



Developing an eco-physiological process-based model of soybean growth and yield (MATCRO-Soy v.1): Model calibration and evaluation

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Abstract. MATCRO-Soy is an eco-physiological process-based crop model for soybean (*Glycine max* L. (Merr.)). It was developed by modifying the parameters of MATCRO-Rice. The original model, MATCRO-Rice, integrates crop growth processes with a land surface model. These modifications were made using data from literature and field experiments across the world. The reliability of the model was validated extensively by observed soybean yield data across the global, national, and grid cell levels. A moderate correlation was observed between the MATCRO-Soy and FAOSTAT yield data with correlation coefficients of 0.81 ($p < 0.001$) for the global average yield and 0.512 ($p < 0.01$) for the global average detrended yield over a 34-year period (1981-2014). Furthermore, the grid-cell level validation revealed that 71 % of the grid cells in the global yield map exhibited a statistically significant correlation between the MATCRO-Soy simulated yield and the reference data derived from observational records. These results highlight the model's ability to reproduce soybean yield under different environmental conditions, integrating soil water availability and nitrogen fertilizer. MATCRO-soy could enhance our understanding of crop physiology, especially, crop response to climate change and reduce uncertainty in climate change impacts on soybeans.

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1 Introduction

Crop growth models have been widely used for yield estimation, agricultural management practice optimization, climate change impact evaluation, and informing decision-making about food security strategies (Adeboye et al., 2021; Cuddington et al., 2013; Hoogenboom, 2000). Given the significant impact of weather variability on global yield changes (Müller et al., 2017; Ray et al., 2015), process-based models can represent physiological processes influenced by key climate factors on the long-term impacts of climate on yield productivity (Boote et al., 2013; Cuddington et al., 2013; Fodor et al., 2017; Jones et al., 2017; Marin et al., 2014; Stöckle and Kemanian, 2020). Process-based models explicitly incorporate crucial eco-physiological processes of photosynthesis and stomatal conductance, improving predictions under varying climate scenarios compared with mechanistic crop models that focus on the direct relationship between absorbed radiation and assimilation through radiation use efficiency (Jin et al., 2018). Hence, crop models are useful for capturing the complexity of soil-crop-climate interactions

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for ensuring food security, optimizing yields, promoting sustainability, and planning adaptation strategies (García-Tejero et al., 2011). Global-scale simulations are essential to enhance these efforts by understanding interactions between physiological processes and environmental factors, supporting adaptive management practices and strengthening agricultural resilience.

45 The Agricultural Model Intercomparison and Improvement Project (AgMIP) has examined the performance of global gridded crop models (GGCMs) in simulating the potential impact of climate change on crop yield (Müller et al., 2017; Kothari et al., 2022). AgMIP efforts have demonstrated that the estimated impacts of environmental factors using a GGCM on crop yields generally align with measurements and that a model ensemble reduces uncertainty (Elliott et al., 2015). However, yield change under future climate change scenarios shows inconsistent results and greater variability in soybean than in other crops due because of model discrepancies (Jägermeyr et al. 2021). Despite being a major crop, soybean (*Glycine max* L. (Merr.)),
50 has been studied less extensively than other crops in terms of crop response to changing environments (Ruane et al., 2017 ; Kothari et al., 2022). Therefore, the development of the new soybean model is crucial for reducing uncertainties in climate change impact assessments.

It is important to utilize a diverse type of crop models and ensure model diversity to accurately understand the uncertainties of simulations, as relying on a single model can lead to biased results. To our knowledge, only five process-based models for
55 global-scale soybean yield estimation with leaf-level photosynthesis and stomatal conductance parameters, including LPJ-GUESS (Ma et al., 2022), LPJmL (Wirth et al., 2024), ORCHIDEE-crop (Wu et al., 2016), PRYSBI2 (Sakurai et al., 2014), and JULES (Leung et al., 2020), making this approach remains relatively uncommon. Thus, further development and validation of process-based models that incorporate leaf-level photosynthesis and stomatal conductance parameters are essential.

MATCRO (Masutomi et al., 2016a), is an ecosystem process-based model for crops embedded into the land surface model
60 of minimal advanced treatments of surface interaction and runoff (MATSIRO; Takata et al., 2003) with a crop growth model for rice, which is further explained in Section 2. MATCRO-Rice uses state variables to exchange information (e.g. temperature, soil moisture, transpiration, leaf area index, and photosynthesis rate) between the land surface model and crop growth model. Crop growth mechanisms that consider photosynthesis and stomatal conductance, which are widely used to assess the impact of greenhouse gases on carbon and water fluxes (e.g. ozone) in Masutomi et al. (2019), have been incorporated. Furthermore,
65 MATCRO-Rice has been applied at the regional scale, and it has been utilized to measure climate impacts, which are important for developing adaptation strategies (Kinose and Masutomi, 2020; Masutomi, et al., 2016b).

We developed a new process-based model for soybean (MATCRO-Soy v.1) that incorporates diverse biological processes and environmental interactions that drive plant growth and adaptation to changing conditions. Adapted from MATCRO Rice, this model is applied for soybeans by parameterizing key processes using experimental data and findings from the literature.
70 The current version of MATCRO-Soy (v.1) was evaluated in a global-scale simulation, following a calibration process that considered essential photosynthesis mechanisms. This paper presents the model description in Section 2, the calibration process in Section 3, and the model evaluation in Sections 4 and 5.

2 Model Description

75 MATCRO-Soy is based on MATCRO-Rice, a process-based model of rice growth and yield, which has been modified for use in soybeans. MATCRO-Rice is initially a combined land surface and crop growth model used to explore land-atmosphere interaction in rice fields. Unlike the MATCRO-Rice v.1 version, MATCRO-Soy focuses on yield simulation only and omits the calculation of sensible and latent heat fluxes in the energy balance to reduce computational complexity while maintaining accuracy in simulating soybean growth and yield.



2.1 Overview of MATCRO-Soy

MATCRO-Soy includes three main modules: phenology, photosynthesis, and carbon partitioning (Figure 1). The photosynthesis and carbon partitioning modules are closely linked with the carbon allocation driven by photosynthetic activity. The phenology module serves as a time dimension based on heat unit accumulation and directs the progression of the processes of carbon assimilation and partitioning by monitoring plant developmental stages from sowing to harvest. The phenology module simulates developmental stages based on developmental rate from sowing to harvest and influences key processes such as glucose production and allocation across plant organs. The photosynthesis module includes the absorbed photosynthetically active radiation (PAR) in the leaf canopy following the concept of de Pury and Farquhar (1997) to produce the net primary product (NPP). These photosynthesis products are stored in glucose and starch reserves. The carbon partitioning module distributes the glucose into each organ (i.e. leaf, stem, root, and storage organ) following the method proposed by the school of De Wit used in MACROS (de Vries et al., 1989). MATCRO accounts for leaf senescence as it influences nutrient cycling, crop productivity, and the leaf area index, which plays an important role in canopy photosynthesis. Leaf senescence is also driven by the phenology module. MATCRO incorporates the amount of nitrogen per leaf area (specific leaf nitrogen) as a key determinant of photosynthetic capacity. Root depth can affect photosynthesis indirectly through the plant's ability to access water and nutrients from soil layers, further influencing plant growth within the model framework.

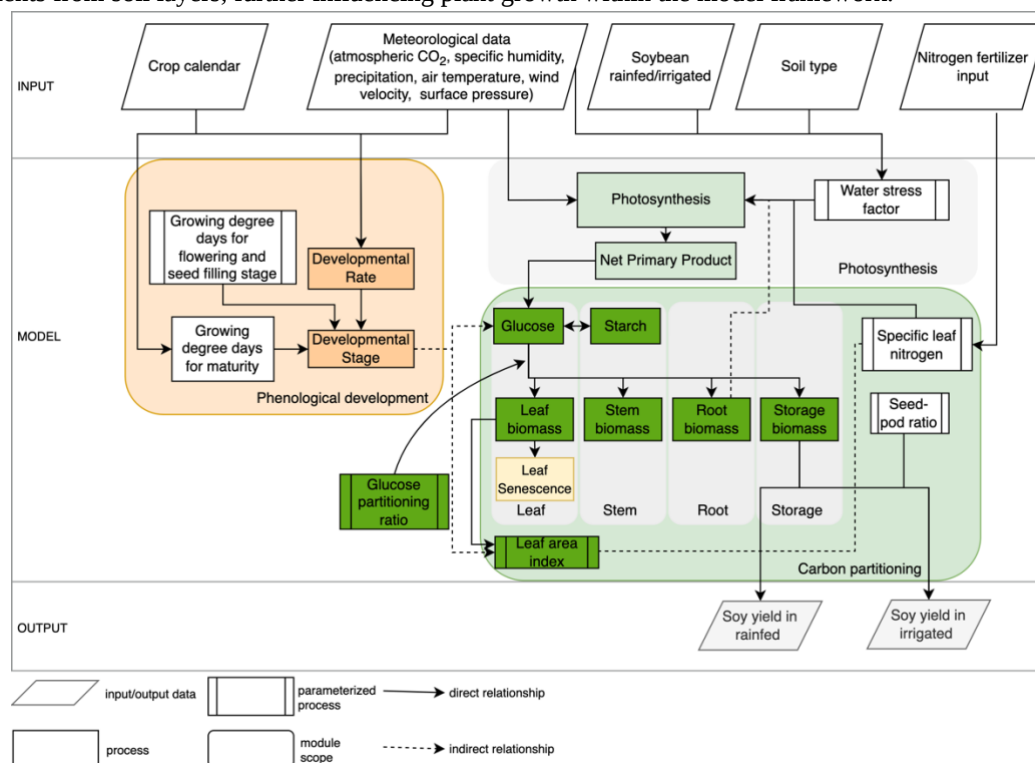


Figure 1. Flowchart diagram of soybean yield simulation by MATCRO-Soy.

The input data consisted of environmental variables obtained from meteorological forcings, soil type classifications, nitrogen fertilizer applications, and agricultural management practices such as irrigation and seed sowing. These inputs are crucial for setting the initial conditions and boundary parameters for the simulations. The output of the MATCRO is the crop yield (kg ha^{-1}) estimated for both irrigated and rainfed conditions on the basis of soil-crop interactions. First, we processed the parameterized growing degree days for maturity using crop calendar data to estimate the harvest time in the phenology module



(see section 2.2). The photosynthesis module includes limiting factors such as nitrogen fertilization and water stress, as detailed in Section 2.3. Then, the module was calibrated (Section 2.4). We conducted a parameterization process encompassing phenological development, carbon partitioning, and photosynthesis limited by water stress and nitrogen uptake. The crop yield was estimated using the parameterized seed-pod ratio (see section 2.5). The adjusted parameters in MATCRO-Soy are described in Section 2.6 where the key dynamic variables are parameterized over time to ensure reliable carbon assimilation in soybean. This comprehensive approach allows MATCRO to account for complex interactions between environmental conditions, crop physiology, and management practices, providing a robust framework for predicting crop yield and assessing agricultural productivity.

