bacterial community organization
114 and the ecological relationships among taxa. However, the carbon sequestration function of karst
115 waters is highly sensitive to environmental variability. Nutrient inputs can reshape phytoplankton
116 community composition (**Caballero et al., 2025; Bao et al., 2022**) and alter microbial metabolic
117 pathways (**Lozupone and Knight, 2007; Bouvy et al., 1998**), thereby potentially modifying the
118 efficiencies of both the BCP and MCP. Such environmental sensitivity presents a substantial
119 challenge for effective water-resource management and the maintenance of carbon sequestration
120 functions in karst regions.

121 Lakes and reservoirs constitute major components of inland aquatic ecosystems and are
122 estimated to account for approximately 38% of carbon emissions from aquatic systems, making
123 them substantial contributors to global carbon fluxes (**Tranvik et al., 2009**). Among these inland
124 water bodies, karst reservoirs are particularly important because they function as transitional
125 interfaces between groundwater and surface water, where carbon cycling is jointly influenced by
126 the characteristics of karst landscapes and aquatic ecosystems (**Goldscheider et al., 2007**). Unlike



127 non-karst lakes and reservoirs, where carbon processing is often strongly influenced by terrestrially
128 derived organic matter transported from surrounding catchments, karst reservoirs are characterized
129 by distinctive carbon storage and greenhouse gas emission patterns associated with substantial
130 endogenous primary production. These unique biogeochemical characteristics may consequently
131 result in carbon cycling dynamics that differ markedly from those of non-karst aquatic systems.
132 Thermal stratification has been observed in some karst reservoirs, generating pronounced vertical
133 gradients in hydrochemical and physical conditions. Such spatial heterogeneity can restructure
134 microbial community composition and functional organization, thereby regulating microbial
135 carbon transformation pathways and ultimately influencing carbon cycling within the reservoir (**Pu**
136 **et al., 2020; Li et al., 2021; Jia et al., 2024**).

137 Despite extensive evidence supporting the MCP as a major pathway for persistent carbon
138 preservation in water bodies, the combined effects of physical stratification and karst-specific
139 geochemical conditions, particularly elevated alkalinity and Ca–P co-precipitation, on carbon
140 retention remain insufficiently resolved. We hypothesized that seasonal thermal stratification in
141 karst reservoirs creates a distinctive physicochemical environment that intensifies phosphorus
142 limitation and alters water-column C:P stoichiometry, thereby driving shifts in microbial
143 metabolism and strengthening the functional coupling between the BCP and MCP. To evaluate this
144 hypothesis, we conducted an integrated investigation focusing on three aspects: (1) the responses
145 of keystone bacterial taxa to stratification-induced hydrochemical gradients; (2) the relationships
146 between microbial co-occurrence network stability and DOM bioavailability; and (3) the regulatory
147 roles of geochemical phosphorus sequestration and nutrient stoichiometry in RDOC accumulation



148 under the distinctive geochemical conditions of karst reservoirs.

149 Southwestern China is characterized by an exceptionally dense distribution of karst reservoirs.
150 Among these water bodies, the Dalongdong Reservoir (DLD), situated in Shanglin County of
151 Guangxi Zhuang Autonomous Region, is primarily sustained by groundwater inputs within a karst
152 landscape. Its distinctive hydrological and geochemical features provide a favorable natural setting
153 for elucidating carbon dynamics within karst aquatic environments. In particular, the absence of
154 direct surface-river inflow minimizes external hydrological and biogeochemical inputs, while the
155 reservoir is characterized by high alkalinity and elevated DIC concentrations. These hydrochemical
156 features are widely observed across karst regions and make the DLD Reservoir a valuable natural
157 analogue for exploring inorganic-carbon-mediated processes in inland aquatic ecosystems at
158 broader spatial scales. Strong water – rock interactions, together with the seasonal establishment
159 and subsequent disruption of thermal stratification, create pronounced temporal and vertical
160 variability in the reservoir environment. These coupled geological and physical processes provide
161 an appropriate framework for examining how geological conditions interact with physical forcing
162 to regulate carbon sequestration in stratified inland waters.

163 Against this background, we focused on the DLD Reservoir and combined high-frequency
164 monitoring of thermal stratification with bacterial community co-occurrence network analysis.
165 Specifically, we sought to determine how thermal stratification, within the context of karst
166 geochemistry, influences RDOC formation through changes in the composition of key bacterial
167 taxa and their ecological interactions. By integrating seasonal physical variability with microbial
168 community organization and carbon transformation, this study seeks to clarify the mechanisms



169 governing the persistence and long-term retention of carbon within karst aquatic systems. More
170 broadly, our findings may provide new insights into the responses of carbon dynamics to ongoing
171 climatic warming across inland aquatic systems characterized by high alkalinity and carbonate-rich
172 geochemistry.

173

174 **2. Materials and methods**

175 **2.1 Study area**

176 Field investigations focused on the DLD Reservoir (23°30'01"–23°40'08" N, 108°30'02"–
177 108°36'04" E), a groundwater-dominated karst water body situated in Shanglin County of Guangxi,
178 southwestern China (**Fig. 1**). Hydrologically, DLD occupies the headwater section of the Qingshui
179 River within the Hongshui River tributary system. Built in 1958 through impoundment of a natural
180 karst valley, the reservoir encompasses a landscape containing interconnected underground cavities,
181 dolines, and rock fractures. Its principal operational purposes are agricultural irrigation and
182 hydroelectric power production, while the associated drainage basin covers approximately 310 km².
183 The impounded water stretches approximately 13 km, generally spans 500–600 m in width, and
184 broadens to more than 1 km at its widest section, corresponding to a water-surface area of 8.05 km².
185 At the lower end of the valley, a 25-m-high earth-rockfill dam impounds approximately 151 million
186 m³ of water at full storage.

187 The DLD Reservoir is situated within a humid subtropical monsoon zone characterized by
188 marked intra-annual climatic variability. Meteorological records for the previous five years indicate
189 a mean annual air temperature of 22.51 °C and annual rainfall averaging 1,584 mm. Rainfall is



190 concentrated primarily between April and September, defining the wetter part of the year, while
191 comparatively drier conditions prevail during the remaining months, extending from October to
192 the following March. Intense rainfall events are particularly common from June to August and are
193 typically characterized by high rainfall intensity over relatively short periods. Hydrological records
194 from the Shanglin County Water Resources Management Bureau indicate that annual precipitation
195 in 2023 was 1,378.5 mm, of which 1,196.1 mm occurred during the wet season, corresponding to
196 approximately 90% of the annual total.

197 Geomorphologically, the DLD Reservoir is situated within a karst peak-cluster depression
198 characterized by canyon-like terrain that generally descends from northwest to southeast. The
199 reservoir receives groundwater primarily through two subterranean rivers, Xialong and
200 Dalongdong. The surrounding watershed is underlain by dolomite of the Yanguan Formation (C1y),
201 limestone of the Liujiang Formation (D2l), and bedded limestone of the Donggangling Formation
202 (D2d). Extensive karstification has produced a complex landscape consisting of peak clusters, karst
203 valleys, dolines, dry valleys, and interconnected cave systems, resulting in highly fragmented
204 terrain and limited areas of level ground. Structurally, the reservoir is located near the convergence
205 of the Damingshan anticline and Qiaoxian syncline. Its shoreline is consequently steep and
206 irregular, with slopes exceeding 45°, making the reservoir particularly susceptible to hydrological
207 fluctuations and erosion processes.

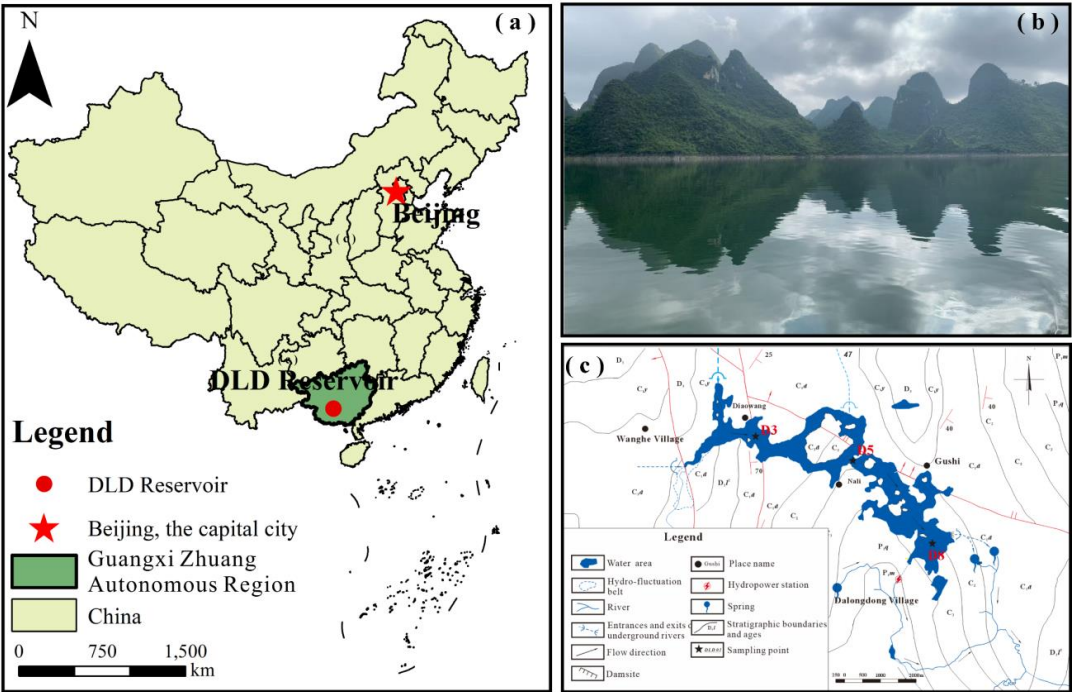


Fig. 1. Overview of the study area and monitoring sites. (a) Location of the DLD Reservoir within China; (b) field photograph of the DLD Reservoir; (c) hydrogeological map of the reservoir area. D3, D5, and D8 denote the monitoring sites positioned in the upper, middle, and lower reaches of the reservoir, respectively.

2.2 Water sample collection and laboratory analysis

2.3.1 Samples collection and pretreatment

Sampling campaigns were carried out during April, August, and November of 2023 across three sections of the DLD Reservoir, with D3, D5, and D8 corresponding to its upper, middle, and lower reaches, respectively (**Fig. 1c**). These sampling campaigns were scheduled to capture variations associated with reservoir operation and seasonal climatic conditions. At each site, a



peristaltic pump was used to obtain water samples along the vertical water-column profile at intervals of 2.5 m. Fieldwork involved three sequential procedures: water collection, on-site measurements, and subsequent sample preparation. In situ physicochemical parameters were determined using a PONSEL Multy 8320 multiparameter water-quality meter (PONSEL), with measurement precisions of ± 0.01 °C for water temperature (WT), ± 0.01 units for pH, and ± 0.01 mg/L for dissolved oxygen (DO). Chlorophyll-a (Chl-a) concentrations were quantified concurrently with an AP-LITE portable fluorometer manufactured by Aquameter (Aquadred, UK); the instrument provides a minimum detection threshold of 0.02 $\mu\text{g/L}$ and a measurement accuracy of $\pm 5\%$. Before each sampling campaign, all sensors were calibrated following the manufacturer's instructions to ensure measurement quality. Specifically, calibration of the pH electrode employed reference solutions with values of 4.00 and 7.00, while air-saturated water served as the calibration medium for the DO sensor. Three replicate measurements were performed at each sampling point to reduce random measurement error, and their mean was used for subsequent analyses. Additional information on water chemistry and its variation is provided in **Text S1 and Fig. S1**.

