



1 **Extreme drought–accelerated dissolved carbon metabolism triggers pulsed CO₂**
2 **outgassing in karst lakes**

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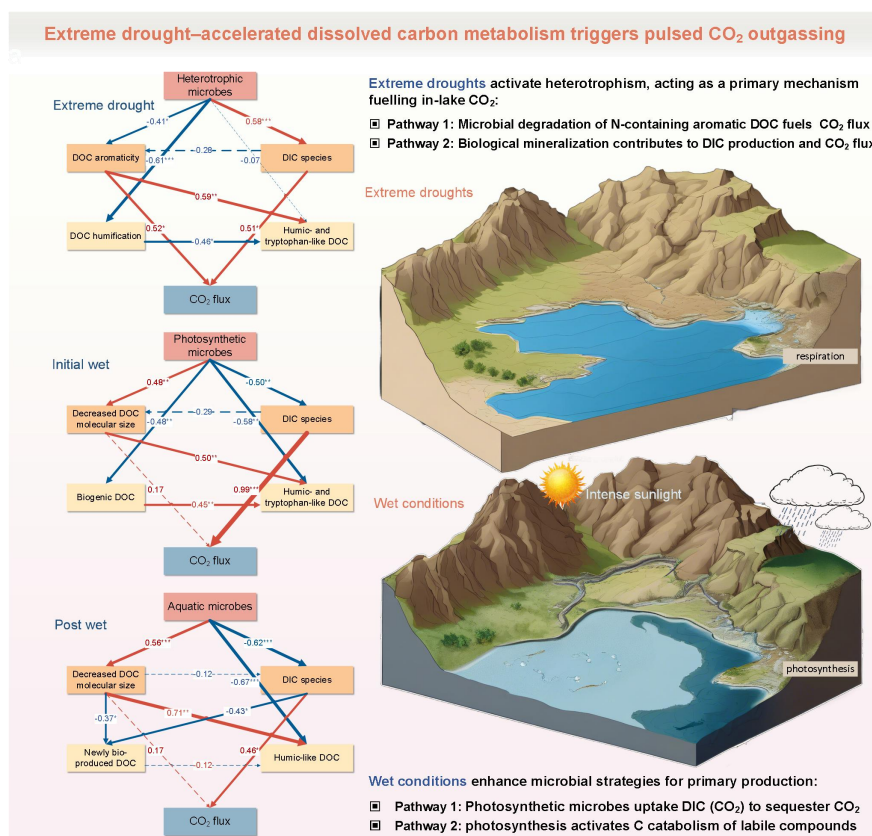
23 **Abstract**

24 Karst aquatic ecosystems are important reservoirs of dissolved carbon (C), supporting
25 dynamic CO₂ fluxes through the biological C pump. However, our current
26 understanding of how sophisticated interactions between aquatic microbiomes and
27 dissolved C turnover constrain the timing of CO₂ sequestration and emission remains
28 limited. Here we capture an extreme drought event and the ensuing relatively wet
29 conditions from systematic investigations in karst lakes, demonstrating that
30 temporally distinct microbiomes are tuned to the metabolic patterns of dissolved C
31 and thereby modulate CO₂ emissions. Specifically, we find that the extreme drought
32 accelerates respiration of dissolved organic C, sharply increasing the CO₂ evasion rate.
33 Wet conditions stimulate photosynthetic uptake of dissolved inorganic C, consuming
34 lake CO₂ while promoting labile organic C formation. We therefore propose that
35 pulses of CO₂ emissions from karst lakes occur after wet conditions end, as a
36 consequence of rapid remineralization of newly produced bioavailable organic C,
37 especially during extreme droughts. Our findings highlight the crucial importance of
38 managing periodic CO₂ outgassing from karst waters under drought conditions for the
39 implementation of region-specific C neutrality strategies.

40 **Keywords:** Karst waters, aquatic microbiome, dissolved carbon, CO₂ flux, extreme
41 drought



42 Graphical Abstract





43 **1. Introduction**

44 Lakes store, metabolize and release large quantities of natural carbon (C),
45 representing a funnel of aquatic C budget (Bogard et al., 2019; Borges et al., 2022;
46 Evans et al., 2017). Therefore, there is mounting evidence that lakes function as
47 “biogeochemical reactors”, receiving terrestrial C and continuously cycling the bulk
48 of aquatic C (Pi et al., 2022; Rodríguez-Cardona et al., 2023). Yet to date, the
49 mechanisms driving lake C cycling are poorly understood, particularly regarding the
50 underlying factors that constrain C emission (Chen et al., 2022; Mendonça et al.,
51 2017). It is estimated that inland lakes, despite covering only ~3% of the land surface,
52 contribute to 38% of aquatic C outgassing (Tranvik et al., 2009). Although the drivers
53 of these emissions are regionally investigated (Maberly et al., 2013; Mu et al., 2023;
54 Serikova et al., 2019), a fundamental factor governing C fixation, mineralization, and
55 subsequent uptake or release remains elusive. Deciphering the cycling of dissolved C
56 offers a promising avenue to address this knowledge gap, as it mediates C turnover
57 between organic and inorganic forms and accounts for ~90% of the global C flux from
58 terrestrial to aquatic ecosystems (Drake et al., 2020), making it essential for
59 understanding aquatic C budget (Raymond and Hamilton, 2018; Song et al., 2018).

60 Evidence suggests that seasonal wetness and drought control the dynamics of
61 dissolved inorganic C (DIC) concentrations and species (Rehn et al., 2023; Tye et al.,
62 2022). These events also alter water-land connectivity and thus the export of dissolved
63 organic C (DOC) to inland waters (Li et al., 2022a; Wang et al., 2024; Yuan et al.,



64 2024). Subsequently, aquatic photosynthesis utilizes DIC, while heterotrophic
65 respiration of DOC generates CO₂ (Guo et al., 2023; Leles and Levine, 2023).
66 Consequently, dissolved C turnover and CO₂ emissions are anticipated to be highly
67 interconnected through hydrological and biological mechanisms (Hu et al., 2022;
68 Kellerman et al., 2014; Monteith et al., 2023). For example, experimental data suggest
69 that droughts can stimulate dissolved C cycling by transiently accelerating primary
70 production and, more persistently, DOC respiration (Harjung et al., 2019).

71 These pervasive turnover and emission of aquatic C, in particular, can be
72 substantial in karst waters. The prevailing view suggests the regional specificity of
73 karst regions, specifically with respect to ecological fragility and the significant role
74 in carbonate C sink (Chen et al., 2023; D'Ettorre et al., 2024). Yet, recent reports also
75 highlight significant dissolved C cycling and CO₂ sequestration in karst aquatic
76 ecosystems, attributed to the “biological C pump” effect (He et al., 2024; Sun et al.,
77 2022; Zhang et al., 2024). Carbonate weathering can couple with photosynthetic
78 uptake of DIC, resulting in self-amplifying CO₂ sink during karst water cycle (Liu et
79 al., 2018; Wang et al., 2022). Nevertheless, primary production may trigger DOC
80 catabolism and rapid cycle of active C (Ni et al., 2023; Ni et al., 2022). These critical
81 processes are associated with aquatic biology, but little is known about how lake
82 microorganisms drive dissolved C turnover and ultimately modulate CO₂ emissions in
83 karst lakes.



