



Bacteriohopanepolyols track past environmental transitions in the Black Sea

Anna Cutmore^{1,2*}, Nora Richter^{1,3*}, Nicole Bale¹, Stefan Schouten^{1,4}, and Darci Rush¹

*Joint first authors

¹NIOZ Royal Netherlands Institute for Sea Research, Texel, the Netherlands

²Durham University, Department of Geography, Durham, United Kingdom

³EAWAG Swiss Federal Institute of Aquatic Science and Technology, Dübendorf, Switzerland

⁴Utrecht University, Utrecht, the Netherlands

Correspondence to: Anna Cutmore, anna.v.cutmore@durham.ac.uk and Nora Richter, nora.richter@eawag.ch

Abstract

Bacteriohopanepolyols (BHPs) are structurally diverse compounds produced by a wide range of bacteria making them ideal candidates as chemotaxonomic biomarkers and indicators of bacterially-driven biogeochemical processes in the geological record. In this study, we characterize changes in the BHP distribution in the Black Sea over the past 20 thousand years (ka), as the basin underwent three distinct environmental phases: (i) an oxic lacustrine phase where the Black Sea was disconnected from the global ocean; (ii) a transition period marked by the initial influx of marine water into the basin; and (iii) a marine phase where the basin was permanently euxinic. During the lacustrine phase we observe a high abundance and diversity of nucleoside BHPs (Nu-BHPs) that are likely derived from elevated terrigenous inputs as well as production of Nu-BHPs in the brackish-to-fresh water column. The transition phase is marked by a decrease in the abundance of most Nu-BHPs and an increase in the abundance of methoxylated-BHPs as well as BHPs such as aminobacteriohopanetriol which are ubiquitous across a wide range of environments including soils as well as marine and freshwater settings. The euxinic marine phase (7.2 ka-present) can be divided into two stages based on changes in BHP composition. The early stage is characterised by a high abundance of aminobacteriohopanetetrol and aminobacteriohopanepentol, which were likely produced by methanotrophs at the oxycline. A shallow oxycline likely allowed for increased transport of these BHPs to the sediment. The later marine phase is characterised by a decline in these BHPs, likely due to a deepening of the oxycline and reduced transport of BHPs from the oxycline to the sediment. The changes in BHP distributions throughout the record may either be attributed to shifts in the bacterial communities or physiological adaptations of bacteria to the changing environment. Throughout the record, diagenetic products of BHPs (e.g., anhydrous-bacteriohopanetetrol) were detected. These degradation products, however, remain a small proportion of the overall BHP composition, indicating excellent preservation conditions throughout the record. This study offers new insights into changes in microbial communities and biogeochemical processes that occurred in the Black Sea during the Last Deglaciation and Holocene in response to substantial shifts in the hydrology and oxygen conditions of the basin.

Keywords: Bacteriohopanepolyols, Black Sea, Holocene, Last Deglaciation, Anoxia, Euxinia



38 1. Introduction

39 Bacteriohopanepolyols (BHPs) are pentacyclic triterpenoids produced by a wide-range of bacteria that are
40 considered to be the precursors to one of the most abundant lipids in the geological record, hopanoids (Rohmer et
41 al., 1984; Ourisson and Albrecht, 1992). BHPs themselves are also abundant in the geological record and are well-
42 preserved in sedimentary archives for >1 million years (Handley et al., 2010). In some cases, BHPs are linked to
43 certain source organisms (Cvejic et al., 2000; Kool et al., 2014; Rush et al., 2014; van Winden et al., 2012) or
44 biogeochemical processes (Coolen et al., 2008; Blumenberg et al., 2013). Consequently, shifts in BHP compositions
45 in the geological record have become a useful tool for palaeoclimate reconstructions to assess changes in redox
46 conditions (Blumenberg et al., 2013; Matys et al., 2017; Rush et al., 2019; Zindorf et al., 2020), salinity (Coolen et al.,
47 2008), methanotrophy (Talbot et al., 2014), and soil inputs (Blumenberg et al., 2013). For example, 35-
48 aminobacteriohopane-31,32,33,34-tetrol (aminotetrol from herein), 35-aminobacteriohopane-30,31,32,33,34-
49 pentol (aminopentol from herein), and methylcarbamate-amino BHPs are primarily associated with aerobic
50 methane-oxidizing bacteria (MOB; Rohmer et al., 1984; Jahnke et al., 1999; Cvejic et al., 2000; Talbot et al., 2001;
51 Zhu et al., 2011; van Winden et al., 2012; Rush et al., 2016) and have been used to reconstruct past changes in
52 methane-oxidation in the Congo River Delta (Talbot et al., 2014; Spencer-Jones et al., 2017), the Baltic Sea
53 (Blumenberg et al., 2013), and in an Antarctic lake (Coolen et al., 2008). Recent studies show that bacteriohopane-
54 32,33,34,35-tetrol-x (BHT-x) is synthesized by anaerobic ammonium oxidation (anammox) bacteria (Rush et al.,
55 2014; Schwartz-Narbonne et al., 2020) and thus has been used to reconstruct changes in N-cycling under low-oxygen
56 conditions in Mediterranean sapropels (Rush et al., 2019; van Kemenade et al., 2023) and the Black Sea (Cutmore et
57 al., 2025). Nucleoside BHPs (Nu-BHPs) occur in high abundance in soils (Seemann et al., 1999; Bravo et al., 2001;
58 Cooke et al. 2008a; Xu et al., 2009; Rethemeyer et al., 2010), and, therefore, have been applied to track the transport
59 of soil organic matter to aquatic sediments (Talbot and Farrimond, 2007; Cooke et al., 2008b). Recent studies,
60 however, have identified potential production of Nu-BHPs along redoxclines in marine oxygen minimum zones
61 (Kusch et al., 2021) and near the oxycline in lakes (Richter et al. 2023). While this highlights a small number of the
62 known BHPs and their potential application to paleo-records, there are still many unknowns surrounding the origin
63 and sources of other BHPs in the environment. Recent analytical advancements in BHP analysis has led to the
64 identification of additional novel BHPs in both cultures and environmental samples (Talbot et al., 2016a; Hopmans
65 et al., 2021; Richter et al., 2023). The large structural diversity of BHPs along with their preservation potential in the
66 geological record, points to a useful new tool in biomarker research that can provide insights into past changes in
67 microbial communities and/or biogeochemical processes.

68 The Black Sea provides a compelling location to study the impact of bacterial community composition and
69 environmental changes on BHP composition due to its unique hydrological history and present-day characteristics.
70 Today, it is the largest permanently stratified anoxic basin in the world with limited connection to the global ocean
71 through the Bosphorus Strait. Over the past 20 thousand years (ka), however, the Black Sea underwent significant
72 hydrological changes (Fig. 1). During the Last Glacial Maximum (LGM), the basin was an oxygenated brackish-to-



freshwater environment (Schrader, 1979), then, during the subsequent deglaciation, the basin experienced many environmental shifts, including changes in temperature (Bahr et al., 2005; 2008; Ion et al., 2022), water-level (Ivanova et al., 2007; Nicholas et al., 2011; Piper and Calvert, 2011), and freshwater input. This freshwater influx was influenced by changes in regional precipitation and melting of Eurasian ice sheets and alpine glaciers (Bahr et al., 2005; 2006; 2008; Badertscher et al., 2011; Shumilovskikh et al., 2012). The basin reconnected with the global ocean around 9.6 ka when post-glacial sea-level rise led to an initial marine inflow (IMI) over the Bosphorus sill (Aksu et al., 2002; Major et al., 2006; Bahr et al., 2008; Ankindinova et al., 2019), leading to enhanced salinity of the water column (Marret et al., 2009; Verleye et al., 2009; Filipova-Marinova et al., 2013) and permanent euxinia developing in the water column after ~7.2 ka (Arthur and Dean, 1998; Eckert et al., 2013).

Several studies have demonstrated the usefulness of lipid biomarkers as records of biogeochemical processes and as indicators of ecological niches within the modern-day water column and surface sediments of the Black Sea (Schubert et al., 2006; Coolen et al., 2007; Wakeham et al., 2007; Schubotz et al., 2009; Sollai et al., 2019; Bale et al., 2021; Kusch et al., 2022). BHPs produced in the lower suboxic zone and upper sulfidic zone, for instance, were shown to record activities of aerobic methanotrophs and anammox bacteria, respectively, demonstrating their reliability as potential biomarkers for microbial communities (Kusch et al., 2022). Furthermore, previous work has explored BHP changes in the Black Sea over the Holocene, however, this applies to a limited number of compounds (Blumenberg et al., 2009a). With significant advances in analytical techniques, there is now the opportunity to explore the BHP geolipidome of the Black Sea in unprecedented detail. Indeed, we recently used the ratio of bacteriohopanetetrol (BHT)-34S and the later eluting stereoisomer BHT-x to trace past anammox activity in the Black Sea over the last 20 ka, complementing other nitrogen cycling biomarker tools (Cutmore 2025). In this study, we now fully characterize the distribution of BHPs in a Black Sea sediment core over the Last Deglaciation and Holocene to shed light on changing environmental conditions and bacterial communities.

2. Methods

Piston core 64PE418 (235 cm length) was recovered from the Black Sea (42°56 N, 30°02 E; 1970 mbsl depth) during the 2017 Pelagia cruise (Fig. 1). The core was subsampled at the Royal Netherlands Institute for Sea Research, and all samples were stored frozen until further analysis. The contemporary Black Sea water column is characterized by pronounced redox gradients. At the location of the core site in the western gyre of the basin, the oxic zone, found between 0 – 75 m depth range, has an oxygen concentration of ~121 µmol/kg at 50 m depth, while sulphide concentrations are not detected (Sollai et al., 2019). The suboxic zone lies below, situated between 75 – 115 m depth range, with traces of sulphide found at the bottom of this layer between 105 – 115 m (Sollai et al., 2019). Beneath this is the euxinic zone, located between 115 – 2000 m depth range, with sulphide concentrations increasing significantly with depth, reaching ~400 µmol/L at 2,000 m water depth (Sollai et al., 2019).

2.1 Lipid extraction and analysis



108 Sediment samples (n=44) were taken at 5 cm intervals throughout the core's depth before freeze drying.
109 Using a modified Bligh and Dyer extraction method (as described previously by Bale et al., 2021), lipids were
110 extracted from the dry sediment samples, as described in detail in Cutmore et al. (2025). Extraction blanks were
111 carried out alongside the sediment extractions, using the same glassware, solvents and extraction methodology.

112 Extract analysis was performed on an ultra-high performance liquid chromatography-high resolution mass
113 spectrometer (UHPLC-HRMS) in reverse phase as described in Hopmans et al. (2021). An Agilent 1290 Infinity I
114 UHPLC, featuring a thermostatted auto-injector and column oven, was utilised, coupled to a Q Exactive Orbitrap MS
115 with Ion Max source with heated electrospray ionization (HESI) probe (Thermo Fisher Scientific, Waltham, MA).
116 Separation was carried out with an Acquity BEH C18 column (Waters, 2.1 × 150 mm, 1.7 µm) maintained at 30°C,
117 using an eluent composition of (1) MeOH/H₂O/formic acid/14.8 M NH₃aq [85:15:0.12:0.04 (v:v)] and (2)
118 IPA/MeOH/formic acid/14.8 M NH₃aq [50:50:0.12:0.04 (v:v)]. The flow rate was 0.2 mL min⁻¹ with an elution program
119 of: 95% A for 3 minutes, followed by a 40% linear gradient at 12 minutes, then 0% A at 50 minutes which was
120 maintained until 80 minutes. The positive ion HESI parameters were as follows: capillary temperature at 300°C;
121 sheath gas (N₂) pressure at 40 arbitrary units (AU); spray voltage at 4.5 kV; auxiliary gas (N₂) pressure at 10 AU; S-
122 lens at 70 V; probe heater temperature at 50°C. Lipid analysis was conducted across a mass range of m/z 350–2000
123 (resolving power 70,000 ppm at m/z 200), followed by data-dependent tandem MS/MS (resolving power 17,500
124 ppm), in which the 10 most abundant masses in the mass spectrum were fragmented successively. Optimal
125 fragmentation was achieved with a stepped normalized collision energy of 22.5 and 40 (isolation width 1.0 m/z) for
126 BHP analysis. The Q Exactive was calibrated to a mass accuracy of ±1 ppm using the Thermo Scientific Pierce LTQ
127 Velos ESI Positive Ion Calibration Solution. During analysis, dynamic exclusion was applied to temporarily exclude
128 masses for 6 s, enabling selection of less abundant ions for MS/MS.

129 BHPs were tentatively identified based on their retention time, exact mass, and fragmentation spectra.
130 Integrations were performed on (summed) mass chromatograms of relevant molecular ions ([M+H]⁺, [M+NH₄]⁺, and
131 [M+Na]⁺). The BHP absolute abundances are all presented as peak area per gram of total organic carbon (TOC) since
132 we lack appropriate standards to correct for differences in ionization efficiency between BHPs (Cutmore et al., 2025).

133

134 2.2. Age Model

135 The age model for core 64PE418 was previously published in Cutmore et al. (2025). Seven bulk organic
136 matter ¹⁴C dates were used in its production. Six of these were from core 64PE418, combined with an additional bulk
137 organic carbon ¹⁴C date from core KNR 134-08 BC17 (Jones and Gagnon, 1994), which is sourced from the same
138 location and water depth as 64PE418. This additional ¹⁴C date is from the widely observed Unit I/II boundary and
139 was employed to refine the age model for the upper section of the core. Variable reservoir-ages, calculated by
140 Kwiecien et al. (2008) for intermediate water depths in the Black Sea over the last ~20 ka, were added to the
141 calibration. The ¹⁴C dates were calibrated using the Marine20 calibration curve (Heaton et al., 2020) for the upper
142 three ¹⁴C dates (reflecting the period after the IMI), while the lower four ¹⁴C dates were calibrated using the IntCal20



143 calibration curve (Reimer et al., 2020), (reflecting the period when then Black Sea was a lacustrine environment).
144 The R-code, CLAM (Blaauw, 2010), was used to create the age–depth model. The BHP record from core 64PE418
145 spans the last 19.5 ka, with an average resolution of ~450 years. The key transitions (identified by colour and
146 elemental changes in the core [Cutmore et al., 2025]) are as follows: the onset of the IMI is dated at 9.6 ka ± 237 yrs;
147 Unit II/III occurs at 7.2 ka ± 202 yrs; and the Unit I/II boundary is dated at 2.6 ka ± 402 yrs.

