

Benthic ostracod diversity and biogeography in an urban semi-enclosed eutrophic riverine bay

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Abstract. The benthic ecosystem has been greatly altered by environmental pressure over the past several decades. Compared to some well-studied large bays, the situation in populated small bay areas is still under investigation. In this study, we investigated the abundance, diversity, composition, and distribution of ostracod (a meiobenthic group) and their interactions with eutrophication and pollution through high resolution sampling of surface sediment in Deep Bay, a small semi-enclosed riverine bay adjacent to two of the world's most populated cities. We found that ostracod abundance and diversity exhibited an inner-outer gradient in Deep Bay, shaped by eutrophication and pollution from human activities. Faunal composition was also characterized by inner-outer difference, as well as difference between Hong Kong and Shenzhen areas, where Hong Kong was more influenced by eutrophication, and Shenzhen was more affected by metal pollution. Ostracod distribution and their environmental preference in Deep Bay were consistent with broader studies in Hong Kong, strongly supporting the idea that ostracod is a useful bioindicator of coastal benthic ecosystems. Our study emphasizes the importance of studying an uncomplicated system with finer sampling, as this approach offers a clear and direct understanding of how benthic ecosystems are shaped by distinct coastal environmental problems.

1 Introduction

Coastal bays that are highly influenced by adjacent populated cities are facing incremental environmental problems and ecosystem degradation due to poor water exchange and intensive human activities (Dai et al., 2023; Nichols et al., 2019; Breitburg et al., 2018). River discharge, aquaculture and maritime activities cause eutrophication and pollution of coastal

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40 areas, ultimately leading to altered marine biological diversity (Glibert, 2017). Deep Bay is a semi-closed bay influenced by several small rivers inside the bay and connected with the Pearl River Estuary outside the bay. The bay is surrounded by Hong Kong and Shenzhen, two of the world's largest urbanized cities with 17.8 and 7.5 million people, respectively. It is also a well-known region for both oyster culture and hinterland mangrove ecosystems (Mohyuddin et al., 2023; Feng et al., 2020). Under the influence of nutrient and pollution inputs from rivers, the aquaculture industry, and large-scale reclamation projects, combined with limited water exchange, Deep Bay is currently experiencing increasing environmental degradation (Xu et al., 2019).

Benthic ecosystem alteration has been recognized as a main consequence of environmental degradation (Dorgham, 2014). Ostracoda (Crustacea), which are abundantly fossilized in marginal-marine sediments, have been used as a useful metazoan group bioindicator (Yasuhara et al., 2007). Previous studies in Hong Kong coast have suggested that ostracods can be used as a bioindicator for the diversity and biogeography of broader soft sediment benthos (Hong et al., 2022; Mamo et al., 2023). Statistical relationships between ostracod and environmental factors have been indicated (Hong et al., 2019). On this topic, some large, populated bays including Chesapeake Bay (Cronin and Vann, 2003), Osaka Bay (Yasuhara et al., 2007) and the Baltic Sea (Frenzel et al., 2010) were well studied; small bays with less population were also investigated before (e.g., Irizuki et al., 2014). However, semi-enclosed, intensively human-influenced small bay areas remain under-studied, especially with high resolution sampling.

Previous studies in Deep Bay have primarily focused on sediment chemistry (Lee, 2000), pollutants (Zhang et al., 2021; Li et al., 2024), land reclamation (Ren et al., 2011) and mangrove restoration (Feng et al., 2020). Some ecological studies were conducted solely in mangrove reserves (Zan et al., 2003). And due to the boundary control of the sea area, sampling has typically been restricted to one side (either Hong Kong area or Shenzhen area only). As a result, detailed biodiversity and biogeographic distributions for marine organisms in the broader area of Deep Bay are largely unknown.

In our study, we provided a comprehensive understanding on benthic community distribution and related environmental factors in Deep Bay, a small semi-enclosed bay adjacent to two of the world's largest cities. We investigated the geographical structure of ostracod abundance, diversity and composition, and how they interacted with eutrophication and pollution through doing high resolution sampling of surface sediments on both Hong Kong and Shenzhen areas of the bay.

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2 Materials and methods

2.1 Site description

Deep Bay (22.41–22.53° N, 113.88–114.00° E) is located between the Shenzhen Special Economic-Zone of China and Hong Kong Special Administrative Region in the eastern Pearl River Estuary (Fig. 1). At 17 km long and 4–10 km wide, it is a semi-enclosed bay with limited water exchange. The main rivers flowing into the bay include the Shenzhen River and Shan Pui River in Hong Kong (Fig. 1). The hydrography of the bay is also strongly influenced by the discharge from the Pearl River. Deep Bay features a semidiurnal tidal system, with an annual tidal range of 1.4 m (Feng et al., 2020). At the north-eastern end of the bay are the Futian and Mai Po mangrove reserves (Fig. 1). Along the Hong Kong coast and in the central bay area, oyster farming rafts have expanded over the last several decades (Bowler et al., 1984, Fig. 1). Deep Bay is experiencing rapid urbanization and population growth, accompanied by intensive human activities such as agricultural, municipal and industrial water discharge into the bay (Yang et al., 2021).

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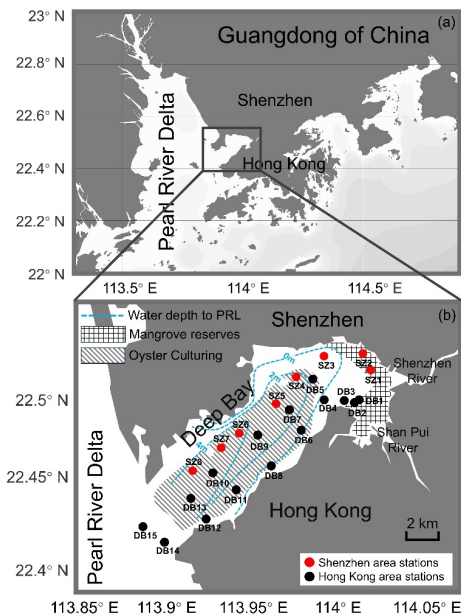
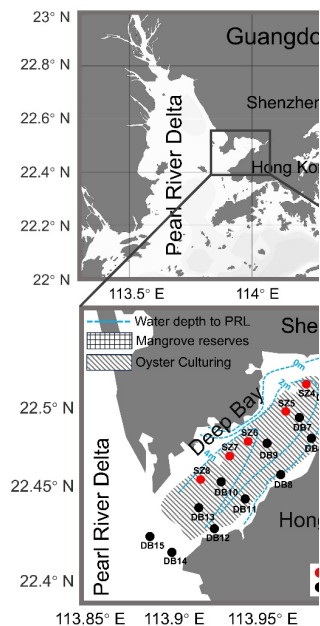


Figure 1. Locality map of Deep Bay and sampling sites. Red dots represent stations in the Shenzhen area, sampled in July 2024. Black dots represent stations in the Hong Kong area, sampled in September 2023. Grid filling represents Futian (north) and Mai Po (south) mangrove reserves (adapted from Feng et al., 2020). Dashed filling represents areas of oyster culturing in Deep Bay (adapted from Google Map). Blue dashed lines represent relative water depth to Pearl River Level (PRL) which reflect local bathymetry (adapted from Ye et al., 2018).

