

## Supplementary material

# Effect of straw retention and mineral fertilization on P speciation and P-transformation microorganisms in water extractable colloids of a Vertisol

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Table S1 Chemical shift of peaks detected in  $^{31}\text{P}$ -NMR spectra of NaOH- $\text{Na}_2\text{EDTA}$  samples

Category <sup>a</sup>	P form or compound class	Chemical Shift (ppm)
Inorganic P		
	Orthophosphate	6.0
	Pyrophosphate	-4.39±0.03
Organic P		
Orthophosphate Monoesters		
	myo-IHP	4.28±0.01, 4.44±0.02, 4.81±0.01, 5.71±0.02
	scyllo-IHP	3.96±0.02
	$\alpha$ -glyc	5.01±0.02
	$\beta$ -glyc	4.66±0.02
	Orthophosphate diesters-- DNA	-0.41±0.04

<sup>a</sup>myo-inositol hexakisphosphate (myo-IHP), scyllo-inositol hexakisphosphate (scyllo-IHP),  $\alpha$ -glycerophosphate ( $\alpha$ -glyc),  $\beta$ -glycerophosphate ( $\beta$ -glyc).

Table S2 The KO number, function descriptions, gene name and classification of the investigated genes referring to KEGG database

Classification	KO number	Details for gene function	Corresponding gene
Genes involved in inorganic P-solubilization	K00117	quinoprotein glucose dehydrogenase	<i>gcd</i>
	K01507	inorganic pyrophosphatase	<i>ppa</i>
	K00937	Polyphosphate kinase	<i>ppk</i>
	K01524	exopolyphosphatase / guanosine-5'-triphosphate,3'-diphosphate pyrophosphatase	<i>ppx</i>
Genes involved in organic P-mineralization	K05774	C-P lyase subunit, ribose 1,5-bisphosphokinase	<i>phnN</i>
	K00111	Glycerol-3-phosphate dehydrogenase	<i>glpA</i>
	K00112	Glycerol-3-phosphate dehydrogenase subunit B	<i>glpB</i>
	K00113	Glycerol-3-phosphate dehydrogenase subunit C	<i>glpC</i>
	K00864	Glycerol kinase glpK	<i>glpK</i>
	K01083	3-Phytase	<i>phy</i>
	K01093	4-phytase	<i>appA</i>
	K01126	glycerophosphoryl diester phosphodiesterase	<i>ugpQ</i>
	K01077	alkaline phosphatase	<i>phoA</i>
	K01113	alkaline phosphatase	<i>phoD</i>
	K09474	acid phosphatase (class A)	<i>phoN</i>
	K03788	acid phosphatase (class B)	<i>aphA</i>
Genes involved in P-starvation response regulation	K07636	two-component system, OmpR family, phosphate regulon sensor histidine kinase PhoR	<i>phoR</i>
	K07657	two-component system, OmpR family, phosphate regulon response regulator PhoB	<i>phoB</i>
Genes involved in P-uptake and transport system	K05813	Glycerol-3-phosphate transporter subunit, periplasmic-binding component	<i>ugpB</i>
	K05814	Glycerol-3-phosphate transporter subunit	<i>ugpA</i>
	K05815	Glycerol-3-phosphate transporter subunit, membrane component	<i>ugpE</i>
	K05816	Glycerol-3-phosphate transporter subunit, ATP-binding component	<i>ugpC</i>
	K02041	phosphonate transport system, ATP-binding component	<i>phnC</i>
	K02042	phosphonate transport system, membrane component	<i>phnE</i>
	K02044	phosphonate transport system, periplasmic-binding component	<i>phnD</i>
	K02036	phosphate transport system, ATP-binding component	<i>pstB</i>
	K02037	phosphate transport system, membrane component	<i>pstC</i>
	K02038	phosphate transport system, membrane component	<i>pstA</i>
	K02040	phosphate transport system, periplasmic-binding component	<i>pstS</i>

