



# Modelling primary production: multitude of theories, or multitude of languages?

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**Abstract.** Marine primary production, converting approximately 50GtC per year, is an important component of the global carbon cycle, and a major determinant of past, present and future climate. Large-scale, long-term estimates of marine primary production rely primarily on two types of models: satellite-based models that make extensive use of remote-sensing data, and ecosystem models providing numerical simulation of ecological processes embedded in general ocean circulation models. Intercomparison exercises of model outputs (both within and across the two model types) have consistently revealed high discrepancies between estimated global ocean primary production, including divergent magnitudes and even opposite trends. Comparisons of model results with in-situ observations have also revealed large uncertainties in marine primary production estimates. These uncertainties limit the applications of these models, especially in the climate context, where an important question is whether climate change will drive significant future changes in regional or global primary production. Both satellite-based and ecosystem model equations rely on a range of fixed parameters, whose values need to be carefully estimated and tested. In this paper, we suggest that such model parameters represent an underappreciated but important source of inter-model differences. With the proliferation of both satellite and in situ observations of relevant variables at global scales and the availability of powerful statistical tools in data assimilation and machine



learning, we argue that time is right to systematically examine model parameters and gain insights into how they may vary spatially and temporally. We emphasize that such spatio-temporal variability can be easily theoretically justified for the models with complexity similar to the satellite models, or the ecosystem models commonly used within Earth System Models (ESMs) in climate studies. We argue that the spatially and temporally varying parameter values provide a strong reason to anticipate unification of models, which would otherwise appear structurally different. A better understanding of model parameter roles could therefore reduce discrepancies among the primary production models and improve the reliability of marine primary production projections.

## 1. Introduction

The climate problem is highly complex, the stakes are very high, and substantial knowledge gaps remain, especially in the ocean biogeochemistry domain (Kwiatkowski *et al.*, 2020). More broadly, the need to address complex issues related to the carbon cycle, ecosystem services and biogeochemistry through Earth System Models (ESMs), e.g. in the context of climate adaptation and resilience, has been highlighted by expert groups in Hewitt *et al.* (2021). Similarly, Jones *et al.* (2024) focus on modelling priorities to support international climate policy. In their assessment, they emphasise the value of “a coordinated, internally consistent set of simulations, data, and knowledge to support Intergovernmental Panel on Climate Change (IPCC) assessments” and outline multiple applications of Coupled Model Intercomparison Project (CMIP) projections. These include investigations of threats to marine ecosystems (which have consequences for the ocean’s ability to buffer climate change, Tjiputra *et al.*, 2025) and downstream services under various climate scenarios and associated risks of tipping points. Jones *et al.* (2024) also state that improving confidence in future projections requires models to reproduce the observed historical period. Furthermore, they identify parameter uncertainty as one of the key elements of uncertainty in climate models. The European Commission (2024) emphasised the need for improved projections of future possible impacts to understand better the relation between physiological processes and environmental variables.

Against this background and in line with the recommendations of expert bodies, this review focuses on the climate priority challenge associated with marine ecosystem and biogeochemistry modelling, with a particular focus on marine primary production. Phytoplankton primary production (PP), the process by which marine autotrophs convert CO<sub>2</sub> into organic matter through photosynthesis, is a major component of the ocean and planetary carbon cycle. Currently estimated at around 50 Pg C y<sup>-1</sup> (Kulk *et al.* 2020, 2021), the magnitude of marine PP is five times the estimated fossil fuel emissions of 10 Pg C y<sup>-1</sup> in 2022 and nearly 20 times the net ocean carbon sink (Friedlingstein *et al.* 2024). Its magnitude is comparable to that of terrestrial primary production (Lurin *et al.* 1994; Longhurst *et al.* 1995; Field *et al.* 1998; Friedlingstein *et al.* 2024). A key question in climate research is whether the current levels of marine PP can be maintained under climate change (Tagliabue *et al.*, 2021), at a time when marine ecosystems are increasingly threatened by a variety of sources, including ocean acidification (Jin *et al.*, 2020, Dai *et al.*, 2025), rising seawater temperatures (Kwiatkowski *et al.*, 2020), intensified storminess over the oceans (Gastineau and Soden, 2009, Young *et al.*, 2019, Gentile *et al.*, 2023, Liu *et al.*, 2024), ocean deoxygenation (Schmidtko *et al.*, 2017), modified current and stratification influencing surface nutrients (Maishal, 2024), as well as changes in aerial nutrient supply (Bergas-Masso *et al.*, 2025), biodiversity loss (Luypaert *et al.*, 2020), and sea-ice loss (Myksovoll *et al.*, 2023). In this review we consider PP estimated from two types of models: “satellite-based models” that utilize remote-sensing data together with statistical algorithms, whose parameters are informed by in situ measurements, to calculate PP, and mechanistic ecosystem



88 models which use numerical methods to solve a set of differential equations representing ecological processes,  
 89 with one output being PP.

90 When discussing marine primary PP, it is important to keep track of its different components. Theoretically,  
 91 PP before any of the loss terms are considered is referred to as gross PP (GPP); once the respiration by marine  
 92 autotrophs is subtracted from GPP we obtain net PP (NPP). GPP can also be partitioned according to whether  
 93 only the organic carbon fixed into particulate material is considered (production of particulate organic carbon), or  
 94 if the exudates (dissolved organic matter) are also included in the estimate (production of total organic carbon,  
 95 Regaudie-de-Gioux *et al.* 2014). While ecosystem models can make explicit distinction between these  
 96 components, it is not always straightforward to measure each of the components, often due to lack of experimental  
 97 methodologies that would differentiate clearly between them (IOCCG, 2022). In this review, we focus mainly on  
 98 GPP as computed in ecosystem models. For satellite-based estimates, we treat primary production derived from  
 99 in-situ experiments measuring short (1-4 hours) incubations as GPP, and those derived from experiments  
 100 measuring longer (12-24 hour) incubations as NPP, while fully recognising that the distinction is not that clear  
 101 cut (e.g., Halsey *et al.* 2011). It should be noted that estimates of the magnitude of losses due to respiration vary  
 102 considerably. Some estimates place it at about 30% of gross primary production (e.g., Platt *et al.* 1991), while  
 103 some other estimates are higher (e.g., 60% according to Halsey *et al.* 2011). Platt and Sathyendranath (1988)  
 104 compared daily water-column primary production computed on the basis of short incubations with those measured  
 105 in situ over daily time scales, and showed the two sets of independent estimates to be comparable, which points  
 106 to low respiration losses. Also, satellite-based estimates of NPP (Behrenfeld and Falkowski 1997) tend to be  
 107 roughly the same or higher than GPP (Longhurst *et al.* 1995). Since, by definition, NPP cannot be greater than  
 108 GPP, these comparisons reveal a great deal of uncertainty in respiration, or in PP computed using different  
 109 approaches, when compared with each other.

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## 112 2. Background

113 Considerable differences exist in model-based estimates (here and elsewhere, we use “models” without a  
 114 qualifier, to mean both satellite-based and ecosystem models) of the current and past global PP in the ocean, and  
 115 in ecosystem-model based projections into the future.

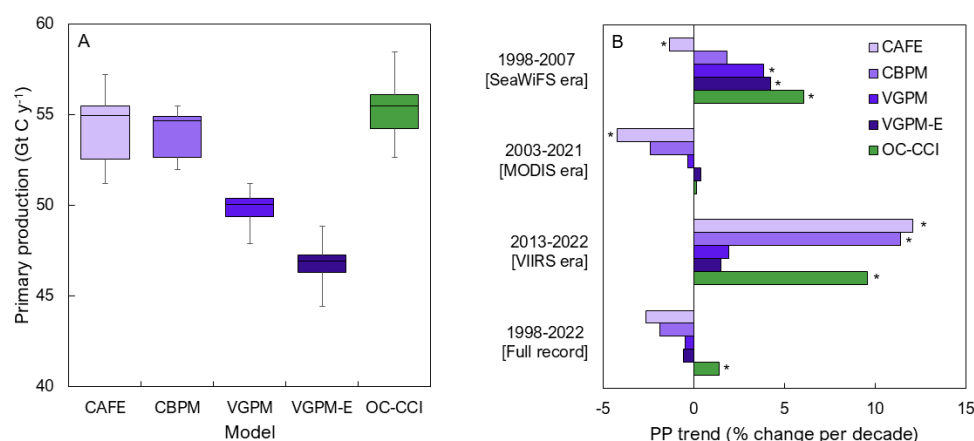
116 Satellite-based estimates of global marine PP are converging around 45-55 Pg C y<sup>-1</sup> (Figure 1A). These  
 117 estimates were obtained from both multi-sensor products of the Ocean Colour Climate Change Initiative (OC-  
 118 CCI; version 6, Sathyendranath *et al.* 2019, Kulk *et al.* 2020, 2021), as well as from single-sensor products of the  
 119 Oregon State University (<http://orca.science.oregonstate.edu/>), which include the Carbon, Absorption, and  
 120 Fluorescence Euphotic-resolving (CAFE) model (Silsbe *et al.* 2016; 2025), Carbon-Based Primary Productivity  
 121 Model (CBPM; Westberry *et al.* 2008), the Vertically Generalised Production Model (VGPM; Behrenfeld and  
 122 Falkowski 1997) and the VGPM-Eppley model (which incorporates the Eppley (1972) temperature function).  
 123 However, we note that much higher values (up to 67 Pg C y<sup>-1</sup>) and lower values (≤45 Pg C y<sup>-1</sup>) have also been  
 124 reported from satellite-based products (Antoine *et al.* 1996; Behrenfeld *et al.* 2005; Carr *et al.* 2006; Uitz *et al.*  
 125 2010) (here we recognize that satellite products may differ in the computed PP components, as noted earlier).

126 Large differences also emerge in the PP trends over the last decades estimated from both the CCI and Oregon  
 127 State University products (Figure 1B), as well as associated reanalyses (e.g. those of Gregg and Rousseaux, 2019).



These differences are strongly impacted by the choice of historical period and the underlying characteristics of the satellite products (e.g. whether they have data-gaps, or not), but the choice of satellite-based PP model does matter. In a recent comparison (Ryan-Keogh 2025) of six satellite-based primary production models applied to a common satellite product (OC-CCI) and a common period (1998-2023), showed that four of them showed declining trends, while the other two showed an increasing trend. Interestingly, the split is along the lines of whether the models incorporated temperature-dependent production parameters, or not. Ryan-Keogh *et al.* (2025) also compared satellite products and outputs of several ecosystem models from the Climate Model Intercomparison Project (CMIP-6), and concluded that, in general, the climate models underestimated the decreasing trends seen in many of the satellite-based models.

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**Figure 1.** Global marine PP computed using the satellite-based model of Platt & Sathyendranath (1988) as updated by Sathyendranath *et al.* (2020) and Kulk *et al.* (2020, 2021) with version 6.0 of Ocean Colour Climate Change Initiative (OC-CCI) data as input (in green), compared with openly available time-series data from four other satellite-based primary production models from the Oregon State University Primary Production website ([http://orca.science.oregonstate.edu/-npp\\_products.php](http://orca.science.oregonstate.edu/-npp_products.php)) based on single-sensors (Sea-viewing Wide Field-of-view Sensor (SeaWiFS; 1998-2007), Moderate Resolution Imaging Spectroradiometer Aqua (MODIS-Aqua; 2003-present), and Visible Infrared Imaging Radiometer Suite (VIIRS; 2013-present)). The panels show the following: A) Global ocean primary production for the five different satellite-based primary production models for the time period between 1998-2022 (i.e., full data record), for all sensors combined; and B) Trends in primary production for the full ocean colour data record and for subsets of the periods during which specific sensors were operational, with stars indicating significant trends ( $p < 0.05$ ), for the five satellite-based primary production models. All latitudes were considered, but coverage at higher latitudes ( $>70^\circ\text{N}$  and S) is typically poor in satellite data.

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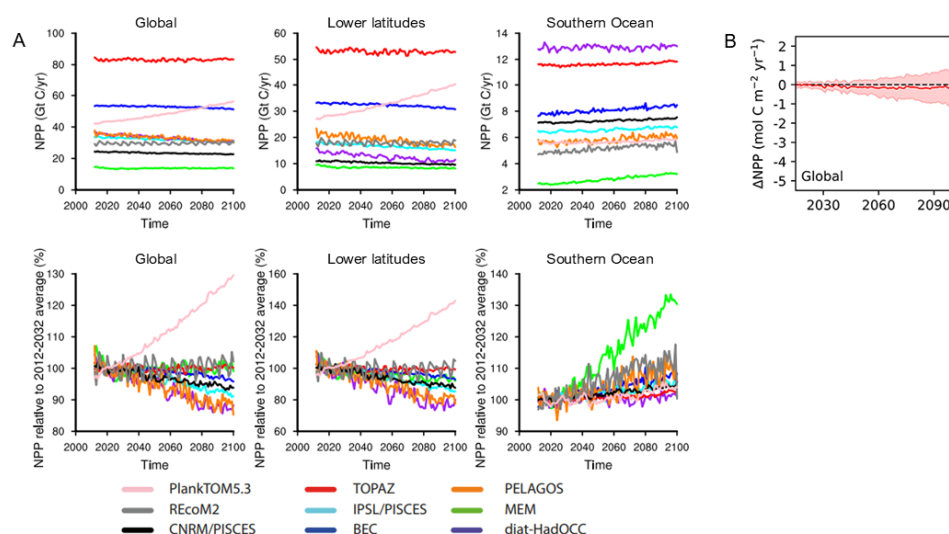
Differences in marine PP and its trends are not limited to satellite-based products. Earth System Model intercomparisons show even considerably larger uncertainty than the satellite models for the annual NPP estimate during the (recent past) “historical” period (i.e., 17-83 Pg C y<sup>-1</sup>; Bopp *et al.* 2013; Doney *et al.* 2014; Laufkötter

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157 *et al.* 2015; Tagliabue *et al.* 2021; see Figure 2), whilst showing weak or no trends over the recent historical period  
 158 (Kwiatkowski *et al.*, 2020). Ecosystem model uncertainties are even higher in future projections where models  
 159 disagree even on the sign of change to the year 2100 under the high emission scenario, although most ecosystem  
 160 models project a decline in global PP. While the uncertainty in annual NPP in the recent past has decreased in the  
 161 CMIP6 (Coupled Model Intercomparison Project phase 6) ensemble compared with CMIP5, the uncertainty in  
 162 projected PP trends has increased significantly in the CMIP6 ensemble compared with CMIP5 (Kwiatkowski *et al.*  
 163 *et al.* 2020). In particular, while the ensemble mean in CMIP5 suggested a significant decrease in PP at the global  
 164 scale of  $-8.06\% \pm 4.83\%$  (where the uncertainties are reported as the inter-model standard deviation), the CMIP6  
 165 ensemble has a much smaller mean and the standard deviation includes the null hypothesis of no trend ( $-1.76\%$   
 166  $\pm 9.01\%$ ). Frölicher *et al.* (2016) have noted that ecosystem model uncertainties (missing/mis-represented  
 167 processes, parameter uncertainties) dominated the total uncertainty in the 21<sup>st</sup>-century projections of PP and their  
 168 relative importance with respect to scenario uncertainty doesn't decrease with projection lead time. Recent studies  
 169 have confirmed this, highlighting the role of uncertainty in the representation of key biogeochemical processes,  
 170 including diazotrophy (Tagliabue *et al.* 2021; Bopp *et al.*, 2022; Doléac *et al.* 2025), bacterial remineralization  
 171 (Kim *et al.*, 2023) and parameter uncertainty (Jones *et al.* 2024), including in zooplankton grazing rates (Rohr *et al.*  
 172 *et al.* 2023). Laufkötter *et al.* (2015) concluded that the projected future changes in marine PP are driven by multiple  
 173 processes, including changes in circulation or mixing, leading to a stronger lateral or vertical loss of biomass;  
 174 increased aggregation or mortality of phytoplankton; or higher grazing pressure. Laufkötter *et al.* (2015) also  
 175 noted that temperature-dependent functions of PP and loss terms can affect the direction of change of PP from  
 176 marine ecosystem models in climate warming scenarios. It should be noted that regional variations in PP are  
 177 especially sensitive to how model represents these wide range of processes (Dutkiewicz *et al.* 2013), and given  
 178 the high uncertainty in their model representation, very few of the models agree on the direction of the trend  
 179 regionally. Furthermore, global models, with their coarse horizontal resolution, will especially struggle to capture  
 180 coastal processes that enhance PP, which makes them also apt to underestimate global PP.