2.2 Materials

2.2.1 Ostracod samples and processing

Surface sediment samples of 100 mL were collected by using a Ekman Grab from twenty three locations that cover the entire Deep Bay area (Fig. 1). Including fifteen sites collected in September, 2023 from the Hong Kong area, eight sites collected in July, 2024 from the Shenzhen area. Several sites, including DB3, DB5, DB9, DB10, DB15, overlapped with the Hong Kong Environmental Protection Department (EPD) monitoring stations DM1 (DS1), DM2, DM3 (DS2), DM4 (DS3), DM5



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(DS4) respectively (DM represents seawater quality monitoring stations and DS represents sediment quality monitoring stations; Environmental Protection Department, 2024).

Sediment samples were washed through a 63 μm sieve and dried at 40 $^{\circ}\text{C}$ in an oven. The dried samples were then sieved using a 150 μm sieve, and ostracod specimens larger than 150 μm were picked out for analysis. All individuals were picked out from the samples if the total number of specimens was less than 200. For samples containing substantially more than 200 specimens, we divided them into fractions using a sample splitter. The sample materials primarily consisted of death assemblages, with some articulated carapaces and a few containing soft parts inside. Each single valve was considered one individual, each carapace was considered two individuals (Yasuhara et al., 2017).

2.2.2 Environmental variables and data compilation

Additional surface sediment samples were collected in the same stations from the same cruise above to examine total organic carbon (TOC), total carbon (TC), total nitrogen (TN), total sulfur (TS) and their isotopes. Sediment chemical oxygen demand and the concentrations of heavy metals, including copper, chromium, barium, cadmium, mercury, and a pollutant total cyanide were obtained from EPD sediment monitoring stations DS1–4 (Environmental Protection Department, 2024). Data on bottom water hypoxia frequency (with threshold of 2 mg/L, 3 mg/L and 4 mg/L respectively), surface nutrient levels of dissolved inorganic nitrogen (DIN) and dissolved inorganic phosphorus (DIP), and physical parameters including bottom temperature and salinity were obtained from EPD seawater monitoring stations DM1–5 (Environmental Protection Department, 2024). As our grab sampler penetrated about 10 cm into the sediments, and the sedimentation rate was approximately 1cm/yr in Deep Bay (Yan et al., 2019), we used recent ten years' average (2012–2022) of the sediment/seawater monitoring result for analysis. We also compiled perfluoroalkyl and polyfluoroalkyl substances (PFAs) and heavy metal data from recent studies (Zhang et al., 2021; Li et al., 2024), selecting sites in proximity to those in our study.

2.3 Methods

2.3.1 Diversity analysis

We used Hill numbers, or the effective numbers of equally abundant species, to estimate ostracod diversity at each station or group of stations. Hill number offers several major advantages and have been increasingly adopted by ecologists (Chao et al., 2020). The order q of Hill numbers represents the sensitivity of the diversity metric to species relative abundance. Specifically, Hill number 0D with order $q=0$ measures species richness; 1D measures the diversity of the abundant species,

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140 and 2D measures the diversity of dominant species (Chao et al., 2014). The Hill numbers 1D and 2D are equivalent to the exponential of Shannon entropy and Simpson diversity index, respectively (Chao et al., 2014).

2.3.2 Multivariate analysis

We applied hierarchical cluster analysis to measure the dissimilarity among ostracod fauna from different sites, using
145 diversity decomposition to obtain dissimilarity measures. We also performed a non-metric Multidimensional Scaling (nMDS) to show faunal similarities among ostracod assemblages in two-dimensional space. We used dissimilarity indices of Sørensen (Sørensen, 1948), Horn (Horn, 1966) and Morisita-Horn (Morisita, 1959), to estimate the effective proportion of unshared species in the ostracod assemblages. While classical Sørensen dissimilarity is presence-absence based, Horn and Morisita-Horn are designed to quantify the compositional dissimilarities of abundant and dominant species. The
150 environmental variables we used for multivariate analysis included TOC, TC, TN, TS in sediment, heavy metals (As, Cd, Cu, Pb, Zn) adapted from Zhang et al (2021), PFAs adapted from Li et al (2024), ten years' average hypoxia frequency, pH, DIN and Chlorophyll-a (*Chla*) calculated from EPD seawater monitoring data (Environmental Protection Department, 2024).

3 Results

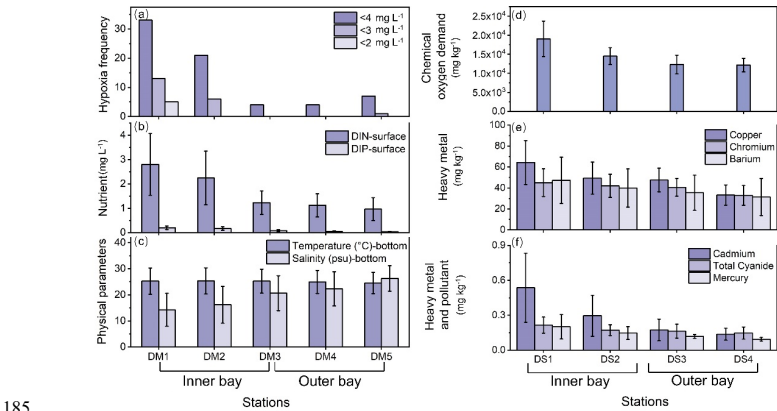
155 3.1 Environmental settings

Deep Bay exhibited a riverine-oceanic salinity gradient from riverine freshwater input in the inner bay to more saline water influenced by the Pearl River Estuary in the outer bay (Fig. 2c). This gradient is also reflected in other related parameters, such as nutrients (Fig. 2b), heavy metal and other pollutants (Fig. 2e, f), and TOC, TN levels (Fig. 3a, c). Hypoxia frequency was much higher in the inner bay than the outer bay, while surface DIN and DIP was higher in inner bay (Fig. 2a, b).
160 Sediment chemical oxygen demand, heavy metal and other pollutants were relatively high in the inner bay and gradually decreased towards the outer bay (Fig. 2d–f). In the inner bay, besides the Shenzhen River and Shau Pui River, mangrove forests expand along the coastline, forming the Futian and Mai Po mangrove reserves (Fig. 1). Oyster shelves expand throughout most of the remaining bay area, particularly in the central and Hong Kong area of the bay (Fig. 1). The bathymetry of Deep Bay is shallower in the inner part and deeper in the outer part (Ye et al., 2018, Fig. 1).

