

Spatial heterogeneity of GHG dynamics across an estuarine ecosystem

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Abstract.

Coastal ecosystems are critical components of the global carbon cycle, exerting a disproportionate influence on the carbon budget despite their limited spatial extent. Although they have gained more attention in the past decade, detailed studies of GHG dynamics across the land-ocean continuum, including shallow-water estuaries, remain relatively scarce even though they are active sources of the three most potent greenhouse gases (GHGs): carbon dioxide (CO₂), methane (CH₄), and nitrous oxide (N₂O). Such shallow coastal ecosystems are highly heterogeneous, shaped by strong physical, biogeochemical, and biological gradients. Together with the spatial variation in coastal biodiversity, these gradients significantly influence carbon cycling at both local and global levels. However, large uncertainties persist due to limited measurements, emphasizing the need for improved constraints on GHG budgets.

15 Surface seawater partial pressure of CO₂ (pCO₂), CH₄, and N₂O concentrations, along with seawater physical and biogeochemical properties, and air-sea gas exchange, were measured at 21 sites in southwest Finland (Baltic Sea). Sampling progressed from estuarine inner bays to the outer archipelago, covering diverse soft-sediment habitats, from sheltered to exposed areas, across a salinity gradient. Seawater pCO₂ and N₂O concentrations ranged from undersaturated (160 ppm and 9 nmol L⁻¹, respectively) to supersaturated (2521 ppm and 25 nmol L⁻¹, respectively), compared to the atmosphere, resulting in an uptake of -36 and -0.0021 mmol m⁻² d⁻¹, and a release up to 220 and 0.0383 mmol m⁻² d⁻¹, respectively. CH₄ concentrations were consistently supersaturated (19 to 469 nmol L⁻¹) compared to the atmosphere, resulting in a net source to the atmosphere from 0.014 to 1.39 mmol m⁻² d⁻¹.

Freshwater input from the Karjaanjoki River and its mixing with seawater shaped the overall spatial patterns of GHGs. However, deviations from this salinity-driven control were seen in sheltered sites within the archipelago, where elevated pCO₂ and CH₄ concentrations likely reflected biological processes, including enhanced organic matter respiration and methanogenesis in warm, late-summer shallow waters, where limited oxidation favoured CH₄ accumulation. At exposed and semi-sheltered sites, mixing processes exerted greater control, resulting in lower GHG concentrations. Our results show that both physical mixing and biological processes influence coastal GHG dynamics, with benthic ecosystems potentially playing a key but still poorly constrained role. The overall budget of air-sea GHG exchanges was dominated by CO₂ fluxes, with CH₄ consistently acting as a source, and N₂O alternating between source and sink. High environmental variability in shallow coastal

systems leads to strong fluctuations in the balance between GHG production and consumption, which needs to be considered when evaluating their role in the global carbon budget.

1. Introduction

Coastal ecosystems are increasingly recognized as critical components of the global carbon cycle (Bauer et al., 2013). Although covering only about 7% of the oceanic area, they play a disproportionately large role in biogeochemical cycling due to their location at the land–sea interface (Gattuso et al., 1998; Wollast, 1998). These environments act as hubs of exchange across biomes (Resplandy et al., 2024), contributing to the global oceanic uptake of anthropogenic carbon by absorbing carbon dioxide (CO₂) from the atmosphere and either burying, transforming, or releasing the carbon that land ecosystems deliver to coastal waters (Regnier et al., 2022). Due to intense inputs of nutrients and carbon from land and open ocean at continental margins, combined with high rates of biological production and degradation, coastal waters are among the most biogeochemically active regions of the biosphere (Wollast, 1998). Shallow coastal waters such as mangroves, salt marshes, and seagrass meadows effectively sequester large amounts of atmospheric carbon through vegetation growth and long-term sediment accumulation, making significant contributions to long-term carbon storage (Lovelock and Duarte, 2019; Macreadie et al., 2021; Mcleod et al., 2011). These so-called “blue carbon” ecosystems have been proposed as nature-based solutions for short-term climate change mitigation (Hoegh-Guldberg et al., 2019; Lovelock and Duarte, 2019).

Vegetated blue carbon ecosystems exemplify the value of healthy habitats in long-term carbon storage. Still, they cover a small part of the coastal oceans (thebluecarboninitiative.org), and measurements from other key habitats remain scarce and largely neglected in carbon cycling budgets (James et al., 2024). Estuarine areas are characterized by strong physical and chemical gradients and host transitional biodiversity that bridges terrestrial and marine ecosystems (Elliott and Whitfield, 2011) and are recognized as hot spots for carbon cycling and GHG exchange with the atmosphere (Borges et al., 2005; 2016; Cai et al., 2013; Frankignoulle et al., 1998; Humborg et al., 2019; Rosentreter et al., 2021a). The global oceans absorb roughly 25% of anthropogenic CO₂ emissions annually (Watson et al., 2020). While coastal waters contribute to this sink by taking up atmospheric CO₂ (Chen and Borges, 2009; Regnier et al., 2022), many nearshore systems like estuaries have been recognized as net heterotrophic ecosystems with negative net ecosystem production (Gattuso et al., 1998; Testa et al., 2012), leading to the production and release of both CO₂ and methane (CH₄) (Abril and Borges, 2005; Bonaglia et al., 2025; Borges et al., 2018). Despite their small size, emissions from near-shore ecosystems such as estuaries could nearly balance the carbon uptake by marginal seas (Borges et al., 2005; Chen and Borges, 2009). Global CO₂ emissions from estuaries range from 0.1 to 0.6 PgC per year (Borges et al., 2018), an amount equivalent to 5-30% of the oceanic CO₂ sink of ~ 2 PgC per year (Le Quéré et al., 2016), while CH₄ emissions range from 1 to 7 Tg per year, mainly driven by organic matter accumulation, anaerobic decomposition in sediments, and production in submerged plants (Borges et al., 2018; Rosentreter et al., 2018). However, CH₄ emissions are probably underestimated due to difficulties in accounting for ebullition and gas flaring (Borges et al., 2016; Humborg et al., 2019). The complexity of these pathways, combined with a lack of systematic, high-resolution, and long-term

measurements, hampers robust global assessments of coastal CH₄ emissions (Roth et al., 2022). Besides CO₂ and CH₄, coastal ecosystems are important but poorly constrained sources of nitrous oxide (N₂O), a potent greenhouse gas with a global warming potential 273 times higher than CO₂ over 100 years (IPCC 2023). N₂O is primarily produced via nitrification (ammonia oxidation) and denitrification (nitrate reduction) in sediments and water columns and is controlled by the availability of dissolved inorganic nitrogen and oxygen (Bange 2006). Eutrophication and hypoxia, often driven by excess nutrient and organic matter inputs, have been shown to promote N₂O emissions (Murray et al., 2015; Brase et al., 2017). Coastal ecosystems are recognized as significant sources of N₂O to the atmosphere (Bange, 2006; Cheung et al., 2025; Resplandy et al., 2024), where denitrification, especially in sediments and on particles, often dominates N₂O production, even in well-oxygenated waters (Wan et al., 2023). However, coastal estimates remain uncertain due to sparse measurements and high spatial heterogeneity (Wan et al., 2023). Given the strong warming potential of N₂O, understanding its fluxes from estuaries is essential for a complete picture of coastal greenhouse gas dynamics and for determining how these emissions, together with those of CH₄ (with a warming potential of 27; IPCC 2023), offset the coastal uptake of CO₂ (Resplandy et al., 2024; Rosentreter et al., 2018; Roth et al., 2023). Such offsets directly influence the net climate mitigation potential of coastal ecosystems, thereby questioning their ability to mitigate rising atmospheric GHG concentrations and associated radiative effects (Kristensen et al., 2025; William and Gattuso, 2022).

We investigate the dynamics of CO₂, CH₄, and N₂O (GHGs) along a salinity gradient and across contrasting coastal habitats within an estuary. We combined detailed field measurements of surface seawater physical and biogeochemical properties with both in situ measurements and calculated estimates of air–sea GHG exchange. This approach enabled us to capture spatial variability and examine the roles of physical drivers and biological processes in the observed changes in GHG concentrations. In doing so, we aim to quantify the contribution of the studied coastal ecosystems to GHG emissions and to provide additional data to improve both regional and global estimates of GHG emissions from the coastal ocean.