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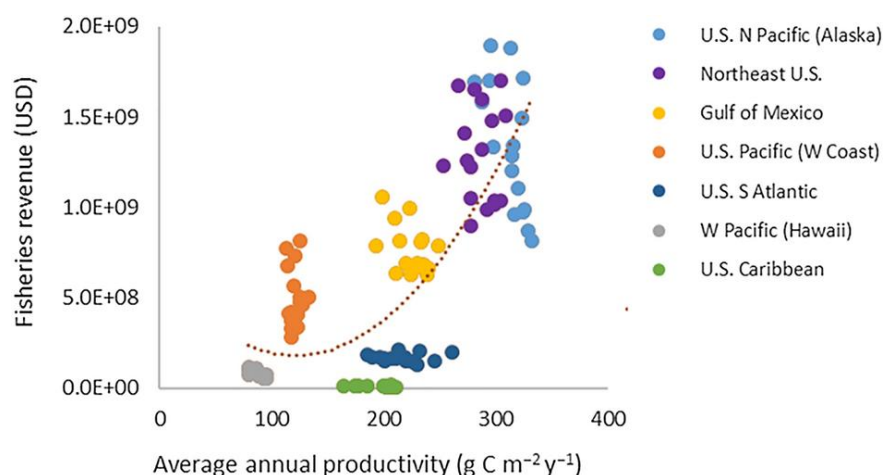


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184 **Figure 2.** Comparison of NPP from marine ecosystem models in CMIP5 comparison projected to the end of this  
 185 century under a high emission scenario. A) From Laufkötter *et al.* (2015) – RCP8.5 (Representative Concentration  
 186 Pathways 8.5 scenario) with from left to right global values, lower latitudes (30°S–30°N) and Southern Ocean (90–  
 187 50°S) in Gt C per year (top) and percent (bottom); and B) Global NPP projections from Tagliabue *et al.* (2021) –  
 188 SSP5-8.5 (Shared Socio-economic Pathways 8.5 scenario). Note that the magnitude of contemporary annual NPP  
 189 ranges from less than 20 to more than 80 Pg C y<sup>-1</sup> in the compilation from Laufkötter *et al.* (2015). Both analyses  
 190 showed negative and positive global trends, though most ecosystem models predict decreasing trends towards the  
 191 year 2100.

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 196 **Figure 3.** The impact of PP on fisheries. Figure reproduced from Marshak and Link (2021). Individual  
 197 observations from different coastal regions of the USA are indicated in different colours.

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199 Several studies have also been carried out to compare estimates from ecosystem models with satellite-based  
 200 products and *in situ* observations, both at global scale (Carr *et al.*, 2006, Steinacher *et al.* 2010; Bopp *et al.* 2013;  
 201 Laufkötter *et al.* 2015; Séférian *et al.* 2020) and at regional scales (Friedrichs *et al.*, 2009, Saba *et al.*, 2010, Lee  
 202 and Yoo 2016; Doléac *et al.* 2024). In some cases these comparisons (e.g., between ecosystem and satellite-based  
 203 models) led to better constrained PP projections, e.g., in the tropics, using an emergent constraint approach  
 204 (Kwiatkowski *et al.*, 2017). However, it is fair to say that overall these comparisons have not led to convergence  
 205 of model outputs that would reduce the uncertainty of marine PP estimates. All previous works have highlighted  
 206 large differences between estimates (e.g., varying from <-60% to >60%; Séférian *et al.* 2020), with highly variable  
 207 spatial patterns (Bopp *et al.* 2013). Tagliabue *et al.* (2021) highlighted the need for stronger constraints on NPP  
 208 using new approaches that include the growing observational coverage from Biogeochemical-Argo (BGC-Argo)  
 209 floats (Claustre *et al.* 2020, for an example of this see Arteaga *et al.*, 2022). Such field observations of PP, typically  
 210 treated as the “truth”, are often compared with model outputs to evaluate model performance. However, this type  
 211 of comparison is confounded by the errors in the field observations themselves, which can be quite high, as well



212 as by the differences in the spatial and temporal scales of the *in situ* observations and the validated models.  
 213 Furthermore, technologies such as BGC-Argo do not measure PP directly, but infer it from other variables.

214 Given these challenges both in remote sensing and ecosystem modelling, the IPCC has assigned low  
 215 confidence to current estimates of marine PP and its trends. The reasons cited include uncertainties in production  
 216 estimates and projections, the short duration of available time series data used in the analyses, and the lack of  
 217 independent validation (IPCC 2019; Gulev *et al.* 2021; Chapter 2 in IPCC AR6, WG I, 2021). This assessment is  
 218 of particular concern as it has major implications for ecosystem service provision, mitigation planning, enhancing  
 219 adaptation and building resilience to climate change (Hewitt *et al.* 2021). These applications often require regional  
 220 to local information as PP determines spatial variability in ecosystem services such as fisheries (Marshak and  
 221 Link 2021; see Figure 3), while uncertainties at these scales are further increased compared to the global estimates  
 222 (Tagliabue *et al.* 2021). Both remote sensing and ecosystem models can, in principle, deliver such regional  
 223 insights, when used with granularity and resolution needed at the appropriate scales. Reducing uncertainty in  
 224 models, ideally through a coordinated and internally consistent set of simulations, data and knowledge, would  
 225 then enable us to discuss downstream services under various climate scenarios and associated risks of tipping  
 226 points (Jones *et al.* 2024). Such improvements would support climate policy, as well as management decisions  
 227 pertaining to mitigation and adaptation, at both international and regional levels.

228 We argue here that efforts to reduce uncertainty in estimates and projections of marine PP should include a  
 229 focus on *investigating model structures and parametrisations*, with the goal of identifying genuine inter-model  
 230 differences and reconciling apparent differences. In this review, we examine both the sources of differences  
 231 between satellite-based and ecosystem models, as well as within these two types of models. We argue that there  
 232 is strong scientific justification for considering how the current model parameterisations could be improved. An  
 233 avenue to improvement might be to allow parameters that are currently treated as constants (e.g. the  $A_i$  parameters  
 234 from Table 1), to acquire different values at different spatial locations and times. Although this increases the  
 235 complexity of functional forms used in PP models, we argue that, at least in the less complex PP models (e.g.  
 236 within satellite models and ecosystem models used in ESMs for climate projections), there are both, good  
 237 scientific reasons to expect such parameter variations to be present, and to assume that absence of such variations  
 238 is responsible for the many apparent differences between the current PP models. Parameter variability might be  
 239 less important for the more complex models with large numbers of phytoplankton types and/or size-classes, but  
 240 for those models it is still essential to focus on the best possible ways of optimizing the existing constant  
 241 parameters. In general, we highlight the importance of correct parameterisations that are valid across multiple  
 242 spatial and temporal scales, and for multiple phytoplankton types. We also discuss the challenges posed by such  
 243 PP model parametrisations, argue that this is the right time to rise to those challenges, and propose strategies to  
 244 overcome them. Finally, we discuss uncertainties in marine PP that persist even when improved model  
 245 parametrisations are adopted.

246

### 247 3. Modelling primary production

248 In this section, we assess how marine PP is treated in satellite-based and ecosystem models, identifying inter-  
 249 model differences.

250 It is useful to consider GPP as the product of a biomass-specific production, say  $P^M$ , where  $M$  is a measure  
 251 of phytoplankton biomass, multiplied by the biomass itself. In other words:





$$P = P^M \times M, \quad (1)$$

such that  $P^M$  carries all the information on the physiological controls on PP, whereas  $M$  accounts for the role of varying phytoplankton concentrations. Since phytoplankton are complex organisms, many options exist for defining biomass, including concentrations of the phytoplankton pigment chlorophyll ( $B$ ), phytoplankton carbon ( $C$ ) or nitrogen content. The choice of biomass often depends on practical considerations (such as data availability) or by the study objectives (for example, carbon is an obvious choice in models designed to investigate the biologically mediated carbon cycle in the ocean). Models can also be classified according to which measure of biomass they track as the main currency in the ecosystem.

Dimensional analysis suggests that, in its simplest form,  $P^M$  can be represented in a canonical form with two parts: a scale factor  $P_m^M$  that carries the same dimensions as  $P^M$ , and a dimensionless function  $f_I$  of the scaled irradiance  $I_*$  available for photosynthesis (Platt and Sathyendranath, 1993), where the scaling factor would be a model parameter with the same dimensions as light. Thus, in such a canonical form,  $P^M$  can be written as:

$$P^M = P_m^M \times f_I(I_*). \quad (2)$$

In this form,  $P_m^M$  is not strictly constant, but implicitly accounts for the effects of other environmental variables on primary production, such as temperature ( $T$ ) and nutrients ( $N$ ), or of changes in species composition. Such dependencies can be made more explicit (removing  $T$  and  $N$  dependence from  $P_m^M$ ), such that Equation (1) becomes:

$$P^M = P_m^M \times F(f_T(T), f_N(N), f_I(I_*)). \quad (3)$$

The function  $F(f_T, f_N, f_I)$  can be specified as a simple product  $f_T \times f_N \times f_I$  (e.g., Laufkötter *et al.* 2015; Kishi *et al.* 2006; Vichy *et al.* 2007; Yool *et al.* 2013; Butenschön *et al.* 2016), representing co-limitation by each variable, or it can follow Liebig's law of the minimum (e.g., Gregoire *et al.* 2008; Daewel and Schrum 2013; Radtke *et al.* 2019) where the most limiting resource dictates the growth rate. Note that in Equations 2 and 3, the functions  $f_i$  are dimensionless, and that all the dimensions are carried by the scaling factor  $P_m^M$ . When models resolve multiple phytoplankton groups or species, then Equation (3) is specified for each group, and their contributions are added to get total PP.

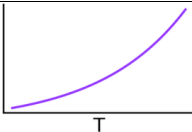
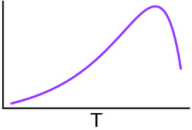
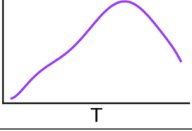
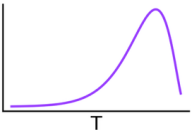
Table 1 summarises commonly used functions in PP models that represent the modulating roles of temperature, nutrients and light. When more than one nutrient is considered, additional terms have to be included for each nutrient. Thus, models differ depending on how many environmental factors are included in the model, and on the explicit functional forms selected for each modulating function. Ideally, the combined function  $F$  would have values within the  $[0,1]$  interval; however, this is often not the case for the temperature  $f_T$  function (as illustrated in Table 1).

In some cases, it is necessary to track multiple measures of phytoplankton biomass within a model. For example, a unit conversion between chlorophyll and carbon might be needed to make the exponent in the light function ( $f_I$ ) dimensionless, or it may be that the model tracks more than one currency. Such a conversion may also be needed to transform modelled phytoplankton carbon fields into chlorophyll fields for comparison with satellite-based chlorophyll products. This is typically achieved using a chlorophyll-to-carbon ratio ( $\theta$ , which varies among phytoplankton and under different environmental conditions and is usually estimated using photo-acclimation models. Commonly used functions in photo-acclimation models are also shown in Table 1. It shows that in those models  $f_I$  often takes a more general form and may also contain a dependence on temperature  $T$  and nutrients  $N$ .

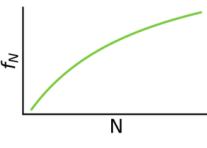
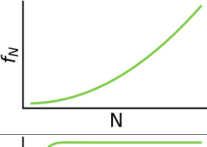
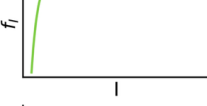
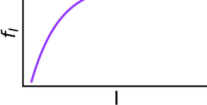
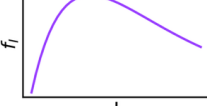

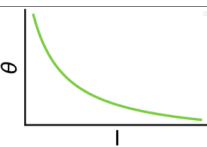




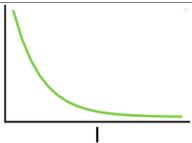
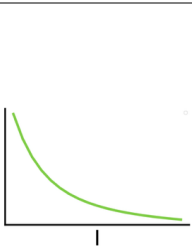
**Table 1.** The different  $f_T$ ,  $f_N$ ,  $f_I$  and  $\theta$  functions used across variety of CMIP and operationally used ecosystem models, as well as satellite models (which however typically do not explicitly use nutrient-limitation function). The ecosystem models explicitly mentioned are Biogeochemical Model for Hypoxic and Benthic Influenced areas (BAHMBI, Gregoire and Soetaert (2010)), Biogeochemical Flux Model (BFM, Vichi et al. 2015), ECOSystem Model (ECOSMO, Daewel and Schrum, 2013), European Regional Seas Ecosystem Model (ERSEM, Butenschön et al. 2016), Hadley Centre Ocean Carbon Cycle (HadOCC, Totterdell, 2013), Model of Ecosystem Dynamics, Sequestration and Acidification (MEDUSA, Yool et al. 2013), Marine Ecosystem Model (MEM, Shigemitsu et al. 2012), North-Pacific Ecosystem Model for Understanding Regional Oceanography (NEMURO, Kishi et al. 2007), PELAgic biogeochemistry for Global Ocean Simulations (PELAGOS, Vichi et al. 2007), Pelagic Interactions Scheme for Carbon and Ecosystem Studies (PISCES, Aumont et al. 2015), Carbon, Ocean Biogeochemistry and Lower Trophics (COBALT, Stock et al., 2020, 2025), DARWIN model (Ward et al. 2012).  $P_h$ , C and N represent the concentrations of nutrients (such as phosphate, carbon, and nitrogen).  $A_i$  and  $D_i$  stand for the different model parameters. In photoacclimation models  $\theta$  is chlorophyll-to-carbon ratio.

Process / Structure	Equation	Description & remarks	Graphical Representation	Examples	Key References
Temperature limitation on photosynthesis	$f_T = e^{A_1 T}$	Exponential temperature dependence on growth rate.		BAHMBI, MEDUSA, NEMURO, BFM, PISCES, PELAGOS, COBALT, DARWIN	Eppeley (1972)
	$f_T = Q_{10} \frac{T-A_2}{A_2} - Q_{10} \frac{T-A_3}{A_4}$	Phytoplankton growth rate increases initially exponentially, with enzyme inhibition above optimal temperature		ERSEM	Blackford et al. (2004)
	$f_T = \sum_{i=0}^7 D_i T^i$	Phytoplankton growth rate is represented as an empirical seventh-order polynomial function, fit to observed data.		Vertically Generalised Production Model (VGPM)	Behrenfeld & Falkowski (1997)
	$f_T = A_5 \times e^{A_6 \times T} \times \left(1 - \frac{T-A_7}{A_8}\right)^2$	Function designed to model individual phytoplankton species or types according to their temperature traits, in multi-species models. It has yet to be used routinely in global-scale simulation models, except in a special case of DARWIN.		A version of DARWIN	Norberg (2004), Thomas et al. (2016); Sauterey et al. 2024); Krinos et al. (2025)
	N/A	No explicit temperature dependence is included in the model structure	N/A	ECOSMO, HadOCC	Yumruktepe, Samuelsen, Daewel (2022)
	Empirical assignment	Indirect methods. An example is province-based assignment of parameters	N/A	Satellite P&S, BICEP	Sathyendranath & Platt (1988), Longhurst et al. (1995), Sathyendranath



					<i>et al. (2020)</i> , Kulk <i>et al. (2020)</i>
N-limitation	$f_N = \left( \frac{\left( \frac{p_h}{c} \right) - A_9}{A_{10} - A_9} \times \left( \frac{N}{c} \right)^{0.5} \right)^{0.5}$	Describes nutrient limitation based on internal nutrient quota for phytoplankton cells. Droop model of cell quota. $0 \leq f_N \leq 1$ depends on the instantaneously calculated internal cell C/N and C/P ratios $\left( \frac{p}{c}, \frac{q}{c} \right)$ and the maximum C/N and C/P ratios ( $A_5, A_7$ ), having subtracted the structural content of the cell ( $A_4, A_6$ ) from each.	N/A	ERSEM PISCES (for iron), DARWIN (quota version)	Droop (1974);
	$f_N = \frac{N}{N + A_{13}}$	Michaelis-Menton Equation. Describes N-limitation as a saturating function of external nutrient concentration, and the half saturation coefficient $A_{13}$ for that nutrient		NEMURO, ECOSMO, BFM, MEDUSA, HadOCC PISCES (for all nutrients except iron), DARWIN (monod version)	Michaelis and Menton (1913), Kovarova-Kovar and Egli (1998), Lee <i>et al.</i> (2015)
	$f_N = \frac{(1-f_A) \times A_{14} \times N}{(1-f_A) \times A_{14} + N}$ $f_A = \frac{1}{1 + \sqrt{\frac{A_{14} \times N}{A_{15}}}}$	Optimal uptake kinetics		MEM	Smith <i>et al.</i> (2009)
Light Limitation	$f_I = I \times e^{1-A_{16} \times I}$	Photosynthesis rate increases then declines at high light intensities due to photoinhibition.		NEMURO	Steele (1962)
	$f_I = (1 - e^{-A_{17} \times I})$	Photosynthesis follows a light saturation curve with no inhibition at high light levels		Satellite	Platt <i>et al.</i> (1980; 1990) Sathyendranath <i>et al.</i> (2020); Kulk <i>et al.</i> (2020)
	$f_I = (1 - e^{-A_{18} \times I}) \times e^{-A_{19} \times I}$	Photosynthesis follows a light saturation curve with inhibition at high light levels.		BAHBI, MEM	Platt <i>et al.</i> (1980)
	$f_I = \tanh(A_{20} \times I)$	Model with no photoinhibition. A hyperbolic tangent function is used to simulate light saturation curve.		ECOSMO	Jassby & Platt (1976)
Photo-acclimation	$\theta = \frac{A_{21}}{\left( 1 + \frac{A_{21} A_{22} I}{2 A_{23}} \right)}$	Photo acclimation model based on the concept of resource allocation, with maximum Chl-to-carbon ratio reached as light approaches zero.		ERSEM, BFM, PISCES, PELAGOS, COBALT, DARWIN	Geider <i>et al.</i> (1997,1998)



	$\theta = 0.022 + (0.045 - 0.022)e^{-3I} - g(N, T)$	The Chl-to-carbon ratio ( $\theta$ is determined by photoacclimation, and also by nutrient and temperature stress.		CBPM	Westberry <i>et al.</i> (2008)
	$\theta = \frac{A_{25}}{I \times A_{24}^{-1}} \times \left(1 - e^{-(I \times A_{24}^{-1})}\right)$	Based on an extended version of Geider <i>et al.</i> (1997, 1998) photoacclimation model. Uses the exact analytic solution to the Guider <i>et al.</i> (1997) model Jackson <i>et al.</i> (2017), extended to account for spectral light effects (Sathyendranath <i>et al.</i> 2020). It incorporates photo-acclimation effects on the chlorophyll-to-carbon ratio		Satellite	Sathyendranath <i>et al.</i> (2020), Jackson <i>et al.</i> (2019), Zheng <i>et al.</i> (2025)

306



307 In the next two sections, we examine in more detail the variety of ways in which these concepts are implemented  
 308 in satellite-based and ecosystem models.