84 Theoretically, microbiome is anticipated to govern internal cycling between DIC
85 and DOC, and this process, in turn, may affect specific CO₂ pathways (Li et al., 2022b;
86 Shanguan et al., 2024). Prior studies found significant DIC uptake by C-fixing
87 microorganisms (Li et al., 2024) and recalcitrant DOC sequestration *via* heterotrophic
88 bacteria in karst aquatic systems (He et al., 2022b; Xu et al., 2023), which potentially
89 regulate CO₂ fixation and outgassing in response to specific pathways of C
90 metabolism. Therefore, we hypothesized that microorganisms will interact with
91 dissolved C to establish distinctive CO₂ drivers in karst lakes. To test this hypothesis,
92 we conducted a two-year investigation, capturing an extreme drought event and its
93 following rainfalls, with the aims of revealing: 1) temporal interactions of aquatic
94 microbiome with dissolved C dynamics; 2) specific pathways governing karst lake
95 CO₂ flux; and 3) microbially-driven dissolved C turnover and the resulting CO₂
96 emission or sequestration. Achieving these aims is expected to uncover the underlying
97 mechanisms of C source-sink transformation in karst water environments, ultimately
98 supporting efforts toward C neutrality.

99 **2. Materials and methods**

100 **2.1. Study area**

101 Observations were performed in the karst lakes Hongfeng (HFH), Laoma (LMH) and
102 Baihua (BBH), located between 25°57' to 26°42'N in latitude and 105°58' to 106°34'E
103 in longitude (Fig. 1). The catchment geology is predominantly karst lithology i.e.,
104 dolomite and limestone. Lake HFH and BHH, two of the largest artificial lakes in



105 Guizhou Province, cover surface areas of 57.2 and 14.5 km², with water volumes of
106 6.01×10^8 and 1.91×10^8 m³, respectively. As a segment of the Maotiao River, the
107 LMH is supplied by several tributaries that connect Lake HFH to BHH. These karst
108 waters are situated in a subtropical monsoon climate zone, with annual temperatures
109 ranging from -7.8°C to 37.5°C. The wet season lasts from May to October, delivering
110 an average annual precipitation of 1130 mm. Specifically, an extreme drought event
111 occurred in 2022–2023, with monthly rainfall <10 mm during the drought (early
112 winter), >100 mm during the initial-wet (early summer), and 50–100 mm during the
113 post-wet (autumn) periods. These lakes acting as drinking water sources, are disrupted
114 by agriculture and domestic sewage.

115 **2.2. Fieldwork and laboratory analysis**

116 During May to November from 2022 to 2023, our sampling captured an extreme
117 drought event, allowing us to clearly identify the specific temporal scales during the
118 drought, initial-wet and post-wet periods. Spatially, fieldworks were designed to
119 incorporate full spectrum of the karst lakes from 32 sampling locations (Fig. 1a).
120 Therefore, a total of 96 water samples were collected from our investigations. In
121 details, we collected surface waters at a depth of ~10 cm and filtered them within 6
122 hours. Filtrates were stored in 1000-mL high-density polyethylene (HDPE) containers
123 designed to eliminate headspace and air bubbles. Water samples were filtered using
124 glass microfiber filters (Whatman, GF/F 47 mm, 0.7-μm) for dissolved C
125 measurement, and polycarbonate membrane filters (Millipore, GF/F 47 mm, 0.22-μm)



126 for microbial determination. Samples were refrigerated at 4°C during transport and
127 stored at -70°C for microbiological analysis.

128 Water temperature and pH were *in-situ* determined with a portable pH meter
129 (PHB-4, Shanghai). Wind velocity was measured using a Testo 410-1 anemometer
130 (Testo, Germany). Total alkalinity was titrated with Alkalinity Test MColortest™
131 (Merck, Germany). DOC concentration was detected using varioTOC cube/select
132 (Elementar, Germany). Chromophoric DOC was determined using a double-beam
133 scanning spectrophotometer (UV-5500PC, Shanghai) with UV–visible absorption
134 spectra ranging from 200 to 700 nm (1-nm interval). Fluorescence DOC was analysed
135 using a RF-6000 Spectrophotometer (Shimadzu, Japan), with excitation and emission
136 wavelengths of 200–450 (5-nm interval) nm and 250–600 nm (1-nm interval),
137 respectively. Molecular DOC was characterized using Fourier transform ion cyclotron
138 resonance mass spectrometry (FT-ICR MS). We combined equal volumes of water
139 samples from each lake site, leaving us three composite samples for FT-ICR MS
140 analysis. Details on sample pretreatment (solid-phase extraction, SPE) for FT-ICR MS
141 are available in Supplementary Text S1.

142 Genomic DNA in sampling waters was extracted using E.Z.N.A. Water DNA Kit
143 (OMEGA, USA) according to the manufacturers' instructions, and detected with 1%
144 agarose gel electrophoresis. It employed universal primers 341F
145 (5'-CCTAYGGGRBGCASCAG-3') and 806R



146 (5'-GGACTACNNGGTATCTAAT-3') for amplifying the V3–V4 hypervariable
 147 region of 16S rRNA, with a GeneAmp PCR System 9700 (ABI GeneAmp, USA). The
 148 PCR products were extracted by 2% agarose gel electrophoresis, and recovered from
 149 AXYPREP DNA Gel Recovery Kit (Axygen Biosciences, USA) with Tris-HCl for
 150 elution. Purified amplicons were pooled in equimolar and sequenced using paired-end
 151 on an Illumina MiSeq PE300 (Illumina, USA) following standard protocols.

152 **2.3. Data processing and calculation**

153 In this study, we employed a carbonate equilibria-based method for estimating
 154 aqueous DIC, using combinations of water chemistry parameters (pH, water
 155 temperature and total alkalinity) through CO₂SYS program (Xu et al., 2017). This
 156 program outputs concentrations of DIC species i.e., total DIC, HCO₃⁻, CO₃²⁻ and
 157 dissolved CO₂, as well as aqueous partial pressure of CO₂ ($p\text{CO}_2$), a crucial indication
 158 for potential CO₂ emissions from aquatic environments. The following thin boundary
 159 layer model was used to calculate areal CO₂ flux (mmol m⁻² d⁻¹) from the karst waters.

$$160 \quad \text{Areal CO}_2 \text{ flux} = (p\text{CO}_{2\text{water}} - p\text{CO}_{2\text{air}}) \times k \times K_h \quad (1)$$

161 This model proposes that the difference between aqueous ($p\text{CO}_{2\text{water}}$, µatm) and
 162 atmospheric $p\text{CO}_2$ ($p\text{CO}_{2\text{air}}$, µatm) can characterize the impetus and direction for CO₂
 163 transfer. Temporal shifts in *in-situ* atmospheric CO₂ levels (dimensionally convertible
 164 to $p\text{CO}_{2\text{air}}$) are available in Fig. S1. By contrast, gas transfer velocity (k , m d⁻¹) can



165 constrain the velocity of water-air CO₂ exchange, which is calibrated from normalized
 166 gas transfer velocity (k_{600} , cm h⁻¹) using water temperature and Schmidt number.
 167 Specifically, k or k_{600} serves as a function of water turbulence, empirically modelled
 168 by wind velocity in lakes and lentic rivers. Henry's constant (K_h , mmol m⁻³ µatm⁻¹)
 169 calibrated from *in-situ* temperature and pressure, characterizes CO₂ equilibrium at
 170 water-air interfaces. Details for chemical calculations of DIC species and thin
 171 boundary layer model see Supplementary Text S2.

