



- 1 <u>Title</u>: Basin-scale connections between reach-scale sediment respiration and point-scale organic-matter
- 2 decomposition

- 4 James C. Stegen^{1,2*}, Morgan Barnes¹, Dillman Delgado¹, Brieanne Forbes¹, Vanessa A. Garayburu-
- 5 Caruso¹, Amy E. Goldman¹, Maggi Laan^{1,3}, Sophia McKever¹, Peter Regier¹, Lupita Renteria¹, Scott D.
- 6 Tiegs⁴
- 7 *Correspondence: <u>James.Stegen@pnnl.gov</u>
- 8 1. Pacific Northwest National Laboratory, Richland, WA, United States
- 9 2. Washington State University, Pullman, WA, United States
- 10 3. University of California, Riverside, CA, United States
- 4. Oakland University, Rochester, MI, United States

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<u>Abstract</u>

- 14 Stream and river ecosystems play a central role in the movement and decomposition of particulate
- 15 organic matter, serving as a conduit between terrestrial hillslopes and coastal environments. Microbial-
- 16 catalyzed decomposition generates simpler organic molecules that fuel respiration, often in the
- 17 sediments of these ecosystems. However, the degree of connection between sediment-associated
- 18 respiration (ER_{sed}) and organic-matter decomposition remains poorly understood. How that relationship
- 19 compares to decomposition's relationship with whole ecosystem (ERtot) and water column (ERwc)
- 20 respiration is also not clear. We examined the link between particulate organic matter decomposition—
- 21 using cellulose-based cotton strips as a standardized substrate—and all three components of respiration
- 22 across 48 sites in the environmentally diverse Yakima River Basin (Washington State, USA). We
- 23 hypothesized that decomposition within sediments would be most strongly related to ER_{sed}, but
- 24 decomposition rates were more closely associated with ER_{tot}, with little connection to ER_{sed} or ER_{wc}. This
- 25 suggests that particulate organic matter decomposition within stream/river sediments reflects
- 26 integrated system respiration rather than processes confined to sediments or the water column alone.
- 27 Further, across the basin, decomposition rates nearly spanned the previously reported global range for
- 28 streams and rivers and were best explained by total dissolved nitrogen (TDN), sediment grain size, and
- 29 aridity of the upstream drainage area. These results highlight the strong influence of land cover and
- 30 basin-scale biophysical variation on sediment-associated decomposition processes and indicate that
- 31 mechanistic models of organic matter decomposition in streams/rivers should account for coupled
- 32 sediment-water-land interactions.

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Introduction

- 35 Stream networks are major components of the global carbon cycle (Cole et al., 2007; Drake et al., 2018;
- 36 Talluto et al., 2024). Whole-stream metabolism is often studied as an integrated outcome of processes
- 37 occurring across the continuum from the air-water interface down through sediments that are below





