

1 **Addition of brackish water to tundra soils does not inhibit**
2 **methane production: implications for Arctic coastal methane**
3 **production**

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14 **Abstract.** In Arctic regions where coastal sediments contain permafrost, global climate change drives processes such
15 as erosion and subsidence. The contribution of these processes to carbon emissions, **especially from ground**
16 **subsidence**, are still uncertain. Relative sea level rise can lead to more waterlogged environments, promoting anoxic
17 degradation of organic matter but it can also lead to a greater exposure of coastal sediments to seawater. This could
18 alter methane (CH₄) production dynamics, although the controls remain poorly understood. For instance, sulfates
19 contained in seawater may have a tampering effect on methanogenesis through competitive inhibition but the increase
20 in microbial abundance could enhance methanogenesis. In this study, we present CH₄ production rates alongside
21 geochemical analyses in a rapidly evolving coastal landscape near the community of Tuktoyaktuk, NWT, Canada,
22 which is located in the continuous permafrost zone. To better constrain CH₄ production dynamics along the land to
23 ocean continuum, sediment **cores were collected from nearshore marine sediments and soil** profiles were collected
24 from the active layer of the coastal (intertidal) zone and inland soils. Anoxic incubations were performed, amended
25 with brackish water to simulate the effect of seawater on the breakdown of organic matter and the production of CH₄.
26 We found marine sediments expectedly led to negligible CH₄ production rates, while the inland sites showed variable
27 rates between null and 35 nmol cm⁻³ d⁻¹. The coastal (intertidal) zone had the highest rates reaching 415 nmol cm⁻³ d⁻¹.
28 Interestingly, sulfate present in brackish water and sediments did not suppress methanogenesis in the incubations of
29 the coastal and inland zones. Analyses of stable carbon isotopes from CH₄ produced in the incubation experiment
30 indicated greater acetotrophy and higher organic matter lability in the coastal zone, possibly contributing to higher
31 CH₄ production rates. This study highlights the potential for significant CH₄ emissions even with high sulfate
32 concentrations which are classically thought to inhibit methanogenesis. This suggests that Arctic coastal microbial
33 CH₄ production might be an understudied source to the atmosphere.

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1 Introduction

Arctic coastal ecosystems are impacted by sea level rise, coastal erosion, land submersion, higher frequency in storm events and permafrost degradation (AMAP 2019; Guimond et al., 2021; Irrgang et al., 2022; Lantuit et al., 2013; Lim et al., 2020). The amplification of coastal environmental changes has impacts on biogeochemical cycles (AMAP, 2017) and on organic matter (OM) degradation processes and fluxes at the land-ocean continuum (Tanski et al., 2021). Furthermore, the progressive thawing of permafrost exposes long frozen organic matter to microbial decomposition (Lapham et al., 2020; Pellerin et al., 2022; Schuur et al., 2015), leading to the release of greenhouse gases like carbon dioxide (CO₂) and methane (CH₄). Inputs and outputs of the Arctic carbon biogeochemical cycle are known to be reshaped by rapid environmental changes (Couture et al., 2018), but processes in coastal settings are still poorly understood.

Rates of coastal change vary according to the morphology of coastal landscapes (Manson et al., 2019). The average rate of land retreat measured in the Tuktoyaktuk Coastlands (North-West Territories, Canada), our study site, between 1985 and 2020 was -1.06 m/yr, while processes of ground subsidence and submersion induced retreat rates higher than -4 m/yr (Costa, 2022) which can inundate large swaths of land. Inundated tundra flats and polygons are widespread landforms in the landscape (Costa, 2022). Polygon tundra flats are characterized by ice-wedge polygons, which are formed by the repeated thermal contraction and expansion of the upper layers of the permafrost (Steedman et al., 2016). At the surface, the polygons are expressed as minor topographic features separated by lower-lying, often wet or inundated channels called troughs (Fig 1). Polygons can be classified as low-centered (with a low, wet center and raised rims) or as high-centered (with well-drained centers and lower well-drained rims) (Fig 1), exhibiting strong thermal, hydrological and geochemical gradients (Vaughn et al., 2016).

During growing season, where atmospheric temperatures allow for active layer to thaw and vegetation to grow, hydrological conditions in polygons play a pivotal role in shaping the pathways of OM decomposition and consequently influence the resulting CO₂ and CH₄ production. Well drained oxic conditions allow microbes to decompose OM rapidly, leading to the production of CO₂ (Jones et al., 2020). Conversely, water saturation restricts oxygen availability, promoting anaerobic respiration and fermentation, inducing both CO₂ and CH₄ production (Lipson et al., 2012; Turetsky et al., 2008). Thus, coastal changes and higher atmospheric temperatures during open-water season can swiftly alter water saturation conditions in polygons, in many cases significantly enhancing fermentation and CH₄ production (Elberling et al., 2013; Holm et al., 2020; Treat et al., 2015).

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Furthermore, coastal changes can also influence the chemistry of the water within soils, which can affect OM degradation. In anaerobic conditions, OM degradation processes follow a sequence of electron acceptors of decreasing energetic yields with nitrate, manganese oxides, iron oxides and sulfate as the most abundant electron acceptors (Froelich et al., 1979). It is when all alternative electron acceptors are depleted that fermentation takes place, leading to the production of CH₄; methanogenesis. For example, it has long been established that in beach, estuarine, and marsh mudflats on the Brittany coast (France), organic matter (OM) degradation is dominated by sulfate reduction, as the high sulfate content of seawater

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inhibits methanogenesis through competitive inhibition (Winfrey and Ward, 1983). In contrast, sediments beneath thermokarst lakes are anoxic and largely devoid of alternative electron acceptors, so OM degradation is almost entirely driven by methanogenesis (Sepulveda-Jauregui et al., 2015). These examples highlight that the chemical composition of the aqueous environment plays a critical role in controlling the pathways of OM degradation. CH₄ produced in soils or sediments can also be oxidized by anaerobic methanotrophic archaea and sulfate-reducing bacteria (Boetius et al., 2000; La et al., 2022) present in the soils or sediment, contributing to lower CH₄ emissions in coastal environments. Thus, on or near the coast, the interaction with seawater, which contains electron acceptors such as sulfate, can shift the OM mineralization pathway and the resulting CO₂ and CH₄ productions. Consequently, a nuanced understanding of biogeochemical processes and their drivers is paramount in determining the magnitude of permafrost carbon emissions, especially from coastal environments.

Numerous CH₄ emissions monitoring programs are in operation, but remote-sensing methods lack the ability to comprehensively capture the microbial, biogeochemical and environmental processes involved. In specific regions, estimates of methane production from the breakdown of OM is possible by carefully studying degradation pathways and production rates (Pellerin et al., 2022; Heslop et al., 2015; Knoblauch et al., 2018; Treat et al., 2014). To reduce the knowledge gap of CH₄ biogeochemistry in coastal permafrost settings, we collected material from the active layer and taliks of water bodies for incubation experiments, which were coupled to physical and chemical characterizations.

The main objective of this study was to assess microbial CH₄ production dynamics in a coastal permafrost setting and apply it at the landscape level, since methane production is well documented in inland thermokarst but is not well understood in a land-ocean interaction context. We hypothesized that methanogenesis in coastal active layer incubations would be suppressed by the addition of sulfate. Consequently, we discuss the influence of environmental conditions on microbial CH₄ production with an emphasis on brackish water addition in coastal soils and sediments along with the microbial pathways involved. We then apply these results at the landscape level to provide an estimate of CH₄ production in the event that a natural process like a storm inputs brackish water over a large area of polygonal patterned ground. We use the region around Tuktoyaktuk as an example.

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2 Methods

2.1 Site description and sampling

Tuktoyaktuk (69°26'24" N, 133°01'52" W) is located in the Inuvik region of the North-West Territories, adjacent to the Arctic Ocean in the Kugmallit Bay, east of the Mackenzie Delta. The region experiences prolonged cold winters, short cool summers, and year-round low precipitation, fostering low-arctic tundra vegetation. Lying in the continuous permafrost zone, its coastal areas feature thick Quaternary and glaciogenic unconsolidated deposits (Rampton, 1988), where permafrost thickness averages 400 m (Hu et al., 2013) and is characterized by prevalent ground ice structures (Mackay and Dallimore, 1992; Martin et al., 2018; Murton, 1996; Rampton, 1988). The area has been ice-free for the past 13,000 years, with evidence indicating that early Holocene summer temperatures were up to 6°C warmer than today, fostering vegetation and peat accumulation (Dallimore et al., 1997; Vardy et al., 1997). During that same period, sea level was considerably lower than it is today and the Tuktoyaktuk area was located approximately 100 km inland (Vardy et al., 1997). Currently, ground subsidence and coastal erosion are major causes of rapid land retreat (Hynes et al., 2014; Lapham et al., 2020; Lim et al., 2020). Combined with sea level rise (Hill et al., 1993), it is projected that a substantial amount of terrestrial soil will become part of the ocean seafloor either by erosion and deposition or by subsidence of land and submersion. Over the past 15 years, extensive studies on Tuktoyaktuk's coastal environment, driven by the region's vulnerability to climate change, highlighted challenges for the Inuvialuit population relying on hunting, fishing, trapping and harvesting (Andrachuk and Smit, 2012).

