

Influences on Chemical Distribution Patterns across the west Greenland Shelf: The Roles of Ocean Currents, Sea Ice Melt, and Freshwater Runoff

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Abstract The west Greenland shelf is a dynamic marine environment influenced by various physicochemical and biological processes. This study provides a comprehensive overview of the main factors affecting the distribution of macronutrients,

15 carbonate system parameters, and dissolved trace elements during July. Key drivers include major ocean currents, melting sea ice, and terrestrial freshwater runoff, each contributing uniquely to the cycling and spatial distribution of chemical constituents.

Major ocean currents, such as the southward-moving Baffin Island Current (BIC) and the northward-moving West Greenland Current (WGC), introduce water masses with distinct chemical signatures that shape the chemical composition of shelf waters.

Melting sea ice serves as an important source of freshwater and dissolved constituents for the marine environment. During the

20 study period, we were able to capture a distinct nutrient gradient following the east-to-west direction of sea ice retreat, with low nutrient levels in highly productive shelf waters and high nutrient levels in areas with prolonged ice cover. This process

also influenced the carbonate system, leading to changes in pH and aragonite saturation states, which is critical to the health of marine organisms. Terrestrial freshwater runoff, particularly from the Greenland Ice Sheet (GIS), replenishes macronutrients

25 in the photic zone, stimulating primary production and creating important CO₂ sinks. However, coastal surface waters become

more susceptible to acidification by the input of poorly buffered glacial freshwater. Understanding these key drivers is essential for predicting future changes in the marine chemistry and ecosystem dynamics on the west Greenland shelf, especially in the context of ongoing climate change within this high-latitude region.

1 Introduction

30 The physical system of the Arctic Ocean is inevitably changing, with profound implications for primary productivity (PP) and the biogeochemical cycling of nutrients and carbon across continental shelves and deep basins (Juranek, 2022). The complex coastal current system that connects the continental shelves of Greenland with the Arctic Ocean, the North Atlantic Ocean, and the Pacific Ocean strongly impacts PP (Vernet et al., 2021), creating an important environment for biota and fisheries (Krawczyk et al., 2021). The major ocean currents on the west side of Greenland are shown in Fig. 1 a. The circulation across
35 the west Greenland shelf and Baffin Bay is dominated by two major currents in the region: the southward-moving Baffin Island Current (BIC) along the Baffin Island continental slope and the northward-moving West Greenland Current (WGC) along the west Greenland continental slope (Rysgaard et al., 2020). On the western side of Baffin Bay, the surface-intensified BIC transports water from the Alaskan shelf and the western Beaufort Sea through the narrow and shallow channels of the Canadian
40 Arctic Archipelago (CAA), including Nares Strait, Jones Sound, and Lancaster Sound, across Davis Strait into the Labrador Sea (Tang et al., 2004; Curry et al., 2011; Aksenov et al., 2016). Arctic waters of Pacific origin that enter Baffin Bay through the CAA have been identified as sources of dissolved trace elements (e.g., dMn, dFe, dCu, dNi) (Colombo et al., 2020; Colombo et al., 2021; Jensen et al., 2022) as well as carbon (Azetsu-Scott et al., 2010; Shadwick et al., 2011b; Burgers et al., 2024). Further south, Davis Strait acts as a gateway for the export of dissolved trace elements (Krisch et al., 2022a) and dissolved inorganic nitrogen (Juranek, 2022) from the Arctic to the North Atlantic Ocean. On the eastern side of Baffin Bay,
45 the West Greenland Current (WGC) flows northward across Davis Strait, consisting of the warm, low-salinity West Greenland Coastal Current (WGCC) on the shelf and the warm, high-salinity West Greenland Slope Current (WGSC) (Curry et al., 2011). These currents transport waters of North Atlantic origin, which have been associated with anthropogenically elevated concentrations of dissolved lead (dPb) (Colombo et al., 2019), as well as enhanced biological productivity and species richness across the west Greenland shelf (Krawczyk et al., 2021).
50 In the last two decades, sea ice concentrations in Baffin Bay and the Labrador Sea have declined significantly due to climate change (Krawczyk et al., 2021), resulting in an earlier melt onset and delayed freeze-up (Stroeve and Notz, 2018; Ballinger et al., 2022). The reduction in sea ice cover, combined with increased inflow of Atlantic-sourced waters (Krawczyk et al., 2021) and elevated freshwater discharge (Møller et al., 2023), has been associated with an overall increase in PP and species richness along the west Greenland shelf (Krawczyk et al., 2021; Møller et al., 2023). In all seasonally ice-covered areas, intense but
55 short-lived phytoplankton blooms develop in low-salinity waters at the edge of melting and retreating sea ice from spring to late summer (Niebauer et al., 1995; Perrette et al., 2011). Phytoplankton growth is triggered by the stabilizing effect of sea ice meltwater-induced stratification and the increased solar irradiance as the ice cover shrinks (Strass and Nöthig, 1996; Perrette et al., 2011). The newly exposed waters are initially nutrient-rich but become rapidly depleted following the onset of the bloom (Niebauer et al., 1995). In early summer, macronutrient distributions (NO_x = nitrate + nitrite; phosphate, PO₄³⁻; silicate, Si(OH)₄) in surface waters of Baffin Bay decrease along a west-to-east gradient, generally following the sea ice coverage (Tremblay et al., 2002; Lafond et al., 2019). Although sea ice meltwater is considered a negligible net source of macronutrients,
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it can significantly enhance trace element concentrations in the receiving waters (Hölemann et al., 1999; Tovar-Sánchez et al., 2010; Kanna et al., 2014; Evans and Nishioka, 2019). Elevated trace element concentrations in Arctic sea ice have been linked to entrained sediments (Measures, 1999) or atmospheric deposition from continuous emissions in highly industrialized countries (McConnell and Edwards, 2008). Sea ice meltwater also influences the carbonate system of Baffin Bay by increasing alkalinity (AT) relative to salinity. When the effect of salinity is removed, sea ice meltwater has been shown to supply additional AT in the form of buffering ions such as carbonate (CO_3^{2-}) (Jones et al., 1983; Fransson et al., 2023). During sea ice formation, ikaite ($\text{CaCO}_3 \cdot 6\text{H}_2\text{O}$) precipitates within the sea ice and is reintroduced to the water column during ice melt. This dissolution process results in higher AT concentrations relative to dissolved inorganic carbon (CT), lower partial pressure of CO_2 ($p\text{CO}_2$), and higher pH and aragonite saturation state (Ω Aragonite) in surface waters (Rysgaard et al., 2011; Rysgaard et al., 2012; Fransson et al., 2013). However, on an annual cycle, sea ice does not serve as a net source or sink of these species to the underlying seawater. Instead, it serves as a mechanism for temporal and spatial redistribution, as these species are trapped during ice formation in autumn and winter, and subsequently released during ice melt – potentially at a different location due to drifting sea ice (Thomas et al., 2011).

The mass loss of the Greenland Ice Sheet (GIS) has increased sixfold since the 1980s, with the region north of Davis Strait experiencing the largest mass loss (Mouginot et al., 2019; Wouters and Sasgen, 2022). The export of glacial runoff from the GIS modifies the delivery of solutes and nutrients to near coastal regions (Hawkings et al., 2015) and the continental shelf (Cape et al., 2019). This glacier-derived freshwater alters marine concentrations of macronutrients (Hawkings et al., 2015; Hawkings et al., 2016; Hawkings et al., 2017; Hendry et al., 2019) and dissolved trace elements (Bhatia et al., 2013; Hawkings et al., 2020; Krause et al., 2021), influences local hydrography, and promotes the upwelling of nutrient-rich deep waters (Juul-Pedersen et al., 2015; Meire et al., 2017). These processes enhance marine productivity and extend the duration of the annual productive season (Oksman et al., 2022). However, glacial freshwater is characterized by low AT and carbonate ion concentration relative to marine waters, thereby reducing the capacity of glacially modified waters to resist changes in pH driven by atmospheric CO_2 uptake (Bates and Mathis, 2009; Chierici and Fransson, 2009; Fransson et al., 2015). Along the Greenlandic coast, AT dilution from glacial meltwater has been observed to drive corrosive conditions in surface waters, resulting in decreased Ω Aragonite and pH (Shadwick et al., 2013; Henson et al., 2023). Photosynthetic CO_2 uptake can partially mitigate this negative effect of AT dilution, as PP reduces CT and $p\text{CO}_2$ concentrations in surface waters and increases the carbonate saturation state (Chierici and Fransson, 2009; Meire et al., 2015; Hopwood et al., 2020).

In this study, we present recent measurements (July 2021) of the marine carbonate system, nutrient concentrations and dissolved (d; $< 0.45 \mu\text{m}$) trace element concentrations across the west Greenland shelf in southern Baffin Bay between 64°N and 71°N (Fig. 1 b). Given the above considerations, the objectives of this study are to: (a) understand the physicochemical processes that influence the internal cycling of chemical constituents within the distinct water masses of the shelf; and (b) resolve the regional and spatial differences in the distribution patterns on the west Greenland shelf during late summer that are driven by (i) major ocean currents, (ii) melting sea ice, and (iii) terrestrial freshwater runoff.

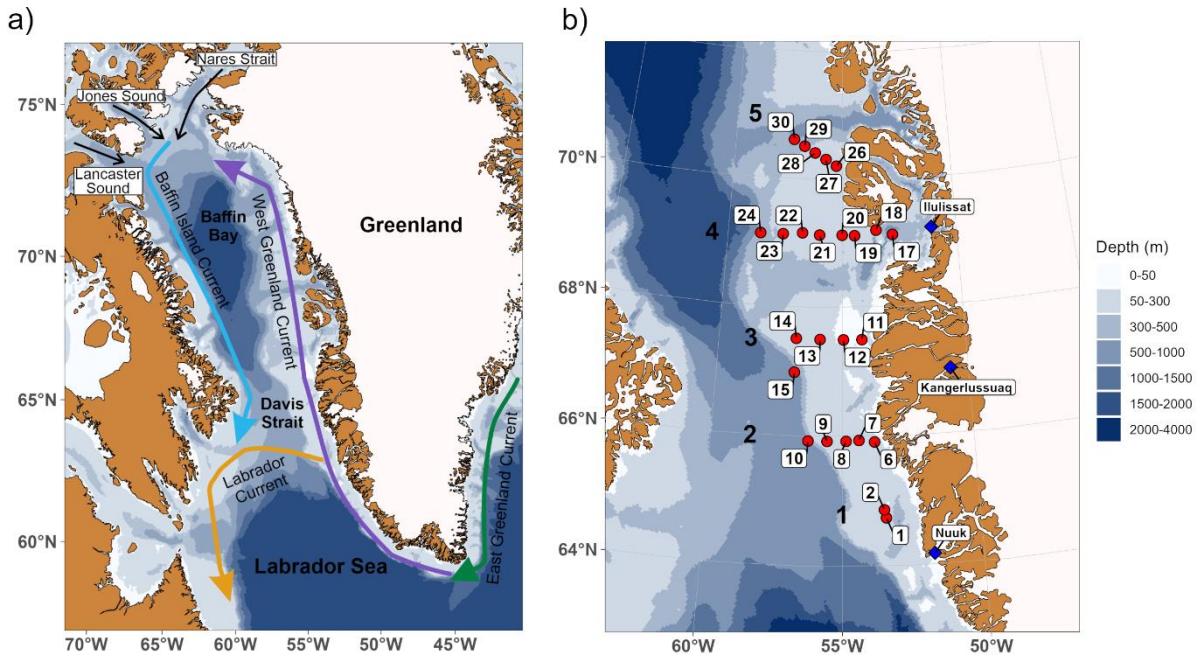


Figure 1: a) Schematic overview of the ocean currents and bathymetry in Baffin Bay and Davis Strait. Colored lines with arrows indicate the direction of different ocean currents. Bathymetry data from the GEBCO 2023 grid (GEBCO Bathymetric Compilation Group 2023). b) Map of the study area during DANA 6/21 with red points indicating the sampling locations and their affiliation to a certain transect (1-5).

100 2 Material and Methods

2.1 Study Area

A map of the study area with stations indicated by red dots is given in Fig. 1 b. General information about the selected stations and samples used in this study is provided in Table S 1. Samples were taken along two transects to the south and three transects to the north of Davis Strait along the west Greenland shelf. Transect 1 was located in a shallow area (< 120 m depth) north of Nuuk. Transects 2 and 3 started close to the mouth of Kangerlussuaq Fjord and Nassuttooq Fjord, respectively. Station 15, which is part of transect 3, had to be moved further south because of the presence of sea ice. Transect 4 started in Disko Bay approximately 70 km from the mouth of the Ilulissat Icefjord, where the Jakobshavn Isbræ terminates. Transects 2, 3, and 4 continued westward away from the coast towards the shelf edge. Transect 5 started close to Disko Island and continued towards the northwest.

110 2.2 Sampling

Water column samples were taken at 25 stations as conductivity-temperature-depth (CTD) casts along five transects during ten consecutive sampling days between 18 and 28 July 2021 on board RV *Dana* (DANA 6/21, Fig. 1 b, Table S 1). A SBE 911 CTD+ unit (Sea-Bird Scientific; Bellevue, USA) was used to collect hydrographic profiles, including temperature, salinity,

pressure, and dissolved oxygen. Two oxygen sensors were part of the CTD system and calibrated prior to the cruise. By cross-referencing the data of the two sensors, we were able to monitor their drift during the cruise and correct the data accordingly. Samples were collected in 10 L metal-free Niskin water samplers (Ocean Test Equipment; Fort Lauderdale, USA). Detailed information about each sample is given in Table S 1, with sample IDs corresponding to the respective station and sampling depth.

2.2.1 Macronutrients

120 Water samples for nutrient analysis were collected in acid-washed 100 mL HDPE (high-density polyethylene) bottles. The bottles were preconditioned three times with sample water before being filled. The samples were stored frozen at -20°C without filtration until analysis.

2.2.2 Dissolved trace elements

125 Bulk samples for multi-element analysis were filled into acid-cleaned 2 L HDPE bottles. The bottles were rinsed three times with sample water before being filled. Immediately after sampling, the water samples were filtered as duplicates using high-purity, metal-free PFA (Perfluoroalkoxy alkane) filtration bombs (Savillex, Eden Prairie, USA) pressurized with nitrogen 5.0 (Air Liquide Danmark; Taastrup, Denmark) and acid-washed polycarbonate filters (0.45 µm, 47 mm diameter, Whatman, UK) in a clean bench (Erlab; Val de Reuil Cedex, France). More information about the filtration procedure can be found in Przibilla *et al.* (2023). The filtered samples were collected in pre-cleaned 250 mL HDPE bottles and stabilized using trace metal grade 130 HCl (Fisher Scientific; Waltham, USA) before storage and shipping.

2.2.3 Carbonate system parameters

135 Samples for AT and CT were collected in 300 mL BOD (biological oxygen demand) bottles (Environmental Express, Charleston, USA) with an addition of 300 µL saturated mercury chloride solution (Carl Roth, Karlsruhe, Germany). The bottles were sealed with ground-glass stoppers (Wheaton Science Products, New Jersey, USA), silicone- and halogen-free grease (type M, Apiezon, Manchester, UK), and plastic caps (Wheaton Science Products, New Jersey, USA), leaving no headspace. The samples were stored in darkness at ambient temperature until analysis.

2.3 Sample preparation and chemical analysis

2.3.1 Macronutrients

140 Nitrite, phosphate, and silicate were measured on a SmartChem 200 discrete analyser (AMS Alliance, Rome, Italy) following the reagent concentrations for the manual method in Hansen and Koroleff (1999), with the inclusion of a 150 g L⁻¹ solution of sodium dodecylsulfate as a dispersant for phosphate analysis. Nitrate was measured following the vanadium chloride reduction

technique described by Schnetger and Lehners (2014). Community reference material for nutrients in seawater supplied by Kanso Technos Co. LTD, Japan, was used for quality control.

2.3.2 Dissolved trace elements

145 All equipment for trace metal analysis was acid-washed prior to use and rinsed with type I reagent-grade water (resistivity: 18.2 MΩ·cm) obtained from a Milli-Q Integral water purification system (Merck; Darmstadt, Germany) to prevent contamination. Dissolved trace elements (dV, dFe, dMn, dCo, dNi, dCu, dCd, and dPb) in seawater were measured with a seaFAST SP2 system (Elemental Scientific; Omaha, USA) coupled online to an inductively coupled plasma mass spectrometer (ICP-MS) (Agilent 7900, Agilent Technologies; Tokyo, Japan). The seaFAST SP2 system contained two columns filled with 150 Nobias chelate-PA1 resin (HITACHI High-Tech Fielding Corporation; Tokyo, Japan). Detailed operating parameters and instrument configurations are given in Table S 2. More information about the analytical procedure can be found in Ebeling *et al.* (2022). For method validation, the certified reference material (CRM) NASS 7 (National Research Council Canada; Ottawa, Canada) as well as an in-house reference material (KBA-QC) mixed from single element standards (Carl Roth GmbH, Karlsruhe, Germany or Sigma-Aldrich, Missouri, USA) and custom-made multi-element standards (all traceable to NIST 155 standards) of different compositions (Inorganic Ventures, Christiansburg, USA) were used. Recovery rates (between 97 % and 119 %) are given in Table S 3.

2.3.3 Carbonate system parameters

160 Measurements of AT and CT were performed using a VINDTA 3 C system (Marianda; Kiel, Germany). AT and CT were determined simultaneously by potentiometric titration using an 800 Dosino (Metrohm; Filderstadt, Germany) with an Aquatrode plus (Metrohm; Filderstadt, Germany) and coulometric titration using a CM5017O coulometer (UIC; Illinois, USA), respectively. Both instruments were calibrated against seawater CRMs (Scripps Institution of Oceanography; San Diego, USA) to ensure a precision of $\pm 1 \mu\text{mol kg}^{-1}$ for each parameter.

2.4 Data analysis

165 The raw CTD data were processed and averaged into 0.5 dbar pressure bins. The processed CTD data were used to construct water depth profiles of potential temperature, salinity, oxygen (Fig. 2), and potential density (Fig. S 1). Calculations were performed using R (version 4.2.2; (R Core Team, 2022)) and RStudio (version 2023.06.0; (Posit team, 2023)). Bivariate, linear data interpolation in combination with a surface approximation using multilevel B-splines with the “interp” function (package akima; (Akima and Gebhardt, 2022)) and the “mba.surf” function (package MBA; (Finley *et al.*, 2022)) was used before plotting.

