

Evaluation of the particulate inorganic carbon export efficiency in the global ocean

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Abstract. The oceanic carbonate pump corresponds to the production and sinking of particulate inorganic carbon (PIC) by calcifying planktonic organisms. In this study, global estimates of PIC standing stock, production derived from ocean colour, and the contribution of calcifying taxa were combined with PIC flux observations from short-term sediment traps deployed over the past decades, covering the global ocean. Coccolithophores are the main calcifying plankton group in the euphotic zone, exhibiting a significant seasonal blooming pattern and a pronounced latitude-dependent seasonal response. The present study highlights that PIC production in the euphotic zone, and the pelagic PIC flux vary among oceanic regions, depths, and seasons. Based on a geographic matchup between PIC flux from sediment traps and remote sensing climatological observations, a correlation between net primary production (NPP) of particulate organic carbon (POC) in the euphotic zone and PIC flux is revealed. However, PIC production in the euphotic zone is not correlated with PIC flux at a global scale, but only in specific ocean basins, such as the North Atlantic and the Southern Ocean. Despite lower PIC production and PIC/POC ratios in the euphotic zone, temperate and subpolar areas are more efficient at exporting PIC compared to equatorial and subtropical regions, which have higher PIC production and PIC/POC ratios in the euphotic zone. Plankton phenology appears to be an important driver of PIC export efficiency (PIC E_{eff}) and PIC transfer efficiency (PIC T_{eff}). This study suggests that the 'packaging factor', which includes the vehicles of the biological carbon pump (e.g., marine snow aggregates, fecal pellets) and the plankton network (e.g., zooplankton communities, microbial loop), largely determines PIC export and transfer efficiencies.

1. Introduction

Through gravitational settling, the biological carbon pump (BCP) transports photosynthetically fixed CO₂ into the deep ocean for decades to centuries to come. Without the BCP, atmospheric CO₂ concentration would be twice as high (Passow and Carlson, 2012). Phytoplankton, through photosynthesis, uptake CO₂ and produce particulate organic carbon (POC). On the other hand, calcifying phytoplankton (such as coccolithophores), produce both POC and particulate inorganic carbon (PIC). Calcification process, often referred to as calcium carbonate (CaCO₃) production, releases CO₂, this is the counter pump effect. Even though calcification contributes to the release of CO₂ (counter effect), all planktonic calcified organisms (such as coccolithophores, foraminifers and pteropods) transport also POC to deep waters through gravitational settling. Field observation of particulate sinking flux (PIC and POC) has been made over many decades to better understand the BCP. To measure a particle flux, the

sediment traps and Thorium-234 activity (^{234}Th activity) are the most widespread techniques, both in terms of time and geography (Savoye et al., 2006, Le Moigne et al., 2014).

35 Coccolithophores contribute to 70-90% of PIC production in the North Pacific Ocean when nutrient and light are available (Ziveri et al., 2023) and more than 90% in the South Atlantic (Kruijt et al., 2026). Coccolithophore are generally considered dominant at global scale (Neukermans et al., 2023). In the global ocean, coccolithophores form mesoscale blooms in high-latitude oceans (Brown and Yoder, 1994; Balch et al., 2005), which are associated with high rates of calcification (Poulton et al., 2007, 2014, Balch et al., 2011; Krumhardt et al., 2017, 2019). The
40 total annual CaCO_3 production by planktonic organisms is characterised by a high uncertainty, with a range of 0.7–4.7 Pg C y^{-1} (Berelson et al., 2007; Buitenhuis et al., 2019; Neukermans et al., 2023). A large proportion of these CaCO_3 produced in the euphotic zone is dissolved within the first 300m of the ocean (Ziveri et al., 2025, Sulpis et al., 2021, Feely et al., 2002; Milliman et al., 1999). This shallow dissolution is not yet clearly explained but considered to be associated with biological and ecological mechanisms such as zooplankton and prokaryotes
45 mediated dissolution (Sulpis et al., 2021, Kwon et al., 2024, Dean et al., 2024). The sedimentation of calcifying organisms constitutes an export flux of CaCO_3 with estimated range of 0.4–1.8 Pg C y^{-1} (Berelson et al., 2007, Neukermans et al., 2023).

At a global scale, particle export efficiency (PE_{eff} , corresponding to the POC sinking flux in the euphotic layer/ POC production) is higher at high latitudes and lower at low latitudes (Henson et al., 2012). The transfer efficiency (T_{eff})
50 corresponds to the proportion of exported organic matter that reaches the deep ocean. T_{eff} is lower at high latitudes and higher at low latitudes (Henson et al., 2012).

Satellite-based estimates of net primary production (NPP) at low latitudes carry substantial uncertainties, potentially biasing PE_{eff} calculations (Henson et al., 2019; Ryan-Keogh et al., 2023; Weber et al., 2016). Recent AI-based models improve the link between deep-ocean particle flux measurements and satellite observations (Picard et al.,
55 2024, 2025). While T_{eff} is not consistently correlated with CaCO_3 export flux (Henson et al., 2012), sediment trap data show that coccoliths and coccospheres are more efficiently transported when incorporated into fecal pellets or marine snow (Honjo, 1976; Pilska and Honjo, 1987; Guerreiro et al., 2021; Liu et al., 2022; Toullec et al., 2022). The “ballast effect,” postulates that biominerals like CaCO_3 and biogenic silica are expected to increase particle density and sinking velocity (Iversen and Ploug, 2010; Laurenceau-Cornec et al., 2020). Hence, BCP is expected
60 to be enhanced by biomineralizing plankton (coccolithophores, diatoms). Nonetheless, the ballast effect remains debated because upper-ocean CaCO_3 export flux is not always linked to particle transfer efficiency, leaving the ballast hypothesis controversial (Henson et al., 2012).

Seasonal influence is an important factor affecting T_{eff} of carbon to the deep sea. During phytoplankton blooms, the exported organic matter is more labile, which likely expect to reduce its transfer efficiency. In low latitudes,
65 which are annually CaCO_3 -productive regions, some modelling and observational syntheses have demonstrated that PE_{eff} tends to be lower while T_{eff} is relatively higher (Henson et al., 2012; Lima et al., 2014, Huang & Fassbender, 2024). However, the most recent studies agree that the structure of the ecosystem is a major driver of PE_{eff} and T_{eff} , even more so than latitude (Henson et al., 2019).

The ‘packaging factor’ theory suggests that CaCO_3 -dominated ecosystems (subtropics and equatorial area) are
70 associated with a complex food web, and CaCO_3 would be more tightly packaged in fast-sinking fecal pellets,

associated with potential ballast effect on the POC (Laurenceau-Cornec et al., 2020). However, particle flux from seasonal, opal-dominated systems (Temperate and sub-Polar ecosystems) would be highly “degradable” formed aggregates, produced by the coagulation of senescent diatoms (Francois et al., 2002). This ‘packaging factor’ as well as the potential associated ballast effect hence should be a strong driver of T_{eff} . Biominerals such as CaCO_3 could be a major driver of POC flux (Lacour et al., 2023) or by physically protecting the more labile POC in aggregates from degradation during gravitational settling (Armstrong et al., 2001).

The biogeographical approach is particularly appealing to understand the structures of plankton communities as well as biogeochemical processes according to the latitude and different ocean basins, also under climate change scenarios (Barton et al., 2013). Indeed, biogeographic patterns are common in macroecology (Kaneko et al., 2023; Thuiller et al., 2015), and BCP understanding (Clements et al., 2023; Ricour et al., 2023; Wang et al., 2023).

In this context, ocean colour data derived from satellite observation is valuable to estimate surface ocean processes over time. Satellite observations of coccolithophore blooms have been available since the emergence of remote sensing of ocean colour techniques (Holligan et al., 1993). Blooms of coccolithophores (e.g., *Gephyrocapsa (Emiliana) huxleyi*) can produce highly reflective patches at the ocean surface and exhibit distinctive optical properties (Balch et al., 1996, 2005; Balch and Mitchell, 2023). These optical signatures can be used to estimate particulate inorganic carbon (PIC) concentrations and production rates at the global scale (Hopkins & Balch, 2018; Hopkins et al., 2019). There remains a gap in our comprehension of processes controlling the transfer of photosynthetically produced organic carbon to the deep (Buesseler et al., 2007; Henson et al., 2012). Nowadays, heterotrophic respiration in sinking aggregates is considered creating a microenvironment supporting dissolution of CaCO_3 in the upper ocean (Morse et al., 2006; Friis et al., 2006; Buitenhuis et al., 2019; Sulpis et al., 2021, Dean et al., 2024). In the deep ocean, dissolution of CaCO_3 is primarily driven by conventional thermodynamics of CaCO_3 solubility with reduced fluxes of CaCO_3 burial to marine sediments. Kwon et al. (2024) showed that without CaCO_3 dissolution in the upper ocean, approximately 20 % more CO_2 would be released to the atmosphere..

Understanding how the surface processes control the export of POC and PIC is still an ongoing challenge in the biogeochemical oceanographic community. In addition, processes such as PIC production, sinking flux, and dissolution are key components of the ocean alkalinity balance, which in turn regulates atmospheric carbon uptake in surface waters (Renforth and Henderson, 2017; Planchat et al., 2023).

This work examines the variability of euphotic zone PIC production and deep PIC flux. In this study, a compilation of existing data sources (e.g. PIC flux from sediment traps, calcified taxa group biomass global estimation) and satellite images, geographically covering the open ocean at different seasonal scales is compared and discussed. The objective of this study is to address the following questions: How spatial and seasonal pattern modulate the PIC export efficiency (PIC E_{eff}) and the PIC transfer efficiency (PIC T_{eff}) at the global scale?

2. Materials and Methods

2.1. PIC production remote sensing-based modelling

PIC production at a global scale was modelled using satellite ocean colour measurements (see Hopkins and Balch, 2018 and reference in here for more details), coupled with physiological constant (growth rate under variable light

intensity and temperature) associated with *Gephyrocapsa (Emiliana) huxleyi*, which is the most cosmopolitan coccolithophore and extensive blooms former (Holligan et al., 1993) across the majority of the world's oceans (Tyrrell and Merico, 2004).

110 2.1.1. Calcification model

The model proposed by Hopkins et Balch (2018) was applied to estimate the calcification rate in the euphotic zone (expressed as EZ PIC production). The model of coccolithophores calcification rate is a function of PIC concentration, growth rate, irradiance, and depth (Equation 1):

$$PIC\ production = f[PIC, \mu, h(surf), g(Z_{eu})] \quad (1)$$

115 In this model, general assumptions are made, such as that PIC production is proportional to the coccolithophores growth rate. The coccolithophores growth rate is a function of temperature and irradiance (parameters established on *G. huxleyi* culture). The calcification rate decreases as a function of light availability through the water column (Hopkins and Balch, 2018). The model generalizes the euphotic zone integrated PIC production rate ($g\ C\ m^{-2}\ d^{-1}$) (Equation 2):

$$120\quad EZ\ PIC\ production = PIC\ concentration \times \mu \times h(I_{surf}) \times g(Z_{eu}) \quad (2)$$

Where μ is a temperature-derived growth rate, $h(I_{surf})$ is a growth limiting irradiance function, and $g(Z_{eu})$ is a depth dependency function (see Hopkins and Balch, 2018 and reference inhere for more details). All the satellite products used in the model are described in the following section.

2.1.2. PIC and POC standing stock

125 Surface PIC and POC satellite-based concentrations were depth integrated to 100 m using empirical relationships based on *in situ* measurements across the Atlantic Ocean (Balch et al., 2018):

$$PIC_{100m} = 40.555 \times PIC_{surface}^{0.560} \quad (3)$$

$$POC_{100m} = 164.376 \times POC_{surface}^{0.617} \quad (4)$$

130 2.1.3. Satellite products collection

The computation was performed with 1° by 1° grid average monthly data (from September 1997 to October 2023) from different sensors merged (Table S1). Satellite ocean color observations are representative the near-surface optical properties (upper 20–40 m in open ocean conditions). The satellite data used in this study (PIC concentration, PAR and Kd_{490nm}) were downloaded from the Globcolour website (<https://hermes.acri.fr/>, last access: 2 February 2024). The Sea surface temperatures were obtained from NASA Ocean Color website (<https://oceancolor.gsfc.nasa.gov/>, last access: 2 February 2024) and the COPERNICUS marine service website (<https://marine.copernicus.eu/fr/>, last access: 2 February 2024). The following product (Table S1) were used in determining the PIC production rate ($g\ C\ m^{-2}\ d^{-1}$): the monthly surface PIC concentration ($mol\ C\ m^{-3}$), the monthly SST (Sea Surface Temperature in $^\circ C$) for the determination of *G. huxleyi* growth rate, the monthly photosynthetic available radiation (PAR; $mol\ photons\ m^{-2}\ d^{-1}$) for the growth rate limiting irradiance, the monthly Kd_{490nm} (Light

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diffuse attenuation coefficient at 490 nm; m^{-1}) used for the determination of the euphotic depth and depth-integrated calcification rate, see details of the methods below. Euphotic zone integrated Primary Production ($mg\ C\ m^{-2}\ d^{-1}$) was obtained from COPERNICUS marine service website (<https://marine.copernicus.eu/fr/>, last access: 2 February 2024, DOI:10.48670/moi-00281), the different variables were obtained from multi-sensors products based on SeaWiFS, MERIS, MODIS-A, MODIS-T, VIIRS-SNPP & JPPS1, OLCI-S3A & S3B). The primary production algorithm was developed by Antoine and Morel (1996).

2.1.4. PIC standing stock evaluation, and taxa contribution

Monthly climatology of depth-integrated coccolithophores PIC standing stock ($g\ C\ m^{-2}$) in the top 100m was obtained from ocean colour remote sensing using empirical relationships between surface ocean PIC concentration and depth-integrated PIC standing stock (eq. 3, Balch et al., 2018). Monthly climatology PIC standing stock for foraminifers and pteropods in the top 200m was obtained from Knecht et al. (2022; see Fig. 4), supplementary data. The carbon biomass concentration for foraminifers and pteropods in the top 200m ($g\ C\ m^{-3}$) was converted into PIC mass concentration (Bednaršek et al., 2012; Knecht et al., 2023; Schiebel and Movellan, 2012). Integrated PIC standing stock ($g\ C\ m^{-2}$) were obtained for foraminifers and pteropods, by multiplying the PIC biomass concentration ($g\ C\ m^{-3}$) by the layer of integration (200m).