165 Eutrophication and pollution were predominant in the inner bay (Figs. 2, 3). Hypoxia frequency and nutrient concentration were elevated at EPD water monitoring stations DM1 and DM2 (Fig. 2a, b). Chemical oxygen demand, heavy metal and other pollutants in sediment were also relatively high at EPD sediment monitoring stations DS1 and DS2 in the inner bay (Fig. 2d–f). Our TOC and TN results showed the same trend that inner bay stations had higher values compared to the outer
170 stations (Fig. 3a, c). DB3 had the highest TOC and TN values, substantially exceeding those of the other sites (Fig. 3a, c).

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180 The lower $\delta^{13}\text{C}$ -TOC and $\delta^{15}\text{N}$ -TN values in the inner bay indicated a predominant organic matter source from terrestrial
 185 detritus, while the higher values in the outer bay suggest a predominant organic matter source of marine origin (Fig. 3a-d).
 The negative linear relationship between $\delta^{13}\text{C}$ -TOC and TOC/TN of the entire bay indicated a mixing between marine algae
 (with higher $\delta^{13}\text{C}$ -TOC and lower TOC/TN value) and C3 terrestrial plants like mangrove (with lower $\delta^{13}\text{C}$ -TOC and higher
 TOC/TN value) (Fig. 3e, f).



185 **Figure 2.** Sediment and seawater properties vary from inner bay to outer bay based on EPD monitoring data. Left panels (a-c)
 190 show bottom water hypoxia frequency (with threshold of 2mg/L, 3mg/L, 4mg/L respectively), surface nutrient DIN and DIP and
 physical parameters bottom temperature and salinity from DM1 to DM5. Right panels (d-f) show sediment chemical oxygen
 demand, and concentrations of heavy metal including copper, chromium, barium, cadmium, mercury and pollutant total cyanide
 from DS1 to DS4. The data are annual averages from 2012 to 2022.

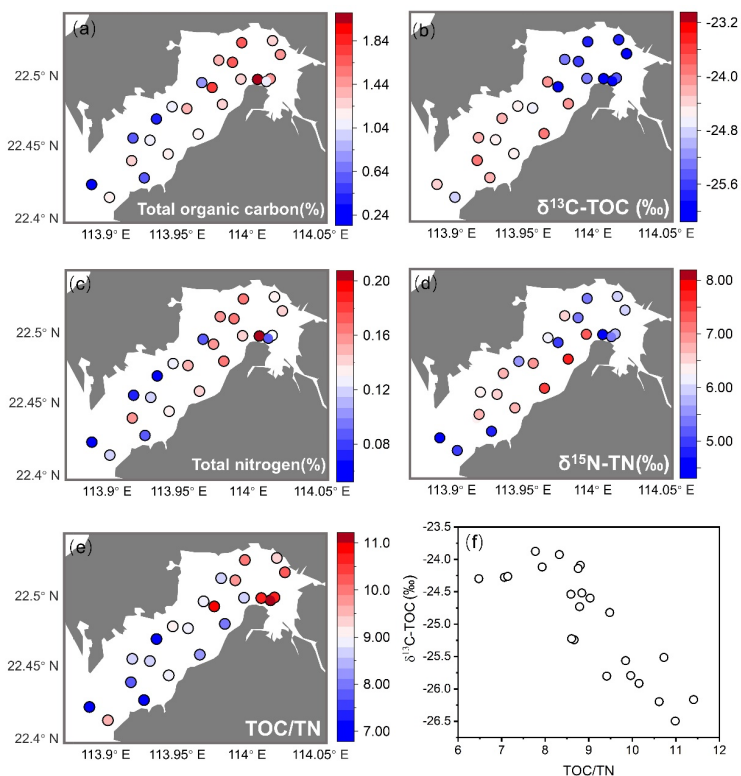


Figure 3. Elemental and their isotope analysis results in the sediments, including total organic carbon (%) (a) and its isotope (b), total nitrogen (%) (c) and its isotope (d), TOC/TN ratio (e), $\delta^{13}\text{C-TOC}$ and TOC/TN ratio relationship (f).

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3.2 Abundance and diversity

Abundance of ostracods in Deep Bay generally followed a gradient corresponding to the environmental conditions from the inner to outer bay. There tended to be lower abundance in the inner bay sites, with abundance increasing exponentially toward the bay mouth (Fig. 4). The lowest abundance was found inside the mangrove conservation area with almost zero

specimens (Fig. 4). This may be due to the acidic conditions in mangrove sediments, which negatively affect the preservation of calcium carbonate ostracod shells (Plazziat et al., 1983). The inner bay, characterized by eutrophication and riverine conditions, generally had ostracod abundance below 100, while the outer bay exhibited much higher abundance, especially at sites DB12, DB14 and DB15 (Fig. 4).

The diversity of ostracods in Deep Bay similarly followed the same inner-outer bay gradient (Fig. 5). The number of species, indicated by Hill Number 0D , was relatively high at the bay mouth sites SZ8, DB10, DB12, DB14 and DB15, ranging from 20 to 33. The other stations mostly had value below 10 (Fig. 5a). Similarly, the diversity of abundant species and dominant species (1D and 2D) were also higher among the stations at the bay mouth (Fig. 5b, c). Notably, DB7 and DB11 exhibited relatively high diversity in both abundant and dominant species (Fig. 5b, c).

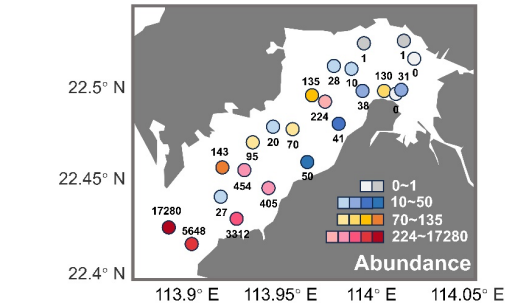
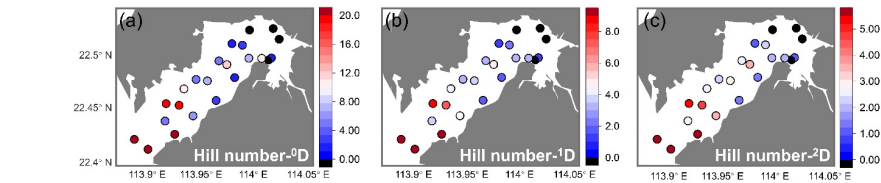


Figure 4. Ostracod abundance (total number of individuals per sample) in deep bay.



