Alpine wetland degradation reshapes the balance between hydrolytic and reductive~~shifts enzyme-mediated~~ soil nitrogen cycling enzyme potentials on the Qinghai–Tibetan Plateau~~from functional coordination to imbalance~~

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Abstract. Alpine wetlands on the Qinghai-~~–~~Tibet Plateau are increasingly degraded, with potentially important consequences for~~threatening~~ soil nitrogen (N) cycling and ecosystem functioning. However, how degradation restructures enzyme-mediated soil N cycling and its functional potential~~balance~~ remains poorly understood. To address this gap, we conducted a three-year field investigation across 2019 to 2021 four degradation stages (i.e.non-degraded (ND),~~,~~ lightly degraded (LD),~~,~~ moderately degraded (MD),~~,~~ and heavily degraded (HD))~~wetlands)~~ in the Gahai Wetland. We quantified four key soil N-cycling enzymes, including~~namely~~ urease, protease, nitrate reductase, and nitrite reductase, and developed a nitrogen cycling functional potential (NCFP~~balance (NCFB~~) index to characterize~~assess~~ shifts in the relative balance ~~coordination~~ between hydrolytic and reductive N-cycling functions potentials. Our results showing that degradation~~. Degradation~~-induced drying and nutrient depletion reduced soil water content, soil organic carbon, total nitrogen, ammonium availability, and microbial biomass N, while increasing soil temperature and nitrate accumulation. Enzyme responses were strongly pathway specific: ~~Degradation increased~~ urease and nitrate reductase activities increased with degradation, whereas ~~but decreased~~ protease and nitrite reductase activities declined. Consequently, NCFP index shifting from slightly positive values in ND and LD to negative values in MD and HD, indicating a reorganization ~~enzyme specific restructuring~~ of ~~soil~~ N- cycling~~. Most enzyme activities peaked in mid-growing season, whereas nitrate reductase peaked later. Soil N cycling shifted from a relatively coordinated state in non-degraded wetlands toward greater~~ functional potential from a relative hydrolytic bias toward a relative reductive bias. Interannual ~~imbalance in degraded wetlands. This reorganization was strongly modulated by~~ hydroclimatic variability further modified this pattern; notably, exceptionally wet~~, with drier~~ conditions in 2021 shifted NCFP index toward negative values across all~~amplifying~~ degradation stages~~, induced divergence in enzyme activities~~. Redundancy analysis and piecewise structural equation modeling showed that soil water content, and organic carbon~~, inorganic N availability, and microbial biomass N~~ were the primary regulators of NCFP index. Alpine wetland degradation reorganized the relative configuration of hydrolytic and reductive N-cycling potentials rather than uniformly suppressing N ~~enzyme variation and N~~ cycling, and this functional balance was jointly regulated by long-term degradation and short-term hydrological variability.~~imbalance.~~ Overall,

thethis study provides new insight into mechanisms of soil N-cycling destabilization in degraded alpine wetlands and informs wetland restoration under a drier future climate.

1 Introduction

Alpine wetlands are among the most important ecosystem types on the Qinghai-–Tibet Plateau (QTP), which is often referred to as the "Third Pole of the Earth", and play indispensable roles in biodiversity conservation, regional climate regulation, water retention, and global carbon (C),) and nitrogen (N) and phosphorus (P) biogeochemical cycling (Li et al., 2021; Wei et al., 2015; Yin et al., 2022). Over recent decades, however, alpine wetlands in this regionacross the QTP have undergone widespread degradation under the combined impactspressures of climate change, hydrological alteration, grazing pressure, and other anthropogenic disturbanceshuman disturbance, particularly overgrazing (Li et al., 2021, 2019; Yuan et al., 2025)(Li et al., 2021, 2019; Yuan et al., 2025). TheThis degradation typically manifests as vegetation succession, biodiversity loss, soil water depletion, and nutrient impoverishment (Alhassan et al., 2018; Du et al., 2024; Ma et al., 2020a; Zhao et al., 2025). Because N cycling is a central process regulating primary productivity, microbial activity, and ecosystem resilience in alpine wetland systems, degradation-induced shifts in soil N cycling are likely to have profound consequences for ecosystem functioning and restoration potential (Chen et al., 2022). Clarifying how soil N cycling responds to alpine wetland degradation is therefore essential for understanding and predicting ecosystem feedback to ongoing global change.

Soil enzymes, which are mainly produced by soil microorganisms and plant roots, mediate essential biogeochemical processes involved in C, Ncarbon, nitrogen, and P-phosphorus cycling (Allison and Vitousek, 2005; Ma et al., 2020b). Owing to their rapid responses to environmental perturbations, soil enzymes have been widely considered recognized as sensitive biochemical indicators of wetland health and soil functional status and nutrient transformation ability (Sinsabaugh et al., 2009; Sinsabaugh and Follstad Shah, 2012; Wang et al., 2022). In N-limited alpine wetland(Sinsabaugh et al., 2009; Sinsabaugh and Follstad Shah, 2012; Wang et al., 2022). In N-limited alpine ecosystems, N-cycling enzymes regulate key steps of organic N depolymerization and inorganic N transformation, thereby influencing N mineralization, denitrification, and potential N loss pathways (Xiang et al., 2024; Zuccarini et al., 2023). In particular, hydrolytic enzymes, such as urease and protease increase the bioavailability of organic N, whereas reductive enzymes such as nitrate reductase and nitrite reductase regulate nitrate and nitrite reduction processes, thereby influencing whether inorganic N is retained within the soil system or further transformed and lost through denitrification-related pathways (Brzostek et al., 2012; Jin et al., 2023; Ma et al., 2021; Priemé et al., 2002; Rajsz et al., 2019; Tabatabai et al., 2010). Despite their central role in regulating soil N transformations, the responses of N-cycling enzymes to alpine wetland degradation remain insufficiently resolved.

The activities of soil N-cycling enzymes are jointly regulated by soil physicochemical properties, plant community composition, temperature, moisture, nutrient availability, and substrate concentration.concentrations (Sardans et al., 2008; Song et al., 2019; Tan et al., 2023). Wetland degradation can substantially reshape these controls by altering vegetation composition, hydrological conditions, soil properties, and microbial habitat quality (Abulaizi et al., 2023; Du et al., 2024; Li et al., 2022). As degradation intensifies, the replacement of hygrophytic species by mesophytic or xerophytic plants generally

reduces plant-derived C and nutrient inputs (Wu et al., 2017), leading to declines in soil organic carbon and total nitrogen pools (Liu et al., 2018). Meanwhile, hydrological drying and associated microclimatic shifts may constrain enzyme diffusion, suppress microbial activity, and reduce catalytic efficiency (Breidenbach et al., 2022; Song et al., 2024). However, the effects of degradation are not uniformly negative across all stages. Although many studies have reported reduced enzyme activity under degraded conditions, others have observed transient stimulation during early degradation, indicating strong context dependence associated with degradation intensity, and resource availability, ~~and grazing pressure~~ (Ma et al., 2020a). Such complexity is especially relevant in the fragile alpine wetlands of the QTP, where degradation may irreversibly alter soil conditions and microbial functioning (Alhassan et al., 2018; Du et al., 2024).

This knowledge gap is further complicated by~~particularly important in alpine wetlands because enzyme-mediated N transformations are highly sensitive not only to degradation intensity but also to~~ strong seasonal and interannual hydroclimatic variability in alpine wetlands. (Deng et al., 2025). On the QTP~~Qinghai–Tibet Plateau~~, short growing seasons, large fluctuations in soil moisture, and increasingly frequent drought events can substantially modify microbial activity (Sorensen et al., 2018), substrate diffusion, and redox conditions, thereby altering both the magnitude and direction of soil N-cycling processes (Asensio et al., 2024; Gao et al., 2021). Although seasonal variation in individual soil enzymes has been reported in alpine ~~and~~ wetland ecosystems (Akinyemi et al., 2020; Bai et al., 2023; Brzostek et al., 2012), most ~~existing~~ studies have focused~~been based~~ on single sampling periods, short-term observations, or isolated enzyme responses. Consequently, we still lack a process-based understanding of whether wetland degradation simply suppresses soil N cycling or instead reorganizes it by differentially affecting distinct enzymatic pathways under contrasting hydroclimatic conditions.

To address this knowledge gap, we conducted a three-year study from 2019 to 2021 ~~field investigation across four stages of alpine wetland degradation~~ in the Gahai Wetland on the northeastern QTP. Using a space-for-time substitution approach, we identified four alpine wetland degradation stages, non-degraded (ND), lightly degraded (LD), moderately degraded (MD), and heavily degraded (HD), under comparable climatic conditions and ~~Rather than treating~~ soil parent material. We measured~~N-cycling enzymes as independent indicators, we selected~~ four representative enzymes associated with hydrolytic and reductive N transformations, including urease, protease, nitrate reductase, and nitrite reductase.~~involved in N hydrolysis and N reduction to test whether wetland degradation drives a functional reorganization of soil N cycling under hydrological drying and nutrient depletion.~~ We further developed a nitrogen cycling functional potential (NCFP~~balance (NCFB~~) index, expressed as the natural logarithm of the ratio between aggregated hydrolytic and reductive potential enzyme activities, to quantify shifts in their~~the~~ relative functional potential. Specifically, we asked: (1) whether alpine wetland degradation induces enzyme-specific rather than uniform changes in potential N-cycling ~~coordination between hydrolytic and reductive~~ functions; (2) how~~, and linked~~ these responses are associated with ~~shifts to~~ changes in soil hydrological conditions, nutrient ~~water status, carbon and nitrogen~~ availability, and microbial N pools~~biomass. Specifically, this study asked: (1) whether degradation alters soil N cycling through enzyme-specific rather than uniform responses; (2) whether hydrological limitation and substrate depletion are the main mechanisms underlying these divergent responses~~; and (3) whether interannual ~~climatic variation, particularly drought, amplifies degradation-induced functional decoupling. By explicitly integrating degradation intensity,~~ hydroclimatic variability

modifies the magnitude of degradation responses among individual N-cycling pathways. By integrating degradation severity, environmental variation, individual enzyme responses, and their relative functional configuration~~, and enzymatic functional balance~~, this study provides~~moves beyond descriptive assessments of enzyme activity and provide~~ a process-oriented~~mechanistic~~ framework for understanding how alpine wetland degradation reorganizes~~reshapes~~ belowground N-cycling potential.