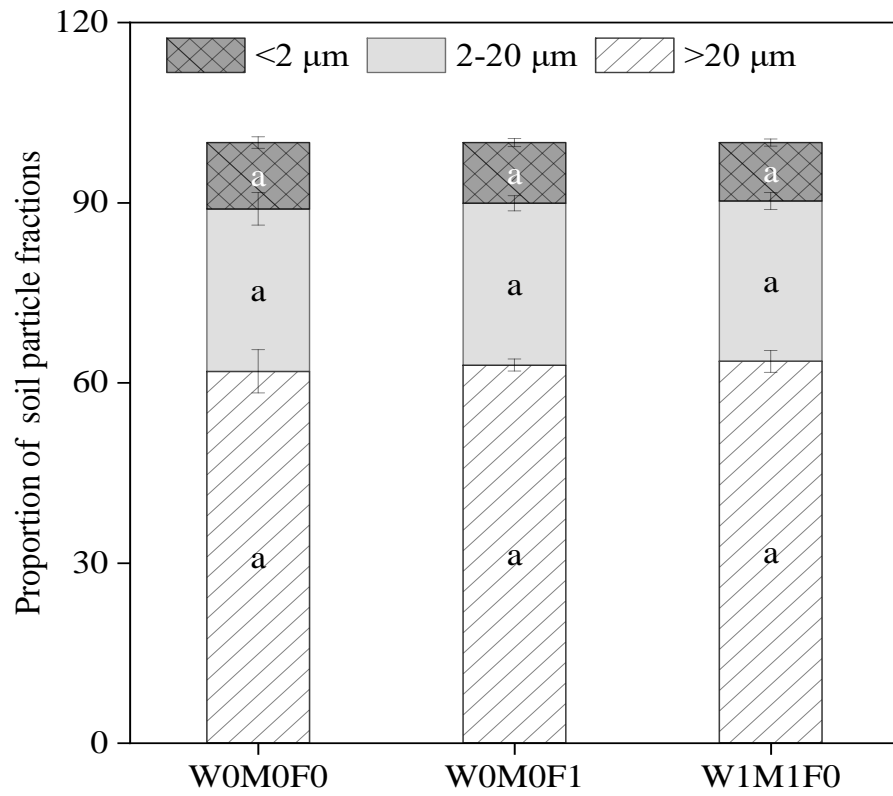


Fig. S1 Proportion of water-dispersible soil particle fractions for the WOM0F0, WOM0F1 and W1M1F0 treatments