172 We analysed UV–visible and fluorescent spectroscopy for understanding DOC
 173 component, origin and fate. Specifically, DOC-normalized absorption coefficient
 174 SUVA₂₅₄ (L mg⁻¹ m⁻¹), an indicator of DOC aromaticity, was computed as the
 175 absorption coefficient (a_{254} , m⁻¹) divided by DOC concentration (mg L⁻¹). Spectral
 176 slope $S_{275-295}$, a proxy for DOC relative molecular weight, was calculated by
 177 nonlinearly fitting an exponential function to the absorption spectrum from 275 to 295
 178 nm. Fluorescence index (FI) increases with intensified biological activity, and
 179 indicates allochthonous (< 1.4) and autochthonous (> 1.9) inputs for aquatic DOC,
 180 which was calculated as the ratio of emission intensity at 470 nm to 520 nm with an
 181 excitation of 370 nm. Biological index (BIX), a proxy for freshness of biologically
 182 produced DOC, was calculated as the ratio of emission intensity at 380 nm to 430 nm,
 183 using an excitation of 310 nm. Humification index (HIX), characterizing to DOC
 184 humification and biodegradability, was expressed as the ratio of total emission
 185 intensities at 435–480 nm divided by 300–345 nm, at an excitation of 254 nm. Parallel



186 factor analysis (PARAFAC) was used to identify primary DOC component by
 187 separating excitation-emission matrices into independent fluorophores. The
 188 PARAFAC modelling employed residual and split-half analyses for component
 189 selection and correspondence validation.

190 FT-ICR MS analysis was conducted to examine dissolved organic matter (DOM,
 191 represents the specific material form of the general DOC) composition using a
 192 molecular formula calculator based on criteria with elemental combinations of
 193 $C_{0-\infty}H_{0-\infty}O_{0-\infty}N_{0-1}S_{0-1}$. Peaks were detected within $S/N > 4$ and a mass accuracy of $\leq \pm 1$
 194 ppm. Van Krevelen diagrams plotting H/C against O/C were employed to visualize
 195 FT-ICR MS data. Seven DOM compositions were extracted based on the elemental
 196 ratios of H/C and O/C (Ni et al., 2024), involving carbohydrates, amino-sugars,
 197 saturated compounds, tannins, lignin, unsaturated hydrocarbons and condensed
 198 aromatic structures. The elements (C, H, O, N, P and S), and formulas (CHO, CHOS,
 199 CHON, CHOP, CHONS, CHONP, CHOSP and CHONSP) were identified based on
 200 molecular exact mass and matched against a molecular formulas database (Yan et al.,
 201 2024). The modified aromaticity index (AI-mod) and nominal oxidation state of
 202 carbon (NOSC) were calculated as follows:

$$AI_{mod} = \frac{1 + C - \frac{1}{2}O - S - \frac{1}{2}(N + H)}{C - \frac{1}{2}O - N - S} \quad (2)$$

$$NOSC = \frac{4C + H - 3N - 2O - 2S}{C} \quad (3)$$



203 Alpha diversity, calculated using Mothur (<https://mothur.org/wiki/calculators/>),
204 was used to assess microbial community through the indices Coverage (community
205 coverage), Chao (community richness) and Shannon (community diversity). We
206 analysed microbial community and its relative abundance, ranking the top 20
207 microorganisms at genus level. Using Tax4Fun, we converted 16S into prokaryotic
208 classification profiles in the KEGG database, enabling KEGG functional annotation
209 for 16S RNA gene sequences. It provides insights into potential microbial functions
210 based on their composition and abundance in aquatic environments, and specifically
211 presents information on KEGG Orthology in this study.

212 **2.4. Quality control**

213 Water collection and analysis followed the standard procedures, as proposed by the
214 American Public Health Association (1985). The pH probe was calibrated with 6.86
215 and 9.18 pH standard solution at 25°C, ensuring ± 0.01 pH unit accuracy. Water
216 temperature and wind velocity were accurate to $\pm 0.5^\circ\text{C}$ and $\pm (0.2 \text{ m/s} + 2\%$ of
217 measurements), respectively. Alkalinity Test MColortestTM measured total alkalinity
218 with $< 3\%$ uncertainty. DIC species calculated from water chemistry may be
219 overestimated due to non-carbonate alkalinity (Liu et al., 2020). We specifically
220 assessed that non-carbonate alkalinity from nitrogen, phosphorus and organic C can
221 result in a maximum 29.5% overestimation of DIC species. Consequently, we
222 corrected this overestimation by using measured DIC concentrations (Fig. S2), which



223 converted systematic errors into instrument errors ($< 2\%$) for all DIC datasets. UV
224 measurements were 10% replicated, suggesting an uncertainty of $< 2\%$. The inner
225 filter effect of fluorescence data can be neglected since absorbances at 254 nm were
226 all below 0.3 (Ohno, 2002). Excitation-emission matrices were corrected for Raman
227 and Rayleigh scatterings through interpolation, and fluorescence intensity (A.U.) was
228 normalized to Raman Unit (R.U.) *via* water Raman peak areas (Ni et al., 2024).
229 FT-ICR MS and microbial analysis were conducted by China National Analytical
230 Centre (Guangzhou, China) and Majorbio Bio-Pharm Technology Co. Ltd. (Shanghai,
231 China), respectively.

232 **2.5. Statistical analysis**

233 Normality and homogeneity of variance were assessed using Kolmogorov-Smirnov
234 test and Levene's test, respectively. Variables were log-transformed as needed to
235 ensure normality assumptions. One-way analysis of variance (ANOVA) with Tukey
236 HSD post hoc was employed to evaluate statistical differences across DIC species,
237 DOC compositions and microbial variables. Correlation analysis was used to assess
238 possible associations within or between dissolved C, CO₂ flux and microbial variables.
239 We introduced a structural equation model (SEM) to examine how aquatic
240 microorganisms interact with dissolved C and ultimately influence CO₂ flux through
241 both direct and indirect pathways. We initially excluded the variables with nonlinear
242 relationships (e.g., alpha diversity and Tax4Fun) to derive comparable standardized
243 path coefficients. We further established and specified 6 latent variables according to



the loadings of their associated observed variables (see Supplementary Text S3). In the SEM, we hypothesized that 1) photosynthetic and heterotrophic microbes will preferentially associate with DIC for anabolism and DOC for catabolism, respectively; and 2) dissolved C dynamics and CO₂ flux are intertwined due to carbonate chemistry, biogenic, and terrestrial regulations. The SEM with partial least squares path modelling was performed using the plspm package in R (Version 4.1.3). Statistical analyses and figures preparing were conducted using OriginPro 2024 and MATLAB 2018.