309

### 330 Satellite-based models

311 In satellite-based PP models, daily water-column production is calculated as a function of phytoplankton biomass  
 312 and light available at the sea surface, obtained from ocean-colour remote-sensing observations, coupled with  
 313 models of photosynthetic response to light. Since the launch of the first ocean-colour satellite, the Coastal Zone  
 314 Color Scanner (CZCS) in the 1970s, scientists have developed various satellite-based PP models that can be  
 315 roughly categorised into three classes: 1) Chlorophyll-based models, 2) Absorption-based models, and 3) Carbon-  
 316 based models (Figure 4). Each of these models differ from each other according to whether they are implemented  
 317 as linear/non-linear, spectral/non-spectral, vertically-uniform/vertically-non-uniform, or as a combination of these  
 318 (Platt & Sathyendranath 1993; Sathyendranath & Platt 2007). Further bifurcations occur, depending on whether  
 319 or not the models are depth-integrated (Friedrichs *et al.* 2009). Most of the satellite-based models do not resolve  
 320 PP by phytoplankton size classes, or functional types, with some exceptions, such as Uitz *et al.* (2010), Brewin *et*  
 321 *al.* (2016) and Tao *et al.* (2017).

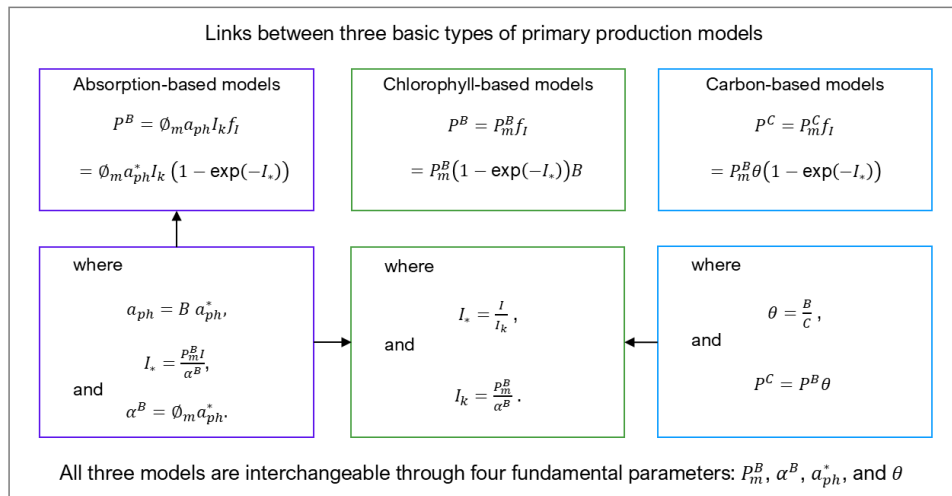
322 Satellite-based model outputs have been compared against in situ data, both globally and regionally  
 323 (Friedrichs *et al.* 2009; Saba *et al.* 2010, Lee *et al.* 2015). No clear winners have emerged from these  
 324 intercomparisons (and perhaps selecting winners was not an objective of the comparisons), and the assignment of  
 325 model parameters remains one of the biggest sources of uncertainty in estimates of primary production from  
 326 remote sensing observations (Platt & Sathyendranath 1993; Sathyendranath & Platt 2007, Sathyendranath *et al.*  
 327 2009; Kulk *et al.* 2020, 2021, Brewin *et al.*, 2023).

328 Interestingly, the types of models described above all converge to the same principles and a common set of  
 329 parameters (Sathyendranath & Platt 2007; Figure 4). Chlorophyll-based (or available-light or photosynthesis-  
 330 irradiance) models typically use the parameters of the photosynthesis-irradiance curve, normalised to  $B$ , the  
 331 concentration of chlorophyll- $a$ , i.e., the initial slope ( $\alpha^B$ ) and the assimilation number ( $P_m^B$ ) of the light saturation  
 332 curve, and the photoacclimation parameter ( $I_k = P_m^B / \alpha^B$ ) derived from the other two (Platt *et al.* 1980;  
 333 Sathyendranath and Platt 2007; Figure 4). Absorption-based (which are also called biomass-independent or  
 334 inherent-optical-property) models use the realised maximum quantum yield ( $\phi_m$ ) and the absorption coefficient  
 335 of phytoplankton ( $a_{ph}$ ) (Kiefer & Mitchell 1983, Lee *et al.* 2015). This model can be shown to be equivalent to  
 336 the photosynthesis-irradiance models by using the identity  $\phi_m = \alpha^B / a_{ph}$  (Platt *et al.* 1988; Sathyendranath and  
 337 Platt 2007; Figure 4). The key parameter in carbon-based (or growth) models is the growth rate ( $g$ ), i.e., the rate  
 338 of change of carbon per unit time normalised to the initial phytoplankton carbon concentration. The chlorophyll-  
 339 to-carbon ratio ( $\theta$ ) can be used to transform growth models to production models and vice versa (Sathyendranath  
 340 & Platt 2007; Sathyendranath *et al.* 2009). Thus, the different types of satellite-based primary production models  
 341 are interchangeable through a common set of parameters: the initial slope ( $\alpha^B$ ) and assimilation number ( $P_m^B$ ) of  
 342 the light saturation curve, the mean specific absorption coefficient of phytoplankton ( $a_{ph}^*$ ), and the chlorophyll-  
 343 to-carbon ratio ( $\theta$ ) (Sathyendranath and Platt 2007; Sathyendranath *et al.* 2009). When the light incident at the  
 344 sea surface exceeds a threshold above which light can damage the photosystems, a photo-inhibition term has to  
 345 be added to the photosynthesis-irradiance equation (Platt *et al.* 1980). This parameter is often not used in satellite-  
 346 based models; a sensitivity analysis on a photosynthesis-irradiance model (Platt *et al.* 1990) showed that



incorporation of realistic values of the photo-inhibition parameter into the model had only small to negligible effect on computed water-column primary production, which lends some justification to why this term is often ignored. But this is a simplification that can be readily dropped, if new evidence suggests that photo-inhibition could be important at large scales.

Spectral models of primary production are designed to capture the wavelength-dependent light penetration underwater, and wavelength-dependent photosynthesis (Sathyendranath and Platt, 1989). In fully-spectral models, the action spectrum of photosynthesis (which describes the wavelength-resolved values of the initial slope  $\alpha^B$  is coupled to the light available at corresponding wavelengths for photosynthesis (Sathyendranath & Platt, 1989; Kyewalyanga *et al.*, 1992), such that the product  $\alpha^B I$  that appears in non-spectral models has to be replaced by the wavelength integral  $\int \alpha^B(\lambda) I(\lambda) d\lambda$ , where  $\lambda$  represents the wavelength, and the integral is taken over the photosynthetically active range (400-700 nm). The spectral form of the action spectrum closely resembles that of the phytoplankton absorption spectrum ( $a_{ph}$ ) (Sathyendranath *et al.* 1989b; Kyewalyanga *et al.* 1997). Spectral effects are generally considered to be not relevant at saturating light levels. Therefore, if the light available is blue-rich, where action spectrum is maximum, then the coupling between light and photosynthesis would be stronger than if the light were green-rich, where the action spectrum typically goes through a minimum. We know from previous studies that spectral and non-spectral models may differ from each other in a systematic manner (Sathyendranath & Platt 2007), because non-spectral models are not able to account for the covariance (or the lack of it) between spectrally-resolved  $\alpha^B$  and  $a_{ph}$ . To some extent, the impact of the spectral effects on water-column primary production could be accommodated into non-spectral models by suitably tuning the parameters of non-spectral models (Platt and Sathyendranath 1991). Typically, therefore, one anticipates systematic differences between spectral and non-spectral models of marine primary production, unless model parameters are adjusted to compensate for the difference.



**Figure 4.** Phytoplankton absorption-, chlorophyll- and carbon-based primary-production models commonly-used in satellite-based approaches, and the parameter transformations between them. Notations: Primary production (P), light-limitation function ( $f_I$ ), as in Table 1, assimilation number of the saturation-light curve, or the light saturation parameter ( $P_m^B$ ), initial slope of the light-saturation curve ( $\alpha^B$ ), mean absorption coefficient of



374 phytoplankton ( $a_{ph}$ ), chlorophyll-to-carbon ratio ( $\theta$ ), chlorophyll-specific absorption coefficient of  
 375 phytoplankton ( $a_{ph}^*$ ), realised maximum quantum yield of photosynthesis ( $\Phi_m$ ), photoacclimation parameter of  
 376 the light-saturation curve ( $I_k$ ), phytoplankton biomass in chlorophyll-a units ( $B$ ), normalised irradiance ( $I_*$ ),  
 377 irradiance ( $I$ ), phytoplankton carbon biomass ( $C$ ), time ( $t$ ), and growth rate ( $g$ ). One of the ( $f_i$ ) functions from  
 378 Table 1 was selected here for illustrative purposes, but other functions have also been used in the literature. As  
 379 shown below (Figure 7), numerically, most of the ( $f_i$ ) functions are almost identical to each other, unless photo-  
 380 inhibition is introduced. Currently, remote-sensing-based primary-production models do not incorporate the  
 381 photo-inhibition term.

382

### 383 3.2 Ecosystem models

384 Ecosystem models differ greatly in their complexity, ranging from simple, three-component Nutrient-  
 385 Phytoplankton-Zooplankton (NPZ) models (Fasham *et al.* 1990; Steele and Henderson 1992; Franks, 2002;  
 386 Gentleman, 2002) to highly complex ones with hundreds of ecosystem components (e.g., Dutkiewicz *et al.*, 2020,  
 387 Fennel *et al.* 2022). Some models use a single nutrient (usually nitrogen, or carbon) as the model currency,  
 388 assuming a fixed stoichiometry (relationship between biogeochemically-important elements), whereas other  
 389 models allow for dynamically resolved stoichiometry within  $f_N$ . In this section, we focus primarily (but not  
 390 exclusively) on marine ecosystem models (here used interchangeably with “marine biogeochemical models”)  
 391 that participate in the Climate Model Intercomparison Project (CMIP) (e.g., Laufkotter *et al.* 2015; Kwiatkowski  
 392 *et al.* 2020), as well as regional ecosystem models that are run operationally by forecasting centres (e.g., Fennel  
 393 *et al.* 2019) for regional climate projections. In these models, primary production is usually estimated along the  
 394 lines of equations 2 and 3, where primary production ( $P$ ) is calculated by multiplying the phytoplankton biomass  
 395 (usually carbon) by its reference growth rate  $g$ , modulated typically by the three functions,  $f_T$ ,  $f_N$  and  $f_I$ .

396 There are also many similarities across the ecosystem models that go beyond the functional form of Equation  
 397 (2), and a few common approaches can be identified in the equations used to express the functions  $f_T$ ,  $f_N$  and  $f_I$ ,  
 398 (Table 1). For instance,  $f_T$  is typically described through an exponential function (originating from Eppley *et al.*  
 399 1971; e.g., see Laufkotter *et al.* 2015), that might include inhibition at temperatures much higher than those  
 400 optimal for the species’ growth (e.g., Norberg 2004; Butenschön *et al.* 2016, Dutkiewicz *et al.*, 2020a), which is  
 401 based on  $Q_{10}$ , a measure of the sensitivity of photosynthesis to temperature. However, most models resolve groups  
 402 of phytoplankton (e.g. diatoms) rather than individual species, and they do not have temperature inhibition  
 403 assuming that there is a spectrum of diatoms that have optima across the full temperature range (see e.g. Anderson  
 404 *et al.* 2020). Furthermore, some models do not have explicit PP temperature dependence at all (e.g., Daewel and  
 405 Schrum 2013). When multiple nutrients are considered, the  $f_N$  function is typically formulated to use Liebig’s law  
 406 of minimum to combine their effects on PP, and is often based either on cell quota of nutrients within the cells  
 407 (Droop 1974), or on the concentrations of the nutrients in the medium (Michaelis and Menten 1913). In some  
 408 cases (e.g., Shigemitsu *et al.* 2012),  $f_N$  is based on the optimal uptake kinetics (Smith *et al.* 2009), which allows  
 409 for parameters in the Michaelis-Menten equation to vary (Table 1). A variety of equations are currently in use to  
 410 describe the light-dependence function ( $f_I$ , see Table 1) in ecosystem models, including those from Platt *et al.*  
 411 (1980), Steele (1962), and Jassby and Platt (1976), some of which account for the effect of photo-inhibition at  
 412 high light, whilst others do not. Furthermore, many of the ecosystem models also include photoacclimation, either  
 413 as part of the  $f_I$  function, or as an additional term, mostly following the model of Geider *et al.* (1997,1998).



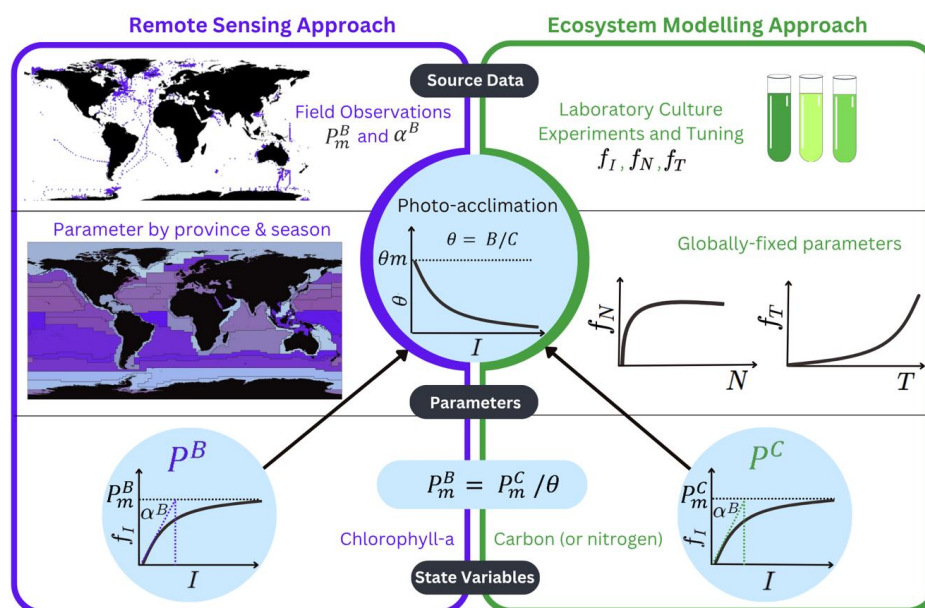
Other significant differences across ecosystem photosynthesis models include the number of phytoplankton functional types and size-classes represented, the number of limiting nutrients included (and the types of equations selected to represent the role of each nutrient), and the number of wavebands considered in representation of irradiance (the level to which light is spectrally and directionally resolved, e.g., see Platt and Sathyendranath 1991, Dutkiewicz et al. 2015, Gregg & Rousseaux, 2016). Practically all ecosystem models include nitrogen limitation. But iron limitation is also considered important, as is silica limitation, especially in those models that include diatoms as a phytoplankton class. Phosphate limitation becomes important as well, in particular when dealing with nitrogen-fixing organisms. Another fundamental difference lies in the representation of the production and remineralisation of particulate and dissolved organic matter which are included in the models as explicit or implicit processes, affecting the model parametrisations up to the formulation of gross primary production which may or may not include exudation (e.g. Butenschön et al 2016, Vichi et al. 2007, Wu et al. 2021).