Following collection, the dissolved fraction was obtained by filtering each water specimen with a 0.45 μm cellulose acetate filter (Millipore, USA), thereby removing suspended particles. Prior to sample processing, both the filtration unit and filters were conditioned sequentially with 100 mL Milli-Q water ($18.2 \text{ M}\Omega\cdot\text{cm}$), followed by 50 mL of the corresponding filtrate to reduce the risk of contamination. Filtrates were then collected in pre-cleaned high-density polyethylene (HDPE) containers. Preparation of these containers involved immersion in 10% HCl for 24 h, five successive washes with Milli-Q water, and three additional rinses using the corresponding filtered



241 water before sample storage. The prepared filtrates were subsequently analyzed for major metal
242 elements, anions, dissolved organic carbon (DOC), and the stable carbon isotope composition of
243 dissolved inorganic carbon ($\delta^{13}\text{C}_{\text{DIC}}$).

244 Preservation and storage protocols were tailored to the specific analytical targets. For major
245 cation determination, aliquots were placed in 15 mL HDPE containers pre-cleaned with HNO_3 ,
246 followed by the addition of 3–5 drops of 7 M high-purity HNO_3 to reduce the pH below 2. For
247 $\delta^{13}\text{C}_{\text{DIC}}$, 60 mL HDPE containers were filled completely, leaving no headspace, to limit isotopic
248 exchange with atmospheric CO_2 . Both DOC and $\delta^{13}\text{C}$ -DIC aliquots received 1–2 drops of saturated
249 HgCl_2 immediately after preparation to inhibit microbial activity. Water intended for chromophoric
250 dissolved organic matter (CDOM) and DOM characterization was stored separately in amber glass
251 containers. Before sampling, these containers underwent three successive washes with potassium
252 dichromate, followed by thorough cleaning with pure water to eliminate residual acid and
253 subsequent air drying. CDOM aliquots were passed through 0.45 μm cellulose acetate filters and
254 subsequently shielded from light to prevent photodegradation. During transportation and storage,
255 all prepared samples were kept refrigerated and protected from light at 4 °C until laboratory
256 measurements were performed.

257

258 **2.3.2 Laboratory analysis and Processing**

259 Cl^- , NO_3^- , and SO_4^{2-} concentrations were determined with a Dionex ICS-900
260 chromatographic system (USA), whereas K^+ , Na^+ , Ca^{2+} , and Mg^{2+} were quantified on an IRIS
261 Intrepid II XSP spectrometer (Thermo Fisher Scientific, USA) based on inductively coupled



262 plasma–optical emission spectrometry (ICP-OES). The cation measurements achieved a precision
263 of 0.01 mg/L. Analytical quality was evaluated by examining the balance between the measured
264 anionic and cationic components, with the relative error maintained below 5%.

265 $\delta^{13}\text{C}_{\text{DIC}}$ were quantified on a MAT253 mass spectrometric platform equipped with a GasBench
266 continuous-flow interface. Following the phosphoric acid procedure, the obtained isotopic
267 signatures were normalized against the Vienna Pee Dee Belemnite (V-PDB) reference and reported
268 with a measurement uncertainty below 0.15‰. Determination of $\delta^{13}\text{C}_{\text{DIC}}$ was undertaken by the
269 Institute of Karst Geology, Chinese Academy of Geological Sciences, through its Karst Geological
270 Resources and Environmental Monitoring and Testing Center affiliated with the Ministry of
271 Natural Resources. Laboratory determinations of major anions and cations were completed at the
272 School of Geography Sciences of Southwest University, whereas DOC concentrations were
273 analyzed by the Third Institute of Oceanography under the Ministry of Natural Resources.

274 TN and TP levels were determined by flow injection analysis with an FIA-6000+ instrument
275 (Jitian Instruments, China). Each determination required a 100 mL aliquot of water without prior
276 filtration, and the analytical procedure achieved a precision of 0.0001 mg/L. These nutrient
277 measurements were undertaken at the School of Geography and Tourism of Chongqing Normal
278 University. For quality assurance, two or three replicates were examined for each specimen;
279 anomalous results were discarded after measurement, and the arithmetic mean of the retained
280 replicates was adopted as the final value for the corresponding parameter.

281 DOM optical properties were characterized by combining three-dimensional fluorescence and
282 UV–visible absorption measurements at the Chongqing Key Laboratory of Carbon Cycling and



283 Carbon Regulation in Mountainous Ecosystems affiliated with Chongqing Normal University. A
284 Hitachi F-7000 spectrofluorometer was employed to acquire excitation–emission matrices (EEMs)
285 for CDOM. During spectral acquisition, excitation (Ex) was scanned over 200–500 nm and
286 emission (Em) over 220–600 nm. The respective sampling increments were set at 5 nm and 1 nm,
287 while identical bandwidths of 5 nm were applied to both Ex and Em channels. Spectra were
288 collected at a scanning speed of 2400 nm/min. Instrument performance was checked against
289 ultrapure water before every measurement.

290 Prior to analysis, all samples were allowed to equilibrate to room temperature and were then
291 placed in a four-window quartz cuvette with a 10 mm optical path length. To minimize cross-
292 contamination, the cuvette was conditioned by rinsing it three times with the corresponding sample
293 before measurement and thoroughly cleaned with ultrapure water three times after each analysis.
294 The acquired EEM spectra were subjected to parallel factor analysis (PARAFAC) using MATLAB
295 2021 to resolve CDOM fluorescent components and quantify their fluorescence intensities and
296 component loadings. The PARAFAC-derived components were further compared with those
297 available in the OpenFluor database for fluorescence component identification (**Table S1, Fig. S2,**
298 **and Fig. S3 in Text S2**). All procedures followed established approaches for CDOM
299 characterization to maintain methodological consistency and facilitate comparison among the
300 resulting data.

301 UV–visible absorbance measurements were performed on an SP-756P spectrophotometric
302 system, with ultrapure water serving as the reference blank. Before each determination, a two-
303 window quartz cuvette with an optical path length of 10 mm was cleaned by three successive rinses



304 with ultrapure water. Absorbance was recorded across the 190–800 nm spectral region with a
305 sampling increment of 1 nm.

306 The absorption coefficient is commonly derived from measured absorbance values to
307 characterize the optical properties of DOM. This conversion is carried out using **Equation (1)**:

$$308 \quad \alpha\lambda = 2.303 A\lambda / L \quad (1)$$

309 $\alpha\lambda$ represents the absorption coefficient at wavelength (λ); L denotes the optical path length of the
310 cuvette (unit: m); $A\lambda$ is the measured absorbance at wavelength λ (dimensionless).

311 The spectral indicators calculated by the three-dimensional fluorescence spectrometer and the
312 ultraviolet spectrophotometer, along with their temporal and spatial variations, are presented in
313 **Text. S3, Fig. S4 and Fig. S5**.

314

315 **2.3.3 Bacterial sequencing and analysis**

316 In prokaryotes, the gene coding for 16S rRNA spans approximately 1,500 bp (**Arnemann,**
317 **2018**) and contains 10 relatively conserved segments separated by nine highly variable regions
318 designated V1–V9. Sequence heterogeneity within these variable regions provides a molecular
319 basis for distinguishing different microbial taxa. Amplicon-based profiling generally involves
320 recovery of genomic DNA from environmental microbial assemblages, PCR amplification
321 targeting selected variable regions (e.g., V3–V4), and high-throughput determination of the
322 resulting amplicons. The sequence profiles obtained through this procedure allow assessment of
323 taxonomic composition, community-level diversity, and relative representation of bacterial taxa,
324 while also supporting inference of their potential functional characteristics in a given environment.



325 Accordingly, this molecular strategy can help clarify associations between bacterial assemblages
326 and surrounding environmental conditions or hosts and provide clues regarding their possible
327 ecological functions (Woo et al., 2008).

328 For bacterial community analysis, amplification targeted the 16S rRNA gene V4 region
329 according to the manufacturer-recommended protocol, employing 515F (5'-
330 GTGYCAGCMGCCGCGGTAA-3') as the forward primer and 806R (5'-
331 GGACTACHVGGGTWTCTAAT-3') as the reverse primer. The amplified products were
332 subsequently sequenced for evaluation of bacterial diversity. Initial processing and quality control
333 of the sequence reads were carried out through the QIIME2 bioinformatic workflow, followed by
334 alignment and comparison of the retained feature sequences. Reads occurring fewer than 20 times
335 across the complete dataset were first excluded, after which operational taxonomic units (OTUs)
336 were delineated from the retained sequences using a sequence identity threshold of 97%. The
337 resulting OTU dataset was subsequently analyzed in RStudio (version 4.4.2) to characterize
338 community composition.

339

340 **2.3.4 Data analysis**

341 Microbial association networks were established from pairwise Spearman correlation
342 coefficients calculated across bacterial OTUs. Before network generation, taxa whose average
343 relative abundance did not reach 0.01% were excluded, and the remaining taxa were subsequently
344 identified manually. Network topology was characterized with the “ggClusterNet” package by
345 calculating the total numbers of nodes and edges, average degree, characteristic path length,



346 maximum network span (diameter), relative modularity, and the average coefficient of clustering.
347 Graphical representations of these associations were produced in Gephi (version 0.10.1). Potential
348 keystone taxa were identified through their positions within the modular network using
349 “ggClusterNet”, based on two node-level parameters: connectivity within a given module (Z_i) and
350 connectivity across different modules (P_i). Beyond these structural descriptors, cohesion metrics
351 were calculated to quantify connectivity at the whole-community level. Positive Cohesion reflected
352 the strength of cooperative associations among community members, whereas Negative Cohesion
353 represented competitive or inhibitory relationships.

354 Associations among DOM components, environmental variables, spectral indices, and
355 bacterial cohesion metrics were evaluated through Spearman correlation analysis and displayed in
356 a correlation heatmap. Mantel tests were additionally applied to examine relationships among the
357 corresponding groups of variables. All correlation analyses were implemented within the R 4.4.3
358 environment, with “corrplot” (version 0.95) employed for graphical presentation. The magnitude
359 and direction of individual correlations were represented by numerical coefficients and color
360 gradients. Significance levels were marked with one, two, or three asterisks, indicating probability
361 thresholds of $p < 0.05$, $p < 0.01$, and $p < 0.001$, respectively.

