

1 **Unifying framework for assessing sensitivity of marine calcifiers to ocean alkalinity**
2 **enhancement categorizes responses and identifies biological thresholds - importance of**
3 **precautionary principle**

4 Nina Bednaršek^{1*}, Hanna van de Mortel^{2, 3}, Greg Pelletier⁴, Marisol García-Reyes⁵, Richard A.
5 Feely⁶, Andrew G. Dickson⁷

6 ^{1*}Cooperative Institute for Marine Ecosystem and Resources Studies, Hatfield Marine Science
7 Center, Oregon State University, 2030 SE Marine Science Drive Newport, OR 97365, USA

8 ²National Institute for Biology, Marine Biological Station, Fornace 41, Piran, Slovenia

9 ³HvdMortel Consulting, Utrecht, NL

10 ⁴Washington Department of Ecology, Olympia, 300 Desmond Dr SE, WA 98503, USA (Emeritus)

11 ⁵Farallon Institute, 101 St. Suite Q, Petaluma, CA 94952, United States

12 ⁶NOAA Pacific Marine Environmental Laboratory, Seattle, WA, 98115 USA

13 ⁷University of California at San Diego, Scripps Institution of Oceanography, 9500 Gilman Drive,
14 La Jolla, CA 92093, USA (Emeritus)

15 *Correspondence to:* nina.bednarsek@oregonstate.edu

16 **Abstract**

17 Ocean alkalinity enhancement (OAE), one of the marine carbon dioxide removal strategies, is
18 gaining recognition in its ability to mitigate climate change and ocean acidification (OA). OAE is
19 based on adding alkalinity to open-ocean and coastal marine systems through a variety of different
20 approaches, which raises carbonate chemistry parameters (such as pH, total alkalinity, aragonite
21 saturation state), and enhances the uptake of carbon dioxide (CO₂) from the atmosphere. There are
22 large uncertainties in both short- and long-term outcomes related to potential environmental
23 impacts, which would ultimately have an influence on the social license and success of OAE as a
24 climate strategy. This paper represents a synthesis effort, leveraging on the OA studies and
25 published data, observed patterns and generalizable responses. We propose a conceptual
26 framework of assessing responses that are predicted under OAE implementation. The synthesis
27 was done using raw experimental OA data based on 65 collected studies, covering 81 species and
28 capturing the responses of eleven biological groups (coralline algae, corals, dinoflagellates,
29 mollusks, gastropods, pteropods, coccolithophores, annelids, crustacean, echinoderms, and
30 foraminifera), using regression analyses to predict biological responses and thresholds to NaOH
31 or Na₂CO₃ additions. Predicted responses were categorized into six different categories (linear
32 positive and negative, threshold positive and negative, parabolic and neutral) to delineate species-
33 and group-specific responders. The results show that 40% of species are predicted to respond
34 positively (N=33), 31% of species negatively (N=25), and 29% (N=24) were found to demonstrate
35 a neutral response upon alkalinity addition. For the majority of negatively impacted species (84%),
36 biological thresholds corresponding to 10 to 500 μmol/kg NaOH addition were found, occurring
37 at much lower values than previously expected. We thus explicitly emphasize the importance of
38 including much lower additions of alkalinity in experimental trials to realistically evaluate *in situ*
39 biological responses. We also found that OAE could offset some of the negative effects of OA, whereby
40 50 to 100 μmol/kg of NaOH addition could compensate for the calcification loss between the pre-
41 industrial and current conditions. The ultimate goal of the study was to provide an assessment of
42 biological rates and thresholds predicted under NaOH/Na₂CO₃ additions that can serve as a tool
43 for delineating OAE risks, guiding and prioritizing future OAE biological research and regional
44 OAE monitoring efforts and communicate the risks with the stakeholders. This is pertinent given
45 the fact that at least some of the current regulatory frameworks likely do not assure safety
46 biological space. With 71% of species showing non-neutral response, a precautionary approach
47 for OAE implementation is warranted, identifying the conditions where potential negative
48 ecological outcomes could happen, which is key for scaling up while also avoiding ecological
49 risks.

