



Reaction-transport modelling of methane cycling beneath the Greenland Ice Sheet

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1 Abstract

Glacial and ice sheet advances have buried large amounts of organic matter (OM), which under anoxic subglacial conditions can be microbially converted into methane (CH₄). Although CH₄ emissions have been observed at glacier margins, the capacity of subglacial environments to sustain such fluxes remains uncertain. To address this, we developed a reaction–transport model (RTM) to simulate CH₄ production, transformation, and transport in sediments beneath warm-based regions of the Greenland Ice Sheet (GrIS) margin. The model explores a wide range of environmental conditions, including sediment thickness, OM quantity and reactivity, O₂ availability, and methanotrophic activity.

Model simulations show that subglacial sediments are largely anoxic. Oxygen (O₂) penetration into subglacial sediments is generally restricted to the upper few tens of centimetres, with an average penetration depth of 22.8 cm. Microbial OM degradation and aerobic CH₄ oxidation (AeOM) represent the main O₂ sinks. Their relative contributions vary with CH₄ availability. AeOM dominates in methane-rich sediments, whereas OM degradation prevails in methane-poor environments. Modelled depth-integrated methanogenesis rates range from 0.1 to 1600 mmol-CH₄ m⁻² yr⁻¹ (mean 73 mmol-CH₄ m⁻² yr⁻¹) and are primarily controlled by OM reactivity, with sediment depth and OM concentration exerting only a small secondary influence. This sensitivity of CH₄ production rates to OM reactivity can produce sharp thresholds, where small decreases in reactivity strongly suppress CH₄ fluxes. A highly variable fraction of the generated CH₄ is consumed by AeOM within the shallow oxygenated zone, and is controlled by OM reactivity and the AeOM rate constant. Resulting net diffusive CH₄ fluxes can range between 0–234.7 mmol m⁻² yr⁻¹. Results show that even shallow sediments (<1 m) can sustain a significant CH₄ release into the subglacial environment when highly reactive OM is available, while oxidation efficiency tends to decline in thick, OM-rich deposits.



Comparison with field measurements of CH₄ export data from southwest GrIS catchments suggests that observed fluxes could be already sustained by subglacial sediments that contain as little as 0.6 wt% of relatively unreactive OM assuming a catchment sediment cover of 10 % with sediment depths of 9 m.

2 Introduction

Fluctuations in glacier extent during past glacial periods repeatedly reshaped high-latitude landscapes by overriding tundra, boreal forest, and peatland ecosystems, burying vast stocks of organic matter (OM) (Weitemeyer and Buffett, 2006; Wadham et al., 2008). Beneath the Greenland Ice Sheet (GrIS), these stocks are estimated to contain 0.5 - 27 Pg of carbon (Wadham et al., 2019). Once isolated beneath the ice, microbial respiration of OM consumes residual oxygen (O₂) and other terminal electron acceptors (TEAs) (Bottrell and Tranter, 2002; Tranter et al., 2005), creating anoxic conditions conducive to methanogenesis, the microbial production of the potent greenhouse gas methane (CH₄) (Wadham et al., 2008, 2012; Stibal et al., 2012). This CH₄ may then accumulate in subglacial sediments as immobile gas hydrates, in the presence of a gas-hydrate stability zone (Wadham et al., 2012; Lamarche-Gagnon et al., 2019), or as free gas that dissolves in porewater. Dissolved CH₄ diffuses into oxidizing sediment layers and, upon entering the subglacial hydrological system (Michaud et al., 2017), can be transported laterally and ultimately released into the atmosphere. The presence of CH₄-oxidizing clades found within microbial assemblages of glacial outlets (Lamarche-Gagnon et al., 2019; Vrbická et al., 2022; Znamínko et al., 2023; Hatton et al., in review, 2025) coupled with direct evidence for active CH₄ oxidation (Dieser et al., 2014; Strock et al., 2024), suggests that these environments may host an internal biological sink that could mitigate the net export of CH₄ from the subglacial environment (Dieser et al., 2014; Michaud et al., 2017).

While field observations confirm that CH₄ is being released from melting glacier and ice sheet margins (Stibal et al., 2012; Dieser et al., 2014; Christiansen and Jørgensen, 2018; Lamarche-Gagnon et al., 2019; Sapper et al., 2023; Hatton et al., in review, 2025), the capacity of these systems to sustain such fluxes remains a critical unknown, mostly due to limited understanding of the physico-chemical conditions within the subglacial environment. Primary among these uncertainties is the nature of the buried OM, specifically its origin (Bierman et al., 2014; Blard et al., 2023), reactivity and age (Kohler et al., 2017; Vinšová et al., 2022), and concentration (Yde et al., 2010; Stibal et al., 2012; Bhatia et al., 2013a), and its spatial distribution under the ice (Maier et al., 2021). Furthermore, the lack of direct measurements of the prevailing redox conditions that govern microbial activity severely limits our ability to assess the spatial variability of CH₄ production and oxidation potential in subglacial environments.

To address this knowledge gap, we utilize a reaction-transport model (RTM) to investigate biogeochemical processes in the GrIS subglacial ecosystem. We assess the potential for biogenic CH₄ production, consumption, and release from subglacial sediments underneath the warm-based margins of the GrIS. The model accounts for the key biogeochemical and physical processes, including microbial CH₄ production (methanogenesis) and CH₄ oxidation, and diffusive transport in and from subglacial sediments. We conduct a sensitivity study with the RTM over a large ensemble of scenarios to explore a wide range



of plausible environmental conditions. These scenarios vary key parameters, including subglacial sediment thickness, OM concentration and reactivity, O₂ concentration, and methanotrophic activity, all of which are constrained by observations from subglacial and other similar environments. We present a case study of two well-studied catchments in SW Greenland where we calculate lateral fluxes and estimate in-stream CH₄ loss. Finally, we discuss model results in the context of available field observations of CH₄ dissolved in meltwater, thus assessing the potential contribution of subglacial CH₄ to regional carbon budgets and cryosphere-climate feedback.

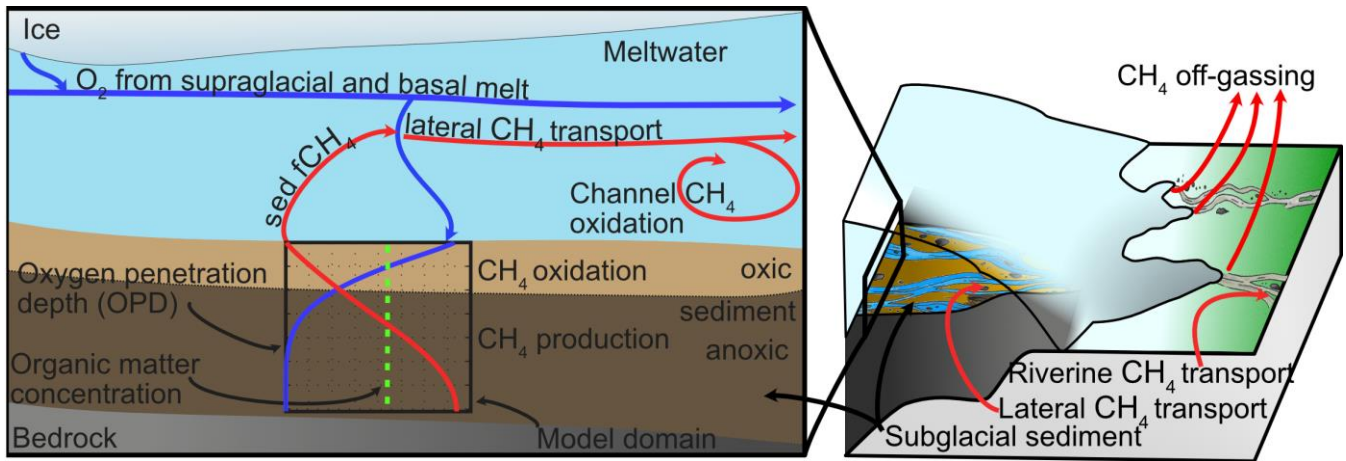


Figure 1: Conceptual diagram of subglacial and proglacial forefield transport and biogeochemical processes influencing CH₄ sources and sinks. Arrows indicate flux directions. The left panel highlights subglacial sediments and hydrological controls on CH₄ dynamics; the right panel shows an overview of subglacial and proglacial zone processes. The sediment biogeochemical model (BRNS) used in this study is outlined in black (left panel). Terminology here is consistent with that used throughout the text. Subglacial sediment CH₄ flux ($\text{sed. } f\text{CH}_4$) is calculated as $F_{\text{CH}_4, \text{sed}} = \int R_{\text{Mgen}} - \int R_{\text{MOx}}$ and glacial outlet flux as $F_{\text{CH}_4, \text{outlet}} = F_{\text{CH}_4, \text{sed}} \cdot A_{\text{catchment}} \cdot (1 - k_{\text{ox}} \cdot T_{\text{transit}})$.

3 Methods

3.1 BRNS - a reaction-transport network

We use a one-dimensional reaction-transport model (RTM), the Biogeochemical Reaction Network Simulator (BRNS, Regnier et al., 2002) and adapt it to subglacial sedimentary environments. BRNS is an adaptive simulation environment, suitable for large, mixed kinetic-equilibrium reaction networks, and has been successfully adopted across a wide range of sedimentary environments (Dale et al., 2008; Thullner, Dale and Regnier, 2009; Puglini et al., 2019). It accounts for methanogenesis, aerobic CH₄ oxidation (AeOM) and for advective and diffusive CH₄ transport to quantitatively assess the potential for biogenic CH₄ production and export from subglacial sediments underneath the GrIS for a wide range of plausible environmental conditions. Below, we describe how the model is set up to simulate subglacial soft sediments and the assumed boundary conditions.



The model solves the set of coupled one-dimensional conservation equations in porous media given by

$$\frac{\partial \sigma C_i}{\partial t} = D_i^* \sigma \frac{\partial^2 C_i}{\partial z^2} + \alpha_i \sigma (C_{i,0} - C_i) + \sigma \sum_j s_i^j R^j$$

where, the first and second terms describe the diffusive and advective transport, respectively, where C_i represents the concentration of species i , t denotes time, and z is the sediment depth. For solid species the porosity term is given by $\sigma = (1 - \phi)$, whereas for dissolved species porosity assumes $\sigma = \phi$. The effective molecular diffusion coefficient of dissolved species i is given by D_i^* ($D_i = 0$ for solid species). The third term, $\sum_j s_i^j R^j$ denotes the sum of consumption/production processes, where the stoichiometric coefficient of species i is given by s_i^j for the kinetically controlled reaction j , with rate R^j . The implemented reaction network (Table C1) considers the most important primary and secondary redox reactions, equilibrium reactions, mineral dissolution and precipitation, and adsorption and desorption processes affecting the dissolved and solid species that are explicitly resolved in the model.

Here, we explicitly simulate the coupled, steady-state reaction/transport dynamics of OM, O_2 , and CH_4 in GrIS marginal subglacial sediments. We assume that modelled diffusive transport in the liquid or gas phase is the dominant transport process in subglacial sediment environments. We assume negligible advective transport of solids and solutes across either the top (ice) or bottom (bedrock) layer. Top layer sediment mixing (cryoturbation) is also assumed to be negligible throughout the sediment column.

The following section provides a short overview of the description of OM, O_2 , and CH_4 dynamics implemented in the model. Table C2 summarizes the range of parameter values and boundary conditions used in the model.

3.2 Transport model

Our transport model assumes diffusive transport is the only dominant transport process in subglacial sediment environments. Molecular diffusion term is described by

$$F|_{diff} = D_i^*(T, S) \cdot \sigma(z) \cdot \frac{\partial C_i}{\partial z}$$

The effective diffusion coefficient, D_i^* , is given by:

$$D_i^*(T, S) = \frac{D_i(T, S)}{1 - \ln(\phi^2)}$$

where $D_i(T, S)$ is the species-dependent molecular diffusion coefficient corrected for in-situ temperature (T , in $^{\circ}C$), salinity (S , in PSU) and tortuosity ($\Phi = 1 - \ln(\phi^2)$) (Boudreau, 1997). $D_i(T, S)$ is linearly interpolated for the in-situ temperature, T (in $^{\circ}C$), using a zero-salinity and zero-degree diffusion coefficient, D_i^0 ($T = 0, S = 0$) and a temperature-dependent diffusion coefficient D_i^T (Soetaert, Herman and Middelburg, 1996).

$$D_i(T, S) = D_i^0(T = 0, S = 0) + D_i^T \cdot T$$



Sediment porosity (Φ) starts at 0.5 and decreases exponentially across the model domain until it reaches 0.25, a range and depth evolution consistent with values reported in (Dow et al., 2013; Lamarche-Gagnon et al., 2019). Similar ranges are found in measurements from western Antarctica (0.4 - 0.5) (Bougamont, Tulaczyk and Joughin, 2003; Kulesa, Hubbard and Brown, 2006).