2. Materials and methods

2.1 Study area

This study was conducted in the Gahai–Zecha National Nature Reserve (102°26′ E, 34°16′ N; 3450 m a.s.l.), located in the northeastern QTP, China (**Figure 1**). The region is characterized by a highland continental monsoon climate. The mean annual temperature of the Gahai wet meadow is 1.2 °C, with the lowest and highest monthly mean temperatures occurring in January (-9.1 °C) and July (10.5 °C), respectively (Du et al., 2025). The mean annual precipitation is approximately 782 mm, of which nearly 80% occurs during the growing season from May to September (**Figure S1**). According to the World Reference Base for Soil Resources (WRB) classification system (Iuss Working Group Wrb, 2022) , the soil type in the study area belongs to Histosols and Gleysols. The dominant plant species include *Carex tibetikobresia*, *Carex meyeriana*, *Thalictrum aquilegifolium*, *Artemisia subulata*, and *Artemisia linearis* (Chang et al., 2023).

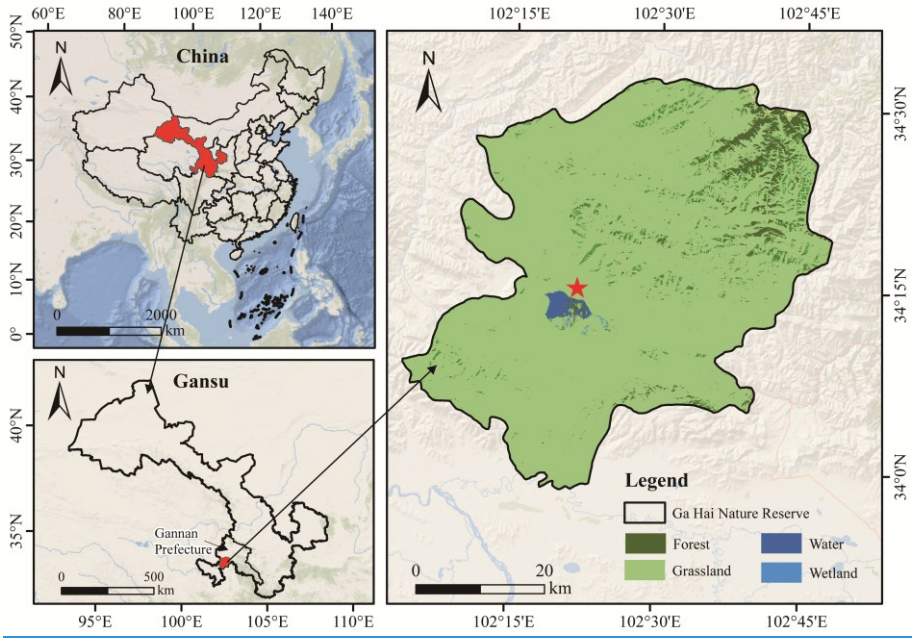


Figure 1 The location of the study area.

Annual precipitation differed markedly among the three study years, with values of 816.4, 666.2, and 1216.4 mm in 2019, 2020, and 2021, respectively (**Figure S2**). Relative to the long-term mean annual precipitation of 782 mm, the corresponding

precipitation anomalies were 4.4%, −14.8%, and 55.6%, respectively. Thus, 2020 represented a relatively dry year, whereas 2021 was exceptionally wet compared with the long-term climatic mean.

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Over the past half-century, temperatures in this region have increased by 0.4 °C per decade, while precipitation has decreased by 2 mm per decade (Masson-Delmotte et al., 2021; Wang et al., 2024). This climatic shift has triggered progressive drying within the Gahai Wetland, significantly diminishing its water storage capacity. Concurrently, under pressure from population growth and socioeconomic activities, a substantial portion of the region's wetlands were drained for conversion to pastureland. From October to April, the Gahai wet meadows are routinely utilized as winter pasture for yaks, sheep, and goats. Long-term overgrazing coupled with inappropriate grazing management practices have greatly exacerbated wetland degradation (Du et al., 2025). Furthermore, high densities of *Ochotona curzoniae* are observed in this area. Consequently, this site presents an ideal setting for investigating the response of N-cycle enzyme activities to wet meadow degradation.

2.2 Experimental design

We referred to the classification criteria for alpine wetland degradation specified in the "Classification of degradation grade of alpine marshes" (DB63/T 1359–2015) and the "Ecosystem health assessment of Alpine Marsh wetland" (DB63/T 1544–2017) and further considered differences in vegetation characteristics and hydrological conditions within the study area (Liu et al., 2025). Using a space-for-time substitution approach, we established a belt transect extending outward from the relatively intact wetland surrounding Gahai Lake. According to the composition of dominant species, vegetation coverage, biomass and groundwater level and other characteristics (Table S1). Sites with relatively flat topography and comparable elevations were selected and classified into~~Based on differences in wetland vegetation characteristics and hydrological conditions in the study area, a space-for-time substitution approach was used to delineate~~ four degradation stages: non-degraded (ND), lightly degraded (LD), moderately degraded (MD), and heavily degraded (HD).~~) wetlands (**Table S1**).~~ Within each degradation stage, three 10 ~~m~~ × 10 m ~~sampling~~ plots were randomly established, resulting in a total of 12 plots. All sampling sites are located within the same local wetland landscape.~~sampling plots.~~ To minimize potential edge effects, a buffer distance of at least 10 m was maintained between adjacent plots. All experimental plots were fenced throughout the study period~~plots belonging~~ to

exclude livestock grazing, including the local winter grazing season from October to the following May. ~~different degradation stages.~~ Detailed information ~~on the vegetation characteristics of the sampling plots~~ has been previously reported in our earlier~~previous~~ studies (Chang et al., 2023; Du et al., 2024).

2.3 Soil samples and analysis

Soil samples were collected during the growing seasons from June to September in 2019, 2020, and 2021. Within each of the 12 established plots, composite soil samples were collected using a five-point sampling method. At each sampling point, soil cores with a diameter of 5 cm were collected from three depths: 0–10, 10–20, and 20–40 cm. Soil samples from the same depth within each plot were thoroughly homogenized to form one composite sample, yielding a total of 432 composite soil samples across the study period (4 × 3 ×12× 3). ~~: 12 sampling events × 4 degradation stages × 3 soil depths × 3 replicates.~~

In addition, undisturbed soil cores were collected from each plot using stainless-steel cutting rings with a volume of 100 cm^3 to determine soil bulk density. Visible roots, plant residues, and organic debris were carefully removed by hand. Fresh soil samples were then passed through a 2-mm mesh sieve to ensure sample homogeneity. Each sieved sample was divided into two subsamples. One subsample was air-dried at room temperature for the analysis of basic soil physicochemical properties, whereas the other was stored at 4 °C in a portable cooler and transported to the laboratory for the analysis of microbial biomass and related biochemical properties. These analyses were completed within two weeks of sample collection.

~~**2.4 Soil physicochemical property analysis**~~

Soil pH was determined in a 1:2.5 soil-to-water suspension using a pH meter (Bao, 2000). Soil organic carbon (SOC) was measured using the potassium dichromate oxidation method. Total nitrogen (TN) was determined using the semi-micro Kjeldahl distillation–titration method (Lu, 2000). Total phosphorus (TP) was measured using the molybdenum–antimony colorimetric method after acid digestion. Soil ammonium nitrogen (NH_4^+-N) and nitrate nitrogen (NO_3^--N) contents were determined by KCl extraction followed by distillation–titration (Min et al., 2011). Microbial biomass nitrogen (MBN) was measured using the chloroform fumigation–extraction method with K_2SO_4 as the extractant (Brookes et al., 2002). Soil water content (SWC) and soil temperature (TEM) were measured using soil moisture/temperature sensors (5TM and EC-TM sensors) connected to a data logger (Em50G, Decagon Devices, Pullman, WA, USA).

2.4 Determination of potential soil nitrogen ~~5 Soil N~~ cycle enzyme activity ~~analysis~~

The activities of four soil N-cycling enzymes, namely urease, protease, nitrate reductase, and nitrite reductase, were determined colorimetrically using urea, casein, nitrate, and nitrite as substrates, respectively (Ma et al., 2021; Ou et al., 2019; Pu et al., 2019). After a 24 -h incubation at a constant temperature, the absorbance of each enzyme assay was measured using a spectrophotometer at the corresponding wavelength: 578 nm for urease, 560 nm for protease, and 520 nm for both nitrate reductase and nitrite reductase. Because these assays were conducted under standardized incubation conditions with defined substrates, the resulting activities are interpreted as potential enzyme activities. Detailed procedures for these enzyme assays have been described previously (Chang et al., 2023).

2.5~~6~~ Soil nitrogen cycling functional potential (NCFP) index~~balance~~

To assess degradation -induced shifts in the internal organization of soil N--cycling, a nitrogen cycling functional potential (NCFP~~balance (NCFB~~) index was calculated as the ~~standardized~~ difference between hydrolytic and reductive enzyme functions: NCFP= ln[(~~NCFB = z(~~urease) + ~~z~~(protease)] − ln[(~~) = [z(~~nitrate reductase) + ~~z~~(nitrite reductase)]. The higher NCFP index value indicates that hydrolysis dominates~~)], where z denotes z-score standardization across all samples. Higher NCFB values indicate a relative predominance of hydrolytic~~ over reduction (> 0),~~reductive functions,~~ whereas a lower NCFP index value indicates that reduction dominates over hydrolysis (<0).~~values indicate a shift toward reductive dominance and greater functional imbalance within soil N cycling.~~

2.6~~7~~ Statistical analyses

All statistical analyses were performed in R version 4.2.2 (R Core Team, 2022). Data normality was assessed using the Kolmogorov–Smirnov test and for homogeneity of variance with Levene's test.~~.~~ Variables that did not meet the assumption of normality were log-transformed before subsequent analyses. One-way analysis of variance (ANOVA) was used to test for differences in soil physicochemical properties and potential soil N-cycling enzyme activities among degradation stages, followed by Tukey's honestly significant difference (HSD) test for multiple comparisons.

