

We sincerely thank the reviewer for their positive assessment of our manuscript and for highlighting the relevance of a nitrite-centered perspective in nitrogen cycling. We greatly appreciate the constructive and detailed feedback provided.

We agree that the manuscript required updating and better integration of recent advances in the field. In particular, we acknowledge the omission of several important and recent studies on nitrite dynamics, isotope characteristics, and modelling approaches. We have carefully incorporated the suggested references and revised the manuscript accordingly to reflect the latest developments and perspectives.

Detailed comments:

P3, L55-56 “Nitrite is the only inorganic nitrogen species produced by both oxidative and reductive processes” – not really, what about N₂O?! it can be produced due to nitrate reduction and as well from ammonia and hydroxylamine oxidation – this nitrification pathway is missing in the Figure 1!

We thank the reviewer for this important clarification. Our intention was to emphasize the central role of nitrite as the key intermediate linking oxidative and reductive nitrogen transformations, rather than to imply that no other nitrogen species can be produced under both regimes. To avoid this ambiguity, we have revised the sentence to: “Nitrite is the central dissolved intermediate linking oxidative and reductive nitrogen transformations and the obligatory precursor to downstream nitrate, ammonium and gaseous nitrogen products.” Lines 57-59. We have also explained in detail the role of hydroxylamine in the caption of Figure 1.

P3 L62 you should also add newer N-cycle reviews here, eg. Denk et al., 2017, Deb et al, 2024.

We have added these recent reviews to the revised manuscript in the section introducing major nitrogen transformation pathways and recent perspectives on nitrogen cycling. Lines 65-66.

P4 L73 also some more recent technical developments of nitrite isotope measurements should be mentioned: Deb and Lewicka-Szczebak, 2025 (doi.org/10.3389/fenvs.2025.1536882), Hu et al., 2026 (doi.org/10.1039/D5JA00363F)

The reviewer is fully right and these interested references have been added. Lines 77-78.

P5 L122 – this was also observed in the recent case study (Deb et al., 2025)

We have now included this reference in the revised manuscript as an additional example supporting the interpretation of simultaneous nitrite oxidation and reduction processes in environmental systems. Lines 134-135.

P5 L 136 – the nitrifier denitrification pathway that you describe here is also omitted in your Fig.1

We have explained in detail the role of hydroxylamine in the caption of Figure 1

P6 Fig 2 – I don't understand the bottom left panel illustrating delta 15N for dominating production, what do the 2 arrows mean? Two following graphs are clear, present the expected possible trends in isotope changes, but the first graph do not show change in time, just as one-point?

When nitrite production is higher than its consumption, the isotopic composition of N-nitrite will be initially depleted in 15N compared to the substrate and then it will evolve towards the initial isotopic 15N of the substrate, if it is limiting. We totally agree that this is not clear in the original version of Figure 2 and we have modified it accordingly in the revised version of the manuscript. We have also distinguished the isotopic effect of consumption during oxidative and reductive pathways.

P7 L198 – the sentence beginning with “Direct determination...” is a totally different topic, you said about analytical techniques before and here the paragraph was about N₂O. Also, a citation for N₂O reduction overprinting its isotopic signal should be added here.

We agree that the sentence discussing direct nitrite isotope determination interrupted the logical flow of the paragraph focused on N₂O isotopic signatures. This sentence has therefore been removed. We have revised the paragraph structure to improve the transition between analytical considerations and N₂O isotope systematics. We also added references discussing isotopic overprinting associated with partial N₂O reduction. Lines 205-227.

P9 Fig.3 – really great figure with perfect presentation of nitrite central role

We thank the reviewer for this positive assessment of Figure 3 and are pleased that the figure effectively conveys the central role of nitrite turnover within the nitrogen cycle framework developed in this perspective.

P10 L262 The oxygen exchange with water is also an important tracer because allows us to estimate the nitrite residence time and rates of biological NO₂ turnover relative to abiotic exchange (Lewicka-Szczebak et al., 2021, Buchwald and Casciotti, 2013)

We thank the reviewer for this important point. We agree that oxygen isotope exchange with water not only constrains interpretation of δ¹⁸O signatures, but may also provide valuable information on nitrite residence times and the relative rates of biological turnover versus abiotic exchange. We have revised the text accordingly. Lines 291-294.

P10 L279 I am missing the chapter on limitation of this approach. Most difficult one is the chemical instability of nitrite and its very low concentrations. If improperly stored and not analysed immediately it is quickly partially oxidised to nitrate and the isotope signal can be totally changed. Therefore, the analysis should be performed immediately, and the samples must be carefully conserved, best with high pH conditions. Because of typically low nitrite concentrations, especially in terrestrial

environments, the development of new analytical methods for lowering the detection limit for its isotope analysis is critical (Deb and Lewicka-Szczebak, 2025). I think these are important points because these analytical challenges are most probably the main reasons why nitrite isotope analyses in terrestrial systems are still very rare.

We agree that analytical limitations are a major factor restricting the broader application of nitrite isotope approaches in environmental systems. We have now added a paragraph discussing nitrite instability, rapid post-sampling transformation, low environmental concentrations, and the need for improved preservation and high-sensitivity analytical methods. This issue was also raised by reviewer 2.

Paragraph added in the Isotopes section (Lines 205-218): “Despite their strong mechanistic potential, nitrite isotope measurements are only feasible when sufficient nitrite accumulates for analysis and remain analytically challenging. Nitrite is chemically unstable and may undergo rapid oxidation or reduction during sampling and storage, potentially altering its isotopic composition before analysis. Reliable preservation therefore requires rapid processing and carefully controlled storage conditions, often under alkaline conditions that minimize transformation rates. In many terrestrial and aquatic systems, low nitrite concentrations restrict isotope analyses to zones where nitrite transiently accumulates. This emphasizes the need for continued development of high-sensitivity analytical approaches (Lewicka-Szczebak et al., 2021; Deb and Lewicka-Szczebak, 2025). For instance, a recent study developed a method for freshwater samples by coupling anion-exchange resin preconcentration with the azide reduction method for nitrite isotopic characterization at concentrations of $0.02 \mu\text{mol L}^{-1}$ or higher (Jiang et al., 2026). These analytical limitations likely contribute to the still limited application of nitrite isotope measurements in environmental studies despite their considerable interpretative potential.”

