



1 Title Page

2 **Biochar addition enhances soil aggregate stability by**
3 **modulating soil internal forces: The critical role of**
4 **readily oxidized organic carbon and particulate**
5 **organic carbon**

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49 **Abstract:** Degradation of soil structure and function is a significant obstacle to
50 sustainable agriculture, yet using biochar is known to be an effective way of
51 increasing soil organic carbon and boosting structure. Aggregate stability is greatly
52 influenced by soil internal forces (SIFs), such as van der Waals, hydration, and
53 electrostatic forces. However, it is still not clear which specific organic carbon
54 components play a significant role in the stability of aggregates and the action of soil
55 internal forces. In this study, a field simulation experiment was conducted using maize
56 straw and wheat straw biochars to investigate how biochar addition affect soil
57 aggregate stability. The findings showed that as the rate of biochar application
58 increased from 0% to 10% over 180 day and 365 days, the contents of soil light
59 fraction organic carbon (LFOC), particulate organic carbon (POC), dissolved organic
60 carbon (DOC), readily oxidized organic carbon (ROC), as well as soil specific surface
61 area and surface charge number, increased. The content of different carbon fractions
62 decreased with prolonged incubation time, while soil electrochemical parameters
63 continued to increase throughout the incubation period. Pearson correlation and
64 redundancy analyses showed that soil active organic carbon fractions were strongly
65 correlated with soil surface electrochemical properties, with ROC and POC being the
66 primary factors. Soil aggregate stability increased with higher biochar application
67 rates, and the beneficial effect of biochar on aggregate stability became increasingly
68 pronounced over time. Based on the quantitative estimates of SIFs, the addition of
69 biochar facilitated soil aggregates stability by increasing van der Waals attractive
70 pressure and decreasing electrostatic repulsive pressure, and finally lowering net



71 repulsive pressure between soil particles. The soil aggregate breaking strength
72 increased quickly at initially and then leveled out as the electrolyte content decreased;
73 $10^{-2} \text{ mol L}^{-1}$ marked the tipping point. The study identified the active organic carbon
74 (ROC and POC) fractions which could mediate the soil surface electrochemical
75 properties and soil internal forces, finally enhance soil aggregate stability. Our
76 research provides a theoretical basis for the targeted use and modifying of biochar to
77 efficiently improve soil structure.

78 **Keywords:** Biochar; Active organic carbon fraction; Soil internal forces; Aggregate
79 stability

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81 **1. Introduction**

82 Soil structural stability is a key indicator for assessing soil erodibility, directly
83 influencing the occurrence and evolution of soil erosion processes (An et al., 2010;
84 Siedt et al., 2021). Soil aggregate formation and stabilization play a fundamental role
85 in shaping soil architecture and underpin a wide range of soil functions (Batista et al.,
86 2023; Li et al., 2016; Rabot et al., 2018). Physical characteristics like porosity,
87 aeration, compaction, and water retention, as well as biochemical characteristics like
88 organic carbon fixation, conversion, and microbial activity, are all impacted by
89 aggregate stability (Liu et al., 2022; Tian et al., 2022). Furthermore, both intrinsic soil
90 characteristics—such as particle composition, organic matter content, and clay
91 mineralogy—and external factors—such as weathering, tillage methods, and
92 erosion—have an impact on aggregate stability (Cao et al., 2016; Das et al., 2014;



93 Zhang et al., 2021). Therefore, improving soil aggregate stability is essential for
94 reducing soil erodibility, improving soil health and sustaining agricultural
95 productivity.

96 The application of biochar as a soil amendment is primarily attributed to its
97 ability to boost organic carbon content and contribute to better soil functionality
98 (Cooper et al., 2020; Ji et al., 2024; Lu et al., 2021; Okebalama and Marschner, 2023;
99 Uchenna et al., 2023; Zhang et al., 2020). Numerous studies have demonstrated that
100 biochar have successfully raised soil organic carbon levels and improved soil
101 aggregate stability (Du et al., 2016; Huang et al., 2018; Ma et al., 2016; Sheng et al.,
102 2023; Šimanský et al., 2018; Sun et al., 2021; Wang et al., 2017). However, Blair et al.
103 (1995) concluded that anthropogenic management practices in agricultural systems
104 primarily affect the reactive organic carbon fraction of soil organic carbon. Soil
105 organic matter has been found to degrade slowly, making short-term changes in its
106 content of limited practical significance. As an early and quick sign of ecosystem
107 management techniques, soil active organic carbon, on the other hand, is less plentiful
108 but more sensitive to environmental changes than soil organic matter (Coleman et al.,
109 1983; Stevenson, 1994; Wander et al., 1994).

110 According to Haynes (2005) and Wei et al. (2014), soil active organic carbon
111 represents the most dynamic fraction of soil organic carbon that has the greatest
112 direct impact on plant nutrient supply; it is highly reactive, has a certain solubility
113 in the soil, and is readily oxidized and mineralized by soil microorganisms. Light
114 fraction organic carbon (LFOC), dissolved organic carbon (DOC), particulate



115 organic carbon (POC), and readily oxidized organic carbon (ROC) are the primary
116 forms of soil active organic carbon (Stewart et al., 2009; Yang et al., 2017; Zhao et
117 al., 2018). Despite making up a small fraction of soil organic carbon, soil active
118 carbon has a direct role in soil biochemical transformation processes and can
119 indicate slight variations in soil organic carbon before changes in total soil carbon
120 (Coleman et al., 1983; Wander et al., 1994). Accordingly, variations in the amount
121 of organic carbon's active fraction in agricultural soils significantly impact on the
122 soil's nutrient cycling and overall quality (Liang et al., 2011; Magill and Aber,
123 2000).

124 Overall, increased soil active organic carbon enhances soil aggregate
125 stability (Bronick and Lal, 2005). It has been demonstrated that soil active organic
126 carbon fractions can affect the nutrient content and bioavailability of the soil and
127 have a significant and positive correlation with the formation and stability of soil
128 macroaggregates (Wang et al., 2018). Aggregate stability was shown to increase
129 positively with organic carbon content, mainly in the light organic matter
130 component (Yagüe et al., 2016). In comparison to soils treated with nitrogen,
131 phosphorus, and potassium (NPK) in combination with green manure or NPK
132 alone, Li et al. (2012) found that soils treated with NPK and animal manure had
133 significantly higher levels of biologically active soil organic carbon and aggregate
134 stability. Microaggregates and silt + clay fractions showed a significant negative
135 association with the mean weight diameter and geometric mean diameter of soil
136 aggregates, while DOC, macroaggregates, macroaggregates-C, and



137 microaggregates-C showed a significant positive correlation (Cao et al., 2024). In
138 trials with simulated rainfall, soil aggregate stability rose when soil organic carbon
139 and LFOC levels increased (Li et al., 2021). Although the role of active organic
140 carbon in enhancing soil aggregate stability has been widely studied, the composition
141 and characteristics of its various forms differ substantially, potentially leading to
142 divergent effects on aggregate stability. Therefore, further research is needed to
143 elucidate how different fractions of soil active organic carbon influence the formation
144 and stabilization of soil aggregates.