38 the stream itself (Battin et al., 2023; Tank et al., 2010). The sediments include the interface between the 39 streambed and water column (i.e., the benthic zone) and the spatial domain below this interface, 40 referred to as the hyporheic zone (Boulton et al., 1998; Krause et al., 2011; Wondzell, 2011). The benthic 41 zone can be highly productive with significant primary producer (e.g., algae) and heterotrophic microbial 42 biomass (Allan et al., 2021). The hyporheic zone is also highly biogeochemically active due, in part, to 43 surface water flowing through it and mixing with groundwater to stimulate heterotrophic microbial 44 activity (Boano et al., 2014; Lewandowski et al., 2019; Zarnetske et al., 2011). Processes occurring on 45 and around sediments across the benthic-to-hyporheic continuum are often jointly responsible for the 46 bulk of biogeochemical activity in stream systems (Burrows et al., 2017; Fellows et al., 2001; Garayburu-47 Caruso et al., 2025; Naegeli and Uehlinger, 1997), with important exceptions in large rivers (Roley et al., 2023). 48 49 Metabolic processes within integrated stream systems are linked to the building of and breaking down 50 of organic matter (Hall and Hotchkiss, 2017; Odum, 1956). Streams are commonly net heterotrophic 51 whereby they release more carbon than they accumulate (Battin et al., 2023; Bernhardt et al., 2022). 52 This emphasizes the importance of understanding organic-matter decomposition in streams and its 53 connection to respiration rates, which are ultimately linked to rates of elemental cycling. Organic-matter 54 decomposition is commonly measured in stream systems by quantifying the breakdown of specific 55 substrates (Benfield et al., 2017; Woodward et al., 2012). Cellulose-based cotton strips are an 56 increasingly common model substrate for such studies as they enable broad comparisons across streams 57 (and other types of ecosystems)(Colas et al., 2019; Filbee-Dexter et al., 2022; Tiegs et al., 2024; Vyšná et 58 al., 2014). A standard approach is to place them in the field for a known amount of time, retrieve them, 59 and measure the loss in tensile strength, as a proxy for the degree of decomposition. This approach has 60 revealed many factors that impact decomposition in streams such as temperature, land use, aqueous 61 chemistry, sediment texture, stream flow, location within the stream network, and canopy cover, 62 among others (Griffiths and Tiegs, 2016; Tiegs et al., 2024). 63 Previous studies have examined the relationship between organic matter decomposition and whole-64 stream respiration (Mancuso et al., 2023; Pingram et al., 2020; Young and Collier, 2009), but have not 65 specifically tied organic matter decomposition within sediments to sediment-associated respiration. This 66 leads to an open question and the focus of our study: To what degree are point-scale rates of organic-67 matter decomposition within sediments linked to reach-scale respiration associated with the whole-68 stream ecosystem (ERtot), the sediments (ERsed), or the water column (ERwc)? ERtot represents reach scale 69 aerobic respiration from autotrophs and heterotrophs across benthic, planktonic, and hyporheic zones. 70 ER_{sed} comprises reach scale sediment-associated respiration from benthic/streambed sediments, 71 rooted/submerged plants, and hyporheic zones that are hydrologically connected to the active channel, 72 and ER_{wc} constitutes reach scale planktonic respiration occurring only in the water column. 73 We specifically tested the hypothesis that reach-scale ER_{sed} rates are strongly linked to point-scale 74 measurements of organic matter decomposition within streambed sediments. More specifically, we 75 tested the prediction that cotton-strip-decay rates will be best explained by ER_{sed}, with little additional 76 variation in decay rates explained by ERtot or ERwc. To test our hypothesis and associated prediction we 77 used field deployments across the Yakima River Basin (YRB). The YRB is an environmentally diverse basin 78 in southeastern Washington State (USA) that is ~16,000 km2 and with a stream network that culminates 79 in the 7th-order Yakima River. To generate the data needed to test our hypothesis, we used a 80 combination of sensors and cotton strips across 48 sites in the YRB that collectively spanned a





continuum from small mountainous streams in coniferous forests with little human impact to a large lowland river in an arid environment surrounded by significant agricultural land use. The resulting patterns help to fill a fundamental knowledge gap in our understanding of how organic matter decomposition relates to respiration and can be used to inform models that aim to mechanistically integrate biogeochemical processes within and across stream networks.

Methods

To evaluate the linkages between organic matter decomposition and stream ecosystem respiration we took advantage of a prior study (Garayburu-Caruso et al., 2025) that separated ER_{tot}, ER_{sed}, and ER_{wc} across environmentally divergent locations in the YRB (Fig. 1). Garayburu-Caruso et al. (2025) used dissolved oxygen (DO) sensors, dark bottle incubations, and the single-station method (Odum, 1956) to estimate these three components of respiration. In addition, that study deployed sensors to log water temperature used here to calculate cumulative degree days, as described below. These and associated contextual data were downloaded from existing data packages (Delgado et al., 2023; Forbes et al., 2023; Garayburu-Caruso et al., 2023), and methods are described in detail by Garayburu-Caruso et al. (2025). In brief, DO timeseries were analyzed via StreamMetabolizer (Appling et al., 2018) to estimate ER_{tot}. To estimate ER_{wc}, 2 L opaque bottles containing a DO sensor were filled with stream water and incubated *in situ*; the rate of DO drawdown was used as the estimate of ER_{wc}. The difference between ER_{tot} and ER_{wc} was used as an estimate of ER_{sed}, which represents all respiration in the stream system that is not directly occurring in the water column. ER_{sed} therefore includes respiration in the hyporheic zone, the streambed surface, and rooted/submerged plants.

To take advantage of the ecosystem respiration study, cotton strips made of Artist fabric (following the protocol of Tiegs et al., 2013) were deployed at the same time as the DO sensors. They were deployed upstream of the DO sensors to capture the upstream reach that influenced the DO sensor readings and prevent disturbance during sensor maintenance.

The cotton strips were deployed at the interface between water and sediments for 35 continuous days at 48 sites across the YRB (Fig. 1). Deployment and retrieval days varied across four days, but all strips were deployed for 35 days: deployments were from July 25th to July 28th, 2022 and retrievals were from August 29th to September 1st, 2022. Cotton strips were cut from bolts of 12-ounce, heavy-weight cotton fabric composed of 95% cellulose (Style 548; Fredrix, Lawrenceville, GA, USA). Each strip was 27 threads wide and cut to 8.0 cm by 2.5 cm. Each cotton strip was laid flat in a stainless-steel mesh cage (10.8 x 4.5 cm, RSV Jumbo Mesh Herb Infuser) to minimize physical damage and feeding by macroinvertebrates, thereby emphasizing microbe-based decomposition. At each site, four cages with one strip each were attached to the underside of clay bricks (20 x 10 x 5.5 cm) with stainless steel wire. The brick/cage/strip setup was nestled into streambed sediment such that the cages/strips were within the sediments and the brick was at the sediment/water interface. The four cages were next to each other. This setup kept the cotton strips out of direct light and within the sediments while allowing water to flow past the cotton strips.