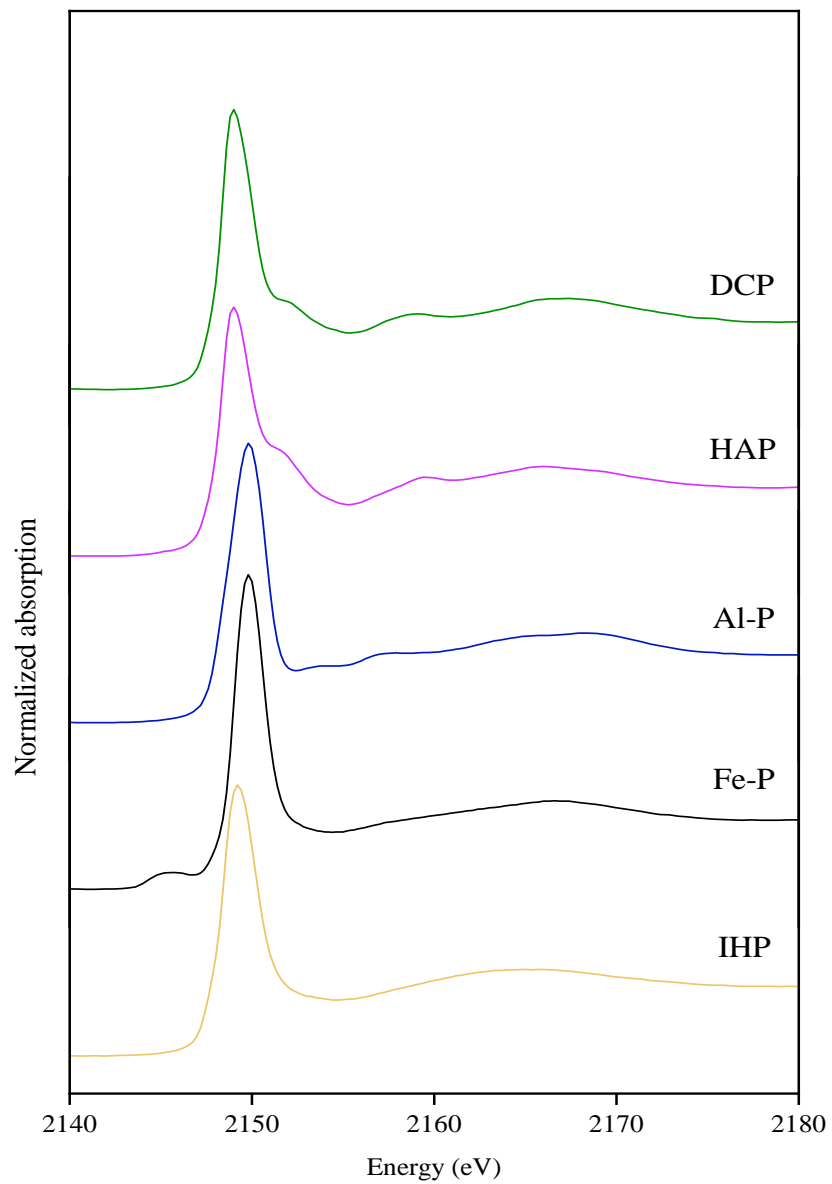


Figure S2 Phosphorus K-edge XANES spectra of P reference compounds

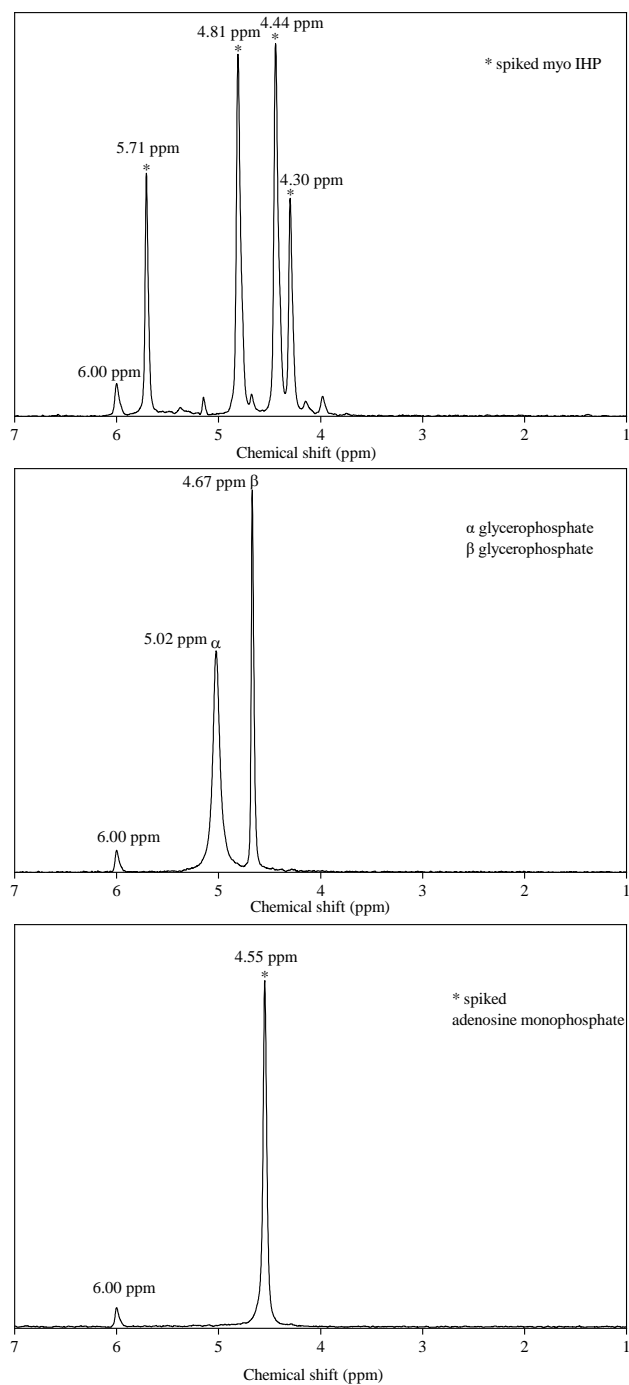


Fig. S3 The solution  $^{31}\text{P}$  NMR spectrum of a NaOH- $\text{Na}_2\text{EDTA}$  extract of the W0M0F0 bulk soil spiked with a) myo-inositol hexakisphosphate (Myo-IHP), b)  $\alpha$ - glycerophosphate and  $\beta$ - glycerophosphate, and c) adenosine monophosphate.

