



1 **Long-term throughfall exclusion reduced soil organic carbon towards higher soil**
2 **microbial carbon use efficiency and lower microbial enzyme activities on *Phyllostachys***
3 ***edulis* plantations**

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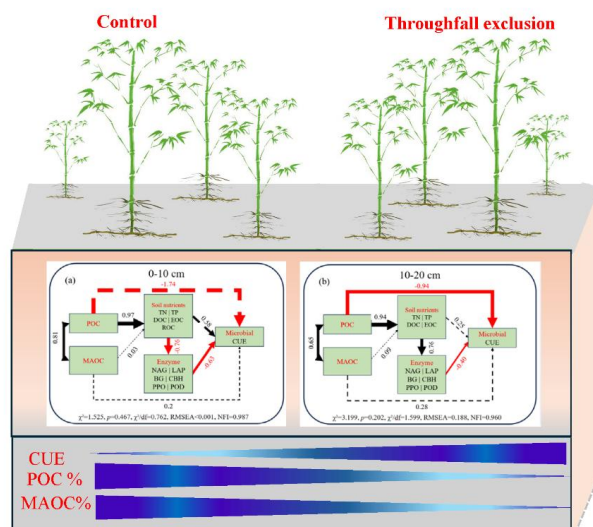
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23 **Abstract:** Soil microbial carbon utilization efficiency (CUE) serves as a crucial metric for
24 evaluating the effectiveness with which microbes assimilate organic carbon, acting as a vital
25 benchmark for assessing the potential of soil carbon sequestration. Previous studies have
26 shown that drought can significantly affect microbial CUE, which is crucial for the carbon
27 cycle in forest ecosystems. However, the mechanisms of microbial CUE on soil carbon
28 stability under drought conditions are not well understood. In this study, a throughfall
29 exclusion experiment was conducted in subtropical *Phyllostachys edulis* plantations (The
30 control, CK; throughfall exclusion, T). The results showed that drought increased microbial
31 CUE by 9.12% ($p < 0.05$) and 2.56% in 0-10 cm and 10-20 cm soil layers, respectively. Soil
32 organic carbon (SOC), soil particulate organic carbon (POC), soluble organic carbon (DOC),
33 and easily oxidized organic carbon (EOC) all significantly decreased, while the proportion of
34 mineral-associated organic carbon (MAOC) in SOC increased by 4.1% and 6.3% ($p < 0.05$) at
35 two layers, respectively. Microbial CUE was positively correlated with POC/SOC ratio and
36 MAOC/SOC ratio, indicating that variation of SOC components substrate quality was an
37 important factor driving microbial physiological changes. Structural equation model (SEM)
38 further showed that soil polyphenol oxidase (PPO) and cellobiohydrolase (CBH) enzymes
39 were the main factors driving changes in microbial CUE. Our results suggested that drought
40 indirectly regulates the storage and transformation of SOC by affecting microbial community
41 structure and function, which would have a profound impact on the carbon cycle of forest
42 ecosystems.

43 **Keywords:** Drought; Microbial CUE; SOC components; Soil enzyme activity; Moso
44 bamboo;



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46

47 1. Introduction

48 Rising global average air temperature and shifting precipitation patterns, both attributable to
 49 human-induced greenhouse gas emissions, will inevitably extend the drought season in most
 50 regions of the world (Peng et al., 2025). This prolongation of the drought season has a
 51 profound impact on various aspects, such as vegetation and composition and soil organic
 52 carbon (SOC), which ultimately affect the carbon cycling processes in terrestrial ecosystems
 53 (Miao et al., 2020; Spinoni et al., 2021). Soil represents the most extensive carbon pool
 54 within terrestrial ecosystems, playing a crucial role in the regulation of carbon cycle
 55 processes and the dynamics of climate change (Crowther et al., 2019). Meanwhile, long-term
 56 drought may lead to a continuous reduction of forest SOC (Peng et al., 2025), which in turn



57 strongly affected the global soil carbon cycle and carbon storage processes (Müchael and
58 Bahn, 2022). In forest ecosystems, the effects of drought on SOC accumulation were mostly
59 negative (Zhou et al., 2019; Kabir et al., 2024). A meta-analysis of the effects of drought
60 (precipitation reduction experiments) on soil carbon cycling in natural ecosystems,
61 particularly forests, showed that prolonged drought led to a -3.3% reduction in SOC content
62 (based on 148 drought-manipulation studies) (Deng et al., 2021).

63 The SOC represents a mixed pool of carbon, which varies with their turnover rates,
64 chemical characteristics and stabilization mechanisms (Lavallee et al., 2020). Generally, SOC
65 generally are divided into active organic carbon and inert organic carbon. Active organic
66 carbon is readily broken down by microorganisms and directly participates in the chemical
67 transformation processes involving soil organisms (Sun et al., 2023). It quickly reflected
68 changes in soil organic matter, with the research concentrated on SOC components (Zhao et
69 al., 2018). Specifically, dissolved organic carbon (DOC), easily oxidizable organic carbon
70 (EOC), and microbial biomass carbon (MBC) all belong to the active carbon components
71 with fast turnover in SOC. However, the impacts of drought on SOC components vary across
72 regions around the world (Deng et al., 2021; Soares et al., 2023). Specifically, drought
73 reduced MBC reservoir size by 2% to 30% (Sun et al., 2020). This reduction leads to an
74 increase in the accumulation of DOC in the topsoil by 30% to 60% (Deng et al., 2021; Brunn
75 et al., 2023). Therefore, active organic carbon storage in different forests respond differently
76 to drought intensity, and the differential mechanism could be clarified under drought.

