

# The 3D submicron-scale skeletal reconstruction of *Nannoconus* (Cretaceous calcareous nannofossil) - Insights into biomineralization

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15 **Abstract.** *Nannoconus* (~5-20 µm) was a major planktonic producer in the Early Cretaceous seas (~150-120 Ma). The heavy calcitic skeletons (~~←(micaliths; ~200-1400 picogram)~~ of this ~~calcareous nannofossil~~extinct nannoplankton genus have contributed to massive carbonate accumulations for over ~30 million years. The ~~skeletal~~micalith microstructure is characterized by an interlocking arrangement of calcitic lamellae spanned around a central canal. The biomineralization process involved in producing the sophisticated ~~skeleton~~micalith is investigated for the first time. Ptychography X-ray

20 computed tomography (PXCT) with synchrotron radiation is applied to an isolated ~~skeleton~~micalith, to obtain a 3D set of tomographic images with ~ 40 nm spatial resolution. This 3D set was processed to virtually segment the individual calcitic lamella and reconstruct the full ~~skeleton~~micalith through constraining different lengths and angles. The lamellae are repetitively stacked in two distinct inclinations, one following the other, and producing segments combined to form the entire ~~skeleton~~micalith. Individual lamellae were calcified in a “template” of organic layer containing amino acid(s)/biomolecule(s),

25 responsible for creating the interlocking arrangement. Our study of *Nannoconus* provides a simple yet potent approach to the analysis of biomineralized microstructures characterized by the repetitive arrangement of calcitic units as commonly seen in the calcareous nannoplankton.

## 1 Introduction

30 Calcareous nannofossils (~1-30  $\mu\text{m}$ ) are biomineralized calcitic remains of marine planktonic unicellular algae (Siesser and Winter, 1994) and are abundant in the sedimentary archives. Initially rare, calcareous nannofossils became increasingly abundant in marine sediments to reach an optimum in the Cretaceous, around ~120 Ma (Suchéras-Marx et al., 2019) and they profoundly altered the dynamics of the carbonate and oceanic carbon cycle (Erba, 2006). Among ~~extinct calcareous nannofossils, nannoplankton~~ *Nannoconus* was the major ~~bi~~carbonate producer in the Early Cretaceous seas (~152-120 Ma) (Bown, 2005) with ~~the largest~~large size (~5-20  $\mu\text{m}$ ) and ~~the heaviest~~heavy mass (~200-1400 picogram); ~~section 4.2, chapter 4~~ of ~~the~~Chowdhury, 2025) for a calcitic micalith (individual piece that composes the skeleton, Aubry, 2025), contributing to massive carbonate accumulations over ~30 million years. ~~These massively produced biocalcites most likely modified the~~The ~~massive production of biocalcite may have significantly altered~~ marine chemistry ~~significantly~~, upon transfer to the oceanic sedimentary record, with possible consequences on the marine biosphere at a time of important planktonic diversification (Hart et al., 2003; Kooistra et al., 2007). Despite the importance of this group, knowledge of the organism ~~that produced the calcitic skeleton of *Nannoconus* is non-existent. A proper understanding of the biomineralization process from the 3D microstructural arrangement of the skeleton, obtained from a finely resolved physical characterization, can help to close this gap, which produced the calcitic micalith is non-existent. Currently, based on 2D scanning electron microscopy (SEM) studies, two different concepts of micalith construction coexist (Aubry, 1974, 2025; Van Niel, 1993). However, neither concept explains the skeletal microstructure and provides information on the parameter(s) controlling the shape of the micalith. High-resolution characterization of the three-dimensional microstructural organization of the micalith, as documented here for *Nannoconus globulus* Brönnimann 1955 provides insights into the biomineralization process.~~

With advances in imaging techniques, several high-resolution characterization methods have been successfully applied to biocalcite produced by extant planktonic algae or preserved as fossils. These techniques include AFM (Henriksen et al., 2004), 3D-FIB-SEM (Hoffmann et al., 2015) and cryo-electron tomography (Walker et al., 2020). In addition, several X-ray scattering methods with synchrotron radiation have been applied, such as 2D (Suchéras-Marx et al., 2016a) and 3D (Walker et al., 2024) X-ray microfluorescence, X-ray Coherent Diffraction (Beuvier et al., 2019). Ptychographic X-ray computed tomography (PXCT) (Dierolf et al., 2010) is a non-destructive tomographic reconstruction imaging method that can be applied to characterize fossilized biocalcite. In this study, we performed the PXCT experiment using synchrotron radiation at the SWING Beamline of the SOLEIL (French synchrotron). Our primary focus was on ~~the~~a well-preserved skeleton/micalith of *Nannoconus* to investigate its microstructural arrangement at the nano-scale resolution. The experiments generated tomographic volumes of ~~the samples~~our specimen with a 3D spatial resolution of approximately 30 nm. Subsequently, we analyzed these volumes using image visualization software and successfully reconstructed the complete 3D skeleton/micalith of a single *Nannoconus* specimen, specifically, *N. globulus*, for the first time. This strategy serves as a methodological framework for reconstructing the ~~*Nannoconus*'s skeleton~~*Nannoconus* micalith structure in general; ~~and to address the validity of the two concepts proposed~~

60 in the literature. The knowledge of the structural constraint of the 3D skeletal micalith arrangement obtained from the  
aforementioned reconstruction has been used to provide perspectives on insights into the *Nannoconus* biomineralization  
process.

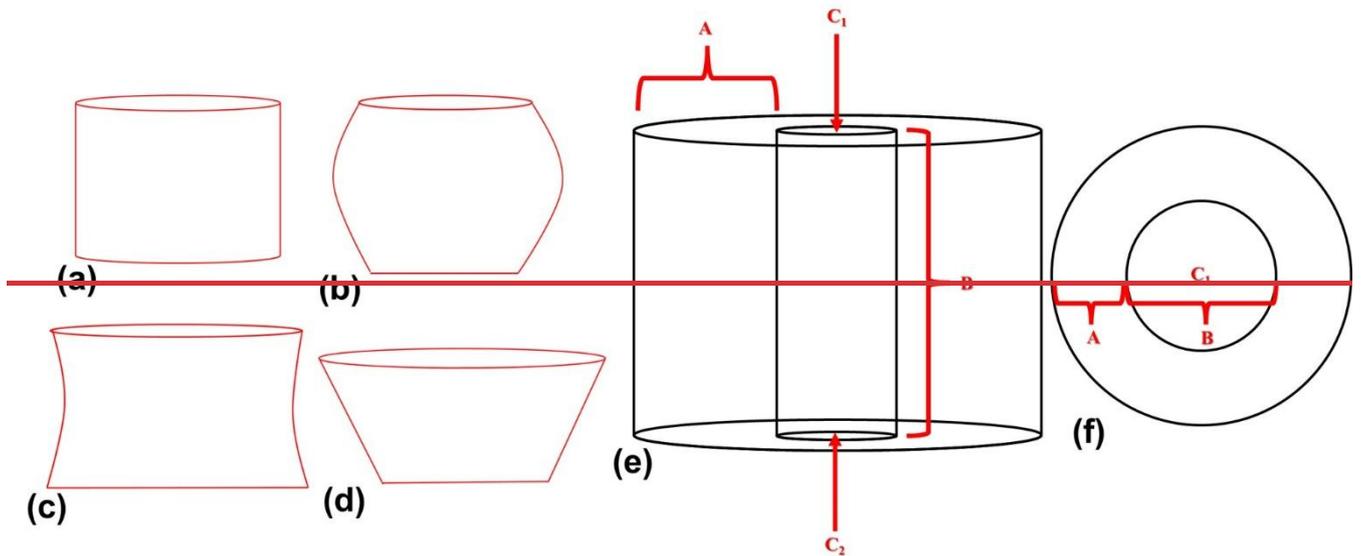
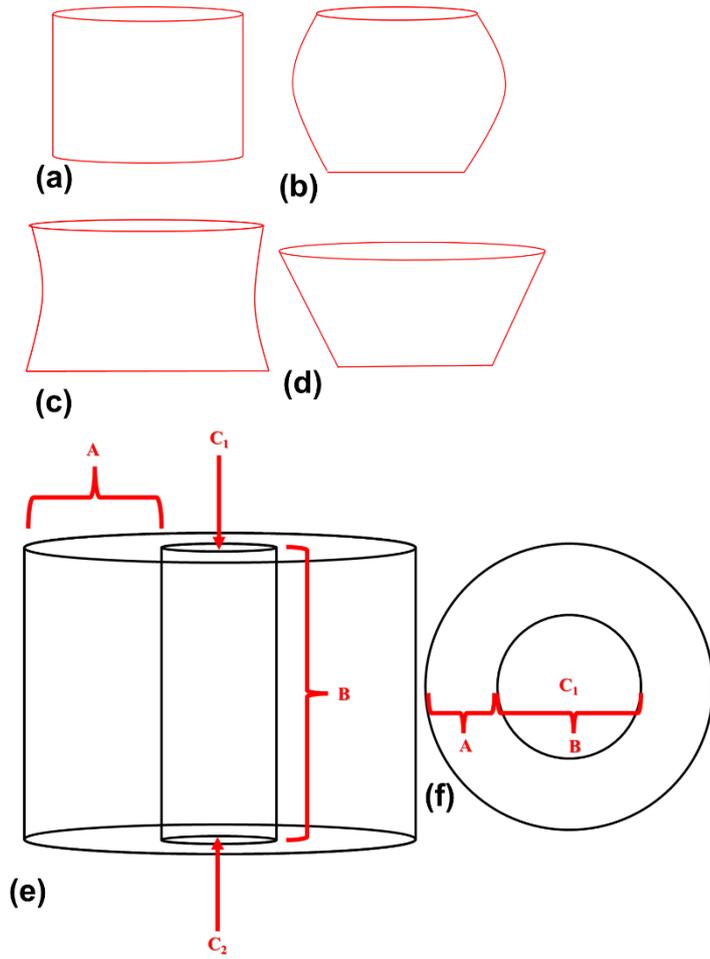
## 2 The *Nannoconus* skeletal micalith

*Nannoconus* is a Ggenus (Kamptner, 1931, emend. Farinacci, 1964) belonging to the Ffamily of *Nannoconaceae* (Reinhardt,  
65 1966) (Table C1 in the Appendix). This Family is included in the Order Lees and Bown (2016) assigned this family to the order  
of *Braarudosphaerales* (Aubry, 2013, Aubry, 2025) by Lees and Bown (2016), which they emended. *Nannoconus* is  
mostly belongs to the micalithophores, a group independent from the coccolithophores, which unites calcareous nannoplankton  
with individual pieces of the skeleton (micaliths) consisting of "identical, adjoining segments that are stacks of lamellae of  
similar shape" (Aubry, 2025, section 3.2.3).

70 *Nannoconus* is a genus of Tethys domain which spans from the latest Jurassic (Tithonian, ~145 Ma) through the Late  
Cretaceous (Campanian, ~72 Ma) (Perch Nielsen, 1985). However, in the Tethys domain, it is only abundant until the end of  
the Aptian (~113 Ma), which is the end of the climax of *Nannoconus* (Erba, 1994). This represents a duration of ~30 Myr,  
during which the skeleton exhibits micaliths exhibit a great morphological diversity (Derès and Achéritéguy, 1980).

### 2.1 Morphology

75 Different Several species of the genus Nannoconus have been described for the Genus Nannoconus and are characterized by  
different shapes of the skeleton, mainly differing in the shape of their micaliths. These include cylindrical, globular, hour-  
glass, and conical shapes (Figs. 1a-1d). Van Niel (1993) introduced several terms to describe it. Some of these terms are used  
in this study their general morphology (Figs. 1e and 1f). The *wall* encloses the *central canal* which runs along the longitudinal  
axis of the skeletal micalith with two terminal openings known as *apertures*. Several morphogroups of *Nannoconus* have been  
80 defined based on the general shape and size of the skeletal micalith (Brönnimann, 1955; Bouché, 1965; Derès and Achéritéguy,  
1972; Aubry, 1974), the shape of the central canal (Brönnimann, 1955; Bouché, 1965), the thickness and construction of the  
wall (Brönnimann, 1955; Bouché, 1965; Derès and Achéritéguy, 1972; Aubry, 1974), the shape and the size of the terminal  
openings (Brönnimann, 1955; Bouché, 1965; Derès and Achéritéguy, 1972; Aubry, 1974).



85 Figure 1: Schematic diagrams illustrating ~~four~~the different ~~skeletal~~micalith morphologies of four ~~different~~ species of *Nannoconus*.

90 These are (a) cylindrical, (b) globular, (c) hour-glass, and (d) conical as observed in *Nannoconus truitii*, *Nannoconus*N. *globulus*, *Nannoconus*N. *abundans*, and *Nannoconus*N. *steinmannii*, respectively. (e and f) Longitudinal and transverse views, respectively, of the schematic illustration of a cylindrical ~~skeleton~~micalith. In this figure, A and B represent the wall and the central canal, respectively. C<sub>1</sub> and C<sub>2</sub> are the two terminal apertures. It should be noted that the shape of the central canal ~~differs~~varies with the general morphology of the ~~skeleton~~micalith.

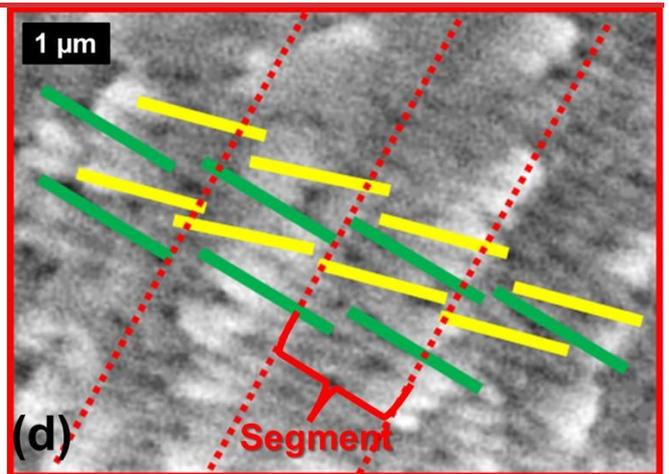
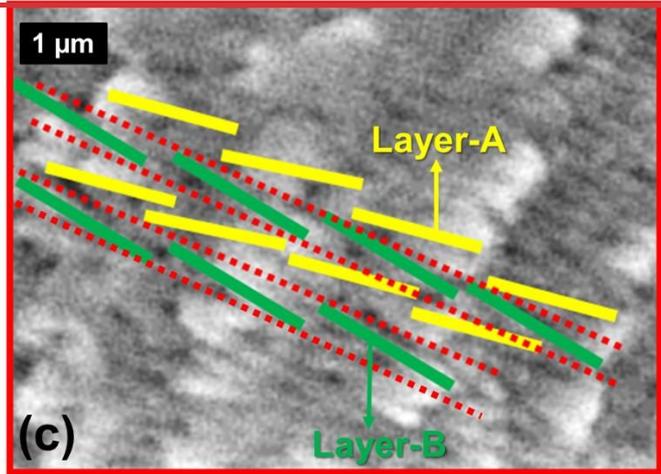
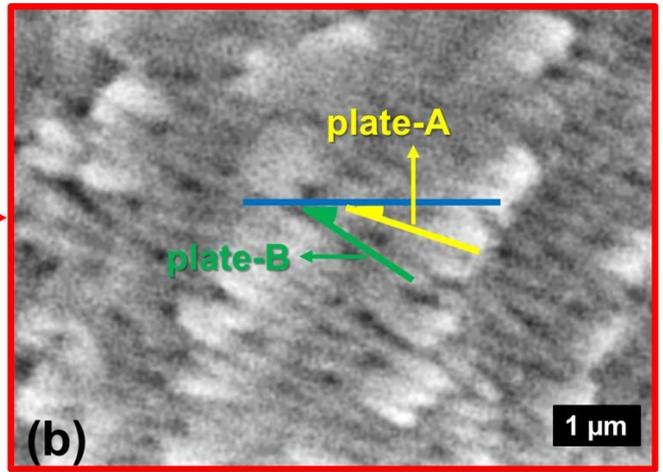
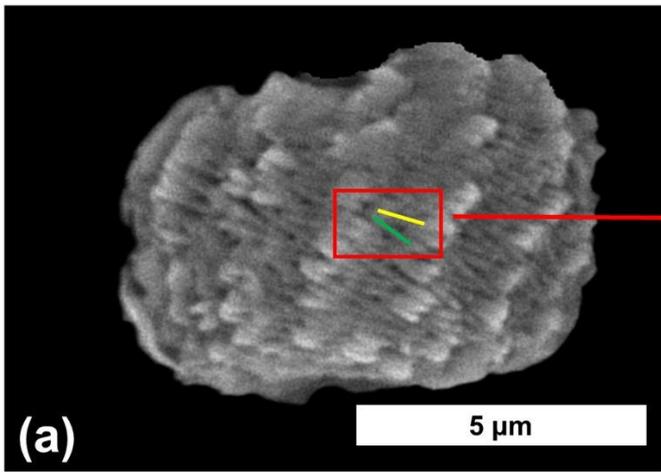
## 2.2 Microstructure

95 ~~The Different terminologies, reviewed by Van Niel (1994), continue to be used to describe the wall of Nannoconus—consists of, “Lamella/ae” is used here in agreement with Aubry (2025). The interlocking sub-rhombohedral (Van Niel, 1994) to lamellae are semi-circular plates—(Stradner and Grun, 1973) (to sub-rhombohedral (Van Niel, 1994) with length ~0.5-1 μm, and thickness ~0.1-0.5 μm) (Kamptner, 1931; Trejo, 1960; Derès and Achéritéguy, 1972; Van Niel, 1993, 1994). Two distinct types of plates/lamellae (green and yellow lines in Fig. 2a) build the skeleton/micalith. Each plate/lamella is characterized by a specific inclination, measured from a horizontal line (blue line in Fig. 2b). These nannofossils have been defined by Aubry (2013) as “homoeoccoliths, i.e., coccoliths consisting of identical, imbricated segments that are stacks of lamellae of similar shape”. Here, the term “lamellae” refers to the plates of Van Niel (1993) and this term is retained in this work. The term “homoeoccolith” has been replaced as “micalith” by Aubry (2025)-2b).~~