2. Materials and methods

2.1 Study area

This study was conducted within a 30 km radius of the Tvärminne Zoological Station (TZS), located at the southwestern tip of mainland Finland. The study area encompasses the coastal waters extending from the cities of Hanko, Tammisaari, and Pohja (Fig. 1), where the coastal environment is characterized by strong spatial heterogeneity due to the intricate bathymetry and geomorphology of the archipelago system (Asmala and Scheinin, 2024) and enhanced human impact. The studied area receives substantial freshwater from the Karjaanjoki river (in Pohja, runoff 0.59 km³ yr⁻¹, Räike et al., 2012), delivering relatively large amounts of allochthonous carbon and nutrients to the system, resulting in eutrophication and elevated concentrations of organic carbon in the water column across the inner archipelago area (Fleming-Lehtinen et al., 2015). Sampling sites (N = 21) were selected to encompass a wide range of soft-sediment habitats (e.g., including both vegetated and non-vegetated sediments with grain size ranging from coarse sand to clay, silt, and mud) and to represent a spatial gradient (50

95 km) from the outer to innermost archipelago. We used the classification by Lammerant et al. (2025), who categorize sampling sites based on their salinity and sediment characteristics to indicate the degree of exposure to environmental forces. Exposed sites (sites 1–4 and 17–21) are likely to experience stronger wave, wind, and current energy, as well as higher water exchange, compared to sheltered sites. Sites within Pojo Bay (sites 1–4) are broadly open to the bay and are directly influenced by wind and water input from the Karjaajoki River and the archipelago. Therefore, those are included with the exposed sites. The
100 sheltered sites (sites 5 and 7–11) are more enclosed and likely to have limited water circulation. Semi-sheltered sites fall between exposed and sheltered environments (sites 6 and 12–16).

Exposed and semi-sheltered sites in the outer archipelago (between Hanko and TZS, sites n°14–21, Fig. 1) are characterized by sandy sediments (96.2–100% sand) with low organic matter content (0.3–1.3%). These sites host both marine and freshwater plant species (Gustafsson and Norkko, 2019; Lammerant et al., 2025), and their faunal biomass is dominated by bivalves and
105 polychaetes (Gammal et al., 2019; Mäkelin et al., 2024). Sheltered and semi-sheltered sites within the archipelago (sites n°5–13) are characterized by sediments containing high proportions of clay/silt (20.8–64.3%) and organic matter (2.4–13.2%; Lammerant et al., 2025). At these sites, fast-growing fresh- and brackish-water macrophytes are often found, while faunal biomass is dominated by gastropods (Mäkelin et al., 2024). In Pojoviken Bay, the 14 km inlet between Tammissaari and Pohja (sites n°1–4), dense vegetation occurs down to ~2 m depth but becomes sparse at greater depths (>3 m) due to light limitation
110 (Lammerant et al., 2025). Deeper sites are characterized by high organic matter content (>13%) in the sediment and low faunal biomass (Gammal et al., 2019). Sampling was conducted from 14 August to 12 September 2023. Sites were visited once during the study period, with sampling conducted every two to three days, depending on weather conditions and boat availability.

2.1 Sampling procedure

Physical and biogeochemical properties of surface seawater, including the partial pressure of dissolved CO₂ (pCO₂) and CH₄
115 concentration, were measured *in situ* using a custom-built flow-through system. Seawater, from a depth of 50 cm, was pumped onboard using a submersible pump and directed into two separate flow-through systems.

The first water flow (3 L min⁻¹) was dedicated to gas extraction using a double showerhead equilibrator (Sunburst) equipped with a temperature probe (Hex fitting thermistor, ThermX, 0.1°C accuracy) and a barometric pressure sensor (BARO-A-4V, All Sensors). From the equilibrator, a continuous air flow (2 L min⁻¹) circulated in a closed loop to an infrared gas analyser
120 (IRGA, LI-COR, LI-7810). The analyser measured the dry mole fractions of CO₂ and CH₄, denoted as xCO₂ and xCH₄, respectively, as the gas passed through a Peltier cooler to remove the excess water before entering the IRGA. Both gas concentration and temperature were recorded every second until an equilibrium was reached. Equilibration between the seawater and gas phases was monitored in real time using a laptop connected to the IRGA. Equilibrium was considered reached when both CO₂ and CH₄ concentrations stabilized at a clear plateau. Depending on the concentration gradient between sites,
125 this equilibration period could vary substantially. While CO₂ typically reached equilibrium within a few minutes, CH₄ required longer times (up to 45 minutes) to reach a stable plateau. The measured xCO₂ (in ppm) was later converted into pCO₂ (in µatm) and corrected for *in situ* temperature using the Matlab CO2SYS v3 (Sharp et al., 2023). Surface seawater CH₄ concentration

(in nmol L^{-1}) was computed from $x\text{CH}_4$ (in ppm), *in situ* temperature, salinity, and the solubility coefficients from Wiesenburg and Guinasso (1979). The IRGA was factory-calibrated, and standard gases for CO_2 (150, 420, and 1500 ppm) and CH_4 (1, 20, and 150 ppm) were passed through the IRGA both before and after deployment. The data were corrected for potential drift. The second water flow (2 L min^{-1}) was directed through a thermosalinograph (SeaBird, TSG045), a fluorometer for chlorophyll *a* (chl-*a*), turbidity, and phycocyanin concentrations (Chelsea, TriLux), a fluorometer for coloured dissolved organic matter (CDOM, Chelsea, UviLux), and an optode for dissolved oxygen concentration and saturation (Aanderaa 4531). Each sensor was factory-calibrated before deployment, and data were logged at 1-second intervals until equilibrium for $x\text{CO}_2$ and $x\text{CH}_4$ was reached (up to 45 min). A 5-second average was then recorded.

Discrete surface water samples were collected for total alkalinity (TA), dissolved inorganic carbon (DIC), methane (CH_4), and nitrous oxide (N_2O) concentration using a peristaltic pump (Cole Palmer, Masterflex Environmental Sampler) equipped with Tygon tubing. Seawater samples were collected into 12 mL gas-tight vials (Exetainers, Labco High Wycombe, UK) for DIC, 60 mL borosilicate vials for TA, and 60 mL serum bottles for dissolved CH_4 and N_2O concentrations. Samples were preserved by adding 1% of the sample volume of a saturated mercuric chloride (HgCl_2) solution. Samples were stored in the dark at room temperature until analysis.

Air-sea exchanges of CO_2 and CH_4 were measured using the accumulation chamber technique (Frankignoulle, 1988). The chamber consists of a polyethylene container (internal diameter: 34 cm, height: 14 cm, total volume = 11.4 L) connected in a closed loop to the IRGA with 1.5m long Tygon tubing, which allows the chamber to move freely on the water surface. The air partial pressure of CO_2 ($p\text{CO}_2$) and CH_4 ($p\text{CH}_4$) within the chamber was recorded every second for 10 minutes. The flux was computed from the slope of the linear regression of $p\text{CO}_2$ and $p\text{CH}_4$ against time ($R^2 > 0.99$) following the method of Frankignoulle (1988), accounting for the air volume enclosed within the chamber. The uncertainty of the flux calculation, based on the standard error on the regression slope, was approximately 3% on average. Before conducting the chamber measurements, the IRGA baseline was established by flushing atmospheric air through the analyser. Based on these measurements, we estimated average mixing ratios of 406 ppm for CO_2 and 2.05 ppm for CH_4 during the survey. Reported fluxes represent the mean of two to three individual chamber measurements performed at each site. Direct measurements of N_2O air-sea exchanges were not possible during our survey. Therefore, the air-sea exchange of N_2O , as well as CO_2 and CH_4 (i.e., F_X , where X represents N_2O , CO_2 , or CH_4 , respectively) was parameterized using the bulk formula:

$$F_X = kK_{0(X)}(pX_{(sw)} - pX_{(atm)}) \quad (1)$$

where k (m s^{-1}) is the gas transfer velocity, K_0 ($\text{mol m}^{-3} \text{ atm}^{-1}$) is the temperature and salinity dependent gas solubility determined from Weiss (1974) for CO_2 , Wiesenburg and Guinasso (1979) for CH_4 , and Weiss and Price (1980) for N_2O , and $pX_{(sw)}$ and $pX_{(atm)}$ are the measured partial pressure of gas in the surface seawater and the air, respectively.

This approach strongly relies on selecting an appropriate parameterization for the gas transfer velocity, k . Numerous theoretical, laboratory, and field studies established that k depends on a variety of parameters, including capillary and breaking waves, boundary layer stability, air bubbles, surfactant surface films, evaporation and condensation, precipitation, water currents (including tides), as well as turbulence at the air-water interface (Borges et al., 2004). The parameterization of k is

most often expressed as a function of wind speed. This relationship is well established for the open ocean, where wind stress is the main source of turbulence (e.g., Wanninkhof, 2014). In estuarine environments, significant regression functions between k and wind speed have also been reported (Borges et al., 2004; Raymond et al., 2000). However, such formulations have been
165 proven to be site-specific and may not fully account for other controlling processes, such as turbulence at the air-water interface (Borges et al., 2004). Since no k -wind relationship has been developed for our study area, we adopted the formulation proposed for the Randers fjord by Borges et al. (2004), as its physical characteristics, particularly tidal amplitude and freshwater discharge, are most comparable to those of our site:

$$k = 1.2 + 2.3u \left(\frac{Sc_{(balt)}}{660} \right)^{-0.5} \quad (2)$$

170 where u is the wind speed (m s^{-1}) and $Sc_{(balt)}$ is the Schmidt number at the sampling site, based on local temperature, salinity, and gas molecules. $Sc_{(balt)}$ is extrapolated from freshwater and seawater ($S = 35$) coefficients from Wanninkhof (2014) for the salinity observed at the sampling site (Roth et al., 2023).