145 The stability of soil aggregates in water is mostly due to soil internal forces
146 (SIFs), which include electrostatic, hydration, and van der Waals attraction (Hu et al.,
147 2015; Ma et al., 2021b). Theoretically, internal forces may reach hundreds of
148 thousands of atmospheric pressures between two adjacent soil particles (Ding et al.,
149 2019; Ma et al., 2022; Xu et al., 2015). The classical double layer theory states that
150 the characteristics of the soil, including its specific surface area (SSA), surface charge
151 density, and cation exchange capacity, as well as external environmental factors
152 (moisture content, solution concentration, etc.), primarily determine the function of
153 SIFs in mesoscale soils (Li et al., 2013; Quirk, 1994). The addition of organic matter
154 modifies SIFs by altering the interfacial properties of soil particles. In a four-year
155 field experiment, the use of weathered coal and biochar had a significant impact on
156 the SSA and surface charge density of soil particles, as well as improving van der
157 Waals attraction, decreasing net soil pressure, and decreasing aggregate fragmentation
158 strength and splash erosion rate (Wang et al., 2024). Changes in the electrochemical



159 characteristics of the soil surface were mostly caused by an increase in the amount of
160 soil organic matter after vegetation restoration (Liu et al., 2020). Nevertheless, few
161 research have examined how various active organic carbon fractions affect SIFs and
162 aggregate stability by altering the surface properties of the soil particles when biochar
163 is applied.

164 In this study, maize straw and wheat straw biochars were applied at different
165 rates in a field simulation experiment to systematically evaluate their effects on soil
166 internal forces. Specifically, we aimed to (i) quantify the changes in SIFs under
167 different biochar types and application rates, (ii) identify the active organic carbon
168 fractions that most strongly regulate soil interfacial electrochemical properties and
169 thereby influence SIFs, and (iii) elucidate the mechanisms and time effects by which
170 biochar-mediated modifications of SIFs contribute to soil aggregate formation and
171 stabilization. This work provides new mechanistic insights into biochar–soil
172 interactions from the perspective of soil internal forces and offers a scientific basis for
173 targeted soil structural improvement and sustainable agricultural management.

174 **2. Materials and methods**

175 **2.1 Study area**

176 This study was conducted in the Yangling Demonstration Zone, Shaanxi
177 Province, China (108°08'47"E, 34°27'21"N). The region has a temperate continental
178 monsoon semi-humid climate, with an average of 2167 hours of sunlight per year,
179 temperatures between 11 and 13 °C, and 500 to 700 mm of rainfall. According to the
180 FAO soil classification, the soil under study was Lou soil, which is categorized as



181 calcareous Cambisol. The basic physicochemical properties of the soil are shown in
182 Table 1.

183 **2.2 Experimental design**

184 The biochar made from wheat and maize straw that was utilized in this
185 investigation was acquired from Zhengzhou Jinbang Environmental Protection
186 Technology Co. An elemental analyzer (Elementar Vario EL III, Germany) was used
187 to determine the elemental composition of the biochar samples. A fully automated
188 specific surface and porosity analyzer (Micromeritics ASAP 2460, USA) was used to
189 measure the SSA of biochar. A Mettler pH meter with a solid-liquid ratio of 1: 20 was
190 used to measure the pH. The amount of ash was measured by placing the biochar in a
191 crucible and burning it for two hours at 800 °C. Table 2 displays the fundamental
192 physical and chemical characteristics of biochar.

193 In mid-December 2021, biochar was added to the top layer of soil (0–20 cm) and
194 tilled lightly until it was incorporated into the soil. In a control treatment (CK), no
195 biochar was used. Two varieties of biochar, manufactured from maize and wheat straw,
196 were treated at five different dose levels: 1%, 3%, 5%, and 10% by mass (designated
197 M1, M3, M5, and M10 for maize straw biochar; W1, W3, W5, and W10 for wheat
198 straw biochar). There were 27 plots in all, each measuring 1.5 m by 1.5 m, and a
199 completely randomized design with three replications was employed (Figure 1). At
200 the end of the 180-day and 365-day experimental periods, five soil auger samples
201 were collected from the 0–20 cm layer of each plot and combined to form a composite
202 soil sample. The samples were then allowed to dry naturally and were ready for use



203 after being cleared of stones, plant roots, and other contaminants. Before determining
204 the soil properties, the unbound biochar particles in the soil samples were floated off
205 in distilled water.

206 **2.3 Determination of soil active organic carbon and electrochemical properties**

207 A carbon and nitrogen analyzer (Multi N/C 3000, Jena, Germany) was used to
208 measure DOC (Qiu et al., 2023). By using potassium permanganate for oxidation,
209 ROC was determined (Zhang et al., 2022). Sulfuric acid-potassium dichromate
210 oxidation with external heating and sodium iodide heavy liquid separation were used
211 to calculate LFOC (Janzen et al., 1992). Sulfuric acid-potassium dichromate oxidation
212 using an external heating technique and sodium hexametaphosphate dispersion were
213 used to determine POC (Ma et al., 2021a).

214 The combined method of surface property determination provided by Li et al.
215 (2011) was used to measure the soil surface charge number (SCN), surface charge
216 density (σ_0), and SSA. In short, soil samples were subjected to exchange equilibrium
217 testing using a combination of NaOH and Ca(OH)₂ after being pre-saturated with H⁺.
218 After measuring the amounts of Na⁺ and Ca²⁺ in the supernatant, the double-layer
219 theory was used to compute the cation exchange capacity, SSA, and σ_0 ; the detailed
220 procedures can be found in our previous work (Hu et al., 2021).

221 **2.4 Quantification of soil internal forces**

222 At the mesoscopic level, the primary SIF interactions among soil particles are
223 van der Waals attractive pressure (P_{vdw}), surface hydration repulsive pressure (P_h),
224 and electrostatic repulsive pressure (P_E). The net inter-particle force (P_{net}) is the



225 combined force of the three, which can be calculated from the soil surface property
226 parameter to obtain the SIFs (Li and Xu, 2008; Liu et al., 2021).

227 **2.5 Soil aggregate stability evaluation**

228 In this experiment, soils were prepared as Na⁺-saturated samples to
229 quantitatively characterize the intrinsic link between SIFs and aggregate stability. Soil
230 aggregate breaking strength (SABS) was used to evaluate the aggregate stability
231 (Xu et al., 2015). It was defined as the proportion of mass released as small particles
232 (less than 5 and less than 10 μm) when macroaggregates broke down. As the
233 percentage mass release increases, soil aggregate stability decreases. The following
234 was the preparation procedure: A 5 L beaker containing 2 kg of air-dried soil was
235 filled with 5 L of NaCl electrolyte solution at a concentration of 0.5 mol L⁻¹. The
236 sample was centrifuged and the supernatant was disposed of after 24 hours of stirring
237 and equilibration. Following three repetitions of the aforementioned procedure, the
238 extra Na⁺ was removed using distilled water and dried through a 1–5 mm sieve.
239 Different concentrations of NaCl solutions (10⁻⁵, 10⁻³, 10⁻², 10⁻¹, 1 mol L⁻¹) were
240 used in the experiment to simulate soil electrolyte solutions. The time required for <
241 10 μm and < 5 μm soil particles to fall at a specific distance was first calculated
242 according to Stokes formula, respectively. About 450 ml of a NaCl solution with
243 concentrations of 1, 10⁻¹, 10⁻², 10⁻³, and 10⁻⁵ mol L⁻¹ was put into a 500 ml
244 measuring cylinder. Weigh 15 g of Na⁺-saturated aggregates and transfer them into a
245 measuring cylinder. Add the same solution to bring the total volume to 500 mL, then
246 seal the cylinder. Allow it to stand for 1 minute. Afterward, hold the cylinder



247 horizontally in your hand and rotate it slowly, inverting it once and allowing it to
248 stand for approximately 30 seconds. Repeat this process four times. Finally, place the
249 cylinder on the tabletop, record the start time, and calculate the end time. A U-shaped
250 tube was used to aspirate the suspension at the conclusion of the allotted period, and it
251 was then moved into an aluminum box. The sample was then weighed after drying at
252 105 °C, following the method of Hu et al. (2015).