3.3 Biogeochemical Reaction Network

3.3.1 Primary redox reactions

Primary redox reactions considered in the reaction network include the OM degradation pathways (see list of the reaction network in Table C1): aerobic respiration (r1) and methanogenesis (r2). The degradation rate of each pathway depends on the concentration of the TEAs and is kinetically limited by the presence of more powerful TEAs, reflecting their sequential utilization in the order of their decreasing yield in Gibbs energy from its half-reaction. Once the TEAs are consumed, the remaining OM is degraded via methanogenesis as the final step.

While this study prioritizes aerobic respiration and methanogenesis as the primary redox pathways, the subglacial environment contains several other TEAs whose inclusion in reactive transport models presents significant challenges. Many TEAs that are energetically more favorable than methanogenesis (such as SO_4^{2-} , NO_3^- , and bioavailable Fe(III)) have been measured in meltwaters (Tranter et al., 2002; Statham, Skidmore and Tranter, 2008; Bhatia et al., 2013; Hawkings et al., 2014, 2018) and their role in reflecting anoxic biogeochemical processes in the subglacial environment is well-documented (Tranter et al., 2002). Extrapolating discharge-weighted TEA concentrations to (dissolved and/or solid) concentrations within subglacial sediment suitable for RTM inputs is highly uncertain. This would introduce model parameters that are difficult to constrain, making the model unnecessarily complex and difficult to verify its results, since in-situ measurements of sediment composition are scarce as they are challenging to sample. Studies have observed both the rapid consumption of nitrate by denitrifying bacteria in basal ice sediment (Yde et al., 2010) and elevated bioavailable iron (Fe(III) and Fe(II)) concentrations in meltwater (Bhatia et al., 2013; Hawkings et al., 2014), highlighting their role in OM degradation via iron reduction. However, translating these findings into applicable boundary conditions is not as straightforward, and additionally the contribution of some processes, like iron reduction, may be minor despite the high TEA concentration (Lenstra et al., 2023). Therefore, given the significant uncertainties in constraining the parameters for these processes, we simplify this reaction network to focus primarily on aerobic respiration and methanogenesis.

Findings from subglacial sediment at the glacier margin indicate CH_4 originates from both acetoclastic methanogenesis (Lamarche-Gagnon et al., 2019; Pain et al., 2021; Christiansen et al., 2021; Hatton et al., in review, 2025) and hydrogenotrophic CH_4 production (Stibal et al., 2012; Telling et al., 2015). Their relative importance under varying environmental conditions, such as substrate availability, carbon oxidation state, pH, temperature, or redox potential in subglacial sediments, is yet to be determined. We thus aggregate methanogenic pathways into a single terminal reaction: $\text{OM} \rightarrow \text{CH}_4 + \text{CO}_2$. The $\text{CH}_4:\text{CO}_2$ ratio of 1 assumes an OM oxidation state of 0 (Burdige et al., 2016), and is usually applicable under optimal methanogenic



conditions (Symons and Buswell, 1933; Nilsson and Öuist, 2013), which are unlikely to be found in subglacial sediments. Nevertheless, this bulk approach, common in models of soils, sediments, and aquatic systems (Thullner et al., 2009; Regnier et al., 2011; Burdige et al., 2016; Puglini et al., 2019), avoids tracking intermediates like acetate or H_2 , which would require constraining additional poorly constrained parameters, thereby introducing additional uncertainty. Since methanogens ferment only products from OM degradation, we assume their activity scales with overall OM degradation ($k \times [OM]$). This simplification efficiently captures observed CH_4 dynamics without compromising model performance. The OM degradation rate, R_{OM} , within the sediment column is described by a reactive continuum model (RCM, see Boudreau and Ruddick, 1991). The RCM assumes a continuous and dynamic distribution of OM fractions over a range of reactivities and encapsulates the decrease in apparent reactivity with depth (and thus age) as the most reactive fractions are successively degraded. R_{OM} is given by:

$$R_{OM} = - \int_0^{\infty} k \cdot om(k, t) dk$$

where $om(k, t)$ denotes a probability density function that determines the concentration of OM having a degradability between k and $k + dk$ at time t , with k being analogous to a first-order rate constant. The initial distribution of organic fractions, $om(k, t = 0)$, may take different mathematical forms and cannot be inferred by observations. It is given by:

$$om(k, t = 0) = \frac{OM(0) \cdot a^{\nu} \cdot k^{\nu-1} \cdot e^{-a \cdot k}}{\Gamma(\nu)}$$

where $OM(0)$ is the initial OM concentration, $\Gamma(\nu)$ is the gamma function, a is the average lifetime of the more reactive fractions of the spectrum of the OM mixture, while ν is a dimensionless scaling parameter shifting the Gamma distribution near $k = 0$ towards a more or less reactive OM mixture. The two free, positive parameters a and ν completely describe the type of OM mixture over the range of k -values and therefore, the overall OM reactivity. High ν and low a values indicate a mixture of OM dominated by more readily degraded OM fractions, i.e. a higher reactivity. On the other hand, low ν and high a values, indicate a larger contribution of less reactive compounds to the total OM pool, i.e. a lower reactivity.

Under these assumptions, the decrease of bulk OM concentration as a function of depth, $OM(z)$, is given by:

$$OM(z) = OM(0) \cdot k(a, \nu) = OM(0) \cdot \left(\frac{a}{a + age(z)} \right)^{\nu}$$

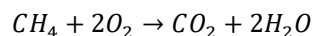
where $age(z)$ denotes the age of the sediment layer at depth z (see sec. 3.6.2 and sec. 3.6.3).

3.3.2 Secondary redox reactions

The upward migration of biologically produced reduced species (e.g. NH_4^+ and CH_4), into more oxidizing sediment layers leads to their re-oxidation via a set of secondary redox reactions, which sustain a continuous recycling of these redox sensitive elements. Specifically, we focus only on aerobic CH_4 oxidation (AeOM, see table C1) as top layers in sediments are likely oxic and acting as a CH_4 sink. The bimolecular rate laws of secondary redox reactions are based on (Wang and Van Cappellen, 1996).



CH₄ is either consumed aerobically or anaerobically (i.e., in the absence of O₂). The former is relevant in oxic and sulfate-poor freshwater sediments, such as wetlands, lakes, or soils (Knoblauch et al., 2008; Oh et al., 2020), while the latter requires an absence of O₂ and the presence of TEAs such as sulfate or ferric iron, which are common in marine sediments (Egger et al., 2018), and in sub-Arctic lake sediments (Martinez-Cruz et al., 2017). The O₂ supply derives from a combination of surface and basal melt and is more important beneath the ablation zone of the ice sheet (see sec. 4.1), where large volumes of O₂-rich surface water reach the subglacial bed through moulins and crevasses, where they subsequently fuel biogeochemical redox reactions. The importance of AeOM relative to anoxic oxidation pathways is supported by an analysis of subglacial meltwater in SW Greenland using clumped CH₄ isotopes (Adnew et al., 2025). The likely minor role of AOM is also supported by the relatively low sulfate concentrations, as reported for subglacial porewater by others (6-26 μM; although values up to 244 μM have also been reported, Christ et al., 2021; Lawson, 2012; Graly et al., 2014), and a recent subglacial sediment core porewater sample collection (~1.7-4 μM SO₄²⁻) under Isunnguata Sermia, SW GrIS (Klímová, Stibal et al., unpublished data). Additionally, unpublished incubation experiments with basal sediments using commonly available TEA (SO₄²⁻, Fe(III) and Mn(IV)) supported the assumption that AOM was negligible (Klímová, Stibal et al., in preparation, 2025). Consequently, in this study, we focused on AeOM rather than AOM, as the primary biological CH₄ sink. We formulate AeOM by aerobic methanotrophs according to the bimolecular reaction equation:



and its rate law is given by

$$r_{AeOM} = k_{AeOM}[CH_4][O_2]$$

where the rate constant for AeOM, k_{AeOM} , is further examined as a model parameter in our sensitivity analysis, since it influences the oxidation efficiency of aerobic methanotrophs.

This equation assumes kinetic and thermodynamic factors, and biomass dynamics are implicitly accounted for in the rate constant (Dale et al., 2006; Regnier et al., 2011). The range for k_{AeOM} is discussed below (sec. 3.6.6).

3.4 Boundary conditions

Dirichlet conditions (fixed concentrations) $c_i(z = 0) = c_{i,0}$ are imposed for dissolved species at the upper boundary (top of the sediment model domain), while for solid species Robin type (constant flux) conditions are set: $J_s = \rho \cdot (1 - \phi) \cdot (v \cdot c_i - D_{bio} \frac{\partial c_s}{\partial z})_{z=0}$, where c_s indicates the concentration of a solid species, ρ is the sediment density (2.0 g cm⁻³) and ϕ is the sediment porosity. At the lower boundary of the model domain, a no-flux ($\frac{\delta c_i}{\delta z} = 0$, Neumann condition) is applied for all species, which implies a negligible influence of biogeochemical processes in sediments underlying the model domain (Boudreau, 1997), i.e. no thermogenic CH₄ diffusing upward through rock cracks.



202 **3.5 Numerical solution**

203 The transport term for dissolved species is discretized using the Crank-Nicolson algorithm (Regnier et al., 1997). The reaction
 204 terms are solved simultaneously by solving the Jacobian Matrix. Transport and reaction terms are solved sequentially in each
 205 time step. The sediment model domain is subdivided into an irregular grid composed of 173 to 579 nodes with increasing grid
 206 resolution from 0.05 cm to 1.25 cm, then to 5.0 cm until z_{\max} . This avoids numerical instabilities that may emerge due to the
 207 succession of strong concentration gradients created by the fast secondary redox reactions in the upper part of the sediment
 208 column. The transient model simulations are run until steady-state is reached using an adaptable time step allowing sufficient
 209 numerical precision if needed, but still permitting considerable speed-up in computation time.

210 **3.6 Setup of the Environmental Model ensemble**

211 To identify the main controls on the depths of the methanogenic zone in subglacial sediments (i.e. $z > O_2$ penetration depth,
 212 OPD), as well as CH_4 flux from subglacial sediments, we ran a model ensemble ($n = 210$) over the below defined plausible
 213 range of subglacial environmental conditions. To this end, we used the Latin hypercube sampling approach to select 210 input
 214 vectors $\chi^j (j = 1, \dots, r)$ each with variations in the model parameters i (i.e., sediment thickness, subglacial OM concentration
 215 and reactivity, dissolved O_2 and aerobic methanotrophic activity), as suggested in the Sensitivity Analysis for Everyone
 216 (SAFE) MATLAB toolbox (Pianosi, Sarrazin and Wagener, 2015). More on the sensitivity analysis below. The following
 217 sections provide an overview and rationale for the chosen parameter ranges, and a description of the sensitivity measures
 218 calculated.

219 **3.6.1 Subglacial Sediment depth**

220 The volume of soft subglacial sediment is critical for creating an environment suitable for microbial methanogenic activity
 221 and its subsequent upward diffusion towards the sediment-water interface and into the subglacial hydrological network.
 222 However, sparse data on both sediment thickness and spatial distribution hinder an accurate assessment. Nevertheless, a
 223 growing body of evidence from along the western margin of the GrIS suggests the ice sheet bed is characterized by a mix of
 224 hard bedrock with only thin coarse sediments (Harper et al., 2017) and layers of unfrozen till/soft sediment (Booth et al., 2012;
 225 Dow et al., 2013; Kulesa et al., 2017; Ruskeenimi et al., 2018) of up to several tens of meters in thickness (Walter et al.,
 226 2014). For instance, under the glacier tongue of Isunnguata Sermia (SW Greenland), an unfrozen till layer of maximum 10 m
 227 thickness was measured (Ruskeenimi et al., 2018), while its neighboring Russell Glacier is also hypothesized to be underlain
 228 with soft sediment (Dow et al., 2013). In this study, we simulate the sediment depth down to 15 m and assume a spatial extent
 229 between 1 - 100% of the total catchment area.