362 Bayesian linear regression models were further developed to assess how environmental
363 conditions, bacterial cohesion, and DOM spectral properties were related to LDOC and RDOC
364 concentrations. The predictor set comprised environmental parameters, Positive and Negative
365 Cohesion metrics, and spectral indicators associated with autochthonous/allochthonous sources
366 and bacterial activity. Model estimation was performed with “rstanarm”, whereas “bayesplot” was

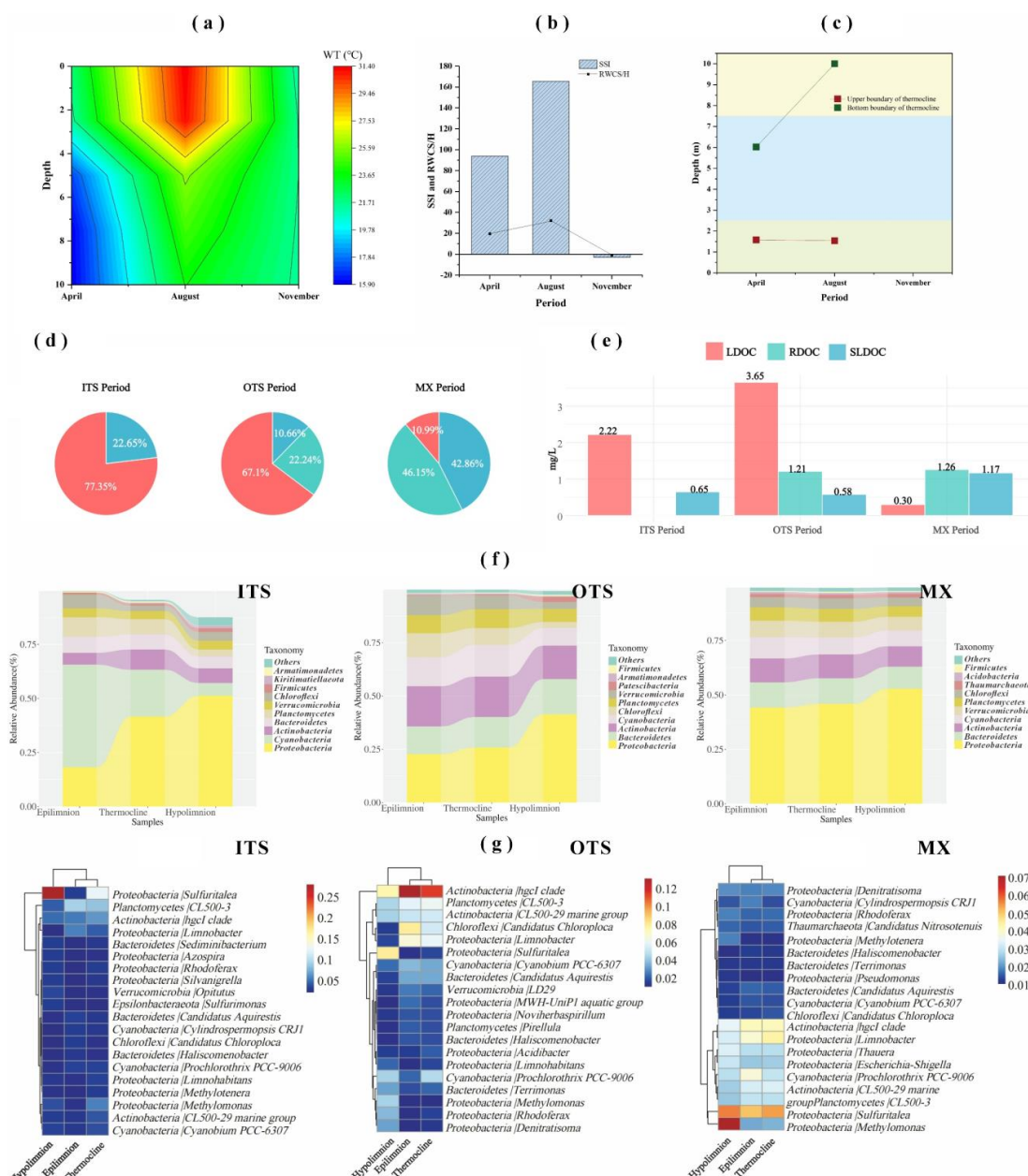


employed to visualize the resulting model outputs. An R^2 prior centered at 0.5 was specified, reflecting an initial assumption that the predictors collectively accounted for roughly half of the observed variability. Posterior regression coefficients were then graphically summarized to compare the direction and magnitude of the associations between individual predictors and LDOC or RDOC concentrations.

3. Results

3.1 Thermal stratification of the DLD Reservoir

Research data indicate that the DLD Reservoir is a warm monomictic reservoir (Hasan et al., 2013), exhibiting distinct seasonal variations in its thermal structure (Jia et al., 2024; Li et al., 2021). Based on water temperature profiles and stratification indices (Schmidt Stability Index (SSI) and RWCS/H (Fig. 2a and b, Text S4), the DLD Reservoir was divided into three periods: the incubating thermal stratification (ITS) period in April, with mild stratification; and the obvious thermal stratification (OTS) period in August, featuring stable layering; and the mixing (MX) period in November (Zhang et al., 2023; Liu et al., 2020). The thermocline was absent in November, while it spanned 1.58–6.02 m in April and 1.53–10 m in August (Fig. 2c). Based on these boundaries and sampling at 2.5 m intervals (Li et al., 2021; Jia et al., 2024), the water column was categorized as: epilimnion/surface (0–2.5 m), thermocline/middle (2.5–7.5 m), and hypolimnion/bottom (≥ 7.5 m).



386

387 **Fig.2 Vertical water temperature profile in the DLD Reservoir during the study period (a),**

388 **temporal variations of SSI and RWCS/H index(negative values of RWCS/H were observed**

389 **during the late mixing period, indicating a temporary density inversion that facilitates rapid**



vertical convective mixing) (b), depths of the upper and lower boundaries of the thermocline (c); the proportions (d) and concentrations (e) of each component of DOM in the water bodies during ITS, OTS, and MX periods; and the top 10 (f) and top 20 (g) bacterial genera and species levels in the water body of the DLD reservoir during the ITS, OTS and MX periods, respectively.

3.2 DOM component characteristics

Parallel factor analysis distinguished five DOC categories across the water-column profiles during the ITS, OTS, and MX phases of the DLD Reservoir. Overall, the resolved fractions comprised three LDOC components, one SLDOC component, and one RDOC component (classification criteria are provided in Text S1). Under ITS conditions, PARAFAC resolved C1, C2, and C3, which were assigned to the LDOC and SLDOC fractions. With the development of OTS, the component profile expanded to C1, C3, C4, and C5, encompassing all three fractions of LDOC, SLDOC, and RDOC. Following transition to the MX phase, C1, C2, and C4 were detected and corresponded to SLDOC, LDOC, and RDOC, respectively (Text S2).

Research has shown that DOC constitutes a substantial proportion of DOM, often accounting for approximately 50% or more of its composition (Moody and Worrall, 2017). Therefore, in this study, DOC concentration was used as a proxy for DOM concentration. Based on the measured DOC concentrations (mg/L) at each sampling site in the DLD Reservoir and the fluorescence intensity contribution rates (%) of various DOM components, the concentrations of RDOC and LDOC (mg/L) at each sampling site were calculated using the equation: $\text{RDOC/LDOC (\%)} \times \text{DOC}$



411 $(\text{mg/L}) = \text{RDOC/LDOC} (\text{mg/L})$.

412 Based on the fluorescence component contribution ratios and DOC concentration calculations,
413 both Thermal stratification (ITS and OTS) periods exhibited higher fluorescence intensity
414 contributions and concentrations of LDOC compared to the MX period (**Fig.2 (d) and (e)**). During
415 the ITS period, LDOC and SLDOC accounted for approximately 76.9% and 23.1% of the total
416 fluorescence intensity, with corresponding concentrations of 2.22 mg/L and 0.65 mg/L, respectively.
417 In the OTS period, LDOC, SLDOC, and RDOC contributed 64.7%, 12.3%, and 23.0% of the total
418 fluorescence intensity, with concentrations of 3.65 mg/L, 0.58 mg/L, and 1.21 mg/L, respectively.
419 During the MX period, the fluorescence intensity contributions of LDOC, SLDOC, and RDOC in
420 the DLD Reservoir were 11.2%, 42.6%, and 46.2%, respectively. Based on the concentration
421 calculation formula, the average concentrations of these DOC categories were 0.30 mg/L, 1.17
422 mg/L, and 1.26 mg/L, respectively. Spatial variations in DOM component concentrations across
423 the different periods are provided in **Text S2 and Fig. S3**.

424

425 **3.3 Bacterial Community Structure and Interactions**

426 **3.3.1 Spatial and Temporal Distribution Pattern of Bacteria**

427 Amplicon sequencing of samples collected during the three thermal phases yielded 411,458
428 high-quality bacterial OTUs after quality control. Taxonomic assignment indicated that the
429 bacterial assemblages primarily comprised *Proteobacteria*, *Cyanobacteria*, *Bacteroidetes*, and
430 *Actinobacteria*. Together, these four most abundant phyla contributed approximately 50%-70% of
431 the overall community. Clear temporal shifts in their relative contributions were observed among



the three phases (**Fig. 2f**). Under ITS conditions, *Proteobacteria* contributed the largest fraction (38% ± 17%), followed by *Cyanobacteria* (24% ± 19%) and *Actinobacteria* (8% ± 3%). With the establishment of OTS, the relative contribution of *Proteobacteria* decreased to 31% ± 9%, whereas *Actinobacteria* and *Bacteroidetes* accounted for 18% ± 2% and 15% ± 4%, respectively; *Cyanobacteria* represented a comparatively smaller fraction of 12% ± 4%. Following the transition to MX conditions, *Proteobacteria* increased to 48% ± 8% of the bacterial assemblage, while *Bacteroidetes* and *Actinobacteria* contributed 11% ± 2% and 10% ± 2%, respectively. By comparison, the contribution of *Cyanobacteria* declined further to 8% ± 2%.

Analysis of genus-level bacterial composition revealed clear spatiotemporal heterogeneity in community distribution. During ITS, however, the proportional representation of individual bacterial genera varied only slightly among water layers. The most abundant genus in the hypolimnion was *Sulfuritalea* (*Proteobacteria*) (29% ± 7%), which also dominated the thermocline (12% ± 11%). In the epilimnion, the genus *CL500-3* (*Planctomycetes*) exhibited the highest relative abundance (8% ± 1%) (**Fig.2 (g)**). In the OTS period, greater variation in bacterial relative abundance was observed among water layers. The *hgcI* clade, affiliated with *Actinobacteria*, showed greater representation in the upper two layers, accounting for 12.9% ± 2.4% in the epilimnion and 10.4% ± 2.5% in the thermocline. In the hypolimnion, *Sulfuritalea* (*Proteobacteria*) remained the most dominant genus (7.4% ± 4.8%) (**Fig.2 (g)**). During the MX period, *Methylomonas* (*Proteobacteria*) reached its highest relative abundance in the hypolimnion (7.9% ± 0.9%), while its distribution was more uniform in the middle and surface layers. *Sulfuritalea* (*Proteobacteria*) showed the second-highest overall abundance (6.2% ± 1.0%) and maintained



453 relatively even distribution across all three water layers (**Fig.2 (g)**).

