

Supplementary material for

## **Methane ebullition as the dominant pathway for carbon sea-air exchange in coastal, shallow water habitats of the Baltic Sea**

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### **Text S1. Extended methods: Habitat classification using HELCOM HUB**

The HELCOM HUB structure is hierarchical, with the first level “A” representing the Baltic Sea, and the second level, also designated as “A”, is for the photic benthic zone. In level three the habitats are classified based on their bottom substrate. If 90 % of the habitat was covered by a single substrate type, it was assigned that specific classification. Otherwise, it was classified as ‘mixed substrate’. To determine substrate type, a visual assessment was conducted, and for areas deemed to have more than 90 % coverage of a specific soft substrate, surface sediment samples were collected and analysed using a sediment sieve. Based on this classification, the seven sampling locations were grouped into four substrate categories: (1) Coarse sediment, in which >30 % of the sediment had a grain size between 2 and 63 mm (AA.I), (2) muddy sediment, if >20 % was less than 63 µm (AA.H), (3) sand, if none of these two criteria were met and grain sizes between 0.063 and 2 mm dominated (AA.J), and (4) mixed substrate (AA.M).

At level four in HELCOM HUB the percentage of vegetation cover is described. All sampling locations in this study were categorized as having either >10 % coverage of macroscopic epibenthic biotic structures, or sparse epibenthic macrocommunities, between 0 – 10 % coverage. Locations with sparse coverage were not further subdivided at this level. Level five describes the type of vegetation community, with classifications determined visually as emergent vegetation, submerged rooted plants, or perennial algae, depending on the dominant group. At level six, the taxa are to be identified, with species contributing over 50 % of the biomass volume within the dominant vegetation community determining the classification. However, in this study, only locations with submerged vegetation where the dominant taxa could be visually determined were classified to this level.

Ultimately, the seven sampling locations were divided into five distinct habitats based on substrate and vegetation cover: Coarse sediment with perennial algae cover (AA.III), sand with sparse epibenthic macrocommunity (AA.J2T), sand with submerged rooted plants (AA.J1C), mixed substrate with submerged rooted plants (AA.M1C) and muddy sediment with emerged plants *Phragmites australis* (AA.H1A1).

## **Text S2. Extended methods: Extrapolation into yearly fluxes**

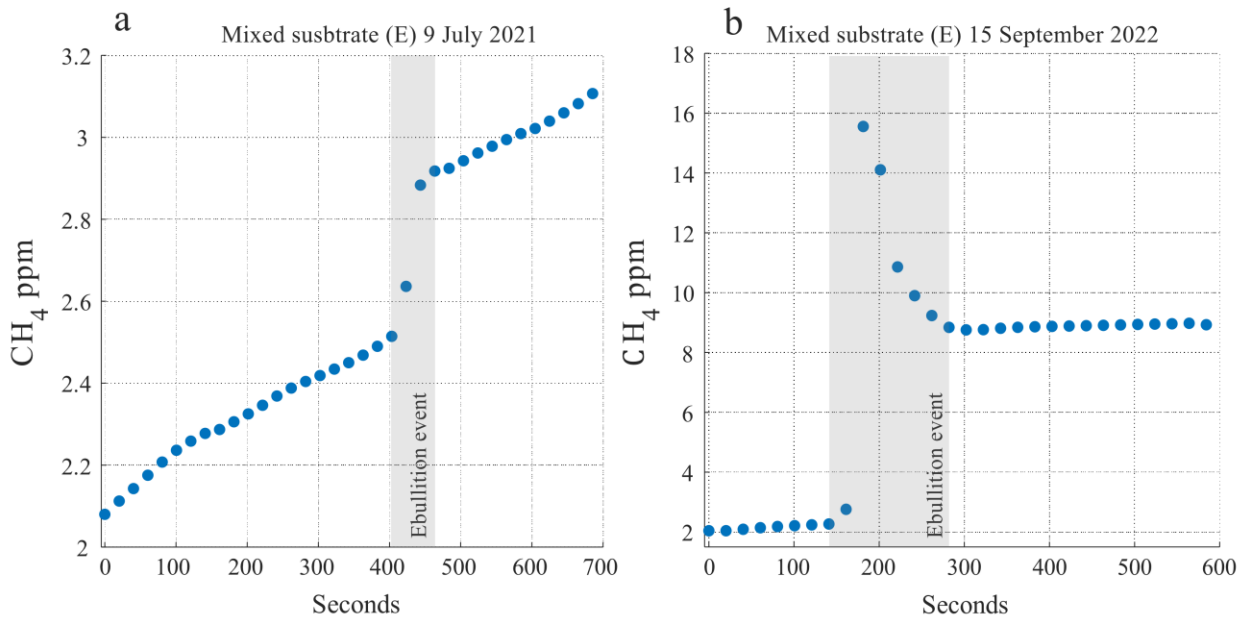
Daily diffusive fluxes were converted to annual fluxes by averaging the daily fluxes from each sampling period to determine monthly fluxes for the month(s) that the sampling period represented. The representativeness of the sampling dates to the month in question was examined by comparison of the locally measured air temperatures and wind speeds with nearby long-term weather observations from the Swedish Meteorological and Hydrological Institute (SMHI) for the years 2020 through 2022 at Skarpö, Gustavsberg and Trosa. Fluxes from unsampled months were linearly extrapolated from adjacent sampled months, except August, which had similar SMHI wind and temperature data as July and was assumed to have the same flux. The monthly average ebullition flux was extrapolated in a similar way as the diffusive fluxes and was assumed to be zero the months where it was not measured. Monthly diffusive and ebullition fluxes were summed to calculate the annual flux at each location.

