

Temporary waterlogging alters CO₂ flux dynamics but not cumulative emissions in cultivated mineral soils

Reija Kronberg^{1,2}, Sanna Kanerva¹, Markku Koskinen^{1,2}, Tatu Polvinen^{1,2}, Tuomas Mattila³, & Mari Pihlatie^{1,2}

5 ¹University of Helsinki, Department of Agricultural Sciences, Unit of Environmental Soil Science, Viikinkaari 9, P.O. Box 56, FI-00014 Helsinki, Finland

²Institute for Atmospheric and Earth System Research (INAR)/Faculty of Agriculture and Forestry, University of Helsinki, Viikinkaari 9, P.O. Box 56, FI-00014 Helsinki, Finland

³Finnish Environment Institute SYKE, Latokartanonkaari 11, 00790, Helsinki, Finland

10

Correspondence to:

Reija Kronberg

E-mail address: reija.kronberg@helsinki.fi

Postal address: Viikinkaari 9, 00790 Helsinki

15

Mari Pihlatie

E-mail address: mari.pihlatie@helsinki.fi

Postal address: Viikinkaari 9, 00790 Helsinki

Abstract

20 Increasingly variable rainfall patterns expose soils to more frequent waterlogging in humid climates. Yet, the effects of waterlogging on soil organic matter decomposition in mineral soils remain uncertain. We studied the impact of off-season waterlogging on carbon dioxide (CO₂) production and dissolved carbon dynamics in controlled greenhouse conditions using 32 monolithic soil columns (hereafter monoliths) (h=63 cm, d=15.2 cm) sampled from two agricultural fields (silty clay, sandy loam) in southern Finland. The 1.5-year study comprised three growth cycles with alternating growing and off-seasons. Spring 25 barley (*Hordeum vulgare*) was grown in all monoliths during the growing seasons. In turn, during all three off-seasons, half of the monoliths were subjected to waterlogging lasting seven weeks, while in the control monoliths soil moisture was maintained below field capacity. Within these water treatment groups, the monoliths were further divided into two plant treatment groups: in half of the monoliths, an overwintering cover crop (*Festuca arundinacea*) was grown, while in the other half soil was left bare for the off-seasons. Soil temperature and moisture were continuously monitored, dissolved organic 30 (DOC) and inorganic carbon (DIC) concentrations in pore water were analyzed at three depths and CO₂ fluxes were measured at the surface. Contrary to our hypothesis, waterlogging did not increase soil DOC content. Instead, on-going microbial/rhizospheric activity promoted an increase in DIC content while CO₂ fluxes declined, indicating an accumulation of respired CO₂ in soil pore water. The sustained CO₂ production could not be explained solely by mobilization of Fe-associated C, as initially hypothesized. After the initiation of drainage, CO₂ fluxes from both soils increased more than predicted based 35 on changes in soil moisture and temperature, likely due to the release of previously accumulated CO₂. These post-waterlogging increases in CO₂ fluxes roughly equaled the earlier decreases during waterlogging. Thus, although off-season waterlogging strongly influenced the temporal dynamics of CO₂ fluxes, it did not alter total cumulative CO₂ emissions from the studied agricultural soils.

Keywords: soil respiration, CO₂ efflux, climate change, soil moisture, dissolved carbon, agricultural soil, cover crop

40 Summary

We studied how off-season waterlogging affects CO₂ fluxes and dissolved carbon dynamics in two cultivated boreal mineral soils. The study was conducted with intact soil profiles in a greenhouse. Waterlogging reduced immediate CO₂ efflux, but CO₂ accumulated in porewater and was released to the atmosphere upon soil drying. Cumulative emissions remained unaltered. Our results suggest that temporary waterlogging does not suppress CO₂ production in mineral soils as much as traditionally 45 assumed.

1 Introduction

Climate change is causing significant alterations in rainfall patterns globally increasing the inter- and intra-seasonal variability, and intensifying the climatic extremes (IPCC, 2023). In humid boreal regions, mild and rainy winters may lead to prolonged and more frequent soil waterlogging outside growing seasons (Mattila and Vihanto, 2024; Ruosteenoja and Jylhä, 2022). Soil moisture is a critical environmental factor influencing soil carbon (C) fluxes (Raich and Schlesinger, 1992). Yet, substantial uncertainties remain concerning the response of soil organic matter (OM) decomposition and carbon dioxide (CO₂) efflux to periods of very high soil moisture (temporary waterlogging) in cultivated mineral soils (Fairbairn et al., 2023; Ghezzehei et al., 2019; Goffin et al., 2015; Huang and Hall, 2017; Moyano et al., 2018). Deepening our understanding of the relationship between soil moisture and C dynamics at or near complete soil water saturation with potentially anaerobic conditions is crucial for improving predictions of soil-climate feedbacks in changing environmental conditions.

As opposed to conventional theory, recent research has shown that temporary anaerobic conditions formed upon waterlogging may even enhance rather than suppress C mineralization in predominantly aerobic mineral soils (e.g. Huang et al., 2021; Huang and Hall, 2017; Liu et al., 2023; Winkler et al., 2019). Although many C cycling and respiration models still predict near zero CO₂ fluxes at water saturation (Ghezzehei et al., 2019; Moyano et al., 2013; Sulman et al., 2014), mineral soils have been shown to sustain substantial CO₂ production in these conditions due to on-going anaerobic respiration (Fairbairn et al., 2023; Huang et al., 2021; Huang and Hall, 2017; Liu et al., 2023; Moyano et al., 2018; Wickland and Neff, 2008; Zheng et al., 2019). This occurs because most of the OM is stabilized by mineral associations (Salonen et al., 2024; von Lützow et al., 2007) which provides protection against biodegradation (Cotrufo et al., 2019; Lavalley et al., 2020), at least in static oxic conditions. In humid soils, redox active iron(III) (Fe(III)) (hydr)oxides represent essential binding agents (Aasano et al., 2018; Rasmussen et al., 2018; Torn et al., 1997). Therefore, if soil saturates with water for prolonged periods and reducing conditions form in the bulk soil, (facultative) anaerobic microbes start utilizing alternative terminal electron acceptors (TEA), such as Fe(III) oxides, instead of oxygen (O₂), to oxidize OM (Fiedler et al., 2007; Jakobsen et al., 1981; Peters and Conrad, 1996; Ponnampertuma, 1985). Upon reduction, the solubility of Fe increases, which leads to the disruption of Fe oxides and the release of OM associated with them, thereby making OM more vulnerable to mineralization (Chen et al., 2020; Fairbairn et al., 2023; Grybos et al., 2009; Huang et al., 2021, 2020; Huang and Hall, 2017).

The influence of anaerobic conditions on C mineralization has, however, not been uniform across soils and experiments. Soil characteristics, such as texture, the amount (Hanke et al., 2013; Liu et al., 2023; Winkler et al., 2019; Zhao et al., 2020) and reducibility (Ginn et al., 2017) of active Fe oxides, and the availability of labile C substrates (e.g. Winkler et al., 2019), influence the soil's response to waterlogging. Naturally, an adequate amount of reducible Fe is a prerequisite for the proposed mechanism. Although in humid boreal soils Fe oxides represent a minority of the clay-sized fraction (Ito and Wagai, 2017), amorphous, low-crystallinity forms constitute a substantial component of the oxide pool (Keskinen et al 2022; Sippola 1974). These short-range-ordered (SRO) Fe and Al oxides play a disproportionately important role in C stabilization thanks to their

high specific surface area and chemical reactivity (Kaiser and Guggenberger, 2003; Yong et al., 1992). Moreover, naturally occurring oscillations in soil oxygen availability, particularly in fine-textured humid soils, may enhance Fe reducibility (Ginn et al., 2017) thereby eventually also influencing C availability. Indeed, a recent large-scale study of arable soils in southern Finland showed that Fe oxides correlated with mineral-associated organic C, particularly in soils with high clay contents (Salonen et al., 2024). However, to our knowledge, studies focusing on Fe–OM interactions in agricultural boreal soils under temporary anoxia are lacking, and it therefore remains unknown whether and how seasonal waterlogging affects C dynamics in these systems. Because soil redox reactions are predominantly microbially catalyzed (Sigg, 2000; Stumm, 1995), the availability of labile C substrates that stimulates microbial activity and O₂ consumption, promotes the formation of reducing conditions and redox transformations of Fe (Khan et al., 2019; Muhammad et al., 2021; Winkler et al., 2019). In natural and cultivated soils, plant roots are a source of readily degradable C for soil microbes (Kuzyakov, 2006; Muhammad et al., 2021). As much as half of the C photosynthetically fixed by plants can be allocated belowground (Chandregowda et al., 2023; Clemmensen et al., 2013; Keiluweit et al., 2015). In agriculture, the cultivation of overwintering cover crops has been recognized as an effective strategy to increase soil C inputs and thus, promote soil C sequestration (Kaye and Quemada, 2017; Poeplau and Don, 2015). However, it has also been observed that root exudates can stimulate decomposition of OM through priming (Bingeman et al., 1953), either directly by increasing microbial activity or indirectly by promoting the mobilization of C associated with soil minerals (Keiluweit et al., 2015; Kuzyakov et al., 2000). This, together with the changing climatic conditions, introduces complexity in assessing the net impact of cover crops on soil C dynamics (Jian et al., 2020; Poeplau and Don, 2015). To our knowledge, the impact of cover crops on C fluxes during periodic waterlogging has gained very little attention, despite the potential for root-derived C inputs to promote the mobilization of stable, mineral-associated C. Thus, it remains unknown whether and to what extent the labile C inputs from roots affect C dynamics in temporarily waterlogged conditions.

Thus far, studies focusing on the effects of water saturation/anaerobic conditions on OM mobilization and CO₂ fluxes in mineral soils have mainly been either laboratory scale incubations (Bhattacharyya et al., 2018; Fairbairn et al., 2023; Huang et al., 2021, 2020; Huang and Hall, 2017; Zhao et al., 2020) or field studies (Jeanneau et al. 2020; Possinger et al. 2020). The middle ground mesocosm experiments considering the essential role of soil structure (Bergström, 1990; Lehmann et al., 2020) and plant cover (Keiluweit, 2015; Liang, 2023) on C flows and transformations, while maintaining the control over environmental conditions, have been rare. In our 1.5-year greenhouse experiment, we focus on this knowledge-gap by studying C dynamics in intact soil profiles under controlled temperature and water conditions to capture processes more representative of field conditions. The data used in this study was collected from a 1.5-year soil monolith experiment which experimental procedure has been described in our preceding publication (Kronberg et al. 2024). In this particular study, we investigate how seasonal waterlogging affects CO₂ fluxes and dissolved C species dynamics in two hydrologically distinct cultivated mineral soil profiles (silty clay, sandy loam) with and without overwintering cover crop (Tall Fescue, *Festuca arundinacea*). . Finally,

an empirical model describing the dependency of soil respiration on temperature and moisture was fitted to the data to assess its ability to simulate observed CO₂ flux dynamics during and after waterlogging. The following hypotheses were set:

- a) Temporary waterlogging mobilizes Fe-associated C leading to increased soil DOC content.
- b) Temporary waterlogging does not reduce cumulative CO₂ emissions from either soil.
- 115 c) Silty clay with higher OM, clay and Fe oxide contents result in higher cumulative CO₂ fluxes than from sandy loam soil.
- d) Soil C input from cover crops promotes C dissolution and CO₂ production in waterlogged conditions.

2 Material and methods

120 2.1 Soil monolith experiment

Cylindrical soil monoliths (d, 15.2 cm; h, 63 cm) were collected with a tractor-mounted soil auger in November 2020 and May 2021 (Uusitalo et al., 2012) from two agricultural fields in Southern Finland classified as Eutric Stagnosol (60°16'23.4"N 24°56'40.6"E) and Eutric Cambisol/Mollic Umbrisol (60°49'07.3"N 23°45'54.8"E) with silty clay and sandy loam USDA texture, respectively. These soils differed in their hydrological characteristics. The dense structure and fine texture in the silty clay soil limited the movement of water through the profile causing water stagnation and repeated alteration of oxidized and reduced conditions resulting in the formation of stagnic properties (see Kronberg et al. 2024). In contrast, the coarse texture in the sandy loam soil facilitated an efficient water movement and transport of solutes into deeper soil layers. In the profile brown colors indicated a release of Fe from primary minerals. The two soils represented typical organic matter (Fernandez-Ugalde et al., 2022; Heikkinen et al., 2013; Lemola et al., 2018) and SRO Fe oxide contents for the region. Selected soil properties are presented in Table 1.

125
130

Table 1. Selected physical and chemical properties of the studied soils. The results accompanied by detailed analytical methods have been previously described in Kronberg et al. (2024).

	Depth (cm)	Soil C & N			pH _{H2O}	Oxides		ρ_b (kg dm ⁻³)	Particle size distribution		
		Total C (%)	WEOC (mg kg ⁻¹)	Total N (%)		Al _{ox} (mmol kg ⁻¹)	Fe _{ox} (mmol kg ⁻¹)		Sand (%)	Silt (%)	Clay (%)
Silty clay	0-10	2.6	48	0.22	7.1	62	112	1.01	9	43	47
	10-20	2.3	50	0.20	7.0	69	117	1.11			
	20-30	1.2	41	0.10	7.0	57	77	1.50	6	34	60
	30-40	0.5	32	0.05	7.2	50	41	1.53			
	40-50	0.3	26	0.03	7.4	48	37	1.48	3	39	58
	50-60	0.3	24	0.02	7.5	35	35	1.43			
Sandy loam	0-10	1.7	33	0.15	6.6	109	51	1.13	67	16	17
	10-20	1.8	29	0.15	6.7	107	50	1.26			
	20-30	1.7	33	0.15	6.7	110	51	1.40	71	15	14
	30-40	1.4	39	0.10	6.8	155	50	1.39			
	40-50	0.7	47	0.03	6.8	171	50	1.31	84	11	5
	50-60	0.5	45	0.02	6.8	152	34	1.39			

Symbols and abbreviations: WEOC, water extractable organic carbon; Al_{ox}, Oxalate extractable aluminum (poorly ordered oxides); Fe_{ox}, Oxalate extractable iron

135 (poorly ordered oxides); ρ_b , soil bulk density.