148

149 2.3. Statistical analyses

150 A principal component analysis (PCA) was used to assess the variability in BHP distributions downcore after
151 applying a Hellinger transformation and scaling the BHP dataset. All analyses were performed and visualized in R
152 (version 4.2.2; R Core Team, 2023) with the vegan package (version 2.6-4; Oksanen et al., 2020), factextra (version
153 1.0.7; Kassambara and Mundt, 2020), FactoMineR (version 2.7; Lê et al., 2008), and ggplot2 (version 3.4.0; Wickham
154 and Chang, 2016).

155

156 4. Results and Discussion

157 Significant changes in total organic carbon (TOC) content, colour and elemental signatures of the core are
158 observed (Cutmore et al., 2025), corresponding to changing environmental conditions in the Black Sea basin over
159 the last 20 ka (Arthur and Dean, 1998; Bahr et al., 2005; Jones and Gagnon, 1994; Ankindinova et al., 2019). Based
160 on this, the core is divided into three widely acknowledged key periods: the oxic lacustrine phase (19.5-9.6 ka) where
161 the basin was disconnected from the global ocean; the transition phase (9.6 - 7.2 ka) after the IMI over the Bosphorus
162 sill at ~9.6 ka, whereby the basin moved towards a marine environment; and the marine phase (7.2 ka to the present)
163 where the Black Sea became a euxinic brackish-to-marine environment (Jones and Gagnon, 1994; Arthur and Dean,
164 1998; Aksu et al., 2002; Major et al., 2006; Bahr et al., 2008; Ankindinova et al., 2019; Cutmore et al., 2025).

165

166 4.1 BHP distribution changes over time

167 In total, 63 BHPs were detected throughout the Black Sea record. A PCA of the relative distribution of BHPs
168 reveals four distinct clusters (Fig. 2), which correspond with the above defined phases of the Black Sea: an oxic
169 lacustrine phase (19.5 - 9.6 ka), a transition phase (9.6 - 7.2 ka), and a marine phase that is further divided into an
170 early marine phase (7.2 - 5 ka) and a late marine phase (5 ka to present). PC1 explains 40.8% of the variance and is
171 positively correlated with Nu-BHPs, while PC2 explains 18.2% of the variance and is negatively correlated with BHPs
172 such as 35-aminobacteriohopane-32,33,34-triol (aminotriol from herein), N-formylated-aminotriol, and
173 ethenolamine-bacteriohopane-32,33,34,35-tetrol (ethenolamine-BHT from herein). BHT-22S, aminotriol and
174 ethenolamine-BHT are present throughout the record (Fig. 3), which is in line with the ubiquitous nature of these
175 BHPs that are found in contemporary marine, freshwater and terrestrial environments (Talbot and Farrimond 2007;
176 Hopmans et al., 2021; Richter et al., 2023).



Each phase of the Black Sea is defined by several notable changes in BHP distributions. Throughout the oxic lacustrine phase, during the Last Deglaciation and early Holocene (19.5 – 9.6 ka), a wider range of BHPs were detected compared to other phases (Fig. 3). This coincides with low TOC levels and low concentrations of elements that accumulate in sediments under anoxic conditions (i.e., uranium [U], vanadium [V], and molybdenum [Mo]) (Cutmore et al., 2025), pointing to a well-oxygenated environment. Nu-BHPs, for instance, are present in higher absolute abundance and demonstrate a more diverse distribution during the lacustrine phase relative to the other periods (Fig. 2). Bacteriohopane-32,33,34-triol (BHtriol), bacteriohopane-31,32,33,34,35-pentol (BHpentol), and bacteriohopane-30,31,32,33,34,35-hexol (BHhexol) are also present in higher absolute abundance during the oxic lacustrine phase, relative to the rest of the record where they are largely absent. The transition phase (9.6 - 7.2 ka) occurs after the IMI at ~9.6 ka and is characterized by a decrease in almost all Nu-BHPs with the exception of an increase in aminotriol, ethenolamine-BHT, and N-formylated-aminotriol.

At 7.2 ka, there is a shift to a permanent euxinic marine system which led to enhanced preservation of organic matter resulting from permanent water column anoxia. Despite the enhanced preservation, previous studies have reported a decrease in both homohopane and BHP concentrations in the Black Sea during this period (Blumenberg et al., 2009a) indicating either a decline in BHP production or a decrease in BHP transport to the sediment, as observed in the modern-day Black Sea water column (Wakeham et al., 2007; Blumenberg et al., 2009a; Kusch et al., 2022). In our record, the early marine phase (7.2 – 5 ka) is marked by an increase in methoxylated-BHT (methoxy-BHT from herein), methoxylated-ethenolamine-BHT (methoxy-ethenolamine-BHT from herein), and propenolamine-BHT. There is a temporary increase in aminotetrol, aminopentol, N-acylated-aminotriols, ethenolamine-BHpentol, and ethenolamine-BHhexol that spans the transition of the early and late marine phases. The absolute abundances of BHPs are considerably lower after ~3.9 ka, with the exception of methoxy-BHT and methoxy-ethenolamine-BHT, which increase from 0.9 ka to present. A decrease in BHP and homohopane concentrations during the late marine phase has also been reported in a previous reconstruction of the Black Sea (Blumenberg et al., 2009a).

201

202 4.2. Diagenetic products of BHPs

BHP preservation was assessed throughout the core by tracking known diagenetic products of BHP degradation: 32,35-anhydrobacteriohopanetetrol (anhydro-BHT) and anhydrobacteriohopanepentol (anhydro-BHpentol) which are formed from the dehydration and cyclization of BHT and BHpentol, respectively (Bednarczyk et al., 2005; Talbot et al., 2005; Schaeffer et al., 2008; 2010). In addition, it has been shown that anhydro-BHT is also a degradation product of composite BHPs, such as BHT-cyclitol ether and adenosylhopane (Schaeffer et al., 2010; Eickhoff et al., 2014). Throughout the record, the absolute abundance of anhydro-BHT and -BHpentol follow opposite trends to each other, with a higher absolute abundance of anhydro-BHpentol and anhydro-BHT during the lacustrine oxic phase and marine phase, respectively (Fig. S4). In a previous study of Black Sea surface sediments, anhydro-BHT decreased below the top 2 cm of the core, suggesting rapid early diagenesis (Kusch et al., 2022). Previous studies



point to the possibility of selective degradation of BHPs, in particular the degradation of Nu-BHPs relative to co-occurring composite BHPs, such as aminotriol (Talbot et al., 2016b; Kusch et al., 2022). However, Nu-BHPs occur in high absolute abundance throughout the record, particularly during the lacustrine oxic phase where we would anticipate increased degradation due to an oxygenated water column relative to the more recent marine phase whereby the water column became anoxic (Schrader, 1979; Arthur and Dean, 1998; Eckert et al., 2013).

BHtriol and unsaturated BHtriol follow a similar trend to anhydro-BHpentol, with all three BHPs present in higher absolute abundance during the lacustrine phase relative to the late transition period and the marine phase, suggesting that BHtriol and unsaturated BHtriol could also be diagenetic products (Fig. S4). Indeed, BHtriol only contains 7 carbons on the side-chain instead of the typical 8 carbons found in other BHPs and in unsaturated BHtriol the unsaturation occurs on the side-chain, indicating that they may have formed from decarboxylation and dehydration of functionalized BHPs. The decline in BHtriol and anhydro-BHpentol throughout the transition period (between 9.6 - 7.2 ka) may indicate reduced degradation due to intermittent anoxia in the water column. It may, however, also indicate a decrease in the abundance of composite BHPs that are precursors to these diagenetic BHPs, with both BHhexol and BHpentol also declining in absolute abundance during the transition period compared to the lacustrine phase. Anhydro-BHT, in contrast, does not decline during the transition period, instead increasing during the transition and marine periods. This could suggest that there is an increase in the source of the diagenetic precursors for anhydro-BHT (i.e., BHT and adenosylhopane; Eickhoff et al., 2014), while the precursors to the other diagenetic BHPs decreased. Overall, there are BHP diagenetic products present throughout the record, and the changes in the absolute abundance of these BHPs are likely associated with the changes in precursor BHP concentrations.

4.3. Sources of Nu-BHPs

Nu-BHPs were particularly abundant and diverse during the lacustrine phase (Fig. 4), with the highest Nu-BHP abundance occurring between 19.5 - 13.8 ka, during the last deglaciation, dominated by adenosylhopane_{HG-diMe} followed by adenosylhopane and adenosylhopane_{HG-Me}. Many of the Nu-BHPs are present in higher abundance early in the record and decline during the latter part of the lacustrine phase. This pattern corresponds with changes in terrigenous input, as indicated by the elemental records of titanium to calcium (Ti/Ca) and of potassium (K), as well as the BIT index based on branched glycerol dialkyl glycerol tetraether (br-GDGT) and crenarchaeol (Hopmans et al., 2004), in this core (pers. comms. B.Yang;). This indicates a high contribution of terrigenous material early in the record and a decline thereafter. Between 18 and 14.8 ka, there were multiple significant meltwater pulses in the Black Sea (Major et al., 2006; Bahr et al., 2008; Badertscher et al., 2011; Yanchilina et al., 2019) followed by enhanced precipitation during the Bølling Allrød (BA) (14.5-13 ka [Shumilovskikh et al., 2012]). Consequently, the increased terrestrial input is likely the result of this enhanced freshwater influx. Many of the Nu-BHPs may therefore be sourced from this enhanced contribution of terrestrial organic matter, with Nu-BHPs primarily associated with soils (Talbot and Farrimond, 2007; Cooke et al., 2008a; Xu et al., 2009; Rethemeyer et al., 2010).



247 The absolute abundance of Nu-BHPs declines after 13 ka, corresponding with low Ti/Ca and K values
248 (Cutmore et al., 2025), which indicate reduced input of terrestrial organic matter. This corresponds with drier
249 regional conditions during the Younger Dryas (YD) and Early Holocene (Shumilovskikh et al., 2012) and reduced influx
250 of meltwater into the Black Sea due to the retreat of ice sheets across Northern Europe (Major et al., 2006). During
251 this time, the composition of the Nu-BHPs also changes with a decline in the absolute abundance of
252 adenosylhopane_{HG-diMe}. After 13 ka, 2Me-adenosylhopane_{HG-diMe} increases, peaking at 10.6 ka and declining abruptly
253 at the IMI at 9.6 ka. While the majority of Nu-BHPs identified in this record are likely linked to soil input and closely
254 follow the terrigenous elements, some, such as 2Me-adenosylhopane_{HG-diMe}, demonstrate a different pattern
255 throughout the core and are likely being produced within the lacustrine water column (Fig. 5). 2Me-
256 adenosylhopane_{HG-diMe}, for example, peaks at the start of the Holocene (11-9 ka), and may therefore be linked to
257 higher temperatures or other factors associated with warming in the Black Sea. Its abundance declines abruptly at
258 the IMI, indicating that it is not produced under anoxic conditions.

259 During the transition period, most Nu-BHPs are still detected in the record (e.g., adenosylhopane_{HG-Me} and
260 2Me-adenosylhopane_{HG-diMe}), but in much lower absolute abundance than during the oxic lacustrine period, likely
261 due to reduced input of terrestrial organic matter, corresponding with low Ti/Ca and K values. Me-
262 adenosylhopane_{HG-diMe}, however, demonstrates a higher abundance during the transition period compared to the
263 end of the oxic lacustrine period (Fig. S8). As this compound has previously been found near the chemocline in lakes
264 (Richter et al., 2023), it is possible that it was being produced near the oxic-anoxic transition zone during periods of
265 water column anoxia.

266 The diversity of Nu-BHPs is low throughout the marine phase with many Nu-BHPs below detection limit
267 throughout this period (e.g., 2Me-Adenosylhopane, 3Me-Adenosylhopane, diMe-adenosylhopane_{HG-Me}, 3Me-
268 Adenosylhopane_{HG-diMe}, 2Me-N1-methyl-inosylhopane and Me-N1-methyl-inosylhopane). The Nu-BHPs that are
269 detected are present in low absolute abundance compared to other phases. This is likely due to the low contribution
270 of terrestrial organic matter to the site, demonstrated by the low Ti/Ca values, which reduced the delivery of soil-
271 sourced Nu-BHPs to this site. Notably, there is an increase in several Nu-BHPs after 1 ka, including adenosylhopane,
272 2Me-adenosylhopane_{HG-Me}, Me-adenosylhopane_{HG-diMe}, inosylhopane, N1-methylinosylhopane and Me-N1-methyl-
273 inosylhopane. If these BHPs are not derived from soil inputs, then they are likely being produced in the water column
274 of the Black Sea. Indeed, adenosylhopane, 2Me-adenosylhopane_{HG-Me}, and inosylhopane are detected in the surface
275 waters of the modern-day Black Sea water column (Kusch et al., 2022).

276 In summary, most Nu-BHPs detected in the Black Sea core are likely sourced from soils, linked to freshwater
277 influx into the basin. Several Nu-BHPs, such as adenosylhopane, 2Me-adenosylhopane_{HG-Me}, Me-adenosylhopane<sub>HG-
278 diMe</sub>, 2Me-adenosylhopane_{HG-diMe}, inosylhopane, N1-methylinosylhopane and Me-N1-methyl-inosylhopane,
279 however, exhibit patterns that suggest these Nu-BHPs were produced in the water column of the Black Sea. As future
280 studies discover more about the sources of Nu-BHPs both in soils and in the water column of lakes and marine
281 environments, it may be possible to expand the interpretation of these records.