253 **2.6 Statistical analysis**

254 Microsoft Excel was used to arrange and compute the data, while SPSS Statistics
255 26 was used to conduct the statistical analysis. Statistical significance was determined
256 using Duncan's test ($P < 0.05$). The relationship between soil surface electrochemical
257 characteristics and soil active carbon fractions was assessed using the Pearson's
258 coefficient test. Soil carbon fractions and electrochemical properties were analyzed
259 redundantly using Canoco 5.0. Origin 2023 software was applied for data plotting.

260 **3. Results**

261 **3.1 Effect of biochar addition on soil active organic carbon fractions**

262 After applying maize straw biochar and wheat straw biochar after 180 days, the
263 overall amount of soil active organic carbon rose as the amount of biochar increased
264 (Figure 2). Both biochar treatments significantly increased soil LFOC concentrations
265 ($P < 0.05$), although slight, non-significant decreases were observed in the M5 and
266 W5 treatments compared to M3 and W3. The M10 and W10 treatments showed the
267 highest increase, 147.9 and 127.0 times compared to CK, respectively. The LFOC
268 under each treatment for the two kinds of straw biochar was only substantially



269 different at the 10% addition rate; at the 3% and 5% biochar application rates, the
270 difference was not significant (Figure 2a). Both biochar treatments significantly
271 enhanced soil POC content ($P < 0.05$). In terms of magnitude, M10 and W10
272 exhibited the largest increases, with POC contents 84.2 and 78.4 times higher than
273 that of the CK, respectively. With the exception of the 5% application rate (Figure 2b),
274 no appreciable variations in the soil POC concentration were noted in relation to the
275 impacts of the two straw biochar treatments at the same application rate. The M10 and
276 W10 treatments increased DOC by 18.6 and 18.1 times, respectively, compared to CK.
277 At the same application rate, there were no discernible variations in DOC between the
278 two biochars (Figure 2c). The M10 and W10 treatments increased ROC by 5.1 and 5.3
279 times, respectively, compared to CK. At the same application rate, there were no
280 discernible variations in ROC between the two biochars (Figure 2d). The contents of
281 different carbon fractions under each treatment decreased with the extension of
282 incubation time, and the data for soil active carbon fractions at 365 days are provided
283 in Supplementary Figure S1.

284 **3.2 Effect of biochar addition on soil surface electrochemical properties**

285 Table 3 shows that as the amount of biochar rose from 0% to 10%, SCN
286 increased considerably ($P < 0.05$). Compared with CK, SCN increased by 23.3%,
287 33.3%, 39.8%, and 42.7% under each treatment of maize straw biochar, and 27.1%,
288 33.3%, 39.4%, and 46.4% under the wheat straw biochar treatment, respectively. The
289 difference between the two biochars at the same addition level was not significant.
290 The soil SSA increased significantly ($P < 0.05$) with increasing biochar application



291 rates; the M1 treatment and CK did not differ significantly, and at 1%, 3%, and 5%
292 application rates, the differences between the treatments were not significant, and
293 M10 and W10 increased by 91.06% and 91.53%, respectively, in comparison to the
294 CK treatment. Surface charge density σ_0 showed no significant change at lower
295 application rates (1%, 3%, 5%) compared to CK. However, σ_0 tended to decline as
296 biochar application rate increased. After 365 days of incubation with biochar, the
297 trends of soil electrochemical parameters were consistent with those observed at 180
298 days, and the values of these parameters were slightly higher than those at 180 days.
299 The data for soil surface electrochemical properties at 365 days are provided in
300 Supplementary Table S1.

301 **3.3 Relationship between soil electrochemical properties and active carbon** 302 **fractions**

303 After 180 days of biochar application, the correlation between soil
304 electrochemical properties and active organic carbon fractions is presented in Figure
305 3a. SCN exhibited a strong positive association with LFOC and POC ($P < 0.05$) and a
306 highly significant positive correlation with DOC and ROC ($P < 0.01$). SSA and DOC
307 ($P < 0.01$), LFOC, POC, and ROC ($P < 0.001$) all showed a substantial positive
308 connection. In contrast, σ_0 was strongly negatively correlated with LFOC, POC, and
309 ROC ($P < 0.001$), and with DOC ($P < 0.01$). The results at 365 days (Fig. 3b) showed
310 that SCN and SSA were significantly and positively correlated with LFOC, POC,
311 DOC, and ROC ($P < 0.001$). In contrast, σ_0 was negatively correlated with LFOC,
312 POC, and ROC ($P < 0.05$), and showed a highly significant negative correlation with



313 DOC ($P < 0.001$).

314 Figure 4 displays the results of redundancy analysis. The results showed that
315 after 180 days of biochar addition, ROC explained 63.6% of the variation in soil
316 electrochemical properties, whereas after 365 days, POC accounted for 66.1% of the
317 variation (Table 4). Therefore, ROC and POC are the primary factors influencing the
318 electrochemical properties of the soil surface. The acute angle formed between the
319 SCN, SSA, and soil active carbon component arrows indicates a strong positive
320 correlation between them (the smaller the acute angle, the stronger the relationship),
321 consistent with the results in Figure 3.

322 **3.4 Effect of biochar addition on soil internal forces**

323 ***3.4.1 Electrostatic repulsive pressure between soil particles***

324 The electrostatic repulsive pressure at various electrolyte concentrations at a
325 distance of 2 nm between soil particles is depicted in Figure 5. The findings
326 demonstrated that as the amount of maize straw and wheat straw biochar applied rose
327 from 1% to 10%, the electrostatic repulsive pressure gradually reduced at the same
328 electrolyte concentration. At a constant biochar application rate, electrostatic repulsive
329 pressure increased sharply under all treatments as the electrolyte concentration
330 decreased progressively from 1 mol L^{-1} to $10^{-2} \text{ mol L}^{-1}$. However, when the
331 electrolyte concentration is further decreased below $10^{-2} \text{ mol L}^{-1}$, the rate of growth
332 of electrostatic repulsive pressure slows down and stabilizes. After 365 days of
333 incubation, the trends in electrostatic repulsive pressure for each treatment were
334 consistent with those at 180 days, and increased compared to the soil after 180 days of



335 incubation.

336 ***3.4.2 Van der Waals attractive pressure and surface hydration repulsive pressure***

337 Table 5 shows that all of the Hamaker constant values were higher than CK and
338 rose as the amount of biochar applied increased. The strength of the van der Waals
339 attractive pressure between soil particles is directly determined by the Hamaker
340 constant, a crucial element in the computation of this pressure. The amount and
341 distribution of the van der Waals attractive pressure and surface hydration repulsive
342 pressure between particles with biochar addition after 180 and 365 days (as seen in
343 Figure 6) may be determined using the formula, in conjunction with the acquired
344 Hamaker constant. The van der Waals attractive pressure (absolute value) and the
345 surface hydration repulsive pressure both sharply decline with increasing soil particle
346 distance. When the spacing between soil particles was smaller than 1.3 nm, the
347 surface hydration repulsive pressure was substantially larger in all treatments than the
348 van der Waals attractive pressure. For instance, under various treatments, the
349 interparticle van der Waals attractive pressure (in absolute value) is less than 100 atm
350 and the interparticle surface hydration repulsive pressure is around 591 atm when the
351 interparticle distance is 0.7 nm. The van der Waals attractive pressure (absolute values)
352 is all below 200 atm, but the surface hydration repulsive pressure is around 1051 atm
353 at an interparticle distance of 0.6 nm. This suggests that the difference between the
354 two rises quickly as the distance decreases. Van der Waals attractive pressure
355 (absolute value) rose with increasing application rate in both biochar treatments. After
356 365 days of incubation, the trends in the Hamaker constant and van der Waals



357 attraction (in absolute value) for each treatment were consistent with those at 180
358 days, and increased compared to the soil after 180 days of incubation.