Active layer samples were collected from two sites: an inland site, Reindeer Point (RP) and a coastal site, Toker Point (TP). Talik sediments were also collected at both RP and TP sites from polygonal troughs and pondlets and sediments from a marine site, Harbor, completed the transect from terrestrial to marine settings. RP was selected as the inland site because it features a polygonal patterned ground typical of the region, and is located in a stable region not directly affected by coastal processes such as storm surges, tides, seawater intrusion, erosion etc. The thermokarst lake margin, about 300m south of RP has remain unchanged since aerial photos began recording the evolution of the landscape in 1947 (Fig S1). TP was selected as the coastal site because of the strong coastal processes such as tides and storm surge that regularly lead to seawater intrusion in this polygonal patterned ground, strongly influenced by ground subsidence. The Harbor site was selected about 400 m offshore in the Harbor of Tuktoyaktuk where total water depth was 20 m and cold marine bottom waters were overlain by a 10 m surface brackish water layer. 25 cm sediment cores were collected using a UWITEC gravity corer. The sediments consisted of recently deposited silty sands originating from the strong erosional processes occurring in the region (Whalen et al., 2022). The site was accessible by small watercraft. At RP and TP sites, soil profiles were extracted from the active layer by digging a soil pit with a shovel. To retain an intact stratigraphic relationship, samples were taken from the wall of the soil pit. Biogenic ebullition gases were collected from pondlets at RP and TP. Pondlets were located within sampled polygonal patterned ground and are defined as small (1 to 3 m²) and shallow standing bodies of water, potentially draining seasonally. Samples were trapped using a plastic funnel attached to a 20 mL glass vial. Surface soil lying at the bottom of the pondlets (Fig. 1) were poked until the vial was filled with gas. Once full, vials were crimped with 20 mm butyl rubber stoppers and aluminum caps. Samples were kept frozen until the time of analyses.

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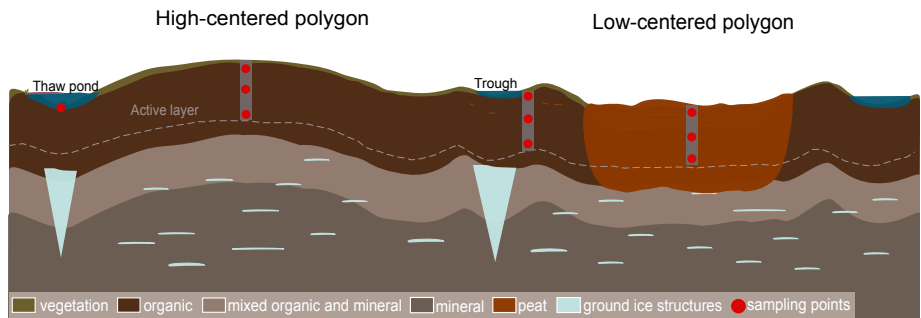


Figure 1. Schematic representation of polygonal tundra with peat accumulation as seen in continuous permafrost environments and sampling design for this study. High-centered polygons are associated with drier conditions, while low-centered polygons, troughs and pondlets are associated with humid or water-saturated conditions. Vegetation cover and OM reflect the hydrology of sites. Not to scale.

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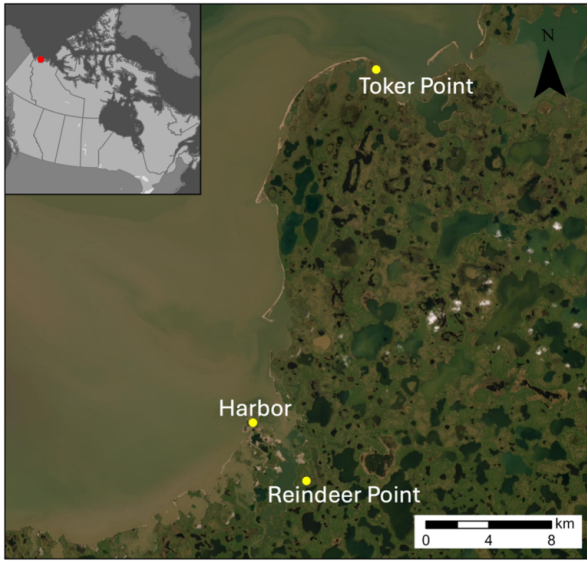


Figure 2. Map of study area indicating the sampled sites with yellow dots (ESRI, 2022). Harbor site is located in the marine waters of Tuktoyaktuk, Toker Point site is located in the coastal (intertidal) zone and Reindeer Point site is located inland. High resolution satellite imagery and pictures of soil profiles for RP and TP sites available in supplementary materials (Fig. S3).

The inland site, (RP), was located 750 m from the coast and 2 km East of Tuktoyaktuk in a polygonal patterned ground. This patterned ground is located in a depression, surrounded by elevated plateaus with observable ground water flowing into the valley. In this area, low-centered polygons exhibited higher moisture levels compared to high-centered polygons. High-centered polygons were colonized by shrubs and small flowering plants like *Ericaceae*, while low-centered polygons were dominated by hydrophilic plants such as grasses and sedges. Wet troughs delimited the polygons, with vegetation reflecting waterlogged conditions. The mean active layer and talik thickness across RP was about 35 cm. Profile 10A was collected from a trough and presented water-saturated conditions with brown OM. Profiles 10B and 10D were collected from high-centered polygons and characterized by unsaturated conditions with dark brown OM and presence of roots until 20 cm depth. Profile 10C was collected from a low-centered polygon and consisted of reddish-brown peat throughout. Profiles 10A, 10B and 10D did not consist of peat.

The coastal site (TP) is located 20 km NW of Tuktoyaktuk, featuring a polygonal patterned ground, largely colonized by *Carex sp.*, a type of graminoid plant common near Arctic coastlines. The mean active layer and talik depths were 35 cm. The site's dynamics are influenced by the twice-daily ebb and flow of tides. Profile 07 was collected from a water-saturated low-centered polygon, located in the intertidal zone. The soil color was very dark greyish black. Profile 08 was collected from a water-saturated polygonal trough not immediately located in the intertidal zone, but which floods during storms. The soil was characterized by dark greyish-brown OM mixed with sand. Finally, profile 09, was collected from the center of a higher-centered polygon situated in the middle intertidal zone. The active layer appeared water unsaturated. The soil from this site consisted of a mixture of black organic-rich material and sand. The sand found in samples from TP appeared to be wind-deposited from nearby dunes.

2.2 Sulfates and chloride concentrations in sediments

The extraction of sulfate and chloride from sediments and soils pore-water was conducted through a leaching experiment following Lacelle (2019). Frozen aliquots of sediments and soils were thawed at 4°C overnight, then weighed, dried in the oven at 60°C for 24 hours and re-weighed to determine the densities. Aliquots of dried material were put in 50 mL falcon tubes with nanopure water following a 1:10 ratio. Tubes were then shaken for one hour to

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promote leaching of anions towards the aqueous phase of the solution. Once the leaching process was done, 2 mL of the aqueous solution was filtered using 0.2 μm pore size Whatman 25 mm GD/X syringe filters and transferred in disposable microtubes. Concentrations of sulfate and chloride were measured by ion chromatography using a Thermo Dionex Integriion at UQAR's Chemistry department facilities with a limit of detection of 0.01 $\mu\text{g}/\text{mL}$. The measured concentrations are expressed in mmol g^{-1} wet-weight¹ of material (mmol g^{-1} wweight¹). Only one measurement per sample was performed as stability tests revealed variability of less than 3% between measured samples. The error on each value was calculated by the least squares method (Skoog et al., 2014).

2.3 Methane production rates in incubations

Long-term sediment and soil incubations under anoxic conditions were used to assess CH_4 production rates over several months by measuring CH_4 accumulation in the vials' headspace. The objective was to simulate the increased connectivity between the land and the ocean in the coastal environment of the Canadian Arctic, which represents an important aspect of the ongoing regional environmental transition. Collected sediments and soil profiles were immediately sub-sampled based on depth, at 5 or 10 cm intervals, according to shifts in sedimentary units. To prepare incubations, about 4 mL of sediment and exactly 2 mL of brackish water (collected from the coast) were immediately transferred into 20 mL glass vials. Incubation vials were crimped with 20 mm blue chlorobutyl rubber stoppers and aluminum caps. The bottles were flushed with nitrogen gas (Alpha Gaz 1) at a rate of 300mL/min for 2 minutes in the field to replace the air with a nitrogen atmosphere. Four incubations were prepared for each sampled depth; 3 were kept for measurements of CH_4 production rates (triplicates) and one served for isotopic analyses. Incubations were kept at a constant temperature of 4°C throughout the entire 339 days incubation period with no fluctuations. Substrate concentrations were not actively controlled or monitored, aside from repeated measurements of headspace methane. For logistical reasons, we were not able to measure CH_4 concentrations from the incubations in the first few weeks and the last measurement was conducted at day 339. The brackish water added to all incubations contained $5.7 \pm 0.0 \text{ mmol g}^{-1}$ wweight¹ of sulfate and $28.7 \pm 0.5 \text{ mmol g}^{-1}$ wweight¹ of chloride.