### 3.3 Comparison of satellite-based and ecosystem models

Satellite-based and ecosystem models for estimating ocean PP have some similarities, but also key distinctions (see Figure 5, also IOCCG 2020). Model parameter assignment provides one clear perspective on a difference between the two types of models. For example, parameters associated with PP models in the satellite-based approach of Platt and Sathyendranath (1988), Kulk et al. (2020) and Sathyendranath *et al.* (2020) are established from field observations, whereas ecosystem model parameters are typically estimated using information from laboratory experiments conducted under controlled conditions, followed by tuning the model towards the available observations. But here also, the distinction is not total: for example, the carbon-based production model of Behrenfeld *et al.* (2005) relies on culture measurements to establish growth rate and carbon-to-chlorophyll ratio. The satellite-based models that do not have explicit nutrient and temperature dependencies, inherently contain those dependencies in the model parameter values, which are allowed to vary across biogeographical provinces and seasons (e.g., see Figure 6; for model described by Longhurst *et al.* 1995 as updated by Sathyendranath *et al.* 2020 and Kulk *et al.* 2020, 2021), representing different nutrient and temperature limiting environments. Ecosystem models typically represent the nutrient and temperature limitation explicitly, with different parameters assigned to each plankton group. Another difference in parameterisation is that many ecosystem models use maximum carbon or nitrogen-specific production rate under optimal conditions as a model parameter and carbon or nutrient biomass is then used to estimate primary production (Figure 5). While carbon-based satellite algorithms for primary production are similar to ecosystem models in this respect, other satellite models rely on bio-optical properties such as chlorophyll concentration or phytoplankton absorption coefficient as the state variable. Some ecosystem models also include a photo-inhibition term, to represent the reduction in photosynthesis under high light intensities, whereas satellite-based models typically account only for the saturating response to light without including photoinhibition. Another process linked to photosynthesis, photo-acclimation, is generally addressed by both approaches and though there are differences, variations of the Geider *et al.* (1997, 1998) models are now common to both satellite-based and ecosystem models.

Finally, the ecosystem models are able to compute depth-resolved PP, as is the case for the satellite-based method proposed by Platt and Sathyendranath (1988), whereas some other satellite-based models are designed to yield vertically integrated production (e.g., Behrenfeld and Falkowski 1997).

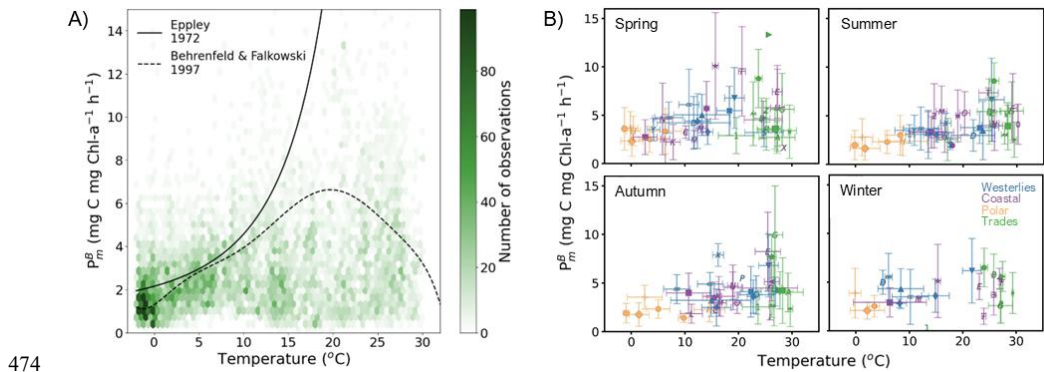




**Figure 5.** Comparison of satellite remote sensing (left) and ecosystem modelling (right) approaches to computing marine primary production, and where they interact (light blue) through the photo-acclimation model which is essential to enable comparison between the models.  $I$  = Light,  $N$  = Nutrient,  $T$ =Temperature. It should be noted that although carbon, or nitrogen, are the most common currency used by the ecosystem models, there are also ecosystem models which use chlorophyll-a as the currency.

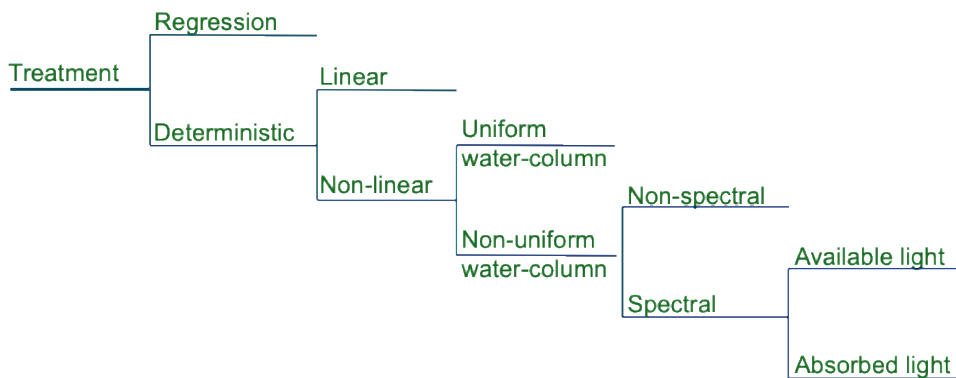
Thus, the differences between satellite-based and ecosystem-based models of primary production are not clear cut. All satellite-based models are data-rich, in the sense that they are designed to exploit satellite observations, typically with global coverage and nominal daily repeat frequency. Some use culture data as auxiliary information; others rely on in situ field observations. Ecosystem models, on the other hand, tend to be data-sparse; even when operated in data assimilation mode, only a fraction of the modelled ecosystem compartments or fluxes are usually constrained by assimilation. The constraints imposed by satellite data availability limit the processes and variables that can be estimated, whereas ecosystem models tend to be rich in outputs they provide.

Platt and Sathyendranath (1997) proposed a hierarchy of primary production models (Figure 7). Almost all the types of models in this hierarchical classification, with the exception of linear models and purely statistical models, are represented in PP models under discussion in this paper. With the exception of absorbed-light models that are in use in satellite-based models, but not in ecosystem models, the different classes of models are common to both. In this regard, the diversity of models within satellite-based or ecosystem-based approaches is no smaller than across those two groups of models.



**Figure 6.** Variability in the photosynthesis-irradiance parameter  $P_m^B$  in the ocean. A) Parameter values from a global in situ dataset (Bouman et al. 2018; Kulk et al. 2020) plotted as a function of temperature. Two commonly-used temperature-dependent equations (Eppley 1972; Behrenfeld and Falkowski 1997) of this parameter are also shown. B) The same data sorted according to ecological provinces of Longhurst (2007) and according to season, with colours representing four different oceanic biomes (Longhurst 2007), showing that some structure and pattern emerge when the data are organised according to oceanic biomes and to a smaller degree seasons.

### Hierarchy of Primary Production Models



**Figure 7.** Hierarchy of primary production models. The models get more complete (and more complex), as we go from left to right, and from the upper to the lower limb of each branch.

#### 4. How similar are the different primary production models?

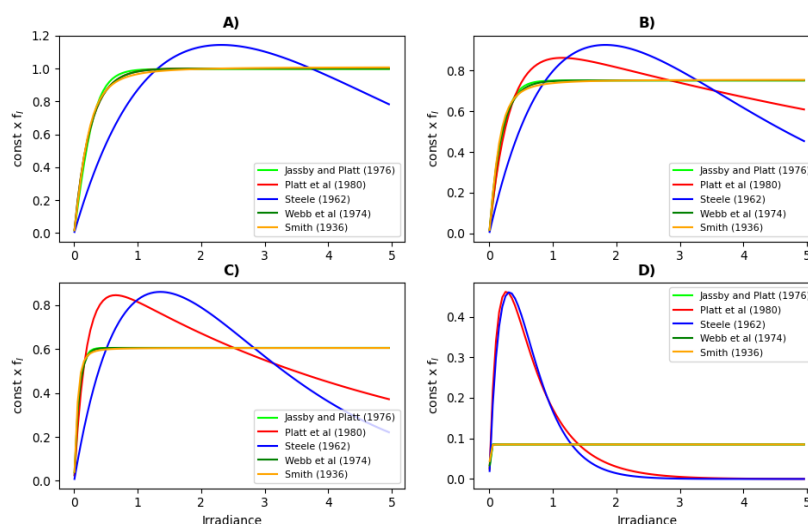
Platt and Sathyendranath (1993) showed that we can anticipate systematic biases between satellite-based models that are structured differently, and we can numerically predict under what conditions the biases will manifest themselves. For example, linear and non-linear models are expected to behave similarly under low-light levels but to diverge as light levels increase. However, they showed that when PP models have similar structures, it is possible to reduce all of them to a common, canonical form, revealing that apparently-different model types (available light models, absorbed light models, chlorophyll-based or carbon-based models) become equivalent when implemented with comparable model parameter values (Platt and Sathyendranath 1993; Sathyendranath and



494 Platt 2007; Sathyendranath *et al.* 2020). Such comparisons also reveal systematic biases between spectral and  
 495 non-spectral models of PP, arising from spectral effects in both underwater light penetration and phytoplankton  
 496 light utilisation. It was demonstrated that biases between spectral and non-spectral PP models can be minimised  
 497 by tuning the diffuse attenuation coefficient of downwelling irradiance, which determines the rate of change of  
 498 available light with depth (Platt and Sathyendranath 1991; Kyewalyanga *et al.* 1992). Similarly, Kovač *et al.*  
 499 (2016a) demonstrated that a locally tuned non-spectral model, with adjusted values of photosynthesis parameters,  
 500 can outperform a spectral model, without locally tuned values of photosynthesis parameters. Such comparisons  
 501 bring to the fore the importance of parameter assessment, assignment, and evaluation to understand model  
 502 performances, uncertainties and divergences, which is at the core of this review.

503 To illustrate the point, let us focus, for example, on the light function ( $f_l$ ), which takes a wide range of forms  
 504 in the literature (see Table 1). Even though the functional forms cannot be analytically transformed into each other  
 505 (they are mathematically different), numerically they could still be very close to each other, in the sense that they  
 506 can all fit the same observations similarly well when the parameters are chosen appropriately, for the range of  
 507 light values commonly encountered at sea (Kovač *et al.*, 2017). These different forms split into two classes: one  
 508 that includes photo-inhibition and the other that does not (Amirian *et al.*, 2025). Figure 8 shows that the  $f_l$  models  
 509 without photo-inhibition (Webb *et al.* 1974; Jassby and Platt 1976; Smith 1936) are all practically identical to  
 510 each other for equivalent parameter values and are therefore basically indistinguishable from each other. It should  
 511 also be noted that the Webb *et al.* (1974) model is a special case of the Platt *et al.* (1980) model for the case of  
 512 zero photo-inhibition. The  $f_l$  model that stands out is the one of Steele (1962), which struggles to match the other  
 513  $f_l$  models under low light conditions. However, when photo-inhibition is important, the  $f_l$  model of Platt *et al.*  
 514 (1980) can again nicely coincide with Steele (1962) for a suitable combination of their parameter values. What  
 515 we learn from Figure 8 is that a lot of the diversity in  $f_l$  models is only apparent, as the diversity can be eliminated  
 516 via model parametrisation.

517



518



519 **Figure 8.** Comparing the functional forms of four  $f_I$  models in different regimes. Since only the functional forms  
 520 are compared, the x and y axes do not necessarily represent realistic values of irradiance and  $f_I$ , or alternatively  
 521 the units in which these variables are expressed are not relevant. The Figure shows the degree to which the five  
 522 different models can be "tuned" to each other through fitting their parameters in a suitable way. The functional  
 523 forms for the  $f_I$  models are introduced in Tab.1, except the model by Webb *et al.* (1974), which is a special case  
 524 of Platt *et al.* (1980) for zero photoinhibition (setting  $A_{14} = 0$ , see Tab.1). However, unlike Tab.1, we include  
 525 here into the free  $f_I$  parameters also a multiplicative constant (hence what is plotted here is  $\text{const} \times f_I$ ), which can  
 526 be always absorbed into the  $P_m^M$  parameter of equations (2-3). The different panels A-D show cases of increasing  
 527 photoinhibition as modelled by the most complex Platt *et al.* (1980) model (A is the lowest, D the highest), with  
 528 the other models calibrated to best fit the curve corresponding to Platt *et al.* (1980). We see that the five models  
 529 essentially split into two families, each representing well a limiting case of either no photoinhibition (Jassby and  
 530 Platt 1976, Webb *et al.* 1974, Smith 1936), or very high photoinhibition (Steele 1962).

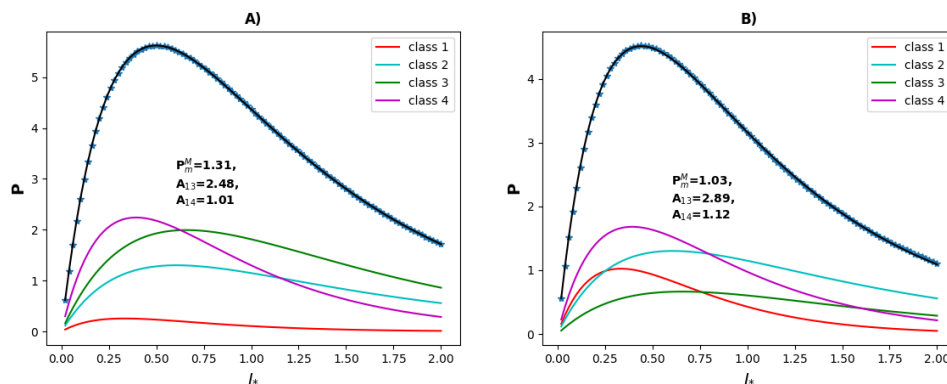
531

532 In general, PP models are designed to represent limitations to phytoplankton growth (whether from light,  
 533 nutrients or temperature) under different environmental conditions. These models have the potential to be  
 534 generalised to deal with additional external conditions (which may not be explicitly included in the model) by  
 535 incorporating spatially and temporally variable parameter values. This flexibility allows models to account for the  
 536 diversity of plankton and the processes responsible for their dynamics, which are not explicitly represented in  
 537 current models. Representing the full diversity of planktonic species is not feasible due to lack of understanding  
 538 and computational demand, which is why models typically rely on the use of plankton classes to represent  
 539 aggregations of multiple species based on shared characteristics or traits, such as body size, life strategies and  
 540 behaviours. This approach captures at best the average or most typical behaviour of each class (e.g., Anderson  
 541 2020; Ratnarajah *et al.* 2023). When aggregating species according to their physiological and functional traits and  
 542 behavioural patterns into a pre-defined number of modelled classes, fixed values are assigned to model parameters  
 543 within each aggregated class. For ecosystem models, many of these parameters have assigned values on the basis  
 544 of laboratory or mesocosm experiments (Geider *et al.* 1998; Schartau *et al.* 2017; Ratnarajah *et al.* 2023), often  
 545 focusing on a small number of carefully-selected species, far from capturing the full diversity of organisms or  
 546 their responses and behaviours that might be expected in the natural environment across large spatio-temporal  
 547 scales (Geider *et al.* 1998; Schartau *et al.* 2017; Ratnarajah *et al.* 2023). In contrast, in the natural environment,  
 548 we can expect parameters to vary in time and space, reflecting both changes in the governing conditions and in  
 549 the unresolved functional diversity in the makeup of planktonic communities (Schartau *et al.* 2017). Such  
 550 parameter variability can be observed in model calibration experiments (e.g., Leeds *et al.* 2011; Mattern *et al.*  
 551 2012, 2014), including those using data assimilation to estimate model parameters jointly with the model state  
 552 (e.g., Pastres *et al.* 2003; Tijjputra *et al.* 2007; Roy *et al.* 2012; Doron *et al.* 2013; Simon *et al.* 2015; Gharamti *et al.*  
 553 2017a,b; Skákala *et al.* 2024).

554 A simple illustration of how parameter variability emerges from aggregating species in classes is provided  
 555 in Figure 9. Although many models differ in the number of phytoplankton classes they resolve, for each  
 556 phytoplankton class they typically use the same functional form to describe photosynthesis, with total  
 557 phytoplankton primary production corresponding to the sum of contributions across all classes. Figure 8  
 558 demonstrates that models with different numbers of classes become equivalent in their description of total PP,



provided that the parameters in models with fewer classes are allowed to vary in space and time. In such way, spatio-temporal parameter variations effectively capture the influence of unresolved diversity in plankton community structure in models with fewer plankton classes. The spatio-temporal model parameter variations are then a consequence of models' inability to sufficiently resolve plankton species, which also means that such parameter variability is expected to be especially relevant for simpler models (e.g. ecosystem models typically used in ESMs). We acknowledge that the most complex models currently in use (e.g. DARWIN, see Ward *et al.*, 2012, Dutkiewicz *et al.*, 2020a) have less reason to adopt spatio-temporally variable parameters, but these models are typically too computationally expensive to be run as part of ESMs in long-term ensemble-based climate projections and even as complex as they are, they still represent a fraction of the real-world diversity. On the other hand, as the models get more complex by incorporating more ecosystem compartments, the challenge shifts to calibrating large numbers of parameters to describe the functions of each of the model components.