77 Despite some progress in understanding the general trends in SOC dynamics under
78 drought conditions, several challenges remain (Brunn et al., 2023). Specifically, one key issue



79 is the lack of a comprehensive understanding of the mechanisms in regulating soil particulate
80 organic carbon (POC) and mineral-associated organic carbon (MAOC) dynamics during
81 drought (Lai et al., 2024). POC was influenced directly by the plant-derived inputs and the
82 functional genes of carbon-degrading fungi with modulated by DOC and total nitrogen (TN).
83 In contrast, MAOC was shaped by the increasing inputs of labile carbon, such as DOC (Gao
84 et al., 2024). Therefore, more research is necessary to explore the role of additional factors—
85 including soil microorganisms, enzyme activities, and other biological factors—in mediating
86 SOC decomposition and stability. For example, in surface soil, MAOC increased
87 proportionally with microbial biomass and POC, fungal necromass carbon had a more
88 contributed to MAOC in Qinling Mountains (Lai et al., 2024). Oppositely, some study
89 reported that the impact of drought on POC and MAOC was limited, but POC exhibited
90 greater vulnerability to drought compared to MAOC (Shi et al., 2024). Therefore, these
91 inconsistent results were verified differently response to drought.

92 Soil microbial carbon utilization efficiency (CUE) denotes to the fraction of carbon
93 incorporated into microbial biomass relative to the overall carbon uptake by microorganisms
94 (Li et al., 2021a). It reflects the carbon allocation between anabolism and catabolism
95 processes in the soil microbial community and directly affects the soil carbon storage
96 capacity (Sun et al., 2025). Moreover, microbial CUE may be an important parameter for
97 determining SOC storage by controlling the proportion of SOC utilization (Geyer et al., 2016;
98 Ullah et al., 2021). Specifically, previous studies have shown that drought may affect the
99 activity of microbes and their CUE, which in turn affects the decomposition and
100 accumulation of POC and MAOC (Duan et al., 2023). For example, higher CUE may



101 accelerate the breakdown of POC, as the generation of microbial byproducts during this
102 process facilitates the formation of MAOC, as shown by [Guenet et al. \(2021\)](#) and [Chen et al.](#)
103 [\(2021\)](#). Further studies are needed to determine whether drought alters CUE and
104 consequently affect soil carbon pools. Understanding how SOC fractions, particularly POC
105 and MAOC, influence CUE could provide insights into the mechanisms through which
106 ecosystems maintain their carbon balance during drought events. Moreover, the relationships
107 between SOC fractions and CUE under drought is poorly understood. To address this gap, we
108 examined the potential correlations among these variables, aiming to provide a more
109 comprehensive perspective on how drought impacts soil carbon dynamics and plant
110 productivity.

111 *Phyllostachys edulis* is a vital plantation in China, thriving abundantly across the
112 subtropical regions, which is the preferred material for “replacing wood with bamboo” and
113 “replacing plastic with bamboo” for its fast natural regeneration. In our study, a drought
114 experiment lasting 9 years was conducted in *P. edulis* forest in southeastern China. We
115 hypothesize that: (a) Throughfall exclusion reduced the soil microbial CUE with higher soil
116 enzyme activities. (b) soil microbial CUE and soil carbon fractions had different responses to
117 drought at soil vertical direction. The main objectives of this study are: (1) to evaluate the
118 effects of throughfall exclusion on soil POC, MAOC and soil microbial CUE; and (2) to
119 reveal how throughfall exclusion influences SOC components by changes in microbial CUE
120 that mediates these impacts. This study helps to clarify the internal link between SOC
121 components and CUE in *P. edulis* plantation under throughfall exclusion as identifying
122 predictors of response to long-term drought, which would provide a reference for the



123 optimization of carbon cycle models in terrestrial ecosystems under drought conditions (Li et
124 al., 2021b).

125

126 2. Materials and methods

127 2.1 Study area

128 The study area locates in Huanggongwang Forest Park (119 56 '~120 02' E, 30 03 '~30 06'
129 N), Hangzhou, Zhejiang Province, which is 10 km away from the city. It exhibits a
130 characteristic subtropical humid monsoon climate, with an average annual temperature of
131 16.1°C, abundant rainfall and annual average rainfall of 1441.9 mm. The soil type in the
132 forest area is slightly acidic red soil. The bamboo forests in experimental areas were planted
133 in the 1960s, which have been used for special experimental areas without agricultural
134 implementations and management such as fertilization, tillage, bamboo shoots harvest or
135 thinning. The physical and chemical properties of soil were first delineated in Table 1.

136 Table caption

137 Table 1 General information of the experimental *P. edulis* stands (mean±SD, n=9)

Factors	Stand 1		Stand 2		Stand 3	
	CK	TE	CK	TE	CK	TE
Altitude /m	136	136	141	141	146	146
Slope / °	15	15	20	20	22	22
Slope aspect	S	S	S	S	S	S
Soil depth /cm	60	60	60	60	60	60
SOC /g·kg ⁻¹	34.37±9.06	36.11±7.03	36.74±8.04	39.78±7.39	37.82±8.21	37.42±9.64
TN/g·kg ⁻¹	2.72±0.79	2.34±0.27	2.72±0.48	2.91±0.52	2.56±0.29	2.52±0.41
SAP / mg·kg ⁻¹	4.25±0.93	4.64±0.81	5.50±1.03	5.94±1.23	6.48±1.09	6.45±2.04
DBH /cm	13.42±2.17	14.72±1.55	14.44±2.14	14.30±1.93	12.99±2.53	14.60±1.26
Height / m	17.28±0.90	16.32±2.28	16.97±1.50	15.97±2.04	15.80±2.41	16.64±0.86



Density/ n·ha ⁻¹	3800	3600	3400	3000	3250	2800
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139 *2.2 The design of throughfall exclusion experiment*

140 In July 2014, three *P. edulis* stands with similar sites, altitude and growth conditions were
141 selected as the experimental plots. To guarantee the scientific rigor and precision of the
142 experiment, two plots, each measuring 10 m×10 m (excluding boundary effect), were
143 designated for the control (CK) and throughfall exclusion (T) treatments, with a completely
144 random allocation. Among them, CK and T treatments were three repetitions.