After the 35-day incubation period, cotton strips were carefully removed from cages and gently brushed with gloved hands in stream water for approximately 10 s to remove large debris. Cleaned cotton strips were placed in 50 mL conical centrifuge tubes with 70% ethanol. Tubes were capped and rolled





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122 approximately 10 times before ethanol was removed, and clean 70% ethanol was added to the 50 mL 123 tube to minimize further microbial-based decomposition. Cotton strips were transported in the ethanol 124 filled tubes on blue ice to Pacific Northwest National Laboratory in Richland, WA. At the laboratory, the 125 ethanol was removed from the tubes and cotton strips were air dried overnight prior to further drying in 126 an oven at 40 °C for at least 24 hours. After drying, cotton strips were stored in air-tight containers with 127 desiccant. 128 Dried cotton strips were shipped to Oakland University for tensile strength analysis following the 129 protocol in Tiegs et al., (2013). A tensiometer was used to estimate tensile strength (Mark 10 MG100 130 with a Chatillon TCM 201 with roller jaws). The tensiometer pulled each cotton strip at a rate of 2 131 cm/min. Some of the cotton strips were completely degraded such that there was no material to 132 measure, while other cages only contained fragments that were too small to measure tensile strength. 133 In both cases, a limit of detection was assigned as the lowest tensile strength calculated in Tiegs et al. 134 (2019) divided by 2, resulting in a final value of 0.05. This was done to avoid statistical artifacts that can 135 arise when simply introducing a value of 0. 136 Tensiometer data were converted into decay rates using tensile loss calculated via Equation 1 (as in 137 Mancuso et al., 2022). $K = \frac{-\ln (T_s/T_{sc})}{Time}$ 138 Equation 1 139 Here, K is the decomposition rate, T_s is the post-incubation tensile strength of the deployed cotton 140 strips, and T_{sc} is the mean tensile strength of control strips that were not incubated in the field. The time 141 variable was calculated as either the number of chronological deployment days (i.e., 35 days) or the 142 number of degree days. Using degree days as the time variable accounts for variation in temperature 143 across field sites and was estimated separately for each site as the sum of mean daily river temperature 144 over the incubation period. We use Kcd and Kdd to represent decay rate per chronological day or per 145 degree day, respectively. The values of Kcd and Kdd were estimated for each individual cotton strip and 146 then replicates were averaged to provide a single site-level value for K_{cd} and K_{dd} . 147 We examined both Kcd and Kdd to evaluate whether the connection between decomposition and reach-148 scale respiration rates depends on accounting for temperature variation across the study basin. This is 149 particularly relevant in the YRB because our field sites ranged from colder headwater streams to warmer 150 low-gradient rivers. To test our hypothesis, we conducted both univariate and multivariate regression-

multiple regression analysis to find an optimized model to explain variation in either K_{cd} or K_{dd} . Further, to explore how other system variables may explain further variation in decomposition rates. a LASSO (Least Absolute Shrinkage and Selection Operator) regression model was built using physical, chemical and environmental variables (Table S1) as inputs, and K_{cd} or K_{dd} as the response variables. Variables were cube root transformed and z-score normalized to reduce the impact of high leverage points in the regression analysis and to equally weight all variables. The LASSO regression was performed over 100 iterations, each with a different random seed using the cv.glmnet function in the glmnet R package (Friedman et al., 2010). β coefficients were normalized to the maximum β coefficient for each iteration, then averaged over the 100 iterations for the final reported value. Both the raw and normalized mean β coefficient and standard deviation are reported in addition to the R2 (Table 2).

based analyses. We used ordinary least squares regression to examine the strength of univariate

correlations between K_{cd} or K_{dd} and ER_{tot}, ER_{wc}, or ER_{sed}. We complemented this univariate analysis with





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Results and Discussion

Decomposition in the Yakima River Basin spans globally reported rates

166 Both K_{cd} and K_{dd} exhibited a wide range of values (Fig. 2), effectively spanning the theoretical maximum 167 of what could have been observed with our deployment setup. This is evidenced by some cotton strips 168 being completely consumed prior to retrieval (Kcd and Kdd maximized), while others were largely intact 169 $(K_{cd} \text{ and } K_{dd} \approx 0)$. This variation is surprising given the relatively small spatial domain sampled by this 170 study, and emphasizes that environmental heterogeneity can surpass the effects of spatial extent 171 (Mancuso et al., 2022). The environments studied here ranged from pristine locations in the 172 mountainous headwaters of the YRB to lowland locations with heavy agricultural influences (Fig. 1; Laan 173 et al. 2025). This emphasizes the value of climatically diverse watersheds like the YRB as useful testbeds 174 to study variation in decomposition rates within single hydrologically connected basins.