The experiment was established in a greenhouse compartment at University of Helsinki in June 2021, and it is described in detail in Kronberg et al. (2024). Briefly, 32 soil monoliths were collected from the two agricultural fields, with 16 from each soil type. These monoliths were assigned to a factorial design with two plant treatments and two water treatments. This setup resulted in eight unique treatment combinations (2 soil types \times 2 plant treatments \times 2 water treatments), each having four treatment replicates (Fig. 1). During the 1.5 year-long experiment, three study cycles with alternating growing and off-seasons were conducted. Repeated cycles were conducted to assess (i) whether the observed responses were consistent over study cycles, thereby increasing the robustness of our findings, and (ii) whether treatment effects, particularly those related to the cover crop, required time to emerge, given that measurable changes in soil properties develop gradually.

Here, only the results from the second and the third study cycles are presented because the first cycle was dedicated for optimization of the experimental conditions, sampling and monitoring methods.

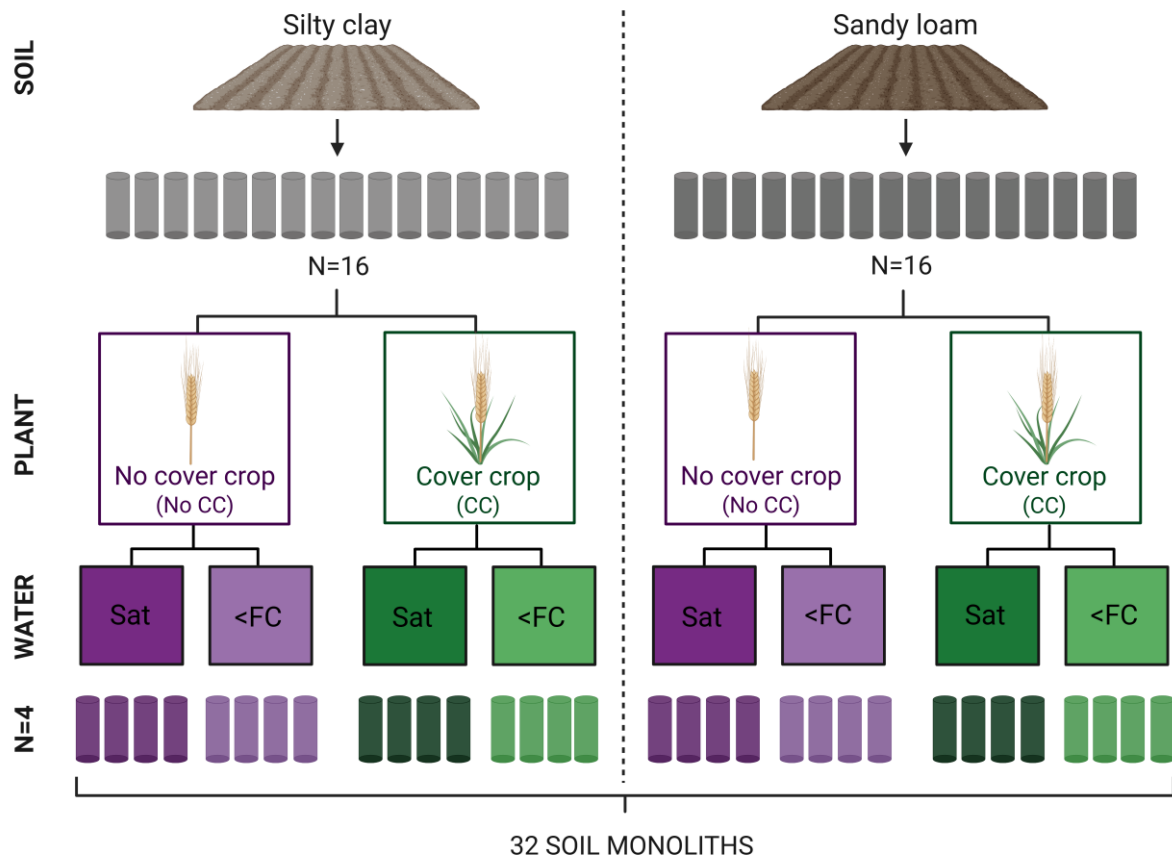


Figure 1. A schematic illustration of the soil monolith experiment study design. The 16 monoliths collected from each agricultural field (silty clay, sandy loam) were assigned to two levels of plant (No CC, No cover crop; CC, Cover crop) and water treatments (Sat, Water saturation; <FC, moisture maintained below field capacity).

150 Soil temperature, volumetric water content (SVWC, $\text{m}^3 \text{m}^{-3}$) and redox potential (E_h) were continuously measured at 10, 30 and 50 cm soil depths in three out of four replicated monoliths (24 in total). Soil temperature and SVWC were measured with Teros12 sensors (METER Group, USA). The soil moisture readings were calibrated to each monolith according to the gravimetric soil moisture content and bulk density determined at the end of the experiment (Kronberg et al., 2024). E_h was measured by platinum electrodes together with Ag/AgCl saturated KCl reference electrodes (Paleo Terra, The Netherlands).
155 Data was logged with a Campbell datalogger (Campbell Scientific CR800 with two AM16/32B multiplexers) (Kronberg et al., 2024). The target seasonal temperatures (day/night) for the greenhouse were 20/15 °C for summer, 15/10 °C for autumn, 10/5 °C for winter and 15/10 °C for spring. The soil temperature was controlled with a separate cooler and an insulation box constructed around the soil monoliths. The realized mean soil temperature at 30 cm depth was 15.1 & 15.3 °C in summer, 10.9 & 11.7 °C in autumn, 5.7 & 7.1 °C in winter and 10.9 & 10.1 °C in spring during study cycles two and three, respectively.
160 During the waterlogging treatment the mean temperature was 6.4 °C in the second and 7.5 °C in the third cycle (Kronberg et al., 2024).

Barley (*Hordeum vulgare*) was grown in all monoliths during the growing seasons while Tall Fescue (*Festuca arundinacea*) was under sown to half of the monoliths as an overwintering cover crop. Both barley and cover crop biomass were harvested and the dry mass was determined by drying at 105 °C for one hour and then overnight at 60 °C (Hakala et al., 2009). Afterwards,
165 the cover crop was allowed to grow over the experimental winter and again harvested before the next cultivation. During each study cycle, all monoliths were fertilized equally using laboratory quality salts with applied nutrient levels corresponding to 90, 110 and 150 kg N ha^{-1} ; 0, 12.5 and 25 kg P ha^{-1} ; and 32, 25 and 100 kg K ha^{-1} in cycles one, two and three, respectively. The fertilization was increased towards the end of the experiment to address nutrient deficiencies observed in barley. Micronutrients were applied as a foliar fertilizer (YaraVita Solatrel) during the second and third study cycles (Kronberg et al.,
170 2024).

During the growing seasons all monoliths received equal irrigation of 17–27 mm week^{-1} via computer controlled Priva automated drip irrigation system (Greenhouse Environmental Control Systems, Priva BV, The Netherlands). Throughout the experiment, the monoliths were irrigated with artificial rainwater which was made according to the ionic composition of typical rainwater in Southern Finland (Vet et al., 2014). After the harvest i.e., during the off-seasons, half of the monoliths received
175 excessive manual irrigation until saturation was reached and water infiltration into the soil ceased (Saturation treatment). In the other half, the topsoil moisture was maintained below soil field capacity (<FC treatment) at ~50 % water filled pore space (Kronberg et al., 2024). Soil moisture in all monoliths was adjusted according to soil moisture readings from Teros12 sensors. The waterlogging treatment was initiated by mimicking a three-day heavy rainfall event (25 mm d^{-1}) that naturally occur in

Southern Finland (Uusitalo et al., 2012; Virtanen et al., 2013). The drainage outlets were closed on the third day of the heavy rainfall event (taken as day 0 of waterlogging treatment). After, these monoliths were manually irrigated with 0–23 mm water per monolith per irrigation, and this was repeated several times per day if needed. Irrigation was stopped/paused once infiltration ceased. Whereas the silty clay soil got waterlogged already during the simulated heavy rainfall event, waterlogging of the sandy loam soil took on average eight days longer due to the larger fraction of air-filled soil pores at the start of the excessive irrigation (Kronberg et al., 2024). In the second study cycle, some unfortunate leakages occurred in some sandy loam monoliths through sensor hole sealings which is why some deviations from complete waterlogging in the topsoil occurred (further discussed in Kronberg et al. 2024). The length of the waterlogging treatment in cycles two and three was 54 and 50 days, respectively. The drainage was initiated by opening the drainage outlets.

2.2 Greenhouse gas flux measurements

Greenhouse gas fluxes (CO_2 , CH_4) were measured with a closed manual chamber system including an opaque cylindrical polypropylene chamber (d, 15.2 cm; height, 20-39 cm), a non-dispersive infrared gas analyzer for CO_2 (Li 850, Li-cor Environmental, USA) and a laser spectroscopy-based gas analyzer for CH_4 (LGR $\text{N}_2\text{OMI-919}$, ABB Group, Switzerland). The measurement system included a controlling unit with a tablet and a Raspberry Pi (Model 3B, Raspberry Pi Ltd., UK) computer running a custom-made Python program, enabling on-line monitoring of the gas mixing ratio development in the chamber headspace. The gas analyzers were installed in parallel in the sample line to prevent problems stemming from their pumps operating at different flow rates. The volume of the used chamber varied according to the plant height and ranged from 3.6 to 7.2 dm^3 . The air inside the chamber was continuously mixed by a fan installed on the chamber sealing and the temperature was continuously monitored using a Pt-100-temperature probe and a Nokeval RMD 680 logger (Nokeval, Finland). The measurements were conducted weekly or bi-weekly during the off-seasons and bi-weekly during growing seasons.

On a measurement day each monolith was measured once for ~ 7 minutes. The junction between the chamber and the monolith was sealed with plastic foil to make the chamber airtight. If any errors or leakages were observed during the closure, the measurement was repeated. The distance of the soil from the chamber edge was measured at least monthly. During the third study cycle, an empty chamber was measured before (and sometimes after) each measurement round to quantify any potential baseline fluxes. Erroneous closures, identified by fluctuations in CO_2 mixing ratio, were excluded from further analysis. For CH_4 , the empty chamber flux did not deviate from zero (Wilcoxon signed-rank test, $p > 0.05$) but the median of the empty chamber flux for CO_2 was $1.25 \text{ mg m}^{-2} \text{ h}^{-1}$ (Q_1 , 0.34; Q_3 , 2.86; n , 25;). Accordingly, the empty chamber CO_2 flux was deducted from the measured CO_2 fluxes.

Gas fluxes were calculated by fitting a linear model to the gas mixing ratio data plotted against the closure time. The model fitting was conducted with a least squares method for the gas concentration change over 6 minutes. The first 30–45 seconds of

210 the chamber closure were omitted to avoid errors caused by the system instabilities and pressure variations at closure (Pavelka et al., 2018). Linear fits of CO₂ mixing ratios against elapsed time were initially examined visually. Clear deviations from linearity were interpreted as evidence of leakage, typically caused by improper chamber insertion. Erroneous measurements were excluded by applying an R2 threshold of 0.84. This threshold effectively removed all failed closures in cases where CO₂ flux exceeded three times the standard deviation of the flux measured in the empty chamber. For lower fluxes, R2 values were
215 generally below 0.84 even when the measurement was successful. These instances were therefore evaluated individually and excluded only when the CO₂ mixing ratio exhibited evident fluctuations indicative of leakage (0.4% of the total data). The fluxes using the slope term dC/dt from the linear fit were calculated with the equation 1:

$$F = \frac{dC}{dt} \frac{M_{\text{gas}} p V}{RTA} \cdot 3600 \quad (1)$$

where F is the gas flux ($\text{g m}^{-2} \text{h}^{-1}$), M_{gas} molar mass of the gas (g mol^{-1}), p air pressure (101325 Pa), V chamber volume (m^3),
220 R ideal gas constant ($8.314 \text{ J K}^{-1} \text{ mol}^{-1}$), T air temperature (K) and A is a monolith surface area (m^2). Positive flux indicates an efflux of gas to the atmosphere.

As only dark chamber measurements were conducted, total ecosystem respiration, including CO₂ originating from both soil and plant components, was obtained. To estimate the fraction of CO₂ deriving specifically from the soil, aboveground cover crop biomass was removed 43 days after waterlogging during the third study cycle. CO₂ fluxes were measured twice on the
225 same day, immediately before and after biomass removal. The difference between the two measurements was assumed to represent respiration from aboveground plant tissues. This approach was applied to ensure that the observed differences between the cover crop and no cover crop treatments did not derive simply from the variation in aboveground biomass.

2.3 Dissolved C and Fe in soil porewater

Soil porewater samples were collected with Rhizon CSS samplers (pore size 0.15-0.2 μm , Rhizosphere Research Products
230 B.V., The Netherlands) from three treatment replicates (monitored monoliths). At each sampling, needles (0.8x40 mm, Becton Dickinson, S.A., Spain) were inserted at the end of the samplers to connect them to 12 ml evacuated glass vials (Labco Limited, England) through rubber septa. Porewater samples were collected only from the monoliths belonging to the saturation treatment because suction was not adequate for sampling the monoliths maintained at soil moisture $< \text{FC}$. In the third cycle, samples were collected only during the waterlogging treatment, whereas in the second cycle, sampling was conducted both
235 during the treatment and after the start of the drainage, with the final sampling done 16 days after. Total dissolved C (TDC), DOC and dissolved inorganic carbon (DIC) concentrations were measured with combustion catalytic oxidation method (TOC-L, Shimadzu Corporation, Japan) and total dissolved Fe concentration was measured colorimetrically with a 1,10-phenantroline method (Hill et al., 1978, see Kronberg et al., 2024). Samples were stored at $-20 \text{ }^\circ\text{C}$ before the analysis.

2.4 Data analysis

240 Data analysis was conducted in R studio software environment with R version 4.4.3 (R Core Team, 2025). The impact of soil
type as well as plant and water treatments on average CO₂ fluxes were tested with linear mixed effects model in glmmTMB
package (Brooks et al., 2017). The effects were tested separately for the two study cycles. Monolith and measurement round
were set as a random intercept to account for the repeated measurements and the non-independency of the data. The flux data
was further divided into two temporal subgroups, during and after the water treatment (but before the next cultivation), and
245 the model was built separately for these time intervals due to their different flux dynamics. The models were first constructed
with the likelihood method (ML), and the selection of interaction terms to include in the final models was done based on
Akaike's information criteria (AIC) with stepAIC function and backwards method in MASS package (Venables and Ripley,
2002). Dropping interaction terms mostly improved the AIC apart from the period after waterlogging in cycle 3 for which the
interaction between water and plant treatment was included. All main terms (soil type, water and plant treatments) were always
250 included in the final model which was constructed with the restricted maximum likelihood method (REML). The assumptions
of normality and homoscedasticity of the model error terms were inspected visually in all analyses. The significance level in
all statistical analyses was set to 0.05.