Analyses of the CH_4 concentrations in the headspace of the vials were performed on a gas chromatograph (Agilent 8900) equipped with a flame ionization detector (GC-FID) at UQAR facilities. The GC-FID is equipped with a 100 μL injection loop to ensure a consistent volume of sample is analyzed. To saturate the injection loop, 300 μL are taken from the headspace of the vials and transferred to the injection loop with a gas-tight syringe. Prior to injection, samples were shaken for 30 seconds to equilibrate headspace and sediment gases. This procedure was done every two weeks for 16 weeks to measure CH_4 accumulation in the headspace. The resulting production rates were calculated from the linear accumulation measured during the incubation period, and values are expressed in nmol of CH_4 per cubic centimeters of wet material per day ($\text{nmol cm}^{-3} \text{ d}^{-1}$). The density of the collected samples varied widely, with some being organic deposits and peat, while others contained higher mineral content. Consequently, the CH_4 production rates were expressed volumetrically to account for these discrepancies which are more representative of the volume they occupy in the soil, sedimentary columns and landscape. The limit of detection of the GC-FID is 0.3 ppm and all samples had higher concentrations. Each value represents the mean of triplicate measurements and the reported uncertainty on the measurement is the standard deviation on triplicates.

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To estimate the potential total active layer CH_4 production (T), the active layer production rates were vertically integrated to obtain the total CH_4 production of each profile. Values are reported in $\text{mmol m}^{-2} \text{ d}^{-1}$ and were calculated using eq. 1.

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$$T = \frac{1}{100} \sum_{i=1}^n P_i \cdot e_i$$

$$T = \frac{1}{100} \sum_{i=1}^n P_i \cdot e_i \quad [\text{mmol m}^{-2} \text{ d}^{-1}]$$

!#\$%

$$T = \frac{1}{100} \sum_{i=1}^n P_i \cdot e_i \quad [\text{mmol m}^{-2} \text{d}^{-1}] \quad \text{eq. 1}$$

Where P_i represents CH_4 production rate in layer i ($\text{nmol cm}^{-3} \text{d}^{-1}$), e_i represents the thickness of layer i (cm) and n represents the numbers of layers in the profile.

Using aerial imagery from 2022, the polygonal tundra at RP was mapped in QGIS, allowing for the discrimination between high-centered polygons, low-centered polygons and throughs (Fig S2). The total area of each geomorphological form was calculated based on the map data (Table S1). Landforms total areas were multiplied by the corresponding potential total active layer methane production (T) to estimate the total CH_4 produced in the polygonal tundra of RP over a day (mol d^{-1}).

2.4 Elemental and isotope composition of the sediment

The total organic carbon (TOC) content of the sediments was measured by combustion using an elemental analyzer (ECS 8020, NC Technologies) combined with a gas chromatograph equipped with a thermal conductivity detector at ULaval facilities (The International Research Laboratory Takuvik). A 100 mg aliquot of sediment was thawed and weighed for each sample. They were then dried in an oven at 60°C for 48 hours and re-weighed to determine their water content. Sediments were then ground using a granite mortar pestle and homogenized using a 1.18 mm pore size sieve to remove roots and rootlets. Instruments were cleaned with ethanol between manipulations. Inorganic carbon was removed from sediments by adding 2.2 mL of 12M HCl in every sample. After reacting for 24 hours, around 8 mg was encapsulated in tin foil capsules. Samples were kept in a desiccator until analyses. Values are expressed as % of carbon contained in the weighed sample (wt. %).

The organic carbon ($\delta^{13}\text{C}$ -TOC) isotopic compositions were measured at UOttawa facilities (Jàn Veizer Stable Isotope Laboratory) using EA-IRMS (Delta Advantage, Thermo Germany). The sample preparation method was the same used for elemental analyses. $\delta^{13}\text{C}$ -TOC values are denoted as $\delta_{\text{‰}} = 10^3 \cdot ((R_{\text{sample}}/R_{\text{standard}}) - 1)$, where R is $^{13}\text{C}/^{12}\text{C}$ and standards refer to the Vienna Pee Dee Belemnite (VPDB).

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2.5 Stable carbon isotopic composition of methane

One incubation vial was analyzed for stable carbon isotopic composition of headspace methane ($\delta^{13}\text{C}-\text{CH}_4$). Stable carbon from methane ebullition samples collected from pondlets were also analyzed. Both types of samples were analyzed with a cavity ring-down spectrometer (PICARRO G2201-i isotopic CO_2/CH_4) equipped with a 16-port distribution manifold and small sample introduction module (SSIM) at McGill (McGill Isotope Biogeochemistry Laboratory). Incubations were kept at 4°C in the dark for 8 months to let the microbial community stabilize and produce sufficient CH_4 for analysis. To stay in the detection range of the analyzer (1.8-1000 ppm CH_4), a small volume of the headspace, proportional to CH_4 concentration in sample, was drawn from the incubation vial (0.2-6 mL). The sample was introduced to the 16-port manifold with a 21G needle connected to a disposable luer lock plastic syringe. Samples were diluted with zero air by the SSIM to reach a volume of 20 mL. Two or three measurements per sample were conducted depending on headspace concentration. Ebullition gases samples were analyzed following the same method. Measured values were corrected with internal certified methane standards (-59 ‰ and -42‰) from AirLiquide and stability of the analyzer was tested with injections of ambient air. Measured values were more precise than $\pm 1.2\%$. All $\delta^{13}\text{C}-\text{CH}_4$ values are expressed relatively to VPDB. While those isotopic analyses results provide valuable insight into methane cycling processes, they should be interpreted with caution in the absence of biological replication.

3 Results

3 Results

3.1 Soil description and composition

TOC content in the sampled soils ranged from 2 to 47 wt. %, with no clear trend in relation to depth (Fig 3, a, c). The RP polygonal patterned ground featured organic soils with TOC content ranging from 14 to 47 wt. % (Fig 3, c). The TP coastal polygonal patterned ground also featured organic soils with TOC content ranging from 2 to 37 wt. % (Fig 3, a).

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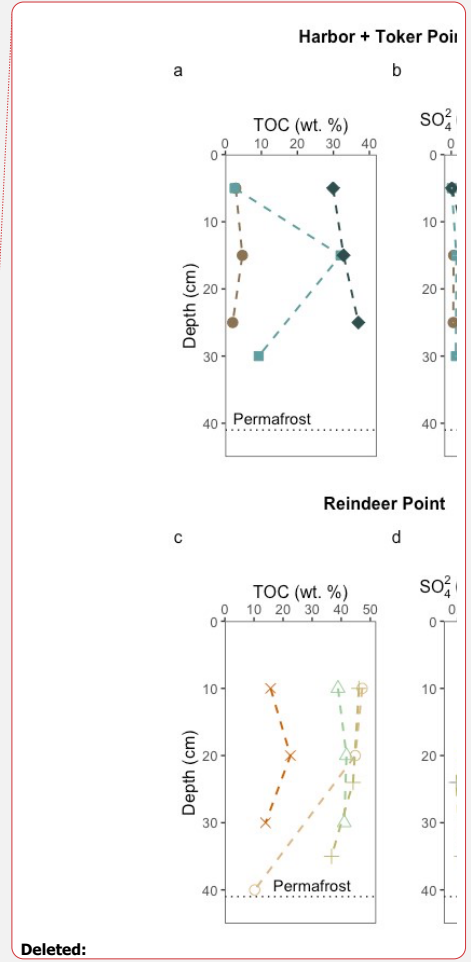
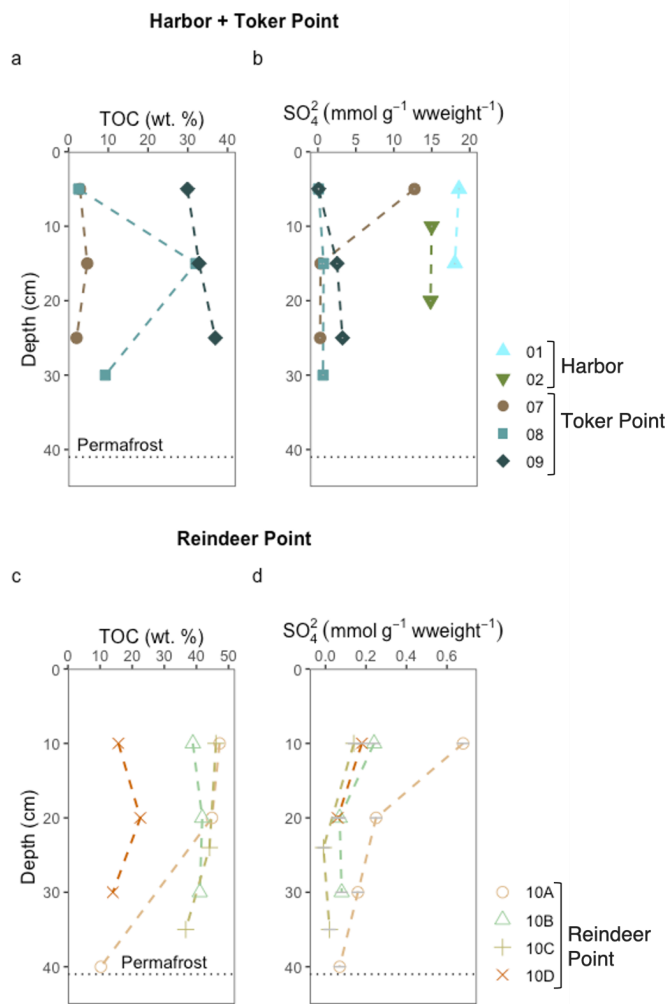
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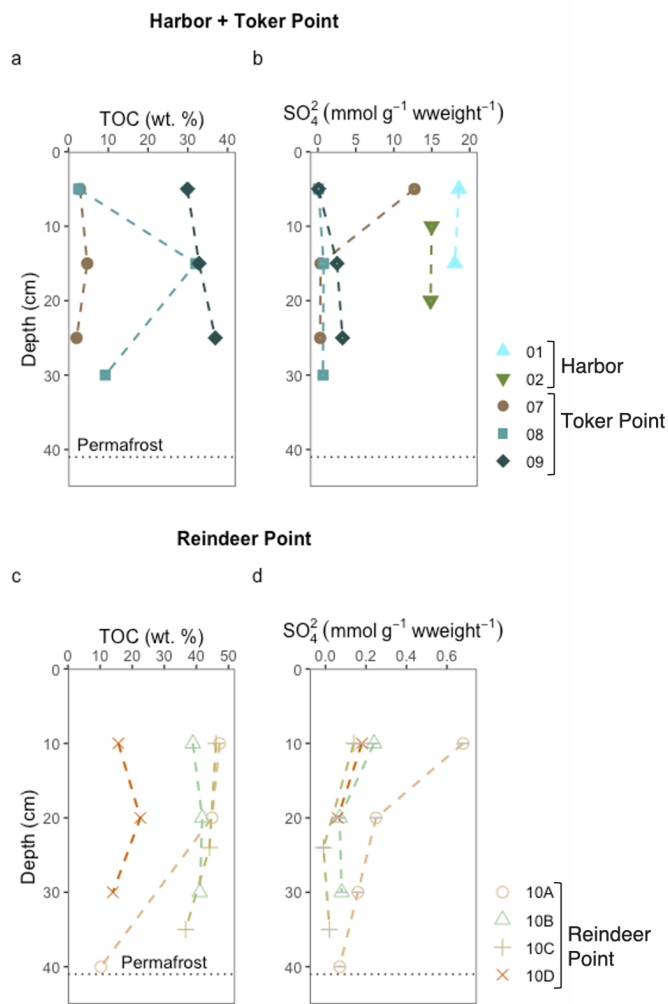