Wind speed (in m s^{-1} , METEK, uSonic-3 Scientific) and air temperature (in $^{\circ}\text{C}$, Vaisala, HMP155) were measured at the 3.2 and 2 m height, respectively, above sea level at the newly established Integrated Carbon Observation System (ICOS) coastal
175 site at TZS (ICOS code FI-Tvm; Fig. 1). Daily averaged wind speed, extrapolated to a height of 10m using the relationship from Hsu et al., (1994), assuming a near-neutral atmospheric stability conditions, was used to calculate the air-sea exchanges of GHGs. Atmospheric concentrations of CO_2 and CH_4 were measured at each site before the air-sea flux measurements. For N_2O , we used the monthly average atmospheric concentration of 0.3367 ppm, measured at Pallas-Sammaltunturi, GAW Station, Finland (PAL, <https://gml.noaa.gov/dv/iadv/graph.php?code=PAL&program=ccgg&type=ts>).

180 The recorded data from the IRGA for both seawater and air-sea flux measurements were filtered to remove measurements taken during the transition period between stations and when switching between ambient air and seawater measurements, as the IRGA requires time to respond to sharp concentration changes. Additionally, data affected by improper functioning (i.e., seawater flow $< 1.5 \text{ L min}^{-1}$) were discarded.

2.3 Sample analysis

185 TA was determined by Gran titration (Gran, 1952) using an AS-ALK2 titration system (Apollo SciTech), where a 15-ml sample was titrated with a standard 0.1 M HCl solution. DIC was measured on a DIC analyser (Apollo SciTech) by acidification of a 0.75 ml subsample with 1 ml 10% H_3PO_4 , and quantification of the released CO_2 with a non-dispersive infrared CO_2 analyser (LI-COR, LI-7000). Results were then converted from $\mu\text{mol L}^{-1}$ to $\mu\text{mol kg}^{-1}$ based on sample density, calculated from salinity and temperature at the time of the analysis. Accuracies of ± 3 and $\pm 2 \mu\text{mol kg}^{-1}$ were determined for
190 TA and DIC, respectively, from routine analysis of certified reference materials (A.G. Dickson, Scripps Institution of Oceanography, San Diego, CA, USA).

Discrete seawater samples for CH_4 and N_2O concentrations were measured via the headspace equilibrium technique (25 ml N_2 headspace in 60 ml serum bottles) and measured with a gas chromatograph (SRI 8610C) with flame ionization detection, and

electron capture detection calibrated with CH₄: CO₂:N₂O: N₂ mixtures (Air Liquide) of 1.0, 10, and 30 ppm CH₄ and 0.2, 1.0, and 6.0 ppm N₂O. The measurement precision was 8 % for CH₄ and 4 % for N₂O.

3. Results

3.1 Atmospheric forcing and seawater physical properties

During the survey period, atmospheric temperatures remained relatively stable, averaging 18.0°C (SD = 1.5, n = 1989; Fig. 1a). A short warm event occurred on the 7th of August, when the air temperature peaked at 26.3°C. In contrast, short cold spells were observed on 5 and 7 September, with temperatures dropping to 13.7 and 11.5°C, respectively. With relatively stable atmospheric temperature, surface seawater temperature (SST) showed limited variation, ranging from 17.2 to 19.5°C (mean = 18.4°C, SD = 0.66, n = 21; Fig. 1c). The Karjaanjoki River, being the main source of freshwater input to the area, created a pronounced salinity gradient, from nearly fresh surface seawater salinity (SSS = 0.07) at the river mouth (in Pohja) to a SSS of 6.36 in Hanko (Fig. 1d).

3.2 Seawater biogeochemical properties

Surface seawater chlorophyll-*a* (chl-*a*) concentration ranged from 3.7 to 37.7 µg L⁻¹ (mean = 14.4, SD = 8.1, n = 21, Fig. 1e), with the highest concentration observed in the sheltered bay around Tammisaari (S5 and S6). Turbidity ranged from 5.0 to 8.4 FTU (mean = 6.1, SD = 0.8, n = 21, Fig. 1f), with the highest values observed in sheltered bays within the archipelago (S8, S9, and S11). Coloured dissolved organic matter (CDOM) ranged from 3.0 to 10.6 µg L⁻¹ (mean = 5.9, SD = 2.6, n = 21, Fig. 1g) and exhibited a salinity-driven gradient, with the highest concentrations observed towards Pohja and the lowest towards Hanko. However, one site in the archipelago (S8) and near Hanko (S19) exhibited elevated CDOM concentrations compared to other locations in their respective area, reaching up to 10.0 and 10.6, respectively. Dissolved oxygen (DO) saturation levels ranged from 74.3 to 112.7 % (mean = 94.1, SD = 9.6, n = 21, Fig. 1h), with most of the sites undersaturated in O₂, excepted for two sites closed by Tammisaari (S3 and S4) and four sites in closed vicinity of TZS (S14, S15, S18, and S19).

Surface water pCO₂ ranged from strong undersaturation (160 ppm) to a significant supersaturation relative to the atmosphere (406 ppm) at S2 (1702 ppm) and up to 2521 ppm at the river mouth (triangle on Fig. 2a). Surface water was consistently supersaturated with respect to CH₄ (from 636 to 14609 %, data not shown) compared to the atmosphere, with concentrations ranging from 19 to 469 nmol L⁻¹ (Fig. 2b). Undersaturated pCO₂ (from 160 to 403 ppm) and low CH₄ concentrations (from 19 to 34 nmol L⁻¹) were mostly observed at open sites between TZS and Hanko (S14 to S21), except for S17, where pCO₂ and CH₄ reached 485 ppm and 48 nmol L⁻¹, respectively. Seawater pCO₂ and CH₄ concentrations increased across the archipelago and Pojoviken Bay, reaching their maximal values at the mouth of the Karjaanjoki River. Local lows were observed in Pojoviken Bay (S3 and S4), where pCO₂ was undersaturated (192 and 238 ppm, respectively), and CH₄ concentrations were relatively low (83 and 67 nmol L⁻¹, respectively). N₂O concentration ranged from 9 to 25 nmol L⁻¹, with saturation level ranging from 93 to 255 %. Higher concentrations and maximum saturation were observed at the mouth of the Karjaanjoki

225 River (Fig. 2c). South of Tammisaari, surface waters were either undersaturated in N₂O (S8, S13, and S16) or close to the atmospheric equilibrium (S6 and S12, data not shown). TA and DIC ranged from 668 to 1631 and from 720 to 1550 $\mu\text{mol kg}^{-1}$, respectively, with minimum TA and DIC concentration measured at the mouth of the Karjaanjoki River. Both TA and DIC exhibited conservative behaviour with changes in salinity (Fig. 3a).

3.3 Air-sea GHG flux densities

230 The area presented contrasts in terms of sink and source of CO₂ and N₂O, but was clearly a source of CH₄. Measured air-sea CO₂ flux densities ranged from -36 to 150 $\text{mmol m}^{-2} \text{d}^{-1}$, where negative values indicate net uptake of atmospheric CO₂ (Fig. 2d). In contrast, the study area acted as a net source of CH₄ to the atmosphere, with flux densities ranging from 0.014 to 1.39 $\text{mmol m}^{-2} \text{d}^{-1}$ (Fig. 2e). Overall, the calculated fluxes showed a similar trend in term of pattern and range compared to the observed values, ranging from -26.4 to 220.3 $\text{mmol m}^{-2} \text{d}^{-1}$ for CO₂ (Fig. 2f) and from 0.03 to 1.1 $\text{mmol m}^{-2} \text{d}^{-1}$ for CH₄ (Fig. 235 2g). Calculated fluxes of N₂O ranged from -0.0021 to 0.0383 $\text{mmol m}^{-2} \text{d}^{-1}$, with sites in the archipelago and towards TZS acting as a slight sink of N₂O (Fig. 2h).

3.4 Statistical analysis

The Kendall's τ coefficient has been calculated to investigate the correlation between surface water pCO₂, CH₄, and N₂O concentration, as well as between physical and biogeochemical parameters and all three GHGs (Fig. 4). Surface pCO₂ and CH₄ 240 concentrations show a moderate ($\tau = 0.5636$) and statistically significant ($p = 0.0165$) positive relationship (Fig. 4a), suggesting that processes driving high pCO₂, such as respiration and organic matter degradation, are likely also contributing to elevated CH₄ levels. N₂O is not correlated with pCO₂ and shows a moderate correlation with CH₄ ($\tau = 0.4182$), although this relationship is only marginally significant ($p = 0.0866$). Among the tested biogeochemical variables, SSS and O₂ saturation show strong, significant negative correlations with pCO₂ and CH₄ (Fig. 4b). N₂O is only strongly negatively correlated with salinity ($\tau = -$ 245 0.8182, $p = 0.00013$) and shows a marginal correlation with CDOM ($\tau = 0.4545$, $p = 0.060$). CDOM is positively correlated with CH₄ ($\tau = 0.4952$, $p = 0.0013$) while chl-*a* is moderately and significantly correlated with CH₄ ($\tau = 0.4095$, $p = 0.009$). SST and turbidity show weak and non-significant relationships with the three gases.