2.2 Crop phenological development

Phenological development defines the timing of developmental events based on environmental inputs. MATCRO calculates crop developmental stages (DVS) using an index indicating the sowing time ($DVS=0$) to maturation time ($DVS=1$) on the basis of the integral of the temperature required to exceed the phenological changes. The module uses a formulation based on Bouman et al. (2001) as outlined in Equations (1) to (4).

$$DVS = GDD / GDD_m \quad (1)$$

$$GDD = \int_0^t DVR dt' \quad (2)$$

$$GDD_m = \int_0^m DVR dt' \quad (3)$$

$$DVR = \begin{cases} 0, & T_m < T_b \mid T_m > T_h \\ T_m - T_b, & T_b < T_m < T_o \\ \frac{(T_o - T_b)(T_h - T_i)}{(T_h - T_o)}, & T_o < T_m < T_h \end{cases} \quad (4)$$

where GDD and GDD_m indicate the growing degree days ($^{\circ}\text{C days}$) used to estimate the development of plants during the growing season at time t and at maturity, respectively. DVR represents the developmental rate at t , whereas T_m represents the temperature at t . The parameters T_b , T_o , and T_h ($^{\circ}\text{C}$) are crop-specific and represent the minimum, optimum, and maximum temperatures for crop development, respectively.

The impact of temperature on phenological stages varies for each crop stage as Boote et al. (1998) observed that cardinal temperatures (T_b, T_h, T_o) may differ for vegetative and reproductive stages. We follow Vries et al. (1989) during the growing season to simplify the calculation input and due to the lack of more detailed data in each phenological stage. This study parameterized the developmental stages at the flowering (DVS_f), seed filling (DVS_s), and maturation (DVS_m) stages on the basis of the experimental datasets by calculating the mean, values listed in Table 2. MATCRO uses these DVS parameters to define the period of leaf dry weight loss due to leaf senescence and the remobilization of starch reserves from the stem (Masutomi et al. 2016a). We assume that this phenological time in soybean is in the middle of the flowering and seed filling stage parameterized in this study as leaf loss started within those periods.

2.3 Carbon assimilation process

In the photosynthesis module of MATCRO-Soy, carbon assimilation is based on the leaf-level photosynthesis calculations in sunlit and shaded conditions (Dai et al., 2004) to predict canopy photosynthesis. The calculation includes the stomatal conductance response to relative humidity (Collatz et al., 1991). The net carbon assimilation (A_n) in MATCRO is calculated using the Farquhar model as further described in Masutomi et al. (2016a), expressed in Eq. (5).

$$A_n = f(PAR, P_a, T_{leaf}, CO_{2leaf}, V_{cmax}, BB_a, BB_b) \quad (5)$$

A_n ($\text{mol}(\text{CO}_2) \text{ m}^{-2} \text{ s}^{-1}$) represents net carbon assimilation that contributes to net primary product for biomass growth. It is a function of the intensity of absorbed photosynthetic active radiation (PAR , in $\text{mol}(\text{photon}) \text{ m}^{-2} \text{ s}^{-1}$), air pressure (P_a , in Pa), leaf



temperature (T_{leaf} , in K), CO_2 concentration at the substomatal chamber (CO_{2leaf} , in $Pa(CO_2) Pa(Air)^{-1}$), maximum Rubisco activity (V_{cmax} , in $mol(CO_2) m^{-2} s^{-1}$), the slope (BB_a , in $mol(H_2O) m^{-2} s^{-1}$) and intercept (BB_b , in $mol(H_2O) m^{-2} s^{-1}$) of Ball-Berry model of the relationship between crop assimilation, stomatal conductance per unit leaf area, relative humidity at the leaf surface, and ambient CO_2 concentration (Ball, 1988). In this study, we assume the leaf temperature is the same as air temperature to reduce the complexity of the calculation.

Rubisco activity (V_{cmax}) is a key variable used to assess the carbon rate entering the photosynthetic pathway, as it catalyzes the crucial initial step of RuBP (Ribulose-1,5-bisphosphate) carboxylation in photosynthetic carbon assimilation for C3 plants (Sage, 2002; Xu et al., 2022). In MATCRO, V_{cmax} is calculated as follows:

$$V_{cmax} = V_{ctop} \exp(-K_n LAI) \quad (6)$$

$$V_{ctop} = \max(aSLN^2 + bSLN + c, V_{ctopmax}) \quad (7)$$

V_{cmax} is the Rubisco activity at the top of the canopy ($\mu mol(CO_2) m^{-2} s^{-1}$) limited by the exponential value of vertical distribution of leaf nitrogen (K_n) and leaf area index (LAI , in $m^2 m^{-2}$). We determined the V_{ctop} for photosynthetic rate limited by the specific leaf nitrogen (SLN) in Eq. (7) for soybean using the relationship between two parameters of rubisco activity and leaf nitrogen from experiments summarized from Ainsworth et al. (2014) in the reproductive stage and Qiang et al. (2022) in the vegetative stage. This relationship is empirically represented with a polynomial quadratic equation limited by maximum value of Rubisco activity at the top canopy (V_{ctop} , in $\mu mol(CO_2) m^{-2} s^{-1}$). a , b , c are quadratic coefficient, linear coefficient, and constant respectively from the relationship of both variables where the data has been digitized from WebPlotDigitizer (Rohatgi, 2023).

MATCRO considers nitrogen fertilization input denoted as N_{fert} (unit: $kg(N) ha^{-1}$) which influences the amount of specific leaf nitrogen (SLN , $g(N) m^{-2}$), particularly under conditions of limited nitrogen availability (Menza et al., 2023; Thies et al., 1995). The change in SLN over the growing period follows the study of (Menza et al., 2023) which measured nitrogen fertilization treatments, as described Eq. (8) and (9).

$$SLN = \begin{cases} SLN_{Y0} + \frac{(SLN_{Y1} - SLN_{Y0})(DVS - SLN_{X1})}{SLN_{X1}}, & \text{if } DVS < SLN_{X1} \\ SLN_{Y2} + \frac{(SLN_{Y2} - SLN_{Y1})(DVS - DVS_f)}{(DVS_f - SLN_{X1})}, & \text{if } SLN_{X1} \leq DVS < DVS_f \\ Y + \frac{(Y - SLN_{Y2})(DVS - DVS_s)}{(DVS_s - DVS_f)}, & \text{if } DVS_f \leq DVS < DVS_s \\ SLN_{Y0} + \frac{(SLN_{Y0} - Y)(DVS - DVS_m)}{(DVS_m - DVS_s)}, & \text{if } DVS_s \leq DVS \leq DVS_m \end{cases} \quad (8)$$

$$Y = SLN_{Y3,l} + \frac{SLN_{Y3,h} - SLN_{Y3,l}}{N_{fert,max}} * N_{fert} \quad (9)$$

SLN values vary across different phenological stages, with the developmental stage (DVS) value of soybean growth ranges from 0 (sow) to 1 (harvest). We define DVS_f , DVS_s , DVS_m , and SLN_{X1} as the start of flowering, seed filling, maturity time, and the point where the SLN pattern started to changes with the parameterized values of 0.4, 0.659, 1, and 0.15 respectively. These growth stages are parameterized based on experimental datasets and align with study from Irmak et al. (2013) using the growth stage classification by Fehr and Caviness (1977). SLN primarily depends on nitrogen derived from biological fixation and soil nitrogen, either from natural sources or applied fertilizers. Nitrogen fixation is implicitly captured through SLN that influence V_{cmax} in Eq. (7) and (8), while the effect of applied fertilizers in Eq. (8) and (9).

2.4 Crop growth dynamics

The products of photosynthesis contribute to glucose reserves, which provide energy for growth during various developmental stages. The crop growth dynamics include a carbon biomass partitioning module to calculate the dry weight of each soybean organ (W_{organ} in $kg ha^{-1}$). This variable is the accumulated value of growth rate of dry weight (G_{organ} in $kg ha^{-1} s^{-1}$) during the time from emergence to harvest. Further details on this module can be found in Masutomi et al. (2016a).



$$W_{organ} = f(G_{organ}) \quad (10)$$

175 We calculate the W_{organ} in each soybean organ (i.e. leaf, stem, pod including the seed, glucose reserves and starch). Growth rate of the dry weight (G_{organ}) is calculated based on the parameters of conversion factor of dry weight from glucose to organ ($F_{glu-organ}$ in $kg\ ha^{-1}(kg\ ha^{-1})^{-1}$) for leaf, stem, pod, root, and starch (listed in Table 1), and ratio of glucose partitioned to organ (P_{organ} in $kg\ ha^{-1}s^{-1}$) for shoot, leaf, and pod (listed in Table 2). G_{organ} for each organ and storage, leaf, pod, root, stem, and starch, are expressed below:

$$180\ G_{glu} = f(W_{leaf}, A_{glu}, R_{glu}) \quad (11)$$

$$G_{leaf} = G_{glu} P_{shoot} P_{leaf} F_{glu-leaf} \quad (12)$$

$$G_{stem} = G_{glu} P_{shoot} (P_{leaf} - P_{pod}) \times (1 - f_{starch}) F_{glu-stem} \quad (13)$$

$$G_{pod} = G_{glu} P_{shoot} P_{pod} F_{glu-pod} \quad (14)$$

$$G_{root} = G_{glu} (1 - P_{shoot}) F_{glu-root} \quad (15)$$

$$185\ G_{starch} = G_{glu} P_{shoot} (P_{leaf} - P_{pod}) f_{starch} F_{glu-starch} \quad (16)$$

G_{glu} is the amount of glucose partitioned to soybean organ and reserve derived from function of dry weight of leaf (W_{leaf} in $kg\ ha^{-1}$), net carbon assimilation in glucose form (A_{glu} in $kg(CH_2O)\ ha^{-1}s^{-1}$), and the remobilization from starch reserve in the stem after conversion to glucose (R_{glu} in $kg\ ha^{-1}s^{-1}$). A_{glu} is A_n that has been already converted using the conversion factor from CO_2 to glucose using the value of $1.08 \times 10^6\ [kg\ ha^{-1}\ h^{-1}(mol\ m^{-2}\ s^{-2})^{-1}]$ that is the physical and chemical constant for the conversion.

190 R_{glu} is the remobilization from starch reserve in the stem after converted to glucose using ratio of remobilization value. This R_{glu} is subtracted from the dry weight of starch reserves (W_{starch}). $f_{starch}\ [kg\ ha^{-1}(kg\ ha^{-1})^{-1}]$ is the fraction of glucose allocated to starch reserves calculated in stem dry weight loss. Each growth rate of dry weight (G_{organ}) is calculated based on the parameters conversion factor of dry weight ($F_{glu-organ}$) and ratio of glucose partitioned to organ (P_{organ}) value as follow in Eq (17) – (19):

$$195\ P_{shoot} = \begin{cases} 1 - P_{root}, & \text{if } DVS < 0 \\ \frac{1 - P_{root}(DVS_m - DVS)}{DVS_m}, & \text{if } 0 \leq DVS < DVS_m \\ 1, & \text{if } DVS \geq DVS_m \end{cases} \quad (17)$$

$$P_{leaf} = \begin{cases} P_{leaf0} + \frac{DVS}{DVS_{leaf1}} (P_{leaf1} - P_{leaf0}), & \text{if } DVS < DVS_{leaf1} \\ P_{leaf2} - \frac{(P_{leaf2} - P_{leaf1})}{DVS_{leaf2} - DVS_{leaf1}} (DVS_{leaf2} - DVS), & \text{if } DVS_{leaf1} \leq DVS < DVS_{leaf2} \\ 0, & \text{if } DVS \geq DVS_{leaf2} \end{cases} \quad (18)$$

$$P_{pod} = \begin{cases} 0, & \text{if } DVS < DVS_{pod1} \\ \frac{DVS - DVS_{pod1}}{DVS_{pod2} - DVS_{pod1}}, & \text{if } DVS_{pod1} \leq DVS < DVS_{pod2} \\ 1, & \text{if } DVS \geq DVS_{pod2} \end{cases} \quad (19)$$

The glucose partitioned in each organ is adjusted during the developmental stage using experimental data in the calibration process, further described in Section 3. However, the dry weight of leaf in this module is reduced due to leaf senescence by calculating loss of leaf dry weight (L_{leaf} in $kg\ ha^{-1}\ s^{-1}$) derived from the calibration of partitioned glucose ratio to the ratio of dead leaf (P_{dleaf} in s^{-1}), as outlined in Eq. (20) and (21).

$$200\ L_{leaf} = \begin{cases} 0, & \text{if } DVS < DVS_{deadleaf1} \\ P_{dleaf} (W_{leaf} - W_{glu}), & \text{if } DVS \geq DVS_{deadleaf1} \end{cases} \quad (20)$$

$$P_{dleaf} = P_{deadleaf2} \frac{(DVS - DVS_{deadleaf1})}{(1 - DVS_{deadleaf1})} \quad (21)$$



Then we calculate the leaf area index (LAI) that serves as a parameter to assess the leaf surface area relative to the ground area.

