**Preparation of test sediment samples**
+ Marine sediments 300 g (wet weight)
+ Milli-Q water 500 mL
(+ 1.0, 0.50, and 0.25 % Na$^+$-Alg aq 20 mL for spike recovery tests)

**Step 1. Enzymatic treatment to remove cellulose, lipids, proteins, and pectin**
+ Amylase 0.80 g (27 ℃, 30 min, pH 6.0)
+ Cellulase 0.80 g (50 ℃, 30 min, pH 6.0)
+ Protease 0.80 g (50 ℃, 30 min, pH 5.5)      The pH was adjusted with
+ Pectinase 0.80 g (38 ℃, 30 min, pH 4.2)   1.0 mol L$^{-1}$ HCl and 3.0 mol L$^{-1}$ NaOH aq

**Step 2. Gelation of Na$^+$-Alg and acidification to remove non-targeted compounds**
+ 10 % CaCl$_2$ aq 100 mL
+ 5.0 mol L$^{-1}$ HCl 15 mL (adjust pH to 1.0)
   (Leave overnight)

Centrifugation and recovery of precipitate

+ 0.10 mol L$^{-1}$ HCl 40 mL to precipitates in each tubes

Centrifugation and recovery of precipitate

**Step 3. Alkaline extraction to dissolve alginate gel**
+ 8.0 % EDTA-2Na aq 500 mL, stir for 1.5 h (pH 4.0)
   (Leave overnight)
+ 3.0 mol L$^{-1}$ NaOH aq (adjust pH to 5.0), stir for 1.5 h
   (Leave overnight)

Centrifugation and recovery of supernatant

**Step 4. Gelation of Na$^+$-Alg into Ca$^{2+}$-Alg**
+ 30 % CaCl$_2$ aq 100 mL
   (Leave overnight)

Centrifugation and recovery of precipitate

**Step 5. Hydrochloric acid washing to remove oligosaccharides**
+ 0.1 mol L$^{-1}$ HCl 20 mL to precipitates in each tubes and shake ←
Centrifugation and recovery of precipitate                  Repeat until almost
                                                            all oligosaccharides
                                                            are removed

**Step 6. Collection of Na$^+$-Alg**
+ 1.0 % NaHCO$_3$ aq 20–100 mL, 60℃ , stir for 1.5 h
   (Leave overnight)

Centrifugation and recovery of supernatant

**Step 7. Prepare the solution for colorimetric analysis**
+ Milli-Q water to dilute 20–100-fold

**Colorimetric analysis by m-hydroxydiphenyl method**

**Figure S1: A complete overview of the modified extraction protocol.**