**Figure 9.** A simple illustration of how unresolved phytoplankton community structure can lead to parameter variability. In both panels, we plot PP expressed as  $P = P_m^M \cdot f_I(I_*)$  with functional form  $f_I$  corresponding to the Platt *et al.* (1980) model (see Tab.1). As in Fig.6, ranges of irradiance,  $I_*$ , and PP values are chosen arbitrarily, purely to demonstrate the essential point made in section 2. Four phytoplankton classes are plotted each with different  $P_m^M, A_{13}, A_{14}$  parameters. The dark blue dots are obtained by summing up the PP across the four classes (this corresponds to PP of total phytoplankton) and the dark blue line is the fit of the points with the same functional form used for the four phytoplankton classes assuming the total phytoplankton concentration is the sum of the concentrations of the four classes. The two panels show two situations where the same total phytoplankton concentration is distributed into classes in different ways (the phytoplankton community structure changes). We can see that if we did not resolve the four classes, we could still use the Platt *et al.* (1980) model (including photoinhibition) for the total phytoplankton, but  $P_m^M, A_{13}, A_{14}$  parameters would vary depending on the (unresolved) variations of the phytoplankton community structure.

## 5. A way forward

Together, these considerations suggest that investigating parameter assignment and parameter variability may be an important route to understand and potentially reduce many of the apparent differences between marine PP models, and hence in the estimated magnitudes of production. Investigation into the role of parameters should be



589 followed by a consistent calibration against the same observational data. To estimate spatially and temporally  
 590 varying parameters in ecosystem models, data assimilation can provide a natural tool for model calibration (e.g.,  
 591 Tjiputra *et al.*, 2007; Singh *et al.*, 2025). However, introducing spatio-temporally variable (or too many constant)  
 592 parameters comes with its own challenges. For example, allowing the (often many) model parameters to vary  
 593 substantially increases model flexibility at the risk of overfitting to observations, particularly if the number of  
 594 model parameters is large or observational data are insufficient. Overfitting may reduce the model ability in  
 595 predicting new phenomena, including future climate-driven changes. It is therefore essential that introducing  
 596 variable parameters takes into account such risks and ensures that reasonable assumptions are made to simplify  
 597 the parameter calibration task. These assumptions would ensure that model calibration is sufficiently constrained,  
 598 so that there are sufficient observations per each calibrated model parameter value. For example, only a carefully  
 599 selected subset of parameters may be calibrated, based on their relevance for primary production (established, for  
 600 example, through sensitivity analysis, e.g. Ciavatta *et al.* 2025) and lack of correlations with other model  
 601 parameters.

602 A key consideration when exploring variable parameters is the spatial and temporal scales at which they  
 603 might vary. For example, it would be important to establish whether seasonal climatological variability in  
 604 parameters alone can capture observed patterns, implying that both inter-annual and sub-seasonal variability could  
 605 be negligible. If so, this would relax the requirement on the volumes of observational data needed for the  
 606 calibration. Hypotheses about temporal variability scales for model parameters can be tested using long time-  
 607 series of measurements at specific stations, such as the Bermuda Atlantic Time-series Study and the Hawaii Ocean  
 608 Time-series, both of which present seasonal cycles in photosynthesis parameters (Kovač *et al.*, 2016b; Kovač *et al.*  
 609 *et al.*, 2018). Another key question is whether parameters vary over fine spatial scales or maintain coherence over  
 610 large scales, such as ocean biomes or Longhurst provinces (Longhurst 2007). Preliminary evidence suggests that,  
 611 at least for the global-scale applications, ecological provinces according to Longhurst might provide an  
 612 appropriate template for mapping parameters (see Figure 6B), and that monthly or seasonal time scales might be  
 613 appropriate for modelling variability in photosynthesis-irradiance parameters (Britten *et al.* 2025). If province-  
 614 based approaches emerge as viable candidates, it would be desirable to avoid sharp discontinuities in parameter  
 615 values at province boundaries, which might require incorporation of smoothing methods to make inter-province  
 616 changes seamless. Moreover, it is essential that model parameter calibration does not compensate for unrelated  
 617 spatio-temporally varying model biases, such as those arising from external forcings or other ecosystem model  
 618 constraints (e.g., boundary conditions). For example, given the importance of underlying physical processes,  
 619 caution should be applied when calibrating parameters in ecosystem models to reproduce the observed PP because  
 620 the model simulation could improve, but for the wrong reasons. Singh *et al.* (2025) illustrate that ecosystem  
 621 parameters in global ocean biogeochemical models are likely calibrated to compensate for biases in their physics  
 622 (see also Loptien and Dietze, 2019). To avoid mixing different sources of ecosystem model errors, parameters  
 623 should be ideally estimated jointly with the model state, e.g., using joint parameter-state data assimilation  
 624 techniques (Schartau *et al.* 2017). Finally, existing knowledge of acceptable ranges of parameter values needs to  
 625 be incorporated into the calibration process to prevent parameters from acquiring unrealistic values.

626 Since parameter spatio-temporal variability results from poorly resolved species types or ecosystem  
 627 processes, interesting insights into its scale and patterns can be also obtained by comparing models of different  
 628 complexity. For example, high complexity models (such as the DARWIN ecosystem model) could be used in



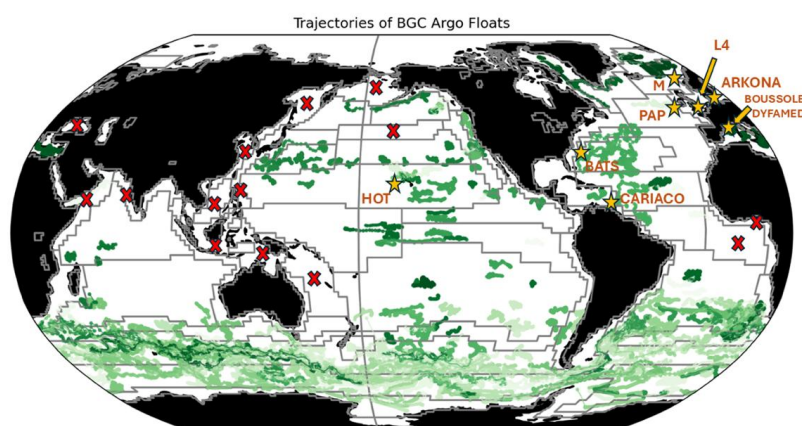
629 some cases to deduce parameter variability of simpler models, or even possibly help inform spatio-temporally  
 630 varying parameter calibration of those models. Comparison studies across models of different complexity would  
 631 be desirable in this case (for some examples see Friedrichs *et al.*, 2007, Xiao and Friedrichs, 2014).

632 It should be noted that even after successfully overcoming the challenges associated with spatio-temporal  
 633 parameter calibration, significant PP uncertainty is likely to remain in both historical estimates and future  
 634 projections. For satellite-based models, this uncertainty is mostly driven by inherent observational biases, e.g.,  
 635 gaps in data due to cloud cover or adverse viewing geometry, or inaccuracies in satellite products associated with  
 636 bio-optical conditions in water. For ecosystem models, additional sources of uncertainty include the forcing data  
 637 and the physical model driving biogeochemical processes, e.g., its vertical and horizontal resolution, and its ability  
 638 to represent currents and mixing responsible for nutrient supply and export of organic material. Further constraints  
 639 are inherent to ecosystem models themselves. Traditionally, plankton are divided into phototrophic phytoplankton  
 640 and phagotrophic zooplankton. However, recent research emphasises ubiquitous presence of mixotrophy in the  
 641 global ocean (Mitra *et al.* 2023), which not only differs in its physiology and ecological role, but can interact with  
 642 other types of plankton in complex ways (Flynn and Mitra 2023). Despite certain commonality in their approach  
 643 to modelling primary production, as discussed above, models differ significantly in their approaches to represent  
 644 various biogeochemical processes such as grazing and associated fluxes, deposition of organic matter and its  
 645 remineralisation. For many of those processes (e.g. zooplankton grazing), lack of data, variability and high  
 646 uncertainty of available data, are a major issue. Focusing on biological ocean carbon storage, Henson *et al.* (2024)  
 647 identified key areas where improved understanding of processes is required to support future modelling efforts.  
 648 For PP, the processes that were ranked highest were: resource limitation for growth, nitrogen fixation, zooplankton  
 649 processes and phytoplankton loss processes. Current ecosystem models differ considerably in their formulation  
 650 and parameterisation of these processes, contributing to uncertainties in model outcomes. Moreover, nitrogen-  
 651 fixation is often not included in these models. Even when these key processes are included, spatial parameter  
 652 estimation through assimilating observed state variables (such as water column nutrients and oxygen) in ocean  
 653 biogeochemical models does not necessarily lead to an improved estimate of primary production, suggesting that  
 654 current ecosystem model parameterizations may still be oversimplified compared to the real world (Singh *et al.*,  
 655 2025). The time is right to address the problem of parameter estimation in PP models, both for ecosystem models  
 656 and satellite-based models. Novel and rapidly expanding observations such as BGC Argo profiles, other types of  
 657 autonomous data collected by in-water vehicles and also large marine mammals (Chai *et al.* 2020; Claustre *et al.*  
 658 2020) have been providing large volumes of biological and bio-optical data that complements in situ data from  
 659 long time series stations and could be harnessed for this purpose (Figure 10). Complementary observations from  
 660 satellite remote sensing, now available over multiple decades and merged into climate-quality, consistent data  
 661 streams (e.g., Sathyendranath *et al.* 2019), is another rich data source, along with novel satellite products from  
 662 emerging capabilities such as geostationary, lidar, cubesat and hyperspectral data. When these are combined with  
 663 more traditional in situ platforms, including long-term gridded climatology from sources such as World Ocean  
 664 Atlas (WOA, e.g., Garcia *et al.*), there is already enough data to support a suitably-constrained spatio-temporally  
 665 varying parameter calibration. This opportunity is further enhanced by new advances in artificial intelligence and  
 666 machine learning, giving us an historically unprecedented capability to exploit these large and growing datasets  
 667 and address long-standing questions about marine PP.





668 However, crucial to this endeavour would be a clear focus on data quality, and on data validation, following  
 669 community-wide accepted protocols and reliable uncertainty characterization. Moreover, some regions, such as  
 670 sea-ice margins, coastal margins, and high latitudes in winter, which are often regions experiencing long-term  
 671 rapid changes and include some of the most productive areas of the global ocean, also tend to be regions that are  
 672 difficult to observe, and hence suffer from sparse data coverage. More observations are needed in such locations  
 673 to understand how model parameters might evolve in the future. Even if constrained spatio-temporally varying  
 674 calibration is possible in these regions with the available data-sets, the importance of further investing in data  
 675 quantity and quality cannot be overemphasized.



676  
 677 **Figure 10.** The global in situ data available for model calibration. The boundaries show ecological provinces  
 678 according to Longhurst (2007). BGC-Argo float trajectories are shown in shades of green, providing sufficiently  
 679 long time-series (since 2008) for calibration. Orange stars mark in situ time series stations with sufficiently long  
 680 time-series records that can also be used for model calibration. The red crosses mark provinces without sufficient  
 681 BGC-Argo data or in situ stations, where the models will need to rely solely on satellite records and compilations  
 682 of in situ observations, such as the World Ocean Atlas.

683

## 684 6. Conclusions

685 We have argued that, given the growing abundance of observations from diverse platforms, such as satellites and  
 686 BGC-Argo, combined with rapidly advancing capabilities in ensemble data assimilation techniques and artificial  
 687 intelligence, the time has now come to address explicitly the importance of parameter assignment in primary  
 688 production models, and in exploring the spatial and temporal variability in the parameters. We have theoretically  
 689 justified why such parameter variability is to be expected both in the satellite-based models (where some models  
 690 already employ variable parameters albeit in a simple fashion) and ecosystem models (where such assignment is  
 691 still quite rare), at least unless the models are of very high complexity. In the case of primary production, the  
 692 number of phytoplankton classes that are included in the model is a key differentiator of the model complexity.  
 693 Relatively simpler models, such as the ecosystem models used as part of ESMs in climate projections, have limited  
 694 capability to resolve phytoplankton communities. For such models, spatio-temporally varying parameters could  
 695 provide a means to account for the unresolved phytoplankton variability and processes.



696 Spatio-temporally variable parameter calibration can shed light on the sources of differences between lower-  
 697 medium complexity ecosystem models used in ESMs and satellite-based primary-production models. Since  
 698 variable parameters can capture, in a simple manner, processes or conditions that are not explicitly included in a  
 699 model, analysing the drivers of parameter variability could help identify how best to overcome current model  
 700 drawbacks. Furthermore, providing those models with spatio-temporally varying parameters could remove many  
 701 apparent differences between models, both potentially reducing the spatial and temporal biases in model parameter  
 702 calibration and enabling the simpler ecosystem models to represent better the effects of unresolved processes or  
 703 plankton classes. We argue that this could reduce the existing high uncertainty both in historical estimates and  
 704 future projections of marine primary production. Due to the importance of primary production for climate  
 705 research, improving its prediction can have a major impact on both climate mitigation and adaptation planning.

706 In the context of our climate, we need to understand how marine ecosystems in general, and phytoplankton  
 707 in particular, respond to change. Three types of changes need investigation: changes in (i) phytoplankton biomass  
 708 (whether they be measured as chlorophyll, carbon or nitrogen concentration, or all of them); (ii) the rates of  
 709 biological processes, with marine primary production being a key process in the global carbon cycle; and (iii)  
 710 community structure. The first of these can be addressed using fixed model parameters, but we would not know  
 711 if the effect of variable parameters could have important impacts, if it were not part of the investigation. The  
 712 second and third objectives both are intimately linked to parameter variability, with the third one in particular  
 713 calling for resolution of parameter variability at the level of major components of the phytoplankton community.

714 For many decades, we have relied on comparisons and analyses of (both satellite and ecosystem) model  
 715 outputs for insights into model performance, and for identifying the way forward. It is now time to shift the  
 716 emphasis toward understanding the behaviour of model parameters, across models, across multiple phytoplankton  
 717 types, and across multiple spatial and temporal scales. This focus has the potential to reduce uncertainties, unify  
 718 divergent model results, and provide a stronger foundation for predicting marine primary production under  
 719 changing climatic conditions.

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721 **Author contributions:** JS organized the writing of the manuscript with substantial input by SS, and all the  
 722 authors contributed ideas, text and Figures.

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 738 **References**



- 739 Amirian, M. M., Finkel, Z. V., Devred, E., Irwin, A. J. (accepted). Parametrization of Photoinhibition for  
 740 Phytoplankton. *Communications Earth and Environment*.  
 741
- 742 Anderson, S.I., A.D. Barton, S. Clayton, S. Dutkiewicz, and T. Rynearson, 2021. Marine phytoplankton functional  
 743 types exhibit diverse responses to thermal change. *Nature Communications*, doi:10.1038/s41467-021-26651  
 744
- 745 Antoine D, Andre J-M, Morel A (1996). Oceanic primary production: 2. Estimation at global scale from satellite  
 746 (Coastal Zone Color Scanner) chlorophyll. *Global Biogeochemical Cycles*, 10:57-69.  
 747 <https://doi.org/10.1029/95GB02832>  
 748
- 749 Arteaga, L.A., Behrenfeld, M.J., Boss, E. and Westberry, T.K., 2022. Vertical structure in phytoplankton growth  
 750 and productivity inferred from biogeochemical-Argo floats and the carbon-based productivity model. *Global*  
 751 *Biogeochemical Cycles*, 36(8), p.e2022GB007389.  
 752
- 753 Behrenfeld, M.J. and Falkowski, P.G. (1997) Photosynthetic Rates Derived from Satellite-based Chlorophyll  
 754 Concentration. *Limnology and Oceanography*, 42, 1-20. <https://doi.org/10.4319/lo.1997.42.1.0001>  
 755
- 756 Behrenfeld MJ, Boss E, Siegel D, and Shea DM (2005) Carbon-based ocean productivity and phytoplankton  
 757 physiology from space. *Global Biogeochemical Cycles* 19. <https://doi.org/10.1029/2004GB002299>.  
 758
- 759 Bergas-Masso, E., Hamilton, D.S., Myriokefalitakis, S., Rathod, S., Gonçalves Ageitos, M. and Pérez García-  
 760 Pando, C., 2025. Future climate-driven fires may boost ocean productivity in the iron-limited North  
 761 Atlantic. *Nature Climate Change*, pp.1-9.  
 762
- 763 Blackford, J. C., Allen, J. I., and Gilbert, F. J.: Ecosystem dynamics at six contrasting sites: a generic modelling  
 764 study, *J. Marine Syst.*, 52, 191–215, 2004  
 765
- 766 Bopp, L., Resplandy, L., Orr, J. C., Doney, S. C., Dunne, J. P., Gehlen, M., Halloran, P., Heinze, C., Ilyina, T.,  
 767 Séférian, R., Tjiputra, J., and Vichi, M.: Multiple stressors of ocean ecosystems in the 21st century: projections  
 768 with CMIP5 models, *Biogeosciences*, 10, 6225–6245, <https://doi.org/10.5194/bg-10-6225-2013>, 2013.  
 769
- 770 Bopp, L., Aumont, O., Kwiatkowski, L., Clerc, C., Dupont, L., Ethé, C., Gorgues, T., Séférian, R., and Tagliabue,  
 771 A.: Diazotrophy as a key driver of the response of marine net primary productivity to climate change,  
 772 *Biogeosciences*, 19, 4267–4285, <https://doi.org/10.5194/bg-19-4267-2022>, 2022  
 773
- 774 Bouman, H.A., Platt, T., Doblin, M., Figueiras, M.G., Gudmundsson, K., Gudfinnsson, H.G., Huang, B.,  
 775 Hickman, A., Hiscock, M., Jackson, T., Lutz, V.A., Mélin, F., Rey, F., Pepin, P., Segura, V., Tilstone, G.H., van  
 776 Dongen-Vogels, V., Sathyendranath, S. (2018) Photosynthesis–irradiance parameters of marine phytoplankton:  
 777 synthesis of a global data set. *Earth Syst. Sci. Data*, 10: 251–266. <https://doi.org/10.5194/essd-10-251-2018>  
 778