Linear mixed-effects models were used to evaluate~~To quantify~~ the effects of degradation stage~~, sampling year,~~ and interannual variation~~sampling month~~ on potential soil N-cycling enzyme activities and NCFP index~~, linear mixed-effects models were fitted using the lme4 package~~ (Bates et al., 2014). Degradation stage, year, and their interaction were included as fixed effects, whereas sampling month and plot identity were included as crossed random intercepts to account for temporal variation and repeated measurements within permanent plots, respectively. No random slopes were included. The model was specified as enzyme activity ~ degradation × year + (1|month) + (1|plot).~~sampling year, and sampling month were treated as fixed effects, whereas sampling block and soil depth were included as random effects. To estimate the relative importance of the main fixed factors, hierarchical partitioning was further performed using the glmm.hp package (Lai et al., 2022), and the independent contributions of degradation stage, sampling year, and sampling month to variation in each enzyme activity were calculated.~~

To evaluate the temporal sensitivity of degradation effects, enzyme responses in degraded wetlands relative to non-degraded wetlands were additionally expressed as logarithmic response ratios (Hedges et al., 1999). Because enzyme responses along the degradation gradient were not expected to be strictly linear, generalized additive mixed models (GAMMs) were fitted to test for potential nonlinear relationships between degradation intensity and each enzyme activity (Wood, 2011).

Redundancy analysis (RDA) was performed to examine the relationships between potential soil N-cycling enzyme activities and soil physicochemical and microbial variables (Oksanen et al., 2007). Prior to ordination, highly collinear explanatory variables were removed based on collinearity diagnostics. The significance and relative importance of the remaining explanatory variables were then evaluated using Monte Carlo permutation tests with 999 permutations. Because some potentially important variables were excluded during collinearity screening, Spearman correlation analysis combined with best-subset multiple regression was further used to quantify the relative contributions of key biotic and abiotic factors to variation in enzyme activities. Finally, based on the key environmental variables identified by the RDA and regression analyses, a piecewise structural equation model was constructed to evaluate the direct and indirect pathways through which wetland

degradation regulated the NCFP~~NCFB~~ index (Lefcheck, 2016). Statistical significance was assessed at $P < 0.05$ unless otherwise stated.

3. Results

3.1 Wetland degradation reshaped the soil hydrological and nutrient environment

Wetland degradation was accompanied by pronounced shifts in soil hydrological and biogeochemical conditions (**Table 1**). Across the degradation gradient, soil water content (SWC), pH, soil organic carbon (SOC), total nitrogen (TN), total phosphorus (TP), ammonium nitrogen (NH_4^+-N), and microbial biomass nitrogen (MBN) generally declined, whereas soil temperature (TEM) and nitrate nitrogen (NO_3^--N) increased. Soil bulk density was also higher in degraded wetlands than in non-degraded wetlands, with the highest value observed under moderate degradation. Among these variables, the strongest degradation-related gradients were observed for SWC, MBN, and inorganic N forms. Non-degraded wetlands maintained substantially higher SWC, NH_4^+-N, and MBN, whereas degraded wetlands, particularly moderately degraded and heavily degraded wetlands, showed lower substrate and microbial N availability but higher NO_3^--N. Overall, these patterns indicated that wetland degradation progressively transformed the soil environment from a wetter and C- and N-rich state toward a drier, warmer, and more nutrient-depleted condition. Principal component analysis (PCA) further demonstrated clear multivariate differentiation in soil physicochemical conditions across the degradation gradient (**Figure S3**), with PC1 and PC2 explaining 63.2% and 14.2% of the total variation, respectively. The degradation stages showed an ordered ND, LD, MD and HD shift primarily along PC1, and PERMANOVA confirmed significant differences in overall soil physicochemical conditions among stages ($P < 0.001$).

Table 1 Basic physical and chemical properties of sample plots.

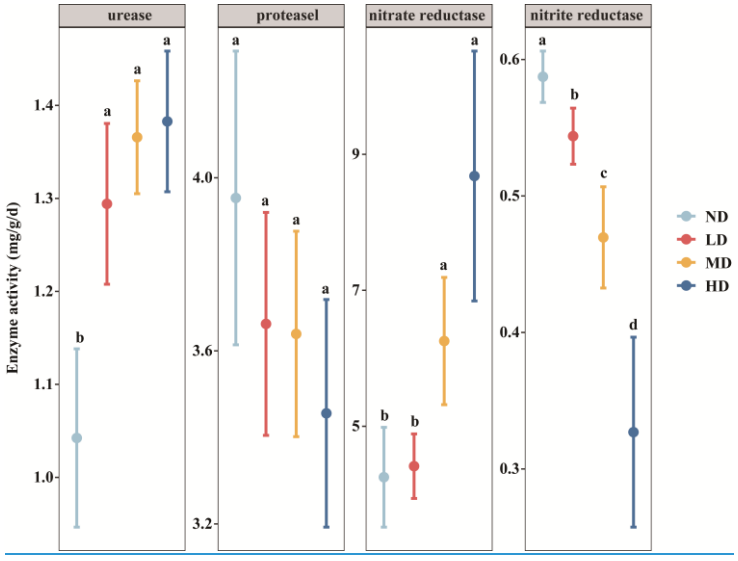
	ND	LD	MD	HD
SWC ($m^3 \cdot m^{-3}$)	0.45±0.12 a	0.31±0.09 a	0.23±0.08 b	0.14±0.03 b
TEM (°C)	9.82±0.10 c	10.35±0.11 b	11.28±0.12 b	13.26±0.05 a
pH	7.92±0.04 a	7.79±0.06 b	7.77±0.08 b	7.76±0.06 b
SOC ($g \cdot kg^{-1}$)	52.55±7.85 a	42.01±5.60 ab	36.45±1.89 ab	32.99±5.99 b
TN ($g \cdot kg^{-1}$)	3.07±0.18 a	2.67±0.20 ab	2.41±0.16 b	1.88±0.21 c
TP ($g \cdot kg^{-1}$)	1.48±0.51 a	1.29±0.30 ab	1.17±0.08 b	1.15±0.22 b
C:N	16.24±0.48 a	15.41±0.72 a	14.75±0.42 b	14.07±0.20 b
C:P	35.53±0.54 a	33.02±0.61 b	31.17±0.51 c	28.67±0.38 d
N:P	2.21±0.02 a	2.16±0.11 a	2.15±0.12 a	1.71±0.07 b
NH_4^+-N ($mg \cdot kg^{-1}$)	6.20±0.51 a	5.87±0.45 b	5.20±0.35 c	4.81±0.21 c
NO_3^--N ($mg \cdot kg^{-1}$)	4.71±0.21 c	5.01±0.18 c	6.32±0.14 b	7.12±0.09 a
MBN ($mg \cdot kg^{-1}$)	45.26±2.25 a	37.35±3.20 b	35.12±3.84 b	29.45±2.24 c

Note: Values are means±standard error (n=3). ~~Note:~~ SWC: soil water content, TEM: temperature. BD: bulk density, SOC: soil organic carbon, TN: total nitrogen, TP: total phosphorus, NH_4^+-N: ammonium nitrogen, NO_3^--N: nitrate nitrogen, MBN, microbial biomass nitrogen. ND, non-degraded; LD, lightly degraded; MD, moderately degraded; HD, heavily degraded. Different lowercase letters in the same row indicate

significant differences between different degrees of degradation (P<0.05). Values are Means ± Standard error (n = 3).~~The data in the table are expressed as average ± standard deviation~~

3.2 Potential soil~~Soil~~ N-cycling enzymes responded divergently to degradation

Wetland degradation induced clear divergence among potential soil N-cycling enzymes~~enzymatic~~ pathways (**Figure 2~~1~~**). Urease and nitrate reductase activities generally increased along the degradation gradient, whereas protease and nitrite reductase activities declined. Although nitrate reductase activity did not differ significantly between non-degraded and lightly degraded wetlands, urease activity differed significantly between these two degradation stages ($P < 0.05$). Soil protease activity decreased with increasing wetland degradation, but the maximum reduction, observed under heavy degradation, was only 13.00% relative to non-degraded wetlands. Accordingly, no significant differences in protease activity were detected among the four degradation stages. In contrast, nitrite reductase activity showed a pronounced decreasing trend, with significant differences observed among degradation stages. Nitrite reductase activity decreased by 95.30% from non-degraded to heavily degraded wetlands. Standardization further highlighted divergent responses among the four enzymes (**Figure S4**). Along the degradation gradient, urease and nitrate reductase shifted progressively toward higher standardized activities, whereas protease and nitrite reductase shifted toward lower values. The strongest relative change was observed for nitrite reductase, which declined from strongly positive in ND to strongly negative in HD, while protease showed the weakest response~~In particular, nitrite reductase activity decreased by 95.30% from non-degraded to heavily degraded wetlands~~.