Comparing decay rates from the YRB to a global dataset from > 500 streams and rivers (Tiegs et al., 2024) showed that YRB rates spanned nearly the entire global range and had substantial overlap with the bulk of the global distributions (Fig. 2a,b). Dominant peaks in the YRB rate distributions were shifted slightly towards faster rates, relative to primary peaks in the global dataset (Fig. 2a,b). This shift towards faster rates and the wide range in rates may be because YRB rates were estimated in later summer, a time of year when decay processes are likely maximized due to relatively high temperatures and slow flows (Collier et al., 2013b; Mancuso et al., 2023). The two decay rates were also closely correlated with each other, though the relationship weakened towards locations with faster decay rates (Fig. 2c). This suggests a weak influence of temperature in the YRB; a strong influence of temperature should lead to a weak relationship between Kcd and Kdd. These results are surprising given previously reported influences of temperature on particulate organic matter decomposition (Benbi et al., 2014; Griffiths and Tiegs, 2016). Temperature-driven decomposition is also expected to lead to a strong relationship between Kcd and summed temperature, but we observed a very weak relationship despite ~4-fold variation in summed temperature (Fig. 2d). This range in temperature among sites would likely be smaller in other seasons, and we do not therefore expect a strong influence of temperature to emerge in the YRB by conducting the study in other seasons. These results emphasize the need to understand factors governing variation in decay rates across the YRB. This is especially true given that rate distributions within this one basin span nearly all globally observed decay rates.

Across the YRB there appears to be potential for some spatial organization for both K_{cd} and K_{dd} (**Fig. 3**). Visual inspection of the maps suggests that the spatial organization may be stronger for K_{cd} than for K_{dd} . To evaluate this possibility more rigorously, we regressed each decay rate against upstream drainage area (Fig. 4). In this case, drainage area is meant to reflect position within the YRB. We used drainage area in preference to stream order because it is a continuous variable directly tied to the spatial domain a given stream integrates, whereas stream order is categorical and primarily reflects stream network topology. Associated regressions were significant (p < 0.05) with both decay rates increasing with drainage area (**Fig. 4**). The relationship with drainage area was stronger, in terms of R^2 , for K_{cd} . Both relationships were, however, relatively weak with R^2 values of 0.22 and 0.14 for K_{cd} and K_{dd} , respectively (**Fig. 4**). Nonetheless, the existence of a significant relationship after controlling for temperature (i.e., for K_{cd}) indicates that spatially structured factors other than temperature influence decay rates. This is not surprising as studies using cotton strips have found several factors that influence decomposition, such as





nutrient concentrations, turbidity, and many others (Collier et al., 2013b; Pingram et al., 2020; Tiegs et al., 2024). Before exploring a broad suite of potential explanatory variables, we tested our hypothesis that decomposition rates will be better explained by sediment-associated respiration (ER_{sed}) than by respiration in the water column (ER_{wc}) or by respiration of the integrated stream system (ER_{tot}).

Respiration in sediments explains little variation in decomposition

Contrary to our hypothesis, we found that both decay rates were most strongly connected to ERtot, less so with ER_{sed}, and not at all with ER_{wc} (Fig. 5). Univariate models using ER_{tot} were better than multivariate models using ER_{sed}, ER_{wc}, and their interaction. This is evidenced by univariate models using ER_{tot} having AIC values more than two units lower than multivariate models containing ER_{sed} and ER_{wc} (Table 1). Multivariate models were not used with ERtot because it contains ERsed and ERwc. These results are surprising, in part, because the cotton strips were deployed within the shallow hyporheic zones (i.e., within the riverbed sediments). This deployment strategy is a key reason we hypothesized that decay rates would be most strongly connected to ER_{sed}. The results indicate that decomposition of particulate organic matter within the shallow hyporheic zone is linked to respiratory processes occurring in both the sediment and water column. We propose that if our deployment configuration was complemented with a simultaneous deployment that enabled growth of benthic algal biofilms on the cotton strips, the combined decomposition from both deployments would capture substantially more of the processes that contribute to ERsed. This would provide a more complete view of sediment-associated biogeochemical function, potentially leading to a stronger correlation between decomposition rates and ER_{sed}. While this remains to be tested, the underlying idea is that primary producers support a large portion of heterotrophic respiration associated with riverbed sediments, which is supported by recent analyses showing a strong link between ERsed and gross primary production across the YRB (Garayburu-Caruso et al., 2025).