Paragraph added in the concluding section (Lines 298-302): “Although nitrite isotope measurements offer strong mechanistic insight, their application is constrained by low concentrations and analytical challenges. Recent advances in high-sensitivity methods are expanding their feasibility in natural systems. Nonetheless, these limitations continue to restrict their widespread use in environmental studies”.

1 **Ideas and perspectives: Nitrite turnover controls nitrogen fate**
2 **across redox gradients**

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16 **Abstract**

17 Reactive nitrogen fate in natural systems remains difficult to predict because pathway
18 partitioning occurs at the stage of nitrite turnover, where rapid and tightly coupled production
19 and consumption processes obscure the underlying fluxes. Concentration-based
20 assessments emphasize the dominant pools — nitrate and ammonium — and pathway
21 divergence is determined at the stage of nitrite turnover, independently of pool size. Nitrite is
22 the only inorganic nitrogen species produced under both oxidative and reductive regimes and
23 the obligatory precursor to all downstream dissolved and gaseous products. Because nitrite
24 rarely accumulates, it has often been treated as a transient intermediate of limited interpretive
25 value. This apparent invisibility reflects rapid, tightly coupled turnover and does not indicate
26 functional insignificance. Nitrogen retention, recycling and losses to the atmosphere are
27 determined during nitrite turnover, where competing pathways partition fluxes according to
28 kinetic and environmental constraints.

29 Observed concentrations integrate formation and consumption into a net signal that masks
30 opposing fluxes when internal cycling is rapid. Coupled $\delta^{15}\text{N}$ – $\delta^{18}\text{O}$ measurements of nitrite
31 constrain simultaneous production and consumption and differentiate biological from abiotic
32 pathways. Partial oxygen isotope exchange with water increases the diagnostic primacy of
33 $\delta^{15}\text{N}$ in resolving hidden turnover. Centering nitrogen-cycle interpretation on nitrite dynamics
34 and isotopic expression across redox gradients from oxic soils to oxygen minimum zones,
35 provides a mechanistic basis for predicting nitrogen budgets, N_2O emissions, and ecosystem
36 sensitivity to increasing redox variability under climate change and land-use intensification.

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39 **Introduction**

40 The nitrogen cycle regulates ecosystem productivity, water quality and climate. Anthropogenic
41 perturbations have profoundly altered nitrogen fluxes across terrestrial and aquatic systems,
42 leading to nitrate (NO_3^-) contamination, eutrophication, and enhanced emissions of nitrous
43 oxide (N_2O), a potent greenhouse gas and ozone-depleting substance (Vitousek et al., 1997;
44 Galloway et al., 2004; Gruber & Galloway, 2008; Callbeck et al., 2026). Uncertainty persists
45 regarding the mechanisms that determine whether reactive nitrogen is retained within
46 ecosystems or transferred to the atmosphere.

47 Conceptual representations of the nitrogen cycle have traditionally emphasized stable pools
48 — ammonium (NH_4^+) and nitrate (NO_3^-) — providing a coherent basis for large-scale budget
49 analyses (Canfield et al., 2010). However, the location within the reaction sequence at which
50 pathway divergence is determined remains unresolved.

51 In reaction networks governed by rapid kinetics and tight coupling, the apparent invisibility of
52 intermediates reflects rapid turnover rather than limited functional relevance. Nitrite (NO_2^-)
53 exemplifies this conceptual blind spot. Described as an ephemeral intermediate that does not
54 accumulate under steady-state conditions (Heil et al., 2016; Wrage et al., 2001), it is often
55 omitted from conceptual frameworks or treated implicitly. This assumption is not consistent
56 with the structure of the nitrogen cycle reaction network.

57 Nitrite is the central stable dissolved intermediate linking oxidative and reductive nitrogen
58 transformations and the obligatory precursor to downstream nitrate, ammonium and gaseous
59 nitrogen products (Burgin & Hamilton, 2007; Lam & Kuypers, 2011; Kraft et al., 2014). Despite
60 NO also participates in both oxidative and reductive branches of the nitrogen cycle it is
61 extremely short-lived in aqueous solution and does not accumulate to measurable
62 concentrations under environmental conditions. No other inorganic nitrogen compound
63 combines this dual role of convergence and divergence. The fate of nitrogen at this junction is
64 governed by the balance between nitrite-producing and nitrite-consuming processes under
65 specific redox, kinetic and environmental constraints (Firestone & Davidson, 1989; Denk et
66 al., 2017; Deb et al., 2024).

67 Concentration measurements of nitrate, ammonium and gaseous products integrate multiple
68 processes, masking mechanistic controls on pathway partitioning: similar nitrate
69 concentrations can arise from fundamentally different combinations of nitrification,
70 denitrification and DNRA (e.g. in riparian zones where both processes operate
71 simultaneously), leading to divergent outcomes in nitrogen retention and N_2O emissions.

72 Stable isotope measurements provide direct constraints on these hidden dynamics. Nitrite
73 integrates oxidative and reductive fluxes; its isotopic composition records concurrent
74 production and consumption even when net concentrations remain unchanged, though
75 disentangling these overlapping signals requires the dual $\delta^{15}\text{N}$ – $\delta^{18}\text{O}$ approach described in