145 The PVC waterproof plates were systematically placed at a height of 1.5 m above the
146 experimental plots to mitigate throughfall, supported by frameworks constructed from
147 galvanized steel pipes with an area of 11 m×11 m. The PVC material had good light
148 transmittance, ensuring that the light conditions in the throughfall exclusion treatments were
149 as consistent as possible with the counterpart. At the same time, to prevent the horizontal
150 transport of water from the surrounding soil to the throughfall exclusion treatments plots,
151 trenches (about 50 cm deep and 20 cm wide) were dug in all around of throughfall exclusion
152 plots, cut off the bamboo whip connected to the throughfall exclusion plots. In the trench,
153 PVC waterproof plates were buried to further cut off the entry water from outside plots. Since
154 August 2014, these samples have officially entered the experimental stage of rain
155 interception. Litter from the exclusion shelters were gathered bi-weekly and allocated to the
156 respective plots, thus mitigating the effects of variability in litter input.

157

158 *2.3 Soil sampling and analysis*



159 Soil samplings were collected in August 2022 with 0–10 cm and 10–20 cm soil depth from all
160 6 plots. Each sample was collected by 5-point sampling and mixed into one sample with the
161 same soil layer. The fresh soil samples after 2 mm soil sieve were divided into two parts, one
162 was stored in a refrigerator (-20°C) with soil β -glucosidase (BG), N-acetyl aminoglycosidase
163 (NAG), leucine aminopeptidase (LAP), microbial carbon (MBC) measurement; one was put
164 in laboratory natural with air drying to screen soil organic carbon (SOC), and total soil
165 nitrogen (TN), etc. Soil samples were promptly returned to the laboratory, where they were
166 sifted through 2-mm sieve and subsequently preserved at 4°C or dried in natural condition for
167 future analysis.

168

169 *2.3.1 Soil properties measurement*

170 Moisture content was determined gravimetrically by utilizing samples dried in an oven for 48
171 h at 105°C . SOC and TN were evaluated utilizing a soil CN elemental analyzer (Elementar
172 Vario MAX, Germany). The concentrations of dissolved organic carbon (DOC) and dissolved
173 organic nitrogen (DON) within the soil were quantified employing a TOC analyzer
174 (Shimadzu, Japan) alongside a continuous flow analytic system (Skalar, Netherlands),
175 maintaining the ratio of soil:water as 1:4. Mineral N levels were ascertained through
176 extraction with 2 M KCl solution, utilizing a continuous flow analytical system
177 (Netherlands). Soil microbial biomass C (MBC) and microbial biomass N (MBN) were
178 quantified from freshly collected soil samples through the chloroform fumigation extraction
179 technique utilizing a 0.5 M K_2SO_4 solution (Vance et al., 1987). The extractable C and N
180 were assessed via a TOC analyzer (Shimadzu, Japan) and continuous flow analytical system



181 (Breda, Netherlands). The MBC and MBN were calculated as the difference in C and N
182 between fumigated and non-fumigated samples and divided by 0.45 and 0.54 (Vance et al.,
183 1987).

184

185 *2.3.2 Soil fractions separation*

186 The measurement of SOC fractionation was according to Degryze et al. (2004). A 60 g
187 sample, air-dried and measuring less than <2 mm, was introduced into 100 mL of a 5%
188 sodium hexametaphosphate solution and agitated for 15 h. Following this, the resulting soil
189 suspension was transferred into a sieve with an aperture of 0.053 mm, the material retained
190 on the sieve (particles larger than 53 µm) was utilized for the assessment of POC quality,
191 while the fractions passed through the sieve (particles smaller than 53 µm) was allocated for
192 the evaluation of MAOC quality. The upper and lower sections of the sieve were gathered
193 separately and placed in a thermostatic oven set at 65°C for drying, after which their
194 respective masses were recorded, and then the total soil mass percentage was calculated.
195 Subsequently, the desiccated soil in the beaker was collected and pulverized through a 0.15
196 mm sieve, with the organic C content determined utilizing the external heating method
197 involving potassium dichromate-concentrated sulfuric acid.

198

199 *2.3.3 Soil enzyme activities measurement*

200 Four soil enzyme activities were assessed in the decomposition of relatively labile C
201 compounds (hydrolytic enzymes such as β-glucosidase, BG), and more resistant C
202 compounds (oxidative enzymes including phenol oxidase: POX, and peroxidase: PER),



203 following the methodologies outlined by [German \(2011\)](#) and [Saiya-Cork et al. \(2002\)](#). In
204 summary, 100 μM methylumbelliferone (MUB)-labeled substrates of α -D-glucopyranoside
205 and β -D-glucopyranoside were utilized for AG and BG, respectively, while a 25 mM 3, 4-
206 dihydroxyphenylalanine (DOPA) substrate was employed for oxidative enzymes. For each
207 soil sample, 2 g of freshly collected soil were measured and subsequently immersed in 2 L of
208 50 mM sodium acetate buffer solution at a pH of 5. For the hydrolytic enzymes (BG), a
209 volume of 200 μl sample suspension and 50 μl of the corresponding enzyme substrates were
210 dispensed into wells of the substrate plate, followed by the addition of suitable standards. In
211 the case of POX and PER, each assay well was filled with 50 μl of 25 mM L-DOPA
212 complemented by 200 μl sample suspension. The fluorescence of hydrolytic enzymes was
213 measured using 365 nm excitation and 450 nm for emission with a microplate reader (BMG
214 LABTECH GmbH, Germany). The activities of hydrolytic and oxidative enzyme were
215 quantified as $\text{nmol}\cdot\text{g}^{-1}\cdot\text{soil}\cdot\text{h}^{-1}$.

216 The enzymatic activities of β -1,4-N-acetyl-glucosaminidase (NAG) and leucine
217 aminopeptidase (LAP) were evaluated utilizing microplate-based fluorometric methodologies
218 ([Sinsabaugh et al., 1997](#)). BG, NAG, and LAP possessed the ability to extract assimilable
219 nutrients from fundamental organic sources of C (e.g., β -linked glucans) and N (e.g., protein
220 and amino polysaccharides) ([Sinsabaugh et al., 2013](#)). The microplates were analyzed
221 through an automated fluorometer (Winooski, VT), employing an excitation wavelength of
222 365 nm and an emission wavelength of 450 nm ([Saiya-Cork et al., 2002](#)).