282

283 4.4. BHPs associated with the N-cycle

284 BHT-x, produced by anammox bacteria, has already successfully been applied to this record in a previous
285 study (Cutmore et al., 2025) to detect anammox activity, which is occurring in the present day Black Sea (Kuypers et
286 al., 2003; Dalsgaard et al., 2012). At 7.2 ka, there is an increase in concentration suggesting expansion of anammox
287 activity, due to the permanently anoxic water column which enabled anammox bacteria to expand their habitat from
288 the anoxic sediments; consequently, during the marine euxinic period, loss of bioavailable nitrogen by anammox
289 activity was enhanced (Cutmore et al., 2025). In contrast, oxazinone-aminotriol was detected throughout the
290 lacustrine and transition phases, and absent during the marine euxinic period. Oxazinone-aminotriol was first
291 identified in cultures of nitrite-oxidizing bacteria and later found in the surface sediments of lakes (Elling et al., 2022;
292 Richter et al., 2023). Its presence in the Black Sea record may therefore indicate that nitrite-oxidation was an
293 important process in the water column when the upper water column was oxygenated during the lacustrine and
294 transition phase, and less so when the water column was euxinic, during the later marine phase.

295 Other BHPs might also be associated with the nitrogen-cycle. For instance, there appears to be a close
296 coupling between the absolute abundance of BHT-CE and crenarchaeol throughout this record (Fig. 5; Cutmore et
297 al., 2025). As crenarchaeol is exclusively produced by the abundant and widespread archaea Thaumarchaeota
298 (Nitrososphaerota) (Sinninghe Damsté et al., 2002), the close coupling of these records indicates an association
299 between the dominant bacterial producer of BHT-CE in the Black Sea over this period and Nitrososphaerota. As
300 Nitrososphaerota plays a crucial role in nitrification in the Black Sea (Lam et al., 2007) by aerobically oxidizing
301 ammonia to nitrite (Könneke et al., 2005; Wuchter et al., 2006), it may be that the dominant bacterial producer of
302 BHT-CE was coupled to this nitrification process. Previous studies have indeed shown associations between archaea
303 and bacteria, with a coupling between ca. *Methanoperedens nitroreducens* and ca. *Methyloirabilis oxyfera* (NC10
304 phylum) in order to perform methane oxidation (Smit et al., 2019), and with anammox bacteria utilising the nitrite
305 produced by both Nitrososphaerota and ammonia-oxidising bacteria (AOB) in the modern-day Black Sea (Kuypers et
306 al., 2003; Lam et al., 2007). For instance, BHT-CE was detected in an enrichment culture from a peat sample for the
307 NC10 bacteria; however, it is unclear whether BHT-CE was being produced by NC10 bacteria or other microbial
308 communities in the enrichment (Kool et al., 2014). Consequently, a coupling may have existed in the Black Sea water
309 column between Nitrososphaerota and unknown bacteria with regards to nitrification. Alternatively, both microbes
310 may have inhabited the same ecological niche and depth habitat, meaning their abundance was similarly affected
311 by changes in the water column conditions.

312 BHPs with a methylation in the second carbon position of the A ring (2Me-BHPs) are precursors for 2Me-
313 hopanes, which were initially proposed as biomarkers for N₂-fixing cyanobacteria (Summons et al., 1999; Kuypers et
314 al., 2004; Talbot et al., 2008). More recently, 2Me-BHPs were shown to also be produced by alpha (α)-proteobacteria
315 involved in the nitrogen cycle (Rashby et al., 2007; Welander et al., 2010; Naafs et al., 2022), such as nitrite-oxidizing
316 bacteria *Nitrobacter vulgaris* (Elling et al., 2020), as well as Actinobacteria and one strain of a terrestrial



317 Acidobacteria (Welander et al., 2010; Sinninghe Damsté et al., 2017; Naafs et al., 2022). In the Black Sea record,
318 2Me-BHT was largely absent during the lacustrine phase, until the early Holocene (10.3 ka) when 2Me-BHT increases
319 and peaks at the start of the transition phase (9.6 ka) followed by a smaller peak during the marine phase (6-4.2 ka)
320 (Fig. S4). As the increase in 2Me-BHT coincides with increasing absolute abundance of hexose HGs, which are known
321 indicators of increased N₂-fixation by freshwater or brackish cyanobacteria (Cutmore et al., 2025), it is possible that
322 2Me-BHT is related to N₂-fixing cyanobacteria in the Black Sea record. The 2Me-BHT record may also be related to
323 changes in the abundance of α -proteobacteria; however, the 2Me-BHT record does not align with oxazinone-
324 aminotriol, which, as previously mentioned, is suggested to be produced by nitrite-oxidizing bacteria (Elling et al.,
325 2022). Therefore, it is more likely that in the Black Sea, 2Me-BHT is tracking changes in N₂-fixing cyanobacteria.

326

327 4.5. BHPs associated with methane-oxidizing bacteria

328 Aminotetrol and aminopentol are present throughout the lacustrine phase of the Black Sea, albeit in low
329 absolute abundance compared to the other phases (Fig. S5). Aminotetrol and aminopentol are associated with MOB
330 (Rohmer et al., 1984; Jahnke et al., 1999; Cvejic et al., 2000; Talbot et al., 2001; Zhu et al., 2011; van Winden et al.,
331 2012; Rush et al., 2016), but are also produced in small proportions by sulfate-reducing bacteria (Blumenberg et al.,
332 2006; 2009b; 2012). In addition, aminopentol was detected in cultured nitrite-oxidizing bacteria, *Nitrospira defluvii*
333 and *Nitrobacter vulgaris* (Elling et al., 2022). Indicators of a well-oxygenated water column during the lacustrine
334 phase (Cutmore et al., 2025), would suggest that methane-oxidizing bacteria are a more likely source of aminotetrol
335 and aminopentol within the lacustrine water column than sulfate-reducing bacteria, which are typically considered
336 to be obligate anaerobes (Hao et al., 1996). Although we cannot rule out possible sulfate-reduction in the sediment,
337 other independent biomarkers suggest that aerobic methanotrophs were active in the basin and are a more likely
338 source for these biomarkers. For instance, high concentrations of 17 β (H)-moret-22(29)-ene with depleted $\delta^{13}\text{C}$
339 values were previously detected in sediments from the lacustrine phase from the Black Sea, and are thought to be
340 sourced from aerobic methanotrophs in either the water column or the surface sediments (Uemura and Ishiwatari,
341 1995; Blumenberg et al., 2009a). Nitrite-oxidizing bacteria are also a potential source for aminopentol (Elling et al.,
342 2022), but as both aminopentol and aminotetrol vary independently of BHPs and other biomarkers associated with
343 the nitrogen-cycle (such as crenarchaeol), it is likely that nitrite-oxidizing bacteria are not a major source of these
344 BHPs. Thus, aminotetrol and aminopentol were likely produced by methane-oxidizing bacteria near the oxycline of
345 the basin, or, if the basin was fully oxygenated, in the surface sediments (Neunlist and Rohmer, 1985a,b; Cvejic et
346 al., 2000; Talbot et al., 2001). The occurrence of temporary anoxia in the bottom waters or surface sediments during
347 the lacustrine phase is supported by the continuous presence of dioxanone-methylaminotriol throughout this
348 period, with the highest absolute abundance occurring between 19.5 and 16.4 ka. Dioxanone-methylaminotriol was
349 previously identified in the surface sediments and bottom waters of basins that experience seasonal hypoxia or
350 anoxia (Richter et al., 2023).



351 Several BHPs increased during the transition period relative to the preceding lacustrine phase (i.e., methoxy-
352 BHT, aminotriol, aminotetrol, EC-aminotriol, ethenolamine-BHT, unsaturated ethenolamine-BHT, ethenolamine-
353 BHpentol, methoxy-ethenolamine, propenolamine-BHT and unsaturated propenolamine-BHT), with several BHPs
354 gradually increasing in absolute abundance over the transition phase (i.e., aminotriol and N-acyl-aminotriols $C_{12:0}$,
355 $C_{14:0}$, $C_{15:0}$, $C_{16:0}$ and $C_{18:0}$) (Fig. S4.2). The continuous presence of aminotetrol and high absolute abundance of
356 aminotriol during this period is likely associated with methane-oxidizing bacteria as the pelagic redoxcline was
357 established. Similarly, in the Baltic Sea, the transition from the Ancyclus Lake phase to the Littorina Sea was marked
358 with a peak in aminotriol and aminotetrol, attributed to methane-oxidizing bacteria (Blumenberg et al., 2013). This
359 interpretation is also supported by the presence of ethenolamine-BHpentol, which has so far only been detected
360 near a terrestrial methane seep and in lakes with low oxygen conditions in the bottom waters (Hopmans et al., 2021;
361 Richter et al., 2023). Similarly, during the transition phase in the Black Sea there is an increase in archaeal lipids
362 derived from anaerobic methanotrophs that might indicate heightened activity by anaerobic methane-oxidizing
363 archaeal communities; alternatively, these lipids could be derived from overprinting in the sediments (Zhu et al.,
364 2024). The presence of MOB-associated BHPs would suggest that aerobic methane-oxidation occurred during this
365 phase, but it is unclear whether this coincided with an increase in anaerobic methane-oxidation.

366 During the marine phase, there is a distinct increase in aminotetrol, aminopentol, ethenolamine-BHpentol,
367 ethenolamine-BHhexol, and propenolamine-BHT. While propenolamine-BHT has been observed near a terrestrial
368 methane seep, as well as in lakes and in the sediments of coastal lagoons, its microbial source remains uncertain
369 (Hopmans et al., 2021; Richter et al., 2023). Methanotrophic bacteria, known to thrive in the oxic-anoxic transition
370 zone in the modern Black Sea water column (Kuypers et al., 2003; Durisch-Kaiser et al., 2005; Schubert et al. 2006),
371 are known sources of aminotetrol and aminopentol (Blumenberg et al., 2007; Wakeham et al., 2007), however, the
372 distributions and concentrations of BHPs detected in the modern-day Black Sea water column are not reflected in
373 the geological record as there is no effective transport mode from the oxic-anoxic transition zone to the sediment
374 (Wakeham et al., 2007; Blumenberg et al., 2009a; Kusch et al., 2022). If the oxic-anoxic transition zone occurs near
375 the photic zone, however, and there is a large accumulation of particles, it is possible for particulate matter to be
376 transported from this zone to the sediments (e.g., Repeta, 1993; Sinninghe Damsté et al., 1993; Coolen et al., 2008).
377 The high absolute abundance of isorenieratene throughout the early marine phase indicates that euxinic conditions
378 reached the photic zone during this period (Cutmore et al., 2025), indicating a shallower oxic-anoxic interface
379 compared to the later marine phase. This likely led to a heightened transport of BHPs from the oxic-anoxic interface
380 to the sediments (Blumenberg et al., 2009a). Between 5 and 3.9 ka, aminotetrol and aminopentol may have also
381 been produced by sulfate-reducing bacteria (SRB) in the water column or sediments. There is, however, also no
382 effective transport of lipids from the deeper waters to the sediments (Schouten et al., 2001; Wakeham et al., 2003).
383 Further, genetic analyses of a Black Sea sediment core shows that although sulfate-reducing bacteria are abundant
384 in the sulfate-methane transition zone, the *Desulfovibrio* genus (which is known to produce BHPs [Blumenberg et
385 al., 2006; 2009b; 2012]) plays a minor role in the Black Sea sediments compared to *Desulfobacter* (Leloup et al.,



2006). *Desulfobacter*, in contrast, does not contain the squalene-hopene cyclase gene (accession no. WP_038942977.1) required for BHP synthesis as confirmed by a protein blast search (NCBI, National Center for Biotechnology Information). Aside from aminotetrol and aminopentol, the only other known BHPs produced by SRB are BHT-22S, BHT-CE, and aminotriol (Blumenberg et al. 2006; 2009b; 2012). Aminotriol is present in low abundance throughout the marine phase. Both BHT-22S and BHT-CE increase during the late marine phase, however, these BHPs could also be derived from other bacterial sources as they are not unique to SRB (see Talbot and Farrimond, 2007). Thus, unless there are previously unknown BHP-producing SRBs in the Black Sea sediments or water column, SRB are likely only a minor source of BHPs during the marine phase of our record (Blumenberg et al., 2009a). Similarly, N-acyl-aminotriols (i.e., C_{12:0}, C_{14:0}, C_{15:0}, C_{16:0}, C_{17:0}, and C_{18:0}) are also present in high absolute abundance during the early marine phase compared to the rest of the core and peak at 6.1 ka. There is no known distinct source for N-acyl-aminotriols; however, they have been detected in a methanotroph culture as well as various environmental settings (e.g., Hopmans et al., 2021; Kusch et al., 2022; Richter et al., 2023). The high abundance of these long-chain compounds might be associated with heightened transport from the oxycline to the sediments and enhanced preservation during this time (Blumenberg et al., 2009a; Cutmore et al., 2025).