454

455 **3.3.2 Distribution Characteristics of Keystone Taxa Regulate DOM Bioavailability**

456 The ecological positions of bacterial OTUs within the reservoir association networks were
 457 evaluated across the three thermal phases using two node-level parameters: connectivity within
 458 individual modules (Z_i) and connectivity across modules (P_i). According to these metrics, all nodes
 459 were assigned to one of four topological categories following previously established criteria
 460 (**Guimerà and Nunes Amaral, 2005; Ma et al., 2021**) (**Fig.3 (a)-(c)**). Network hubs occurred
 461 exclusively during OTS period, with two such nodes identified in this phase. Because Connectors,
 462 Module hubs, and Network hubs represent highly connected positions that contribute substantially
 463 to the structural integrity of microbial networks, taxa belonging to these three categories were
 464 regarded as keystone members. Across ITS, OTS, and MX, the corresponding networks contained
 465 1, 4, and 8 Module hubs and 22, 10, and 3 Connectors, respectively, whereas Peripheral nodes
 466 numbered 329, 533, and 421 (**Table 1**). Further information on the taxonomic composition and
 467 abundance of keystone members identified in the DLD Reservoir during each thermal period is
 468 provided in **Text S5** and **Table 1**.

Fig.3 Node classification for identifying putative keystone taxa in the networks of the (a) ITS period, (b) OTS period, and (c) MX period. Each symbol represents an OTU derived from detailed modular analysis of the two networks. Categories include: Module hubs (nodes with high connectivity within their own module, defined as $Z_i > 2.5$ and $P_i < 0.62$), Connectors (nodes with high connectivity between different modules, defined as $Z_i < 2.5$ and $P_i > 0.62$),



475 **Network hubs (nodes with high connectivity across the entire network, defined as $Z_i > 2.5$**
476 **and $P_i > 0.62$), and Peripherals (nodes with low connectivity both within and between**
477 **modules, defined as $Z_i < 2.5$ and $P_i < 0.62$). Bacterial names in the figure correspond to the**
478 **genus-level taxonomy represented by each OTU node. Heatmap showing the Spearman**
479 **correlation between the relative abundance of bacterial genera in the key bacterial groups of**
480 **the water bodies and the concentrations of LDOC, SLDOC and RDOC during the thermal**
481 **stratification period (d), thermocline (e), bottom water layer (f), and mixed period (g).Linear**
482 **and non-linear regression analyses of positive ((h) and (j)) and negative ((i) and (k)) cohesion**
483 **with LDOC and RDOC of bacteria in DLD reservoir.**

484

485 **Table 1. Detailed phylogenetic relationships of Keystone taxa in the Zi-Pi diagram**

Networks	Node type	Number of OTUs	Phylum	Genus
Incubating thermal stratification period	Module hubs	1	<i>Proteobacteria</i> (1)	<i>Acidibacter</i> (1)
			<i>Proteobacteria</i> (7)	<i>Paludibacter</i> (1),
			<i>Bacteroidetes</i> (4)	<i>Unclassified</i> (5),
			<i>Planctomycetes</i> (3)	<i>Fluviicola</i> (2),
			<i>Firmicutes</i> (2)	<i>Flavobacterium</i> (1),
			<i>Cyanobacteria</i> (1)	<i>Limnothrix</i> (1),
	Connectors	22	<i>Verrucomicrobia</i> (1)	<i>Pirellula</i> (1), <i>CL 500-</i>
			<i>Hydrogenedentes</i>	3 (2), <i>LD 29</i> (1),
			(1)	<i>Longilinea</i> (1),
			<i>Chloroflexi</i> (1)	<i>Mycobacterium</i> (1),
			<i>Actinobacteria</i> (1)	<i>Sulfurimonas</i> (1),
			<i>Epsilonbacteraeota</i>	<i>Sterolibacterium</i> (1),



Obvious thermal stratification period	Connectors	10	(1)	<i>Denitratisoma</i> (1), <i>Candidatus Nitrotoga</i> (1) <i>Rubrivivax</i> (1), <i>MWH-UniPI aquatic</i> <i>group</i> (1)
			<i>Actinobacteria</i> (4)	<i>CL 500-29 marine</i>
			<i>Proteobacteria</i> (2)	<i>group</i> (4),
			<i>Cyanobacteria</i> (1)	<i>Cyanobium PCC-</i>
			<i>Bacteroidetes</i> (1)	<i>6307</i> (1), <i>Brevifollis</i>
			<i>Verrucomicrobia</i> (1)	(1), <i>Limnobacter</i> (1),
			<i>Planctomycetes</i> (1)	<i>Unclassified</i> (3)
				<i>Cyanobium PCC-</i>
	Module hubs	4	<i>Cyanobacteria</i> (2)	<i>6307</i> (1), <i>Nodosilinea</i>
			<i>Proteobacteria</i> (2)	(1), <i>Sphaero tilus</i> (1),
				<i>Unclassified</i> (1)
	Network hubs	2	<i>Actinobacteria</i> (1)	<i>hgcI clade</i> (1)
			<i>Bacteroidetes</i> (1)	<i>Terrimonas</i> (1)
			<i>Proteobacteria</i> (2)	<i>Paucibacter</i> (1),
			<i>Dependentiae</i> (1)	<i>Unclassified</i> (2)
Mixing period	Connectors	3		<i>Candidatus</i>
			<i>Proteobacteria</i> (2)	<i>Methylopumilus</i> (1),
			<i>Bacteroidetes</i> (3)	<i>Limnobacter</i> (1),
			<i>Verrucomicrobia</i> (2)	<i>Chthonibacter</i> (1),
	Module hubs	8	<i>Actinobacteria</i> (1)	<i>Unclassified</i> (5)

486 Spearman coefficients were calculated to examine associations of genus-level bacterial
 487 abundances with LDOC, SLDOC, and RDOC concentrations among water layers under stratified
 488 conditions (TS, comprising ITS and OTS) and during MX in the DLD Reservoir. Because vertical
 489 thermal stratification disappeared under MX conditions, the entire water column was considered
 490 without further division into individual layers. The relationships linking keystone bacterial taxa to



491 RDOC and LDOC at different depths are presented in **Fig. 3(d-g)**.

492 Clear depth-dependent associations with RDOC emerged during OTS. Within the epilimnion,
493 higher abundances of LD29, MWH-UniP1 aquatic group, *Acidibacter*, *Nodosilinea* PCC 7104,
494 *Limnobacter*, and *hgcI* clade coincided significantly with elevated RDOC concentrations ($p < 0.05$).
495 Even stronger positive associations were detected for *Terrimonas*, *Cyanobium* PCC 6307, and
496 *CL500-29* marine group ($p < 0.01$). In deeper hypolimnetic waters, RDOC formation was
497 associated only with LD29, *Flavobacterium*, *Candidatus Nitrotoga*, *Pirellula*, *Brevifollis*, and
498 *Nodosilinea*. After transition to MX conditions, *Limnobacter* was the sole genus exhibiting a
499 positive relationship with RDOC ($p < 0.05$).

500 Associations involving LDOC differed markedly among vertical zones under stratified
501 conditions. *Brevifollis* was the only keystone genus related to LDOC within the epilimnion. At
502 thermocline depths, increasing abundances of *hgcI* clade and *Fluviicola* corresponded to
503 significant decreases in LDOC, whereas *Pirellula* displayed the opposite pattern, with a significant
504 positive association. No keystone taxon exhibited a statistically detectable relationship with LDOC
505 either in the hypolimnion under TS conditions or throughout the unstratified water column during
506 MX.

507 Relationships involving SLDOC were restricted to the epilimnion under thermal stratification.
508 Greater representation of *CL500-3* and *Limnothrix* was associated with higher SLDOC
509 concentrations ($p < 0.05$), whereas LD29, *CL500-29* marine group, *hgcI* clade, and *Terrimonas*
510 displayed inverse associations with SLDOC ($p < 0.05$). Taken together, the spatial and temporal
511 patterns of these Spearman relationships suggest that keystone taxa are more closely associated



512 with processes governing RDOC production and persistence than with those responsible for LDOC
513 and SLDOC generation and degradation in the aquatic environment.

514

515 **3.3.3 Driving Effects of Bacterial Community Cohesion on DOM Bioavailability**

516 Positive cohesion represents facilitative microbial interactions, while negative cohesion
517 reflects competitive relationships (Yuan et al., 2021; Herren and McMahon, 2017). The spatio-
518 temporal variations in the average values of positive and negative bacterial cohesive values in the
519 DLD reservoir are calculated as shown in **Table S2** and **Text S6**.

520 A regression analysis was conducted between the overall RDOC and LDOC concentrations in
521 the water column of DLD Reservoir and the corresponding positive and negative cohesion values
522 at each sampling point (**Fig.3 (h)-(k)**). As shown in **Fig. 3(h)**, a non-linear correlation was observed
523 between bacterial positive cohesion and RDOC concentration ($p < 0.05$), with the scatter points
524 forming a U-shaped pattern. On the left side of the U-shape, all sampling points within the 95%
525 confidence interval were from the MX period, indicating that RDOC concentration decreased non-
526 linearly with increasing positive cohesion during this period. On the right side of the U-shape, most
527 sampling points from the OTS period fell within the 95% confidence interval, suggesting that
528 RDOC levels increased with rising positive cohesion during this phase. In contrast to the linear
529 relationships observed in **Fig. 3(i)-(k)**, the correlation between positive cohesion and RDOC (**Fig.**
530 **3(h)**) was best characterized by a quadratic (U-shaped) model. This non-linearity reflects a
531 functional shift in microbial facilitative interactions between hydrological periods. In the MX
532 period, the negative correlation suggests that increased bacterial cooperation may be directed



533 toward carbon consumption under low-temperature, low-productivity conditions. Conversely, in
534 the OTS period, positive cohesion appears to promote RDOC production via the MCP effect. This
535 bimodal response distinguishes RDOC accumulation from the consistent linear trends observed for
536 LDOC (**Fig. 3(j), (k)**).

537 By comparison, positive cohesion increased linearly with LDOC concentration ($r = 0.69$, $p <$
538 0.01) (**Fig. 3(j)**). A similar increasing trend was observed between bacterial negative cohesion and
539 RDOC concentration ($r = 0.49$, $p < 0.01$) (**Fig. 3(i)**), while the relationship between negative
540 cohesion and LDOC was inverse ($r = -0.63$, $p < 0.01$) (**Fig. 3(k)**). Collectively, these patterns
541 suggest that facilitative bacterial associations in the DLD Reservoir favored LDOC formation, with
542 greater positive cohesion corresponding to enhanced LDOC production. In contrast, competitive
543 and inhibitory associations appeared to constrain LDOC formation and potentially accelerate its
544 utilization, while simultaneously favoring RDOC production and retention.