Figure 3. Total organic carbon and sulfate (SO_4^{2-}) concentrations in sediment or soil of different sites in this study. The datasets are separated into two for clarity. The upper part of the figure (panels A and B) displays the data of the marine site Harbor (profile 01 and 02), and the coastal site, Toker Point (profile 07, 08 and 09). The lower part of the figure (panel C and D) displays the data from the inland site Reindeer Point (profile 10A, 10B, 10C and 10D). The black horizontal dotted line in each graph represents the permafrost-active layer or talik interface except for the Harbor

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site, where the talik is much deeper but not measured. TOC data from Harbor site is not available. A uniform color pattern is used throughout this manuscript.

RP, the inland site, had low sulfate and chloride concentrations relative to TP, the coastal site (Fig 3 (b), (d) and S2). Sulfates at RP ranged from null concentrations to $0.68 \pm 0.03 \text{ mmol g}^{-1} \text{ wweight}^{-1}$, while at TP, profiles exhibited varying concentrations and patterns in relation to depth. Sulfate concentrations, ranged from 0.07 ± 0.03 to $12.72 \pm 0.03 \text{ mmol g}^{-1} \text{ wweight}^{-1}$. Profile 07, the low-centered polygon, exhibited the highest sulfates concentrations of all TP site at its surface ($12.72 \pm 0.03 \text{ mmol g}^{-1} \text{ wweight}^{-1}$), with concentrations decreasing drastically with depth, reaching $0.29 \pm 0.03 \text{ mmol g}^{-1} \text{ wweight}^{-1}$ at 25 cm (Fig 3, a). In profile 09, the high-centered polygon, sulfate concentrations increased with depth ranging from $0.09 \pm 0.03 \text{ mmol g}^{-1} \text{ wweight}^{-1}$ at 5 cm to $3.2 \pm 0.03 \text{ mmol g}^{-1} \text{ wweight}^{-1}$ at 25 cm. Finally, profile 08, characterized as a polygonal trough, had sulfate concentrations ranging from 0.07 ± 0.03 to $0.75 \pm 0.03 \text{ mmol g}^{-1} \text{ wweight}^{-1}$. The highest sulfate concentrations measured in this study were found in the sediments of the Harbor site, with a mean value of $16.6 \text{ mmol g}^{-1} \text{ wweight}^{-1}$ (Fig 3, a).

3.2 Methane production

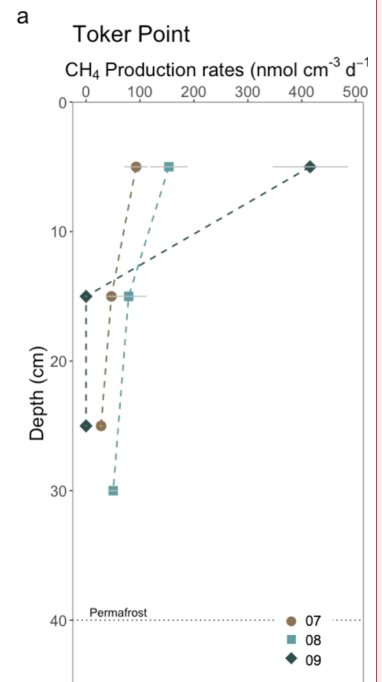
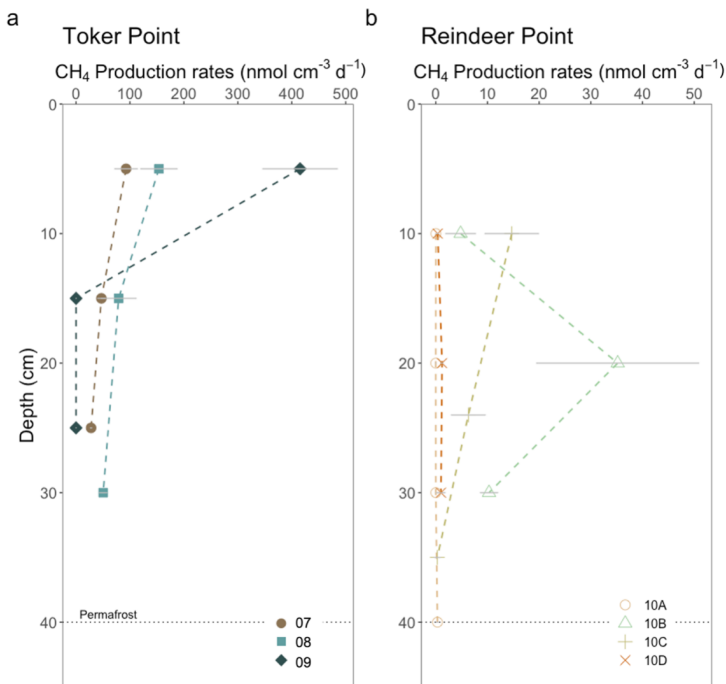
Rates of CH_4 production in incubations of sediment and soil with brackish water were undertaken at the three studied sites: RP, TP and Harbor. Production rates ranged from null to $415.4 \pm 69.2 \text{ nmol cm}^{-3} \text{ d}^{-1}$ (Fig 4) throughout all samples in this study. At RP, the maximum CH_4 production rate of $35.2 \pm 15.7 \text{ nmol cm}^{-3} \text{ d}^{-1}$ was measured in the trough profile (10B) at a depth of 20 cm. Lower values were obtained for the surface and at the talik-permafrost interface. The low-centered polygon (10C) had its maximum CH_4 production rate in the surface, decreasing with depth. High-centered polygons (10A and 10D) had very low production rates along their depth profiles ranging between null to $1.2 \pm 0.2 \text{ nmol cm}^{-3} \text{ d}^{-1}$. Both water-saturated trough and low-centered polygon (10B,10C) had relatively high CH_4 production rate compared with the high-centered polygon profiles (10A, 10D), which were water-unsaturated.

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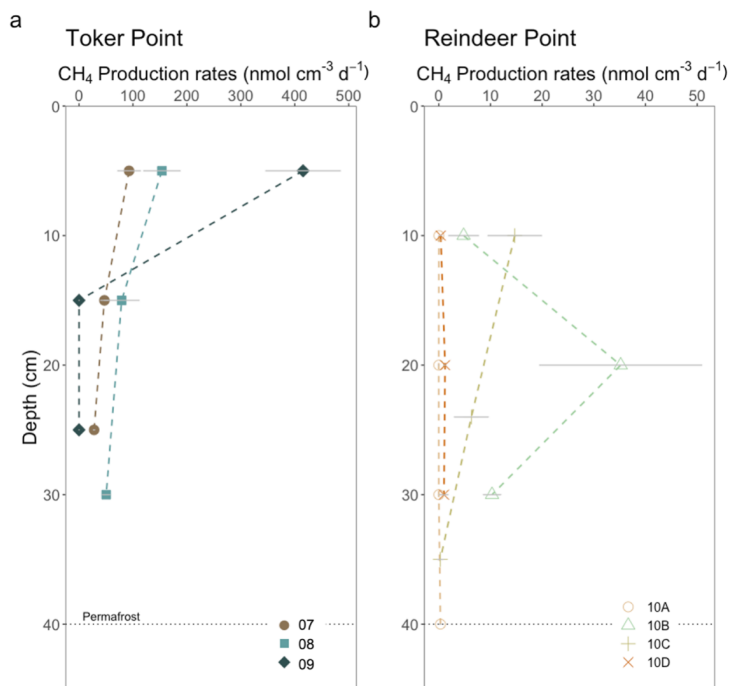


Figure 4. CH₄ production in incubations of soil and sediment with brackish water from (a) TP and (b) RP. Each datapoint represent the mean value of three incubations. The error bars in grey lines are equal to the standard deviation of the three separate incubations. Each profile corresponds to a specific landform. At Toker Point (panel A), profile 07 is from a low-centered polygon, profile 08 is from a trough and profile 09 is from a high-centered polygon. At Reindeer point (panel B), profile 10A is from a high-centered polygon, profile 10B is from a trough, profile 10C is from a low-centered polygon and profile 10D is from a high-centered polygon.