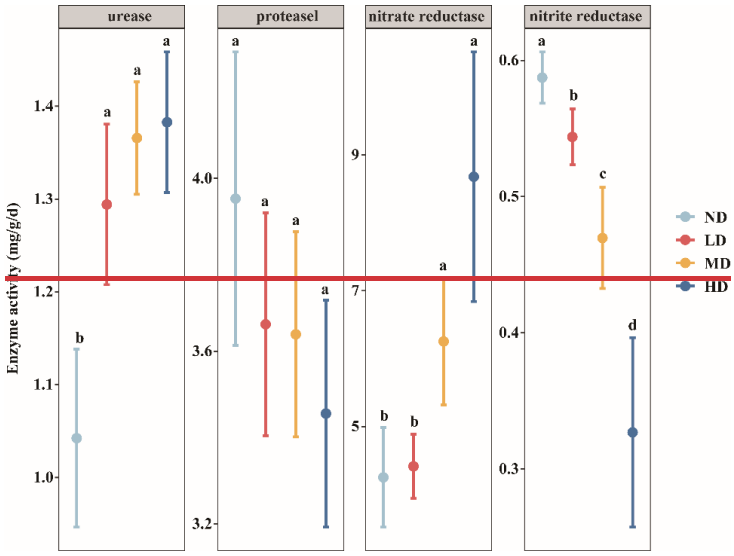


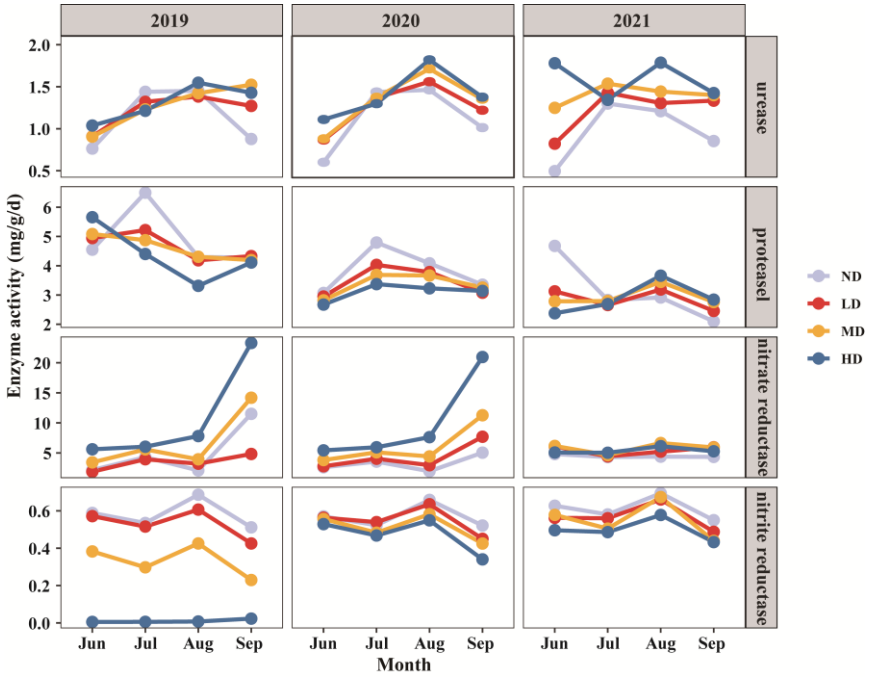
Figure 2. Soil enzyme ~~1. Enzyme~~ specific responses of four soil N~~nitrogen~~-cycling enzymes across the alpine wetland degradation gradient. Activities of (a) urease, ~~(URE),~~ (b) protease, ~~(PRO),~~ (c) nitrate reductase, ~~(NR),~~ and (d) nitrite reductase, ND, ~~(NIR) are shown for~~ non-degraded; LD, ~~(ND),~~ lightly degraded; MD, ~~(LD),~~ moderately degraded; HD, ~~(MD), and~~ heavily degraded ~~(HD) wetlands~~. Different lowercase letters indicate significant differences among degradation stages at $P < 0.05$.

3.3 Temporal variation modulated the strength of degradation effects

Potential soil~~Soil~~ N-cycling enzyme activities exhibited clear seasonal and interannual variation (**Figure 3~~2~~**). Urease activity generally increased from the early to the middle growing season and then declined toward September, whereas protease activity showed a similar seasonal trajectory but remained relatively low late in the growing season. Nitrate reductase activity was typically lowest in June and increased thereafter, whereas nitrite reductase activity displayed a more conserved seasonal pattern, with peak values generally occurring in August. Potential soil N-cycling enzyme activities also differed among years, coinciding with marked interannual variation in precipitation, particularly the exceptionally wet conditions in 2021 (**Figure S5**). These patterns indicated that the timing of maximum activity differed among enzymatic pathways, suggesting different temporal controls on hydrolytic and reductive processes.

Linear mixed-effects models ~~further~~ showed significant effects of~~that~~ degradation stage and sampling time on ~~month significantly affected~~ all four enzymes, whereas the effects ~~of sampling year~~ were enzyme-specific (**Table 2**). Significant degradation × year interactions occurred for ~~Degradation stage explained the largest proportion of variation in~~ urease, nitrate reductase, and nitrite reductase, but ~~activities, sampling month explained most of the variation in nitrate reductase activity, and sampling year dominated variation in protease activity (**Figure S2**). Interannual contrasts also showed that degradation effects were~~ not for protease.~~constant over time.~~ Urease showed the strongest~~responses to~~ degradation differentiation~~were weak~~ in ~~2019 but became stronger in 2020 and~~ 2021, nitrate reductase differed significantly among~~especially under moderate and heavy~~ degradation stages in 2019 and 2020. ~~Protease activity generally declined with degradation, with the clearest reductions occurring in the drier year. Nitrate reductase and nitrite reductase activities also showed year-dependent differences in the magnitude,~~ but not in 2021, and the decline in nitrite reductase ~~the overall direction, of degradation effects. Together,~~

~~these results showed that temporal variation regulated not only absolute enzyme activity but also the strength with which~~ ~~degradation~~ was strongest in 2019.~~functionally expressed.~~ Generalized additive mixed models (GAMMs) indicated nonlinear and enzyme-specific responses along the degradation gradient (**Figure S6**).~~further showed that~~ the responses of soil N-cycling enzymes to degradation were enzyme-specific and were not uniformly linear along the alpine wetland degradation gradient.~~(Figure S3).~~ Urease and nitrate reductase activities showed stronger nonlinear increases with degradation, whereas protease and nitrite reductase activities declined more gradually or showed year-specific rates of decline. Overall, interannual variation modulated degradation effects differently among the four N-cycling enzymes.



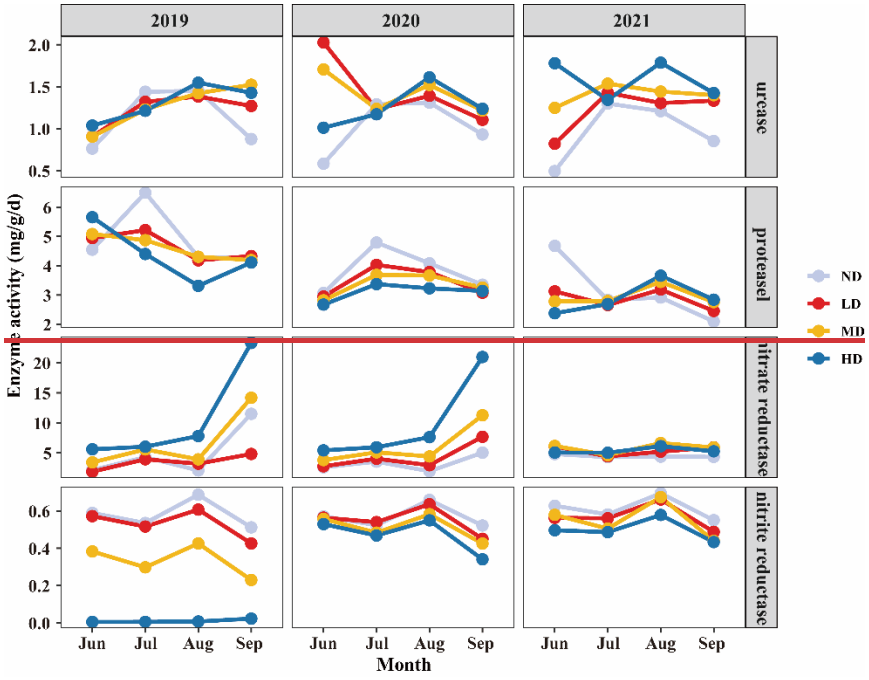


Figure 3~~2~~. Temporal changes in potential soil N-cycling~~cycle~~ enzyme activities across degradation~~at different~~ levels ~~of degradation~~ during 2019, 2020, and 2021. Values are means of three independent replicate plots (n = 3) for the integrated 0–40 cm soil layer. URE, ~~and 2021.~~ urease; PRO, ~~(URE),~~ protease; ~~(PRO), nitrate reductase (~~NR, nitrate reductase; NIR,~~), and~~ nitrite reductase. ~~(NIR).~~ ND, non-degraded; LD, lightly degraded; MD, moderately degraded; HD, heavily degraded.

Table 2. Fixed effects ~~Effects~~ of degradation level~~stage~~, year, and their interaction~~sampling time~~ on potential soil N-cycling enzyme activities based on linear mixed-effects models.

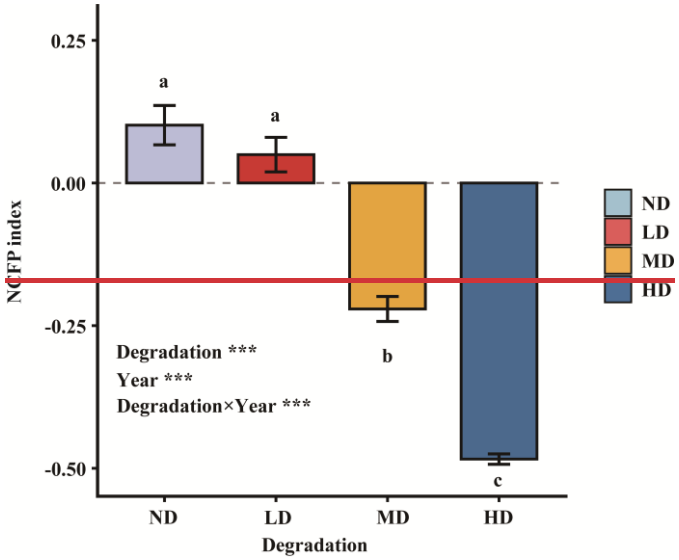
	df	Urease		Protease	Nitrate reductase	Nitrite reductase	
Degradation	(3,129)	~~14.90~~**	15.53**	4.17~~40~~**	18.41~~29.13~~**	349.08~~87.44~~**	
Year	(2,129)	~~1.516~~		0.94~~196.45~~**	98.72**~~6.74~~*	2.14~~164.11~~**	382.88**
Degradation × Year~~Month~~	(6,129)	~~12.60~~**		3.91~~7.73~~**	0.42~~52.03~~**	4.23~~30.28~~**	141.06**

Note: Degradation level, year~~Summary of linear mixed model analyzing the effects of degradation and time on four soil N cycle enzyme activities, using degradation, time~~, and their interaction were included as fixed effects, whereas sampling month~~depth~~ and plot identity~~block~~ were included~~taken~~ as crossed~~a~~ random intercepts. F tests for fixed effects were evaluated using Satterthwaite-approximated denominator degrees of freedom. Values shown are F statistics; *P < ~~factor. The numeric value represents the size of the F value, * and ** representing p<~~0.05 and P < ~~p<~~0.01~~, respectively~~.