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Based on 2D scanning electron microscopy (SEM) studies, some authors (Aubry, 1974, 2013, 2025; Van Niel, 1993) attempted to explain the ~~3D~~micalith microstructural arrangement. ~~Currently,; two different concepts of skeletal construction are have been proposed.~~ The first concept is based on a layering of two types of ~~plates/lamellae~~ (Van Niel, 1993; Lees and Bown, 2016), the second concept is based on an organization of lamellae in to segments (Aubry, 2013, 2025) ~~of Nannoconus.~~ They can be summarized ~~as~~by the following two models:

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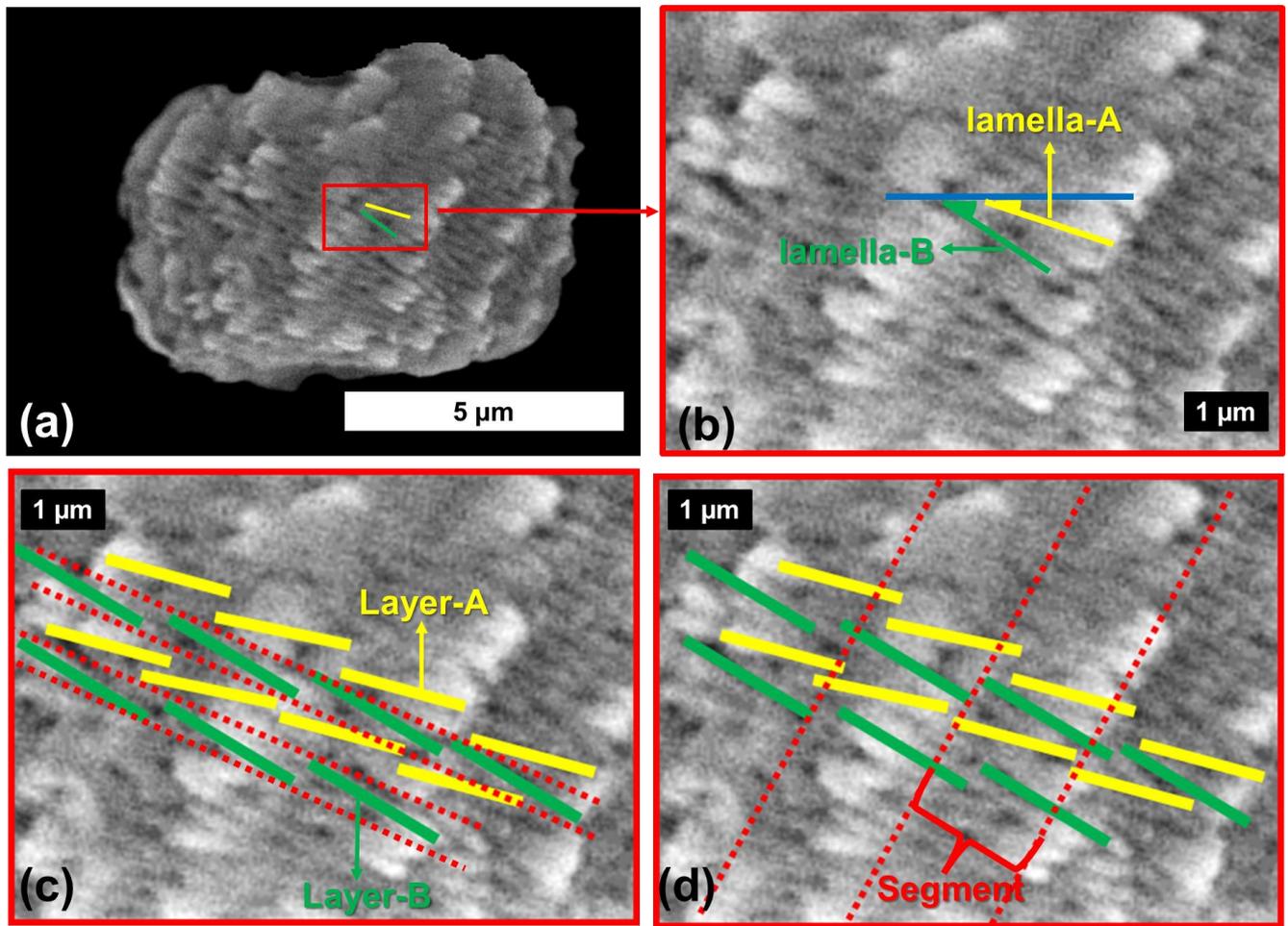


Figure 2: Scanning electron microscopic (SEM) image of *Nannoconus globulus*. (a) Longitudinal view. The solid green and yellow lines inside the red box highlight the two distinct types of plate lamellae, lamella-A and -B. (b) Closer view of the plate lamella -A and plate lamella -B shown in (a), with the inclinations measured from the blue horizontal line. (c) Arrangement of plate lamellae -A and -B in two layers (-A is yellow and -B is green). The red-dotted line separates the two consecutive layers. (d) Oblique arrangement of the segments composed of alternating plate lamellae-A (yellow) and -B (green). Consecutive segments are individualized by red dotted lines. “Plates” are used following Lamella/ae is the term selected to describe the structure of micoliths (Aubry, 2025). It is the equivalent of “plate” in the terminology of Stradner and Grun (1973) and Van Niel (1993); they have been referred to as “lamellae” by Aubry (2013). The term “lamellae” is retained throughout this work.

### 2.2.1 Model-1

In Van Niel (1993), the plate lamellae with a lower angle of inclination (yellow) are termed plate lamellae-A and the plate lamellae with a higher angle of inclination (green) are termed plate lamellae-B. Plate Lamellae-A and -B form two distinct types of layers (Fig. 2c) named referred to here as layer-A and layer-B, respectively. The skeleton micolith can be understood as a combination of these two types of spiralling layers of plate lamellae alternatively arranged one after another. This can be explained in Fig. 2c: first, a layer-B (green lines) is placed, followed by a layer-A (yellow lines). Over layer-A, another layer-

B is again placed. Delimitation of layers is marked by red dotted lines (Fig. 2c). Van Niel (1993) distinguished nine different morphogroups among *Nannoconus* (Table C1 in the Appendix) of *Nannoconus*, based on the various inclination(s) of plate lamella-B and the width(s) between two consecutive layers of plate lamella-A. In a SEM image of the species *Nannoconus* *N. globulus* (Brönnimann, 1955) (selected for the present study), in Covington and Wise (1987, pl. 12, Fig. 2) it is possible to count six plate lamellae with two distinct angles of inclination in the part where the plate lamellae are well-preserved (Figs. B1a and B1b in the Appendix). This part constitutes about half of the circular aperture (180°). Thus, the total number of plate lamellae in the full circular aperture (360°) would be = 12, with six plate lamellae-A and six plate lamellae-B. Considering that each of these plate lamellae belongs to a separate layer, there are six layer-A and six layer-B, i.e., a total of 12 layers in the whole skeleton micalith.

### 2.2.2 Model-2

As suggested by Aubry (2013, 2025), the skeleton micalith is a combination of identical, imbricated segments. These segments are formed by stacking lamellae (Fig. 2d) of similar shape (Fig. 12d ~~in~~ Aubry, 2025). Lamella-B (green line) and lamella-A (yellow line) are organized two by two to form duos. Duos of lamellae are stacked to compose a segment. Therefore, the spiral combination of several such segments forms the entire skeleton micalith. Three such segments are delineated by red dotted lines in Fig. 2d. In a SEM image (Fig. B1c in the Appendix) of *N. globulus* ~~described~~ referred to above (in model-1), six segments ~~in~~ can be delineated from the ~~part where the skeleton is well-preserved, can be detected~~ part of the micalith. As this part constitutes about 1/2 of the circular aperture (180°), the total number of segments in the full circular aperture would be  $6 * 2 = 12$ . Therefore, the total number of the segments in the full ~~skeleton of *Nannoconus* = 12. It should be noted that these two concepts only explain the skeletal microstructure and do not provide any information about the parameter(s) controlling the shape of the skeleton. To address the validity of the two models and also to decipher the parameter(s) influencing the skeletal shape and microstructure from the 3D skeletal reconstruction, a synchrotron-based PXCT was conducted.~~ micalith of *Nannoconus* = 12.

## 3 Materials and Methods

### 3.1 Material

Six specimens representing five different species of *Nannoconus* have been used for the Ptychographic X-ray Computed Tomography (PXCT) experiment. They ~~come~~ are from the Aptian ~~(Covington and Wise, 1987)~~ calcareous sediments of the DSDP Leg-93-Site 603 (continental rise of the western margin of the North Atlantic) ~~e~~ Core 44, interval 115-116 cm ~~(Covington and Wise, 1987)~~. They have been selected for their good state of preservation as determined through scanning electron microscopy (SEM) images ~~of *Nannoconus* are~~ and shown in Fig. B2 in the Appendix ~~).~~. Here, “good state of

preservation” refers to ~~skeletons~~micaliths which are structurally fully intact without any breakage and devoid of post-depositional growth of calcite, that often obscures the microstructure of nanofossils. We focus ~~our~~this study on the species *N. globulus* to illustrate the microstructural arrangement and the 3D reconstruction of the ~~skeleton~~micalith. This globular ~~skeleton~~micalith exhibits a well-defined interlocking pattern of ~~the~~ lamellae.

## 155 3.2 Sample Preparation

The PXCT experiment required the picking of an individual specimen of *N. globulus* with a silica needle (Figs. B3a and B3b in the Appendix) (modified from Suchéras-Marx et al., 2016b). The needle was then placed in a metal holder (Fig. B3c in the Appendix), directly placed at the experimental station of the beamline.

## 3.3 Ptychography computed X-ray tomography (PXCT)

160 Ptychography is an advanced X-ray imaging technique that involves capturing a series of far-field diffraction patterns from overlapping areas of a sample. These diffraction patterns are then used in a computational phase retrieval algorithm to reconstruct a high-resolution two-dimensional image. In the field of PXCT, ptychography is combined with tomography. This process involves performing ptychographic experiment at different tomography angles, which produce highly spatially resolved, detailed tomographic projections. As a result, PXCT offers a thorough three-dimensional (3D) representation of the  
165 microstructure of ~~the~~a sample, achieving nanometric resolution. This enhances the ability to analyse complex materials at a microstructural level. After obtaining the 3D images, it is essential to examine the resulting high-resolution tomographic volume of biocalcite using advanced 3D image analysis techniques. Methods such as image segmentation and visualization (Reznikov et al., 2020) are necessary to extract relevant microstructural properties from the sample.

The experiments were performed at the SWING beamline at SOLEIL-Synchrotron, Paris, France. The photon energy was of  
170 8.00 keV, and the sample-to-detector position was set to 6.484 m. The ptychography experiments were carried out with a beam size of about 4  $\mu\text{m}$  at the sample position and a scanning step size of 1.0  $\mu\text{m}$ . ~~A number of 164~~One hundred and sixty-four positions were collected and distributed as concentric circles within a rectangular field of view of 20  $\mu\text{m}$  x 10  $\mu\text{m}$  (HxV). We used a region of interest of 1000 x 1000 pixels of an EIGER-4M in-vacuum detector from DECTRIS with a pixel size of 75  $\mu\text{m}$ . For the PXCT experiments (Dierolf et al., 2010), 632 tomographic projections were acquired between 0° and 180°, with  
175 exposure time of 100 ms, as 8 interlaced sub-tomograms. The resulting diffraction patterns were transformed into real space projections using the Ptycho-Shelves (Wakonig et al., 2020) suite from the cSAXS team at Paul Scherrer Institute, with a starting probe model previously determined experimentally using a Siemens Star ptychographic scan. The final volume was determined from the tomographic projections using the Matlab tomography package within PtychoShelves.

## 4 Results

## 180 4.1 Descriptive parameters

The combination of the tomographic image slices, obtained from PXCT, reveals the external and internal views of the *N. globulus* ~~skeleton~~micalith. These observations are presented here to understand the shape of the ~~skeleton~~micalith and the arrangement of the lamellae and to propose a set of standard parameters that can be used to describe other *Nannococonus* morphogroups. In the following, cylindrical coordinates are used to describe the ~~skeleton~~micalith microstructure in terms of  
185 the positions and orientations of the constituting lamellae (Figs. 3a-3d). For this geometric description, the lowest and the highest points of the micalith in longitudinal orientation are defined as base and apex, respectively (Fig. 3a). The z-axis is parallel to the longitudinal view of the ~~skeleton~~micalith, passing through the central canal. The total vertical length between the base and apex is given by L (Fig. 3a). ~~The Radius~~The Radial axis,  $r$ , corresponds to a vector going from the center of mass of ~~the single~~a lamella to the center of the central canal, and it is perpendicular to the z-axis. For this geometric description, the lowest and the highest points of the skeleton in longitudinal orientation are defined as base and apex, respectively (Fig. 3a).  
190 ~~The~~ The total vertical length between the base and apex is given by L (Fig. 3a). ~~The length of the~~ radius (r) is the length of the radial axis, measured from the ~~central~~z axis to the ~~end~~center of mass of the ~~w~~lamella (Fig. 3a). It changes throughout the whole ~~skeleton~~micalith from the base to the apex. It is minimum at the base of the micalith (Fig. 3a). Starting from the base, the radius increases, reaching a maximum and decreasing towards the apex, giving the ~~skeleton~~micalith a globular shape. This  
195 also indicates that the radius ~~value~~ is maximum at  $2/3^{\text{rd}}$  of the length (measured longitudinally) between the base and the apex. ~~The minimum value of the radius is observed at the base of the skeleton (Fig.~~

~~3a).~~ Two angles define the spatial orientation of each lamella. ~~The~~ Based on the SEM images, the first angle was previously defined by Van Niel (1993), based on the SEM images and is) is the Inclination ( $\delta$ ). ~~In~~, which, in the cylindrical coordinates (described before, ~~it~~), corresponds to the angle (Fig. 3b) formed between ~~the~~a lamella and an axis corresponding to the radius (blue line in Fig. 3b). ~~Two~~3b). On the basis of this inclination angle ( $\delta$ ), two distinct sets of lamellae with different inclinations are shown in Fig. (Fig. 3c) can be distinguished: the lamella with low inclination ( $\delta_A$ , yellow line given by VW), is named lamella-A. The lamella with high inclination ( $\delta_B$ , green line given by XY), is termed ~~as~~ lamella-B (Fig. 3c). Lamella-A ~~and~~ lamella-B formed together form twin lamellae. The term “twin lamellae” describes the repeating pair of lamellae (i.e., lamella-A and lamella-B), with alternating inclinations that together construct the full ~~skeleton~~micalith of *Nannococonus*. The  
200 ~~second~~other angle (Fig. 3d), called Tilt ( $\tau$ ), corresponds to the angle formed between a lamella and an axis ~~of rotation~~ perpendicular to the radius (red line in Fig. 3d). Tilt values are of opposite sign for lamellae placed in the upper (high z) or lower (low z) parts of the ~~skeleton~~micalith. ~~As mentioned above, the arrangement of the lamellae has been described in two models in the literature. Here, we will show that the components (the layers in model 1 and the segments in model 2) of the two models can be verified using the abovementioned radius and angles.~~micalith (Fig. 3d).