Slower rates of Kcd and Kdd are in streams with coarse sediments, set within wet forests

Given that our hypothesis was rejected and that ER $_{tot}$ explained only about 29% and 16% of K $_{cd}$ and K $_{dd}$, respectively (Table 1, Figure 5), we used a discovery-based approach to explore other system variables that may explain further variation in decomposition rates. LASSO-based modeling indicated that total dissolved nitrogen (TDN) and the median grain size of sediment texture (D50) were most important for explaining Kdd, while TDN and aridity were most important for explaining K $_{cd}$ (Table 2). Other variables were retained in the LASSO models (Table S1), but we interpreted TDN, D50, and aridity as the most important because they consistently had the largest normalized coefficients. This interpretation is based on these variables having mean normalized coefficients above 0.5—in terms of absolute value—meaning they were at least 50% as important as the most important variable in the 100 LASSO model runs. Further, the LASSO coefficients for these variables had a coefficient of variation less than 0.5, meaning that across the 100 LASSO model runs the values of their normalized coefficients were relatively stable (Table S1). The LASSO modeling also confirmed a relatively weak influence of temperature, evidenced by relatively small and highly variable β coefficients for summed temperature in the Kcd model (Table S1); temperature was not used in the Kdd model.





245 Both decomposition rates increased with higher TDN concentrations, while Kdd decreased with larger 246 D50 and Kcd decreased with higher aridity index values (Table 1). To more deeply interpret these 247 relationships, we examined Pearson-based univariate correlations between these three explanatory 248 variables and other variables included in the LASSO models. This is important because of strong 249 collinearity among some explanatory variables (Fig. S1). In this case, variables identified as being the 250 most important may be acting as proxies for one or more additional variables. We found that TDN was 251 most strongly correlated with percent agricultural land cover of the upstream drainage area and sulfate 252 concentrations in the stream water (Fig. S1). Increases in both decomposition rates with TDN may, 253 therefore, reflect agricultural inputs of nutrients that increase overall microbial activity of the stream 254 ecosystems we studied. D50 was most strongly correlated with the aridity index, which was most 255 strongly correlated with percent forest cover; the correlation between D50 and aridity is unlikely to 256 reflect a causal connection, while aridity and forest cover most likely are causally linked. If the 257 relationship between decomposition and D50 is causal, it could be mediated by the total surface area 258 available for microbial attachment. Coarser sediments have much less surface area, potentially limiting 259 overall microbial activity.

Considering the directionality of the univariate relationships, in context of the LASSO outcomes, suggests slower decomposition—for both rates—in streams with relatively coarse sediments and set within relatively wet forests. This contrasts with Clapcott and Barmuta (2010) who found faster decomposition in coarser sediments. The discrepancy is likely because we excluded macroinvertebrates while they did not, and they interpreted the link to sediment texture as due to greater habitat availability for macroinvertebrates in coarser sediments. Locations with slower decomposition should, thus, primarily be in higher elevation, relatively pristine parts of the YRB, while faster decomposition occurs at lower elevations impacted by agricultural inputs. These results are consistent with previous work showing greater cotton strip decomposition in impaired streams (Young and Collier, 2009), those with little natural land cover (Collier et al., 2013a; Webb et al., 2019), and in streams with higher nutrient concentrations (Ferreira et al., 2015; Pingram et al., 2020; Tiegs et al., 2013). In addition to differences in nutrient concentrations between higher and lower elevation sites, we expect less light penetration to streams in higher elevation sites because of more forest cover and smaller streams. Though not measured here, less light could suppress autotrophic production which will limit heterotrophic respiration (Bernhardt et al., 2022; Mulholland et al., 2001; Young and Huryn, 1999) and, in turn, indirectly lead to slower decomposition.

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Decomposition is linked to processes across the sediment-water continuum and land features

Our results collectively indicate that to study shallow hyporheic zone decomposition processes, it is not sufficient to conceptualize organic-matter decomposition by only considering sediment or water column processes; one must examine the integrated system. The implication of our analyses is that organic matter decomposition rates are linked to the integrated system more strongly than they are linked to individual components or direct interactions between components of that integrated system. Mechanistic models of stream ecosystem respiration, therefore, need to account for sediment processes, water-column processes, and land-cover influences from beyond the stream. Focusing exclusively on the hyporheic zone is insufficient, even in systems for which the hyporheic zone accounts for most reactions (Boano et al., 2014; Burrows et al., 2017; McClain et al., 2003). This is further