545

546 **3.3.4 Regulation of DOM Bioavailability by Environmental Factors in Stratified Water**

547 **Columns**

548 Across the TS phases, encompassing both ITS and OTS, significant associations between
549 environmental variables and DOM components were detected in the epilimnion of the DLD
550 Reservoir. SLDOC concentrations increased significantly with DO, TN, and FI ($p < 0.05$), whereas
551 an inverse relationship was detected between positive cohesion and SLDOC ($p < 0.05$) (**Fig. 4(a)**).
552 LDOC was positively associated with pH but varied inversely with $p\text{CO}_2$ ($p < 0.05$). For RDOC,
553 higher concentrations coincided significantly with increases in TP and the Simpson index ($p < 0.05$),



554 while HCO_3^- showed the opposite relationship with RDOC levels ($p < 0.05$).

555 Within the thermocline (**Fig. 4(b)**), LDOC increased significantly with both WT and FI ($p <$
556 0.05). For RDOC, higher levels were significantly associated with WT, the Simpson index, and
557 HIX, while an inverse association was detected with FI ($p < 0.05$).

558 Within the hypolimnion (**Fig. 4(c)**), LDOC was significantly associated with fewer
559 environmental parameters, with positive cohesion being the only variable showing a positive
560 relationship ($p < 0.05$). SLDOC likewise increased significantly with positive cohesion ($p < 0.05$).
561 At the same depth, higher RDOC concentrations were significantly associated with elevated HCO_3^- ,
562 WT, TP, and TN ($p < 0.05$).

563 Under MX conditions (**Fig. 4(d)**), LDOC varied inversely with the Simpson index ($p < 0.05$).
564 Higher HCO_3^- , pCO_2 , and HIX values were significantly associated with lower concentrations of
565 both SLDOC and RDOC ($p < 0.05$), whereas increasing pH corresponded significantly to higher
566 levels of these two DOM fractions ($p < 0.05$). For RDOC specifically, its concentration increased
567 significantly in association with DO, Chl-a, $\delta^{13}\text{C-DIC}$, and the $\beta:\alpha$ ratio ($p < 0.05$).

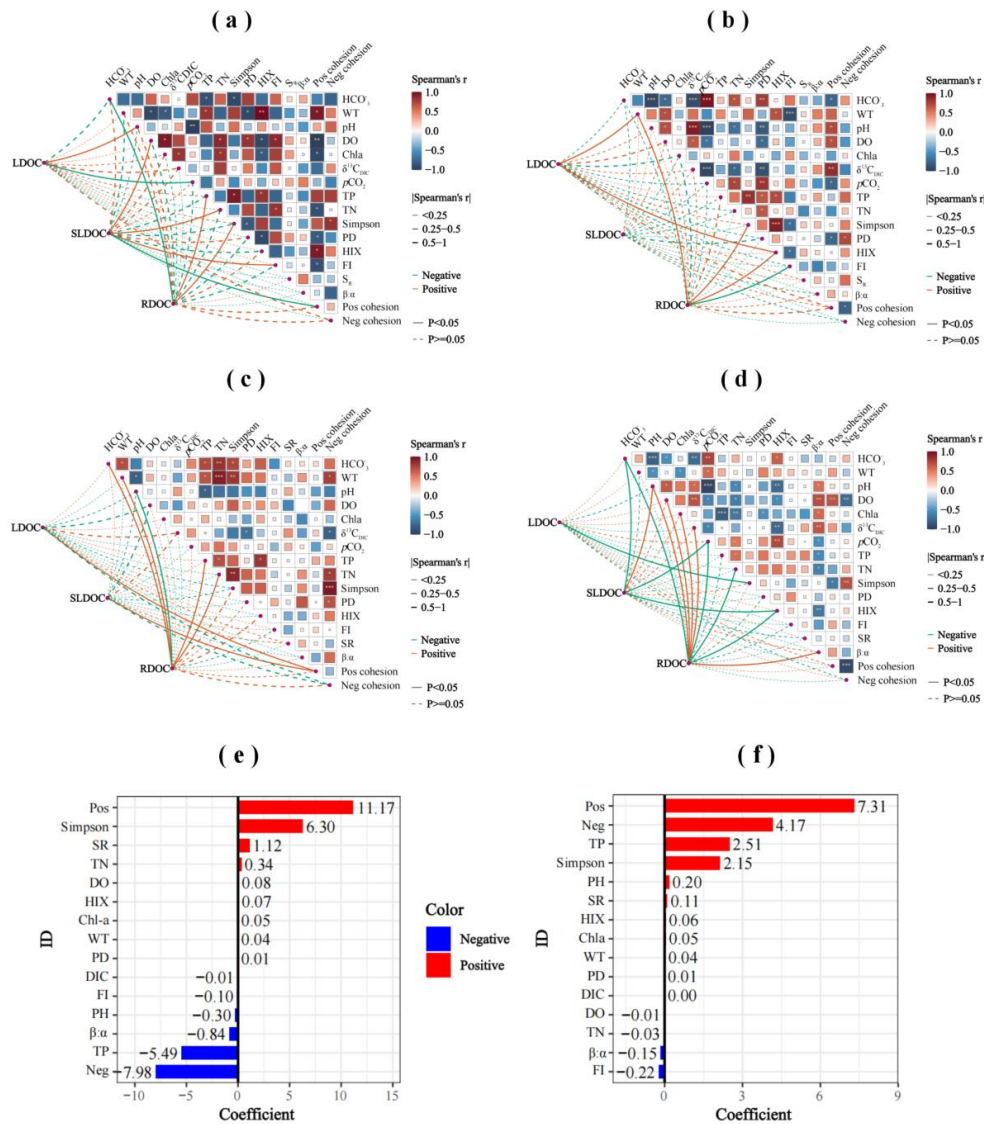


Fig.4 (a-d) Heatmaps of Spearman correlation coefficients from Mantel tests illustrating relationships among DOM, bacterial positive/negative cohesion, hydrochemical parameters, and spectral indices in DLD Reservoir: (a) epilimnion during TS periods, (b) thermocline during TS periods, (c) hypolimnion during TS periods, and (d) MX period (*p < 0.05, **p < 0.01, *p < 0.001). (e-f) Bayesian linear regression analyses of environmental and spectral**



574 **indicators with (e) LDOC and (f) RDOC concentrations (red indicates positive coefficients,**
575 **blue indicates negative coefficients; R^2 values for the models are 0.530 and 0.655, respectively).**

576 Furthermore, as shown in **Fig. 4**, bacterial cohesion exhibited spatiotemporal correlations with
577 environmental factors. In the epilimnion of the TS periods, positive cohesion correlated positively
578 with WT and HIX, but negatively with DO, Chl-a, TN, PD, and FI. In the thermocline, it was
579 positively linked to pH, DO, and $\delta^{13}\text{C}_{\text{DIC}}$, and negatively to HCO_3^- , $p\text{CO}_2$, and PD. In the
580 hypolimnion, only negative cohesion showed significant correlations. During the MX period, DO
581 strongly promoted positive cohesion but suppressed negative cohesion, while the Simpson index
582 showed the opposite pattern.

583 Bayesian linear regression revealed distinct predictive effects on LDOC and RDOC (**Fig.4 (e)**
584 **and (f)**). For LDOC, positive cohesion and the Simpson index were promotive predictors, while
585 negative cohesion and TP were inhibitory. Bacterial activities and alkalinity (represented by alpha
586 diversity and pH in **Fig.4 (e)**) exhibited inhibitory predictive effects on LDOC concentration—that
587 is, higher values of these indicators predicted lower LDOC levels. In contrast, Chl-a showed a
588 slight promotive predictive effect on LDOC, with higher values predicting increased LDOC
589 concentration. For RDOC, positive cohesion, negative cohesion, and TP consistently exhibited
590 promotive effects, while FI and the $\beta:\alpha$ ratio showed slight inhibition. Overall, cohesion metrics,
591 nutrient status, and microbial alpha diversity dominated DOM bioavailability.

592

593 **4. Discussion**

594 **4.1 Thermal Stratification Drives Spatiotemporal Differentiation in Microbial Interaction**



595 **Networks and RDOC Production**

596 Thermal stratification provides the primary physical framework underlying temporal and
597 vertical variations in reservoir microbial associations and RDOC formation. Suppression of water-
598 column mixing during stratification generates pronounced vertical hydrochemical heterogeneity,
599 with oxygen-rich conditions in surface waters and oxygen depletion at depth, which in turn
600 promotes depth-specific organization of microbial assemblages (Li et al., 2021; Jia et al., 2025).
601 Within this stratified environment, keystone taxa assume central ecological roles by shaping the
602 composition and functional attributes of microbial assemblages (Banerjee et al., 2018) and
603 contributing directly to RDOC formation across different depths (Shi et al., 2025). Evidence from
604 network and correlation analyses (Fig. S6) further demonstrates close associations between
605 keystone-taxon abundance and key hydrochemical variables, including water temperature, DO, and
606 pH. Such relationships suggest that environmental gradients may regulate spatial patterns of RDOC
607 formation through their effects on these ecologically influential taxa. These keystone taxa occupy
608 central positions within microbial interaction networks and engage in multiple interaction pathways
609 with both undefined genera and other keystone taxa (Fig. 5), further supporting their role in shaping
610 the spatiotemporal patterns of RDOC production through complex microbial interactions. The
611 manifestation of these complex interactions is most evident in the functional differentiation
612 characteristics of microorganisms across water layers, where distinct functional segregation is
613 evident (Fig. 6).

614 In the epilimnion (Fig. 6(a)): *Limnothrix* is extensively involved in aerobic carbon
615 transformation; *Flavobacterium* shows a positive correlation with photosynthetic functions, while



616 *Mycobacterium* exhibits a negative correlation; *Brevifollis* is negatively correlated with functions
617 such as fermentation and methane oxidation, suggesting it may play an inhibitory role in the carbon
618 cycle. *CL500-3* and *Fluviicola* exhibit opposite correlations in xylan and chitin degradation,
619 reflecting niche differentiation in complex organic matter degradation. In the thermocline (**Fig.**
620 **6(b)**), chloroplast-related functions, chitin degradation, and anaerobic photoautotrophy show
621 significant correlations with numerous bacterial groups, demonstrating strong functional coupling
622 between groups. In the hypolimnion (**Fig. 6(c)**), photosynthesis-related functions showed
623 significant positive correlations with *Terrimonas* and *Acidibacter*; xylan degradation was
624 negatively correlated with multiple bacterial groups, while cellulose and chitin degradation were
625 positively correlated with specific groups. In methane oxidation, *Fluviicola* showed negative
626 correlations with oxidative functions, whereas *Brevifollis* and *Flavobacterium* often exhibited
627 positive correlations. In methane production, hydrogenotrophic methanogenesis showed
628 significant positive correlations with *Bacillus longus* and *Limnobacter*. During the Mixing period
629 (**Fig. 6(d)**), *Candidatus Methylopumilus* and *Chthoniobacter* correlated positively with carbon
630 degradation and methane oxidation, while *Limnobacter* correlated negatively with nearly all
631 metabolic functions, suggesting that its increased abundance may be accompanied by decreased
632 functional activity.