2.2. PIC flux

PIC productions in the euphotic zone are expressed in $g\ C\ m^{-2}\ d^{-1}$ (as detailed in section 2.1.1). PIC fluxes for the different depth of integration are expressed in $g\ C\ m^{-2}\ d^{-1}$, considering the deployment duration (in day), the sediment trap collecting surface area (m^2) and the quantity of collected matter (g).

2.2.1. Sediment trap data collection

PIC flux from sediment trap data was obtained from public repositories and published data (<https://www.pangaea.de/>, see Table S2; Fig. 1).

To ensure that the time scale of the PIC flux corresponds as closely as possible to the products derived from the satellites, analysis on a subset of sediment traps deployed 31 days or less (monthly), for a total of 6057 PIC flux observations (sediment traps deployed from 1983 to 2012) were performed. PIC flux observation was then aggregated into seven different layers of depth (0-100m, 100-500m, 500-1000m, 1000-2000m, 2000-3000m, 3000-4000m and > 4000m), as presented in Table 1. Analysis on monthly deployed sediment flux were matched with monthly remote-sensing climatology (see details in the following section). On the other hand, a possible bias associated with the interaction of hydrodynamics, the capture of zooplankton, organic particle remineralization, and PIC dissolution could happen for a longer time of deployment.

Table 1: Layer of the depth of the subset used in the analysis.

Depth layer	Number of PIC flux observation	Number of sediment traps (location according to 1° by 1° grid map)
0-100m	101	84
100-500m	996	62
500-1000m	1175	54

1000-2000m	1405	64
2000-3000m	730	64
3000-4000m	1077	35
> 4000m	573	28

A total of 262 sediment trap locations were used in the analysis. Note that the total of sediment traps for each layer of depth is equal to 391 (multiple sediment traps may have been deployed on the same mooring line), due to the multiple depth deployments at the same geographic location. Moreover, multiple PIC flux observations can be attributed to the same sediment trap deployments (time series, e.g.: PAP, ALOHA station). As for EZ PIC production model output, the analysis and interpretation of observed PIC flux were applied to the open ocean only, excluding data obtained from a water column of less than 200 m depth.

2.2.2. Global data flux estimates

Total global EZ PIC production (Pg C yr^{-1}) was determined by multiplying integrated PIC production by the latitudinally varying area of each 1° by 1° pixel. The latitude variation pixel surface (m^2) was computed using equation 5:

$$Surface (m^2) = \left[\frac{40\,075.017 \times \cos\left(\frac{\pi}{180}(Latitude)\right)}{360} \times \frac{40\,007.864}{180} \right] \times 10^6 \quad (5)$$

Where the equatorial earth circumference = $40\,075.017 \text{ km}^2$ and the meridional earth circumference = $40\,007.864 \text{ km}^2$. The data downloaded were a matrix of 180 pixels of latitude by 360 pixels of longitude (based on cylindrical projection).

The analysis and interpretation of the EZ PIC production model output were focus on the open ocean only, excluding data obtained from bathymetry lower than 200 m depth.

2.2.3. Biogeochemical regions

RECCAP2 biogeochemical regions (second REgional Carbon Cycle Assessment and Processes) aim to accurately assess land and ocean CO_2 sources and sinks through the efforts of hundreds of scientists around the globe (Hauck et al., 2023). The overall aim of RECCAP2 is to support the Global Carbon Project (<https://www.globalcarbonproject.org/>) and the stocktaking of greenhouse gases by providing a reliable scientific basis for the transport of carbon between land, ocean and atmosphere. RECCAP2 biogeochemical regions mask was used to aggregate data from sediment traps and remote sensing according to relevant geographical regions (Fig.1, Table S3).

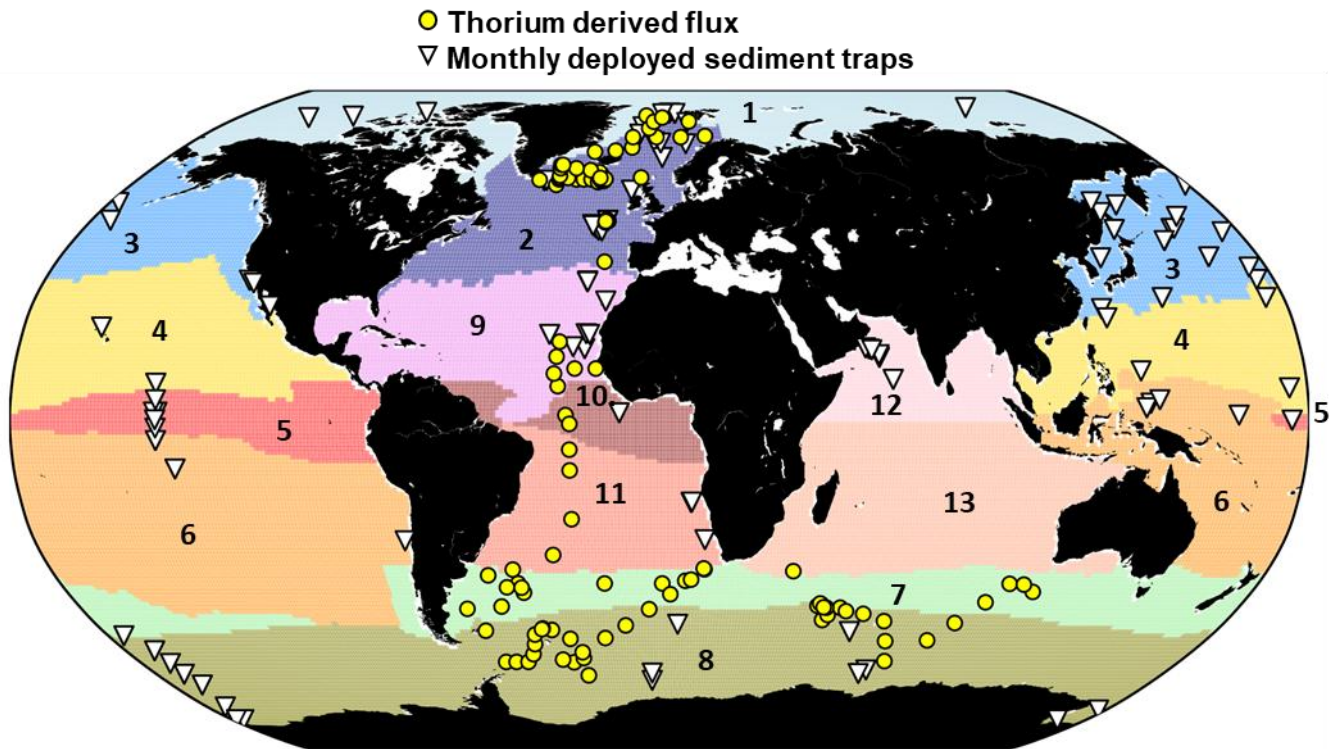


Figure 1: Location of sediment traps measuring PIC flux within 13 biogeochemical regions (RECCAP 2 regions). Region 1 = Arctic (Ar); 2 = North Atlantic (NA); 3 = North Pacific (NP); 4 = North Subtropics Pacific (NSTP); 5 = Equatorial Pacific (EP); 6 = South Subtropics Pacific (SSTP); 7 = Subantarctic (SAZ); 8 = Antarctic (AAZ); 9 = North Subtropics Atlantic (NSTA); 10 = Equatorial Atlantic (EA); 11 = South Subtropics Atlantic (SSTA); 12 = North Indian Ocean (NI) and 13 = South Indian Ocean (SI). The monthly deployed sediment traps are depicted by white triangles and Thorium-derived PIC flux is depicted by yellow dots. 54 stations were out of the RECCAP2 mask (open ocean station) and then have been removed from the 6057 PIC flux observations subset.

2.2.4. Euphotic zone integrated satellite-derived production and deep PIC flux matchup

The majority of sediment trap deployments occurred before the launch of ocean colour observation from satellite, from 1983 to 2012 (SeaWiFS observation started in September 1997). 75.7% of PIC flux data used in the analysis (4588 observations over a total of 6057 observations) were collected before the satellite record. Hence, a match-up between the PIC flux data and EZ PIC production monthly climatology were performed (monthly mean from September 1997 to November 2023).

In the analysis, 7 different layers of depth were aggregated (0-100m, 100-500m, 500-1000m, 1000-2000m, 2000-3000m, 3000-4000m and > 4000m). Discrete observed PIC flux values obtained from sediment traps were aggregated over the depth layer of interest and matched geographically with the monthly climatology-modelled EZ PIC production and NPP on the $1^{\circ} \times 1^{\circ}$ grid. PIC flux values were therefore matched to the same 1° pixel and then associated to the same EZ PIC production. The monthly climatology average of EZ PIC production was established on a period from September 1997 to November 2023 on the $1^{\circ} \times 1^{\circ}$ grid map. Pearson's correlation test was

performed between the monthly climatology average of EZ PIC production and PIC flux values for the depth layers of interest. The process has been performed for the 13 RECCAP2 biogeochemical regions, covering the global ocean.

EZ PIC production values $< 0.1 \text{ mg m}^{-2} \text{ d}^{-1}$ were removed from the dataset to exclude values close to zero or below the detection limit, which may result from a lack of satellite coverage. (e.g. winter months in the north hemisphere above 40°N).

2.2.5. Supporting dataset

1° by 1° grid map of fecal pellets and aggregates contribution to the total particles export were obtained from model ensemble output from Nowicki et al., 2022 (FigShare database: <https://doi.org/10.6084/m9.figshare.19074521>).

3. Results

3.1. NPP, PIC production and residence time seasonality

At the global scale, annual EZ PIC production of $1.65 \pm 0.36 \text{ Pg C y}^{-1}$ was estimated (monthly mean $\pm \sigma$, 1997-2023 annual mean, 302 months of observation), which is congruent with previous estimation based on the same calcification model (ca. $1.42 \pm 1.69 \text{ Pg C y}^{-1}$ in Hopkins and Balch, 2018). The difference between the global estimates reported by Hopkins & Balch (2018) and those obtained in this study may be explained using a merged satellite product (see table S1), which provides broader ocean coverage.

The most NPP annual productive areas are located along the continental margins, above 40°N and within equatorial upwelling ecosystems (Fig. 2a). In contrast to the NPP, the most PIC productive areas are located within subtropical gyres, in the Southern Ocean (along the “Great Calcite Belt”), in the North Atlantic, but not in the Northern Indian Ocean, and less productive within equatorial upwelling ecosystems (Fig. 2b). NPP (POC production, Fig 2) is driven by nutrient availability (nitrate, phosphate, iron). Upwelling zones or high-latitude oceans have high nutrient supply, so phytoplankton production is high. Whereas PIC production, especially by *G. huxleyi*, is favoured in nutrient-poor, stable waters (oligotrophic subtropical gyres), where coccolithophores can thrive and calcify efficiently. The seasonal variation of NPP and PIC production follows the phytoplanktonic bloom phenology, with higher seasonal variation above 40°N and below 40°S (Fig. 2c and d). It is noticeable that the PIC production seasonal bias amplitude is higher than NPP seasonal bias (Fig. 2c and d). Both residence time (the integrated stock divided by the integrated production) and amplitude variation are higher for PIC than for POC (Fig. 2e and f). The amplitude of spatial variation of POC residence time is lower than PIC residence time (Fig. 2e and f). The residence time is obtained by dividing the euphotic layer integrated standing stock (g m^{-2}) by the euphotic layer integrated production rate ($\text{g m}^{-2} \text{ d}^{-1}$). POC residence time highest values reach 20 days in the Southern Ocean and over 20 days in arctic regions (Fig. 2e). However, PIC residence time values reach more than 30 days at high latitudes regions (the Southern Ocean and above 40°C). PIC residence time values over 10 days are also observed within equatorial upwelling (Indian Ocean and West Pacific equatorial). Both PIC and POC residence times reflect a balance between sinking and removal processes such as dissolution or remineralization. For either to sink efficiently, aggregation into marine snow or incorporation into fecal pellets is generally required. PIC observation (ocean colors based) results mainly from free-floating coccoliths, which can accumulate at the surface to a greater extent than

POC. In contrast, POC is readily consumed by bacteria and zooplankton, leading to rapid remineralization in the upper ocean. PIC (CaCO_3), however, is chemically more resistant to biological degradation in surface waters and typically dissolves only under specific conditions, such as in undersaturated carbonate waters, in the deep ocean.