3.4 Wetland degradation shifted the hydrolytic to reductive~~induced imbalance in soil~~ N-cycling potential~~functions~~

The nitrogen cycling functional potential (NCFP~~balance (NCFB~~) index declined~~decreased~~ significantly along the wetland~~with increasing~~ degradation gradient~~intensity~~ (**Figure 4~~3~~**). Relative to ND, NCFP decreased ~~the NCFB index declined~~ by approximately 0.052~~465~~, 0.322~~660~~, and 0.585 units~~775~~ in LD, MD, and HD, respectively. No significant difference~~The NCFB index~~ was detected between~~positive in~~ ND and ~~wetlands (0.432), approached zero under~~ LD, whereas all other pairwise comparisons were significant ($P < 0.01$). Degradation stage, year, and their interaction all had highly significant effects on NCFP. The relatively small difference between ND and LD, followed by pronounced declines~~became negative~~ under MD and

HD ~~(-0.18 and -0.30, respectively). ANOVA showed that ND differed significantly from all degraded stages, including LD, MD, and HD, confirming a strong effect of wetland degradation on soil N cycling functional balance.~~



~~**Figure 3. Response of N cycling functional balance (NCFB) index to wetland degradation. ND, non-degraded; LD, lightly degraded; MD, moderately degraded; HD, heavily degraded.**~~

. Changes in the relationship between hydrolytic and reductive functions further supported this functional reorganization (**Figure** S7~~S4~~). ND, LD, and MD showed negative associations between hydrolytic and reductive functions, whereas HD showed a positive association. The strongest pattern occurred under light degradation, where the two functional groups were significantly negatively correlated (ρ = -0.534). The suggests an apparent transition in N-cycling functional potential once degradation progressed beyond the lightly degraded stage~~By contrast, the relationships in ND (ρ = -0.215), MD (ρ = -0.242), and HD (ρ = 0.265) were not significant. Overall, these results indicated that wetland degradation altered not only the magnitude of enzyme activity but also the coupling structure between major soil N cycling pathways~~.

NCFP index also varied significantly among years (**Figure S8**). In 2019, NCFP decreased by 0.728 units from ND to HD ($P \leq 0.001$), whereas the corresponding decline reached 0.987 units in the relatively dry year of 2020 ($P < 0.001$). In contrast, no significant differences among degradation stages were detected in the exceptionally wet year of 2021, when the ND to HD difference was only 0.04 units. These results indicate that the degradation response of NCFP was strongly modulated by interannual variation, with pronounced degradation gradients in 2019 and 2020 but little differentiation among degradation stages in 2021.

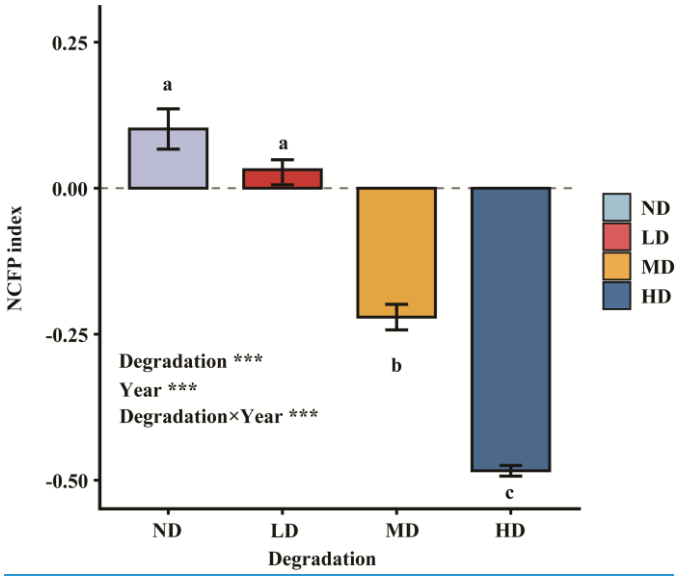
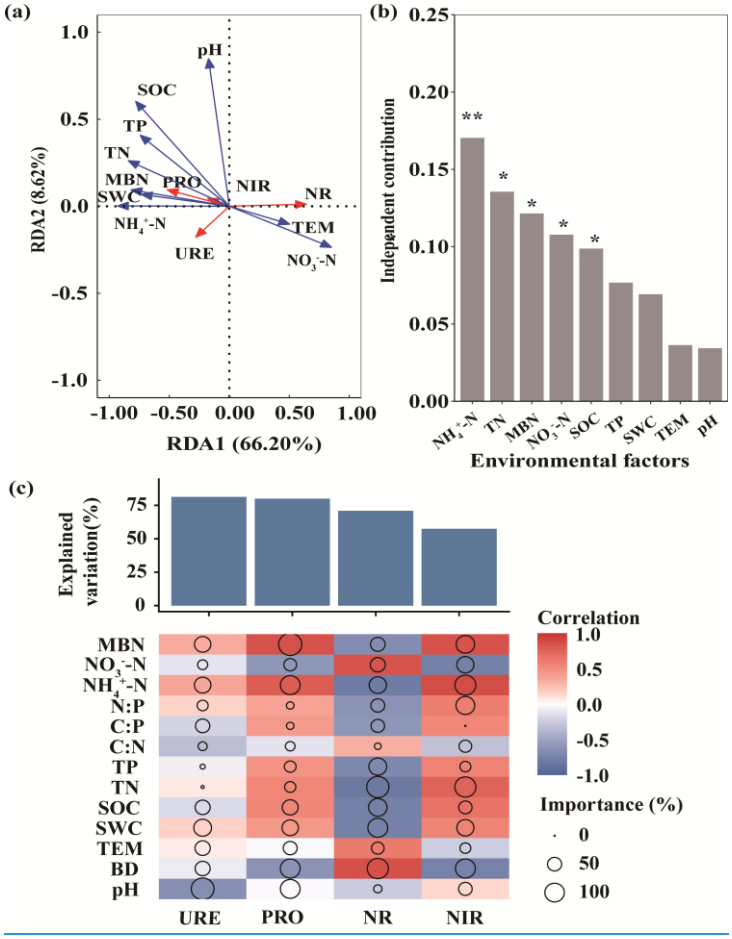


Figure 4. Response of nitrogen cycling functional potential (NCFP) index to wetland degradation. ND, non-degraded; LD, lightly degraded; MD, moderately degraded; HD, heavily degraded.

3.5 Relationships between potential soil N-cycling enzyme activities and potential environmental drivers

Redundancy analysis (RDA) indicated that soil environmental factors strongly structured variation in potential soil N-cycling enzyme activities, with RDA1 and RDA2 explaining 66.20% and 8.62% of the total variation, respectively (**Figure** 5). RDA1 represented the dominant environmental gradient, separating relatively wetter and more resource-rich conditions, characterized by higher SWC, SOC, TN, TP, MBN, and NH_4^+-N, from warmer and more nitrate-rich conditions, characterized by higher TEM and NO_3^--N. In contrast, RDA2 represented a weaker secondary environmental gradient that was primarily associated with pH.~~4).~~ Urease and protease activities were primarily~~mainly~~ associated with the resource-rich side of the ordination~~SOC, TN, TP, MBN, SWC, and NH_4^+ N~~, whereas itrate~~nitrate~~ reductase activity was more closely aligned~~associated~~ with higher TEM and NO_3^--N. Nitrite reductase activity showed comparatively weaker associations with individual environmental variables. Permutation tests with 999 permutations identified NH_4^+-N as the most important independent explanatory variable, followed by TN, MBN, NO_3^--N, and SOC. Correlation analysis combined with best-subset multiple regression further confirmed that TN, MBN, NO_3^--N, and SOC were the major contributors to variation in enzyme activities, with greater explanatory power for urease and protease than for nitrate reductase and nitrite reductase.


(a)
pH
SOC
TP
TN
MBN
PRO
SWC
NIR
NR
NH4+-N
TEM
URE
NO3--N
RDA2 (8.62%)
RDA1 (66.20%)
(b)
Independent contribution
Environmental factors
NH4+-N
TN
MBN
NO3--N
SOC
TP
SWC
TEM
pH
(c)
Explained variation(%)
MBN
NO3--N
NH4+-N
N:P
C:P
C:N
TP
TN
SOC
SWC
TEM
BD
pH
URE
PRO
NR
NIR
Correlation
Importance (%)