3. Results

3.1. DIC species and CO₂ emissions

The significant carbonate kinetics in karst waters present an opportunity to understand the dynamics of aqueous DIC species. This study captured strong seasonality in water chemistry (Table 1), particularly with lower pH and water temperature during the extreme-drought period ($p < 0.001$ by ANOVA). *In-situ* measured total alkalinity (range: 2178.7–6210.8 $\mu\text{eq L}^{-1}$), along with DIC (range: 1538.7–6206.0 $\mu\text{mol L}^{-1}$) and HCO₃⁻ (range: 918.4–5970.6 $\mu\text{mol L}^{-1}$), had no temporal variations across the periods ($p > 0.05$). Extreme drought caused lower aqueous CO₃²⁻ but higher dissolved CO₂ levels in comparison to wet conditions ($p < 0.001$, Fig. S3). It should be noted that we corrected concentrations of DIC species by eliminating systematic errors from non-carbonate alkalinity, reducing average uncertainties of 6.8% for DIC, 7.0% for HCO₃⁻, and 38.7% for dissolved CO₂ (Fig. S2).



265 To estimate potential CO₂ emissions from the study lakes, we calculated aqueous
 266 $p\text{CO}_2$, k and areal CO₂ flux (Fig. 1). These lakes had a broad range of aqueous $p\text{CO}_2$
 267 levels, spanning from 20 to 13479 μatm across sampling locations (Fig. 1b). We found
 268 that extreme drought ($4437 \pm 3468 \mu\text{atm}$) unexpectedly caused a ~five-fold increase
 269 in mean $p\text{CO}_2$ relative to initial- ($804 \pm 1080 \mu\text{atm}$) and post-wet periods (928 ± 1728
 270 μatm , $p < 0.001$), which can escalate to ~ten-fold when considering median $p\text{CO}_2$ (Fig.
 271 1b). Indeed, our dataset reveals that 61% of samples were oversaturated with CO₂
 272 relative to atmospheric equilibrium (Fig. S1), attributable to high $p\text{CO}_2$ levels during
 273 extreme droughts. Gas transfer velocity k showed no significant temporal shifts ($p >$
 274 0.05), ranging from $0.37\text{--}2.28 \text{ m d}^{-1}$ with a mean of $0.62 \pm 0.26 \text{ m d}^{-1}$ (Fig. 1c). This is
 275 slightly lower than previously reported global average of 0.74 m d^{-1} from a similar
 276 empirical model based on wind speed (Raymond et al., 2013). We estimated areal CO₂
 277 flux to be $42 \pm 79 \text{ mmol m}^{-2} \text{ d}^{-1}$ (range: $-24\text{--}459 \text{ mmol m}^{-2} \text{ d}^{-1}$) from the karst lakes.
 278 However, it is also noted that the bulk of samples during wet periods were
 279 undersaturated (Fig. 1d). We show that extreme drought increased areal CO₂ efflux
 280 sharply ($111 \pm 104 \text{ mmol m}^{-2} \text{ d}^{-1}$), relative to initial- ($6 \pm 22 \text{ mmol m}^{-2} \text{ d}^{-1}$) and
 281 post-wet periods ($10 \pm 26 \text{ mmol m}^{-2} \text{ d}^{-1}$) ($p < 0.001$). Our findings, therefore, may
 282 partially deviate from the previous understanding of aquatic karst carbon sinks (An et
 283 al., 2015; Binet et al., 2020; Liu et al., 2010), indicating that karst lakes could be a
 284 significant source of CO₂ driven by extreme drought events.



285 **3.2. DOC composition, origin and fate**

286 We used spectroscopic and molecular methods to assess DOC composition, origin and
 287 fate (Fig. 2 and 3). Aqueous DOC concentrations had a ~30-fold variation (range:
 288 0.76–22.1 mg L⁻¹) across the samplings (Fig. 2a). Extreme drought apparently
 289 elevated DOC concentrations to an average of 12.4 ± 3.7 mg L⁻¹, relative to initial-
 290 (5.4 ± 2.3 mg L⁻¹) and post-wet periods (5.7 ± 1.6 mg L⁻¹) ($p < 0.001$). DOC
 291 aromaticity descended significantly across the periods, with initial-wet > post-wet >
 292 extreme drought ($p < 0.01$), as suggested by SUVA₂₅₄. By contrast, relative molecular
 293 weight of DOC, indicated by S₂₇₅₋₂₉₅, was higher during extreme-drought period than
 294 wet periods ($p < 0.05$). We were able to identify DOC fluorescent component through
 295 PARAFAC, demonstrating two distinct humic-like (C1) and one tryptophan-like DOC
 296 (C2) across the periods (Fig. 2b). The temporal abundances differed significantly ($p <$
 297 0.001), with %C1 > %C2 during the extreme drought yet %C1 < %C2 during wet
 298 conditions (Fig. S4). These karst waters received both allochthonous and
 299 autochthonous inputs, with 80% of samples falling within the FI range of 1.4–1.9 (Fig.
 300 2c). As suggested by BIX, we found biologically produced young DOC was more
 301 abundant during initial-wet than extreme-drought period ($p < 0.05$). However, DOC
 302 humification proxied by HIX was notably greater during the extreme drought than
 303 that in the wet conditions ($p < 0.01$).

304 It captured the molecular formulas for a total of 8060, 8636 and 8403
 305 FT-ICR-MS compounds in lake LMH, HFH and BHH, respectively (Fig. 3). The



306 molecular formulas were classified into distinct categories based on H/C and O/C
 307 distributions, as well as exact mass in van Krevelen Diagram (Fig. 3a). The study
 308 lakes showed a similar range of DOM exact mass, varying from 114.031694 to
 309 1121.181055 Da. We suggest that these lakes were governed by low-molecular-weight
 310 DOM, with 80%, 74%, and 77% of the detected molecules for LMH, HFH, and BHH
 311 having an exact mass < 500 Da, respectively. The summed intensity was highly
 312 variable for each DOM molecular category, notably with lignin compounds (or
 313 carboxy-rich acyclic molecules) comprising 83%–84%, followed by saturated
 314 compounds at 11%–13% across lakes. Amino-sugars also represented a notable 1.8%–
 315 2.3%, while carbohydrate, tannins, unsaturated hydrocarbons and condensed aromatic
 316 structures each occupied $\leq 1\%$ across all molecular categories. These DOM molecules
 317 were largely comprised of atomic groups CHO (44.8%–49.2%), CHOS (17.2%–
 318 25.5%), CHON (19.2%–26.6%) and CHONS (5.6%–6.3%) (Fig. 3b). Each karst lake
 319 shared a comparable average AI_{mod} across FT-ICR-MS compounds ($p > 0.05$), while
 320 NOSC was higher in lake BHH (-0.29 ± 0.48) than that in LHM (-0.32 ± 0.48) ($p <$
 321 0.01 , Fig. 3c).