287 emphasized by previous work showing that most respiration occurs in the water column of large rivers 288 (Gardner and Doyle, 2018; Roley et al., 2023). Garayburu-Caruso et al. (2025) also show that fractional 289 contributions of ER_{sed} to ER_{tot} is often high, but that there is significant variation in those fractional 290 influences across the YRB. This variability is due to ER_{wc} being fast enough, in some locations, to account 291 for more than 80% of ERtot (Garayburu-Caruso et al., 2025). Similarly, Laan et al. (2025) found substantial 292 overlap between the distribution of ER_{wc} rates from the YRB and ER_{tot} from across the contiguous United 293 States. The overall picture is that decomposition is the result of integrated processes occurring across 294 the sediment-water continuum and influenced by external factors tied to land cover and land use. We 295 infer that these integrated processes are influenced by biophysical variation across the YRB (Laan et al., 296 2025), leading to decomposition rates within this single basin that resemble global rate distributions and 297 nearly span the global range of observed rates (Tiegs et al., 2024). Other basins that contain only one 298 ecoregion or have homogeneous land cover may be expected to have a narrow range of decomposition 299 rates (Webb et al., 2019). Nonetheless, models applied to any stream network that aim to predict 300 spatiotemporal variation in decomposition rates would be well served by considering processes 301 throughout the integrated watershed system. 302 Code and data availability 303 Data and scripts used to generate the main findings within this manuscript can be found at https://github.com/river-corridors-sfa/rcsfa-ST-2B-SSS-cotton-strip. Upon acceptance of this 304 305 manuscript, they will be published on the U.S. Department of Energy's Environmental System Science Data Infrastructure for a Virtual Ecosystem (ESS-DIVE) repository Other data collected during the field 306 307 efforts (i.e., sensor data; surface water chemistry data; and geospatial information, metadata, and maps 308 for 2021 Spatial Study sampling event) can be accessed on ESS-DIVE at https://data.ess-309 dive.lbl.gov/datasets/doi:10.15485/1987520 (Garayburu-Caruso et al., 2023), https://data.ess-310 dive.lbl.gov/datasets/doi:10.15485/1969566 (Delgado et al., 2023), and https://data.essdive.lbl.gov/datasets/doi:10.15485/1923689 (Forbes et al., 2023). 311 312 **Author contributions** 313 Conceptualization: JCS, MB, VAGC, PR, ST 314 Data Curation: MB, JCS, BF, ML, SM, DD, LR and AEG 315 Formal Analysis: MB, MML, BF, and JCS 316 **Funding Acquisition: JCS** Investigation: MB, DD, BF, VAGC, AEG, ML, SM. PR, LR, ST and JCS 317 318 Methodology: MB, DD, BF, VAGC, AEG, ML, SM. PR, LR, ST and JCS 319 Project Administration: MB, VAGC and JCS 320 Resources: MB, DD, BF, VAGC, AEG, ML, SM. PR, LR, ST and JCS 321 Software: MB, MML, BF, and JCS 322 Supervision: MB, VAGC and JCS 323 Validation: MB and JCS





324 Visualization: MB, MML, BF, and JCS 325 Writing - Original Draft Preparation: MB, VAGC, ST and JCS Writing - Review & Editing: MB, DD, BF, VAGC, AEG, ML, SM. PR, LR, ST and JCS 326 327 **Competing interest** 328 The authors declare that they have no conflict of interest. 329 Acknowledgements 330 This work was supported by the River Corridor Science Focus Area (RC-SFA) at the Pacific Northwest 331 National Laboratory (PNNL). The RC-SFA is supported by the United States Department of Energy, Office 332 of Biological and Environmental Research (BER), Environmental System Science (ESS) Program. PNNL is 333 operated by Battelle Memorial Institute for the United States Department of Energy under contract no. 334 DE-AC05-76RL01830. We thank the United States Forest Service, Washington Department of Natural 335 Resources, Washington Department of Fish and Wildlife, Confederated Tribes and Bands of the Yakama 336 Nation, and Cowiche Canyon Conservancy for access to field locations where these samples were 337 collected. We also thank the Confederated Tribes and Bands of the Yakama Nation Tribal Council and 338 Yakama Nation Fisheries for working with us to facilitate sample collection and optimization of data 339 usage according to their values and worldview. We thank the field team including: Dillman Delgado, 340 Morgan Barnes, Brandon T. Boehnke, Yunxiang Chen, Kali Cornwell, Brianna I. Gonzalez, Samantha 341 Grieger, Glenn E. Hammond, Peishi Jiang, Bing Li, Zhi Li, Xinming Lin, Sophia A. McKever, Maruti K. 342 Mudunuru, Katherine A. Muller, Opal Otenburg, Aaron Pelly, Kelsey Peta, Alan Roebuck, Joshua M. 343 Torgeson, and Jianqiu Zheng. 344 345 References 346 Allan, J. D., Castillo, M. M., and Capps, K. A.: Stream Ecology: Structure and Function of Running Waters, 347 Springer International Publishing, Cham, https://doi.org/10.1007/978-3-030-61286-3, 2021. 348 Appling, A. P., Hall Jr., R. O., Yackulic, C. B., and Arroita, M.: Overcoming Equifinality: Leveraging Long 349 Time Series for Stream Metabolism Estimation, Journal of Geophysical Research: Biogeosciences, 123, 350 624-645, https://doi.org/10.1002/2017JG004140, 2018. 351 Battin, T. J., Lauerwald, R., Bernhardt, E. S., Bertuzzo, E., Gener, L. G., Hall, R. O., Hotchkiss, E. R., 352 Maavara, T., Pavelsky, T. M., Ran, L., Raymond, P., Rosentreter, J. A., and Regnier, P.: River ecosystem 353 metabolism and carbon biogeochemistry in a changing world, Nature, 613, 449-459, 354 https://doi.org/10.1038/s41586-022-05500-8, 2023. 355 Benbi, D. K., Boparai, A. K., and Brar, K.: Decomposition of particulate organic matter is more sensitive to 356 temperature than the mineral associated organic matter, Soil Biology and Biochemistry, 70, 183–192, 357 https://doi.org/10.1016/j.soilbio.2013.12.032, 2014. 358 Benfield, E. F., Fritz, K. M., and Tiegs, S. D.: Chapter 27 - Leaf-Litter Breakdown, in: Methods in Stream 359 Ecology (Third Edition), edited by: Lamberti, G. A. and Hauer, F. R., Academic Press, 71–82,