4. Discussion

Surface water pCO₂, CH₄, and N₂O concentrations showed significant spatial variation, spanning several orders of magnitude 250 (Fig. 2). Humborg et al. (2019) reported pCO₂ and CH₄ levels following the salinity gradient driven by the Karjaanjoki River, with values reaching up to 1583 μatm and 70 nmol L^{-1} , respectively. However, these measurements were conducted in the deeper part of the archipelago's main channel, and they did not explore Pojoviken Bay. In contrast, our study focused on shallow (< 4m depth) nearshore ecosystems. Asmala and Scheinin (2024) observed a similar range of pCO₂ and CH₄ concentrations across a 2000 km^2 coastal region surrounding the Hanko Peninsula, with the large magnitude of both gases

255 associated with excessive organic matter loads, elevated primary production, trapping and accumulation of allochthonous organic matter, and sedimentary conditions favourable to CH₄ production. Focusing on nearshore shallow (< 4m depth) coastal habitats around the island of Askö (Sweden, northern Baltic Sea), Roth et al. (2023) reported strong spatial variability in surface water pCO₂ and CH₄ concentrations and emphasized that their dynamics were highly habitat-specific. The N₂O concentrations we observed were in the same range as those observed by Aalto et al. (2021) in the same area, with
260 concentrations ranging from 25 to 50 nmol L⁻¹ within Pojoviken Bay and from 10 to 30 nmol L⁻¹ within the archipelago. Aalto et al. (2021) reported that higher N₂O concentrations were associated with higher nitrate concentrations and inputs of allochthonous carbon, while lower N₂O concentrations were associated with efficient internal recycling of N.

4.1 Salinity gradient

The Karjaanjoki river is the main source of freshwater into the studied area, delivering large amounts of allochthonous carbon and nutrients to the system (Fleming-Lehtinen et al., 2015). Changes in SSS have been suggested to strongly influence the
265 concentration of all three GHGs (Fig. 4b), with terrestrial runoff likely contributing to elevated CH₄ concentrations in the area (Asmala and Scheinin, 2024). At a first glimpse, only N₂O exhibited a strong relationship with salinity ($R^2 = 0.95$), while pCO₂ ($R^2 = 0.58$) and CH₄ ($R^2 = 0.39$) exhibited weaker correlations (Fig. 5, dotted black line). However, when focusing on exposed and semi-sheltered sites, the correlation with salinity strengthened considerably, reaching values up to 0.79 for pCO₂, 0.94 for
270 CH₄, and 0.96 for N₂O (Fig. 5, dashed red line). The impact of water mixing on surface-water pCO₂ can be assessed by estimating the variation in pCO₂ resulting only from the physical mixing of Karjaanjoki River water ($S = 0$, $TA = 668 \mu\text{mol kg}^{-1}$, $DIC = 720 \mu\text{mol kg}^{-1}$) with a seawater endmember (black symbols in Fig. 3, $S = 6.42$, $TA = 1635 \mu\text{mol kg}^{-1}$, $DIC = 1586 \mu\text{mol kg}^{-1}$). The seawater endmember was collected on 18 September 2023 at 59.77393 °N, 23.2607167 °E, about 7.5 km south of TZS. This calculation excludes the effects of biological activity, gas exchange, and potential precipitation or
275 dissolution of calcium carbonate. Assuming that changes in SSS reflect a mixing ratio between the Karjaanjoki River and the seawater endmembers and based on the observed linear relationships between salinity and both TA and DIC (Fig. 3a), representing conservative mixing between the two water masses, TA and DIC can be estimated based on salinity. The estimated TA and DIC are then used to compute pCO₂ (blue dashed line, Fig. 5a) using CO2SYS v3 (Sharp et al., 2023), applying the carbonic acid dissociation constants (K_1 and K_2) of Millero et al. (2006) and the KHSO₄ formulation of Dickson (1990). While
280 TA and DIC behaved conservatively during mixing, pCO₂ exhibited a pronounced nonlinear response, similar to the one observed for exposed and semi-sheltered sites (Fig. 5a). Such behaviour has already been reported in estuaries (Abril et al., 2021; Cai et al., 2013). This suggests that the input of CO₂, CH₄, and N₂O supersaturated water from the Karjaanjoki River is a major source for all three GHGs across the study area, with dilution playing a dominant role in GHG dynamics at exposed and semi-sheltered sites. While changes in SSS appear to explain most of the variability observed in pCO₂, CH₄, and N₂O
285 concentrations, deviations from the expected salinity-driven pattern, such as those observed within the archipelago, at sheltered sites, likely reflect additional local processes.

4.2 Biological drivers

Many processes affecting the carbonate system are best described by examining the associated changes in DIC and TA (Zeebe and Wolf-Gladrow, 2001). However, most of the changes in TA and DIC in surface seawater appeared to be driven by salinity changes, with estimated endmember ($S=0$) values of 662 and 712 $\mu\text{mol kg}^{-1}$ for TA and DIC, respectively (Fig. 3a). These estimates closely match the measured TA and DIC at the mouth of the Karjaanjoki River (668 and 720 $\mu\text{mol kg}^{-1}$, respectively). Therefore, to discard the impact of freshwater inputs, TA and DIC were normalized to the average surface seawater salinity of 5 (denoted as nTA and nDIC, respectively) using the normalization of Friis et al. (2003). In Figures 3b, c, and d, the dotted lines illustrate the expected responses of TA and DIC to different biogeochemical processes. Biological activity (photosynthesis/respiration) affects both TA and DIC in a ratio of -0.16 (Lazar and Loya, 1991), air-sea exchange of CO_2 only affects DIC, and the precipitation-dissolution of calcium carbonate affects TA and DIC in a 2:1 ratio. However, the effect of calcifying primary producers in the carbon pool can be neglected in the Baltic Sea, except in the benthic zone (Tyrrell et al., 2008). The sampling sites are compared to the seawater endmember ($S = 6.42$, $\text{TA} = 1635 \mu\text{mol kg}^{-1}$, $\text{DIC} = 1586 \mu\text{mol kg}^{-1}$, black symbol in Fig. 3). Figures 3b and c, show how exposed sites, undersaturated in pCO_2 and low CH_4 concentrations, follow the theoretical trend of photosynthesis and CO_2 release. However, an uptake of CO_2 was both measured and estimated for those sites (Fig. 2), suggesting that gas exchange as release towards the atmosphere could not explain the observed changes in the carbonate system. In contrast, sheltered sites, with supersaturated pCO_2 and elevated CH_4 concentrations, were more scattered around the theoretical respiration line and CO_2 uptake. Gas exchange, as CO_2 uptake, can also be ruled out, as both measured and estimated air-sea fluxes pointed to a net release of CO_2 into the atmosphere. Altogether, this suggests that biological processes—primary production at undersaturated pCO_2 sites and respiration at supersaturated sites—mainly influence the carbonate system, while the direct role of air-sea exchange of CO_2 appears to be minor in shaping the inorganic carbonate dynamics across the study area. The similar pattern observed for both pCO_2 and CH_4 concentrations illustrated the positive relationship between the two gases (Fig. 4), where processes responsible for higher pCO_2 , such as respiration and/or organic matter degradation, also contribute to higher CH_4 concentrations (Reeburgh, 2007). No clear pattern could be observed between the nTA: nDIC ratio and N_2O concentration, as expected by the poor Kendall's coefficient of correlation (Fig. 4). Elevated chl-*a* concentrations (up to 37.7 $\mu\text{g L}^{-1}$, Fig. 1) in surface seawater were reported across the studied area. Chl-*a* measurements are typically considered as a proxy for pelagic primary production (Cloern et al., 2014). Therefore, elevated chl-*a* could indicate active photosynthesis production, resulting in pelagic O_2 production and pelagic CO_2 uptake. To investigate the relationship between photosynthetic activity and surface water pCO_2 , CH_4 , and N_2O concentrations, apparent oxygen utilization (AOU), calculated as the difference between oxygen saturation and measured oxygen concentration, was examined with chl-*a* concentrations. However, no clear relationship was observed between chl-*a* and AOU, and negative AOU values, indicative of net photosynthesis, were associated with relatively low chl-*a* concentrations within the observed range (Fig. 6a). These negative AOU values corresponded to undersaturated pCO_2 and low CH_4 concentrations (Fig. 6b, c). In contrast, positive AOU values, representing net respiration, were associated with supersaturated pCO_2 and elevated CH_4

320 concentrations, both of which increased with higher AOU. A strong correlation exists between AOU and surface water pCO₂ (Fig. 6b), which improves when focusing on exposed and semi-sheltered sites (red dashed line), suggesting that primary production strongly affects the carbonate system, as already indicated by the nDIC: nTA ratio (Fig. 3). Primary production could take place either in the pelagic or the benthic realm. However, the lack of correlation between chl-*a* and AOU and the low chl-*a* concentration associated with negative AOU suggests that pelagic primary production may not be solely responsible
325 for the observed changes in pCO₂. Pelagic primary production may be limited by the shallow water column, which restricts phytoplankton biomass and its productivity at the time, as this study was conducted at the end of the summer bloom (Uth et al., 2024). The correlation between AOU and CH₄ is weaker than for CO₂ ($R^2 = 0.51$, black dotted line) but improves significantly when only exposed and semi-sheltered sites are considered ($R^2 = 0.78$, red dashed line). O₂ depletion, associated with supersaturated pCO₂ and high CH₄ concentration, could be related to organic matter degradation, as large amounts of
330 carbon are turned over during the summer (Attard et al., 2019). However, here as well, limited phytoplankton biomass in the shallow water column could restrict the impact of the pelagic realm on CH₄ concentration. N₂O exhibited a clear positive correlation with AOU ($R^2 > 0.7$). This correlation is known to reflect the coupling between O₂ consumption (from organic matter remineralization) and microbial nitrogen cycling processes that produce N₂O (Kock et al., 2016; Carrasco et al., 2017). Therefore, the absence of a correlation between chl-*a* and AOU, despite AOU correlating with all three GHGs, suggests that
335 pelagic processes alone cannot explain the observed changes in GHG concentrations. Instead, processes beyond the pelagic realm, such as benthic community production and organic-matter degradation, are likely contributing to the observed surface-water pCO₂, CH₄, and N₂O concentrations (e.g. Roth et al. 2022, 2023).