205 It directly influences the plant ability to intercept solar radiation for photosynthesis.

LAI is computed using the adjusted specific leaf weight (SLW in kg ha⁻²) expressed as:

$$LAI = \frac{W_{leaf} + W_{glu}}{SLW} \quad (22)$$

LAI is calculated from the estimated leaf weight (W_{leaf} , in kg ha⁻²) and glucose weight (W_{glu} , in kg ha⁻²) divided with parameterized SLW, a key parameter that links leaf dry weight to surface area. The value of SLW dynamically changed during the developmental stage following exponential relationship:

$$SLW = SLW_{max} + (SLW_{min} - SLW_{max}) \exp(-SLW_x DVS) \quad (23)$$

210 SLW_{max} , SLW_{min} , and SLW_x represent the maximum, minimum, and slope parameters, respectively, that define the values observed in the exponential relationship based on experimental dataset in Table 3. In addition to LAI, photosynthesis is also indirectly affected by the root depth (z_{root} , in m) that determines the plant capacity for water and nutrient uptake. Root depth is calculated as follow:

$$z_{root} = f(r_{root}, z_{rootmax}) \quad (24)$$

z_{root} is the accumulative value from growth rate of root depth (r_{root} in mm day⁻¹) limited by maximum possible root depth ($z_{rootmax}$, in meter).

2.5 Soybean yield estimation

220 The soybean yield is calculated from the pod dry weight at harvest ($W_{podharvest}$) via the seed-pod ratio (SR) in MATCRO-Soy. The yield is further affected by water stress ($f_{wstress}$) in Eq. (25).

$$Yield = f(W_{podharvest}, SR, f_{wstress}, T) \quad (25)$$

The yield was calculated using the parameter SR, which is the ratio of yield (seed, kg ha⁻¹) to the storage organ of the pod ($W_{podharvest}$, kg ha⁻¹) at harvest time and was derived from experimental datasets in Table 3. T is the temperature (Kelvin) that limits heat and cold damage to the yield of soybean. The water stress factor ($f_{wstress}$) was determined on the basis of the fraction of available soil water at the soil layer -i (FAW_i) over crop yield, based on a previous study on the relationship between the soybean transpiration ratio and transpirable soil water conducted by Ray and Sinclair (1998), given in Eq (26).

$$f_{wstress} = \begin{cases} \frac{1}{0.5} FAW_i, & \text{if } FAW_i \leq 0.5 \\ 1, & \text{if } FAW_i > 0.5 \end{cases} \quad (26)$$

230 The value of $f_{wstress}$ depends on soil water availability at soil layer-i (FAW_i), which is the estimated soil water content based on the water flux between the soil layers (Masutomi et al., 2016a) calculated via Eq. (27):

$$FAW_i = \frac{WSL_i - WSL_{wilt}}{WSL_{FC} - WSL_{wilt}} \quad (27)$$

where WSL_i , WSL_{wilt} , and WSL_{FC} represent the water level in the soil layer -i, wilting point, and field capacity, respectively. A value of $f_{wstress}$ equal to 1 indicates no water stress as the fraction of available soil water is adequate for crop growth. Hence, yield is calculated as the potential yield constrained by water stress.

2.6 Soybean-specific parameters

240 MATCRO-Soy shares several parameters with MATCRO-Rice as both are C3 species. However, soybean differs from cereal crops because of its nitrogen-fixing ability. This characteristic is represented through specific leaf nitrogen during the crop growth, as described in Eqs. (8) and (9). The crop-specific parameters reflect the unique physiological and chemical processes involved in soybean growth. but still align with the general framework of MATCRO-Rice. Key parameter adjustments are outlined in **Table 1** as MATCRO employs a set of specific parameters to simulate crop growth and yield.



These parameters include factors related to carbon allocation, root growth characteristics, and crop development based on cardinal temperatures. By accurately representing the unique physiological and biochemical characteristics of soybeans, these parameters contribute to the ability of the model to predict crop yield with greater precision.

245 MATCRO-Soy aims for simulations applicable to a global scale; hence, it uses a single global parameterization as a standardized set of parameters applied worldwide. It uses a unified approach for modelling crop behaviour across different regions. It was assumed that the parameter values from the different treatments and cultivars were independent. Table 2 contains a list of variables parameterized within the model, including the glucose partitioning, nitrogen parameters, and photosynthetic capacity. Through the parameterization of these variables, the model can be adapted for various growing conditions and employed to assess the sensitivity of crop performance to different factors. These parameters are commonly used to evaluate
250 the crop model sensitivity to environmental changes and require further attention, as highlighted by simulations from other crop model as wells (Battisti et al., 2018).

Table 1. Crop-specific parameters used for MATCRO-Soy

Parameters	Description	Value	Units	Source	Eq.
$F_{glu-leaf}$	conversion factor of dry weight from glucose to leaf	0.871	$\text{kg ha}^{-1} (\text{kg ha}^{-1})^{-1}$	Penning de Vries et al. (1989)	(12)
$F_{glu-stem}$	conversion factor of dry weight from glucose to stem	0.810	$\text{kg ha}^{-1} (\text{kg ha}^{-1})^{-1}$	Penning de Vries et al. (1989)	(13)
$F_{glu-root}$	conversion factor of dry weight from glucose to root	0.857	$\text{kg ha}^{-1} (\text{kg ha}^{-1})^{-1}$	Penning de Vries et al. (1989)	(15)
$F_{glu-pod}$	conversion factor of dry weight from glucose to pod	0.759	$\text{kg ha}^{-1} (\text{kg ha}^{-1})^{-1}$	Penning de Vries et al. (1989)	(14)
$F_{glu-starch}$	carbon fraction in the dry matter of starch	0.9	$\text{kg ha}^{-1} (\text{kg ha}^{-1})^{-1}$	Physical and chemical constant	(15)
K_N	vertical distribution of leaf nitrogen	0.11	-	Bonan et al. (2011)	(6)
r_{root}	rate of root depth increase	0.03	mm day^{-1}	Ordóñez et al. (2018) ; Nakano et al. (2021)	(24)
$Z_{rootmax}$	maximum root depth	1.7	m	Penning de Vries et al. (1989)	(24)
T_b	base temperature for crop development	10	°C	Penning de Vries et al. (1989)	(4)
T_h	highest temperature for crop development	34	°C	Penning de Vries et al. (1989)	(4)
T_o	optimum temperature for crop development	27	°C	Penning de Vries et al. (1989)	(4)

255 **Table 2.** Parameterized variables for soybean in MATCRO

Variables	Value	Units	Description
a	-18.516	-	coefficient at relationship of rubisco activity and leaf nitrogen in Eq. (7)
b	114.33	-	coefficient at relationship of rubisco activity and leaf nitrogen in Eq. (7)
c	-73.336	-	constant at relationship of rubisco activity and leaf nitrogen in Eq. (7)
$DVS_{deadleaf1}$	0.6	-	1 st DVS point where the dead leaf ratio pattern changes
$DVS_{deadleaf2}$	1	-	2 nd DVS point where the dead leaf ratio pattern changes
DVS_f	0.4	-	developmental stage on initial flowering stage
DVS_{leaf1}	0.25	-	1 st DVS point where the leaf partitioning pattern changes
DVS_{leaf2}	0.659	-	2 nd DVS point where the leaf partitioning pattern changes
DVS_m	1	-	developmental stage at maturity time



Variables	Value	Units	Description
DVS_{pod1}	0.53	-	1 st DVS point where the pod partitioning pattern changes
DVS_{pod2}	0.72	-	2 nd DVS point where the pod partitioning pattern changes
DVS_s	0.659	-	developmental stage to start seed filling stage
DVS_{SLN1}	0.4	-	1 st DVS point where the specific leaf nitrogen changes along with DVS
DVS_{SLN2}	0.4	-	2 nd DVS point where the specific leaf nitrogen changes along with DVS
DVS_{SLN3}	0.659	-	3 rd DVS point where the specific leaf nitrogen changes along with DVS
f_{starch}	0.18	-	fraction of glucose allocated to starch reserves
SR	0.68	-	seed-pod ratio (SR) accounting harvest index from storage organ
$N_{fert,high}$	300	$kgNha^{-1}$	nitrogen fertilizer value used in high nitrogen fertilizer in Menza et al. (2023)
P_{leaf0}	0.38	-	glucose partitioning ratio of leaf toward shoot in the initial DVS point
P_{leaf1}	0.6	-	glucose partitioning ratio of leaf toward shoot in the 1 st DVS point
P_{leaf2}	0	-	glucose partitioning ratio of leaf toward shoot in the 2 nd DVS point
$P_{deadleaf1}$	0	s^{-1}	dead leaf ratio value in the 1 st DVS point
$P_{deadleaf2}$	0.000001	s^{-1}	dead leaf ratio value the 2 nd DVS point
SLN_{Y0}	0.75	gNm^{-2}	initial specific leaf nitrogen
SLN_{Y1}	2.25	gNm^{-2}	specific leaf nitrogen value in the 1 st DVS point
SLN_{Y2}	1.7	gNm^{-2}	specific leaf nitrogen value in the 2 nd DVS point
$SLN_{Y3,h}$	0.75	gNm^{-2}	specific leaf nitrogen value in the 3 rd DVS point when using high nitrogen fertilizer
$SLN_{Y3,l}$	1.8	gNm^{-2}	specific leaf nitrogen value in the 3 rd DVS point when using low nitrogen fertilizer
SLW_{max}	600	kgm^{-2}	maximum specific leaf weight
SLW_{min}	250	kgm^{-2}	minimum specific leaf weight
SLW_x	2.5	-	exponential slope of specific leaf weight to the developmental stage
$V_{ctopmax}$	103	$\mu mol(CO_2)m^{-2}s^{-1}$	maximum Rubisco capacity at the canopy top in Eq. (7)

3 Model Calibration

The model parameters were tuned to represent the observed phenology and seasonality of biomass development. Once calibration is complete, the model continues to simulate crop growth, which encompasses phenological development, carbon assimilation, assimilate partitioning, and crop yield. We conducted calibrations from various environmental conditions and soybean varieties documented in previous experimental studies as detailed in 3.1 and **Table 3**. The model calibration included parameterizing the dynamic biomass growth for each organ, leaf senescence, and specific leaf weight denoted as P_{organ} during the developmental stage denoted as DVS. Other calibrations using the experimental dataset included the phenological stage, and the seed-pod ratio (SR). The crucial phenological stage (e.g. flowering and seed filling) was calculated as the average value of the reported values in the experimental dataset. MATCRO applies this crop growth module following the method proposed by the school of de Wit used in MACROS (Vries et al., 1989), and compares biomass growth with the observed values during



developmental stages. Shifts in partitioning and growth patterns were identified and used as reference points in the parameterization.