359 **3.4.3 Net pressure between soil particles**

360 Figure 7 shows the net pressure of soil particles at 2 nm for different electrolyte
361 concentrations under biochar addition after 180 and 365 days. Under both biochar
362 types, net pressure increased as electrolyte concentration decreased. From 1 mol L^{-1} to
363 $10^{-2} \text{ mol L}^{-1}$, net pressure rose markedly; below $10^{-2} \text{ mol L}^{-1}$, the rate of increase
364 plateaued. At the same electrolyte concentration, net pressure decreased as biochar
365 application increased. After 365 days of incubation, the trends in the net interparticle
366 force were consistent with those at 180 days, and decreased compared to the soil after
367 180 days of incubation.

368 **3.5 Effect of biochar addition on soil aggregate stability**

369 Figure 8 shows SABS ($< 5 \mu\text{m}$, $< 10 \mu\text{m}$) at different electrolyte concentrations
370 under biochar addition after 180 days. As the electrolyte concentration increased, the
371 SABS decreased. The change in SABS was relatively smooth when the electrolyte
372 concentration was less than $10^{-2} \text{ mol L}^{-1}$, but it significantly decreased for all
373 treatments when the electrolyte concentration went from 10^{-2} to 1 mol L^{-1} . A
374 concentration of $10^{-2} \text{ mol L}^{-1}$ represents the critical threshold for aggregate
375 disintegration. SABS dropped as biochar application increased under the same
376 electrolyte concentration, suggesting that biochar application improved aggregate
377 stability. After 365 days of incubation, the trends in aggregate breaking strength for
378 each treatment were consistent with those at 180 days, and the values decreased



379 compared to the soil after 180 days of incubation. The data for soil aggregate breaking
380 strength at 365 days are provided in Supplementary Figure S2.

381 **3.6 Relationship between soil internal forces and aggregate stability**

382 Figure 9 depicts the relationship between the net pressure at a 2 nm distance
383 between soil particles and SABS under biochar addition after 180 and 365 days. The
384 figure reveals a significant positive correlation between SABS and net pressure across
385 all treatments, with SABS exhibiting an exponential increase in response to rising net
386 pressure.

387 **4. Discussion**

388 **4.1 Response of soil active carbon fraction and electrochemical properties to** 389 **biochar application**

390 The application of biochar enhanced soil active carbon fractions and had a
391 considerable impact on soil organic carbon (Han et al., 2020; Li et al., 2024; Lu et al.,
392 2021; Yang et al., 2017; Zhao et al., 2018). According to our findings, the soil active
393 carbon percentage rose when both types of biochar were added (Figure 2 and Figure
394 S1). The reason can be explained: the biochar itself contains more carbon, especially
395 more resistant humic acid and condensed aromatic carbon structures. Secondly,
396 biochar possesses a large surface area, which may facilitate the accumulation of
397 specific organic carbon fractions in the soil by adsorbing them onto its surface.
398 Biochar has a highly porous structure that also protects some organic carbon
399 components from microbial degradation (Feng et al., 2021; He et al., 2023). In a field
400 experiment, Qiu et al. (2023) applied biochar to 0%, 1%, 2%, and 4% of the dry



401 weight of topsoil. After 8 and 12 months, they observed a positive correlation between
402 the amount of particulate organic carbon and the amount of biochar applied, as well as
403 significant increases in organic carbon, particulate organic carbon, and DOC content.
404 In comparison to the control, soil organic carbon, ROC, and POC levels rose by 7.09–
405 38.08%, 1.62–39.80%, and 9.52–62.30%, respectively, when Jiang et al. (2022)
406 applied 0, 5, 15, 20, and 40 t ha⁻¹ of biochar to the soil. As a result, soil active carbon
407 fractions were significantly impacted by biochar additions.

408 With the extension of incubation time from 180 days to 365 days, the content of
409 soil active carbon fractions decreased, which may be attributed to several factors
410 (Stewart et al., 2009; Yang et al., 2017; Zhao et al., 2018). Firstly, as the incubation
411 period increased, organic carbon in the soil was gradually degraded and mineralized
412 by microorganisms, leading to a decrease in the content of ROC and DOC. Secondly,
413 the long-term application of biochar may have transformed some organic carbon into
414 more stable forms, which are less susceptible to microbial decomposition, resulting in
415 a decline in the active organic carbon fractions. Additionally, changes in the soil
416 environment, such as fluctuations in moisture, temperature, and pH, may have
417 affected the availability and turnover rate of organic carbon, further contributing to
418 the reduction in active organic carbon content.

419 Both biochar applications considerably raised the SSA and SCN of soil particles
420 in this investigation (Table 2 and Table S1). According to Liang et al. (2006), this
421 phenomenon is caused by the numerous negative charges that are carried on the
422 surface of biochar. These charges come from the oxygen-containing functional groups,



423 like -COOH, -OH, etc., that are widely distributed on the surface of biochar and
424 which effectively improve the cation exchange of the soil. The high SSA of biochar
425 itself directly contributes to an increase in soil SSA, improves cation adsorption, and
426 thus enhances exchange capacity (Lone et al., 2015). Consistent with Hu et al. (2021),
427 our findings show that increasing the biochar application rate enhances soil SSA and
428 SCN (Table 2 and Table S1). However, the increase in soil organic carbon
429 (particularly the active carbon component) brought about by the addition of biochar
430 may have contributed to the rise in SSA and SCN. The study's findings demonstrated
431 that adding two different kinds of biochar raised the amount of active carbon in the
432 soil and that there was a significant relationship between soil active carbon and soil
433 electrochemical characteristics, with ROC and POC being the primary determinant of
434 the soil surface's electrochemical characteristics (Table 4; Figures 3–4). According to
435 Meyer et al. (1994) and Pennell et al. (1995), soil organic matter, particularly humic
436 acid, has a cation exchange capacity of up to 100–700 cmol(c) kg⁻¹ and an SSA of up
437 to 500–800 m² g⁻¹. Vertisol soil SSA (about 35.3%) and SCN (approximately 29.3%)
438 were shown to decrease with the removal of soil organic matter (Yu et al., 2020).
439 Compared with more stable organic carbon pools, ROC represents the relatively
440 amorphous, chemically reactive fraction that is enriched in oxygen-containing
441 functional groups (e.g. -COOH, -OH, -C=O) and is preferentially adsorbed onto or
442 co-precipitated with mineral surfaces. Such functional groups provide abundant
443 negatively charged sites and enhance the development of the diffuse double layer,
444 thereby increasing soil SSA, SCN and surface charge density and modifying the



445 balance between electrostatic repulsion and van der Waals attraction at the particle–
446 water interface (He et al., 2021). In contrast, POC is typically found in smaller soil
447 particles and influences the electrochemical properties of the soil surface through
448 physical adsorption. The role of POC primarily involves its interaction and stability
449 with minerals, which increases the charge and physical surface characteristics of the
450 soil, further regulating the soil's electrochemical behavior. ROC and POC jointly
451 influence the electrochemical properties of the soil, with ROC being more sensitive to
452 changes in soil electrochemical properties in the short term, while POC affects the
453 soil's electrochemical reactions over a longer period through stable mineral-organic
454 carbon complexes. These organic carbon fractions have been repeatedly identified as
455 a sensitive indicator of short-term management effects and is closely linked to key
456 fertility parameters, including effective CEC and other electrochemical properties
457 (Culman et al., 2012). Therefore, soil active organic carbon (especially ROC and POC)
458 is closely related to the electrochemical properties of the soil surface.