High-rates of methane-oxidation in the modern-day Black Sea are mediated by anaerobic methanotrophs in the euxinic water column, with lower rates of anaerobic methane oxidation occurring in the sediments (Reeburgh et al., 1991; Lin et al., 2006; Schubert et al., 2006). MOB occur at the oxic-anoxic transition zone of the modern-day Black Sea water column, however, aerobic methane-oxidation rates are several orders of magnitude lower compared to the anoxic water column (Reeburgh et al., 1991; Schubert et al., 2006). Consequently, the limited transport of suspended particulate matter from the chemocline and euxinic bottom waters to the sediments, leads to an underrepresentation of these lipids in the geological record and restricts any lipid-based interpretations of methane-oxidizing microbial communities or associated sulfate-reducing bacteria in the Black Sea water column during the late marine phase (Schouten et al. 2001; Wakeham et al., 2003; Schubert et al., 2006). Studies of lipid biomarkers associated with anaerobic methanotrophs (e.g., compound-specific carbon isotope compositions of archaeal lipids) confirm that there was minimal overprinting from active methane-oxidizing archaea in the sediments and fossil lipids derived from anaerobic methanotrophs were only a minor portion of the archaeal lipid assemblage during the marine phase (Zhu et al., 2024). In summary, MOB-associated BHPs in our record indicate that aerobic methane-oxidation occurred throughout the Black Sea record. However, archaeal lipids and modern studies demonstrate that anaerobic methane-oxidation became a more significant “methane sink” in the Black Sea water column during the marine phase relative to the lacustrine phase. Additional studies are needed to confirm this hypothesis by quantifying the contributions of aerobic versus anaerobic methanotrophs and identifying sulfate-reducing bacteria during the lacustrine and transition phase.

418

419 *4.6. BHPs as indicators for surface salinity*



420 Methoxy-BHT has previously been detected in marine environments and at present, has not been detected
421 in lake samples (Richter et al., 2023). In the Black Sea record, we detect methoxy-BHT intermittently during the oxic
422 lacustrine phase, possibly due to the slightly brackish conditions that existed during this period (Huang et al., 2021).
423 After the IMI, the absolute abundance of this BHP increases, possibly due to the influx of saline water. The highest
424 abundance of methoxy-BHT occurs between 6.1 and 4.2 ka, when surface water salinities reach a peak (Filipova-
425 Marinova et al., 2013; Huang et al., 2021). The reduction in methoxy-BHT between 3.8 and 1.5 ka coincides with the
426 absence of isorenieratene which likely resulted from the erosion of the chemocline (Sinninghe Damsté et al., 1993)
427 due to enhanced freshwater input, with a short reoccurrence of freshwater/brackish species occurring at this time
428 (Filipova-Marinova et al., 2013; Huang et al., 2021). Consequently, the lower salinity of the surface waters may have
429 also led to the decline in methoxy-BHT. Methoxy-BHT and methoxy-ethanolamine-BHT show similar patterns
430 throughout this period (Fig. 5), both peaking between 6.1 and 4.2 ka, declining between 3.8 and 1.5 ka and increasing
431 towards the end of the record. This indicates that both BHPs may be a specific adaptation of BHP compositions to
432 saline environments or are linked to specific microbes adapted to brackish to marine systems.

433

434 5. Conclusions

435 The Black Sea record shows substantial shifts in BHP absolute abundance and composition over the last 20 ka, which
436 correspond to major environmental transitions. During the lacustrine phase (19.5 – 9.6 ka), high Nu-BHP absolute
437 abundance and diversity likely reflects enhanced terrigenous input during the Last Deglaciation and/or in situ
438 production in the water column. The transition phase (9.6 – 7.2 ka) is marked by a decrease in many BHPs, such as
439 Nu-BHPs, and an increase in methoxylated-BHPs and more ubiquitous BHPs (e.g., aminotriol), likely linked to
440 changing oxygenation and salinity of the water column. The early euxinic marine phase (7.2 ka – 5 ka) is characterized
441 by a high aminotetrol and aminopentol absolute abundance, likely transported with sinking particulate matter to
442 the sediment from a shallow oxycline. The late marine phase (5 ka – present) is marked by a decline in these BHPs,
443 likely due to a deepening of the oxycline. BHP distribution changes throughout the record are attributed either to
444 microbial adaptations to shifts in the oxygen levels or hydrology of the basin, or are associated with specific groups
445 of bacteria suited to these conditions. For instance, BHT-CE was closely associated with crenarchaeol throughout the
446 record, indicating that these compounds are either produced in a similar ecological niche or by related microbes.
447 Aminotetrol, aminopentol, ethanolamine-BHPentol and -BHHexol were detected in the Black Sea record as likely
448 biomarkers for methane-oxidizing bacteria, while methoxylated-BHPs, detected only during the transition and
449 marine phases, may be associated with higher salinities or marine environments. The changes in the Nu-BHP
450 distribution likely reflects changing terrestrial input over this period, with the exception of some Nu-BHPs (e.g., 2Me-
451 adenosylhopane_{HG-diMe} and Me-adenosylhopane_{HG-diMe}) that are likely produced in the water column. BHP
452 degradation products were detected as a minor component of the overall BHP distribution, suggesting good BHP



453 preservation throughout the record. This new BHP record provides valuable insights into ecological changes as well
454 as changes in the nitrogen and methane cycle in the Black Sea over the last 20 ka.

455

456

457 **Data Availability**

458 All data generated for this study are archived and publicly available via the Mendeley Data repository online at
459 <http://doi.org/10.17632/m6v5mj5gtp.1> (Cutmore and Richter et al., 2025).

460

461 **Author Contribution**

462 AC- Conceptualization, Formal analysis, Investigation, Data Curation, Visualization, Writing - Original Draft, Writing
463 – Review and Editing. NR- Conceptualization, Formal analysis, Investigation, Data Curation, Visualization, Writing -
464 Original Draft, Writing – Review and Editing. DR- Supervision, Writing – Review and Editing. NB- Supervision, Writing
465 – Review and Editing. SS- Supervision, Funding acquisition, Writing – Review and Editing.

466

467 **Competing Interests**

468 The authors declare that they have no conflict of interest.

469

470 **Acknowledgements**

471 We thank the Chief Scientist Prof. Laura Villanueva and Dr. Rick Hennekam, as well as the captain and crew of the
472 *R/V Pelagia* for the collection of core 64PE418. For laboratory support we thank Anhelique Mets, Denise Dorhout
473 and Monique Verweij. Thank you to Diana Sahonero Canavesi for helpful discussions and Bingjie Yang for providing
474 helpful insight into the BIT index of this core. This study was funded by the Netherlands Earth System Science Centre
475 (024.002.001) from the Dutch Ministry of Education, Culture and Science (OCW) and the Swiss National Science
476 Foundation (SNSF) Ambizione Fellowship (PZ00P2_216050).

477

478

479 **References:**

480

481 Aksu, A., Hiscott, R.N., Kaminski, M.A., Mudie, P.J., Gillespie, H., Abrajano, T and Yasar, D.: Last glacial–Holocene
482 paleoceanography of the Black Sea and Marmara Sea: stable isotopic, foraminiferal and coccolith evidence, *Mar.*
483 *Geol.*, **190**, 119–149, doi.org/10.1016/S0025-3227(02)00345-6, 2002.

484

485 Ankindinova, O., Hiscott, R.N., Aksu, A.E and Grimes, V.: High-resolution Sr-isotopic evolution of Black Sea water
486 during the Holocene: Implications for reconnection with the global ocean, *Mar. Geol.*, **407**, 213–228,
487 doi.org/10.1016/j.margeo.2018.11.004, 2019.



488
489 Arthur, M.A and Dean, W.E.: Organic-matter production and preservation and evolution of anoxia in the Holocene
490 Black Sea, *Paleoceanogr. Paleoclimatol.*, **13**, 395-411, doi.org/10.1029/98PA01161, 1998.
491
492 Badertscher, S., Fleitmann, D., Cheng, H., Edwards, R.L., Göktürk, O.M., Zumbühl, A., Leuenberger, M and Tüysüz, O.:
493 Pleistocene water intrusions from the Mediterranean and Caspian seas into the Black Sea, *Nat. Geosci.*, **4**, 236–239,
494 doi.org/10.1038/ngeo1106, 2011.
495
496 Bahr, A., Lamy, F., Arz, H., Kuhlmann, H and Wefer, G.: Late glacial to Holocene climate and sedimentation history in
497 the NW Black Sea, *Mar. Geol.*, **214**, 309-322, doi.org/10.1016/j.margeo.2004.11.013, 2005.
498
499 Bahr, A., Arz, H., Lamy, F and Wefer, G.: Late glacial to Holocene paleoenvironmental evolution of the Black Sea,
500 reconstructed with stable oxygen isotope records obtained on ostracod shells, *Earth Planet. Sc. Lett.*, **241**, 863-875,
501 doi.org/10.1016/j.epsl.2005.10.036, 2006.
502
503 Bahr, A., Lamy, F., Arz, H., Major, C., Kwicien, O and Wefer, G.: Abrupt changes of temperature and water chemistry
504 in the late Pleistocene and early Holocene Black Sea, *Geochem., Geophys., Geosy.*, **9**, 1-16,
505 doi.org/10.1029/2007GC001683, 2008.
506
507 Bale, N., Ding, S., Hopmans, E.C., Arts, M.G.I., Villanueva, L., Boschman, C., Haas, A.F., Schouten, S and Sinninghe
508 Damsté, J.S.: Lipidomics of Environmental Microbial Communities. I: Visualization of Component Distributions Using
509 Untargeted Analysis of High-Resolution Mass Spectrometry Data, *Front. Microbiol.*, **12**, 1-15,
510 doi.org/10.3389/fmicb.2021.659302, 2021.
511
512 Bednarczyk, A., Hernandez, T. C., Schaeffer, P., Adam, P., Talbot, H. M., Farrimond, P., Riboulleau, A., Largeau, C.,
513 Derenne, S., Rohmer, M., and Albrecht, P.: 32,35-Anhydrobacteriohopanetetrol: an unusual bacteriohopanepolyol
514 widespread in recent and past environments, *Org. Geochem.*, **36**, 673–677,
515 doi.org/10.1016/j.orggeochem.2004.10.014, 2005.
516
517 Bissere, P., Zundel, M., and Rohmer, M.: Prokaryotic triterpenoids, *Eur. J. Biochem.*, **150**, 29–34,
518 doi.org/10.1111/j.1432-1033.1985.tb08982.x, 1985.
519
520 Blaaw, M.: Methods and code for ‘classical’ age-modelling of radiocarbon sequences, *Quat. Geochron.*, **5**, 512-518,
521 doi.org/10.1016/j.quageo.2010.01.002, 2010.
522



- 523 Blumenberg, M., Krüger, M., Nauhaus, K., Talbot, H. M., Oppermann, B. I., Seifert, R., Pape, T., and Michaelis, W.:
524 Biosynthesis of hopanoids by sulfate-reducing bacteria (genus *Desulfovibrio*), *Environ. Microbiol.*, **8**, 1220–1227,
525 doi.org/10.1111/j.1462-2920.2006.01014.x, 2006.
- 526
- 527 Blumenberg, M., Seifert, R., and Michaelis, W.: Aerobic methanotrophy in the oxic–anoxic transition zone of the
528 Black Sea water column, *Org. Geochem.*, **38**, 84–91, doi.org/10.1016/j.orggeochem.2006.08.011, 2007.
- 529
- 530 Blumenberg, M., Hoppert, M., Krüger, M., Dreier, A., and Thiel, V.: Novel findings on hopanoid occurrences among
531 sulfate reducing bacteria: Is there a direct link to nitrogen fixation?, *Org. Geochem.*, **49**, 1–5,
532 doi.org/10.1016/j.orggeochem.2012.05.003, 2012.
- 533
- 534 Blumenberg, M., Seifert, R., Kasten, S., Bahlmann, E and Michaelis, W.: Euphotic zone bacterioplankton sources
535 major sedimentary bacteriohopanepolyols in the Holocene Black Sea, *Geochim. Cosmochim. Ac.*, **73**, 750–766,
536 doi.org/10.1016/j.gca.2008.11.005, 2009a.
- 537
- 538 Blumenberg, M., Oppermann, B. I., Guyoneaud, R., and Michaelis, W.: Hopanoid production by *Desulfovibrio bastinii*
539 isolated from oilfield formation water, *FEMS Microbiol. Lett.*, **293**, 73–78, doi.org/10.1111/j.1574-
540 6968.2009.01520.x, 2009b.
- 541
- 542 Blumenberg, M., Berndmeyer, C., Moros, M., Muschalla, M., Schmale, O and Thiel, V.: Bacteriohopanepolyols record
543 stratification, nitrogen fixation and other biogeochemical perturbations in Holocene sediments of the central Baltic
544 Sea, *Biogeosciences*, **10**, 2725–2735, doi:10.5194/bg-10-2725-2013, 2013.
- 545
- 546 Bravo, J.-M., Perzl, M., Härtner, T., Kannenberg, E. L and Rohmer, M.: Novel methylated triterpenoids of the
547 gammacerane series from the nitrogen-fixing bacterium *Bradyrhizobium japonicum* USDA 110, *Eur. J. Biochem.*, **268**,
548 1323–1331, doi.org/10.1046/j.1432-1327.2001.01998.x, 2001.
- 549
- 550 Coolen, M.J.L., Abbas, B., Van Bleijswijk, J., Hopmans, E.C., Kuypers, M.M.M., Wakeham, S.G and Sinninghe Damsté,
551 J.S.: Putative ammonia-oxidizing Crenarchaeota in suboxic waters of the Black Sea: a basin-wide ecological study
552 using 16S ribosomal and functional genes and membrane lipids, *Environ. Microbiol.*, **9**, 1001–1016,
553 doi.org/10.1111/j.1462-2920.2006.01227.x, 2007.
- 554
- 555 Coolen M.J.L., Talbot, H.M., Abbas, B., Ward, C., Schouten, S., Volkman, J.K and Sinninghe Damsté, J.S.: Sources for
556 sedimentary bacteriohopanepolyols as revealed by 16S rDNA stratigraphy, *Environ. Microbiol.*, **10**, 1783–1803,
557 doi.org/10.1111/j.1462-2920.2008.01601.x, 2008.