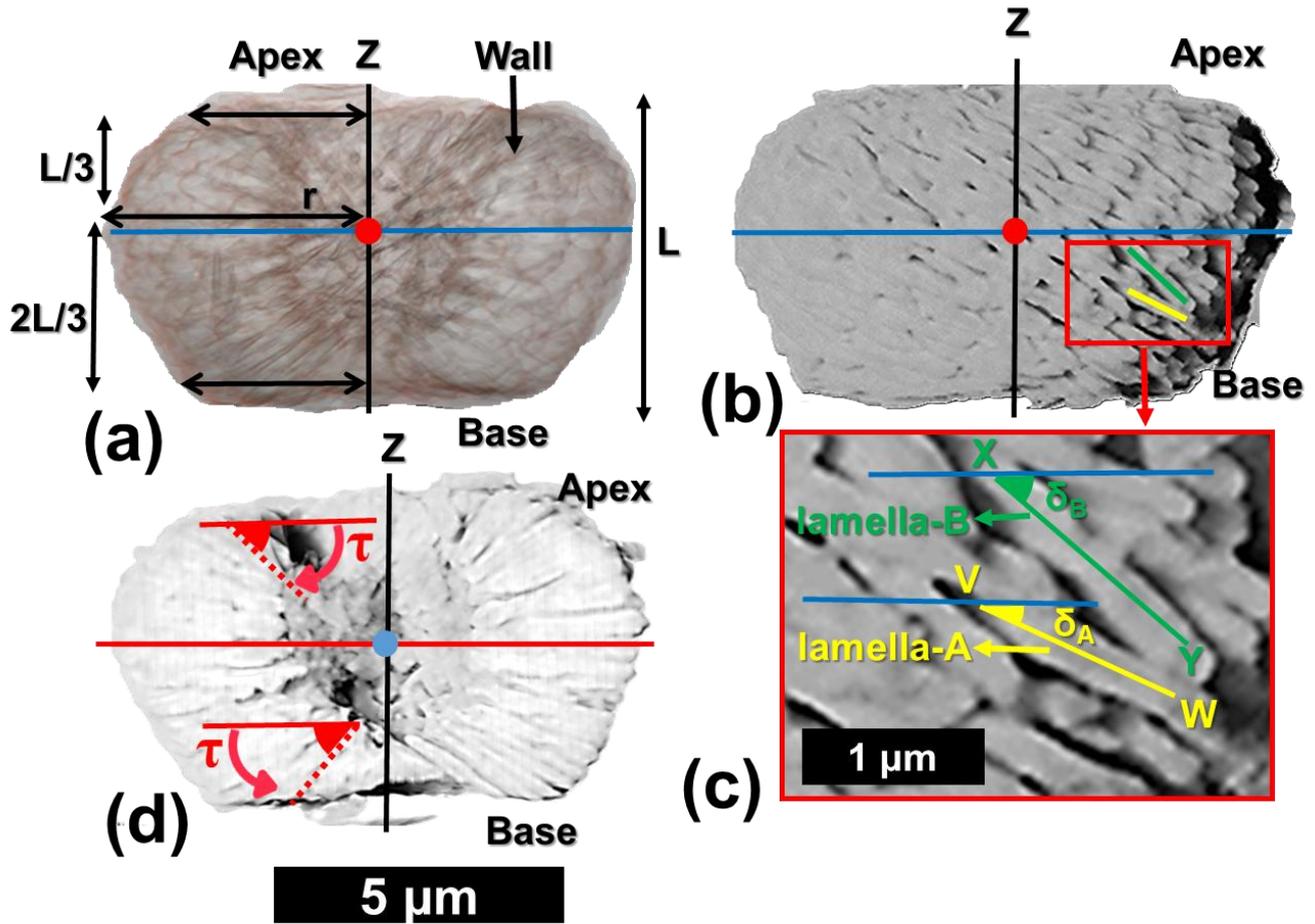
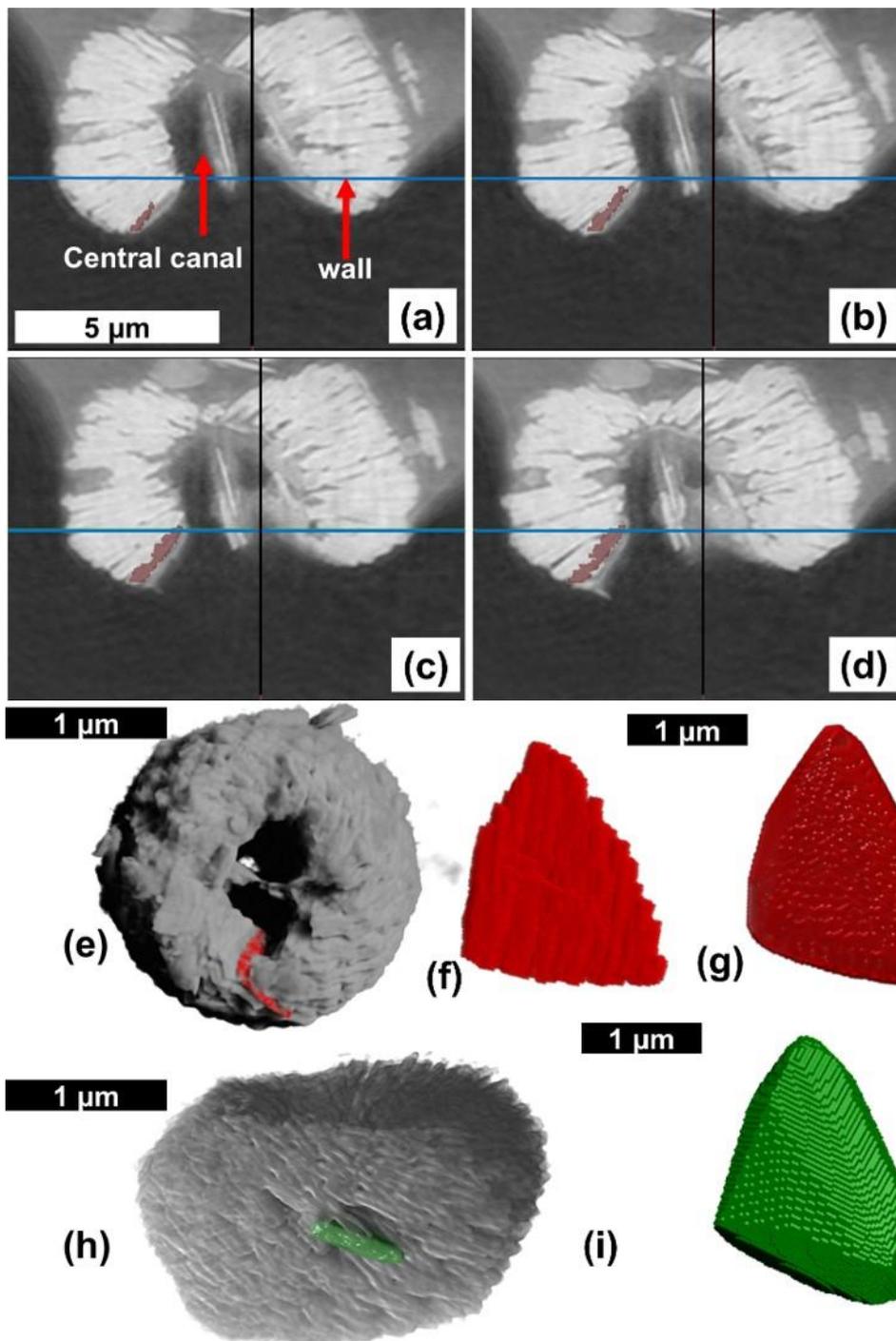


Figure 3: Description of the parameters for the reconstruction of the 3D skeletal microstructure of *N. globulus* using the results of the PXCT experiment. (a) Internal view of *N. globulus*. The microstructure is interpreted using a cylindrical coordinate system. The red dot indicates the radial axis (perpendicular to the plane of the paper), and the blue line represents an axis perpendicular to the radius. The z-axis, marked by the black line, passes longitudinally through the central canal. The length of the red arrows corresponds to the radius, measured from the z-axis to the end of the wall of the skeletal microstructure. The radius increases initially from the base for 2/3<sup>rd</sup> of the length (L) of the *Nannoconus*, and then decreases towards the apex for the remaining 1/3<sup>rd</sup> of the length (L). (b) External view of the same specimen of *N. globulus*, in the cylindrical coordinate system. (c) Magnified skeletal microstructure of the part delineated by the red box in (b), illustrating the two lamellae types: lamella-A (yellow) and lamella-B (green). Their inclinations,  $\delta_A$  and  $\delta_B$  respectively, correspond to the angles measured between the blue line (axis perpendicular to the radius) and each lamella. (d) Cross-section of *N. globulus* across the length. The angle measured between the lamella and the red line (radial axis (red line)) defines the tilt. The tilts of the lamellae at the base (lower z) are opposite in direction with respect to the lamellae at the apex (upper z).

#### 4.2 Segmentation of a lamella

The first step in our analysis of the PXCT data was to isolate one lamella (Figs. 4a-4g). The experiment resulted in a series of tomographic image slices for the whole skeletal microstructure. These images reveal overlapping lamellae within the interior of the skeletal microstructure, likely resulting from diagenetic alteration ~~(a process involving the dissolution of the biocalcite, produced~~

during the biomineralization and subsequent reprecipitation). Although the specimen was considered “well-preserved” based on SEM images, the high spatial resolution of the PXCT reveals clear diagenetic effects within the *Nannoconus*'s skeleton. *Nannoconus micalith*. As a consequence result, individual lamellae are sometimes difficult to segment (i.e., virtually separate) in the tomographic images. Also, despite having a The spatial resolution of the PXCT data (~30–40 nm, this resolution) is at of the same scale of as the thickness of the lamellae themselves and is therefore insufficient for distinctly segmenting a single lamella. To overcome this problem, all the images were filtered using Contrast Limited Adaptive Histogram Equalization (CLAHE) (Reza, 2004). This increased the contrast of the images and hence helped to delineate the boundaries between the lamellae more clearly. After this contrast enhancement, a single lamella was segmented by selecting by hand regions of interest for each slice. Four such slices are shown in (Figs. 4a-4d). This results in a 3D volume of the lamella that was exported and used to reconstruct the structure of the skeleton *micalith* using our two models (Figs. 4e and 4f). Given the high amount of data, the convex hull (Table C1 in the Appendix; Ky, 1959) of the 3D volume of the segmented lamella (Fig. 4g) was generated using Python routines and utilized for the reconstruction of the models. However, this manual segmentation approach is not applicable to the entire *Nannoconus* because: (Aa) It is extremely time consuming to manually segment numerous lamellae of *Nannoconus*; from ~300 tomographic image slices. (Bb) The lamellae frequently overlap with each other, due to post-depositional overgrowth (a process that dissolves and reprecipitates the calcite); making them difficult to distinguish for segmentation. Hence, we have segmented a distinctly identifiable lamella and utilized it as a unit to reconstruct the entire skeleton *micalith*. As described earlier, *Nannoconus* belongs to *Braarudosphaerale* Order (order *Braarudosphaerales* Aubry, 2013, 2025); which is defined (Aubry, 2013) as “consisting of identical, imbricated segments that are stacks of lamellae of similar shape”. Therefore, it can be inferred that *Nannoconus*'s lamellae are also of “similar” shape/morphology. To check verify this, we have segmented another lamella (Fig. 4h) from a different part of the skeleton (different from Fig. 4e), whose *micalith*; its convex hull of this lamella is given in Figure 4i. The Both segmented lamellae present a similar shape/morphology (Figs. 4g and 4i). They are of triangular, flat shape, ~2.00 μm long and ~0.50 μm thick. Both of the lamellae We conclude from this present a similar shape/morphology (Figs. 4g and 4i). Based on the morphological similarity between two lamellae taken from different parts of the skeleton, we concluded *micalith*, that the entire skeleton *micalith* of *N. globulus* is composed of morphologically similar lamellae.



255 Figure 4: Procedures describing the segmentation/virtual separation of a lamella from the result of the PXCT analysis of *N. globulus*. (a-d) Four consecutive tomographic images obtained from the PXCT analysis of *N. globulus* with the wall and central canal. The part/object present within the central canal is possibly a lamella detached from the wall. (e) The marked lamella (red colour) is presented/shown within the skeleton/micalith. (f) Isolated individual lamella after the segmentation. (g) Convex hull of the lamella, with smooth edges and surfaces. Convex hull (See Table C1 in the Appendix) is the mathematical representation of an envelope

covering the segmented lamella. (h) Another segmented lamella (green colour) ~~is marked~~located within the ~~skeleton~~micalith. (i) Convex hull of the latter segmented lamella, ~~marked~~(i.e., seen in (h)).

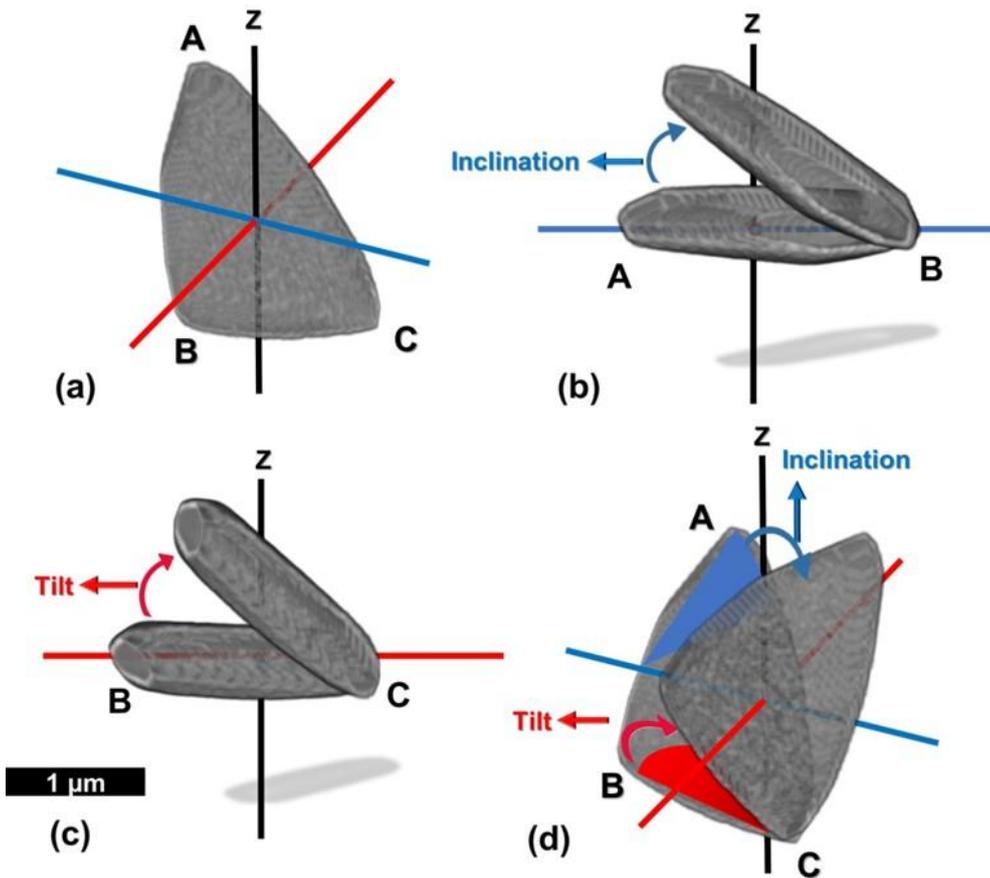
## 260 4.3 Reconstruction

We developed a code ([Chowdhury et al., 2025](#)) using Python script (Van Rossum and Drake, 2009) directly in a console attached to the image visualization software to reconstruct the ~~skeleton~~micalith using the segmented lamella. The process of the ~~skeleton~~micalith reconstruction from the segmented lamella can be described as:

### 4.3.1 Generation of inclination and tilt

265 The lamella (given by ABC in Fig. 5a), ~~which has a~~of triangular, and flat shape, is rotated in two perpendicular directions in a cylindrical coordinate system to set values for inclination and tilt. The procedure is as follows:

1. The rotation with the radial axis as the axis of rotation generates the inclination,  $\delta$ , of the lamella (Figs. 5b and 5d).
2. The rotation with the axis perpendicular to the radial axis (Figs. 5c and 5d) as the axis of rotation generates the tilt,  $\tau$ , of the lamella.



270

Figure 5: Generation of the inclination and tilt in the segmented lamella. (a) The segmented lamella, given by ABC, is placed in the cylindrical coordinate system with one vertical z-axis (black line) and two radial axes (red and blue lines). (b) The rotation of the lamella, where the radius (red line) is the axis of rotation, creates inclination, and the rotation angle is measured between the lamella and the blue line. (c) The rotation of the lamella, where the axis perpendicular to the radius (blue line) is the axis of rotation, creates tilt, and the rotation angle is measured between the lamella and the red line. It should be noted that the axes of rotation for creating inclination and tilt are perpendicular to the lines from which the individual angle of inclination and tilt are measured. (d) The combination of two perpendicular rotations in the cylindrical coordinate, creates the inclination and tilt in the segmented lamella.

275

#### 4.3.2 Formation of points in spiral axes

The skeletonamicalith being conceptualized as a combination of spiral layers/segments, it is now necessary to create the layers/segments utilizing the segmented lamellae. A series of points forming spiral axes was generated to place individual lamellae, creating layers/segments. The points of a series are defined following cylindrical coordinates (Fig. 6a):

280

(i) The radius,  $r$ , given by the (=distance from the centralz axis to the center of mass of the lamella-

), (ii) ~~The~~ azimuth,  $\theta$ , given by the (=angle around the centralz axis-

(iii) The vertical distance,  $z$ , (perpendicular to the radius and along the axis of rotation of the azimuth).

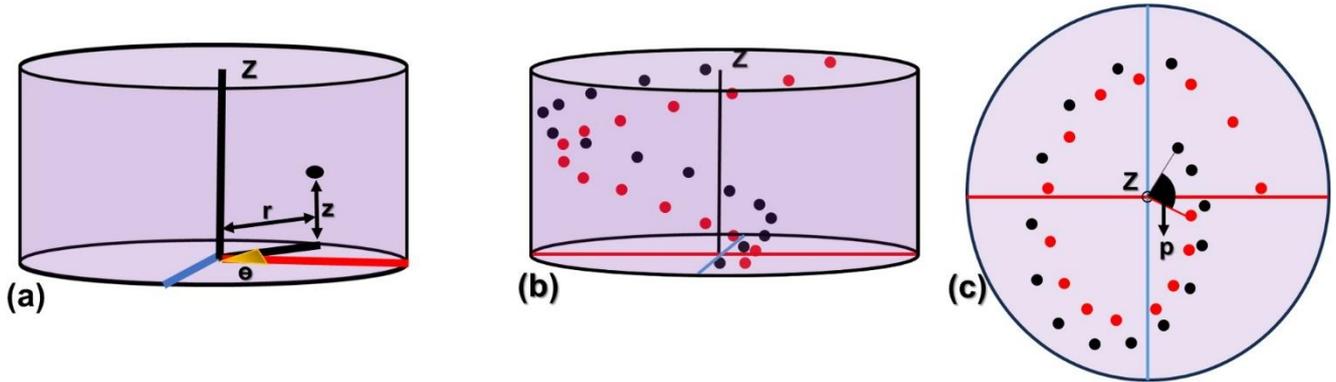


Figure 6: Presentation of the series of points in spiral axes to form the layers and segments in the cylindrical coordinate system. (a) Radius ( $r$ ), azimuth ( $\theta$ ), and vertical distance ( $z$ ) that define a point in the cylindrical coordinate system. The red line represents the radial axis, and the blue line represents a line perpendicular to it. (b, c) Longitudinal and transverse views, respectively, of the two series of points defined by the black and red dots in the cylindrical coordinate system, representing two consecutive layers/segments. Here, “ $p$ ” marks the angular separation between the two series of points.

Values of  $r$ ,  $\theta$ , and  $z$  are generated so that the lamellae fully occupy the space, forming a compact skeletonomicolith in visual accordance with the 3D reconstruction obtained from the PXCT data. Here, the first and last points represent the base and the apex of the skeletonomicolith, respectively. Considering that the total number of points in the series is  $n$ , an  $i^{\text{th}}$  point ( $r_i, \theta_i, z_i$ ), [where  $i < n$ ] in the series is represented by the following equations:

$$r_i = R + i * r_{step} \text{ [if } i \leq 2 * n / 3 \text{] (radius increases for the first } 2 * n / 3 \text{ points) and } r_i = r(i - 1) - i * r_{step} \text{ [if } i > 2 * n / 3 \text{] (radius decreases the last } n / 3 \text{ points) (1). This results in a globular shape for the } \underline{\text{skeletonomicolith}}.$$

$$R = \text{Initial radius, } r_{step} = (\text{Maximum radius} - \text{Minimum radius}) / n.$$

$$\theta_i = (\theta + i * \theta_{step}) \text{ (2)}$$

$$\theta = \text{Initial azimuth, } \theta_{step} = \text{angular separation between two consecutive lamellae} = (\text{Final azimuth} - \text{Initial azimuth}) / n$$

$$z_i = (Z + i * z_{step}) \text{ (3)}$$

$$Z = \text{Initial vertical distance, } z_{step} = (\text{Final vertical distance} - \text{Initial vertical distance}) / n$$

This results in a series of points with individual parameters, given by:  $[(r_1, \theta_1, z_1), (r_2, \theta_2, z_2), \dots, (r_i, \theta_i, z_i), \dots, (r_n, \theta_n, z_n)]$ .