223

224 *2.3.4 CUE estimation*



225 We employed soil enzyme activity, readily organic matter, and the C:N ratio of microbial
226 biomass to determine the CUE based on stoichiometric modelling (Geyer et al., 2019;
227 Sinsabaugh et al., 2016). Labile organic matter was assessed through the measurement of
228 DOC and N extracted from non-fumigated samples (Geyer et al., 2019). The CUE was
229 derived from stoichiometric models based on the growth and respiration of bacterial and
230 fungal (Sinsabaugh et al., 2016). We determined microbial efficiency in utilizing C and N
231 through the lens of ecological stoichiometry. The method was described as follows:

$$232 \quad CUE = CUE_{max} \left[S_{C:N} / (S_{C:N} + K_N) \right]$$

233 Where $S_{C:N}$ indicates the enzyme activity allocation offset the difference between the
234 existing available resource element composition (DOC/DN ratio) and the microbial biomass
235 composition (MBC/MBN ratio). The half-saturation constant K_N was established at 0.5. The
236 maximum efficiency of carbon utilization denoted the pinnacle of C that could be harnessed
237 for microbial proliferation, which is determined to be 0.6 in accordance with the
238 thermodynamic constraints. The ratio of the C: N enzyme (i. e. EEAC: N) activity was
239 calculated using $BG / (NAG + LAP)$. $L_{C:N}$ is the carbon to nitrogen ratio of unstable organic
240 matter.

241

242 *2.4 Statistical analysis*

243 All statistical evaluations were performed utilizing SPSS 20 or Origin version 12.0. Normal
244 distribution of data and homogeneity of variance were checked using Shapiro-Wilk and
245 Levene's tests, respectively. One-way ANOVA was employed to examine the influence of soil
246 depth and drought treatments on soil indicators. To evaluate the effect of a singular variable,



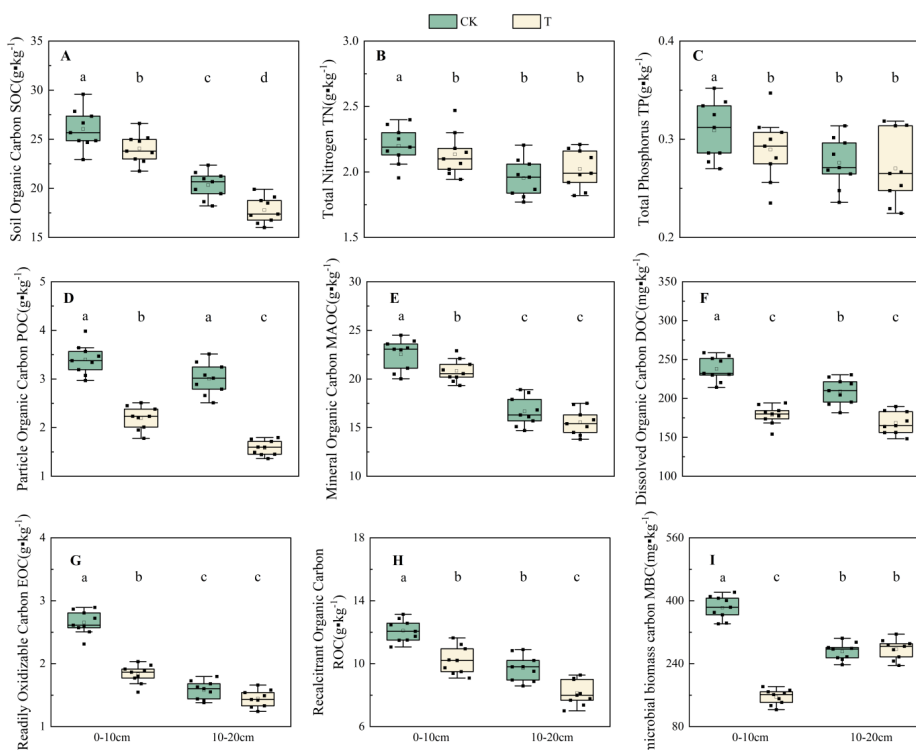
247 one-way ANOVA was employed for the data that followed a normal distribution, while the
248 Kruskal–Wallis tests was utilized for those that did not conform to normally. To investigate
249 the impact of environmental factors on microbial-related parameters, Pearson and Spearman
250 correlations were used for data that followed a normal distribution and those that deviated
251 from it, respectively. Differences and correlations considered at a significant level of $p <$
252 0.05.

253

254 **3. Results**

255 *3.1 Effect of the throughfall exclusion on SOC fractions*

256 Throughfall exclusion significantly reduced SOC, POC, DOC and ROC ($p < 0.05$), but did not
257 affect MAOC ($p > 0.05$) (Fig. 1). SOC and soil carbon fractions varied significantly across soil
258 layers (Fig. 1). Compared to the control, EOC and MBC in throughfall exclusion treatments
259 were significantly reduced by 53.5% and 61.6% at the 0-10 cm soil layer, respectively, but
260 they had no significant change at the 10-20 cm soil layer. In vertical direction, SOC, MAOC,
261 EOC, and ROC content in control treatment decreased by 21.8%, 23.7%, 26.7% and 51.1%
262 with depth increased ($p < 0.05$), respectively. In vertical direction, SOC, MAOC, EOC, and
263 ROC in throughfall exclusion treatment decreased by 18.2%, 25.1%, and 34.1% with soil
264 depth increased ($p < 0.05$), respectively.



265

266 Fig. 1 Effect of throughfall exclusion on soil organic carbon fractions. Different lowercase
 267 letters indicate significant differences across treatments ($p < 0.05$).