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55 **1. Introduction**

56 Anthropogenic carbon dioxide (CO₂) emissions have increased at an unprecedented rate and have
57 contributed to global climate change and negative ecological and biogeochemical impacts in the
58 oceans (Feely et al., 2004; Gattuso et al., 2018), to the extent of crossing six different planetary
59 boundaries (Richardson et al., 2023). Oceans play a crucial role in attenuating the increase in
60 atmospheric CO₂ through the absorption of the excess atmospheric CO₂ of roughly a quarter of
61 anthropogenic carbon dioxide (CO₂) emissions, drawing down around 2–3 Pg C yr⁻¹ in recent
62 decades (Friedlingstein et al., 2022). However, without substantial CO₂ emissions abatement and
63 CO₂ removal strategies, profound repercussions on climate, extreme weather events, and
64 socioeconomic implications will follow. Ocean-based CO₂ removal and sequestration strategies
65 (broadly referred to as marine CDR) are among the proposed CDR approaches that remove CO₂
66 and store it for geologically relevant times (National Academies of Sciences, Engineering, and
67 Medicine, 2021). These mCDR approaches only complement CO₂ emission reductions and
68 contribute to the portfolio of climate response strategies needed to meet the global goal of limiting
69 warming to well below 2°C as established by the Paris Agreement. Various mCDR approaches
70 have unique benefits and costs but differ in their value depending on their state of implementation,
71 and whether they act globally and/or locally (Oschlies et al., 2023).

72 Ocean alkalinity enhancement (OAE) has the potential to mitigate climate change through
73 increasing ocean uptake of CO₂, while simultaneously reversing ocean acidification (OA), and
74 improving marine habitats. Despite mostly being in the concept stage, OAE is viewed with a high
75 level of confidence as to its effectiveness: medium on environmental risk, but low on the
76 underlying knowledge base (Eisaman et al., 2023; Gattuso et al., 2021; National Academies of
77 Sciences, Engineering, and Medicine, 2021). The primary concerns for OAE are large
78 uncertainties in both short- and long-term OAE outcomes related to potential environmental
79 impacts of OAE (Kheshgi, 1995; Bach et al., 2019), especially if OAE were to induce novel
80 conditions in the marine systems that are outside the range of the natural variability, exposing
81 organisms to conditions not experienced in their evolutionary history. The outcome of OAE as a
82 successful climate strategy depends on a thorough and advanced understanding of the impacts of
83 OAE implementation while avoiding negative biological effects.

84 **1.1 Leveraging ocean acidification research on marine calcifiers**

85 Increased CO₂ uptake, which initially is absorbed by the ocean as dissolved CO₂, causes a decline
86 in pH, shoaling of the saturation state horizon (Ω_{ar}) and reduced carbonate ion amount content in
87 a process termed ocean acidification (Feely et al., 2004), causing negative consequences to marine
88 biota, especially marine calcifiers, the structure and function of the vulnerable marine ecosystem,
89 and alteration of the carbon cycle. On the other hand, chemical changes induced by OAE are
90 inherently linked to reversing the OA process: increasing pH, shifting carbonate chemistry

91 speciation towards lower aqueous carbon dioxide ($p\text{CO}_2$) and higher carbonate ion (CO_3^{2-}) amount
92 contents, as well as higher aragonite saturation state (Ω_{ar}). Such changes could either be within the
93 ranges of the variability of the natural systems to which species are acclimatized, or outside them,
94 creating novel conditions for which species might not have developed suitable acclimation
95 strategies. As such, the biological outcomes are, due to their complexity, highly unpredictable.