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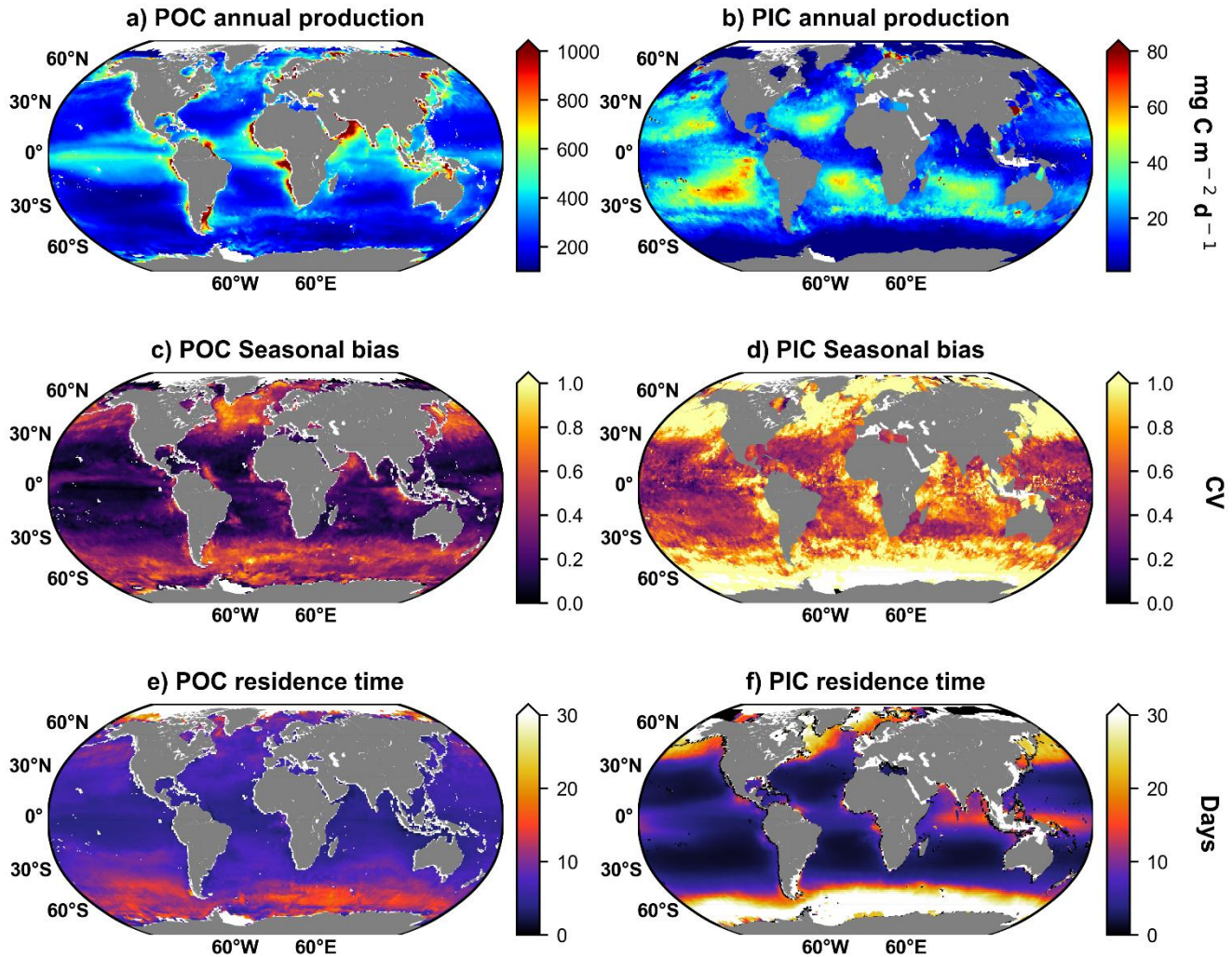


Figure 2: Global maps of (a) POC production (NPP) annual mean ($\text{mg C m}^{-2} \text{d}^{-1}$, 1997-2023 annual mean), (b) PIC production annual mean ($\text{mg C m}^{-2} \text{d}^{-1}$, 1998-2023 annual mean), (c) NPP seasonal bias expressed as coefficient of variation (σ/μ) of monthly NPP climatology, (d) PIC production seasonal bias expressed as coefficient of variation (σ/μ) of monthly PIC production climatology, (e) POC residence time (day, 1997-2023 annual mean) and (f) PIC residence time (day, 1998-2023 annual mean). POC and PIC residence time is obtained by dividing the annual standing integrated stock (mg m^{-2}) by annual production ($\text{mg C m}^{-2} \text{d}^{-1}$), results are expressed in days.

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3.2. Taxa contribution to the PIC standing stock estimation

270 At the global scale, coccolithophores PIC standing stock dominates the total estimated PIC standing stock,
except in few areas of Equatorial Atlantic (EA), Equatorial Pacific (EP), North Atlantic (NA) and North Pacific (NP),
where the pteropods PIC standing stock reach almost 50% (Fig. 3). Foraminifers PIC standing stock represent less
than 10% of the PIC standing stock, with higher value in the North Atlantic (NA). Regarding the seasonal variation
index, coccolithophores are characterised by high seasonal variation in high latitudes ($>30^{\circ}\text{N}$ and $<30^{\circ}\text{S}$), while
275 pteropods' seasonal index is higher only $>30^{\circ}\text{N}$, and in the Equatorial Pacific (EP). Foraminifers tend not to reveal
any seasonal variation at a global scale, except in the southeastern North Atlantic (NA) (Fig. 3). Note that the depth
of integration is different for coccolithophore (100m) and zooplankton taxa (200m for Pteropods and Foraminifers,
from Knecht et al., 2023). Although coccolithophore integration to 100m over 200m for pteropods and foraminifers
leads to an underestimation of coccolithophore contribution to the global PIC standing stock in a 200m layer, the
280 general statement that coccolithophore dominates the standing stock remains unchanged. In addition, the
foraminifers maximum abundance peak between 0-100m depth (Chaabane et al., 2024).

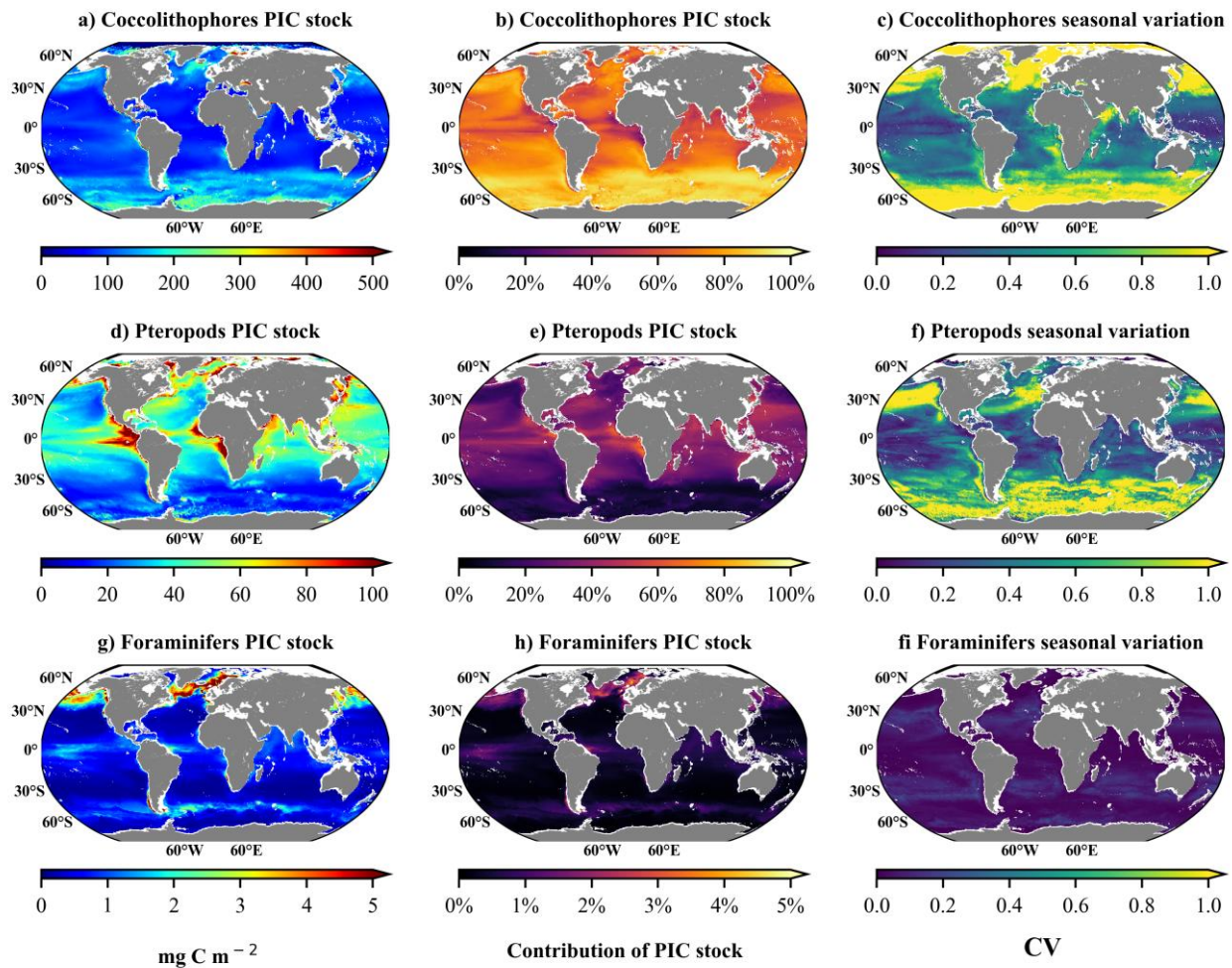
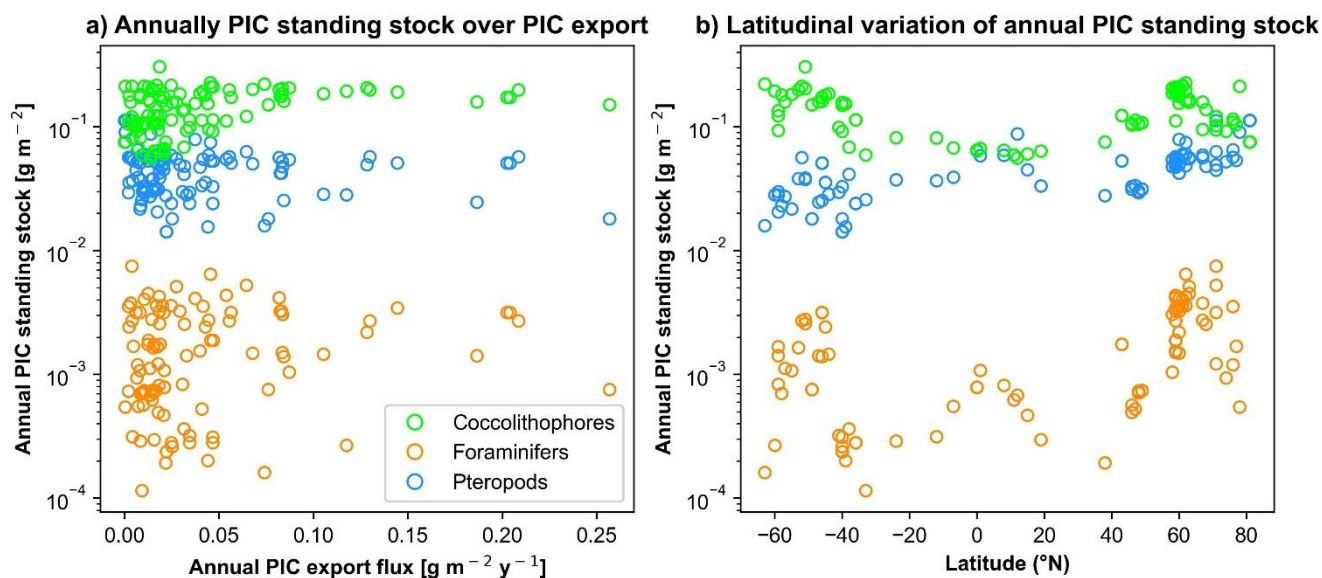


Figure 3: Maps of taxa contribution to the total PIC standing stock regrouping the 3 calcifying taxa: Coccolithophore (a, b and c) Pteropods (d, e and f) and Foraminifers (g, h and i). a, d, and g) Maps of annual PIC standing stock contribution for the 3 calcifying taxa. b, e and h) Maps of annual PIC standing stock for the 3 calcifying taxa. c, f and i) Maps of temporal variability (seasonal index) of PIC standing stock as measured by the seasonal bias (SB) expressed as coefficient of variation (σ/μ) of monthly standing stock for the 3 calcifying taxa. Note that the depth of integration is different for coccolithophore (100m) and zooplankton taxa (200m for Pteropods and Foraminifera, from Knecht et al., 2023).

This study highlights the importance of rapid calcification events. Satellite observations of coccolithophore blooms, which typically last less than 30 days, suggest that PIC fluxes from sediment traps should be integrated over short deployments rather than longer periods. Only including short term sediment traps (less than 30 days), give us a picture of relatively fast sedimentation events, this way, coupling with monthly satellite climatology provides greater meaning regarding potential process involved. Despite pteropods and foraminifers contribute to PIC production and deep flux, their seasonality pattern and residence time were not coupled with PIC flux observation, considering short time deployed sediment traps (less than 30 days). In the other hand, other pelagic contributors (Pteropods and foraminifers) are expected to be more peripheral in the export efficiency of the PIC in this study, regarding their

300 respective residence time (more than 30 days for foraminifers and months to year for pteropods). This aspect is the main constraint in the present study. It is assumed that *G. huxleyi* blooms would be present in the water for an average of 30 days (Hopkins et al., 2015), hence, monthly PIC production climatology coverage map represents the mean monthly conditions. Pelagic calcifiers are characterized by different surface stock and seasonality (Fig. 3) and residence time: less than a month for coccolithophore (Hopkins et al., 2015), a month for foraminifers (Schiebel and A. Movellan., 2012) and months to years for pteropods (Lalli & Gilmer., 1989, Bednaršek et al., 2012).