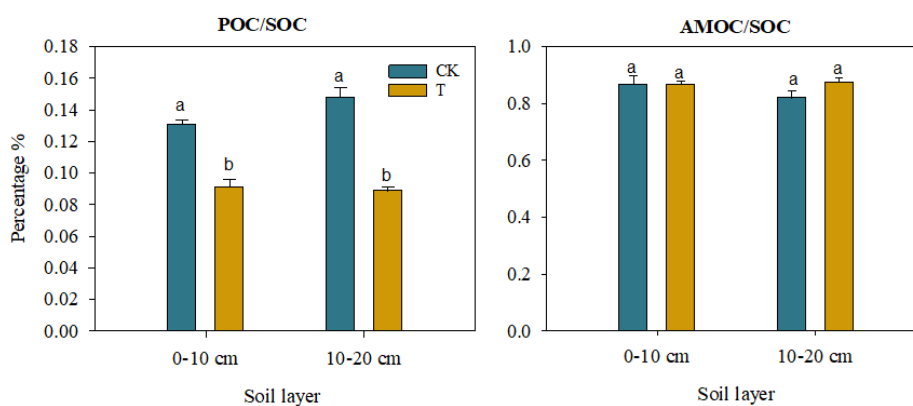
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269 *3.2 Effect of throughfall exclusion on the proportion of POC and MAOC*

270 The throughfall exclusion treatments had a significant impact on the proportion of POC and
 271 MAOC in SOC ($p < 0.05$) (Fig.2). Throughfall exclusion treatments, in contrast to CK, led to a
 272 4.0% reduction in POC, which in turn caused a significant increase of MAOC within SOC.
 273 Specifically, the POC/SOC ratio diminished by 4.1% in the 0-10 cm soil layers and by 6.3%
 274 in the 10-20 cm soil layer. The vertical distribution of POC/SOC exhibited a slight increase of
 275 2.6% with depth in the control treatments ($p < 0.05$), while throughfall exclusion treatments



276 showed a negligible effect of merely 0.4%. The one-way ANOVA revealed that the
277 throughfall exclusion treatment had a significant impact on the proportion of POC and
278 MAOC in SOC ($p<0.01$), whereas the soil depth remained unaffected.
279



280

281 Fig. 2 Effect of throughfall exclusion on the proportion of POC and MAOC in SOC.

282

283 3.3 Effect of throughfall exclusion on soil enzyme activities and CUE

284 Compared with the control, PPO activities in throughfall exclusion treatments were
285 significantly reduced by 30.1% and 27.9% in the two soil layers, respectively (Fig.3). The
286 activities of BG and CBH in the 10-20 cm soil layer decreased by 34.6% and 38.1%,
287 respectively, whereas no significant differences were detected in the 0-10 cm soil layer. On
288 the contrary, throughfall exclusion treatments significantly increased the activities of NAG
289 and POX by 14.9% and 29.4% in 0-10 cm soil layers, 18.3% and 29.1% in 10-20 cm soil
290 layers ($p<0.05$), respectively. Additionally, the activities of PPO exhibited a notable decline
291 ($p<0.05$) as the increase of soil depth, whereas the activities of CBH and NAG remained no



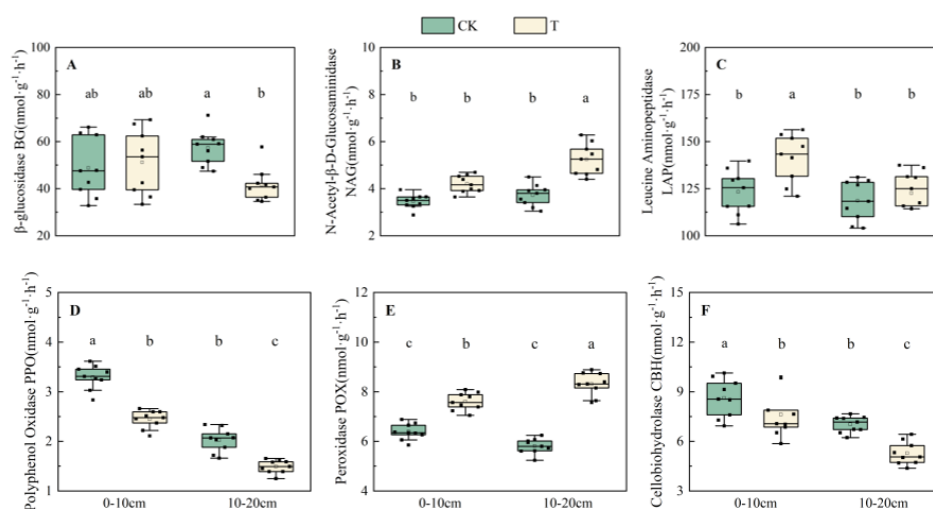
292 significant change with the increase of soil deep (Fig.3).

293 Throughfall exclusion treatments significantly decreased carbon pool activity (CA) by

294 31.0% at 0-10 cm (Fig.4), however, no significant change was observed at 10-20 cm (Fig.4a).

295 Conversely, throughfall exclusion treatments significantly enhanced microbial CUE by 9.1%

296 at 0-10 cm ($p < 0.05$), and by 2.6% at 10-20 cm (Fig.4).

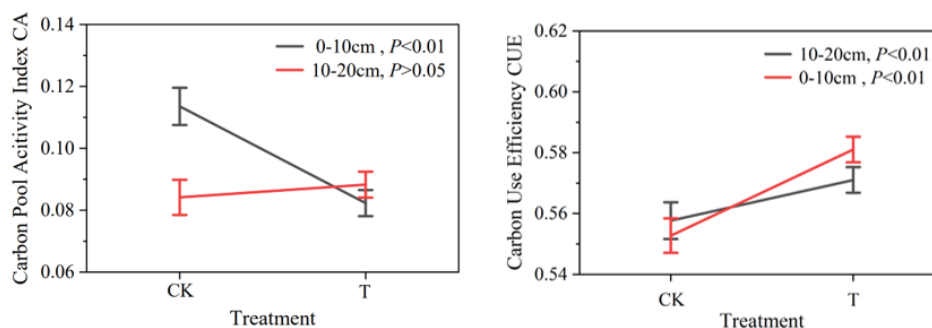


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298 Fig.3 Effect of throughfall exclusion on soil enzyme activity at 0-20 cm soil layer. Different

299 lowercase letters indicate significant differences across treatments ($p < 0.05$).