215 **Figure 5. Diversity results of deep bay ostracods. Distribution of Hill numbers 0D (q=0, species richness) (a), 1D (q=1, exponential Shannon) (b) and 2D (q=2, inversed Simpson) (c). Black dots represent stations with low abundance which are not included in the diversity calculation.**

3.3 Multivariate analysis

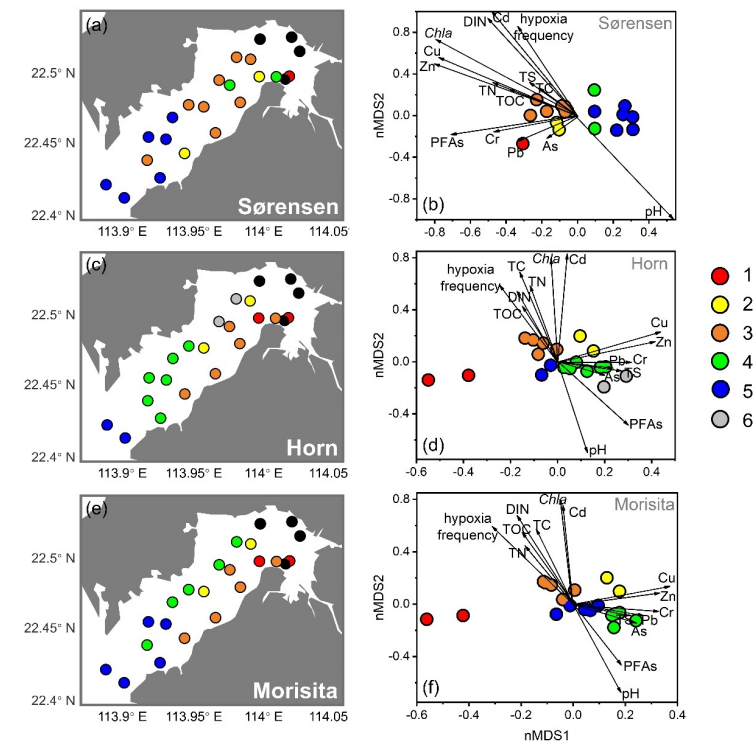
220 Cluster analysis based on Sørensen, Horn and Morisita-Horn dissimilarities delineated biofacies considering faunal composition in terms of species occurrence, relative abundance of abundant species, and relative abundance of dominant species, respectively. These three dissimilarities all showed a relatively clear separation both in geographical distribution and nMDS results (Fig. 6). The nMDS results showed a separation of five biofacies based on Sørensen and Morisita-Horn and six biofacies based on Horn dissimilarity (Fig. 6a, c, e).

225 Sørensen dissimilarity showed a separation between inner bay and outer bay of Deep Bay into Biofacies 3 and Biofacies 5, respectively (Fig. 6a). The inner bay (Biofacies 3) was related to almost all the environmental parameters we used (Fig. 6b), which showed that the inner bay was strongly influenced by eutrophication, hypoxia and pollutants. Biofacies 3 was characterized by the presence of *Cytherois* sp., *Loxoconcha sinensis*, *Paracypris* sp. and *Paracytherois* sp. as well as the absence of species that characterized other biofacies such as *Spinileberis bella* and *Spinileberis quadriculeata* (Table 1).
 230 Biofacies 5 was characterized by the presence of marine species *Alocopocythere goujoni*, *Keijiella kloempritsensis*, and *Xestoleberis hanaii* (Table 1). Except the two main groups above, inner bay Biofacies 1 was characterized by the presence of *Spinileberis bella* and *Sinocytheridea impressa* and the absence of other species (Table 1). Biofacies 2 was characterized by the occurrence of brackish water species *Spinileberis bella*, *Spinileberis quadriculeata*, *Loxoconcha ocellata*, and *Paracathacythere* sp. (Table 1). Biofacies 4 was similar to Biofacies 2 but distinguished by the occurrence of
 235 *Stigmatocythere bona* and *Propontocypris* sp. (Table 1).

Horn and Morisita-Horn dissimilarities revealed a distinction between Hong Kong area stations and Shenzhen area stations, i.e., the Hong Kong area group (Biofacies 3) and the Shenzhen area group (Biofacies 4) (Fig. 6c, e). The Hong Kong area group was more eutrophication-influenced and characterized by the dominance of *Sinocytheridea impressa* (Table 1), while
 240 the Shenzhen area group was more related to heavy metal and other pollutants (Fig. 6d, f) and characterized by the dominance of *Neomonceratina delicata* (Table 1). Other biofacies were distributed in the bay mouth (Biofacies 5) and inner bay (Biofacies 1, 2, and 6) (Fig. 6c, e). The bay mouth Biofacies 5 was unrelated to all eutrophication/pollution environmental parameters (Fig. 6), and characterized by moderately high abundance of both *Sinocytheridea impressa* and *Neomonceratina delicata* without any distinctively dominant species (Table 1). The inner-most bay Biofacies 1 was
 245 distinguished by the dominance of *Spinileberis bella* (Table 1). Biofacies 2 was distinguished by the dominance of

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Bicornucythere bisanensis s.l. (Table 1). Biofacies 6 in Horn dissimilarity was less diverse and characterized by the dominance of *Neomonoceratina delicata* and *Neosinocythere elongata* (Table 1).



270 **Figure 6.** Distribution map in left panels (a, c, e) and nMDS ordination in right panels (b, d, f) showing biofacies based on Sørensen, Horn and Morisita-Horn dissimilarities. Vectors on the nMDS ordination show the strength and direction of relation between environmental variables and the nMDS axes. Black dots represent stations with low abundance which are not included in calculation.

275 **Table 1.** List of top 10 species with the highest (%) mean relative abundance for Biofacies 1~6 based on Sørensen, Horn and Morisita-Horn dissimilarities. Specific biofacies among different dissimilarities may not be correspondingly related. Circular marks in the table represents corresponding biofacies showed in Fig. 6.