3.1 Description of the site data for calibration

The calibration process used experimental datasets from previous studies collected from field experiments across six different sites in four countries: Frederico Westphalen and Piracicaba (Brazil), Ya'an (China), Champaign (United States of America, US), Morioka and Tsukubamirai (Japan), as seen in Table 3. The soybean cultivars grown at these experimental sites represented different maturity groups. A variety of management practices related to water management and nutrients were utilized in the experiments. Nitrogen fertilizers were applied in most experiments, but mineral nitrogen was used in the soil at sites in Brazil and the US.

Weather data were derived from the records at the meteorological station nearest to the experimental site. The climates at the respective sites were as follows. The ranges of daily mean air temperatures during the growing season was 18-30°C in Frederico Westphalen (Brazil), 19-31 °C in Piracicaba (Brazil), 17-27 °C in Tsukubamirai (Japan), 14-25 °C in Morioka (Japan), 18-26 °C in Ya'an (China), and 15-28 °C in Champaign (US). The seasonal precipitation (mm) for the sites were 1669 mm in Frederico Westphalen (Brazil), 679 mm in Piracicaba (Brazil), 453 mm in Morioka (Japan), 865 mm in Tsukubamirai (Japan), 1012 mm in Ya'an (China), and 787 mm in Champaign (US). The amount of solar radiation also differed among the experimental sites where China received lowest solar radiation and Brazil received highest solar radiation during the experimental period (Supplementary file Figure S1). These data represent diverse climatic conditions in soybean-producing countries. The field data used for calibration were collected across multiple crop seasons, specifically from 2002, 2003 to 2007 and from 2013 to 2016. These time periods were expected to capture the current climatic and environmental variability.

Table 3. Information on field-experimental data of location, crop season, variety, maturity group, water management, and nitrogen fertilizer, as well as the number of experiments for calibrating glucose partitioning ratio and evaluating the soybean yield simulations.

Location	Crop season	Variety (RMG*)	Water management, Nitrogen fertilizer (g N m ⁻²)	Experiments (n)	Reference
Brazil (Frederico Westphalen)	2013-2014	BRS284 (6)	Rainfed, 0	5	(Battisti et al., 2017a)
Brazil (Piracicaba)	2013-2014	BRS284 (6)	Irrigated and Rainfed, 0	6	(Battisti et al., 2017a)
China (Ya'an)	2014	15 cultivars (5-8)	Irrigated, NA	15	(Wu et al., 2019)
	2014-2016	Texuan13 (7), Jiuyuehang (5), Nandou12 (6)		9	
United States (Champaign)	2002, 2004-2007	Pioneer93B15 (3)	Rainfed, 0	8	(Morgan et al., 2005; Ainsworth et al., 2007)
Japan (Tsukubamirai)	2013-2015	Enrei (2), Fukuyutaka (4), Ryuhou (2)	Rainfed, 25-27	16	(Nakano et al., 2021)
Japan (Morioka)	2013-2016	Ryuhou (2)	Rainfed, 25-30	12	(Kumagai, 2018; Kumagai, 2021)

*Relative maturity group

3.2 Biomass partitioning and specific leaf weight

This model represents carbon assimilation by incorporating the carbon fraction in dry matter and glucose allocation to various plant organs. The glucose ratio for each organ is parameterized based on measurements of leaf weight, leaf senescence, stem weight, pod weight, and specific leaf weight across different developmental stages. To simulate glucose partitioning, we used Eq. (17) – (24) to fit the segmented linear models to the experimental dataset (Figure 2 and Figure 4) and used the parameter values as shown in Table 2, as this value is used to obtain the average value of soybean partitioning behaviour. The calibrated glucose partitioning ratio varied across the varieties and environmental conditions and was derived by converting biomass growth into glucose allocation as outlined in Eqs. (11)–(16). However, our parameterization reflected the observation data, as well as the linear growth of leaves and pods during the developmental stages. The dataset from Morioka (Japan) was not included in this biomass partitioning. However, it was utilized for seed-pod ratio and phenology parameterization along with evaluation after the calibration step (Supplementary file Figure S2). The dashed lines in Figure 2 and 3 indicate the estimated flowering and seed filling stages by calculating the average in all experimental datasets.

Carbon assimilation primarily occurs with subsequent allocation to other parts of the plant. Compared with varieties from other sites, the soybean varieties observed in the experimental dataset from Tsukubamirai (Japan) tended to have lower partitioning to the stem during the vegetative stage. The ratio of glucose to leaves in Sichuan (China) was unexpectedly high near maturity in 2016, resulting in partitioning to pods at a low level due to low temperature and drought conditions. The storage organ biomass increases in the reproductive stage to produce pods and seeds, whereas the shoot will senesce at the end of the maturity period. Hence, yield is estimated using seed weight (as determined by the storage organ weight) and the parameterized seed-pod ratio. Pod partitioning in Champaign (US) tended to occur early in pod initiation in early maturation varieties, and the dry weight of pods before the seed filling stage is relatively high (Kawasaki et al., 2018). Early pod initiation has also been observed in the Ryuhou variety in Tsukubamirai in 2013 (Nakano et al., 2021).

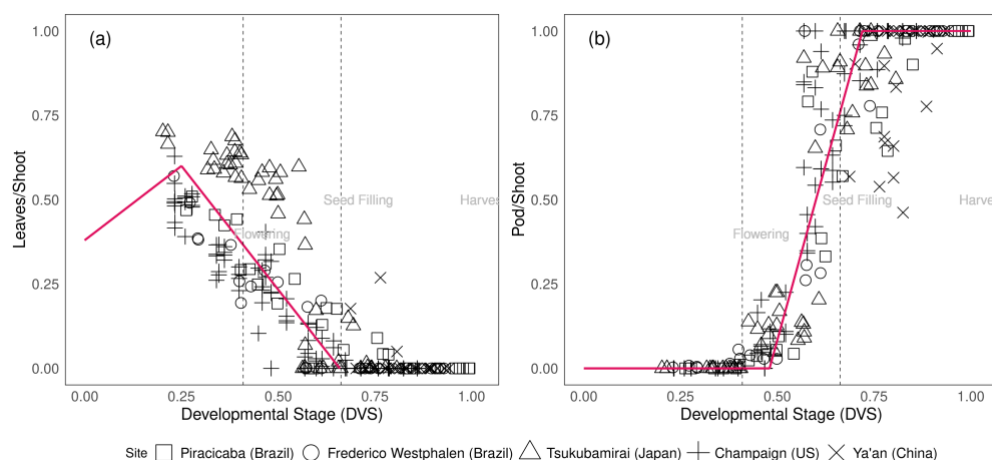


Figure 2. Glucose partitioning ratio to leaves (a) and pod (b) compared with the shoot during the developmental stage (DVS = 0 - 1) in the experimental sites shown by shaped points (square: Piracicaba, circle: Frederico Westphalen, triangle: Tsukubamirai, plus: Champaign, cross: Ya'an). The red lines are the fitted model of segmented lines used for glucose partitioning in MATCRO-Soy. The dashed line marks the averaged flowering, seed filling, and harvest time from the experimental datasets.

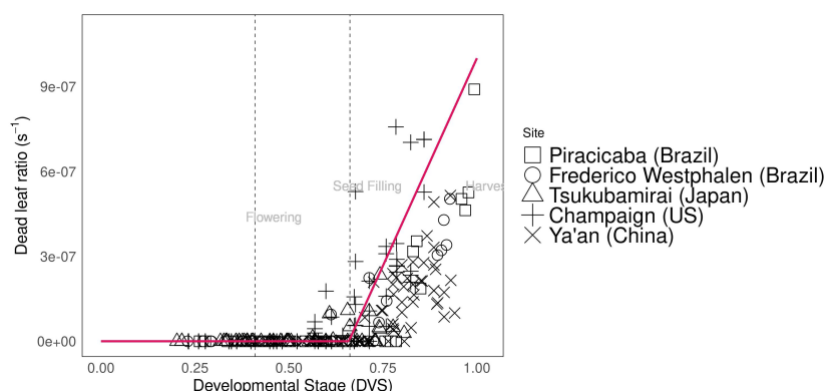


Figure 3. Dead leaf ratio (s^{-1}) during the developmental stage (DVS = 0 - 1). Similar with Figure 2.

The dead leaf ratio parameter in Figure 3 shows the degree of leaf senescence after the seed filling stage due to the leaf process. The dead leaf ratio is calculated from the amount of leaf loss observed during the growing season. The specific leaf weight (SLW) is a significant parameter in crop growth parameterization and has been calibrated to follow the observation data pattern shown in Figure 4. We used the measured leaf weight and leaf area index data from the experimental datasets described in 2.4 and Eq. (23) to calculate the ratio of leaf weight to leaf area (SLW) during different phenological stages. These ratios change over time with distinct values as they vary across different growing seasons and cultivars (Thompson et al., 1996; Slattery et al., 2017). In the figure, SLW from Champaign (US) was excluded because of discrepancies in the timing of the measurements in leaf area and leaf weight biomass. While the specific leaf weight varied among the sites, we fit the model of SLW assuming a saturating exponential function of the developmental stage (red line in Figure 4). This pattern aligns well with the biological process as SLW initially increases due to rapid biomass accumulation but saturates as leaves mature.

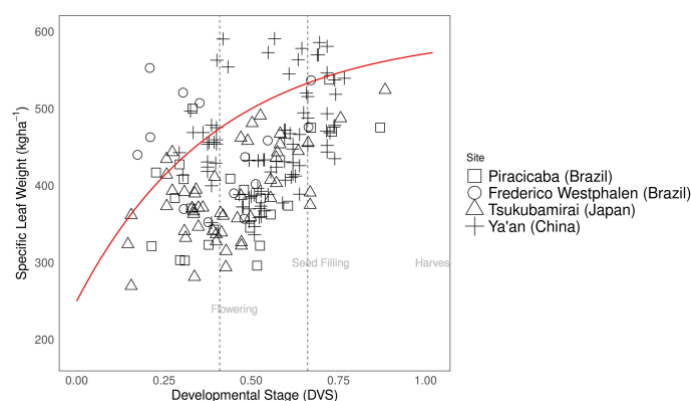


Figure 4. Specific leaf weight ($kg\ ha^{-1}$) during the developmental stage (DVS = 0 - 1). Similar with Figure 2.

4 Model Evaluation Setup

MATCRO was developed in FORTRAN and coupled with the global climate models output, simulated at a spatial resolution of $0.5^\circ \times 0.5^\circ$ and hourly-daily temporal resolution. The output of the model is gridded crop yield ($kg\ ha^{-1}$) as stored in netCDF file format in a global map with one-time harvest simulated in one year. We perform the model evaluation for global, country, and grid cell levels for 34 years (1981 – 2014) at 0.5° spatial resolution and yearly harvested yield output. The



accuracy of the simulated yield was assessed using reference global and country-level data from the Food and Agriculture Organization (FAOSTAT, 2023), while the grid cell level yield was compared with the Global Dataset of Historical Yield (GDHY) data which is derived from statistical records, FAO data, and remote sensing data (Iizumi, 2019).