322 **3.3. Aquatic microbiome in the karst lakes**

323 To evaluate microbial processes in the karst lakes, we examined temporal variability
 324 of microbiome from 89 samples (Fig. 4). For the microbial diversity, a total of 121–
 325 351 ASV (14382 sequences), 236–479 ASV (40251 sequences) and 143–445 ASV
 326 (42507 sequences) were assigned during the extreme-drought, initial-wet and post-wet



327 periods, respectively (Fig. S5). Rainfall increased microbial richness and diversity in
 328 the aquatic environment (Fig. 4a), as indicated by Chao (initial wet > post wet >
 329 extreme drought, $p < 0.05$) and Shannon (initial wet > post wet and extreme drought,
 330 $p < 0.05$). However, microbial coverage was higher in the extreme-drought period (p
 331 < 0.01). The top 3 microorganisms were *Acinetobacter*, *CL500-29_marine_group* and
 332 *Cyanobium_PCC-6307* at genus level, with the average relative abundances of 11.6%
 333 $\pm 10.3\%$, $10.3\% \pm 5.58\%$ and $5.68\% \pm 3.91\%$, respectively (Fig. 4b). *Acinetobacter*
 334 and *CL500-29_marine_group*, important for C catabolism, had increased relative
 335 abundance during the extreme drought. By contrast, photosynthetic microorganisms
 336 *Cyanobium_PCC-6307* and *g_norank_f_norank_o_Chloroplast* were more
 337 abundant during the initial-wet period (Fig. 4b). Therefore, we found that amino acid
 338 catabolism was significant under extreme droughts, while carbohydrate anabolism
 339 was predominant under wet conditions (Fig. 4c, $p < 0.001$).

340 4. Discussion

341 4.1. Temporal interactions of aquatic microbiome with dissolved C dynamics

342 Karst aquatic ecosystems, with rapid kinetics of carbonate chemistry and biological
 343 metabolism involved, may develop associated strategies for dissolved C turnover (He
 344 et al., 2024; Ni et al., 2023; Xi et al., 2024). Here, our results show the consistent
 345 shifts of dissolved C with aquatic microbiome in karst lakes. Extreme droughts, for
 346 instance, set the stage for substantial proliferation of heterotrophic microbes (Fig. 4b)
 347 and thus microbial degradation of enriched DOC (Fig. 2a). As labile DOC is



348 exhausted, these microbes are compelled to metabolize typically more refractory
349 DOC. This aligns with recent observations in karst and thermokarst lakes (Hu et al.,
350 2023; Ni et al., 2022), as also evidenced by the parallel increase in CO₂ levels (Fig. 1b)
351 and decrease in DOC aromaticity (Fig. 2a), suggesting that in-lake aromatic
352 compounds are partially labile during extreme droughts. By analysing DOC molecular
353 composition, we specifically found that N-containing, rather than S-containing
354 aromatics are more bioavailable for heterotrophic microorganisms (Fig. S6 and
355 Supplement table S1).

356 Rainfall accelerates atmospheric CO₂ uptake and CO₃²⁻ generation in karst
357 aquatic environments (Zhao et al., 2024), which, combined with photosynthetic
358 uptake of DIC, provides conditions for the enhancement of photosynthetic
359 microorganisms (Fig. 4b). This explains the consistently low CO₂ levels during wet
360 periods (Fig. 1b), aside from the known dilution effect (Ni et al., 2019). Here, we
361 suggest that initial rainfall following extreme droughts substantially boosts microbial
362 richness and diversity (Fig. 4a), possibly due to amplified terrestrial inputs and
363 rejuvenated aquatic biology (Fig. S7). Moreover, we were able to examine the
364 associations between DOC molecular compositions and microorganisms, revealing
365 that photosynthetic microorganisms generate substantial quantities of biodegradable
366 DOC e.g., carbohydrate and lipid-like compounds (Fig. S6 and Supplement table S1).
367 Therefore, our results indicate season-specific microbial strategies for dissolved C
368 metabolism in karst lakes.



369 **4.2. Specific pathways governing karst lake CO₂ flux**

370 Microbiome-driven dissolved C turnover can further alter CO₂ dynamics, especially
371 in lakes with long-term hydraulic retention and active biological processes (Lindström
372 and Bergström, 2004). Our results show that the photosynthesis-driven associations
373 between microbiome, dissolved C, and CO₂ flux emerged substantially during
374 initial-wet periods (Fig. S8). Studies previously predicted self-amplifying
375 photosynthesis driven by carbonate dissolution in karst waters (He et al., 2022a; Liu
376 et al., 2010). Here, we present direct evidence that photosynthetic microorganisms
377 dominate the microbial communities during the initial-wet period (Fig. 4b). Extreme
378 drought, however, decouples the linkages between dissolved C and CO₂ flux from
379 photosynthetic microorganisms. Respiration of DOC e.g., tryptophan-like component
380 in turn fuels lake CO₂ during the extreme-drought period (Fig. S8). Our data allow us
381 to extrapolate that amino acids, aliphatic compounds and small molecular organic
382 acids serve as primary substrates driving CO₂ production and emissions in these lakes
383 (Fig. 9 and Supplement table S1).

384 Using SEM, we clearly demonstrate the specific pathways governing lake CO₂
385 transfer across the periods (Fig. 5). Our observations reinforce that extreme droughts
386 stimulate microbial metabolism of aromatic DOC and contribute to CO₂ outgassing
387 from the study lakes (Fig. 5a). Nevertheless, it emerged an alternative C metabolism
388 pathway during the initial-wet period (Fig. 5a): photosynthetic microorganisms



389 constrain DIC species and subsequent CO₂ flux in lakes. This photosynthetic pathway
390 persisted during the post-wet period, yet we found a specific pathway involving
391 aquatic microbes that promotes a decrease in molecular size and subsequent increases
392 in humic-like abundance (Fig. 5a). This observation aligns with a previous study in
393 karst aquatic ecosystems (Xia et al., 2022), suggesting that intense photosynthesis
394 produces labile compounds and subsequently activates the catabolism of DOC pool,
395 ultimately resulting in the selective accumulation of recalcitrant compounds. These
396 findings, complement prior evidence from biodegradation assays (Ni et al., 2022),
397 providing a new insight into photosynthetic products triggering significant microbial
398 activities and dissolved C turnover in karst lakes.

399 **4.3. Implications for biologically driven C cycling in karst waters**

400 Karst aquatic ecosystems typically represent strong interactions between microbial
401 metabolism and dissolved C turnover, driven by the known “biological C pump” (Yi
402 et al., 2021; Zhang et al., 2024). While the specific C sink from this mechanism is
403 well-documented (Cao et al., 2018; Chen et al., 2023; Sun et al., 2021), the causes of
404 periodic C evasion from karst waters are still questionable. In this study, we propose
405 that extreme droughts yield large CO₂ emissions from the karst lakes, increasing >10
406 times on average relative to wet conditions (Fig. 1d). Droughts apparently accelerate
407 heterotrophic respiration of DOC, acting as a primary mechanism fuelling in-lake CO₂.
408 Drought-enriched heterotrophic microorganisms extensively deplete labile DOC, and
409 even resort to metabolizing previously recognized recalcitrant compounds. Aromatic



410 DOC, for instance, characterized by microbial resistance, has recently been reported
411 to be activated through photochemistry (Hu et al., 2023). Here, our results
412 demonstrate a clear mechanism by which microbial degradation of N-containing
413 aromatic compounds contributes to C metabolism (Fig. S6), indicating that extreme
414 droughts suppress biological C sink in karst lakes.