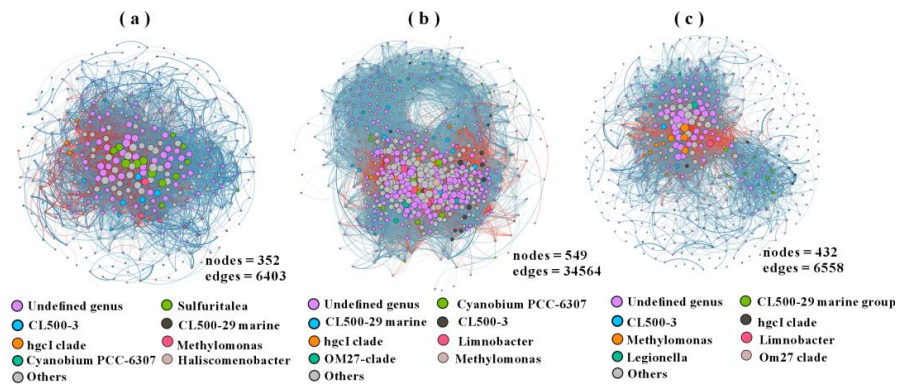
633 This overarching functional segregation provides the ecological template upon which carbon
634 transformation functions are dominated by distinct keystone taxa across different periods and layers.
635 During the ITS period, *Fluviicola*, acting as a Connector in the co-occurrence network, indirectly
636 influenced RDOC formation through heterotrophic metabolism and interspecies interactions.



637 Studies indicate that *Fluviicola* is ubiquitously distributed in freshwater environments, where its
638 heterotrophic metabolic activity may indirectly modulate aquatic carbon cycling. Moreover,
639 synergistic interaction with *Fluviicola* and *Polynucleobacter* are implicated in the RDOC
640 formation and persistence of RDOC (Shi et al., 2025). In the OTS period, the Cyanobacteria
641 *Nodosilinea* PCC-7104 and *Cyanobium* PCC-6307 functioned as Module hubs, fixing CO₂ through
642 photosynthesis and serving as primary producers of LDOC. These two Cyanobacterial taxa utilize
643 photosynthesis and carbon concentration mechanisms (CCMs) to achieve carbon fixation. CCMs
644 enable them to efficiently concentrate CO₂ under CO₂-limited conditions, thereby sustaining
645 efficient carbon fixation catalyzed by ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco)
646 (Kupriyanova et al., 2023). Their carbon fixation capacity is regulated by light intensity, as light
647 energy drives the photosynthetic electron transport chain, influencing the production of ATP and
648 NADPH, which in turn modulates carbon fixation efficiency (Tu et al., 2023). These two bacterial
649 taxa contribute substantially to carbon fixation within surface waters under thermally stratified
650 conditions. Furthermore, in this period (Fig. 5 and 6), *hgcI* clade (Network hubs) and *CL500-29*
651 *marine group* (Connectors) not only connected different modules and nodes in the bacterial
652 interaction network but also contributed to the transformation of both terrestrially derived and
653 autochthonously produced LDOC, leveraging their carbon metabolic capabilities. The *CL500-29*
654 *marine group* is recognized as a core species in warm lakes (Guo et al., 2023; Gómez-Consarnau
655 et al., 2012), capable of degrading various carbon- and nitrogen-containing substrates (Zhang et
656 al., 2020) and showing a positive correlation with rising water temperatures (Guo et al., 2023),
657 which aligns with our findings (Fig. S6). The *hgcI* clade represents a prevalent bacterial lineage



658 across diverse freshwater ecosystems (**Newton et al., 2011**). Members of this facultatively aerobic
659 lineage are capable of light-supported metabolism (**Garcia et al., 2013; Ghylin et al., 2014**) and
660 exhibit considerable capacity for the utilization of carbon compounds (**Ghylin et al., 2014**).
661 Meanwhile, *Terrimonas*, another Network hub during the OTS period, is closely associated with
662 denitrification processes (**Yan et al., 2022**) and may indirectly regulate carbon metabolism by
663 influencing the nitrogen cycle. During the MX period, when the water column of the DLD
664 Reservoir was fully mixed, *Limnobacter* emerged as the sole Module hub in the co-occurrence
665 network, and its relative abundance increased significantly with RDOC levels (**Fig. 3(d)-(g)**). This
666 bacterium is affiliated with β -Proteobacteria and represents a typical thiosulfate oxidizer
667 (**Xamxidin et al., 2022**). Energy production can be sustained through sulfide oxidation, a process
668 that facilitates electron-acceptor utilization by anaerobic microorganisms (**Bruser et al., 2000**) and
669 provides favorable conditions for anaerobic phototrophs that exploit reducing equivalents derived
670 from sulfide to support CO₂ fixation and anoxygenic phototrophic growth (**Bryant et al., 2011**). In
671 addition, its chemoautotrophic metabolism enables energy acquisition through oxidative
672 transformation of reduced inorganic sulfur species, with oxidized nitrogen compounds serving as
673 electron acceptors. This metabolic pathway establishes a biogeochemical linkage among sulfur
674 oxidation, carbon turnover, and nitrogen transformation (**Hu et al., 2018**). This explains why,
675 during the MX period when DIC concentrations were the highest, *Limnobacter*—as the only
676 keystone taxa significantly positively correlated with RDOC concentration (**Fig. 3(g)**) — played a
677 critical role in RDOC production by directly utilizing inorganic carbon sources and fixing CO₂ to
678 synthesize organic carbon.



679

680 **Fig. 5. Genus-level bacterial association networks in the DLD Reservoir under (a) ITS, (b)**

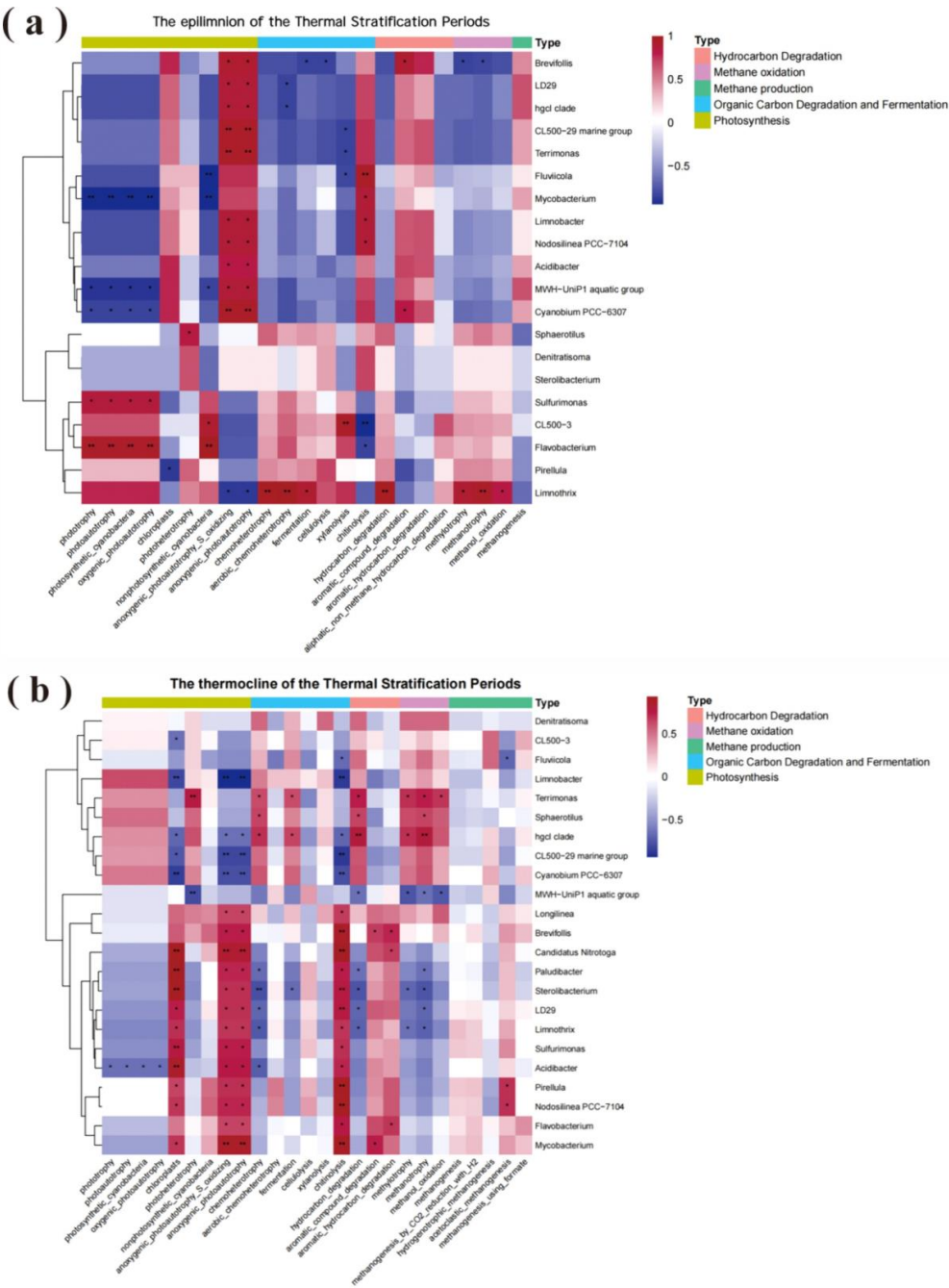
681 **OTS, and (c) MX conditions. Red and blue links denote negative and positive associations,**

682 **respectively. “Nodes” refers to the total number of taxa included in each network, whereas**

683 **“Edges” denotes the total number of connections. Node size reflects the relative**