4.3 Exposed and semi-sheltered vs sheltered sites

The inputs of freshwater supersaturated in CO₂, CH₄, and N₂O represent a major source of GHGs to the study area. Freshwater
340 mixing with seawater appears to control the concentration of all three GHGs at exposed and semi-sheltered sites, as much of the variability in surface seawater concentrations can be explained by salinity. In sheltered sites of the archipelago, deviations from the salinity-driven pattern indicate the influence of additional processes. Biological activity is likely the key driver, with pCO₂ undersaturation associated with primary production and pCO₂ supersaturation linked to respiration. However, pelagic production alone is unlikely to be the main contributor to the observed changes. Benthic processes such as benthic community
345 metabolism (i.e., the balance between production and respiration), organic matter degradation, and sediment-water interactions are likely to contribute to both CO₂ and CH₄ dynamics (Attard et al., 2019; Roth et al., 2023). For CH₄, riverine inputs and subsequent dilution largely determine large-scale patterns, whereas elevated concentrations at sheltered sites within the archipelago reflect localized production and limited oxidation. Large amounts of carbon are turned over in habitats from the studied area during the summer months (Attard et al., 2019), and macrophyte tissue may become a direct component of local
350 sediment organic matter pools, which favour local CH₄ production (Roth et al., 2022; Wallenius et al., 2021). Anoxic degradation of organic-rich sediments (methanogenesis), exacerbated by warm late-summer waters (Roth et al., 2022; Yvon-Durocher et al., 2014), combined with short water residence times that limit oxidation in both sediment and the overlying water

column (Reeburgh, 2007), creates favourable conditions for CH₄ production. Due to the sheltered nature of the sites and limited water exchange, the produced CH₄ can accumulate. Similarly, enhanced respiration of organic carbon in shallow ecosystems elevates CO₂ concentrations (Humborg et al., 2019). While macrophyte growth during summer can draw down pCO₂ through photosynthetic CO₂ uptake, the subsequent decomposition of deposited tissues may instead enhance CO₂ and CH₄ release. Spatial variability of N₂O mirrors the findings of Aalto et al. (2021), who linked higher N₂O concentrations near the Karjaanjoki River to higher nitrate inputs and allochthonous carbon inputs. They suggested that the ratio between nitrate and autochthonous organic carbon controls the balance between N-removing denitrification and N-recycling through Dissimilatory Nitrate Reduction to Ammonium (DNRA), as well as the end-product of denitrification (Aalto et al., 2021). Within the archipelago, where riverine influence is limited, DNRA can produce significant amounts of bioavailable ammonium, enhancing nitrogen recycling between sediments and surface water, especially in summer, when autochthonous biomass production and sedimentation are highest.

4.4 Air-sea flux densities

Given the strong spatial variability observed in GHG concentrations across the study area, strong variations were also observed in the corresponding air-sea fluxes. At exposed sites between Hanko and TZS, where surface seawater was undersaturated in CO₂ and low in CH₄ concentration, an uptake of atmospheric CO₂ associated with small releases of CH₄ was observed. In contrast, within the archipelago and across Pojoviken Bay, where surface waters were supersaturated in CO₂ and exhibited elevated CH₄ concentrations, releases of both CO₂ and CH₄ were observed. Sites 3 and 4 represented exceptions, being the only locations undersaturated in CO₂, thus acting as a CO₂ sink. Daily CH₄ emissions of $\geq 0.1 \text{ mmol m}^{-2}$ across all habitats are comparable to, or even higher than, CH₄ fluxes reported from similar (Lundevall-Zara et al., 2021; Roth et al., 2023) or other vegetated coastal ecosystems (Al-Haj and Fulweiler, 2020; Rosentreter et al., 2021a). Regarding N₂O, Pojoviken Bay is estimated to act as a source to the atmosphere. In sheltered sites of the archipelago and exposed sites near TZS, N₂O concentrations close to or slightly below atmospheric equilibrium result in small emissions and occasional uptake, respectively. Our estimated air-sea exchanges overall mirrored the spatial trends of chamber-based fluxes and exhibited a similar range of air-sea exchange. Both chamber-based and estimated fluxes may underestimate the air-sea fluxes, as possible ebullition events are not considered. Humborg et al. (2019) suggested that air-sea CH₄ flux is likely dominated by frequent bubbling from the sediment. This process could be dominant in shallow coastal environments, as shallow seafloor depth promotes a short residence time for CH₄ in the water column, which prevents its potential for full oxidation, which, in contrast, been observed in the deep open ocean (Reeburgh, 2007). Bubbling was, however, not observed during our chamber-based measurements (a comparison between chamber-based fluxes and estimated air-sea exchange can be found in the supplementary material). The overall contribution of the study area to atmospheric GHG budgets can be assessed by combining the air-sea flux of CO₂ with the CO₂-equivalent fluxes of CH₄ and N₂O. These equivalents are calculated using their respective sustained-flux global warming potential (SGWP), as a greenhouse gas metric to describe the relative radiative impact of a standardized amount of

385 gas over a defined time horizon. Specifically, over a 100-year time horizon, the SGWP of CH₄ and N₂O is 27 and 273 times, respectively, greater than that of CO₂ (IPCC 2023), on a mass basis, based on:

$$F_{CO_2-eq(X)} = F_{(X)}SGWP_{(X)} \quad (3)$$

where the CO₂-eq flux of X (CH₄ or N₂O), $F_{CO_2-eq(X)}$, is the product of the flux of the gas ($F_{(X)}$) and its respective SGWP (i.e., 27 or 273) over the time horizon of 100 years. As direct measurements of air-sea N₂O fluxes were not available at the time, and to ensure consistency across all three gases, we chose to use the estimated fluxes rather than the chamber-based measurements.

Overall, the study area acted as a net source of GHGs to the atmosphere, with an average release of 1.6 g of CO₂-eq m⁻² d⁻¹ over the study period, and ranging from -1.0 g of CO₂-eq m⁻² d⁻¹ at S14 to 10.6 g of CO₂-eq m⁻² d⁻¹ at the mouth of the Karjaanjoki River (Fig. 7d). Exposed sites between Hanko and TZS generally acted as GHGs sink, primarily due to the substantial CO₂ uptake compensating for CH₄ emissions. An exception was S21, where the small uptake of CO₂ (-0.008 g of CO₂ m⁻² d⁻¹) was offset by the release of CH₄ (0.016 g of CO₂-eq m⁻² d⁻¹), resulting in a net source. In Pojoviken Bay, the river mouth and S2 were acting as a strong source of GHGs to the atmosphere, with N₂O fluxes in the same order of magnitude as those of CH₄. However, S3 and S4, both sites undersaturated in pCO₂, remained net GHG sinks, despite releasing similar amounts of CH₄ and N₂O. In the archipelago, all sheltered sites remained a net source of GHGs to the atmosphere. If, from a climate mitigation perspective, it may seem sufficient to focus on CO₂ (as Figure 7a and d are pretty similar), as in the blue carbon approach, it has been shown that CO₂ uptake can be offset by CH₄ emissions (Roth et al., 2023). Our findings demonstrate that, if CH₄ matters in coastal, nearshore environments, so does N₂O, since N₂O fluxes are not only of the same order of magnitude as CH₄ fluxes but can either reinforce the warming effect of CH₄ or partially counterbalance it.

5. Conclusions

405 Shallow coastal ecosystems are highly heterogeneous, with their spatial structure and temporal dynamics of benthic communities shaping ecosystem functions (Snelgrove et al., 2014). This heterogeneity drives strong spatial variability in coastal biogeochemical processes, which exert an important influence on the carbon cycle at both local and global scales (Ward et al., 2020). Yet, despite their importance, identifying the locations and processes regulating coastal CO₂, CH₄, and N₂O fluxes remains uncommon, and global estimates still fail to capture the heterogeneous and dynamic nature of these environments (Rosentreter et al., 2021b). In particular, northern temperate coastal habitats are poorly represented in current GHG budgets (Resplandy et al., 2024), despite their relative importance: shallow waters (<5 m) in the Baltic Sea cover ~30,000 km² (HELCOM, 2013), an area comparable to 20% of the global distribution of mangroves (Bunting et al., 2018) or seagrass meadows (McKenzie et al., 2020).