At TP, a maximum CH₄ production rate was recorded in profile 09, the high-centered polygon at 415.4 ± 69.2 nmol cm⁻³ d⁻¹ at the uppermost depth but it quickly decreased in the subsurface. Profile 08, the low-centered polygon, had lower sub-surface CH₄ production rates, but rates decreased less drastically with depth with values being relatively high at the permafrost-talik and permafrost-active layer interface, respectively. Profile 07 had values ranging from 27.9 ± 1.5 nmol cm⁻³ d⁻¹ to 92.8 ± 21.2 nmol cm⁻³ d⁻¹ and profile 08 had values ranging from 50.4 ± 7.2 nmol cm⁻³ d⁻¹ and 153.7 ± 33.9 nmol cm⁻³ d⁻¹ (Fig 4). In general, at TP, the coastal site, much higher CH₄ production rates were measured than at RP, the inland site (Fig 4). The mean CH₄ production rate measured at RP was 5.7 nmol cm⁻³ d⁻¹, while at TP it was 96.2 nmol cm⁻³ d⁻¹. The incubations with silty-clay Harbor sediments did not have measurable CH₄ production rates (Fig S5).

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Estimated total CH_4 production rates were calculated for each geomorphological landforms of RP and TP sites. At RP, the total CH_4 production estimated for the high-centered polygons (profile 10A and 10D), low-centered polygon (profile 10C) and trough (profile 10B) were $0.3 \pm 0.1 \text{ mmol m}^{-2} \text{ d}^{-1}$, $2.4 \pm 1.0 \text{ mmol m}^{-2} \text{ d}^{-1}$ and $5 \pm 2 \text{ mmol m}^{-2} \text{ d}^{-1}$, respectively (Fig 5). At TP, the total CH_4 production estimated for the high-centered polygon (profile 09), the low-centered polygon (profile 07) and the through (profile 08) were $41.5 \pm 6.9 \text{ mmol m}^{-2} \text{ d}^{-1}$, $16.8 \pm 3.0 \text{ mmol m}^{-2} \text{ d}^{-1}$ and $28.3 \pm 7.4 \text{ mmol m}^{-2} \text{ d}^{-1}$, respectively (Fig 5). In all landforms, the total CH_4 production rates were higher in the coastal site, TP, than the inland site, RP.

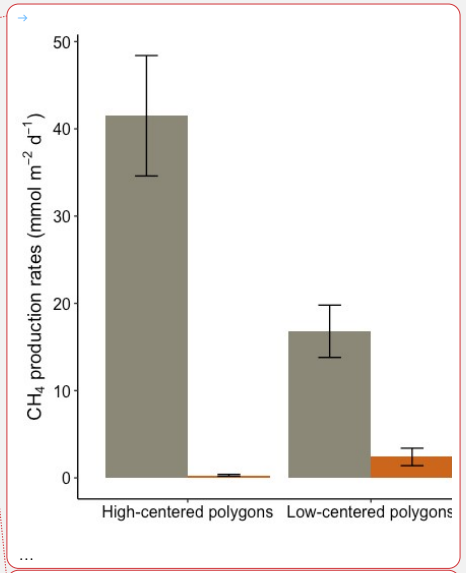
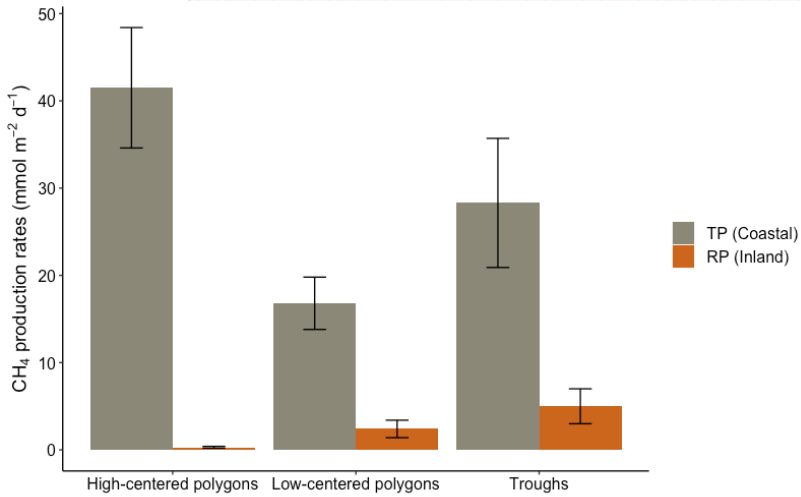


Figure 5. Total CH_4 production rates (T) at Toker Point (Coastal) and Reindeer Point (Inland) organized by geomorphological forms of high-centered polygons, low-centered polygons and troughs. High-centered polygons at RP is the mean of two

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profiles. All other landforms at RP and TP are one profile. The uncertainty on T is propagated from the uncertainty of individual CH_4 production rates, not averages from replicate sites.

3.3 Isotopic composition of $^{13}C-CH_4$

In parallel with CH_4 production rates, one incubation vial per depth was used to measure the stable carbon isotopic composition of the CH_4 produced. At RP, the $\delta^{13}C-CH_4$ of the first sampled depth (10 cm) ranged from -81.3‰ to -89.4‰. At TP, the coastal site, the $\delta^{13}C-CH_4$ signature of the first sampled depth (5 cm) ranged from -47.1‰ and -51.6‰. The values cluster together based on site, suggesting surface OM degradation processes are most similar within sites than between sites (Fig. 6). Profiles at RP became progressively enriched in ^{13}C with depth, except for profile 10C where a more depleted value was observed at 35 cm. Conversely, at TP, profiles became depleted in ^{13}C with depth, except for profile 08 where an enrichment was measured between 15 and 30 cm.

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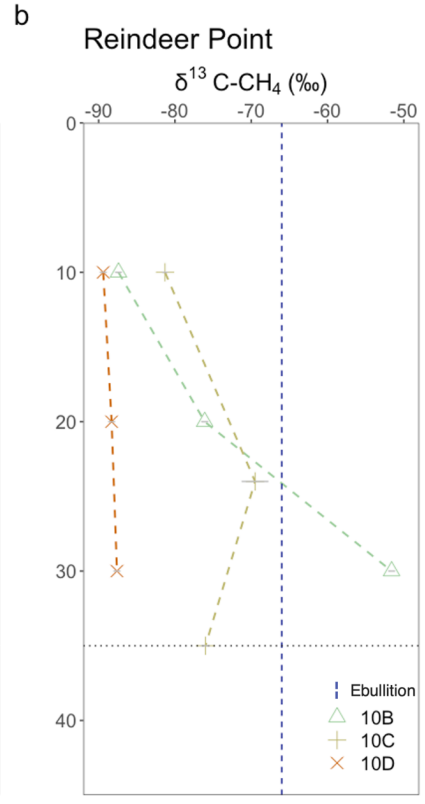
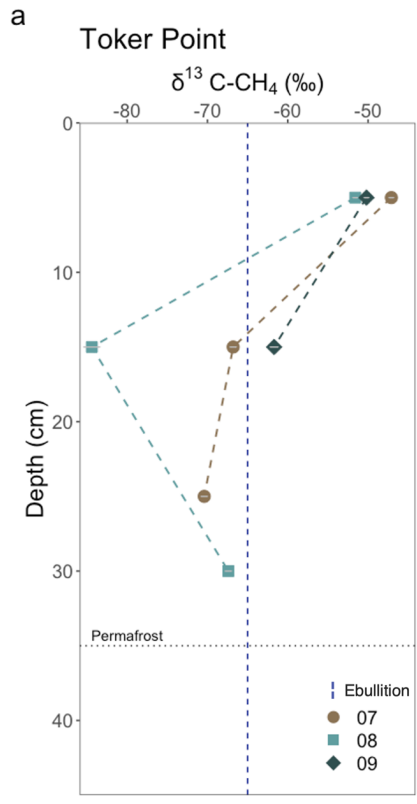
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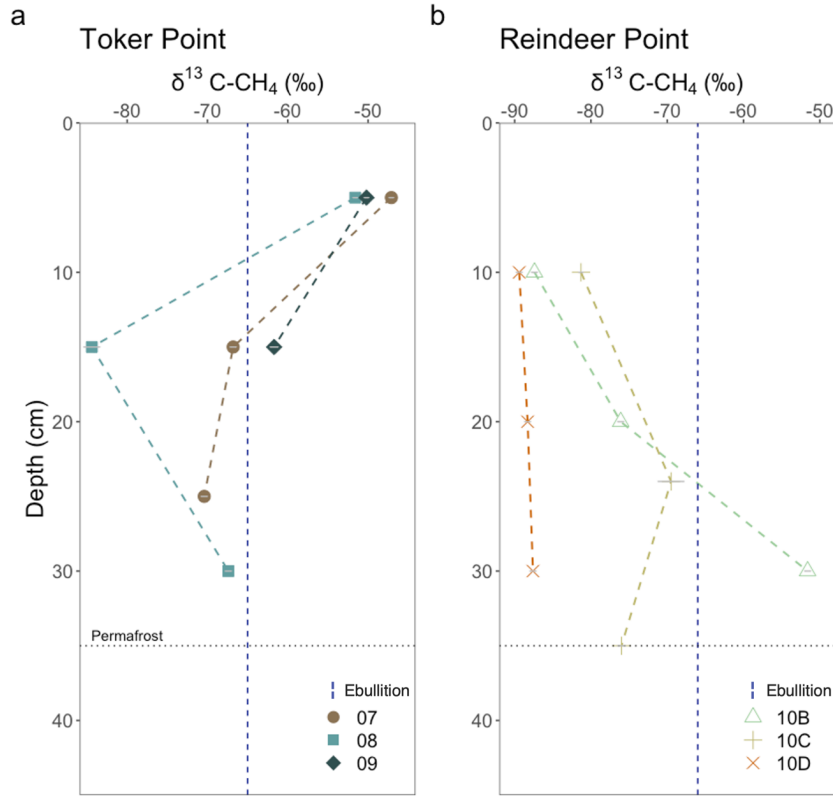