4.1 Simulation settings and data inputs

The parameters were set as shown in Table 4, covering the period of the sowing year from 1980 to 2014, with a various planting time across different regions. This model incorporated global daily climate data (86400 s) as input data. While the simulation framework was inherited from the established MATCRO-Rice v.1 (Masutomi et al. 2016b), several modifications were made to enhance its applicability at a global scale. Notably, the temporal resolution was adjusted from half-hourly (1800 s) to hourly (3600 s), allowing the model to maintain consistency in capturing critical processes such as diurnal variations in photosynthesis and transpiration, while optimizing computational efficiency. These adjustments ensured that the model remained suitable for large-scale simulations while preserving essential physiological processes.

The model simulates soybean yield using input data as described in Table 5. It uses global input data as follows: crop calendar from the Global Gridded Crop Model Intercomparison (GGCMI), which separates the rainfed and irrigated systems, atmospheric CO₂ and climate data from the Inter-Sectoral Impact Model Intercomparison Project (ISIMIP) that provides bias-adjusted climate input data for historical data (GSWP3-W5E5 v2.0), soil classification from the Harmonized World Soil Database (HWSD v1.2), and nitrogen fertilization for C3 fixing crops of the ISIMIP, which is derived from the land use dataset (Hurt et al., 2020). We use ISIMIP bias-adjusted data to maintain uniformity in the climate impact data across sectors and scales in their framework. This dataset, which is provided by ISIMIP, has a spatial resolution of 0.5°. To determine the growing degree days for maturity, we considered the phenological maturity time from the GGCMI crop calendar for harvest time and global ISIMIP climate data over 10 years (2000-2010) to capture the variability shifts in the current evaluation years.

Table 4. Parameter settings for simulation

Variable	Value	Unit	Description
Year _{sow}	varied	Year	year of sowing day
DOY _{sow}	varied	DOY	day of year of sowing day
RES _{time}	3600	s	time resolution for simulation
RES _{climate}	86400	s	time resolution for climate forcing data
RES _{we/ns}	0.5	degree	spatial resolution north to south or west to east
Soil layer	5.0	-	number of simulated soil layer to calculate soil water content
WSL	1.0	-	soil water content at emergence
W _{leaf0}	1.0	kg ha ⁻¹	dry weight of leaf at emergence
W _{stem0}	1.0	kg ha ⁻¹	dry weight of stem at emergence
W _{root0}	1.0	kg ha ⁻¹	dry weight of root at emergence
W _{glu0}	0.5	kg ha ⁻¹	dry weight of glucose reserve at emergence
Z _a	3.0	m	reference height at which wind speed is observed
Z _{max}	4.0	m	depth of soil layer
Z _t	0.05	m	depth of topsoil layer
Z _b	2.0	m	depth from the soil surface to the upper bound of the most bottom layer of soil

Table 5. Data input for MATCRO simulation.

Variable	Unit	Data source	Spatial Resolution
<i>Daily time-step</i>			
Precipitation	kgm ⁻² s ⁻¹		0.5° × 0.5°



Near-surface specific humidity	kg kg ⁻¹	GSWP3 – W5E5 (Kim, 2017; Cuchi et	0.5° × 0.5°
Maximum, minimum, and mean temperature	Kelvin	al., 2020; Lange, 2019; Lange et al.,	0.5° × 0.5°
Surface downwelling shortwave radiation	W m ⁻²	2021)	0.5° × 0.5°
Near-surface wind speed	m s ⁻¹		0.5° × 0.5°
Surface air pressure	Pa		0.5° × 0.5°
<i>Yearly time-step</i>			
Atmospheric CO2 concentration	ppm	ISIMIP (Büchner and Reyer, 2022)	-
Nitrogen fertilizer	kg ha ⁻¹	ISIMIP (Volkholz and Ostberg, 2022)	0.5° × 0.5°
<i>Constants</i>			
Latitude and longitude	°	-	-
Agricultural management	Irrigated or rainfed	MIRCA2000 (Portmann et al., 2010)	0.5° × 0.5°
Sowing time	Julian day	GGCMI (Jägermeyr et al., 2021)	0.5° × 0.5°
Growing degree days for harvest time	°C days	Parameterized in this study	0.5° × 0.5°
Soil type	-	HWSD (Volkholz and Müller, 2020)	0.5° × 0.5°

4.2 Global yield evaluation methods

In this study, we assessed the statistical relationship between simulated yields and reference data using common metrics of Pearson correlation coefficient (corr) in Eq (28) with the significancy levels (p-values), agreement between the simulation and observation using root mean square error (RMSE) in Eq. (29), and bias in Eq. (30) for the time-series yield data.

$$corr = \frac{\sum_{i=1}^n (X_i - \bar{X})(Y_i - \bar{Y})}{\sqrt{\sum_{i=1}^n (X_i - \bar{X})^2 (Y_i - \bar{Y})^2}} \quad (28)$$

$$RMSE = \sqrt{\frac{1}{n} \sum_{i=1}^n (X_i - Y_i)^2} \quad (29)$$

$$relative\ bias = \frac{1}{n} \sum_{i=1}^n |X_i - Y_i| \times \frac{1}{\bar{Y}} \quad (30)$$

where X_i and Y_i indicated simulated and observed values in each measurement, while $\bar{X}$ and $\bar{Y}$ denotes the mean of simulated and observed values for the harvested year annually. The i and n shows the i -th data point and total number of data, respectively. We use $n = 34$ years for global-scale data, while output after calibration is evaluated in point-scale using n ranged from 14-122 of the available experimental datasets.

Furthermore, we evaluated yield fluctuation on the detrended yield and long-term yield trend separately using mean standard deviation (MSD) and its component to provide more clear interpretation of the model evaluation error (Gauch et al., 2003; Kobayashi and Salam, 2000) in Eq. (31)

$$MSD_y = SB_y + SDSD_y + LCS_y \quad (31)$$

Mean squared deviation (MSD_y) is the square of RMSE for long-term yield trend or detrended yield. Detrended yield is calculated based on the value after yield is reduced by its long-term trend. Its components included mean squared bias (SB_y), difference in the magnitude of fluctuation namely squared difference between standard deviations ($SDSD_y$), and the lack of positive correlation weighted by the standard deviations (LCS_y) as proposed by Kobayashi and Salam (2000) calculated in Eq (32) – (37) below:

$$SB_y = (\bar{X} - \bar{Y})^2 \quad (32)$$

$$SDSD_y = (SD_X - SD_Y)^2 \quad (33)$$

$$SD_X = \sqrt{\frac{1}{n} \sum_{i=1}^n (X_i - \bar{X})^2} \quad (34)$$



$$SD_Y = \sqrt{\frac{1}{n} \sum_{i=1}^n (Y_i - \bar{Y})^2} \quad (35)$$

$$LCS_y = SD_X SD_Y (1 - corr) \quad (36)$$

Higher SB_y , $SDSD_y$, and LCS_y indicate that model failed to simulate mean of the measurement, magnitude of fluctuation around the mean, and pattern of fluctuation across the n measurements, respectively, of the yield. SD_X and SD_Y denotes the standard deviation of simulated (X) and observed values (Y), while LCS_y depends on the correlation coefficient ($corr$).

5 Model Performance Evaluation

We calculated soybean yield in a global-scale map based on the gridded data of irrigated and rainfed area from MIRCA2000 dataset, which represents global agricultural land use around the year 2000 (Portmann et al., 2010), to get the actual yield value. We evaluated yield during the period of 1981-2014 as the MIRCA dataset was available within that period. The simulated yield at the country and global scales for regional comparison was determined by aggregating grid cell data to compute the mean soybean harvested area within each country grid as described below in Eq (37):

$$Yield_{region} = \frac{\sum_{i=1}^n [(Yield_{rf})_i (Area_{rf})_i + (Yield_{ir})_i (Area_{ir})_i]}{\sum_{i=1}^n [(Area_{rf})_i + (Area_{ir})_i]} \quad (37)$$

where $Yield_{region}$ is the aggregated yield at a given region (country or global-scale) in $kg\ ha^{-1}$ from the grid cell number (i) range from 1 to n (total number of grid cells in the region). The estimated yield under rainfed and irrigated conditions are denoted by $Yield_{rf}$ and $Yield_{ir}$, respectively. While the soybean rainfed and irrigated area (ha) used in the simulations are $Area_{rf}$ and $Area_{ir}$, respectively.

5.1 Model output yield as evaluated at the global and national scales

Figure 5a shows a time-series comparison from 1981 to 2014 between the global mean yields reported by FAOSTAT and those simulated by MATCRO-Soy. The results indicated that the model captures the upwards trend in yields over the period with smaller slope compared with the reported yield data. The correlation coefficient is 0.81, which is significant ($p < 0.01$). The errors were $362\ kg\ ha^{-1}$ and 0.15 for the RMSE and relative bias, respectively. Notably, the simulated linear increase contributed to the higher coefficient correlation for the yield trends.

Figure 5b shows the comparison between the detrended global mean yield observed by FAOSTAT and the simulated value by MATCRO-Soy after the long-term linear trend across the study period used in the yield trend was removed. The correlation coefficient decreased to 0.512, which was significant ($p < 0.01$). The model reproduced the interannual variations well with an RMSE of $124\ kg\ ha^{-1}$ and a relative bias of 0.37. Specifically, according to observations, there were significant yield reductions in the years 1983, 1988, 2009, and 2012. Among these, the model successfully reproduced the yield reductions in three years (1983, 1988, and 2012), excluding 2009. These years are reported to have experienced severe droughts, and the model's ability to capture these events is noteworthy.

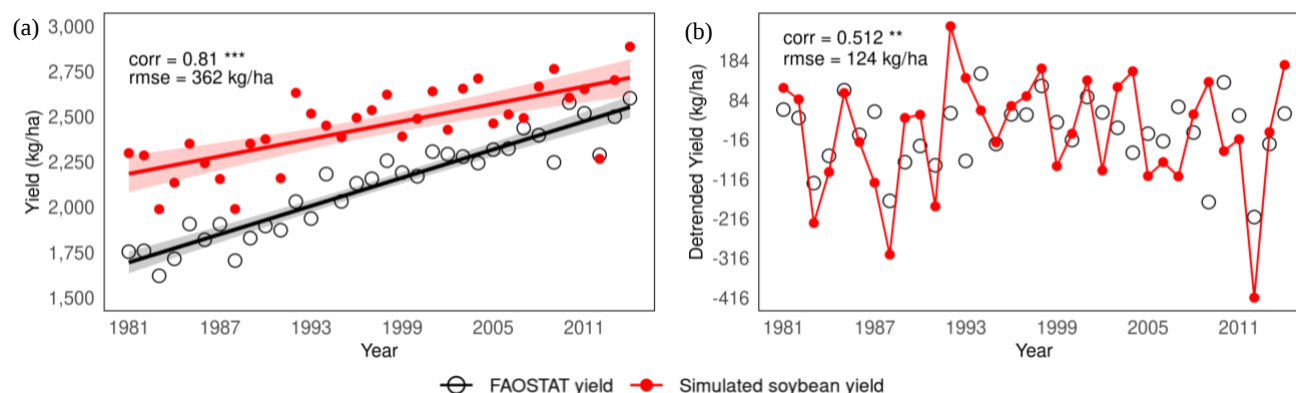


Figure 5. Time-series comparison between simulated yield by MATCRO-Soy and FAOSTAT reported yield data in global long-term trend (a), and detrended (b) yield during 1981-2014. The correlation for detrended yield is calculated after removing the linear trend. The symbols ***, **, and * denote $p < 0.001$, 0.01 , and 0.05 , respectively.