96 Scientific progress in over 20 years of OA research has brought substantial insights into the
97 biological effects, with the most fundamental outcome being that calcifying organisms would be
98 primarily affected (Riebesell and Gattuso, 2015), with the calcification process being one of the
99 most susceptible pathways, underpinned by species differences in calcification mechanisms (Ries
100 et al., 2009; 2011; Bach et al., 2013; 2015; Leung et al., 2022). However, OA focused heavily on
101 investigating biological effects on the more acidic range of the carbonate chemistry conditions
102 predicted under future scenarios and most of the studies focused on manipulating the level of $p\text{CO}_2$
103 rather than alkalinity. This resulted in poor understanding of the biological effects at the higher pH
104 end of the carbon chemistry range (Renforth and Henderson, 2017). Some biological inferences
105 can be made based on the understanding of the physiological mechanisms underlying the
106 calcification mechanisms (Bach et al., 2019), but such insights are not adequate to provide
107 sufficient understanding. Despite the lack of biological data at the upper ranges of pH and Ω_{ar} , this
108 study builds on the premise that previous OA studies could be leveraged for assessment of
109 biological responses under OAE. Comparative experimental work, meta-analyses, and the
110 threshold work (Kroeker et al., 2013; Leung et al., 2022; Bednaršek et al., 2019; 2021b,c) have
111 indicated that even very diverse responses can be grouped into categorical responses.

112 Calcification is a primary pathway of the organismal sensitivity to OA, which can act as an early
113 warning response, and is directly implicated in growth and (abnormal) development across most
114 of the marine calcifiers, while it also underlies the ecological success of numerous marine
115 calcifiers. Calcification also underlies the ecological success of numerous marine calcifiers
116 because it directly addresses the level of susceptibility to predation, which could lead to altered
117 size of the overall population. Studies also clearly show that the threshold for calcification occurs
118 at similar pH/ Ω values as the thresholds for metabolic and energy metabolism processes (Lutier et
119 al., 2022; Bednaršek et al., 2019; 2021b,c). It is also directly implicated in the carbon export with
120 significant biogeochemical implications that could also impact OAE efficiency. This study aims
121 to systematically assess species responses predicted under carbonate-based OAE compound
122 additions and categorize them based on calcification rate responses.

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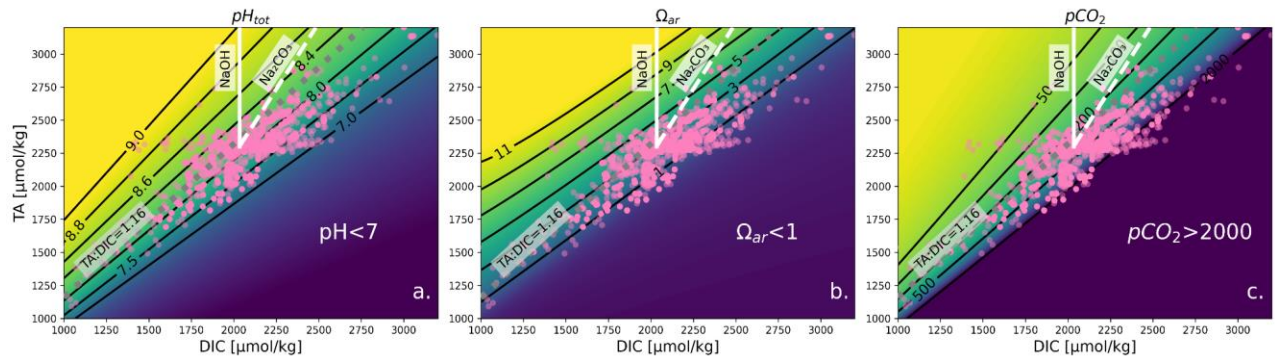
124 **1.2. Complex carbonate chemistry changes induced by various OAE compounds**

125 Various OAE compounds added to the water change carbonate chemistry in a multifaceted way
126 and require complex calculations of a multi-parameter problem. As the values of TA and DIC
127 change, a variety of other parameters, such as pH, CO_3^{2-} , Ω_{ar} , and $p\text{CO}_2$, exhibit approximately

128 linear relationships, with slopes that vary along these lines (see Fig. 1). This means that if TA and
 129 DIC vary in proportion to one another, then the values of these displayed parameters hardly change
 130 at a particular salinity, temperature, and pressure. With TA, DIC and the hydrographic conditions
 131 (salinity, temperature and pressure), one can fully constrain the carbonate system. Our method
 132 requires us to have *one* variable constraining the entire carbonate system. TA and DIC have the
 133 benefit that they can both be directly measured with high accuracy or calculated from other
 134 carbonate parameters. They are also both directly linked to OAE, as we are enhancing the TA
 135 which then allows DIC to increase over time due to the gradual uptake of atmospheric CO₂.