To calculate cumulative CO₂ fluxes, we first generated daily flux values for each monolith by linearly interpolating between
temporally consecutive measurement events using the na.approx function in the zoo package (Zeileis and Grothendieck, 2005).
255 Next, the cumulative sum was calculated starting either from crop cultivation (beginning of the season) or from the beginning
of the waterlogging treatment and calculated until next cultivation. The cumulative CO₂ fluxes calculated over the entire cycles
were also further normalized for cumulative above ground biomass. The effect of soil, water and plant treatment on cumulative
CO₂ fluxes was tested with linear model separately for the study cycles two and three. The assumptions of normality and
homoscedasticity of the model residuals were visually evaluated by diagnostic plots. Biomass normalized cumulative CO₂
260 fluxes did not fulfill the assumption of residual normality and hence, Box-Cox transformation was applied (Box and Cox,
1964). Tukey's post hoc tests were further applied to test the differences in between the treatments by using emmeans function
in emmeans package (Russell, 2023). The comparison was done among the levels of water and plant treatments for each soil
type and study cycle.

We also tested how well could a simple empirical model predict the CO₂ efflux during and after waterlogging events. CO₂
265 efflux was modelled in the monitored monoliths with an empirical equation utilizing an exponential temperature function
(Kätterer et al., 1998) combined with a parabolic WFPS function (Luo and Zhou, 2006; Moyano et al., 2013) as follows:

$$F_{CO_2} = Q_{10}^{\frac{T-10}{10}} \cdot (-a \cdot WFPS^2 + b \cdot WFPS) \quad (2)$$

F_{CO_2} is the soil CO₂ efflux (g m⁻² d⁻¹), Q_{10} is the CO₂ temperature sensitivity factor, T is the measured soil temperature (°C) at
10 cm depth, $WFPS$ is the water-filled pore space (%) calculated as the proportion of the measured SVWC at 10 cm from the

maximum SVWC ($SVWC_{max}$) in each monolith belonging to the saturation treatment. In <FC treatment, an average of the
 270 $SVWC_{max}$ in the saturated monoliths in each soil type was used. In the equation, a , b and Q_{10} are parameters fitted for the two
 soils and two plant treatments separately. Residual standard error, adjusted R^2 + slope from the linear fit between the modelled
 and measured fluxes as well as the model residuals were used to evaluate the model performance. The constructed models
 were used to calculate the daily CO_2 flux for each monitored monolith with daily average values of soil T and $WFPS$ ($n=3$).
 275 Obtained fluxes were used to calculate the cumulative CO_2 fluxes similarly than with measured values starting from the
 beginning of the water treatment until next cultivation. The difference in cumulative fluxes calculated based on measured vs.
 modelled values was tested with pairwise t-test.

To study the relationship between dissolved C and Fe concentrations in soil porewater, repeated measures correlation analysis,
 where monolith was set as a participant/subject, was performed with rmcrr package (Bakdash & Marusich, 2024). Then, the
 concentrations of dissolved C species in soil pore water (c , $mg\ l^{-1}$) were further converted to the amount of C per liter soil (mg
 280 l^{-1} soil). This was done to eliminate the apparent effect of SVWC on soil C contents in changing moisture conditions. In each
 monolith, the amount of accumulated DIC, i.e., the change in soil DIC storage (ΔDIC , $mg\ C\ m^{-2}$), during waterlogging was
 calculated with the equation 3.

$$\Delta DIC = \frac{\sum_{Layer=1}^3 V \cdot (\theta_{end} \cdot c_{end} - \theta_{start} \cdot c_{start})_{Layer}}{A} \quad (3)$$

In the equation, V , is the volume of each soil layer ($3.54\ dm^3$), A is the monolith surface area ($0.018\ m^2$), c_{end} and c_{start} are the
 soil porewater DIC concentrations ($mg\ l^{-1}$) at the end and at the first sampling of the waterlogging period, respectively, and
 285 θ_{end} and θ_{start} are the corresponding volumetric soil water contents. In cases where the c_{start} was not available for a specific soil
 monolith (no porewater obtained), the average of the first sampling round in each soil type and depth calculated based on
 available monoliths was used. The equilibrium porewater CO_2 concentration at the start was assumed to be similar within the
 same soil type and depth. The concentration was also assumed to be similar within each 20 cm soil layer around the sampling
 depth.

290 Characteristic of Finnish soils, neither of the studied soils contained significant amounts of carbonates (verified with 0.2 M
 HCl, “Fizz test”) and thus, ΔDIC was assumed to consist mainly of dissolved CO_2 deriving from soil respiration. Thus, the
 total amount of respired CO_2 during the waterlogging was estimated by adding the ΔDIC to the cumulative CO_2 fluxes from
 each monolith. Only the waterlogging period from which porewater samples were collected was considered (Cycle 2:
 waterlogging days 0-49; cycle 3: days 7-45). In <FC treatment, only the cumulative CO_2 flux was considered because no
 295 porewater was obtained and the ΔDIC was assumed to be negligible because of the gas exchange with the atmosphere. The
 effect of time (days), soil type, plant treatment and depth on the concentration of TDC, DIC, DOC and total CO_2 production
 were tested with linear mixed effects model and nlme package (Pinheiro et al., 2023). A stepwise (backwards) model selection
 was performed with stepAIC function in MASS package (Venables and Ripley, 2002). The effect of waterlogging on total

respired/produced CO₂ (in comparison to the <FC treatment) was further tested by pairwise comparisons by emmeans function
300 (Russell, 2023). The comparison was done separately for the two cycles within soil type and plant treatment.

3 Results

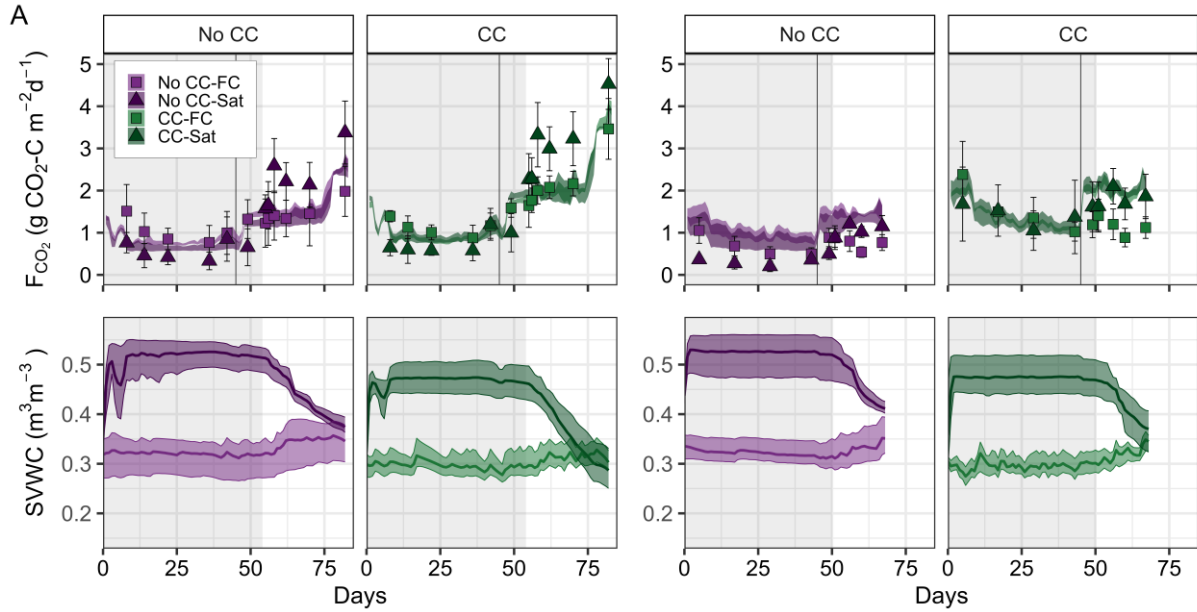
3.1 Measured CO₂ and CH₄ fluxes

During the waterlogging treatment the CO₂ fluxes were significantly ($p < 0.01$) lower in the saturated soil monoliths than in the control monoliths kept below FC (Fig. 2, Table A1). CO₂ fluxes from the saturated monoliths without cover crops were mostly
305 around (cycle 2) or below (cycle 3) $0.5 \text{ g C m}^{-2} \text{ d}^{-1}$, and fluxes in the monoliths with cover crop were significantly higher (+
0.18 and + 0.87 $\text{g C m}^{-2} \text{ d}^{-1}$ in cycles 2 and 3, respectively (Table A1). Upon drainage, the fluxes from the saturated monoliths increased to significantly higher level than in the <FC treatment ($p < 0.001$, Fig. 2). The observed CO₂ pulse upon drainage was steeper in sandy loam compared to the silty clay soil (Fig. 2). After the CO₂ pulse, the differences between the two water treatments levelled out within two weeks in sandy loam soil. In silty clay soil, CO₂ fluxes from the previously saturated
310 monoliths remained higher than from the non-saturated treatment until the next cultivation which was four weeks after drainage.

Cycle 2: 9 March - 31 May 2022

Cycle 3: 5 Sep - 13 Nov 2022

Silty clay:



Sandy loam:

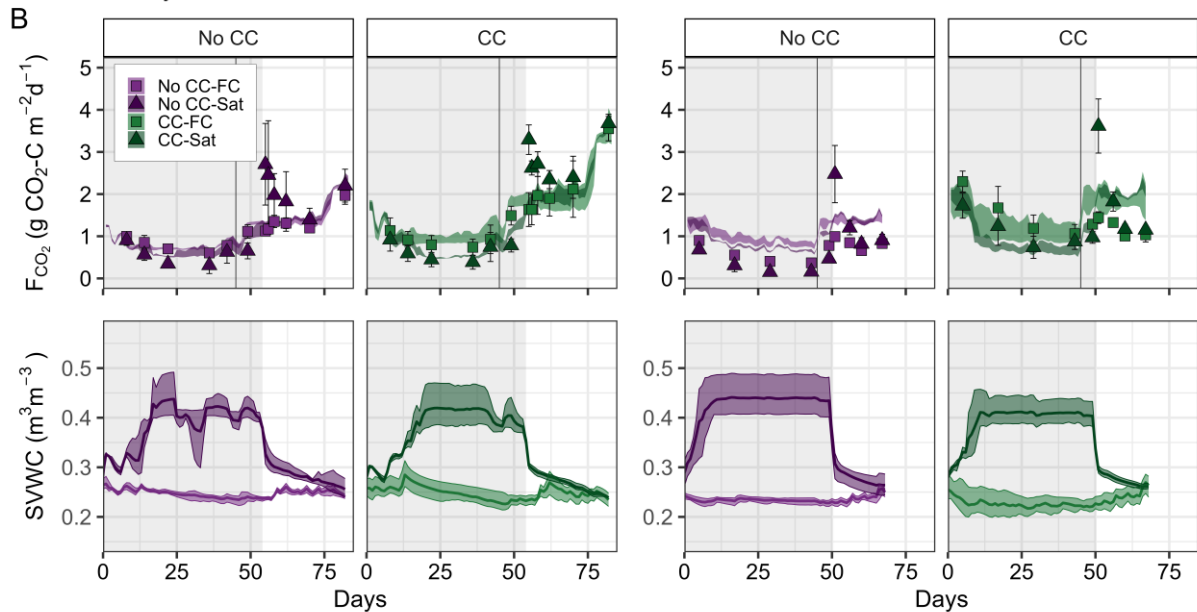


Figure 2. Measured (mean \pm standard deviation, $n=4$) and modelled (shaded ribbons, range, $n=3$) CO₂ efflux and soil volumetric water content at 10 cm depth (SVWC, mean \pm standard deviation, $n=3$) during and after waterlogging in A. silty

315 clay and B. sandy loam soil during study cycles two and three CC, Cover crop; No CC, No cover cro ; FC, < field capacity; Sat, Water saturation). The gray shaded area represents the duration of the waterlogging event and the vertical solid line the day when the soil temperature was increased from 5 °C to 10 °C for simulated spring. The x-axis represents the number of days since the onset of the respective waterlogging event.

Waterlogging did not have a significant impact on off-season cumulative CO₂ fluxes in either of the study cycles, plant
320 treatments or soils (Fig. 3). In the third cycle, however, the mean cumulative CO₂ fluxes in the saturated soils without the cover crop (silty clay 34 ± 5.0 g C m⁻², sandy loam 40 ± 4.9 g C m⁻²) were slightly lower than in the <FC treatment (silty clay 49 ± 11.3 g C m⁻², sandy loam 46 ± 7.0 g C m⁻²). In turn, according to the linear model, the cover crop significantly increased the cumulative fluxes (p<0.01), while the soil type did not have a significant impact on them (p=0.06 or 0.87 in cycle two and three, respectively). The difference between the two plant treatments was larger in the third than in the second cycle (Fig. 3)
325 and the post hoc tests revealed that in the second cycle the difference between the CC and No CC monoliths was barely insignificant in both water treatments in sandy loam soil (p=0.08/0.09) and in the <FC treatment in silty clay (p=0.09). The treatment-wise pattern in cumulative CO₂ fluxes over the entire study cycles (growing and off-seasons combined) remained consistent with those calculated for the off-seasons alone, but only when the fluxes were normalized for above-ground biomass (Fig. 3). Finally, cumulative CO₂ fluxes from the last off-season were also normalized per root biomass determined at the end
330 of the experiment (see Kronberg et al. 2024 for details of root biomass determination). Then, the cumulative fluxes in the monoliths without the cover crop were 3.7 to 6.0 times higher than the fluxes in the monoliths with the cover crop under the corresponding soil type and water treatment (Fig. S2).