We evaluated the model performance for 10 major soybean-producing countries, Argentina, Brazil, China, India, Paraguay, United States, Italy, Russia, Bolivia, and Canada, consisting of 96% of all global soybean production (based on total average production from 2012 to 2021 in FAOSTAT). Figure 6 compared between the simulated country averaged yields and reported country averaged yields of FAOSTAT for 1981-2014 with the ellipsoid indicating the distribution of the simulated yield values within the 90% confidence range. The results indicate that the model reproduces the national average yield levels well in the top 10 producing countries, as indicated by a correlation coefficient of 0.502 ($p < 0.001$) and an RMSE of 995 kg ha^{-1} . Significant correlation coefficients were observed for six countries (Argentina, Brazil, India, Italy, Paraguay, and the United States; see Supplementary file Figure S3 for further evaluation for these six countries). Focusing on the top three producing countries (the United States, Brazil, and Argentina), which account for 69% of global production, the model's accuracy further improves, with a correlation coefficient of 0.81 and an RMSE of 362 kg ha^{-1} . However, when all countries are considered, the correlation coefficient decreases to 0.263, although it remains statistically significant. These results demonstrate that the model achieves particularly high accuracy in reproducing yields for countries with relatively high production levels.

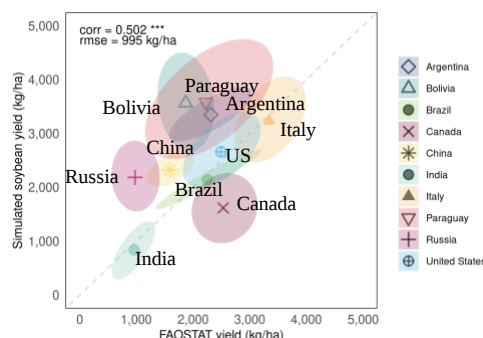
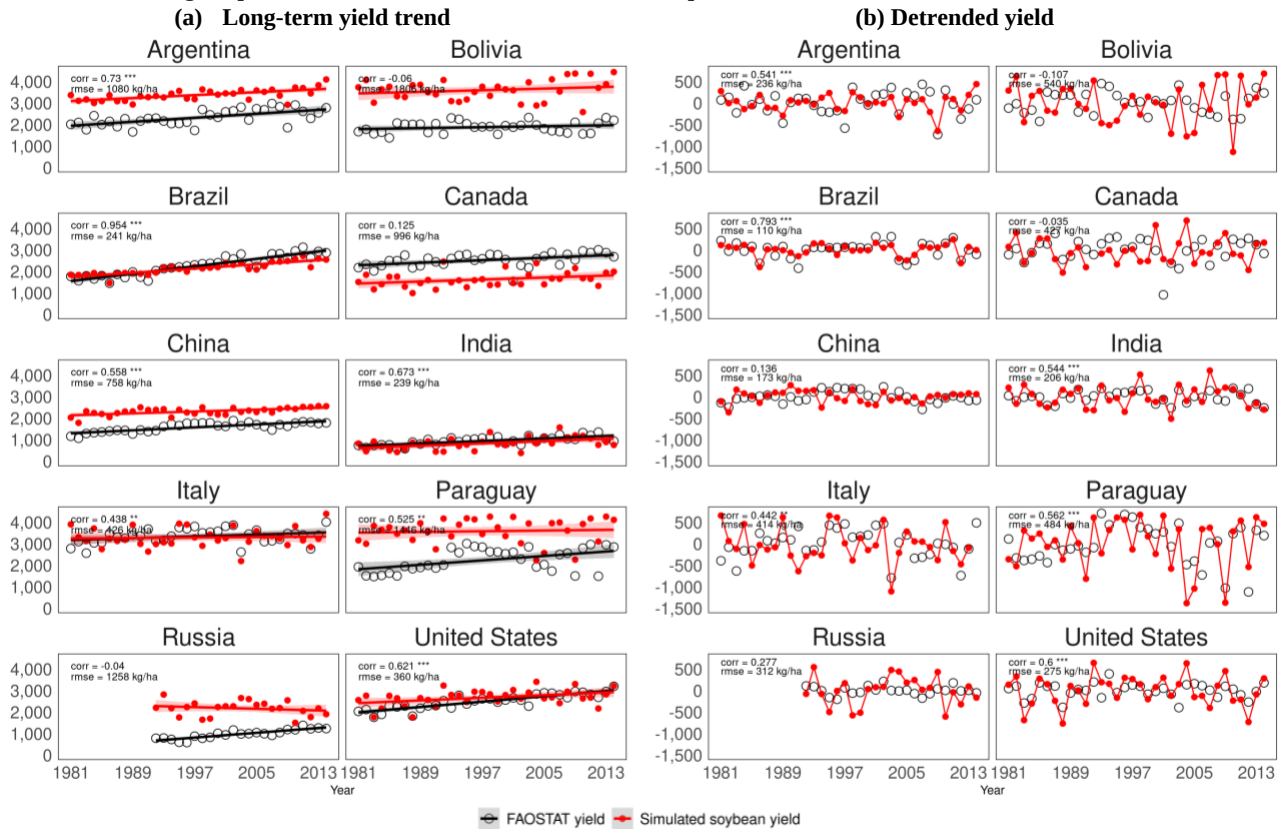


Figure 6. Comparison between simulated yield by MATCRO-Soy and FAOSTAT of the country mean yield during 1981-2014 in 10 major soybean producing countries. Ellipsoid shows 90% confidence range of annual yield.

A time series comparison of country averaged yields focusing on the major producing countries is shown in **Error! Reference source not found.** An evaluation of the long-term trend (Figure 7a) revealed that MATCRO-Soy effectively captured the increasing trend. Brazil demonstrated the strongest agreement, followed by Argentina at 0.73 and the United States at 0.62. For detrended yield (Figure 7b), the interannual variability in Brazil presented the highest correlation coefficient



440 at 0.79, followed that in the United States at 0.6 and that in Paraguay at 0.562. On the other hand, the lowest correlation was observed for China at 0.136 and Bolivia at -0.107. These findings suggest that the model tends to perform with greater accuracy for countries with higher production levels, even in time series comparisons at the national level.



445 **Figure 7.** Time-series comparison between simulated yield by MATCRO-Soy (red circle) and FAOSTAT yield (open circle) in 10 top soybean producer countries during 1981-2014 for long-term yield trend shown by solid line (a) and detrended yield after removing the linear trend (b). The correlation and RMSE based on yield (a) and detrended yield (b) data. The symbols ***, **, and * denote $p < 0.001$, 0.01, and 0.05, respectively. The shading near solid line is the standard error with confidence interval of 95%.

450 5.2 Temporal trends and variability

Model performance was further assessed with the mean squared deviation (MSD) components for the yield and separated by yield, long-term yield trend, and detrended yield for both the global (Supplementary file Table S1) and country scales (Supplementary files Table S2, S3, and S4). Figure 8 shows the source of error based on the MSD components of squared bias (SB), the sum of the difference in standard deviation (SDSD), and the lack of positive correlation (LCS) in the top 6 soybean-producing countries. SBs are the primary source of error in countries with high MSDs: Argentina, China, and Paraguay.

455 The highest MSD in Paraguay was largely driven by SB, with a minor contribution from LCS. In contrast, the lowest MSD in Brazil was largely driven by SDSD and LCS. The SDSD is the primary contributor to the MSD for Brazil and the United States, and a small SB was observed in both countries, indicating that the model effectively simulated the mean yield but poorly captured the trends. These results highlighted the model strength in simulating the mean yield in the top 2 major soybean producing countries (Brazil and the United States) with the largest soybean-growing areas.

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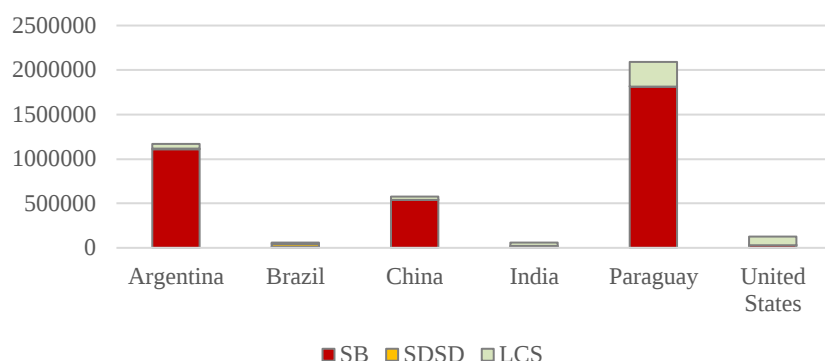


Figure 8. Mean standard deviation components of squared bias (SB), sum of difference in standard deviation (SDSD), lack of positive correlation (LCS) for yield error in top 6 soybean producing countries.

5.3 Model performance at the grid-cell level variation

465 We evaluated MATCRO-Soy at the grid-cell level, by comparing simulated yields with observed ones from Global Dataset of Historical Yield (GDHY) dataset by Iizumi (2019). Figure 9a and b show the simulated and observed yields averaged over 34 years, and Figure 9c shows relative bias between them. Figure 10 shows interannual correlation between simulated and observed yields for 34 years. The simulated yield was calculated for soybean-growing areas from the MIRCA2000 dataset, which offers broad spatial coverage where yield data for certain regions, including Canada, Russia, Australia, and many European and Asian countries, are missing in the GDHY dataset (Iizumi and Sakai, 2020). The density plot of the simulated yield showed more variability than did the GDHY data in Figure 9. However, both datasets exhibited a density peak of approximately 3,000 kg ha⁻¹ and the simulated yield mostly overestimated the yield value. Figure 9 a, b, and c also show the distribution of simulated and observed yields.

475 The relative bias map (Figure 9c) highlights that overestimation was prominent in parts of South America (particularly Argentina) and China, whereas underestimation was observed in South Africa and India. These results aligned with the trends observed at the national scale, which are influenced by the aggregation process. During aggregation, the national-scale results represented the average performance across all grid cells, weighted by the number of grids within each region. Most grids were within a relative bias of -0.2 to 0.2, accounting for 33 % of the total grid area. The grey area was found to be statistically insignificant. The density plot in simulated yield showed more variability compared to the GDHY data. However, both data exhibited the density peak around 3,000 kg ha⁻¹ and simulated yield mostly overestimated the yield value. The correlation between the simulated yield and the GDHY dataset for interannual variation after removing the moving-average (Figure 10) reveals that 71 % of the grid cells are significantly correlated ($p < 0.05$).

