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Seasonal stratification regulates carbon allocation between particulate and dissolved pathways in the Gulf of Aqaba

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Abstract

Water column stratification exerts fundamental control on microbial carbon cycling in oligotrophic areas of the ocean, yet its impact on the partitioning and fate of newly fixed carbon remains insufficiently resolved. Here, we investigated carbon fluxes in the Northern Red Sea, Gulf of Aqaba from 5 cruises conducted during the stratified period. We measured ^{14}C-based measurements of primary production partitioned into particulate (>0.7 µm; PP_{POC}) and dissolved (<0.7 µm; PP_{DOC}) fractions, bacterial production (BP), community and bacterial respiration across the euphotic zone (0-100 m). As stratification intensifies and nutrient supply from depth diminishes, depth-integrated PP_{POC} declined from 1.26 to 0.35 g C m^{-2} d^{-1} while the relative contribution of dissolved carbon pathways increases. The fraction of newly fixed carbon released as dissolved organic carbon (extracellular release; PER) increased from 2.5% to >7% of total PP, indicating that a larger fraction of photosynthetically fixed carbon was released into the dissolved C pool. PP_{DOC} (0.02-0.03 g C m^{-2} d^{-1}) was positively correlated with BP (0.08-0.16 g C m^{-2} d^{-1}), suggesting that recently released dissolved substrates contribute to sustaining heterotrophic microbial activity. Despite declining primary production, BR remained substantial (0.23-0.52 g C m^{-2} d^{-1}), resulting in low to moderate bacterial growth efficiency (13-35%) and indicating that most processed carbon was respired rather than incorporated into biomass. These findings indicate that summer stratification enhances the relative importance of dissolved carbon release and microbial recycling, thereby reducing the efficiency of carbon transfer to depth in the Gulf of Aqaba and likely other oligotrophic systems.

Key words

Microbial carbon cycling; Oligotrophic seas; Primary production; Bacterial growth efficiency (BGE); Extracellular carbon release (PER).

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1. Introduction

Much of the open ocean is characterized by chronic oligotrophy, which shapes the pathways and efficiency of carbon transformation in surface waters (Marañón et al., 2003; Polovina et al., 2008). Under such nutrient-depleted conditions, primary production (PP) tends to be low/negligible and is predominantly supported by small-celled phytoplankton that can cope persistent resource limitation (Dore et al., 2008; Pennington et al., 2006; Reich et al., 2022). Under such conditions, newly fixed carbon is largely retained and remineralized within the microbial loop, where strong coupling between phytoplankton and bacteria controls its subsequent fate (Azam and Malfatti, 2007; Zhang et al., 2025). As a result, the efficiency with which carbon is transferred to higher trophic levels or exported to depth is often low (Armengol et al., 2019; Alkalay et al., 2020; Roshan and DeVries, 2017; Torfstein et al., 2020).

The onset and relaxation of stratification largely determine how nutrients are delivered to surface waters and thus govern microbial dynamics in oligotrophic environments (Dave and Lozier, 2010; Reich et al., 2026; Signorini et al., 2015). With strengthening stratification, the exchange between deep, nutrient-rich waters and the surface diminishes, causing nutrients in the euphotic zone to become progressively depleted (Chen et al., 2021; Hazan et al., 2018; Lozier et al., 2011). This imposes strong physiological constraints on phytoplankton growth and activity due to limited nutrient availability (Rahav et al., 2016; Zohary et al., 2005). Under these conditions, a considerable portion of newly produced organic carbon can be exuded by phytoplankton as dissolved material via extracellular release (Kang et al., 2022; Morana et al., 2014; Thornton, 2014). The resulting dissolved organic carbon provides a substrate for heterotrophic bacteria and forms an important conduit between autotrophic production and heterotrophic metabolism. This recently fixed dissolved carbon can be reused and account for a substantial proportion of total carbon fixation under nutrient-limited conditions (abovementioned references).

The fate of dissolved organic carbon in the ocean is largely determined by heterotrophic bacteria, whose metabolic activity often controls whether carbon is retained within the euphotic layer or transferred to depth (Baetge et al., 2021; LaBrie et al., 2022; Mentges et al., 2019). Bacterial carbon uptake and respiration together define the efficiency of bacterial organic carbon utilization, typically expressed as bacterial growth efficiency, BGE (del Giorgio and Cole, 1998). Oligotrophic waters commonly exhibit low BGE, indicating that much of the assimilated carbon is respired rather than incorporated into cellular biomass (Alothman et al., 2025; Anderson and Turley, 2003; del Giorgio et al., 1997). Such conditions promote fast

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carbon cycling within the surface ocean and further bias the system toward recycling rather than export (del Giorgio and Cole, 1998). The relative magnitudes of PP and community respiration (CR) therefore serve as a useful indicator of the system's metabolic status and offers insight into whether the system is characterized by net carbon accumulation or rapid remineralization (Izett et al., 2024; Stanley et al., 2010).

The Gulf of Aqaba (northern Red Sea) is an oligotrophic basin whose physical and biogeochemical properties are highly affected by seasonal and interannual variability (Laiolo et al., 2014; Rahav et al., 2015; Reich et al., 2024). During winter, deep convective mixing injects nutrients into the surface layer, often leading to increased phytoplankton biomass and productivity (Lindell and Post, 1995; Avrahami et al., 2025). Differently, in summer, a strongly stratified water column results in nutrient depletion (Meder et al., 2012) and subsequently to low phytoplankton biomass and activity (Lindell and Post, 1995; Reich et al., 2024). Despite extensive characterization of the physical and chemical seasonal cycles in this system (Biton and Gildor, 2011), the corresponding shifts in microbial carbon processing, particularly the partitioning of the carbon fixed during the stratified season between particulate matter, dissolved carbon released from cells, and heterotrophic utilization of this carbon, remain insufficiently resolved. While the dominance of microbial recycling in oligotrophic systems has long been recognized (e.g., low *f-ratios*, Alothman et al., 2025), the explicit partitioning of newly fixed carbon between particulate and dissolved pathways, and its direct linkage to heterotrophic metabolism across seasonal stratification, remains poorly constrained.