415 Wet conditions in karst lakes, in contrast, show a distinct pathway governed by
416 aquatic photosynthesis (Fig. 5b), aligning with the prior theory regarding karst
417 biological C sink (Liu et al., 2018). This stimulates a substantial uptake of CO₂ and
418 thus limit lake C emission. Our observations provide direct evidence that
419 photosynthetic microorganisms produce carbohydrates and promote the formation of
420 lipid-like (Fig. S6) as well as other low-molecular-weight compounds (Fig. S8). These
421 labile DOC quickly engage in subsequent biological processes, accelerating the
422 proliferation of heterotrophic microorganisms. Our efforts to upscale this mechanism
423 demonstrate a clear causality: 1) catabolism of photosynthetically derived labile DOC
424 stimulates heterotrophic microorganism growth; 2) recalcitrant DOC accumulates
425 relatively as heterotrophic respiration increases, even though aromatic compounds are
426 partially utilized; 3) a priming effect is triggered, promoting both labile DOC
427 mineralization to CO₂ and the sequestration of recalcitrant DOC. These findings
428 highlight that biologically driven organic C turnover critically determine the CO₂
429 uptake or release in karst aquatic ecosystems.



430 Overall, as we initially hypothesized, aquatic microbiome interacts with
431 dissolved C dynamics, establishing the distinctive CO₂ driving mechanisms that
432 particularly fluctuate with biochemical timing in karst lakes. It is anticipated that
433 pulses of CO₂ emissions will occur after wet conditions end, particularly under
434 drought conditions, as a result of rapid decay of photosynthetically derived organic C.
435 Therefore, managing C sink in karst aquatic systems depends heavily on mitigating
436 the sudden bursts of DOC respiration, especially the dramatic increased CO₂ release
437 during extreme droughts. Our results indicate that wet conditions significantly
438 enhance biological strategies for CO₂ uptake in karst aquatic ecosystems, highlighting
439 the crucial role of mitigating climate change-induced droughts in strengthening karst
440 C sink.

441 **4. Conclusion**

442 Karst lakes represent distinctive microbial pathways for dissolved C metabolism and
443 subsequent CO₂ sequestration or emission. Here, we specifically reveal the temporal
444 dynamics of CO₂ fluxes driven by periodic interactions between aquatic microbiome
445 and dissolved C turnover in the study karst lakes. We show that extreme droughts
446 accelerate the proliferation of heterotrophic microbes and thus rapid respiration of
447 DOC, resulting in a sharp increase in CO₂ emissions. By contrast, wet conditions
448 stimulate aquatic photosynthesis, which consumes DIC and sequesters CO₂ within the
449 lakes. We highlight that exhausted DOC compels heterotrophic microorganisms to
450 metabolize refractory N-containing aromatic compounds, while photosynthetic



451 microorganisms promote the formation of labile DOC compounds. These specific
452 microbial strategies indicate that pulsed CO₂ outgassing from karst lakes occurs after
453 periods of high photosynthetic activity, with the magnitude being significantly
454 amplified during extreme droughts. Therefore, we propose that managing periodic
455 CO₂ outgassing, particularly during droughts, is essential for developing C neutrality
456 in karst waters.

457 **Code/Data availability**

458 The full dataset used for the evaluation of this study is publicly available at
459 <https://figshare.com/s/7475752c5f7b19718199>.

460 **Declaration of Competing Interest**

461 All authors agree this submission and the authors declare that there is no conflict of
462 interests regarding the publication of this article

463 **Author contributions**

464 The project was conceived by M.N. and Z.K. M.N. directed and managed the study,
465 as well as prepared the first draft of the manuscript. Z.K. collected and analysed
466 samples, contributed to data interpretation and paper writing. G.Z. specified the
467 experimental conditions and measured CO₂ data. W.L. specified the experimental
468 conditions and contributed to CO₂ data analysis and interpretation. J.P. conceptualized
469 dissolved carbon turnover in this study, and contributed to data interpretation and



470 paper writing. J.C. and J.Z. and X.W. provided comments on the manuscript. All
471 authors reviewed and contributed to the final manuscript.

472 **Acknowledgments**

473 This study was financially supported by the National Natural Science Foundation of
474 China (NSFC grant no. U2244216, 42167050, 42407094), National Special Support
475 Plan for High-Level Talent to Junbing Pu (Young Talent Plan, 2022), Natural Science
476 Foundation of Chongqing, China (CSTB2022NSCQ-LZX0022,
477 2024NSCQ-MSX3061), Guizhou Provincial Platform and Talent Program (YQK
478 [2023]017), Guizhou Provincial Science and Technology Projects (ZK[2024]061),
479 Science and Technology Plan for High-Level Young Talent of Guizhou Education
480 Department ([2024]325) and the Second Tibetan Plateau Scientific Expedition and
481 Research Program (Sub-item: 2019QZKK0601-3).



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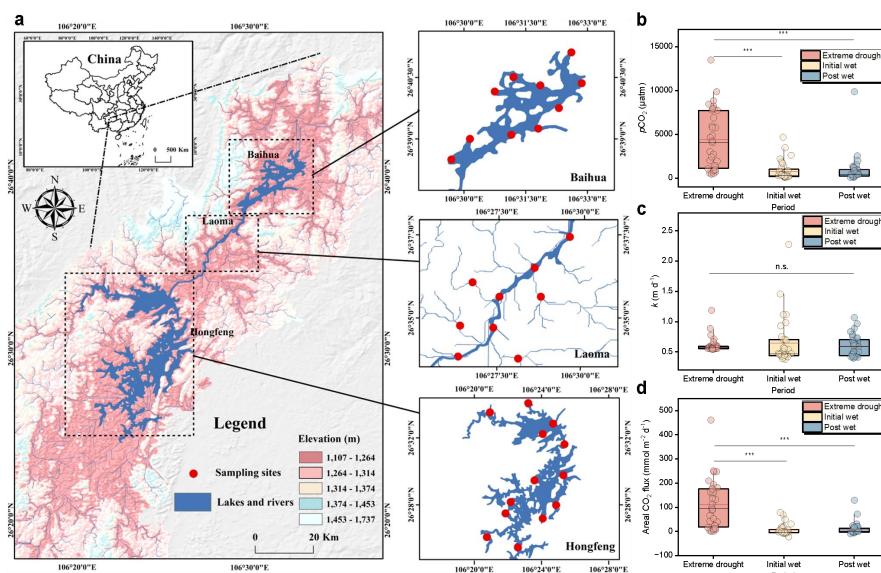
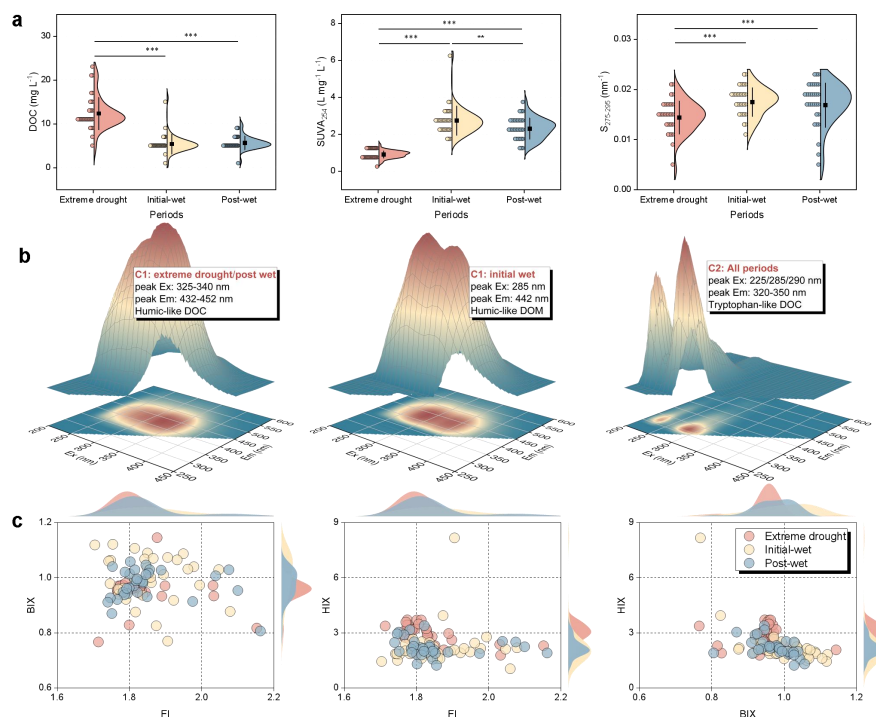