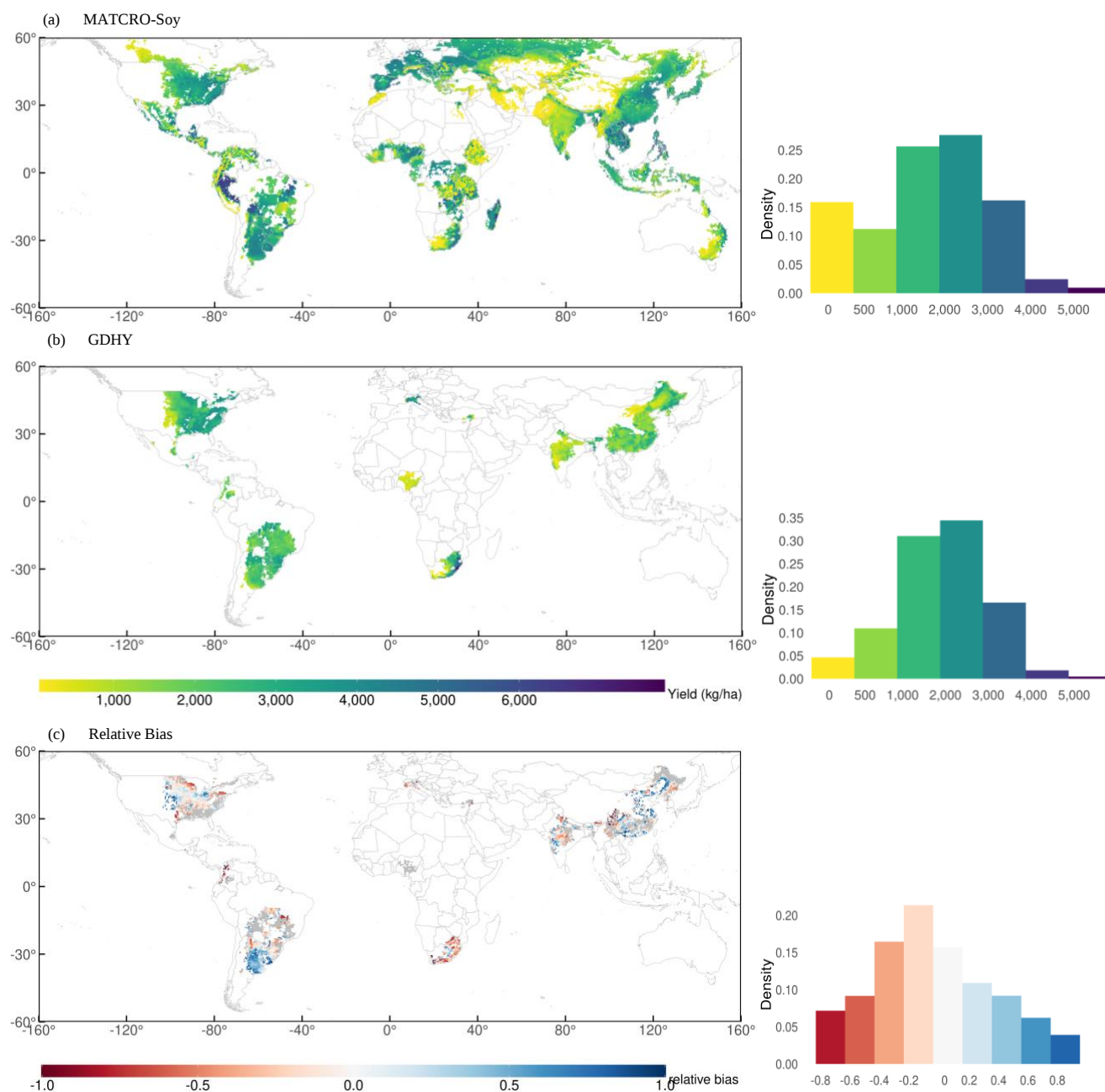


Figure 9. Global map of 34-year averaged (1981-2014) yield of GDHY dataset (a), simulated by MATCRO-Soy (b), and relative bias (c) with each density plot distribution. In figure c, grey colour depicts the correlation with no significance ($p > 0.05$) in the map.

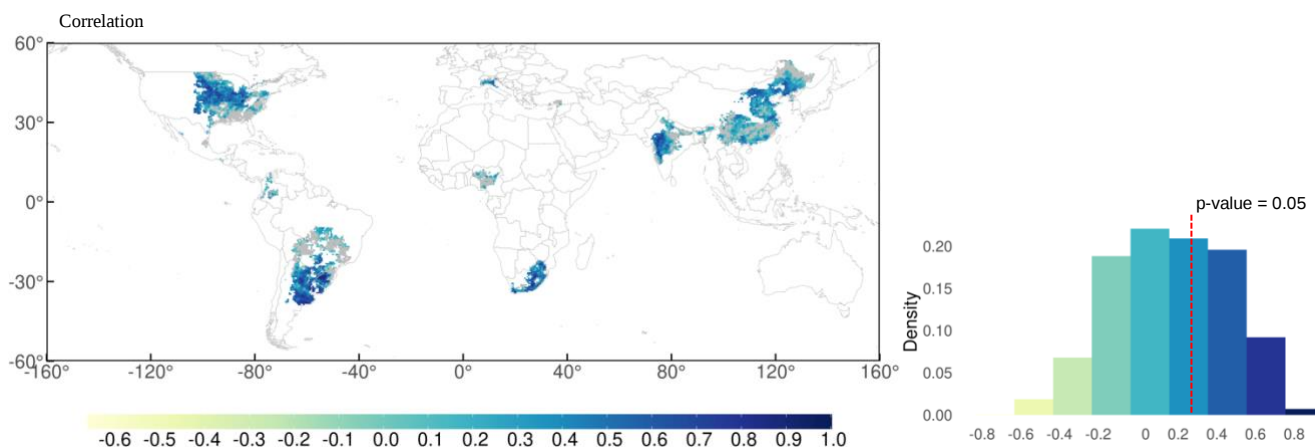


Figure 10. Time-series correlation between simulated and observed yield in 1981-2014 after removing trends from 5-year moving average (c). Grey colour depicts the correlation with no significance ($p > 0.05$) in the map while the red dashed line shows the border of $p = 0.05$ for the number of n year (34) in the density distribution plot.

5.4 Model performance at the leaf-level

We simulated the leaf-level variation in V_{max} for the United States (largest soybean producing country) at the site scale of the Champaign for the 2002 growing season using the global parameterization of MATCRO-Soy (Figure 11). These leaf-level simulated V_{max} values align closely with the observation data from Bernacchi et al. (2005) during the vegetative stage with some deviations during the flowering to seed-filling stages, as shown by the dashed line in the developmental stage of Figure 11. This alignment highlighted the ability of the model to represent essential photosynthetic processes influenced by leaf nitrogen content.

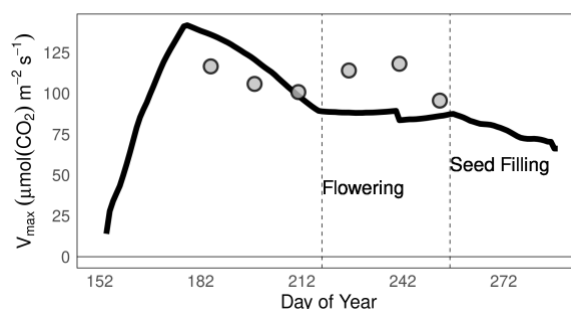


Figure 11. The maximum carboxylation capacity of Rubisco ($\mu\text{mol}(\text{CO}_2)\text{m}^{-2}\text{s}^{-1}$) during the growing period of simulation using MATCRO-Soy (black line) and observation data (grey dots) from Bernacchi et al. (2005) in Champaign (US) year 2002.

6 Discussions

6.1 Validation of MATCRO-Soy

In prior studies, soybean yield predictions often faced challenges in capturing crop responses to climatic variables. The MATCRO-Soy model effectively captures the linear trend in soybean yields, with higher accuracy for long-term trends ($\text{corr} = 0.81$) than for detrended yields ($\text{corr} = 0.512$), as shown in Figure 5. This result of the global detrended yield is improved



compared with that of benchmark studies conducted by Müller et al. (2017), indicating less variation among the process-based models based on its statistical correlation, where another crop model, PRYSBI2, reaches significant correlations of 0.57 ($p < 0.050$) if trends are not removed. However, the accuracy is enhanced when using site-specific parameters are used, as demonstrated in regional scale evaluations from previous studies, which were used for parameterization in this global simulation (Battisti et al., 2017b; Kumagai, 2018, 2021; Morgan et al., 2005; Nakano et al., 2021; Wu et al., 2019). These studies have shown that integrating factors of cultivar differences, ensembles of multiple crop models, nitrogen content, and more accurate measurement method allows for a more reliable representation of local growing conditions and climate variability.

When examining the 10 largest soybean-producing countries, the model performance (Figure 6) has an RMSE of 0.9 ton ha^{-1} (average yield of 34 years), which is comparable with the RMSE of another study using LPJ-GUESS coupled with biological nitrogen fixation (Ma et al., 2022) of approximately 0.8 ton ha^{-1} (average yield of 10 years). The grid-cell level evaluation simulated by MATCRO-Soy, as shown in Figure 9, revealed that 71% of the grid cells were significantly correlated ($p < 0.05$) with most grids falling within 0.2–0.6. These findings align with other studies that show that time-series correlations in GGCM simulated soybean yields range from 0.25 to 0.65 due to discrepancies in benchmark studies (Müller et al., 2017).

The correlation values between yield and detrended yield in Figure 5 and Figure 6 indicate that the increased correlation in model performance was due to the long-term yield trend. MATCRO-Soy could capture the trend of increased atmospheric CO_2 and nitrogen fertilizer inputs, despite of the interannual variability in climate conditions. The MSD calculation revealed that the sum of the differences in the standard deviation (SDSD) was the major contributor error in Brazil, and Italy within the 10 top soybean producing countries (Supplementary file Table S2). Both countries have small squared biases (SBs), suggesting that MATCRO-Soy accurately represents the average productivity in despite of the inability to capture the variability or amplitude of the yield trend over time within the region. Factors such as changes in sowing date, land use, pest management, cultivar maturity group, and planting density may contribute to discrepancies in soybean yield under climate change (Battisti et al., 2018; Marin et al., 2022). Hence, there is a need for improved parameterization to better represent the dynamics of yield variability in countries such as Brazil and Italy.

The high yields in Argentina and Paraguay reflect the consistency of favourable growing conditions (Figure 8a), particularly the alignment of daily temperatures and seasonal precipitation with critical growth stages, suggesting that these regions are less susceptible to interannual variability along with the geographic locations to receive more radiation for photosynthesis sources. The comparison of simulations and observations at the grid-cell level (Figure 11) reveals weak correlations with no statistical significance in high-latitude countries (e.g., Canada and Russia). The models that lack sensitivity to daylength are observed to contribute to more uncertainty (Battisti et al., 2018). Moreover, the low simulated yield in India, which has a hot climate characterized by high mean daily temperatures of 27–28 °C (Supplementary file Figure S4) and low soil moisture during the growing season, highlights the model capacity of the model to capture regional climatic challenges that impact productivity. These climatic challenges likely exacerbate heat stress during critical phenological stages, such as flowering and pod development, leading to reduced yields (Sinclair, 1986; Egli and Bruening, 2004). The contrasting regions of high and low soybean yields underscore the ability of the model to capture the complex interplay between climate and crop yields across diverse agroecological zones.

6.2 Model strength and application

We developed MATCRO-Soy v.1, a process-based eco-physiological model that uses the Farquhar equation to simulate the leaf-level photosynthesis. The Farquhar equation is a widely recognized framework in plant physiology that simulates the biochemical mechanisms of photosynthesis by describing the relationships among light intensity, CO_2 assimilation, and Rubisco enzyme activity (Farquhar et al., 1980; Scafaro et al., 2023). Through the integration of this equation into a gridded global crop model, MATCRO-Soy enhances the simulation of soybean growth and productivity under environmental changes



to atmospheric CO₂, temperature, and water scarcity. These factors are important for predicting and understanding the mechanism of the impact of climate change on productivity. The calibration of MATCRO-Soy successfully represented the response of soybean growth to a wide range of climatic conditions, resulting in reliable global yield simulations using a single parameterization. While simplification may introduce errors, global tuning effectively minimizes these discrepancies in specific regions as this similar result also shown by Smith et al. (2014).

