

Supplementary material

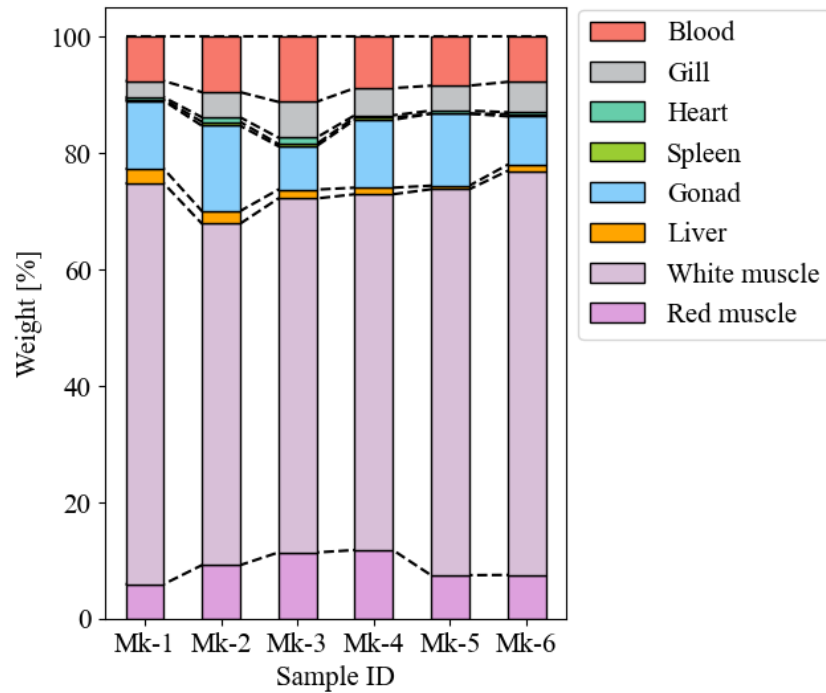


Figure S1: Proportion of wet weight among chub mackerel tissues.

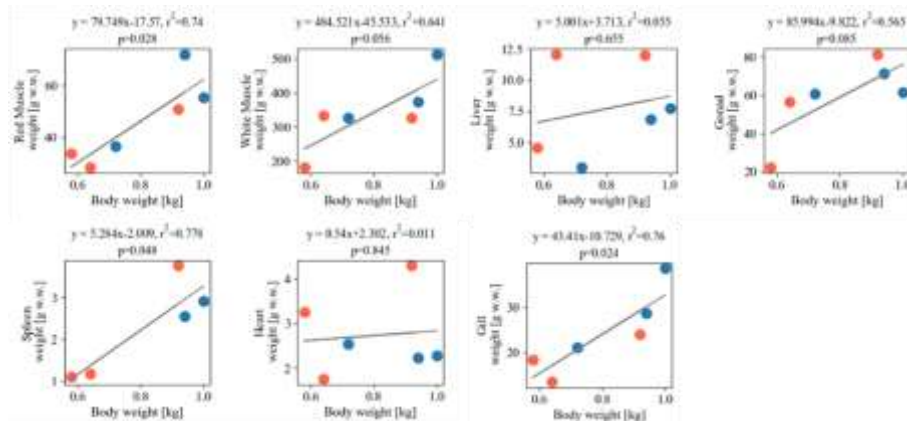


Figure S2: Correlation between tissue weight and total body weight. Red circles represent females, and blue circles represent males. Black lines show the linear regression of all data points.