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 137 To demonstrate the changes of carbonate system in the experimental system, Figure 1 shows the
 138 changes in carbonate parameters with the addition of two OAE compounds, i.e. NaOH (black line)
 139 and Na₂CO₃ (dotted line) to seawater. When NaOH is added, only TA increases and when Na₂CO₃
 140 is added, TA and DIC increase at a 2:1 ratio. This results in corresponding changes in pH (Fig.
 141 1a), Ω_{ar} (Fig. 1b) and pCO₂ (Fig. 1c) and shows how much of a change is required to bring the
 142 system back to equilibrium with respect to the atmosphere.

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 147 **Figure 1:** The effect of changes in TA and DIC on the properties of seawater ($S = 34.68$, $T = 16^\circ\text{C}$,
 148 $[\text{SiO}_2] = 50 \mu\text{mol/kg}$, $[\text{PO}_4^{3-}] = 0.5 \mu\text{mol/kg}$, $\text{TA} = 2303 \mu\text{mol/kg}$, $\text{DIC} = 2034 \mu\text{mol/kg}$), adapted
 149 from Schulz et al. (2023). Pink dots represent experimental TA and DIC data used in our synthesis.
 150 Subfigures show pH_{tot} , aragonite saturation state and pCO_2 . Calculations were carried out with
 151 the Python version of CO2SYS (Humphreys et al., 2022) using the stoichiometric dissociation
 152 constants for carbonic acid from Sulpis et al. (2020), for sulfuric acid by Dickson et al. (1990) and
 153 for total boron from Uppström (1974). The dotted gray line represents the pre-industrial TA:DIC
 154 of 1.16. The solid white line indicates the effect of adding NaOH and the dashed white line
 155 indicates the effect of adding Na₂CO₃. This grouping of lines can be translated so that its initial
 156 position moves elsewhere to visualize different initial conditions. Note that at $\text{TA} < 1000 \mu\text{mol/kg}$
 157 and $\text{DIC} < 500 \mu\text{mol/kg}$ this correspondence no longer holds true when considering Ω_{ar} , however,
 158 such conditions are rare in the ocean and not widely applicable. The same contour plot utilizing
 159 GLODAP data plotted instead of experimental data is shown in Supplement Figure 1.

160

161 **1.3 Testable conceptual framework based on the existing OA studies**

162 Based on Ries et al. (2009), calcification responses can be categorized into six categories (Fig. 2):
163 linear positive or negative response; threshold positive or negative response (exponential fit);
164 parabolic response; and neutral (no significant) response. We hypothesize that these categories of
165 responses based on ocean acidification data and delineated by Ries et al. (2009, 2011), could also
166 be applicable to OAE dosing. For this meta-analysis, we have undertaken three steps: first,
167 synthesize carbonate chemistry data at regional and global scales to obtain TA, DIC and Ω_{ar}
168 correlations; second, conduct a literature review and collect available data from OA literature
169 related to the calcification rate responses across the species of eleven groups of marine calcifiers;
170 and third, run regression analyses and determine the category of calcification rate response to
171 TA:DIC, further extending it with addition of NaOH and Na₂CO₃.

172 The most accurate way of predicting the responses to OAE addition is done based on the
173 mechanistic understanding of calcification response to specific carbonate chemistry parameter(s).
174 The hypothesis was that if mechanistic relationships with identified carbonate chemistry driver(s)
175 are available for species, calcification rate under various feasible OAE scenarios can be predicted
176 with greater accuracy and lower uncertainty. We further focused on investigating if the empirical
177 results were consistent with mechanistic calcification predictions for a few selected species for
178 which the mechanisms were known.