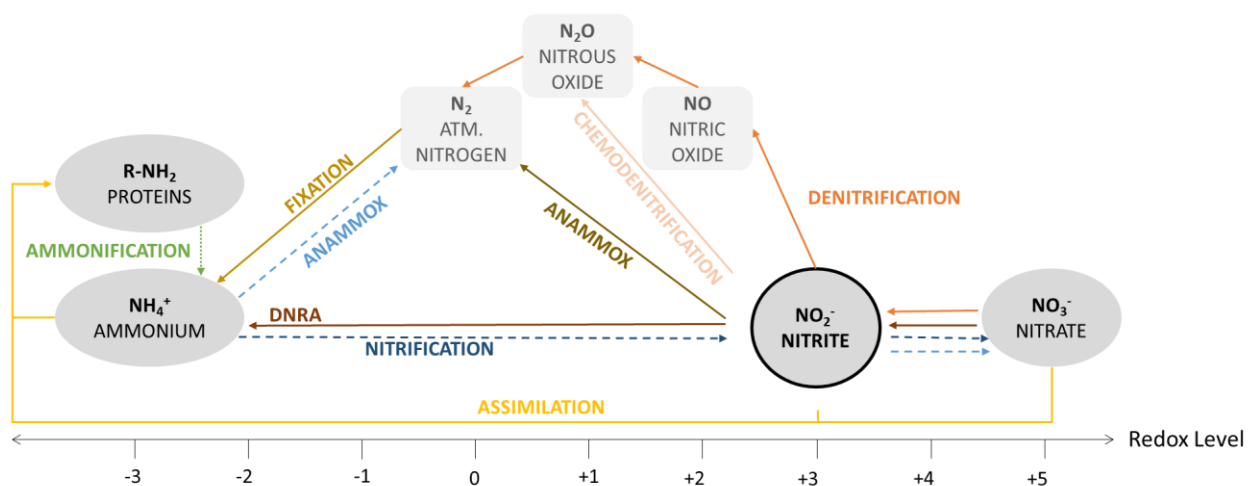
76 detail in the Isotopic constraints section below. Available methods advances permit direct
 77 determination of natural-abundance $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$ in nitrite (McIlvin & Altabet, 2005; Sebiló et
 78 al., 2019; Deb & Lewicka-Szczebak, 2025; Hu et al., 2026). Oxygen isotopes of nitrite are
 79 partially reset due to equilibrium with oxygen isotopes of ambient water, modifying the primary
 80 biological signal and thereby increasing the diagnostic importance of $\delta^{15}\text{N}$.
 81 Resolving nitrogen fate requires shifting analytical focus from accumulated pools to the
 82 intermediate at which pathway divergence is decided. The reaction network structure, kinetics
 83 of nitrite turnover, and stable isotope constraints together provide a mechanistic basis for
 84 predicting nitrogen budgets and N_2O emissions. This basis relies on considering nitrite as a
 85 central component of the nitrogen cycle rather than a transient residual.

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88 Nitrite as the structural branching node

89 The central role of nitrite emerges directly from the reaction network architecture (Fig. 1).
 90 Nitrite does not constitute a regulatory control point in a biochemical sense, but rather the
 91 reaction network node at which pathway partitioning is resolved. Under oxic conditions, nitrite
 92 is produced during ammonium oxidation by ammonia-oxidizing bacteria and archaea and
 93 subsequently oxidized to nitrate, or diverted into alternative pathways depending on oxygen
 94 availability and enzyme kinetics (Casciotti, 2016). In comammox organisms, both oxidation
 95 steps occur within a single metabolic framework (Daims et al., 2015; van Kessel et al., 2015),
 96 further constraining accumulation while maintaining high gross turnover.

97



98

99 **Figure 1. Nitrite-based nitrogen cycling reactions across redox gradients.** Oxidative pathways are
 100 presented in blue arrows, reductive ones in orange/brown and those involving organic matter in
 101 yellow/green. To simplify the scheme, hydroxylamine has not been included in the figure.
 102 Hydroxylamine is also involved in oxidative and reductive pathways. It can be produced during ammonia
 103 oxidation and nitrite reduction. It is primarily oxidized by nitrifying microorganisms to nitrite, with minor

104 side production of nitric oxide and nitrous oxide, and it can be reduced to ammonium. However, it is
105 highly reactive, it is usually rapidly oxidized before accumulating in the environment, limiting its influence
106 at the ecosystem scale.

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109 Under oxygen-limited or anoxic conditions, nitrite is produced during nitrate reduction and
110 partitioned among denitrification, DNRA, anammox, and abiotic reduction. Each pathway
111 channels nitrite toward distinct fates, from nitrogen retention to irreversible gaseous loss
112 (Zumft, 1997; Lam & Kuypers, 2011). Abiotic reactions extend this network: reduction of nitrite
113 by ferrous iron generates NO and N₂O independently of enzymatic control, competing with
114 biological pathways (Jones et al., 2015; Grabb et al., 2017; Robinson et al., 2021).

115 The defining characteristic of this convergence-divergence architecture is redistribution, not
116 accumulation. Nitrite operates as a flux junction, a node at which inputs from multiple upstream
117 pathways are portioned among competing downstream transformations, so that its
118 concentration reflects the balance between upstream formation and downstream
119 consumption, while its turnover rate determines the direction and magnitude of nitrogen
120 transfer. In spatially heterogeneous environments, nitrite produced in one microdomain may
121 be consumed in an adjacent zone within short diffusion distances (Firestone & Davidson,
122 1989). Accumulation arises primarily when this coupling is disrupted by kinetic limitation, redox
123 fluctuation, or imbalance in electron donor and acceptor supply.

124

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126 **Kinetic control of nitrite turnover and gaseous nitrogen speciation**

127 If the structural position of nitrite determines where pathway divergence occurs, the kinetic
128 balance between gross production and gross consumption determines how. The direction and
129 magnitude of nitrogen redistribution at the nitrite node depend on the kinetic balance between
130 gross production and gross consumption. When these proceed at comparable rates, net
131 concentration change approaches zero while flux through the intermediate remains
132 substantial; pool size therefore provides limited information about pathway activity (Margalef-
133 Marti et al., 2026). Such tightly coupled nitrite turnover has recently been observed in nitrate-
134 rich groundwater systems using combined isotope and microbial approaches (Deb et al.,
135 2025).

136 Accumulation and persistence along time of nitrite is determined by the degree of kinetic
137 coupling between sequential reactions. Tight coupling in oxic systems constrains
138 accumulation and shortens residence time; disruption by fluctuating oxygen supply, transport
139 limitation, or electron donor imbalance prolongs residence time and increases the probability
140 of accumulation (see Sensitivity across environmental gradients section). Shifts in

141 environmental conditions reorganize flux distribution at the nitrite node even when nitrate or
142 ammonium pools exhibit minimal change.

143 Gaseous nitrogen (i.e., NO, N₂O and N₂) production represents a relevant downstream
144 outcome of nitrite turnover. In microbial denitrification, the relative gross rates of successive
145 reductions from nitrite to NO, N₂O, and ultimately N₂, determine the N₂O:N₂ ratio. Sustained
146 electron donor supply (such as organic matter) and active nitrous oxide reductase favour
147 completion of the sequence and N₂ dominance; partial decoupling enhances expression of
148 intermediate products, particularly N₂O (Firestone & Davidson, 1989; Zumft, 1997; Lewicka-
149 Szczebak et al., 2020). During oxygen-limited nitrification, nitrite generated from ammonium
150 oxidation may be partially reduced within ammonia oxidizers, producing N₂O under fluctuating
151 oxygen conditions (Wrage et al., 2001). Abiotic reduction of nitrite by ferrous iron and reduced
152 mineral phases generates NO and N₂O independently of enzymatic control (Jones et al., 2015;
153 Grabb et al., 2017).