459 **4.2 Response of soil internal forces to biochar application**

460 Soil particles in close proximity experience mutual repulsion due to like charges,
461 generating electrostatic repulsive pressure. Because soil texture, organic matter
462 content, and mineral composition all affect the particles' surface charge, variations in
463 these characteristics will result in variations in electrostatic repulsive pressure (Hu et
464 al., 2015; Liu et al., 2021). At the same electrolyte concentration, soil electrostatic
465 repulsive pressure decreased under both biochar treatments as biochar application
466 increased from 1% to 10% (Figure 5). This phenomenon is caused by the substantial



467 alterations in the electrochemical characteristics of the soil surface, which are shown
468 by the increase in SCN and SSA, the decrease in the intensity of the surface electric
469 field and charge density, and the effective control of electrostatic repulsive pressure.
470 Both forms of biochar reduced electrostatic repulsive pressure similarly, which was
471 essentially in line with the trend of soil surface electrochemical parameters under
472 varying rates of biochar application, which is in line with Wang et al. (2024) findings.

473 Biochar application also significantly increased van der Waals attractive pressure
474 between soil particles, particularly at higher application rates (Figure 6). Van der
475 Waals pressure is directly related to the Hamaker constant, which itself depends on
476 the properties of the materials and the intervening medium (Bergström, 1997; Hu et
477 al., 2021; Yu et al., 2020). The observed increases in Hamaker constants (Table 4)
478 suggest that biochar incorporation introduced more organic complexes into the soil
479 (Resurreccion et al., 2011). Additionally, our results revealed that net interparticle
480 pressure decreased as biochar application increased (Figure 7), consistent with the
481 findings of Yu et al. (2017). These results suggest that the influence of biochar on net
482 pressure is primarily through increased van der Waals attraction and reduced
483 electrostatic repulsion.

484 **4.3 Response of soil aggregate stability to biochar application**

485 In this study, SABS values decreased with increasing biochar application (Figure
486 8 and Figure S2), indicating improved aggregate stability. This aligns with previous
487 findings that biochar enhances soil aggregation (Fu et al., 2021; He et al., 2020; Luo
488 et al., 2020; Sun et al., 2021; Wang et al., 2022). The observed decline in net pressure



489 with increasing biochar (Figure 7) supports this conclusion, as lower internal forces
490 correspond to greater structural stability. This is consistent with prior reports by Yu et
491 al. (2020) and Hu et al. (2021), who also found that biochar reduces internal particle
492 forces and enhances aggregate stability.

493 We also observed that SABS increased with decreasing electrolyte concentration
494 and then plateaued, with a threshold near 10^{-2} mol L⁻¹, indicating a critical transition
495 in aggregate disintegration. This turning point corresponded to a similar trend in net
496 interparticle pressure, reinforcing the connection between soil internal forces and
497 aggregate stability. Regression analysis confirmed a significant positive correlation
498 between SABS and net pressure (Figure 9), highlighting the central role of
499 interparticle forces in determining aggregate stability.

500 The effectiveness of biochar in improving soil aggregation depends on
501 environmental conditions, soil type, and feedstock characteristics (Du et al., 2016;
502 Fungo et al., 2017; Jien and Wang, 2013; Pituello et al., 2018; Yang and Lu, 2021;
503 Zhang et al., 2015). Overall, there were no significant differences in SIF distribution
504 and aggregate stability between the two biochar treatments in this study. This may be
505 attributed to the fact that the basic properties of the two types of straw biochar are
506 relatively similar (Table 2). Additionally, there were no significant changes in the
507 active carbon components and surface electrochemical properties of the soil after the
508 two types of biochar were added. In addition, our study found that the improvement in
509 soil aggregate stability was greater at 365 days compared to 180 days following the
510 application of both types of biochar. This is because, compared to 180 days, biochar



511 addition at 365 days increased both soil charge density and the Hamaker constant,
512 which in turn enhanced both the electrostatic repulsive force and molecular attraction
513 at the mesoscale between soil particles. However, the increase in molecular attraction
514 was much greater than that of electrostatic repulsion, resulting in a lower net force.
515 Therefore, soil aggregate stability was stronger after 365 days of biochar addition.

516 Although this study focused on short-term (365-day) effects, biochar is known
517 for its long-term stability in soil environments. Previous research has shown that
518 biochar can enhance soil aggregation, nutrient retention, and carbon sequestration for
519 years to decades (Jiang et al., 2022; Sun et al., 2021). However, these long-term
520 benefits may be affected by aging processes, environmental variability, and
521 management practices. Future long-term field experiments are needed to better
522 understand the persistence and dynamics of biochar effects in diverse agroecosystems.

523 **5. Conclusions**

524 After 180 and 365 days of maize straw biochar and wheat straw biochar addition,
525 the contents of soil active carbon fractions increased with higher biochar application
526 rates. Soil active carbon fractions were closely related to soil electrochemical
527 properties, with ROC and POC being the primary influencing factors. The application
528 of biochar markedly modified the SSA and SCN of soil particles, thereby increasing
529 van der Waals attractive forces, reducing net repulsive pressure, and ultimately
530 enhancing aggregate stability. These beneficial effects were further amplified as the
531 application rate increased. As the electrolyte concentration decreased, the SABS first
532 increased and then tended to level off, and $10^{-2} \text{ mol L}^{-1}$ was the turning point for the



533 change. Although this study utilized Na⁺-saturated soil samples, which do not fully
534 reflect natural soil conditions, the findings quantitatively assessed the intensity and
535 distribution of soil internal forces (SIFs) and elucidated the mechanistic pathway by
536 which biochar-induced increases in ROC and POC, along with associated changes in
537 soil surface electrochemistry, enhance aggregate stability. Collectively, these findings
538 offer a theoretical basis for the targeted use of biochar to manipulate soil internal
539 forces, strengthen aggregate structure, and mitigate soil erosion in agricultural
540 systems.

541

542 **Code and data availability**

543 No custom code was used in this study. The experimental data that support the
544 findings of this study are available from the corresponding author upon reasonable
545 request.