340

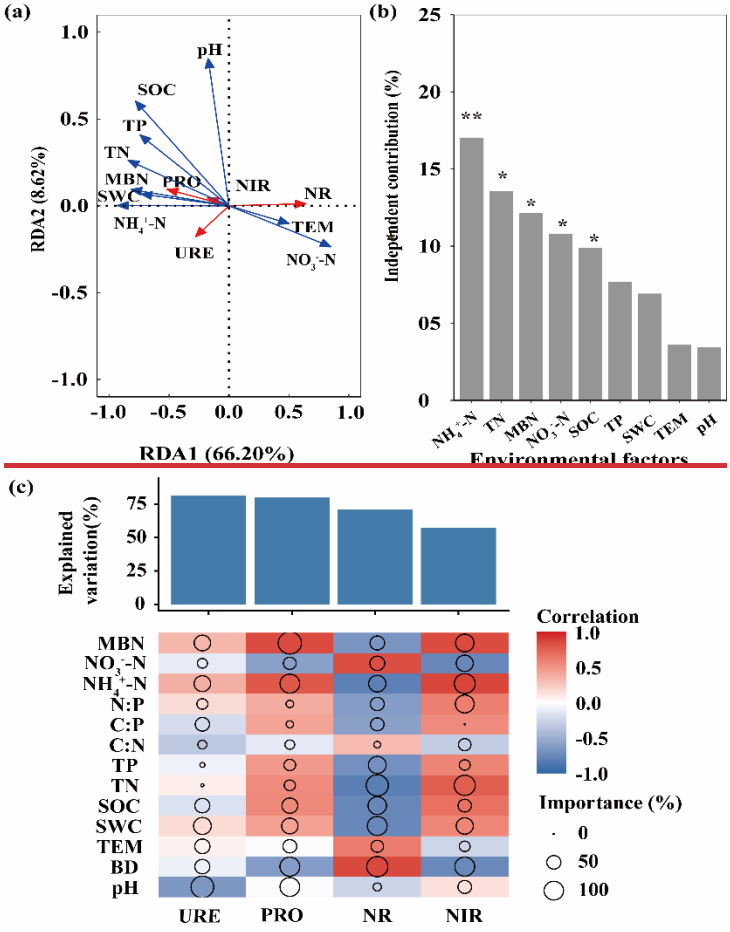


Figure 54. Contributions of soil biotic and abiotic factors to potential soil N-cycling enzyme activities. (a) Redundancy analysis (RDA) ordination of enzyme activities and environmental variables. (b) Permutation-based ranking of the independent contributions of environmental factors identified by RDA. (c) Relative importance of soil variables for enzyme activities based on correlation analysis and optimal multiple regression modeling. Circle size represents~~indicates~~ the relative contribution/importance (%) estimated from the optimal regression model~~of each variable~~, and color indicates the Spearman correlation coefficient. URE, urease; PRO, protease; NR, nitrate reductase; NIR, nitrite reductase; BD, bulk density; TEM, temperature; SWC, soil water content; SOC: soil organic carbon; TN, total nitrogen; TP: total phosphorus; NH_4^+-N, ammonium nitrogen; NO_3^--N, nitrate nitrogen; MBN, microbial biomass nitrogen.

Based on the key environmental factors identified by the RDA and regression analyses, we constructed a piecewise structural equation model (piecewise-SEM) to elucidate how wetland degradation affected the NCFP~~NCFB~~ index through hydrological limitation, carbon resource status, soil N availability, and microbial biomass (**Figure 65**). Wetland degradation had significant positive~~negative~~ effects on SWC and SOC, and SWC showed positive pathways to TN, NH_4^+-N, and NO_3^--N. NH_4^+-N exerted a significant positive effect on NCFP~~NCFB~~, suggesting that greater ammonium availability was associated with a relative predominance of hydrolytic over reductive N-cycling functions. In contrast, SOC had a direct positive~~negative~~ effect on NCFP~~NCFB~~. Overall, the piecewise SEM~~path~~ analysis identified hydrological limitation as a dominant control on

NCFP~~NCFB~~ imbalance, with SWC exerting both direct and indirect effects through associated changes in soil N availability and microbial biomass.

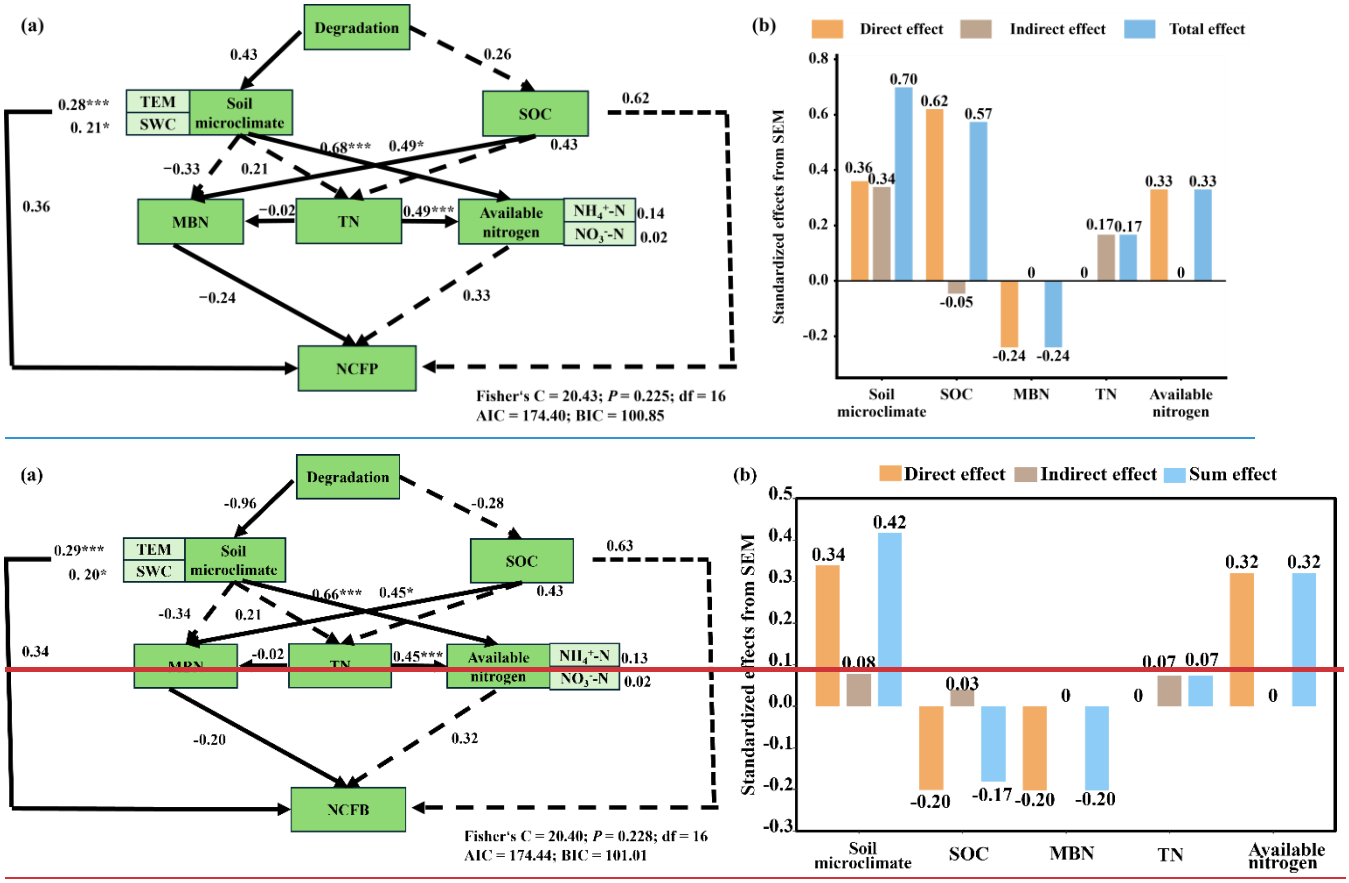


Figure 6~~5~~. Piecewise structural equation model showing how wetland degradation influences nitrogen-cycling functional imbalance through hydrological drying, substrate depletion, and microbial biomass constraints. a) Structural equation modeling shows the influences of N cycling functional potential~~balance~~. b) Direct effects, indirect effects and standardized total effect values of structural equation models. TN, total nitrogen; NH_4^+-N, ammonium nitrogen; NO_3^--N, nitrate nitrogen; MBN, microbial biomass nitrogen; SWC, soil water content; TEM, temperature.

4. Discussion

4.1 Wetland degradation redefines the hydrological and resource environment of soil N cycling

From a process-based perspective, hydrological drying appears to be the primary trigger linking vegetation degradation to belowground functional change. Soil water content declined by 69% from ND to HD (Table 1), and water availability regulates oxygen diffusion, redox conditions, solute transport, and enzyme substrate contact. Progressive drying may therefore shift the soil environment from relatively water-saturated and diffusion-limited conditions toward greater aeration and a more oxic state, thereby altering the environmental constraints imposed on individual N-transformation pathways. This hydrological shift was accompanied by substantial changes in soil N availability and microbial N storage. Declines in SOC and NH_4^+-N indicated

reductions in soil C resources and available N, whereas lower MBN indicated~~As wetland soil loses water and becomes more aerated, substrate diffusion, oxygen availability, and nutrient accessibility are altered (**Table 1**). At the same time, declines in SOC and NH_4^+ N suggest a progressive reduction in readily available C and N substrates, whereas lower MBN indicates~~ reduced microbial biomass and potentially weakened microbial N retention capacity (Cai et al., 2024; Li et al., 2024). Together, these changes created~~create~~ a more resource-limited and increasingly oxic soil environment, in which degradation progressively reduced substrate supply and biological N retention. Rather than uniformly suppressing microbial functioning, reduced water availability, contraction of the microbial N pool, altered substrate availability, and enhanced soil aeration created a fundamentally different physicochemical environment in which individual microbial pathways were subject to different constraints. RDA further revealed differential environmental regulation of potential N-cycling processes. Microorganisms~~microorganisms~~ may therefore~~need to~~ reallocate enzymatic investment between~~to maintain~~ N acquisition and N transformation in response to changing resource availability and redox conditions (Li et al., 2025). Thus,~~Therefore, the first mechanistic consequence of~~ alpine wetland degradation primarily restructured~~is the restructuring of~~ the environmental context in which soil N-cycling operates, rather than uniformly~~a uniform~~ weakening ~~of~~ all microbial functions.