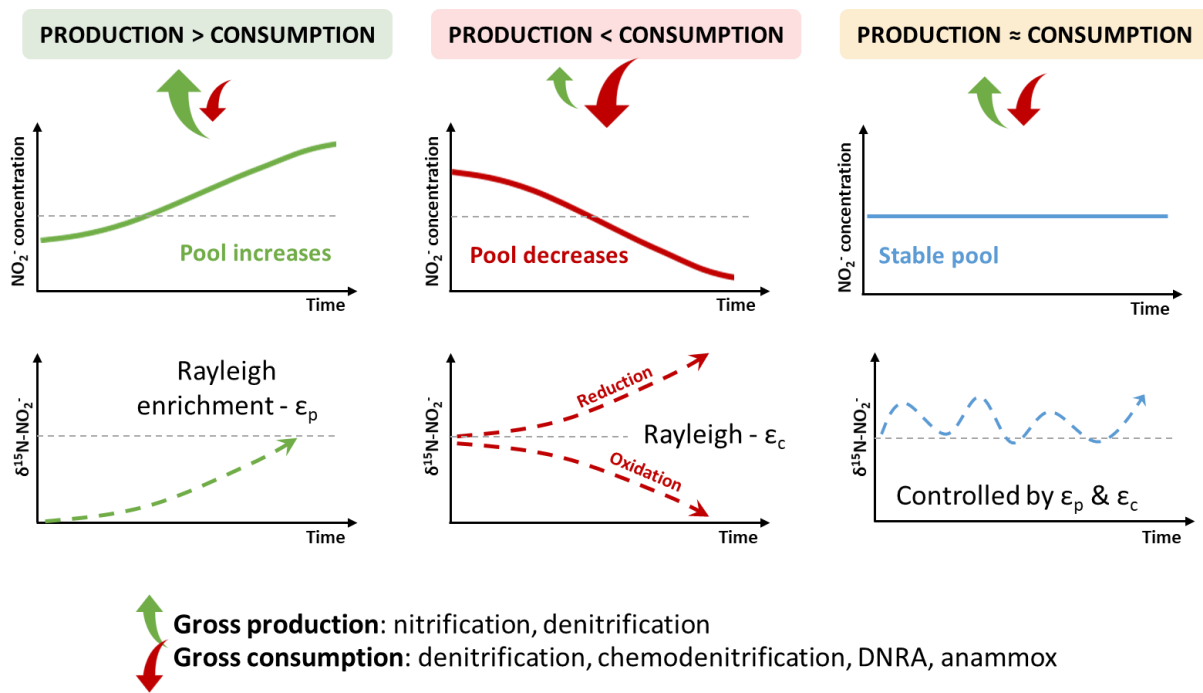
154 Although N₂O is not a primary product of canonical anammox metabolism — hydrazine
155 oxidation yields N₂ directly — anammox competes with denitrifiers for the available nitrite
156 pool, thereby influencing the partitioning of nitrite between N₂-producing and N₂O-producing
157 pathways in mixed-metabolism environments (Kartal et al., 2011). Therefore, the N₂O:N₂ ratio,
158 and the relative production of N₂ versus N₂O more broadly, is controlled by turnover intensity
159 and kinetic coupling at the nitrite stage rather than by pool size.

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162 **Isotopic constraints on gross nitrite turnover**

163 Stable isotope measurements of nitrite provide direct constraints on nitrogen transformations
164 at the stage where flux redistribution occurs: the nitrite node. Isotopic composition responds
165 to gross production and gross consumption rather than to net pool change alone. Constant
166 concentration does not imply constant $\delta^{15}\text{N}$: progressive isotopic shifts may occur under
167 steady-state pool conditions, revealing turnover intensity undetectable in concentration data
168 (Fig. 2).

169



Note: δ¹⁸O-NO₂⁻ influenced by equilibration with δ¹⁸O-H₂O

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171 **Figure 2. Isotopic expression of gross nitrite turnover under contrasting production–**
 172 **consumption regimes.** Top panels show nitrite concentration ([NO₂⁻]) as a function of time, reflecting
 173 the net balance between gross production (P) and gross consumption (C): accumulation when P > C,
 174 depletion when P < C, and quasi-steady state when P ≈ C despite high internal fluxes. Bottom panels
 175 illustrate conceptual trajectories of δ¹⁵N(NO₂⁻). In contrast to concentration, isotopic composition
 176 responds to gross turnover rather than net change. δ¹⁵N(NO₂⁻) evolves according to the isotopic
 177 signatures of contributing sources and the combined effects of isotope fractionation during production
 178 (ε_P) and consumption (ε_C). The direction of isotopic change is therefore process-dependent and not
 179 universal (e.g. inverse isotope effects during nitrite production by nitrification). Under conditions of
 180 balanced production and consumption (P ≈ C), δ¹⁵N(NO₂⁻) may drift despite constant concentration,
 181 reflecting ongoing gross turnover. Oxygen isotope signals (δ¹⁸O) may be partially overprinted by
 182 exchange with water, increasing the diagnostic primacy of δ¹⁵N.

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185 Each pathway imposes a characteristic kinetic isotope fractionation. During ammonia
 186 oxidation, inverse nitrogen isotope effects may enrich newly formed nitrite relative to its
 187 ammonium source (Casciotti et al., 2003; Santoro & Casciotti, 2011). During nitrite oxidation
 188 to nitrate, preferential removal of lighter isotopes enriches the residual pool in both ¹⁵N and
 189 ¹⁸O (Casciotti, 2009; Buchwald et al., 2012). Reductive pathways generate distinct
 190 fractionation patterns associated with nitrite reductase activity (Brunner & Bernasconi, 2005;
 191 Casciotti et al., 2010). Abiotic reduction by ferrous iron produces additional nitrogen isotope
 192 effects governed by surface-mediated electron transfer (Jones et al., 2015; Grabb et al., 2017).
 193 Overlapping fractionation factors across biological and abiotic pathways preclude simple end-
 194 member mixing and require integration with environmental and redox context. The direction of
 195 δ¹⁵N-NO₂⁻ evolution is not universal and depends on the relative contributions of concurrent

196 production and consumption pathways, their associated isotope effects (ϵ_P , ϵ_C), and the
197 isotopic composition of the source substrate.