300



301



302 Fig.4 Effect of throughfall exclusion on soil microbial CUE across 0-20 cm soil layer.

303

304 *3.4 Effect of throughfall exclusion on the relationships between CUE and SOC fractions*

305 CUE was significantly correlated with POC and DOC across both two soil layers (Fig.5 and

306 Fig. 6). CUE significantly affected by POC, DOC, EOC, MBC, MBN in 0-10 cm soil layer

307 ($p < 0.05$), CUE extremely significantly affected by POC, DOC, MBN, BG in 10-20 cm soil

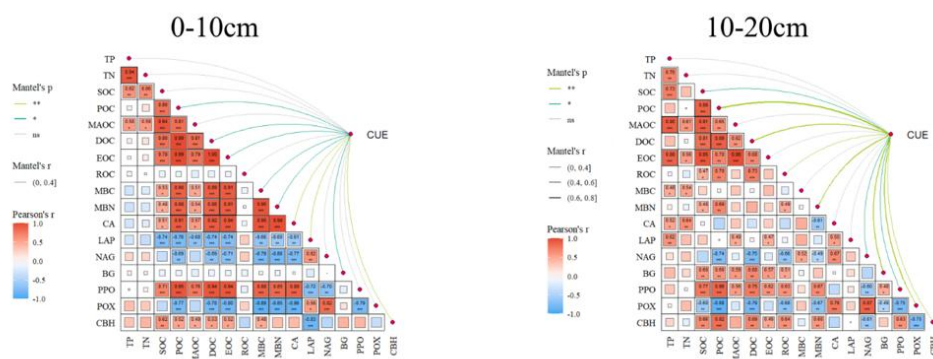
308 layer ($p < 0.01$) and significantly affected by SOC, NAG and PPO ($p < 0.05$). POC showed a

309 highly significant positive correlation with CBH, PPO as well as soil nutrients (Fig. 5), and a

310 significant negative correlation with LAP (0-10 cm) and POX (10-20 cm). There was

311 significantly difference relationships between CUE and POC across two soil layers, as well as

312 soil nutrients and soil enzyme (Fig.6).



313

314 Fig.5 Correlation between CUE and soil chemical properties as well as soil enzymes across

315 two soil layers. CUE: Soil microbial carbon utilization efficiency; CA: Carbon Pool Activity

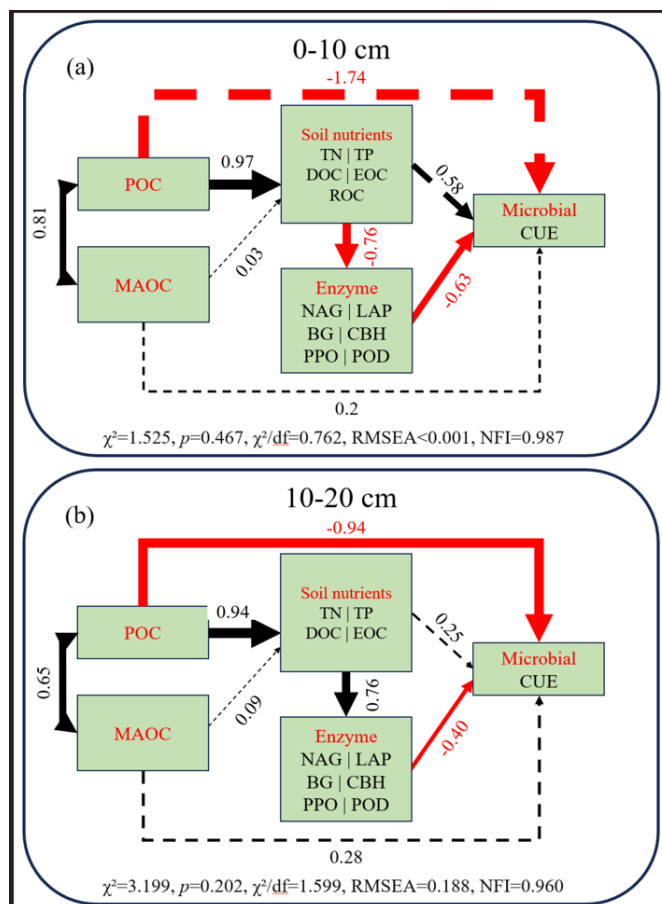
316 Index; SOC: Soil organic carbon (SOC); POC: soil particulate organic carbon; MAOC:

317 mineral-associated organic carbon; DOC: soluble organic carbon; ROC: easily oxidized

318 organic carbon; EOC: MBC: microbial carbon; PPO: polyphenol oxidase; PER: peroxidase



319 CBH: cellobiohydrolase; BG: β -glucosidase; NAG: N-acetyl aminoglycosidase; LAP: leucine
 320 aminopeptidase.



321
 322 Fig. 6 The structural equation model (SEM) depicts the direct and indirect effects of CUE,
 323 soil microorganisms and soil nutrients under two treatments at 0-10 cm and 10-20 cm.
 324 Notes: Principal component analysis was conducted for soil enzyme data from DOC, EOC,
 325 ROC, TN, TP; Principal component analysis was conducted for soil enzyme data from NAG,
 326 LAP, NG, CBH, PPO, POD. DOC: soluble organic carbon, EOC: readily oxidizable organic
 327 carbon, ROC: recalcitrant organic carbon, TN: total nitrogen, TP: total phosphorus. PPO:



328 polyphenol oxidase; PER: peroxidase CBH: cellobiohydrolase; BG: β -glucosidase; NAG: N-
329 acetyl aminoglycosidase; LAP: leucine aminopeptidase. The black arrows signify a
330 positive effect, while the red lines denote a negative effect. The thickness of the arrow
331 is directly proportional to the magnitude of the effect.