546

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553

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Table 1 Basic physicochemical properties of the soil

	Bulk	Organic	Total	Available	Available
pH	density	carbon	nitrogen	phosphorus	potassium
	(g cm ³)	(g kg ⁻¹)	(g kg ⁻¹)	(mg kg ⁻¹)	(mg kg ⁻¹)
7.53	1.33	9.02	0.82	20.79	165.52

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Table 2 Basic properties of biochar

Biochar	pH	C (%)	H (%)	O (%)	N (%)	H/C	O/C	SSA (m ² g ⁻¹)	Ash (%)
Maize	6.30	32.67	0.92	11.72	0.48	0.03	0.36	14.48	7.51
Wheat	6.08	76.10	2.18	24.21	0.67	0.03	0.32	15.17	3.52

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869 **Table 3** Soil electrochemical properties under biochar treatment for 180 days

Treatment	SCN (cmol(c) kg ⁻¹)	SSA (m ² g ⁻¹)	σ ₀ (c m ⁻²)
CK	13.12±0.36e	32.23±2.42c	0.39±0.04a
M1	16.18±0.72d	40.63±7.67bc	0.39±0.08a
M3	17.50±0.14c	45.69±3.56b	0.37±0.03abc
M5	18.35±0.08b	49.94±0.57b	0.35±0.01abc
M10	18.72±0.03ab	61.58±7.24a	0.30±0.03c
W1	16.67±0.36d	42.26±7.04b	0.39±0.07ab
W3	17.50±0.09c	45.70±3.39b	0.37±0.03abc
W5	18.29±0.40b	47.85±5.43b	0.37±0.05abc
W10	19.21±0.16a	61.73±7.40a	0.30±0.04bc

870 Note: M1, M3, M5, M10, add 1%, 3%, 5%, 10% maize straw biochar; W1, W3, W5, W10, add

871 1%, 3%, 5%, 10% wheat straw biochar; CEC: cation exchange capacity, SSA: specific surface

872 area, σ₀: surface charge density; Values are means ± stand deviation (n = 3).

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876 **Table 4** Redundancy analysis of soil surface electrochemical properties and active
877 carbon fractions after 180 and 365 days.

	Soil active carbon fractions	Explanation rate (%)	<i>F</i>	<i>P</i>
180 d	ROC	63.6	43.7	0.002
	DOC	1.7	1.2	0.322
	LFOC	1.5	1	0.37
	POC	1.7	1.2	0.348
365 d	POC	66.1	48.7	0.002
	DOC	0.5	0.4	0.614
	LFOC	0.4	0.3	0.762
	ROC	0.1	0.1	0.878

878 Note: ROC: readily oxidized organic carbon; DOC: dissolved organic carbon; LFOC: Light

879 fraction organic carbon; POC: particulate organic carbon.

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881 **Table 5** Changes in soil Hamaker constant under different treatments after 180 and
882 365 days.

Treatment	Hamaker (J) 180 d	R^2	Hamaker (J) 365 d	R^2
CK	$4.78 \times 10^{-20} \pm 5.91 \times 10^{-21}$	0.82	$4.79 \times 10^{-20} \pm 4.63 \times 10^{-21}$	0.90
M1	$6.01 \times 10^{-20} \pm 6.05 \times 10^{-21}$	0.82	$8.89 \times 10^{-20} \pm 7.27 \times 10^{-21}$	0.85
M3	$6.08 \times 10^{-20} \pm 7.99 \times 10^{-21}$	0.73	$9.47 \times 10^{-20} \pm 8.29 \times 10^{-21}$	0.83
M5	$6.06 \times 10^{-20} \pm 7.71 \times 10^{-21}$	0.78	$9.78 \times 10^{-20} \pm 6.28 \times 10^{-21}$	0.91
M10	$6.15 \times 10^{-20} \pm 6.90 \times 10^{-21}$	0.78	$1.06 \times 10^{-19} \pm 3.44 \times 10^{-21}$	0.98
W1	$4.96 \times 10^{-20} \pm 5.95 \times 10^{-21}$	0.81	$7.38 \times 10^{-20} \pm 7.57 \times 10^{-21}$	0.75
W3	$5.02 \times 10^{-20} \pm 6.67 \times 10^{-21}$	0.75	$8.39 \times 10^{-20} \pm 7.80 \times 10^{-21}$	0.79
W5	$5.19 \times 10^{-20} \pm 6.08 \times 10^{-21}$	0.82	$8.73 \times 10^{-20} \pm 3.78 \times 10^{-21}$	0.95
W10	$5.22 \times 10^{-20} \pm 5.55 \times 10^{-21}$	0.84	$8.85 \times 10^{-20} \pm 2.69 \times 10^{-21}$	0.98

883 Note: M1, M3, M5, M10, add 1%, 3%, 5%, 10% maize straw biochar; W1, W3, W5, W10, add
884 1%, 3%, 5%, 10% wheat straw biochar; Values are means \pm stand deviation (n = 3).

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Figure Captions

888 **Figure 1** Experimental plot design. Note: W is wheat straw biochar and M is maize
889 straw biochar; 0, 1, 3, 5, and 10 represent 0%, 1%, 3%, 5%, and 10% biochar
890 additions, respectively; A, B, and C represent three replicates, respectively.

891 **Figure 2** Effect of biochar addition on soil active organic carbon fractions after 180
892 days. M1, M3, M5, M10, add 1%, 3%, 5%, 10% maize straw biochar; W1, W3,
893 W5, W10, add 1%, 3%, 5%, 10% wheat straw biochar.

894 **Figure 3** Pearson correlation analysis between soil active carbon fractions and soil
895 electrochemical properties after 180 and 365 days. * indicates significant
896 correlation at the 0.05 level; ** indicates significant correlation at the 0.01
897 level; and *** indicates significant correlation at the 0.001 level.

898 **Figure 4** Redundancy analysis of soil active carbon fractions and soil electrochemical
899 properties after 180 and 365 days.

900 **Figure 5** Electrostatic repulsive pressure between soil particles at an interparticle
901 distance of 2 nm under different electrolyte concentrations after 180 and 365
902 days. M1, M3, M5, M10, add 1%, 3%, 5%, 10% maize straw biochar; W1, W3,
903 W5, W10, add 1%, 3%, 5%, 10% wheat straw biochar.

904 **Figure 6** Distributions of van der Waals attractive pressure and surface hydration
905 repulsive pressure under biochar addition after 180 and 365 days. M1, M3, M5,
906 M10, add 1%, 3%, 5%, 10% maize straw biochar; W1, W3, W5, W10, add 1%,
907 3%, 5%, 10% wheat straw biochar.

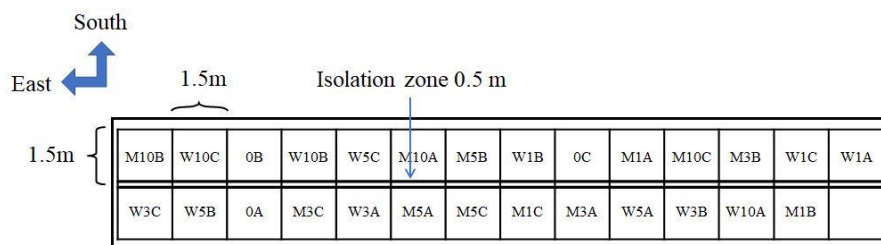
908 **Figure 7** Net pressure of soil particles at 2 nm for different electrolyte concentrations
909 under biochar addition after 180 and 365 days. M1, M3, M5, M10, add 1%,
910 3%, 5%, 10% maize straw biochar; W1, W3, W5, W10, add 1%, 3%, 5%, 10%
911 wheat straw biochar.

912 **Figure 8** Soil aggregate breaking strength ($< 5 \mu\text{m}$, $< 10 \mu\text{m}$) at different electrolyte
913 concentrations under biochar addition after 180 days. M1, M3, M5, M10, add
914 1%, 3%, 5%, 10% maize straw biochar; W1, W3, W5, W10, add 1%, 3%, 5%,
915 10% wheat straw biochar.