Here, we investigate microbial carbon cycling during the summer stratified period in the Gulf of Aqaba, focusing on how progressive nutrient depletion influences the balance between carbon fixation, release, and remineralization. To this end, we combined measurements of primary production, dissolved organic carbon release, and heterotrophic bacterial activity from 5 monthly cruises, to assess how the structure and efficiency of carbon processing evolve over the course of the stratified season.

2. Material and methods

2.1. Study site

Seawater was collected across the euphotic zone (0-100 m) at 20 m depth intervals at an offshore station in the northern Gulf of Aqaba, Red Sea (29.47°N, 34.92°E). Sampling was conducted monthly from May to September 2023, spanning the transition from late spring to

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late summer. Measurements included a comprehensive set of microbial activity measurements, including particulate primary productivity (>0.7 µm PP_{TOT}), dissolved primary production (PP_{DOC}, <0.7 µm), bacterial production (BP), and community respiration (CR). Ancillary data comprised CTD profiles (Sea-Bird SBE 19plus), inorganic nutrients concentration, and chlorophyll *a* measurements.

2.2. Particulate and dissolved primary production and extracellular release

Autotrophic activity was assessed by measuring particulate primary productivity (PP_{POC}) and carbon fixed present in the dissolved fraction (PP_{DOC}), enabling a detailed partitioning of carbon flow pathways within the microbial food web. To this end, seawater was collected in triplicate 50 ml acid-washed Falcon tubes. The collected samples were spiked with $NaH^{14}CO_3$ (Perkin Elmer, specific activity 56 mCi $mmol^{-1}$) at a final radioisotope dilution of $1:10^4$ *v:v* (1 µCi/10 ml) and incubated for 24 h under ambient temperature and light. Incubations were stopped by filtration of the ^{14}C-enriched water samples through GF/F filters (0.7 µm nominal pore size, PP_{POC}) using low vacuum pressure (<50 mmHg). Filtrate material samples (5-10 ml, PP_{DOC}) were also collected in 15 ml Faclon tubes. Filters were rinsed with 5 ml of sterile-filtered seawater collected from the same site (<0.2 µm) and the samples were subsequently acidified with 50 µl of 37% hydrochloric acid to remove inorganic carbon overnight. Filters were transferred into glass vials and 5 ml of scintillation cocktail (Ultima Gold) was added. To determine PP_{DOC}, the filtrate (10 ml) was transferred to scintillation vials, acidified with 50 µl hydrochloric acid and 5ml scintillation cocktail was added as above. Samples were counted using a TRI-CARB 4810 TR (Packard) liquid scintillation counter. Blank samples (for both PP_{POC} and PP_{DOC}) collected from each depth were spiked with $NaH^{14}CO_3$ and filtered immediately without incubation. These blanks yielded negligible counts, and their reads were subtracted from their respective samples read. Added activity of the radiolabeled 'working solution' was obtained in each campaign by removing 50 µl from random samples immediately after spiking and before incubation (usually one per sampling depth), placing it on a new GF/F filter, adding 50 µl ethanolamine (a strong base that prevent bicarbonate evaporation) and scintillation liquid, and counting immediately. The variability between the 'added activity' measurements was usually negligible for the same work solution (prepared freshly before each campaign), and we therefore used the average value of all collected samples/depths on each cruise.

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PP_{POC} and PP_{DOC} rates were calculated based on the Bermuda Atlantic Time-series Study (BATS) protocol (https://www.bco-dmo.org/dataset/893182/description#acquisition) using the equation:

$$(1)\ \text{Primary productivity} = \frac{(\text{DPM} - \text{blank})}{\text{V}} \times \text{DIC} \times \frac{\text{AA vol}}{\text{TDPM}} \times \text{f} \times \frac{1}{\text{t}}$$

Where, DPM equals the disintegrations per minute, V = the filtered volume, DIC is the dissolved inorganic carbon in seawater calculated from total alkalinity; ~25 µg C L^{-1}), AA vol = Added activity volume, TDPM = Total ^{14}C disintegration per minute, t = incubation time, and f = factor correcting the uneven uptake of ^{14}C due to fractionation (1.05).

The percentage of extracellular release (PER; %) was calculated as follows:

$$(2)\ \text{PER}\ (\%) = \frac{\text{PP\_DOC}}{\text{PP\_POC+PP\_DOC}} \text{x}\ 100$$

2.3. Bacterial productivity (BP)

Triplicate seawater samples (1.7 ml) from each sampling depth were incubated in the dark under ambient temperature with 10 nmol L^{-1} of ^{3}H-leucine (Perkin Elmer; specific activity 123 Ci $mmol^{-1}$) for 4 h at *in situ* temperature, following the protocol of Simon et al., (1990). Incubations were terminated by the addition of 100 µl of 100% trichloroacetic acid, and samples were subsequently processed using the microcentrifugation method (Smith and Azam, 1992). After processing, 1 ml of scintillation cocktail (Ultima Gold) was added to each vial, and radioactivity was measured using a TRI-CARB 4810 TR (Packard) liquid scintillation counter. Random 'killed' controls, containing ^{3}H-leucine and trichloroacetic acid added prior to incubation were used to account for background activity. Leucine incorporation rates were converted to carbon production using a conversion factor of 3 kg C mol^{-1} leucine, assuming an isotopic dilution factor of 2 (Simon and Azam, 1989).