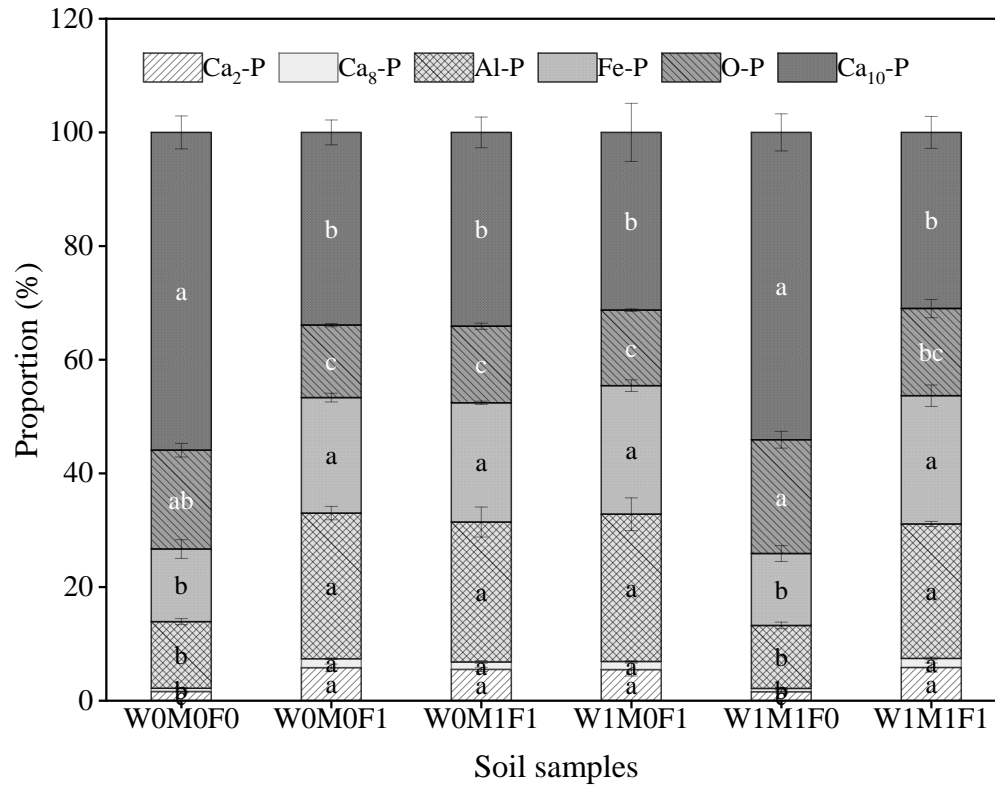


Fig. S4 Proportion of different soil inorganic phosphorus fractions in bulk soil

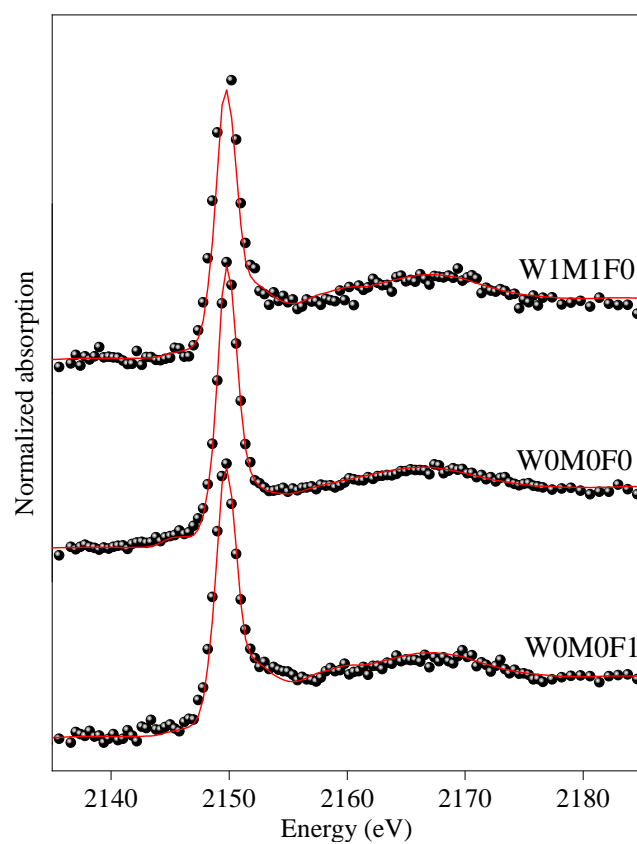
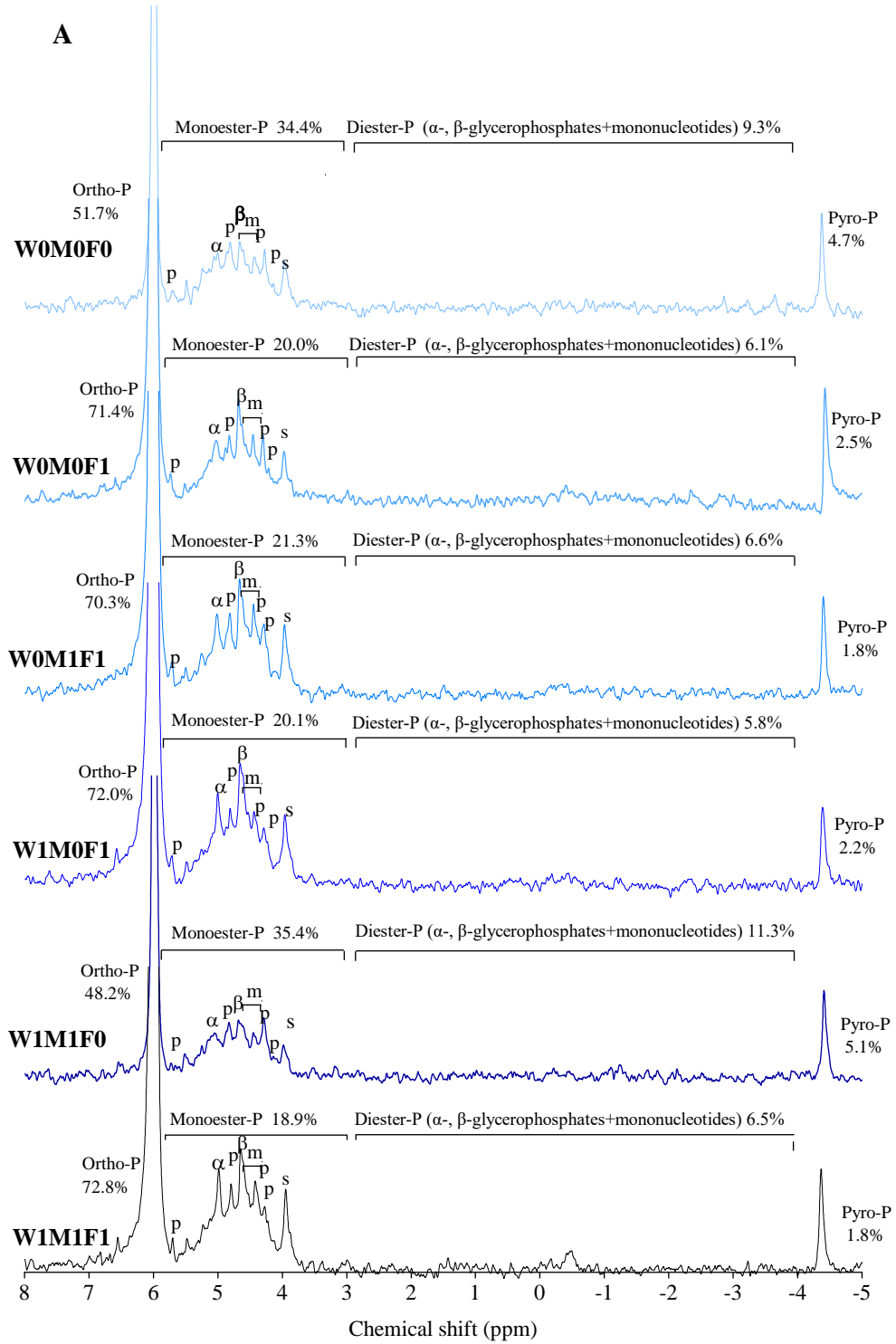


Fig. S5 Results of the linear combination fitting (LCF) of the P K- edge XANES spectra for water-extractable colloids (WECs) of the W0M0F0, W0M0F1 and W1M1F0 treatments. The raw data is represented by points and the fitting results are shown by solid lines.