The difference between the final and the initial azimuths of the points of the series, is expressed as the angle of rotation ( $S$ ).

As a series of points is equivalent to a layer/segment of the skeletonomicolith, the angle of rotation of the series also represents

the angle of rotation of a layer/segment. This angle of rotation ( $S$ ) of a series/layer, is given by:

$$S = \theta_{step} * n \quad (4)$$

310 A set of spiral axes (i.e., a series of points), each with the same number (n) of points, is needed to create the other layers/segments for the entire skeletonmicalith reconstruction. Two such consecutive series of points (in red and black colors respectively) are presented in Figs. 6b and 6c. The points (r,  $\theta$ , z) of all consecutive series are defined following the Eqs. (1), (2), and (3). The initial radius (R) and the vertical distance (V) is the same for all the consecutive series of points. However, the initial azimuth ( $\Theta$ ) is different for each of the series and depends on the angular separation between two consecutive series of points. If the angular separation between two consecutive series of points is given by  $p$ , then, the value of “p” can be calculated as:

$$p = 360^\circ / N \quad (5)$$

315 where N is the total number of series (i.e., the total number of layers/segments) for the full skeletonmicalith reconstruction. As the azimuth of the first point in the first series is  $\theta_1$ , therefore the azimuth of the first point of the second series will be  $(\theta_1 + p)$ . The I<sup>th</sup> series of points in N numbers of series (I < N) can be represented as:

$$\text{Series-I: } [(r_1, (\theta_1 + (I-1)*p), z_1), (r_2, (\theta_2 + (I-1)*p), z_2), \dots, (r_i, (\theta_i + (I-1)*p), z_i), \dots, (r_n, (\theta_n + (I-1)*p), z_n)].$$

### 4.3.3 Creation of a layer and a segment

320 The segmented lamella is assigned to each point in the series, creating thea layer and segment composed of lamellae. The inclination of the lamella is added by rotating it, as described in Figs. 5a and 5b. In the case of forming a layer (following model-1 of the microstructural arrangement), all the lamellae (n) are assigned a single value of angle of inclination ( $\delta_A$  or  $\delta_B$ ). This suggests that rotating all the lamellae of the series with an inclination of  $\delta_A$  or  $\delta_B$  formed layer-A or layer-B, respectively. For generating a segment (following model-2 of the microstructural arrangement), the lamellae are alternately assigned two  
325 distinct angles ( $\delta_A$  and  $\delta_B$ ) of inclination, indicating rotation of the alternative lamellae of the series with inclinations of  $\delta_A$  and  $\delta_B$ .

Considering that the first and the last points of the layer/segment represent the base and the apex of the skeletonmicalith, the tilt associated with the two points are  $\tau_b$  and  $\tau_a$ , respectively. The tilt values consistently change from  $\tau_b$  at the base to  $\tau_a$  at the apex. Thus, the value of tilt for i<sup>th</sup> point (i < n) of the series (i.e., layer/segment) is given by the following equation:

$$330 \quad \tau_i = \tau(i-1) + i * \text{tilt-increment} \quad (6), \text{ tilt-increment} = (\tau_b - \tau_a) / n.$$

The value of the tilt is given to each lamella in the layer/segment by rotating it as described in Figs. 5a and 5c.

The descriptions of all parameters lead to the next step of ~~reconstructing in~~ the ~~reconstruction of the full~~ ~~skeleton~~ ~~micalith~~. The two ~~skeleton~~ ~~micalith~~ reconstruction models of the microstructural arrangement have been termed ~~as~~ the layer model and the segment model, respectively. It is ~~described~~ ~~explained~~ earlier (in sections 2.2.1 and 2.2.2) ~~that~~ ~~how~~ the total number (N) of layers/segments in the whole ~~skeleton~~ ~~micalith~~ of *N. globulus* is ~~calculated~~ ~~determined~~ as 12. Thus, the angular separation between two consecutive layers/segments will, therefore, be  $p = 360^\circ/12 = 30^\circ$  (from Eq. (5)). However, the values of initial and final azimuths, initial radius and vertical distance,  $r_{\text{step}}$ ,  $Z_{\text{step}}$ , and the total number of lamellae (n) in a layer/segment are impossible to calculate without the segmentation of a layer/segment. As it is ~~really~~ ~~very~~ difficult to separate one layer/segment of lamellae from the ~~skeleton~~ ~~micalith~~, we have applied several trials with different combinations of these values and have taken those values that ~~give~~ ~~create~~ a space-filling layer/segment with the separated lamella. Importantly, the physical overlap between two consecutive lamellae is effectively close to zero. After the trials, the values of different angles and lengths are taken such that the lamellae merely touch each other without any physical overlap. Therefore, a single voxel contains only one lamella. The results suggest that the total number of lamellae in one layer/segment (n) is 15. All the values of the parameters used to reconstruct the layers/segments, following different equations given in section 4.3.2, are given in Table 1.

**Table 1: Values of the parameters for reconstructing layers/segments of *N. globulus*.**

Parameters	Values for layer model	Values for segment model
$\delta_A$	25°	25°
$\delta_B$	40°	40°
$\tau_a$	-35°	-35°
Tilt-increment	4.37°	4.37°
$\tau_b$	30.38°	30.38°
$\theta$	0°	0°
$\theta_{\text{step}}$	36°	6°
p	30°	30°
R	0.005 $\mu\text{m}$	0.005 $\mu\text{m}$
$r_{\text{step}}$	0.139 $\mu\text{m}$	0.139 $\mu\text{m}$
Z	0 $\mu\text{m}$	0 $\mu\text{m}$
$Z_{\text{step}}$	0.251 $\mu\text{m}$	0.251 $\mu\text{m}$
n	15	15
N	12	12

#### 4.3.4 Layer Model

~~Values~~ The values of each of the cylindrical coordinates ( $r_i$ ,  $\theta_i$ ,  $z_i$ ), inclinations ( $\delta_A$  and  $\delta_B$ ), and tilt ( $\tau_i$ ) for individual lamella in the first layer (layer-A) are provided in Table 2. The relative changes of these values for each of the lamellae across the length

350 (L) are graphically represented in Fig. B4 in the Appendix. A total of 12 layers (six layer-A and six layer-B) are created, alternatively putting  $I = 0$  to  $I = 11$  in series I and using the values of parameters (layer model) of Table 1. Two successive layers of -A and -B and the full ~~skeletal~~micalith reconstruction are shown in Figs. 7a-7d.

**Table 2: Values of the radius, azimuth, vertical distance, inclination, and tilt of all the 15 lamellae of the first reconstructed layer-A of *N. globulus*. The lamellae are labelled sequentially from A1 to A15.**

355

Lamellae	Radius (r) ( $\mu\text{m}$ )	Azimuth ( $\theta$ ) (degree)	Vertical distance (z) ( $\mu\text{m}$ )	Inclination ( $\delta$ ) (degree)	Tilt ( $\tau$ ) (degree)
A1	2.09	0	0.47	20	-35
A2	2.23	36	0.72	20	-30.33
A3	2.37	72	0.98	20	-25.66
A4	2.51	108	1.23	20	-20.99
A5	2.65	144	1.48	20	-16.32
A6	2.79	180	1.73	20	-11.65
A7	2.93	216	1.98	20	-6.98
A8	3.07	252	2.23	20	-2.31
A9	3.21	288	2.48	20	2.36
A10	3.35	324	2.73	20	7.03
A11	3.49	360	2.98	20	11.7
A12	3.35	396	3.23	20	16.37
A13	3.21	432	3.49	20	21.04
A14	3.07	468	3.74	20	25.71
A15	2.93	504	3.99	20	30.38

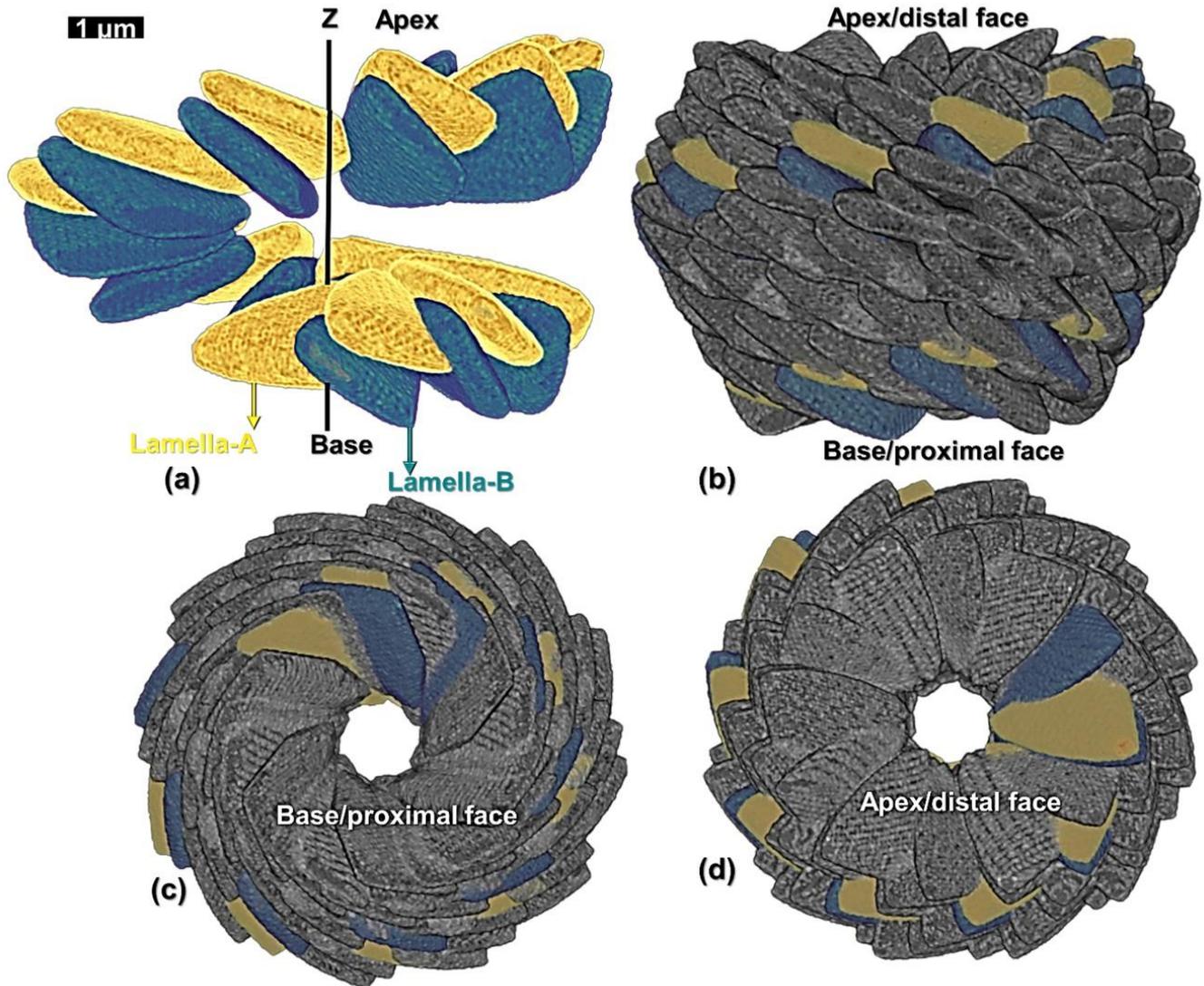


Figure 7: Reconstruction of the skeleton-micalith of *N. globulus* using the layer model. (a) Longitudinal view of the two layers formed by lamella-A (yellow) and -B (green). (b, c, and d) Longitudinal, basal and apical views, respectively, of the combination of 12 such reconstructed layers forming the entire skeleton-micalith. The first two layers of lamellae-A and B are also shown. “Proximal” and “distal” (Aubry, 2013) terms refer to the orientation of the skeleton-micalith on the cell. Proximal face: face closest to the cell membrane and distal face: face farthest from the cell membrane.

#### 4.3.5 Segment Model

Values of each of the cylindrical coordinates ( $r_i$ ,  $\theta_i$ ,  $z_i$ ), inclination ( $\delta_A$  and  $\delta_B$ ), and tilt ( $\tau_i$ ) for individual lamella of the first reconstructed segment are given in Table 3. The graphical representation of their relative changes across the length ( $L$ ) are presented in Fig. B5 in the Appendix. A total of 12 segments were generated using  $I = 0$  to  $I = 11$  in series I and the values of

parameters (segment model) of Table 1. A segment with the lamella -A and -B and the full skeletal reconstruction are shown in Figs. 8a-8d.

**Table 3: Values of the radius, azimuth, vertical distance, inclination, and tilt of all the 15 lamellae of the first reconstructed segment of *N. globulus*. The lamellae-A and -B are sequentially marked from A1 to A8 and B1 to B7, respectively.**

Lamellae	Radius (r) ( $\mu\text{m}$ )	Azimuth ( $\theta$ ) (degree)	Vertical distance (z) ( $\mu\text{m}$ )	Inclination ( $\delta$ ) (degree)	Tilt ( $\tau$ ) (degree)
A1	2.09	0	0.47	20	-35
B1	2.23	36	0.72	20	-30.33
A2	2.37	72	0.98	20	-25.66
B2	2.51	108	1.23	20	-20.99
A3	2.65	144	1.48	20	-16.32
B3	2.79	180	1.73	20	-11.65
A4	2.93	216	1.98	20	-6.98
B4	3.07	252	2.23	20	-2.31
A5	3.21	288	2.48	20	2.36
B5	3.35	324	2.73	20	7.03
A6	3.49	360	2.98	20	11.7
B6	3.35	396	3.23	20	16.37
A7	3.21	432	3.49	20	21.04
B7	3.07	468	3.74	20	25.71
A8	2.93	504	3.99	20	30.38

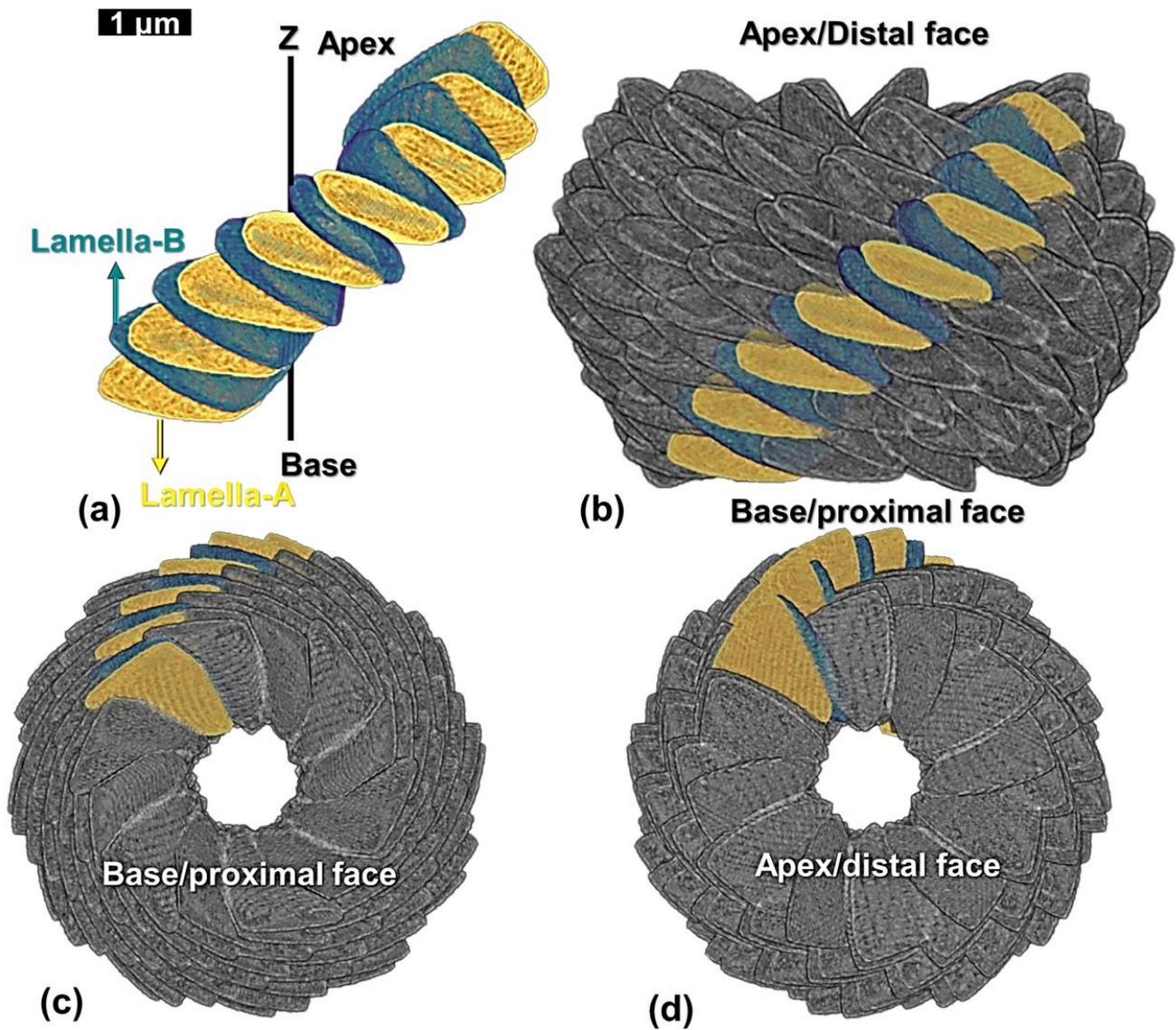


Figure 8: Reconstruction of the skeleton-micalith of *N. globulus* using the segment model. (a) Longitudinal view with alternating, properly oriented, lamella-A (yellow) and -B (green). (b, c, and d) Longitudinal, basal and apical views, respectively, of the combination of 12 such reconstructed segments forming the entire skeleton-micalith. A segment containing alternatively placed lamella-A and -B is highlighted. “Proximal” and “distal” (Aubry, 2013) terms refer to See Fig. 7 for terminology regarding the orientation of the skeleton on the cell. Proximal face: face closest to the cell and distal face: face farthest from the cell-micalith.