170 For trace elements analysis, filtration duplicates were obtained and measured as triplicates ($n = 6$). Outliers were identified using a Dean-Dixon outliers test and removed from further analysis. Mean concentrations (μ) and standard deviation (SD ; σ) were estimated, considering twice the SD to represent $\mu \pm 2\sigma$. Limits of detection (LOD) and limits of quantification (LOQ)

are given in Table S 3 and were calculated according to DIN 32645:2008-11, based on measurements of filtrations blanks ($n = 8$). The LOD is defined as three times the standard deviation ($3 \times \text{SD}$) of the blank measurements and represents the 175 lowest concentration that can be reliably distinguished from background noise, though not necessarily quantified with precision. The LOQ is defined as ten times the standard deviation ($10 \times \text{SD}$) and represents the lowest concentration that can be quantitatively determined with acceptable accuracy and precision (DIN e.V., 2008).

The sum parameter NOx for nitrate and nitrite (NOx = nitrate + nitrite) was established and used throughout the discussion. The LOQs for the nutrient analysis are given in Table S 3.

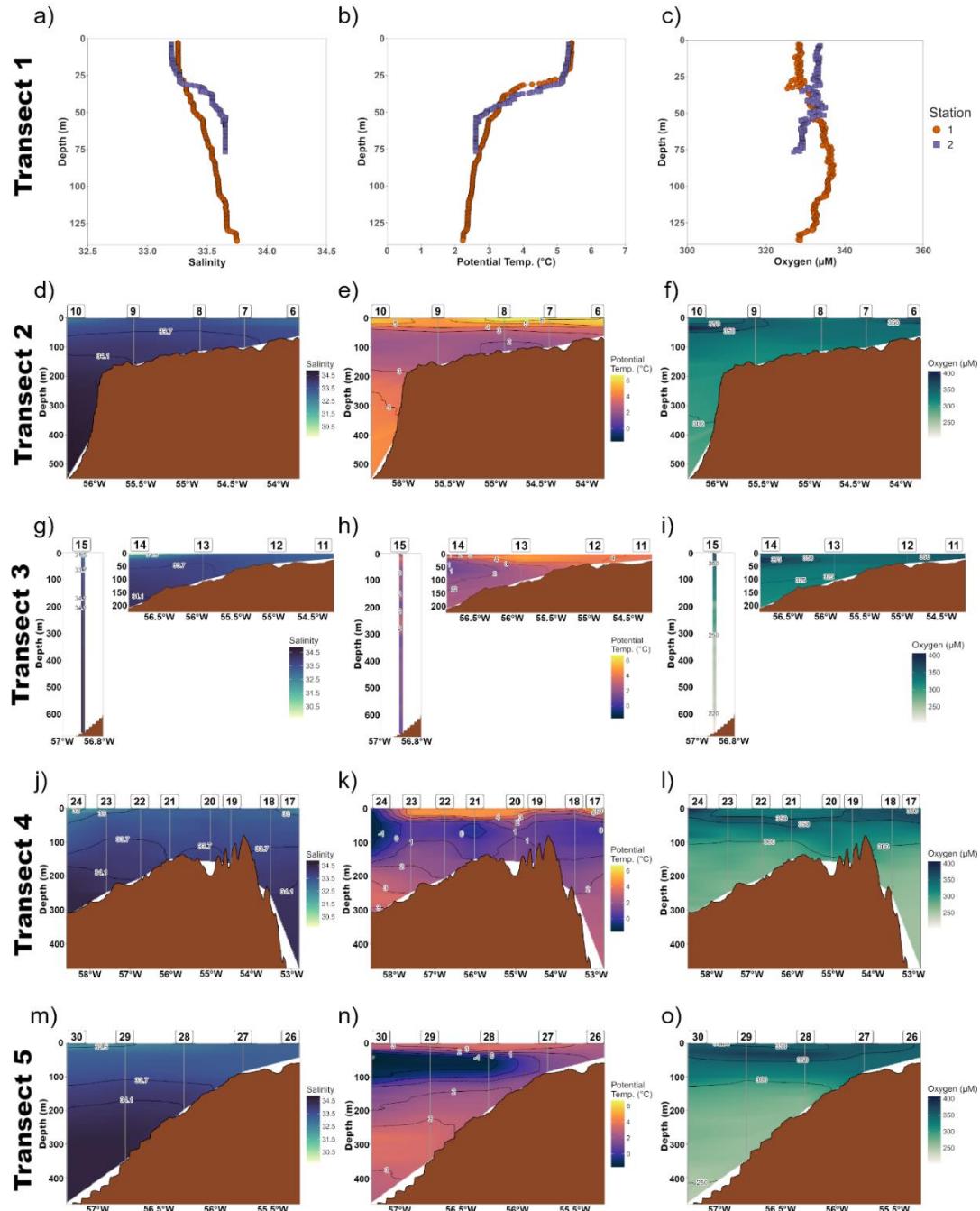
180 Parameters of the carbonate system (pH, $p\text{CO}_2$, Revelle factor, and Ω Aragonite) were calculated using the CO2Sys Macro (Pierrot et al., 2011) with salinity, temperature, AT and CT as input variables. As input parameters, we used the dissociation constant by Mehrbach et al. (1973), refit by Dickson and Millero (1987), the HSO_4^- dissociation constant by Dickson (1990) and the seawater pH-scale (Dickson, 1990; Dickson and Millero, 1987; Mehrbach et al., 1973). We used the CO2Sys Macro to calculate temperature-normalized CT (CT_{temp}) according to Wu *et al.* (2019). The calculation was based on the median 185 potential temperature of WGSW (3.23°C , $n = 465$) from the CTD profiles, filtered for a depth range of 30 to 50 m. Further input parameters were observed salinity, AT, and the in situ $p\text{CO}_2$.

For statistical analysis, the data of the different parameters were merged according to station and sampling depth, obtaining 190 one data matrix with 17 parameters and 41 observations. As a minor proportion of the total observations ($< 3\%$) were below the LOD or LOQ, a single imputation method was applied (Helsel, 2005; Jain, 2016). Values below the LOD were replaced with a random value between zero and the LOD, while values below the LOQ were replaced with a random value between the LOD and LOQ. Each parameter was tested for normality (Shapiro-Wilk test, $\alpha = 0.01$) and transformed (log- or Box Cox- 195 transformation) if the normality criteria was not met (Table S 4). The data were standardized, and principal component analysis (PCA) with varimax rotation was performed using the “principal” function (package psych; (Revelle, 2024)). Three principal components (PCs) were selected based on the Guttman-Kaiser criterion and the scree plot, retaining only those with eigenvalues greater than 1 (Bro and Smilde, 2014). A broken stick analysis was performed to evaluate the significance of variable loadings and identify which variables are associated with specific PCs (Peres-Neto *et al.*, 2003).

3 Results

The distributions of basic oceanographic parameters (salinity, potential temperature, oxygen, apparent oxygen utilization (AOU)), macronutrients (NOx, silicate, phosphate), dissolved trace elements (dV, dMn, dFe, dCo, dNi, dCu, dCd, and dPb), 200 and carbonate system parameters (AT, CT, pH, $p\text{CO}_2$, Revelle factor, Ω Aragonite) are presented in Sect. 3.1 to 3.4. In Fig. 2, detailed water column profiles of salinity, potential temperature, and oxygen are provided. An overview of minimum and maximum values of macronutrients, carbonate system parameters, and trace elements are presented in Table 1. Figures 3 to 5 illustrate the vertical distributions of macronutrients, dissolved trace elements, and carbonate system parameters for each station and transect. Surface water concentration plots for individual parameters are provided in the supplementary material

205 (Fig. S 2 to Fig. S 4). In Sect. 3.5, we classify the water masses present across the west Greenland shelf based on characteristic
salinity and potential temperature ranges.



210 **Figure 2: Water depth profiles illustrating the spatial distribution of salinity, potential temperature, and oxygen along transect 1 (a-c), transect 2 (d-f), transect 3 (g-i), transect 4 (j-l), and transect 5 (m-o). For transect 1, data are not shown as interpolated profiles
due to limited data availability. For transect 3, station 15 is displayed separately because of its southerly location (refer to Fig. 1 b).**

3.1 Basic oceanographic parameters

The distribution of salinity, potential temperature, and oxygen are shown as water depth profiles for each transect in Fig. 2. Ranges of minimum and maximum values for salinity, potential temperature, oxygen and AOU are summarized in Table S 5. Salinity was generally low in surface waters at stations close to the coast (stations 6, 11 and 17) and towards the west of each transect (stations 14, 15, 24, 29 and 30), with a minimum value of 29.70 recorded at the surface of station 14 (Fig. 2 g). The highest salinity (34.89) was observed in deep waters at station 10, between 530 and 558 m depth (Fig. 2 d). Potential temperature was highest in surface waters in the southeastern part of Davis Strait, particularly at transect 2 (stations 6, 7, 8, and 10), and at station 17 in Disko Bay. Lower potential temperatures were found further offshore in the northwestern part of Davis strait. The minimum potential temperature of -1.63 °C was recorded in intermediate waters (40 to 55 m) at station 30 (Fig. 2 n), while the maximum value of 6.7 °C occurred in surface waters at station 8 (Fig. 2 e). In general, higher oxygen concentrations and lower AOU values were found in the photic zone (1.5 to 40 m), either near the Greenlandic coast at the beginning of transects (stations 6 and 17) or at the western ends (stations 10, 14, 15, and 24). The lowest oxygen concentration (204 $\mu\text{mol L}^{-1}$) and highest AOU (147 $\mu\text{mol L}^{-1}$) were observed in deep waters at station 15 (645 to 673 m) (Fig. 2 i). Conversely, the highest oxygen concentration (406 $\mu\text{mol L}^{-1}$) and lowest AOU (-72 $\mu\text{mol L}^{-1}$) were measured in intermediate waters (34 to 37 m) at station 10 (Fig. 2 f).

Table 1: Minimum and maximum values of macronutrients, carbonate system parameters and trace elements across the study area.

Parameter	Unit	Minimum		Maximum		Literature data
		Value	Station [Depth in m]	Value	Station [Depth in m]	
NOx	$\mu\text{mol L}^{-1}$	0.21	13 [30]	17.85	15 [667]	2015: 0.032 – 24.3 ¹ 2016: 0.05 – 28.5 ²
Silicate	$\mu\text{mol L}^{-1}$	0.51	22 [3]	35.95	15 [667]	2015: 0.32 – 103.7 ¹ 2016: 0.11 – 131.4 ²
Phosphate	$\mu\text{mol L}^{-1}$	0.12	22 [3]	1.43	15 [667]	2015: 0.46 – 1.97 ¹ 2016: 0.06 – 2.2 ²
AT	$\mu\text{mol kg}^{-1}$	2062	15 [3]	2305	10 [551]	2015: 2156 – 2294 ⁴ 2016: 2176 – 2310 ³
CT	$\mu\text{mol kg}^{-1}$	1949	15 [3]	2214	15 [667]	2015: 2008 – 2255 ⁴ 2016: 1937 – 2277 ³
CT:AT		0.907	17 [3]	0.968	15 [667]	NA
pH		7.86	15 [667]	8.20	15 [32]	2015: 7.40 – 7.81 ¹
$p\text{CO}_2$	μatm	249	15 [32]	559	15 [667]	NA
Revelle factor		11.85	17 [3]	17.80	15 [667]	NA
\mathcal{Q} Aragonite		0.90	15 [667]	2.17	17 [3]	NA
dV	ng L^{-1}	1270 ± 30	24 [3]	1810 ± 60	10 [551]	NA
dMn	ng L^{-1}	54.8 ± 1.8	24 [317]	503 ± 20	12 [37]	$13.5 \pm 1.7 – 306.0 \pm 2.6$ ¹
dFe	ng L^{-1}	56.9 ± 2.5	27 [15]	630 ± 30	18 [256]	$20.0 \pm 0.6 – 112 \pm 8$ ¹
dCo	ng L^{-1}	3.81 ± 0.15	10 [551]	14.7 ± 0.6	11 [3]	NA
dNi	ng L^{-1}	232 ± 7	6 [3]	381 ± 4	24 [35]	$240 \pm 12 – 376 \pm 5$ ¹
dCu	ng L^{-1}	85.4 ± 2.5	10 [551]	206 ± 3	24 [3]	$115 \pm 10 – 333.0 \pm 0.7$ ¹
dCd	ng L^{-1}	3.1 ± 0.7	8 [4]	52.2 ± 1.2	15 [667]	$20.0 \pm 0.5 – 49.0 \pm 0.6$ ¹
dPb	ng L^{-1}	0.99 ± 0.08	28 [186]	4.04 ± 0.13	10 [551]	$0.52 – 6.0$ ¹

Note. The measurement uncertainty of trace elements corresponds to 2 SD. Literature data from Geotraces cruise GN02 in 2015 (station BB1, BB2 and BB3) and GreenEdge cruise in 2016 (stations G100 to G3000). ¹(GEOTRACES Intermediate Data Product Group, 2023). ²(Bruyant et al., 2022). ³(Miller et al., 2020). ⁴(H. Thomas, personal communication, June 28, 2024).

3.2 Macronutrients

The overall nutrient distribution is shown as vertical profiles in Fig. 3 and as sea surface concentration plots in Fig. S 2. In 235 surface waters, nutrient concentrations were uniformly low across the shelf. NO_x concentrations were particularly low (< LOQ = 0.21 $\mu\text{mol L}^{-1}$) in surface waters and shallow shelf waters. Exceptions to this trend were observed at station 17 in Disko Bay and at stations 14, 15, 24 and 30 towards the west of each transect, where surface waters with higher nutrient concentrations were present (refer to Fig. S 2). Below the biological productive zone, nutrient concentrations gradually increased with depth, reaching maximum concentrations at station 15 (667 m) (refer to Table 1). Sherwood et al. (2021) 240 reported similarly high nutrient concentrations in these deep waters, which are further discussed in Sect. 4.1.2.

In comparison to data collected during 2015 (GEOTRACES Intermediate Data Product Group, 2023) and 2016 (Bruyant et al., 2022), maximum nutrient concentrations of this study are lower, which we attribute to regional differences in the study 245 areas, particularly with respect to the maximum sampling depth. We measured maximum values at the shelf edge at 667 m, whereas the referenced studies reported maximum values at stations located in central Baffin Bay, where sampling extended to significantly greater depths.

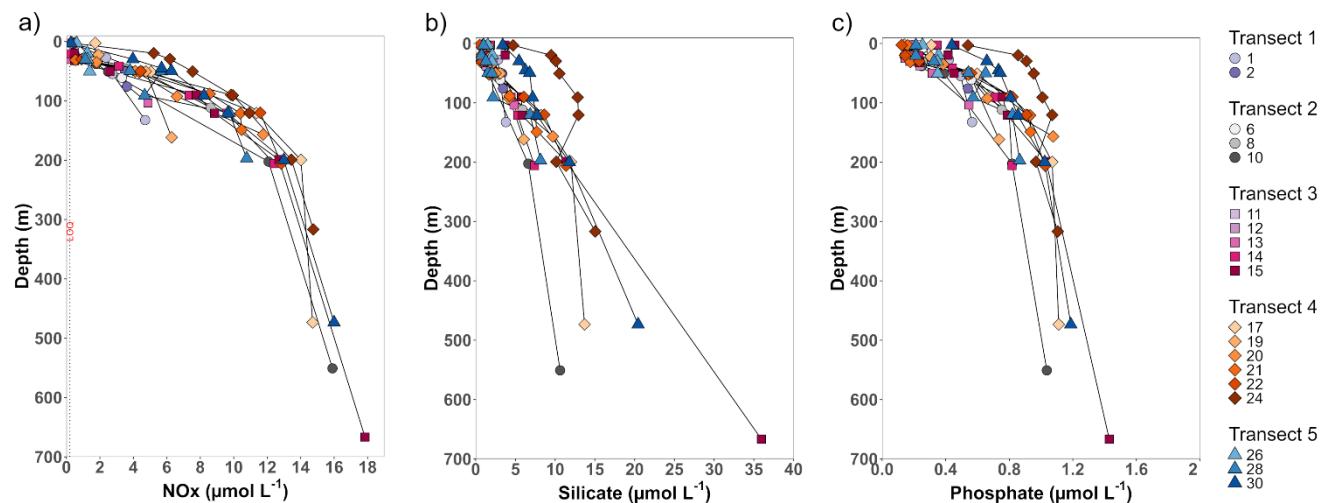


Figure 3 Vertical profiles of nutrient concentrations across the study area. Each data point is identified by shape (transect number) and color (station number).

3.3 Dissolved trace elements

250 The distributions of trace elements are shown as vertical profiles in Fig. 4 and surface water concentrations in Fig. S 3. The vertical profiles of dV and dCu showed opposing trends (Fig. 4 a and f). With depth, dV concentrations increased, while dCu concentrations decreased. The dV concentrations ranged from $1270 \pm 30 \text{ ng L}^{-1}$ (station 24; 3 m) to $1810 \pm 60 \text{ ng L}^{-1}$ (station 10; 551 m), while dCu concentrations ranged from $206 \pm 3 \text{ ng L}^{-1}$ (station 24; 3 m) to $85.4 \pm 2.5 \text{ ng L}^{-1}$ (station 10; 551 m), both matching minimum and maximum occurrences of salinity. Surface waters close to the coast of Greenland and at the 255 western end of each transect showed lower dV concentrations (Fig. S 3 a) and higher dCu concentrations (Fig. S 3 f),

respectively. The distributions of dFe, dNi and dCd showed low concentrations in surface and subsurface waters, which gradually increased with depth (Fig. 4 c, e, and g; Fig. S 3 c, e, and g). Maximum concentrations were observed at depth, with dFe reaching $630 \pm 30 \text{ ng L}^{-1}$ (station 18; 256 m), dNi reaching $307 \pm 9 \text{ ng L}^{-1}$ (station 15; 667 m) and dCd reaching $52.2 \pm 1.2 \text{ ng L}^{-1}$ (station 15; 667 m). Minimum concentrations of dFe, dNi and dCd were present in surface waters (refer to 260 Table 1), particularly at stations with some distance to the coast. At several stations, dFe concentrations in surface waters were below the LOQ ($< 50 \text{ ng L}^{-1}$). Surface concentrations of dFe and dNi (Fig. S 3 c and e) were elevated near the coast, decreased with increasing distance along each transect and increased again towards the west of each transect. In contrast, surface dCd concentrations were rather uniform across the shelf, with a slight increase toward the westernmost stations (Fig. S 3 g). The 265 concentrations of dMn and dCo decreased from surface to subsurface waters and increased again with depth at stations located on the shelf (Fig. 4 b and d; Fig. S 3 b and d). In contrast, at the shelf edge, dMn and dCo concentrations remained stable with depth, resulting in minimum values of $54.8 \pm 1.8 \text{ ng L}^{-1}$ for dMn (station 24; 317 m) and of $3.81 \pm 0.15 \text{ ng L}^{-1}$ for dCo (station 10; 551 m). Elevated surface concentrations of both dMn and dCo were observed at station 17 in Disko Bay and at stations 11 and 12, located near the mouth of the Nassuttooq Fjord (Fig. 4 b and d; Fig. S 3 b and d). Surface concentrations of dMn and 270 dCo generally decreased with increasing distance from the coast. The depth profile of dPb did not exhibit a consistent vertical pattern (Fig. 4 h). South of Davis Strait, elevated dPb concentrations were observed across transect 2 (refer to Fig. S 3 h), including a maximum dPb concentration of $4.04 \pm 0.13 \text{ ng L}^{-1}$ in the deep waters of station 10. In contrast, north of Davis Strait, dPb concentrations were generally lower, falling below the LOQ ($< 0.98 \text{ ng L}^{-1}$) in the deep waters of station 15 and the intermediate to deep waters of Disko Bay (stations 17 and 18). The minimum dPb concentration of $0.99 \pm 0.08 \text{ ng L}^{-1}$ was observed in the deep waters of station 28.