Across the study area, freshwater inputs from the Karjaanjoki River and subsequent mixing with seawater largely determined the overall spatial patterns of surface water pCO₂, CH₄, and N₂O concentrations. However, important deviations from this salinity-driven control were observed at the local scale. In sheltered sites within the archipelago, elevated CH₄ concentrations

and supersaturated $p\text{CO}_2$ likely reflected biological processes, including enhanced organic matter respiration and methanogenesis in warm, late-summer shallow waters and sediments, where limited oxidation favoured CH_4 accumulation. By contrast, at semi-sheltered and exposed sites, mixing processes exerted stronger control, leading to lower GHG concentrations. N_2O patterns followed riverine influence with higher concentrations near the river mouth associated with inputs of allochthonous carbon and elevated nitrate availability, and lower concentrations within the archipelago reflecting reduced riverine input and active nitrogen recycling between sediments and surface waters (Aalto et al., 2021). Together, these results underline the dual importance of large-scale physical mixing and local biological processes in shaping the spatial heterogeneity of GHG dynamics. Our findings suggest that benthic ecosystems likely play a pivotal role in regulating GHG dynamics, especially in shallow coastal environments, not only through production but also through consumption pathways, which remain insufficiently constrained in current budgets (Rodil et al., 2021; Roth et al., 2022; Wallenius et al., 2021).

When translated into $\text{CO}_2\text{-eq}$, air–sea GHG fluxes were dominated by CO_2 , while CH_4 and N_2O contributed comparably but in different ways. CH_4 consistently acted as a source, whereas N_2O partially offset the CH_4 release through uptake. This interaction highlights that the balance between production and consumption processes, especially within different seafloor habitats, is critical for understanding coastal contributions to the global carbon budget. There is a critical need to quantify CH_4 and N_2O exchanges more accurately and to deepen our understanding of the environmental and management factors that control their production and consumption. This would help make global estimates less sensitive to statistical assumptions and reduce uncertainties in blue carbon potential estimates (Rosentreter et al. 2021a).

While coastal ecosystems are often recognized as CO_2 sinks, recent work shows that they are also significant sources of CH_4 and N_2O that can offset a substantial portion of the climate benefit of taking up CO_2 (Rosentreter et al., 2021a; Roth et al., 2023; Resplandy et al., 2024). Given their extent, northern temperate coastal ecosystems represent a relevant but overlooked source of GHGs, with the potential to amplify the global ocean carbon budget by increasing net greenhouse gas emissions to the atmosphere. Our results highlight the urgent need for research that integrates GHG with biodiversity, benthic–pelagic interactions, and microbial processes, and that resolves temporal variability across seasonal cycles. Such knowledge is essential to improve predictions of how coastal ecosystems will mediate carbon–climate feedback under future environmental change.

Data availability

The dataset will be available at the Bolin Centre Database. The datasets are currently being validated.

Author contribution

NXG, AN, and AV designed the hypothesis field sampling. Field measurements were carried out by NXG and AV, while laboratory and data analyses were performed by NXG and BD. NXG prepared the manuscript with input from all co-authors. All authors approved the final version of the manuscript for submission.

Competing interests

The authors declare no competing interests.

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Figure 1: Atmospheric conditions: **a)** air temperature ($^{\circ}\text{C}$) and **b)** wind speed (m/s) recorded at the ICOS station FI-Tvm established at the Tvärminne Zoological Station. Spatial variation of **c)** surface seawater temperature (SST, $^{\circ}\text{C}$), and **d)** salinity (SSS), **e)** chlorophyll-*a* (Chl-*a*, $\mu\text{g L}^{-1}$) concentration, **f)** turbidity (FTU), **g)** coloured dissolved organic matter (CDOM, $\mu\text{g L}^{-1}$), and **h)** dissolved oxygen saturation (%) across the sampling area. Triangles represent the surface water conditions at the mouth of the Karjaanjoki River.

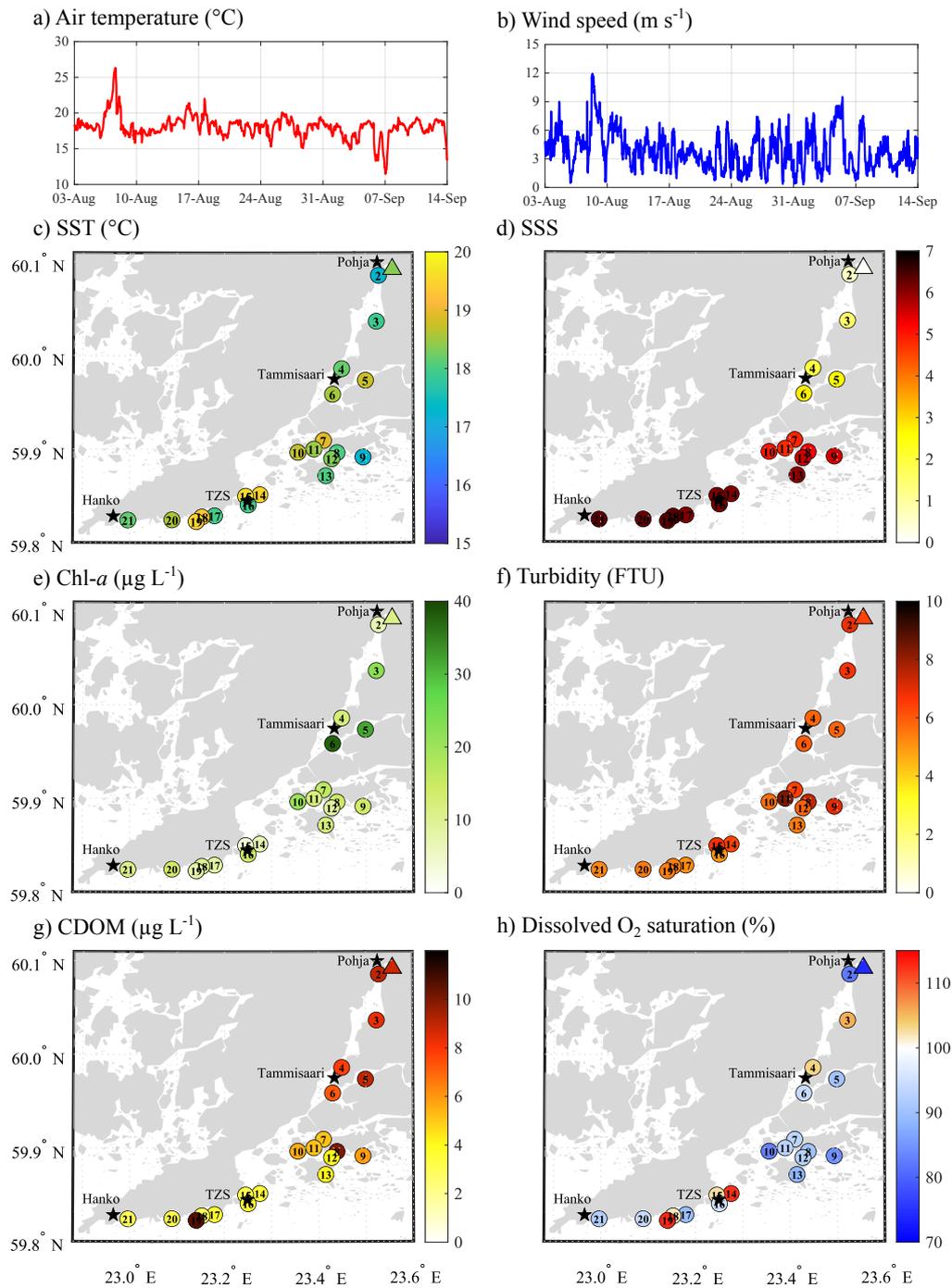


Figure 2: Left column presents the spatial variation of surface seawater **a)** $p\text{CO}_2$ (μatm), **b)** CH_4 (nmol L^{-1}), and **c)** N_2O concentration (nmol L^{-1}). The middle column presents measured air-sea fluxes of **d)** CO_2 ($\text{mmol m}^{-2} \text{d}^{-1}$) and **e)** CH_4 ($\text{mmol m}^{-2} \text{d}^{-1}$). The right column presents the calculated air-sea fluxes of **f)** CO_2 ($\text{mmol m}^{-2} \text{d}^{-1}$), **g)** CH_4 ($\text{mmol m}^{-2} \text{d}^{-1}$), and **h)** N_2O ($\text{mmol m}^{-2} \text{d}^{-1}$). Positive values indicate a release from the sea to the atmosphere, while negative values represent an uptake by the sea. Triangles represent the surface water conditions at the mouth of the Karjaanjoki River.