## 5 Discussion

### 5.1 Which of the two models best explains the *Nannoconus* skeletonmicalith?

PXCT ~~alone is insufficient to conclusively distinguish between the two models. However, it indeed~~ provides the successful  
380 3D reconstruction of the *Nannoconus* skeletonmicalith structure. ~~The, whatever the considered model (layer and/or-segment  
models are based on two different concepts of skeletal microstructure of *N. globulus*. Both models result in).~~ Indeed, similar  
reconstructions (Figs. 9a and 9b) of the skeletonmicalith were obtained, which structurally resembles the real specimen (Fig.  
9c). ~~Each~~ However, each of these two models of the microstructural arrangement suggests a distinct biomineralization process  
to form the entire skeletonmicalith. The first model implies that the skeletonmicalith is formed by lamellae organized in  
385 successive layers, whereas the second one involves ~~that the skeleton is formed a~~ primarily of organization of lamellae in  
segments, each defined by the stacking of lamellae. ~~However, the following observations/arguments show that the layer model  
is insufficient to account for the structure~~ To resolve this issue, we supplement our study with careful observation of SEM  
images of other *Nannoconus*, and that the segment model is the right one. The species available in the literature. In particular,  
on SEM images of one of the youngest species, *N. funiculus* (reported at ~90 Ma; Lees and Bown, 2016), the layered structure  
390 of *Nannoconus* is distinctively visible in scanning electron microscope (SEM) images, but they layers are clearly organized in  
segments, as it is clearly visible on one of the youngest species, *N. funiculus* (reported at ~90 Ma; Lees and Bown, 2016; Fig.  
B6a, in the aAppendix). Clear segment boundaries are also observed in several species (Figs. B6a-d in the aAppendix), from  
the oldest one (*N. compressus*, reported at ~140 Ma; Bralower et al., 1989; Fig. B6d, in the aAppendix) to the youngest one  
(Fig. B6a, in the aAppendix). In many recrystallized or overgrown specimens, the individual layers are no longer visible; they  
395 have fused to form sorts of bricks, but the organisation in segments remains (Fig. B6d, in the appendix). AsAppendix). Another  
argument favoring the segment model is that, as described earlier, the Ggenus *Nannoconus* belongs to the Ffamily  
*Nannoconaceae* (Reinhardt, 1966), which is included in the Oorder *Braarudosphaerales* (Aubry, 2013; Lees and Bown, 2016).  
This order also includes the Ffamily *Braarudosphaeraceae*, which shares a strong evolutionary link with *Nannoconaceae*;  
and therefore, with *Nannoconus*, as described by Lees and Bown (2016) and Aubry (2025). According to Lees and Bown  
400 (2016), the pentaliths of the family *Braarudosphaeraceae* are characterized by “five segments formed from stacks of non-  
imbricated laminae/elements”, whereas *Nannoconaceae* are defined by “numerous stacked, imbricating elements.” Here,  
“laminae/elements” refers to lamellae. *Braarudosphaera bigelowii*, an extant species of *Braarudosphaeraceae*, with fossil  
representatives first occurring 100 Myr ago, calcifies a pentagonal skeletonmicalith with the combination of five trapezoidal  
segments (Fig. 4 in Hagino et al., 2016) composed of parallel “lamellae stacked in segments” (Aubry, 2013). Given the close  
405 evolutionary relationship between these two Ffamilies, it is reasonable to hypothesize that *Nannoconaceae* (and therefore,  
*Nannoconus*) may have calcified via a similar process, where imbricating lamellae were indeed stacked into segments.

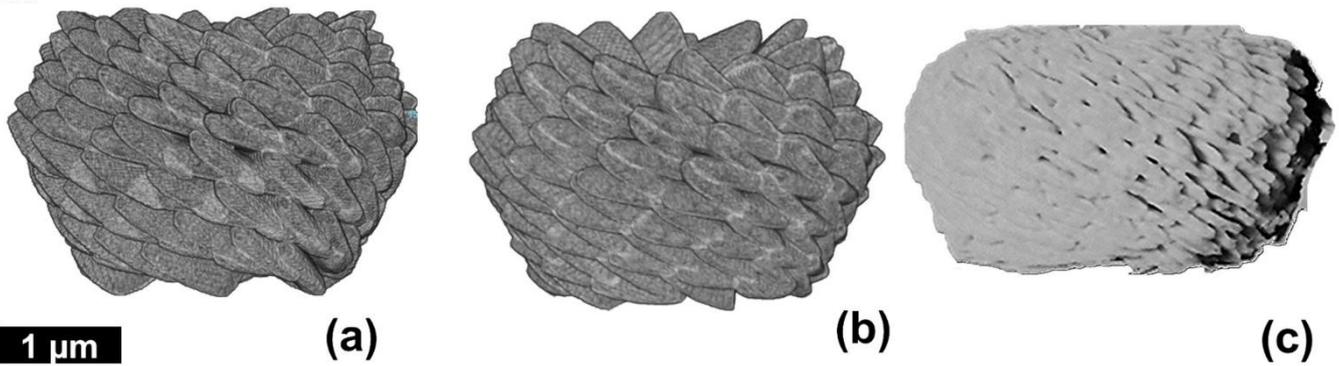


Figure 9: Two different reconstructions of the entire skeletal micalith of *N. globulus*. (a) reconstruction based on the layer model and (b) reconstruction based on the segment model. Similar skeletal reconstruction is also proposed (at low resolution) by Aubry (2025), Fig. 12d. (c) External view of the specimen of *N. globulus* used in the PXCT experiment (see also Fig. 3b).

## 5.2 Which parameters control *Nannoconus*'s skeletal micalith morphology?

Considering the segment model, we now try to show which parameters control the inter-specific morphological variability observed within the *Nannoconus* group during the ~30 Myr of the Early Cretaceous. The parameters that can potentially control the morphology of the skeletal micalith are the angle of rotation of the segment and the radius. The angle of rotation of each segment (Eq. (4) in section 4.3.2) is considered as set at 90° in the discussed reconstruction by the segment model. With the same number of total numbers of segments ( $N = 12$ ) and total lamellae in each segment ( $n = 15$ ) used in the segment model, we have applied three different angles of rotations to individual segments: 30°, 60° and 180°. All of the angles of rotation we have used, resulted in reconstructed skeletons (Figs. 10a–10d) micaliths of similar shape. So, it can (Figs. 10a - 10d). It may be inferred from this that the angle of rotation of the segments does not affect the general shape of *Nannoconus* the micalith.

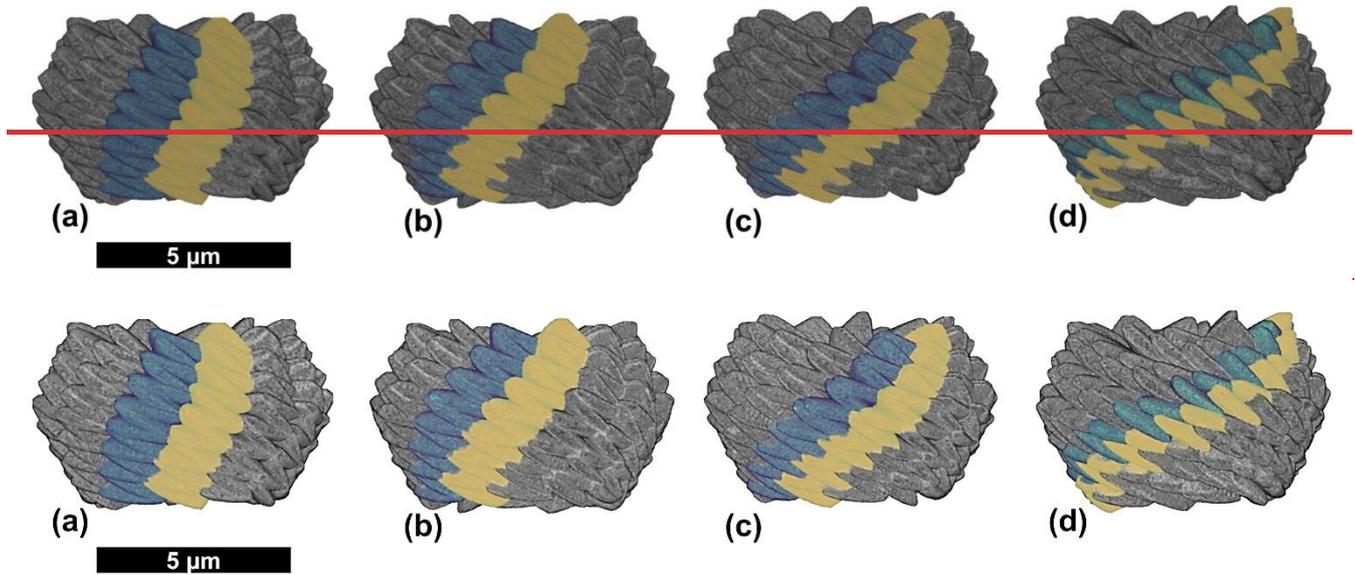


Figure 10: Four complete reconstructed *Nannoconus* skeletal micaliths based on the segment model with four different angles of rotation of each segment. (a-d) These angles of rotation are 30°, 60°, 90°, and 180° respectively. Two consecutive segments of each reconstructed skeletal micalith are marked in green and yellow.

The other parameter that can potentially affect the morphology of the skeletal micalith is the radius. As we discussed earlier, the particular specific way of changing the radius with the length (see Eq. (1)) resulted in the globular shape of the skeletal micalith. Thus, different ways of changing the radius would give rise to different skeletal micalith morphologies. Five different morphologies (globular, hour-glass, conical, barrel-shaped, cylindrical; Fig. 11) of the skeletal micaliths have been reconstructed by applying the same segment model and using for each, the same values of the differential parameters (Table 1) except for the radius (Table 4); we obtained reconstructions that resemble the morphologies of *Nannoconus* real species. Thus, it is determined that the radius is the parameter that controls the inter-specific morphological variability of *Nannoconus*. However, other than radius, the total number of the segments forming the entire skeletal micalith could also vary for different species of *Nannoconus*. It should be kept in mind that in this discussion, the number of segments is considered accepted to be 12 based on a SEM image of only one species, that is *N. globulus*. A comparative study of SEM images of different species of *Nannoconus* could be useful to investigate whether the number of the segments in the skeletal micaliths is species-dependent. More generally, with

Applying the model of reconstruction described model developed in this study, we have succeeded in creating successfully produced a 3D skeletal three-dimensional micalith of *Nannoconus*; that means that we could. More generally, this approach will enable us to constrain changing skeletal variations in skeletal morphology not only at the genus level of the Genus, but also at the ordinal level of the Order of (*Braarudosphaerales*), thereby allowing us to investigate whether a particular skeletal morphology had been favoured during an assessment of whether specific skeletal morphologies were preferentially selected

over the long evolutionary history (~150 Ma) of this order. (~150 Ma). Recent research (Aubry, 2025) suggests that the importance of this Order lies in its distinct skeletal structure, indicating both a taxonomic specificity and a unique biological position within the calcareous nannoplankton group.

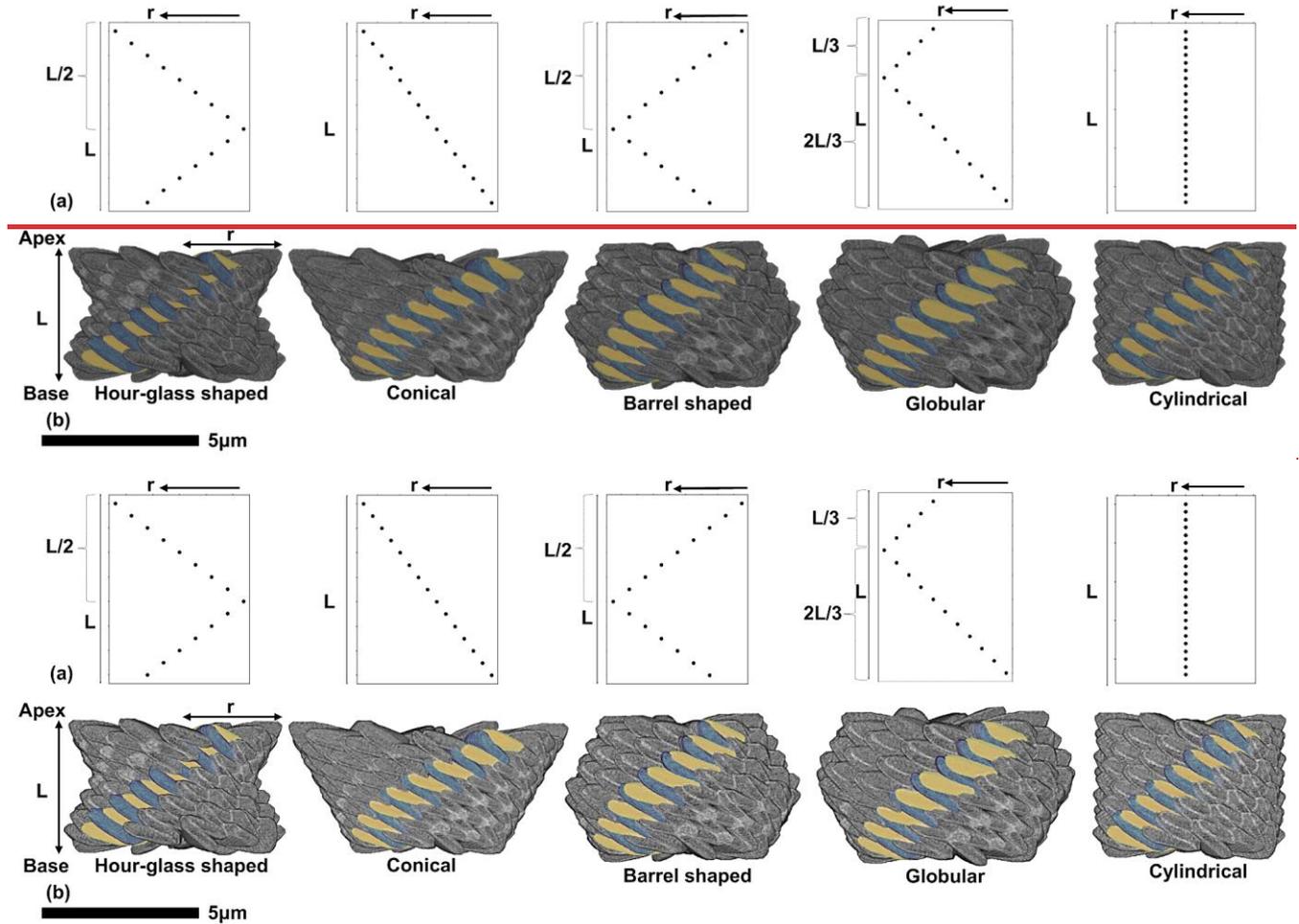


Figure 11: Five different **skeletal** morphologies of *Nannoconus*, each with distinct variation of radius (r) across the length (L). (a) Graphical expressions of five different variations of radius along the length of the reconstructed **skeletal**, as shown in Table 4. Each of the points on the graphs indicate individual lamella of a segment. (b) **Skeletal** reconstructed following each of the possibilities, using the segment model, with each **skeletal** highlighting a segment containing both types of lamellae-A (yellow) and -B (green).

Table 4: Five possible variations in radius along the length (from base to apex) of the reconstructed **skeletal** of *Nannoconus* are presented, each associated with distinct skeletal morphologies (as shown in Fig. 11). Corresponding species of *Nannoconus* exhibiting these respective morphologies are also provided.