198 Oxygen isotopes introduce an additional dimension. Exchange between nitrite oxygen atoms
199 and ambient water may proceed on timescales comparable to biological turnover (Buchwald
200 & Casciotti, 2010; Casciotti et al., 2010; Granger & Wankel, 2016), attenuating the
201 independence of $\delta^{18}\text{O}$ as a pathway tracer. Nitrogen isotopes do not undergo analogous
202 exchange, preserving sensitivity to pathway-specific fractionation. Under conditions of
203 significant oxygen isotope exchange, $\delta^{15}\text{N}$ provides the more robust constraint on gross
204 turnover.

205 Despite their strong mechanistic potential, nitrite isotope measurements are only feasible
206 when sufficient nitrite accumulates for analysis and remain analytically challenging. Nitrite is
207 chemically unstable and may undergo rapid oxidation or reduction during sampling and
208 storage, potentially altering its isotopic composition before analysis. Reliable preservation
209 therefore requires rapid processing and carefully controlled storage conditions, often under
210 alkaline conditions that minimize transformation rates. In many terrestrial and aquatic systems,
211 low nitrite concentrations restrict isotope analyses to zones where nitrite transiently
212 accumulates. This emphasizes the need for continued development of high-sensitivity
213 analytical approaches (Lewicka-Szczebak et al., 2021; Deb and Lewicka-Szczebak, 2025).
214 For instance, a recent study developed a method for freshwater samples by coupling anion-
215 exchange resin preconcentration with the azide reduction method for nitrite isotopic
216 characterization at concentrations of $0.02 \mu\text{mol L}^{-1}$ or higher (Jiang et al., 2026). These
217 analytical limitations likely contribute to the still limited application of nitrite isotope
218 measurements in environmental studies despite their considerable interpretative potential.

219 Isotopic signatures of nitrite propagate to gaseous products. The bulk $\delta^{15}\text{N}$ of N_2O reflects
220 fractionation associated with nitrite reductases and kinetic coupling among successive
221 reduction steps. Intramolecular ^{15}N site preference (SP) provides additional mechanistic
222 resolution: because SP is largely independent of the isotopic composition of precursor nitrite,
223 it records enzyme-specific reaction pathways involved in N_2O formation (Brunner &
224 Bernasconi, 2005; Toyoda et al., 2017). Variations in SP discriminate among nitrifier-
225 denitrification, canonical denitrification, and partial reduction sequences downstream of nitrite.
226 Isotopomer signatures must account for subsequent N_2O reduction and mixing, which may
227 overprint primary fractionation signals (Toyoda et al., 2017; Lewicka-Szczebak et al., 2020).

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229

230 **Sensitivity across environmental gradients**

231 Environmental systems differ not in whether nitrite turnover operates, but in how sensitively
232 flux redistribution at the nitrite node responds to perturbation. Sensitivity is highest where

233 oxidative and reductive metabolisms overlap at oxic-anoxic transition zones, and attenuates
234 where reaction coupling is constrained to a single redox regime. Small shifts in oxygen supply,
235 electron donor availability, or hydrological residence time can reorganize pathway partitioning
236 at the nitrite node without producing detectable changes in bulk nitrogen pools. The nitrogen
237 cycle is therefore most vulnerable to mechanistic misinterpretation precisely where nitrite
238 turnover is most intense.

239 In fully oxic environments, nitrite is produced during ammonia oxidation and rapidly oxidized
240 to nitrate, reflecting tight coupling between nitrification steps (Casciotti, 2016). Redox
241 transition zones — oxic–anoxic interfaces in soils, sediments, riparian zones and stratified
242 water columns — represent the most dynamic settings (Burgin et al., 2011). Simultaneous
243 production from ammonia oxidation and nitrate reduction, combined with kinetically
244 constrained consumption, promotes nitrite accumulation and amplifies sensitivity to
245 environmental change (Buchwald & Casciotti, 2010; Bristow et al., 2016). These zones are
246 recognized as hotspots of N₂O production, consistent with the central role of nitrite in
247 regulating the N₂O:N₂ ratio (Butterbach-Bahl et al., 2013; Babbin et al., 2020). In persistently
248 anoxic environments — deep sediments, saturated soils and oxygen minimum zones — nitrite
249 is predominantly generated via nitrate reduction and consumed through denitrification, DNRA
250 or anammox (Zumft, 1997; Lam & Kuypers, 2011; Ward et al., 2009; Dalsgaard et al., 2012;
251 Kalvelage et al., 2013; Denk et al., 2017; Deb et al., 2024). Hydrological and transport
252 processes further modulate this coupling across all settings: long residence times favor
253 complete turnover, whereas rapid transport can decouple production from consumption,
254 allowing accumulation or downstream export (Sebilo et al., 2006).

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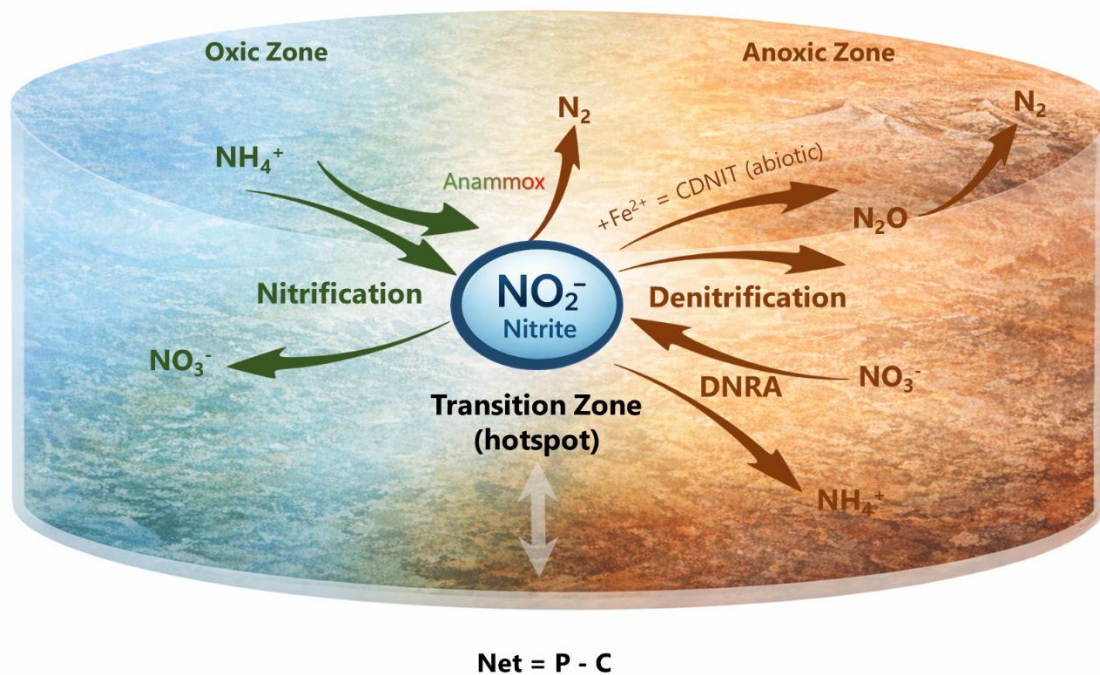
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257 **Nitrite as the control point of nitrogen fate**