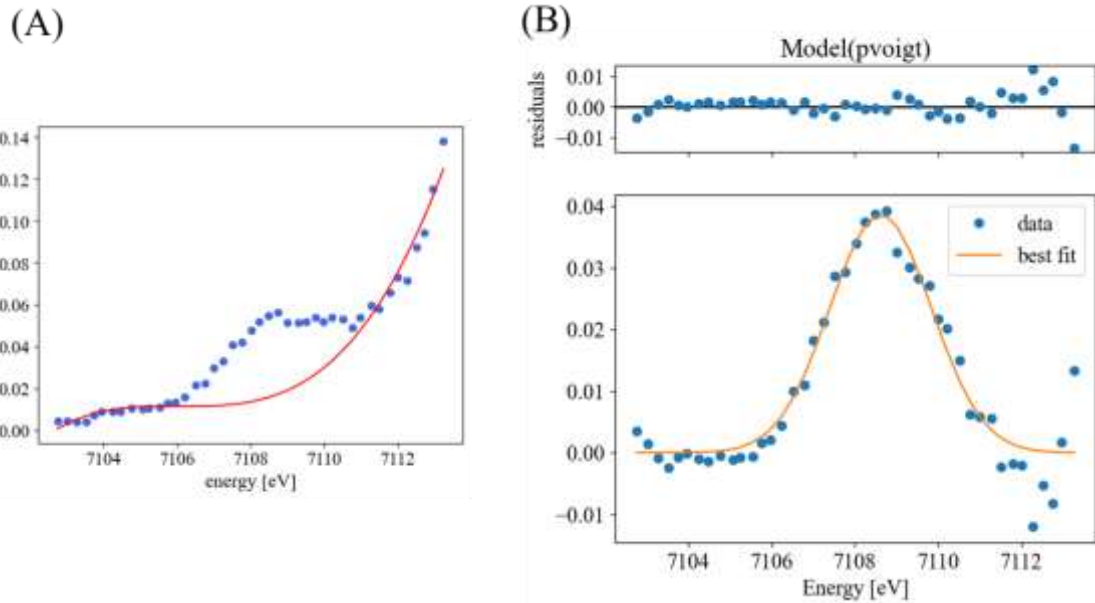


Figure S3: Example of pre-edge curve fitting. Blue dots in panel (A) represent the raw spectra around the pre-edge peak. The red line is a spline curve fitted to the raw spectra excluding the pre-edge region. The dots in panel (B) show the data obtained by subtracting the spline curve from the raw spectra. The orange line represents the fitted pseudo-Voigt model.

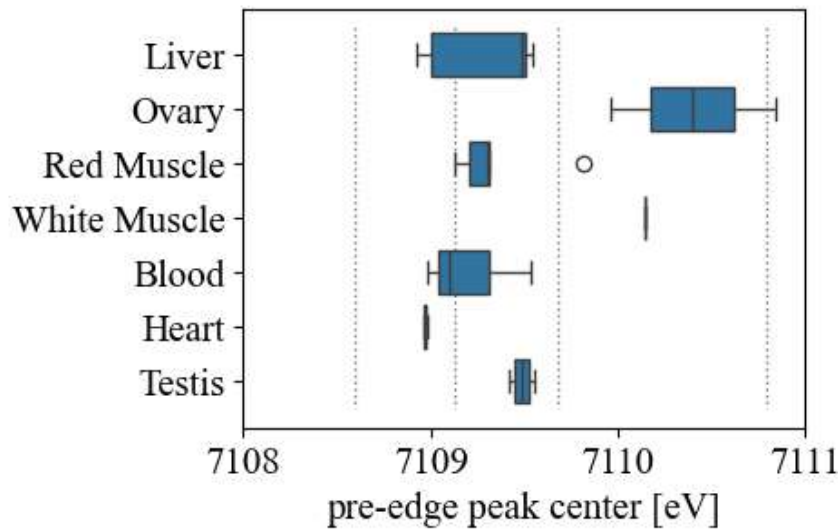


Figure S4: Pre-edge peak energies in the spectra of chub mackerel tissues. Gray dotted lines indicate the values for the following standard materials: horse spleen ferritin (Hs-Ft; 7110.8 eV), bovine blood methemoglobin (Bb-metHb; 7109.7 eV), bovine blood oxyhemoglobin (Bb-oxyHb; 7109.1 eV), and bovine deoxyhemoglobin (Bb-deoxyHb; 7108.6 eV)

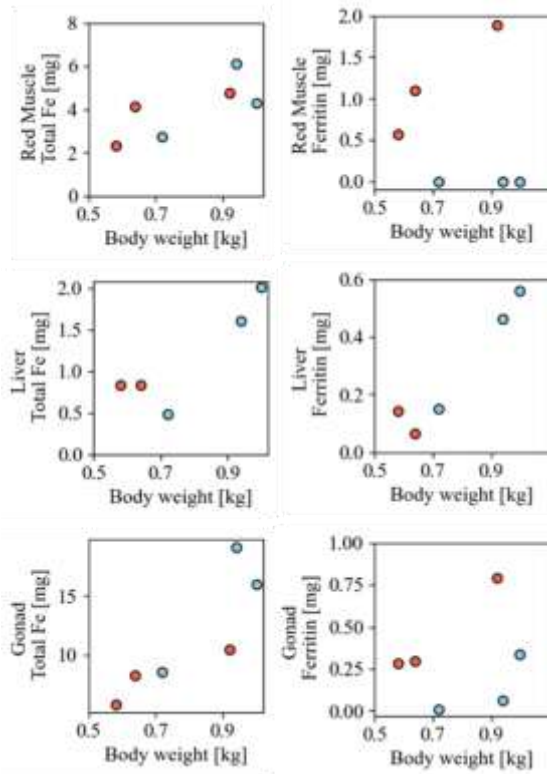


Figure S5: Correlations between body weights and iron content in either the total tissue compartment or the ferritin reservoir. Red circles represent females, and blue circles represent males.