2.4. Community and bacterial respiration

Community respiration (CR) rates were estimated from dark triplicate incubations per depth under ambient temperature. Dissolved O_2 concentrations were measured using an optical oxygen meter (FireSting, PyroScience GmbH) equipped with fiber-optic sensors. Prior to the incubations, optical sensor spots (OXSP5, PyroScience) were affixed to the inner wall of Winkler bottles (300 ml) according to the manufacturer's instructions. Bottles were carefully filled to avoid bubble formation and sealed with ground-glass stoppers. Samples were incubated for 24 h under continuous, gentle stirring using magnetic stir bars to ensure

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homogeneous O_2 distribution without introducing air bubbles or disturbing the sensor spot. The system was calibrated prior to measurements using a two-point calibration (air-saturated seawater and zero-oxygen solution) following the manufacturer's guidelines. CR was calculated as the temporal change in O_2 concentration (T_{24} minus T_0) normalized to the incubation duration (Cheung et al., 2024). The detection limit was 0.5 µmol O_2 L^{-1}. CR was converted into bacterial respiration (BR) using the linear regression of Aranguren-Gassis et al., (2012):

$$(3)\ BR = 0.3 \times CR^{1.22} - 0.013$$

A respiratory quotient of 1 was used to convert O_2 consumption into carbon respiration (del Giorgio and Cole, 1998). Bacterial gross efficiency (BGE) was calculated as follows (del Giorgio and Cole, 1998):

$$(4)\ BGE\ (\%) = \frac{BP}{BP + BR} \times 100$$

2.5. Inorganic nutrients

Samples were collected in acid-washed plastic Falcon tubes and kept frozen at -20 °C until analysis within a few months. One sample per depth was collected. Nitrate+nitrite (NO_2+NO_3), orthophosphate (PO_4) and $Si(OH)_4$ were measured with a Seal Analytical AA-3 system (Kress and Herut, 2001). The limits of detection (twice the standard deviation of the blank) were 0.03 µM for nitrate+ nitrite, 0.008 µM for orthophosphate, and 0.05 µM for silicic acid. Ammonium (NH_4) concentrations were measured fluorometrically using the orthophthaldialdehyde (OPA) method (Holms et al., 1999) using a Turner Designs (Trilogy) fluorometer equipped with 365-nm excitation and 460-nm emission filters. The detection limit was 0.01 µM. The quality of the nutrient measurements is regularly confirmed by inter-comparison exercises (e.g., QUASIMEME program).

2.6. Chlorophyll a

Seawater (300 ml) were filtered through GF/F and stored at -20 °C in a dark box. One sample per depth was collected. Samples were extracted overnight in 5 ml of 90% acetone at 4 °C in the dark (Welschmeyer, 1994). Chlorophyll concentrations were determined using a Turner Designs (Trilogy) fluorometer with 436-nm excitation and 680-nm emission filters.

2.7. Statistical analyses

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Pairwise relationships among environmental and biological variables were assessed using Pearson correlation analysis. All data points from the upper 0–100 m across the five sampling campaigns were included. Prior to analysis, data were screened for missing values, and only complete datasets were retained. Pearson correlation coefficients (r) and corresponding p-values were calculated to assess the strength and significance of pairwise relationships. Statistical significance was evaluated at two levels ($p<0.05$ and $p<0.01$). Correlation matrices were visualized using a lower-triangle format to improve readability, with color gradients representing correlation strength and overlaid values indicating correlation coefficients and their significance levels. Analyses and figure generation were performed using Python (version 3.13).

3. Results

Surface waters (0-20 m) were consistently warm throughout the study period (25-28 °C) and showed a clear seasonal increase in temperature from early to mid-summer (Figure 1A). The mixed layer depth (MLD), estimated from a temperature threshold criterion (ΔT = 0.2 °C from surface values, de Boyer Montégut et al., 2004), shoaled progressively from 45 m in early summer (May) to 15-20 m during peak stratification (July-August), before slightly deepening again in September (28 m). A pronounced vertical temperature gradient was evident throughout the sampling period, with temperatures decreasing to 23-24 °C at 100 m (Figure 1A). Inorganic nutrient concentrations were generally low throughout the upper water column and increased with depth across the 0-100 m layer (Figure 1B-D). Nitrate+nitrite (NO_3+NO_2) concentration ranged from below detection level (<0.03 µmol L^{-1}) to 0.14 µmol L^{-1} (Figure 1B), resulting in integrated values of 4.5-11.2 µmol m^{-2} (Table 1). NO_3+NO_2 concentrations were only slightly higher below the MLD (Figure 1B), indicating restricted nutrient supply from the nutricline to the upper euphotic zone during summertime. Ammonium (NH_4) concentrations were also low, ranging from 12 to 65 nmol L^{-1}, and increased with depth (Figure 1C). Depth-integrated NH_4 concentrations were higher during mid- to late summer (3.0-4.6 µmol m^{-2}) compared to early summer (1.7 µmol m^{-2}; Table 1). Orthophosphate (PO_4) was usually low throughout the water column and across all sampling periods; 0.01-0.03 µmol L^{-1}, with the exception of a single elevated surface value of 0.07 µmol L^{-1} observed during the September cruise (Figure 1D). The integrated PO_4 was low and ranged from 2.1-4.3 µmol m^{-2} (Table 1). The resulting N:P ratio was below the canonical Redfield ratio of 16:1 (Redfield, 1934) and ranged from 1.8:1 to 7.5:1 (Table 1), suggesting N limiting conditions for the microbial populations (Tanioka et al., 2022). Silicic acid ($Si(OH)_4$) concentrations were