3.6.2 Organic matter concentration and depth-profile

OM concentration is a key parameter influencing the subglacial biogeochemical processes. Several studies have reported bulk OM concentrations in subglacial sediments under the GrIS, yet only a few focus on thawed-bed areas of glaciers, where unfrozen till under the marginal ablation zones of the GrIS is assumed to be biologically active. The reported range from basal ice at glacier margins is 0.01 - 1.52 wt%, with a mean value of 0.21 wt% (Yde et al., 2010; Stibal et al., 2012; Graly, Drever and Humphrey, 2017). A recent pan-Arctic review of marginal subglacial OM (~0.5 wt%, Vinšová et al., 2022) corroborates these low concentrations found previously. We are aware that higher values of OM concentration up to 1.7 wt% at GISP2 (Bierman et al., 2014) and even 47.7 wt% at Camp Century (Christ et al., 2021) have been reported; however, these sediments were collected frozen from a cold-based part of the GrIS, suggesting good preservation of frozen OM and are not necessarily representative of marginal sediments underlying warm-bed glaciers, in which microbes play an active role in transforming OM (Christiansen et al., 2021; Vinšová et al., 2022).

We assume a constant OM concentration with subglacial sediment depth, due to the intense reworking of soils in proglacial areas during the glacier re-advance. Therefore, we prescribe the depth profile once as constant across the model domain. While this approach likely overestimates the depth-integrated OM content, which in turn affects all redox pathways, the current lack of OM concentration depth profiles in marginal subglacial sediment leaves too much room for speculation regarding the nature of a decreasing profile.

3.6.3 Organic matter reactivity

The RCM requires the parameters a and ν to fully describe the OM reactivity (i.e. its quality, or bioavailability); however, these are yet to be constrained for subglacial systems due to the scarcity of observational data. As revealed by plant fossils found in soil/sediment samples from several sites across Greenland (Dye 3, GRIP, John Evans Glacier, Camp Century; Willerslev et al., 2007) and in biomarker analysis of freshly thawed basal sediment (Vinšová et al., 2022), environments such as boreal forest and tundra dominated ice free areas in Greenland during the Holocene Climate Optimum. Linking paleo-vegetation analogues to present-day ecosystems is a first step to estimate the likely range in OM reactivities stored in subglacial sediments.

A recent effort to constrain the RCM parameter using incubation experiments of OM in high latitude soils (i.e., permafrost, boreal forest, and tundra vegetation), puts the range of parameter ν between 0.0012 and 1.7 with a median of 0.04; and a mean of 0.21, while parameter a is narrowed down to 0.0065 - 3.75 years; median 0.18; mean 0.9 years (Arndt, in review, 2025). Ranges are limited to their 5% and 95% quantiles respectively to prevent a skewed distribution due to outliers. The uncertainty in constraining the range in OM reactivity is further reduced by limiting parameter a to its mean value, instead of the full range, such that parameter ν solely determines the overall reactivity, k (see Eq. OM k). This simplification applies when parameter a is much lower ($a_{\max} = 14.6$ yrs) than the age of the OM in subglacial sediments (age = 4000 yrs, see Eq. OM k). The mean age of the OM exported from the subglacial environment in the Kangerlussuaq sector of the SW GrIS range between



5000 - 9000 years depending on subglacial drainage system development (Kohler et al., 2017) and reflects a mix of Holocene and Eemian materials.

Fig. 2 a shows the temporal evolution of OM k as a function of parameter a , expressed as $k = f(a, \nu = \text{const})$. Here, the selected scenarios illustrate plausible values for parameter a in high-latitude soils ($a_{\min} = 10^{-9}$ yrs; $a_{\max} = 14$ yrs), yet also include a case with $a = 1000$ years to demonstrate how OM k stabilizes to a time-invariant value after a few hundred years (red line in Fig. 2 a). Thus, we chose the median value of 0.2 years for parameter a for our model setup. This simplification holds for the time scales considered relevant here (i.e. the Holocene) and would not be applicable for OM buried during the Eemian period. On the other hand, Fig. 2 b demonstrates the considerable influence of parameter ν on OM k , $k = f(a = \text{const}, \nu)$, for the range of values found in high latitudes (Arndt, in review, 2025). Higher values of ν indicate a greater proportion of reactive compounds in the initial OM mixture. We therefore varied ν across the entire range of plausible values, i.e. between 0.0012 and 0.9.

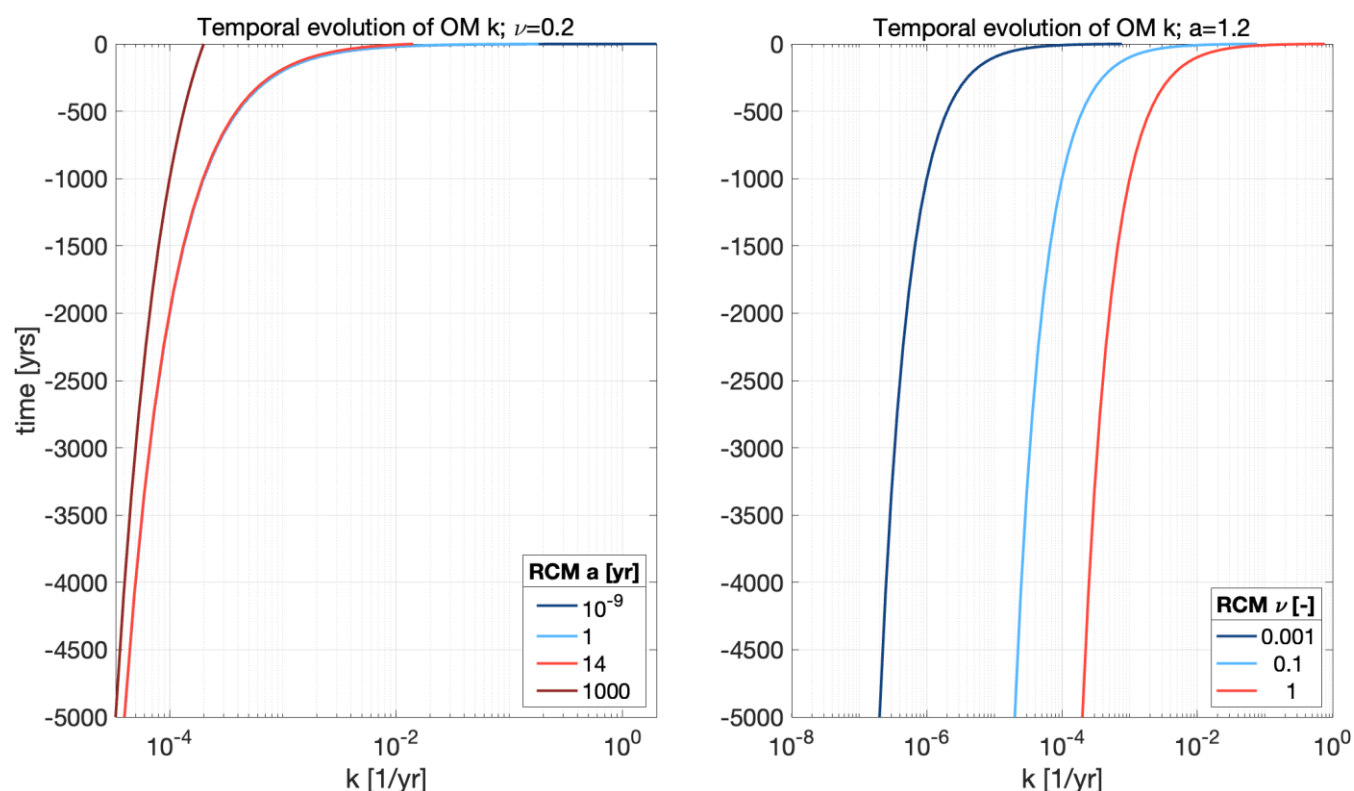


Figure 2: Change in organic matter reactivity (OM k) over time, modelled with the Reactive Continuum Model (RCM) using the equation $k = \nu / (a + t)$ from 5 kyr ago till present-day. Left panel: The effect of parameter a on OM reactivity. Parameter $\nu = 0.2$, while parameter a varies from 10^{-9} to 1000 years. OM k converges to similar values once the elapsed time (t or age) is much greater than a . The curve for $a = 1,000$ years deviates significantly because the 5,000-year timescale is not long enough to satisfy the $t \gg a$ condition. Right panel: The effect of parameter ν on OM reactivity, when parameter $a = 1.2$ years, while ν varies from 0.001 to 1. This panel



shows that an increase in parameter ν leads to an overall higher reactivity, as a larger ν implies a greater contribution of highly reactive compounds in the initial OM mixture.

3.6.4 Temperature

Temperature in the subglacial sediments is assumed stable throughout the year at $-0.5\text{ }^{\circ}\text{C}$. This represents the mean of several temperature measurements of basal ice at the glacier margin and inland of the Isunnguata Sermia glacier (Harrington, Humphrey and Harper, 2015), and we assume these are reflective of the general thermal state of soft subglacial sediments. We ignore the influence of a geothermal heat gradient on the redox chemistry and assume a constant temperature throughout the sediment column, which, given the relatively shallow simulated sediments (0.2 - 15 m) is assumed reasonable, i.e. assuming a moderate geothermal heat flux of 56 mWm^{-2} would lead to less than $0.5\text{ }^{\circ}\text{C}$ increase over simulated 15 m (Colgan et al., 2022).

3.6.5 Redox conditions in the subglacial drainage system

Redox conditions can vary within the drainage system, and there is evidence for both oxic and anoxic environments (Bottrell and Tranter, 2002; Tranter et al., 2002). The redox state of the subglacial environment depends on the supply of O_2 and its consumption rates within the seasonally developing drainage system. Underneath the GrIS, O_2 is supplied through supraglacial or basal meltwaters. Supraglacial meltwater saturated with O_2 enters the subglacial environment through moulins, increasing subglacial O_2 concentration and creating oxic conditions. In addition, basal melt also supplies O_2 trapped in air bubbles in the melting basal ice overlaying subglacial sediments. Assuming basal melt rates of $6.1 - 2.1\text{ mm yr}^{-1}$ (Dahl-Jensen et al., 2003; Buchardt and Dahl-Jensen, 2007; Harper et al., 2021) and O_2 concentrations of $291 - 446\text{ }\mu\text{M}$ in basal ice (gas bubble concentration from Herron, Hoar and Langway, 1979), this process could supply a baseline flux of $1.7 - 8.9\text{ mmol m}^{-2}\text{ yr}^{-1}$. The rates of O_2 consumption within the subglacial drainage system remain poorly constrained, but the presence of subglacial sediments likely increases local O_2 consumption due to microbial degradation of OM and oxidation of reduced species, such as CH_4 .

The spatial extent of oxic environments under the ice and its temporal evolution over the melt season remain unassessed. Nevertheless, integrated water chemistry signals measured at the glacier margins strongly suggest that anoxic conditions prevail over large parts of the subglacial drainage system over prolonged periods of time (Bottrell and Tranter, 2002; Tranter et al., 2005). To fully cover the spectrum of plausible redox conditions at the sediment-water/ice interface, we here explore the full range of dissolved subglacial water O_2 ($T = -0.5\text{ }^{\circ}\text{C}$, $S = 0\text{ PSU}$) concentrations from anoxia ($0\text{ }\mu\text{M}$) to fully oxic conditions ($456\text{ }\mu\text{M}$). It is important to note that we neglect here seasonality in O_2 concentration, i.e. O_2 replenishing supraglacial meltwater, hence assuming a given redox state prevails over the entire simulation time. Potential caveats of this are discussed below.



3.6.6 Rate constant for aerobic CH₄ oxidation

Soil microbial processes, including AeOM, are regulated by microbial biomass and community composition, with additional control from environmental factors such as temperature, CH₄ availability, moisture, pH, and soil type (Murguía-Flores et al., 2018). The effects of these drivers on AeOM are poorly constrained and rarely represented explicitly in models. Instead, rate constants are calibrated against observations, thereby implicitly accounting for factors that are not explicitly described in the model formulation and tend to show a strong variability across different environments.

The rate constant for the AeOM (k_{AeOM}) is a key parameter that exerts significant control on AeOM and thus the estimated diffusive CH₄ flux. Previously published constants in Arctic soils vary between $[2.7 - 3.4] \times 10^9 \text{ cm}^3 \text{ mol}^{-1} \text{ yr}^{-1}$ (Murguía-Flores et al., 2018), while in coastal marine sediments they span $(9.4 - 150) \times 10^7 \text{ cm}^3 \text{ mol}^{-1} \text{ yr}^{-1}$ (Mao et al., 2022). In contrast, much higher constants have been used for shallow Arctic Ocean shelf sediments ($10^{13} \text{ cm}^3 \text{ mol}^{-1} \text{ yr}^{-1}$; Puglini et al., 2019). Here, we adopt a conservative approach to avoid speculation on the oxidation efficiency, limiting the range of range of published k_{AeOM} to $[10^6 - 10^{10}] \text{ cm}^3 \text{ mol}^{-1} \text{ yr}^{-1}$.