179

180 Here, we demonstrate the TA:DIC relationship with biological outcomes and show the application
181 for the TA:DIC thresholds beyond which the responses become negative. Ultimately, we
182 synthesize which calcifying species or groups are predicted to benefit or loss due to OAE, what
183 constitutes a species-specific safe operating space related to the OAE and we delineate what
184 experiments are most urgently needed to fill in critical knowledge gaps before massive OAE field
185 implementation can be considered.

186 **2. Methodology**

187 **2.1 Literature review of data on marine calcification impact by OA**

188 To assess the impact of OAE on a range of marine calcifiers, we used existing studies on marine
189 species calcification response that had aligned raw biological (calcification rate) data along with
190 the carbonate chemistry. We searched within Scopus, Web of Science, and PubMed and then used
191 the datasets that were archived in NCEI, OA-ICC and Pangaia. Through personal correspondence,
192 we have additionally contacted lead authors of the studies, whose data are not or are insufficiently
193 archived, mostly to validate the predicted response. If we received data through this procedure, we
194 explicitly acknowledged this in the dataset. These searches for the biological datasets related to
195 calcification rate and corresponding carbonate chemistry were carried out until November 2023
196 and cover 70 existing studies, with the aim to cover a wide range of calcifying organisms across
197 various functional groups and 84 species. For several functional groups data was easy to find

198 (algae, coccolithophores, corals, foraminifera, mollusks and dinoflagellates), so no new studies
199 were added after 10 to 15 studies were found. Seven studies were found for pteropods, four for the
200 echinoderm and gastropod groups, three for crustaceans and one for annelids. When reviewing the
201 literature, we included data from the OA experimental studies related to the physical-chemical
202 parameters (temperature, salinity, DIC, TA) and biological data related to calcification rate.

203 **2.2 Use of TA:DIC instead of Ω_{ar}**

204 Understanding the change in carbonate chemistry upon the OAE addition is essential for the
205 biological experimentalists who are conducting biological assessments to report on the OAE
206 effects. However, complex changes in the carbonate chemistry induced by the OAE addition are
207 not intuitive or straightforward; in fact, they are multi-parameter problems that require complex
208 carbonate chemistry calculations. Using the TA:DIC ratio is a more practical way of looking at the
209 impacts of the OAE treatment instead of using a single carbonate parameter because of the highly
210 correlative relationship between TA:DIC and other carbonate system parameters (see Fig. 1).

211 With TA, DIC and the hydrographic conditions (salinity, temperature and pressure), one can fully
212 constrain the carbonate system. Our method allows *one* variable constraining the entire carbonate
213 system. TA and DIC have the benefit that they can both be directly measured or calculated from
214 other carbonate parameters (e.g., pCO₂, pH). They are also both directly linked to OAE, as we are
215 enhancing the TA which then allows DIC to increase over time due to the gradual uptake of
216 atmospheric CO₂ (Fig. 1 shows the changes in the carbonate chemistry system upon NaOH and
217 Na₂CO₃ additions).

218 As such, our focus was on simplifying the steps to express the results when conducting the
219 experimental work, and subsequently, the reporting of the responses, with the aim to reduce the
220 multi-parameter problem into a one-parameter simplification. This step reduces multiple degrees
221 of freedom into just two, i.e. TA and DIC, with the ratio allowing us to consider this as a 1-
222 parameter problem. As such, TA:DIC is a simplistic and convenient way of describing the system,
223 where we only need to understand the change in TA and DIC ratio, which is feasible for every
224 OAE compound added to the experimental system. In addition, TA:DIC is also the best
225 approximation for the CO₃²⁻ concentration. The insights from multiple biological experimental
226 studies show that the CO₃²⁻ concentration is the representative driver of the calcification process
227 for multiple calcifying groups, although not all, compared to aragonite saturation state (Ω_{ar}), which
228 represents an empirical approximation. Furthermore, by using TA:DIC we do not have to choose
229 a particular parameter to describe the changes in calcification. It could also work for the species in
230 which other parameters drive the calcification, e.g. bicarbonate in autotrophic, Ω_{ar} in bivalves and
231 H⁺ flux in foraminifera. In that way, we standardize all the parameters that would otherwise
232 influence the carbonate system and come up with a more uniform way to express the experimental
233 conditions, which would then be useful for easier comparisons among the conducted experiments.