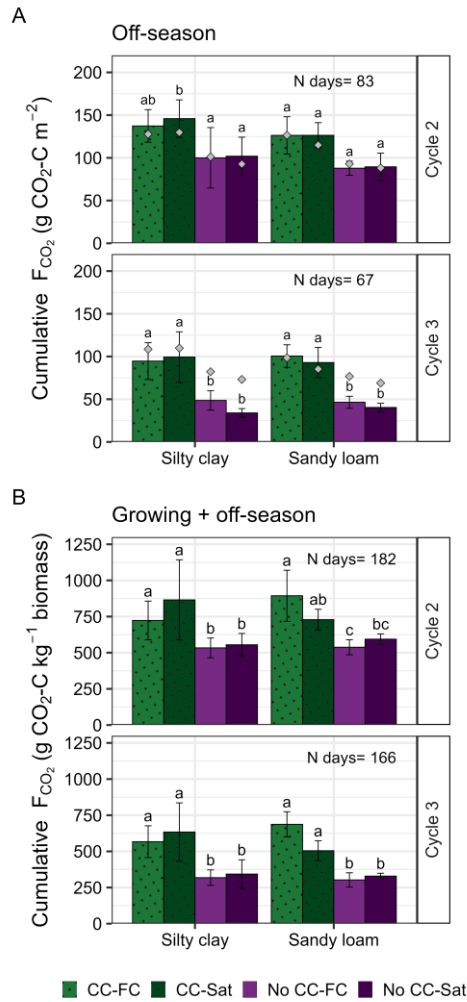


Figure 3. Cumulative CO_2 fluxes (as $g\ CO_2-C$) calculated for the off-season starting from the initiation of the waterlogging treatment until the next cultivation (Cycle 2) or until the end of the experiment (Cycle 3) (A), and for the entire study cycle (Cycle 2, Cycle 3) normalized to the cumulative above ground biomass (kg^{-1} aboveground biomass) (B). Mean \pm standard deviation ($n=4$) of the replicated monoliths (CC-FC, Cover crop & <field capacity; CC-Sat, Cover crop & Water saturation; No CC-FC, No cover crop & <field capacity; No CC-Sat, No cover crop & Water saturation). Gray squares illustrate the mean ($n=3$) cumulative flux from the empirical temperature and moisture dependency model. *N days* represents the number of days included in the calculation of the cumulative sums, and lower-case letters the statistical differences within each soil type and cycle.

In the <FC treatment, the amount of above ground cover crop respiration, determined as the difference between the measured fluxes before and after above ground biomass removal, was on average $0.6 \pm 0.42\ g\ CO_2\ m^{-2}\ h^{-1}$ and $0.5 \pm 0.55\ g\ CO_2\ m^{-2}\ h^{-1}$

in silty clay and sandy loam monoliths, respectively. This represented 15% and 12% of the total CO₂ flux in silty clay and
 345 sandy loam, respectively. However, the averages were not statistically different from zero in either of the soils. Instead of a
 decrease, the above ground biomass removal induced an increase in CO₂ fluxes from saturated monoliths. In silty clay soil, the
 fluxes increased on average $0.6 \pm 0.88 \text{ g m}^{-2} \text{ d}^{-1}$ while the increase in sandy loam soil was modest, on average only 0.1 ± 0.59
 $\text{mg m}^{-2} \text{ d}^{-1}$. However, because of the large deviation between the replicate monoliths, the increases did not deviate from zero
 significantly.

350 3.2 CO₂ efflux modelled with the empirical model

In the empirical temperature and moisture dependency model fit to our data, soil temperature dominated over moisture as the
 predictor for CO₂ efflux as indicated by the high Q_{10} and low a and b parameter values (Table 2). The modelled fluxes were
 very similar in the waterlogged and in <FC treatment especially in silty clay soil (Fig. 2). The CO₂ flux dynamics and response
 to soil moisture were better captured (Fig. 2) and the model performance was overall better in sandy loam soil which is
 355 illustrated by the smaller RSE of the model and a larger R² of the linear model fitted for the predicted vs. measured data than
 in silty clay soil (Table 2). In both soils, model performed better with the cover crop than without the cover crop as indicated
 by higher R² (Table 2).

Towards the end of the waterlogging, the model overestimated the fluxes in the saturation treatment. In turn, the model could
 not reproduce the CO₂ peak measured after the initiation of drainage which led to underestimation of the fluxes (positive
 360 residuals) upon drainage in the monoliths belonging to the saturation treatment (Fig. 2, Fig B1). During the third waterlogging,
 the model overestimated the CO₂ fluxes in the monoliths without the cover crop (Fig. 2). In cycle two, the cumulative CO₂
 efflux calculated with modelled data was in a good agreement with those calculated from the measured data with no statistically
 significant difference between the two (Fig. 3, Table S3). During cycle three, the same was true only for the monoliths with
 the cover crop. Without cover crops, cumulative CO₂ efflux obtained from the modelled CO₂ fluxes was significantly higher
 365 in both soils and water treatments (Fig. 3, Table S3).

Table 2. Results of the temperature (T) and water filled pore space (WFPS) dependency model fit to CO₂ flux data from the
 monitored monoliths during the off-seasons. Also, the adjusted R², slope and intercept from the linear fit between the modelled
 and measured fluxes, used to evaluate model performance, are presented.

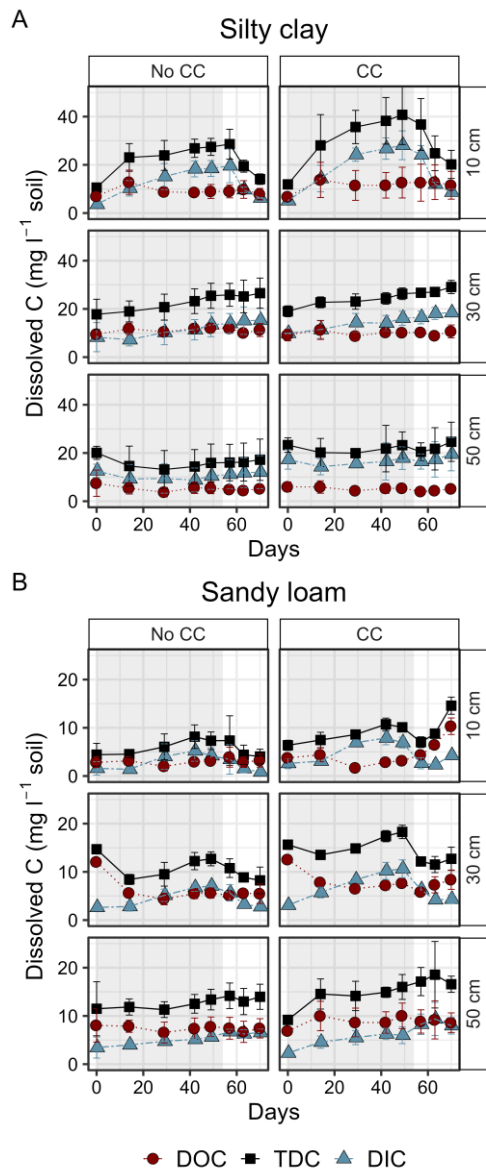
		FCO ₂ ~ T, WFPS					Predicted ~ Measured		
		RSE	Q10	a	b	N	Intercept	Slope	R ²
No cover crop	Silty clay	2.24	3.49	0.0009	0.13	145	2.73	0.38	0.39
	Sandy loam	1.88	3.31	0.0010	0.13	150	2.42	0.41	0.45
Cover crop	Silty clay	2.22	4.79	0.0008	0.14	150	2.58	0.60	0.59
	Sandy loam	1.93	3.79	0.0018	0.21	150	2.14	0.65	0.66

370 RSE=Residual standard error of the model ($\text{g C m}^{-2} \text{d}^{-1}$), Q_{10} =fitted CO_2 flux temperature sensitivity parameter, a & b = fitted model parameters, N=number of observations, Intercept=intercept of the linear regression fitted for modelled and measured CO_2 fluxes, Slope=slope of the linear regression, R^2 =adjusted coefficient of determination of the linear regression.

3.3 Dissolved C dynamics in porewater

375 Total dissolved C (TDC) content increased during waterlogging at 10 and 30 cm depths in both soils and under both plant treatments (Fig. 4). The observed increase was attributed to the increased dissolved inorganic carbon (DIC) content while dissolved organic carbon (DOC) content remained relatively unchanged. The increases in TDC and DIC contents during waterlogging at 30 cm depth were more pronounced in sandy loam than in silty clay, whereas especially in the second cycle the increase in DIC content in the topsoil was steeper in silty clay (Table A5 & S6, Fig. 4). In sandy loam, waterlogging induced an increase in DIC content also at 50 cm depth (Fig. 4).

380 In the topsoil the contents of all dissolved C species (TDC, DIC, DOC) were significantly higher in silty clay than in sandy loam soil as illustrated by the significant soil term (for TDC, DIC, DOC) as well as the significant interaction of soil and waterlogging days (for TDC, DIC) in the mixed effects models (Table A4-6). Cover crops increased the TDC content in the silty clay topsoil while in sandy loam the increase was more pronounced at 30 cm depth (Fig. 4). In the third cycle, the average DOC content at 50 cm depth was significantly higher in the monoliths with the cover crop than without in sandy loam soil ($p=0.01$) (Fig. B2), while there was no statistically significant difference during the second cycle (Table A6).



385

Figure 4. Timeseries of dissolved organic carbon (DOC), inorganic carbon (DIC) and total dissolved carbon (TDC) contents as mg C l^{-1} soil (mean \pm standard deviation, $n=3$) during (shaded area) and after waterlogging in silty clay (A) and sandy loam (B) soil profiles with and without cover crops (No CC, No cover crops; CC = Cover crops) in cycle two. Days on the x-axis represent the days since the beginning of waterlogging. Note the differing scales on y-axis in the two soils. Data from cycle 390 three is presented in supplementary Fig. S1.

During the second off-season, porewater TDC concentration correlated with dissolved Fe concentration at 10 and 30 depths in silty clay soil both with (10 cm: $r= 0.51$, $p=0.023$; 30 cm: $r=0.58$, $p=0.005$) and without the cover crop (10 cm: $r= 0.46$, $p=0.049$; 30 cm: $r=0.58$, $p=0.006$), and in sandy loam with the cover crop (10 cm: $r= 0.58$, $p=0.009$; 30 cm: $r=0.59$, $p=0.07$) (Table A9). In silty clay soil, the correlation with dissolved concentration was attributed to the changes in DIC concentration in both plant treatments (Fig. 5A), whereas in sandy loam Fe concentration correlated with DOC instead, however, only in the topsoil (Fig. 5D). During the third cycle the porewater C and Fe concentrations were only measured during the waterlogging (excluding the drainage phase) and then, the DOC concentration did not exhibit positive correlation with Fe in either soil at any depth (Table A9). During this cycle, the correlation between DIC and Fe concentrations, in turn, was significant in both soils with the cover crop at 30 cm (silty clay: $r=0.88$, $p<0.001$; sandy loam: $r=0.71$, $p=0.022$) and in silty clay at 10 cm with the cover crop ($r=0.75$, $p=0.013$) (Table A9).

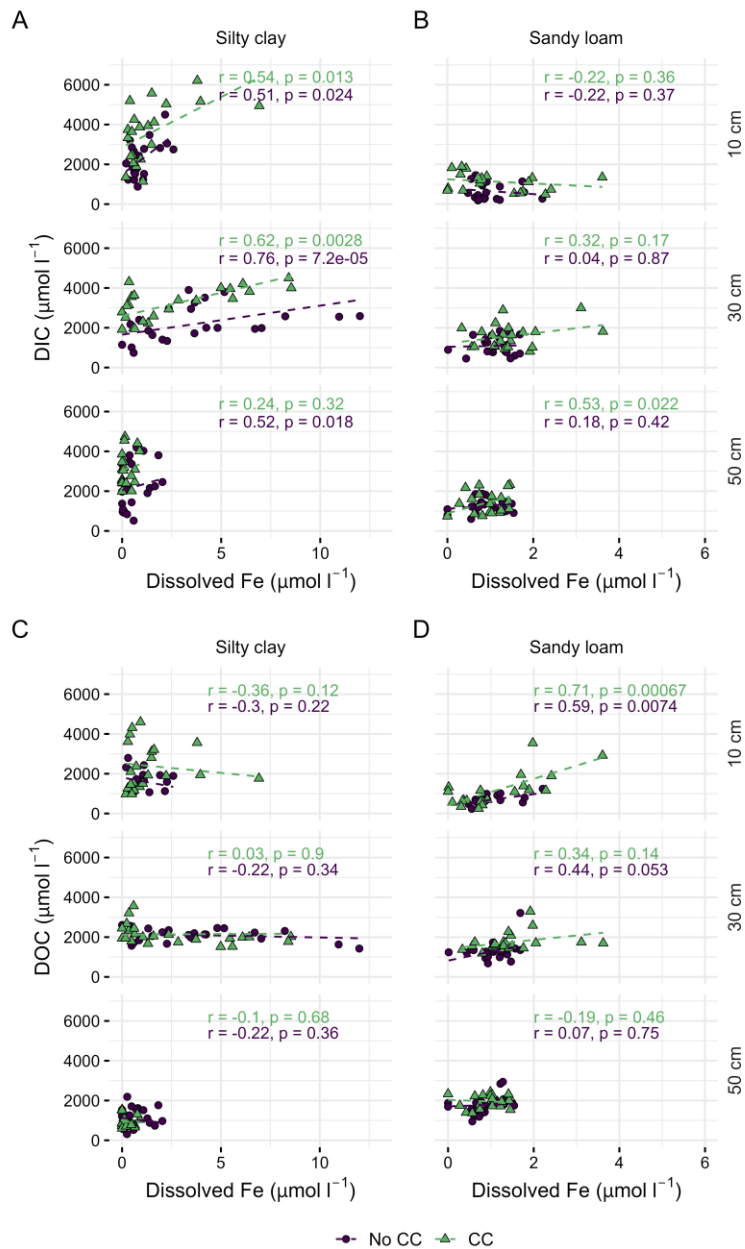


Figure 5. Relationship between porewater dissolved Fe and dissolved inorganic carbon (DIC) (A-B) and dissolved organic carbon (DOC) (C-D) in the soils belonging to the saturation treatment, and with (CC) and without the cover crop (No CC) at different depths. The regression lines, and the correlation coefficients (r) together with the corresponding p -values were obtained from repeated measures correlation analysis. Observe the differing x-axis scales in the two soils.

3.4 Total CO₂ production as the sum of cumulative CO₂ fluxes and ΔDIC

Results from the linear model showed that waterlogging decreased the total CO₂ production (cumulative CO₂ flux + ΔDIC) statistically significantly during the second (p=0.002) but not during the third (p=0.10) waterlogging (Fig. 6, Table A7). However, according to the pairwise comparisons, the means of the two water treatments within each soil type and plant treatment were only statistically significantly different in silty clay soil in cycle two without the cover crop (Fig. 6, Table A8). In the third cycle CO₂ production was statistically significantly higher with the cover crop than without (+35 g C m⁻², p<0.01) while there was no difference in the second cycle (p=0.1). Soil type was significant in cycle two where the total CO₂ production during waterlogging was lower in sandy loam than in silty clay (-8.3 g C m⁻², p=0.05) (Fig. 6). Soil type was not significant in cycle three.