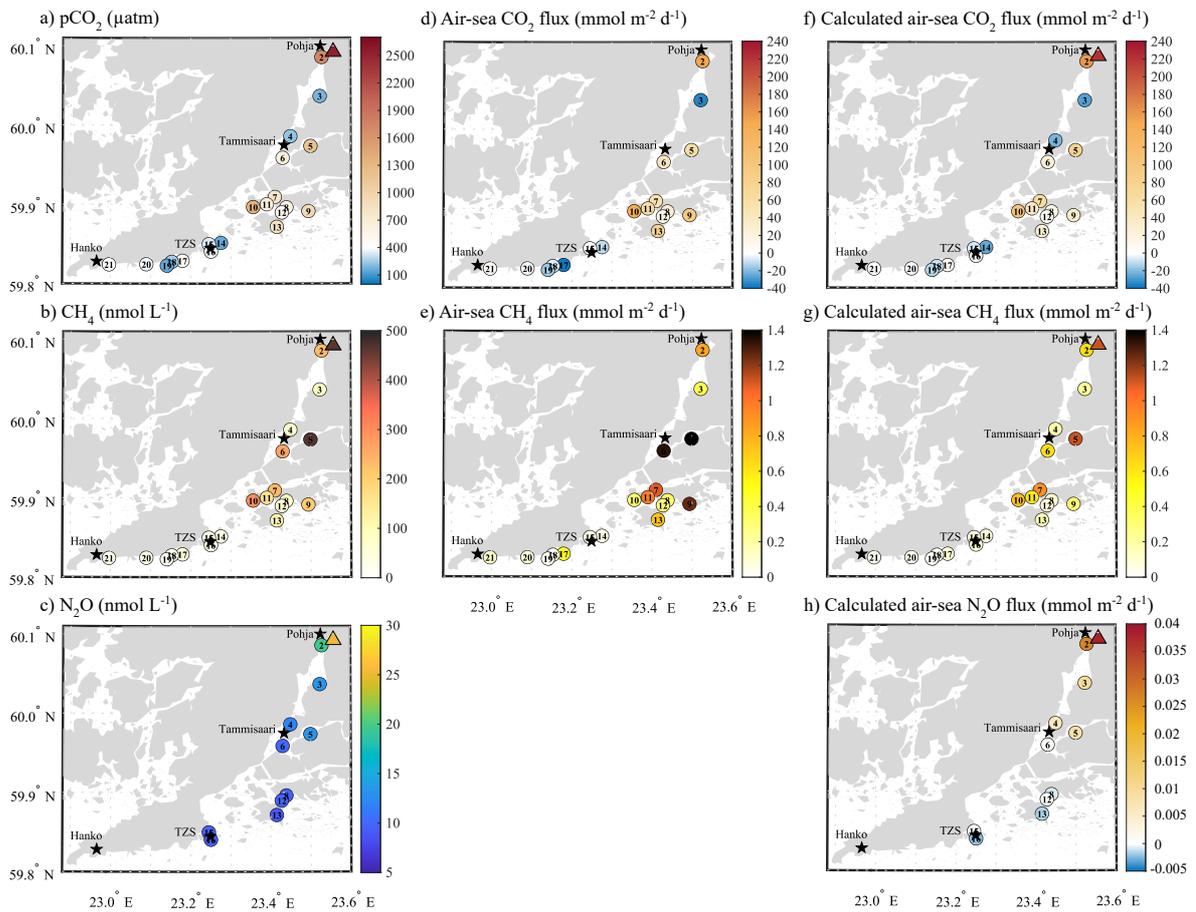


Figure 3: **a)** Relationship between salinity, TA ($\mu\text{mol kg}^{-1}$, circles) and DIC ($\mu\text{mol kg}^{-1}$, squares). Relationship between nDIC and nTA, and their association with **b)** pCO_2 (μatm), **c)** CH_4 (nmol L^{-1}), and **d)** N_2O (nmol L^{-1}) concentrations. Black symbols represent the seawater endmember. Exposed and semi-sheltered sites are represented by circles, while sheltered sites are represented by square symbols.

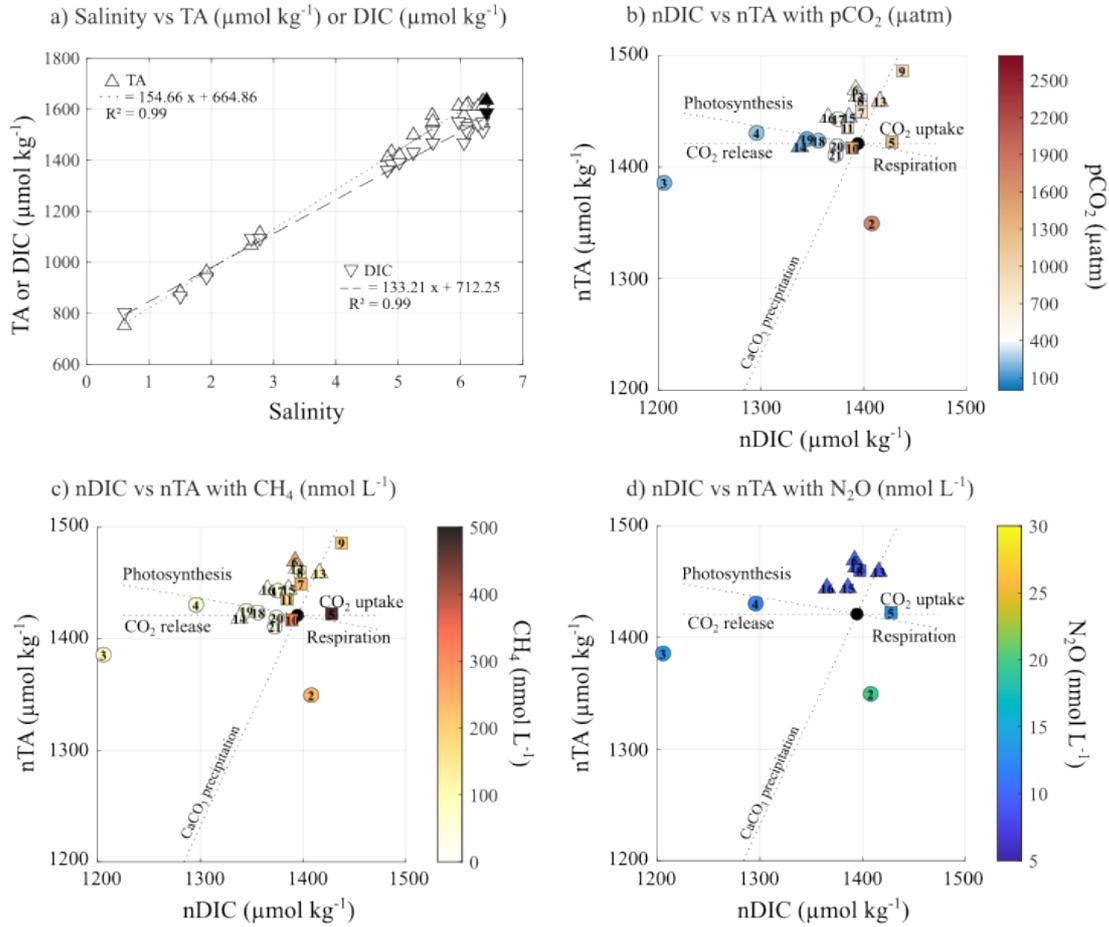


Figure 4: Heatmap of Kendall correlation coefficient **a)** between surface water pCO₂, CH₄, and N₂O concentration, and **b)** between main seawater biogeochemical properties (SST, SSS, chl-*a* concentration, colored dissolved organic carbon, turbidity, and O₂ saturation) and pCO₂, CH₄, and N₂O concentration.

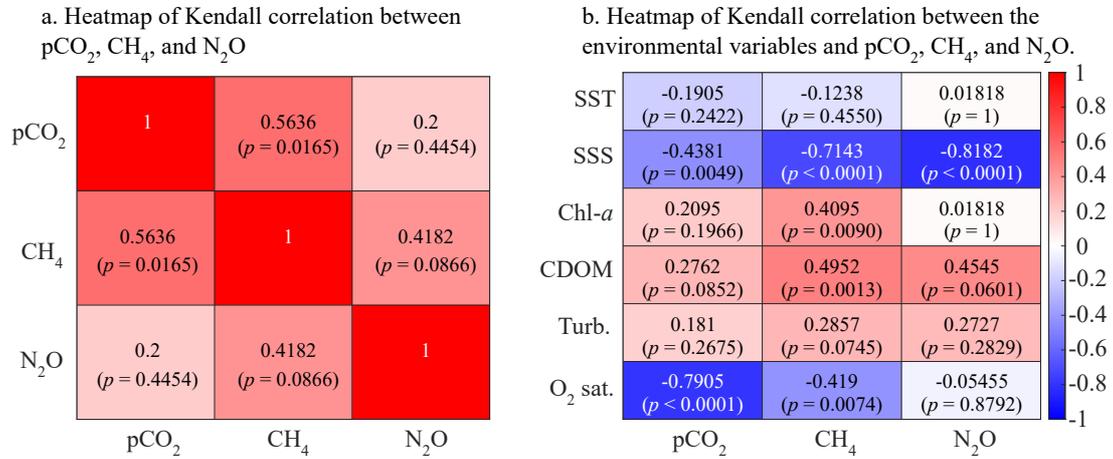


Figure 5: Relationship between surface seawater salinity and **a)** pCO₂ (μatm), **b)** CH₄ (nmol L⁻¹), and **c)** N₂O (nmol L⁻¹) concentrations. In each panel, the dotted line represents the regression across all sites, while the red dashed line shows the regression limited to exposed and semi-sheltered sites. The blue line in panel **a)** represents the estimated pCO₂ resulting only from the mixing of the Karjaanjoki River water (S = 0) with seawater (S = 6.42).