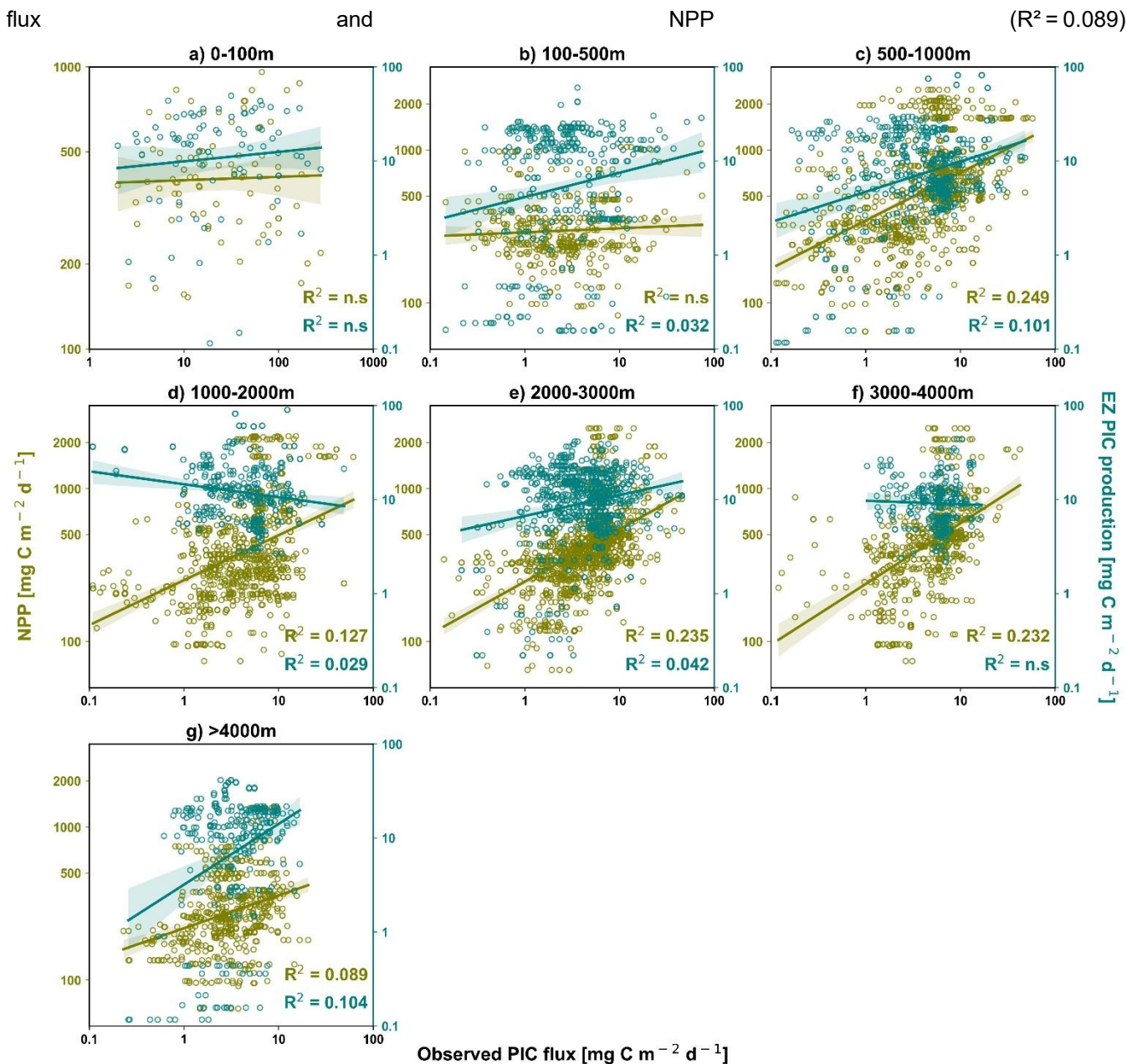
305 The estimate of calcified taxa PIC stock, including coccolithophores, pteropods, and foraminifera, is not correlated with annual PIC export flux on a global scale (Fig. 4a). These estimates show that coccolithophores dominate the PIC standing stock, followed by pteropods and then foraminifera (Fig. 4). The latitudinal variation in PIC standing stock indicates an overlap between coccolithophore and pteropod PIC standing stock at the equator (Fig. 3b, 3e & Fig. 4b). Coccolithophores and pteropods can contribute roughly equally to the PIC standing stock, but coccolithophores dominate at latitudes above 40°.



315 **Figure 4:** a) Calcifying taxa annual PIC standing stock (g m^{-2}) over annual ^{234}Th -derived PIC flux ($\text{g m}^{-2} \text{y}^{-1}$). b) Latitudinal variation of annual PIC standing stock (g m^{-2}) for each calcifying taxa. Only locations where PIC export flux data is shown in this figure. Coccolithophore PIC standing stock is derived from ocean colours (Balch et al., 2005). Pteropods and foraminifers are extracted from Knecht et al. (2023) supplementary data.

3.3. PIC production, NPP and PIC flux

320 No significant correlation is observed between log-transformed PIC flux and log-transformed NPP in the upper 100 m (Fig. 5). On average, although Pearson's correlation coefficients are low ($R^2 < 0.25$), the correlation between PIC flux and NPP is generally higher than that between PIC flux and PIC production (Fig. 5). At the deepest layer (>4000 m), however, the correlation between PIC flux and PIC production ($R^2 = 0.104$) exceeds that between PIC



325 **Figure 5:** Matchup between observed sediment traps PIC flux (x-axis, mg PIC $m^{-2} d^{-1}$) and NPP (mg POC $m^{-2} d^{-1}$) in green, and satellite-derived PIC production (mg PIC $m^{-2} d^{-1}$) in blue (Hopkins et Balch, 2018 model). The different windows correspond to the different layers of depth. Non-significant correlations (p -value > 0.5 , Pearson test) are indicated by "n.s.".

3.4. Correlation between EZ PIC production and deep PIC flux

330 Most of the RECCAP2 biogeochemical regions have no observations of PIC flux or are insufficient to perform
correlation in shallow waters (0-100m depth). PIC flux established in shallow waters (0-100m depth) in the North
Atlantic was collected during the productive period (May, June, July and August) and does not reflect the EZ PIC
production seasonal variation (a non-significant correlation between EZ PIC production and PIC flux has been
established, see Table S4). At the global scale, the linear regression and correlation test between EZ PIC
335 production and PIC flux are displayed for the 6 layers of depth and regarding the RECCAP2 biogeochemical regions
in the Fig. 6 (see Table S4, for the linear regression parameters). At the global scale that the EZ PIC production is
not correlated with PIC flux in the upper ocean. However, considering distinct oceanic bioregions (RECCAP2, Fig.
6), in the mesopelagic layer and deeper, significant correlations between EZ PIC production and deep PIC flux are
observed in the North Atlantic (100 to 4000m) and the Southern Ocean (100-500m and 1000-2000m). North Indian
340 Ocean regions (subtropical areas) are also characterized by PIC production positively correlated with deep PIC
flux (500to 4000m, Fig. 6).

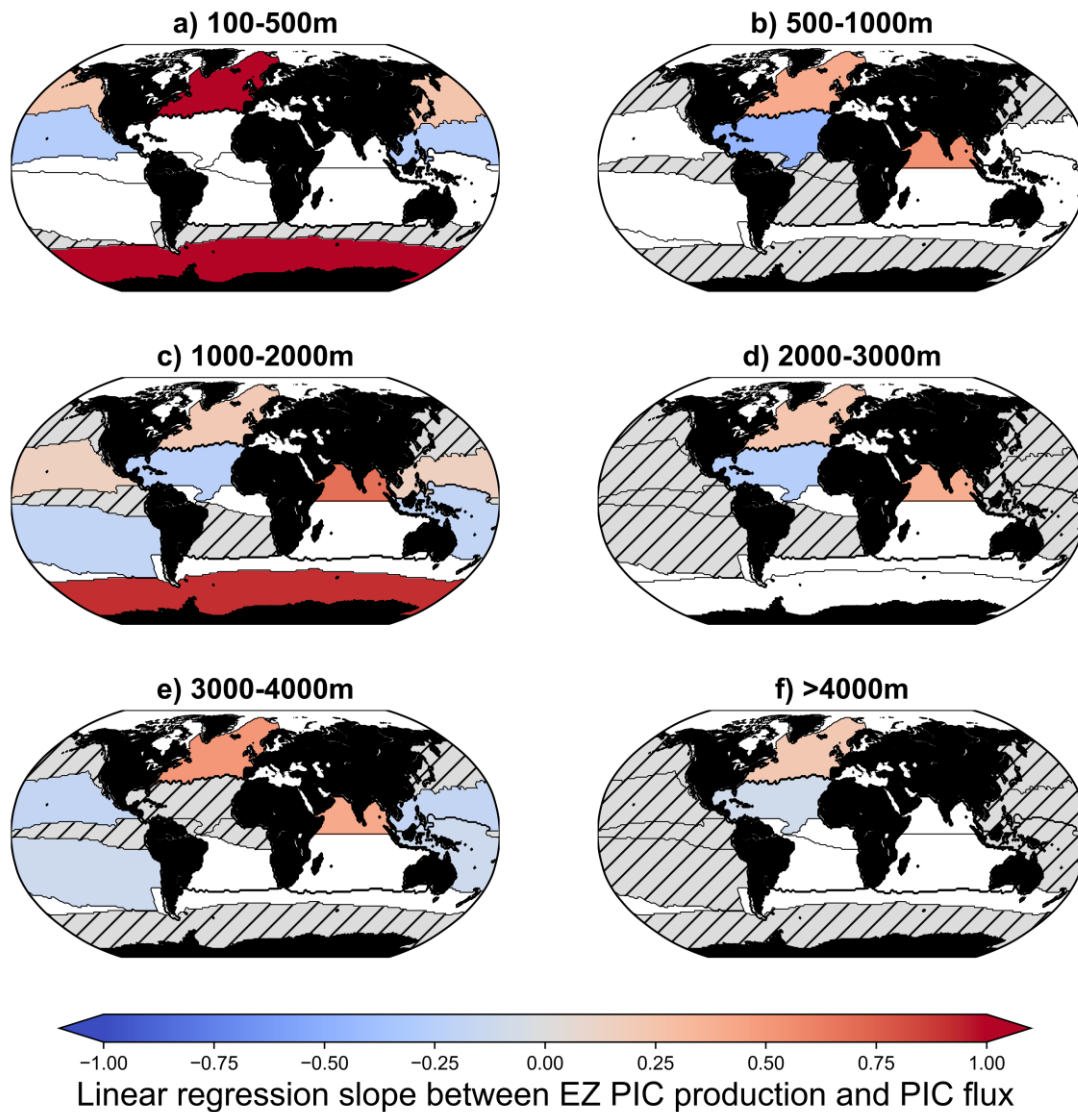


Figure 6: Map of linear regression slope between EZ PIC production and PIC flux according to the RECCAP2 regions and depth layers. Hatched grey areas correspond to regions and/or depths where the correlation coefficient is non-significant at a 95 % threshold (p -value < 0.05). White areas correspond to regions where no data are available.

345

The PIC export efficiency ($PIC E_{eff}$) corresponds to the PIC export flux divided by the EZ PIC production. A latitudinal variability trend could be overlapped to both NPP and PIC production seasonal bias (Fig. 7a). Regions above 40° , are characterized by higher $PIC E_{eff}$ and lower $PIC T_{eff}$ compared to subtropics and equatorial areas (Fig. 7a). The higher PIC production seasonal bias in the euphotic layer coincides with higher normalized $PIC E_{eff}$, but lower $PIC T_{eff}$ (Fig. 7a). The model output of Nowicki et al. (2022), estimated that the contribution to the gravitational carbon pump of zooplankton fecal pellets and sinking phytoplankton aggregates were respectively of 85% and 15%,

350

(Nowicki et al., 2022). Once mapped of 1° by 1° grid map, zooplankton fecal pellets and sinking phytoplankton aggregates contribution are characterized by a significant latitudinal pattern (Fig. 7 in Nowicki et al., 2022). When
355 mapped onto a $1^\circ \times 1^\circ$ grid, the contributions of zooplankton fecal pellets and sinking phytoplankton aggregates exhibit a significant latitudinal pattern (Fig. 7 in Nowicki et al., 2022). The seasonality of NPP and PIC production can be overlapped with aggregates' contribution to the gravitational pump (estimated by Nowicki et al., 2022), as shown in Fig. 7b. The contribution of fecal pellets to the export (estimated by Nowicki et al., 2022) increases continuously from 40° to the equator, while aggregates' contribution to the export decreases. The higher
360 contribution of fecal pellets to the export coincides with low normalized PIC E_{eff} , but high PIC T_{eff} (Fig. 7b).