234 **2.3 Experimental biological and biogeochemical data**

235 Based on the collected data, the range of pH and Ω_{ar} experimental conditions used and their
236 TA:DIC relationship was determined (Supplement Fig. 2). Most studies covered pH conditions
237 from 7.5 to 8.5 and Ω_{ar} up to 5, with a few studies increasing pH up to 9 and exceeding Ω_{ar} of 10.
238 This indicates the potential of leveraging such experimental studies as a baseline for predictive
239 regression models of biological responses to a range of Ω_{ar} conditions, as expected under OAE
240 studies.

241 Once biological data was compiled, units were standardized where possible. The main issue when
242 compiling data was the lack of standardization of the calcification rates. A variety of calcification
243 rate units were used across different studies. Where possible, the units were converted to mmol of
244 $\text{CaCO}_3 \text{ g weight}^{-1} \text{ hr}^{-1}$. However, the data required to do so was not always readily available. Other
245 units used for calcification rate were mmol of $\text{CaCO}_3 \text{ m}^{-2} \text{ h}^{-1}$ and mmol of $\text{CaCO}_3 \text{ m}^{-3} \text{ hr}^{-1}$, and there
246 was also data used as an indication of calcification rate with units mmol/# h^{-1} , mmol h^{-1} , mmol/#
247 h^{-1} , mmol cm^{-2} , % h^{-1} , where ‘#’ indicates one individual. Growth rates and PIC production rates
248 were used as indicators of calcification rate for single-cell organisms. For some species, direct
249 calcification rates were not reported in the literature, instead only relevant parameters related to
250 calcification (shell length, density, thickness) over the time were available from the experimental
251 studies. The decision was made to also collect these additional datasets because the statistical
252 analyses of this study focus on the trend in the absolute numbers and would not change by being
253 transformed into the rates. Data were analyzed on a species level. Where there were multiple
254 studies available for the calcification rate of one species using the same rate units, the data were
255 combined (for example, the coccolithophores).

256 **2.4 Sorting species-specific responses into categories per calcification response**