558
559 Cooke, M. P., Talbot, H. M and Farrimond, P.: Bacterial populations recorded in bacteriohopanepolyol distributions
560 in soils from Northern England, *Org. Geochem.*, **39**, 1347–1358, doi.org/10.1016/j.orggeochem.2008.05.003, 2008a.
561
562 Cooke, M.P., Talbot, H.M and Wagner, T.: Tracking soil organic carbon transport to continental margin sediments
563 using soil-specific hopanoid biomarkers: a case study from the Congo fan (ODP site 1075), *Org. Geochem.*, **39**, 965-
564 971, <https://doi.org/10.1016/j.orggeochem.2008.03.009>, 2008b.
565
566 Cutmore, A., Bale, N., Hennekman, R., Yang, B., Rush, D., Schouten, Reichart, G.-J., Hopmans, E and S, Schouten.:
567 Impact of deoxygenation and hydrological changes on the Black Sea nitrogen cycle during the Last Deglaciation and
568 Holocene, *Clim. Past*, doi.org/10.5194/cp-2024-59, 2025.
569
570 Cvejic, J.H., Bodrossy, L., Kovács, K.L and Rohmer, M.: Bacterial triterpenoids of the hopane series from the
571 methanotrophic bacteria *Methylocaldum* spp.: phylogenetic implications and first evidence for an unsaturated
572 aminobacteriohopanepolyol, *FEMS Microbiol. Lett.*, **182**, 361–365, doi.org/10.1111/j.1574-6968.2000.tb08922.x,
573 2000.
574
575 Dalsgaard, T., Thamdrup, B., Fariás, L and Revsbech, N.P.: Anammox and denitrification in the oxygen minimum zone
576 of the eastern South Pacific, *Limnol. Oceanog.*, **57**, 1331-1346, doi.org/10.4319/lo.2012.57.5.1331, 2012.
577
578 Durisch-Kaiser, E., Klauser, L., Wehrli, B., and Schubert, C.: Evidence of Intense Archaeal and Bacterial
579 Methanotrophic Activity in the Black Sea Water Column, *Appl. Environ. Microbiol.*, **71**, 8099–8106,
580 doi.org/10.1128/AEM.71.12.8099-8106.2005, 2005.
581
582 Deutzmann, J.S., Stief, P., Brandes, J and Schink, B.: Anaerobic methane oxidation coupled to denitrification is the
583 dominant methane sink in a deep lake, *Proc. Nat. Acad. Sci. USA.*, **111**, 18273-18278,
584 doi.org/10.1073/pnas.1411617111, 2014.
585
586 Eckert, S., Brumsack, H.-J., Severmann, S., Schnetger, B., März, C and Fröllje, H.: Establishment of euxinic conditions
587 in the Holocene Black, *Geology*, **41**, 431–434, doi.org/10.1130/G33826.1, 2013.
588
589 Eickhoff, M., Birgel, D., Talbot, H. M., Peckmann, J., and Kappler, A.: Diagenetic degradation products of
590 bacteriohopanepolyols produced by *Rhodopseudomonas palustris* strain TIE-1, *Org. Geochem.*, **68**, 31–38,
591 doi.org/10.1016/j.orggeochem.2014.01.002, 2014.
592



- 593 Elling, F. J., Hemingway, J. D., Evans, T. W., Kharbush, J. J., Spieck, E., Summons, R. E and Pearson, A.: Vitamin B12 -
594 dependent biosynthesis ties amplified 2-methylhopanoid production during oceanic anoxic events to nitrification,
595 *Proc. Nat. Acad. Sci. USA.*, **117**, 32996–33004, doi.org/10.1073/pnas.2012357117, 2020.
596
- 597 Elling, F. J., Hemingway, J. D., Kharbush, J. J., Becker, K.W., Polik, C.A and Pearson, A.: Linking diatom-diazotroph
598 symbioses to nitrogen cycle perturbations and deep-water anoxia: Insights from Mediterranean sapropel events,
599 *Earth Planet. Sc. Lett.*, **571**, 1-11, doi.org/10.1016/j.epsl.2021.117110, 2021.
600
- 601 Elling, F. J., Evans, T. W., Nathan, V., Hemingway, J. D., Kharbush, J. J., Bayer, B., Spieck, E., Husain, F., Summons, R.
602 E and Pearson, A.: Marine and terrestrial nitrifying bacteria are sources of diverse bacteriohopanepolyols,
603 *Geobiology*, **20**, 399-420, doi.org/10.1111/gbi.12484, 2022.
604
- 605 Ettwig, K. F., Butler, M. K., Le Paslier, D., Pelletier, E., Mangenot, S., Kuypers, M. M. M., Schreiber, F., Dutilh, B. E.,
606 Zedelius, J., De Beer, D., Gloerich, J., Wessels, H. J. C. T., Van Alen, T., Luesken, F., Wu, M. L., Van De Pas-Schoonen,
607 K. T., Op Den Camp, H. J. M., Janssen-Megens, E. M., Francoijs, K.-J., Stunnenberg, H., Weissenbach, J., Jetten, M. S.
608 M., and Strous, M.: Nitrite-driven anaerobic methane oxidation by oxygenic bacteria, *Nature*, **464**, 543–548,
609 doi.org/10.1038/nature08883, 2010.
610
- 611 Filipova-Marinova, M., Pavlov, D., Coolen, M and Giosan, L.: First high-resolution marinopalynological stratigraphy
612 of Late Quaternary sediments from the central part of the Bulgarian Black Sea area, *Quatern. Int.*, **293**, 170-183,
613 doi.org/10.1016/j.quaint.2012.05.002, 2013.
614
- 615 Handley, L., Talbot, H.M., Cooke, M.P., Anderson, K.E and Wagner, T.: Bacteriohopanepolyols as tracers for
616 continental and marine organic matter supply and phases of enhanced nitrogen cycling on the late Quaternary Congo
617 deep sea fan, *Org. Geochem.*, **41**, 910-914, doi.org/10.1016/j.orggeochem.2010.04.016, 2010.
618
- 619 Hao, O.J., Chen, J.M., Huang, L and Buglass, R.L.: Sulfate- reducing bacteria, *Crit. Rev. Env. Sci. Tec.*, **26**, 155-187,
620 doi.org/10.1080/10643389609388489, 1996.
621
- 622 Heaton, T.J., Köhler, P., Butzin, M., Bard, E., Reimer, R.W., Austin, W.E.N., Bronk Ramsey, C., Grootes, P.M., Hughen,
623 K.A., Kromer, B., Reimer, P.J., Adkins, J., Burke, A., Cook, M.S., Olsen, J and Skinner, L.C.: Marine20—The Marine
624 Radiocarbon Age Calibration Curve (0–55,000 cal BP), *Radiocarbon*, **62**, 779–820, doi.org/10.1017/RDC.2020.68,
625 2020.
626



- 627 Hiscott, R.N., Aksu, A.E., Mudie, P.J., Marret, F., Abrajano, T., Kaminski, M.A., Evans, J., Çakiroğlu, A.İ., Yaşar, D.: A
628 gradual drowning of the southwestern Black Sea shelf: Evidence for a progressive rather than abrupt Holocene
629 reconnection with the eastern Mediterranean Sea through the Marmara Sea Gateway, *Quatern. Int.*, **167–168**, 19–
630 34, doi.org/10.1016/j.quaint.2006.11.007, 2007.
- 631
- 632 Hopmans, E.C., Weijers, J.W.H., Schefuß, E., Herfort, L., Sinninghe Damsté, J.S and Schouten, S.: A novel proxy for
633 terrestrial organic matter in sediments based on branched and isoprenoid tetraether lipids, *Earth Planet. Sc. Lett.*,
634 **224**, 107–116, doi.org/10.1016/j.epsl.2004.05.012, 2004.
- 635
- 636 Hopmans, E.C., Smit, N.T., Schwartz-Narbonne, R., Sinninghe Damsté, J.S and Rush, D.: Analysis of non-derivatized
637 bacteriohopanepolyols using UHPLC-HRMS reveals great structural diversity in environmental lipid assemblages,
638 *Org. Geochem.*, **160**, 1–17, doi.org/10.1016/j.orggeochem.2021.104285, 2021.
- 639
- 640 Huang, Y., Zheng, Y., Heng, P., Giosan, L and Coolen, M.J.L.: Black Sea paleosalinity evolution since the last
641 deglaciation reconstructed from alkenone-inferred Isochrysidales diversity, *Earth Planet. Sc. Lett.*, **564**, 1–9,
642 doi.org/10.1016/j.epsl.2021.116881, 2021.
- 643
- 644 Ion, G., Briceag, A., Vasiliu, D., Lupaşcu, N and Melinte-Dobrinescu, M.: A multiproxy reconstruction of the Late
645 Pleistocene-Holocene paleoenvironment: New insights from the NW Black Sea, *Mar. Geol.*, **443**, 1–19,
646 doi.org/10.1016/j.margeo.2021.106648, 2022.
- 647
- 648 Ivanova, E.V., Murdmaa, I.O., Chepalyga, A.L., Cronin, T.M., Pasechnik, I.V., Levchenko, O.V., Howe, S.S., Manushkina,
649 A.V and Platonova, E.A.: Holocene sea-level oscillations and environmental changes on the Eastern Black Sea shelf,
650 *Palaeogeogr. Palaeoclimatol. Palaeoecol.*, **246**, 228–259, doi.org/10.1016/j.palaeo.2006.09.014, 2007.
- 651
- 652 Jahnke, L.L., Summons, R., Hope, J.M and Des Marais, D.J.: Carbon isotopic fractionation in lipids from
653 methanotrophic bacteria II: the effects of physiology and environmental parameters on the biosynthesis and isotopic
654 signatures of biomarkers, *Geochim. Cosmochim. Ac.*, **63**, 79–93, doi.org/10.1016/S0016-7037(98)00270-1, 1999.
- 655
- 656 Jones, G.A and Gagnon, A.R.: Radiocarbon chronology of Black Sea sediments, *Deep-Sea Res. Pt. 1.*, **41**, 531–557,
657 doi.org/10.1016/0967-0637(94)90094-9, 1994.
- 658
- 659 Kassambara, A and Mundt, F.: Factoextra: Extract and Visualize the Results of Multivariate Data Analyses, R Package
660 Version 1.0.7, <http://CRAN.R-project.org/package=factoextra>, 2020.
- 661



- 662 Köneke, M., Bernhard, A.E., de la Torre, J.R., Walker, C.B., Waterbury, J.B and Stahl, D.A.: Isolation of an autotrophic
663 ammonia-oxidizing marine archaeon, *Nature*, **437**, 543–546, doi.org/10.1038/nature03911, 2005.
- 664
- 665 Kool, D.M., Talbot, H.M., Rush, D., Ettwig, K and Sinninghe Damsté, J.S.: Rare bacteriohopanepolyols as markers for
666 an autotrophic, intra-aerobic methanotroph, *Geochim. Cosmochim. Ac.*, **136**, 114-125,
667 doi.org/10.1016/j.gca.2014.04.002, 2014.
- 668
- 669 Kulkarni, G., Busset, N., Molinaro, A., Gargani, D., Chaintreuil, C., Silipo, A., Giraud, E and Newman, D. K.: Specific
670 Hopanoid Classes Differentially Affect Free-Living and Symbiotic States of *Bradyrhizobium diazoefficiens*. *mBio*, **6**,
671 e01251-15, doi.org/10.1128/mBio.01251-15, 2015.
- 672
- 673 Kusch, S., Sepúlveda, J and Wakeham, S.G.: Origin of Sedimentary BHPs Along a Mississippi River–Gulf of Mexico
674 Export Transect: Insights From Spatial and Density Distributions, *Front. Mar. Sci.*, **6**, 1-14,
675 doi.org/10.3389/fmars.2019.00729, 2019.
- 676
- 677 Kusch, S., Wakeham, S. G and Sepúlveda, J.: Diverse origins of “soil marker” bacteriohopanepolyols in marine oxygen
678 deficient zones, *Org. Geochem.*, **151**, 104150, doi.org/10.1016/j.orggeochem.2020.104150, 2021.
- 679
- 680 Kusch, S., Wakeham, S.G and Sepúlveda, J.: Bacteriohopanepolyols across the Black Sea redoxcline trace diverse
681 bacterial metabolisms, *Org. Geochem.*, **172**, 1-18, doi.org/10.1016/j.orggeochem.2022.104462, 2022.
- 682
- 683 Kuypers, M.M.M., Sliekers, A.O., Lavik, G., Schmid, M., Barker Jørgensen, B., Kuenen, J.G., Sinninghe Damsté, J.S.,
684 Strous, M and Jetten, M.S.M.: Anaerobic ammonium oxidation by anammox bacteria in the Black Sea. *Nature*, **422**,
685 608-611, doi.org/10.1038/nature01472, 2003.
- 686
- 687 Kuypers, M. M. M., Van Breugel, Y., Schouten, S., Erba, E and Sinninghe Damsté, J. S.: N₂-fixing cyanobacteria
688 supplied nutrient N for Cretaceous oceanic anoxic events, *Geology*, **32**, 853, doi.org/10.1130/G20458.1, 2004.
- 689
- 690 Kwiecien, O., Arz, H.W., Lamy, F., Wulf, S., Bahr, A., Röhl, U and Haug, G.H.: Estimated Reservoir Ages of the Black
691 Sea Since the Last Glacial, *Radiocarbon*, **50**, 99-118, doi.org/10.1017/S0033822200043393, 2008.
- 692
- 693 Lam, P., Jensen, M.M., Lavik, G., McGinnis, D.F., Müller, B., Schubert, C.J., Amann, R., Thamdrup, B., and Kuypers,
694 M.M.M.: Linking crenarchaeal and bacterial nitrification to anammox in the Black Sea, *Proc. Nat. Acad. Sci. USA.*,
695 **104**, 7104-7109, doi.org/10.1073/pnas.061108110, 2007.
- 696