Dissimilarity	Species	1	2	3	4	5	6
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Sorensen	<i>Alocopocythere goujoni</i>			3.10		5.27	
	<i>Aurila</i> sp.				0.83		
	<i>Bicornucythere bisanensis</i> s.l.	5.92		20.33	17.72	8.94	
	<i>Cytherois</i> sp.			1.85			
	<i>Keijella kloempitensis</i>					5.60	
	<i>Loxoconcha sinensis</i>			0.63			
	<i>Loxoconcha ocellata</i>	3.29					
	<i>Neomonceratina delicata</i>	17.76		40.93	14.64	34.53	
	<i>Neosinocythere elongata</i>	0.16		4.43	0.89	2.11	
	<i>Paracathacythere</i> sp.	1.32					
	<i>Paracypris</i> sp.			0.54			
	<i>Paracytherois</i> sp.			0.18			
	<i>Pistocythereis bradyformis</i>				0.99	1.63	
	<i>Pistocythereis bradyi</i>	1.32				3.40	
	<i>Propontocypris</i> sp.				2.33		
	<i>Sinocytheridea impressa</i>	3.23	25.99	28.01	55.27	19.93	
	<i>Spinileberis bella</i>	96.77	41.61		0.89		
	<i>Spinileberis quadriaculeata</i>		2.63		1.12		
	<i>Stigmatocythere bona</i>				1.34	2.19	
	<i>Xestoleberis hanaii</i>					2.28	
Horn	<i>Alocopocythere goujoni</i>		0.71		7.82		2.96
	<i>Aurila</i> sp.					1.42	
	<i>Bicornucythere bisanensis</i> s.l.	1.32	54.29	9.73	12.51	6.71	7.65
	<i>Cytherois</i> sp.				3.59		
	<i>Hemicytheridea reticulata</i>				1.13		
	<i>Keijella kloempitensis</i>				0.99	13.84	
	<i>Loxoconcha sinensis</i>				1.24		
	<i>Loxoconcha ocellata</i>			1.32			
	<i>Neomonceratina delicata</i>	3.95	19.29	17.11	46.93	20.58	72.35
	<i>Neosinocythere elongata</i>		1.43	0.42		3.41	16.30
	<i>Paracathacythere</i> sp.	1.32					
	<i>Paracypris</i> sp.		2.14				
	<i>Paracytherois</i> sp.		0.71				

	<i>Pistocythereis bradyformis</i>			0.40		3.27
	<i>Pistocythereis bradyi</i>	1.32				7.71
	<i>Propontocypris</i> sp.			0.93		2.85
	<i>Sinocytheridea impressa</i>	6.88	21.43	63.87	13.89	24.33
	<i>Spinileberis bella</i>	82.60		3.32		
	<i>Spinileberis quadriaculeata</i>	2.63		0.45		
	<i>Stigmatocythere bona</i>			0.54	1.55	1.92
	<i>Xestoleberis hanaii</i>				1.80	
Morisita	<i>Alocopocythere goujoni</i>		0.71		6.56	4.43
	<i>Bicornucythere bisanensis</i> s.l.	1.32	54.29	9.73	11.71	9.05
	<i>Cytherois</i> sp.				3.17	
	<i>Keijella kloempritis</i>				0.42	6.30
	<i>Loxoconcha sinensis</i>				1.21	
	<i>Loxoconcha ocellata</i>			1.32		
	<i>Neocyprideis agilis</i>				0.21	
	<i>Neomonoceratina delicata</i>	3.95	19.29	17.11	64.05	29.44
	<i>Neosinocythere elongata</i>		1.43	0.42	6.94	2.11
	<i>Paracathacythere</i> sp.	1.32				
	<i>Paracypris</i> sp.		2.14			
	<i>Paracytherois</i> sp.		0.71			
	<i>Pistocythereis bradyformis</i>			0.40	0.42	
	<i>Pistocythereis bradyi</i>	1.32				4.07
	<i>Propontocypris</i> sp.			0.93		1.71
	<i>Sinocytheridea impressa</i>	6.88	21.43	63.87	5.09	21.60
	<i>Spinileberis bella</i>	82.60		3.32		
	<i>Spinileberis quadriaculeata</i>	2.63		0.45		
	<i>Stigmatocythere bona</i>			0.54		2.42
	<i>Xestoleberis hanaii</i>					2.73

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3.4 Biogeography

Geographical distribution of individual species were showed in Figs. 7 and 8. *Neomonoceratina delicata* was mostly distributed in the Shenzhen [area](#) (Fig. 7a). *Sinocytheridea impressa* had higher abundance in the Hong Kong [area](#) (Fig. 7b). *Spinileberis bella* was concentrated in the river mouth (Fig. 7c). *Bicornucythere bisanensis* s.l. was distributed with high

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abundance in the inner bay eutrophic/polluted area (Fig. 7d). *Alocopocythere goujoni* was distributed mostly in the outer bay, Shenzhen area (Fig. 7e). *Neosinocythere elongata* was abundant in an inner bay station SZ5 (Fig. 7f). *Keijiella kloempricensis* was abundant only in the stations DB14 and DB15 located near the bay mouth (Fig. 7g). *Pistocythereis bradyi* was abundant in a bay mouth station DB15 (Fig. 7h). *Cythereis* sp. occurred mostly in an outer bay station DB13 (Fig. 7i). *Stigmatocythere bona* was abundant in outer bay stations SZ7 and SZ8 (Fig. 8a). *Xestoleberis hanaii* was abundant in outer-central bay stations, Shenzhen area, including SZ7, SZ8, and DB9 (Fig. 8b). *Propontocypris* sp. was relatively broadly distributed from the inner bay to the outer bay, with higher abundance observed in the outer region (Fig. 8c). *Pistocythereis bradyformis* tended to show higher abundance in the outer bay and at the bay mouth, while it was absent in most stations of the inner bay and at the Hong Kong area stations (Fig. 8d). *Spinileberis quadriaculeata* was abundant in central-outer bay stations SZ7 and SZ8 (Fig. 8e). *Hemicytheridea reticulata* was abundant in central bay stations SZ6, SZ7 and DB11 (Fig. 8f). *Loxoconcha sinensis* was distributed in the middle of the bay, Shenzhen area, with high abundance observed in a relatively inner station SZ4 and one outer station SZ8 (Fig. 8g). *Loxoconcha ocellata* was abundant in outer bay stations SZ7 and SZ8 (Fig. 8h). *Aurila* cf. *disparata* was mostly found in station DB11 (Fig. 8i). Shenzhen area stations SZ7 and SZ8 had a distinct characteristics with relatively high abundance of *Stigmatocythere bona*, *Xestoleberis hanaii*, *Spinileberis quadriaculeata* and *Loxoconcha ocellata*. The bay mouth, particularly the oceanic-influenced stations DB14 and DB15, exhibited a distinctly high abundance of *Keijiella kloempricensis* and *Pistocythere bradyi*. These stations also recorded highest total ostracod abundance.