Figure 6. Isotopic composition of CH_4 produced in brackish water incubations from (a) TP and (b) RP. Each datapoint corresponds to the mean value of two or three measurements done on one incubation, depending on the headspace concentration. The dashed vertical lines correspond to in situ ebullition CH_4 collected in pondlets at each sampling site ($n=1$). These values give information on the pathways used by the soil microbes to produce CH_4 . $\delta^{13}\text{C}$ between -65‰ and -50‰ is typically associated with acetoclastic methanogenesis, while $\delta^{13}\text{C}$ between -110‰ and -60‰ is associated with hydrogenotrophic methanogenesis (Hornibrook et al., 1997, 2000). The grey error bars on each point represents the analytical uncertainty on the measured value. If not visible, the uncertainty is smaller than the point.

Ebullition samples from pondlets were also measured for stable isotopes. The ebullition samples represent the net $\delta^{13}\text{C}$ signature of methane produced in the sediments of pondlets at RP and TP. At RP, CH_4 ebullition from a sampled thaw pond had a $\delta^{13}\text{C}$ of -66.1‰ (fig 6). At TP, CH_4 ebullition from a sampled pondlet had a $\delta^{13}\text{C-CH}_4$ of -65.0‰ (fig 6).

4 Discussion

4 Discussion

4.1 Addition of brackish water to anoxic incubations did not strongly suppress methanogenesis

Before discussing the effects of brackish water addition in incubation experiments, it is important to clarify the role of sulfate measured in situ within soil and sediment profiles. Across the studied sites, sulfate concentrations varied

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47 with depth and between landforms; however, this spatial variability did not show a consistent relationship with
48 methane production rates measured in the incubations (Fig S4). A few layers clearly contained higher sulfate amounts.
49 However, layers characterized by higher or lower sulfate concentrations did not systematically correspond to lower or
50 higher CH₄ production, indicating that in situ sulfate availability alone does not explain the observed patterns in
51 methane production across profiles. This interpretation is subject to important limitations. Sulfate and chloride
52 concentrations were measured at single points within each profile and were not replicated across multiple locations
53 within the same landform, preventing resolution of fine-scale spatial heterogeneity in electron-acceptor availability.
54 As a result, sulfate concentrations are interpreted here as first-order indicators of geochemical context rather than as
55 spatially representative or mechanistic controls on methane production. Given these constraints, we focus the
56 following discussion on the experimental addition of sulfate via brackish water during anoxic incubations, which
57 evaluate how episodic marine influence may affect methane production potential in coastal permafrost environments.

58 Despite the addition of brackish water containing sulfate to incubations, the range of CH₄ production rates
59 measured in this study is consistent with reports for anaerobic incubations of recently thawed permafrost soils,
60 suggesting that the input of brackish water to some coastal systems may not inhibit CH₄ production. For example, in
61 the talik of Big Trail Lake, a young thermokarst lake in the interior of Alaska, CH₄ production rates based on
62 incubations ranged between 4.7 and 16.1 nmol cm⁻³ d⁻¹ (Pellerin et al., 2022), while in incubations from Vault Lake,
63 another thermokarst lake in the interior of Alaska, CH₄ production rates varied between 11.1 and 275 nmol cm⁻³ d⁻¹
64 (Heslop et al., 2015). In active layer incubations from the Yamal Peninsula in NW Siberia (Russia), CH₄ production
65 rates of incubations varied between 0.1 and 33.8 nmol cm⁻³ d⁻¹ (Heyer et al., 2002). This indicates that overall, the
66 CH₄ production rates measured at both TP and RP are within the range observed in typical ice-rich permafrost settings
67 and reasonable for the environment studied (Fig. 4). ~~We note that our experimental design did not include parallel~~
68 ~~incubations without brackish water or with sulfate concentration gradients; therefore, our interpretation relies in part~~
69 ~~on comparison with previous incubations of Tuktoyaktuk soils conducted without brackish water addition (Lapham et~~
70 ~~al., 2020) and should be regarded as exploratory rather than definitive.~~ Lapham et al. (2021) conducted sediment
71 incubation experiments using a core collected from the coast of the Tuktoyaktuk Peninsula. In their study, CH₄
72 production was measured under anaerobic conditions at 15 °C, without the addition of water, over a 35-day period.
73 The reported CH₄ production rate was 0.07 nmol cm⁻³ d⁻¹. Although their incubations were performed over a shorter
74 duration and at a significantly higher temperature than those in the present study, the measured rate reflects CH₄
75 production under relatively natural, unamended conditions. This value is comparable to the lowest CH₄ production
76 rates measured in our incubations at both the coastal (TP) and inland (RP) sites and provides a useful reference for
77 CH₄ production under unamended conditions.

78 The novel aspect of this study is that it attempts to understand marine influence on OM degradation by addition
79 of brackish water to sediment and soil incubations of a fully marine site (Harbor), one that is periodically submerged
80 (TP) and never submerged (RP). This simulates the input of seawater to the active layer and taliks of tundra soils (RP)
81 as well as providing reference sites with a high marine influence (Harbor and TP). We hypothesized that the addition
82 of locally obtained brackish water, which contained sulfate (5.7 mmol L⁻¹), to the incubations, would suppress CH₄
83 production in RP, the inland site and potentially also at TP, the coastal site. This reasoning is because supplying sulfate
84 to low sulfate organic-rich sediment would promote sulfate reduction, which is thermodynamically more favorable
85 than methanogenesis, thereby competitively inhibiting it (Lovely and Klug, 1983; Oremland and Polcin, 1982). This
86 hypothesis is also consistent with field observations; organic matter mineralization in brackish wetlands is consistently
87 dominated by bacterial sulfate reduction (Bridgman et al., 2013; Torres-Alvarado et al., 2005) where little to no
88 CH₄ emissions are observed (Pönisch et al., 2022; Petersen et al., 2023; Kroeger et al., 2017). However, recent field
89 studies show that in coastal permafrost soils, inundation and low sulfate concentrations do not necessarily suppress
90 methanogenesis (Jenrich et al., 2025; Jenrich et al., 2024; Yang et al., 2023). These contrasting observations reveal a
91 key knowledge gap in how marine influence controls carbon mineralization pathways in permafrost systems. By
92 experimentally testing brackish water additions across sites with contrasting marine exposure, our study provides new
93 mechanistic insight into the regulation of OM degradation and CH₄ production under ongoing Arctic coastal change.

94 RP had low sulfate concentrations before addition of the brackish water but so did many of the profiles from TP
95 (Fig 3). In the Harbor sediments, no methane production was observed (Fig S5). This is consistent with the competitive
96 inhibition of methanogenesis by energetically favorable redox reactions with electron acceptors like oxygen, nitrate,
97 iron oxides or sulfate that is typical of marine systems e.g. (Martens and Berner, 1974) as well as the potential for
98 anaerobic oxidation of methane (AOM). Given that the Harbor sediments already had high sulfate concentrations, the
99 lack of methane production with addition of brackish water was expected. However, strong CH₄ production was
100 observed in the incubations of both the coastal site TP and the inland site RP, indicating that CH₄ production was not
101 halted by the addition of sulfate via the brackish water addition at those sites. While sulfate reduction rates were not
102 measured and therefore not demonstrated directly in our incubations, a strong sulfide smell was recorded when

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113 opening most of the incubations at the end of the experiment. ~~This observation may indicate~~ the coexistence of sulfate
114 reduction and methanogenesis during the incubations. ~~However, to rigorously assess this observation, future studies~~
115 ~~should include tracer-based sulfate reduction assays and microbial functional gene analysis.~~