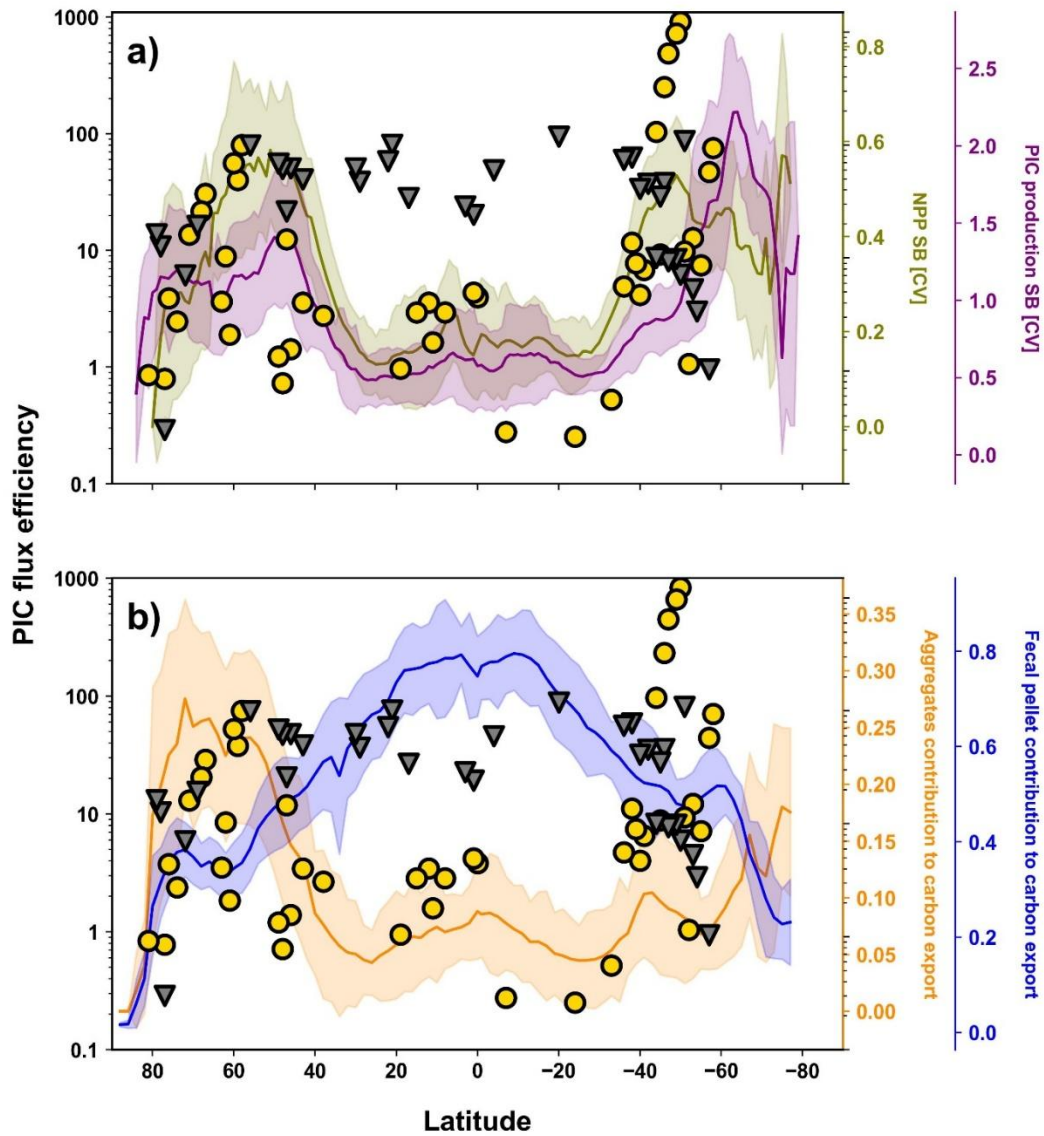


Figure 7: a) Annual PIC production seasonal bias (C.V) in purple and NPP seasonal bias (C.V) in green. Depicted in the left y-axis: The yellow circles represent the latitudinal annual mean PIC Export efficiency (PIC E_{eff}) from shallow sediment traps (0-100m) and ^{234}Th -derived flux over the latitude (x-axis). The grey triangles represent the latitudinal annual mean PIC Transfer efficiency (PIC T_{eff}) from mesopelagic sediment traps (100-1000m). b) Annual Fecal pellets contribution to the total carbon export (Nowicki et al., 2022) in blue and Annual aggregates contribution to the total carbon export (Nowicki et al., 2022) in orange. Depicted in the left y-axis: The yellow circles represent the latitudinal annual mean PIC Export efficiency (PIC E_{eff}) from shallow sediment traps (0-100m) and ^{234}Th -derived flux over the latitude (x-axis). The grey triangles represent the latitudinal annual mean PIC Transfer efficiency (PIC T_{eff}) from mesopelagic sediment traps (100-1000m).

370 4. Discussion

4.1. Mesopelagic PIC Flux and Ballast Effect Hypothesis

The ballast hypothesis originates from correlations between POC flux and mineral fluxes (opal and CaCO_3) in deep sediment traps (Klaas and Archer, 2002). However, CaCO_3 export flux in the upper ocean does not correlate with transfer efficiency (Henson et al., 2012), suggesting that CaCO_3 does not significantly protect POC from degradation at mesopelagic depths. Ecosystem structure, rather than mineral ballast, might be the primary control on the biological carbon pump. François et al. (2002) proposed the “packaging factor” theory, suggesting that highly CaCO_3 -productive systems also contain organisms that produce sinking fecal pellets, which efficiently deliver organic carbon to deep waters (e.g., Nowicki et al., 2022). In subtropical and equatorial upwelling regions, export flux is not always associated with mineral ballast (Le Moigne et al., 2014), highlighting spatial variability in biomineral inclusion and supporting the role of ecosystem structure and phytoplankton phenology. On a global scale, our results demonstrate that EZ PIC production is not correlated with PIC flux in the upper ocean. However, in specific bioregions (RECCAP2), significant correlations exist between EZ PIC production and deep PIC flux (North Atlantic, Southern Ocean, and North Indian Ocean; Fig. 6; Table S4). These observations suggest that ecosystem structure and phenology are more important than the ballast effect in controlling PIC E_{eff} and PIC T_{eff} .

385 4.2. Taxa Contribution to Global PIC Stock and Production

Global CaCO_3 production estimates remain uncertain, ranging from 0.7 to 4.7 Pg C yr^{-1} (Berelson et al., 2007; Buitenhuis et al., 2019; Lee, 2001). Contributions from coccolithophores, foraminifers, and pteropods vary widely. Pteropods are estimated to contribute 0.87–4.2 Pg C yr^{-1} , representing 20–89% of global CaCO_3 production (Gangstø et al., 2008; Lebrato et al., 2010; Buitenhuis et al., 2019). Foraminifers contribute 0.036–0.14 Pg C yr^{-1} , corresponding to 2–4% of global CaCO_3 production (Schiebel, 2002; Lebrato et al., 2010; Buitenhuis et al., 2019). Coccolithophores are estimated to account for ~90% of CaCO_3 production (Ziveri et al., 2023; Kruijt et al., 2026). Deep sediment traps recover significant amounts of foraminifers and pteropods (Table 1; Fig. 3 in Neukermans et al., 2023), whereas coccolithophores dominate surface stocks and production. These observations remain poorly understood in terms of taxon-specific contributions and highlight the need for additional proof-of-concept and process-based studies to better quantify ecosystem-specific controls on PIC production and export. This study demonstrates that, at the global scale, coccolithophores dominate the PIC standing stock. Moreover, seasonal variability in coccolithophore production and biomass appears to play a central role in regulating PIC export efficiency (Fig. 7).

4.3. Influence of Ecosystem Structure on PIC Export

400 The fraction of exported phytoplankton production that is remineralized is mainly influenced by ecosystem structure, which is linked to the seasonal amplitude of NPP (Fig. 7a). Blooms of diatoms and coccolithophores (e.g., *G. huxleyi*), which are expected to induce intense particle sedimentation, occur mostly in regions with high annual mean NPP and large seasonal amplitudes (Fig. 7a). In contrast, nanoplankton and picoplankton dominate global production in oligotrophic areas (low latitudes) characterized by low seasonal NPP amplitude (Lima et al., 2014).

405 The ballast effect hypothesis, driven by biomineral inclusion (calcite and biogenic silica), has long been considered
a mechanism enhancing particle export efficiency (PE_{eff}). In this study, PIC export efficiency ($PIC E_{\text{eff}}$; the proportion
of PIC production exported from the surface) is generally higher north of 40°N and south of 40°S (temperate and
subpolar regions). PIC transfer efficiency ($PIC T_{\text{eff}}$; the proportion of exported PIC reaching the deep ocean) is
410 higher between 40°N and 40°S (subtropical regions) and exhibits a pattern like that of zooplankton fecal pellet
contributions to the gravitational pump (Fig. 7b). Considering the main particle types involved in the gravitational
pump (Nowicki et al., 2022), phytoplankton aggregates may enhance $PIC E_{\text{eff}}$, whereas zooplankton fecal pellets
may enhance $PIC T_{\text{eff}}$. The following sections explore the mechanisms underlying these patterns.

4.4. “Biological Gatekeeper” of the mesopelagic PIC Flux

4.4.1. Packaging Factor and Aggregate Contribution

415 The packaging factor theory (François et al., 2002) suggests that subtropical and equatorial CaCO_3 -rich
ecosystems produce fast-sinking fecal pellets, thereby enhancing PIC export. In this study, the particle-dependent
export model (Nowicki et al., 2022; Fig. 7b) shows that fecal pellet contributions are higher in these CaCO_3 -rich
ecosystems (subtropical and equatorial regions). However, observed PIC fluxes are lower in subtropical and
equatorial regions despite higher production, challenging the hypothesis that CaCO_3 packaged in fecal pellets is
420 protected from dissolution. Opal-dominated systems (temperate and subpolar regions) exhibit high PIC fluxes,
suggesting that labile aggregates may also enhance PIC transfer and export efficiency ($PIC E_{\text{eff}}$). Overall, PIC
production in the euphotic layer appears to be decoupled from PIC export and transfer efficiencies ($PIC E_{\text{eff}}$ and
 $PIC T_{\text{eff}}$). Upper-ocean PIC loss is primarily attributed to biologically mediated dissolution (Morse et al., 2006; Friis
et al., 2006; Buitenhuis et al., 2019; Sulpis et al., 2021; Dean et al., 2024). Although zooplankton and bacterial
425 activity decrease with depth (Hernández-León et al., 2020), epipelagic and mesopelagic grazing still appears to
contribute to PIC loss.

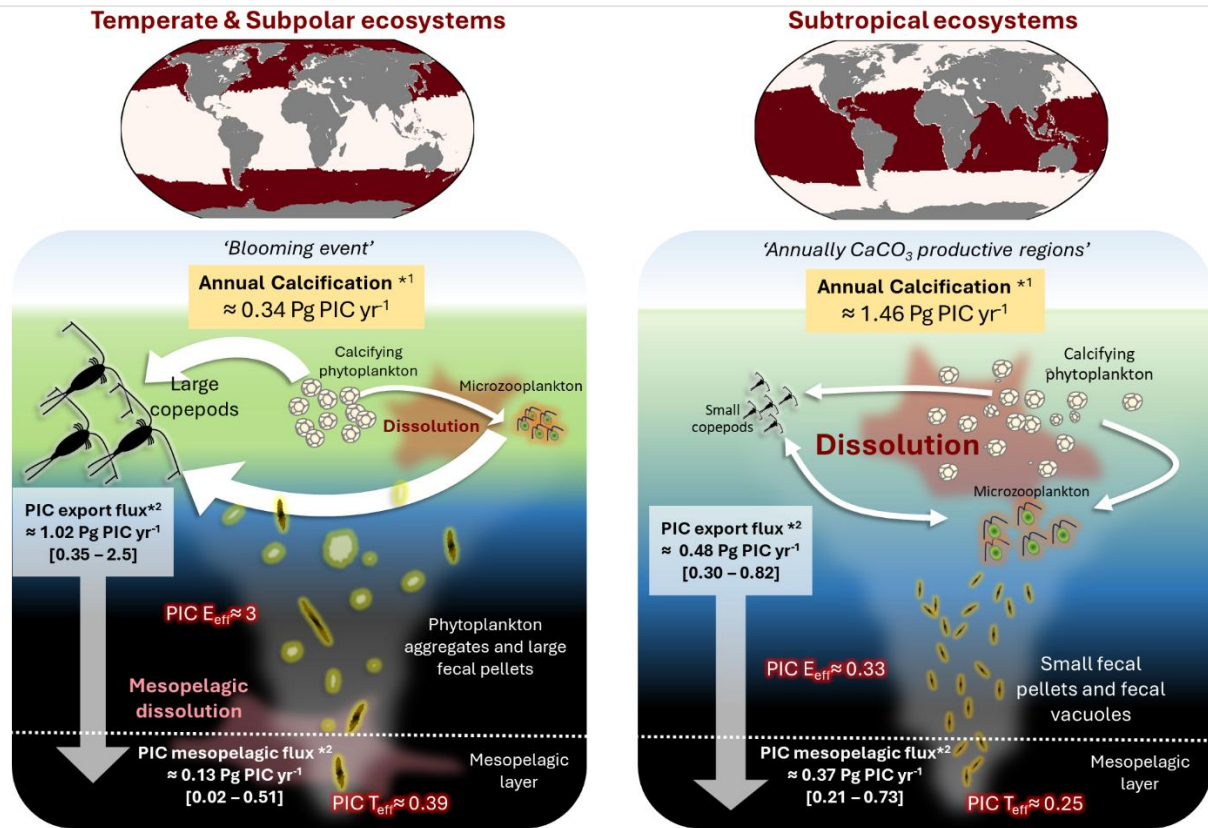
4.4.2. Hypothetical Processes of Biological-Mediated PIC Dissolution

Heterotrophic bacteria colonizing CaCO_3 particles appear to induce minimal dissolution, suggesting a limited role
430 in PIC loss during sinking (Bissett et al., 2011). Similarly, the increase in hydrostatic pressure experienced by *G.*
huxleyi aggregates during sedimentation does not significantly enhance calcite dissolution (Tamburini et al., 2021).
Experimental and modeling studies also show that calcite is largely preserved during zooplankton gut passage,
with dissolution generally low or negligible across various species and conditions (Harris, 1994; Honjo, 1976; Roth
et al., 1975; Langer et al., 2007; Jansen and Wolf-Gladrow, 2001; Antia et al., 2008; Toullec et al., 2022; Dean et
435 al., 2024). Despite lower contributions of fecal pellets to the gravitational pump at high latitudes (Fig. 7b), PIC export
efficiency ($PIC E_{\text{eff}}$) remains elevated in these regions (temperate and subpolar areas), suggesting that additional
factors, likely related to plankton community composition and phenology, play an important role in controlling PIC
preservation and export.