258 Nitrogen cycling is commonly interpreted through the distribution of dominant inorganic pools.
259 This perspective captures accumulation and export, it does not identify where transformation
260 trajectories are decided. The reaction network places that decision at the stage of nitrite
261 turnover. Environmental conditions regulate this distribution by modulating the rates of nitrite
262 production and consumption.

263 This reframing resolves several persistent ambiguities. Similar nitrate or ammonium
264 concentrations can arise from fundamentally different internal configurations of production and
265 consumption. N₂O emissions cannot be predicted from pool size alone because their
266 magnitude depends on the completeness of reduction downstream of nitrite. Redox transition
267 zones emerge as hotspots not because they contain larger pools, but because they intensify
268 flux redistribution at this intermediate. A nitrite-centred framework shifts emphasis from
269 storage to flux (Fig. 3), from accumulation to coupling, and from static pools to dynamic

270 turnover, linking microbial metabolism, abiotic reactivity, redox heterogeneity, and
 271 atmospheric exchange within a single mechanistic perspective. Because both production and
 272 reduction of N_2O depend on nitrite availability and turnover, resolving dynamics at this node
 273 provides a direct mechanistic link between microbial processes and climate-relevant gas
 274 fluxes.



275
 276 **Figure 3. Conceptual synthesis positioning nitrite turnover as the control point linking redox**
 277 **gradients, gross flux redistribution, isotopic expression, and nitrogen fate.**
 278 Nitrite integrates oxidative and reductive processes across environmental gradients. Gross production
 279 and gross consumption determine residence time and flux partitioning among retention, recycling, and
 280 gaseous loss. The $\delta^{15}N$ of NO_2^- records turnover intensity, whereas the $\delta^{18}O$ may be modified by
 281 exchange with water. Isotopic signals propagate to N_2O and inform pathway attribution. Nitrogen fate
 282 is therefore governed at the nitrite stage rather than by the size of accumulated nitrate or ammonium
 283 pools.
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288 Concentration reflects net balance between production and consumption. $\delta^{15}\text{N}$ records the
289 imprint of concurrent production and consumption even when pool size remains constant.
290 Oxygen isotope exchange constrains interpretation but reinforces the diagnostic primacy of
291 nitrogen isotopes. At the same time, the extent of oxygen exchange with water may provide
292 information on nitrite residence times and the relative rates of biological turnover versus abiotic
293 exchange (Buchwald and Casciotti, 2013; Lewicka-Szczebak et al., 2021). Incorporating nitrite
294 isotopes converts an otherwise unobservable internal turnover into a measurable quantity.
295 Translating this framework into practice requires prioritising direct nitrite isotope
296 measurements alongside conventional concentration measurements, particularly in redox-
297 dynamic systems where net fluxes are most ambiguous. Dual $\delta^{15}\text{N}$ – $\delta^{18}\text{O}$ approaches,
298 combined with isotopomer analysis of N_2O , offer the most diagnostic power. Although nitrite
299 isotope measurements offer strong mechanistic insight, their application is constrained by low
300 concentrations and analytical challenges. Recent advances in high-sensitivity methods are
301 expanding their feasibility in natural systems. Nonetheless, these limitations continue to
302 restrict their widespread use in environmental studies.
303 At the modelling scale, representing nitrite explicitly as a state variable, rather than collapsing
304 nitrification and denitrification into net transformations, would improve mechanistic fidelity in
305 biogeochemical models applied to nitrogen budgets and greenhouse gas inventories. The
306 measurement and modelling strategies that follow from this reorientation are those organised
307 around the node where nitrogen fate is actually decided.
308 As environmental systems experience increasing redox variability under climate change and
309 land-use intensification, sensitivity at the nitrite stage is likely to amplify. Understanding
310 nitrogen cycling at Earth-system scale requires resolving where and how nitrite flux is
311 redistributed. Without explicit consideration of turnover at this branching node, interpretations
312 of nitrogen budgets, greenhouse gas emissions, and isotopic signals remain incomplete.

313

314

315 **Author contribution**

316 MS and RM jointly conceived the perspective and wrote the manuscript.

317

318 **Competing interests**

319 The authors declare that they have no conflict of interest.

320

321 **Data and code availability**

322 No new data or code were generated for this study. The perspective is based exclusively on
323 bibliographic sources, all of which are cited in the reference list.