916 **Figure 9** Relationship between the net pressure at a 2 nm distance between soil
917 particles and aggregate breaking strength under biochar addition after 180 and
918 365 days. M1, M3, M5, M10, add 1%, 3%, 5%, 10% maize straw biochar; W1,
919 W3, W5, W10, add 1%, 3%, 5%, 10% wheat straw biochar.

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Figure 1 Experimental plot design.

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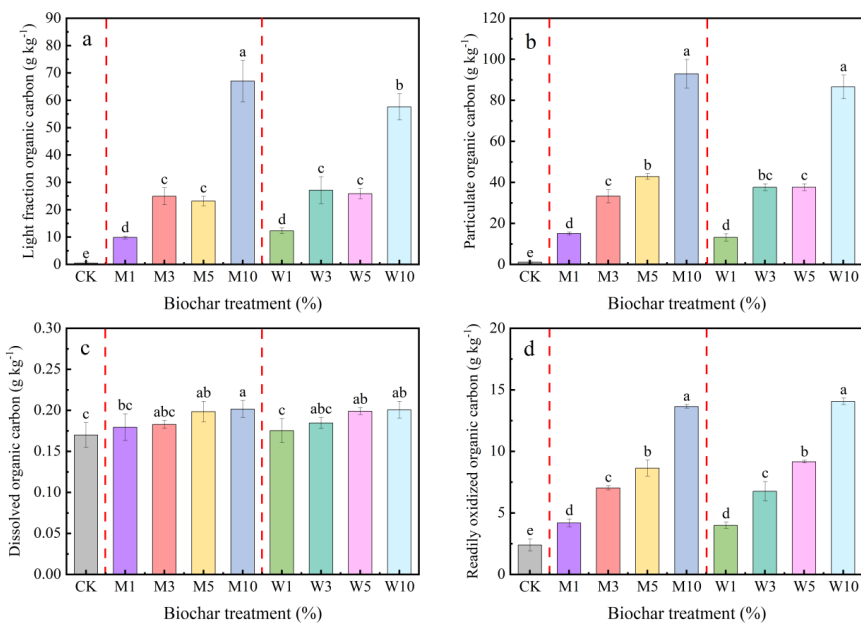
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Fig. 2. Effect of biochar addition on soil active organic carbon fractions after 180

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days.

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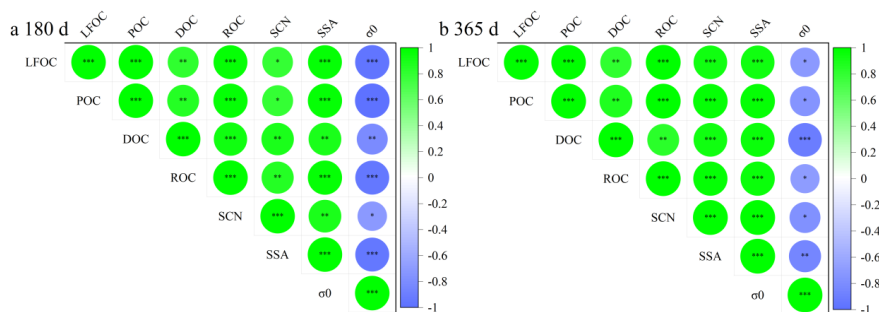
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Fig. 3. Pearson correlation analysis between soil active carbon fractions and soil

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electrochemical properties after 180 and 365 days.

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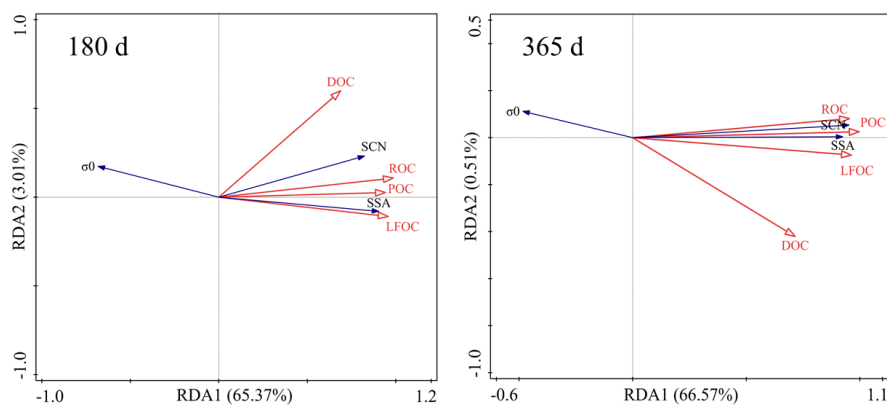
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968 **Fig. 4.** Redundancy analysis of soil active carbon fractions and soil electrochemical

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properties after 180 and 365 days.

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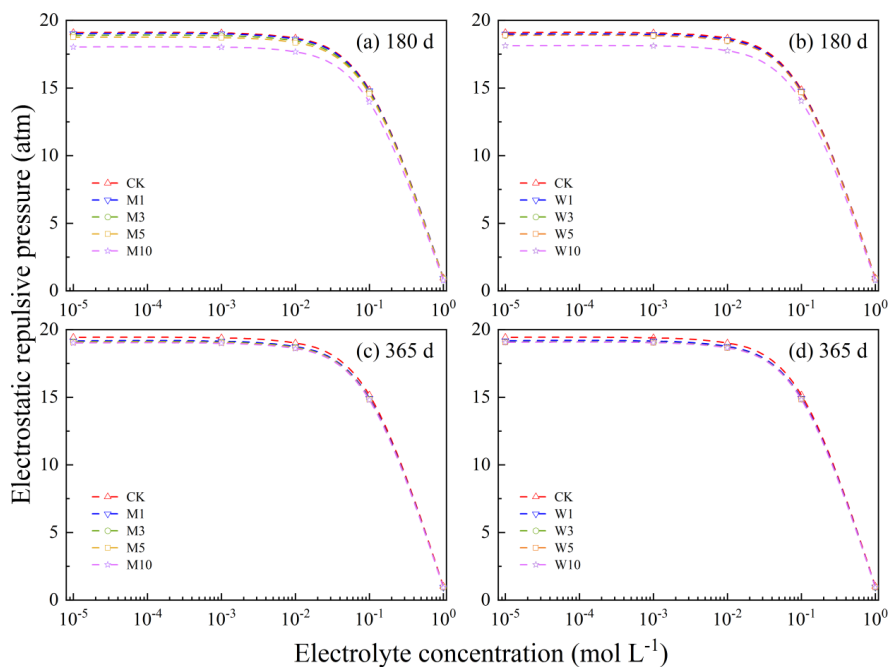
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Fig. 5. Electrostatic repulsive pressure between soil particles at an interparticle

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distance of 2 nm under different electrolyte concentrations after 180 and 365 days.