4.5. Ecosystem control on PIC flux

440 Microzooplankton (<200 μm) play a central role in regulating primary producer biomass and facilitating carbon
export via fecal vacuoles or aggregates (McNair et al., 2021; Calbet and Landry, 2004). Their grazing intensity
exhibits strong latitudinal variation, ranging from 59% of annual primary production in temperate-polar regions to
75% in tropical-subtropical regions (Calbet and Landry, 2004). These patterns align with the observed latitudinal
445 distribution of PIC export efficiency (PIC E_{eff}) and PIC transfer efficiency (PIC T_{eff}) (Fig. 7). In the North Atlantic,
microzooplankton consume 288–589 $\text{mg C m}^{-2} \text{ day}^{-1}$ in the mixed layer during mid-summer, representing 39–
115% of local phytoplankton production (Burkill et al., 1993), highlighting their substantial contribution to particle
processing and carbon flux. Blooming coccolithophores, such as *G. huxleyi*, can temporarily escape
microzooplankton grazing during bloom onset through predation-avoidance traits, including colony formation,
increased cell size, spines, or toxin production (Irigoien et al., 2005; Monteiro et al., 2016). Conversely, subtropical
450 and equatorial regions, characterized by low seasonal variability in coccolithophore biomass and continuous
grazing pressure, experience sustained PIC loss, possibly due to dissolution within microzooplankton vacuoles
(Antia et al., 2008; Dean et al., 2024). Large zooplankton may indirectly enhance PIC preservation by suppressing
microzooplankton biomass and repackaging coccoliths into fast-sinking fecal pellets (Nejstgaard et al., 1994).
Indeed, a mesocosm study demonstrated that ingestion rates of the large copepod *Calanus finmarchicus* were
455 similar during blooms of diatoms and *G. huxleyi* (Nejstgaard et al., 1994). However, *C. finmarchicus* biomass
increased threefold more in mesocosms dominated by *G. huxleyi* compared to those dominated by diatoms at
similar algal biomass (Nejstgaard et al., 1994). The authors suggested that, during bloom conditions, copepods
preferentially graze on microzooplankton. The incorporation of coccoliths into large fecal pellets (produced by
mesozooplankton) likely results from passive, non-selective feeding behavior (e.g., current feeding), rather than
460 from selective grazing on coccolithophores. Our dataset reveals a positive correlation between PIC production and
PIC flux in the North Atlantic across all depth layers (Fig. 6), emphasizing the role of zooplankton-mediated carbon
transfer (Hernández-León et al., 2020). Zooplankton functional traits vary among bioregions (Benedetti et al., 2023):
temperate and subpolar regions are dominated by large, detritivorous or omnivorous copepods that feed passively
(current- or cruise-feeders; Fig. 5 in Benedetti et al., 2023), whereas subtropical and equatorial regions are
465 dominated by smaller, carnivorous copepods that feed actively (ambush- or current-ambush-feeders). Grazing by
these distinct functional groups shapes phytoplankton biomass and community structure, ultimately influencing the
efficiency and depth of PIC export (Le Quéré et al., 2016; Vallina et al., 2014; Fig. 8). The present study suggests
that, in temperate and subpolar ecosystems, large copepods may enhance PIC export efficiency in two ways: (1)
by repackaging coccoliths into fecal pellets through passive current feeding; and (2) by exerting strong grazing
470 pressure on microzooplankton, thereby indirectly reducing CaCO_3 -mediated dissolution within microzooplankton
vacuoles (Dean et al., 2024; Fig. 8). In contrast, marine snow aggregates may create microenvironments that
promote PIC dissolution in the mesopelagic layer, potentially explaining the observed decrease from PIC E_{eff} to
PIC T_{eff} in temperate ecosystems (Fig. 8). Subtropical regions, on the other hand, exhibit continuous grazing,
efficient nutrient recycling, and more complex food webs. Microzooplankton strongly regulate primary producer
475 biomass and particulate organic carbon transfer, a fraction of which may subsequently be exported as fecal pellets

or aggregates (McNair et al., 2021). This ecological context may favor CaCO_3 -mediated dissolution within microzooplankton vacuoles (Fig. 8), potentially reducing PIC export efficiency ($\text{PIC } E_{\text{eff}}$).



^{*1} Satellite based NPP, excluding continental margin (200m depth) and interior sea

^{*2} Estimates based on the sediment trap annual PIC flux ($\text{g C m}^{-2} \text{ yr}^{-1}$) extended to the total surface area. Results are expressed by the median and interquartile [q25 – q75]

Figure 8: Synthesis of the potential PIC pathway through the water column, in two distinct ecosystems: a) Subtropical ecosystems (subtropical gyres and equatorial upwellings). b) Temperate zone (North Atlantic, North Pacific and subpolar regions). The white arrows represent the trophic transfer between the different planktonic compartments (Predator prey), and double arrow means that both compartments could be both prey and predator each other. Small copepods correspond to individual body sizes ranging from 200 μm to 2 mm; Microzooplankton (mostly protists, < 200 μm) represent the flagellates and ciliates community; Large copepods correspond to individual body sizes larger than 2 mm (mostly large calanoid). Note that microzooplankton could be heterotrophic, autotrophic or mixotrophic.

This study integrates the main conceptual frameworks currently proposed in the field to interpret the observed patterns in PIC export efficiency ($\text{PIC } E_{\text{eff}}$) and biologically mediated CaCO_3 dissolution. Our results highlight that variability in $\text{PIC } E_{\text{eff}}$ and the mechanisms regulating CaCO_3 dissolution remain insufficiently constrained, particularly across contrasting biogeographical regimes. These findings underscore the need for targeted proof-of-concept and process-based studies to better quantify ecosystem-specific controls on PIC export. Improving this

mechanistic understanding is essential for refining predictions of the oceanic carbon cycle under ongoing environmental changes.

5. Conclusion and perspectives

495 In this study, the “packaging factor” theory is proposed as an important driver of the discrepancy between estimated PIC production and export flux across distinct oceanic regions. Despite the PIC/POC production ratio being twice as high in subtropical regions compared to temperate and subpolar regions, PIC export efficiency (PIC E_{eff}) is estimated to be tenfold lower and PIC transfer efficiency (PIC T_{eff}) 1.5-fold lower. Our results suggest that zooplankton functional diversity and biogeography may explain the contrasting patterns of CaCO_3 export efficiency, through differences in dissolution and preservation within particles. Such processes could substantially contribute to total downward carbon export and associated nutrient fluxes. However, only a limited number of experimental studies have directly demonstrated the effect of zooplankton functional diversity on CaCO_3 dissolution. To test and refine the hypothesis proposed here, there is a strong need for experimental data and in situ observations addressing both mesozooplankton and microzooplankton grazing dynamics and CaCO_3 fluxes.

500
505 In the context of surface ocean warming and acidification, phytoplankton losses due to microzooplankton grazing in eutrophic waters are expected to increase (Chen et al., 2012), potentially attenuating the carbonate pump in temperate regions. In addition, POC export is unlikely to respond uniformly across high-latitude regions to future changes in ballast availability, which may also affect the biological carbon pump (BCP). Data compilations and model outputs indicate that coccolithophores tend to become less calcified relative to growth under increasing CO_2 concentrations (Krumhardt et al., 2017, 2019). End-of-century CO_2 projections suggest an approximately 11% decline in global oceanic calcification relative to preindustrial levels (Krumhardt et al., 2019). Such changes in surface ocean dynamics are expected to alter the future balance of surface alkalinity and CO_2 exchange between the ocean and atmosphere (Planchat et al., 2024; Tyrrell, 2008; Volk and Hoffert, 1985).

Code and Data availability. The author confirms that the data supporting the findings of this study are available within the article and its Supplement.

Author’s contribution. Jordan Toullec: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Validation, Visualization, Writing original draft, study and editing.

Competing interest. The author declares no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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6. References

- 525 Armengol, L., Calbet, A., Franchy, G., Rodríguez-Santos, A., & Hernández-León, S. Planktonic food web structure and trophic transfer efficiency along a productivity gradient in the tropical and subtropical Atlantic Ocean. *Scientific reports*, 9(1), 2044. 2019.
- 530 Armstrong, R. A., Lee, C., Hedges, J. I., Honjo, S., & Wakeham, S. G. A new, mechanistic model for organic carbon fluxes in the ocean based on the quantitative association of POC with ballast minerals. *Deep Sea Research Part II: Topical Studies in Oceanography*, 49(1-3), 219-236. 2001
- Antia, A. N., Suffrian, K., Holste, L., Müller, M. N., Nejtgaard, J. C., Simonelli, P., Carotenuto, Y., and Putzeys, S.: Dissolution of coccolithophorid calcite by microzooplankton and copepod grazing, *Biogeosciences discussion*, <https://doi.org/10.5194/bgd-5-1-2008>, 2008.
- 535 Antoine, D. and Morel, A.: Oceanic primary production: 1. Adaptation of a spectral light-photosynthesis model in view of application to satellite chlorophyll observations, *Global Biogeochemical Cycles*, 10, 43–55, <https://doi.org/10.1029/95GB02831>, 1996.
- Balch, W. M. and Mitchell, C.: Remote sensing algorithms for particulate inorganic carbon (PIC) and the global cycle of PIC, *Earth-Science Reviews*, 239, 104363, <https://doi.org/10.1016/j.earscirev.2023.104363>, 2023.
- 540 Balch, W. M., Kilpatrick, K. A., Holligan, P., Harbour, D., and Fernandez, E.: The 1991 coccolithophore bloom in the central North Atlantic. 2. Relating optics to coccolith concentration, *Limnology and Oceanography*, 41, 1684–1696, <https://doi.org/10.4319/lo.1996.41.8.1684>, 1996.
- Balch, W. M., Gordon, H. R., Bowler, B. C., Drapeau, D. T., and Booth, E. S.: Calcium carbonate measurements in the surface global ocean based on Moderate-Resolution Imaging Spectroradiometer data, *Journal of Geophysical Research: Oceans*, 110, <https://doi.org/10.1029/2004JC002560>, 2005.
- 545 Balch, W. M., Drapeau, D. T., Bowler, B. C., Lyczkowski, E., Booth, E. S., and Alley, D.: The contribution of coccolithophores to the optical and inorganic carbon budgets during the Southern Ocean Gas Exchange Experiment: New evidence in support of the “Great Calcite Belt” hypothesis, *Journal of Geophysical Research: Oceans*, 116, <https://doi.org/10.1029/2011JC006941>, 2011.
- 550 Balch, W. M., Bowler, B. C., Drapeau, D. T., Lubelczyk, L. C., and Lyczkowski, E.: Vertical Distributions of Coccolithophores, PIC, POC, Biogenic Silica, and Chlorophyll a Throughout the Global Ocean, *Global Biogeochemical Cycles*, 32, 2–17, <https://doi.org/10.1002/2016GB005614>, 2018.
- Barton, A. D., Pershing, A. J., Litchman, E., Record, N. R., Edwards, K. F., Finkel, Z. V., Kiørboe, T., and Ward, B. A.: The biogeography of marine plankton traits, *Ecology Letters*, 16, 522–534, <https://doi.org/10.1111/ele.12063>, 2013.
- 555 Bednaršek, N., Možina, J., Vogt, M., O’Brien, C., and Tarling, G. A.: The global distribution of pteropods and their contribution to carbonate and carbon biomass in the modern ocean, *Earth System Science Data*, 4, 167–186, <https://doi.org/10.5194/essd-4-167-2012>, 2012.
- 560 Bendif, E. M., Probert, I., Archontikis, O. A., Young, J. R., Beaufort, L., Rickaby, R. E., & Filatov, D. Rapid diversification underlying the global dominance of a cosmopolitan phytoplankton. *The ISME Journal*, 17(4), 630-640. 2023

- Benedetti, F., Wydler, J., and Vogt, M.: Copepod functional traits and groups show divergent biogeographies in the global ocean, *Journal of Biogeography*, 50, 8–22, <https://doi.org/10.1111/jbi.14512>, 2023.
- 565 Berelson, W. M., Balch, W. M., Najjar, R., Feely, R. A., Sabine, C., and Lee, K.: Relating estimates of CaCO₃ production, export, and dissolution in the water column to measurements of CaCO₃ rain into sediment traps and dissolution on the sea floor: A revised global carbonate budget, *Global Biogeochemical Cycles*, 21, <https://doi.org/10.1029/2006GB002803>, 2007.
- Berner, R. A. and Morse, J. W.: Dissolution kinetics of calcium carbonate in sea water; IV, Theory of calcite dissolution, *American Journal of Science*, 274, 108–134, <https://doi.org/10.2475/ajs.274.2.108>, 1974.
- 570 Bissett, A., Neu, T. R., and Beer, D. de: Dissolution of Calcite in the Twilight Zone: Bacterial Control of Dissolution of Sinking Planktonic Carbonates Is Unlikely, *PLOS ONE*, 6, e26404, <https://doi.org/10.1371/journal.pone.0026404>, 2011.
- 575 Brandão, S. N., Hoppema, M., Kamenev, G. M., Karanovic, I., Riehl, T., Tanaka, H., Vital, H., Yoo, H., and Brandt, A.: Review of Ostracoda (Crustacea) living below the Carbonate Compensation Depth and the deepest record of a calcified ostracod, *Progress in Oceanography*, 178, 102144, <https://doi.org/10.1016/j.pocean.2019.102144>, 2019.
- Brown, C. W., & Yoder, J. A. Coccolithophorid blooms in the global ocean. *Journal of Geophysical Research: Oceans*, 99(C4), 7467-7482. 1994.
- Buitenhuis, E. T., Le Quéré, C., Bednaršek, N., and Schiebel, R.: Large Contribution of Pteropods to Shallow CaCO₃ Export, *Global Biogeochemical Cycles*, 33, 458–468, <https://doi.org/10.1029/2018GB006110>, 2019.
- 580 Burkill, P. H., Edwards, E. S., John, A. W. G., and Sleight, M. A.: Microzooplankton and their herbivorous activity in the northeastern Atlantic Ocean, *Deep Sea Research Part II: Topical Studies in Oceanography*, 40, 479–493, [https://doi.org/10.1016/0967-0645\(93\)90028-L](https://doi.org/10.1016/0967-0645(93)90028-L), 1993.
- 585 Burridge, A. K., Goetze, E., Wall-Palmer, D., Le Double, S. L., Huisman, J., and Peijnenburg, K. T. C. A.: Diversity and abundance of pteropods and heteropods along a latitudinal gradient across the Atlantic Ocean, *Progress in Oceanography*, 158, 213–223, <https://doi.org/10.1016/j.pocean.2016.10.001>, 2017.
- Buesseler, K. O., Lamborg, C. H., Boyd, P. W., Lam, P. J., Trull, T. W., Bidigare, R. R., Bishop, J. K. B., Casciotti, K. L., Dehairs, Elskens, M., Honda, M., Karl, D. M., Siegel, D. A., Silver, M. W., Steinberg, D. K. Valdes, J., Van Mooy, B. & Wilson, S. Revisiting carbon flux through the ocean's twilight zone. *science*, 316(5824), 567-570. 2007
- 590 Calbet, A. and Landry, M. R.: Phytoplankton growth, microzooplankton grazing, and carbon cycling in marine systems, *Limnology and Oceanography*, 49, 51–57, <https://doi.org/10.4319/lo.2004.49.1.0051>, 2004.
- Chaabane, S., de Garidel-Thoron, T., Meilland, J., Sulpis, O., Chalk, T. B., Brummer, G. J. A., ... & Schiebel, R.: Migrating is not enough for modern planktonic foraminifera in a changing ocean. *Nature*, 1-7. <https://doi.org/10.1038/s41586-024-08191-5>, 2024.
- 595 Chaffron, S., Delage, E., Budinich, M., Vintache, D., Henry, N., Nef, C., Ardyna, M., Zayed, A. A., Junger, P. C., Galand, P. E., Lovejoy, C., Murray, A. E., Sarmiento, H., Tara Oceans coordinators, Acinas, S. G., Babin, M., Iudicone, D., Jaillon, O., Karsenti, E., Wincker, P., Karp-Boss, L., Sullivan, M. B., Bowler, C., de Vargas, C. & Eveillard, D. Environmental vulnerability of the global ocean epipelagic plankton community interactome. *Science Advances*, 7(35), eabg1921. 2021.