275 Compared to literature values, our results for trace element concentrations are generally consistent with data collected during the 2015 Geotraces cruise GN02 (GEOTRACES Intermediate Data Product Group, 2023). Variations of minimum and maximum values are attributed to differences in sampling locations. While the present study took place on the shelf, capturing the influence of coastal runoff and sea ice meltwater, the Geotraces cruise focused on offshore stations located further north in central Baffin Bay.

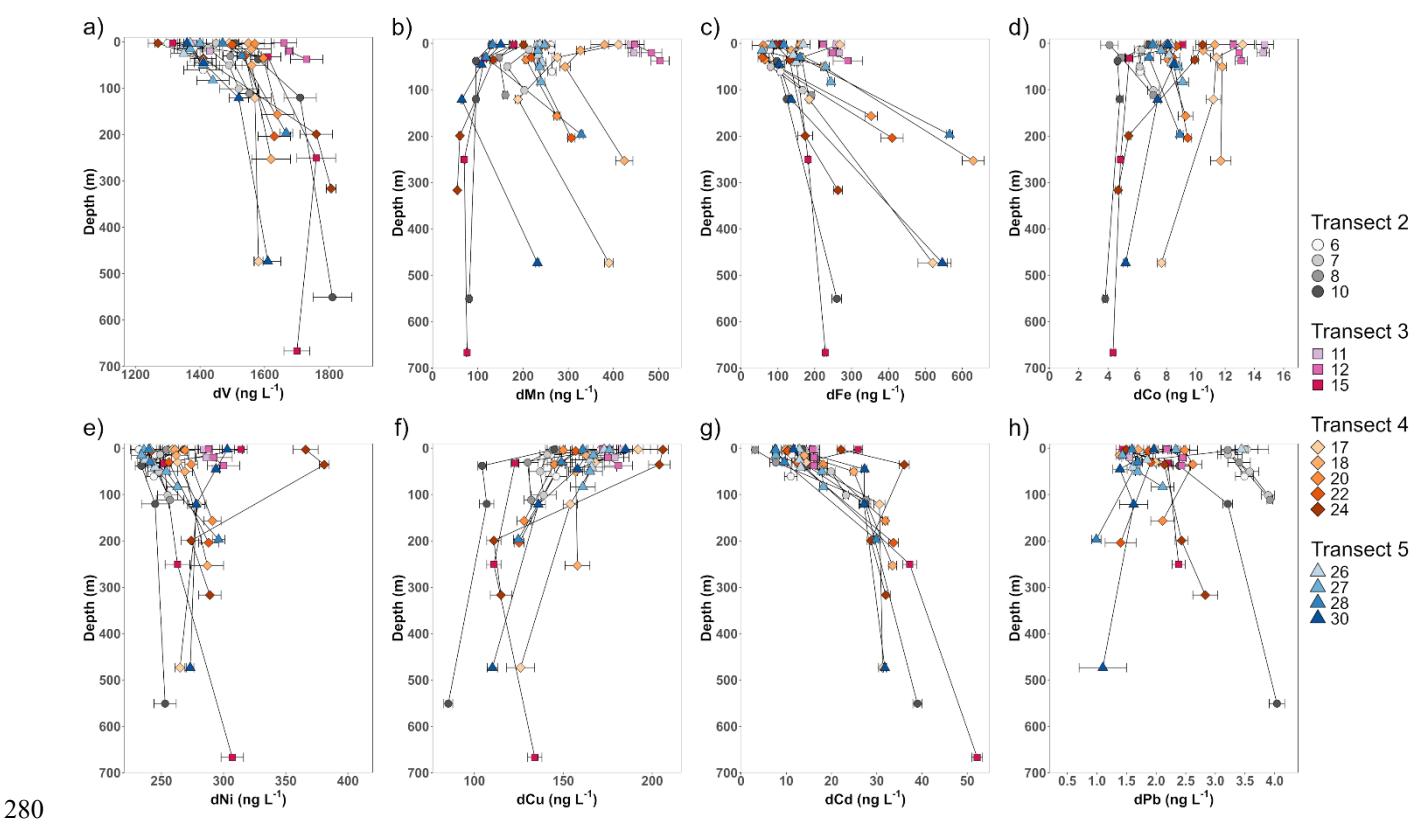


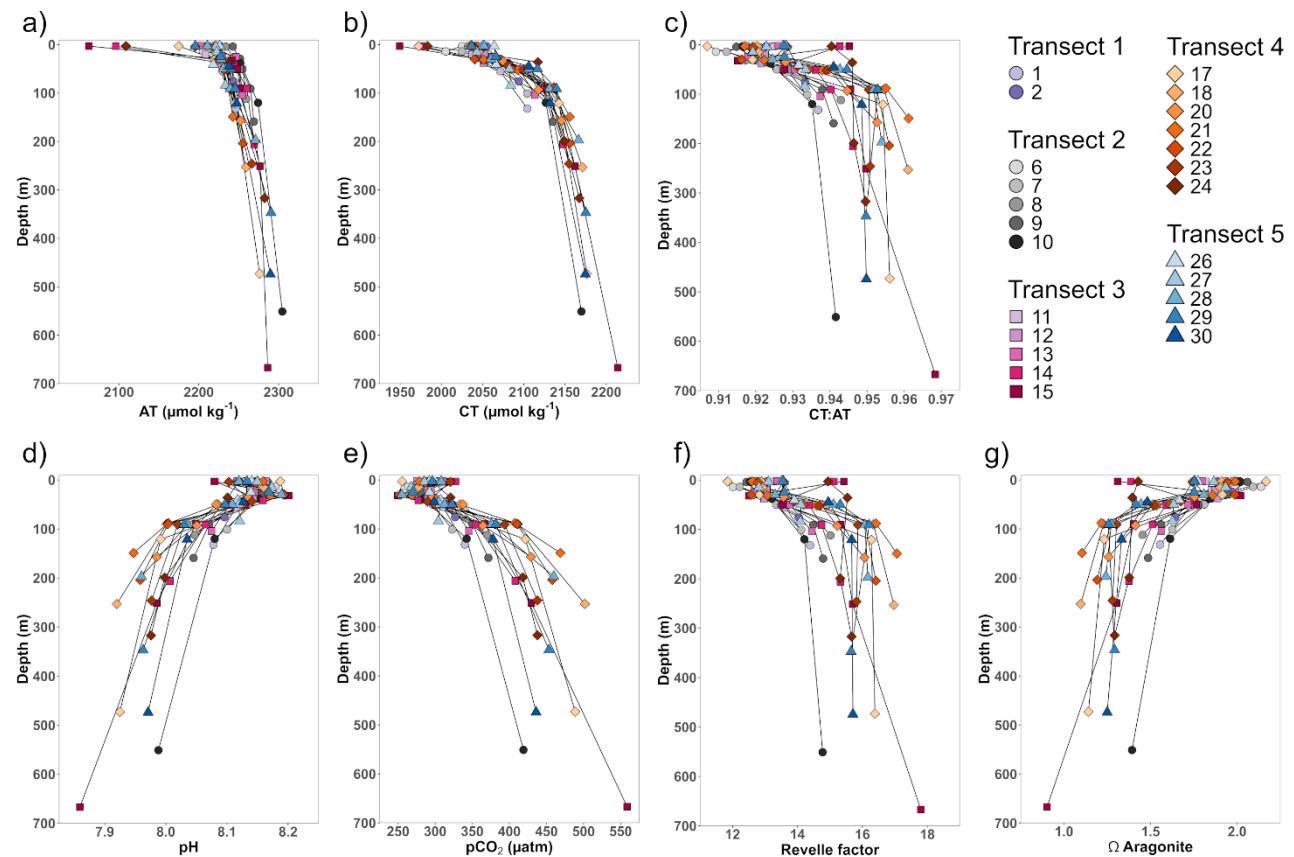
Figure 4 Vertical profiles of trace element concentrations across the study area. Each data point is identified by shape (transect number) and color (station number). Error bars correspond to 2 SD.

3.4 Carbonate system parameters

The vertical profiles of AT and CT (Fig. 5 a and b) show a gradual increase with depth. Minimum concentrations of AT and 285 CT were present in the low-salinity surface waters at station 15 (AT: 2062 $\mu\text{mol kg}^{-1}$; CT: 1494 $\mu\text{mol kg}^{-1}$). While maximum CT concentrations of 2214 $\mu\text{mol kg}^{-1}$ were observed in the deep waters of station 15, AT concentrations followed the salinity gradient, reaching a maximum value of 2305 $\mu\text{mol kg}^{-1}$ in the deep waters of station 10. The vertical profiles of pH and $p\text{CO}_2$ (Fig. 5 d and e) show opposing trends, with pH increasing and $p\text{CO}_2$ decreasing towards intermediate waters of the photic zone. Below the photic zone, pH gradually decreases and $p\text{CO}_2$ increases with depth. This trend results in a pH maximum 290 (8.20) and a $p\text{CO}_2$ minimum (249 μatm) in the photic zone of station 15 at 32 m, while the deep waters of the same station show a pH minimum (7.86) and a $p\text{CO}_2$ maximum of (559 μatm). The vertical profiles of the CT:AT ratio, Revelle factor and Ω Aragonite (Fig. 5 c, f and g) show that these parameters remain relatively stable within the upper 25 m of the water column. Below this depth, the CT:AT ratio and Revelle factor gradually increase, while Ω Aragonite decreases. The maximum CT:AT 295 ratio (0.968) was observed in the deep waters of station 15, coinciding with a maximum Revelle factor of 17.80 and a minimum Ω Aragonite value of 0.90.

The surface water concentrations of the carbonate system parameters are given in Fig. S 4. In general, stations closest to the Greenland coast exhibited lower AT and CT values, which increased with distance to the coast. Following this trend, surface water pH increased and $p\text{CO}_2$ values decreased along each transect with increasing distance from the coast. In coastal surface waters at stations 6 and 11, higher CT:AT ratios (0.920 and 0.925) coincided with higher Revelle factors (12.8 and 13.3) and lower Ω Aragonite values (1.93 and 1.82), indicating a reduced buffering capacity in coastal waters. In contrast, surface waters in Disko Bay (station 17; 3 m) exhibited a lower CT:AT ratio (0.907), driven by a low CT concentration of $1972 \mu\text{mol kg}^{-1}$ and a corresponding low $p\text{CO}_2$ value of $255 \mu\text{atm}$. This was accompanied by a high pH (8.19), a low Revelle factor (11.85), and a maximum Ω Aragonite value of 2.17, indicating the higher buffering capacity of this water. In the westernmost surface waters of each transect, AT and CT values were again lower, but CT:AT ratios were among the highest observed. As a result, $p\text{CO}_2$ values and Revelle factors were elevated at these stations, while pH and Ω Aragonite values were reduced.

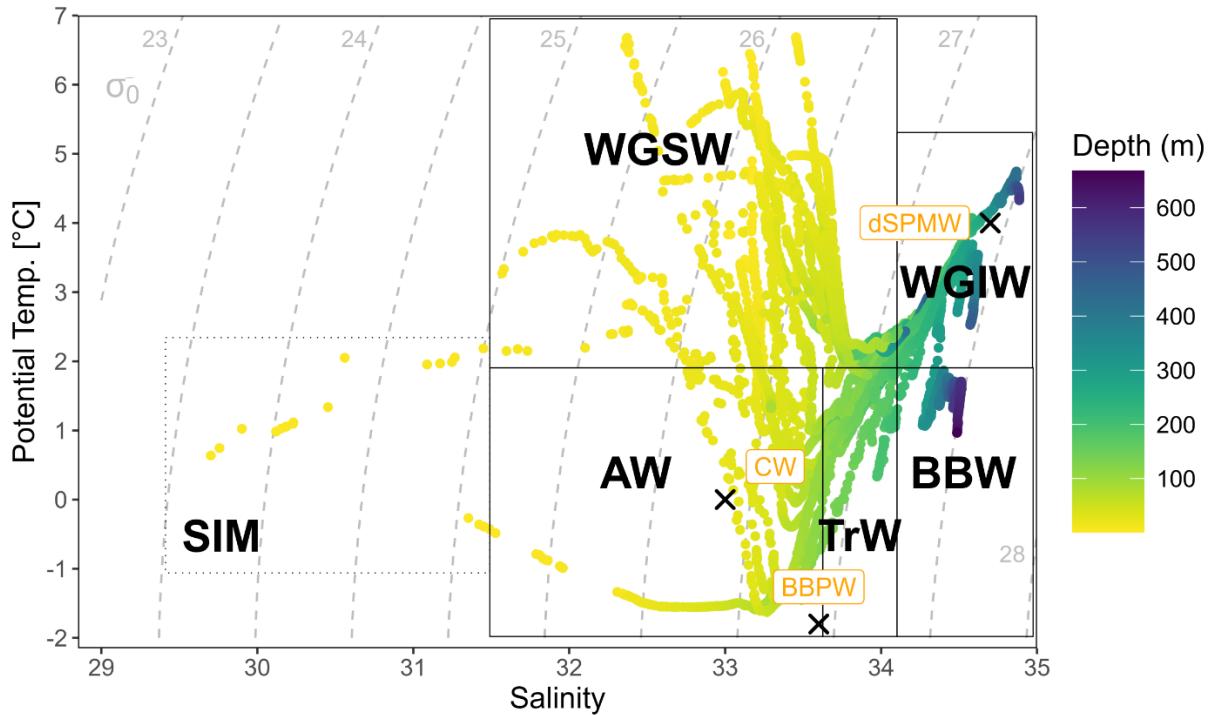
Compared to AT and CT data collected in 2015 (H. Thomas, personal communication, June 28, 2024) and 2016 (Miller et al., 2020), our results fall within a similar range. However, the minimum values observed in this study are slightly lower, likely due to the influence of sea ice meltwater, which was captured during our sampling.



310 **Figure 5** Vertical profiles of carbonate system parameters across the study area. Each data point is identified by shape (transect number) and color (station number).

3.5 Characterization of water masses

The distribution of water masses along the west coast of Greenland has been the subject of considerable discussion over the past decades, leading to the development of multiple classification systems and a wide range of nomenclatures. In this study, 315 we identify water masses based on characteristic salinity and potential temperature ranges, as illustrated in Fig. 6 (black boxes; based on Tang et al., 2004; Curry et al., 2011, 2014; Sherwood et al., 2021). This terminology is applied consistently throughout the discussion. For comparison, a more recent interpretation of water mass structure off the west Greenland coast, as proposed by Rysgaard et al. (2020), is also included in Fig. 6 (orange labels). The West Greenland Shelf Water (WGSW; $\Theta < 7^\circ\text{C}$; S < 34.1) was the dominant water mass in the study area, occurring in each transect across the shelf above a water depth of 320 200 m. The WGSW is also referred to as Southwest Greenland Coastal Water (CW), originating from the East Greenland Current and modified by freshwater input from Greenlandic glacial runoff. The West Greenland Intermediate Water (WGIW; $\Theta > 2^\circ\text{C}$; S > 34.1) was present in deeper waters along the slope below 200 m. This water mass corresponds to the Subpolar Mode Water (SPMW), of which we identified a colder and less saline variant – the deep Subpolar Mode Water (dSPMW) – along the slope. Arctic Water (AW; $\Theta \leq 2^\circ\text{C}$; S ≤ 33.7) was detected north of Davis Strait at transects 3, 4, and 5, forming an 325 intermediate layer between WGSW and WGIW above 100 m on the western side of the transects. These cold, low-salinity AW pockets can be observed in Fig. 2 h, k, and n. At transect 4, AW was found at station 17 approximately 70 km off the coast of Greenland, extending into Disko Bay. The updated classification refers to this water mass as Baffin Bay Polar Water (BBPW), of which a diluted form was found west of transects 4 and 5. The Transitional Water (TrW; $\Theta \leq 2^\circ\text{C}$; S > 33.7) was primarily 330 observed at transects 4 and 5 between 100 and 200 m, separating AW from WGIW. At station 15, the waters below 300 m were defined as Baffin Bay Water (BBW; $\Theta \leq 2$, S > 34.1), a shallower extension of the Baffin Bay Deep Water, presenting as a distinct tail on the Θ -S diagram (Sherwood et al., 2021). Due to very low surface salinity values at stations 14, 15, and 24 (S < 31.5), we introduced sea ice meltwater (SIM) as an additional source water type to distinguish areas affected by sea ice melt.