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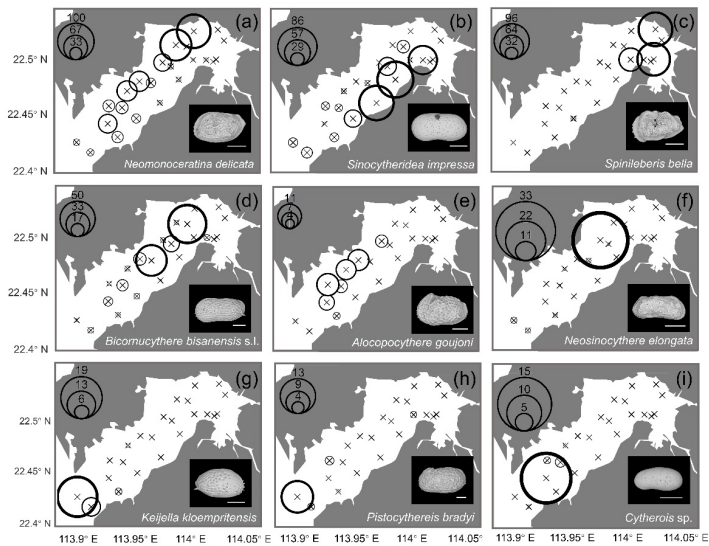


Figure 7. Spatial distribution of the relative (%) abundance for the top 9 abundant species (relative abundance over than 1% among all species) in Deep Bay. Including (a) *Neomonoceratina delicata*, (b) *Sinocytheridea impressa*, (c) *Spinileberis bella*, (d) *Bicornucythere bisanensis* s.l., (e) *Alocopocythere goujoni*, (f) *Neosinocythere elongata*, (g) *Keijella kloempitensis*, (h) *Pistocythereis bradyi*, and (i) *Cythereis* sp.. The scale bar represents 200 μ m for scanning electron microscope images at the lower-right corner of each subplot.

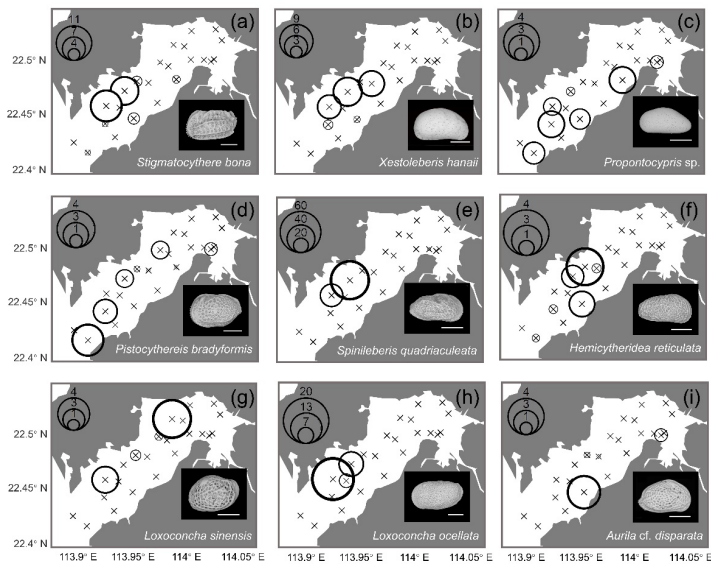


Figure 8. Spatial distribution of the relative (%) abundance for top 10–18 abundant species (relative abundance over than 0.3% among all species) in Deep Bay. Including (a) *Stigmatocythere bona*, (b) *Xestoleberis hanaii*, (c) *Propontocypris* sp., (d) *Pistocythereis bradyformis*, (e) *Spinileberis quadriaculeata*, (f) *Hemicytheridea reticulata*, (g) *Loxoconcha sinensis*, (h) *Loxoconcha ocellata*, and (i) *Aurila* cf. *disparata*. The scale bar represents 200 μm for scanning electron microscope images at the lower-right corner of each subplot.

4 Discussion

4.1 Abundance and diversity

Both ostracod abundance and diversity distribution followed a riverine-oceanic gradient in Deep Bay, driven by riverine input of excessive human waste leading to eutrophication and pollution (Note that the bay mouth area of Deep Bay is a part of Pearl River estuary but is still much more oceanic compared to the inner Deep Bay) (Figs. 4, 5). Recent studies showed that large amounts of nitrogen and phosphorus are transported to the inner-most Deep Bay by rainwater-runoff mainly via Shenzhen River from urban catchments, inducing eutrophication (Xu et al., 2019). Hypoxia frequency and nutrient

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concentration were much higher in the inner-bay, river-influenced sites as shown in the 10 years averaged EPD data (Fig. 2a, b), indicating that eutrophication and hypoxia already became a long-term environmental problem in Deep Bay. In Asian urban marginal marine areas, eutrophication has been identified as one of the main drivers altering benthic ostracod community, both contemporarily and historically (Yasuhara et al., 2007; Cheung et al., 2019; Hong et al., 2022). Ostracod abundance and diversity (Hill number 1D and 2D) of the entire bay had a significantly negative correlation with TOC content in sediments which also supported our perspective (Fig. 10). Metal pollution was investigated to be the dominant environmental factors structuring meiobenthic diversity in Hong Kong coastal waters (Hong et al., 2021; Hong et al., 2022). Relevant investigations in Deep Bay also showed a riverine-oceanic gradient on heavy metal pollutants and PFAs (Zhang et al., 2021; Li et al., 2024), suggesting that the riverine-oceanic distribution of ostracod abundance and diversity may result from the combined effect of both eutrophication and pollution. We compared Hill Number values between riverine and oceanic sites (Figs. 9b, 11), we found that the diversity in oceanic sites was substantially higher when Hill Number order was low. The diversity between riverine and oceanic sites was almost the same in higher order Hill Numbers, indicating that eutrophication did not influence much of the structure of the dominant species among stations. This difference in diversity between riverine and oceanic sites was mostly because of rare and abundant species. Low salinity and very shallow intertidal water depth could also be a reason for the lower diversity in riverine sites (Fig. 2c), as brackish-water environment is commonly characterized by much lower diversity compared to full-marine environment (Frenzel and Boomer, 2005; Whitfield et al., 2012) and aerial exposure at low tides could be stressful for marine organisms (Tomanek and Helmuth, 2002).

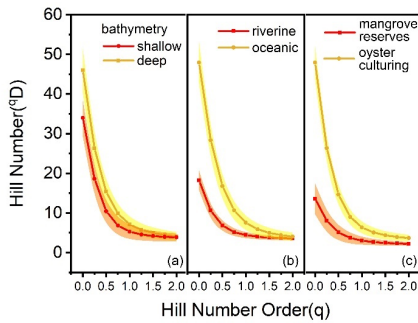


Figure 9. Hill number profile of deep-shallow bathymetry, riverine-oceanic area and mangrove reserves-oyster culturing zone. The shaded areas show 95% confidence interval of the profile. (a) Deep bathymetry indicates topography lower than PRL while shallow bathymetry indicates higher than PRL. (b) Riverine stations included stations inner than DB9 (DM3) while oceanic included stations outer and DB9 itself. (c) Stations in mangrove reserves included DB1–4 and SZ1–3 while other stations were regarded as oyster culturing zone.