Change of radius	Final morphology	Species
------------------	------------------	---------

Radius decreases until $\frac{1}{2}$ of the total length from the base and then increases to the apex	Hour-glass shaped	<i>N. abundans</i>
Radius constantly increases from the base to the apex	Conical	<i>N. steinmannii</i>
Radius increases until $\frac{1}{2}$ of the total length from the base and then decreases to the apex	Barrel shaped	<i>N. circularis</i>
Radius increases until $\frac{2}{3}$ <sup>rd</sup> of the total length from the base and then decreases to the apex	Globular	<i>N. globulus</i>
Radius is constant from the base to the apex	Cylindrical	<i>N. truttii</i>

### 5.3 How were the segments of the *Nannoconus* ~~skeleton~~micalith biomineralized?

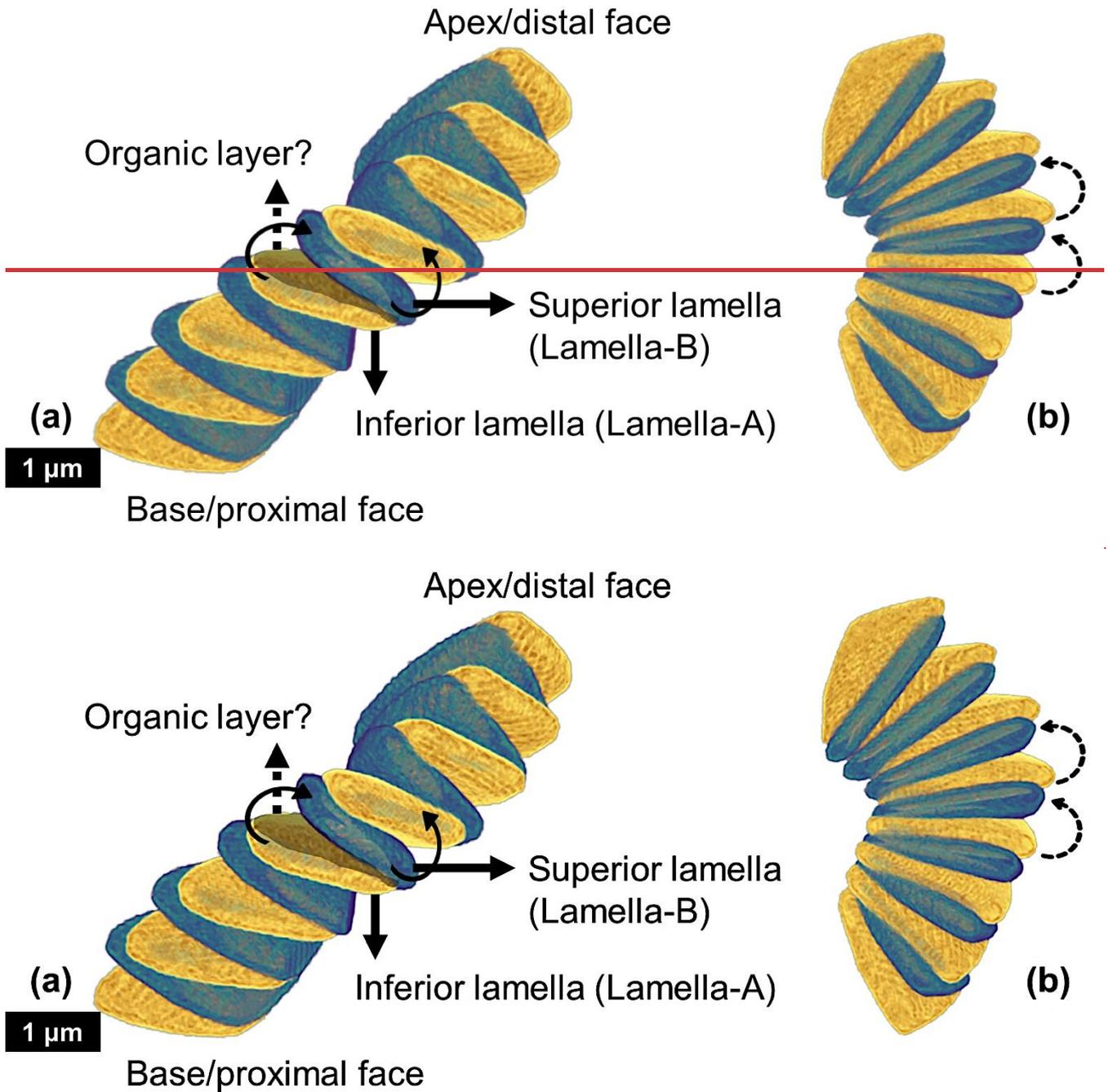
It is now important to infer the process of biomineralization of the segments of the *Nannoconus*. Inferences can be provided based on the comparison of the biomineralization process of *B. bigelowii*, as documented in Hagino et al., (2016). In an uncalcified living cell of *B. bigelowii*, an organic substrate is reported that resembles the shape of the pentagonal skeleton (Fig. B7a in the Appendix). The organic substrate is divided into five parts, with each of the parts mimicking the shape of a segment (Fig. B7b in the Appendix). In the vertical segments of the calcified skeleton produced by living *B. bigelowii*, thin organic layers are observed between two successive lamellae. Thus, the organic substrate and layers are seen to act as a “template” for the calcification of the segments and lamellae. This would indicate that the lamellae are stacked in each of the five parts of the organic substrate, templated by the organic layers, creating five segments. We suggest a similar process of calcification for *Nannoconus* with 12 segments; the calcification occurred on an organic “substrate” divided into 12 parts (since 12 segments are considered) (Fig. B7c in the Appendix). Additionally, lamellae in the segments of *Nannoconus* are placed with inclination and tilt, creating their interlocking arrangement, whereas they are parallel in the segments of *B. bigelowii*. A mineralization process involved in the formation of a *Nannoconus* micalith would likely include four components: (a) the lay out of the duos of calcitic lamellae and (b) their organization into segments, and, also, their acquisition of (c) a tilt and of (d) an inclination. The extant species *Braarudosphaera bigelowii* provides a model for the layout of the mineral lamellae and their organization into segments. The mineralization of its pentaliths is an extracellular process mediated by an organic template divided into five symmetrical parts in the shape of the segments and by extremely thin layers of organic matter between the calcitic lamellae (Hagino et al., 2016; Fig. B7a, b). Applying this to *Nannoconus globulus*, we may hypothesize that the mineralization would have involved an organic template divided into 12 symmetrical parts (each a segment), with very thin layers of organic matter controlling the formation of the lamellae (fig. B7c). A complication in *Nannoconus* compared to *Braarudosphaera* is the association of the lamellae in duos with different inclination and tilt.

The inclination and tilt are explained by two mutually perpendicular rotations of the lamellae (see section 4.3.1) ~~of in the~~ *Nannoconus micalith*. Such ~~rotations with~~ specific rotational directions (i.e., clockwise and anti-clockwise) are commonly  
480 observed in biomineralized skeletons produced by marine organisms; for example, the coiling of gastropod's shell (Ueshima and Asami, 2003), the overlapping of the chambers generated by foraminifera (Schiebel and Hemleben, 2017), microstructural imbrications of the calcite units in the ~~skeletons~~coccoliths produced by calcareous nannoplankton (Young et al., 1999; Aubry, 2013, 2025). ~~It is implied that these~~These rotations are influenced by biomolecules such as proteins, amino acids, and polysaccharides (Young and Henriksen, 2003; Yu et al., 2005; Jiang et al., 2017, 2018, 2019). ~~Was it the case for Nannoconus?~~  
485 ~~To discuss this point, we presented here~~An in vitro experiment to examine the ~~process~~acquisition of ~~mineralization of vaterite~~ ~~(a polymorph~~chirality by skeletons consisting of calcium ~~carbonate, CaCO<sub>3</sub>)~~induced by amino acid carbonates (Jiang et al., 2018). ~~provides insightful means by which the Nannoconus lamellae may have acquired inclination and tilt. These authors have observed that the addition of chiral acidic amino acid Aspartine to a supersaturated calcium- and carbonate-containing growth solution results in the formation of "small vaterite helicoid-shaped suprastructures with subunit inclinations and spiralling curves chiral platelets oriented in the right-handed direction" (op. cit., p.2).~~ The growth ~~of the superstructure~~ occurs layer-by-layer with ~~the~~ amino acid intervening between two consecutive ~~layers to form the microstructure of the layered vaterite. The two layers are termed ("mother" and "daughter" layers. The amino acid (i.e., Aspartic acid) present between the mother and the daughter layers, rotates")~~ mineral layers, and also rotating the daughter layer clockwise (Figs. 6A and 6B in Jiang et al., 2018), ~~creating~~ which produces a difference in inclination ~~of the platelets~~ between the ~~two~~coupled layers. Thus,  
490 because of the presence of amino acid between ~~the~~ two consecutive layers, the "inclination" of the platelets in the new layers ~~added during growth,~~ increases continuously. ~~during growth.~~

The amino acid induced rotation of the layered vaterite structurally resembles the interlocking arrangement of the lamellae of ~~a Nannoconus micalith. (the lamellae of the micalith are the equivalent of the "platelets" of the suprastructure in the experiment above).~~ However, the reconstruction of the ~~skeleton of Nannoconus showed~~micalith shows that not only one but two rotations  
500 of the lamellae in two mutually perpendicular directions are necessary to develop the interlocking pattern (as discussed in section 4.3.1). ~~Based on the comparison with~~By comparing the layer-by-layer grown vaterite, we may hypothesize on the ~~probable~~ presence of a layer of amino acid(s) or any other biomolecule(s) to ~~explain~~have controlled the two rotations between two successive lamellae of a Nannoconus micalith, creating the tilt and the inclination. These ~~successive~~consecutive lamellae could correspond, respectively, to the mother ~~and~~ daughter layers in the vaterite experiment. For two such  
505 ~~successive~~consecutive lamellae ~~of in~~ a segment we ~~call~~refer to the preceding (~~similar~~equivalent to "mother layer" of Jiang et al., 2018) and succeeding (~~similar~~equivalent to "daughter layer" of Jiang et al., 2018) lamellae as the "inferior" and "superior" lamellae, respectively (Fig. 12). The ~~sense of the rotations~~rotational directions to generate the inclination and the tilt are ~~marked~~shown in the inferior and superior lamellae in Fig. 12. The required arrangements for these two lamellae are: (1) for the inclination, the opposite directions (both clock and anti-clockwise) of successive rotations (~~given~~shown by the black bold

510 arrows in Fig. 12a), and, (2) for the tilt, the same direction (anti-clockwise) of successive rotations (~~given shown~~ by the black dashed arrows in Fig. 12b). The opposite ~~direction of rotations~~ rotational directions in the inferior and superior lamellae ~~actually~~ generate the two lamellar types i.e., lamella-A and -B. Therefore, both the clock and anti-clockwise ~~rotations~~ rotational directions occur simultaneously during biomineralization of the superior lamella ~~on over~~ the inferior lamella, generating the interlocking arrangement of the constituting lamellae segments. These ~~possible hypothetical~~ explanations strengthen our initial  
515 hypothesis of the presence of an amino acid(s)/biomolecule(s) ~~containing rich~~ layer, between two ~~successive consecutive~~ lamellae. ~~This layer, containing the~~ An organic substrate of amino acid(s)/biomolecule(s), ~~could correspond to the organic substrate occurring as the~~ “template” (see above) for the calcification of the segments/micaliths of *Nannoconus*.

These organic layers, if ~~they were originally~~ present in the *Nannoconus* skeleton/micalith, are very unlikely to be preserved in the fossil record. ~~However, specimens of *Nannoconus* have been studied. Yet, a preliminary study~~ with confocal Raman micro-  
520 spectroscopy, ~~and the obtained spectra have shown a~~ on micaliths of *Nannoconus* yielded a spectral signal ~~that could be related to~~ (Fig. B8 in the Appendix), possibly indicative of preserved organic matter within ~~the skeleton~~ (Fig. B8 in the Appendix). The confocal Raman spectra of *Nannoconus* are characterized by a number of peaks, with distinct intensities at specific wavenumbers. The spectra present two distinct parts: one from 150 to 1200  $\text{cm}^{-1}$  with characteristic peaks of the skeletal  $\text{CaCO}_3$ , and the second one from 1200 to 2000  $\text{cm}^{-1}$ , which could correspond to organic matter signature preserved within the  
525 skeleton. Indeed, in a study focused on the identification of organic matter in bioclastic grains encountered in marine carbonate rocks, Moyathem (Giraud et al., (2023) performed confocal Raman micro spectroscopy and detected ~~on the spectra the fingerprints of organic matter in the higher wavenumber region (i.e., wavenumber >1200  $\text{cm}^{-1}$ ).~~ Additionally, Raman signals related to protein and polysaccharide are also reported in the same spectral region from biocalcite produced by extant calcareous nannoplankton (Silvestri et al., 2020). Therefore, a future investigation ~~research in progress~~. Further investigations  
530 using tomographic technique with a resolution finer than the PXCT, such as atom probe tomography (resolution  $\sim 0.5$  nm), could be useful is required to confirm the ~~preservation~~ presence of in situ organic matter ~~preserved~~ within the skeleton/micalith of *Nannoconus*.



535 Figure 12: Reconstructed segment of *N. globulus* with the proposed inferior and superior lamellae, along with the hypothesized intervening organic layer in between ~~them~~. (a) The two black bold arrows show the two opposite directions of rotation (clock and anti-clockwise), which create distinct inclinations in the two successive lamellae, generating the lamella-A (yellow) and -B (green). (b) Same segment, with two dashed black arrows presenting anti-clockwise rotations in two successive lamellae, generating the tilt in them.

540 The *Nannoconus* skeleton is reconstructed from a specimen of *N. globulus*. Although the specimen is generally well preserved  
as seen in SEM images, PXCT observations at higher resolution reveal finer diagenetic alterations within the skeletal interior.  
Nevertheless, our reconstruction process enables the segmentation of individual lamellae and the generation of a 3D skeleton  
using geometric parameters, including tilt and inclination angles as well as radial length. To ensure accuracy, these geometric  
parameters i.e., radius, inclination, and tilt, were measured repeatedly, and the 3D skeleton was iterated through multiple trials  
545 to obtain the closest possible resemblance to the real skeleton. As a result, the internally detected diagenetic effects do not  
affect the validity of the 3D reconstruction allowing subsequent interpretations of biomineralization. The strategy is therefore  
independent of diagenetic alteration, and thus, applicable to explain the skeletal microstructure of *Nannoconus* specimens even  
with poor state of preservation. It is noteworthy that the 2D SEM images of various *Nannoconus* species from different  
geological settings and ages show that their skeletons consist of interlocking lamellae, and species differ in overall skeletal  
550 morphology. The PXCT based 3D reconstruction of *N. globulus* as obtained from the segment model, clarifies this  
microstructure as: the interlocking pattern results from lamellae stacked at two distinct inclinations, and changes in radius  
control the general morphology. Thus, a single specimen provides sufficient geometric parameters to interpret 3D  
microstructure across species, and the resulting biomineralization inferences are applicable to *Nannoconus* species through  
different geological intervals. We have hypothesized above that the biomineralization of the lamellae of the *Nannoconus*  
555 segments is “templated” by an organic layer containing amino acid(s)/biomolecule(s). The rotations induced by the amino  
acid(s)/biomolecule(s) created the inclination and tilt associated with the lamellae, hence generating the interlocking  
arrangement of the lamellae. Both the clock and anti-clockwise rotations are inferred in the same skeleton of *Nannoconus*.  
Such “biomolecule(s)” influenced natural biomineralized 3D microstructure with repetitive arrangement of units like the  
calcite lamellae as observed in *Nannoconus*, are often studied to synthesize biomimetic materials (Jiang et al., 2019). These  
560 materials, engineered for enhanced physical (e.g., hardness) and chemical (e.g., solubility) properties (Cho et al., 2023), have  
applications in research fields like catalysis and biomedicine. Therefore, the 3D skeleton reconstruction of *Nannoconus* can  
definitely provide significant insights into biomimetic material design, expanding its relevance beyond Palaeontology.

## 6 Conclusion

Among calcareous nannofossils, the *Nannoconus* group of the Order *Braarudosphaerales* (fossil and extant representatives)  
565 presents sophisticated massive skeletons, organized in an interlocking arrangement of calcitic lamellae stacked in segments,  
forming a wall around a central canal. They contributed to huge carbonate accumulations over ~30 million years in the Early  
Cretaceous seas. The biomineralization process that led to the production of these massive skeletons was unknown. The aim  
of the present study was to obtain a proper understanding of this process from a 3D reconstruction of the skeleton. The aim of  
this study was to obtain a sound understanding of the structure of the *Nannoconus* micolith as a first step towards understanding  
570 the mineralization process in an extinct genus that dominated the Early Cretaceous Tethys epicontinental nannoplankton  
communities for over 30 Myr. A set of Ptychography X-ray computed tomography (PXCT) with synchrotron radiation at the

SWING Beamline of SOLEIL (French synchrotron) was applied on several well-preserved micaliths of different *Nannoconus* skeletons species to understand and document the microstructural arrangement of the crystal components (lamellae) at the nanometer level (finer than the thickness of a single lamella). ~~The result~~ For the first time, a 3D micalith reconstruction has been achieved for one specimen of *Nannoconus*. We present here the result of the experiment obtained from a hand-picked specimen of *Nannoconus globulus*. The result was a series of tomographic image slices (3D resolution ~40 nm) for the ~~skeleton~~ micalith. Then, one lamella, virtually separated from the image slices, ~~is~~ was used to reconstruct the entire ~~skeleton~~ micalith of *Nannoconus* in the ORS-Dragonfly software. ~~For the first time, a 3D skeletal reconstruction has been achieved for *Nannoconus globulus*.~~ The interlocking arrangement of the lamellae results from two different angles, termed inclination and tilt, while changing radius (minimum ~2.1  $\mu\text{m}$  to maximum ~3.5  $\mu\text{m}$ ) controls the micalith morphology. Thus, a single specimen provides sufficient geometric parameters to interpret 3D microstructure across species, and the resulting biomineralization inferences are applicable to *Nannoconus* species through different geological intervals. ~~skeletal morphology.~~ ~~The 3D model of skeleton~~ The 3D model of micalith reconstruction developed in the present study could be further applied to the reconstruction of the ~~skeleton~~ micalith of other ~~G~~ genera within the ~~O~~ order of *Braarudosphaerales* to investigate if a particular ~~skeleton~~ micalith morphology has been favoured during the long evolutionary history of this order. ~~The biomineralization~~ PXCT 3D reconstruction of the *Nannoconus* micalith structure, combined with existing 2D SEM observation, supports the interpretation of the *Nannoconus* micalith as being composed of “segments”. Further research can now be undertaken based on the hypothesis that the biomineralization of the lamellae of the *Nannoconus* segments is hypothesized to be “templated” by organic layers containing amino acid(s)/biomolecule(s). During ~~We hypothesize that during~~ the biomineralization of the lamellae, ~~the amino acid(s)/biomolecule(s) are proposed to~~ have induced clockwise and anti-clockwise rotations, creating the inclination and tilt and therefore generating the interlocking arrangement of the lamellae. ~~PXCT 3D reconstruction of the *Nannoconus* skeletal structure, combined with existing 2D SEM observation, as well as biomineralization hypotheses, supports the interpretation of the *Nannoconus* skeleton as being composed of “segments” in the same way as *Braarudosphaera* is and probably other Mesozoic nannofossils (Aubry 2025).~~ Biomolecule(s)-driven naturally produced microstructures characterized by the repetition of units such as the 3D arrangement of the lamellae of ~~*Nannoconus*'s skeleton~~ the *Nannoconus* micalith, are often extensively investigated for designing biomimetic materials. Such materials with artificially enhanced physico-chemical properties (e.g., hardness, solubility) are implemented in research with specialisation to catalysis and biomedicine, therefore making this publication relevant and important, outside the study of calcareous ~~nannofossils~~ nannoplankton.