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relatively homogeneous throughout the water column (Figure 1E). However, higher levels were observed in May-June (1 µmol L^{-1}, 75-89 µmol m^{-2}), followed by a decline during July-September (<0.6 µmol L^{-1}, 55 µmol m^{-2}) (Figure 1E, Table 1), suggesting a transition from a diatom-influenced system in spring/early summer to an oligotrophic, small-cell-dominated community under more stratified conditions (Avrahami et al., 2025).

Chlorophyll *a* exhibited a pronounced vertical structure across all sampling periods (Figure 2A). Where surface concentrations were typically low, ranging from 0.05 to 0.15 µg L^{-1}, and increased with depth to form a well-defined deep chlorophyll maximum (DCM). The DCM was consistently located between 60 and 100 m, coinciding with the lower euphotic zone. Chlorophyll concentrations at the DCM ranged from 0.3 to 0.7 µg L^{-1}, representing a several-fold increase relative to surface waters. Vertically integrated chlorophyll biomass (0-100 m) ranged from 16 to 28 mg m^{-2} across the study period (Table 1). Integrated biomass was lowest in early summer (May-June; 16-20 mg m^{-2}) and increased toward mid/late-summer (July-September; 25-28 mg m^{-2}).

Particulate primary production (PP_{POC}) exhibited a pronounced vertical gradient across all sampling periods, with highest rates observed in surface waters and a progressive decline with depth (Figure 2B). Surface PP_{POC} ranged from 25 µg C L^{-1} d^{-1} in early summer and decreased to 8 µg C L^{-1} d^{-1} during mid- to late summer (Figure 2B). Below 40-60 m, PP_{POC} declined substantially in all stations and ranged from 8 µg C L^{-1} d^{-1} in May to 1 µg C L^{-1} d^{-1} in June-September to (Figure 2B). Depth-integrated PP_{POC} ranged from 0.28 to 1.26 g C m^{-2} d^{-1} (Table 1), with the highest PP_{POC} values observed in early summer (May), followed by a marked decline toward mid-summer (June-August), and a slight increase in September. This seasonal decrease in integrated PP_{POC} coincided with the progressive shoaling of the mixed layer (Table 1, Figure 1A) and reduced nutrient availability in surface waters (Figure 1B-E), and is in agreement with previous observations in the area (Iluz et al., 2009; Reich et al., 2024). PP_{DOC} generally displayed its highest rates in the upper water column (Figure 2C). In June-September, PP_{DOC} was elevated in surface and near-surface waters, typically ranging from 0.2 to 0.5 µg C L^{-1} d^{-1}, and then declined progressively with depth. Below 80 m, rates were consistently lower than 0.10 µg C L^{-1} d^{-1}. In contrast, the May profile was uniform throughout the water column (0.4 µg C L^{-1} d^{-1}). Despite these differences in vertical distribution, depth-integrated PP_{DOC} varied only slightly across months, ranging from 0.02 to 0.03 g C m^{-2} d^{-1} (Table 1). The percentage of extracellular organic carbon release (PER) increased over the course of the summer and showed a clear depth dependence (Figure 2D). In May, PER was

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generally low and relatively uniform (<5%), consistent with the homogeneous distribution of PP_{DOC} (Figure 2C). In contrast, from June onward PER was lower in the upper water column (<10%) and increased with depth, with the highest values typically observed below 40-60 m, near the base of the euphotic zone reaching as high as 20%. Integrated PER rose from 2.5% in May to 5.7-6.4% during June-August and reached 7.4% in September (Table 1).

Heterotrophic prokaryotic (bacterial) production (BP) exhibited moderate vertical and temporal variability (Figure 3A). Where BP rates were generally highest in the upper water column (0-40 m), ranging from 0.8 to 2.5 µg C L^{-1} d^{-1}, and declined with depth. Below 60-80 m, BP decreased substantially, with values commonly <1 µg C L^{-1} d^{-1} near the base of the euphotic zone. Depth-integrated BP ranged from 0.08 to 0.16 g C m^{-2} d^{-1} across the study period, with the higher values observed during mid- to late summer (Table 1). Bacterial respiration (BR) was high and relatively homogeneous throughout the water column in May-June, with rates typically around 4-8 µg C L^{-1} d^{-1} (Figure 3B), summing to 0.4-0.5 g C m^{-2} d^{-1} over the upper 100 m (Table 1). Differently, during July-August, BR exhibited a stronger vertical variability with higher respiration rates recorded in the upper 40-60 m (4-5 µg C L^{-1} d^{-1}) and lower at the bottom of the euphotic layer around 0-40 m (1-2 µg C L^{-1} d^{-1}). In September, BR again showed relatively low vertical variability, with rates mostly between 1 and 4 µg C L^{-1} d^{-1}, corresponding to 0.25 g C m^{-2} d^{-1}.