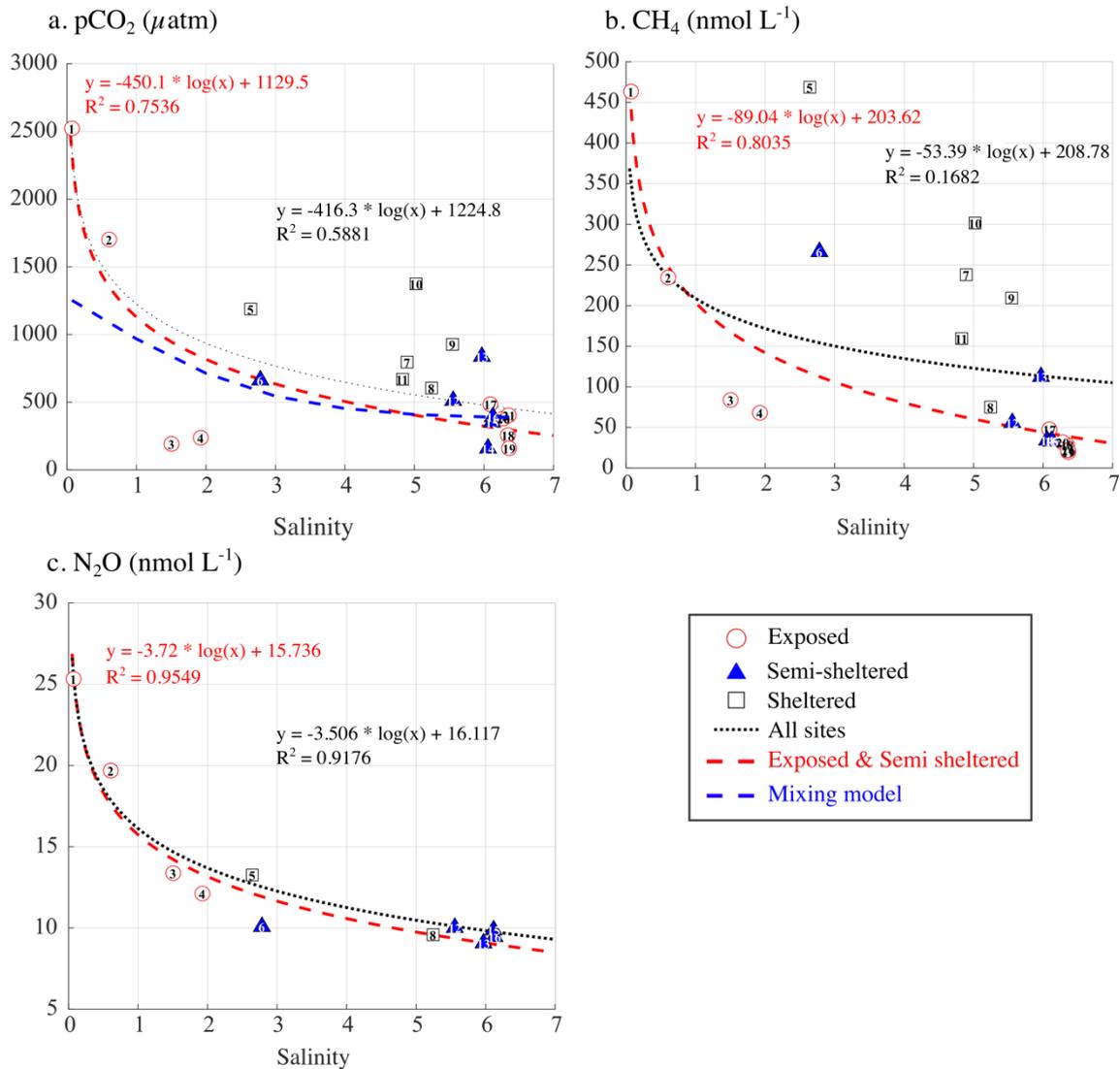


Figure 6: Relationship between the apparent oxygen utilization (AOU) and **a)** surface water Chl-*a* concentration ($\mu\text{g L}^{-1}$), **b)** surface water pCO_2 (in μatm), **c)** CH_4 (nmol L^{-1}), and **d)** N_2O (nmol L^{-1}) associated with the Chl-*a* concentration ($\mu\text{g L}^{-1}$). In each panel, the dotted line represents the regression across all sites, while the red dashed line shows the regression limited to exposed and semi-sheltered sites. Exposed and semi-sheltered sites are represented by circles, while sheltered sites are represented by square symbols.

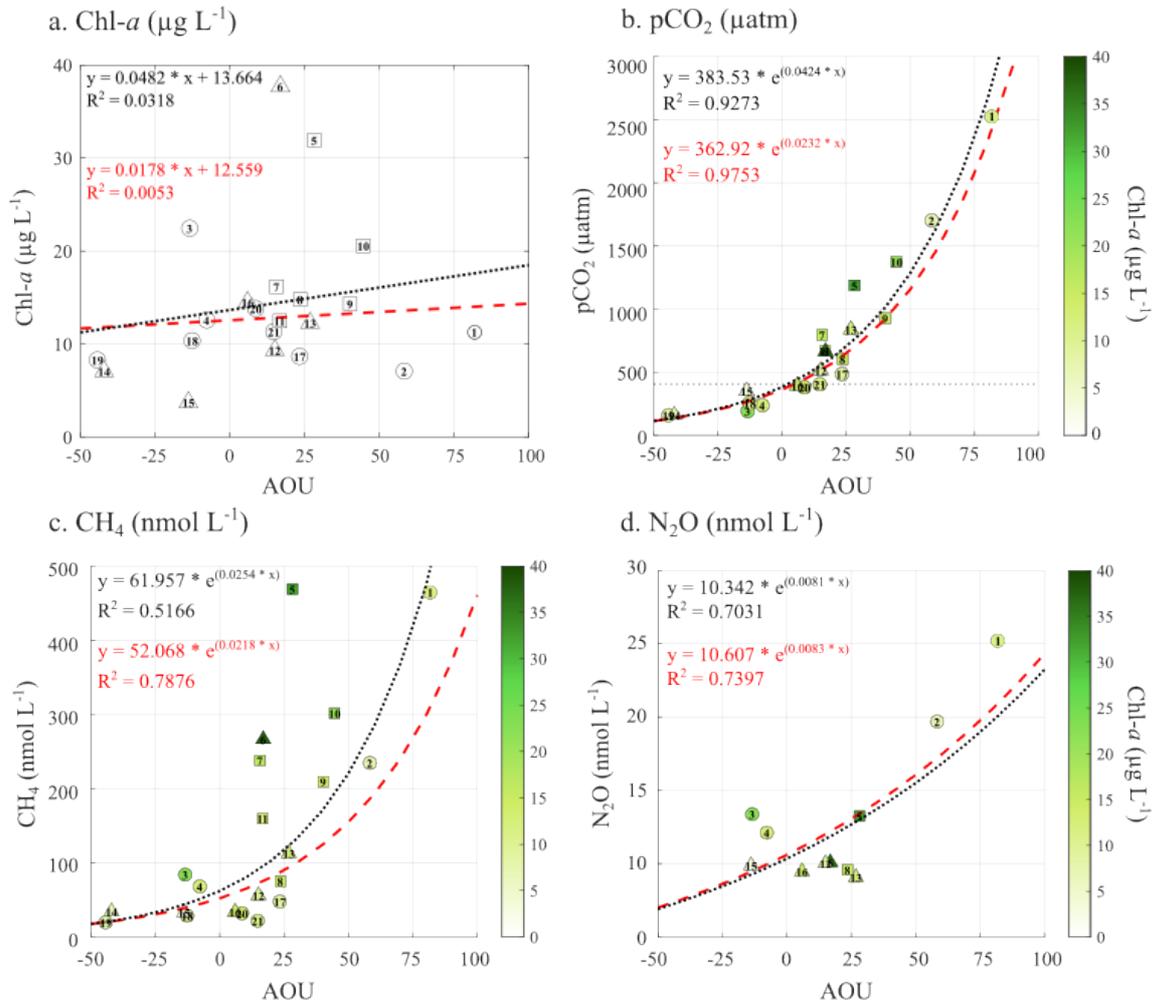


Figure 7: Air-sea fluxes of **a)** CO_2 ($\text{g m}^{-2} \text{d}^{-1}$), and CO_2 -equivalent fluxes of **b)** CH_4 ($\text{g m}^{-2} \text{d}^{-1}$), and **c)** N_2O ($\text{g m}^{-2} \text{d}^{-1}$) estimated using their respective sustained-flux warming potential (SGWPs) over a 100-year time horizon ($\text{CH}_4 = 27$; $\text{N}_2\text{O} = 273$, relative to $\text{CO}_2 = 1$, IPCC 2023), and **d)** the overall budget for all three GHGs ($\text{g m}^{-2} \text{d}^{-1}$).