324

325 **References**

- 326 Babbin, A. R., Buchwald, C., Morel, F. M. M., Wankel, S. D., and Ward, B. B.: Nitrite oxidation
327 exceeds reduction and fixed nitrogen loss in anoxic Pacific waters, *Mar. Chem.*, 224, 103814,
328 2020.
- 329
- 330 Bristow, L. A., Dalsgaard, T., Tiano, L., Mills, D. B., Bertagnolli, A. D., Wright, J. J., Hallam, S.
331 J., Ulloa, O., Canfield, D. E., and Revsbech, N. P.: Ammonium and nitrite oxidation at
332 nanomolar oxygen concentrations in oxygen minimum zone waters, *Proc. Natl Acad. Sci.*
333 *USA*, 113, 10601–10606, 2016.
- 334
- 335 Brunner, B., and Bernasconi, S. M.: A revised isotope fractionation model for dissimilatory
336 sulfate reduction in sulfate reducing bacteria, *Geochim. Cosmochim. Acta*, 69, 4759–4771,
337 2005.
- 338
- 339 Buchwald, C., and Casciotti, K. L.: Oxygen isotopic fractionation and exchange during
340 bacterial nitrite oxidation, *Limnol. Oceanogr.*, 55, 1064–1074, 2010.
- 341
- 342 Buchwald, C., and Casciotti, K. L.: Isotopic ratios of nitrite as tracers of the sources and age
343 of oceanic nitrite, *Nature Geoscience*, 6(4), 308–313, 2013.
- 344 Buchwald, C., Santoro, A. E., McIlvin, M. R., and Casciotti, K. L.: Oxygen isotopic composition
345 of nitrate and nitrite produced by nitrifying cocultures and natural marine assemblages, *Limnol.*
346 *Oceanogr.*, 57, 1361–1375, 2012.
- 347
- 348 Burgin, A. J., and Hamilton, S. K.: Have we overemphasized the role of denitrification in
349 aquatic ecosystems? A review of nitrate removal pathways, *Front. Ecol. Environ.*, 5, 89–96,
350 2007.
- 351
- 352 Burgin, A. J., Yang, W. H., Hamilton, S. K., and Silver, W. L.: Beyond carbon and nitrogen:
353 how the microbial energy economy couples elemental cycles in diverse ecosystems, *Front.*
354 *Ecol. Environ.*, 9, 44–52, 2011.
- 355
- 356 Butterbach-Bahl, K., Baggs, E. M., Dannenmann, M., Kiese, R., and Zechmeister-Boltenstern,
357 S.: Nitrous oxide emissions from soils: how well do we understand the processes and their
358 controls?, *Philos. Trans. R. Soc. B*, 368, 20130122, 2013.
- 359

360 Callbeck, C.M., Mazzoli, A.P., Paulus, T.J., Frey, C., Bürgmann, H., Schuber, C.J., and
361 Lehmann, M.F.: Seasonality of lake microbial denitrification and its sensitivity to climate
362 warming, *Nature microbiology*, 2026.
363
364 Canfield, D. E., Glazer, A. N., and Falkowski, P. G.: The evolution and future of Earth's
365 nitrogen cycle, *Science*, 330, 192–196, 2010.
366
367 Casciotti, K. L.: Inverse kinetic isotope fractionation during bacterial nitrite oxidation, *Geochim.
368 Cosmochim. Acta*, 73, 2061–2076, 2009.
369
370 Casciotti, K. L.: Nitrogen and oxygen isotopic studies of the marine nitrogen cycle, *Annu. Rev.
371 Mar. Sci.*, 8, 379–407, 2016.
372
373 Casciotti, K. L., McIlvin, M. R., and Buchwald, C.: Oxygen isotopic exchange and fractionation
374 during bacterial ammonia oxidation, *Limnol. Oceanogr.*, 55, 753–762, 2010.
375
376 Casciotti, K. L., Sigman, D. M., Hastings, M. G., Böhlke, J. K., and Hilkert, A.: Measurement
377 of the oxygen isotopic composition of nitrate in seawater and freshwater using the denitrifier
378 method, *Anal. Chem.*, 75, 4905–4912, 2003.
379
380 Daims, H., Lebedeva, E. V., Pjevac, P., Han, P., Herbold, C., Albertsen, M., Jehmlich, N.,
381 Palatinszky, M., Vierheilig, J., Bulaev, A., Kirkegaard, R. H., von Bergen, M., Rattei, T.,
382 Bendinger, B., Nielsen, P. H., and Wagner, M.: Complete nitrification by *Nitrospira* bacteria,
383 *Nature*, 528, 504–509, 2015.
384
385 Dalsgaard, T., Thamdrup, B., Farías, L., and Revsbech, N. P.: Nitrogen cycling in a deep-
386 water oxygen-deficient zone of the Arabian Sea, *Limnol. Oceanogr.*, 57, 1331–1346, 2012.
387
388 Deb, S. and Lewicka-Szczebak, D.: Simplified bacterial denitrification method using
389 *Stenotrophomonas nitritireducens* for nitrite dual isotope analysis in low-concentration
390 environmental samples, *Front. Environ. Sci.* 13:1536882, 2025.
391
392 Deb, S., Espenberg, M., Well, R., Bucha, M., Jakubiak, M., Mander, Ü., Jędrysek, M.O.,
393 Lewicka-Szczebak, D.: N transformations in nitrate-rich groundwaters: combined isotope and
394 microbial approach, *Biogeosciences*, 22 (19), 5535-5556, 2025.
395

396 Deb, S., Lewicka-Szczebak, D., and Rohe, L.: Microbial nitrogen transformations tracked by
397 natural abundance isotope studies and microbiological methods: a review, *Sci. Total Environ.*,
398 926, 172073, 2024.
399

400 Denk, T.R.A., Mohn, J., Decock, C., Lewicka-Szczebak, D., Harris, E., Butterbach-Bahl, K.,
401 Kiese, R., Wolf, R.: The nitrogen cycle: A review of isotope effects and isotope modeling
402 approaches, *Soil Biol. Biochem.*, 105, 121-137, 2017.
403

404 Firestone, M. K., and Davidson, E. A.: Microbiological basis of NO and N₂O production and
405 consumption in soil, in: *Exchange of Trace Gases between Terrestrial Ecosystems and the*
406 *Atmosphere*, edited by: Andreae, M. O. and Schimel, D. S., Wiley, New York, 7–21, 1989.
407

408 Galloway, J. N., Dentener, F. J., Capone, D. G., Boyer, E. W., Howarth, R. W., Seitzinger, S.
409 P., Asner, G. P., Cleveland, C. C., Green, P. A., Holland, E. A., Karl, D. M., Michaels, A. F.,
410 Porter, J. H., Townsend, A. R., and Vörösmarty, C. J.: Nitrogen cycles: past, present, and
411 future, *Biogeochemistry*, 70, 153–226, 2004.