335 **Figure 6: Water masses in the study area were characterized using potential temperature and salinity: West Greenland Shelf Water (WGSW; $\Theta < 7^\circ\text{C}$; $S < 34.1$), West Greenland Intermediate Water (WGIW; $\Theta > 2^\circ\text{C}$; $S > 34.1$), Transitional Water (TrW; $\Theta \leq 2^\circ\text{C}$; $S > 33.7$), Arctic Water (AW; $\Theta \leq 2^\circ\text{C}$; $S \leq 33.7$) and Baffin Bay Water (BBW; $\Theta \leq 2^\circ\text{C}$, $S > 34.1$), according to Tang *et al.* (2004), Curry *et al.* (2011), Curry *et al.* (2014), and Sherwood *et al.* (2021). Surface water with $S < 31.5$ is indicative of sea ice meltwater (SIM). An updated perspective on the water masses was provided by Rysgaard *et al.* (2020): Baffin Bay Polar Water (BBPW; $\Theta = -1.8^\circ\text{C}$, $S = 33.6$), Southwest Greenland Coastal Water (CW; $\Theta = 0^\circ\text{C}$, $S = 33$), and deep Subpolar Mode Water (dSPMW; $\Theta = 4^\circ\text{C}$, $S = 34.7$).**

340

4 Discussion

4.1 Influence of physicochemical processes on biogeochemical properties

The extensive data set was processed using PCA to find physicochemical processes that alter the distribution of carbon, 345 macronutrients, and trace elements. The results of the PCA analysis are summarized in Table S 6, where significant PC loadings of each parameter are highlighted in bold. Overall, 82 % of the total variance in the normalized data set can be explained by three PCs. The majority of variance is explained by PC 1 with 52 %, followed by PC 2 with 18 % and PC 3 with 12 %. Salinity, depth, oxygen, AT, CT, dV, and dCu significantly load on PC 1; potential temperature, AOU, NO_x, silicate, phosphate, dNi, dCd, and dPb significantly load on PC 2; and dFe, dMn, dCo, dNi, and dCu significantly load on PC 3. In 350 Fig. 7, the results of the PCA as biplots with colors indicating the water mass affiliation of each sample are provided.

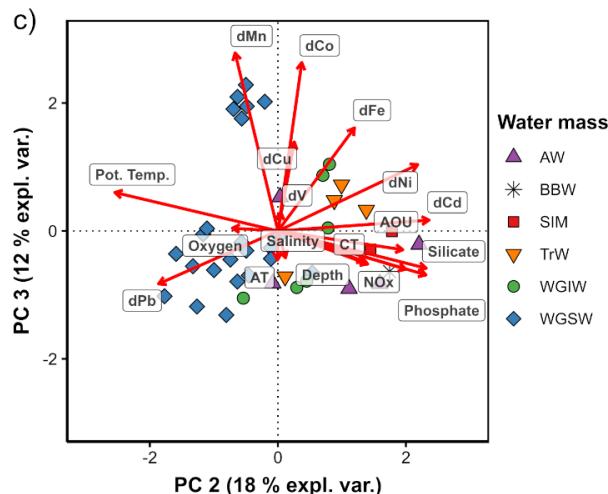
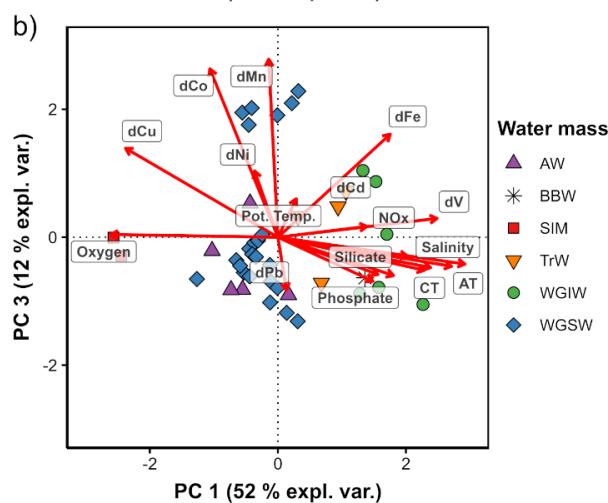
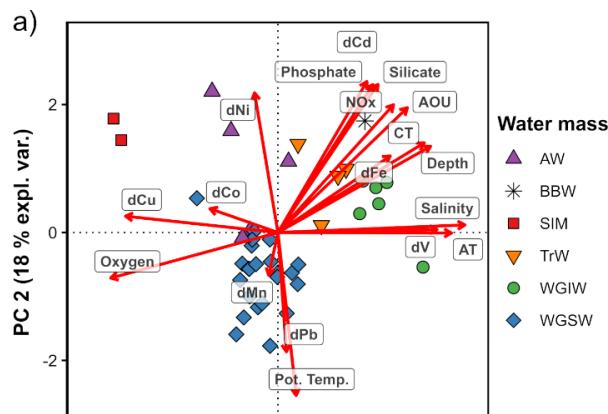


Figure 7: Biplots according to PCA results for a) PC 1 score vs PC 2 score; b) PC 1 score vs PC 3 score; and c) PC 2 score vs PC 3 score, with colors indicating the water mass affiliation of each sample. The loading of each parameter is indicated by a red arrow.

355 **4.1.1 Conservative mixing along the salinity gradient (PC 1)**

The comparison of low PC 1 scores in surface water samples with lower salinity (WGSW, AW, and SIM) and high PC 1 scores in deeper, more saline water samples (WGIW, TrW, and BBW) indicates that PC 1 resolves the salinity gradient of the study area (Fig. 7 a and b). The freshwater character of WGSW, AW, and SIM is clearly reflected in their low PC1 scores. The WGSW integrates freshwater contributions from the GIS (Foukal and Pickart, 2023), while AW represents a mixture of winter-cooled water entering Baffin Bay on the eastern side of Davis Strait and cold, fresh Arctic Ocean water of Pacific origin flowing through the CAA (Curry et al., 2014). SIM contributes to the formation of a low-salinity surface layer that remains concentrated in the upper part of the water column (Haine et al., 2015).

The statistical results illustrate the importance of freshwater input on the chemical properties of the study area. The high loadings of AT, CT, dV, and dCu on PC 1 indicate that these parameters behave conservatively, with freshwater being either a source (dCu) or a cause for dilution (AT, CT, and dV) of parameter concentrations. The positive loadings of AT and CT on PC 1 are consistent with the general understanding that freshwater inputs from sea ice and glacial melt are characterized by low AT and CT concentrations (Bates and Mathis, 2009; Chierici and Fransson, 2009; Fransson et al., 2015). The positive correlations of AT and CT against salinity are illustrated in Fig. S 5 a and b. Similarly, the distribution of dV shows a positive correlation with salinity (Fig. S 5 c), as both sea ice melt and coastal freshwater inputs are associated with low dV concentrations (Marsay et al., 2018; Whitmore et al., 2019). The distribution of dCu correlated negatively with salinity and was primarily influenced by coastal freshwater input (Fig. S 5 d). Sea ice meltwater also contributed to elevated dCu concentrations in surface waters (Fig. S 3 f), particularly toward the western ends of Transects 4 and 5. Cu-binding ligands play an important role in maintaining Cu in the dissolved phase (Ruacho et al., 2022), thereby facilitating the transport of dCu across the Arctic (Arnone et al., 2023). This is reflected in the consistent lateral decrease of dCu concentrations away from the coastal sources along the shelf transects (refer to Sect. 3.3). The influence of freshwater input on the chemical properties of the water column is further discussed in Sect. 4.3, which focuses on sea ice meltwater, and in Sect. 4.4, which addresses coastal runoff.

4.1.2 Biological uptake and recycling (PC 2)

The high positive loading of AOU, NOx, silicate, phosphate, dNi, and dCd on PC 2 suggests that PC 2 describes distributions driven by biological cycling. The nutrient-type behavior of dNi and dCd, as discussed in Sect. 3.3, aligns well with the PCA results. Additionally, dFe also shows a high loading on PC 2, reflecting its nutrient-type character. This observation is consistent with previously published data for the Nansen Basin and the Barents Sea (Gerringa et al., 2021), the Chukchi Sea (Vieira et al., 2019), the Fram Strait (Krisch et al., 2022a) and the Canadian Arctic (Colombo et al., 2020).

The anticorrelation between nutrients (positive PC 2 loading) and potential temperature (negative PC 2 loading) reflects the spatial distribution of these parameters across the study area. Low PC 2 scores of WGSW indicate that the warmer shelf waters are highly productive, resulting in low nutrient concentrations. In contrast, the colder deep waters of TrW, WGIW and BBW

exhibit high PC 2 scores, suggesting remineralization processes that return nutrients and trace elements to the water column. Similarly, AW and SIM water samples also show high PC 2 scores, characterized by higher nutrient concentrations alongside lower temperatures. The difference in nutrient concentrations between AW and WGSW is further explored in Sect. 4.2, while 390 the influence of the retreating sea ice is discussed in Sect. 4.3.

The remineralization of organic matter (OM) led to elevated concentrations of NO_x, silicate, phosphate, and dCd, along with maximum AOU values (refer to Sect. 3.1 and 3.2) in the deep waters of BBW (640 to 667 m; station 15). Although BBW was only encountered at this station, our results are consistent with Sherwood *et al.* (2021), who reported similar AOU and nutrient concentrations in BBW. The accumulation of remineralized nutrients at depth is attributed to the long residence time of deep 395 and bottom waters in Baffin Bay, combined with the region's enclosed bathymetry, which restricts water circulation (Sherwood *et al.*, 2021; Lehmann *et al.*, 2019). In addition to nutrients, CT also shows a high loading on PC 2, whereas AT loads exclusively on PC 1. This distinction highlights the different biogeochemical behavior of AT and CT. While AT is primarily influenced by freshwater dynamics, CT is also significantly affected by biological processes. Consequently, elevated CT concentrations were observed in BBW (refer to Sect. 3.4), where restricted circulation and remineralization contribute to more 400 corrosive conditions for Baffin Bay shelf sediments and benthic ecosystems. Similar to other studies in Baffin Bay (Beaupré-Laperrière *et al.*, 2020; Burgers *et al.*, 2024), OM respiration in deep waters led to undersaturation with respect to Ω Aragonite, accompanied by increased $p\text{CO}_2$ and decreased pH (refer to Sect. 3.4).

Alongside potential temperature, dPb exhibited a negative loading on PC 2, reflecting the latitudinal trend of potential temperature and dPb concentrations (refer to Sect. 3.1 and 3.3). The distribution of dPb is primarily controlled by the mixing 405 of warm Atlantic-origin waters with high dPb signatures and cold Arctic-origin waters with low dPb signatures (Colombo *et al.*, 2019). As a result, dPb concentrations were highest in the warmer waters southeast of Davis Strait and decreased progressively toward the colder northwest.

4.1.3 Scavenging and benthic processes (PC 3)

The third PC is associated with the trace elements dMn, dFe, dCo, dNi, and dCu. It is known that during estuarine mixing, 410 dCo, dNi and dCu are co-cycled with dMn and dFe through their incorporation into (oxy)hydroxide mineral phases (Smrzka *et al.*, 2019). We suggest that PC 3 indicates particle-driven mixing of these trace elements with coastal runoff as a source. High PC 3 scores (Fig. 7 b and c) of surface samples from stations 11 and 12 (mouth of Nassuttoq Fjord) as well as station 17 415 (Disko Bay) highlight the dominant influence of coastal freshwater inputs on the distribution of these trace elements. The role of coastal runoff in shaping trace element distribution is further discussed in Sect. 4.4.

415 The biplots (Fig. 7 b and c) reveal a notable feature of deep water samples (111 to 667 m), identified as TrW and WGIW. At these shelf stations, elevated concentrations of dMn, dFe, dCo, and dNi were observed in deep water samples (refer to Sect. 3.2), which we attribute to benthic processes occurring in surface sediments, i.e., reversible scavenging linked to mineral dissolution and the remineralization of organic material. Benthic release of dCo, dFe, and dMn has been documented in the Barents Sea (Gerringa *et al.*, 2021), the Chukchi Sea (Vieira *et al.*, 2019), the Canadian Arctic (Colombo *et al.*, 2020) and

420 throughout the Arctic basin (Bundy et al., 2020), while benthic dNi release has been reported in the Bering Sea and Chukchi Sea (Jensen et al., 2022). In contrast, dCu did not exhibit signs of a benthic source or evidence of biological cycling. Instead, its distribution remained dominated by freshwater runoff from coastal sources, consistent with observations by Jensen *et al.* (2022).

425 A striking feature of these deep shelf water samples is the divergence in PC3 scores: some stations (8, 10, 15, 24, 30) show negative scores, while others (17, 20, 22, 28) exhibit positive scores. We interpret this pattern as a reflection of spatial variability in return fluxes at the sediment-water interface. Stations with positive PC 3 scores are located closer to the coast, where greater export of biogenic and terrigenous material enhances recycling in deep waters. In contrast, offshore stations receive less particulate input, resulting in lower trace element concentrations and negative PC3 scores. A similar pattern was observed by Seo et al. (2022) in the East Sea, where significantly higher concentrations of dMn, dFe, and dCo were found on 430 the slopes and in the bottom layer of the Ulleung Basin compared to the Japan Basin. The authors attributed this to enhanced sedimentary release driven by the high OM content of shelf sediments (Seo et al., 2022).

4.2 Influence of major current systems on biogeochemical properties

435 The spatial distribution of parameters revealed significant differences in eastern and western shelf waters. To investigate these differences, we conducted a direct comparison of the three major water masses present along the west Greenland shelf: the northward-moving WGSW (1 to 220 m) and WGIW (130 to 560 m) with the southward-moving AW (2 to 165 m). For each parameter, the median and the mean \pm SD values are summarized in Table 2. The median is used throughout the data discussion because of its robustness against outliers.

Table 2: Comparison of the median and mean \pm SD concentrations between WGSW, WGIW and AW. The number of samples (n) used for the calculation is given in brackets.

	Parameter	WGSW (Median; Mean \pm SD)	WGIW (Median; Mean \pm SD)	AW (Median; Mean \pm SD)
Basic oceanographic parameters (WGSW: $n = 3021$; WGIW: $n = 3034$; AW: $n = 2301$)	Salinity	33.57 33.52 ± 0.42	34.45 34.45 ± 0.23	33.48 33.42 ± 0.27
	Pot. Temp. ($^{\circ}$ C)	2.68 3.20 ± 1.15	2.93 3.16 ± 0.73	0.38 0.15 ± 1.02
	Oxygen ($\mu\text{mol L}^{-1}$)	336 344 ± 23	273 278 ± 21	332 332 ± 23
	AOU ($\mu\text{mol L}^{-1}$)	-4 0 ± 26	62 55 ± 23	36 30 ± 24
Macronutrients	NOx	1.7	13.5	5.7
(WGSW: $n = 54$; WGIW: $n = 9$; AW: $n = 29$)	($\mu\text{mol L}^{-1}$)	2.4 ± 2.8	13.7 ± 1.7	6.0 ± 2.8
	Silicate	1.5	10.6	4.2
	($\mu\text{mol L}^{-1}$)	2.0 ± 1.8	11.5 ± 4.1	5.2 ± 3.5
	Phosphate	0.23	1.02	0.66
	($\mu\text{mol L}^{-1}$)	0.31 ± 0.18	0.99 ± 0.13	0.68 ± 0.21
Carbonate system parameters (WGSW: $n = 50$; WGIW: $n = 12$; AW: $n = 25$)	AT ($\mu\text{mol kg}^{-1}$)	2226 2228 ± 19	2275 2278 ± 11	2237 2237 ± 6
	CT ($\mu\text{mol kg}^{-1}$)	2051 2056 ± 28	2164 2159 ± 16	2104 2104 ± 27
	CT:AT	0.922 0.923 ± 0.007	0.950 0.948 ± 0.006	0.941 0.941 ± 0.010
	pH	8.14 8.14 ± 0.03	7.99 7.99 ± 0.04	8.10 8.09 ± 0.06
	$p\text{CO}_2$ (μatm)	292 295 ± 22	433 425 ± 37	321 332 ± 52
	Ω Aragonite	1.88 1.86 ± 0.15	1.30 1.34 ± 0.12	1.48 1.49 ± 0.20

Continuation of Table 2: Comparison of the median and mean \pm SD concentrations between WGSW, WGIW and AW. The number of samples (n) used for the calculation is given in brackets.

Parameter	WGSW (Median; Mean \pm SD)	WGIW (Median; Mean \pm SD)	AW (Median; Mean \pm SD)
Trace elements	1490	1735	1480
(WGSW: $n = 31$; WGIW: $n = 8$; AW: $n = 8$)	1486 ± 234	1713 ± 121	1478 ± 127
dV (ng L^{-1})	116	262	162
dFe (ng L^{-1})	138 ± 67	329 ± 55	174 ± 26
dMn (ng L^{-1})	237	89	186
	261 ± 71	164 ± 13	184 ± 20
dCo (ng L^{-1})	8.0	5.0	9.0
	8.5 ± 2.5	5.7 ± 0.5	9.2 ± 0.8
dNi (ng L^{-1})	254	269	266
	258 ± 51	270 ± 22	280 ± 20
dCu (ng L^{-1})	161	111	160
	158 ± 44	111 ± 13	162 ± 16
dCd (ng L^{-1})	13	31.6	21.6
	13 ± 6	32.2 ± 2.7	22.0 ± 2.3
dPb (ng L^{-1})	2.4	2.4	1.7
	2.5 ± 0.8	2.4 ± 0.5	1.8 ± 0.4

The median values of salinity and potential temperature reflect the origin of the water masses and the freshwater dynamics of the area. In the upper water column, AW is slightly fresher (33.48) and colder (0.38°C) than WGSW (33.57; 2.68°C). This east–west gradient in temperature and salinity has also been described by Tang *et al.* (2004) between Baffin Island and the Greenland coast. As the warmer WGSW flows cyclonically around Baffin Bay and mixes with adjacent waters from the CAA, it becomes progressively colder and fresher, eventually returning southward as AW (Tang *et al.*, 2004). The WGIW is part of the deep water WGSC that transports warm (2.93°C) and salty (34.45) waters from the North Atlantic (Curry *et al.*, 2014). The AOU concentrations are reflective of respiration and the biological activity of the water (Sarmiento and Gruber, 2006). As AOU is a depth-dependent variable, we examined a consistent depth range of 30 to 50 m to enable a more meaningful comparison between WGSW and AW. Within this range, the median AOU values are $-6 \mu\text{mol L}^{-1}$ for WGSW and $7 \mu\text{mol L}^{-1}$ for AW. This suggests that warmer shelf waters (WGSW) are more biologically productive than colder AW, as indicated by greater oxygen production in the upper water column. Although primary production was not directly measured in this study, this interpretation is supported by Krawczyk *et al.* (2021), who reported the highest chlorophyll *a* concentrations along the

460 southwest Greenland coast and the lowest values along Baffin Island. This pattern is consistent with overall lower nutrient concentrations observed in WGSW compared to AW (refer to Sect. 4.1.2 and Table 2), as nutrients are removed from the water column during PP. In Fig. 8 a, NO_x concentrations are plotted against longitude within the 30 to 50 m depth range, including literature data from the Green Edge cruise in 2016 (Bruyant et al., 2022). A clear spatial gradient is evident, with lower NO_x concentrations in WGSW on the shelf and higher NO_x concentrations in AW further offshore, particularly in areas that were still ice-covered. This nutrient gradient reflects the east-to-west progression of sea ice retreat and has been linked to enhanced 465 PP and species richness along the southern coast of west Greenland and in Disko Bay, compared to the more nutrient-rich but less productive regions near Baffin Island and northwest Greenland (Lafond et al., 2019; Krawczyk et al., 2021). We consider the timing of our sampling during the mid-summer season to be critical in capturing these pronounced differences in water mass composition. By this time, shelf waters had already become depleted in nutrients, whereas offshore regions were only beginning to experience increased biological processes following the retreat of sea ice (refer to Sect. 4.3). With increasing 470 depth, respiration returns nutrients to the water column, as evidenced by high AOU values in WGIW (62 $\mu\text{mol L}^{-1}$) and overall higher nutrient concentrations at depth (refer to Table 2).