CH₄ becomes inaccessible to microbial degradation once it solidifies into immobile CH₄ gas hydrates, which forms when high concentrations of CH₄ meet high pressures and low temperatures. Here we assume that the saturation concentration is 50 mM assuming $T=0^\circ\text{C}$, $S=0$ PSU and a pressure equivalent to 600 m of ice thickness above the sediments, which is an average ice thickness above marginal subglacial sediments.

3.7 Global sensitivity analysis

To assess the sensitivity of CH₄ fluxes and OPD from/in subglacial sediments to variations in model parameters, we used the ‘Elementary Effect Test’ (Morris, 1991; Saltelli et al., 2007), which takes the mean of finite differences as a measure of global sensitivity of input parameter i :

$$S_i = \frac{1}{r} \sum_{j=1}^r EE^j = \frac{1}{r} \sum_{j=1}^r \frac{g(\chi_1^j, \dots, \chi_i^j + \Delta_i^j, \dots, \chi_M^j) - g(\chi_1^j, \dots, \chi_i^j, \dots, \chi_M^j)}{\Delta_i^j} c_i$$

Where $g()$ is our diagenetic model, which transforms the vector of the input factors $\chi^j = (\chi_1^j, \dots, \chi_M^j)$ into the output space - specifically the simulated CH₄ fluxes and OPD at the sediment-water/ice interface. Δ_i represents the variation of the input parameter i (i.e. sediment thickness, subglacial OM concentration, and its reactivity, dissolved O₂ at the sediment-water/ice interface and aerobic methanotrophic activity) and output j . We calculate the standard deviation of the EEs, which measures the degree of interaction of input parameter i with the other input parameters. Both sensitivity indices are relative measures, hence their values do not have a specific meaning and are only used to rank the influence of the input parameters.

To compute the mean and standard deviation of the EEs for M input parameters ($M = 5$), a total of $N = r \cdot (M + 1)$ model evaluations are required. To evaluate the robustness of the sensitivity indices and determine whether they are independent of the specific input-output sample, we derived bootstrapping-based confidence limits for the indices. Following recommendations in the literature ($r > 30$, e.g., Pianosi et al., 2016), we computed $r = 35$ finite differences, a number sufficient



to distinguish between influential and non-influential parameters and to estimate reasonable confidence intervals for the sensitivity indices (Pianosi et al., 2016). Overall, for our global sensitivity analysis (GSA) we conducted $N = r \cdot (M + 1) = 210$ model simulations using varying input parameter values.

3.8 Setup for two case studies in SW Greenland

The model-derived subglacial sediment CH_4 fluxes generated across the entire range of environmental conditions (i.e., OM content and reactivity, sediment depth, and redox conditions) establish quantitative relationships between environmental parameters and resulting CH_4 fluxes, effectively creating a multidimensional response function. Comparing the observed laterally transported CH_4 with the simulated subglacial sediment CH_4 fluxes helps identify the combinations of OM availability, reactivity, sediment thickness, and oxygen availability that could reproduce the measured signal. Thus, the modelled CH_4 fluxes serve as a quantitative diagnostic framework for constraining the range of plausible subglacial environmental conditions, including the minimum OM content and its reactivity consistent with observed CH_4 export under various assumptions of catchment sediment cover.

We use the well-studied glaciers of Isunnguata Sermia (IS) and Leverett Glacier (LG) (SW Greenland) to illustrate in our case studies to extrapolate diffusive subglacial sediment CH_4 fluxes out of subglacial sediments into seasonal lateral fluxes and dissolved CH_4 concentrations in meltwater (Fig. 5 a) for different assumptions of subglacial sediment covers and CH_4 oxidation scenarios during lateral transport, as well as their loss during lateral transport. We used modelled mean cumulative discharge between May and September 2012 - 2022 ($5.5 \pm 2.4 \text{ km}^3$ for IS; $1.4 \pm 0.32 \text{ km}^3$ for LG, Mankoff et al., 2020) and compare our results to observed lateral CH_4 transport from these catchments measured between 2015 and 2023 (Lamarche-Gagnon et al., 2019; Hatton et al., in review, 2025).

3.9 Model Output

We applied our RTM to model key porewater and solid-phase depth profiles, and production and reduction rates of O_2 and CH_4 , the integrated reaction rates and the resulting diffusive fluxes. Subsequently, we extrapolated our mass balance calculation to the IS and LG catchments and compared results to previously measured dissolved CH_4 concentrations and estimated diffusive and lateral fluxes of CH_4 .

4 Results and Discussion

4.1 O_2 dynamics in subglacial sediments

Model results show that subglacial sediments in the warm-bed areas of the GrIS are predominantly anoxic, with O_2 penetration depths (OPD) ranging from 1.8-184cm and averaging 22.8 cm even with fully oxygenated overlying waters (Fig. 3). Global sensitivity analysis (GSA) results reveal that OM reactivity (parameterized by the RCM parameter ν) exerts the dominant



control over OPD, with OM content and aerobic oxidation of CH_4 (k_{AeOM}) playing a lesser role (Fig. 3). Highly reactive OM drives high O_2 demand through OM respiration and AeOM (see sec. 4.3), thus restricting penetration depth and favoring anoxic conditions. Conversely, less reactive OM reduces O_2 demand, enabling deeper penetration and ultimately limiting CH_4 production. When the subglacial water O_2 concentration is $<50 \mu\text{M}$ ($\sim 15\%$ saturation), OPD remains shallow ($<30 \text{ cm}$) regardless of other sediment properties (Fig. 3). Above $50 \mu\text{M}$, OPD scales with overlying-water O_2 deepening the oxic layer. Simulated O_2 fluxes into the sediment range from 1.6 to $240.3 \text{ mmol m}^{-2} \text{ yr}^{-1}$, with a mean of 44.0 ± 47.0 (mean \pm SD) $\text{mmol m}^{-2} \text{ yr}^{-1}$. AeOM is an important O_2 sink, accounting for up to $\sim 67\%$ ($0.03 - 66.9\%$; $23.7 \pm 18.1\%$, mean \pm SD) of the total O_2 consumption, a process that highlights a key escape path for CH_4 through the sediment-water/ice interface. Heterotrophic respiration of OM contributes between $24.9-64.6\%$ ($38.7 \pm 9.6\%$, mean \pm SD).

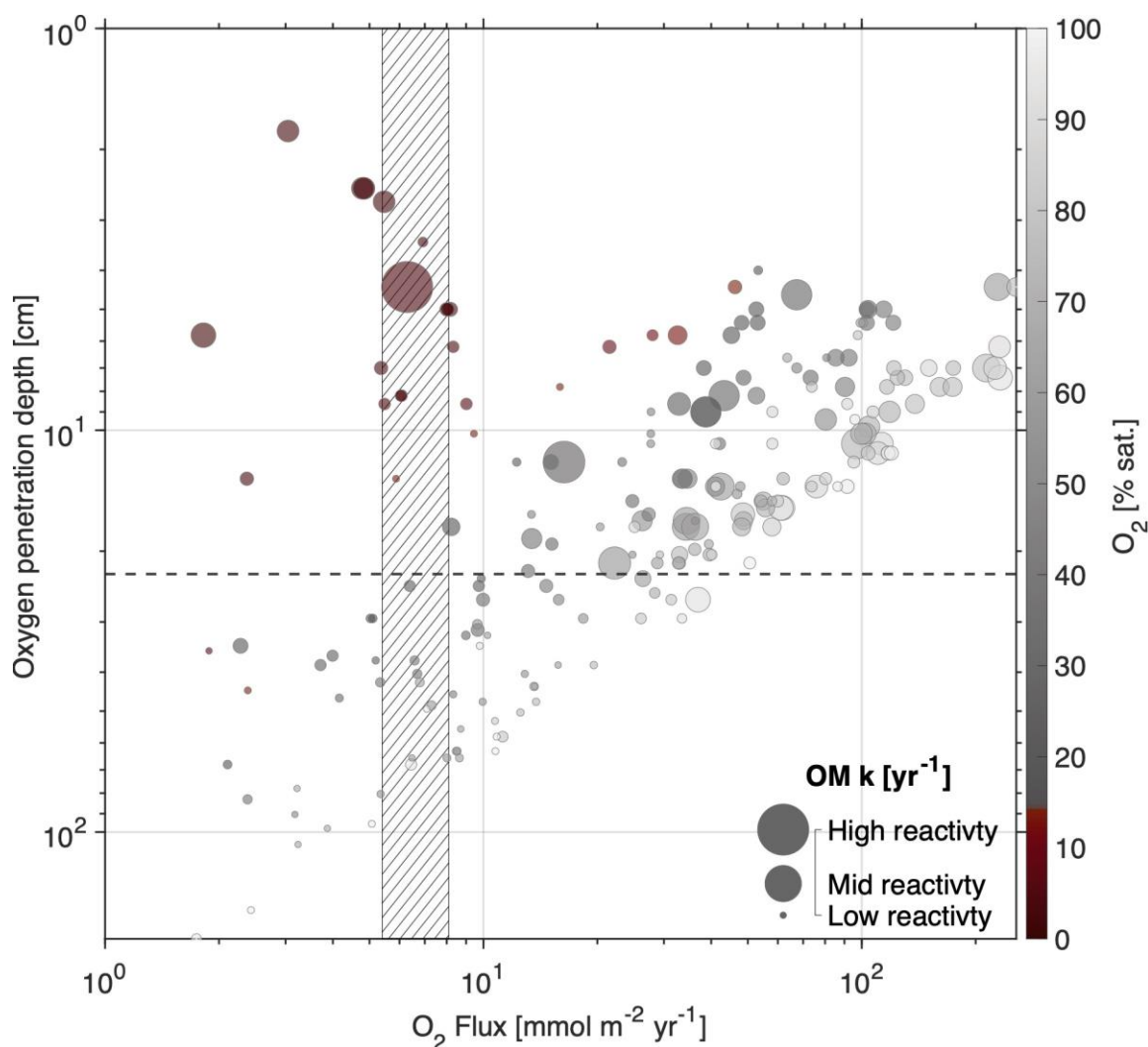


Figure 3: Modelled O_2 fluxes across the sediment-water/ice interface versus O_2 penetration depths (OPD) in GrIS subglacial sediments. Higher O_2 fluxes indicate higher microbial activity (heterotrophic respiration and/or CH_4 oxidation), reducing O_2



penetration. Anoxic conditions ($O_2 < 15\%$ saturation, shaded in red which equates to $50 \mu M$) are associated with shallower O_2 penetration and increased OM reactivity (OM k) (circle size). The hatched vertical bar indicates baseline O_2 flux delivered by basal melting (see sec. 3.6.5). The dashed line is the average OPD of 22.8 cm.

4.2 CH_4 production in subglacial sediment

The extensive anoxic zones within the GrIS subglacial sediment create favorable conditions for methanogenesis. Model results reveal CH_4 production rates range from 0.6×10^{-10} to 1.1×10^{-5} mol g.d.w⁻¹ yr⁻¹ with a mean of $(0.61 \pm 8.0) \times 10^{-7}$ mol g.d.w⁻¹ yr⁻¹ (mean \pm SD). These rates fit the range of rates measured in incubation experiments with subglacial sediment from Greenland ($(4.5 \pm 3.7) \times 10^{-10}$ mol g.d.w⁻¹ yr⁻¹ (mean \pm SD); Stibal et al., 2012), Canadian Arctic ($(2.3 \pm 3.3) \times 10^{-8}$ mol g.d.w⁻¹ yr⁻¹ (mean \pm SD); Boyd et al., 2010; Stibal et al., 2012), and Antarctica ($(2.9 \pm 2.6) \times 10^{-7}$ mol g.d.w⁻¹ yr⁻¹ (mean \pm SD); Stibal et al., 2012). Fig. 5 shows the comparison between modelled and measured rates across different studies and cryospheric environments. The highest modelled production rates are comparable to rates measured in incubation experiments of Arctic soil/sediments with much higher OM concentrations (up to 30 wt%) than those assumed here (Yedoma lakes; 3.56×10^{-4} mol g.d.w⁻¹ yr⁻¹; De Jong et al., 2018, and permafrost peat soils; $5.0 \pm 4.0) \times 10^{-6}$ mol g.d.w⁻¹ yr⁻¹; Kotsyurbenko et al., 2004). Results from the GSA suggest that CH_4 production rate is largely controlled by O_2 concentration, with OM content and reactivity as secondary drivers.