415 The amount of the accumulated DIC in the saturated monoliths corresponded to on average 6–29% of the total CO₂ production during waterlogging. In the second cycle, the percentage was on average 11.5 ± 5.7% (No CC) and 13.5 ± 0.4% (CC) in silty clay, and 6.0 ± 1.7% (No CC) and 8.8 ± 3.9% (CC) in sandy loam. In the third cycle, the percentage was larger in No CC treatment (silty clay 28.8 ± 2.5%, sandy loam 19.8 ± 2.6%) but it remained at a similar level in the CC treatment (silty clay 9.4 ± 1.4%, sandy loam 8.9 ± 2.4%).

420

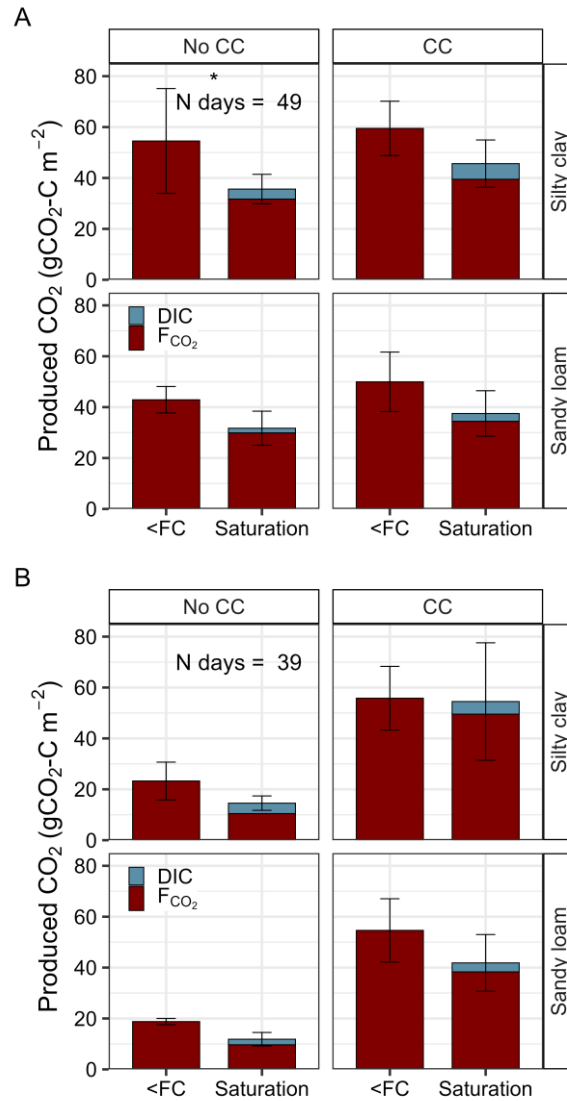


Figure 6. Total produced CO₂-C as the sum of cumulative CO₂ flux and accumulated DIC in soil pore water during the second (A) and third (B) waterlogging in silty clay and sandy loam soil with (CC) and without (No CC) cover crops. Statistically significant differences between the two water treatments are marked with asterisks.

4 Discussion

4.1 Impact of waterlogging on measured CO₂ and CH₄ fluxes

We studied the impact of periodic off-season waterlogging on soil C dissolution, and CO₂ emissions in a controlled mesocosm study with intact soil profiles with and without Tall Fescue as a cover crop. In our experiment, temporary soil waterlogging lasting ~50 days at an average soil temperature of ~7 °C did not significantly alter cumulative CO₂ emissions calculated over waterlogging and subsequent drainage period. Our results align with findings from several incubation studies suggesting that temporary anaerobic conditions may exert only a minor impact on C decomposition and cumulative greenhouse gas emissions (e.g. Bhattacharyya et al., 2018; Hanke et al., 2013; Huang et al. 2021; Tete et al., 2015; Winkler et al., 2019). However, previous studies have reported effects in both directions, depending on soil characteristics, and the nature and duration of redox conditions.

While static anoxic conditions have suppressed C losses in several studies (Fairbairn et al., 2023; Huang et al., 2020; Tete et al. 2015), as traditionally believed, oxic-anoxic fluctuations have led to equal or greater C losses in the form of dissolved organic C (DOC), CO₂ and CH₄ (Huang et al. 2021; Jeanneau et al., 2020; Tete et al., 2015). In the laboratory incubations by Huang et al. (2021) and Tete et al. (2015), reduced decomposition during anaerobic period(s) was compensated by increased CO₂ efflux during the following aerobic period(s) in weekly to monthly timescale. This pattern is consistent with our results: the cumulative CO₂ fluxes remained unaffected by waterlogging but the temporal flux dynamics differed between the two water treatments. The reduced CO₂ fluxes during waterlogging were compensated for by the higher fluxes after the drainage in both soils. Based on these observations, during transiently anaerobic events, most CO₂ fluxes tend to occur during the subsequent drainage/aerobic phase which makes the calculation of cumulative emissions sensitive to the chosen observation period. Thus, our results underscore the importance of considering the following drying period in assessing the impacts of waterlogging on soil CO₂ efflux under fluctuating moisture conditions.

According to Gin et al. 2017, repeated anoxic-oxic cycles increase soil Fe reducibility, which may, in turn, enhance C availability and promote C mineralization. Among the soils used in our experiment, the silty clay soil had experienced fluctuating redox conditions in field as indicated by its stagnic properties. It also contained a higher Fe oxide content than the sandy loam. Therefore, we expected that waterlogging would enhance C availability and mineralization more strongly in silty clay soil. Yet, contrary to our hypothesis, the cumulative CO₂ fluxes (calculated over the waterlogging and the following drainage period), and their response to waterlogging, were similar in the two soils. Only a slight difference between the two soils was observed in CO₂ production during waterlogging in the second study cycle. This lack of clear differences between the two soils contradicts with the higher DOC and DIC contents measured in the silty clay than in the sandy loam topsoil during waterlogging, implying a higher mineralization rate in the silty clay during both study cycles. The dissolved C dynamics in deeper soil layers, however, help to explain the lack of difference in the aggregated CO₂ fluxes from the whole profile. The

marked increase in DIC content at 30 cm depth in the sandy loam soil, and a more modest increase in the silty clay suggests that while CO₂ production was lower in the sandy loam topsoil compared to silty clay, higher microbial activity at 20–40 cm
460 depth in sandy loam increased the total respiration per area, thereby narrowing the difference in cumulative CO₂ emissions between the two soils. Most studies on soil C dynamics focus exclusively on changes in the top 0–20 cm soil layer (Poeplau and Don, 2015). However, it has been demonstrated that examining the entire soil profile can yield conclusions that greatly differ from those based solely on topsoil analysis (Tautges et al., 2019). Thus, our findings emphasize that C dynamics in deeper soil layers may play a significant role in overall C dynamics and should therefore not be overlooked in future studies.

465 Previous incubation studies have shown that methane production can contribute substantially to soil C mineralization under anaerobic conditions in mineral soils (e.g. Huang et al. 2021). Although CH₄ production was not in the focus of this study, it warrants a mention that methane was also measured and waterlogging did not induce CH₄ production in either soil. This aligns with the measured soil reduction-oxidation (redox) potentials (reported in our previous paper), whose lowest average values were -110 mV and -60 mV in silty clay and sandy loam soil, respectively (Kronberg et al., 2024). This indicates that conditions
470 did not reach the strongly reducing levels associated with methane production (Reddy et al., 2022

4.2 Belowground C dynamics during waterlogging

4.2.1 Waterlogging causes a discrepancy between respired CO₂ and measured flux

The response of CO₂ efflux to soil moisture reflects not only the moisture sensitivity of heterotrophic respiration but also the associated effects on gas transport. When soil pores become filled with water, diffusive gas movement is substantially impeded
475 (Greenway et al., 2006). Accordingly, respired CO₂ becomes trapped within water-filled soil pores, leading to a discrepancy between CO₂ production and measured soil CO₂ efflux (Jeanneau et al., 2020; Maier et al., 2011, 2010; Sánchez-Cañete et al., 2018). In our study, increasing soil DIC concentrations—reflecting the amount of CO₂ retained within soil pores—indicated that during waterlogging, C mineralization continued at a higher rate than would have been inferred from CO₂ efflux measurements alone.

480 Our observations demonstrate that, contrary to convention, momentary soil CO₂ efflux should not be considered equivalent to soil respiration as the two are decoupled in periodic high soil moisture events (Maier et al., 2011; Ryan and Law, 2005; Sánchez-Cañete et al., 2018). In the long term, the two can usually be considered equal as all respired CO₂ will eventually be released into the atmosphere (Maier et al., 2011). However, momentary fluxes can significantly deviate from respired CO₂ (Bond-Lamberty et al., 2024; Maier et al., 2011, 2010; Sánchez-Cañete et al., 2018). In our experiment, accumulated CO₂
485 (Δ DIC) during waterlogging was at most ~30% of the total CO₂ production (sum of cumulative CO₂ fluxes and Δ DIC) being in a good agreement with previously reported values (Maier et al., 2011; Sánchez-Cañete et al., 2018).

The discrepancy between CO₂ production and CO₂ efflux measured at the soil surface caused the empirical CO₂ efflux model to fail in simulating the flux dynamics and capturing the post-drainage CO₂ pulse. As a result, the model was unable to

reproduce the CO₂ pulse upon drainage because the pulse did not represent momentary soil respiration but was largely a
490 consequence of the release of previously respired CO₂. Thus, the observed decrease in CO₂ efflux likely stems from a decreased
gas transport rather than production only. These results support previous evidence that simple models may underestimate the
CO₂ production during water saturated, anaerobic conditions (e.g. Fairbairn et al., 2023).

To account for the difference in measured CO₂ efflux and soil respiration, Maier et al. (2011, 2010) and Hirsch et al. (2004)
have proposed an incorporation of a storage flux term in soil moisture dependency models. They defined the storage flux as
495 the flux resulting from changes in the amount of CO₂ stored in soil (Hirsch et al., 2004; Maier et al., 2011, 2010). We believe
that incorporating a storage flux term, could have also improved model performance in our study. However, a robust
implementation would have required higher spatial resolution of DIC measurements across the soil profile. Overall, to
accurately model C dynamics during transient or temporary waterlogging, process-based approaches that account for anaerobic
CO₂ production (Fairbairn et al., 2023) and changes in gas transport should be considered.

500 4.2.2 Processes contributing to the increase in dissolved C content

We hypothesized that soil waterlogging would promote mobilization of the Fe associated C and therefore result in an increase
in soil DOC content, as commonly observed in incubation studies (e.g. Chen et al., 2020; Huang and Hall, 2017; Pan et al.,
2016; Winkler et al., 2019). As opposed to our hypothesis, soil DOC content did not increase in either soil during waterlogging;
the only clear increase was observed in sandy loam topsoil after drainage, likely due to enhanced OM decomposition and plant
505 root exudation. However, TDC content increased during waterlogging, suggesting that an immediate microbial mineralization
of DOC to CO₂ may have masked any increases in DOC content, with the effect instead becoming apparent as an increase in
DIC (Fairbairn et al., 2023). In addition to mobilization of mineral associated C, in the presence of plant cover, the
accumulating CO₂ (DIC) may have also originated from root respiration or from the decay of root litter (Kuzyakov, 2006).
The above-mentioned processes and their contributions to increased TDC/DIC are discussed next.

510 Our results suggest that the contribution of Fe dissolution and the mobilization of associated C in the overall increase in
TDC/DIC content was likely small. Waterlogging appeared to induce only slight reductive Fe dissolution since the Fe
concentrations in porewater remained low throughout the study (max ~10 μmol l⁻¹) (Kronberg et al., 2024). In silty clay topsoil,
for instance, the highest average DIC concentration during the second cycle (~3 mmol l⁻¹) was three orders of magnitude higher
than that of Fe (~2 μmol l⁻¹). Thus, although soil DIC concentration correlated with Fe concentration during waterlogging,
515 particularly in silty clay soil, the dissolution of Fe-OC alone can not explain the observed increases in DIC/TDC. Vice versa,
the higher dissolved C content, reflecting soil microbial/rhizospheric activity, may have rather enhanced Fe dissolution slightly,
as also seen in previous studies (e.g. Winkler et al. 2019). Overall, substrate availability was likely not substantially enhanced
by waterlogging, as cumulative CO₂ fluxes did not increase relative to the <FC treatment.

Root respiration, including the activity of roots and closely associated microbes, can represent between 10 and 90% of the total soil respiration (Hanson et al., 2000). In our experiment DIC increased only slightly more with the cover crop than without despite the ~10 times higher root biomass at 0–20 cm depth with the cover crop (Kronberg et al., 2024). Thus, if a substantial fraction of the accumulated DIC in soil would have derived from root respiration, we would have expected a larger difference in the produced DIC. In fact, during waterlogging, cover crops had a more pronounced effect on measured CO₂ fluxes than on DIC, and thus, we think that a significant fraction of the CO₂ respired by cover crop roots was released to the atmosphere directly through their stems. Indeed, it has previously been demonstrated that a substantial amount root-derived CO₂ can be transported to the atmosphere via transpiration stream in plant xylem (Aubrey and Teskey, 2021, 2009). Although this mechanism could not be verified with our experimental setup, the transport of CO₂ through Tall Fescue stems seems highly likely, as the species has been shown to develop aerenchyma under waterlogged conditions, enhancing gas exchange between the rhizosphere and the atmosphere (Mui et al., 2021). This feature is specific to Tall Fescue and has not been shown to apply to many other commonly used cover crop species, such as annual ryegrass for example.