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533 Figures

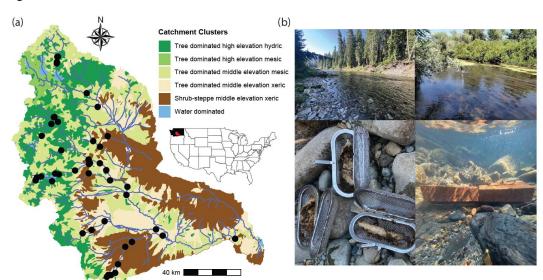


Figure 1. Biophysical clusters, sampling locations, and example conditions across the YRB. (a) The inset map shows the location of the YRB within the contiguous United States, with black indicating Washington State and red indicating the YRB. The YRB is shown with multiple colors, which correspond to biophysical clusters, as presented in Laan et al. (2025) and summarized briefly in the legend. Black circles are locations where decay rates were estimated. (b) Photos provide examples of the breadth of conditions studied across the YRB, post-incubation states of cotton strips, and deployment of the cotton strips.





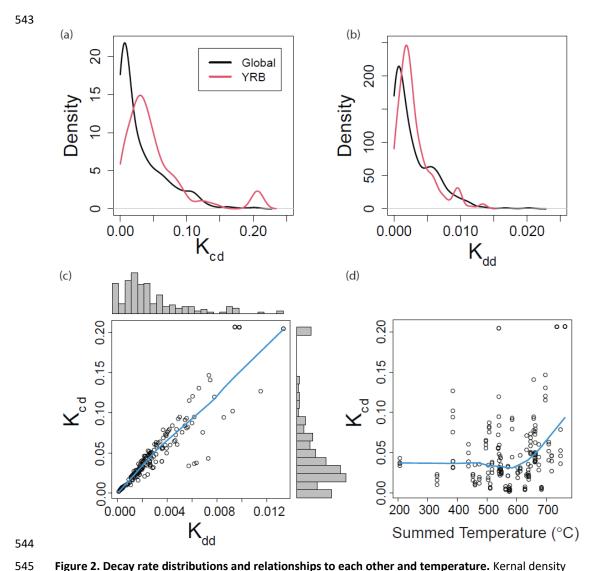


Figure 2. Decay rate distributions and relationships to each other and temperature. Kernal density functions for (a) K_{cd} and (b) K_{dd} from a global streams/rivers dataset and from the YRB. (c) Scatterplot correlating K_{cd} to K_{dd} . Histograms summarizing the distribution of each rate are provided on the outer boundaries. (d) K_{cd} related to temperature summed across the deployment period; summed temperature was used to calculate K_{dd} . Blue lines represent lowess spline fits as regression analysis was not required for interpretation.