The months, or parts of the month, where ice cover prevented sampling, were assumed to have zero flux. The mixed substrate habitat D was ice-covered in January and February, and reed habitat F and submerged plant-covered habitat C were ice-covered in January, February, and half of March. Mixed substrate habitat E and the macroalgae habitat A were ice-covered during the first half of January and the sand habitat B was ice-covered for the entire January and for half of February. During these periods, these sampling locations could not be sampled and in the yearly calculations they have been counted as zero fluxes. The channel in reed bed G was never fully ice-covered due to flowing water.

This simple extrapolation method was chosen over a more complex approach using drivers such as temperature or wind speed due to the generally low and heterogeneous relationship between the fluxes and temperature and/or wind (Table S1).

**Table S1.** R-square values for linear least-squared relationships between CH<sub>4</sub> or CO<sub>2</sub> and the environmental parameters wind speed ( $U_{10}$ ), water temperature, water depth and distance to shore. For relationships between  $U_{10}$  and CO<sub>2</sub>, absolute CO<sub>2</sub> values have been used. Bold R-squares indicate statistically significant ( $p < 0.05$ ) relationships. Negative relationships are marked with (-).

Habitats	Locations	$U_{10}$		Water temp		Water depth		Distance to shore		Salinity	
		CH <sub>4</sub>	CO <sub>2</sub>	CH <sub>4</sub>	CO <sub>2</sub>	CH <sub>4</sub>	CO <sub>2</sub>	CH <sub>4</sub>	CO <sub>2</sub>	CH <sub>4</sub>	CO <sub>2</sub>
All		<b>0.02</b>	<b>0.03</b>	<b>0.13</b>	<b>0.03 (-)</b>	0.00	0.00	0.01	<b>0.04 (-)</b>	0.00	<b>0.18 (-)</b>
Coarse sed. - Macroalgae	A	<b>0.60</b>	0.09	<b>0.13</b>	<b>0.33 (-)</b>	0.01	0.00	0.03	0.06	0.01	0.00
Sand – sparse veg.	B	<b>0.34</b>	<b>0.33</b>	<b>0.08</b>	<b>0.45 (-)</b>	0.04	0.04	0.00	0.02	0.00	0.04
Sand – subm. plants	C	<b>0.37</b>	<b>0.23</b>	<b>0.16</b>	0.02	0.00	0.03	0.01	0.06	<b>0.13 (-)</b>	<b>0.15</b>
Mixed sub. – subm. plants	D	0.02	0.00	<b>0.22</b>	0.01	<b>0.04</b>	0.03	0.08	0.00	0.06	0.00
	E	<b>0.26</b>	0.06	0.01	<b>0.17 (-)</b>	0.02	0.08	0.01	0.03	<b>0.16 (-)</b>	0.07
Muddy sed. - Reeds	F	0.03	0.02	<b>0.37</b>	0.04	0.00	0.03	0.04	0.04	0.05	<b>0.21 (-)</b>
	G	0.01	<b>0.14</b>	<b>0.13</b>	0.01	0.05	<b>0.13</b>	0.00	0.01	0.02	0.05



**Figure S1.** Raw data from the GGA showing two (A and B) ebullition events, both captured in sampling location E. The blue dots indicate how the concentration of  $\text{CH}_4$  in the chamber system changes as a function of seconds elapsed since the start of the incubation. The abrupt changes marked in grey are interpreted as ebullition events and indicate the start and end time of the ebullition used for calculating the flux.