116 Coexistence of sulfate and methanogenesis within complex sediment systems such as estuarine, coastal and salt
117 marsh sediments, ~~as well as thermokarst lasens~~ has been widely reported (Lovely et al., 1982; Oremland and Taylor,
118 1978; Sela-Adler et al., 2017; [Yang et al., 2023](#)). Two main mechanisms are invoked to explain this co-existence in
119 our incubation experiment: (1) noncompetitive methanogenesis (i.e. methylotrophic methanogenesis) and (2)
120 syntrophic methanogenesis. (1) Noncompetitive substrates are substrates like methanol and methylamines that are
121 used by methanogens alone and cannot be used with electron acceptors like sulfate (Lovley and Klug, 1983; Oremland
122 et al., 1982). Noncompetitive substrates are thus microbially converted to CH₄, even in sediments with high sulfate
123 concentrations (Maltby et al., 2018; Yuan et al., 2019). For example, in salt marshes, where high sulfate concentrations
124 are often found, elevated CH₄ emissions are suggested to mainly stem from noncompetitive methanogenesis (Comer-
125 Warner et al., 2022; Poffenbarger et al., 2011; Yuan et al., 2019). In the sulfate reducing zone of sediments from the
126 Baltic Sea, where ample sulfate is found in the porewaters, seasonal methanogenesis rates were measured up to 1.3
127 nmol cm⁻³ d⁻¹ due to noncompetitive substrates (Maltby et al., 2018). In permafrost soils, methanol, methylamines
128 and the microorganisms capable of degrading them have been observed but their concentrations are typically low
129 (Coolen and Orsi, 2015; Kramshoj et al., 2018). However, our study sites are on a coast undergoing a rapid
130 transgression which may be driving imbalances between substrate supply and microbial abundances. The rates of
131 methane production observed at RP and TP of up to 154 nmol cm⁻³ d⁻¹ contrast with reported values for
132 methylotrophic methanogenesis (Maltby et al., 2018). Based on these numbers, noncompetitive substrates likely play
133 a small role in the total methane production at our study sites but further investigation into methylotrophic methane
134 production in coastal environments will allow to document the overall role of methylotrophic methane production in
135 coastal permafrost settings. ▽

136 (2) Syntrophic methanogenesis occurs when molecular hydrogen produced by acetoclastic sulfate-reducing
137 bacteria is used by hydrogenotrophic methanogens. In this syntrophy, the chemical energy is shared via interspecies
138 hydrogen transfer (Ozuolmez et al., 2015). For instance, in permafrost soils of Sweden, it was demonstrated that
139 syntrophic methanogenesis was favored in anoxic and water-saturated soils by an elevated abundance in methanogens
140 and their syntrophic partners (Keuschnig et al., 2022). As the incubation experiment in our study at RP and TP featured
141 water-saturated and anoxic environments, syntrophic methanogenesis could participate in the co-occurrence of ~~sulfate-~~
142 ~~reduction~~ and methanogenesis. This mechanism is consistent with most incubations producing methane with a $\delta^{13}\text{C}$
143 value in the range of hydrogenotrophic methanogenesis (see below). ▽

144 Measuring methane production through incubations inherently has limitations as they prevent continuous inputs
145 of microorganisms, fresh OM and nutrients that would occur in the natural environment. This can create a “bottle
146 effect”, which leads to restrictions in microbial community composition, limits the input of nutrients and leads to the
147 accumulation of metabolites which would normally be degraded (Ionescu et al., 2015). Typically, overestimation of
148 microbial processes rates is observed compared to *in situ* data (Sherr et al., 1999). The overestimation of CH₄
149 production rates by incubations relative to the *in situ* rates are difficult to assess because of a lack of data in permafrost
150 environments (Heslop et al., 2020). Furthermore, a lag time between the start of anaerobic incubations and maximum
151 CH₄ production rate is widely documented, which appears to be the case for both active layer and thawed permafrost
152 incubations (Holm et al., 2020; Knoblauch et al., 2018; Knoblauch et al., 2013; Roy Chowdry et al., 2015). Drier or
153 water-unsaturated conditions lead to a longer lag time before the onset of maximum CH₄ production (Treat et al.,
154 2015). Microbial community composition in the soil or sediment also exerts a strong control on the organic carbon
155 degradation and has been shown to change throughout the incubations (Holm et al., 2020). Low initial methanogen
156 population in soils can contribute to this lag time, but other factors such as disturbance of sediment during sampling,
157 substrate availability and redox state can also contribute to the observed lag time in some incubations (Treat et al.,
158 2015; Roy Chowdry et al., 2015). ▽

159 Furthermore, it is also possible that a “priming effect” from the addition of brackish water in incubations could
160 have supercharged OM degradation with marine organic carbon, nutrients and microorganisms (Bianchi, 2011), which
161 may have enhanced CH₄ production. However, this priming effect was not observed in the Harbor sediments which
162 were amended with the same brackish water. Furthermore, CH₄ ebullition samples collected from pondlets adjacent
163 to RP and TP exhibited broadly similar $\delta^{13}\text{C}$ values to methane produced in incubations (Fig 6), suggesting a similitude
164 in microbial degradation pathways to methane *in situ* and in the incubations. Despite these uncertainties, our dataset
165 shows clear depth trends and landscape-level variations, indicating that even under brackish water addition, local
166 conditions will strongly influence CH₄ production. ▽

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175 **4.2 CH₄ production pathways depend on hydrology and organic matter lability.**
 176 The addition of brackish water resulted in incubation conditions being water-saturated in all cases, but it
 177 appears that biological and hydrological conditions of the polygonal patterned grounds influenced the magnitude of
 178 CH₄ production, nonetheless.

179 In all landforms, CH₄ production rates were lower at the inland site, RP than at TP, the coastal site (Fig 4).
 180 Inland, low-centered polygons and troughs have typically higher CH₄ fluxes than unsaturated landforms like high-
 181 centered polygons (Roy Chowdry et al., 2015; Martin et al., 2018; Zheng et al., 2018) which indicates they may also
 182 have higher CH₄ production rates. Within sites in our study, brackish water amended incubations of high-centered
 183 polygon soils had lower CH₄ production rates, while brackish water amended incubations of troughs and low-centered
 184 polygons had higher CH₄ production rates (Fig 4). This indicates that for the degradation of organic matter into CH₄
 185 in tundra soils, increasing seawater interactions through coastal processes, such as submersion due to subsidence or
 186 increased storm severity, resulting in the input of seawater in terrestrial soils, does not halt CH₄ production. It also
 187 shows that landforms and local hydrology remain important in controlling the microbial communities which affects
 188 the resulting CH₄ production. Differences among sites and along landforms in each specific sites are generally large
 189 and exceeded the range of variability as shown by the uncertainty (Fig 5), supporting the use of means with standard
 190 deviations to convey contrasts without formal statistical tests. This approach allowed us to highlight general
 191 differences in CH₄ production potential and geochemical context across coastal and inland sites.

192 Marine OM and nutrient inputs from tides and storm surges may contribute to the higher lability of OM and
 193 could fuel greater fermentation (Valdemarsen and Kristensen, 2010). It was reported that 8.7% of the organic carbon
 194 in nearshore sediments of Herschel Island, Beaufort Sea, came from marine sources (Couture et al., 2018). This is
 195 relevant for the TP site because while $\delta^{13}\text{C}$ signature of soils showed that terrestrial OM is dominant (Fig S6), marine
 196 OM may get transported and deposited in coastal soils during high tides and storm surges. Although our analyses
 197 could not detect the presence of marine OM in TP soils, the higher CH₄ production rates recorded in the incubations
 198 of TP, relatively to those of RP could in part be explained by marine OM and nutrient inputs. Interestingly, the high-
 199 centered polygon at TP, profile 09 (Fig 4), did not behave in a predictable manner, since it had very high CH₄
 200 production rates on the surface. This elevated methane production rate coincided with the presence of substantial
 201 goose fecal deposits at TP, profile 09. While this observation suggests a potential local input of labile organic matter
 202 and nutrients (e.g., N and P) and possibly a distinct surface microbial community, no direct measurements were
 203 conducted to establish a mechanistic link. This site-specific observation is therefore reported as contextual field
 204 information rather than evidence of causation. Lower in the profile, CH₄ production rates were very low, characteristic
 205 of the CH₄ production rates observed in water-unsaturated high-centered polygons (Fig 4). Therefore, in this instance,
 206 proximity with the coast may have influenced CH₄ production through the presence of fauna.

207 Stable carbon isotopic signature of CH₄ provides insights on the microbial processes involved in
 208 methanogenesis and on substrates used. $\delta^{13}\text{C}$ -CH₄ between -65‰ and -50‰ is typically associated with acetoclastic
 209 methanogenesis, while $\delta^{13}\text{C}$ -CH₄ between -110‰ and -60‰ is associated with hydrogenotrophic methanogenesis
 210 (Hornibrook et al., 1997, 2000). The stable isotopic signature of methylotrophic methanogenesis is between -83‰ and
 211 -72‰ (Penger et al., 2012), which overlaps with the hydrogenotrophic interval, precluding us from separating these
 212 two metabolic pathways. At RP, except for profile 10B, $\delta^{13}\text{C}$ -CH₄ had more negative values, consistent with the
 213 processing of recalcitrant organic matter through the hydrogenotrophic production pathway (Heffernan et al., 2022;
 214 Hodgkins et al., 2014). Profile 10B, a polygonal trough, had less negative ^{13}C -CH₄ values more consistent with
 215 acetoclastic methanogenesis (Hornibrook et al., 1997). At TP, the coastal polygonal tundra, $\delta^{13}\text{C}$ -CH₄ at 5 cm depth
 216 is less negative, consistent with methanogenesis with more labile organic carbon and the acetoclastic production
 217 pathway (Hodgkins et al., 2014), transitioning to more negative values, associated to hydrogenotrophic production
 218 with depth. This shift suggests an input of labile OM in TP surface and sub-surface soils. This may be due to the labile
 219 OM from abundant geese fecal matter that was observed in the surface. It is also possible that *Carex sp.*, the dominant
 220 plant species of the site, may be a source of labile fermentation precursors (Galand et al., 2010; Liebner et al., 2015).
 221 To evaluate whether $\delta^{13}\text{C}$ -CH₄ covaried with other geochemical properties measured in this study, $\delta^{13}\text{C}$ -CH₄ values
 222 were examined alongside TOC content and sulfate concentrations; however, no consistent relationships were observed
 223 across landforms or depths (Fig. S4), indicating that methanogenic pathway signatures are not straightforwardly
 224 predicted by bulk TOC or sulfate availability at the scale investigated. However, it is clear that future work should
 225 integrate measurements of organic matter degradation, microbial community composition, and pore water chemistry
 226 to better resolve the mechanisms driving spatial variability in methane production.