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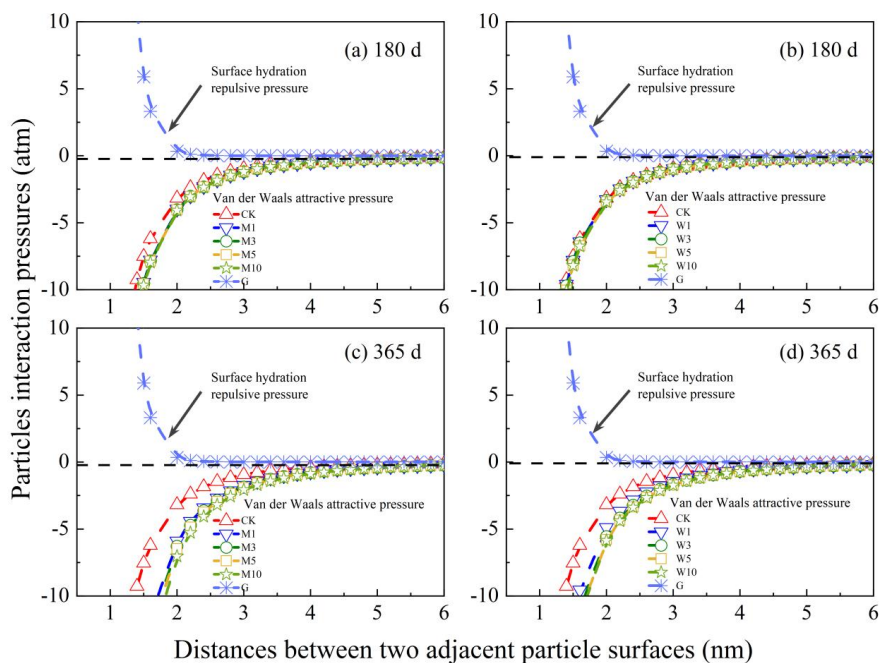
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Fig. 6. Distributions of van der Waals attractive pressure and surface hydration

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repulsive pressure under biochar addition after 180 and 365 days.

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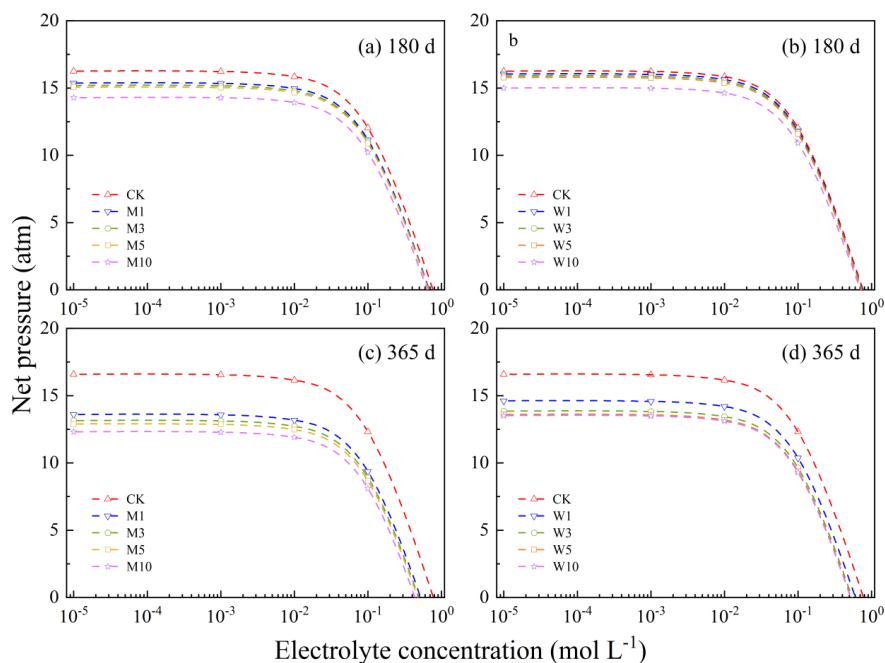
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Fig. 7. Net pressure of soil particles at 2 nm for different electrolyte concentrations

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under biochar addition after 180 and 365 days.

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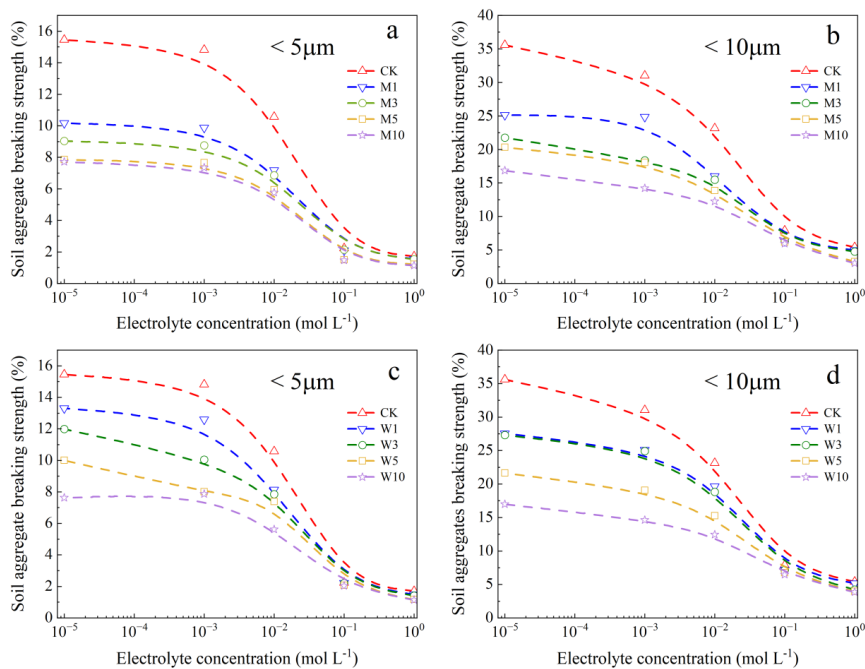
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Fig. 8. Soil aggregate breaking strength (< 5 μm, < 10 μm) at different electrolyte

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concentrations under biochar addition after 180 days.

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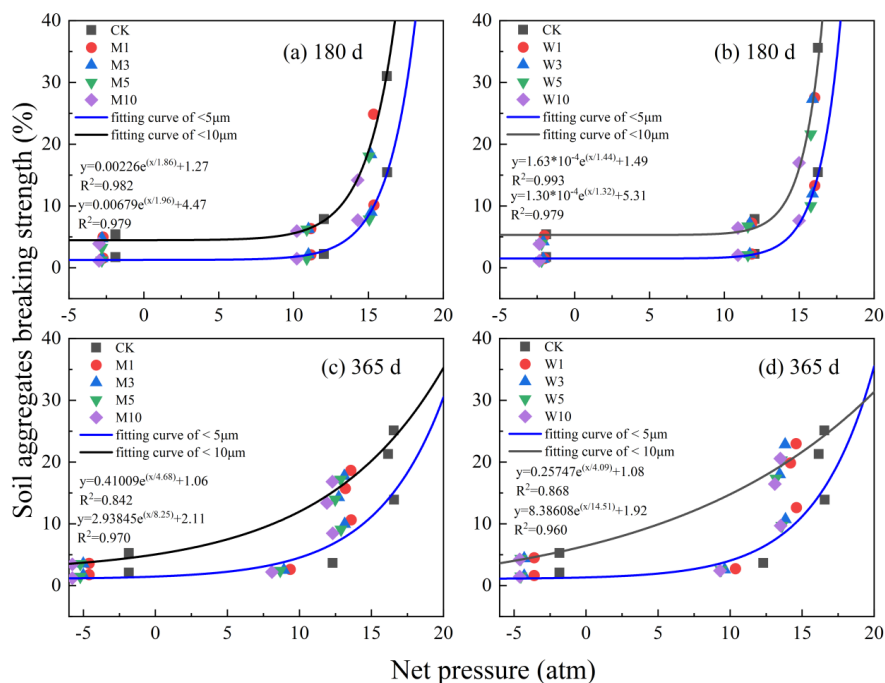
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1030 **Fig. 9.** Relationship between the net pressure at a 2 nm distance between soil particles

1031 and aggregate breaking strength under biochar addition after 180 and 365 days.