Improving photosynthetic efficiency is one of the key improvements, particularly through enhancing stomatal conductance and modifying Rubisco, the enzyme responsible for carbon fixation (Xu et al., 2022). We used V_{cmax} as a photosynthetic parameter as it quantifies the Rubisco activity that is responsible for catalysing the conversion of carbon dioxide into organic compounds. The peak Rubisco activity observed during the reproductive stage corresponds with trends in specific leaf nitrogen and implicitly affected by the additional nitrogen fertilizer (Menza et al., 2023). The consideration of nitrogen fixation is important as it is sensitive to adverse environmental conditions, flooding, water deficit, and inadequate temperatures, all of which reduce N₂ fixation (Santachiara et al., 2019).

The simulated yield, LAI, aboveground biomass, and pod biomass from MATCRO-Soy were further compared at the point-scale level with experimental datasets with distinct datasets used for each step of calibration and evaluation (**Table 3**) prior to global-scale evaluation (Supplementary file Figure S2). While point-scale simulations employed the unified global parameters, the results demonstrated reasonable agreement with a p value < 0.001 and a bias of 20–60 % for harvested yield, the seasonal leaf area index, aboveground biomass, and pod biomass. The highest bias was observed for the seasonal LAI, which aligns with the underestimation of V_{cmax} during critical growth stages. MATCRO-Soy can reproduce photosynthesis parameters comparable to those of the observation data in site-scale analysis with overestimation in the reproductive stage (Figure 11).

MATCRO-Soy effectively uses high-quality climate data, soil information, and nitrogen fertilizer data to capture biophysical processes involved in soybean growth and yield formation based on previous studies. Its flexibility in spatial resolution enables its application across various scales, from local studies to global assessments. Moreover, the structure of MATCRO is easily coupled with climate models and atmospheric CO₂ to increase the accuracy of yield predictions through high-quality data input. This adaptability also enables integration with other land models, making it a valuable tool in both ecological and agricultural research. MATCRO-Soy can be continuously refined with new data and plant physiological knowledge, ensuring that it remains robust and adaptable. This adaptability makes it a valuable for researchers and policy-makers working towards sustainable agriculture and global food security.

Furthermore, the integration of crop models with remote sensing data will open new possibilities for monitoring and predicting crop productivity at finer spatial scales (Basso et al., 2001). Climate change may shift favourable conditions for high yields in the United States or worsen the challenge of low yields in India for producing soybean yields, making it essential for the model to project these trends accurately for future agricultural planning. In addition to climatic factors, variations in yield may be attributed to technological advancements, shifts in agricultural practices, and changes in crop management strategies that have not been considered in the model. The consideration of more detailed mechanisms of soybean growth can be considered for more accurate results as climate change affects the pest populations (Chen and Mccarl, 2001). However, it is important to acknowledge the limitations of the model, particularly its ability to predict yield variations under extreme or rapidly changing climatic conditions. Continuous updates of the experimental dataset are necessary to maintain its relevance and accuracy in predicting future soybean yields.

6.3 Model challenges and future directions

In the evaluation process, it is important to recognize the interannual variability and spatial variability. There are many grid cells that have a low correlation (nonsignificant) of soybean yield between the simulated and observed values in Brazil when considered in each single cell (Figure 9), but the correlation at the national-scale level is high (Figure 7). This means that local climatic factors affect soybean yield in Brazil. However, MATCRO-Soy is able to recognize broader regional trends leading



to its aim at representing yield behaviour. Figure 12 presents the relative RMSE (RMSE value compared with the observation value) between the simulation and GDHY datasets for the detrended yield at the grid-scale. High relative RMSE values are observed in some parts of South America (particularly in Argentina and Paraguay), the United States, India, and China. Lower relative RMSE values are evident in regions such as Brazil. However, low RMSEs are found in Brazil and the United States at the national-level, although higher relative RMSEs are found in some grid cells of the United States. Detailed information on the spatial variation in the error components contributors is provided in Supplementary File Figure S5 for the long-term yield trend and Figure S6 for the detrended yield. These findings highlighted that the number of grids significantly influences model performance, with regions containing fewer grids being more sensitive to localized factors and spatial heterogeneity during aggregation. These emphasize the importance of considering spatial resolution and representation when evaluating model performance.

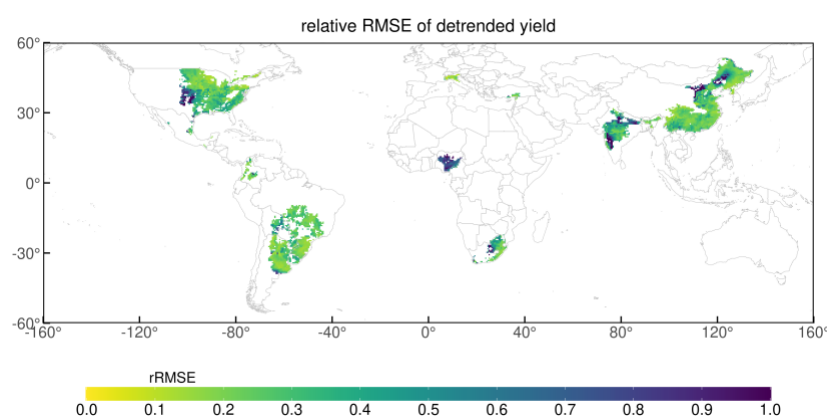


Figure 12. Relative RMSE calculation between simulated and observed yield for detrended yield in grid-cell level.

Uncertainty in MATCRO-Soy is reflected through the challenges in global-scale model evaluation related to the model assumptions of crop cultivars being homogenous globally and the upscaling parameters due to the lack of parameterization, making it is unrealistic to reproduce the variability at the regional-scale with very high accuracy (Müller et al., 2017; Zaehle and Friend, 2010). This uncertainty is notably pronounced in the global aggregation of yield simulations at the grid-cell scale. Global aggregation can escalate substantially for specific combinations of aggregation units, crop model limitations, and years (Porwollik et al., 2017). Future assessments of models and projections of crop yields will require careful consideration of the significant contrast between different aggregation approaches used for individual countries or regions. To address this, we used harmonized ISIMIP data to minimize methodological bias and emphasize the importance of flexible model development for reducing uncertainty (Yin, 2013).

We found a large underestimation in 2002, and overestimation in 2009 when comparing the soybean yield simulated using bias-corrected climate data was compared with FAO data (Figure 5). One possibility for these discrepancies in the interannual variability may be attributed to the influence of unaccounted extreme climatic events. Climatic events indicated by Oceanic Niño index, a three-month running mean of SST anomalies in the Niño 3.4 region, show that La Niña was present at the end of 2002 and that El Niño occurred at the end of 2009 (NOAA, 2024). Some regions within major soybean-producing countries are significantly affected by El Niño events, further influencing yield variability (Anderson et al., 2017; Iizumi et al., 2014). Another possibility for the interannual variation in MATCRO-Soy tends to overestimate the long-term yield trend because of the sensitive effect of the CO₂ concentration on the carbon assimilation module.

The simulated yield increases throughout the year, driven by the positive effects of increased atmospheric CO₂, a phenomenon known as the CO₂ fertilization effect, as observed in studies by Long et al. (2005) and Sakurai et al. (2014). Compared with simulations using statistical radiation use efficiency, process-based models have this tendency because of the



625 greater effect of CO₂ on the photosynthesis process (Ai and Hanasaki, 2023). This result is expected, as most of the simulated yield values were overestimated compared with the reference data, except for the yield in Canada, which was due to the low-temperature conditions.

Comparative studies with other soybean models and refining the MATCRO-Soy on the basis of these findings will contribute to a more comprehensive understanding of its capabilities and limitations. Incorporating additional datasets will
630 further enhance the model representation of real-world conditions. McCormick et al. (2021) suggested that integrating machine learning models could improve accuracy through the calibration process with numerous datasets. However, the use of mechanistic models embedded in MATCRO to simplify the process has proven valuable for understanding and predicting the impacts of environmental factors on agricultural systems. This model can be used to identify potential adaptation strategies, such as changes in planting dates or the development of new crop varieties, to mitigate the adverse effects of climate change
635 on soybean production. However, the application of this model at the field-scale requires high-quality data input and local parameter data.

7 Conclusions

We utilize MATCRO which incorporates carbon assimilation modules based on the C3 photosynthesis of the Farquhar model, to simulate global soybean yield in terms of eco-physiological integrated gridded data inputs of climate, soil type, and
640 nitrogen fertilizer. This study used experimental datasets and literature from previous studies to MATCRO-Soy to represent soybean growth under various environmental conditions. An evaluation of the global mean yield revealed a statistical correlation of 0.81 (p -value < 0.001) between the simulated and reported FAOSTAT data before the long-term yield trend was removed. The correlation value decreased after the long-term yield trend was removed, with a Pearson correlations of 51.2 % ($p < 0.050$), 50.2 % ($p < 0.001$), and 71 % grid cells statistically greater than the significant value ($p > 0.05$) over 34 years
645 (1981-2014) for the global, top 10 countries, and grid cell levels, respectively. The model successfully captured long-term trends and interannual variability, demonstrating its capacity to reflect the impacts of climate factors. Moreover, MATCRO-Soy also modelled reasonable photosynthetic processes in site-scale study, which shows a strong ability to represent the temporal variation. This result highlights the model's reliability and adaptability as a tool for understanding soybean growth and yield dynamics.

650 While MATCRO-Soy presents a valuable framework for understanding the impacts of climate change on global soybean production, many localized factors that influence soybean yield due to the shifts in climate (e.g., pests and diseases) can lead to discrepancies in yield prediction. This highlights the need for high-quality data input. The integration of CO₂ dynamics in MATCRO enhances crop response modelling while providing the carbon fertilization effect in process-based models, warranting further investigation along with the effects of other greenhouse gases. The model may benefit for further refinement, particularly in its treatment of temperature extremes, transpirable soil water, and nitrogen uptake during the photosynthesis
655 process. Integrating MATCRO with other environmental models would enhance its applicability in agricultural management, while emphasizing the necessity for field-scale calibration to improve the model's reliability. MATCRO-Soy provides an opportunity to estimate changes in global soybean production under future land-use or climate change scenarios to address the complexities of climate interactions with agricultural systems. Overall, the MATCRO-Soy has proven to be useful in
660 understanding eco-physiological processes at both the global scale and the country and grid cell levels, providing valuable insights for agricultural management and climate change adaptation.



Code and data availability

This study used the model simulated by source code of MATCRO (Yusara et al, 2025) archived at <https://doi.org/10.5281/zenodo.14881385>.

Author Contributions

TK supervised this study and acquired the funding. YM developed the model source code and supervised this study. AY adjusted the source code for the new parameterization and carried out the analysis and result's figures. RB, LA, YW, SN, and EK provided the data for model calibration. AY prepared the original draft with further editing by YM, TK, LA, YT, KK, RB, EK, SN, and YW. KK supervised the statistical analysis of MSD and provided result's figure. YM, TK, KK, YT, LA, RB, EK, SN, and YW reviewed it. Final manuscript was written by AY and approved by all authors.

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Competing interests. There are no conflicts of interest to declare.

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