- 600 Chen, B., Landry, M. R., Huang, B., and Liu, H.: Does warming enhance the effect of microzooplankton grazing on marine phytoplankton in the ocean?, *Limnology and Oceanography*, 57, 519–526, <https://doi.org/10.4319/lo.2012.57.2.0519>, 2012.
- Clements, D. J., Yang, S., Weber, T., McDonnell, A. M. P., Kiko, R., Stemmann, L., and Bianchi, D.: New Estimate of Organic Carbon Export From Optical Measurements Reveals the Role of Particle Size Distribution and Export Horizon, *Global Biogeochemical Cycles*, 37, e2022GB007633, <https://doi.org/10.1029/2022GB007633>, 2023.
- 605 Dean, C. L., Harvey, E. L., Johnson, M. D., & Subhas, A. V. Microzooplankton grazing on the coccolithophore *Emiliana huxleyi* and its role in the global calcium carbonate cycle. *Science Advances*, 10(45), eadr5453. 2024
- Feely, R. A., Sabine, C. L., Lee, K., Millero, F. J., Lamb, M. F., Greeley, D., ... & Wong, C. S. In situ calcium carbonate dissolution in the Pacific Ocean. *Global Biogeochemical Cycles*, 16(4), 91-1. 2002.
- 610 Fischer, G., Karstensen, J., Romero, O., Baumann, K.-H., Donner, B., Hefter, J., Mollenhauer, G., Iversen, M., Fiedler, B., Monteiro, I., and Körtzinger, A.: Bathypelagic particle flux signatures from a suboxic eddy in the oligotrophic tropical North Atlantic: production, sedimentation and preservation, *Biogeosciences*, 13, 3203–3223, <https://doi.org/10.5194/bg-13-3203-2016>, 2016.
- Francois, R., Honjo, S., Krishfield, R., and Manganini, S.: Factors controlling the flux of organic carbon to the bathypelagic zone of the ocean, *Global Biogeochemical Cycles*, 16, 34-1-34–20, <https://doi.org/10.1029/2001GB001722>, 2002.
- 615 Friis, K., Najjar, R. G., Follows, M. J. & Dutkiewicz, S. Possible overestimation of shallow-depth calcium carbonate dissolution in the ocean. *Glob. Biogeochem. Cycles* <https://doi.org/10.1029/2006gb002727>. 2006.
- Gangstø, R., Gehlen, M., Schneider, B., Bopp, L., Aumont, O., and Joos, F.: Modeling the marine aragonite cycle: changes under rising carbon dioxide and its role in shallow water CaCO₃ dissolution, *Biogeosciences*, 5, 1057–1072, <https://doi.org/10.5194/bg-5-1057-2008>, 2008.
- 620 Gifford, D. J., Fessenden, L. M., Garrahan, P. R., and Martin, E.: Grazing by microzooplankton and mesozooplankton in the high-latitude North Atlantic Ocean: Spring versus summer dynamics, *Journal of Geophysical Research: Oceans*, 100, 6665–6675, <https://doi.org/10.1029/94JC00983>, 1995.
- 625 Guerreiro, C. V., Baumann, K. H., Brummer, G. J. A., Valente, A., Fischer, G., Ziveri, P., Vanda, B. & Stuetz, J. B. W. Carbonate fluxes by coccolithophore species between NW Africa and the Caribbean: Implications for the biological carbon pump. *Limnology and Oceanography*, 66(8), 3190-3208. 2021.
- Harris, R. P.: Zooplankton grazing on the coccolithophore *Emiliana huxleyi* and its role in inorganic carbon flux, *Marine Biology*, 119, 431–439, <https://doi.org/10.1007/BF00347540>, 1994.
- 630 Hauck, J., Gruber, N., Ishii, M., and Müller, J. D.: Constraining regional and global ocean carbon fluxes in RECCAP2, Copernicus Meetings, <https://doi.org/10.5194/egusphere-egu23-12786>, 2023.
- Henson, S. A., Sanders, R., and Madsen, E.: Global patterns in efficiency of particulate organic carbon export and transfer to the deep ocean: Export and transfer efficiency, *Global Biogeochem. Cycles*, 26, n/a-n/a, <https://doi.org/10.1029/2011GB004099>, 2012.
- 635 Henson, S., Le Moigne, F., & Giering, S. Drivers of carbon export efficiency in the global ocean. *Global biogeochemical cycles*, 33(7), 891-903. 2019.

- Hernández-León, S., Koppelman, R., Fraile-Nuez, E., Bode, A., Mompeán, C., Irigoien, X., Olivar, M. P., Echevarría, F., Fernández de Puellas, M. L., González-Gordillo, J. I., Cózar, A., Acuña, J. L., Agustí, S., and Duarte, C. M.: Large deep-sea zooplankton biomass mirrors primary production in the global ocean, *Nat Commun*, 11, 6048, <https://doi.org/10.1038/s41467-020-19875-7>, 2020.
- 640 Holligan, P. M., Fernández, E., Aiken, J., Balch, W. M., Boyd, P., Burkill, P. H., Finch, M., Groom, S. B., Malin, G., Muller, K., Purdie, D. A., Robinson, C., Trees, C. C., Turner, S. M., and van der Wal, P.: A biogeochemical study of the coccolithophore, *Emiliania huxleyi*, in the North Atlantic, *Global Biogeochemical Cycles*, 7, 879–900, <https://doi.org/10.1029/93GB01731>, 1993.
- 645 Honjo, S.: Coccoliths: Production, transportation and sedimentation, *Marine Micropaleontology*, 1, 65–79, [https://doi.org/10.1016/0377-8398\(76\)90005-0](https://doi.org/10.1016/0377-8398(76)90005-0), 1976.
- Honjo, S. and Roman, M.: Marine copepod fecal pellets: Production, preservation and sedimentation, *Journal of Marine Research*, 36, 1978.
- Hopkins, J., Henson, S. A., Painter, S. C., Tyrrell, T., & Poulton, A. J. Phenological characteristics of global coccolithophore blooms. *Global Biogeochemical Cycles*, 29(2), 239-253. 2015.
- 650 Hopkins, J. and Balch, W. M.: A New Approach to Estimating Coccolithophore Calcification Rates From Space, *Journal of Geophysical Research: Biogeosciences*, 123, 1447–1459, <https://doi.org/10.1002/2017JG004235>, 2018.
- Irigoien, X., Flynn, K. J., and Harris, R. P.: Phytoplankton blooms: a 'loophole' in microzooplankton grazing impact?, *Journal of Plankton Research*, 27, 313–321, <https://doi.org/10.1093/plankt/fbi011>, 2005.
- 655 Iversen, M. H. and Ploug, H.: Ballast minerals and the sinking carbon flux in the ocean: carbon-specific respiration rates and sinking velocity of marine snow aggregates, *Biogeosciences*, 7, 2613–2624, <https://doi.org/10.5194/bg-7-2613-2010>, 2010.
- Jansen, H. and Wolf-Gladrow, D.: Carbonate dissolution in copepod guts: a numerical model, *Mar. Ecol. Prog. Ser.*, 221, 199–207, <https://doi.org/10.3354/meps221199>, 2001.
- 660 Kaneko, H., Endo, H., Henry, N., Berney, C., Mahé, F., Poulain, J., Labadie, K., Beluche, O., El Hourany, R., Chaffron, S., Wincker, P., Nakamura, R., Karp-Boss, L., Boss, E., Chris Bowler, de Vargas, C., Tomii, K., and Ogata, H.: Predicting global distributions of eukaryotic plankton communities from satellite data, *ISME COMMUN.*, 3, 1–9, <https://doi.org/10.1038/s43705-023-00308-7>, 2023.
- 665 Klaas, C. and Archer, D. E.: Association of sinking organic matter with various types of mineral ballast in the deep sea: Implications for the rain ratio, *Global Biogeochemical Cycles*, 16, 63-1-63–14, <https://doi.org/10.1029/2001GB001765>, 2002.
- Knecht, N. S., Benedetti, F., Elizondo, U. H., Bednaršek, N., Chaabane, S., de Weerd, C., Peijnenburg, K. T. C. A., Schiebel, R., and Vogt, M.: The impact of zooplankton calcifiers on the marine carbon cycle, *Global Biogeochemical Cycles*, n/a, e2022GB007685, <https://doi.org/10.1029/2022GB007685>, 2023.
- 670 Kruijt, A. L., van Dijk, R., Sulpis, O., Beaufort, L., Lassus, G., Brummer, G.-J., van der Burg, A. D., Cala, B. A., Ourradi, Y., Peijnenburg, K. T. C. A., Humphreys, M. P., Chaabane, S., Sluijs, A., and Middelburg, J. J.: The contributions of various calcifying plankton to the South Atlantic calcium carbonate stock, *Biogeosciences*, 23, 531–563, <https://doi.org/10.5194/bg-23-531-2026>, 2026.

- 675 Krumhardt, K. M., Lovenduski, N. S., Iglesias-Rodriguez, M. D., and Kleypas, J. A.: Coccolithophore growth and calcification in a changing ocean, *Progress in Oceanography*, 159, 276–295, <https://doi.org/10.1016/j.pocean.2017.10.007>, 2017.
- Krumhardt, K. M., Lovenduski, N. S., Long, M. C., Lévy, M., Lindsay, K., Moore, J. K., & Nissen, C. Coccolithophore growth and calcification in an acidified ocean: Insights from community Earth system model simulations. *Journal of Advances in Modeling Earth Systems*, 11(5), 1418-1437. 2019
- 680 Kwon, E. Y., Dunne, J. P., & Lee, K. Biological export production controls upper ocean calcium carbonate dissolution and CO₂ buffer capacity. *Science Advances*, 10(13), ead10779. 2024
- Lacour, L., Llort, J., Briggs, N., Strutton, P. G., & Boyd, P. W. Seasonality of downward carbon export in the Pacific Southern Ocean revealed by multi-year robotic observations. *Nature Communications*, 14(1), 1278. 2023.
- 685 Langer, G., Nehrke, G., and Jansen, S.: Dissolution of *Calcidiscus leptoporus* coccoliths in copepod guts? A morphological study, *Mar. Ecol. Prog. Ser.*, 331, 139–146, <https://doi.org/10.3354/meps331139>, 2007.
- Lalli, C. M., & Gilmer, R. W. Pelagic snails: the biology of holoplanktonic gastropod mollusks. Stanford University Press. 1989.
- Laurenceau-Cornec, E. C., Le Moigne, F. A. C., Gallinari, M., Moriceau, B., Toullec, J., Iversen, M. H., Engel, A., and De La Rocha, C. L.: New guidelines for the application of Stokes' models to the sinking velocity of marine aggregates, *Limnology and Oceanography*, 65, 1264–1285, <https://doi.org/10.1002/lno.11388>, 2020.
- 690 Le Moigne, F. A. C., Pabortsava, K., Marcinko, C. L. J., Martin, P., and Sanders, R. J.: Where is mineral ballast important for surface export of particulate organic carbon in the ocean?, *Geophysical Research Letters*, 41, 8460–8468, <https://doi.org/10.1002/2014GL061678>, 2014.
- Le Quéré, C., Buitenhuis, E. T., Moriarty, R., Alvain, S., Aumont, O., Bopp, L., Chollet, S., Enright, C., Franklin, D. J., Geider, R. J., Harrison, S. P., Hirst, A. G., Larsen, S., Legendre, L., Platt, T., Prentice, I. C., Rivkin, R. B., Salliey, S., Sathyendranath, S., Stephens, N., Vogt, M., and Vallina, S. M.: Role of zooplankton dynamics for Southern Ocean phytoplankton biomass and global biogeochemical cycles, *Biogeosciences*, 13, 4111–4133, <https://doi.org/10.5194/bg-13-4111-2016>, 2016.
- 700 Lebrato, M., Iglesias-Rodríguez, D., Feely, R. A., Greeley, D., Jones, D. O. B., Suarez-Bosche, N., Lampitt, R. S., Cartes, J. E., Green, D. R. H., and Alker, B.: Global contribution of echinoderms to the marine carbon cycle: CaCO₃ budget and benthic compartments, *Ecological Monographs*, 80, 441–467, <https://doi.org/10.1890/09-0553.1>, 2010.
- Lee, K.: Global net community production estimated from the annual cycle of surface water total dissolved inorganic carbon, *Limnology and Oceanography*, 46, 1287–1297, <https://doi.org/10.4319/lo.2001.46.6.1287>, 2001.
- 705 Leising, A. W., Pierson, J. J., Halsband-Lenk, C., Horner, R., and Postel, J.: Copepod grazing during spring blooms: Can *Pseudocalanus newmani* induce trophic cascades?, *Progress in Oceanography*, 67, 406–421, <https://doi.org/10.1016/j.pocean.2005.09.009>, 2005.
- Lima, I. D., Lam, P. J., and Doney, S. C.: Dynamics of particulate organic carbon flux in a global ocean model, *Biogeosciences*, 11, 1177–1198, <https://doi.org/10.5194/bg-11-1177-2014>, 2014.