If the dissolution of Fe-associated C and root respiration can account only for small portions of the observed increases in TDC, what are the remaining potential sources? In addition to direct release of C from Fe associations, the commonly observed increase in C solubility upon flooding has been attributed to pH-driven OC desorption from mineral phases (Grybos et al., 2009; Pan et al., 2016), dispersion of soil colloids (Buettner et al., 2014) and accumulation of microbial metabolites and fermentation products in anaerobic conditions (Fairbairn et al., 2023; Tete et al., 2015). The circumneutral initial pH of the soils in our experiment resulted in a slight decrease rather than an increase in pH during waterlogging which was likely caused by an accumulation of CO₂ as carbonic acid and bicarbonate (H₂CO₃, HCO₃⁻) (Kronberg et al., 2024). Thus, desorption driven by the pH increase (Grybos et al., 2009) was unlikely. In soil, fermentation and Fe reduction are often coupled (Lovley 2011; Snoeyenbos-West et al. 2000). Had fermentation proceeded at a much higher rate than Fe reduction in our system, we would have expected a more pronounced initial accumulation of organic metabolites and fermentation products, reflected as an increase in DOC content. Finally, we also want to bring up the possibility that O₂ entrapped in soil pores upon water saturation could have formed aerobic microenvironments (Williams and Oostrom, 2000) where aerobic respiration could go-on and contribute to an increased soil DIC and TDC content. Huang & Hall (2017) speculated that anoxic-oxic interfaces may play an important role in the observed increases in C mineralization during waterlogging periods. The porous and heterogenous soil matrix often leads to significant spatiotemporal variability in soil redox conditions (Fiedler, 2000). Thus, despite the low redox potentials measured in our study (Kronberg et al., 2024), aerobic microsites could have also facilitated aerobic instead of anaerobic respiration during waterlogging.

4.3 Impact of the cover crop on C mineralization

We expected that Tall Fescue as a cover crop would alter the response of soil CO₂ production to waterlogging because of increased substrate availability promoting C mobilization. As opposed to our hypothesis, the response of cumulative CO₂

fluxes to waterlogging was similar in both plant treatments. This suggests that cover crop root C inputs did not significantly promote mobilization of OM from mineral phases as reported by e.g. Winkler et al. (2019), although higher dissolved C content in the cover crop treatment appeared to slightly enhance Fe solubility. Despite careful soil moisture monitoring, slight oscillations in soil moisture occurred due to cover crop transpiration in the sandy loam topsoil. This introduced a minor confounding effect on soil redox state, as noted in our preceding study (Kronberg et al., 2024). While we consider the impact on C fluxes to be small, it may have facilitated more efficient gas transport from soil to the atmosphere. Furthermore, the specific feature of Tall Fescue being able to form aerenchyma, which facilitates O₂ transport into the roots, may have mitigated its effects on Fe-associated C by accelerating the drop in soil redox potential less than anticipated (Kronberg et al. 2024). Therefore, these findings should not be generalized to other cover crop species.

The applied methodology does not allow us to distinguish soil respiration from total ecosystem respiration which unfortunately prevents a direct assessment of waterlogging effects on soil versus plant respiration. However, measurements taken during the third waterlogging event, following the removal of aboveground cover crop biomass, showed that cover crop respiration was relatively low (0.6 g C m⁻² d⁻¹) compared to the average difference in CO₂ fluxes between monoliths with and without the cover crop (3.2 g C m⁻² d⁻¹). This indicates that the overall higher CO₂ emissions from the monoliths with the cover were likely driven by belowground processes rather than by aboveground autotrophic respiration. Furthermore, higher cumulative CO₂ emissions with the cover crop persisted when normalized to cumulative above ground biomass further suggesting that the higher fluxes were not resulting from above ground autotrophic respiration. In fact, cover crop biomass was minor compared to barley biomass (~12%, data not shown) while the root biomass was significantly larger under the cover crop treatment as presented in our previous publication (Kronberg et al., 2024).

Normalizing cumulative CO₂ fluxes from the last off-season to root biomass in the top 20 cm of soil showed that the fluxes per gram of root biomass were approximately three times higher without cover crops than with them (Fig. 3). This suggests that although the cover crop increased the CO₂ emissions per m², the plant cover had a net positive effect on soil C sequestration because emissions per soil C input (i.e. root biomass) were smaller. Thus, potential priming effects of the cover crop on OM (not directly measured) did not appear to be substantial enough to offset the positive contributions of the cover crop to soil C storage. Hence, our results tentatively suggest that cover crops had a net positive effect on soil C sequestration, even under periodic waterlogging, which is in line with studies that have promoted cover crops as effective strategy to improve soil C sequestration (Heikkinen et al., 2022; Kaye and Quemada, 2017; Poeplau et al., 2015; Poeplau and Don, 2015).

4.5 Environmental implications and research outlook

Our results suggest that temporary waterlogging outside growing seasons might not significantly affect total CO₂ emissions from cultivated mineral soils in cool, humid climate, which is good news from the climate change perspective. A recent study demonstrated that a substantial fraction of agricultural fields in Finland may suffer from periodic waterlogging during late autumns when transpiration is low and rainfall is high (Mattila and Vihanto, 2024). This highlights the urgent need to apply

mechanistic insights on the effects of temporary waterlogging and anaerobic conditions on C dynamics from laboratory incubations to field scale, with our mesocosm study acting as a crucial bridge between the two. However, our study focused
585 merely on the effects of altered soil moisture on C, CO₂ fluxes and dissolved C dynamics, without accounting for potential impacts on C inputs, soil N dynamics or on production of another potent greenhouse gas, nitrous oxide (N₂O). It is important to recognize that climate change-induced shifts in e.g. off-season soil temperatures and freeze-thaw cycles are also likely to affect greenhouse gas emissions and soil C sequestration capacity (Heikkinen et al., 2022; Liu et al., 2024). Thus, studies that evaluate the net impacts from these factors in an integrated manner are warranted in future.

590 From the methodological perspective, we want to highlight that the changes in soil CO₂ storage during and after high soil moisture events should not be overlooked as neglecting these changes could lead to biased view of momentary soil respiration. In modelling, model-data synthesis is used to match modelled results with the measured fluxes facilitating the calibration of C cycling simulation models (Nevalainen et al., 2022) and enabling the assessment of individual weather parameters' effects on the outcome (Heimsch et al., 2024). The lag between respiration and the release of the respired CO₂ to the atmosphere will
595 make such model-data synthesis challenging, as the measured CO₂ efflux may result from respiration over the preceding weeks. Incorporating the temporary storage of dissolved CO₂ into models would improve the accuracy of the simulation of short-term dynamics and ensure that the respiration terms would be fitted based on the correct time period under when they occur.

We conclude that because surface CO₂ fluxes reflect not only production but also temporary accumulation or release of CO₂ within the soil profile, simple empirical models are unable to capture momentary soil respiration. Moreover, the continued
600 CO₂ production observed under anaerobic, water-saturated conditions supports previous findings that temporary waterlogging does not suppress CO₂ production in mineral soils to near zero. However, despite the important role of short-range-ordered Fe-oxides in C stabilization in boreal agricultural soils, the dissolution and mobilization of associated C did not alone explain the sustained CO₂ production. Furthermore, root C inputs from Tall Fescue, used as an overwintering cover crop, did not promote a substantial release of Fe-associated C, as indicated by low dissolved Fe concentrations and an unaltered soil CO₂
605 efflux response to waterlogging in the presence of plant cover. Overall, our results suggest that off-season waterlogging in cool, humid climate may not significantly affect the cumulative C efflux from mineral soils to the atmosphere.

Author contributions

R.K.: Conceptualization, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration, Visualization, Writing – original draft, Writing – review & editing. **S.K.:** Conceptualization, Methodology, Supervision,
610 Writing – review & editing. **M.P.:** Conceptualization, Methodology, Supervision, Funding acquisition, Project administration, Writing – review & editing. **T.P.:** Methodology, Software. **M.K.:** Methodology, Software, Writing – review & editing. **T.M.:** Supervision, Writing – review & editing.

Acknowledgements

This research was funded by Strategic Research Council at the Research Council of Finland (Multi-benefit solutions to climate-
615 smart agriculture, MULTA) (grant number 337549, 328309, 327236), Research Council of Finland post-doctoral researcher
funding (grant number 339489), Drainage Foundation sr. (grant number H-13-2022-9.1), and Maa- ja Vesiteknikan Tuki ry
(grant number 44875). R.K. acknowledges the University of Helsinki Doctoral Programme in Sustainable Use of Renewable
Natural Resources (AGFOREE) for financial support as a doctoral scholarship. We are grateful for Prof. Jussi Heinonsalo
(University of Helsinki) for his valuable contributions to the experimental conceptualization and planning. We also want to
620 thank Rauna Lilja and Lisa Leinonen for their contributions to establishing and maintaining the experiment. Figure 1 was
created in BioRender. Kronberg, R. (2025) (<https://BioRender.com/pg6cizz>).

Competing interests

There are no competing interests to declare.

Declaration of Generative AI and AI-assisted technologies in the writing process

625 During the preparation of this work R.K. used ChatGPT 4o to improve the readability and language. Sentences were given to
ChatGPT after which R.K. reviewed and edited the content as needed. Authors take full responsibility for the publication
content.

Data availability

The data have been published in Zenodo (<https://doi.org/10.5281/zenodo.14438980>).

630 References

- Asano, M., Wagai, R., Yamaguchi, N., Takeichi, Y., Maeda, M., Suga, H., & Takahashi, Y.: In Search of a Binding Agent: Nano-Scale Evidence of Preferential Carbon Associations with Poorly-Crystalline Mineral Phases in Physically-Stable, Clay-Sized Aggregates, *Soil Syst.*, 2(2), 32, <https://doi.org/10.3390/soilsystems2020032>, 2018.
- 635 Aubrey, D.P., Teskey, R.O.: Xylem transport of root-derived CO₂ caused a substantial underestimation of belowground respiration during a growing season, *Glob. Change Biol.*, 27(12), 2991–3000, <https://doi.org/10.1111/gcb.15624>, 2021.
- Aubrey, D.P., Teskey, R.O.: Root-derived CO₂ efflux via xylem stream rivals soil CO₂ efflux, *New Phytol.* 184, 35–40, <https://doi.org/10.1111/j.1469-8137.2009.02971.x>, 2009.
- Bakdash, J. Z., & Marusich, L. R.: rmcrr: Repeated Measures Correlation, <https://CRAN.R-project.org/package=rmcrr>, 2024
- 640 Bergström, L. Use of lysimeters to estimate leaching of pesticides in agricultural soils. *Environ. Pollut.*, 62(4), 325-347, 10.1016/0269-7491(90)90070-S, 1990.
- Bhattacharyya, A., Campbell, A.N., Tfaily, M.M., Lin, Y., Kukkadapu, R.K., Silver, W.L., Nico, P.S., Pett-Ridge, J.: Redox Fluctuations Control the Coupled Cycling of Iron and Carbon in Tropical Forest Soils, *Environ. Sci. Technol.*, 52, 14129–14139, <https://doi.org/10.1021/acs.est.8b03408>, 2018.
- 645 Bond-Lamberty, B., Ballantyne, A., Berryman, E., Fluet-Chouinard, E., Jian, J., Morris, K.A., Rey, A., Vargas, R.: Twenty Years of Progress, Challenges, and Opportunities in Measuring and Understanding Soil Respiration. *J. Geophys. Res. Biogeosci.* 129, e2023JG007637, <https://doi.org/10.1029/2023JG007637>, 2024.
- Box, G.E.P., Cox, D.R.: An Analysis of Transformations. *J. R. Stat.: Series B (Methodological)*, 26, 211–243. <https://doi.org/10.1111/j.2517-6161.1964.tb00553.x>, 1964.
- 650 Brooks, M.E., Kristensen, K., van Benthem, K.J., Magnusson, A., Berg, C.W., Nielsen, A., Skaug, H.J., Machler, M., Bolker, B.M.: glmmTMB balances speed and flexibility among packages for zero-inflated generalized linear mixed modelling, *The R journal*, 9, 378–400, <https://doi.org/10.3929/ethz-b-000240890>, 2017.
- Buettner, S.W., Kramer, M.G., Chadwick, O.A., Thompson, A.: Mobilization of colloidal carbon during iron reduction in basaltic soils. *Geoderma*, 221–222, 139–145. <https://doi.org/10.1016/j.geoderma.2014.01.012>, 2014.
- 655 Burgin, A.J., Loecke, T.D.: The biogeochemical redox paradox: how can we make a foundational concept more predictive of biogeochemical state changes? *Biogeochemistry*, 164, 349–370, <https://doi.org/10.1007/s10533-023-01036-9>, 2023.

- Buyse, P., Flechard, C.R., Hamon, Y., Viaud, V.: Impacts of water regime and land-use on soil CO₂ efflux in a small temperate agricultural catchment, *Biogeochemistry*, 130, 267–288, <https://doi.org/10.1007/s10533-016-0256-y>, 2016.
- 660 Chandregowda, M.H., Tjoelker, M.G., Pendall, E., Zhang, H., Churchill, A.C., Power, S.A.: Belowground carbon allocation, root trait plasticity, and productivity during drought and warming in a pasture grass, *J. Exp. Bot.*, 74, 2127–2145, <https://doi.org/10.1093/jxb/erad021>, 2023.
- Chen, C., Hall, S.J., Coward, E., Thompson, A.: Iron-mediated organic matter decomposition in humid soils can counteract protection, *Nat. Commun.*, 11, 2255, <https://doi.org/10.1038/s41467-020-16071-5>, 2020.
- 665 Clemmensen, K.E., Bahr, A., Ovaskainen, O., Dahlberg, A., Ekblad, A., Wallander, H., Stenlid, J., Finlay, R.D., Wardle, D.A., Lindahl, B.D.: Roots and Associated Fungi Drive Long-Term Carbon Sequestration in Boreal Forest, *Sci.*, 339, 1615–1618, <https://doi.org/10.1126/science.1231923>, 2013.
- Cotrufo, M.F., Ranalli, M.G., Haddix, M.L., Six, J., Lugato, E.: Soil carbon storage informed by particulate and mineral-associated organic matter, *Nat. Geosci.*, 12, 989–994, <https://doi.org/10.1038/s41561-019-0484-6>, 2019.
- 670 Davidson, E.A., Janssens, I.A., Luo, Y.: On the variability of respiration in terrestrial ecosystems: moving beyond Q₁₀, *Glob. Change Biol.*, 12, 154–164, <https://doi.org/10.1111/j.1365-2486.2005.01065.x>, 2006.
- Fairbairn, L., Rezanezhad, F., Gharasoo, M., Parsons, C.T., Macrae, M.L., Slowinski, S., Van Cappellen, P.: Relationship between soil CO₂ fluxes and soil moisture: Anaerobic sources explain fluxes at high water content, *Geoderma*, 434, 116493, <https://doi.org/10.1016/j.geoderma.2023.116493>, 2023.
- 675 Fernandez-Ugalde, O., Scarpa, S., Orgiazzi, A., Panagos, P., Van Liedekerke, M., Marechal, A., Jones, A.: LUCAS 2018 soil module: presentation of dataset and results. (No. JRC129926). Publications Office of the European Union, Luxembourg, 2022
- Fiedler, S.: In Situ Long-Term-Measurement of Redox Potential in Redoximorphic Soils, *Redox: Fundamentals, Processes and Applications*, Schüring, J., Schulz, H.D., Fischer, W.R., Böttcher, J., Duijnisveld, W.H.M., Springer, Berlin, Heidelberg, 81–94, https://doi.org/10.1007/978-3-662-04080-5_7, 2000.
- 680 Fiedler, S., Vepraskas, M.J., Richardson, J.L.: Soil Redox Potential: Importance, Field Measurements, and Observations, *Advances in Agronomy*, Sparks, D.L., Academic Press, 1–54, [https://doi.org/10.1016/S0065-2113\(06\)94001-2](https://doi.org/10.1016/S0065-2113(06)94001-2), 2007.
- Freeman, C., Ostle, N., Kang, H.: An enzymic “latch” on a global carbon store, *Nat.*, 409, 149–149, <https://doi.org/10.1038/35051650>, 2001.