- 697 Lê, S., Josse, J and Husson, F.: FactoMineR: An R Package for Multivariate Analysis, *J. Stat. Softw.*, **25**(1), 1–18,
698 doi.org/10.18637/jss.v025.i01, 2008.
- 699
- 700 Leloup, J., Loy, A., Knab, N.J., Borowski, C., Wagner, M. and Jørgensen, B.B.: Diversity and abundance of sulfate-
701 reducing microorganisms in the sulfate and methane zones of a marine sediment, Black Sea, *Environ. Microbiol.*, **9**,
702 131-142, doi.org/10.1111/j.1462-2920.2006.01122.x, 2007.
- 703
- 704 Lin, X., Wakeham, S. G., Putnam, I. F., Astor, Y. M., Scranton, M. I., Chistoserdov, A. Y and Taylor, G. T.: Comparison
705 of Vertical Distributions of Prokaryotic Assemblages in the Anoxic Cariaco Basin and Black Sea by Use of Fluorescence
706 In Situ Hybridization, *Appl. Environ. Microbiol.*, **72**, 2679–2690, doi.org/10.1128/AEM.72.4.2679-2690.2006, 2006.
- 707
- 708 Major, C., Goldstein, S., Ryan, W., Lericolais, G., Piotrowski, A.M and Hajdas, I.: The co-evolution of Black Sea level
709 and composition through the last deglaciation and its paleoclimatic significance, *Quaternary Sci. Rev.*, **25**, 2031-2047,
710 doi.org/10.1016/j.quascirev.2006.01.032, 2006.
- 711
- 712 Marret, F., Mudie, P., Aksu, A and Hiscott, R.N.: A Holocene dinocyst record of a two-step transformation of the
713 Neoeuxinian brackish water lake into the Black Sea, *Quatern. Int.*, **197**, 72-86, doi.org/10.1016/j.quaint.2007.01.010,
714 2009.
- 715
- 716 Matys, E.D., Sepúlveda, J., Pantoja, S., Lange, C.B., Caniupán, M., Lamy, F and Summons, R. E.: Bacteriohopanepolyols
717 along redox gradients in the Humboldt Current System off northern Chile, *Geobiology*, **15**, 844-857,
718 doi.org/10.1111/gbi.12250, 2017.
- 719
- 720 Naafs, B. D. A., Bianchini, G., Monteiro, F. M and Sánchez-Baracaldo, P.: The occurrence of 2-methylhopanoids in
721 modern bacteria and the geological record, *Geobiology*, **20**, 41–59, doi.org/10.1111/gbi.12465, 2022.
- 722
- 723 Neunlist, S. and Rohmer, M.: Novel hopanoids from the methylotrophic bacteria *Methylococcus capsulatus* and
724 *Methylomonas methanica*. (22S)-35-aminobacteriohopane-30,31,32,33,34-pentol and (22S)-35-amino-3 beta-
725 methylbacteriohopane-30,31,32,33,34-pentol, *Biochemical Journal*, **231**, 635–639, doi:10.1042/bj2310635, 1985a.
- 726
- 727 Neunlist, S. and Rohmer, M.: The Hopanoids of '*Methylosinus trichosporium*': Aminobacteriohopanetriol and
728 Aminobacteriohopanetetrol, *Microbiology*, **131**, 1363–1367, doi.org/10.1099/00221287-131-6-1363, 1985b.
- 729



- 730 Nicholas, W.A., Chivas, A.R., Murray-Wallace, C.V and Fink, D.: Prompt transgression and gradual salinisation of the
731 Black Sea during the early Holocene constrained by amino acid racemization and radiocarbon dating, *Quaternary*
732 *Sci. Rev.*, **30**, 3769-3790, doi.org/10.1016/j.quascirev.2011.09.018, 2011.
- 733
- 734 Oksanen, J., Simpson, G. L., Blanchet, F. G., Kindt, R., Legendre, P., Minchin, P. R., O'Hara, R. B., Solymos, P., Stevens,
735 M. H. H., Szoecs, E., Wagner, H., Barbour, M., Bedward, M., Bolker, B., Borcard, D., Carvalho, G., Chirico, M., Caceres,
736 M. D., Durand, S., Evangelista, H. B. A., FitzJohn, R., Friendly, M., Furneaux, B., Hannigan, G., Hill, M. O., Lahti, L.,
737 McGlinn, D., Ouellette, M.-H., Cunha, E. R., Smith, T., Stier, A., Braak, C. J. F. T., and Weedon, J.: vegan: Community
738 Ecology Package, 2022.
- 739
- 740 Ourisson, G and Albrecht, P.: Hopanoids. 1. Geohopanoids: the most abundant natural products on Earth?, *Accounts*
741 *Chem. Res.*, **25**, 398-402, doi.org/10.1021/ar00021a003, 1992.
- 742
- 743 Piper, D.Z and Calvert, S.E.: Holocene and late glacial palaeoceanography and palaeolimnology of the Black Sea:
744 Changing sediment provenance and basin hydrography over the past 20,000 years, *Geochim. Cosmochim. Ac.*, **75**,
745 5597-5624, doi.org/10.1016/j.gca.2011.07.016, 2011.
- 746
- 747 R Core Team.: R: A Language and Environment for Statistical Computing, Vienna, Austria, [https://www.R-](https://www.R-project.org/)
748 [project.org/](https://www.R-project.org/), 2023.
- 749
- 750 Rashby, S. E., Sessions, A. L., Summons, R. E and Newman, D. K.: Biosynthesis of 2-methylbacteriohopanepolyols by
751 an anoxygenic phototroph, *Proc. Nat. Acad. Sci. USA.*, **104**, 15099–15104, doi.org/10.1073/pnas.0704912104, 2007.
- 752
- 753 Reeburgh, W. S., Ward, B. B., Whalen, S. C., Sandbeck, K. A., Kilpatrick, K. A and Kerkhof, L. J.: Black Sea methane
754 geochemistry, *Deep-Sea Res. Pt. 1.*, **38**, S1189–S1210, doi.org/10.1016/S0198-0149(10)80030-5, 1991.
- 755
- 756 Reimer, P.J., Austin, W.E.N., Bard, E., Bayliss, A., Blackwell, P.G., Bronk Ramsey, C., Butzin, M., Cheng, H., Edwards,
757 R.L., Friedrich, N., Grootes, P.M., Guilderson, T.P., Hajdas, I., Heaton, T.J., Hogg, A.G., Hughen, K.A., Kromer, B.,
758 Manning, S.W., Muscheler, R., Palmer, J.G., Pearson, C., van der Plicht, J., Reimer, R.W., Richards, D.A., Scott, E.M.,
759 Southon, J.R., Turney, C.S.M., Wacker, L., Adolphi, F., Büntgen, U., Capano, M., Fahrni, S.M., Fogtmann-Schulz, A.,
760 Friedrich, R., Köhler, P., Kudsk, S., Miyake, F., Olsen, J., Reinig, F., Sakamoto, M., Sookdeo, A and Talamo, S.: The
761 IntCal20 Northern Hemisphere Radiocarbon Age Calibration Curve (0–55 cal kBP), *Radiocarbon*, **62**, 725–757,
762 doi.org/10.1017/RDC.2020.41, 2020.
- 763



- 764 Rethemeyer, J., Schubotz, F., Talbot, H.M., Cooke, M.P., Hinrichs, K.-U., Mollenhauer, G.: Distribution of polar
765 membrane lipids in permafrost soils and sediments of a small high Arctic catchment, *Org. Geochem.*, **41**, 1130–1145,
766 doi.org/10.1016/j.orggeochem.2010.06.004, 2010.
- 767
- 768 Repeta, D.J.: A high resolution historical record of Holocene anoxygenic primary production in the Black Sea,
769 *Geochim. Cosmochim. Ac.*, **57**, 4337–4342, doi.org/10.1016/0016-7037(93)90334-S, 1993.
- 770
- 771 Řezanka, T., Siristova, L., Melzoch, K and Sigler, K.: N-Acylated Bacteriohopanehexol-Mannosamides from the
772 Thermophilic Bacterium *Alicyclobacillus acidoterrestris*, *Lipids*, **46**, 249–261, doi.org/10.1007/s11745-010-3482-4,
773 2011.
- 774
- 775 Ricci, J. N., Coleman, M. L., Welander, P. V., Sessions, A. L., Summons, R. E., Spear, J. R., and Newman, D. K.: Diverse
776 capacity for 2-methylhopanoid production correlates with a specific ecological niche, *ISME J.*, **8**, 675–684,
777 doi.org/10.1038/ismej.2013.191, 2014.
- 778
- 779 Richter, N., Hopmans, E.C., Mitrovic, D., Raposeiro, O.M., Gonçalves, P., Costa, A.C., Amaral-Zettler, L.A., Villanueva,
780 L and Rush, D.: Distributions of bacteriohopanepolyols in lakes and coastal lagoons of the Azores Archipelago,
781 *Biogeosciences*, **20**, 2065–2098, doi.org/10.5194/bg-20-2065-2023, 2023.
- 782
- 783 Rohmer, M., Bouvier-Nave, P and Ourisson, G.: Distribution of Hopanoid Triterpenes in Prokaryotes, *Microbiology*,
784 **130**, 1137–1150, doi.org/10.1099/00221287-130-5-1137, 1984.
- 785
- 786 Rush, D., Sinninghe Damsté, J.S., Poulton, S.W., Thamdrup, B., Garside, A.L., Acuña González, J., Schouten, S., Jetten,
787 M.S.M and Talbot, H.M.: Anaerobic ammonium-oxidising bacteria: A biological source of the bacteriohopanetetrol
788 stereoisomer in marine sediments, *Geochim. Cosmochim. Ac.*, **140**, 50–64, doi.org/10.1016/j.gca.2014.05.014, 2014.
- 789
- 790 Rush, D., Osborne, K.A., Birgel, D., Kappler, A., Hirayama, H., Peckmann, J., Poulton, S.W., Nickel, J.C., Mangelsdorf,
791 K., Kalyuzhnaya, M., Sidgwick, F.R and Talbot, H.M.: The Bacteriohopanepolyol Inventory of Novel Aerobic Methane
792 Oxidising Bacteria Reveals New Biomarker Signatures of Aerobic Methanotrophy in Marine Systems, *PLOS One*, **11**,
793 1–27, doi.org/10.1371/journal.pone.0165635, 2016.
- 794
- 795 Rush, D., Talbot, H.M., van der Meer, M.T.J., Hopmans, E.C., Douglas, B and Sinninghe Damsté, J.S.: Biomarker
796 evidence for the occurrence of anaerobic ammonium oxidation in the eastern Mediterranean Sea during Quaternary
797 and Pliocene sapropel formation, *Biogeosciences*, **16**, 2467–2479, doi.org/10.5194/bg-16-2467-2019, 2019.



- 798 Schaeffer, P., Schmitt, G., Adam, P., and Rohmer, M.: Acid-catalyzed formation of 32,35-
799 anhydrobacteriohopanetetrol from bacteriohopanetetrol, *Org. Geochem.*, **39**, 1479–1482,
800 doi.org/10.1016/j.orggeochem.2008.07.007, 2008.
- 801
- 802 Schaeffer, P., Schmitt, G., Adam, P., and Rohmer, M.: Abiotic formation of 32,35-anhydrobacteriohopanetetrol: A
803 geomimetic approach, *Org. Geochem.*, **41**, 1005–1008, doi.org/10.1016/j.orggeochem.2010.04.013.
- 804
- 805 Schrader, H.-J. 1979. Quaternary Paleoclimatology of the Black Sea basin, *Sedimentary Geol.*, **23**, 165-180,
806 doi.org/10.1016/0037-0738(79)90013-7, 2010.
- 807
- 808 Schouten, S., Wakeham, S. G and Damsté, J. S. S.: Evidence for anaerobic methane oxidation by archaea in euxinic
809 waters of the Black Sea, *Org. Geochem.*, **32**, 1277–1281, doi.org/10.1016/S0146-6380(01)00110-3, 2001.
- 810
- 811 Schubert, C. J., Coolen, M. J. L., Neretin, L. N., Schippers, A., Abbas, B., Durisch-Kaiser, E., Wehrli, B., Hopmans, E. C.,
812 Damste, J. S. S., Wakeham, S., and Kuypers, M. M. M.: Aerobic and anaerobic methanotrophs in the Black Sea water
813 column, *Environ. Microbiol.*, **8**, 1844–1856, doi.org/10.1111/j.1462-2920.2006.01079.x, 2006.
- 814
- 815 Schubotz, F., Wakeham, S. G., Lipp, J. S., Fredricks, H. F., and Hinrichs, K.: Detection of microbial biomass by intact
816 polar membrane lipid analysis in the water column and surface sediments of the Black Sea, *Environ. Microbiol.*, **11**,
817 2720–2734, doi.org/10.1111/j.1462-2920.2009.01999.x, 2009.
- 818
- 819 Schwartz-Narbonne, N., Schaeffer, P., Hopmans, E.C., Schenese, M., Charlton, E.A., Jones, D.M., Sinninghe Damsté,
820 J.S., Farhan, M., Haque, U., Jetten, M.S.M., Lengger, S.K., Murrell, J.C., Normand, P., Nuijten, G.H.L., Talbot, H.M and
821 Rush, D.: A unique bacteriohopanetetrol stereoisomer of marine anammox, *Org. Geochem.*, **143**, 1-10,
822 doi.org/10.1016/j.orggeochem.2020.103994, 2020.
- 823
- 824 Seemann, M., Bissert, P., Tritz, J.-P., Hooper, A. B and Rohmer, M.: Novel bacterial triterpenoids of the hopane
825 series from *Nitrosomonas europaea* and their significance for the formation of the C35 bacteriohopane skeleton,
826 *Tetrahedron Lett.*, **40**, 1681–1684, doi.org/10.1016/S0040-4039(99)00064-7, 1999.
- 827
- 828 Shatz, M., Yosief, T and Kashman, Y.: Bacteriohopanehexol, a New Triterpene from the Marine Sponge *Petrosia*
829 *Species*, *J. Nat. Prod.*, **63**, 1554–1556, doi.org/10.1021/np000190r, 2000.
- 830
- 831 Shumilovskikh, L.S., Tarasov, P., Arz, H.W., Fleitmann, D., Marret, F., Nowaczyk, N., Plessen, B., Schlütz, F and Behling,
832 H.: Vegetation and environmental dynamics in the southern Black Sea region since 18 kyr BP derived from the marine