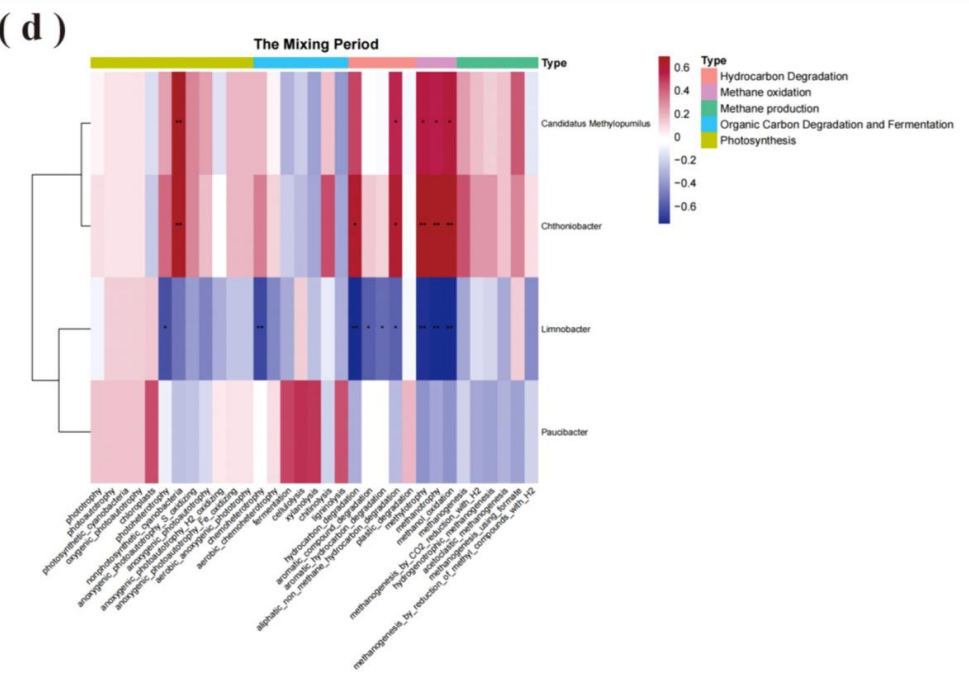
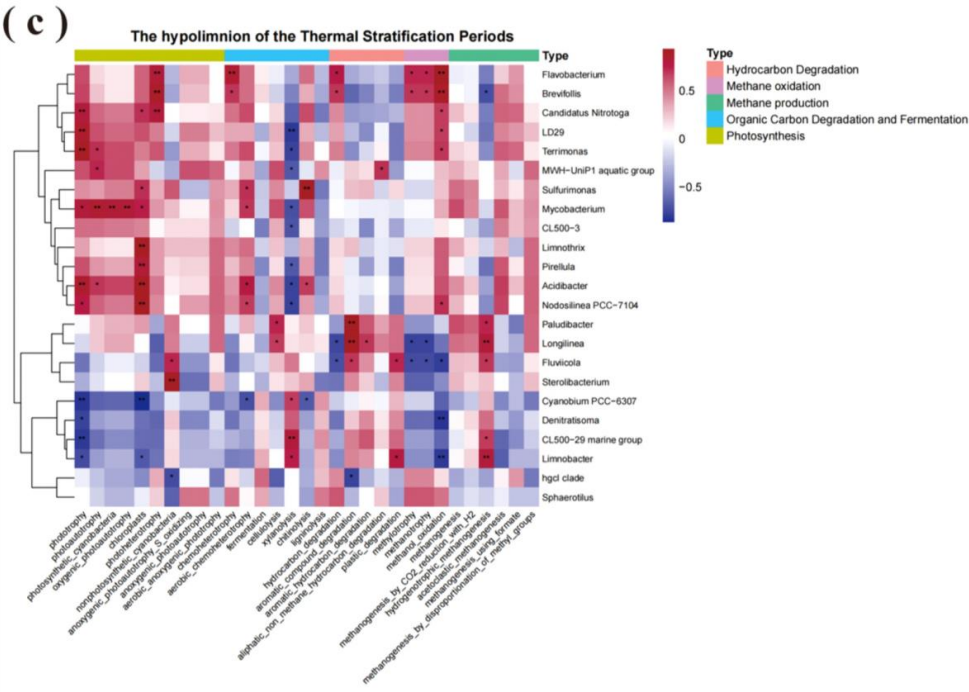
684 **representation of each bacterial genus within the community.**

685



686

687





689 **Fig. 6 Spearman correlation analysis of relative abundance and functional prediction of key**
690 **taxa during the thermal stratification periods in the epilimnion (a), thermocline (b),**
691 **hypolimnion (c), and Mixing period (d).**

692

693 **4.2 Bacterial Cohesion Reflects the MCP Mechanism**

694 The succession of thermal stratification drives shifts in key environmental parameter. These
695 environmental changes subsequently determine the balance between positive and negative bacterial
696 cohesion across different water layers. Ultimately, this shift in microbial interaction patterns
697 regulates the efficiency of the MCP and the resulting production of RDOC.

698 During ITS, enhanced light availability together with elevated water temperature favors
699 phytoplankton proliferation, leading to substantial inputs of readily bioavailable DOM, within
700 which carbohydrates and amino acids occur in relatively high proportions (**Meon and Kirchman,**
701 **2001; Amon et al., 2001**). The resulting organic matter supplies heterotrophic bacteria with readily
702 accessible carbon resources (**Wen et al., 2022**), creating favorable conditions for facilitative
703 associations among bacterial taxa and consequently enhancing microbial carbon pump (MCP)
704 activity. Within these microbial networks, certain bacterial taxa may facilitate the proliferation of
705 neighboring microorganisms or aquatic vegetation by producing various bioactive substances, such
706 as B₁₂ and other vitamins, extracellular enzymes, and compounds that promote growth (**Hu et al.,**
707 **2025**). Such biological exchanges can enhance LDOC supply while simultaneously reinforcing
708 positive interspecific associations, thereby providing additional support for MCP-mediated carbon
709 transformation. Consequently, the rise in positive cohesion during this period is accompanied by



710 an increase in RDOC concentration (**Fig.3(h)**). Studies have shown that algal–bacterial interactions
711 further enhance RDOC production: algae provide dimethylsulfoniopropionate (DMSP) as a carbon
712 source for bacteria, while bacteria reciprocate with vitamin B₁₂, enhancing algal photosynthetic
713 carbon fixation and LDOC secretion. Through the MCP effect, approximately 20% of the biomass
714 can be transformed into persistent RDOC, and horizontal gene transfer of sulfur metabolism genes
715 also contributes to stabilizing algal-derived organic carbon (**Hu et al., 2025**). As a result, LDOC
716 concentration exhibits a linear increase with rising positive cohesion (**Fig.3(j)**). However, in
717 environments with high bacterial diversity and intensified resource competition, the struggle for
718 oxygen and organic matter leads to increased LDOC consumption. Under these conditions, LDOC
719 decreases with increasing negative cohesion, while RDOC increases (**Fig.3(i) and (k)**), indicating
720 that competitive negative cohesion also contributes to MCP-mediated carbon accumulation.

721 The scenario during the MX period reflects a legacy of the preceding thermal cycle, triggered
722 by the complete water column overturn that redistributes dissolved organic matter. Although
723 RDOC concentrations remain relatively high due to this vertical mixing, they represent a complex
724 mixture: while allochthonous humic-like components persist, a significant portion is derived from
725 the autochthonous RDOC that was efficiently generated and accumulated during the thermal
726 stratification cycle (formation—stabilization—decline) through the DIC-enhanced MCP
727 mechanism. Bacterial degradation capacity is reduced at lower temperatures, resulting in lower
728 carbon conversion efficiency, and positive cohesion shows an inverse relationship with RDOC
729 (**Fig.3 (h)**). The weakening of thermal stratification (**Hasan et al., 2013**) facilitates the sedimentation
730 of POC thereby constraining bacterial transformation and RDOC formation. Although bacteria



731 increase cohesion to share limited resources under these conditions, the overall MCP effect remains
732 relatively weak.

733 In aquatic environments, both DO and bacterial diversity jointly regulate the type of cohesion
734 and the pathway of the MCP. High DO generally promotes the complexity of microbial networks
735 and facilitative interactions (**Hu et al., 2023**), and stimulates the synthesis of extracellular
736 polymeric substances (EPS), enhancing bacterial aggregation (**Bowen et al., 2018**). However, our
737 results revealed an inverse relationship between epilimnetic DO and positive cohesion, potentially
738 attributable to intensified resource competition under conditions of high bacterial diversity (**Fig.4**
739 **(a)**). In the thermocline, the uneven vertical distribution of DO correlate positively with positive
740 cohesion and negatively with negative cohesion. Within the oxygen-depleted hypolimnion, intense
741 competition for oxygen occurred, while negative cohesion declined as bacterial diversity increased.
742 Following the transition to MX conditions, the homogenization of DO throughout the water column
743 alleviated competitive pressure and favored cooperative interactions and metabolic activity among
744 aerobic bacteria (**Yue et al., 2022; Zhou et al., 2020**). Accordingly, higher DO levels were
745 associated with greater positive cohesion but lower negative cohesion ($p < 0.05$). Similarly,
746 increased salinity has been shown to enhance competitive negative correlations (**Ji et al., 2019**),
747 further supporting the role of resource pressure in regulating the type of bacterial cohesion.

748 Given the relatively shallow water depth of the DLD Reservoir and the seasonal development
749 of hypolimnion under ITS and OTS conditions, sedimentary inputs of DOM cannot be excluded as
750 a potential source. Nevertheless, multiple observations from our dataset indicate that benthic
751 release is unlikely to dominate RDOC accumulation. First, analysis of the vertical distribution



revealed no significant RDOC enrichment in deeper waters compared with the surface layer, which deviates from the vertical trend anticipated under a sediment-dominated RDOC source scenario (**Fig. S3**). Second, optical indices further discount a major benthic contribution: the HIX remained low in the hypolimnion, whereas sediment-derived DOM is typically characterized by high humic-like features (**Fig. S5**). Furthermore, the FI shows no significant correlation with RDOC in most layers, and was even negatively correlated in the thermocline (**Fig. 4(a)-(d)**), suggesting that the RDOC pool does not predominantly mirror allochthonous-like signals from the sediment.

Most importantly, the relationship between RDOC and metabolic indicators supports a water-column origin. RDOC concentrations were not significantly associated with DO across the stratified layers, whereas higher RDOC levels coincided significantly with increasing DO under MX conditions (**Fig. 4(a)-(d)**). Such covariation under oxygenated conditions, rather than the inverse relationship anticipated from anaerobic sedimentary release, is consistent with microbial processing of autochthonous carbon. Furthermore, RDOC increased significantly in association with $\delta^{13}\text{C}_{\text{DIC}}$ under fully mixed conditions (**Fig. 4(d)**), suggesting a potential mechanistic connection mediated by microbial carbon fixation. The preferential uptake of $^{12}\text{CO}_2$ by chemoautotrophic taxa for biosynthesis causes a progressive ^{13}C -enrichment in the residual DIC pool. Concurrently, the enhanced carbon fixation flux produces substantial labile organic precursors. These precursors are subsequently channeled through the microbial food web and transformed into refractory compounds via the MCP. Therefore, the observed positive correlation is not a direct stoichiometric coupling, but rather a rate-coupling signature: higher in-situ CO_2 fixation rates simultaneously elevate both the $\delta^{13}\text{C}$ signal in the remaining DIC and the



773 accumulation of newly-formed RDOC, confirming that the RDOC pool originates primarily from
774 water-column internal carbon processing rather than external inputs. Collectively, these results
775 demonstrate that while sediment-water exchange is a physical reality in shallow systems, the
776 RDOC dynamics in DLD Reservoir are fundamentally governed by the DIC-enhanced MCP within
777 the water column rather than internal loading from the benthos.