- 685 Ghezzehei, T.A., Sulman, B., Arnold, C.L., Bogie, N.A., Berhe, A.A.: On the role of soil water retention characteristic on aerobic microbial respiration, *Biogeosciences*, 16, 1187–1209, <https://doi.org/10.5194/Biogeosciences-16-1187-2019>, 2019.
- Ginn, B., Meile, C., Wilmoth, J., Tang, Y. & Thompson, A.: Rapid Iron Reduction Rates Are Stimulated by High-Amplitude Redox Fluctuations in a Tropical Forest Soil, *Environ. Sci. Technol.* 51, 3250–3259, 2017.
- 690 Goffin, S., Wylock, C., Haut, B., Maier, M., Longdoz, B., Aubinet, M.: Modeling soil CO₂ production and transport to investigate the intra-day variability of surface efflux and soil CO₂ concentration measurements in a Scots Pine Forest (*Pinus Sylvestris*, L.), *Plant Soil*, 390, 195–211, <https://doi.org/10.1007/s11104-015-2381-0>, 2015.
- Greenway, H., Armstrong, W., Colmer, T.D.: Conditions Leading to High CO₂ (>5 kPa) in Waterlogged–Flooded Soils and Possible Effects on Root Growth and Metabolism, *Ann. Bot.* 98, 9–32, <https://doi.org/10.1093/aob/mcl076>, 2006.
- 695 Grybos, M., Davranche, M., Gruau, G., Petitjean, P., Pédrot, M.: Increasing pH drives organic matter solubilization from wetland soils under reducing conditions, *Geoderma*, 154, 13–19, <https://doi.org/10.1016/j.geoderma.2009.09.001>, 2009.
- Hakala, K., Keskitalo, M., Eriksson, C., Pitkänen, T.: Nutrient uptake and biomass accumulation for eleven different field crops, *Agr. Food Sci.*, 18, 366–387, 2009.
- 700 Hanke, A., Cerli, C., Muhr, J., Borken, W., Kalbitz, K.: Redox control on carbon mineralization and dissolved organic matter along a chronosequence of paddy soils. *Eur. J. Soil Sci.*, 64, 476–487, <https://doi.org/10.1111/ejss.12042>, 2013.
- Hanson, P.J., Edwards, N.T., Garten, C.T., Andrews, J.A.: Separating root and soil microbial contributions to soil respiration: A review of methods and observations, *Biogeochemistry*, 48, 115–146, <https://doi.org/10.1023/A:1006244819642>, 2000.
- 705 Heikkinen, J., Ketoja, E., Nuutinen, V., Regina, K.: Declining trend of carbon in Finnish cropland soils in 1974–2009. *Glob. Change Biol.* 19, 1456–1469., <https://doi.org/10.1111/gcb.12137>, 2013.
- Heikkinen, J., Keskinen, R., Kostensalo, J., Nuutinen, V.: Climate change induces carbon loss of arable mineral soils in boreal conditions. *Glob. Change Biol.*, 28, 3960–3973, <https://doi.org/10.1111/gcb.16164>, 2022.
- Heimsch, L., Vira, J., Fer, I., Vekuri, H., Tuovinen, J.-P., Lohila, A., Liski, J., Kulmala, L.: Impact of weather and management practices on greenhouse gas flux dynamics on an agricultural grassland in Southern Finland, *Agric., Ecosyst. Environ.* 374, 109179, <https://doi.org/10.1016/j.agee.2024.109179>, 2024.
- 710 Hill, A. G., Bishop, E., Coles, L. E., McLauchlan, E. J., Meddle, D. W., Pater, M. J., Watson, C. A., & Whalley, C.: Standardised General Method for the Determination of Iron with 1,10-Phenanthroline, *Analyst*, 103, 391–396, 1978.

- 715 Hirsch, A.I., Trumbore, S.E., Goulden, M.L.: The surface CO₂ gradient and pore-space storage flux in a high-porosity litter layer, *Tellus B: Chem. Phys. Meteorol.*, 56, 312–321, <https://doi.org/10.3402/tellusb.v56i4.16449>, 2004.
- Huang, W., Hall, S.J.: Elevated moisture stimulates carbon loss from mineral soils by releasing protected organic matter. *Nat. Commun.*, 8, 1774, <https://doi.org/10.1038/s41467-017-01998-z>, 2017.
- Huang, W., Wang, K., Ye, C., Hockaday, W.C., Wang, G., Hall, S.J.: High carbon losses from oxygen-limited soils challenge biogeochemical theory and model assumptions, *Glob. Change Biol.*, 27, 6166–6180, <https://doi.org/10.1111/gcb.15867>, 2021.
- 720 Huang, W., Ye, C., Hockaday, W.C., Hall, S.J.: Trade-offs in soil carbon protection mechanisms under aerobic and anaerobic conditions, *Glob. Change Biol.*, 26, 3726–3737, <https://doi.org/10.1111/gcb.15100>, 2020.
- IPCC: Sections 2-3, *Climate Change 2023: Synthesis Report.*, Contribution of Working Groups I, II and III to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change, H. Lee and J. Romero, 2023.
- 725 Jakobsen, P., Patrick, W.H.J., Williams, B.G.: Sulfide and methane formation in soils and sediments, *Soil Sci.*, 132, 279, 1981.
- Jeanneau, L., Buysse, P., Denis, M., Gruau, G., Petitjean, P., Jaffrézic, A., Flechard, C., Viaud, V.: Water Table Dynamics Control Carbon Losses from the Destabilization of Soil Organic Matter in a Small, Lowland Agricultural Catchment, *Soil Syst.*, 4, 2, <https://doi.org/10.3390/soilsystems4010002>, 2020.
- Jia, B., Niu, Z., Wu, Y., Kuzyakov, Y., Li, X.G.: Waterlogging increases organic carbon decomposition in grassland soils, *Soil Biol. Biochem.*, 148, 107927, <https://doi.org/10.1016/j.soilbio.2020.107927>, 2020.
- 730 Jian, J., Du, X., Reiter, M.S., Stewart, R.D.: A meta-analysis of global cropland soil carbon changes due to cover cropping. *Soil Biol. Biochem.*, 143, 107735, <https://doi.org/10.1016/j.soilbio.2020.107735>, 2020.
- Kaiser, K. & Guggenberger, G.: Mineral surfaces and soil organic matter. *Eur. J. Soil Sci.*, 54, 219–236, 2003).
- Khan, I., Fahad, S., Wu, L., Zhou, W., Xu, P., Sun, Z., Salam, A., Imran, M., Jiang, M., Kuzyakov, Y., & Hu, R.: Labile organic matter intensifies phosphorous mobilization in paddy soils by microbial iron (III) reduction, *Geoderma*, 352, 185–196, <https://doi.org/10.1016/j.geoderma.2019.06.011>, 2019.
- 735 Kätterer, T., Reichstein, M., Andrén, O., Lomander, A.: Temperature dependence of organic matter decomposition: a critical review using literature data analyzed with different models, *Biol Fertil Soils*, 27, 258–262, <https://doi.org/10.1007/s003740050430>, 1998.
- 740 Kaye, J.P., Quemada, M.: Using cover crops to mitigate and adapt to climate change. A review, *Agron. Sustain. Dev.*, 37, 4, <https://doi.org/10.1007/s13593-016-0410-x>, 2017.

- Keiluweit, M., Bougoure, J.J., Nico, P.S., Pett-Ridge, J., Weber, P.K., Kleber, M.: Mineral protection of soil carbon counteracted by root exudates, *Nat. Clim. Change* 5, 588–595, <https://doi.org/10.1038/nclimate2580>, 2015.
- 745 Keskinen, R., Hillier, S., Liski, E., Nuutinen, V., Nyambura, M., & Tiljander, M.: Mineral composition and its relations to readily available element concentrations in cultivated soils of Finland. *Acta. Agr. Scand. Section B — Soil & Plant Sci.*, 72, 751-760, <https://www.tandfonline.com/doi/abs/10.1080/09064710.2022.2075790>, 2022.
- Koizumi, H., Kontturi, M., Mariko, S., Nakadai, T., Bekku, Y., Mela, T.: Soil Respiration in Three Soil Types in Agricultural Ecosystems in Finland, *Acta. Agr. Scand. Section B — Soil & Plant Sci.*, 49, 65–74, <https://doi.org/10.1080/09064719950135560>, 1999.
- 750 Kronberg, R., Kanerva, S., Koskinen, M., Polvinen, T., Heinonsalo, J., Pihlatie, M.: Controlled soil monolith experiment for studying the effects of waterlogging on redox processes, *Geoderma*, 452, 117110, <https://doi.org/10.1016/j.geoderma.2024.117110>, 2024.
- Kuzyakov, Y.: Sources of CO₂ efflux from soil and review of partitioning methods, *Soil Biol. Biochem.*, 38, 425–448, <https://doi.org/10.1016/j.soilbio.2005.08.020>, 2006.
- 755 Kuzyakov, Y., Friedel, J.K., Stahr, K.: Review of mechanisms and quantification of priming effects, *Soil Biol. Biochem.*, 32, 1485–1498, [https://doi.org/10.1016/S0038-0717\(00\)00084-5](https://doi.org/10.1016/S0038-0717(00)00084-5), 2000.
- LaCroix, R.E., Tfaily, M.M., McCreight, M., Jones, M.E., Spokas, L., Keiluweit, M.: Shifting mineral and redox controls on carbon cycling in seasonally flooded mineral soils, *Biogeosciences*, 16, 2573–2589, <https://doi.org/10.5194/Biogeosciences-16-2573-2019>, 2019.
- 760 Lavalley, J.M., Soong, J.L., Cotrufo, M.F.: Conceptualizing soil organic matter into particulate and mineral-associated forms to address global change in the 21st century, *Glob. Change Biol.*, 26, 261–273, <https://doi.org/10.1111/gcb.14859>, 2020.
- Lehmann, J., Hansel, C.M., Kaiser, C., Kleber, M., Maher, K., Manzoni, S., Nunan, N., Reichstein, M., Schimel, J.P., Torn, M.S., Wieder, W.R., Kögel-Knabner, I.: Persistence of soil organic carbon caused by functional complexity. *Nat. Geosci.* 13, 529–534, <https://doi.org/10.1038/s41561-020-0612-3>, 2020.
- 765 Lemola, R., Uusitalo, R., Hyväluoma, J., Sarvi, M., Turtola, E.: Suomen peltojen maalajit, multavuus ja fosforipitoisuus: Vuodet 1996–2000 ja 2005–2009. Luonnonvarakeskus, 2018.
- Liang, Z., Rasmussen, J., Poeplau, C. & Elsgaard, L.: Priming effects decrease with the quantity of cover crop residues – Potential implications for soil carbon sequestration, *Soil Biol. Biochem.*, 184, 109110, 2023.

- 770 Lin, Y., Campbell, A.N., Bhattacharyya, A., DiDonato, N., Thompson, A.M., Tfaily, M.M., Nico, P.S., Silver, W.L., Pett-Ridge, J.: Differential effects of redox conditions on the decomposition of litter and soil organic matter, *Biogeochemistry*, 154, 1–15, <https://doi.org/10.1007/s10533-021-00790-y>, 2021.
- Lind, S.E., Shurpali, N.J., Peltola, O., Mammarella, I., Hyvönen, N., Maljanen, M., Rätty, M., Virkajärvi, P., Martikainen, P.J.: Carbon dioxide exchange of a perennial bioenergy crop cultivation on a mineral soil, *Biogeosciences*, 13, 1255–1268, <https://doi.org/10.5194/Biogeosciences-13-1255-2016>, 2016.
- 775 Liu, Y., Jia, B., Zhang, Y., Cui, H., Li, X.G.: The Effect of Waterlogging on Soil Organic Carbon Decomposition Is Dependent on Its Biochemistry, *J. Soil Sci. Plant Nutr.*, 23, 4609–4619, <https://doi.org/10.1007/s42729-023-01377-2>, 2023.
- Liu, Y., Wang, X., Wen, Y., Cai, H., Song, X., Zhang, Z.: Effects of freeze-thaw cycles on soil greenhouse gas emissions: A systematic review. *Environ. Res.*, 248, 118386, <https://doi.org/10.1016/j.envres.2024.118386>, 2024.
- 780 Lohila, A., Aurela, M., Regina, K., Laurila, T.: Soil and total ecosystem respiration in agricultural fields: effect of soil and crop type, *Plant and Soil*, 251, 303–317, <https://doi.org/10.1023/A:1023004205844>, 2003.
- Lovley, D. R.: Dissimilatory Fe(III) and Mn(IV) reduction, *Microbiol. Rev.*, 55(2), 259–287, <https://doi.org/10.1128/mr.55.2.259-287.1991>, 1991.
- Luo, Y., Zhou, X.: *Soil Respiration and the Environment*, Academic Press, London, UK, 2006.
- 785 Maier, M., Schack-Kirchner, H., Hildebrand, E.E., Holst, J.: Pore-space CO₂ dynamics in a deep, well-aerated soil, *Eur. J. Soil Sci.*, 61, 877–887, <https://doi.org/10.1111/j.1365-2389.2010.01287.x>, 2010.
- Maier, M., Schack-Kirchner, H., Hildebrand, E.E., Schindler, D.: Soil CO₂ efflux vs. soil respiration: Implications for flux models, *Agr. Forest Meteorol.*, 151, 1723–1730, <https://doi.org/10.1016/j.agrformet.2011.07.006>, 2011.
- Mattila, T.J., Vihanto, N.: Agricultural limitations to soil carbon sequestration: Plant growth, microbial activity, and carbon stabilization, *Agr. Ecosyst. Environ.*, 367, 108986, <https://doi.org/10.1016/j.agee.2024.108986>, 2024.
- 790 Mikutta, C., Niegisch, M., Thompson, A., Behrens, R., S. Schnee, L., Hoppe, M., Dohrmann, R.: Redox cycling of straw-amended soil simultaneously increases iron oxide crystallinity and the content of highly disordered organo-iron(III) solids, *Geochim. Cosmochim. Acta.*, 371, 126–143, <https://doi.org/10.1016/j.gca.2024.02.009>, 2024.
- Moyano, F.E., Manzoni, S., Chenu, C.: Responses of soil heterotrophic respiration to moisture availability: An exploration of processes and models, *Soil Biol. Biochem.*, 59, 72–85, <https://doi.org/10.1016/j.soilbio.2013.01.002>, 2013.
- 795 Moyano, F.E., Vasilyeva, N., Menichetti, L.: Diffusion based modelling of temperature and moisture interactive effects on carbon fluxes of mineral soils (preprint), *Biogeosciences*, 2018.