The spatial and vertical distribution of bacterial growth efficiency (BGE, Figure 3C) broadly mirrored that of BR (Figure 3B) and was consistent with patterns observed for BP (Figure 3A). In all months, relatively higher BGE values were found in the upper 0-60 m, whereas lower values were typically observed at greater depths (Figure 3C). Temporally, BGE was lower during May-June, when values throughout the water column were typically 10-20% (integrated value 13%, Table 1), reflecting the relatively high and vertically homogeneous BR during this period (Figure 3C). In contrast, from July onward, BGE increased in the upper water column, commonly reaching 20-40% in the 0-60 m layer and 10-20% below 60 m (Figure 3C), summing 28-30% over the whole euphotic layer (Table 1). In September, BGE exhibited reduced vertical variability, with values generally ranging between 15 and 30% across the water column (Figure 3C).

Pearson correlation analysis revealed that depth and temperature were the dominant structuring variables, with depth negatively correlated with temperature ($r=-0.83$, $p<0.01$) and strongly positively correlated with chlorophyll *a* ($r=0.89$, $p<0.01$), reflecting the vertical

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separation between surface waters and the DCM (Figure 4). Depth was also negatively correlated with both PP_{POC} ($r=-0.54$, $p<0.01$) and PP_{DOC} ($r=-0.63$, $p<0.01$) (Figure 4), indicating that autotrophic activity was mainly concentrated in the upper euphotic zone (Figure 2B,C). As expected, PP_{POC} positively correlated with PP_{DOC} ($r=0.69$, $p<0.01$) (Figure 4), demonstrating that the release of dissolved organic carbon scales with total photosynthetic activity. At the same time, PP_{POC} showed a significant negative correlation with chlorophyll *a* ($r=-0.46$, $p<0.05$) (Figure 4), suggesting a decoupling between biomass accumulation and photosynthetic rates as found for chlorophyll *a* (Figure 2). Interestingly, PP_{DOC} exhibited several key relationships that highlight its central role in carbon cycling in the Gulf of Aqaba during summertime. For example, PP_{DOC} was positively correlated with PO_4 ($r=0.47$, $p<0.05$), and negatively correlated with chlorophyll *a* ($r=-0.58$, $p<0.01$) (Figure 4). These patterns indicate that PO_4 may be preferentially regenerated from following the DOC released from the cells. Furthermore, PP_{DOC} was positively correlated with BP ($r=0.50$, $p<0.01$) (Figure 4), supporting the role of PP_{DOC} as an important substrate fueling heterotrophic activity in the Gulf of Aqaba. In contrast, inorganic nitrogen species (e.g., NO_3+NO_2, NH_4) showed weak or non-significant relationships with PP_{POC} or PP_{DOC} (Figure 4), suggesting these N species are utilized as fast as they released from cells or that they are not released as much as carbon and orthophosphate.

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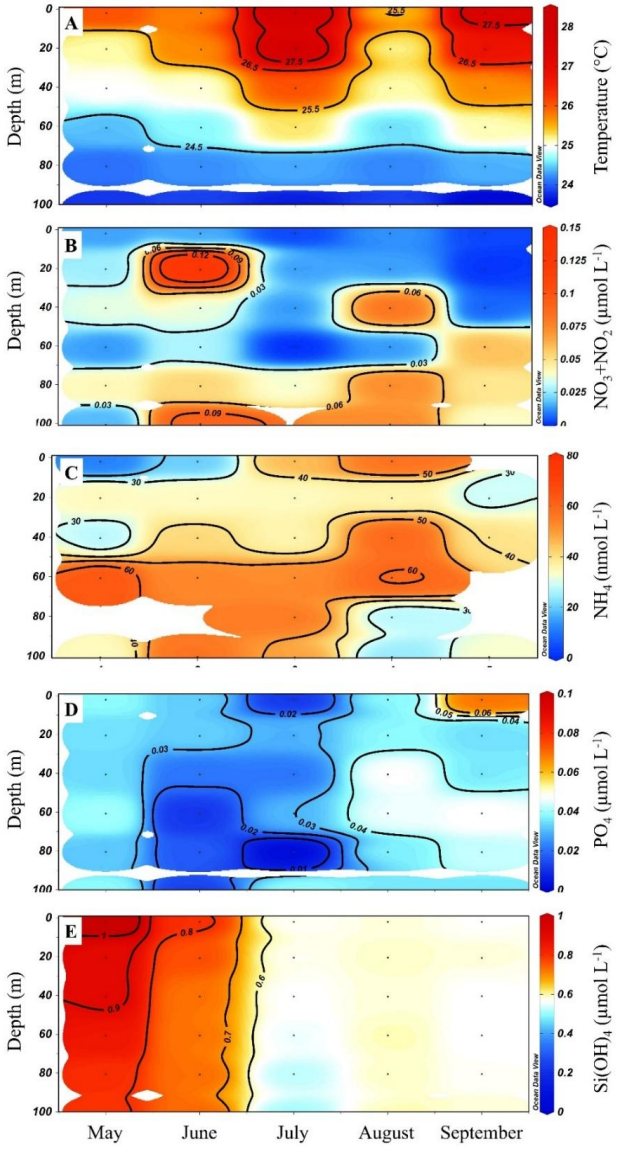

315

316 **Figure 1**: Depth distribution of temperature (A), NO_2+NO_3 (B), NH_4 (C), PO_4 (D) and $Si(OH)_4$
317 (E) in the upper water column (0-100 m) of the Gulf of Aqaba during summertime (May to
318 September).