- 833 core 22-GC3, *Palaeogeogr. Palaeoclimatol. Palaeoecol.*, **337–338**, 177–193, doi.org/10.1016/j.palaeo.2012.04.015,
834 2012.
- 835
- 836 Sinninghe Damste, J.S., Wakeham, S.G., Kohnen, M.E.L., Hayes, J.M., de Leeuw, J.W.: A 6,000–year sedimentary
837 molecular record of chemocline excursions in the Black Sea, *Nature*, **362**, 827–829, doi.org/10.1038/362827a0,
838 1993.
- 839
- 840 Sinninghe Damsté, J. S., Rijpstra, W. I. C., Dedysh, S. N., Foesel, B. U and Villanueva, L.: Pheno- and Genotyping of
841 Hopanoid Production in Acidobacteria, *Front. Microbiol.*, **8**, 1–20, doi.org/10.3389/fmicb.2017.00968, 2017.
- 842
- 843 Sinninghe Damsté, J.S., Schouten, S., Hopmans, E.C., van Duin, A.C.T and Geenevasen, A.J.A.: Crenarchaeol, *J. Lipid*
844 *Res.*, **43**, 1641–1651, doi:10.1194/jlr.M200148-JLR200, 2002.
- 845
- 846 Sollai, M. Villanueva, L., Hopmans, E.C., Reichart, G.-J and Sinninghe Damsté, J.S.: A combined lipidomic and 16S
847 rRNA gene amplicon sequencing approach reveals archaeal sources of intact polar lipids in the stratified Black Sea
848 water column, *Geobiology*, **17**, 91–109, doi.org/10.1111/gbi.12316, 2019.
- 849
- 850 Spencer-Jones, C.L., Wagner, T and Talbot, H.M.: A record of aerobic methane oxidation in tropical Africa over the
851 last 2.5 Ma, *Org. Geochem.*, **89–90**, 1–13, doi.org/10.1016/j.gca.2017.08.042, 2017.
- 852
- 853 Summons, R. E., Jahnke, L. L., Hope, J. M and Logan, G. A.: 2-Methylhopanoids as biomarkers for cyanobacterial
854 oxygenic photosynthesis, *Nature*, **400**, 554–557, doi.org/10.1038/23005, 1999.
- 855
- 856 Talbot, H. M., Watson, D. F., Murrell, J. C., Carter, J. F and Farrimond, P.: Analysis of intact bacteriohopanepolyols
857 from methanotrophic bacteria by reversed-phase high-performance liquid chromatography–atmospheric pressure
858 chemical ionisation mass spectrometry, *J. Chromatogr. A.*, **921**, 175–185, doi.org/10.1016/S0021-9673(01)00871-8,
859 2001.
- 860
- 861 Talbot, H. M., Farrimond, P., Schaeffer, P., and Pancost, R. D.: Bacteriohopanepolyols in hydrothermal vent biogenic
862 silicates, *Org. Geochem.*, **36**, 663–672, doi.org/10.1016/j.orggeochem.2004.10.015, 2005.
- 863
- 864 Talbot, H.M and Farrimond, P.: Bacterial populations recorded in diverse sedimentary biohopanoid distributions,
865 *Org. Geochem.*, **38**, 1212–1225, <https://doi.org/10.1016/j.orggeochem.2007.04.006>, 2007.
- 866



- 867 Talbot, H. M., Summons, R. E., Jahnke, L. L., Cockell, C. S., Rohmer, M., and Farrimond, P.: Cyanobacterial
868 bacteriohopanepolyol signatures from cultures and natural environmental settings, *Org. Geochem.*, 39, 232–263,
869 doi.org/10.1016/j.orggeochem.2007.08.006, 2008.
- 870
- 871 Talbot, H.M., Handley, L., Spencer-Jones, C.L., Dinga, B.J., Schefuß, E., Mann, P.J., Poulsen, J.R., Spencer, R.G.M.,
872 Wabakanghanzi, J.N and Wagner, T.: Variability in aerobic methane oxidation over the past 1.2 Myrs recorded in
873 microbial biomarker signatures from Congo fan sediments, *Geochim. Cosmochim. Ac.*, 133, 387–401,
874 doi.org/10.1016/j.gca.2014.02.035, 2014.
- 875
- 876 Talbot, H.M., Sidgwick, F.R., Bischoff, J., Osborne, K.A., Rush, D., Sherry, A and Spencer-Jones, C.L.: Analysis of non-
877 derivatised bacteriohopanepolyols by ultrahigh-performance liquid chromatography/tandem mass spectrometry.
878 *Rapid Commun. Mass Sp.*, **30**, 2087–2098, doi.org/10.1002/rcm.7696, 2016a.
- 879
- 880 Talbot, H.M., McClymont, E.L., Inglis, G.N., Evershed, R.P and Pancost, R.D.: Origin and preservation of
881 bacteriohopanepolyol signatures in Sphagnum peat from Bissendorfer Moor (Germany), *Org. Geochem.*, **97**, 95–110,
882 doi.org/10.1016/j.orggeochem.2016.04.011, 2016b.
- 883
- 884 Uemura, H. and Ishiwatari, R.: Identification of unusual 17 β (H)-moret-22(29)-ene in lake sediments, *Org. Geochem.*,
885 **23**, 675–680, doi.org/10.1016/0146-6380(95)00036-E, 1995.
- 886
- 887 van Kemenade, Z.R., Cutmore, A., Hennekam, R., Hopmans, E.C., van der Meer, M.T.J., Mojtahid, M., Jorissen, F.J.,
888 Bale, N.J., Reichart, G.-J., Sinninghe Damsté, J.S and Rush, D.: Marine nitrogen cycling dynamics under altering redox
889 conditions: insights from deposition of sapropels S1 and the ambiguous S2 in the Eastern Mediterranean Sea,
890 *Geochim. Cosmochim. Ac.*, **354**, doi.org/10.1016/j.gca.2023.06.018, 2023.
- 891
- 892 van Winden, J.F., Talbot, H.M., Kip, N., Reichart, J.-G., Pol, A., McNamara, N.P., Jetten, M.S.M., Op den Camp, H.J.M
893 and Sinninghe Damsté, J.S.: Bacteriohopanepolyol signatures as markers for methanotrophic bacteria in peat moss,
894 *Geochim. Cosmochim. Ac.*, **77**, 52–61, doi.org/10.1016/j.gca.2011.10.026, 2012.
- 895
- 896 Verleye, T.J., Mertens, K.N., Louwye, S and Arz, H.W.: Holocene salinity changes in the southwestern black sea: A
897 reconstruction based on dinoflagellate cysts, *Palynology*, **33**, 77–100, doi:10.1080/01916122.2009.9989666, 2009.
- 898
- 899 Wakeham, S. G. and Beier, J. A.: Fatty acid and sterol biomarkers as indicators of particulate matter source and
900 alteration processes in the Black Sea, *Deep-Sea Res. Pt. 1.*, **38**, S943–S968, doi.org/10.1016/S0198-0149(10)80018-
901 4, 1991.



- 902 Wakeham, S. G., Beier, J. A., and Clifford, C. H.: Organic matter sources in the Black Sea as inferred from hydrocarbon
903 distributions, *in: Black Sea Oceanography*, edited by: İzdar, E., and Murray, J.W., Dordrecht: Springer Netherlands,
904 319-341, https://doi.org/10.1007/978-94-011-2608-3_20, 1991.
- 905 Wakeham, S. G., Lewis, C. M., Hopmans, E. C., Schouten, S., and Damsté, J. S. S.: Archaea mediate anaerobic oxidation
906 of methane in deep euxinic waters of the Black Sea, *Geochim. Cosmochim. Ac.*, **67**, 1359-1374,
907 [doi.org/10.1016/S0016-7037\(02\)01220-6](https://doi.org/10.1016/S0016-7037(02)01220-6), 2003.
- 908
- 909 Wakeham, S.G., Amann, R., Freeman, K.H., Hopmans, E.C., Barker Jørgensen, B., Putnam, I.F., Schouten, S., Sinninghe
910 Damsté, J.S., Talbot, H.M and Woebken, D.: Microbial ecology of the stratified water column of the Black Sea as
911 revealed by a comprehensive biomarker study, *Org. Geochem.*, **38**, 2070-2097,
912 doi.org/10.1016/j.orggeochem.2007.08.003, 2007.
- 913
- 914 Welander, P. V., Coleman, M. L., Sessions, A. L., Summons, R. E., and Newman, D. K.: Identification of a methylase
915 required for 2-methylhopanoid production and implications for the interpretation of sedimentary hopanes, *Proc.*
916 *Nat. Acad. Sci. USA.*, **107**, 8537–8542, doi.org/10.1073/pnas.0912949107, 2010.
- 917
- 918 Welte, C.U., Rasigraf, O., Vaksmaa, A., Versantvoort, W., Arshad, A., Op den Camp, H.J.M., Jetten, M.S.M., Lücke, C.
919 and Reimann, J.: Nitrate- and nitrite-dependent anaerobic oxidation of methane, *Environ. Microbiol. Rep.*, **8**, 941-
920 955, doi.org/10.1111/1758-2229.12487, 2016.
- 921
- 922 Wickham, H and Chang, W.: Package 'ggplot2': Create elegant data visualisations using the grammar of graphics. R-
923 package Version 3.4.0, <https://ggplot2.tidyverse.org/reference/ggplot2-package.html>, 2016.
- 924
- 925 Wuchter, C., Abbas, B., Coolen, M.J.L., Herfort, L., van Bleijswijk, J., Timmers, P., Strous, M., Teira, E., Herndl, G.J.,
926 Middelburg, J.J., Schouten, S and Sinninghe Damsté, J.S.: Archaeal nitrification in the ocean, *Proc. Nat. Acad. Sci.*
927 *USA.*, **103**, 12317-12322, doi.org/10.1073/pnas.0600756103, 2006.
- 928
- 929 Xu, Y., Cooke, M.P., Talbot, H.M and Simpson, M.J.: Bacteriohopanepolyol signatures of bacterial populations in
930 Western Canadian soils, *Org. Geochem.*, **40**, 79-86, doi.org/10.1016/j.orggeochem.2008.09.003, 2009.
- 931
- 932 Yanchilina, A. G., Ryan, W. B. F., Kenna, T. C., and McManus, J. F.: Meltwater floods into the Black and Caspian Seas
933 during Heinrich Stadial 1, *Earth-Sci. Rev.*, **198**, 102931, doi.org/10.1016/j.earscirev.2019.102931, 2019.
- 934
- 935 Zindorf, M., Rush, D., Jaeger, J., Mix, A., Penkrot, M.L., Schnetgere, B., Sidgwick, F.R., Talbot, H.M., van der Land, C.,
936 Wagner T., Walczak, M and März, C.: Reconstructing oxygen deficiency in the glacial Gulf of Alaska: Combining



937 biomarkers and trace metals as paleo-redox proxies, *Chem. Geol.*, **558**, 1-11,
938 doi.org/10.1016/j.chemgeo.2020.119864, 2020.

939

940 Zhao, N., Berova, N., Nakanishi, K., Rohmer, M., Mougnot, P., and Jürgens, U. J.: Structures of two
941 bacteriohopanoids with acyclic pentol side-chains from the cyanobacterium *Nostoc* PCC 6720, *Tetrahedron*, **52**,
942 2777–2788, doi.org/10.1016/0040-4020(96)00013-0, 1996.

943

944 Zhu, Q.-Z., Elvert, M., Meador, T. B., Schröder, J. M., Doeana, K. D., Becker, K. W., Elling, F. J., Lipp, J. S., Heuer, V. B.,
945 Zabel, M and Hinrichs, K.-U.: Comprehensive molecular-isotopic characterization of archaeal lipids in the Black Sea
946 water column and underlying sediments, *Geobiology*, **22**, e12589, doi.org/10.1111/gbi.12589, 2024.

947

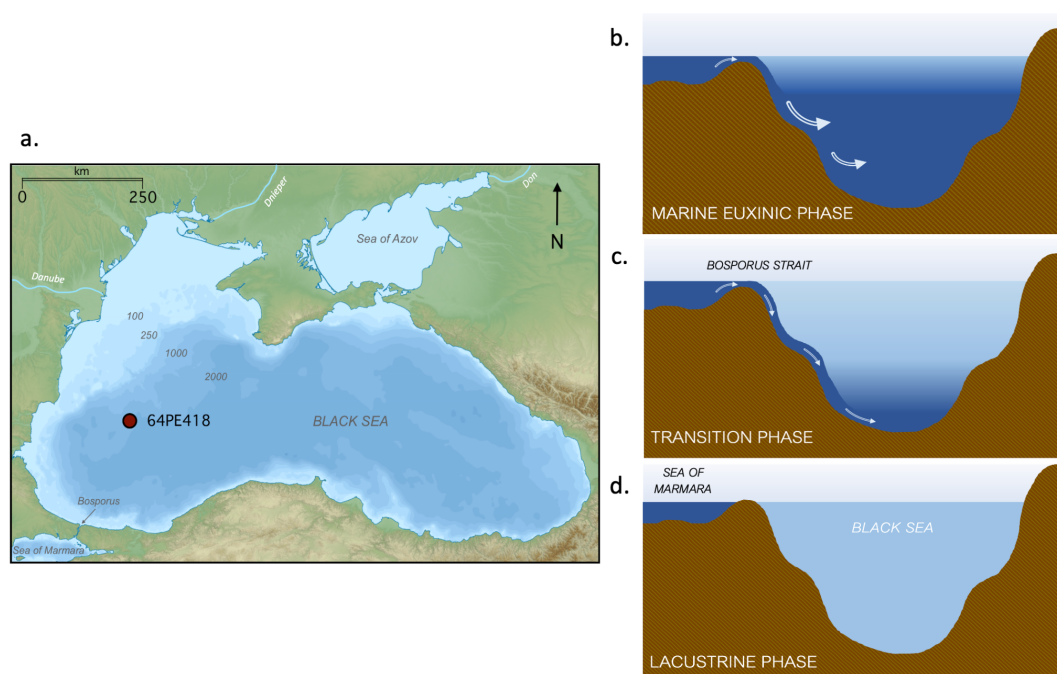
948 Zhu, C., Talbot, H. M., Wagner, T., Pan, J.-M., and Pancost, R. D.: Distribution of hopanoids along a land to sea
949 transect: Implications for microbial ecology and the use of hopanoids in environmental studies, *Limnol. Oceanog.*,
950 **56**, 1850–1865, doi.org/10.4319/lo.2011.56.5.1850, 2011.