Fig. 1. Distribution of sampling locations and temporal patterns of CO₂ emissions in the karst lakes. (a) Map showing DEM information and observation sites. Variations of $p\text{CO}_2$ (b), k (c) and areal CO₂ flux (d) across extreme drought, initial wet and post wet periods. The boxes with bars represent 25%–75% percentiles with 5%–95% percentiles. Black lines, white lines and dots show median, mean and all data, respectively. Asterisks indicate statistical significance: * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.



670

671 **Fig. 2.** Spectroscopic characteristics of DOC in the karst lakes. (a) Temporal patterns

672 of DOC, SUVA₂₅₄ and S₂₇₅₋₂₉₅ over the study periods. Violins with dots represent

673 Kernel Smooth distribution with all data. Black boxes with white bars show mean

674 with standard deviation (s.d.). Asterisks indicate statistical significance: * p < 0.05, **

675 p < 0.01, *** p < 0.001. (b) 3D view of primary DOC fluorophores identified by

676 PARAFAC analysis. (c) Distributions of FI, BIX and HIX over the study periods.

677 Dots correspond to all data of these fluorescent parameters. The waves show Kernel

678 Smooth distributions of the data.

679

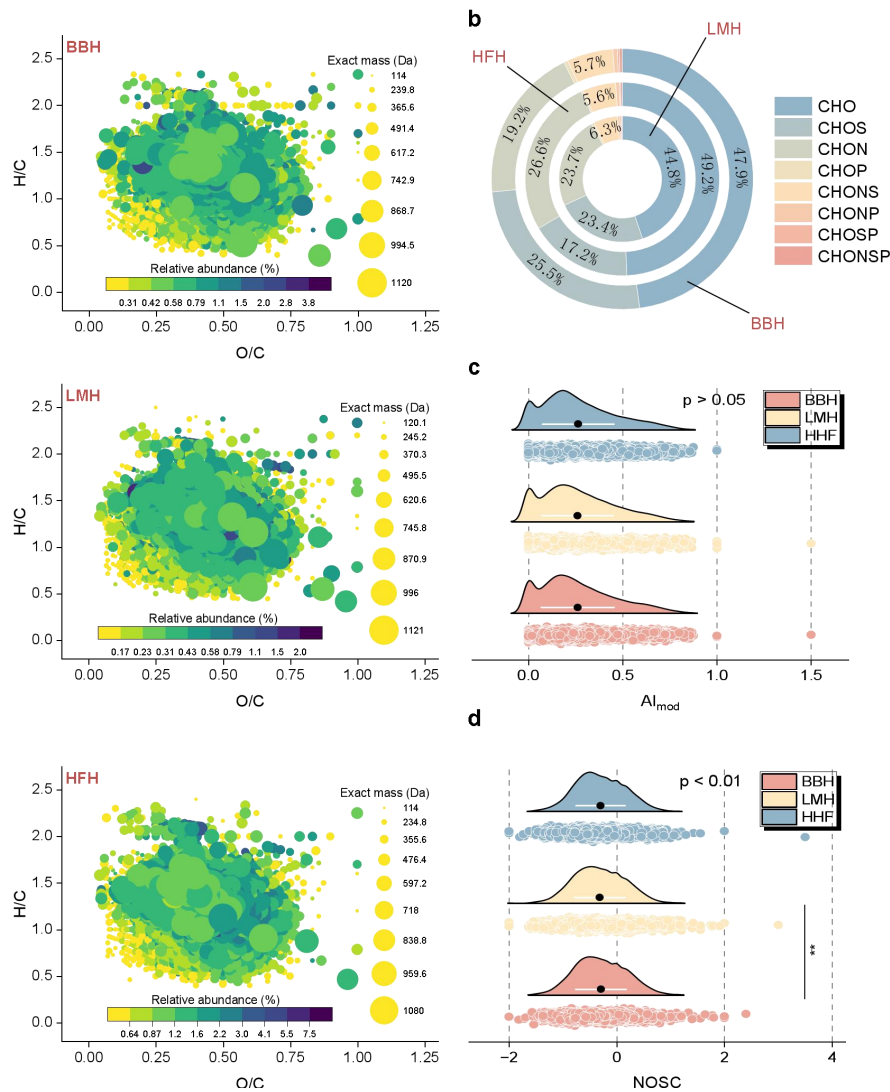


Fig. 3. Molecular characteristics of DOC in the karst lakes. (a) van Krevelen
Diagrams plotting H/C against O/C with exact mass information of DOC molecules.
(b) Proportions of primary DOC molecular formulas across the lakes. The AI-mod (c)
and NOSC (d) across lakes. Violins with dots represent Kernel Smooth distribution
with all data. Black spots with white bars show mean with standard deviation (s.d.).