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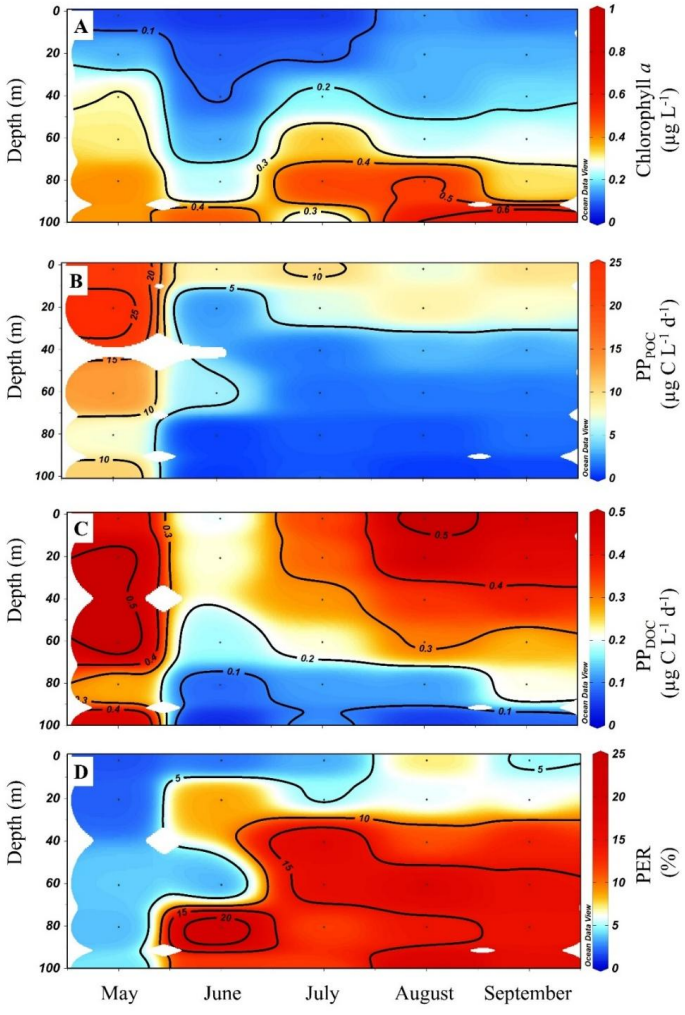

Figure 2: Depth distribution of chlorophyll *a* (A), PP_{POC} (B) PP_{DOC} (C) and the percentage of extracellular release, PER (D) in the upper water column (0-100 m) of the Gulf of Aqaba during summertime (May to September).

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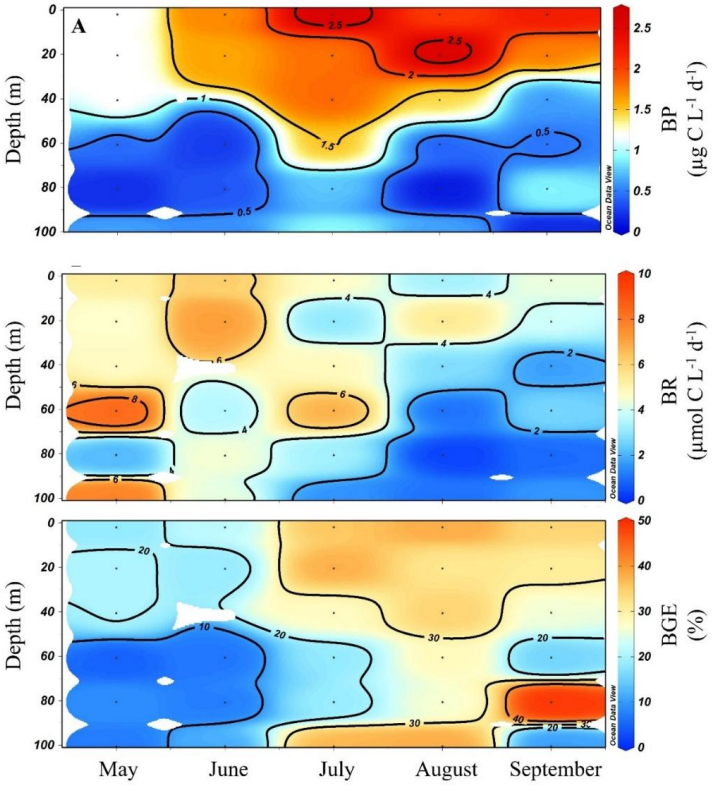

Figure 3: Depth distribution of bacterial productivity, BP (A), bacterial respiration, BR (B), and bacterial gross efficiency, BGE (C) in the upper water column (0-100 m) of the Gulf of Aqaba during summertime (May to September).

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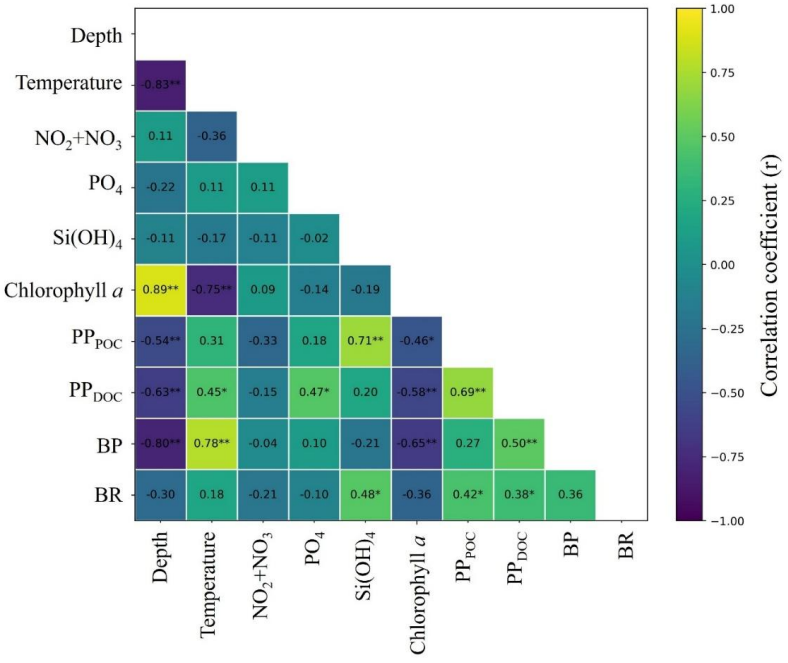

Figure 4: Pearson correlation matrix of physical, chemical, and biological variables measured in the upper 0–100 m of the water column. Colores indicate the strength and direction of the correlation (Pearson's r), and values within each cell represent correlation coefficients. Statistical significance is indicated as *p<0.05 and **p<0.01. The analysis is based on all depth-resolved measurements across the five cruises (May to September).