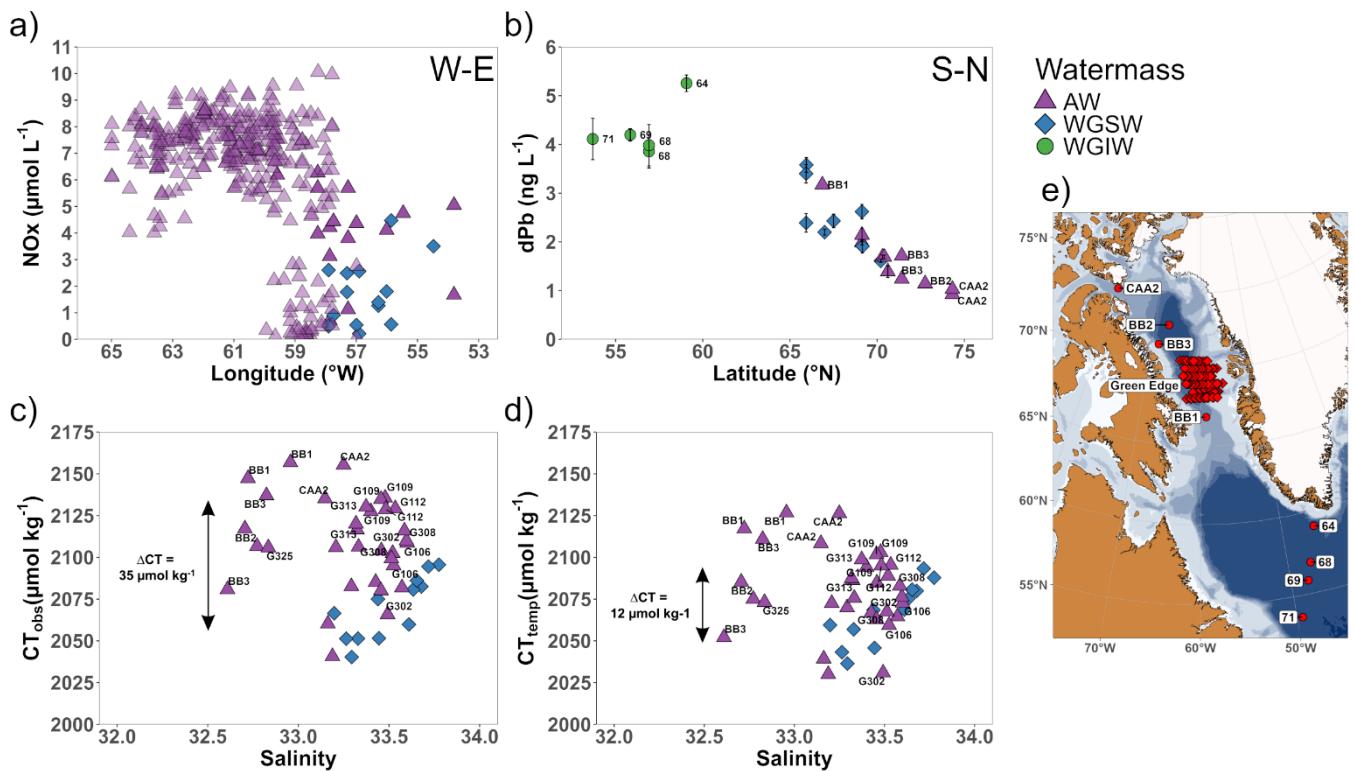


Figure 8: a) NO_x vs. longitude between 30 and 50 m with results of this study in solid colors and results of the Green Edge cruise in transparent colors (Bruyant et al., 2022); b) dPb vs. latitude between 30 and 50 m with results of this study (unlabeled) in comparison to results of the Geotraces GA01 cruise in 2014 (stations 64-71) and Geotraces GN02 in 2015 (BB1-BB3, CAA2), both: (GEOTRACES Intermediate Data Product Group, 2023); c) CT_{obs} and d) CT_{temp} vs. salinity between 30 and 50 m with results of this study (unlabeled) in comparison to results of the Green Edge cruise (Miller et al., 2020) and Geotraces GN02 in 2015 (BB1-BB3, CAA2).

CAA2) (H. Thomas, personal communication, June 28, 2024); e) location of the stations used as comparison. Different water mass categories are given in the legend.

480 A comparison of the carbonate system parameters in WGSW and AW reveals that both AT and CT median concentrations were lower in WGSW. This observation aligns with a study by Burgers *et al.* (2024), who reported that the dominant source of freshwater for WGSW in eastern Baffin Bay is glacial meltwater, characterized by a significantly lower AT endmember ($390 \pm 140 \mu\text{mol kg}^{-1}$) compared to AW in western Baffin Bay, where the AT endmember is $1170 \pm 120 \mu\text{mol kg}^{-1}$. The higher median CT:AT ratio in AW (0.941) compared to WGSW (0.922) is attributed to the CAA outflow, which carries Pacific-origin
485 water characterized by elevated CT concentrations (Azetsu-Scott *et al.*, 2010; Shadwick *et al.*, 2011b). In contrast, Atlantic-derived waters transported to eastern Baffin Bay tend to have lower CT:AT ratios (Burgers *et al.*, 2024). These slightly higher CT:AT ratios in AW are also reflected in a higher median $p\text{CO}_2$ concentration, lower median pH value and lower median Ω Aragonite concentration (refer to Table 2).

To minimize depth-related variability, we further examined CT values within a consistent depth range of 30 to 50 m. It is well
490 established that colder sea surface temperatures at high latitudes contribute significantly to the latitudinal gradient in surface CT, due to the increased CO_2 solubility of colder waters (Li and Tsui, 1971; Wu *et al.*, 2019). To assess the extent to which temperature differences between AW and WGSW influence CT concentrations, we calculated temperature-normalized CT (CT_{temp}) following the method described by Wu *et al.* (2019) (see Sect. 2.4). This normalization was based on the median potential temperature of WGSW (3.23°C , $n = 465$) within the 30 to 50 m depth range. The observed CT (CT_{obs}) and CT_{temp}
495 values for AW and WGSW are shown in Fig. 8 c and d. For comparison, we included literature data from the Green Edge cruise (Miller *et al.*, 2020) and Geotraces GN02 in 2015 (BB1-BB3, CAA2) (H. Thomas, personal communication, June 28, 2024), filtered to the same depth range of 30 to 50 m. Including literature data, the difference between the median CT values of AW and WGSW is $\Delta\text{CT}_{\text{obs}}^{\text{WGSW-AW}} = 35 \mu\text{mol kg}^{-1}$. After temperature normalization, the difference is significantly reduced to $\Delta\text{CT}_{\text{temp}}^{\text{WGSW-AW}} = 12 \mu\text{mol kg}^{-1}$. This indicates that the majority of the CT difference in AW and WGSW is driven by the
500 temperature difference between the two water masses. The remaining CT difference may be attributed to higher biological productivity in WGSW, which reduces CT concentrations, and to the mixing of AW with the CAA outflow, which is known to transport additional CT to Baffin Bay (Azetsu-Scott *et al.*, 2010; Shadwick *et al.*, 2011b; Burgers *et al.*, 2024).

In the deeper waters of WGIW, both AT and CT median concentrations increase, following the salinity gradient and reflecting the contribution of OM respiration to the release of CT to the water column. This is also evident in higher median $p\text{CO}_2$
505 concentrations and lower median Ω Aragonite concentrations in WGIW (refer to Table 2), consistent with observations by Burgers *et al.* (2024).

All median concentrations of trace elements exhibiting a nutrient-type profile (dFe, dNi, and dCd) were lower in WGSW compared to AW. This difference is attributed to biological uptake during PP, which we found to be higher in WGSW. The return of these elements to the water column through the remineralization of OM is evident in the elevated median
510 concentrations observed in WGIW (refer to Table 2). In addition to biological cycling, variations in source water composition also contribute to differences in trace element concentrations. AW exhibited elevated concentrations for dFe, dMn, dCo, and

dNi (refer to Table 2), which are likely linked to the CAA outflow into Baffin Bay. This outflow has been shown to have higher dFe and dMn concentrations due to sediment resuspension in the benthic boundary layer (Colombo et al., 2020; Colombo et al., 2021), as well as higher dNi and dCu concentrations associated with Pacific-origin waters advected from the 515 Canada Basin through the CAA (Jensen et al., 2022). In contrast to other trace elements, higher dPb median concentrations were observed in WGSW and WGIW compared to AW (refer to Table 2). As mentioned in Sect. 3.3, a latitudinal gradient in dPb concentrations exists across Davis Strait, with dPb concentrations decreasing along the west coast of Greenland. This trend is illustrated in Fig. 8 b, where dPb concentrations are plotted against latitude within the 30 to 50 m depth range. The figure combines our data with literature values from the Geotrades GA01 cruise in 2014 (stations 64 to 71) and Geotrades 520 GN02 in 2015 (BB1-BB3, CAA2) (both: GEOTRACES Intermediate Data Product Group, 2023). The elevated dPb concentrations in WGSW and WGIW are likely influenced by freshwater discharge from the GIS (Krisch et al., 2022b) and the advection of North Atlantic waters (Colombo et al., 2019), both enriched in dPb. In contrast, AW exhibited much lower dPb concentrations, which we attribute to the influence of the CAA outflow, as this region is relatively isolated from anthropogenic sources of dPb (Colombo et al., 2019). Our results demonstrate that the distribution of dPb is primarily 525 controlled by the mixing of water masses with distinct dPb signatures. As such, dPb can serve as a useful tracer for illustrating the gradual mixing and transformation of water masses along the west Greenland shelf.

4.3 Influence of retreating sea ice on biogeochemical properties

Baffin Bay and its surrounding areas are typically ice-free between July and October (Bi et al., 2019). Our statistical analysis highlighted the significant influence of SIM on the chemical composition of surface waters (refer to Sect. 4.1.1). To visualize 530 the decline in sea ice concentration during the sampling period, we used the ASI sea ice concentration product provided by the University of Bremen (Spreen et al., 2008). In Fig. 9 a, the change in sea ice concentration between 19 July and 28 July 2021 is illustrated. At the beginning of our sampling period on 19 July 2021, two extensive areas with high sea ice concentration were present between 66°N-68°N and 70°N-74°N. By 28 July 2021. These areas had disappeared almost completely, indicating a rapid and complete sea ice retreat within just 10 days. This melt event significantly affected sea surface salinity and potential 535 temperature, as shown in Fig. 9 b and c. Both parameters decreased towards the west along transects 3, 4 and 5, with salinity reaching a minimum of 29.70 (station 14; 3 m) and potential temperature of -0.36°C (station 24; 3 m). The category SIM (refer to Sect. 3.5) was identified in the upper 4 m of stations 14 and 24, and in the upper 8 m of station 15. Although not formally classified as SIM, a similar trend of reduced surface salinity and potential temperature values was observed at stations 10, 13, 23, 28, 29, and 30, adjacent to SIM stations (refer to Fig. 2). We suggest that this trend reflects the gradual advection of SIM 540 from central Baffin Bay towards the southeast.

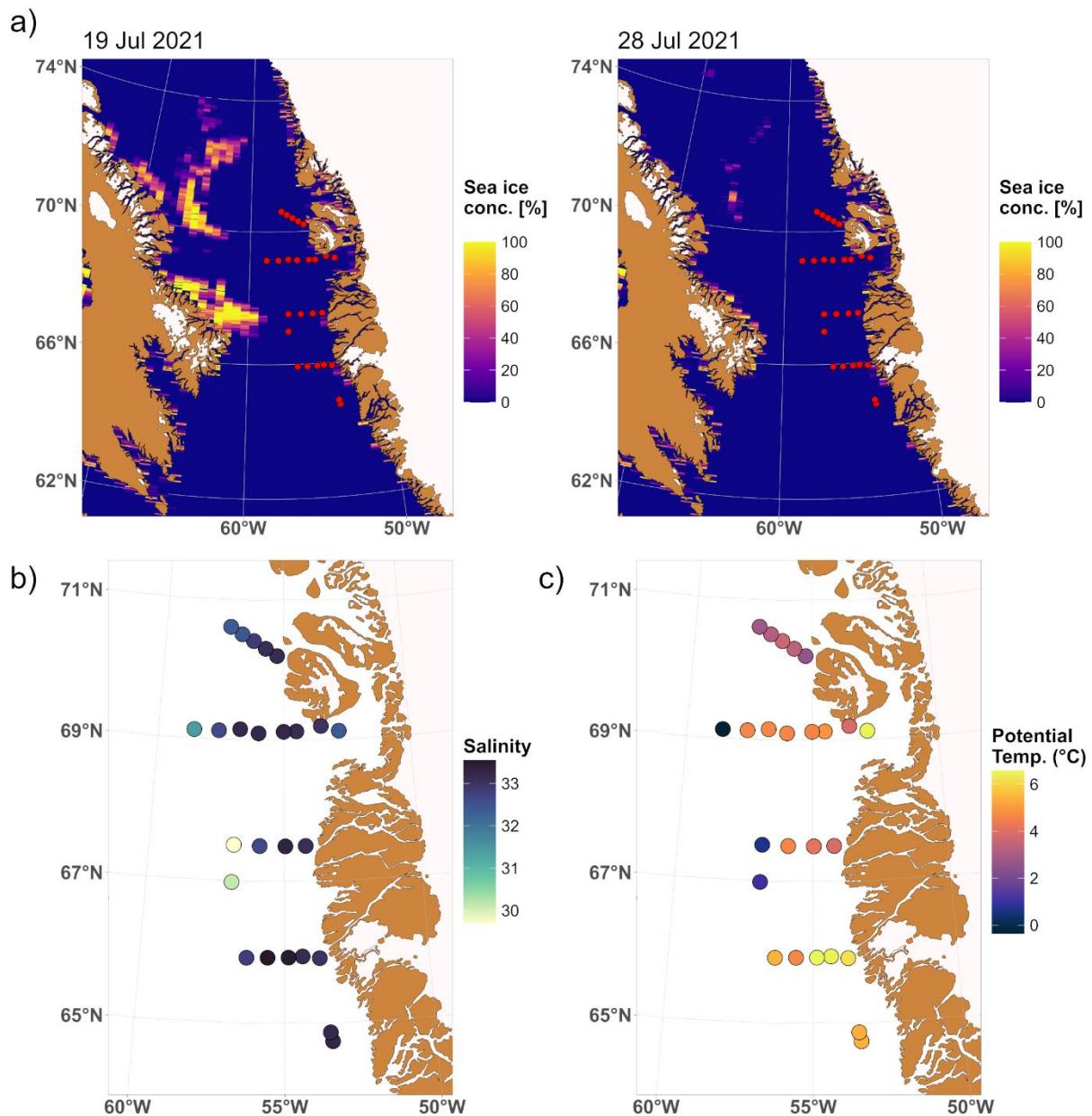


Figure 9: a) Sea ice concentration (%) in the study area during the sampling period on 19 July and 28 July 2021. Sea ice concentration data (ASI data, ASMR2, version 5.4, 3.125 km grid) was downloaded from the data archive of the University of Bremen (Spreen et al., 2008). Sea surface b) salinity and c) potential temperature from CTD sensor data.

545 In Baffin Bay, Lafond *et al.* (2019) observed elevated surface nutrient concentrations at stations covered by sea ice, in contrast to open-water stations located east of the ice edge, where nutrients were nearly depleted. In this study, the distribution of surface nutrients follows a similar trend (refer to Sect. 3.2). Along each transect, nutrient concentrations were consistently higher towards the west, coinciding with the recent retreat of sea ice. These western areas also exhibited elevated oxygen

concentrations and low AOU values (refer to Sect. 3.1), suggesting the onset of a phytoplankton bloom. The timing of the
550 study enabled us to observe a transitional phase in the seasonal cycle: nutrients in the western shelf waters had not yet been fully consumed, while waters farther east had already become depleted due to earlier biological activity.