Table S1: Tissue wet weights (g) of chub mackerel.

Sample ID	Mk-1	Mk-2	Mk-3	Mk-4	Mk-5	Mk-6	Mean
All internal organs	106	131	73	128	120	127	114
Red muscle	31.1	52.7	36.5	73.8	39.1	57.7	48.5
White muscle	333	326	182	374	326	513	342
Liver	13.7	13.6	6.1	10.1	5.3	10.6	9.90
Ovary	58.6	84.3	23.7	NA	NA	NA	55.5
Testis	NA	NA	NA	75.7	65.7	67.6	69.7
Spleen	2.7	4.8	2.8	5	NA	5.2	4.10
Heart	4	5.6	5.1	4.6	5.6	3.5	4.73
Blood	5.1	4.6	1.8	5.5	6.1	12.5	5.93
Gill	14.7	25.6	NA	31	21.1	40.9	26.7
Gut	25.2	31.5	32.3	29.5	25.2	42.4	31.0
Carcass	161	386	278	299	174	270	261

Table S2: Iron concentrations, amounts, and stable isotope ratios in tissues of chub mackerel.

Sample ID	Tissue	$\delta^{56}\text{Fe} \pm 2 \text{ S.E.}$	Fe conc. [$\mu\text{g/g w.w.}$]	Fe conc. [$\mu\text{g/g d.w.}$]	Total Fe [mg]
Mk-1	Red Muscle	-1.5 ± 0.05	144.36	317.57	4.12
Mk-1	White Muscle	-1.39 ± 0.05	4.01	12.18	1.33
Mk-1	Liver	-1.21 ± 0.04	68.95	283.11	0.83
Mk-1	Gonad	-1.48 ± 0.09	9.34	22.38	0.53
Mk-1	Spleen	-1.26 ± 0.04	720.77	3210.5	0.85
Mk-1	Heart	-1.4 ± 0.05	120.72	569.19	0.21
Mk-1	Blood	-1.29 ± 0.04	222.27	1201.54	8.29
Mk-1	Gill	-1.2 ± 0.01	96.15	329.5	1.3
Mk-2	Red Muscle	-1.56 ± 0.03	93.45	194.25	4.77
Mk-2	White Muscle	-1.5 ± 0.04	4.5	10.36	1.47
Mk-2	Liver	-1.25 ± 0.002	80.49	342.72	0.97
Mk-2	Gonad	-1.21 ± 0.06	12.32	26.03	1
Mk-2	Spleen	-1.46 ± 0.04	431	2306.69	1.62
Mk-2	Heart	-1.51 ± 0.06	70.43	393.93	0.3
Mk-2	Blood	-1.42 ± 0.04	196.19	1105.47	10.51
Mk-2	Gill	-1.34 ± 0.07	63.07	228.04	1.51
Mk-3	Red Muscle	-1.53 ± 0.04	68.94	279.89	2.34
Mk-3	White Muscle	-1.53 ± 0.04	4.77	20.29	0.87
Mk-3	Liver	-1.2 ± 0.04	181.92	695.04	0.83
Mk-3	Gonad	-1.5 ± 0.06	16.08	46.54	0.36
Mk-3	Spleen	-1.39 ± 0.04	891.42	3178.31	0.99
Mk-3	Heart	-1.42 ± 0.04	262.33	1283.99	0.86
Mk-3	Blood	-1.39 ± 0.03	174.24	1085.33	5.89
Mk-3	Gill	-1.43 ± 0.04	73.61	324.04	1.35
Mk-4	Red Muscle	-1.66 ± 0.04	84.66	217.74	6.09
Mk-4	White Muscle	-1.58 ± 0.03	3.64	12.44	1.36
Mk-4	Liver	-1.07 ± 0.04	234.24	858.21	1.61
Mk-4	Gonad	-1.04 ± 0.06	10.68	47.63	0.76
Mk-4	Spleen	-1.43 ± 0.04	872.5	3425.53	2.22
Mk-4	Heart	-1.49 ± 0.03	120.75	516.76	0.27
Mk-4	Blood	-1.28 ± 0.05	349.12	1704.59	19.12
Mk-4	Gill	-1.37 ± 0.04	68.96	253.94	1.97

Table S2: Continued.