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Table 1: Summary of integrated values (0-100 m) measured in the Gulf of Aqaba during the summer.

	May	June	July	August	September
Mixed layer depth (m)	45	31	21	15	28
NO_3+NO_2 (µmol m^{-2})	4.5	11.2	7.5	7.8	6.0
NH_4 (µmol m^{-2})	1.7	4.5	4.6	3.4	3.0
PO_4 (µmol m^{-2})	3.5	2.1	2.1	4.1	4.3
DIN/PO_4 (ratio)*	1.8	7.5	5.8	2.7	2.1
$Si(OH)_4$ (µmol m^{-2})	89	75	55	59	57
PP_{POC} (g C m^{-2} d^{-1})	1.26	0.28	0.34	0.37	0.39
PP_{DOC} (g C m^{-2} d^{-1})	0.03	0.02	0.02	0.03	0.03
PER (%)	2.5	5.7	5.6	6.4	7.4
BP (g C m^{-2} d^{-1})	0.08	0.08	0.16	0.12	0.10
CR (g C m^{-2} d^{-1})	1.03	1.01	0.86	0.50	0.53
BR (g C m^{-2} d^{-1})	0.52	0.51	0.42	0.23	0.24
BGE (%)	13.5	13.1	27.7	34.1	29.8
PP_{TOT}/CR (ratio)**	1.26	0.30	0.42	0.78	0.78

* DIN = NH_4+NO_3+NO_2

** PP_{TOT} = PP_{POC}+PP_{DOC}

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4. Discussion

4.1. Summer stratification shifts carbon cycling toward dissolved pathways

The Gulf of Aqaba exhibits strong seasonal hydrographic variability with deep winter mixing followed by stable stratification afterward (Biton and Gildor, 2011), leading to nutrient-depleted surface water throughout the summer months (Laiolo et al., 2014; Meeder et al., 2012). The shift from deeply mixed winter conditions to a strongly stratified summer water column substantially reshapes autotrophic production, heterotrophic activity, and the efficiency with which carbon is exported from the surface (Karl et al., 2021). Understanding how microbial communities adjust their carbon-use strategies under nutrient-poor conditions is therefore important for predicting the fate of newly fixed carbon in stratified and oligotrophic oceans.

The progressive shoaling of the mixed layer from early to late summer (Table 1), together with persistently low nutrient concentrations and reduced nutrient inventories (Figure 1B-E), indicates increasing isolation of the euphotic zone from the nutricline, typically located at depths >150 m in this region (Landou et al., 2023; Meeder et al., 2012; Rahav et al., 2015). As stratification intensified, depth-integrated primary production declined from early to mid-summer, while chlorophyll biomass became increasingly concentrated within a pronounced DCM (Figure 2A). This decoupling between biomass and productivity is consistent with a community persisting under chronic low nutrient availability, maintaining low standing stocks but operating at reduced photosynthetic efficiency (Marañón et al., 2010). At the same time, the vertical distributions of PP_{DOC} and PER indicate a shift in carbon allocation within the water column (Figure 2). Where PP_{DOC} remains concentrated in the upper water column and is relatively stable over the season, while PER increases both with depth and over time, reaching its highest values under the strongest stratification (Figure 2). This pattern implies that an increasing fraction of newly fixed carbon was released as DOC in the upper 100 m, especially as summer progressed and as imbalances between carbon fixation and nutrient assimilation limited biomass synthesis and 'favored' the exudation of photosynthates (Mueller et al., 2016; Ofaim et al., 2021). Under such conditions, excess carbon leaks or excreted as low-molecular-weight compounds, thus elevating PER and enhancing the supply of labile dissolved organic substrates to heterotrophic prokaryotes (Engel et al., 2017, 2014, 2004). Consequently, a larger share of PP_{TOT} was routed into the microbial loop rather than retained in particulate autotrophic biomass (i.e., PP_{POC}), enhancing carbon recycling and reducing the efficiency of export to depth (Table 1; Carlson et al., 2010).