The model suggests that CH_4 saturation of porewaters is rarely reached and CH_4 remains thus primarily dissolved in porewater, which can diffuse upwards and become available for other microbial processes. However, immobile gas hydrates, which are a function of pressure (ice thickness above the sediments), temperature, and salinity, are present in approximately a third of our model results, formed once CH_4 exceeds saturation (50 mM; see methods), thus making up a large CH_4 pool inaccessible to microbial degradation. Depth-integrating the CH_4 production rate yields a range from 0.1 to 1600 mmol m⁻² yr⁻¹, with a mean of 73 ± 136 mmol m⁻² yr⁻¹ (mean \pm SD). Results from the GSA indicate that depth-integrated CH_4 production rates are controlled by OM reactivity, subglacial sediment depth, and OM concentration (in this order, see Fig. B1). The primary driver, OM reactivity, directly controls (together with OM concentration) the rate at which OM is transformed into CH_4 (see reaction r2 in table C1). Sediment depth indirectly influences the depth-integrated methanogenesis rate by increasing the overall thickness of the methanogenic zone; so, a thicker sediment column contains a larger anoxic zone, leading to greater total CH_4 production. Additionally, subglacial sediments less than two meters deep show a positive correlation between sediment depth and depth-integrated CH_4 production rate. However, this relationship breaks down in deeper sediments (see dotted line in Fig. 5), and the drivers (i.e., OM concentration and reactivity) gain more influence. Finally, our GSA ranks k_{AcOM} and O_2 as the weakest controls on subglacial depth-integrated CH_4 production rate.

4.3 CH_4 oxidation in subglacial sediment

AeOM is generally restricted to the uppermost tens of centimeters of sediment, where O_2 is available (i.e. above the OPD). Simulated AeOM rates span over several orders of magnitude, ranging from 1.5×10^{-18} to 3.4×10^{-6} mol cm⁻³ yr⁻¹, with mean



values of $4.4 \pm 6.2 \times 10^{-7}$ (mean \pm SD) $\text{mol cm}^{-3} \text{yr}^{-1}$ for the model ensemble. Our simulated mean rates compare well to the measured rates in incubation experiments using water samples from a subglacial outflow in Greenland ($1.2 \times 10^{-7} \text{mol cm}^{-3} \text{yr}^{-1}$; Dierer et al., 2014), but are two orders of magnitude higher than AeOM measured in Antarctic subglacial lake sediments ($3.07 \times 10^{-9} \text{mol cm}^{-3} \text{yr}^{-1}$; Michaud et al., 2017).

AeOM rates and AeOM efficiencies (i.e. ratio of depth-integrated AeOM rate to depth-integrated methanogenesis rate) are primarily controlled by OM reactivity and k_{AeOM} ; O_2 and OM contents also play a significant yet secondary role. An increase in k_{AeOM} narrows the CH_4/O_2 transition zone and shifts closer to the surface, resulting in a shallower OPD.

Across the GSA ensemble, AeOM efficiency is highly variable, ranging from near-zero to 100 %, with a mean of 51 %. AeOM efficiency is negatively correlated with OM content and its reactivity, because the rapid O_2 consumption via heterotrophic respiration reduces O_2 availability for AeOM. Conversely, k_{AeOM} and O_2 concentration are positively correlated, with k_{AeOM} exerting primary control over the oxidation efficiency. Therefore, high oxidation efficiencies within sediments (>90 %), such as those observed in Antarctic subglacial lakes (Michaud et al., 2017), are only attainable when OM reactivity is low ($v < 0.06$), k_{AeOM} is high ($> 10^{8.8} \text{cm}^3 \text{mol}^{-1} \text{yr}^{-1}$), and under prevailing oxic conditions ($\text{O}_2 > 50 \mu\text{M}$, above 15 % sat.). Sediment depth is not a primary driver of oxidation efficiency. However, across our GSA most boundary conditions lead to oxidation efficiencies much lower than the 90 % of the depth-integrated subglacial CH_4 production rate. This is particularly true for subglacial sediment characterized by low O_2 concentrations ($\text{O}_2 < 50 \mu\text{M}$) in overlying subglacial waters, but also for thick sediments ($> 5 \text{m}$) with high contents of comparatively reactive OM ($v \sim 0.2$). Therefore, our results indicate that GrIS subglacial sediment tends to act as a weak filter for diffusive CH_4 fluxes, and that even sediments underlying fully oxygenated waters can thus be a source of CH_4 to the subglacial drainage system, unlike for what has been observed in Antarctic subglacial lakes (Michaud et al., 2017).

4.4 Diffusive CH_4 fluxes from subglacial sediment

The imbalance between CH_4 production and consumption results in a net flux across the sediment-water/ice interface into the subglacial hydrological system, where it dissolves in meltwater (Fig. 4). Simulated fluxes range between $[0 - 234.7] \text{mmol m}^{-2} \text{yr}^{-1}$ (Fig. 5) with mean fluxes of 33.7 ± 32.5 (mean \pm SD) $\text{mmol m}^{-2} \text{yr}^{-1}$. The majority of the modelled fluxes exceeds calculated estimates in SW Greenland, e.g. Isunnguata Sermia (IS) $0.0024 - 0.58 \text{mmol m}^{-2} \text{yr}^{-1}$ (Hatton et al., in review, 2025) and Leverett Glacier (LG) span $0.289 - 0.966 \text{mmol m}^{-2} \text{yr}^{-1}$ (Lamarche-Gagnon et al., 2019), but are comparable to those reported for Russell Glacier (RU) $25.7 - 51.5 \text{mmol m}^{-2} \text{yr}^{-1}$ (Dierer et al., 2014). Diffusive fluxes for IS and RU were calculated by normalizing reported laterally transported CH_4 by the glaciers' respective total catchment area. Our modelled diffusive fluxes cover observed fluxes across a variety of different environments from Antarctic subglacial lake at $6.8 \pm 1.8 \text{mmol m}^{-2} \text{yr}^{-1}$ (Michaud et al., 2017) to groundwater fluxes in proglacial lakes in southwest Greenland around $951.3 \pm 2123.8 \text{mmol m}^{-2} \text{yr}^{-1}$ (Olid et al., 2022). The difference between our modelled fluxes and those calculated using literature data reflects methodological differences (simple mass balance vs. RTM) and underscore the difficulties of extrapolating laterally transported CH_4 to area-normalized fluxes, as done here and in Lamarche-Gagnon et al. (2019). Along with a better spatial



coverage of subglacial soft sediments, the acquisition of subglacial sediment cores with depth-resolved concentrations of OM and TEA would provide critical constraints to resolve this discrepancy.

GSA identifies OM reactivity as the primary control on diffusive CH_4 fluxes. OM quantity and sediment depth exert weak positive influence, whereas k_{AeOM} and O_2 concentration show a weakly inhibitory effect. Statistical analysis further reveals that the diffusive CH_4 flux is shaped by the interaction of these drivers. For instance, OM quantity and reactivity act synergistically, driving particularly large diffusive fluxes when both are high, although k_{AeOM} consistently reduces this positive effect. Conversely, the negative effect of O_2 concentration increases with depth and in combination with k_{AeOM} , but is partially offset by high OM reactivity.

In sediments shallower than 2 m, depth is positively correlated with diffusive CH_4 flux (dotted line in Fig. 5). However, this trend disappears below the maximum OPD, where anoxic conditions promote methanogenesis and upward diffusion becomes unaffected by AeOM. Interestingly, even shallow sediments (<1 m) produce diffusive fluxes large enough to cross the oxic filter ($24.4 \text{ mmol m}^{-2} \text{ yr}^{-1}$), under conditions of moderate OM quantity (0.2 wt%) with very high reactivity ($\nu=1.15$) when k_{AeOM} is low ($10^{6.7} \text{ cm}^3 \text{ mol}^{-1} \text{ yr}^{-1}$), regardless of O_2 concentrations at the sediment-water/ice interface. However, most fluxes in shallow sediments (< 2 m) are lower (4.9 ± 8.1 (mean \pm SD) $\text{mmol m}^{-2} \text{ yr}^{-1}$) with low oxidation efficiencies. By contrast, sediments deeper than 2 m that contain OM of very low reactivity cannot produce sufficient CH_4 to overcome oxidative losses, thus limiting the CH_4 flux into subglacial meltwater. These results demonstrate that while sediment depth is relevant, OM remains the strongest driver of diffusive CH_4 flux.

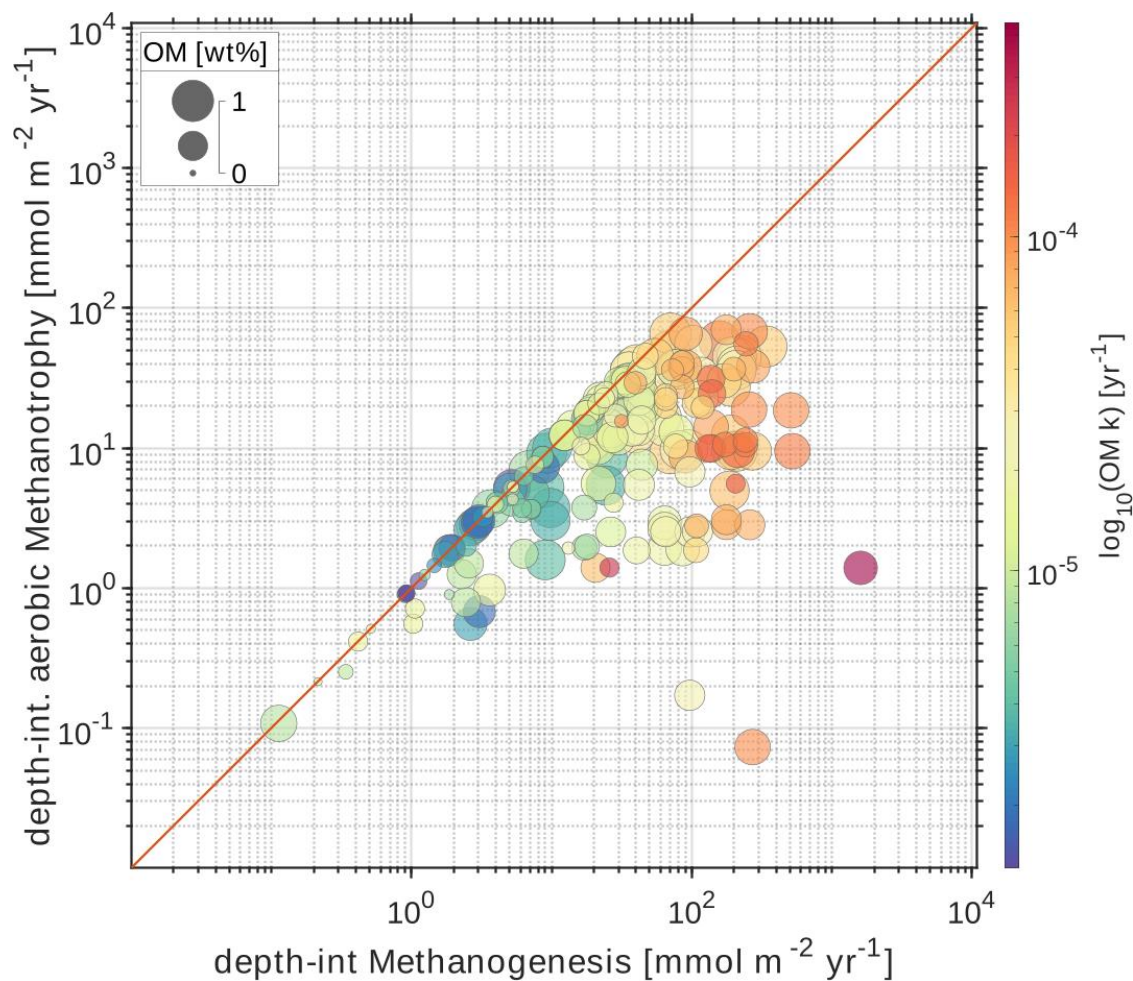


Figure 4: Scatter plot illustrating the relationship between CH_4 production and consumption, with bubble size representing OM content and color indicating reactivity (OM k). The 1:1 line indicates that depth-integrated CH_4 production rates equal consumption rates, which prevents diffusive flux across the sediment-water interface.