412 Grabb, K. C., Buchwald, C., Hansel, C. M., and Wankel, S. D.: A dual nitrite isotopic
413 investigation of chemodenitrification by mineral-associated Fe(II) and its production of nitrous
414 oxide, *Geochim. Cosmochim. Acta*, 196, 388–402, 2017.
415

416 Granger, J., and Wankel, S. D.: Isotopic overprinting of nitrification on denitrification as a
417 ubiquitous and unifying feature of environmental nitrogen cycling, *Proc. Natl Acad. Sci. USA*,
418 113, E6391–E6400, 2016.
419

420 Gruber, N., and Galloway, J. N.: An Earth-system perspective of the global nitrogen cycle,
421 *Nature*, 451, 293–296, 2008.
422

423 Heil, J., Vereecken, H., and Brüggemann, N.: A review of chemical reactions of nitrification
424 intermediates and their role in nitrogen cycling and nitrogen trace gas formation in soil, *Eur. J.*
425 *Soil Sci.*, 67, 23–39, 2016.
426

427 Hu, J., Yao, S.J., Li, Y., Ren, H., Yang, K., and Liu, W.: Accurate isotopic analysis of nitrate
428 and nitrite in freshwater: integration of ion exchange and bacterial denitrification methods, *J.*
429 *Anal. At. Spectrom.*, 41, 1774-1783, 2026.
430

431 Jiang, M., Koba, K., Ono, M., Hayashi, K.: Improved Isotopic Analysis of Low-Concentration
432 Freshwater Nitrite by Anion-Exchange Resin Enrichment and Azide Reduction, *Anal. Chem.*
433 98, 4, 2956–2967, 2026.
434

435 Jones, L. C., Peters, B., Pacheco, J. S. L., Casciotti, K. L., and Fendorf, S.: Stable isotopes
436 and iron oxide mineral products as markers of chemodenitrification, *Environ. Sci. Technol.*,
437 49, 3444–3452, 2015.
438

439 Kalvelage, T., Jensen, M. M., Contreras, S., Revsbech, N. P., Lam, P., Günter, M., LaRoche,
440 J., Lavik, G., and Kuypers, M. M. M.: Nitrogen cycling driven by organic matter export in the
441 South Pacific oxygen minimum zone, *Nat. Geosci.*, 6, 228–234, 2013.
442

443 Kartal, B., Kuenen, J. G., and van Loosdrecht, M. C. M.: Molecular mechanism of anaerobic
444 ammonium oxidation, *Nature*, 479, 127–130, 2011.
445

446 Kraft, B., Strous, M., and Tegetmeyer, H. E.: Microbial nitrate respiration — genes, enzymes
447 and environmental distribution, *J. Biotechnol.*, 155, 104–117, 2011.
448

449 Lam, P., and Kuypers, M. M. M.: Microbial nitrogen cycling processes in oxygen minimum
450 zones, *Annu. Rev. Mar. Sci.*, 3, 317–345, 2011.
451

452 Lewicka-Szczebak, D., Lewicki, M. P., and Well, R.: N₂O isotope approaches for source
453 partitioning of N₂O production and estimation of N₂O reduction, *Biogeosciences*, 17, 5513–
454 5537, 2020.
455

456 Lewicka-Szczebak, D., Jansen-Willems, A., Müller, C., Dyckmans, J., and Well, R.: Nitrite
457 isotope characteristics and associated soil N transformations, *Sci. Rep.*, 11, 5008, 2021.
458

459 Margalef-Marti, R., Bourbonnais, A., Knöller, K., Mayer, B., Altabet, M., and Sebiló, M. Using
460 N and O isotope fractionation for evaluating denitrification in aquatic systems, *TrAC Trends*
461 *Anal. Chem.*, 194, 118527, 2026.
462

463 McIlvin, M. R., and Altabet, M. A.: Chemical conversion of nitrate and nitrite to nitrous oxide
464 for nitrogen and oxygen isotopic analysis in freshwater and seawater, *Anal. Chem.*, 77, 5589–
465 5595, 2005.
466

467 Robinson, T. C., Latta, D.E., Notini, L., Schilling, K.E. and Scherer, M.M.: Abiotic reduction of
468 nitrite by Fe(II): a comparison of rates and N₂O production, *Environ. Sci. Process. Impacts*,
469 23, 1531–1541, 2021.

470

471 Santoro, A. E., and Casciotti, K. L.: Enrichment and characterization of ammonia-oxidizing
472 archaea from the open ocean, *ISME J.*, 5, 1796–1808, 2011.

473

474 Sebilo, M., Billen, G., Grably, M., and Mariotti, A.: Isotopic composition of nitrate-nitrogen as
475 a marker of riparian and benthic denitrification at the scale of the whole Seine River system,
476 *Biogeochemistry*, 75, 35–51, 2006.

477

478 Sebilo, M., Aloisi, G., Mayer, B., Perrin, E., Vaury, V., Mothet, A. and Laverman, A.M.: Controls
479 on the isotopic composition of nitrite ($\delta^{15}\text{N}$ and $\delta^{18}\text{O}$) during denitrification in freshwater
480 sediments, *Scientific Reports*, 9, 2019.

481

482 Toyoda, S., Yoshida, N., and Koba, K.: Isotopocule analysis of biologically produced nitrous
483 oxide in various environments, *Mass Spectrom. Rev.*, 36, 135–160, 2017.

484

485 van Kessel, M. A. H. J., Speth, D. R., Albertsen, M., Nielsen, P. H., Op den Camp, H. J. M.,
486 Kartal, B., Jetten, M. S. M., and Lückner, S.: Complete nitrification by a single microorganism,
487 *Nature*, 528, 555–559, 2015.

488

489 Vitousek, P. M., Aber, J. D., Howarth, R. W., Likens, G. E., Matson, P. A., Schindler, D. W.,
490 Schlesinger, W. H., and Tilman, D. G.: Human alteration of the global nitrogen cycle: sources
491 and consequences, *Ecol. Appl.*, 7, 737–750, 1997.

492

493 Ward, B. B., Devol, A. H., Rich, J. J., Chang, B. X., Bulow, S. E., Naik, H., Pratihary, A., and
494 Jayakumar, A.: Denitrification as the dominant nitrogen loss process in the Arabian Sea,
495 *Nature*, 461, 78–81, 2009.

496

497 Wrage, N., Velthof, G. L., van Beusichem, M. L., and Oenema, O.: Role of nitrifier
498 denitrification in the production of nitrous oxide, *Soil Biol. Biochem.*, 33, 1723–1732, 2001.

499

500 Zumft, W. G.: Cell biology and molecular basis of denitrification, *Microbiol. Mol. Biol. Rev.*, 61,
501 533–616, 1997.

502