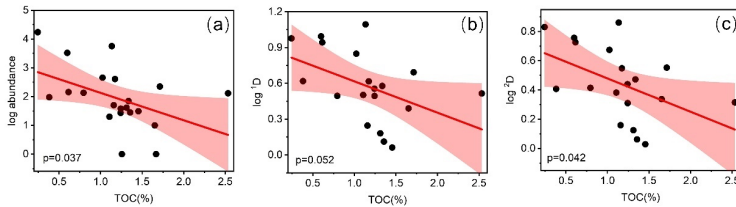


Figure 10. Relationship between eutrophication [total organic carbon (TOC) content in sediment] and abundance/diversities (Hill number 1D and 2D). (a) Total abundance vs. TOC. (b) Hill number 1D vs. TOC. (c) Hill number 2D vs. TOC. Shaded area shows 95% confidence interval.

4.2 Faunal distribution

In our study, three eutrophication/hypoxia tolerant species *Neomonoceratina delicata*, *Sinocytheridea impressa* and *Bicornucythere bisanensis* s.l. (Irizuki et al., 2003; Yasuhara et al., 2007; Yasuhara et al., 2012) were abundantly distributed in most of the inner bay stations where TOC was high (Figs. 3a, 7). The Sørensen dissimilarity result showed a distinction between riverine and oceanic (inner bay and middle-outer bay) stations (Fig. 6a). The outer-bay Biofacies 5 in Sørensen dissimilarity was characterized by the lowest TOC and TN values, along with highest abundance and highest diversity. This biofacies exhibited oceanic characteristic and was less related to riverine eutrophication and pollution (Figs. 3a, c, 4, 5, 6a). In contrast, the inner-bay Biofacies 3 in Sørensen dissimilarity was composed of brackish water species and related to almost all the eutrophication and pollution parameters that we used in this study, accompanied with high TOC and TN value, low abundance and low diversity (Figs. 3a, b, 4, 5, 6a, b). These results indicated that eutrophication and pollution were the drivers that differentiated the inner bay fauna from the outer bay ones. The inner-most bay Biofacies 1 (of all dissimilarities) was probably a river mouth, brackish-water fauna, because it was located nearby Shenzhen River mouth and characterized by low salinity (Figs. 2, 6a, c, e). The dominant species in this biofacies was *Spinileberis bella* that is most likely a brackish-water species (Hou and Gou, 2007), as most *Spinileberis* species are known as brackish-water inhabitants (e.g., Zhao and Wang, 1988).

In addition to this major faunal trend of the inner-outer bay gradient, another major faunal trend was the separation of Hong Kong area and Shenzhen area faunas, clearly represented as Biofacies 3 and 4, respectively, in the Horn and Morisita-Horn dissimilarity results (Fig. 6c, d). Correspondingly, Hong Kong area stations were more eutrophication-influenced while Shenzhen area stations were more affected by heavy metal pollution (Fig. 6d, f). Biofacies 3 showed the highest abundance

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in eutrophication-tolerant *Sinocytheridea impressa* (Hong et al., 2019) in both Horn and Morisita-Horn based cluster analysis (Table 1), indicating that the Hong Kong *area* benthic ecosystem was influenced more by eutrophication. In contrast, *Bjofacies* 4 was characterized by a very high abundance in metal-pollution-tolerant *Neomonoceratina delicata* (Hong et al., 2019) and a much lower abundance of *Sinocytheridea impressa* (Table 1), indicating that the Shenzhen *area* benthic ecosystem was influenced more by metal pollution. Most eutrophication parameters were related to *Bjofacies* 3 in the Hong Kong *area*, including hypoxia frequency, *Chla*, DIN, and related sediment eutrophication parameters including TC, TN and TOC (Fig. 6d, f). Pollutant PFAs and heavy metal Cu, Zn, Pb, Cr, As were related to *Bjofacies* 4 in the Shenzhen *area* (Fig. 6d, f), and TS was also related to *Bjofacies* 4, potentially because heavy metal availability affects sediment sulfur cycle (Jørgensen et al., 2019).

The reason for the distinction between the Hong Kong *area* and Shenzhen *area* remained unclear due to limited evidence. However, there are several possibilities. Stations in *Bjofacies* 3 (Hong Kong *area*) were shallower than staitons in *Bjofacies* 4 (Shenzhen *area*) (Zhang and Mao, 2015) (Also see Fig. 1). With shallower water depth in the Hong Kong *area*, organic matter could be less decomposed in the water column with its short distance from the surface water to the bottom sediment, resulted in higher sedimentation of organic materials in the sediments compared to the deeper Shenzhen *area*. It is also shown in sedimentary TOC and TN result, with shallower Hong Kong *area* stations having relatively high TOC and TN concentrations compared to the Shenzhen *area* stations (Fig. 3a, c), indicating a more eutrophic condition in the Hong Kong *area*. Notably, α diversity was lower at shallower water depth (Figs. 9a, 11), potentially because of the stress from eutrophication. Another difference is that the Shenzhen *area* of Deep Bay is surrounded by Futian and Nanshan district, which are two of the most populated districts of Shenzhen City (Zhou et al., 2020). Comparably, Hong Kong *area* of Deep Bay is close to Hong Kong rural area. Moreover, secondary industry was transferred from Hong Kong to Shenzhen after 1978 (Sit et al., 2020). Now secondary industry accounts for 37.6% GDP in Shenzhen which is almost 5 times higher than 6.5% in Hong Kong (data from the website of the Greater Bay Area <https://www.bayarea.gov.hk/sc/home/index.html>). More industrial waste could be the reason of pollutant-driven ostracod fauna distribution in Shenzhen *area*. Possible roles of oyster aquaculturing were uncertain, because oyster aquaculture rafts distribute almost cross the entire Deep Bay except at the inner-most bay, mangrove area (Fig. 1). Although diversity was much higher in the oyster aquaculture sites compared to the inner-most bay mangrove sites (Figs. 9c, 11), it might be complicated by the fact that mangrove sites were characterized by low salinity, acidic condition, and intertidal water depth as discussed earlier.