227 To evaluate whether $\delta^{13}\text{C}$ -CH₄ covaried with other geochemical properties measured in this study, $\delta^{13}\text{C}$ -CH₄ values
 228 were examined alongside TOC content and sulfate concentrations; however, no consistent relationships were observed
 229 across landforms or depths (Fig. S4), indicating that methanogenic pathway signatures are not straightforwardly

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- Commented [RLA25]: Addition of supplementary figure... [24]

279 predicted by bulk TOC or sulfate availability at the scale investigated. However, it is clear that future work should
280 integrate measurements of organic matter degradation, microbial community composition, and pore water chemistry
281 to better resolve the mechanisms driving spatial variability in methane production.

282 4.3 Total CH₄ production rates are comparable to the net CH₄ fluxes measured in similar environments.

283 In a polygonal terrain of the Tuktoyaktuk Coastlands, net CH₄ fluxes from the center of high-centered polygons
284 and troughs derived from flux chambers were measured to be 1.9 ± 20.4 mmol m⁻² d⁻¹ and 13.0 ± 20.4 mmol m⁻² d⁻¹
285 respectively (Martin et al., 2018). These overlap with values of estimated total CH₄ production derived from the
286 brackish water amended incubation experiments (Fig. 5). It is clear from the large variations in measured CH₄
287 emissions from the study of Martin et al., (2018) that incubations to estimate total active layer CH₄ production rates
288 can discern small differences due to local variations that stem mostly from the polygonal features. For example, at
289 RP a comparable polygonal terrain located in the same study area of Martin et al., (2018), the total CH₄ production of
290 high-centered polygons and trough were 0.3 ± 0.1 mmol m⁻² d⁻¹ and 5.0 ± 2.0 mmol m⁻² d⁻¹ (Fig. 5), respectively which
291 are significantly different. This indicates the role of polygonal forms in controlling the activity of microbial
292 communities which controls CH₄ production and the potential to scale more accurately CH₄ production at the
293 landscape level based on landform distributions.

294 Interestingly, TP, the coastal site, had an estimated total CH₄ production rate comparable to emissions of a St.
295 Lawrence estuary salt marsh which had a CH₄ flux of 24 ± 14.4 mmol m⁻² d⁻¹ (Comer-Warner et al., 2022). The St.
296 Lawrence estuary salt marshes are affected by freeze-thaw cycles associated with seasons comparable to the freeze-
297 thaw cycles observed in the active layer of Tuktoyaktuk coastlands despite lacking some characteristics features of
298 our site like the presence of permafrost and rapid coastal erosion rates. CH₄ emissions and production within areas of
299 coastal influence thus appear of similar magnitude. By comparison, mangrove forests, which are a major global source
300 of CH₄ but a very different environment from coastal Arctic polygon terrain, had average CH₄ fluxes to the atmosphere
301 of 0.3 ± 0.1 mmol m⁻² d⁻¹ (Rosentreter et al., 2018). In another study, the average measured CH₄ flux from a Yangtze
302 Estuary (China) tidal salt marsh, with a subtropical monsoon climate, was 2.4 mmol m⁻² d⁻¹ (Li et al., 2021). These
303 reported values are similar to our study as well as other studies in the region. When considered alongside the global
304 distribution of coastal wetlands, this similarity in flux magnitude becomes particularly relevant. Tropical coastal
305 wetlands are dominated by mangroves (~147,000 km²), whereas Arctic wetlands cover approximately 3.5 million km²
306 (Wortington et al., 2024). Even if only a small fraction of Arctic wetlands is located within coastal zones, their total
307 extent is comparable to the global mangrove area (Wortington et al., 2024), suggesting that permafrost Arctic coastal
308 wetlands could represent a non-negligible component of the global CH₄ budget and warrant further investigation.

309 The calculated total methane production rates (T) from TP and RP do not take into account aerobic and anaerobic
310 oxidation of CH₄, which will most likely reduce fluxes of CH₄ from these sites. Studies and models of Arctic soils
311 emissions have highlighted that aerobic methanotrophy could consume more than half of the CH₄ produced in soils,
312 greatly limiting surface emissions (Oh et al., 2020; Zheng et al., 2018). Furthermore, AOM has been shown to play
313 an important role in attenuating CH₄ production in soils and sediments (Segarra et al., 2013; Winkel et al., 2019) but
314 did not appear to influence significantly CH₄ production in incubations with thermokarst lake sediments (Lotem et al.,
315 2023). While AOM represents a major sink for CH₄ in marine sediments (Knittel and Boetius, 2009; Reeburgh, 2007),
316 the very different biogeochemical and hydrological characteristics of our coastal sites suggest that the role of AOM in
317 these environments may diverge from that observed in fully marine systems. Recent work in coastal thermokarst
318 lagoons, which can present key similarities to our coastal study sites due to episodic or persistent brackish water
319 intrusion, have been shown to exhibit strong AOM control on CH₄ dynamics, particularly in sulfate-rich settings where
320 AOM may constitute a major CH₄ sink (Yang et al., 2023). For the discussion of this study, we compared results of
321 brackish water incubations to CH₄ emissions measured in other landscapes. Such comparisons provide valuable
322 context by comparing long-term microbial production processes with net atmospheric fluxes. However, we emphasize
323 that CH₄ production rates cannot be directly equated to CH₄ emissions.

324
325
326
327
328
329
330 **Table 1.** Total methane production in a context of brackish water addition in high-centered polygons, low-centered
331 polygons and troughs during growing season applied to the spatial scale of the polygonal landscape of RP. Two
332 samples were taken for the high-centered polygon. The mean active layer and talik depth of the region is 35 cm. The
333 error represents the propagation of the analytical uncertainty from the incubations results. Geomorphological form.

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Relative area of each landform (km²)

Estimated Total CH₄ production (mol d⁻¹)

High-centered polygons

Low-centered polygons

Troughs

5 Conclusions

The primary hypothesis for this study was that an increase in waterlogged environments due to coastal flooding and inundation processes would not enhance CH₄ production because of sulfate present in coastal waters. However, our incubation experiment revealed high CH₄ production rates in the presence of sulfates. Additionally, waterlogged conditions attributed to the ebb and flow of tides, seems to favor anoxic OM degradation and may potentially provide inputs of fresh OM and nutrients from marine sources, contributing to the elevated CH₄ production rates measured in the coastal setting of TP. Moreover, no conclusive explanation for the co-occurrence of sulfate reduction and methanogenesis in our brackish water incubations was identified but based on evidence, we suggest syntrophic methanogenesis could support this co-occurrence. More investigation on methylotrophic methanogenesis in coastal soils are needed as it can be an important process in saline environments (Conrad, 2020). Future studies should investigate CH₄ oxidation processes in greater detail, as they could provide crucial insights into Arctic coastal carbon cycling in sediments and soils affected by changing sea level.

Data availability

All raw data of incubation experiment and other analyses performed and generated by study are available as supplementary information.

Competing interests

The authors declare that they have no conflict of interest.

Author contributions

AP designed the experiment; ARL executed the experiments and analyses. AP, ARL, DW, RL participated in the fieldwork. PMJD provided lab space, equipment and insights for the stable carbon isotopes analyses on incubation CH₄. RL performed all GIS analyses and maps. ARL performed the data interpretation and generated all figures. AP provided expertise on the writing and interpretation of figures. All authors reviewed and edited the manuscript.

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431 sediments. We also thank the community of Tuktoyaktuk for providing wildlife monitors with insightful information
432 on the territory during field sampling. This research was funded by NSERC Discovery Grant and Northern Supplement
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435 **Financial support**

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436 Support funds and grant agreement numbers are listed as specified upon manuscript registration.

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Page 4: [13] Commented [ARL6] **Roy-Lafontaine Alexie** **12/29/25 12:01:00 PM**

Différenciation between active layer and taliks to answer R1 comment.

Page 4: [14] Commented [ARL7] **Roy-Lafontaine Alexie** **12/29/25 12:37:00 PM**

Core length added to address R1 comment.

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Page 4: [16] Commented [ARL6] **Roy-Lafontaine Alexie** **12/29/25 12:01:00 PM**

Différenciation between active layer and taliks to answer R1 comment.

Page 4: [17] Commented [ARL7] **Roy-Lafontaine Alexie** **12/29/25 12:37:00 PM**

Core length added to address R1 comment.

Page 4: [18] Commented [ARL6] **Roy-Lafontaine Alexie** **12/29/25 12:01:00 PM**

Différenciation between active layer and taliks to answer R1 comment.

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Page 20: [24] Commented [RLA25] **Roy-Lafontaine Alexie** **1/20/26 9:00:00 AM**

Addition of supplementary figure that addresses R2 comment 6 with emphasis on limitation of scale studied.

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