779 RJW Brewin, GH Tilstone, T Jackson, T Cain, PI Miller (2017) Modelling size-fractionated primary production  
 780 in the Atlantic Ocean from remote sensing. *Progress in Oceanography*. 158: 130-149, ISSN 0079-6611,  
 781 <https://doi.org/10.1016/j.pocean.2017.02.002>.  
 782  
 783 Brewin, R.J., Sathyendranath, S., Kulk, G., Rio, M.H., Concha, J.A., Bell, T.G., Bracher, A., Fichot, C., Frölicher,  
 784 T.L., Galí, M. and Hansell, D.A., 2023. Ocean carbon from space: Current status and priorities for the next  
 785 decade. *Earth-science reviews*, 240, p.104386.  
 786  
 787 Butenschön, M., Clark, J., Aldridge, J.N., Allen, J.I., Artioli, Y., Blackford, J., Bruggeman, J., Cazenave, P.,  
 788 Ciavatta, S., Kay, S., Lessin, G. *et al.*, 2016. ERSEM 15.06: a generic model for marine biogeochemistry and the  
 789 ecosystem dynamics of the lower trophic levels. *Geoscientific Model Development*, 9(4), pp.1293-1339.  
 790  
 791 Carr, M.E., Friedrichs, M.A., Schmeltz, M., Aita, M.N., Antoine, D., Arrigo, K.R., Asanuma, I., Aumont, O.,  
 792 Barber, R., Behrenfeld, M. and Bidigare, R., 2006. A comparison of global estimates of marine primary production  
 793 from ocean color. *Deep Sea Research Part II: Topical Studies in Oceanography*, 53(5-7), pp.741-770.  
 794  
 795 Chai, F., Johnson, K.S., Claustre, H., Xing, X., Wang, Y., Boss, E., Riser, S., Fennel, K., Schofield, O. and Sutton,  
 796 A., 2020. Monitoring ocean biogeochemistry with autonomous platforms. *Nature Reviews Earth &*  
 797 *Environment*, 1(6), pp.315-326.  
 798  
 799 Ciavatta, S., Lazzari, P., Álvarez, E., Bertino, L., Bolding, K., Bruggeman, J., Capet, A., Cossarini, G., Daryabor,  
 800 F., Nerger, L., Popov, M. *et al.*, 2025. Control of simulated ocean ecosystem indicators by biogeochemical  
 801 observations. *Progress in Oceanography*, 231, p.103384.  
 802  
 803 Claustre, H. and Johnson, KS, and Takeshita, Y (2020) Observing the Global Ocean with Biogeochemical-Argo.  
 804 *Annual Review of Marine Science*, 12: 23-48. <https://doi.org/10.1146/annurev-marine-010419-010956>  
 805  
 806 Daewel, U. and Schrum, C., 2013. Simulating long-term dynamics of the coupled North Sea and Baltic Sea  
 807 ecosystem with ECOSMO II: Model description and validation. *Journal of Marine Systems*, 119, pp.30-49.  
 808  
 809 Dai, R., Wen, Z., Hong, H., Browning, T.J., Hu, X., Chen, Z., Liu, X., Dai, M., Morel, F.M. and Shi, D., 2025.  
 810 Eukaryotic phytoplankton drive a decrease in primary production in response to elevated CO<sub>2</sub> in the tropical and  
 811 subtropical oceans. *Proceedings of the National Academy of Sciences*, 122(11), p.e2423680122.  
 812  
 813 Doléac, S., Lévy, M., El Hourany, R., and Bopp, L.: Toward more robust net primary production projections in  
 814 the North Atlantic Ocean, *Biogeosciences*, 22, 841–862, <https://doi.org/10.5194/bg-22-841-2025>, 2025.  
 815  
 816 Doney, S., Bopp, L., Long, M (2014) Historical and Future Trends in Ocean Climate and Biogeochemistry.  
 817 *Oceanography*. 27 (1), pp.108-119. [ff10.5670/oceanog.2014.14](https://doi.org/10.5670/oceanog.2014.14)ff. [ffhal-03211060f](https://doi.org/10.5670/oceanog.2014.14)  
 818



819 Doron M, Brasseur P, Brankart JM, Losa SN, Melet A. Stochastic estimation of biogeochemical parameters from  
 820 Globcolour ocean colour satellite data in a North Atlantic 3D ocean coupled physical–biogeochemical model.  
 821 Journal of Marine Systems. 2013 May 1;117:81-95.  
 822

823 Droop, M. R.: The nutrient status of alga cells in continuous culture, J. Mar. Biol. Assoc. UK, 54, 825–855,  
 824 doi:10.1016/0924-7963(94)00031-6, 1974  
 825

826 Dutkiewicz, S., J.R. Scott, and M.J. Follows, 2013, Winners and Losers: Ecological and Biogeochemical Changes  
 827 in a Warming Ocean. Global Biogeochemical Cycles, 27, 463–477, doi: 10.1002/gbc.20042  
 828

829 Dutkiewicz, S., Hickman, A.E., Jahn, O., Gregg, W.W., Mouw, C.B. and Follows, M.J., 2015. Capturing optically  
 830 important constituents and properties in a marine biogeochemical and ecosystem model. *Biogeosciences*, 12(14),  
 831 pp.4447–4481.  
 832

833 Dutkiewicz, S., Cermenio, P., Jahn, O., Follows, M.J., Hickman, A.E., Taniguchi, D.A. and Ward, B.A., 2020.  
 834 Dimensions of marine phytoplankton diversity. *Biogeosciences*, 17(3), pp.609–634.  
 835

836 Fasham, MJR, Ducklow, HW, McKelvie, SM (1990) A nitrogen-based model of plankton dynamics in the oceanic  
 837 mixed layer. Journal of Marine Research, 48: 591–639.  
 838

839 Fennel, K., Gehlen, M., Brasseur, P., Brown, C.W., Ciavatta, S., Cossarini, G., Crise, A., Edwards, C.A., Ford,  
 840 D., Friedrichs, M.A. and Gregoire, M., 2019. Advancing marine biogeochemical and ecosystem reanalyses and  
 841 forecasts as tools for monitoring and managing ecosystem health. *Frontiers in Marine Science*, 6, p.89.  
 842

843 Fennel, K., Mattern, J.P., Doney, S.C., Bopp, L., Moore, A.M., Wang, B. and Yu, L., 2022. Ocean biogeochemical  
 844 modelling. *Nature Reviews Methods Primers*, 2(1), p.76.  
 845

846 Field, CB, Behrenfeld, MJ, Randerson, JT, Falkowski, P (1998) Primary Production of the Biosphere: Integrating  
 847 terrestrial and oceanic Components. Science 281, 237–240. DOI: 10.1126/science.281.5374.237  
 848

849 Flynn, K.J. and Mitra, A., 2023. Feeding in mixoplankton enhances phototrophy increasing bloom-induced pH  
 850 changes with ocean acidification. *Journal of Plankton Research*, 45(4), pp.636–651.  
 851

852 Franks, P.J.S., 2002. NPZ models of plankton dynamics: their construction, coupling to physics, and application.  
 853 J. Oceanogr. 58, 379–387. DOI: 10.1023/a:1015874028196.  
 854

855 Friedlingstein, P., O'Sullivan, M., Jones, M. W., Andrew, R. M., Bakker, D. C. E., Hauck, J., Landschützer, P.,  
 856 Le Quéré, C., Luijkx, I. T., Peters, G. P., Peters, W., Pongratz, J., Schwingshackl, C., Sitch, S., Canadell, J. G.,  
 857 Ciais, P., Jackson, R. B., Alin, S. R., Anthoni, P., Barbero, L., Bates, N. R., Becker, M., Bellouin, N., Decharme,  
 858 B., Bopp, L., Brasika, I. B. M., Cadule, P., Chamberlain, M. A., Chandra, N., Chau, T.-T.-T., Chevallier, F., Chini,



- 859 L. P., Cronin, M., Dou, X., Enyo, K., Evans, W., Falk, S., Feely, R. A., Feng, L., Ford, D. J., Gasser, T., Ghattas,  
 860 J., Gkritzalis, T., Grassi, G., Gregor, L., Gruber, N., Gürses, Ö., Harris, I., Hefner, M., Heinke, J., Houghton, R.  
 861 A., Hurtt, G. C., Iida, Y., Ilyina, T., Jacobson, A. R., Jain, A., Jarníková, T., Jersild, A., Jiang, F., Jin, Z., Joos, F.,  
 862 Kato, E., Keeling, R. F., Kennedy, D., Klein Goldewijk, K., Knauer, J., Korsbakken, J. I., Körtzinger, A., Lan,  
 863 X., Lefèvre, N., Li, H., Liu, J., Liu, Z., Ma, L., Marland, G., Mayot, N., McGuire, P. C., McKinley, G. A., Meyer,  
 864 G., Morgan, E. J., Munro, D. R., Nakaoka, S.-I., Niwa, Y., O'Brien, K. M., Olsen, A., Omar, A. M., Ono, T.,  
 865 Paulsen, M., Pierrot, D., Pocock, K., Poulter, B., Powis, C. M., Rehder, G., Resplandy, L., Robertson, E.,  
 866 Rödenbeck, C., Rosan, T. M., Schwinger, J., Séférian, R., Smallman, T. L., Smith, S. M., Sospedra-Alfonso, R.,  
 867 Sun, Q., Sutton, A. J., Sweeney, C., Takao, S., Tans, P. P., Tian, H., Tilbrook, B., Tsujino, H., Tubiello, F., van  
 868 der Werf, G. R., van Ooijen, E., Wanninkhof, R., Watanabe, M., Wimart-Rousseau, C., Yang, D., Yang, X., Yuan,  
 869 W., Yue, X., Zaehle, S., Zeng, J., and Zheng, B.: Global Carbon Budget 2023, *Earth Syst. Sci. Data*, 15, 5301–  
 870 5369, <https://doi.org/10.5194/essd-15-5301-2023>, 2023.
- 871
- 872 Garcia, H.E., Weathers, K.W., Paver, C.R., Smolyar, I., Boyer, T.P., Locarnini, M.M., Zweng, M.M., Mishonov,  
 873 A.V., Baranova, O.K. and Seidov, D., 2019. World ocean atlas 2018. Vol. 4: Dissolved inorganic nutrients  
 874 (phosphate, nitrate and nitrate+ nitrite, silicate).
- 875
- 876 Gharamti ME, Samuelsen A, Bertino L, Simon E, Korosov A, Daewel U. Online tuning of ocean biogeochemical  
 877 model parameters using ensemble estimation techniques: Application to a one-dimensional model in the North  
 878 Atlantic. *Journal of Marine Systems*. 2017 Apr 1;168:1-6.
- 879
- 880 Gharamti ME, Tjiputra J, Bethke I, Samuelsen A, Skjelvan I, Bentsen M, Bertino L. Ensemble data assimilation  
 881 for ocean biogeochemical state and parameter estimation at different sites. *Ocean Modelling*. 2017 Apr 1;112:65-  
 882 89.
- 883
- 884 Gregg, W.W. and Rousseaux, C.S., 2016. Directional and spectral irradiance in ocean models: Effects on  
 885 simulated global phytoplankton, nutrients, and primary production. *Frontiers in Marine Science*, 3, p.240.
- 886
- 887 Grégoire, M. and Soetaert, K., 2010. Carbon, nitrogen, oxygen and sulfide budgets in the Black Sea: A  
 888 biogeochemical model of the whole water column coupling the oxic and anoxic parts. *Ecological Modelling*,  
 889 221(19), pp.2287-2301.
- 890
- 891 Eppley, R., 1971. Temperature and phytoplankton growth in the sea. *Fishery bulletin*, 70(4), p.1063
- 892
- 893 Friedlingstein, P., O'sullivan, M., Jones, M.W., Andrew, R.M., Hauck, J., Landschützer, P., Le Quéré, C., Li, H.,  
 894 Luijkx, I.T., Olsen, A. and Peters, G.P., 2024. Global carbon budget 2024. *Earth System Science Data*  
 895 *Discussions*, 2024, pp.1-133.
- 896
- 897 Friedrichs, M.A., Dusenberry, J.A., Anderson, L.A., Armstrong, R.A., Chai, F., Christian, J.R., Doney, S.C.,  
 898 Dunne, J., Fujii, M., Hood, R. and McGillicuddy Jr, D.J., 2007. Assessment of skill and portability in regional



899 marine biogeochemical models: Role of multiple planktonic groups. *Journal of Geophysical Research: Oceans*,  
 900 112(C8).  
 901  
 902 Friedrichs, M.A.M., Carr, M.-E., Barber, R.T., Scardi, M., Antoine, D., Armstrong, R. A., *et al.* (2009) Assessing  
 903 the uncertainties of model estimates of primary productivity in the tropical Pacific Ocean. *J. Mar. Syst.* 76, 113-  
 904 133. doi:10.1016/j.jmarsys.2008.05.010  
 905  
 906 Frölicher, T. L., K. B. Rodgers, C. A. Stock, and W. W. L. Cheung (2016), Sources of uncertainties in 21st century  
 907 projections of potential ocean ecosystem stressors, *Global Biogeochem. Cycles*, 30, 1224–1243,  
 908 doi:10.1002/2015GB005338.  
 909  
 910 Galli, G., Wakelin, S., Harle, J., Holt, J., Artioli, Y., 2024. Multi-model comparison of trends and controls of near-  
 911 bed oxygen concentration on the northwest European continental shelf under climate change. *Biogeosciences* 21,  
 912 2143–2158. <https://doi.org/10.5194/bg-21-2143-2024>  
 913  
 914 Gastineau, G., & Soden, B. J. (2009). Model projected changes of extreme wind events in response to global  
 915 warming. *Geophysical Research Letters*, 36(10). <https://doi.org/10.1029/2009GL037500>  
 916  
 917 Geider, RJ, Macintyre, HL, and Kana, TM (1997) Dynamic model of phytoplankton growth and acclimation:  
 918 responses of the balanced growth rate and the chlorophyll a: carbon ratio to light, nutrient imitation and  
 919 temperature,” *Mar. Ecol. Prog. Ser.* 148, 187–200.  
 920  
 921 Geider RJ, MacIntyre HL, Kana TM. (1998) A dynamic regulatory model of phytoplankton acclimation to light,  
 922 nutrients, and temperatures. *Limnol. Oceanogr.*, 43(4), 679-694.  
 923  
 924 Gentile, E. S., Zhao, M., & Hodges, K. (2023). Poleward intensification of midlatitude extreme winds under  
 925 warmer climate. *Npj Climate and Atmospheric Science*, 6(1), 1–10. <https://doi.org/10.1038/s41612-023-00540-x>  
 926  
 927 Gentleman, W., 2002. A chronology of plankton dynamics in silico: how computer models have been used to  
 928 study marine ecosystems. *Hydrobiologia* 480, 69–85. DOI: 10.1023/A:1021289119442.  
 929  
 930 Gregg W.W and Rousseaux C.S. 2019. Global ocean primary production trends in the modern ocean color satellite  
 931 record (1998–2015). *Environ. Res. Lett.* 14 124011  
 932  
 933 Grégoire, M., Raick, C. and Soetaert, K., 2008. Numerical modeling of the central Black Sea ecosystem  
 934 functioning during the eutrophication phase. *Progress in Oceanography*, 76(3), pp.286-333.  
 935  
 936 Grégoire, M. and Soetaert, K., 2010. Carbon, nitrogen, oxygen and sulfide budgets in the Black Sea: A  
 937 biogeochemical model of the whole water column coupling the oxic and anoxic parts. *Ecological Modelling*,  
 938 221(19), pp.2287-2301.