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The increasing release of newly fixed carbon as dissolved organic matter thus provides a key link between autotrophic production and sustained heterotrophic activity within the euphotic zone, even as primary production declines over the summer (Table 1, Figure 2). In oligotrophic systems such as the Gulf of Aqaba, heterotrophic microbial communities are often tightly coupled to DOC supply derived from contemporaneous primary production as well as semi-labile DOC that accumulates over seasonal timescales (Hansell et al., 2009; Santinelli, 2015). The persistence of BP in our study suggests that microbial metabolism is supported by a combination of freshly released DOC, recycled organic matter, and/or external nutrient inputs such as those added from aerosol deposition (Paytan et al., 2009; Rahav et al., 2018). Similar accumulated DOC and associated impacts on the microbial community have been observed in other stratified regions, where semi-labile DOC mitigates temporal imbalances between production and consumption (Hansell et al., 2009; Hansell and Carlson, 2002). In contrast, a recent study in the Eastern Tropical North Atlantic reported that BP and BR closely follow PP_{TOT} concentrations, which are dominated by semi-labile DOC, with PP_{DOC} supply often exceeded bacterial carbon demand (Devresse et al., 2022). There, heterotrophic activity was largely controlled by and coupled to contemporaneous production, resulting in a production-driven carbon cycle. In the Gulf of Aqaba, however, the weaker coupling between PP_{TOT} and heterotrophic activity points to a recycling-dominated regime in which microbial communities increasingly depend on internally regenerated carbon and are not directly linked to contemporaneous production. Under sustained stratification, reduced nutrient supply constrains autotrophic growth while promoting DOC release and accumulation, thereby maintaining BR even as primary production declines (Hansell et al., 2009; Hansell, 2013). The low BGE observed (Table 1, Figure 3C) further support this view, indicating that most of the consumed carbon is respired rather than converted into biomass, a characteristic of strongly recycling systems in oligotrophic ocean settings (del Giorgio and Cole, 1998). Indeed, the vertical structure of BGE closely mirrors that of BR, indicating that most of the organic carbon processed by heterotrophic prokaryotes was respired rather than incorporated into new biomass. These low growth efficiencies are consistent with the energetic constraints typical of oligotrophic waters, indicating that bacterial carbon use was directed mainly toward respiration rather than growth, with much of the organic carbon rapidly returned to dissolved inorganic form (del Giorgio and Cole, 1998).

4.2. Potential drivers of microbial carbon processing in the Gulf of Aqaba during summertime

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The observed vertical patterns in microbial carbon cycling are primarily structured by gradients in light availability/nutrient supply and stratification intensity, which all co-vary with depth (Figure 4). Where, chlorophyll *a* peaks at the DCM, reflecting enhanced biomass accumulation at depth, while rates of PP_{TOT}, PP_{DOC}, and BP are highest in the upper, well-lit layers. This pattern suggests that while biomass is concentrated near the DCM, much of the carbon fixation, release, and recycling occur in the upper euphotic zone. Within this framework, a strong coupling between PP_{POC} and PP_{DOC} would suggest that dissolved carbon release scales directly with photosynthetic activity, consistent with extracellular release as an inherent component of primary production under oligotrophic conditions. However, the negative relationship between both PP_{POC} and PP_{DOC} with chlorophyll *a* (Figure 4) indicates that carbon fixation and release are not directly linked to standing stock of biomass, but rather to physiological activity in the upper water column (Figure 2). Importantly, PP_{DOC} showed a stronger association with BP than PP_{POC} does, emphasizing the central role of dissolved organic carbon in sustaining heterotrophic metabolism. This supports the idea that the microbial loop in the Gulf of Aqaba is primarily fueled by recently produced dissolved substrates rather than by particulate pathways (abovementioned references). In contrast, the weak/no correlations between nutrients and biological rates, attributed by their rapid uptake, suggest that nutrient distributions are largely governed by vertical mixing rather than biological uptake.

Together, these patterns indicate that carbon cycling in the Gulf of Aqaba is structured by two complementary processes: (i) vertical decoupling between biomass accumulation and carbon cycling activity driven by stratification, and (ii) tight coupling between dissolved carbon production and heterotrophic utilization (Figure 4). That is in the surface layer fixed carbon fuels biomass activity of both fixation and respiration with no increase in biomass due to rapid recycled of the carbon while deeper in the euphotic zone at the DCM fixed carbon contributes to increase in biomass and particulate carbon sinking to depth. This duality reinforces the view of a recycling-dominated system in which dissolved pathways, rather than particulate export, regulate the fate of fixed carbon in the upper photic zone.

5. Conclusions

Our observations provide a quantitative and process-based framework for understanding how stratification restructures carbon flow, showing that the Gulf of Aqaba system shifts from particulate production toward sustained dissolved carbon release that maintains heterotrophic metabolism despite declining primary production. As climate-driven warming intensifies the density barriers of low- and mid-latitude pelagic systems (Duarte et

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al., 2013), nutrient recycling rates are expected to become more prevalent, potentially enhancing DOC turnover, thus lowering carbon export efficiency, and strengthening the role of the microbial loop in regulating air-sea CO_2 exchange. Thus, increased stratification may lead to longer residence times of organic carbon in the surface ocean (through multiple regeneration and utilization cycles), tighter microbial coupling to DOC, and an expansion of BR relative to BP, with implications for nutrient regeneration, and ecosystem structure, including shifts toward smaller phytoplankton and more efficient recyclers.

Future work should explicitly link physical forcing, nutrient inputs, and microbial carbon partitioning across seasonal to interannual scales in the Gulf of Aqaba and other oligotrophic basins. Long-term observations and targeted experiments that manipulate stratification, nutrient supply, and temperature will be essential to predict how microbial communities adjust DOC release, BGE, and BR under progressing oligotrophication. Integrating field measurements with trait-based and ecosystem models could help quantify how shifts in microbial carbon dynamics under enhanced stratification alter the efficiency of the biological carbon pump and feedback to climate, including potential interactions with aerosol inputs and emerging stressors such as acidification and deoxygenation.

Data Availability Statement

All the data is presented in the graphs/table/text and will be made available in excel format upon request.

Credit Authorship Contribution Statement

Conceptualized and conducted the field measurements; ER. Data curation, formal analysis, and visualization; ER, and AP. The paper was prepared by ER and AP.

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