Our observation of the carbonate system indicates that surface waters of stations influenced by SIM exhibited lower AT and CT concentrations, along with higher CT:AT ratios. These conditions resulted in higher $p\text{CO}_2$ values and lower Ω Aragonite values (refer to Sect. 3.4). Similar results of sea ice meltwater increasing $p\text{CO}_2$ and decreasing CaCO_3 mineral saturation states
555 have been reported along the east and west coast of Greenland (Henson et al., 2023), the Fram Strait (Tynan et al., 2016), and in other Arctic regions such as the Canada Basin (Bates et al., 2009). To investigate the effect of sea ice on the upper 50 m of the water column, which is the typical penetration depth of sea ice meltwater (Jones et al., 1983), we calculated salinity-normalized AT ($\text{AT}_{\text{sal}} = 35\text{AT/S}$) and temperature-salinity-normalized CT ($\text{CT}_{\text{temp}}^{\text{sal}} = 35\text{CT}_{\text{temp}}/\text{S}$), according to Wu *et al.* (2019) (refer to Sect. 2.4. and 4.2). These normalizations account for the effects of temperature and salinity on CO_2 solubility,
560 while remaining sensitive to biological processes, air-sea gas exchange, and water mass mixing (Shadwick et al., 2011a). For AT_{sal} (Fig. 10 a), most surface water concentrations ranged between 2330 and 2350 $\mu\text{mol kg}^{-1}$. We observed an AT addition of ≈ 25 to 55 $\mu\text{mol kg}^{-1}$ for SIM stations and of ≈ 5 to 45 $\mu\text{mol kg}^{-1}$ for SIM adjacent stations. These values are consistent with Jones *et al.* (1983), who reported an AT addition of $\approx 100 \mu\text{mol kg}^{-1}$ further north in Baffin Bay, where the influence of sea ice meltwater is more pronounced. This AT enrichment is attributed to the release of alkalinity during the melting of sea ice that
565 contains ikaite, as described by Rysgaard *et al.* (2011 and 2012) and Jones *et al.* (1983). The expected increase in CT by ikaite dissolution should be half the corresponding change in AT (Jones et al., 1983), or approximately 15 to 30 $\mu\text{mol kg}^{-1}$. The $\text{CT}_{\text{temp}}^{\text{sal}}$ values are shown in Fig. 10 b, with most surface water concentrations ranging between 2140 and 2170 $\mu\text{mol kg}^{-1}$. At SIM stations, we observed an increase in $\text{CT}_{\text{temp}}^{\text{sal}}$ of ≈ 60 to 75 $\mu\text{mol kg}^{-1}$, and at SIM adjacent stations of ≈ 10 to 60 $\mu\text{mol kg}^{-1}$. Hence, the observed CT addition exceeds the expected contribution from ikaite dissolution alone. Possible explanations for
570 the observed surplus in CT include the release of CT from organic carbon remineralization under the ice (Bates et al., 2009; Tynan et al., 2016), or within the ice itself, as sea ice is known to contain higher concentrations in OM than the underlying seawater (Vancoppenolle et al., 2013). Additionally, the inflow of CT-enriched waters transported through the CAA into Baffin Bay may contribute to the elevated CT levels (Azetsu-Scott et al., 2010; Shadwick et al., 2011b; Henson et al., 2023). Our results indicate that the dissolution of ikaite in sea ice acted as a source of AT, providing a small geochemical buffer to surface
575 waters (Jones et al., 2021). However, the inflow of Pacific-origin waters and remineralization under and within the ice contributed to elevated CT:AT ratios, resulting in higher $p\text{CO}_2$ and lower pH and Ω Aragonite values in surface waters. There is evidence that once the sea ice melt facilitated bloom fully develops and CT is removed from seawater by PP, higher pH and Ω Aragonite values can be expected along the sea ice edge (Tynan et al., 2016). This process is reflected in the photic zone of station 15 at 32 m, where an oxygen maximum and AOU minimum indicate high PP, resulting in a pH maximum (8.20) and
580 $p\text{CO}_2$ minimum (249 μatm) (refer to Sect. 3.4).

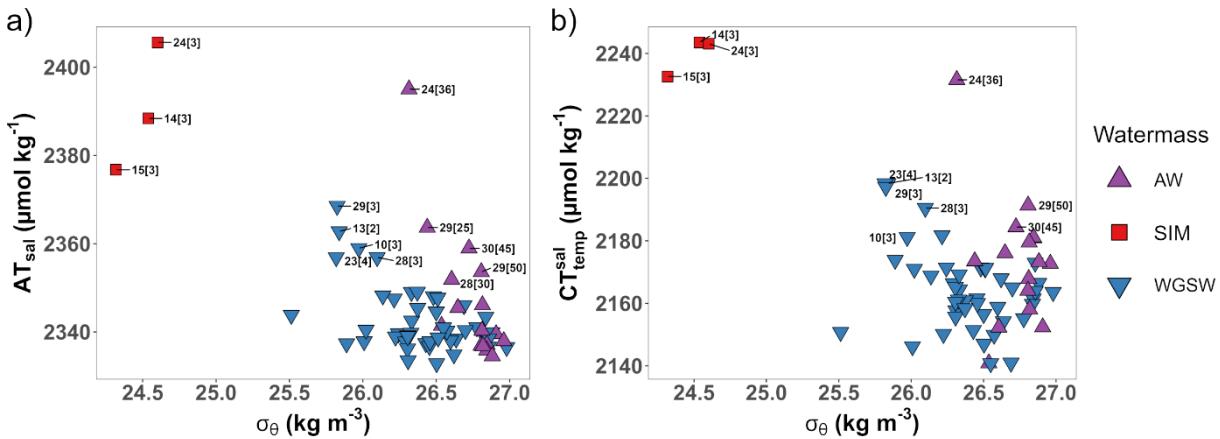


Figure 10: First 50 m of the water column for a) salinity-normalized AT values against pot. density and b) temperature-salinity-normalized CT values against pot. density. Different water mass categories are given in the legend. SIM affected stations (14, 15, 24) and stations adjacent to those (10, 13, 23, 28, 29, 30) are labeled. The number in brackets corresponds to the sampling depth.

We observed elevated surface water concentrations of dFe, dCo, dNi, dCu, and dCd in areas recently covered by sea ice, particularly towards the west of each transect (refer to Sect. 3.3). This observation aligns with a study by Tovar-Sánchez *et al.*

(2010), who observed an enrichment of these trace elements in sea ice relative to surface waters, suggesting that sea ice meltwater is a significant source of these trace elements to surface waters in Baffin Bay. In particular, the additional input of

dFe from melting sea ice may play a biologically important role in maintaining ice-edge blooms, as previously observed, e.g., in the Bering Sea (Aguilar-Islas *et al.*, 2008). In addition to sea ice input, we suggest that, similar to macronutrients, trace elements had not yet been removed from surface waters by biological uptake at the time of sampling. We hypothesize that

following the development of a bloom, concentrations of both macronutrients and trace elements would decline, as already observed in surface waters at shelf stations closer to the Greenlandic coast. Consistent with findings from other Arctic regions

(Nakanowatari *et al.*, 2007; Kanna *et al.*, 2014), the progressive melting and retreat of sea ice in Baffin Bay and Davis Strait

appears to significantly influence the biogeochemical cycling of carbon and trace elements, as well as the biological productivity of the upper water column.

4.4 Influence of freshwater runoff from coastal sources on biogeochemical properties

The influence of coastal freshwater runoff on the chemical composition of surface waters was demonstrated by statistical analysis (refer to Sect. 4.1.3) and is evident in minimum salinity values observed at stations 6, 11, and 17 (refer to Sect. 3.1).

Stations 6 and 11 are located near to the mouths of the Kangerlussuaq Fjord and the Nassuttooq Fjord, both of which receive glacial and riverine freshwater from the GIS (Monteban *et al.*, 2020). Station 17 is located in Disko Bay, approximately 70 km

from the mouth of the Ilulissat Icefjord/Jakobshavn Isbræ system, where glacially modified waters form a buoyant coastal current outside the fjord (Beaird *et al.*, 2017). These low-salinity surface waters coincide with elevated macronutrient concentrations (refer to Sect. 3.2). A modeling study by Møller *et al.* (2023) demonstrated that a portion of the macronutrient input into Disko Bay is directly linked to freshwater runoff. Additional mechanisms that transport marine-sourced nutrients

from deeper waters into the photic zone include vertical mixing fueled by wind and tide (Møller et al., 2023), as well as estuarine upwelling circulation forced by the glacier's freshwater input (Williams et al., 2021). Our results suggest that the continued replenishment of macronutrients into the photic zone of Disko Bay can sustain PP into the summer season. This is
610 evidenced by high oxygen and low AOU concentrations in surface waters at station 17 (refer to Sect. 3.1). These conditions have significant implications for the carbonate system and the uptake of CO₂, as reflected by low Revelle factors and low *p*CO₂ values in surface waters of Disko Bay (refer to Sect. 3.4). A similar pattern was observed in the Godthåbsfjord system in southern Greenland, where biological processes were identified as the primary drivers of CO₂ uptake, resulting in low surface water *p*CO₂ (Meire et al., 2015). Based on these findings and supporting literature, we propose that nutrient cycling in Disko
615 Bay is strongly influenced by GIS-derived freshwater input. This input stimulates PP and establishes Disko Bay as an important seasonal sink for atmospheric CO₂ well into the summer.

Apart from the Disko Bay area, surface waters at stations closest to the Greenland coast exhibited lower Ω Aragonite values with lower buffering capacities (refer to Sect. 3.4). Glacial freshwater discharge from the GIS is known to have a strong dilution effect on AT relative to CT, which reduces the buffering capacity of surface waters, resulting in lower Ω Aragonite
620 saturation states and pH values (Henson et al., 2023; Burgers et al., 2024). However, surface water acidification can be mitigated by PP, which removes CT from the photic zone, consequently increasing both pH and Ω Aragonite (Chierici and Fransson, 2009), as observed in Disko Bay. In stations along the coast, surface waters exhibited negative AOU values, indicating CT drawdown via PP. Simultaneously, extremely low NO_x values (< LOQ) were detected, suggesting that PP was approaching nutrient limitation. This nutrient depletion likely constrained further CT uptake, thereby no longer compensating
625 the AT dilution effect caused by the GIS freshwater input. As a result, surface waters exhibited lower pH values coupled with higher Revelle factors and *p*CO₂ values. Our findings suggest that as the summer season progresses and PP declines, surface waters along the west Greenland coast become increasingly vulnerable to acidification due to the input of poorly buffered glacial freshwater. These observations are consistent with Henson *et al.* (2023), who observed that AT dilution from glacial meltwater drives corrosive conditions in Greenlandic coastal surface waters, leading to a general decline in both Ω Aragonite
630 and pH (refer to Sect. 3.4).

In addition to influencing the cycling of macronutrients and carbon, coastal freshwater input significantly affects the distribution of trace elements. Surface water concentrations of dMn, dFe, dCo, dNi, and dCu were generally highest close to the coast and decreased with increasing distance along each transect (refer to Sect. 4.2.3). This trend was especially pronounced at stations 11 and 12 (mouth of Nassuttooq Fjord), and station 17 (Disko Bay, Ilulissat Icefjord). While surface water
635 concentrations of dNi in Disko Bay remained relatively constant, elevated concentrations were observed at the mouth of Nassuttooq Fjord. Similar spatial variability in dNi has been reported by Krause *et al.* (2021), who related these trends to the presence of outcrops of Ni-rich minerals along the Greenland coast. Overall, our work is consistent with previous studies from various regions around Greenland, which identified GIS-derived freshwater as a source of trace elements including dMn, dFe, dCo, dNi, and dCu (Colombo et al., 2020; Hawkins et al., 2020; Krause et al., 2021; Chen et al., 2022; Krisch et al., 2022a;).
640 The input of bioactive trace elements, e.g., dMn, dFe, and dCo, via GIS freshwater runoff has been suggested to enhance

primary productivity in adjacent shelf areas (Bhatia et al., 2013; Hawkings et al., 2020). Our study provides further evidence that freshwater from the GIS may support biological processes by supplying bioactive trace elements to surface waters.

5 Summary and Conclusion

The distribution of macronutrients, carbonate system species, and dissolved trace elements on the west Greenland shelf is highly dynamic and influenced by the complex interplay between physicochemical drivers, major ocean currents, sea ice melting, and terrestrial freshwater runoff from the GIS.

The biogeochemical cycling of these parameters is governed by several key mechanisms. First, conservative mixing along the salinity gradient distinguishes fresher surface waters (WGSW, AW, SIM) from more saline deep waters (WGIW, TrW, BBW). While AT, CT, and dV show positive correlations with salinity due to dilution by freshwater, dCu shows a negative correlation, indicating coastal and sea ice meltwater as sources. Second, biological processes regulate the vertical fluxes of macronutrients, CT, and trace metals such as dFe, dNi, and dCd by photosynthetic uptake and OM remineralization. In the deep waters of Baffin Bay, the long residence time and the enclosed bathymetry promote the accumulation of remineralized nutrients and CT. This accumulation drives corrosive conditions, resulting in \mathcal{Q} Aragonite undersaturation in deep waters of southern Baffin Bay. Such conditions pose a potential threat to the benthic ecosystem, particularly by creating an unfavorable environment for shell-forming marine organisms. Third, benthic fluxes across the sediment-water interface contribute to elevated concentrations of certain trace elements (e.g., dMn, dFe, dCo, and dNi) in bottom waters of shelf stations. We hypothesize that the magnitude of this return flux decreases with increasing distance from the Greenlandic coast, likely due to reduced deposition of biogenic and terrigenous material available for remineralization and reversible scavenging. Further investigation of Baffin Bay sediments is necessary to quantify benthic fluxes of trace elements and geochemical reactions occurring within the sediment. Our study provides an initial assessment of the role of benthic inputs in trace element cycling on the west Greenland shelf and Baffin Bay, highlighting their potential significance in shaping regional biogeochemistry.

The distribution of chemical constituents in the water column across the west Greenland shelf is primarily influenced by the opposing directions of the BIC and the WGC and their respective water masses. The BIC transports the southward-moving AW, which combines winter-cooled water from Baffin Bay with Pacific-origin Arctic Ocean water entering through the CAA.

We characterized this water mass as cold and fresh, with elevated nutrient concentrations and high CT:AT ratios, reflecting both the lower biological activity and the additional CT input from the CAA. Elevated concentrations of dFe, dMn, dCo, dNi, and dCu were observed in AW, likely due to the influence of trace element-rich outflows from the CAA into Baffin Bay. In contrast, the WGSW, transported northward by the WGC, integrates freshwater of Arctic-origin from East Greenland with glacial meltwater from the GIS. This water mass dominated the upper water column on the shelf and is characterized as warm and fresh, with high biological productivity and low nutrient concentrations. Beneath the WGSW lies the WGIW, a warm and salty water mass of North Atlantic origin. This water mass is enriched in nutrients and carbon due to the remineralization of

OM. The gradual mixing of these three water masses along the shelf was further illustrated by the latitudinal distribution of dPb, which is influenced by elevated dPb waters from the North Atlantic and low dPb waters from the CAA.

Our study provides evidence that the progressive melting and retreat of sea ice alters the biological productivity and the
675 biogeochemical cycling of carbon and trace elements in surface waters of southern Baffin Bay. The east-to-west retreat of sea ice during spring and summer created a significant gradient in nutrient concentrations. Nutrients were lowest in the highly productive shelf waters along the eastern side of the west Greenland coast, while higher concentrations were observed towards the west of southern Baffin Bay, where sea ice cover persisted for a longer duration. The timing of this study enabled us to capture the onset of a phytoplankton bloom in areas previously covered by sea ice, where nutrients had not yet been depleted.

680 Additionally, sea ice meltwater provided additional AT to surface waters, likely through the dissolution of ikaite. However, this small geochemical buffer was offset by elevated CT concentrations, potentially derived from the remineralization of OM under and within the ice and the inflow of CT-rich Pacific-origin waters via the CAA to Baffin Bay. Overall, the combined influence of sea ice meltwater and the inflow of Pacific-origin waters reduced AT relative to CT, resulting in elevated $p\text{CO}_2$ and decreased pH and Ω Aragonite values in surface waters. As the ice-edge bloom progresses and CT is removed by PP, an
685 increase in pH and Ω Aragonite values can be expected along the retreating ice margin. Furthermore, we identified sea ice meltwater as a source of bioactive trace elements, including dFe, dCo, dNi, dCu, and dCd to surface waters of southern Baffin Bay. This additional input of micronutrients may sustain and prolong the duration of ice-edge blooms, with implications for regional productivity and biogeochemical cycling.

Freshwater runoff from the GIS significantly influenced the chemical composition of coastal waters along west Greenland,
690 particularly in Disko Bay and at the mouth of the Nassuttooq Fjord. In the Disko Bay area, GIS-derived freshwater input replenished macronutrients in the photic zone, stimulating PP and creating an important sink for CO_2 long into the summer season. The supply of bioactive trace elements via GIS freshwater runoff, could further support biological processes in the surrounding shelf areas. However, in coastal areas where PP approached nutrient limitation, surface waters became more susceptible to acidification due to the input of poorly buffered glacial freshwater.

695 This work successfully captured a high-resolution, large-scale snapshot of various water column parameters across the west Greenland shelf during July. Nevertheless, we were unable to resolve seasonal variability, which requires observations at higher temporal resolution. We emphasize the need for sustained time-series observations to fully assess the consequences of climate change in this climate-sensitive region.

700 **Data availability statement**

The data set for the carbonate system and dissolved trace elements can be found online at: <https://doi.org/10.5281/zenodo.14235091> (Schmidt et al., 2024).

The data set for the nutrients can be found online at: <https://doi.org/10.5281/zenodo.15590652> (Stedmon et al., 2025).

Supplement link

705 The supplements to this article can be found online at: XXX

Author contribution

CES: Conceptualization, Formal Analysis, Investigation, Methodology, Data curation, Visualization, Writing – original draft

TZ: Writing – review & editing

KK: Investigation, Writing – review & editing

710 DP: Resources, Supervision, Writing – review & editing

HT: Funding acquisition, Project administration, Resources, Writing – review & editing

Declaration of competing interest

The authors declare that they have no conflict of interest.

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References

Aguilar-Islas, A. M., Rember, R. D., Mordy, C. W., and Wu, J.: Sea ice-derived dissolved iron and its potential influence on
725 the spring algal bloom in the Bering Sea, *Geophysical Research Letters*, 35, doi:10.1029/2008GL035736, 2008.

Akima, H., and Gebhardt, A.: *akima: Interpolation of Irregularly and Regularly Spaced Data*, CRAN [code], <https://CRAN.R-project.org/package=akima>, 2022.

Aksenov, Y., Karcher, M., Proshutinsky, A., Gerdes, R., de Cuevas, B., Golubeva, E., Kauker, F., Nguyen, A. T., Platov, G.
A., Wadley, M., Watanabe, E., Coward, A. C., and Nurser, A. J.: Arctic pathways of Pacific Water: Arctic Ocean Model
730 Intercomparison experiments, *Journal of geophysical research: Oceans*, 121, 27-59, doi:10.1002/2015jc011299, 2016.

Arnone, V., Santana-Casiano, J. M., González-Dávila, M., Planquette, H., Sarthou, G., Gerringa, L. J. A., and González, A.
G.: Natural copper-binding ligands in the Arctic Ocean. The influence of the Transpolar Drift (GEOTRACES GN04), *Frontiers in Marine Science*, 10, doi:10.3389/fmars.2023.1306278, 2023.

Azetsu-Scott, K., Clarke, A., Falkner, K., Hamilton, J., Jones, E. P., Lee, C., Petrie, B., Prinsenberg, S., Starr, M., and Yeats,
735 P.: Calcium carbonate saturation states in the waters of the Canadian Arctic Archipelago and the Labrador Sea, *Journal of Geophysical Research: Oceans*, 115, doi:10.1029/2009JC005917, 2010.

Ballinger, T. J., Moore, G. W. K., Garcia-Quintana, Y., Myers, P. G., Imrit, A. A., Topál, D., and Meier, W. N.: Abrupt Northern Baffin Bay Autumn Warming and Sea-Ice Loss Since the Turn of the Twenty-First Century, *Geophysical Research Letters*, 49, e2022GL101472, doi:10.1029/2022GL101472, 2022.