Sample ID	Tissue	$\delta^{56}\text{Fe} \pm 2 \text{ S.E.}$	Fe conc. [$\mu\text{g/g w.w.}$]	Fe conc. [$\mu\text{g/g d.w.}$]	Total Fe [mg]
Mk-5	Red Muscle	-1.36 ± 0.05	74.99	279.26	2.75
Mk-5	White Muscle	-1.26 ± 0.08	6.28	23.29	2.05
Mk-5	Liver	-1.3 ± 0.05	161.98	615.63	0.49
Mk-5	Gonad	-1.26 ± 0.03	5.29	26.83	0.32
Mk-5	Spleen	NA	NA	NA	NA
Mk-5	Heart	-1.24 ± 0.06	144.24	676.72	0.37
Mk-5	Blood	-1.4 ± 0.06	206.17	1038.39	8.65
Mk-5	Gill	-1.34 ± 0.04	65.12	262.66	1.37
Mk-6	Red Muscle	-1.65 ± 0.05	77.27	233.16	4.28
Mk-6	White Muscle	-1.47 ± 0.05	3.85	13.17	1.98
Mk-6	Liver	-1.11 ± 0.05	259.06	992.82	2.01
Mk-6	Gonad	-0.92 ± 0.0001	9.17	51.65	0.56
Mk-6	Spleen	-1.3 ± 0.05	875.43	3531.84	2.56
Mk-6	Heart	-1.53 ± 0.05	185.29	865.38	0.42
Mk-6	Blood	-1.58 ± 0.04	275	1480.91	16.02
Mk-6	Gill	-1.56 ± 0.05	94.75	349.44	3.65

Principal Component Analysis of XANES

Evaluation by Linear combination fitting (LCF) carries the risk of incorrectly identifying components not originally present in the sample when too many components are included in the fitting. To address this, the number of contributing components was determined using principal component analysis (PCA) and target transformation analysis, as described by (Ressler et al., 2000) and (Martini et al., 2020). PCA of XANES spectra was performed using the singular value decomposition (SVD) algorithm, expressed as follows:

$$[A] = [E] \cdot [V] \cdot [w]^t \quad (S - 1)$$

This means that any $m \times n$ matrix A (where $m \geq n$) can be decomposed into the product of an $m \times n$ column-orthogonal matrix E , an $n \times n$ diagonal matrix V , and the transpose of an $n \times n$ orthogonal matrix w . The matrix A was constructed so that each column represented a sample XANES spectrum interpolated onto the same energy grid. The singular values, represented by the diagonal elements of V , were determined so that the experimental spectra can be reconstructed by the minimal number of components which is equal to the principal component. The singular values of SVD results for the XANES spectra of the mackerel tissue samples indicated that approximately three to four components were sufficient to reconstruct the spectra (Figure S6).

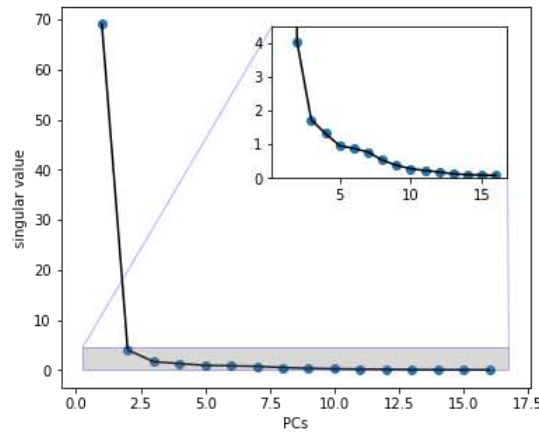


Figure S6: SVD result of the tissue spectra.

Next, sample spectra were reconstructed using one to three singular values from matrix V , as expressed as:

$$[E] \cdot [V^*] \cdot [w]^t = [A^*] \quad (S - 2)$$

where the matrix A^* represents the reconstructed sample spectra, and V^* is an $n \times n$ diagonal matrix whose diagonal elements consist of a selected subset of elements from V . For example, if two singular values are sufficient for reconstruction, V^* is given by:

$$[V^*] = \begin{bmatrix} v_{11} & 0 & 0 & \cdots & 0 \\ 0 & v_{22} & 0 & \cdots & 0 \\ 0 & 0 & 0 & \cdots & 0 \\ \vdots & \vdots & \vdots & \ddots & \vdots \\ 0 & 0 & 0 & \cdots & 0 \end{bmatrix} \quad (S - 3)$$

The reconstructed spectra of fish tissue samples are shown in Figure B. The results indicate subtle differences between the original sample spectra and the reconstructed spectra when using V^* with only one or two singular values (i.e. one or two

principal components), whereas using three principal components reproduces the sample spectra accurately (Figure S7).