4.2 Differential responses of soil N-cycling enzymes to alpine wetland degradation

Against this altered environmental background, the four soil N-cycling enzymes did not respond synchronously. Instead, wetland degradation promoted a clear divergence among enzymatic pathways, indicating that soil N cycling was reorganized rather than uniformly suppressed (**Figure 2~~1~~**). Urease and nitrate reductase activities increased with degradation, whereas protease and nitrite reductase activities declined. The increase in urease activity can be interpreted as a shift toward faster access to relatively simple N compounds under resource-constrained conditions (Burns et al., 2013). As soil drying and nutrient depletion progress, microorganisms may rely more heavily on enzymatic routes that rapidly release inorganic N from accessible substrates, especially when the decomposition of complex organic matter becomes increasingly constrained (Geisseler et al., 2010). As the enzyme that directly catalyzes the hydrolysis of urea to ammonium(Krajewska, 2009), urease is closely linked to substrate turnover, pH conditions, and soil aeration (Pan et al., 2020; Wang et al., 2023). In contrast, the decline in protease activity indicates a reduced capacity to depolymerize more complex organic N pools, which is consistent with lower substrate supply, reduced microbial activity, and poorer diffusion conditions in degraded soils(Song et al., 2014; Vranova et al., 2013). At the same time, declines in SOC, TN, NH_4^+-N, and MBN suggest progressive depletion of both substrate resources and microbial metabolic capacity (Greenfield et al., 2021), thereby reducing microbial investment in the acquisition of complex organic N (Brockett et al., 2011; Chen et al., 2018; Ma et al., 2021; Vranova et al., 2013)~~(Brockett et al., 2011; Chen et al., 2018; Ma et al., 2021; Vranova et al., 2013)~~. This interpretation is consistent with findings from studies of alpine wetland degradation in Zoige (Huo et al., 2013). Thus, within the hydrolytic component of soil N cycling, degradation appears to shift function away from complex organic N acquisition and toward rapid-turnover pathways.

A similar internal divergence occurred within the reductive component of N cycling. Nitrate reductase activity increased, whereas nitrite reductase activity declined sharply with degradation (**Figure 2~~1~~**). This decoupling is consistent with a drying-induced shift in soil redox status: lower water tables and greater aeration can stimulate nitrification and nitrate accumulation

(Szajdak and Gaca, 2010; Li et al., 2015), thereby supporting the initial step of nitrate reduction while simultaneously suppressing the anaerobic microsites required for sustained downstream nitrite reduction (Hefting et al., 2004). In other words, degradation appears to create a functional bottleneck in the N-reduction pathway, in which upstream substrate supply is maintained or enhanced (Ayiti and Babalola, 2022) whereas downstream oxygen-sensitive reduction is constrained (Jin et al., 2019) This interpretation is also supported by the observed increase in NO_3^--N and by the stronger association of nitrate reductase activity with TEM and NO_3^--N in the RDA results. Therefore, wetland degradation did not simply weaken soil N cycling overall~~,~~ but reorganized it by enhancing some rapid-turnover processes while suppressing enzymes associated with complex organic N depolymerization and oxygen-sensitive reduction steps.

It should be emphasized that the enzyme activities measured here represent potential enzymatic capacities rather than realized in situ N-transformation rates. Standard soil enzyme assays quantify activity under controlled substrate and incubation conditions, whereas field expression is additionally regulated by substrate availability, moisture, temperature, diffusion, and redox conditions (Burns, 2002; German et al., 2011; Wallenstein and Weintraub, 2008). Despite this distinction, potential enzyme activities are often related to realized N-transformation processes. Soil protease and other hydrolytic enzyme activities have been associated with organic-N solubilization and gross N mineralization rates (Darby et al., 2020), and coordinated changes in enzyme activities and net N mineralization and nitrification have also been reported in wetland soils (Jia et al., 2020). Conversely, studies of wetland denitrification enzyme activity emphasize that potential assays are most appropriately used to compare relative functional capacities among sites or treatments rather than as direct measurements of in-situ denitrification (Hartzog et al., 2017). Consistent with these observations, an independent three-year in situ incubation study conducted in the same Gahai wetland showed degradation-induced decreases in net ammonification and mineralization but an increase in net nitrification, and further identified N-cycling enzyme activities as important predictors of field N-transformation dynamics (Du et al., 2025). These findings indicate that potential enzyme activities can serve as informative indicators of the biochemical capacity for soil N transformations, although the strength and direction of enzyme–process relationships depend on environmental context and the specific N pathway considered.

4.3 Hydroclimatic variability modulated the magnitude of degradation-induced functional divergence

Our multi-year analysis from 2019 to 2021 revealed pronounced seasonal and interannual patterns in potential soil N-cycling enzyme activities (**Figure 3~~2~~**). At the seasonal scale, urease, protease, and nitrite reductase, reached peak activity during the middle of the growing season, particularly in July and August, whereas nitrate reductase activity showed a delayed peak in September. Such temporal separation likely reflects the combined effects of plant phenology, substrate availability, and seasonal hydrothermal conditions. The midsummer maxima were consistent with the period of relatively favourable~~favorable~~ environmental conditions on the QTP, when higher temperatures, adequate soil moisture, and intensified plant growth jointly stimulate microbial metabolism and enzyme production (Kivlin and Treseder, 2014). In particular, July and August correspond to the peak period of plant vegetative growth, during which enhanced root exudation supplies labile C to the rhizosphere (Brzostek et al., 2011).~~In particular, July and August correspond to the peak period of plant vegetative growth, during which enhanced root exudation supplies labile C to the rhizosphere,~~ thereby stimulating microbial activity and promoting enzyme

synthesis (Brzostek et al., 2011; Meier et al., 2017). The interpretation is also supported by independent field measurements from the same wetland, where net ammonification and net N mineralization generally reached their highest rates between July and August, broadly coinciding with the seasonal maxima in hydrolytic enzyme potential (Du et al., 2025).~~.~~ By contrast, the delayed peak in nitrate reductase activity in September likely reflected a seasonal shift in plant–microbe competition for N. As plant senescence progresses toward the end of the growing season, plant N uptake declines, allowing residual nitrate (NO_3^-) to accumulate in soil (Hefting et al., 2004; Keiluweit et al., 2017). The~~This~~ substrate accumulation, together with increased soil moisture and the formation of localized anaerobic microsites under cooler autumn conditions, may create a favourable~~favorable~~ microenvironment for nitrate reductase induction (Kotroczó et al., 2014). The later seasonal peak of nitrate reductase relative to nitrite reductase is particularly important because it indicates that even the reductive components of N-cycling were internally asynchronous rather than operating as a single coordinated pathway. Accordingly, the distinct seasonal maxima of these enzymes are better interpreted as evidence of seasonal functional partitioning among N-cycling pathways, jointly driven by vegetation phenology, substrate dynamics, and pathway-specific sensitivities to hydrothermal and redox conditions~~These seasonal differences highlight the complexity of soil N-cycling enzyme regulation under wetland degradation and indicate that individual enzymes respond to distinct substrate and microenvironmental controls~~.

Interannual hydroclimatic variability significantly modulated the effects of wetland degradation on potential soil N-cycling enzyme activities, but this modulation was strongly enzyme-specific rather than indicative of a consistent drought-amplification effect. Significant degradation × year interactions for urease, nitrate reductase, and nitrite reductase demonstrated that the magnitude of degradation effects varied among years (**Table 2**). Although urease and nitrate reductase generally increased with degradation, whereas protease and nitrite reductase declined, their interannual responses were asynchronous. Degradation-related differentiation in urease was strongest in the exceptionally wet year of 2021; nitrate reductase increased significantly in heavily degraded wetlands in 2019 and in the relatively dry year of 2020, but this degradation gradient disappeared in 2021 (**Figure S5**). In contrast, the decline in nitrite reductase was most pronounced in 2019, whereas the negative response of protease was strongest in 2020. These contrasting patterns suggest that changes in water availability may impose different constraints on individual N-transformation pathways by altering soil aeration, redox conditions, substrate diffusion, resource availability, and microbial N retention, rather than uniformly intensifying all degradation effects during dry years ~~More importantly, the effects of wetland degradation on soil N-cycling enzymes were not static, but were strongly modulated by interannual hydroclimatic variation, particularly under drought conditions. The marked differences in annual precipitation during the study period appeared to determine the extent to which structural degradation was translated into functional divergence. Urease and nitrate reductase activities generally showed positive responses to degradation relative to ND, indicating that these two enzymes remained consistently higher in degraded wetlands than in non-degraded wetlands, with the contrast being especially pronounced in MD and HD during the dry year of 2020 (**Figure S2**). This pattern suggests that drought may amplify the relative advantage of rapid N hydrolysis and the initial step of nitrate reduction in degraded wetlands, likely because degradation-enhanced aeration, stronger redox differentiation, and altered N transformation pathways become more evident under water deficit~~ (Asensio et al., 2024; Steinweg et al., 2012). Accordingly, future climate change is more

likely to influence alpine wetland N cycling through pathway-specific and potentially nonlinear interactions between degradation and hydroclimatic variability. In this context, maintaining wetland hydrological integrity may be critical for reducing functional instability in N cycling and sustaining the ecological resilience of alpine wetlands.