951

952

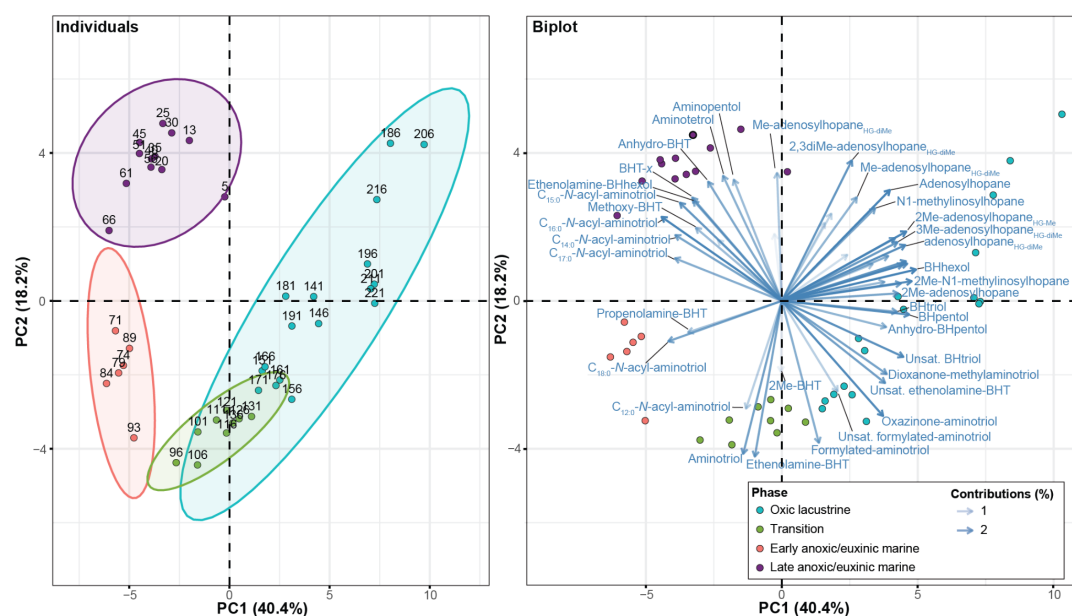
953 Figures

954



955

960



967

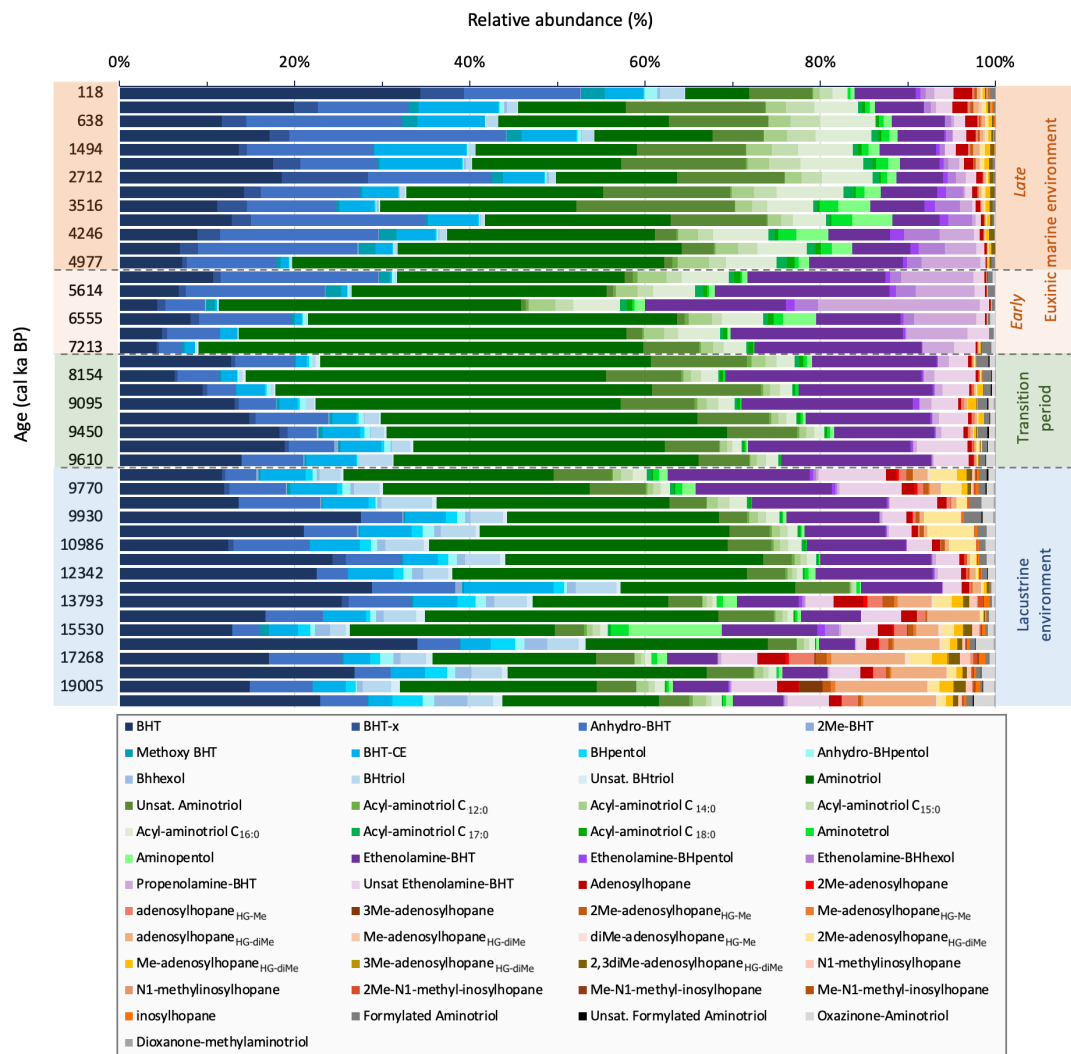


Figure 3: Relative abundance of peak areas (%) of all BHPs identified in core 64PE418 over the last 19.5 ka. Changes in abundance of each BHP (peak area per g TOC) over time is shown in the supplementary material.

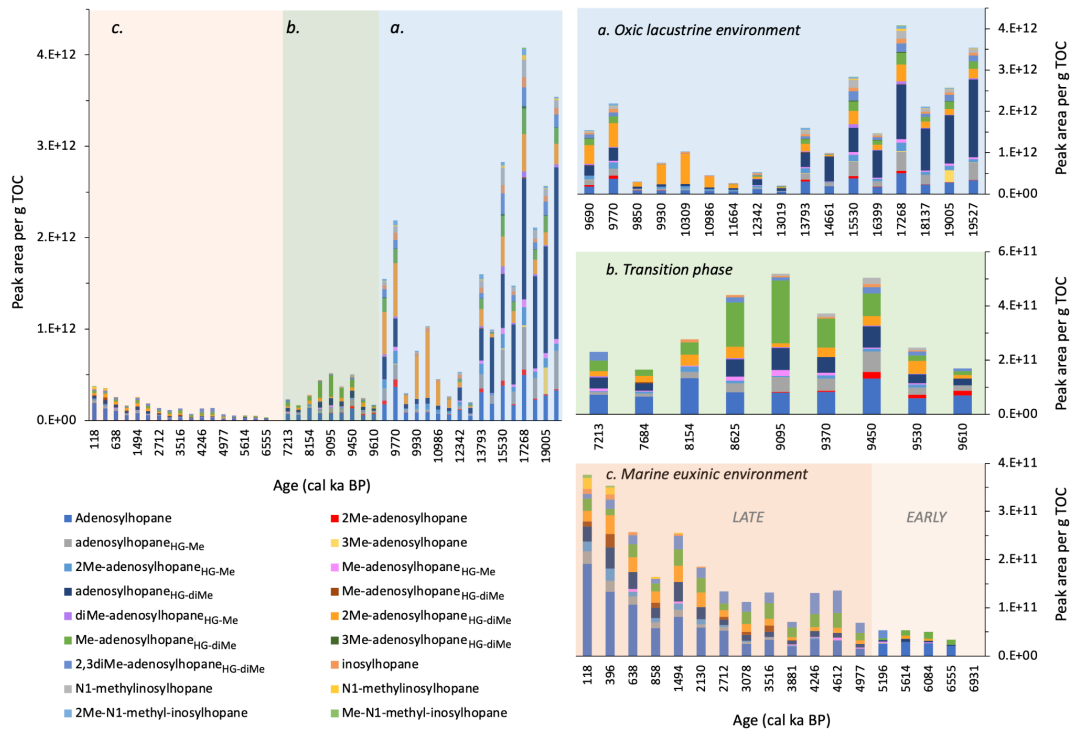
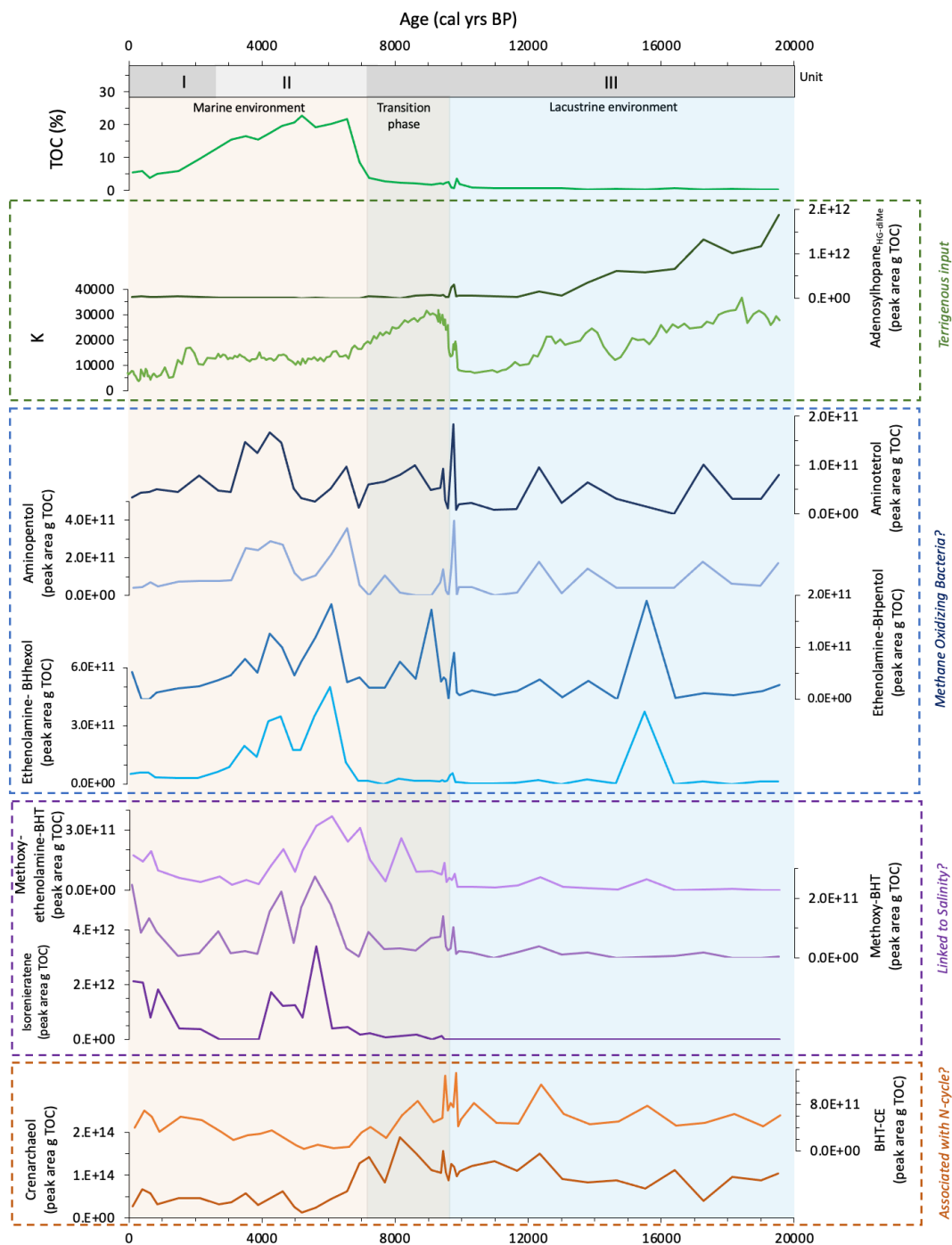


Figure 4: Changes in the peak area per g TOC of Nu-BHPs in Black Sea core 64PE418 during: **a)** the oxic lacustrine phase; **b)** the transition phase; and **c)** the euxinic marine phase. Note the different scales on the y-axes.



980
981 **Figure 5:** Changes in the peak area per g TOC of a number of potential diagnostic BHPs.