- 939
- 940 Gulev, S.K., P.W. Thorne, J. Ahn, F.J. Dentener, C.M. Domingues, S. Gerland, D. Gong, D.S. Kaufman, H.C.  
 941 Nnamchi, J. Quaas, J.A. Rivera, S. Sathyendranath, S.L. Smith, B. Trewin, K. von Schuckmann, and R.S. Vose,  
 942 2021: Changing State of the Climate System. In *Climate Change 2021: The Physical Science Basis. Contribution*  
 943 *of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change* [Masson-  
 944 Delmotte, V., P. Zhai, A. Pirani, S.L. Connors, C. Péan, S. Berger, N. Caud, Y. Chen, L. Goldfarb, M.I. Gomis,  
 945 M. Huang, K. Leitzell, E. Lonnoy, J.B.R. Matthews, T.K. Maycock, T. Waterfield, O. Yelekçi, R. Yu, and B.  
 946 Zhou (eds.)]. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, pp. 287–422,  
 947 doi: 10.1017/9781009157896.004.
- 948
- 949 Halsey, K.H., Milligan, A.J. and Behrenfeld, M.J., 2011. Linking time-dependent carbon-fixation efficiencies in  
 950 *Dunaliella Tertiolecta* (Chlorophyceae) to underlying metabolic pathways 1. *Journal of Phycology*, 47(1), pp.66-  
 951 76.
- 952
- 953 Henson, S., Baker, C.A., Halloran, P., McQuatters-Gollop, A., Painter, S., Planchat, A. and Tagliabue, A., 2024.  
 954 Knowledge gaps in quantifying the climate change response of biological storage of carbon in the ocean. *Earth's*  
 955 *Future*, 12(6), p.e2023EF004375.
- 956
- 957 Hewitt, C. D., and Coauthors, 2021: Recommendations for Future Research Priorities for Climate Modeling and  
 958 Climate Services. *Bull. Amer. Meteor. Soc.*, 102, E578–E588, <https://doi.org/10.1175/BAMS-D-20-0103.1>.
- 959
- 960 IOCCG (2020). Synergy between Ocean Colour and Biogeochemical/Ecosystem Models. Dutkiewicz, S. (ed.),  
 961 IOCCG Report Series, No. 19, International Ocean Colour Coordinating Group, Dartmouth, Canada.  
 962 <http://dx.doi.org/10.25607/OBP-711>
- 963
- 964 IOCCG Protocol Series (2022). Aquatic Primary Productivity Field Protocols for Satellite Validation and Model  
 965 Synthesis. Balch, W.M., Carranza, M., Cetinić, I., Chaves, J.E., Duhamel, S., Fassbender, A., Fernandez-Carrera,  
 966 A., Ferrón, S., García-Martín, E., Goes, J., Gomes, H., Gundersen, K., Halsey, K., Hirawake, T., Isada, T., Juranek,  
 967 L., Kulk, G., Langdon, C., Letelier, R., López-Sandoval, D., Mannino, A., Marra, J.F., Neale, P., Nicholson, D.,  
 968 Silsbe, G., Stanley, R.H., Vandermeulen, R.A. IOCCG Ocean Optics and Biogeochemistry Protocols for Satellite  
 969 Ocean Colour Sensor Validation, Volume 7.0, edited by R.A. Vandermeulen, J. E. Chaves, IOCCG, Dartmouth,  
 970 NS, Canada. doi:<http://dx.doi.org/10.25607/OBP-1835>
- 971
- 972 IPCC (2019): IPCC Special Report on the Ocean and Cryosphere in a Changing Climate [H.-O. Pörtner, D.C.  
 973 Roberts, V. Masson-Delmotte, P. Zhai, M. Tignor, E. Poloczanska, K. Mintenbeck, A. Alegria, M. Nicolai, A.  
 974 Okem, J. Petzold, B. Rama, N.M. Weyer (eds.)].
- 975
- 976 IPCC, 2021: Climate Change 2021 - the Physical Science Basis, Contribution of Working Group I to the Sixth  
 977 Assessment Report of the Intergovernmental Panel on Climate Change [Masson-Delmotte, V., P. Zhai, A. Pirani,  
 978 S.L. Connors, C. Péan, S. Berger, N. Caud, Y. Chen, L. Goldfarb, M.I. Gomis, M. Huang, K. Leitzell, E. Lonnoy,



979 J.B.R. Matthews, T.K. Maycock, T. Waterfield, O. Yelekçi, R. Yu, and B. Zhou (eds.)). Cambridge University  
 980 Press, In Press, Published: 9 August 2021.  
 981  
 982 Jackson, T, Sathyendranath, S, and T. Platt, T (2017) An exact solution for modeling photoacclimation of the  
 983 carbon-to-chlorophyll ratio in phytoplankton, *Front. Mar. Sci.* 4, 283  
 984  
 985 Jassby, A.D. and Platt, T., 1976. Mathematical formulation of the relationship between photosynthesis and light  
 986 for phytoplankton. *Limnology and oceanography*, 21(4), pp.540-547.  
 987  
 988 Jin, P., Hutchins, D.A. and Gao, K., 2020. The impacts of ocean acidification on marine food quality and its  
 989 potential food chain consequences. *Frontiers in Marine Science*, 7, p.543979.  
 990  
 991 Jones, C. G., Adloff, F., Booth, B., Cox, P., Eyring, V., Friedlingstein, P., Frieler, K., Hewitt, H., Jeffery, H.,  
 992 Joussaume, S., Koenigk, T., Lawrence, B. N., O'Rourke, E., Roberts, M., Sanderson, B., Séférian, R., Somot, S.,  
 993 Vidale, P.-L., van Vuuren, D., Acosta, M., Bentsen, M., Bernardello, R., Betts, R., Blockley, E., Boé, J.,  
 994 Bracegirdle, T., Braconnot, P., Brovkin, V., Buontempo, C., Doblus-Reyes, F. J., Donat, M. G., Epicoco, I.,  
 995 Falloon, P., Fiore, S., Froelicher, T., Fuckar, N., Gidden, M., Goessling, H., Graversen, R. G., Gualdi, S.,  
 996 Gutiérrez, J. M., Ilyina, T., Jacob, D., Jones, C., Juckes, M., Kendon, E., Kjellström, E., Knutti, R., Lowe, J. A.,  
 997 Mizielinski, M., Nassisi, P., Obersteiner, M., Regnier, P., Roehrig, R., Salas y Melia, D., Schleussner, C.-F.,  
 998 Schulz, M., Scoccimarro, E., Terray, L., Thiemann, H., Wood, R., Yang, S., and Zaehle, S.: Bringing it all  
 999 together: Science and modelling priorities to support international climate policy, EGU sphere [preprint],  
 1000 <https://doi.org/10.5194/egusphere-2024-453>, 2024.  
 1001  
 1002 Kiefer, D.A. and Mitchell, B.G., 1983. A simple, steady state description of phytoplankton growth based on  
 1003 absorption cross section and quantum efficiency 1. *Limnology and Oceanography*, 28(4), pp.770-776.  
 1004  
 1005 Kim, H.H., Laufkötter, C., Lovato, T., Doney, S.C. and Ducklow, H.W., 2023. Projected 21st-century changes in  
 1006 marine heterotrophic bacteria under climate change. *Frontiers in microbiology*, 14, p.1049579.  
 1007  
 1008 Kishi, M.J., Kashiwai, M., Ware, D.M., Megrey, B.A., Eslinger, D.L., Werner, F.E., Noguchi-Aita, M., Azumaya,  
 1009 T., Fujii, M., Hashimoto, S. and Huang, D., 2007. NEMURO—a lower trophic level model for the North Pacific  
 1010 marine ecosystem. *Ecological Modelling*, 202(1-2), pp.12-25.  
 1011  
 1012 Kovač, Ž., Platt, T., Sathyendranath, S., Morović, M., Jackson, T. (2016a). Recovery of photosynthesis parameters  
 1013 from in situ profiles of phytoplankton production. *ICES Journal of Marine Science*, 73 (2), 275–285. DOI:  
 1014 10.1093/icesjms/fsv204.  
 1015  
 1016 Kovač, Ž., Platt, T., Sathyendranath, S., Morović, M. (2016b). Analytical solution for the vertical profile of daily  
 1017 production in the ocean. *Journal of Geophysical Research: Oceans*, 121. DOI: 10.1002/2015JC011293.  
 1018



- 1019 Kovač, Ž., Platt, T., S., S., Antunović, S. (2017). Models for estimating photosynthesis parameters from in situ  
 1020 production profiles. *Progress in Oceanography*, 159, 255-266. doi:10.1016/j.pocean.2017.10.013.  
 1021
- 1022 Kovač, Ž., Platt, T., Sathyendranath, S., Lomas, M. W. (2018). Extraction of photosynthesis parameters from time  
 1023 series measurements of in situ production: Bermuda atlantic time-series study. *Remote Sensing*, 10, 915. DOI:  
 1024 10.3390/rs10060915.  
 1025
- 1026 Kovárová-Kovar, K. and Egli, T., 1998. Growth kinetics of suspended microbial cells: from single-substrate-  
 1027 controlled growth to mixed-substrate kinetics. *Microbiology and molecular biology reviews*, 62(3), pp.646-666.  
 1028
- 1029 Krinos, A.I., Shapiro, S.K., Li, W., Haley, S.T., Dyhrman, S.T., Dutkiewicz, S., Follows, M.J. and Alexander, H.  
 1030 (2025), Intraspecific Diversity in Thermal Performance Determines Phytoplankton Ecological Niche. *Ecology*  
 1031 *Letters*, 28: e70055. <https://doi.org/10.1111/ele.70055>  
 1032
- 1033 Kulk, G, Platt, T, Dingle, J, Jackson, T, Jönsson, BF, Bouman, HA, Babin, M, Brewin, RJW, Doblin, M, Estrada,  
 1034 M, Figueiras, FG, Furuya, K, González-Benítez, N, Gudfinnsson, HG, Gudmundsson, K, Huang, B, Isada, T,  
 1035 Kovač, Ž, Lutz, VA, Marañón, E, Raman, M, Richardson, K, Rozema, PD, Poll, WH, Segura, V, Tilstone, GH,  
 1036 Uitz, J, Dongen-Vogels, V, Yoshikawa, T, Sathyendranath, S (2020) Primary Production, an Index of Climate  
 1037 Change in the Ocean: Satellite-Based Estimates over Two Decades. *Remote Sensing*, 12, 826.  
 1038 <https://doi.org/10.3390/rs12050826>  
 1039
- 1040 Kulk, G.; Platt, T.; Dingle, J.; Jackson, T.; Jönsson, B.F.; Bouman, H.A.; Babin, M.; Brewin, R.J.W.; Doblin, M.;  
 1041 Estrada, M.; *et al.* Correction: Kulk *et al.* Primary Production, an Index of Climate Change in the Ocean: Satellite-  
 1042 Based Estimates over Two Decades. *Remote Sens.* 2020, 12, 826. *Remote Sens.* 2021, 13, 3462.  
 1043 <https://doi.org/10.3390/rs13173462>  
 1044
- 1045 Kyewalyanga, M., Platt, T., & Sathyendranath, S. (1992). Ocean primary production calculated by spectral and  
 1046 broadband models. *Marine Ecology Progress Series*, 85, 171–185. DOI: 10.3354/meps085171.  
 1047
- 1048 Kyewalyanga, MN, Platt, T, Sathyendranath, S (1997) Estimation of the photosynthetic action spectrum:  
 1049 implications for primary production models. *Mar. Ecol. Prog. Ser.* 146: 207-223.  
 1050
- 1051 Kwiatkowski, L., Bopp, L., Aumont, O., Ciais, P., Cox, P.M., Laufkötter, C., Li, Y. and Séférian, R., 2017.  
 1052 Emergent constraints on projections of declining primary production in the tropical oceans. *Nature Climate*  
 1053 *Change*, 7(5), pp.355-358.  
 1054
- 1055 Kwiatkowski, L., Torres, O., Bopp, L., Aumont, O., Chamberlain, M., Christian, J.R., Dunne, J.P., Gehlen, M.,  
 1056 Ilyina, T., John, J.G., Lenton, A., Li, H., Lovenduski, N.S., Orr, J.C., Palmieri, J., Santana-Falcón, Y., Schwinger,  
 1057 J., Séférian, R., Stock, C.A., Tagliabue, A., Takano, Y., Tjiputra, J., Toyama, K., Tsujino, H., Watanabe, M.,  
 1058 Yamamoto, A., Yool, A., Ziehn, T., 2020. Twenty-first century ocean warming, acidification, deoxygenation, and



upper-ocean nutrient and primary production decline from CMIP6 model projections. *Biogeosciences* 17, 3439–3470. <https://doi.org/10.5194/bg-17-3439-2020>

Laufkötter, C., Vogt, M., Gruber, N., Aita-Noguchi, M., Aumont, O., Bopp, L., Buitenhuis, E., Doney, S. C., Dunne, J., Hashioka, T., Hauck, J., Hirata, T., John, J., Le Quéré, C., Lima, I. D., Nakano, H., Seferian, R., Totterdell, I., Vichi, M., and Völker, C. (2015) Drivers and uncertainties of future global marine primary production in marine ecosystem models, *Biogeosciences*, 12, 6955–6984, <https://doi.org/10.5194/bg-12-6955-2015>

Lee, Z. Veronica P. Lance, VP, Shang, S, Vaillancourt, R, Freeman, S, Lubac, B, Hargreaves, BR, Del Castillo, C, Richard Miller, R, Twardowski, M, Wei, G (2011) An assessment of optical properties and primary production derived from remote sensing in the Southern Ocean (SO GasEx). *J. Geophys. Res.* 116, C00F03 (2011). doi:10.1029/2010JC006747.

Lee Z, Marra J, Perry MJ, Kahru M (2015). Estimating oceanic primary productivity from ocean color remote sensing: A strategic assessment, *Journal of Marine Systems*. <http://dx.doi.org/10.1016/j.jmarsys.2014.11.015>

S. and Yoo, S. (2016) ‘Interannual variability of the phytoplankton community by the changes in vertical mixing and atmospheric deposition in the Ulleung Basin, East Sea: A modelling study’, *Ecological Modelling*, 322, pp. 31–47. Available at: <https://doi.org/10.1016/j.ecolmodel.2015.11.012>.

Lee, Y. J., P. A. Matrai, M. A. M. Friedrichs, V. S. Saba, D. Antoine, M. Ardyna, I. Asanuma, M. Babin, S. Belanger, M. Benoit-Gagne, E. Devred, M. Fernandez-Mendez, B. Gentili, T. Hirawake, S.-H. Kang, T. Kameda, C. Katlein, S.H. Lee, Z. Lee, F. Melin, M. Scardi, T.J. Smyth, S. Tang, K.R. Turpie, K.J. Waters, and T.K. Westberry (2015). An assessment of ocean color model estimates of primary productivity in the Arctic Ocean. *J. Geophys. Res. Oceans*, FAMOS SI, 120:6508–6541, DOI 10.1002/2015JC011018.

Liu, H., Li, D., Chen, Q., Feng, J., Qi, J., & Yin, B. (2024). The multiscale variability of global extreme wind and wave events and their relationships with climate modes. *Ocean Engineering*, 307, 118239. <https://doi.org/10.1016/j.oceaneng.2024.118239>

Longhurst, A, Sathyendranath, S, Platt, T, Caverhill, C (1995) An estimate of global primary production in the ocean from satellite radiometer data. *J. Plankton Res.* 17: 1245-1271.

Longhurst, A.R. *Ecological Geography of the Sea*, 2nd ed.; Elsevier Academic Press: Cambridge, MA, USA, 2007; p. 542.

Löptien, U. and Dietze, H., 2019. Reciprocal bias compensation and ensuing uncertainties in model-based climate projections: pelagic biogeochemistry versus ocean mixing. *Biogeosciences*, 16(9), pp.1865-1881.



- 1098 Lurin, B., Rasool, S.I., Cramer, W. and Moore, B. (1994) Global terrestrial net primary production. *Glob. Change*  
 1099 *NewsL (IGBP)*, 19, 6-8.  
 1100
- 1101 Luypaert, T., Hagan, J.G., McCarthy, M.L. and Poti, M., 2020. Status of marine biodiversity in the  
 1102 Anthropocene. *YOUMARES*, 9, pp.57-82.  
 1103
- 1104 Maishal, S., 2024. Decadal changes in global Oceanic Primary Productivity and its drivers. *Ocean-Land-*  
 1105 *Atmosphere Research*, 3, p.0066.  
 1106
- 1107 Marshak, A.R., Link, J.S. Primary production ultimately limits fisheries economic performance. *Sci Rep* 11,  
 1108 12154 (2021). <https://doi.org/10.1038/s41598-021-91599-0>  
 1109
- 1110 Michaelis L, Menten ML. Die kinetik der invertinwirkung. *Biochem. z.* 1913 Feb;49(333-369):352.  
 1111
- 1112 Mitra, A., Caron, D.A., Faure, E., Flynn, K.J., Leles, S.G., Hansen, P.J., McManus, G.B., Not, F., do Rosario  
 1113 Gomes, H., Santoferrara, L.F. and Stoecker, D.K., 2023. The Mixoplankton Database (MDB): Diversity of photo-  
 1114 phago-trophic plankton in form, function, and distribution across the global ocean. *Journal of Eukaryotic*  
 1115 *Microbiology*, 70(4), p.e12972.  
 1116
- 1117 Myksvoll, M.S., Sandø, A.B., Tjiputra, J., Samuelsen, A., Yumruktepe, V.Ç., Li, C., Mousing, E.A., Bettencourt,  
 1118 J.P. and Ottersen, G., 2023. Key physical processes and their model representation for projecting climate impacts  
 1119 on subarctic Atlantic net primary production: A synthesis. *Progress in Oceanography*, 217, p.103084.  
 1120
- 1121 Norberg J., Biodiversity and ecosystem functioning: A complex adaptive systems approach, *Limnology and*  
 1122 *Oceanography*, 4, part 2, doi: 10.4319/lo.2004.49.4\_part\_2.1269. 2004.  
 1123
- 1124 T. Parsons, M. Takahashi, B. Hargrave., *Biological Oceanographic Processes* (Third edition), Pergamon  
 1125 International Library of Science, Technology, Engin, Pergamon (1984)  
 1126
- 1127 Pastres, R., Ciavatta, S. and Solidoro, C., 2003. The Extended Kalman Filter (EKF) as a tool for the assimilation  
 1128 of high frequency water quality data. *Ecological modelling*, 170(2-3), pp.227-235.  
 1129
- 1130 Platt, T., Gallegos, C.L. and Harrison, W.G., 1980. Photoinhibition of photosynthesis in natural assemblages of  
 1131 marine phytoplankton.  
 1132
- 1133 Platt, T., Sathyendranath, S (1988) Oceanic primary production: Estimation by remote sensing at local and  
 1134 regional scales. *Science* 241: 1613-1620.  
 1135
- 1136 Platt, T., Sathyendranath, S, Ravindran, P (1990) Primary production by phytoplankton: analytic solutions for  
 1137 daily rates per unit area of water surface. *Proc. R. Soc. Lond. Ser. B* 241: 101-111.