740 Bates, N. R., and Mathis, J. T.: The Arctic Ocean marine carbon cycle: evaluation of air-sea CO₂ exchanges, ocean acidification impacts and potential feedbacks, *Biogeosciences*, 6, 2433-2459, doi:10.5194/bg-6-2433-2009, 2009.

Bates, N. R., Mathis, J. T., and Cooper, L. W.: Ocean acidification and biologically induced seasonality of carbonate mineral saturation states in the western Arctic Ocean, *Journal of Geophysical Research: Oceans*, 114, doi:10.1029/2008JC004862, 2009.

745 Beaird, N., Straneo, F., and Jenkins, W.: Characteristics of meltwater export from Jakobshavn Isbræ and Ilulissat Icefjord, *Annals of Glaciology*, 58, 107-117, doi:10.1017/aog.2017.19, 2017.

Beaupré-Laperrière, A., Mucci, A., and Thomas, H.: The recent state and variability of the carbonate system of the Canadian Arctic Archipelago and adjacent basins in the context of ocean acidification, *Biogeosciences*, 17, 3923-3942, doi:10.5194/bg-17-3923-2020, 2020.

750 Bhatia, M. P., Kujawinski, E. B., Das, S. B., Breier, C. F., Henderson, P. B., and Charette, M. A.: Greenland meltwater as a significant and potentially bioavailable source of iron to the ocean, *Nature Geoscience*, 6, 274-278, doi:10.1038/ngeo1746, 2013.

Bi, H., Zhang, Z., Wang, Y., Xu, X., Liang, Y., Huang, J., Liu, Y., and Fu, M.: Baffin Bay sea ice inflow and outflow: 1978–1979 to 2016–2017, *The Cryosphere*, 13, 1025-1042, doi:10.5194/tc-13-1025-2019, 2019.

755 Bro, R., and Smilde, A. K.: Principal component analysis, *Analytical Methods*, 6, 2812-2831, doi:10.1039/C3AY41907J, 2014.

Bruyant, F., Amiraux, R., Amyot, M.-P., Archambault, P., Artigue, L., Barbedo de Freitas, L., Béchu, G., Bélanger, S., Bourgoin, P., Bricaud, A., Brouard, E., Brunet, C., Burgers, T., Caleb, D., Chalut, K., Claustre, H., Cornet-Barthaux, V., Coupel, P., Cusa, M., Cusset, F., Dadaglio, L., Davelaar, M., Deslongchamps, G., Dimier, C., Dinasquet, J., Dumont, D., Else, B., Eulaers, I., Ferland, J., Filteau, G., Forget, M.-H., Fort, J., Fortier, L., Galí-Tapías, M., Gallinari, M., Garbus, S.-E., Garcia, 760 N., Gérikas Ribeiro, C., Gombault, C., Gourvil, P., Goyens, C., Grant, C., Grondin, P.-L., Guillot, P., Hillion, S., Hussherr, R., Joux, F., Joy-Warren, H., Joyal, G., Kieber, D., Lafond, A., Lagunas, J., Lajeunesse, P., Lalande, C., Larivière, J., Le Gall, F., Leblanc, K., Leblanc, M., Legras, J., Levesque, K., Lewis, K.-M., Leymarie, E., Leynaert, A., Linkowski, T., Lizotte, M., Lopes dos Santos, A., Marec, C., Marie, D., Massé, G., Massicotte, P., Matsuoka, A., Miller, L., Mirshak, S., Morata, N., Moriceau, B., Morin, P.-I., Morisset, S., Mosbech, A., Mucci, A., Nadaï, G., Nozais, C., Obernosterer, I., Paire, T., 765 Panagiotopoulos, C., Parenteau, M., Pelletier, N., Picheral, M., Quéguiner, B., Raimbault, P., Ras, J., Rehm, E., Ribot Lacosta, L., Rontani, J.-F., Saint-Béat, B., Sansoulet, J., Sardet, N., Schmechtig, C., Sciandra, A., Sempéré, R., Sévigny, C., Toullec, J., Tragin, M., Tremblay, J.-E., Trottier, A.-P., Vaulot, D., Vladoiu, A., Xue, L., Yunda-Guarin, G., and Babin, M.: The Green Edge cruise: following the evolution of the Arctic phytoplankton spring bloom, from ice-covered to open waters, SEANOE [data set], doi:10.17882/86417, 2022.

770 Bundy, R. M., Tagliabue, A., Hawco, N. J., Morton, P. L., Twining, B. S., Hatta, M., Noble, A. E., Cape, M. R., John, S. G., Cullen, J. T., and Saito, M. A.: Elevated sources of cobalt in the Arctic Ocean, *Biogeosciences*, 17, 4745-4767, doi:10.5194/bg-17-4745-2020, 2020.

Burgers, T. M., Azetsu-Scott, K., Myers, P. G., Else, B. G. T., Miller, L. A., Rysgaard, S., Chan, W., Tremblay, J.-É., and 775 Papakyriakou, T.: Unraveling the Biogeochemical Drivers of Aragonite Saturation State in Baffin Bay: Insights From the West Greenland Continental Shelf, *Journal of Geophysical Research: Oceans*, 129, e2024JC021122, doi:10.1029/2024JC021122, 2024.

Cape, M. R., Straneo, F., Baird, N., Bundy, R. M., and Charette, M. A.: Nutrient release to oceans from buoyancy-driven upwelling at Greenland tidewater glaciers, *Nature Geoscience*, 12, 34-39, 10.1038/s41561-018-0268-4, 2019.

780 Chen, X.-G., Krisch, S., Al-Hashem, A., Hopwood, M. J., Rutgers van der Loeff, M. M., Huhn, O., Lodeiro, P., Steffens, T., and Achterberg, E. P.: Dissolved, Labile, and Total Particulate Trace Metal Dynamics on the Northeast Greenland Shelf, *Global Biogeochemical Cycles*, 36, e2022GB007528, doi:10.1029/2022GB007528, 2022.

Chierici, M., and Fransson, A.: Calcium carbonate saturation in the surface water of the Arctic Ocean: undersaturation in freshwater influenced shelves, *Biogeosciences*, 6, 2421-2431, doi:10.5194/bg-6-2421-2009, 2009.

785 Colombo, M., Rogalla, B., Myers, P. G., Allen, S. E., and Orians, K. J.: Tracing Dissolved Lead Sources in the Canadian Arctic: Insights from the Canadian GEOTRACES Program, *ACS Earth and Space Chemistry*, 3, 1302-1314, doi:10.1021/acsearthspacechem.9b00083, 2019.

Colombo, M., Jackson, S. L., Cullen, J. T., and Orians, K. J.: Dissolved iron and manganese in the Canadian Arctic Ocean: On the biogeochemical processes controlling their distributions, *Geochimica et Cosmochimica Acta*, 277, 150-174, doi:10.1016/j.gca.2020.03.012, 2020.

790 Colombo, M., Rogalla, B., Li, J., Allen, S. E., Orians, K. J., and Maldonado, M. T.: Canadian Arctic Archipelago Shelf-Ocean Interactions: A Major Iron Source to Pacific Derived Waters Transiting to the Atlantic, *Global Biogeochemical Cycles*, 35, e2021GB007058, doi:10.1029/2021GB007058, 2021.

Curry, B., Lee, C. M., and Petrie, B.: Volume, Freshwater, and Heat Fluxes through Davis Strait, 2004–05, *Journal of Physical Oceanography*, 41, 429–436, doi:10.1175/2010JPO4536.1, 2011.

795 Curry, B., Lee, C. M., Petrie, B., Moritz, R. E., and Kwok, R.: Multiyear Volume, Liquid Freshwater, and Sea Ice Transports through Davis Strait, 2004–10, *Journal of Physical Oceanography*, 44, 1244–1266, doi:10.1175/JPO-D-13-0177.1, 2014.

Dickson, A. G., and Millero, F. J.: A comparison of the equilibrium constants for the dissociation of carbonic acid in seawater media, *Deep Sea Research Part A. Oceanographic Research Papers*, 34, 1733–1743, doi:10.1016/0198-0149(87)90021-5, 1987.

Dickson, A. G.: Standard potential of the reaction: $\text{AgCl}(s) + 12\text{H}_2(g) = \text{Ag}(s) + \text{HCl}(aq)$, and the standard acidity constant 800 of the ion HSO_4^- in synthetic sea water from 273.15 to 318.15 K, *The Journal of Chemical Thermodynamics*, 22, 113–127, doi:10.1016/0021-9614(90)90074-Z, 1990.

DIN e.V.: DIN 32645: Chemical analysis—Decision limit, detection limit and determination limit under repeatability conditions—Terms, methods, evaluation, doi:10.31030/1465413, 2008.

Ebeling, A., Zimmermann, T., Klein, O., Irreger, J., and Pröfrock, D.: Analysis of Seventeen Certified Water Reference 805 Materials for Trace and Technology-Critical Elements, *Geostandards and Geoanalytical Research*, 46, 351–378, doi:10.1111/ggr.12422, 2022.

Evans, L. K., and Nishioka, J.: Accumulation processes of trace metals into Arctic sea ice: distribution of Fe, Mn and Cd associated with ice structure, *Marine Chemistry*, 209, 36–47, doi:10.1016/j.marchem.2018.11.011, 2019.

Finley, A., Banerjee, S., and Hjelle, Ø.: MBA: Multilevel B-Spline Approximation, CRAN [code], <https://CRAN.R-project.org/package=MBA>, 2022.

Foukal, N. P., and Pickart, R. S.: Moored Observations of the West Greenland Coastal Current along the Southwest Greenland Shelf, *Journal of Physical Oceanography*, 53, 2619–2632, doi:10.1175/JPO-D-23-0104.1, 2023.

Fransson, A., Chierici, M., Miller, L. A., Carnat, G., Shadwick, E., Thomas, H., Pineault, S., and Papakyriakou, T. N.: Impact 815 of sea-ice processes on the carbonate system and ocean acidification at the ice-water interface of the Amundsen Gulf, Arctic Ocean, *Journal of Geophysical Research: Oceans*, 118, 7001–7023, doi:10.1002/2013JC009164, 2013.

Fransson, A., Chierici, M., Nomura, D., Granskog, M. A., Kristiansen, S., Martma, T., and Nehrke, G.: Effect of glacial drainage water on the CO₂ system and ocean acidification state in an Arctic tidewater-glacier fjord during two contrasting years, *Journal of Geophysical Research: Oceans*, 120, 2413–2429, doi:10.1002/2014JC010320, 2015.

Fransson, A., Chierici, M., Granskog, M. A., Dodd, P. A., and Stedmon, C. A.: Impacts of glacial and sea-ice meltwater, 820 primary production, and ocean CO₂ uptake on ocean acidification state of waters by the 79 North Glacier and northeast Greenland shelf, *Frontiers in Marine Science*, Volume 10 - 2023, doi:10.3389/fmars.2023.1155126, 2023.

GEBCO Bathymetric Compilation Group 2023: The GEBCO_2023 Grid - a continuous terrain model of the global oceans and land, NERC EDS British Oceanographic Data Centre NOC [data set], doi:10.5285/f98b053b-0cbc-6c23-e053-6c86abc0af7b, 2023.

825 GEOTRACES Intermediate Data Product Group: The GEOTRACES Intermediate Data Product 2021 version 2 (IDP2021v2), NERC EDS British Oceanographic Data Centre NOC [data set], doi:10.5285/ff46f034-f47c-05f9-e053-6c86abc0dc7e, 2023.

Gerringa, L. J. A., Rijkenberg, M. J. A., Slagter, H. A., Laan, P., Paffrath, R., Bauch, D., Rutgers van der Loeff, M., and Middag, R.: Dissolved Cd, Co, Cu, Fe, Mn, Ni, and Zn in the Arctic Ocean, *Journal of Geophysical Research: Oceans*, 126, e2021JC017323, doi:10.1029/2021JC017323, 2021.

830 Haine, T. W. N., Curry, B., Gerdes, R., Hansen, E., Karcher, M., Lee, C., Rudels, B., Spreen, G., de Steur, L., Stewart, K. D., and Woodgate, R.: Arctic freshwater export: Status, mechanisms, and prospects, *Global and Planetary Change*, 125, 13-35, doi:10.1016/j.gloplacha.2014.11.013, 2015.

Hansen, H. P., and Koroleff, F.: Determination of nutrients, in: *Methods of Seawater Analysis*, 159-228, doi:10.1002/9783527613984.ch10, 1999.

835 Hawkins, J., Wadham, J., Tranter, M., Telling, J., Bagshaw, E., Beaton, A., Simmons, S.-L., Chandler, D., Tedstone, A., and Nienow, P.: The Greenland Ice Sheet as a hot spot of phosphorus weathering and export in the Arctic, *Global Biogeochemical Cycles*, 30, 191-210, doi:10.1002/2015GB005237, 2016.

Hawkins, J. R., Wadham, J. L., Tranter, M., Lawson, E., Sole, A., Cowton, T., Tedstone, A. J., Bartholomew, I., Nienow, P., Chandler, D., and Telling, J.: The effect of warming climate on nutrient and solute export from the Greenland Ice Sheet,

840 *Geochemical Perspectives Letters*, 1, 94-104, doi:10.7185/geochemlet.1510, 2015.

Hawkins, J. R., Wadham, J., Benning, L., Hendry, K., Tranter, M., Tedstone, A., Nienow, P., and Raiswell, R.: Ice sheets as a missing source of silica to the polar oceans, *Nature Communications*, 8, 14198, doi:10.1038/ncomms14198, 2017.

Hawkins, J. R., Skidmore, M. L., Wadham, J. L., Priscu, J. C., Morton, P. L., Hatton, J. E., Gardner, C. B., Kohler, T. J., Stibal, M., Bagshaw, E. A., Steigmeyer, A., Barker, J., Dore, J. E., Lyons, W. B., Tranter, M., and Spencer, R. G. M.: Enhanced

845 trace element mobilization by Earth's ice sheets, *Proceedings of the National Academy of Sciences*, 117, 31648-31659, doi:10.1073/pnas.2014378117, 2020.

Hendry, K. R., Huvenne, V. A. I., Robinson, L. F., Annett, A., Badger, M., Jacobel, A. W., Ng, H. C., Opher, J., Pickering, R. A., Taylor, M. L., Bates, S. L., Cooper, A., Cushman, G. G., Goodwin, C., Hoy, S., Rowland, G., Samperiz, A., Williams, J. A., Achterberg, E. P., Arrowsmith, C., Alexander Brearley, J., Henley, S. F., Krause, J. W., Leng, M. J., Li, T., McManus, J.

850 F., Meredith, M. P., Perkins, R., and Woodward, E. M. S.: The biogeochemical impact of glacial meltwater from Southwest Greenland, *Progress in Oceanography*, 176, 102126, doi:10.1016/j.pocean.2019.102126, 2019.

Henson, H. C., Holding, J. M., Meire, L., Rysgaard, S., Stedmon, C. A., Stuart-Lee, A., Bendtsen, J., and Sejr, M.: Coastal freshening drives acidification state in Greenland fjords, *Science of The Total Environment*, 855, 158962, doi:10.1016/j.scitotenv.2022.158962, 2023.

855 Helsel, D. R.: More Than Obvious: Better Methods for Interpreting Nondetect Data, *Environmental Science & Technology*, 39, 419A-423A, doi:10.1021/es053368a, 2005.

Hölemann, J. A., Schirmacher, M., and Prange, A.: Dissolved and Particulate Major and Trace Elements in Newly Formed Ice from the Laptev Sea (Transdrift III, October 1995), in: *Land-Ocean Systems in the Siberian Arctic: Dynamics and History*, edited by: Kassens, H., Bauch, H. A., Dmitrenko, I. A., Eicken, H., Hubberten, H.-W., Melles, M., Thiede, J., and Timokhov, 860 L. A., Springer Berlin Heidelberg, Berlin, Heidelberg, 101-111, doi:10.1007/978-3-642-60134-7_11, 1999.

Hopwood, M. J., Carroll, D., Dunse, T., Hodson, A., Holding, J. M., Iriarte, J. L., Ribeiro, S., Achterberg, E. P., Cantoni, C., Carlson, D. F., Chierici, M., Clarke, J. S., Cozzi, S., Fransson, A., Juul-Pedersen, T., Winding, M. H. S., and Meire, L.: Review article: How does glacier discharge affect marine biogeochemistry and primary production in the Arctic?, *The Cryosphere*, 14, 1347-1383, doi:10.5194/tc-14-1347-2020, 2020.

865 Jain, R. B.: On the consequence of substituting maximum likelihood estimates for the observations below the limit of detection, *Chemosphere*, 144, 2044-2051, doi:10.1016/j.chemosphere.2015.10.113, 2016.

Jensen, L. T., Cullen, J. T., Jackson, S. L., Gerringa, L. J. A., Bauch, D., Middag, R., Sherrell, R. M., and Fitzsimmons, J. N.: A Refinement of the Processes Controlling Dissolved Copper and Nickel Biogeochemistry: Insights From the Pan-Arctic, *Journal of Geophysical Research: Oceans*, 127, e2021JC018087, doi:10.1029/2021JC018087, 2022.

870 Jones, E. M., Chierici, M., Menze, S., Fransson, A., Ingvaldsen, R. B., and Lødemel, H. H.: Ocean acidification state variability of the Atlantic Arctic Ocean around northern Svalbard, *Progress in Oceanography*, 199, 102708, doi:10.1016/j.pocean.2021.102708, 2021.

Jones, E. P., Coote, A. R., and Levy, E. M.: Effect of sea ice meltwater on the alkalinity of seawater, ISSN: 0022-2402, 1983.

Juranek, L.: Changing Biogeochemistry of the Arctic Ocean: Surface Nutrient and CO₂ Cycling in a Warming, Melting North, 875 *Oceanography*, doi:10.5670/oceanog.2022.120, 2022.

Juul-Pedersen, T., Arendt, K. E., Mortensen, J., Blicher, M. E., Søgaard, D. H., and Rysgaard, S.: Seasonal and interannual phytoplankton production in a sub-Arctic tidewater outlet glacier fjord, SW Greenland, *Marine Ecology Progress Series*, 524, 27-38, <https://www.int-res.com/abstracts/meps/v524/p27-38/>, 2015.

Kanna, N., Toyota, T., and Nishioka, J.: Iron and macro-nutrient concentrations in sea ice and their impact on the nutritional 880 status of surface waters in the southern Okhotsk Sea, *Progress in Oceanography*, 126, 44-57, doi:10.1016/j.pocean.2014.04.012, 2014.