4.3 Comparison with broader Hong Kong study

Species distribution of ostracod species has been investigated in the broader Hong Kong region (Hong et al., 2019). While *Alocopocythere goujoni* was common in Hong Kong's western water, albeit with low abundance (Hong et al., 2019), we found that this species was most abundant in the middle-outer Deep Bay (Fig. 7e) among Hong Kong's waters (Hong et al.,

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2019). Hong et al. (2019) suggested that this species is a high salinity indicator. Our study partially supports this finding, as *Alocopocythere goujoni* were abundant in the middle-outer Deep Bay. However, other factors may also be at play, considering that Deep Bay is not the area with highest salinity in Hong Kong (Fig. 2c; Hong et al., 2019). *Neosinocythere elongata* was favourable to high turbidity condition in Hong Kong broadly (Hong et al., 2019), and was indeed distinctly abundant in inner-bay Biofacies 6 in Horn dissimilarity in Deep Bay (Table 1). *Neomonoceratina delicata* broadly distributed in all of Hong Kong waters (Hong et al., 2019), and it also broadly distributed with relatively high abundance in Deep Bay (Fig. 7a). *Sinocythereidea impressa* and *Bicornucythere bisanensis* s.l. were more abundant in semi-enclosed bay areas, such as Tolo harbour and inner Port Shelter in Hong Kong (Hong et al., 2019). Consistently, we found that they were also abundant mostly in the inner Deep Bay (Fig. 7b, c). In contrast, *Keijella kloempricensis* and *Pistocythereis bradyi* were distributed more in open waters, such as the southern and eastern part of Hong Kong, as they preferred deep waters and high salinity environments, respectively (Hong et al., 2019). Consistently, they distributed mostly at the bay mouth of Deep Bay (Fig. 7g, h). *Xestoleberis hanaii* had a preference of more transparent water condition in Hong Kong (Hong et al., 2019) and, in our Deep Bay study, were located more in the deeper and less eutrophic Shenzhen area (Fig. 8b). *Propontocypris* sp., which preferred lower productivity in Hong Kong (Hong et al., 2019), was distributed more in the outer Deep Bay (Fig. 8c). In summary, we found consistent spatial trends of species distribution between our Deep Bay study and the previous broader Hong Kong study of Hong et al (2019). Phytal species such as *Cytherois* sp., *Xestoleberis hanaii* and *Aurila* cf. *disparata* were mostly distributed in oyster aquaculture areas (Irizuki et al., 2008) (Figs. 1, 7i, 8b, i), which may be related to phytal habitats associated with oyster rafts. However, this is speculative, as oyster aquaculture covers almost entire area of Deep Bay and not just the inner bay and mangrove areas (Fig. 1).

The wide distribution of ostracod biofacies in Hong Kong is complex and cannot be explained straightforwardly by one or two environmental factors (Hong et al., 2022). This complexity likely arises from the region's multiple enbayments, complicated coastline, numerous small and large islands, and varied environmental regimes influenced by multiple sources of pollutants, nutrients, freshwater, etc. However, our high-resolution study in much simpler, smaller system as an important part of Hong Kong, i.e., Deep Bay, clearly confirmed that the river-influenced eutrophication is a critical factor controlling marginal marine benthic community. While intermediate level of eutrophication may be beneficial to soft sediment benthos, the excess food supply caused by eutrophication, along with the consequent bottom deoxygenation, can lead to the dominance of a few opportunistic species, finally resulting in low diversity, as observed in East Asia and globally (Yasuhara et al., 2007, 2012). Riverine metal pollution may amplify this benthic community change and contributed to form the gradient of opportunistic species under stress in the inner bay to healthier fauna in the outer bay in Deep Bay.

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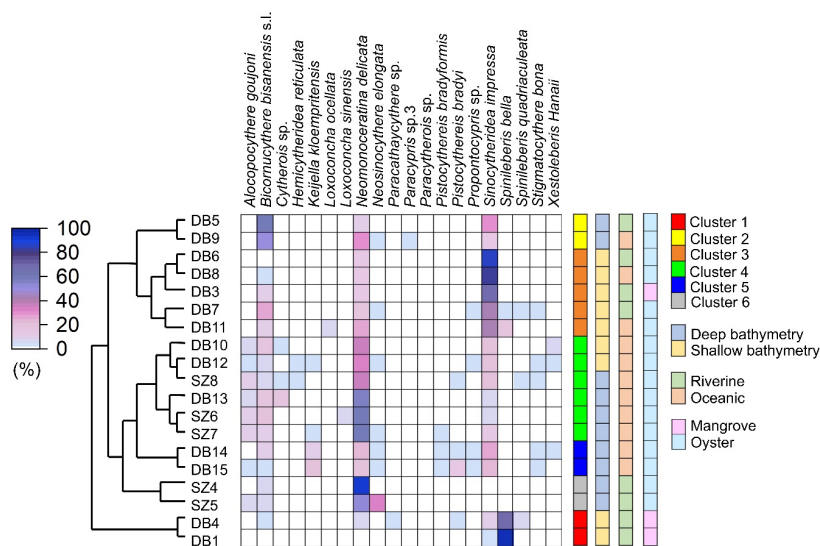


Figure 11. Heatmap with Horn dissimilarity among samples which shows top 10 species of highest mean relative abundance in each cluster. The side panel shows cluster details, bathymetry difference and riverine-oceanic circumstances of each sample.

500 5 Conclusion

Ostracod abundance, diversity, composition, distribution, as well as thier interactions with eutrophication and pollution, were investigated through high-resolution sampling of surface sediments in Deep Bay, a small semi-enclosed bay adjacent to two of the most populated cities in the world. We found that:

- 505 1. Eutrophication and pollution were the most dominant drivers of ostracod abundance and diversity, creating a pronounced inner-outer bay gradient in Deep Bay.

2. Faunal composition in Deep Bay was characterized by difference between the inner and outer bays, as well as between the Hong Kong and Shenzhen [areas](#). The inner-outer difference was controlled by eutrophication along with pollution from rivers. The Hong Kong-Shenzhen [area](#) difference was associated with eutrophication in the Hong Kong [area](#) and metal pollution in the Shenzhen [area](#). However, the mechanism and dominant factor driving the Hong Kong-Shenzhen gradient remained unclear.

3. Our results in Deep Bay were consistent with broader studies on ostracod diversity and distribuiton in Hong Kong, strongly supporting the idea that ostracod is an useful bioindicator of coastal benthic ecosystems. Moreover, in our smaller urban bay case, our investigation provided strong evidence of how eutrophication shapes benthic ostracod communities—an effect that may not be as apparent at larger spatial scales, especially in the regions with complicated geographic settings like Hong Kong, which features with multiple embayments, complex coast lines, and numerous islands. This underscores the importance of fine-scale sampling for regional studies. Eutrophication and pollution in small urban bay area have already caused profound consequences.

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[Data availability. The data are available in Zenodo open data repository \(Huang et al., 2025\).](#)

[Author contributions. J.H., M.Y., and H.W. designed the research; J.H., H.W., P.J. and J.L. performed the research; J.H. analyzed data; J.H. and M.Y. wrote the paper with intellectual contributions from all authors.](#)

[Competing interests. The authors declare they have no competing interests.](#)

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