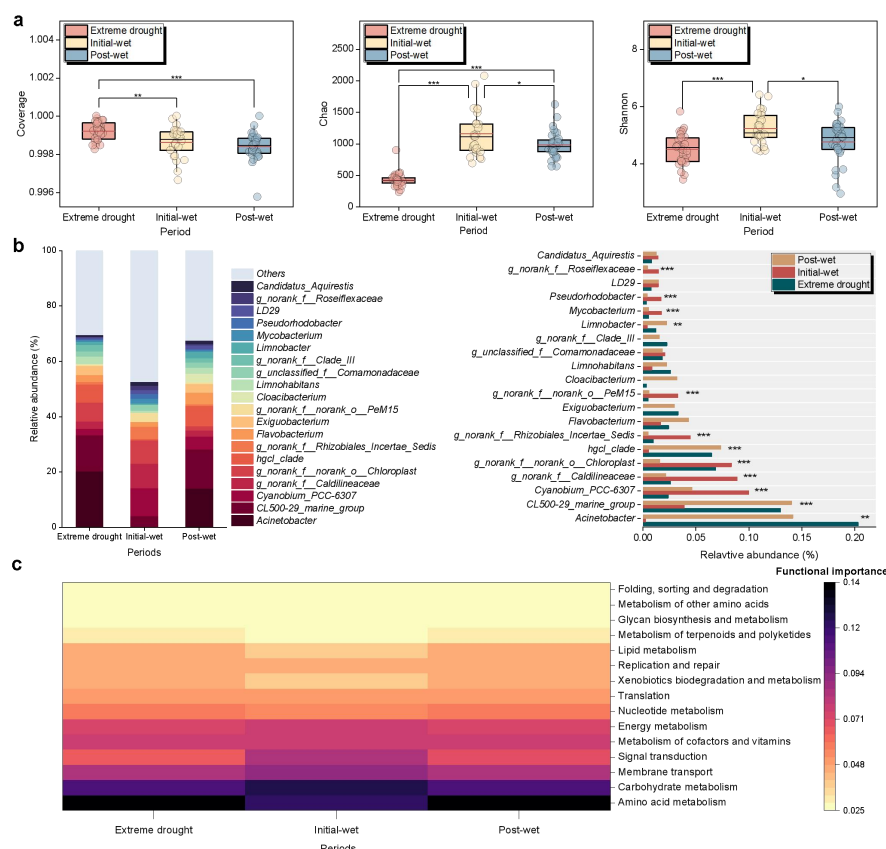


Fig. 4. Temporal variability of microbial communities and functions in the karst lakes.

(a) Alpha diversity indices regarding Coverage, Chao and Shannon over the study

periods. The boxes with bars represent 25%–75% percentiles with 5%–95%

percentiles. Black lines, white lines and dots show median, mean and all data,

respectively. Asterisks indicate statistical significance: * $p < 0.05$, ** $p < 0.01$, *** p

< 0.001 . (b) Comparison of the top 20 genera identified by relative abundance. (c)

Potential microbial functions predicted by Tax4Fun based on the KEGG database.

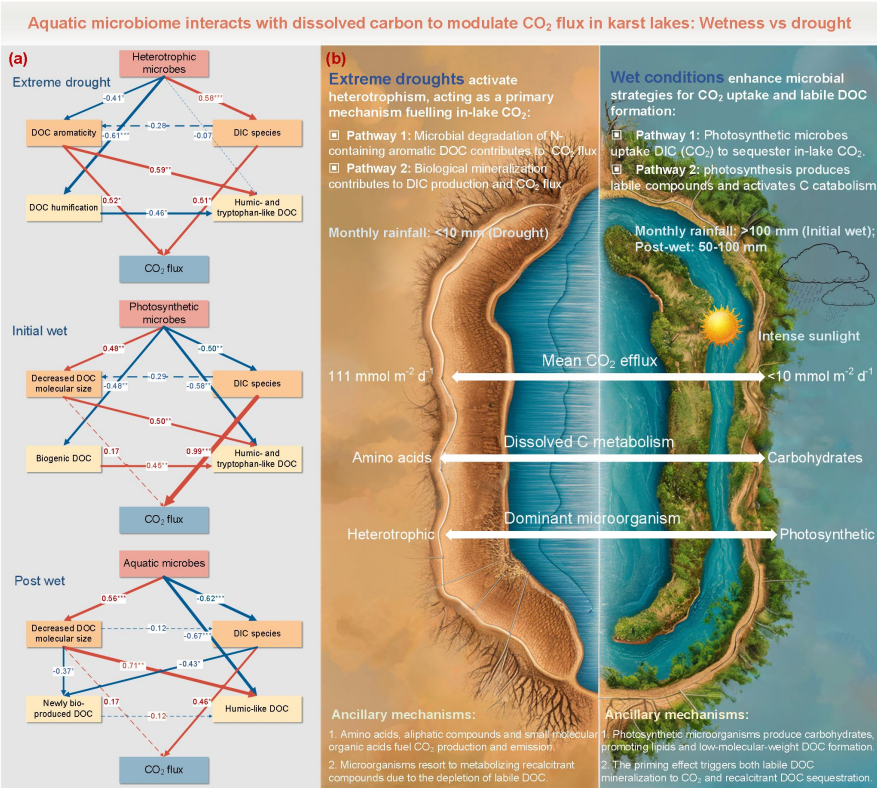


Fig. 5. Temporal pathways of aquatic microbiome-dissolved C interactions involved in CO₂ emission modulation in karst lakes. (a) The structural equation model showing temporal pathways of CO₂ flux driven by microbial communities and DIC-DOC turnover. Red and blue arrows represent positive and negative effects, respectively. Path coefficients are shown along the arrows. (b) A conceptual framework illustrating how microbiome interacts with dissolved carbon to modulate CO₂ emissions.



Table 1. Temporal patterns of pH, water temperature, total alkalinity and DIC species in the karst lakes.

	n	Min	Max	Median	Mean	Std. Dev
Extreme drought						
pH	32	6.71	8.30	7.20	7.40	0.48
Water temperature (°C)	32	14.5	19.5	16.2	16.2	0.99
Total alkalinity (µeq L ⁻¹)	32	2325.7	4940.9	2726.5	2960.5	703.2
DIC (µmol L ⁻¹)	32	2405.0	5272.5	2828.5	3114.2	668.8
HCO ₃ ⁻ (µmol L ⁻¹)	32	2290.3	4895.5	2645.1	2878.6	655.0
CO ₃ ²⁻ (µmol L ⁻¹)	32	3.95	244.6	16.0	40.9	51.3
Dissolved CO ₂ (µmol L ⁻¹)	32	18.0	602.4	177.0	194.7	153.5
Initial-wet						
pH	32	7.36	9.12	8.33	8.25	0.38
Water temperature (°C)	32	23.9	31.0	26.6	26.9	1.75
Total alkalinity (µeq L ⁻¹)	32	2178.7	6210.8	2895.3	3235.3	978.4
DIC (µmol L ⁻¹)	32	1538.7	6206.0	2688.8	3014.9	1042.5
HCO ₃ ⁻ (µmol L ⁻¹)	32	918.4	5970.6	2428.0	2745.2	1073.0
CO ₃ ²⁻ (µmol L ⁻¹)	32	35.7	752.7	218.2	243.5	155.1
Dissolved CO ₂ (µmol L ⁻¹)	32	0.58	160.3	10.4	26.2	36.0
Post-wet						
pH	32	6.80	9.15	8.22	8.18	0.47
Water temperature (°C)	32	12.9	29.5	27.1	25.8	3.45
Total alkalinity (µeq L ⁻¹)	32	2239.7	5435.0	2795.5	3133.6	862.9
DIC (µmol L ⁻¹)	32	2096.8	5180.7	2576.5	2908.9	777.2
HCO ₃ ⁻ (µmol L ⁻¹)	32	1546.7	4863.6	2441.5	2623.8	758.1
CO ₃ ²⁻ (µmol L ⁻¹)	32	6.27	887.9	195.5	253.5	222.5
Dissolved CO ₂ (µmol L ⁻¹)	32	1.04	315.9	14.3	31.6	57.1