- Muhammad, I., Wang, J., Sainju, U.M., Zhang, S., Zhao, F., Khan, A.: Cover cropping enhances soil microbial biomass and affects microbial community structure: A meta-analysis, *Geoderma*, 381, 114696, <https://doi.org/10.1016/j.geoderma.2020.114696>, 2021.
- 800
- Nevalainen, O., Niemitalo, O., Fer, I., Juntunen, A., Mattila, T., Koskela, O., Kukkamäki, J., Höckerstedt, L., Mäkelä, L., Jarva, P., Heimsch, L., Vekuri, H., Kulmala, L., Stam, Å., Kuusela, O., Gerin, S., Viskari, T., Vira, J., Hyväluoma, J., Tuovinen, J.-P., Lohila, A., Laurila, T., Heinonsalo, J., Aalto, T., Kunttu, I., Liski, J.: Towards agricultural soil carbon monitoring, reporting, and verification through the Field Observatory Network (FiON), *Geosci. Instrum. Meth.*, 11, 93–109, <https://doi.org/10.5194/gi-11-93-2022>, 2022.
- 805
- Pan, W., Kan, J., Inamdar, S., Chen, C., Sparks, D.: Dissimilatory microbial iron reduction release DOC (dissolved organic carbon) from carbon-ferrihydrite association, *Soil Biol. Biochem.*, 103, 232–240, <https://doi.org/10.1016/j.soilbio.2016.08.026>, 2016.
- Pavelka, M., Acosta, M., Kiese, R., Altimir, N., Brümmer, C., Crill, P., Darenova, E., Fuß, R., Gielen, B., Graf, A., Klemetsson, L., Lohila, A., Longdoz, B., Lindroth, A., Nilsson, M., Jiménez, S.M., Merbold, L., Montagnani, L., Peichl, M., Pihlatie, M., Pumpanen, J., Ortiz, P.S., Silvennoinen, H., Skiba, U., Vestin, P., Weslien, P., Janous, D., Kutsch, W.: Standardisation of chamber technique for CO₂, N₂O and CH₄ fluxes measurements from terrestrial ecosystems, *Int. Agrophys.*, 32, 569–587, <https://doi.org/10.1515/intag-2017-0045>, 2018.
- 810
- Peters, V., Conrad, R.: Sequential reduction processes and initiation of CH₄ production upon flooding of oxic upland soils. *Soil Biol. Biochem.*, 28, 371–382, [https://doi.org/10.1016/0038-0717\(95\)00146-8](https://doi.org/10.1016/0038-0717(95)00146-8), 1996.
- 815
- Pinheiro, J., Bates, D., R Core Team: nlme: Linear and Nonlinear Mixed Effects Models. R package version 3.1-164, 2023.
- Poeplau, C., Aronsson, H., Myrbeck, Å., Kätterer, T.: Effect of perennial ryegrass cover crop on soil organic carbon stocks in southern Sweden, *Geoderma Reg.*, 4, 126–133, <https://doi.org/10.1016/j.geodrs.2015.01.004>, 2015.
- Poeplau, C., Don, A.: Carbon sequestration in agricultural soils via cultivation of cover crops – A meta-analysis, *Agric. Ecosyst. Environ.*, 200, 33–41, <https://doi.org/10.1016/j.agee.2014.10.024>, 2015.
- 820
- Ponnamperuma, F.N.: Chemical kinetics of wetland rice soils relative to soil fertility, *Wetland Soils: Characterization, Classification, and Utilization*, IRRI, 71–89, 1985.
- Possinger, A.R., Bailey, S.W., Inagaki, T.M., Kögel-Knabner, I., Dynes, J.J., Arthur, Z.A., Lehmann, J.: Organo-mineral interactions and soil carbon mineralizability with variable saturation cycle frequency, *Geoderma* 375, 114483, <https://doi.org/10.1016/j.geoderma.2020.114483>, 2020.
- 825
- R Core Team.: R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, <https://www.R-project.org/>, 2025.

- Raich, J.W., Schlesinger, W.H.: The global carbon dioxide flux in soil respiration and its relationship to vegetation and climate, *Tellus B*, 44, 81–99, <https://doi.org/10.1034/j.1600-0889.1992.t01-1-00001.x>, 1992.
- 830 Rasmussen, C., Heckman, K., Wieder, W. R., Keiluweit, M., Lawrence, C. R., Berhe, A. A., Blankinship, J. C., Crow, S. E., Druhan, J. L., Hicks Pries, C. E., Marin-Spiotta, E., Plante, A. F., Schädel, C., Schimel, J. P., Sierra, C. A., Thompson, A., & Wagai, R.: Beyond clay: Towards an improved set of variables for predicting soil organic matter content, *Biogeochemistry*, 137(3), 297–306, <https://doi.org/10.1007/s10533-018-0424-3>, 2018.
- Reddy, K.R., DeLaune, R. D., Inglett P. W.: *Biogeochemistry of Wetlands: Science and Applications*. CRC Press, Boca Raton, 89–119, <https://doi.org/10.1201/9780429155833>, 2022.
- 835 Ruosteenoja, K., Jylhä, K.: Projected climate change in Finland during the 21st century calculated from CMIP6 model simulations, *Geophysica*, 56, 39–69, 2022.
- Russell, V.L.: *Estimated Marginal Means, aka Least-Squares Means*, R package version 1.8.5., 2023.
- Ryan, M.G., Law, B.E.: Interpreting, measuring, and modeling soil respiration, *Biogeochemistry*, 73, 3–27, <https://doi.org/10.1007/s10533-004-5167-7>, 2005.
- 840 Salonen, A.-R., de Goede, R., Creamer, R., Heinonsalo, J., Soinne, H.: Soil organic carbon fractions and storage potential in Finnish arable soils, *Eur. J. Soil Sci.* 75, e13527, <https://doi.org/10.1111/ejss.13527>, 2024.
- Sánchez-Cañete, E.P., Barron-Gafford, G.A., Chorover, J.: A considerable fraction of soil-respired CO₂ is not emitted directly to the atmosphere, *Sci. Rep-UK.*, 8, 13518, <https://doi.org/10.1038/s41598-018-29803-x>, 2018.
- 845 Sigg, L.: *Redox Potential Measurements in Natural Waters: Significance, Concepts and Problems*, *Redox: Fundamentals, Processes and Applications*, Schüring, J., Schulz, H. D., Fischer, W. R., J. Böttcher, & Duijnisveld, W. H. M., 81–94, Springer, https://doi.org/10.1007/978-3-662-04080-5_7, 2000.
- Sippola, J.: Mineral composition and its relation to texture and to some chemical properties in Finnish subsoils, *Ann. Agric. Fenn.* 13: 169-234, 1974.
- 850 Snoeyenbos-West, O. L., Nevin, K. P., Anderson, R. T., & Lovley, D. R.: Enrichment of *Geobacter* Species in Response to Stimulation of Fe(III) Reduction in Sandy Aquifer Sediments, *Microb. Ecol.*, 39(2), 153–167, <https://doi.org/10.1007/s002480000018>, 2000.
- Stumm, W.: *Aquatic Chemistry: Chemical Equilibria and Rates in Natural Waters*, John Wiley & Sons, Incorporated. <http://ebookcentral.proquest.com/lib/helsinki-ebooks/detail.action?docID=7103403>, 1995.

- 855 Sulman, B.N., Phillips, R.P., Oishi, A.C., Shevliakova, E., Pacala, S.W.: Microbe-driven turnover offsets mineral-mediated storage of soil carbon under elevated CO₂, *Nat. Clim. Change*, 4, 1099–1102, <https://doi.org/10.1038/nclimate2436>, 2014.
- Sun, S.-Q., Cai, H.-Y., Chang, S.X., Bhatti, J.S.: Sample storage-induced changes in the quantity and quality of soil labile organic carbon, *Sci. Rep-UK.*, 5, 17496, <https://doi.org/10.1038/srep17496>, 2015.
- 860 Tautges, N.E., Chiartas, J.L., Gaudin, A.C.M., O’Geen, A.T., Herrera, I., Scow, K.M.: Deep soil inventories reveal that impacts of cover crops and compost on soil carbon sequestration differ in surface and subsurface soils, *Glob. Change Biol.*, 25, 3753–3766, <https://doi.org/10.1111/gcb.14762>, 2019.
- Tete, E., Viaud, V., Walter, C.: Organic carbon and nitrogen mineralization in a poorly-drained mineral soil under transient waterlogged conditions: an incubation experiment, *Eur. J. Soil Sci.*, 66, 427–437, <https://doi.org/10.1111/ejss.12234>,
865 2015.
- Torn, M.S., Trumbore, S.E., Chadwick, O.A., Vitousek, P.M., Hendricks, D.M.: Mineral control of soil organic carbon storage and turnover, *Nat.*, 389, 170–173, <https://doi.org/10.1038/38260>, 1997.
- Uusitalo, R., Ylivainio, K., Hyväluoma, J., Rasa, K., Kaseva, J., Nylund, P., Pietola, L., Turtola, E.: The effects of gypsum on the transfer of phosphorus and other nutrients through clay soil monoliths. *Agr. Food Sci.*, 21, 260–278,
870 <https://doi.org/10.23986/afsci.4855>, 2012.
- Venables, W.N., Ripley, B.D.: Random and Mixed Effects, *Modern Applied Statistics with S*, Venables, W.N., Ripley, B.D. (Eds.), Springer, New York, 271–300, https://doi.org/10.1007/978-0-387-21706-2_10, 2002.
- Vet, R., Artz, R. S., Carou, S., Shaw, M., Ro, C.-U., Aas, W., Baker, A., Bowersox, V. C., Dentener, F., Galy-Lacaux, C., Hou, A., Pienaar, J. J., Gillett, R., Forti, M. C., Gromov, S., Hara, H., Khodzher, T., Mahowald, N. M., Nickovic, S.,
875 Rao, P. S. P., Reid, N. W.: A global assessment of precipitation chemistry and deposition of sulfur, nitrogen, sea salt, base cations, organic acids, acidity and pH, and phosphorus. *Atmos. Environ.*, 93, 3–100. <https://doi.org/10.1016/j.atmosenv.2013.10.060>, 2014.
- Virtanen, S., Simojoki, A., Knuutila, O., Yli-Halla, M.: Monolithic lysimeters as tools to investigate processes in acid sulphate soil. *Agric. Water Manag.*, 127, 48–58, <https://doi.org/10.1016/j.agwat.2013.05.013>, 2013.
- 880 von Lützow, M., Kögel-Knabner, I., Ekschmitt, K., Flessa, H., Guggenberger, G., Matzner, E., Marschner, B.: SOM fractionation methods: Relevance to functional pools and to stabilization mechanisms, *Soil Biol. Biochem.*, 39, 2183–2207, <https://doi.org/10.1016/j.soilbio.2007.03.007>, 2007.
- Wickland, K.P., Neff, J.C.: Decomposition of soil organic matter from boreal black spruce forest: environmental and chemical controls, *Biogeochemistry*, 87, 29–47, <https://doi.org/10.1007/s10533-007-9166-3>, 2008.

- 885 Williams, M.D., Oostrom, M.: Oxygenation of anoxic water in a fluctuating water table system: an experimental and numerical study. *J. Hydrol.*, 230, 70–85, [https://doi.org/10.1016/S0022-1694\(00\)00172-4](https://doi.org/10.1016/S0022-1694(00)00172-4), 2000.
- Winkler, P., Kaiser, K., Jahn, R., Mikutta, R., Fiedler, S., Cerli, C., Kölbl, A., Schulz, S., Jankowska, M., Schloter, M., Müller-Niggemann, C., Schwark, L., Woche, S.K., Kümmel, S., Utami, S.R., Kalbitz, K.: Tracing organic carbon and microbial community structure in mineralogically different soils exposed to redox fluctuations. *Biogeochemistry*, 890 143, 31–54, <https://doi.org/10.1007/s10533-019-00548-7>, 2019.
- Yong, R. N., Mohamed, A. M. O. & Wang, B. W.: Influence of amorphous silica and iron hydroxide on interparticle action and soil surface properties, *Can. Geotech. J.*, 29, 803–818 , 1992.
- Zeileis, A., Grothendieck, G.: zoo: S3 Infrastructure for Regular and Irregular Time Series, <https://doi.org/10.48550/arXiv.math/0505527>, 2005.
- 895 Zhao, Q., Dunham-Cheatham, S., Adhikari, D., Chen, C., Patel, A., Poulson, S.R., Obrist, D., Verburg, P.S.J., Wang, X., Roden, E.R., Thompson, A., Yang, Y.: Oxidation of soil organic carbon during an anoxic-oxic transition, *Geoderma*, 377, 114584, <https://doi.org/10.1016/j.geoderma.2020.114584>, 2020
- Zheng, J., Thornton, P.E., Painter, S.L., Gu, B., Wullschleger, S.D., Graham, D.E.: Modeling anaerobic soil organic carbon decomposition in Arctic polygon tundra: insights into soil geochemical influences on carbon mineralization. *Biogeosciences*, 16, 663–680, <https://doi.org/10.5194/Biogeosciences-16-663-2019>, 2019. 900
- Zhou, W., Hui, D., Shen, W.: Effects of Soil Moisture on the Temperature Sensitivity of Soil Heterotrophic Respiration: A Laboratory Incubation Study, *PLOS ONE* 9, e92531, <https://doi.org/10.1371/journal.pone.0092531>, 2014.