Krause, J., Hopwood, M. J., Höfer, J., Krisch, S., Achterberg, E. P., Alarcón, E., Carroll, D., González, H. E., Juul-Pedersen, T., Liu, T., Lodeiro, P., Meire, L., and Rosing, M. T.: Trace Element (Fe, Co, Ni and Cu) Dynamics Across the Salinity Gradient in Arctic and Antarctic Glacier Fjords, *Frontiers in Earth Science*, 9, doi:10.3389/feart.2021.725279, 2021.

885 Krawczyk, D. W., Kryk, A., Juggins, S., Burmeister, A., Pearce, C., Seidenkrantz, M. S., Moros, M., Hoyer, J. L., Kuijpers, A., and Witkowski, A.: Spatio-temporal changes in ocean conditions and primary production in Baffin Bay and the Labrador Sea, *Palaeogeography, Palaeoclimatology, Palaeoecology*, 563, 110175, doi:10.1016/j.palaeo.2020.110175, 2021.

Krisch, S., Hopwood, M. J., Roig, S., Gerringa, L. J. A., Middag, R., Rutgers van der Loeff, M. M., Petrova, M. V., Lodeiro, P., Colombo, M., Cullen, J. T., Jackson, S. L., Heimbürger-Boavida, L.-E., and Achterberg, E. P.: Arctic – Atlantic Exchange of the Dissolved Micronutrients Iron, Manganese, Cobalt, Nickel, Copper and Zinc With a Focus on Fram Strait, *Global Biogeochemical Cycles*, 36, e2021GB007191, doi:10.1029/2021GB007191, 2022a.

Krisch, S., Huhn, O., Al-Hashem, A., Hopwood, M. J., Lodeiro, P., and Achterberg, E. P.: Quantifying Ice-Sheet Derived Lead (Pb) Fluxes to the Ocean; A Case Study at Nioghalvfjerdssbræ, *Geophysical Research Letters*, 49, e2022GL100296, doi:10.1029/2022GL100296, 2022b.

Lafond, A., Leblanc, K., Queguiner, B., Moriceau, B., Leynaert, A., Cornet, V., Legras, J., Ras, J., Parenteau, M., Garcia, N., Babin, M., and Tremblay, J.-E.: Late spring bloom development of pelagic diatoms in Baffin Bay, *Elementa-science Of The Anthropocene*, 7, doi:10.1525/elementa.382, 2019.

Lehmann, N., Kienast, M., Granger, J., Bourbonnais, A., Altabet, M. A., and Tremblay, J.-É.: Remote Western Arctic Nutrients Fuel Remineralization in Deep Baffin Bay, *Global Biogeochemical Cycles*, 33, 649-667, doi:10.1029/2018GB006134, 2019.

Li, Y.-H., and Tsui, T.-F.: The solubility of CO₂ in water and sea water, *Journal of Geophysical Research (1896-1977)*, 76, 4203-4207, doi:10.1029/JC076i018p04203, 1971.

Marsay, C. M., Aguilar-Islas, A., Fitzsimmons, J. N., Hatta, M., Jensen, L. T., John, S. G., Kadko, D., Landing, W. M., Lanning, N. T., Morton, P. L., Pasqualini, A., Rauschenberg, S., Sherrell, R. M., Shiller, A. M., Twining, B. S., Whitmore, L. M., Zhang, R., and Buck, C. S.: Dissolved and particulate trace elements in late summer Arctic melt ponds, *Marine Chemistry*, 204, 70-85, doi:10.1016/j.marchem.2018.06.002, 2018.

McConnell, J. R., and Edwards, R.: Coal burning leaves toxic heavy metal legacy in the Arctic, *Proceedings of the National Academy of Sciences*, 105, 12140-12144, doi:10.1073/pnas.0803564105, 2008.

Measures, C. I.: The role of entrained sediments in sea ice in the distribution of aluminium and iron in the surface waters of the Arctic Ocean, *Marine Chemistry*, 68, 59-70, doi:10.1016/S0304-4203(99)00065-1, 1999.

Mehrbach, C., Culberson, C. H., Hawley, J. E., and Pytkowicz, R. M.: MEASUREMENT OF THE APPARENT DISSOCIATION CONSTANTS OF CARBONIC ACID IN SEAWATER AT ATMOSPHERIC PRESSURE1, *Limnology and Oceanography*, 18, 897-907, doi:10.4319/lo.1973.18.6.0897, 1973.

Meire, L., Søgaard, D. H., Mortensen, J., Meysman, F. J. R., Soetaert, K., Arendt, K. E., Juul-Pedersen, T., Blicher, M. E., and Rysgaard, S.: Glacial meltwater and primary production are drivers of strong CO₂ uptake in fjord and coastal waters adjacent to the Greenland Ice Sheet, *Biogeosciences*, 12, 2347-2363, doi:10.5194/bg-12-2347-2015, 2015.

Meire, L., Mortensen, J., Meire, P., Juul-Pedersen, T., Sejr, M. K., Rysgaard, S., Nygaard, R., Huybrechts, P., and Meysman, F. J. R.: Marine-terminating glaciers sustain high productivity in Greenland fjords, *Glob Chang Biol*, 23, 5344-5357, doi:10.1111/gcb.13801, 2017.

Miller, L., Davelaar, M., Caleb, D., Mucci, A., Burgers, T., Ahmed, M., and Irish, V.: Dissolved inorganic carbon (DIC), total alkalinity, stable oxygen isotope (O-18), temperature, salinity, dissolved oxygen and other parameters measured from discrete samples and profile observations during the Canadian Coast Guard Ship Amundsen ArcticNet cruise (EXPOCODE

18DL20160603, Leg 1 and Leg 2) in the Eastern Canadian Arctic, Baffin Bay, Nares Strait, Lancaster Sound, Barrow Strait and Coronation Gulf from 2016-06-03 to 2016-08-23 (NCEI Accession 0217304), NOAA National Centers for Environmental Information [data set], doi:10.25921/719e-qr37, 2020.

925 Møller, E. F., Christensen, A., Larsen, J., Mankoff, K. D., Ribergaard, M. H., Sejr, M., Wallhead, P., and Maar, M.: The sensitivity of primary productivity in Disko Bay, a coastal Arctic ecosystem, to changes in freshwater discharge and sea ice cover, *Ocean Sci.*, 19, 403-420, doi:10.5194/os-19-403-2023, 2023.

Monteban, D., Pedersen, J. O. P., and Nielsen, M. H.: Physical oceanographic conditions and a sensitivity study on meltwater runoff in a West Greenland fjord: Kangerlussuaq, *Oceanologia*, 62, 460-477, doi:10.1016/j.oceano.2020.06.001, 2020.

930 Mouginot, J., Rignot, E., Bjørk, A. A., van den Broeke, M., Millan, R., Morlighem, M., Noël, B., Scheuchl, B., and Wood, M.: Forty-six years of Greenland Ice Sheet mass balance from 1972 to 2018, *Proceedings of the National Academy of Sciences*, 116, 9239-9244, doi:10.1073/pnas.1904242116, 2019.

Nakanowatari, T., Ohshima, K. I., and Wakatsuchi, M.: Warming and oxygen decrease of intermediate water in the northwestern North Pacific, originating from the Sea of Okhotsk, 1955–2004, *Geophysical Research Letters*, 34, 935 doi:10.1029/2006GL028243, 2007.

Niebauer, H. J., Alexander, V., and Henrichs, S. M.: A time-series study of the spring bloom at the Bering Sea ice edge I. Physical processes, chlorophyll and nutrient chemistry, *Continental Shelf Research*, 15, 1859-1877, doi:10.1016/0278-4343(94)00097-7, 1995.

Oksman, M., Kvorning, A. B., Larsen, S. H., Kjeldsen, K. K., Mankoff, K. D., Colgan, W., Andersen, T. J., Nørgaard-Pedersen, 940 N., Seidenkrantz, M. S., Mikkelsen, N., and Ribeiro, S.: Impact of freshwater runoff from the southwest Greenland Ice Sheet on fjord productivity since the late 19th century, *The Cryosphere*, 16, 2471-2491, doi:10.5194/tc-16-2471-2022, 2022.

Peres-Neto, P. R., Jackson, D. A., and Somers, K. M.: GIVING MEANINGFUL INTERPRETATION TO ORDINATION AXES: ASSESSING LOADING SIGNIFICANCE IN PRINCIPAL COMPONENT ANALYSIS, *Ecology*, 84, 2347-2363, doi:10.1890/00-0634, 2003.

945 Perrette, M., Yool, A., Quartly, G. D., and Popova, E. E.: Near-ubiquity of ice-edge blooms in the Arctic, *Biogeosciences*, 8, 515-524, doi:10.5194/bg-8-515-2011, 2011.

Pierrot, D., Wallace, D., and Lewis, E.: MS Excel program developed for CO₂ system calculations, Carbon Dioxide Information Analysis Center [code], doi:10.3334/CDIAC/otg.CO2SYS_XLS_CDIAC105a, 2011.

Posit team: RStudio: Integrated Development Environment for R, Posit Software, PBC [code], <http://www.posit.co/>, 2023.

950 Przibilla, A., Iwinski, S., Zimmermann, T., and Pröfrock, D.: Impact of storage temperature and filtration method on dissolved trace metal concentrations in coastal water samples, *Water Environment Research*, 95, e10922, doi:10.1002/wer.10922, 2023.

R Core Team: R: A Language and Environment for Statistical Computing, R Foundation for Statistical Computing [code], <https://www.R-project.org/>, 2022.

Revelle, W.: psych: Procedures for Psychological, Psychometric, and Personality Research, CRAN [code], <https://CRAN.R-project.org/package=psych>, 2024.

Ruacho, A., Richon, C., Whitby, H., and Bundy, R. M.: Sources, sinks, and cycling of dissolved organic copper binding ligands in the ocean, *Communications Earth & Environment*, 3, 263, doi:10.1038/s43247-022-00597-1, 2022.

Rysgaard, S., Bendtsen, J., Delille, B., Dieckmann, G. S., Glud, R. N., Kennedy, H., Mortensen, J., Papadimitriou, S., Thomas, D. N., and Tison, J.-L.: Sea ice contribution to the air–sea CO₂ exchange in the Arctic and Southern Oceans, *Tellus B: Chemical and Physical Meteorology*, doi:10.1111/j.1600-0889.2011.00571.x, 2011.

Rysgaard, S., Glud, R. N., Lennert, K., Cooper, M., Halden, N., Leakey, R. J. G., Hawthorne, F. C., and Barber, D.: Ikaite crystals in melting sea ice – implications for CO₂ and pH levels in Arctic surface waters, *The Cryosphere*, 6, 901-908, doi:10.5194/tc-6-901-2012, 2012.

Sarmiento, J. L., and Gruber, N.: *Ocean Biogeochemical Dynamics*, Princeton University Press, ISBN: 9780691017075, 2006.

Schmidt, C. E., Pröfrock, D., and Thomas, H.: ECOTIP Dana Cruise July 2021 - Alkalinity (AT), dissolved inorganic carbon (CT) and dissolved trace elements, Zenodo [data set], doi:10.5281/zenodo.14235091, 2024.

Schnetger, B., and Lehnert, C.: Determination of nitrate plus nitrite in small volume marine water samples using vanadium(III)chloride as a reduction agent, *Marine Chemistry*, 160, 91-98, doi:10.1016/j.marchem.2014.01.010, 2014.

Seo, H., Kim, G., Kim, T., Kim, I., Ra, K., and Jeong, H.: Trace elements (Fe, Mn, Co, Cu, Cd, and Ni) in the East Sea (Japan Sea): Distributions, boundary inputs, and scavenging processes, *Marine Chemistry*, 239, 104070, doi:10.1016/j.marchem.2021.104070, 2022.

Shadwick, E. H., Thomas, H., Chierici, M., Else, B., Fransson, A., Michel, C., Miller, L. A., Mucci, A., Niemi, A., Papakyriakou, T. N., and Tremblay, J.-É.: Seasonal variability of the inorganic carbon system in the Amundsen Gulf region of the southeastern Beaufort Sea, *Limnology and Oceanography*, 56, 303-322, doi:10.4319/lo.2011.56.1.0303, 2011a.

Shadwick, E. H., Thomas, H., Gratton, Y., Leong, D., Moore, S. A., Papakyriakou, T., and Prowe, A. E. F.: Export of Pacific carbon through the Arctic Archipelago to the North Atlantic, *Continental Shelf Research*, 31, 806-816, doi:10.1016/j.csr.2011.01.014, 2011b.

Shadwick, E. H., Trull, T. W., Thomas, H., and Gibson, J. A. E.: Vulnerability of Polar Oceans to Anthropogenic Acidification: Comparison of Arctic and Antarctic Seasonal Cycles, *Scientific Reports*, 3, 2339, doi:10.1038/srep02339, 2013.

Sherwood, O. A., Davin, S. H., Lehmann, N., Buchwald, C., Edinger, E. N., Lehmann, M. F., and Kienast, M.: Stable isotope ratios in seawater nitrate reflect the influence of Pacific water along the northwest Atlantic margin, *Biogeosciences*, 18, 4491-4510, doi:10.5194/bg-18-4491-2021, 2021.

Smrzka, D., Zwicker, J., Bach, W., Feng, D., Himmeler, T., Chen, D., and Peckmann, J.: The behavior of trace elements in seawater, sedimentary pore water, and their incorporation into carbonate minerals: a review, *Facies*, 65, 41, doi:10.1007/s10347-019-0581-4, 2019.

Spreen, G., Kaleschke, L., and Heygster, G.: Sea ice remote sensing using AMSR-E 89-GHz channels, *Journal of Geophysical Research: Oceans*, 113, doi:10.1029/2005JC003384, 2008.

Stedmon, C., Svensen, C., and Wiedmann, I.: Nutrient Data for cruise DANA 6/21 (18 - 28 July 2021), Zenodo [data set], doi:10.5281/zenodo.15590652, 2025.

990 Strass, V. H., and Nöthig, E. M.: Seasonal shifts in ice edge phytoplankton blooms in the Barents Sea related to the water column stability, *Polar Biology*, 16, 409-422, doi:10.1007/BF02390423, 1996.

Stroeve, J., and Notz, D.: Changing state of Arctic sea ice across all seasons, *Environmental Research Letters*, 13, 103001, doi:10.1088/1748-9326/aade56, 2018.

995 Tang, C. C. L., Ross, C. K., Yao, T., Petrie, B., DeTracey, B. M., and Dunlap, E.: The circulation, water masses and sea-ice of Baffin Bay, *Progress in Oceanography*, 63, 183-228, doi:10.1016/j.pocean.2004.09.005, 2004.

Thomas, H., Shadwick, E., Dehairs, F., Lansard, B., Mucci, A., Navez, J., Gratton, Y., Prowe, F., Chierici, M., Fransson, A., Papakyriakou, T. N., Sternberg, E., Miller, L. A., Tremblay, J.-É., and Monnin, C.: Barium and carbon fluxes in the Canadian Arctic Archipelago, *Journal of Geophysical Research: Oceans*, 116, doi:10.1029/2011JC007120, 2011.

1000 Tovar-Sánchez, A., Duarte, C. M., Alonso, J. C., Lacorte, S., Tauler, R., and Galbán-Malagón, C.: Impacts of metals and nutrients released from melting multiyear Arctic sea ice, *Journal of Geophysical Research: Oceans*, 115, doi:10.1029/2009JC005685, 2010.

Tremblay, J.-É., Gratton, Y., Carmack, E. C., Payne, C. D., and Price, N. M.: Impact of the large-scale Arctic circulation and the North Water Polynya on nutrient inventories in Baffin Bay, *Journal of Geophysical Research: Oceans*, 107, 26-21-26-14, doi:10.1029/2000JC000595, 2002.

1005 Tynan, E., Clarke, J. S., Humphreys, M. P., Ribas-Ribas, M., Esposito, M., Rérolle, V. M. C., Schlosser, C., Thorpe, S. E., Tyrrell, T., and Achterberg, E. P.: Physical and biogeochemical controls on the variability in surface pH and calcium carbonate saturation states in the Atlantic sectors of the Arctic and Southern Oceans, *Deep Sea Research Part II: Topical Studies in Oceanography*, 127, 7-27, doi:10.1016/j.dsrr.2016.01.001, 2016.

Vancoppenolle, M., Meiners, K. M., Michel, C., Bopp, L., Brabant, F., Carnat, G., Delille, B., Lannuzel, D., Madec, G., 1010 Moreau, S., Tison, J.-L., and van der Merwe, P.: Role of sea ice in global biogeochemical cycles: emerging views and challenges, *Quaternary Science Reviews*, 79, 207-230, doi:10.1016/j.quascirev.2013.04.011, 2013

Vernet, M., Ellingsen, I., Marchese, C., Bélanger, S., Cape, M., Slagstad, D., and Matrai, P. A.: Spatial variability in rates of net primary production (NPP) and onset of the spring bloom in Greenland shelf waters, *Progress in Oceanography*, 198, 102655, doi:10.1016/j.pocean.2021.102655, 2021.

1015 Vieira, L. H., Achterberg, E. P., Scholten, J., Beck, A. J., Liebetrau, V., Mills, M. M., and Arrigo, K. R.: Benthic fluxes of trace metals in the Chukchi Sea and their transport into the Arctic Ocean, *Marine Chemistry*, 208, 43-55, doi:10.1016/j.marchem.2018.11.001, 2019.

Whitmore, L. M., Morton, P. L., Twining, B. S., and Shiller, A. M.: Vanadium cycling in the Western Arctic Ocean is influenced by shelf-basin connectivity, *Marine Chemistry*, 216, 103701, doi:10.1016/j.marchem.2019.103701, 2019.

1020 Williams, P. L., Burgess, D. O., Waterman, S., Roberts, M., Bertrand, E. M., and Bhatia, M. P.: Nutrient and Carbon Export From a Tidewater Glacier to the Coastal Ocean in the Canadian Arctic Archipelago, *Journal of Geophysical Research: Biogeosciences*, 126, e2021JG006289, doi:10.1029/2021JG006289, 2021.

Wouters, B., and Sasgen, I.: Increasing freshwater fluxes from the Greenland Ice Sheet observed from space, *Oceanography*, 35, 103-105, doi:10.5670/oceanog.2022.125, 2022.

1025 Wu, Y., Hain, M. P., Humphreys, M. P., Hartman, S., and Tyrrell, T.: What drives the latitudinal gradient in open-ocean surface dissolved inorganic carbon concentration?, *Biogeosciences*, 16, 2661-2681, doi:10.5194/bg-16-2661-2019, 2019.