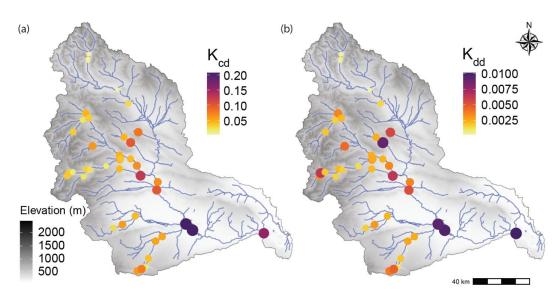


Figure 3: Spatial distribution of decay rates across the YRB. Each map shows elevation profiles and either K_{cd} (A) or K_{dd} (B). Colored circles are field locations where rates were estimated. The color of each circle is related to decay rate as indicated in the legends, and circle size is scaled to decay rate to further facilitate visual interpretability.



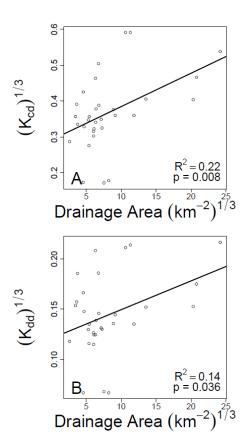


Figure 4. Decay rates increase with drainage area. K_{cd} (A) and K_{dd} (B) regressed against upstream drainage area and fit with an ordinary least squares linear regression. Associated regression models are shown as solid black lines and statistics are provided on each panel. Data were cube root transformed to improve normality before analysis.





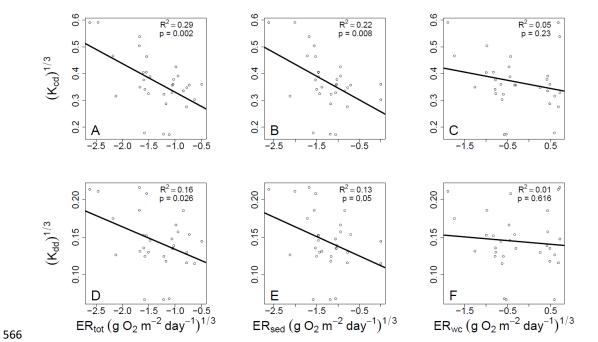


Figure 5: Decay rates related to each of the three aspects of stream ecosystem respiration. Both K_{cd} (A-C) and K_{dd} (D-F) show strongest relationships with ER_{tot} (A,D), weaker relationships with ER_{sed} (B,E), and non-significant relationships with ER_{wc} (C,F). Ordinary least squares linear regression models are shown and solid black lines and associated statistics are provided on each panel. All variables were cube-root transformed to improve normality prior to regression analysis.



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573 Tables

Table 1. Comparison of univariate and multivariate regression models explaining variation in K_{cd} and K_{dd} . Model structures are indicated, along with change in AIC relative to the best model. ER_{tot} was not used in multivariate models because $ER_{tot} = ER_{sed} + ER_{wc}$. Regression statistics for the univariate models are provided in Figure 5; only the best performing univariate models, in terms of R^2 , are shown. The models with ER_{sed} and ER_{wc} , but not the interaction term, are effectively the same as the ER_{tot} model because $ER_{tot} = ER_{sed} + ER_{wc}$. They are included as a point of reference for the model that also includes the $ER_{sed} * ER_{wc}$ interaction term.

| Model | Δ AIC |
|--|-------|
| K _{cd} ~ ER _{tot} | 0 |
| $K_{cd} \sim ER_{sed} + ER_{wc}$ | 3.8 |
| $K_{cd} \sim ER_{sed} + ER_{wc} + ER_{sed} *ER_{wc}$ | 5.7 |
| K _{dd} ~ ER _{tot} | 0 |
| $K_{dd} \sim ER_{sed} + ER_{wc}$ | 3.0 |
| $K_{dd} \sim ER_{sed} + ER_{wc} + ER_{sed} *ER_{wc}$ | 4.8 |

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Table 2. Regression coefficients from LASSO models explaining variation in K_{dd} and K_{cd} . Explanatory variables were cube root transformed to reduce influence from high leverage data points and standardized as z-scores to enable direct comparison of the regression coefficients. LASSO models were fit over 100 seeds. Regression coefficients (β) and R^2 values were averaged across the 100 seeds. Normalized regression coefficients were calculated by dividing each β coefficient by the maximum β coefficient in each seed. Standard deviation (sd) is reported for each variables' coefficient over the 100 seeds and used to calculate the coefficient of variation (cv). Variables shown have an absolute value of mean normalized β of > 0.5 and cv < 0.5 to emphasize variables that were consistently important across seeds. Results for all variables, both normalized and not normalized, can be found in Table S1.

Response Variable **Predictor Variable** Mean Normalized Regression Coefficient (β) sd C۷ 0 K_{dd} Water TDN 1 D50 -0.6990.087 -0.124Mean R² 0.502 (sd = 0.0673) K_{cd} Aridity -0.9590.065 -0.067 0.805 Water TDN 0.156 0.193 Mean R² 0.883 (sd = 0.083)

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