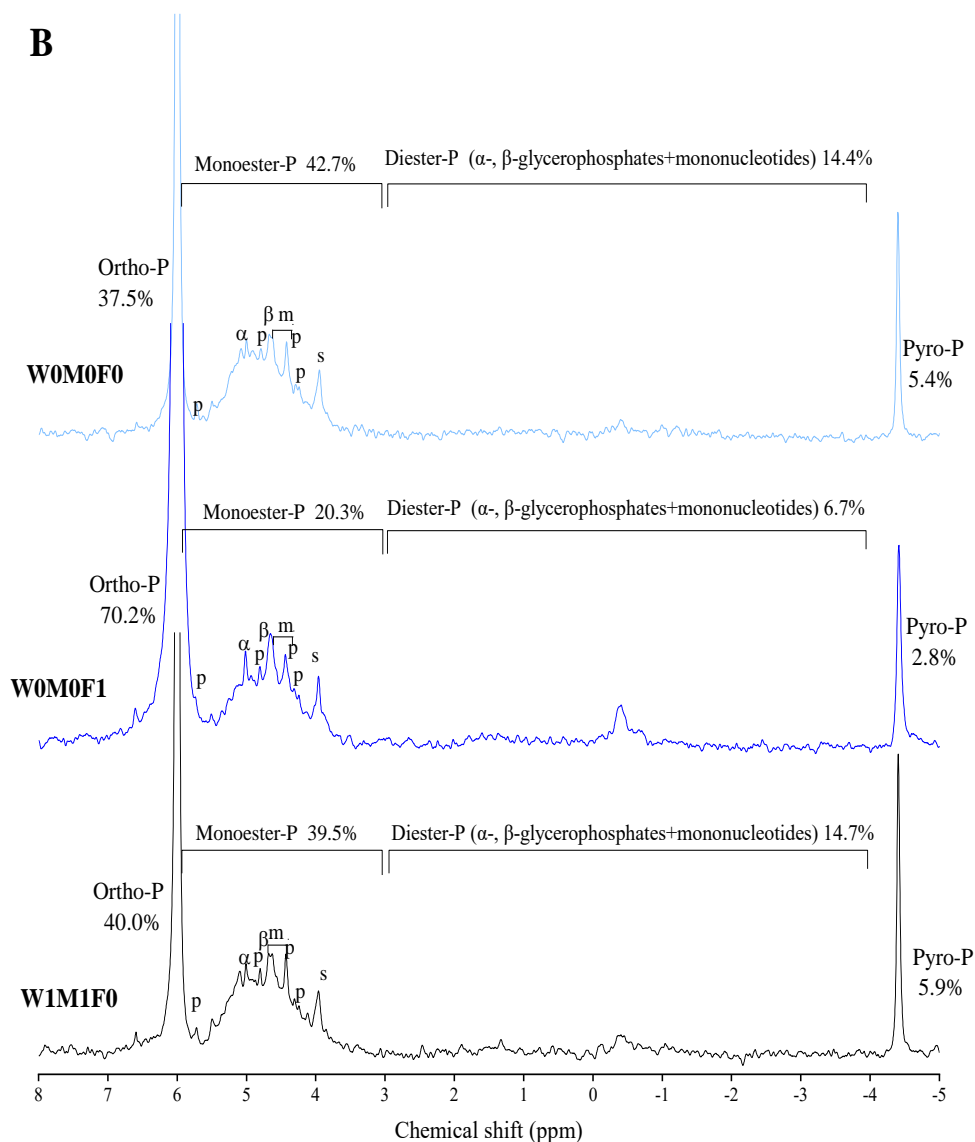


Fig. S6 Solution  $^{31}\text{P}$  NMR spectra of NaOH–Na<sub>2</sub>EDTA extracts of bulk soil (a) and water-extractable colloids (WECs, b)

The signals were assigned as follows: orthophosphate (Ortho-P); orthophosphate monoester (Monoester-P); orthophosphate diesters (Diesters-P); pyrophosphate (Pyro-P); myo-inositol hexakisphosphate (p); syllo-inositol hexakisphosphate (s); mononucleotides (m);  $\alpha$ - glycerophosphate ( $\alpha$ );  $\beta$ - glycerophosphate ( $\beta$ ). Calculation by including diester degradation products (i.e., Glyc+nucl:  $\alpha/\beta$ - glycerophosphate and mononucleotides) with Diesters-P rather than Monoesters-P. The percentage indicated the proportion of each phosphorus compounds fraction to the extracted-P.