## 600 Appendix A

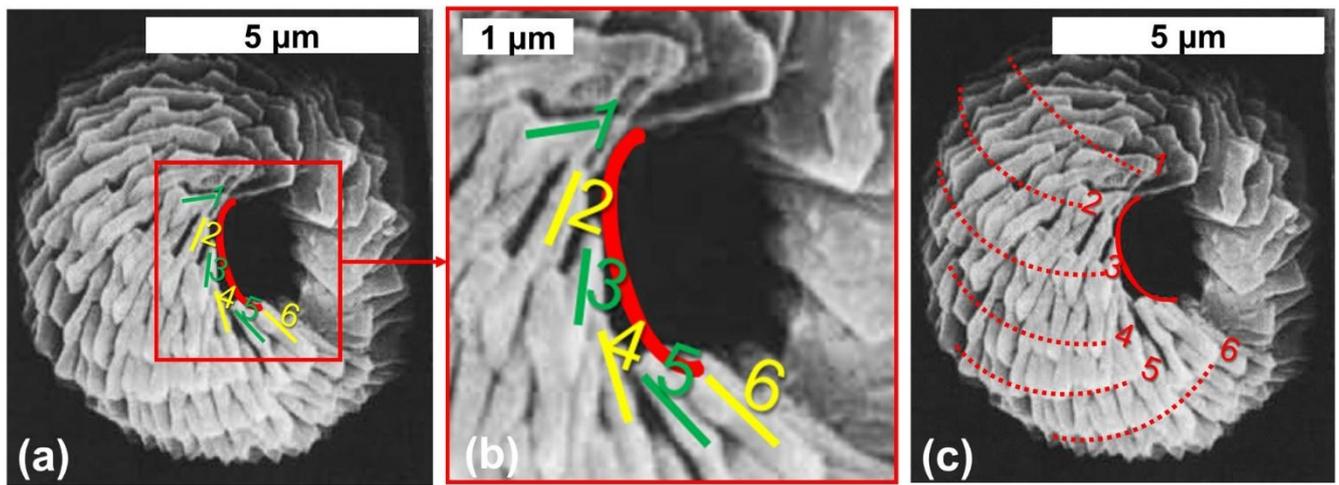
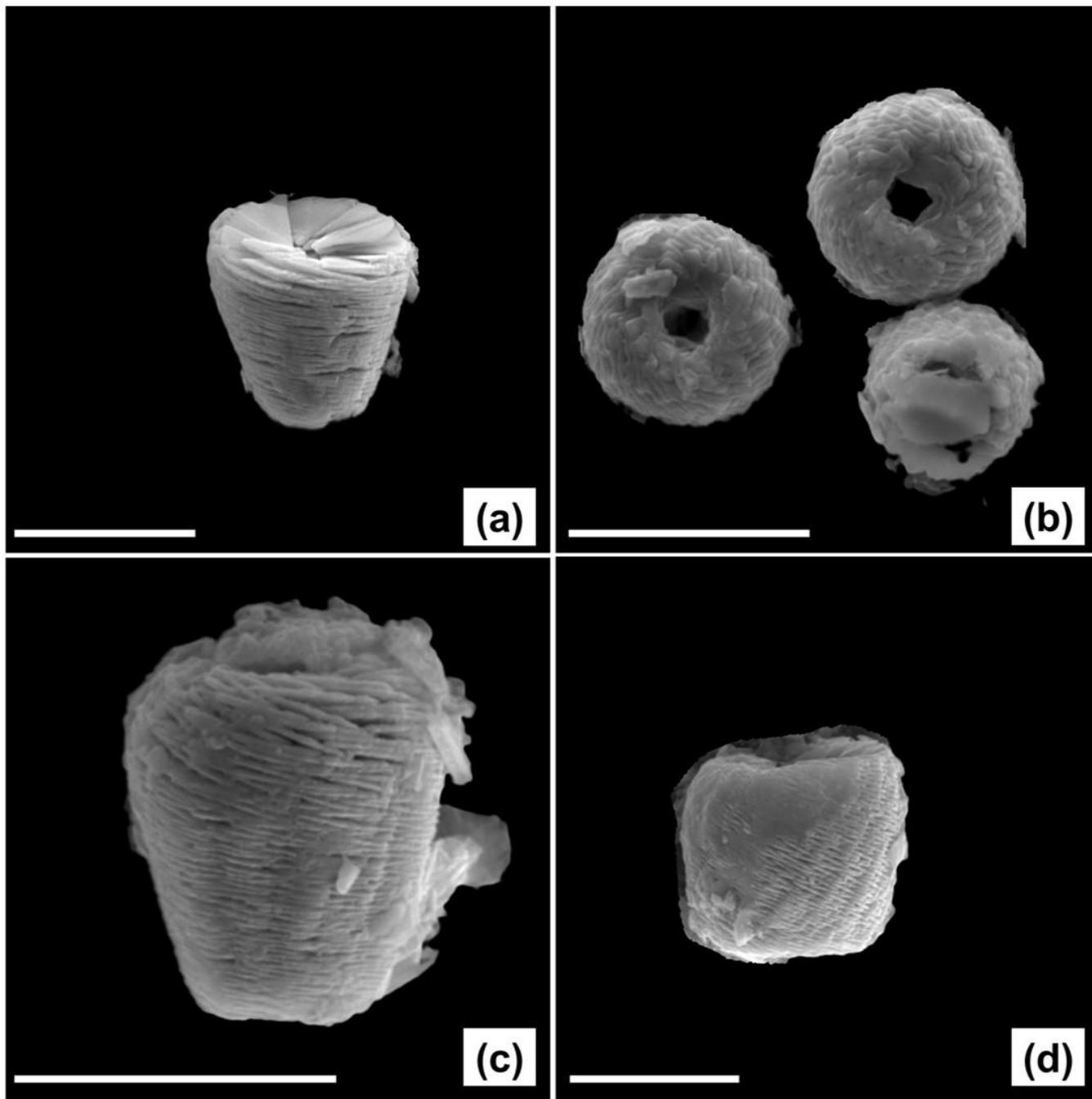
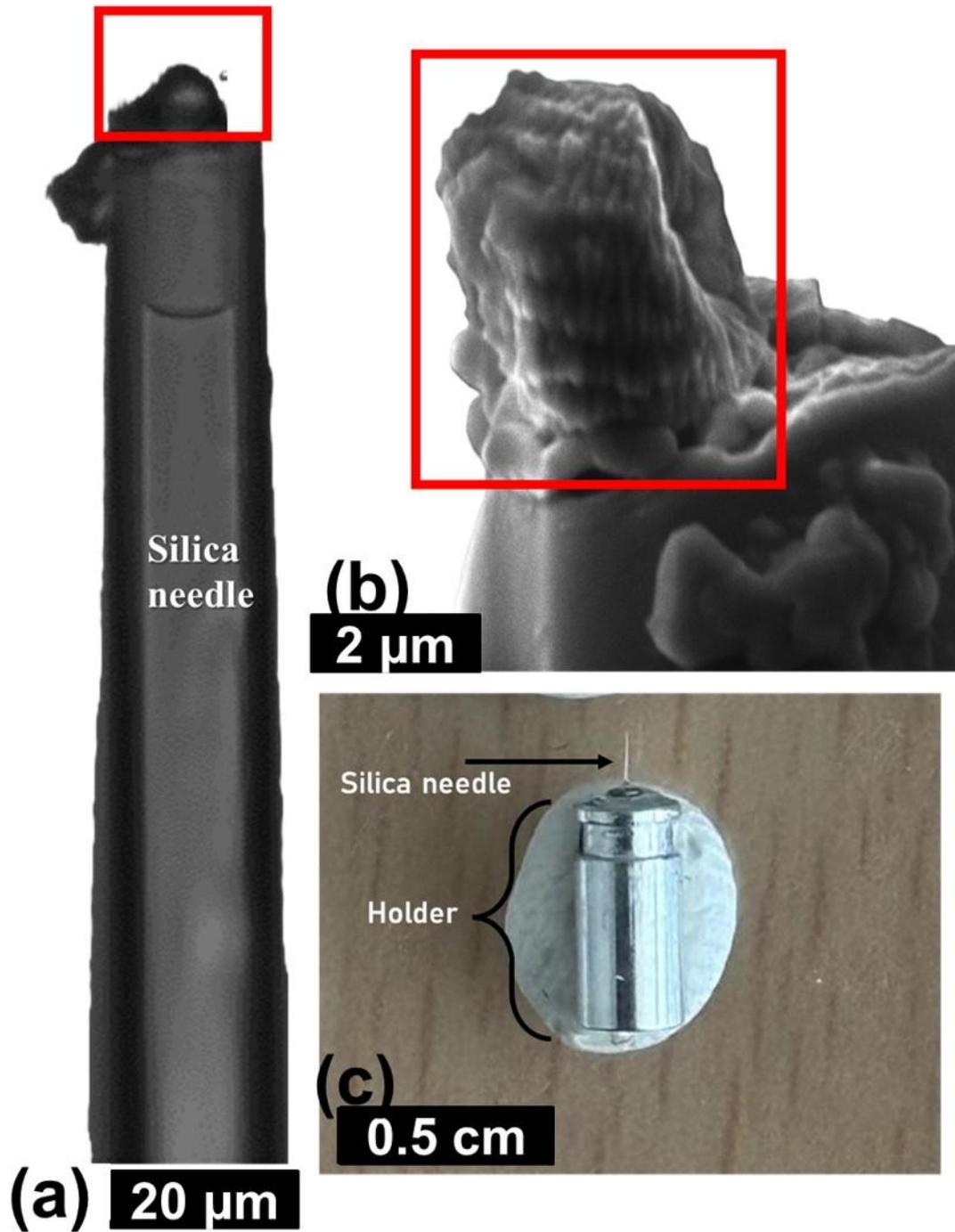


Figure B1: Three-quarter view of the distal face of *N. globulus* in scanning electron microscopy (SEM). (a) Six lamellae in the 180° transect of the apex (given by the red curve) are highlighted. Three of lamellae-A and three of lamellae-B are numbered in green and yellow colours, respectively. (b) Magnified view of the part delineated by the red box in (a) with all the six lamellae: three lamellae-A (yellow) and three lamellae-B (green). (c) Boundaries between six segments highlighted by red-dotted lines in the same 180° transect of the apex of the *N. globulus*. SEM image modified from Covington and Wise, 1987.

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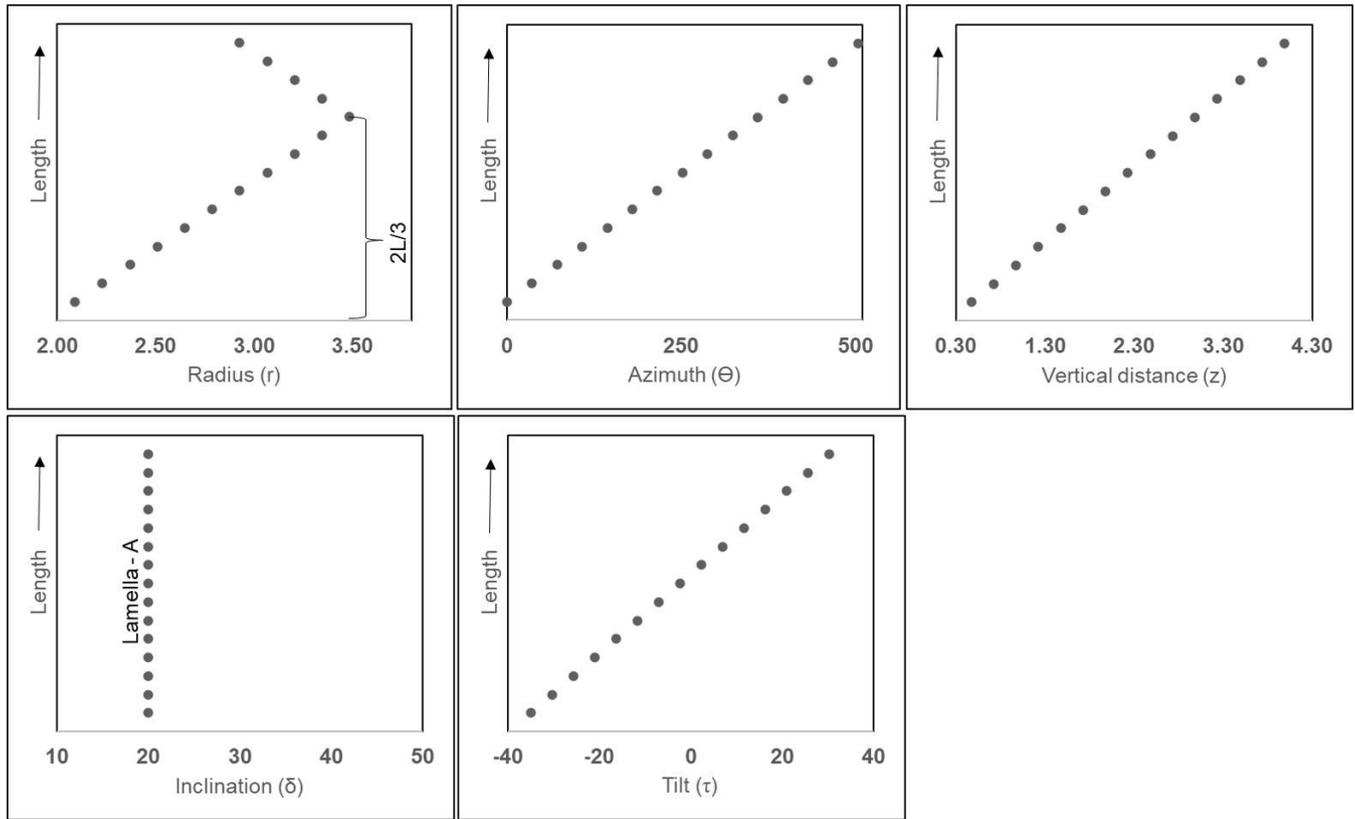


610 Figure B2: SEM images of well-preserved specimens of *Nannoconus*, from DSDP Leg-93-Site 603 (continental rise of the western margin of the North Atlantic) eCore 44, interval 115-116 cm. Individual lamellae can be clearly distinguished in each of the specimens thanks to their good preservation. Identification: (a) *N. steinmannii*, (b) *N. globulus*, (c) *N. kamptneri*, (d) *N. circularis*. The white line represents 10  $\mu\text{m}$ .

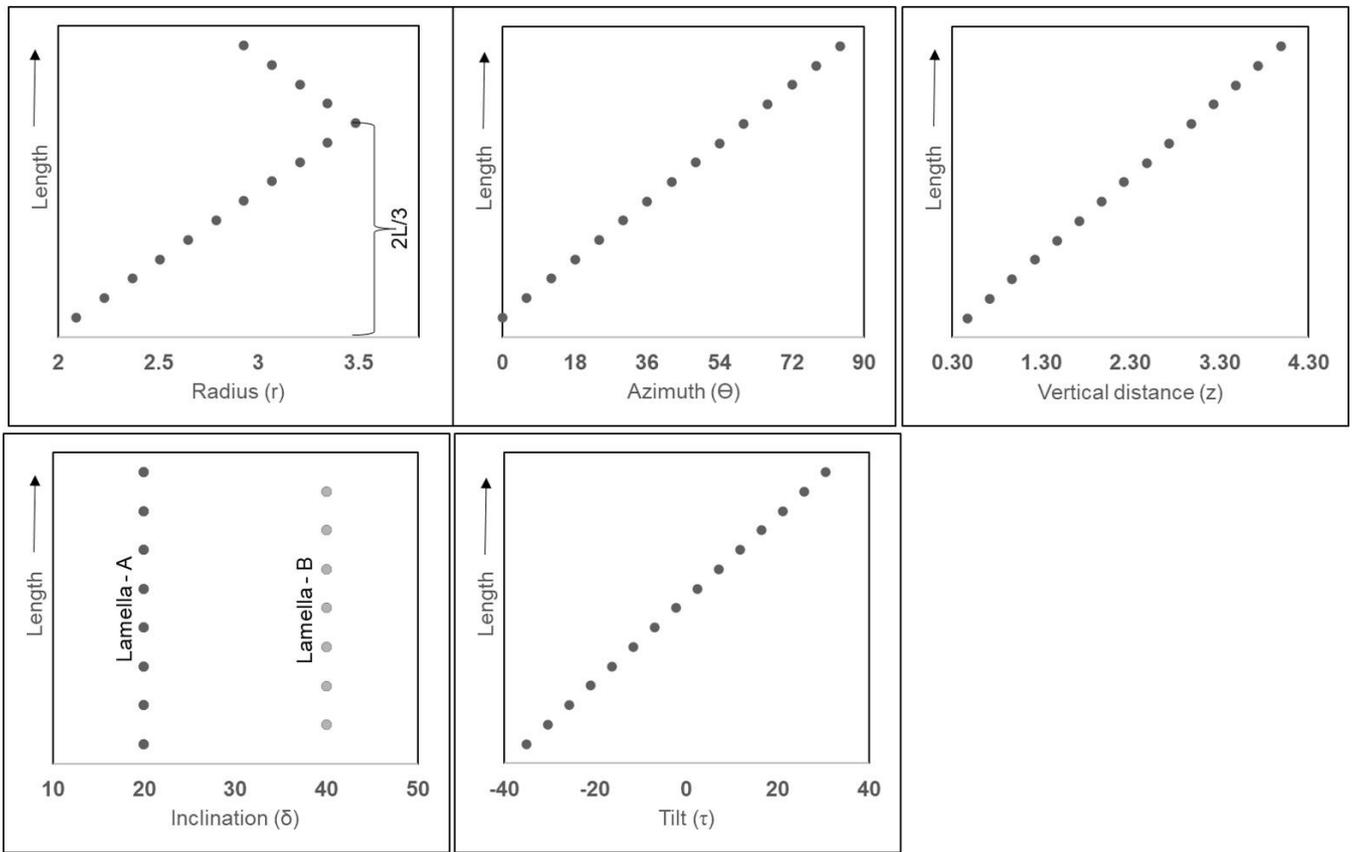


615 Figure B3: Procedures used for the sample preparation of synchrotron-based ptychography X-ray computed tomography experiment, applied on *N. globulus*. (a) A specimen of *N. globulus* (inside the red box) is manually picked with a silica needle. (b)