- 710 Li, J., Liu, Z., Lin, B., Zhao, Y., Cao, J., Zhang, X., Ling, C., Ma, P. & Wu, J. Zooplankton fecal pellet characteristics and contribution to the deep-sea carbon export in the southern South China Sea. *Journal of Geophysical Research: Oceans*, 127(12), e2022JC019412. 2022
- Millero, F. J., Lee, K., & Roche, M. Distribution of alkalinity in the surface waters of the major oceans. *Marine Chemistry*, 60(1-2), 111-130. 1998.
- 715 Milliman, J. D., Troy, P. J., Balch, W. M., Adams, A. K., Li, Y.-H., and Mackenzie, F. T.: Biologically mediated dissolution of calcium carbonate above the chemical lysocline?, *Deep Sea Research Part I: Oceanographic Research Papers*, 46, 1653–1669, [https://doi.org/10.1016/S0967-0637\(99\)00034-5](https://doi.org/10.1016/S0967-0637(99)00034-5), 1999.
- Monteiro, F. M., Bach, L. T., Brownlee, C., Bown, P., Rickaby, R. E. M., Poulton, A. J., Tyrrell, T., Beaufort, L., Dutkiewicz, S., Gibbs, S., Gutowska, M. A., Lee, R., Riebesell, U., Young, J., and Ridgwell, A.: Why marine phytoplankton calcify, *Sci. Adv.*, 2, e1501822, <https://doi.org/10.1126/sciadv.1501822>, 2016.
- 720 Morse, J. W., Andersson, A. J. & Mackenzie, F. T. Initial responses of carbonate-rich shelf sediments to rising atmospheric pCO₂ and “ocean acidification”: role of high Mg-calcites. *Geochim. Cosmochim. Acta* 70, 5814–5830. 2006.
- Mouw, C. B., Barnett, A., McKinley, G. A., Gloege, L., and Pilcher, D.: Global ocean particulate organic carbon flux merged with satellite parameters, *Earth System Science Data*, 8, 531–541, <https://doi.org/10.5194/essd-8-531-2016>, 2016.
- 725 Nejtgaard, J., Gismervik, I., and Solberg, P.: Feeding and reproduction by *Calanus finmarchicus*, and microzooplankton grazing during mesocosm blooms of diatoms and the coccolithophore *Emiliana huxleyi*, *Mar. Ecol. Prog. Ser.*, 147, 197–217, <https://doi.org/10.3354/meps147197>, 1997.
- 730 Nejtgaard, J. C., Witte, H. J., van der Wal, P., and Jacobsen, A.: Copepod grazing during a mesocosm study of an *Emiliana huxleyi* (Prymnesiophyceae) bloom, *Sarsia*, 79, 369–377, <https://doi.org/10.1080/00364827.1994.10413568>, 1994.
- Neukermans, G., Bach, L. T., Butterley, A., Sun, Q., Claustre, H., and Fournier, G. R.: Quantitative and mechanistic understanding of the open ocean carbonate pump - perspectives for remote sensing and autonomous in situ observation, *Earth-Science Reviews*, 104359, <https://doi.org/10.1016/j.earscirev.2023.104359>, 2023.
- 735 Nowicki, M., DeVries, T., and Siegel, D. A.: Quantifying the Carbon Export and Sequestration Pathways of the Ocean’s Biological Carbon Pump, *Global Biogeochemical Cycles*, 36, e2021GB007083, <https://doi.org/10.1029/2021GB007083>, 2022.
- Passow, U. and Carlson, C.: The biological pump in a high CO₂ world, *Mar. Ecol. Prog. Ser.*, 470, 249–271, <https://doi.org/10.3354/meps09985>, 2012.
- 740 Picard, T., Gula, J., Fablet, R., Collin, J., and Mémerly, L.: Predicting particle catchment areas of deep-ocean sediment traps using machine learning, *Ocean Sci.*, 20, 1149–1165, <https://doi.org/10.5194/os-20-1149-2024>, 2024.
- 745 Picard, T., Baker, C. A., Gula, J., Fablet, R., Mémerly, L., and Lampitt, R.: Estimating the variability of deep-ocean particle flux collected by sediment traps using satellite data and machine learning, *Biogeosciences*, 22, 4309–4331, <https://doi.org/10.5194/bg-22-4309-2025>, 2025.

- Pilskaln, C. H. and Honjo, S.: The Fecal Pellet fraction of biogeochemical particle fluxes to the deep sea, *Global Biogeochem. Cycles*, 1, 31–48, <https://doi.org/10.1029/GB001i001p00031>, 1987.
- 750 Planchat, A., Kwiatkowski, L., Bopp, L., Torres, O., Christian, J. R., Butenschön, M., ... & Stock, C.: The representation of alkalinity and the carbonate pump from CMIP5 to CMIP6 Earth system models and implications for the carbon cycle. *Biogeosciences*, 20(7), 1195-1257. 2023
- Planchat, A., Bopp, L., Kwiatkowski, L., & Torres, O. The carbonate pump feedback on alkalinity and the carbon cycle in the 21st century and beyond. *Earth System Dynamics*, 15(3), 565-588. 2024
- 755 Poulton, A. J., Adey, T. R., Balch, W. M., & Holligan, P. M. Relating coccolithophore calcification rates to phytoplankton community dynamics: Regional differences and implications for carbon export. *Deep Sea Research Part II: Topical Studies in Oceanography*, 54(5-7), 538-557. 2007.
- Poulton, A. J., Stinchcombe, M. C., Achterberg, E. P., Bakker, D. C. E., Dumoussaud, C., Lawson, H. E., Lee, G. A., Richier, S., Suggett, D. J., and Young, J. R.: Coccolithophores on the north-west European shelf: calcification rates and environmental controls, *Biogeosciences*, 11, 3919–3940, <https://doi.org/10.5194/bg-11-3919-2014>, 2014.
- 760 Renforth, P., & Henderson, G.: Assessing ocean alkalinity for carbon sequestration. *Reviews of Geophysics*, 55(3), 636-674. 2017.
- Ricour, F., Guidi, L., Gehlen, M., DeVries, T., and Legendre, L.: Century-scale carbon sequestration flux throughout the ocean by the biological pump, *Nat. Geosci.*, 1–9, <https://doi.org/10.1038/s41561-023-01318-9>, 2023.
- 765 Romero, O. E., Fischer, G., Karstensen, J., and Cermeño, P.: Eddies as trigger for diatom productivity in the open-ocean Northeast Atlantic, *Progress in Oceanography*, 147, 38–48, <https://doi.org/10.1016/j.pocean.2016.07.011>, 2016.
- Rosengard, S. Z., Lam, P. J., Balch, W. M., Auro, M. E., Pike, S., Drapeau, D., and Bowler, B.: Carbon export and transfer to depth across the Southern Ocean Great Calcite Belt, *Biogeosciences*, 12, 3953–3971, <https://doi.org/10.5194/bg-12-3953-2015>, 2015.
- 770 Roth, P. H., Mullin, M. M., and Berger, W. H.: Coccolith Sedimentation by Fecal Pellets: Laboratory Experiments and Field Observations, *Geol Soc America Bull*, 86, 1079, [https://doi.org/10.1130/0016-7606\(1975\)86<1079:CSBFPL>2.0.CO;2](https://doi.org/10.1130/0016-7606(1975)86<1079:CSBFPL>2.0.CO;2), 1975.
- Ryan-Keogh, T. J., Thomalla, S. J., Chang, N., & Moalusi, T. A new global oceanic multi-model net primary productivity data product. *Earth System Science Data*, 15(11), 4829-4848. 2023.
- 775 Samtleben, C. and Bickert, T.: Coccoliths in sediment traps from the Norwegian Sea, *Marine Micropaleontology*, 16, 39–64, [https://doi.org/10.1016/0377-8398\(90\)90028-K](https://doi.org/10.1016/0377-8398(90)90028-K), 1990.
- Sarmiento, J. L.: *Ocean Biogeochemical Dynamics*, in: *Ocean Biogeochemical Dynamics*, Princeton University Press, <https://doi.org/10.1515/9781400849079>, 2013.
- 780 Savoye, N., Benitez-Nelson, C., Burd, A. B., Cochran, J. K., Charette, M., Buesseler, K. O., ... & Elskens, M.: ²³⁴Th sorption and export models in the water column: A review. *Marine Chemistry*, 100(3-4), 234-249. 2006.

- Schiebel, R. and Movellan, A.: First-order estimate of the planktic foraminifer biomass in the modern ocean, *Earth System Science Data*, 4, 75–89, <https://doi.org/10.5194/essd-4-75-2012>, 2012.
- Sulpis, O., Jeansson, E., Dinauer, A., Lauvset, S. K., and Middelburg, J. J.: Calcium carbonate dissolution patterns in the ocean, *Nat. Geosci.*, 14, 423–428, <https://doi.org/10.1038/s41561-021-00743-y>, 2021.
- 785 Tamburini, C., Garel, M., Barani, A., Boeuf, D., Bonin, P., Bhairy, N., Guasco, S., Jacquet, S., Le Moigne, F. A. C., Panagiotopoulos, C., Riou, V., Veloso, S., Santinelli, C., and Armougom, F.: Increasing Hydrostatic Pressure Impacts the Prokaryotic Diversity during *Emiliana huxleyi* Aggregates Degradation, *Water*, 13, 2616, <https://doi.org/10.3390/w13192616>, 2021.
- 790 Thuiller, W., Pollock, L. J., Gueguen, M., and Münkemüller, T.: From species distributions to meta-communities, *Ecology Letters*, 18, 1321–1328, <https://doi.org/10.1111/ele.12526>, 2015.
- Torres Valdés, S., Painter, S. C., Martin, A. P., Sanders, R., and Felden, J.: Data compilation of fluxes of sedimenting material from sediment traps in the Atlantic Ocean, *Earth System Science Data*, 6, 123–145, <https://doi.org/10.5194/essd-6-123-2014>, 2014.
- 795 Toullec, J., Vincent, D., Frohn, L., Miner, P., Le Goff, M., Devesa, J., and Moriceau, B.: Copepod Grazing Influences Diatom Aggregation and Particle Dynamics, *Frontiers in Marine Science*, 6, <https://doi.org/10.3389/fmars.2019.00751>, 2019.
- Toullec, J., Delegrange, A., Perruchon, A., Duong, G., Cornille, V., Brutier, L., and Hermoso, M.: Copepod Feeding Responses to Changes in Coccolithophore Size and Carbon Content, *Journal of Marine Science and Engineering*, 10, 1807, <https://doi.org/10.3390/jmse10121807>, 2022.
- 800 Tyrrell, T. and Merico, A.: *Emiliana huxleyi*: bloom observations and the conditions that induce them, in: *Coccolithophores: From Molecular Processes to Global Impact*, edited by: Thierstein, H. R. and Young, J. R., Springer, Berlin, Heidelberg, 75–97, https://doi.org/10.1007/978-3-662-06278-4_4, 2004.
- Tyrrell, T. Calcium carbonate cycling in future oceans and its influence on future climates. *Journal of plankton research*, 30(2), 141-156. 2008.
- 805 Vallina, S. M., Follows, M. J., Dutkiewicz, S., Montoya, J. M., Cermeno, P., and Loreau, M.: Global relationship between phytoplankton diversity and productivity in the ocean, *Nat Commun*, 5, 4299, <https://doi.org/10.1038/ncomms5299>, 2014.
- 810 Volk, T., & Hoffert, M. I. Ocean carbon pumps: Analysis of relative strengths and efficiencies in ocean-driven atmospheric CO₂ changes. *The carbon cycle and atmospheric CO₂: Natural variations Archean to present*, 32, 99-110. 1985
- Wang, W.-L., Fu, W., Le Moigne, F. A. C., Letscher, R. T., Liu, Y., Tang, J.-M., and Primeau, F. W.: Biological carbon pump estimate based on multidecadal hydrographic data, *Nature*, 1–7, <https://doi.org/10.1038/s41586-023-06772-4>, 2023.
- 815 Wassmann, P.: Retention versus export food chains: processes controlling sinking loss from marine pelagic systems, in: *Eutrophication in Planktonic Ecosystems: Food Web Dynamics and Elemental Cycling: Proceedings of the Fourth International PELAG Symposium, held in Helsinki, Finland, 26–30 August 1996*, edited by: Tamminen, T. and Kuosa, H., Springer Netherlands, Dordrecht, 29–57, https://doi.org/10.1007/978-94-017-1493-8_3, 1998.

820 Weber, T., Cram, J. A., Leung, S. W., DeVries, T., & Deutsch, C. Deep ocean nutrients imply large latitudinal variation in particle transfer efficiency. *Proceedings of the National Academy of Sciences*, 113(31), 8606-8611. 2016.

Ziveri, P., Langer, G., Chaabane, S., de Vries, J., Gray, W. R., Keul, N., ... & Mortyn, P. G.: Calcifying plankton: From biomineralization to global change. *Science*, 390(6771), <https://doi/abs/10.1126/science.adq8520>. 2025.

825 Ziveri, P., Gray, W. R., Anglada-Ortiz, G., Manno, C., Grelaud, M., Incarbona, A., Rae, J. W. B., Subhas, A. V., Pallacks, S., White, A., Adkins, J. F., and Berelson, W.: Pelagic calcium carbonate production and shallow dissolution in the North Pacific Ocean, *Nat Commun*, 14, 1–14, <https://doi.org/10.1038/s41467-023-36177-w>, 2023.

Zöllner, E., Hoppe, H.-G., Sommer, U., and Jürgens, K.: Effect of zooplankton-mediated trophic cascades on marine microbial food web components (bacteria, nanoflagellates, ciliates), *Limnology and Oceanography*, 54, 262–275, <https://doi.org/10.4319/lo.2009.54.1.0262>, 2009.