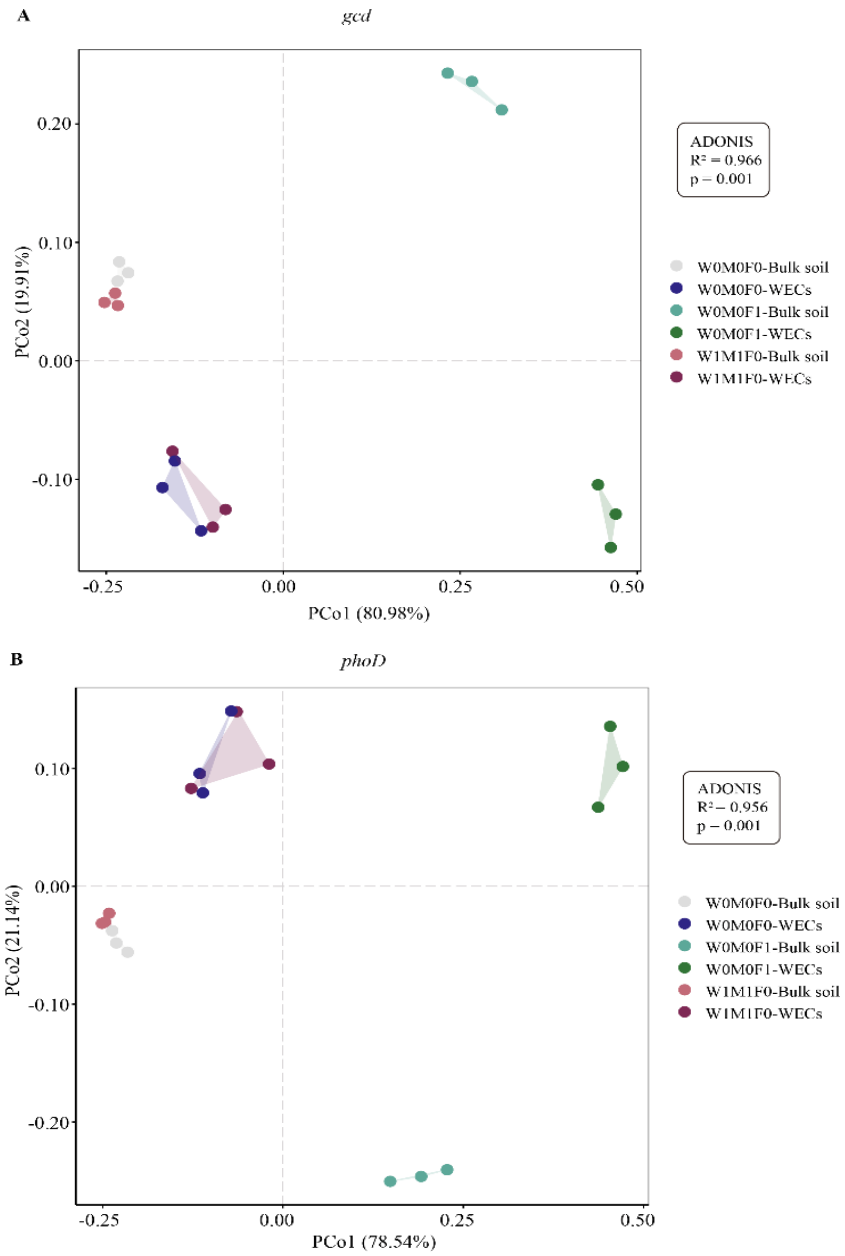


Fig. S7 Principal coordinate analysis (PCoA) of microbial community for *gcd* (A) and *phoD* (B) gene in bulk soils and water-extractable colloids (WECs)