778

779 **4.3 Nutrient Imbalance-Driven Carbon Sequestration: Coupling Mechanisms Between BCP** 780 **and MCP in Karst Reservoirs**

781 In the carbon cycle of karst reservoirs, BCP initiates the primary production of endogenous
782 organic carbon, whilst MCP is the key factor determining whether these carbon sources can be
783 retained over the long term and converted into carbon sequestration. RDA analysis results (**Fig. 7**)
784 indicate that during thermocline and mixing periods, environmental factors such as DIC, pH, WT
785 and DO were significantly correlated with key microbial community structures ($R^2 > 0.3$, $p <$
786 0.05). This suggests that, within a karst weathering context, the cyclical evolution of reservoir
787 thermocline structures shapes specific microbial community patterns by modulating the water
788 chemistry. Concurrently, the persistent strong correlations between Chl-a, TN, TP and key
789 microbial communities highlight the regulatory role of nutrient status on community structure.
790 Driven by these environmental factors, the regulated microbial communities further alter the
791 bioavailability of DOM via the MCP effect (**Fig. 3 (d)-(g)**).

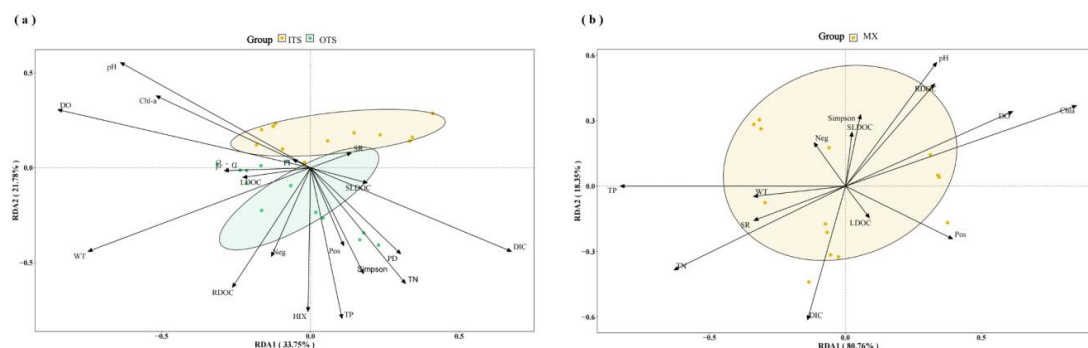


Fig. 7 RDA analysis using keystone taxa from the Thermal Stratification periods (a) and Mixing period (b) as response variables, with other indicators as predictor variables.

This study found that this transition from production to sequestration is strongly influenced by the unique geochemical environment of karst landscapes. Among these environmental filters, the carbonate-rich geochemical background exerts a unique regulatory effect on nutrient stoichiometry, which in turn acts as the primary switch for the BCP-MCP transition. A strongly significant quadratic relationship ($y = -1.7091 + 0.3601x - 0.0016x^2$, $R^2 = 0.395$, $p < 0.01$) is observed between Ca^{2+} concentration and the N:P ratio in **Fig.8(a)**, indicating a progressively accelerating positive response of N:P to increasing Ca^{2+} across the studied gradient. This nonlinear positive pattern likely reflects the enhanced removal of phosphate via calcium-phosphate precipitation (e.g., hydroxyapatite, HAP) under high-pH conditions (**Borkiewicz et al., 2010; Syers et al., 1973**). This physicochemical barrier effectively reduces available phosphorus (SRP) in the water column, leading to a systemic stoichiometric imbalance in the C:P ratio (**Lu et al., 2023; Murphy and Hall, 1983**). The significance of phosphorus is further supported by the Bayesian model (**Fig. 4(f)**), where TP is identified as a primary positive predictor of RDOC concentration. Although TP is essential for the initial carbon production through the BCP, its

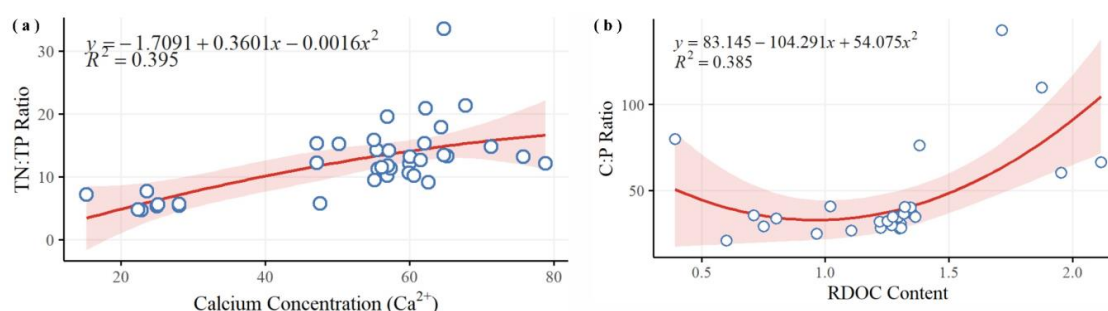


809 positive correlation with RDOC indicates that in karst reservoirs, the total nutrient level does not
810 reflect what is actually available for microbial use. Due to the geochemical immobilization of
811 phosphorus, the bioavailable portion remains low even as the total nutrient pool increases, which
812 maintains the high C:P ratios that are necessary for carbon to persist in the water column.

813 Under phosphorus-limited conditions, RDOC increased significantly along the C:P gradient,
814 following a quadratic polynomial relationship ($y = 83.145 - 104.291x + 54.075x^2$, $p < 0.05$; **Fig.**
815 **8(b)**), with the magnitude of the RDOC response becoming progressively stronger toward higher
816 C:P values. The underlying biological mechanism can be explained from the following two
817 perspectives: first, according to the theory of ecometrics, microbial growth is strictly limited by
818 nutrient ratios (**Sterner and Elser, 2002**). Studies have shown that when phosphorus supply is
819 insufficient to sustain ribosomal RNA and ATP synthesis (**Li et al., 2018; Schuhmacher et al.,**
820 **2014**), microorganisms are unable to effectively convert the high-throughput carbon source
821 produced by BCP into biomass, resulting in a significant decline in carbon use efficiency (CUE)
822 (**Li et al., 2025**). In such cases, microorganisms are forced to release excess carbon in the form of
823 EPS or secondary metabolites (**Zevin et al., 2015**). These metabolites are often enriched in
824 structurally intricate aromatic moieties and exhibit strong resistance to chemical degradation, as
825 exemplified by humin-like substances. Their limited bioavailability restricts subsequent utilization
826 by heterotrophic microorganisms, ultimately favoring the persistence and accumulation of RDOC
827 in aquatic environments (**Jiao et al., 2010; Jiao et al., 2024**). Furthermore, nutrient stress also
828 inhibits extracellular enzyme activity, slowing down the rate of organic matter mineralization and
829 accelerating its conversion into humus. The degradation of organic matter relies on extracellular



830 hydrolases secreted by microorganisms, and the synthesis of such enzymes itself consumes
831 significant amounts of nitrogen and phosphorus. Under phosphorus-limiting conditions,
832 microorganisms tend to downregulate the expression of energy-intensive enzymes to conserve
833 resources (Bilyera et al., 2021; Sun et al., 2023). This results in the failure of pre-existing active
834 organic matter to be decomposed in a timely manner, causing it to persist in the water column for
835 extended periods. Over long scales, these organic compounds undergo photochemical oxidation
836 and repeated microbial reprocessing, transforming through molecular rearrangement into a
837 reservoir of RDOC with more complex structures and greater biological resistance (Liu et al., 2026;
838 Obernosterer and Benner, 2004). In summary, the Ca-P co-precipitation effect, which is unique
839 to karst reservoirs, amplifies phosphorus limitation in the water body. By reducing microbial
840 carbon utilization efficiency and inhibiting enzymatic degradation, it synergistically drives the
841 coupled transition from BCP to MCP. The higher the C:P ratio, the more pronounced this
842 phosphorus-limitation-induced carbon reduction effect becomes, thereby reinforcing the function
843 of karst reservoirs as regional carbon sinks.



844

845 **Fig. 8 Linear correlation analysis of calcium ion concentration with TN:TP (a), and linear**

846 **correlation analysis of C:P in water bodies with RDOC concentration (b).**



847

848 **5. Conclusion**

849 In summary, this research establishes a multi-dimensional framework to elucidate the
850 mechanisms by which karst reservoirs function as effective regional carbon sinks. Central to this
851 framework is the role of thermal stratification, which provides the structural foundation for carbon
852 transformation. By restricting vertical mixing, stratification creates partitioned ecological niches
853 where keystone bacterial taxa facilitate the conversion of labile organic matter into refractory forms;
854 this accumulation of RDOC is independent of benthic inputs. This physical structure is further
855 modulated by microbial dynamics, which regulate the efficiency of the MCP. The fluctuating
856 balance between facilitative and competitive interactions across hydrological periods dictates
857 whether organic carbon is remineralized or sequestered, with competitive interactions under
858 environmental stress proving essential for the long-term persistence of the RDOC pool. Most
859 significantly, the geochemical “Ca-P co-precipitation” effect acts as the primary catalyst for
860 carbon sequestration. By depleting bioavailable phosphorus, this process imposes a severe
861 stoichiometric constraint that decouples carbon uptake from biomass synthesis. This nutrient-
862 limitation forcing inhibits enzymatic mineralization and promote microbial metabolism, effectively
863 shunting excess carbon into stable, chemically complex RDOC.

864 Ultimately, the synergy between geochemical nutrient-locking and microbial processing
865 intensifies the coupling of the BCP and MCP. This study underscores that the carbon sink capacity
866 of karst reservoirs is fundamentally enhanced by their unique alkaline-calcium geochemistry,
867 which facilitates carbon sequestration by imposing severe restrictions on phosphorus



868 bioavailability for the microbial community. This integrated mechanism provides a robust scientific
869 foundation for incorporating karst-specific aquatic processes into global carbon budget models.

870

871 **Author Contribution**

872 **Yikun Jia:** Conceptualization, Methodology, Formal analysis, Software, Data Curation, Writing
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880 **Zeng:** Resources, Data Curation, Writing - Review & Editing, Project administration.

881

882 **Conflict of Interest**

883 The authors declare no conflicts of interest relevant to this study

884

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891

892 **Data Availability Statement**

893 The microbial data, water chemical indicators, and the code used to analyze the relationship
894 between them are available in a figShare repository at **Jia (2025)**
895 <https://doi.org/10.6084/m9.figshare.30855587>. The code can also be found at the Github link
896 (<https://github.com/YikunJia/-/releases/tag/MCP>). All data were plotted using Origin 2021,
897 MATLAB, Arcgis 10.8 and R 4.4.3. The DOM analysis method: Parallel factor analysis was plotted
898 using the MATLAB software, and the source code was referenced from **Colin (2008)**
899 <https://doi.org/10.4319/lom.2008.6.572>.

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