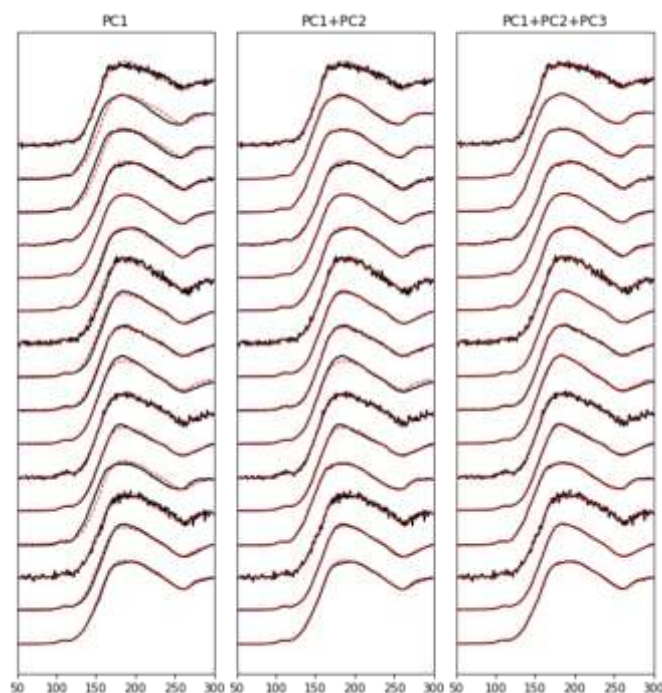


Figure S7: Reconstructed sample spectra from PCA results. Black lines represent the measured spectra, and red lines represent the reconstructed spectra.

To identify the standard materials that best fit the sample spectra, target transformation analysis was conducted. This analysis uses matrix E from the SVD results as follows:

$$[T^*] = [E] \cdot [E]^t \cdot [T] \quad (S - 4)$$

where T is a vector constructed from the XANES spectrum of standard materials. If the reconstructed vector T^* closely matches T within experimental error, T is confirmed as a principal component of the sample spectra A. The target transformation results for various standard materials (Hs-ferritin, Bb-metHb, Bb-oxyHb, Bb-deoxyHb, Fe_3O_4 , ferrihydrite, FeS, and hematite) are shown in Figure S8. These results indicated that ferritin and hemoglobin derivatives were principal components of the sample spectra, while other materials were not well reproduced. Because ferritin contains ferrihydrite-like nanoparticles within its cavity (Harrison et al., 1967; Theil, 1987), the similarity between the XANES spectra of ferritin and ferrihydrite allows for accurate reconstruction of the ferrihydrite spectrum from the sample spectra.

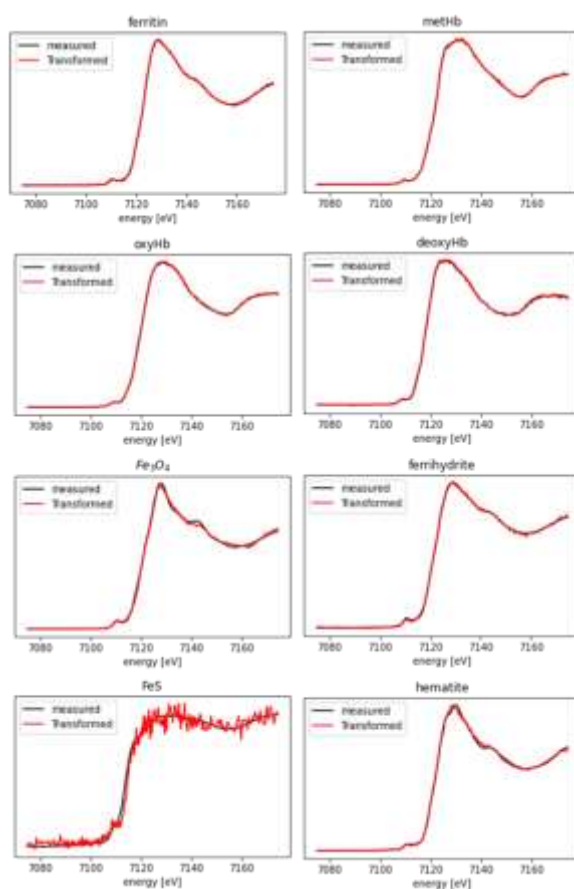


Figure S8: Results of target transformation of standard materials. Black lines represent the measured spectra, and red lines represent the transformed

References

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