4.4 Effects of functional decoupling on soil N-cycling

The NCFP index revealed a clear reorganization of the relative balance between hydrolytic and reductive N-cycling enzyme potentials along the wetland degradation gradient (**Figure 4**). NCFP progressively declined from ND to HD, shifting from slightly positive values in ND and LD to negative values in MD and HD. This pattern indicates a gradual shift in N-cycling enzyme function from a relative bias toward hydrolytic potential to a relative bias toward reductive potential, reflecting an adaptive reorganization of microbial resource allocation as habitat degradation progresses. Urease and protease mediate the hydrolysis and depolymerization of organic N substrates such as urea and proteins and therefore represent important components of organic N mineralization and microbial N acquisition (Marx et al., 2005). Accordingly, the relatively high NCFP values observed in ND and LD may reflect the greater availability of organic matter and microbial substrates in less-degraded wetlands, which could sustain a relatively strong potential for organic N hydrolysis. According to microbial resource allocation theory, extracellular enzyme production entails substantial energetic and nutrient costs (Sinsabaugh et al., 2009; Sinsabaugh and Moorhead, 1994), and microbial investment in enzyme synthesis is regulated by both substrate availability and cellular stoichiometric demand (Sinsabaugh and Follstad Shah, 2012). In contrast, the pronounced declines in soil moisture and nutrient availability associated with wetland degradation may alter substrate supply, microbial metabolic requirements, and the relative importance of different enzyme-mediated N transformations ~~. In contrast, protease and nitrite reductase activities showed consistently negative responses across the three years, indicating that degraded wetlands persistently exhibited lower capacities for complex organic N depolymerization and downstream nitrite reduction than ND wetlands (Priemé et al., 2002). Notably, the negative response of protease activity was strongest in 2020 but became weaker under the wetter conditions of 2021, suggesting that this enzyme was particularly sensitive to soil moisture and resource availability. Drought likely intensified degradation induced constraints on substrate diffusion, microbial activity, SOC availability, and MBN (**Figure 5**). By comparison, the negative response of nitrite reductase activity was more directionally stable across years, despite variation in magnitude, implying that this process was intrinsically more sensitive to degradation and could not readily recover even under wetter conditions (Giles et al., 2012). Together, these findings suggest that degradation created a vulnerable soil environment, whereas drought intensified the same constraints that pushed the system toward greater functional divergence. This hydroclimatic dependence also helps explain why enzyme responses should not be interpreted as fixed, trait-like outcomes of degradation stage alone. In wetter years or wetter periods of the growing season, partial rewetting may buffer some differences among degradation stages by improving substrate diffusion and microbial activity. However, this buffering should not be mistaken for functional recovery. Rather, it indicates that the expression of degradation effects depends strongly on the hydrological background. The key implication is that future climatic drying is likely to increase, rather than reduce, the ecological importance of wetland degradation for soil N cycling on the QTP.~~

~~**4.4 Functional decoupling of soil N cycling and its implications for wetland stability and restoration**~~

~~The progressive decline in the nitrogen-cycling functional balance (NCFB) index provides an integrative summary of these enzyme-specific responses. As degradation intensified, the NCFB index shifted from positive values in non-degraded wetlands toward near-zero or negative values in degraded wetlands (**Figure 3**), indicating that the relative coordination between hydrolytic and reductive functions was progressively weakened. This interpretation is consistent with the broader ecoenzymatic framework, in which the relative activities of enzymes involved in nutrient acquisition and transformation reflect shifts in microbial resource allocation and coupled biogeochemical functioning (Luo et al., 2017; Moorhead et al., 2023). However, this shift should not be interpreted as a uniform weakening of all hydrolytic processes or a uniform strengthening of all reductive processes. Rather, it reflects pathway-specific reallocation, whereby increased urease and nitrate reductase activities were accompanied by declines in protease and nitrite reductase activities. Such reallocation is consistent with the concept of microbial economic allocation, whereby microorganisms adjust enzymatic investment according to resource demand, substrate accessibility, and environmental constraints~~ (Malik et al., 2019). The stage-dependent changes in the relationship between hydrolytic and reductive functions further support this interpretation (**Figure S7**), suggesting that wetland degradation does not uniformly enhance or suppress all N-cycling processes (Fujita et al., 2018), but instead modifies the coupling among different functional components.

More importantly, strong interannual hydrological variability further modulated the expression of this degradation effect (**Figure S8**). In the relatively dry year of 2020, ND and LD generally maintained higher NCFP values, whereas in the exceptionally wet year of 2021, NCFP became negative across all degradation stages. This pattern indicates that the hydrolytic-to-reductive functional balance is sensitive not only to degradation intensity but also to short-term hydrological variability. Soil moisture is a major regulator of microbial N transformations because it influences substrate diffusion, microbial activity, and soil aeration, whereas water saturation constrains O_2 diffusion and promotes the development of anoxic or reducing microsites (Alldred and Baines, 2016). Notably, NCFP in ND also shifted from positive values in normal or relatively dry years to negative values in the wet year, indicating that degradation and interannual hydrological variability may influence the hydrolytic–reductive functional balance through different mechanisms. Long-term degradation likely restructures N-cycling functions through persistent changes in soil organic matter, inorganic N availability, microbial resources, and soil moisture, whereas anomalously high precipitation may rapidly modify soil O_2 diffusion and redox conditions, thereby reconfiguring the relative expression of hydrolytic and reductive enzyme potentials. Thus, NCFP reflects a dynamic balance in soil N-cycling functional potential jointly shaped by long-term ecosystem degradation and short-term hydrological variability.

~~. Thus, the decline in NCFB indicates that the internal balance of enzyme-mediated soil N cycling became increasingly skewed under degraded conditions.~~

~~This interpretation is further supported by the stage-dependent changes in the relationship between hydrolytic and reductive functions (**Figure S4**). The association between these two functional groups shifted in both strength and direction along the degradation gradient, suggesting that wetland degradation altered not only enzyme activity levels but also the coupling structure among major N-cycling pathways. In particular, the strongest negative relationship occurred in lightly degraded wetlands, indicating that mild degradation may generate a pronounced functional mismatch between hydrolytic and reductive~~

~~processes. This mismatch may arise because early-stage degradation differentially affects substrate availability, soil moisture, and microbial allocation strategies before the system shifts toward a more uniformly constrained state under severe degradation (Song et al., 2024). By the heavily degraded stage, soil N cycling no longer appeared to be organized around a relatively balanced internal turnover regime; instead, its functional configuration became increasingly skewed toward specific stress-responsive pathways. Given that alpine wetland degradation can induce soil nutrient deficiencies, alter stoichiometric balance, and reshape microbial diversity and function, such stage-dependent decoupling is ecologically plausible and provides a mechanistic explanation for the observed decline in NCFB.~~

~~From a land degradation and restoration perspective, these findings suggest that the principal consequence of alpine wetland degradation is not merely a reduction in belowground biological activity, but a reorganization of soil N cycling functions toward a less balanced and potentially less stable state. Restoration efforts should therefore focus not only on aboveground vegetation recovery but also on re-establishing hydrological conditions that support substrate diffusion, microbial biomass maintenance, and the internal coordination of N cycling.~~

4.5 Limitations of the Study

Although this study provides valuable insights into the effects of alpine wetland degradation on soil N cycling, several limitations should be considered when interpreting our results. First~~First, the study was conducted within a specific landscape on the northeastern QTP. Although this region is representative of alpine wetlands, the observed patterns may not fully capture the variability of alpine wetland ecosystems across different geographical and climatic conditions. Second~~, the enzyme activities measured in this study represent potential activities under standardized laboratory conditions rather than direct estimates of in situ transformation rates. All four enzymes were assayed under controlled incubation conditions with specific substrates, which facilitated comparisons among degradation stages but reduced the influence of field constraints such as substrate diffusion, temperature fluctuations, substrate limitation, oxygen availability, and soil redox conditions. This distinction is particularly important for nitrate reductase and nitrite reductase. Second~~, who's in situ activities are strongly regulated by oxygen availability, redox status, and the formation of anaerobic microsites. Finally~~, although we examined key soil physicochemical properties and plant nutrient characteristics, the role of microbial communities in mediating nutrient cycling was not directly assessed. Because soil microorganisms play central roles in C, N, and P~~nutrient~~ transformations, their responses to alpine wetland degradation~~incorporating microbial community and functional analyses~~ could further clarify the mechanisms underlying plant–soil interactions. Third, the lack of measurements of labile and biologically relevant carbon pools, such as dissolved organic carbon, particulate organic carbon, and microbial biomass carbon, limits our ability to determine how carbon availability regulates N-cycling functional potential~~ during alpine wetland degradation~~. Addressing these limitations in future studies will improve our understanding of N cycling in degraded alpine wetlands and provide a stronger scientific basis for developing~~the development of~~ effective wetland restoration strategies.

5. Conclusions

Alpine wetland degradation did not uniformly suppress soil N-~~nitrogen~~ cycling but~~;~~ instead the relative balance between hydrolytic and reductive soil N-~~, it reorganized soil N~~ cycling ~~by differentially affecting~~ enzyme potentials.~~ mediated transformation pathways.~~ With increasing degradation intensity, s urease and nitrate reductase activities increased, whereas protease and nitrite reductase activities declined. Despite these enzyme-specific responses, the nitrogen cycling~~soil N cycling~~ functional potential (NCFP) index showed an overall decreasing trend, indicating~~shifted from~~ a progressive shift ~~relatively coordinated state~~ in the relative~~non degraded wetlands toward a functionally decoupled~~ configuration of hydrolytic and reductive functions.~~, characterized by enhanced rapid turnover processes but weakened complex organic N depolymerization and downstream oxygen sensitive reduction.~~ This functional reorganization was primarily associated with~~linked to~~ degradation-induced hydrological drying and resource depletion, as declines~~. Declines~~ in soil water ~~content and carbon and nitrogen~~ availability and carbon resources altered the environmental context for microbial resource allocation.~~biochemical environment in which microorganisms allocated enzymatic investment, thereby favoring some N transformation pathways while constraining others.~~ Interannual hydroclimatic variability further modified~~modulated~~ this pattern~~process~~, with exceptionally wet~~drier~~ conditions shifting NCFP toward negative values across all degradation stages, suggesting~~amplifying the functional divergence between degraded and non degraded wetlands. These findings suggest~~ that the hydrolytic–reductive balance is jointly regulated by long-term ecosystem degradation and short-term hydrological fluctuations. Therefore, the ecological consequences~~consequence~~ of alpine wetland degradation extend beyond reductions in soil moisture and nutrient availability to include a restructuring of~~is not merely a reduction in soil biochemical activity, but a shift toward a less balanced and potentially less stable~~ belowground N-cycling functional balance and its sensitivity to hydrological variability.~~system.~~ Restoration of degraded alpine wetlands should thus the recovery of wetland hydrological conditions and soil resource availability to promote a more balanced configuration of N-cycling functions~~therefore prioritize hydrological recovery and microbial functional capacity, rather than focusing solely on vegetation cover or nutrient status~~.

Data availability

The data used in this paper can be accessed via the Zenodo: https://zenodo.org/records/20739760 (Chang et al., 2026).

Author contributions

Conceptualization, Methodology, Data curation, Software, Formal analysis, Writing - original draft: WC. Supervision, Project administration, Funding acquisition, Writing - review & editing: WM. Writing - review & editing: JW. Investigation: LS. Methodology: JD. Investigation: WH. Resources, Writing - review: GL.

Competing interests

The contact author has declared that none of the authors has any competing interests.

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