332 **4. Discussion**

333 *4.1 The influence of throughfall exclusion on soil POC and MAOC*

334 These findings indicated that drought resulted in a decline in soil organic carbon reserves,
335 and the volatile carbon pools experienced a more pronounced impact compared with the
336 stable carbon pools (Fig.1). The proportion increase of stable soil carbon pools may reflect an
337 adaptive adjustment to drought conditions. In this context, soil microorganisms were inclined
338 to utilize more resilient carbon sources to maintain their biological functions (Jones et al.,
339 2019). Analyzing the response of SOC to climate change, without taking account the POC
340 and MAOC pools, may be misleading. In this study, eight years of throughfall exclusion
341 significantly decreased soil POC and had a limited impact on MAOC (Fig.2), indicating that
342 as the percentage of MAOC in SOC escalated, the proportion of stable SOC components
343 increased. The significant increase of stable soil carbon pools under drought conditions may
344 reflect an adaptive adjustment response, while soil microorganisms tend to use more stable
345 carbon sources to maintain their metabolic activities (Jones et al., 2017). If the POC and
346 MAOC pools were omitted from consideration, examining the response of SOC to climate
347 change in isolation could be misleading, as the responses of both surface and subsurface
348 carbon pools to diminish throughfall exhibited significant differences.

349 The distinctions in SOC components between topsoil and subsoil may be intricately



350 linked to (i) the diverse environmental and microbial properties inherent to different soil
351 layers and (ii) the diminished nitrogen availability in the subsoil under drought conditions, in
352 contrast to the topsoil. Subsoil microorganisms were recognized for their susceptibility to
353 deficiencies in accessible carbon or energy (Jones et al., 2018), exhibiting a more sensitive to
354 fluctuations in labile carbon supply when compared to topsoil microorganisms (Fontaine et
355 al., 2007; Jones et al., 2019). According to microbial economic theory, when labile carbon is
356 available, microorganisms can minimize their metabolic energy expenditure and reduce
357 extracellular enzyme production (Bradford, 2013). Thus, increased root deposition and
358 unstable carbon supply may inhibit the microbial production of hydrolytic enzymes in subsoil
359 (Wei et al., 2019). The elevated root density of moso bamboo may exacerbate the competition
360 between the plant and microbes for nutrients located in the subsoil (Hill & Jones, 2019; Liu et
361 al., 2018). This may further limit the metabolism of subsoil microorganisms and the synthesis
362 of extracellular enzymes (Schimel & Weintraub, 2003), and the drought stress caused by the
363 throughfall exclusion treatment (Burns et al., 2013; Mcdaniel et al, 2013; Sardans &
364 Penuelas, 2005).

365 As soil depth increased, there was a subtle reduction in the soil POC, which could be
366 attributed to the implementation of the throughfall exclusion treatments, which involved the
367 installation of shelters beneath the forest canopy. While these structures diminished the
368 amount of rainfall reaching the soil, they permitted both the canopy and trunk of the *P. edulis*
369 forest to continue absorbing precipitation. Furthermore, throughfall exclusion treatments in
370 this study were relatively moderate compared to methods employed in previous studies.
371 Forest ecosystems were resilient to throughfall exclusion, therefore, detecting significant



372 alterations in POC content, as highlighted by [Canarini et al. \(2017\)](#).

373

374 *4.2 Effect throughfall exclusion on soil enzyme and microbial CUE*

375 Soil enzymes play an indispensable role in nutrient mineralization, functioning as a distinct
376 indicator for forecasting the available nutrient of plants ([Aon and Colaneri, 2001](#); [Baum et](#)
377 [al., 2003](#)). In this study, soil enzyme activity exhibited different sensitivity to drought
378 conditions, throughfall exclusion significantly reduced PPO activities (27.9% to 68.4%)
379 ($p<0.05$), CBH activity (16.2% to 38.6%) ($p<0.05$) and POX enzyme activity (18.2% to
380 51.0%) ($p<0.05$), respectively. These changes indicated soil moisture played a critical role in
381 regulating microbial enzyme activity in subtropical forest soils. In the long run, alterations in
382 the activity of soil enzymes involved in carbon and nitrogen cycles will affect soil available
383 nutrients, thereby affecting the nutrient supply to plants ([Roldán et al., 2025](#)). In this study,
384 drought reduced the activity of carbon-related enzymes (BG, CBH) and increased the activity
385 of nitrogen-related enzymes (NAG, LAP), which could be explained by the limiting effect of
386 nitrogen in subtropical forest soils, as reported in previous studies ([Mayo et al., 1994](#)). The
387 limitation of nitrogen implied a higher sensitivity to throughfall exclusion conditions. The
388 most pronounced impact of drought in our study was observed in oxidases (POX, PPO),
389 which were produced by microorganisms that function as essential decomposers of substrates
390 oxidation, such as lignin ([Tian et al., 2025](#)). This resulted indicated microorganisms played a
391 crucial role in the decomposition of organic carbon in soil, with POX involved in humus
392 transformation and PPO promoting lignin and humus degradation ([Wang et al., 2008](#)).

393 In this study, drought reduced the activity of polyphenol oxidase, slowed organic



394 matter decomposition and with the accumulation of SOC, indicating that drought may
395 become the limiting factor for microbial community nutrients, affecting the growth of related
396 microorganisms and reducing MBC content (Tian et al., 2014). Hydrolytic enzymes,
397 exemplified by BG, can facilitate the breakdown of macroglycemic sugars and active organic
398 matter, transforming them into nutrients available to both plants and microorganisms (Grandy
399 et al., 2007). In this study, we found that drought improved the activity of nitrogen-related
400 hydrolase and reduced the DOC content, enhanced hydrolase activity promoted the
401 decomposition of active organic matter, expediting the microbial utilization of DOC (Fan et
402 al., 2015). The activity of the six enzymes decreased with soil depth, consistent with Chen
403 (2003), due to ground litter and better soil ventilation leading to higher soil enzyme activity
404 in the 0-10 cm layer compared to the 10-20 cm lower.