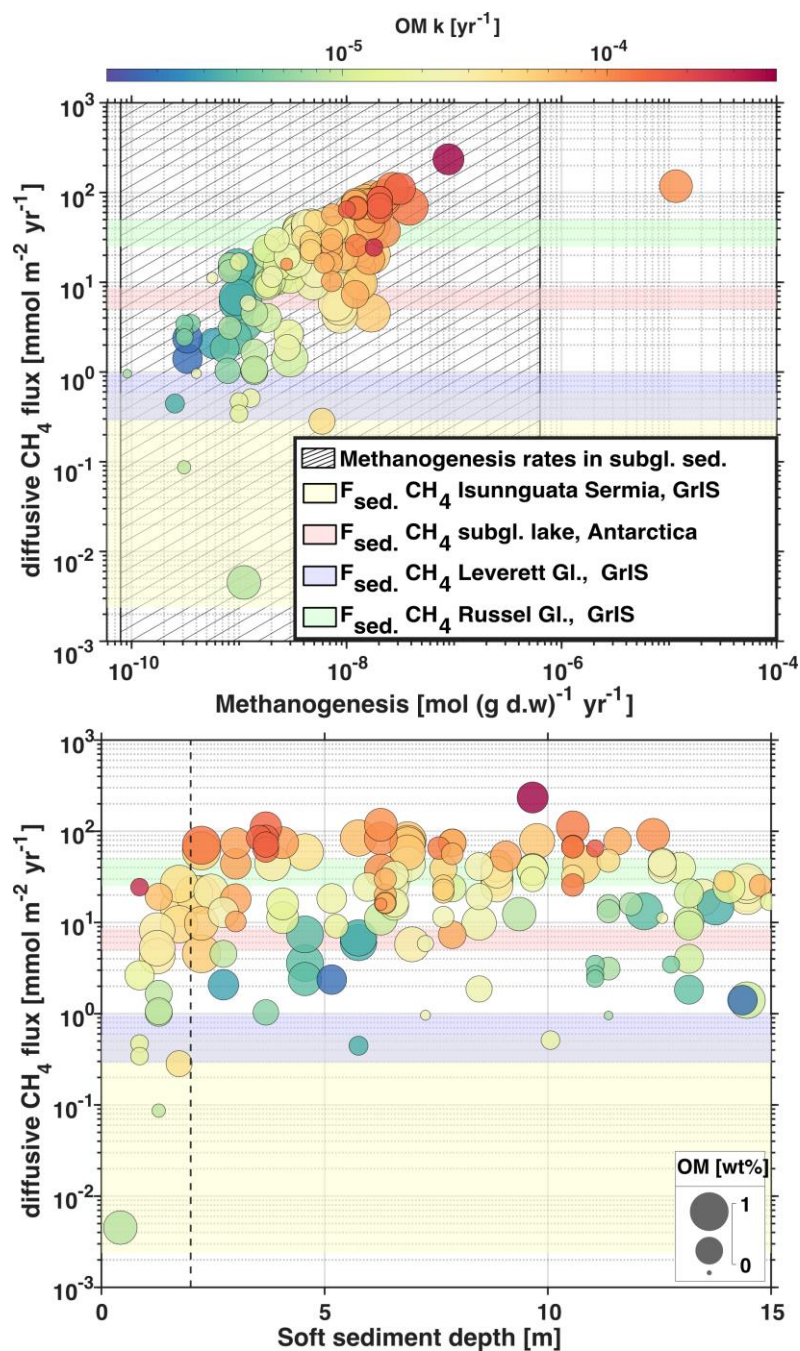


Figure 5: Modelled results for diffusive CH_4 flux out of the subglacial sediment as functions of sediment depth (left panel) and rate of methanogenesis (right panel). Left: Main drivers of simulated diffusive CH_4 flux from subglacial sediments into the subglacial environment. OM reactivity (denoted as OM k, circle color) is the strongest driver, followed by OM content (circle size), and sediment depth (x-axis). Sediment depth of less than 2 m drives CH_4 flux, whereas deeper sediments exert no significant influence.



Right: Simulated methanogenesis rates driving diffusive CH₄ flux. Simulated rates are compared to measured rates and fluxes through the sediment-water interface (SWI) from various cryospheric environments. The grey shaded area is the range in measured methanogenesis rates from incubation experiments from glaciers in Greenland, Antarctica, and Canada (Boyd et al., 2010; Stibal et al., 2012). Horizontal colored areas show comparative diffusive fluxes from sediments in a subglacial lake in Antarctica (Michaud et al., 2017), and extrapolated/calculated area yield fluxes (i.e. lateral export divided by reported catchment area) in GrIS at Isunnguata Sermia (Hatton et al., in review, 2025), Leverett Glacier (Lamarche-Gagnon et al., 2019), and Russell Glacier (Dieser et al., 2014).

4.5 Inferring subglacial environmental conditions: A case study on Isunnguata Sermia and Leverett Glacier (SW Greenland)

The comparison of simulated and measured CH₄ concentrations in subglacial meltwater (CH_{4(aq)}) allows inferring plausible combinations of subglacial environmental conditions within the catchment areas, able to sustain observed lateral CH₄ fluxes from two well-studied outlets of the SW GrIS (IS and LG). To this end, we estimate the potential lateral CH₄ fluxes by combining modelled diffusive sediment fluxes with hydrological scaling, oxidation, and off-gassing rates.

IS and LG drain catchments of approximately 14,000 km² and 1200 km², respectively (Mankoff et al., 2020). However, the hydrologically active subglacial areas are likely much smaller, yet critical for upscaling and estimating lateral fluxes. For instance, LG is thought to have a hydrologically active subglacial area approximately half its full catchment size (Cowton et al., 2012; Hawkings et al., 2021). Because glacier surface slope drives the hydraulic gradient, the surface catchment provides a first-order approximation of subglacial meltwater routing (Palmer et al., 2011; Cowton et al., 2012). Based on this approach, Cowton et al. (2012) put the active area at 600 km² at LG, while we estimated ~1300 km² at IS. However, the extent to which these areas are underlain by soft sediment remains unknown.

We then spatially upscale the simulated sediment CH₄ fluxes (0 - 234.7 mmol m⁻² yr⁻¹) to fractions of the hydrologically active and total area assumed to be underlain by subglacial sediments (see Fig. 6 a-b). Under these assumptions, IS could produce lateral fluxes of 0.03 - 3.3 × 10⁹ mol CH₄ (0.39 - 39.5 Gg) per year, while LG 0.0028 - 0.27 × 10⁹ mol CH₄ (0.0339–3.4 Gg) per year, and are comparable to lateral fluxes in the proglacial rivers at both sites of 0.002–28.9 Mg per year (IS; Hatton et al. in review) and 2.8 - 9.3 Mg per year (LG, Lamarche-Gagnon et al., 2019). Multiplying the lateral flux with annual discharge (IS: 5.5 ± 2.4 km³ yr⁻¹; LG: 1.4 ± 0.32 km³ yr⁻¹; see Mankoff et al., 2020), results in average dissolved CH₄ concentrations in meltwaters over a season ranging between 0.0001 - 593.4 μM for IS and 0.0003 - 200.2 μM for LG. These results suggest that dissolved CH₄ concentrations in meltwater are similar despite the generally large difference in catchment size.

Depending on the soft sediment cover present in the hydrologically active catchment area, the simulated CH_{4(aq)} may exceed, match, or fall below measured CH_{4(aq)} in proglacial rivers at the glacier outlet (see grey shading in Fig. 6 a-b; IS: 0.006–0.13 μM (Hatton et al., in review, 2025); LG: 0.036 - 0.7 μM (Lamarche-Gagnon et al., 2019)). In a few cases, the simulated CH_{4(aq)} falls below the measured CH_{4(aq)}, meaning such combinations of OM, sedimentary characteristics, and sediment cover are unlikely to result in measurable CH_{4(aq)} at the glacier margin. However, in cases where the simulated CH_{4(aq)} fall within the observed range, we postulate that no or minimal CH₄ oxidation or off-gassing occurs during lateral transport in the subglacial



hydrological system. Fig. 6 a-b shows that even a low subglacial sediment coverage (i.e., < 1% and <10% of the hydrologically active area), with a modest concentration (~0.5 wt%) of relatively unreactive ($\nu > 0.02$) OM supports observed CH_4 concentrations. In contrast, exceeding observed concentrations requires OM with either higher OM content (~0.65 wt%) and/or higher reactivity ($\nu > 0.14$), which in turn implies a loss of CH_4 before reaching the glacier outlet, either by in-stream oxidation or off-gassing. If we instead assume a 10% coverage of subglacial sediment, 78% for IS and 56% for LG of simulated $\text{CH}_{4(\text{aq})}$ exceed observed $\text{CH}_{4(\text{aq})}$, suggesting additional removal processes. Simulated $\text{CH}_{4(\text{aq})}$ exceeding measured ranges (IS: 0.13 - 6 μM , LG: 0.7 - 10 μM) imply substantial subglacial in-transit losses, possibly through microbial oxidation and/or off-gassing. Fig. 6 c-d highlights the necessary OM concentration and reactivity to produce such excess $\text{CH}_{4(\text{aq})}$ (circles with white border). Conversely, some combinations of boundary conditions result in $\text{CH}_{4(\text{aq})}$ within the measured range (circles with blue border), indicating no or minimal loss during lateral transport to the glacier outlet. Finally, if sediment covers more than 50% of the active hydrological area, almost all estimated concentrations (even for low OM and low reactivity) exceed observed concentrations.

These findings highlight that even limited subglacial sediment coverage and low contents of relatively unreactive OM already suffice to support observed subglacial $\text{CH}_{4(\text{aq})}$. Because subglacial sediment cover is likely above 1%, they also suggest that a fraction of the CH_4 produced in subglacial sediments is either stored in the subglacial environment or is oxidized or evades during transport in the subglacial hydrological network. Estimating the potential extent of additional CH_4 oxidation in the subglacial hydrological network will further help delineate the subglacial sediment cover, OM content and reactivity required to support observed concentrations and fluxes.

To this end, we postulate that $\text{CH}_{4(\text{aq})}$ is rapidly consumed in the channel's sediments in a process similarly observed in proglacial streams (Lamarche-Gagnon et al., 2019; Strock et al., 2024). We use an oxidation rate reported from a nearby subglacial environment (0.32 $\mu\text{M day}^{-1}$; Dierer et al., 2014) to calculate how quickly the excess $\text{CH}_{4(\text{aq})}$ at IS (6 μM) and LG (10 μM) decreases to the observed levels (Fig. 6 e-f). Because $\text{CH}_{4(\text{aq})}$ oxidation is time-dependent, we use the meltwater residence time from LG, which ranges from hours near the outlet to several days further upstream (grey shading in Fig. 6 e-f; Chandler et al., 2013). We combine these assumptions to model to focus on modelled $\text{CH}_{4(\text{aq})}$ exceeding measured concentrations resulting from 10 % subglacial sediment cover under the hydrologically active area and simulate their O_2 -unconstrained oxidation during transit in the channelized drainage system. Fig. 6 e-f illustrates that $\text{CH}_{4(\text{aq})}$ are still above observed $\text{CH}_{4(\text{aq})}$ by the time the melt water exits the subglacial environment, suggesting that oxidation does not suffice as CH_4 sink within the subglacial system. To remain within the observed $\text{CH}_{4(\text{aq})}$ and the residence time of 72 hours, soft sediments under IS would have to produce fluxes ranging from 5.9 to 7.4 $\text{mmol m}^{-2} \text{yr}^{-1}$ and at LG 16.8 - 41.0 $\text{mmol m}^{-2} \text{yr}^{-1}$. In both catchments, sediments should be deep (9.2 ± 3.6 m) and have substantial OM concentrations (0.64 ± 0.27 wt%) of low reactivity ($\nu=0.15 \pm 0.1$ for IS and $\nu=0.08 \pm 0.12$). Higher OM reactivity ($\nu=0.22 \pm 0.22$ at LG and $\nu=0.37 \pm 0.3$ at LG) would yield $\text{CH}_{4(\text{aq})}$ concentrations exceeding measured values even after 72 hours of water residence time, suggesting out-gassing or higher oxidation rates are required.