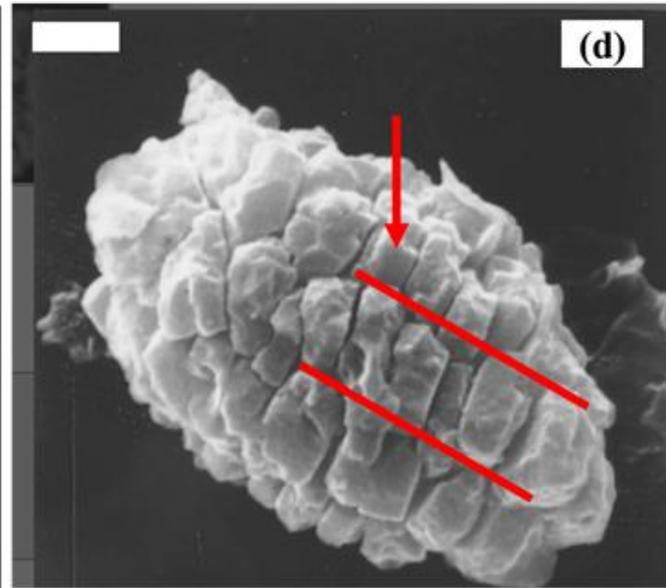
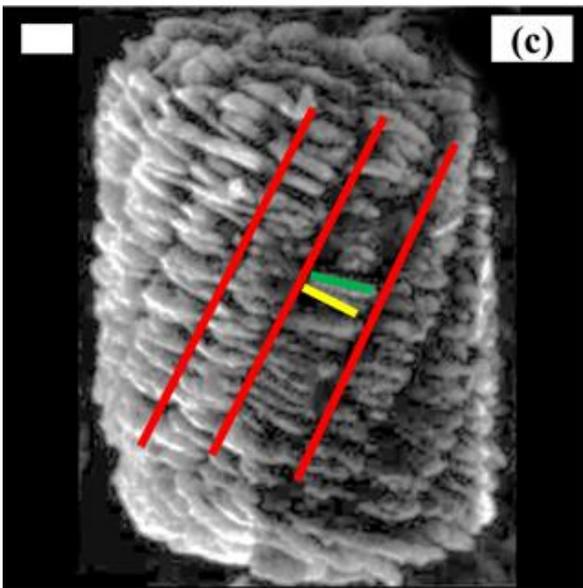
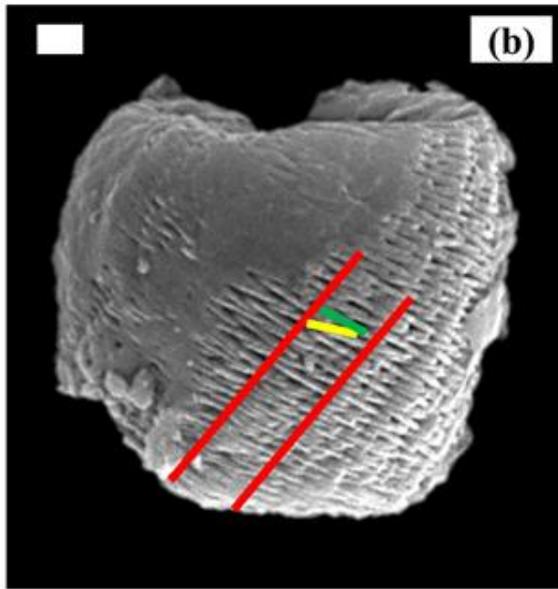
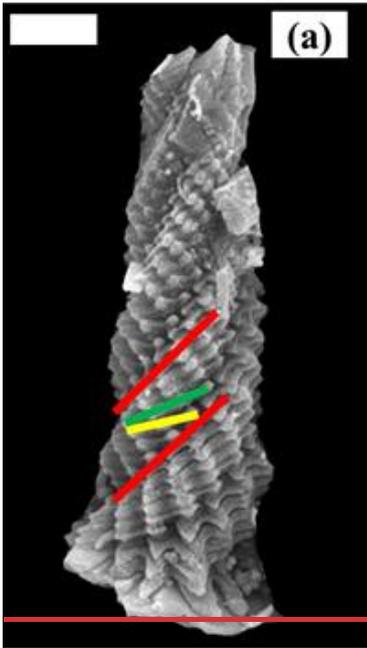
SEM image of the specimen. (c) The needle with a *Nannoconus* resting at its tip is placed in a metal holder that fits in the experimental station of the beamline of the synchrotron radiation centre.

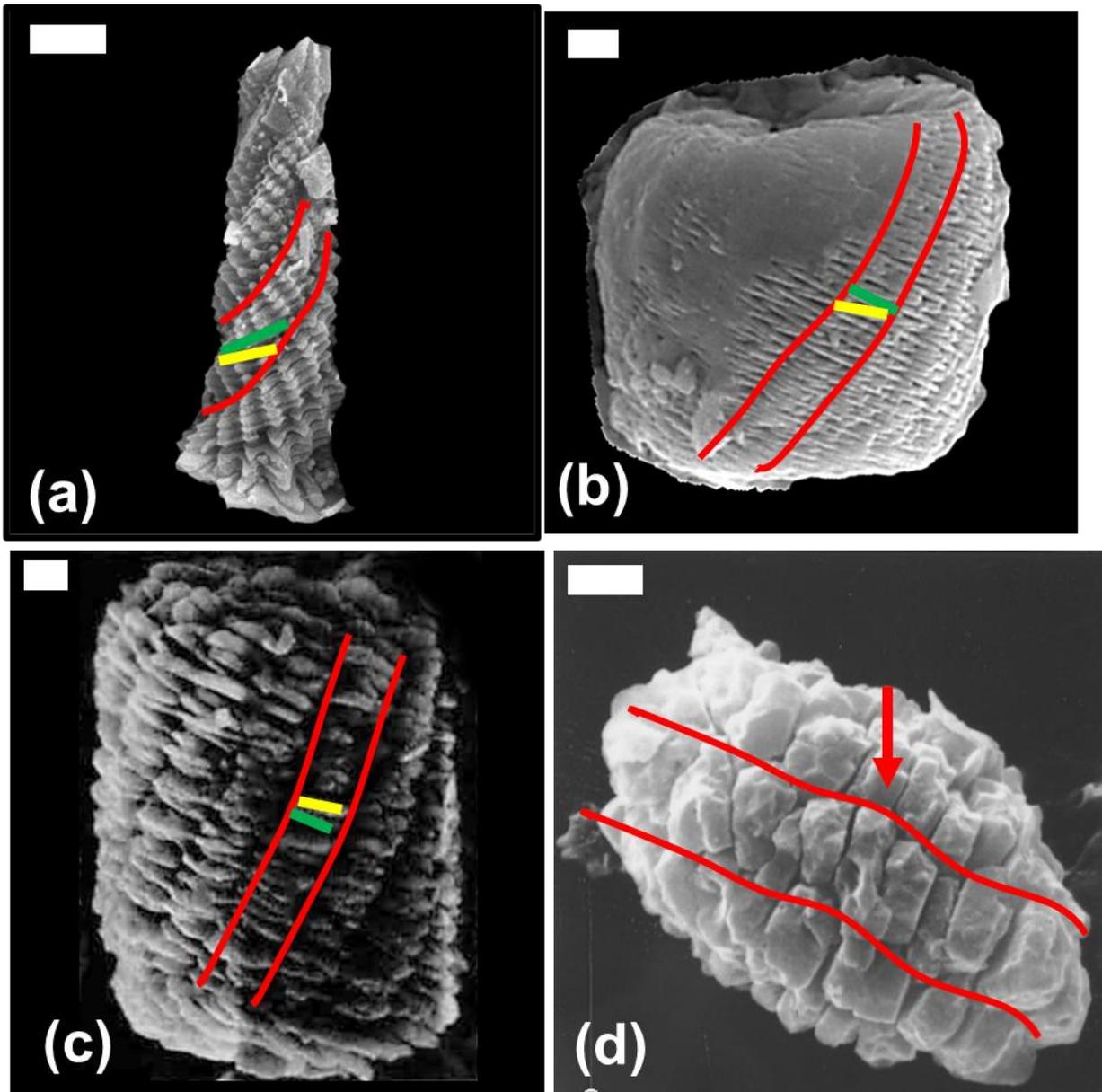


620 **Figure B4:** Graphical representation showing the variation of parameters along the length (L) of the first reconstructed layer (layer-A) composed of lamella-A, of *N. globulus*. The black points in the graphs, mark individual lamellae.



**Figure B5:** Graphical representation showing the variation of parameters along the length ( $L$ ) of the first reconstructed segments composed of lamella-A and -B of *N. globulus*. The black points in the graphs, mark individual lamellae.

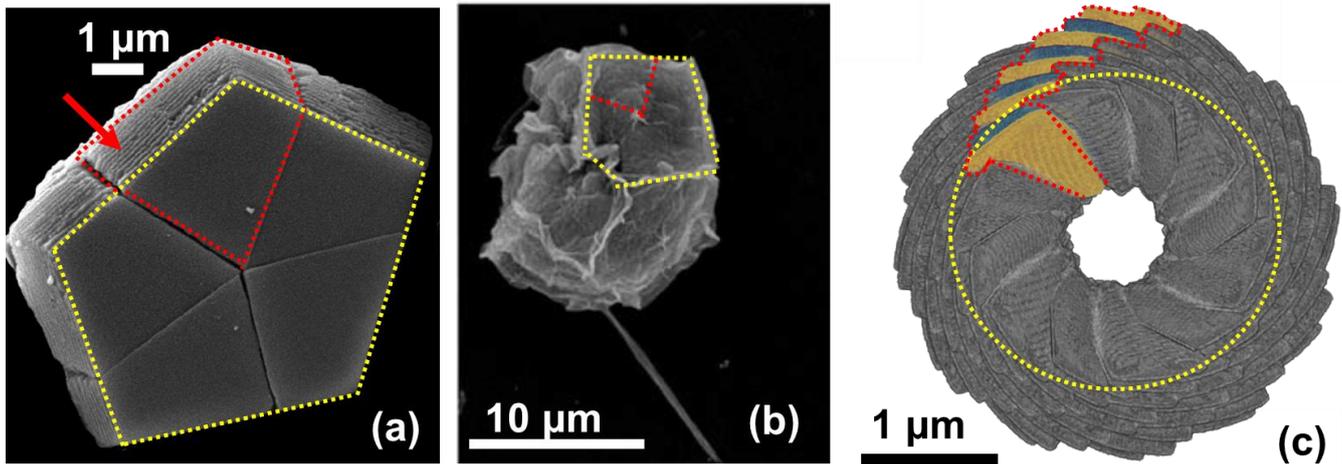




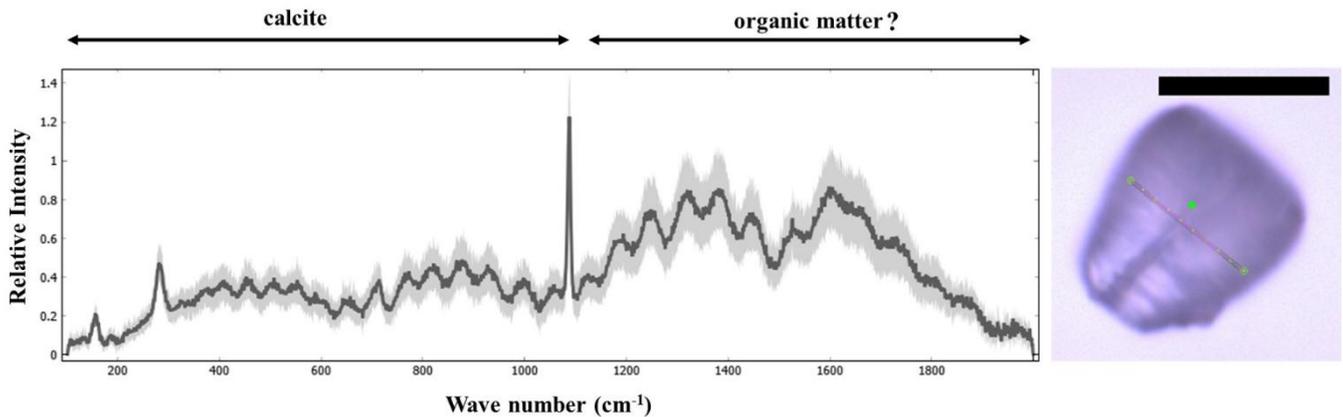
625  
 Figure B6: SEM images of different species of *Nannoconus*, with the segments, marked in red lines along with lamella-A (yellow line) and -B (green line). (a) *N. funiculus*; adapted from Lees and Bown (2016). (b) *N. globulus*, from DSDP Leg-93-Site 603 (continental rise of the western margin of the North Atlantic;  $\epsilon$ Core 44, interval 115-116 cm) presented in Fig. B2d in this study. (c) *N. truttii*; adapted from Hattner and Wise (1980), as available in the Nannotax3 website (Young et al., 2022). (d) *N. compressus*; adapted from Bown and Cooper (1998). The red arrows indicate lamellae that have fused and thickened due to overgrowth. The two types of lamellae (i.e., -A and -B) cannot be distinguished separately. This specimen, however, has preserved the organization of the segments. The white bar represents 1  $\mu$ m.

625

630



635 Figure B7: Comparison between the *B-a pentagonal pentalith of B. bigelowii* and the reconstructed *micolith of N. globulus*. (a) SEM  
 640 image of the *pentagonal skeleton of B. bigelowii pentalith*, adapted from Bown, 2010. The yellow dotted lines mark the boundaries of the  
 645 *pentagonal skeleton pentalith* and the red dotted lines represent a segment. The red arrow indicates the parallel stacking of the  
 lamellae, forming a segment. (b) The organic substrate, acts as template for the calcification of *B. bigelowii*. The yellow dotted lines  
 650 represent the boundary of the substrate that resembles the *pentagonal skeleton pentalith*. The substrate is divided into five parts,  
 with one part marked by the red dotted lines, mimicking the shape of the segment. Image adapted from Hagino et al., 2016. (c) The  
 basal view of the reconstructed *skeleton micolith* of *N. globulus*. The yellow dotted circle represents the “probable” hypothetical  
 655 organic substrate, acting as template for the calcification of *Nannoconus the micolith*. The substrate is divided into 12 parts (as  
*Nannoconus micolith* is considered to have 12 segments), with the boundaries of one part marked by the red dotted lines.



645 Figure B8: Confocal Raman spectra of *Nannoconus* displaying two regions corresponding to the compositional  $\text{CaCO}_3$  and the  
 “organic matter” preserved within the *skeleton micolith*. The photomicrograph in optical microscopy (PPL) of the *Nannoconus N.*  
 650 *steinmannii* is given on the right, with a red line showing the scanning path for the spectra. The black scale bar represents 10  $\mu\text{m}$ .  
 PPL: plane-polarized light. The Raman spectra were obtained using a LabRAM Soleil Horiba Raman micro spectrometer with 532  
 655 nm wavelength laser-excitation and 1  $\mu\text{m}$  spot size. A laser power of 1.1 mW was used to confirm that there is no damage to  
 the specimens induced by the laser. The laser beam was focused on the sample through a hole of 50  $\mu\text{m}$  using an objective at 100X  
 magnification. The specimen was subjected to linear scans along the surface (5-10  $\mu\text{m}$ ) and also inside the specimen (4-5  $\mu\text{m}$ ). A total  
 25 scans were recorded, each with an acquisition time of 2 seconds. Peaks were fitted in the spectrum using Gaussian functions in  
 Quasar software (Toplak et al., 2017) as a part of the analyses. The spectra have been normalized to the maximum intensity. The  
 black spectrum represents the average of all 25 recorded spectra of the *Nannoconus*.

**Table C1: Glossary for different terms used in this publication.**

<u>Term</u>	<u>Description</u>
<u>Taxonomy</u>	<u>A formal hierarchical biological classification scheme of any organism, also known as Linnaean Taxonomy. This is given by Life&lt;Domain&lt;Kingdom&lt;Phylum&lt;Class&lt;Order&lt;Family&lt;Genus&lt;Species</u>
<u>Order</u>	<u>The 6<sup>th</sup> rank of the taxonomic classification.</u>
<u>Genus</u>	<u>The 8<sup>th</sup> rank in the taxonomic classification, the plural of genus is genera.</u>
<u>Species</u>	<u>The 9<sup>th</sup> and the final rank of the taxonomic classification.</u>
<u>Morphogroups</u>	<u>An informal term to classify organisms based on similar morphological criteria.</u>
<u>Convex Hull</u>	<u>A mathematical representation of an envelope/elastic cover covering a large set of data, defining a shape in space.</u>

**Code availability**

660 The code for the ~~skeletal~~micalith reconstruction of *Nannoconus globulus* for both the layer and segment models is available in the repository: <https://doi.org/10.5281/zenodo.14925063> (Chowdhury et al., 2025). The segmentation, reconstruction, and visualization of the results have been done in Dragonfly software version 2024.2 for [windows]. Comet Technologies Canada Inc., Montreal, Canada; software available at <https://www.theobjects.com/dragonfly>.

**Sample availability**

665 Rock samples and associated nannofossil samples used for the experiment are curated at the Collections de Géologie de l'Observatoire des Sciences de l'Univers de Grenoble (OSUG), with an appropriate UJF-ID number. OSUG-COLLECTIONS is a database of rocks, minerals, and fossils, <https://web.collections.osug.fr>, OSUG, UGA. doi:10.17178/OSUG-COLLECTIONS.all.

**Author contributions**

670 **RC** prepared the sample with inputs from **BSM**, **FG**, **AK**, and **AFM**, applied the image segmentation, co-developed the reconstruction code with **AFM**, and wrote the original draft. **JCS**, **JLH**, and **AK** helped in the initial conceptualization of the experiment. **MPA** intensely contributed to the conceptualization of the segment model of the reconstruction. **RC**, **RB**, **BSM**, **MD**, **AK**, **JP**, **FG**, and **AFM** performed the experiments in the synchrotron radiation centre. **FG** and **AFM** jointly supervised **RC** in this work and acquired all the relevant funding. All the co-authors contributed significant inputs for the review and  
675 editing.

## Competing interests

The authors declare that they have no conflict of interest.

## Acknowledgments

680 We acknowledge SOLEIL for providing access to synchrotron radiation on the SWING beamline through proposals 20211643 and 20221681. We are ~~thankful~~indebted to ~~Jeremy Young, Angela Fraguas,~~ Isaline Demangel, ~~one~~Angela Fraguas, Jeremy Young, and an anonymous referee, ~~and the editor~~ for their ~~valuable remarks and constructive~~ comments ~~to improve on~~ this ~~publication work, and to the editor for improving its presentation.~~

## Financial support

This work was supported by the Tellus Program of CNRS-INSU, OSUG@2020, and IODP-France.

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