405 Drought altered soil carbon and nitrogen cycling and reduced the nutrient utilization
406 efficiency of soil microorganisms (Guo et al., 2020). In this study, drought improved soil
407 microbial CUE, but significantly decreased CA, suggesting CUE was negatively associated
408 with some enzyme activities under drought conditions. It was possible that the microbial
409 community was carbon-limited and thus secreted less extracellular enzymes with higher CUE
410 (Ullah et al., 2021). This secretion accelerated the decomposition, absorption and utilization
411 of organic carbon, thereby increasing CUE and enhancing microbial carbon sequestration.
412 However, CA decreased with SOC, DOC, POC and MBC, indicating a threat posed by
413 drought to soil carbon sequestration. The intensification of nitrogen limitation and its effects
414 on soil enzyme activity most likely underlay the changes in CUE under drought conditions
415 (Allison & Vitousek, 2005; Bicharanloo et al., 2020). This conclusion was supported by the



416 strong positive correlation between CUE and leucine aminopeptidase activity in soil.
417 Furthermore, although CUE increased as LAP rose in the subsoil, NAG did not exhibit
418 similar change. Since microbial residues, including aminoglycans, were nitrogen-rich organic
419 substrates, they played a crucial role in soil nitrogen cycling (Heuck et al., 2015),
420 microorganisms may recover or decompose them, especially under nitrogen-limiting
421 conditions (Cui et al., 2020).

422

423 *4.3 The influence of microbial CUE on soil carbon fractions*

424 POC is a functional soil component that contributes to the stabilization of SOC (Witzgall et
425 al., 2021) and has weaker physical and chemical protection than MAOC, making it more
426 susceptible to microbial utilization and mineralization (Tang et al., 2023). Higher CUE could
427 potentially expedite the breakdown of the POC (Averill et al., 2018). In this study, soil
428 microbial CUE demonstrated a significant and positive correlation with SOC, DOC and POC
429 (Fig. 5), indicated that CUE was responsible for the most alteration in both the quantity and
430 composition of SOC (Shi et al., 2025). The study of Liu et al (2018) concluded that CUE
431 mediated carbon loss via microbial respiration and strongly influenced the rate of SOC
432 formation. In this study, the diminished accessibility of POC to microorganisms may foster an
433 enhancement in CUE for survival, elucidating the favorable correlation observed between
434 CUE and POC in our findings (Fig. 5). Furthermore, the study by Stone et al (2023) showed
435 that drought could reduce the NH_4^+ -N and NO_3^- -N, thereby decreasing nitrogen use efficiency
436 and consequently reducing CUE. In this study, TN and MBC both decreased (Fig.1), because
437 lower microbial CUE resulted in less accumulation of MBC and microbial residues, thereby



438 reducing the MAOC repertoire. Thus, changes in MAOC concentrations may result from
439 alterations in microbial N cycling in response to drought. These findings did not support our
440 observation that drought decreased CUE and promoted MAOC reduction, but demonstrated
441 the microbial CUE was affected by SOC fractions with substrate available C.

442 In addition, studies have shown that long-term drought reduced soil available N, which
443 may cause a significant increase in soil C/N ratio, resulting in microorganisms experiencing
444 N limitation with a decrease in microbial biomass (Chen et al., 2020). When soil
445 microorganisms were relatively limited by a specific element, which were induced to
446 synthesize the corresponding element acquisition enzymes (Wang et al., 2009). This
447 enzymatic response also contributed to the rapid improvement of LAP activity observed in
448 our study (Fig. 3). However, Koyama et al. (2025) showed microbial CUE significantly
449 decreased by the increase of C availability. Ullah et al. (2021) showed that drought caused
450 small increases in CUE across season on grassland in Australia. Fuchslueger et al. (2019)
451 showed microbial CUE decreased with temperature change independent of drought by
452 substrate respiration. Therefore, drought may affect microbial CUE differently over different
453 time scales, substrate quantity and quality change of C availability (Koyama et al., 2025) and
454 temperature sensitivity (Fuchslueger et al., 2019). There is a need to further quantify the
455 effects of drought on long-term C sequestration and the microbial mechanisms underlying
456 this process. The drought duration of our study was 9 years, to better understand how SOC
457 components respond to droughts of varying durations, further studies should observe the
458 long-term effects of drought over different time scales. The increase of CUE with SOC
459 reduction mainly depended on scenarios such as forest vegetation restoration under global



460 change (Shi et al., 2025).

461

462 **5. Conclusion**

463 We found that throughfall exclusion significantly decreased POC and MAOC and increased
464 soil microbial CUE. Furthermore, the proportion reduction of POC/SOC under drought was
465 mainly due to the improvement of CUE. POC subjected to fewer soil aggregate protection
466 mechanisms was more variable under drought, which reflected the adaptive adjustment of the
467 soil carbon pools under drought stress. Under drought conditions, changes in enzyme activity
468 played a key role in understanding how the increase of CUE leads to changes in soil carbon
469 components. This change provides an important explanation for SOC dynamics. The results
470 showed that soil microbial CUE primarily influenced soil microbial biomass, which in turn
471 affected SOC components. These findings helped to better understand how throughfall
472 exclusion impacts on SOC in *P. edulis* plantations. Drought may have variable effects on
473 CUE over different time periods and substrate C quality. To gain a deeper understanding of
474 this complex relationship, it is imperative to assess the precise impacts of drought on long-
475 term carbon sequestration processes, along with the microbial mechanisms underpin these
476 processes.

477

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483

484 **Declaration of interests**

485 The authors declare that they have no known competing financial interests or personal

486 relationships that could have appeared to influence the work reported in this paper.

487

488 **Availability of data and materials**

489 The data that support the findings of this study are available from the corresponding author

490 upon reasonable request.

491

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