539 The results point to several key implications for subglacial CH₄ cycling. First, CH₄ fluxes from subglacial sediments in SW
 540 GrIS may be substantially larger than previously calculated, with the spatial extent of soft sediments in hydrologically active
 541 zones acting as a critical control. The sensitivity of modelled CH_{4(aq)} to assumptions about sediment cover highlights the need
 542 for direct mapping of subglacial sediment beneath the margins of major GrIS catchments. Secondly, in-stream CH₄ loss
 543 depends strongly on the assumed oxidation rate, which may vary seasonally as methanotrophs adapt to substrate availability
 544 (i.e. due to seasonal changes in CH_{4(aq)} concentrations from changes in dilution from supraglacial meltwaters). Additionally,
 545 in-stream CH₄ loss via evasion through turbulent flow around the glacier outlet might substantially increase the required
 546 diffusive CH₄ flux and in turn the CH₄ production in subglacial sediments to match the observed range. Because we do not
 547 account for this process, our flux estimates are likely conservative. Third, hydrological constraints, through their control on
 548 water residence time, emerge as a first-order determinant of whether CH₄ is preserved or oxidized before reaching the margin.
 549 Fourth, while we do not currently account for off-gassing as meltwater travels within the unpressurised drainage system
 550 (Chandler et al., 2013), we recognize that it may represent a significant loss of CH₄ during lateral transport. We hypothesize
 551 that in glacier hydrological systems characterised by upwelling, where meltwater surges at the glacier front, i.e. Isunnguata
 552 Sermia, subglacial pressurization changes may drive and redirect evaded CH₄ towards marginal outlets where high(er) CH₄
 553 concentrations (CH_{4(aq)} = 5 µM) have been recorded (e.g. Christiansen and Jørgensen, 2018). As such, CH_{4(aq)} measurements
 554 at the upwelling might not fully capture the entirety of the subglacially produced CH₄. We hypothesize that (seasonal)
 555 hydrological pressure changes might reroute CH₄ to lateral/marginal outlets (e.g. Christiansen and Jørgensen, 2018) and
 556 therefore that lateral fluxes measured at the main glacial outlets and in proglacial rivers may underestimate the overall
 557 CH₄ fluxes.

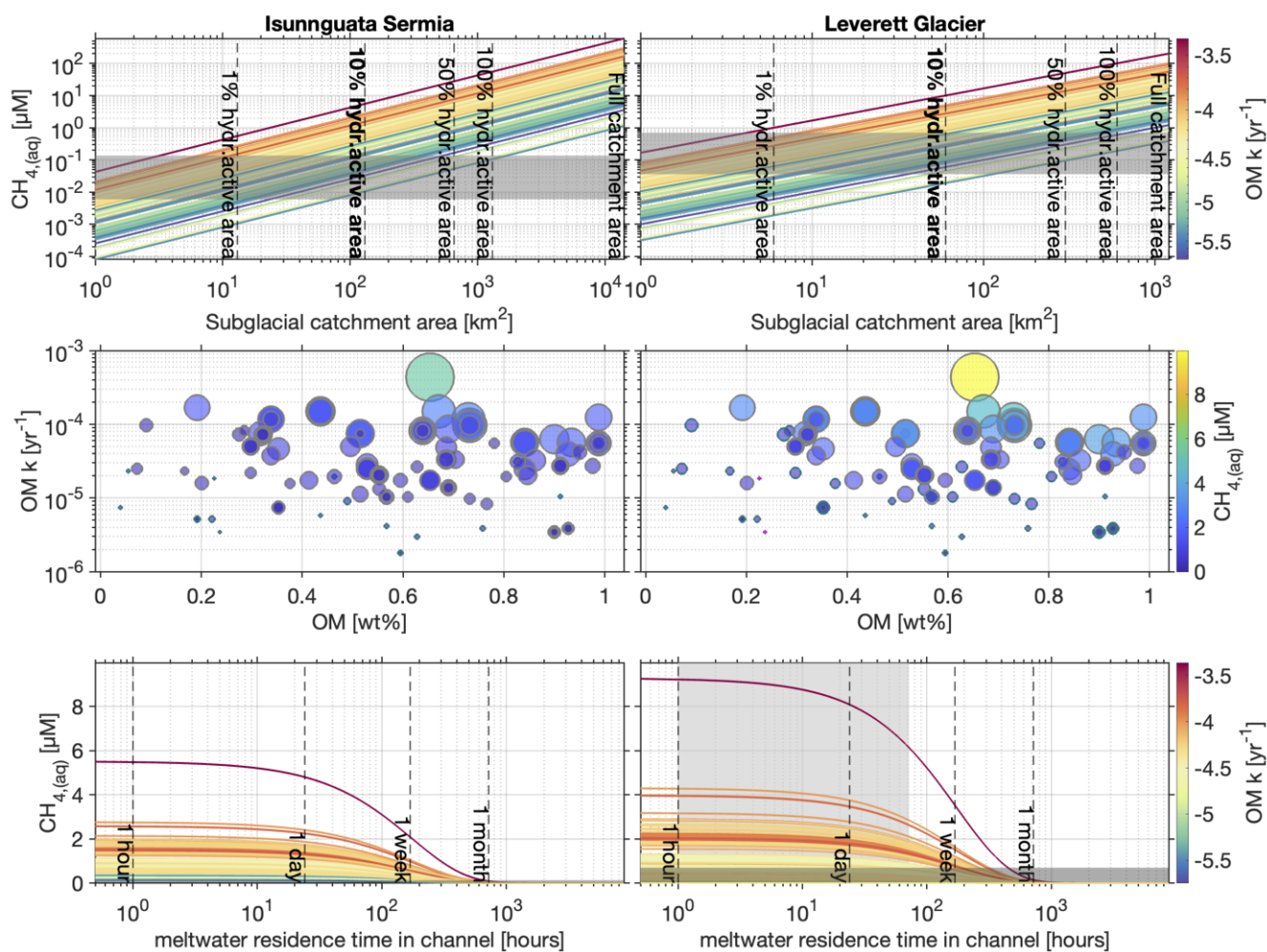


Figure 6: Case studies at Isunnguata Sermia and Leverett Glacier, Southwest Greenland, to estimate the fate of dissolved CH_4 once it enters the subglacial drainage system. Top row: in-stream CH_4 concentrations (discharge-weighted and extrapolated to increasingly hydrologically active catchment areas) leaving the glacier margin, its main drivers and its subsequent loss (oxidation) during lateral transport reaching the glacier outlet. Line color denotes OM reactivity. Grey areas are measured dissolved CH_4 concentrations in the proglacial rivers at the glacial outlets of IS (Hatton et al., in review, 2025), and LG (measurements at 1.5 km from the margin) (Lamarche-Gagnon et al., 2019). Middle row: subdivision of main drivers (OM concentration and its reactivity) by CH_4 concentration depending on whether it is above (grey-border circles), within (green-border circles) or below (magenta-border circles) site-specific measurements. Bottom row: melt water residence time vs. melt water CH_4 concentration as it decreases due to in-stream oxidation. Vertical light grey area indicates the range of residence times of meltwater at LG (Chandler et al., 2013), while horizontal dark grey area indicates the site-specific measurements.



5 Conclusion and future outlook

In this study, we used a reaction-transport model approach to conduct an environmental parameter sensitivity study. This approach established a quantitative framework that provides information on the magnitude and controls of subglacial CH₄ production and release and thus allows contextualizing measured CH₄ in meltwater outflow.

Our results indicate that subglacial sediments remain largely anoxic ($z > 23$ cm) even under fully O₂-saturated waters. Aerobic oxidation of methane (AeOM) efficiency—defined as the ratio of depth-integrated AeOM to methanogenesis—varies from nearly 0 to 100 % (mean = 51 %), reflecting its strong dependence on organic matter (OM) availability and reactivity. Sediments rich in highly reactive OM exhibit low AeOM efficiency, whereas less abundant or less reactive OM yields higher efficiency. Thus, sediments with limited or less reactive OM generate and release less CH₄ overall. When subglacial water O₂ concentrations drop below 50 μM (~15 % saturation), the OPD remains shallow (< 30 cm) regardless of sediment characteristics; above this threshold, OM reactivity becomes the dominant control.

The GSA identifies OM reactivity as the primary control on diffusive CH₄ fluxes. OM quantity and sediment depth exert a weak positive influence, whereas k_{AeOM} and O₂ concentration show a weakly inhibitory effect. Depth is positively correlated with diffusive CH₄ flux in sediments shallower than 2 m, but at greater depths, low-reactivity OM limits CH₄ production and prevents fluxes from overcoming oxidative losses. Thus, while sediment depth contributes to flux variability, OM reactivity ultimately dominates. These dynamics emphasize that CH₄ fluxes emerge from coupled biogeochemical controls rather than isolated parameters.

Nevertheless, our sensitivity study shows that nearly one-quarter of parameter combinations yield negligible CH₄ flux, even when conditions appear favorable (e.g. deep anoxic sediments). Our simulated sediment CH₄ fluxes range from 0 to 234.7 mmol m⁻² y⁻¹ (Fig. 5), with mean fluxes of 33.7 ± 32.5 mmol m⁻² y⁻¹ (mean ± SD). Most modeled fluxes exceed calculated estimates for southwestern Greenland, including Isunnguata Sermia (0.0024–0.58 mmol m⁻² y⁻¹; Hatton et al., in review, 2025) and Leverett Glacier (0.289–0.966 mmol m⁻² y⁻¹; Lamarche-Gagnon et al., 2019), but are comparable to values reported for Russell Glacier (25.7–51.5 mmol m⁻² y⁻¹; Dieser et al., 2014).

This work also reveals a critical biogeochemical trade-off governing net CH₄ export. Based on the case studies at Isunnguata Sermia and Leverett Glacier (SW GrIS), achieving previously reported CH₄ export (Lamarche-Gagnon et al., 2019; Hatton et al., in review, 2025) requires a narrow balance of conditions. While high OM reactivity drives methanogenesis, it also promotes rapid O₂ consumption via heterotrophic respiration (by offering an easily degradable source of substrate) and CH₄ oxidation. Consequently, simulations approximating measured CH₄ concentrations in meltwaters at glacier termini emerge from a very specific set of boundary conditions, i.e., sediment layers of 9 ± 3 m, moderate OM content (0.64 ± 0.27 wt%), and low OM reactivity ($v = 0.08$ – 0.15), permitting methanogenesis to outpace oxidation. In contrast, shallow sediments (< 1 m) are unlikely to be significant CH₄ sources, unless rich in highly reactive OM ($v > 1.0$).

Our analysis provides a first-order approximation by focusing on biogeochemical processes within the static sediment column. The model neglects several potentially important redox processes (iron and nitrate cycles), which likely influence CH₄ cycling.



602 Additionally, a more sophisticated formulation of the transport and transformation of CH₄ within the dynamic subglacial
603 hydrological system is a likely candidate for improvement. The model also neglects transient processes, including seasonal
604 hydrological flushing, sediment erosion, and changes in O₂ supply, which likely constitute an additional control over the CH₄
605 emissions. Such processes also differentiate the GrIS environments from Antarctic subglacial lakes, where stagnant, O₂-rich
606 waters sustain > 90% oxidation.

607 These findings reveal the critical consequences for subglacial exploration, indicating that drilling strategies should prioritize
608 areas with deep sediment to locate active methanogenic communities and their substrates. Additionally, future modelling work
609 would also greatly benefit from depth-resolved measurements of solid and dissolved species, such as OM and terminal electron
610 acceptors (e.g., sulfate and bioavailable iron) to expand our reaction network and, thus, our understanding of subglacial
611 biogeochemistry. And finally, future work should also integrate reactive transport with subglacial hydrology to capture CH₄
612 cycling and quantify its contribution to the global CH₄ budget.

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614
615 *Code and data availability.* Boundary conditions for the Global sensitivity analysis and model output Model outputs are
616 available from the Zenodo repository: <https://doi.org/10.5281/zenodo.17512276>

617
618 *Author contributions.* PA, MS and SA conceived the idea. PP built, implemented and performed model runs, created the
619 figures, and did the data curation. PP wrote the paper with guidance from MS and SA. MS and SA acquired funding. GLG and
620 PK provided input data and field expertise for the discussion. All authors reviewed and edited the paper.

621
622 *Competing interests.* The authors declare that they have no conflict of interest.