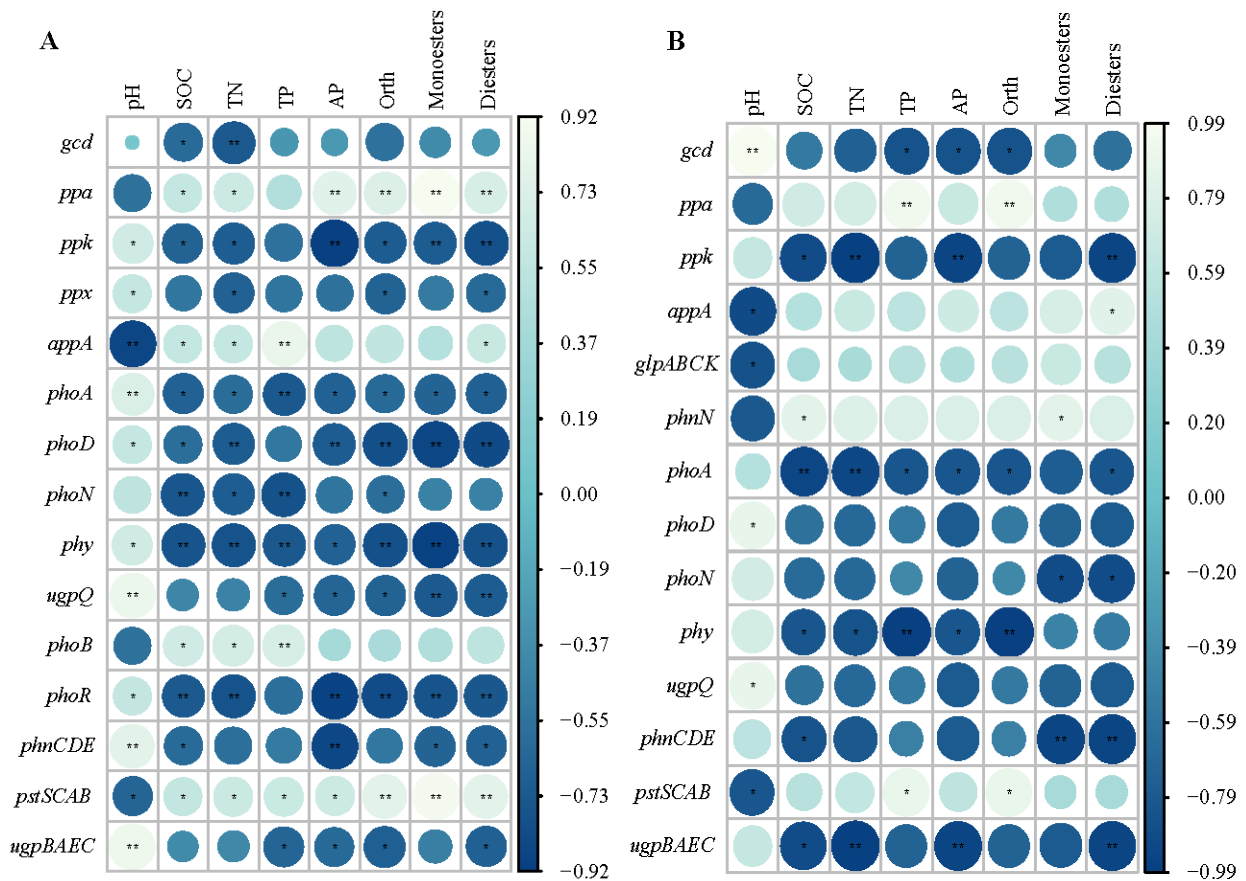


Fig. S8 Spearman correlation heatmap of P-cycling genes and soil nutrients in bulk soils (A) and water-extractable colloids (WECs, B)

The relative abundance of *phn* transporter systems was calculated as the average abundances of gene *phnC*, *phnD*, and *phnE*; the *pst* transporter systems was calculated as the average abundances of gene *pstS*, *pstC*, *pstA*, and *pstB*; The *ugp* transporter systems was calculated as the average abundances of gene *ugpB*, *ugpA*, *ugpE*, and *ugpC*. Soil properties include soil pH, soil organic carbon (SOC), total nitrogen (TN), total phosphorus (TP), available P (AP), orthophosphate (Orth), orthophosphate monoesters (Monoesters), orthophosphate diesters (Diesters). Asterisks indicate a significant association between the abundance of a given functional gene and particular soil properties: \*\*,  $p < 0.01$ ; \*,  $p < 0.05$ .