- 1138  
 1139 Platt, T, Sathyendranath, S (1991) Biological production models as elements of coupled, atmosphere-ocean  
 1140 models for climate research. *J. Geophys. Res.* 96: 2585-2592.  
 1141  
 1142 Platt, T, Sathyendranath, S (1993) Estimators of primary production for interpretation of remotely sensed data on  
 1143 ocean color. *J. Geophys. Res.* 98: 14,561-14,576.  
 1144 Platt, T, Sathyendranath, S (1997) Modelling primary production IV (in Japanese). *Aquabiology* 19: 229-232.  
 1145  
 1146 Radtke, H., Lipka, M., Bunke, D., Morys, C., Woelfel, J., Cahill, B., Böttcher, M.E., Forster, S., Leipe, T., Rehder,  
 1147 G. and Neumann, T., 2019. Ecological ReGional Ocean Model with vertically resolved sediments (ERGOM SED  
 1148 1.0): coupling benthic and pelagic biogeochemistry of the south-western Baltic Sea. *Geoscientific Model*  
 1149 *Development*, 12(1), pp.275-320.  
 1150  
 1151 Ratnarajah L, Abu-Alhaija R, Atkinson A, Batten S, Bax NJ, Bernard KS, Canonico G, Cornils A, Everett JD,  
 1152 Grigoratou M, Ishak NH. Monitoring and modelling marine zooplankton in a changing climate. *Nature*  
 1153 *Communications*. 2023 Feb 2;14(1):564.  
 1154  
 1155 Regaudie-de-Gioux, A., Lasternas, S., Agustí, S., Duarte, C.M. 2019. Comparing marine primary production  
 1156 estimates through different methods and development of conversion equations. *Frontiers in Marine Science*, 1.  
 1157 URL: <https://www.frontiersin.org/journals/marine-science/articles/10.3389/fmars.2014.00019>.  
 1158 DOI=10.3389/fmars.2014.00019  
 1159  
 1160 Rohr T, Richardson AJ, Lenton A, Chamberlain MA, Shadwick EH. Zooplankton grazing is the largest source of  
 1161 uncertainty for marine carbon cycling in CMIP6 models. *Communications Earth & Environment*. 2023 Jun  
 1162 14;4(1):212.  
 1163  
 1164 Roy S, Broomhead DS, Platt T, Sathyendranath S, Ciavatta S. Sequential variations of phytoplankton growth and  
 1165 mortality in an NPZ model: A remote-sensing-based assessment. *Journal of Marine Systems*. 2012 Apr  
 1166 1;92(1):16-29.  
 1167  
 1168 Ryan-Keogh, T.J., Tagliabue, A. & Thomalla, S.J. Global decline in net primary production underestimated by  
 1169 climate models. *Commun Earth Environ* 6, 75 (2025). <https://doi.org/10.1038/s43247-025-02051-4>  
 1170  
 1171 Saba, V.S., Friedrichs, M.A.M., Carr, M.-E., Antoine, D., Armstrong, R.A., Asanuma, I., *et al.* (2010) Challenges  
 1172 of modeling depth-integrated marine primary productivity over multiple decades: a case study at BATS and HOT.  
 1173 *Glob. Biogeochem. Cycle* 24, GB3020.doi:10.1029/2009GB003655  
 1174  
 1175 Sathyendranath, S., & Platt, T. (1989a). Computation of aquatic primary production: Extended formalism to  
 1176 include the effect of angular and spectral distribution of light. *Limnology and Oceanography*, 34, 188–198. DOI:  
 1177 10.4319/lo.1989.34.1.0188.



1178  
 1179 Sathyendranath, S, Platt, T, Caverhill, CM, Warnock, RE, Lewis, MR (1989b) Remote sensing of oceanic primary  
 1180 production: Computations using a spectral model. *Deep-Sea Res. I* 36: 431-453.  
 1181  
 1182 Sathyendranath, S, Platt, T (2007) Spectral effects in bio-optical control on the ocean system. *Oceanologia* 49: 5-  
 1183 39.  
 1184  
 1185 Sathyendranath, S, Stuart, V, Nair, A, Oka, K., Nakane, T, Bouman, H, Forget, M-H, Maass, H, Platt, T (2009)  
 1186 Carbon-to-chlorophyll ratio and growth rate of phytoplankton in the sea. *Mar. Ecol. Prog. Ser.* 383: 73–84, doi:  
 1187 10.3354/meps07998  
 1188  
 1189 Sathyendranath, S., Brewin, R., Brockmann, C., Brotas, V., Calton, B., Chuprin, A., *et al.* (2019). An ocean-  
 1190 colour time series for use in climate studies: The experience of the Ocean-Colour Climate Change Initiative (OC-  
 1191 CCI). *Sensors*, 19(19), 4285. <https://doi.org/10.3390/s19194285>  
 1192  
 1193 Sathyendranath, S, Platt, T, Kovač, Ž, Dingle, J, Jackson, T, Brewin, R JW, Franks, P, Marañón, E, Kulk, G, and  
 1194 Bouman, HA (2020) Reconciling models of primary production and photoacclimation [Invited]. *Applied Optics*,  
 1195 59: C100-C114. <https://doi.org/10.1364/AO.386252>  
 1196  
 1197 Sauterey B., Le Gland G., Cermeño P., Aumont O., Lévy M., Vallina S.M., Phytoplankton adaptive resilience to  
 1198 climate change collapses in case of extreme events – A modeling study. *Ecological Modelling*, Volume 483, 2023,  
 1199 110437, ISSN 0304-3800, <https://doi.org/10.1016/j.ecolmodel.2023.110437>  
 1200  
 1201 Séférian, R., Berthet, S., Yool, A., Palmieri, J., Bopp, L., Tagliabue, A., Kwiatkowski, L., Aumont, O., Christian,  
 1202 J., Dunne, J., Gehlen, M., Ilyina, T., John, J.G., Li, H., Long, M.C., Luo, J.Y., Nakano, H., Romanou, A.,  
 1203 Schwinger, J., Stock, C., Santana-Falcón, Y., Takano, Y., Tjiputra, J., Tsujino, H., Watanabe, M., Wu, T., Wu,  
 1204 F., Yamamoto, A., 2020. Tracking Improvement in Simulated Marine Biogeochemistry Between CMIP5 and  
 1205 CMIP6. *Curr Clim Change Rep* 6, 95–119. <https://doi.org/10.1007/s40641-020-00160-0>  
 1206  
 1207 Schartau, M., Wallhead, P., Hemmings, J., Löptien, U., Kriest, I., Krishna, S., Ward, B. A., Slawig, T., and  
 1208 Oschlies, A.: Reviews and syntheses: parameter identification in marine planktonic ecosystem modelling,  
 1209 *Biogeosciences*, 14, 1647–1701, <https://doi.org/10.5194/bg-14-1647-2017>, 2017.  
 1210  
 1211 Schmidt S., Stramma L., Visbeck M. (2017). Decline in global oceanic oxygen content during the past five  
 1212 decades. *Nature* 542, 335–339. doi: 10.1038/nature21399  
 1213  
 1214 Shigemitsu, M., Okunishi, T., Nishioka, J., Sumata, H., Hashioka, T., Aita, M.N., Smith, S.L., Yoshie, N., Okada,  
 1215 N. and Yamanaka, Y., 2012. Development of a one-dimensional ecosystem model including the iron cycle applied  
 1216 to the Oyashio region, western subarctic Pacific. *Journal of Geophysical Research: Oceans*, 117(C6).  
 1217





- 1218 Silsbe, G.M., Behrenfeld, M.J., Halsey, K.H., Milligan, A.J. and Westberry, T.K., 2016. The CAFE model: A net  
 1219 production model for global ocean phytoplankton. *Global Biogeochemical Cycles*, 30(12), pp.1756-1777.  
 1220
- 1221 Silsbe, G.M., Fox, J., Westberry, T.K. *et al.* Global declines in net primary production in the ocean color era. *Nat*  
 1222 *Commun* 16, 5821 (2025). <https://doi.org/10.1038/s41467-025-60906-y>  
 1223
- 1224 Simon E, Samuelsen A, Bertino L, Mouysset S. Experiences in multiyear combined state–parameter estimation  
 1225 with an ecosystem model of the North Atlantic and Arctic Oceans using the Ensemble Kalman Filter. *Journal of*  
 1226 *Marine Systems*. 2015 Dec 1;152:1-7.  
 1227
- 1228 Singh, T., Counillon, F., Tjiputra, J. and Wang, Y., 2025. A novel ensemble-based parameter estimation for  
 1229 improving ocean biogeochemistry in an Earth system model. *Journal of Advances in Modeling Earth*  
 1230 *Systems*, 17(2), p.e2024MS004237.  
 1231
- 1232 Skákala, J., Wakamatsu, T., Bertino, L., Teruzzi, A., Lazzari, P., Alvarez, E., Cossarini, G., Spada, S., Nerger, L.,  
 1233 Vliegen, S., Brankart, J. M., and Brasseur, P.: SEAMLESS Target indicator quality in CMEMS MFCs (D6.1),  
 1234 <https://doi.org/10.5281/zenodo.10522305>, 2024.  
 1235
- 1236 Smith, S.L., Yamanaka, Y., Pahlow, M. and Oschlies, A., 2009. Optimal uptake kinetics: physiological  
 1237 acclimation explains the pattern of nitrate uptake by phytoplankton in the ocean. *Marine Ecology Progress Series*,  
 1238 384, pp.1-12.  
 1239
- 1240 Steele, J.H., 1962. Environmental control of photosynthesis in the sea. *Limnology and oceanography*, 7(2),  
 1241 pp.137-150.  
 1242
- 1243 Steele, J.H, Henderson, E.W. (1992) The role of predation in plankton models. *Journal of Plankton Research*,  
 1244 14(1): 157-172.  
 1245
- 1246 Steinacher, M., Joos, F., Frölicher, T. L., Bopp, L., Cadule, P., Cocco, V., Doney, S. C., Gehlen, M., Lindsay, K.,  
 1247 Moore, J. K., Schneider, B., and Segschneider, J.: Projected 21st century decrease in marine productivity: a multi-  
 1248 model analysis, *Biogeosciences*, 7, 979–1005, <https://doi.org/10.5194/bg-7-979-2010>, 2010  
 1249
- 1250 Stock, C.A., Dunne, J.P., Fan, S., Ginoux, P., John, J., Krasting, J.P., Laufkötter, C., Paulot, F. and Zadeh, N.,  
 1251 2020. Ocean biogeochemistry in GFDL's Earth System Model 4.1 and its response to increasing atmospheric CO<sub>2</sub>.  
 1252 *Journal of Advances in Modeling Earth Systems*, 12(10), p.e2019MS002043.  
 1253
- 1254 Stock, C.A., Dunne, J.P., Luo, J.Y., Ross, A.C., Van Oostende, N., Zadeh, N., Cordero, T.J., Liu, X. and Teng,  
 1255 Y.C., 2025. Photoacclimation and photoadaptation sensitivity in a global ocean ecosystem model. *Journal of*  
 1256 *Advances in Modeling Earth Systems*, 17(6), p.e2024MS004701.  
 1257



- 1258 Tagliabue A, Kwiatkowski L, Bopp L, Butenschön M, Cheung W, Lengaigne M and Vialard J (2021) Persistent  
 1259 Uncertainties in Ocean Net Primary Production Climate Change Projections at Regional Scales Raise Challenges  
 1260 for Assessing Impacts on Ecosystem Services. *Front. Clim.* 3:738224. doi: 10.3389/fclim.2021.738224  
 1261
- 1262 Tao, Zui & Wang, Yan & Ma, Sheng & Lv, Tingting & Zhou, Xiang. (2017). A Phytoplankton Class-Specific  
 1263 Marine Primary Productivity Model Using MODIS Data. *IEEE Journal of Selected Topics in Applied Earth*  
 1264 *Observations and Remote Sensing*. PP. 1-10. 10.1109/JSTARS.2017.2747770.  
 1265
- 1266 Tjiputra, J.F., Polzin, D. and Winguth, A.M., 2007. Assimilation of seasonal chlorophyll and nutrient data into an  
 1267 adjoint three-dimensional ocean carbon cycle model: Sensitivity analysis and ecosystem parameter  
 1268 optimization. *Global biogeochemical cycles*, 21(1).  
 1269
- 1270 Tjiputra, J.F., Couespel, D. and Sanders, R., 2025. Marine ecosystem role in setting up preindustrial and future  
 1271 climate. *Nature Communications*, 16(1), p.2206.  
 1272
- 1273 Thomas, M.K., Kremer, C.T. and Litchman, E. (2016), Phytoplankton temperature trait biogeography. *Global*  
 1274 *Ecology and Biogeography*, 25: 75-86. <https://doi.org/10.1111/geb.12387>  
 1275
- 1276 Uitz J, Claustre H, Gentili B, Stramski D. (2010). Phytoplankton class-specific primary production in the world's  
 1277 oceans: Seasonal and interannual variability from satellite observations. *Global Biogeochemical Cycles* 24.  
 1278 <https://doi.org/10.1029/2009GB003680>  
 1279
- 1280 Vichi, M., Pinardi, N. and Masina, S., 2007. A generalized model of pelagic biogeochemistry for the global ocean  
 1281 ecosystem. Part I: Theory. *Journal of Marine Systems*, 64(1-4), pp.89-109.  
 1282
- 1283 Ward, B.A., Dutkiewicz, S., Jahn, O. and Follows, M.J., 2012. A size-structured food-web model for the global  
 1284 ocean. *Limnology and Oceanography*, 57(6), pp.1877-1891.  
 1285
- 1286 Webb, W.L., Newton, M. and Starr, D., 1974. Carbon dioxide exchange of *Alnus rubra*: a mathematical model.  
 1287 *Oecologia*, 17, pp.281-291  
 1288
- 1289 Westberry, T., Behrenfeld, M.J., Siegel, D.A. and Boss, E., 2008. Carbon-based primary productivity modeling  
 1290 with vertically resolved photoacclimation. *Global Biogeochemical Cycles*, 22(2).  
 1291
- 1292 Wu, Z., S. Dutkiewicz., O. Jahn, D. Sher, A. White, and M.J. Follows, 2021. Modeling photosynthesis and  
 1293 exudation in the subtropical oceans. *Global Biogeochemical Cycles*, 35, doi:10.1029/2021GB006941  
 1294
- 1295 Xiao, Y. and Friedrichs, M.A., 2014. Using biogeochemical data assimilation to assess the relative skill of multiple  
 1296 ecosystem models in the Mid-Atlantic Bight: effects of increasing the complexity of the planktonic food web.  
 1297 *Biogeosciences*, 11(11), pp.3015-3030.



- 1298  
1299 Yool, A., Popova, E.E. and Anderson, T.R., 2013. MEDUSA-2.0: an intermediate complexity biogeochemical  
1300 model of the marine carbon cycle for climate change and ocean acidification studies. *Geoscientific Model*  
1301 *Development*, 6(5), pp.1767-1811.  
1302  
1303 Young, I. R., & Ribal, A. (2019). Multiplatform evaluation of global trends in wind speed and wave height.  
1304 *Science*, 364(6440), 548–552. <https://doi.org/10.1126/science.aav9527>  
1305  
1306 Yumruktepe, V.Ç., Samuelsen, A. and Daewel, U., 2022. ECOSMO II (CHL): a marine biogeochemical model  
1307 for the North Atlantic and the Arctic. *Geoscientific Model Development*, 15(9), pp.3901-3921.  
1308  
1309 Zheng, Q., Viljoen, J.J., Sun, X., Kovač, Ž., Sathyendranath, S. and Brewin, R.J., 2025. Simulating vertical  
1310 phytoplankton dynamics in a stratified ocean using a two-layered ecosystem model. *Biogeosciences*, 22(13),  
1311 pp.3253-3278.  
1312 .