623
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Appendix A: Examples of depth-profiles of concentration and rate for relevant species/rates as model output
Depth 225 TOC 0.568 O₂ 243 nu 0.0411 AeOM 7.71

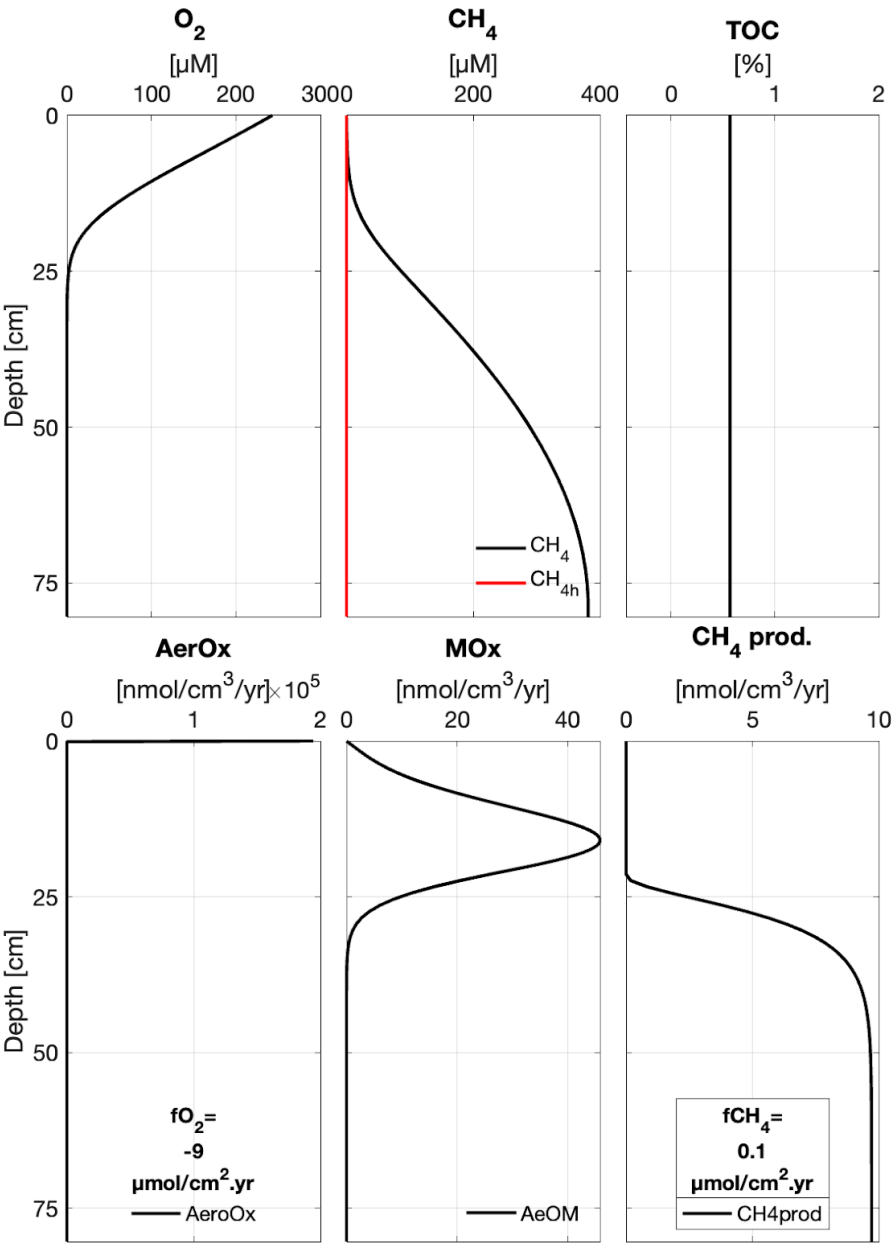


Figure A1: Depth-profiles for (constant) OM, for dissolved species and for porosity for shallow subglacial sediment depths (0.75m).
Top: concentration profiles for the key species. Bottom: rate profiles. Panel on right hand side is the superposition of the primary redox reaction rates normalized by their total contribution to OM degradation.



Depth 397 TOC 0.9 O₂ 238 nu 0.0137 AeOM 6.1

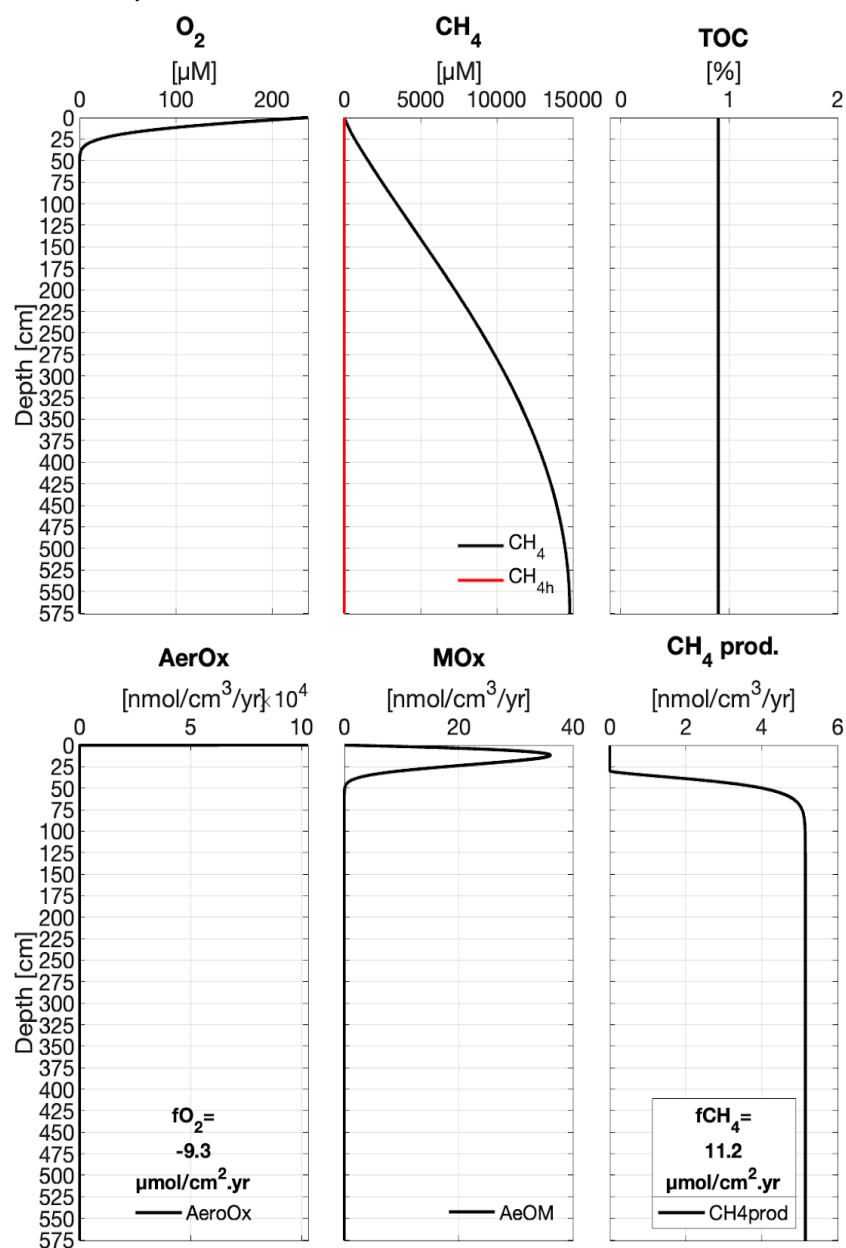


Figure A2: Depth-profiles for (constant) OM, for dissolved species and for porosity for shallow subglacial sediment depths (5.75 m).
Top: concentration profiles for the most important species. Bottom: rate profiles. Panel on right hand side is the superposition of
the primary redox reaction rates normalized by their total contribution to OM degradation.

Appendix B: Sensitivity analyses for diffusive CH₄ flux and oxygen penetration depth

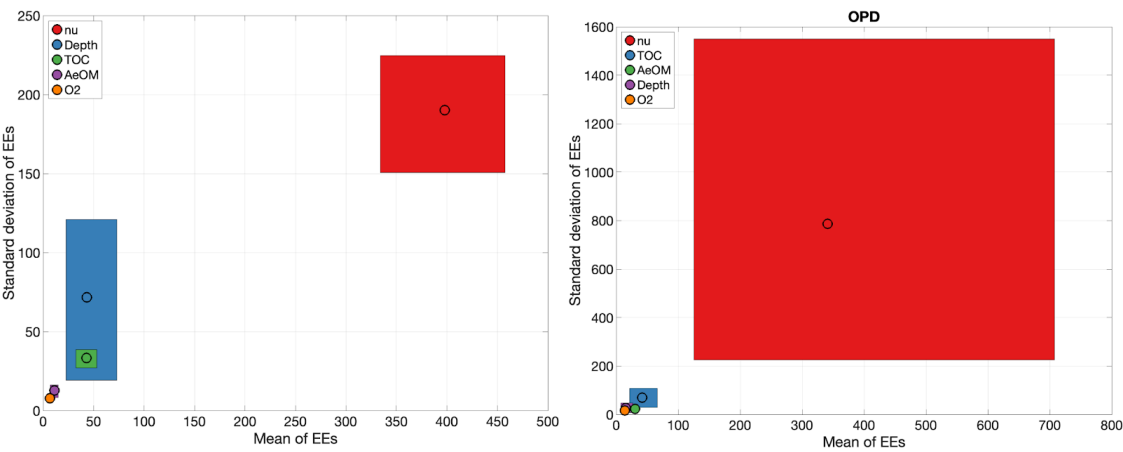


Figure B1: Sensitivity analyses for diffusive CH₄ flux out of the sediment (left panel) and for oxygen penetration depth into the sediment (right panel). Mean of Elementary Effects (EEs) versus their standard deviation for five key model parameters: $\nu = \nu$ is one of the two RCM reactivity parameters, depth represents the subglacial sediment depth, TOC denotes the OM quantity, AeOM is the rate constant for the aerobic oxidation of CH₄, and O₂ is the oxygen concentration at the SWI. Confidence bounds were derived via bootstrapping around the mean and standard deviation of the EEs.

Appendix C: Reaction network and list of symbols used for the model

Table C1: Reaction network governing heterotrophic organic matter degradation and CH₄ oxidation in subglacial sediments implemented in the Reaction-Transport Model (Adapted from Aguilera et al., 2005; Thullner et al., 2009).

| Reaction Pathway | Stoichiometry | Reaction rate |
|---|--|--|
| | Primary redox reactions | |
| Aerobic OM degradation (r1) | $(CH_2O)_x(NH_3)_y(H_3PO_4)_z + (x + 2y)O_2 \xrightarrow{+(y + 2z)HCO_3^-} (x + y + 2z)CO_2 + yNH_4^+ + zHPO_4^{2-} + (x + 2y + 2z)H_2O$ | $r1 = \frac{v}{(a + age)} \cdot CH_2O \cdot fO_2$ |
| Methanogenesis (r2) | $(CH_2O)_x(NH_3)_y(H_3PO_4)_z + (y - 2z)H_2O \xrightarrow{\frac{x - 2y + 4z}{2}CO_2 + (y - 2z)HCO_3^- + yNH_4^+ + zPO_4^{2-}} \frac{x}{2}CH_4$ | $r2 = \frac{v}{(a + age)} \cdot CH_2O \cdot fCH_4$ |
| | Secondary reaction | |
| CH ₄ oxidation by O ₂ | $CH_4 + 2O_2 \rightarrow CO_2 + 2H_2O$ | $r3 = kAeOM \cdot CH_4 \cdot O_2$ |
| CH ₄ dissolution | $kdis \begin{cases} [CH_4(h)]([CH_4] - [CH_4]^*) & \text{if } [CH_4] < [CH_4]^* \\ 0 & \text{if } [CH_4] > [CH_4]^* \end{cases}$ | |
| CH ₄ hydrate formation | $khyd \begin{cases} ([CH_4] - [CH_4]^*) & \text{if } [CH_4] > [CH_4]^* \\ 0 & \text{if } [CH_4] < [CH_4]^* \end{cases}$ | |

Table C2: Reaction parameter values implemented in the RTM. Effective molecular diffusion coefficients (D_i) are given for T = 0 °C and are corrected by subglacial environment specific temperature, salinity, and tortuosity. GSA input parameters are highlighted in **bold**.

| Parameter | Unit | Value | Reference |
|----------------------------------|------|-------------|---|
| Subglacial sediment depth | m | 1 - 15 | This study |
| x/y/z | – | 106/16/1 | Redfield (1934) |
| TOC | wt% | 0.02 — 1 | This study; Yde et al. (2010); Stibal et al. (2012) |
| RCM ν | – | 0.005 - 1.7 | Boudreau and Ruddick (1991) |
| RCM α | yr | 0.2 | Boudreau and Ruddick (1991) |
| age | yr | 4000 | Hatton et al. In review |



| | | | |
|-------------------------|-------------------------|------------------|--|
| O_2 | μM | 0 - 456 | This study |
| kAeOM | M yr | $10^6 - 10^{10}$ | Dieser et al. (2014); Murguia-Flores et al. (2018) |
| $D(O_2)$, μO_2 | $cm^2 yr$ | 380.45, 0.06 | Wang and Van Cappellen (1996) |
| $D(CH_4)$ $\mu CH_4(g)$ | $cm^2 yr$ | 5000.0, 0.0 | Wang and Van Cappellen (1996) |
| $[CH_4]^*$ | mM | 59.36 | This study; calculated for 600 m ice thickness |
| kdis | $cm^3 mol^{-1} yr^{-1}$ | 0 | Wang and Van Cappellen (1996) |
| khyd | yr^{-1} | 789.0 | Puglini et al. (2019) |

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672 References

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