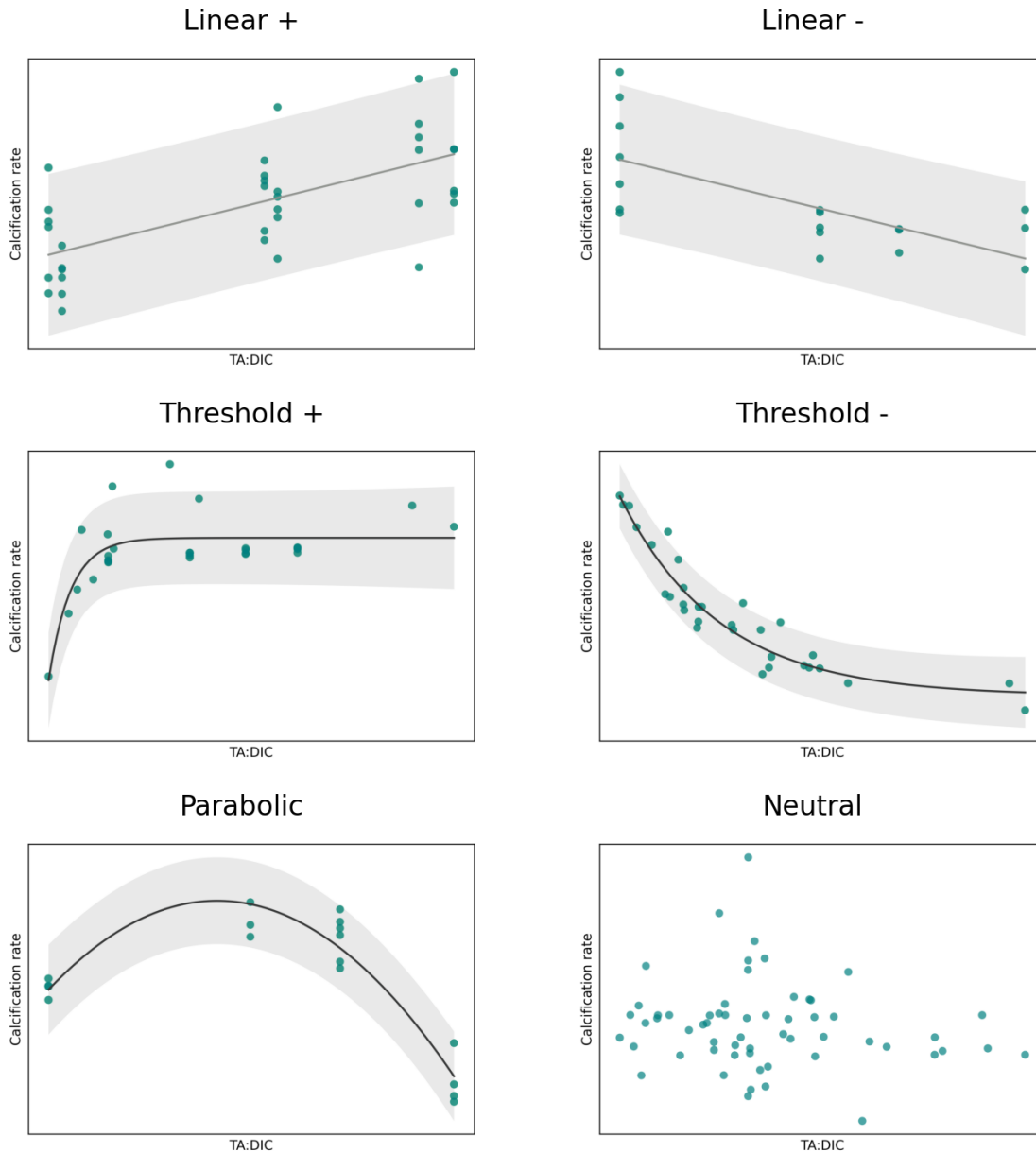
257 Responses were split into 6 categories: linear positive and linear negative, parabolic, threshold
258 positive and negative, and neutral, whatever was the best as determined by their respective
259 regression models (using the ordinary least squares method in Statsmodels for Python, see Seabold
260 et al. (2010)). See Fig. 2 for an overview of these responses of calcification rate to increasing
261 TA:DIC ratio.

262 The final response for each species was determined by the regression with the lowest p-value. This
263 method is in contrast with the Ries et al. (2009) study where they chose the regression analysis
264 that yielded the lowest square root of the mean squared error (RMSE) for a given species and that
265 was statistically significant ($p \leq 0.05$). When applying their method to our data, parabolic and
266 exponential regressions were always favored over linear regressions. When examining these
267 regressions, we found that choosing the best fit based on the lowest p-value yielded better fits, as
268 this method prevents overfitting due to noise in the data. Where a linear regression had the best fit,
269 we assigned a linear response, which could be either positive or negative based on the slope. The
270 species with a significant exponential fit were categorized as threshold positive (+) or threshold
271 negative (-), which was distinguished from the parabolic response with the fitted parabolic curve.

272 The best fit regression was assigned to each species and plotted, but only if the p-value was
273 considered significant, i.e. lower than 0.05. These regressions were plotted along with a 90%
274 prediction interval, which accounts for the variability of the experimental data. The species with a
275 p-value > 0.05 were categorized as having no correlation.

276 We note that if there were multiple datasets obtained from different studies for the same species
277 and rate units could not be combined, we report p-value and RMSE for each of the studies and
278 take all responses across the studies into consideration. Even if there were different observed
279 calcification responses for the same species in different studies, we avoided making a judgment
280 on the studies by choosing an overall response since the heterogeneity among the studies can be
281 true, especially when considering that species from different regional settings (as represented in
282 the studies) might be differentially physiological acclimatized and genetically adapted to the range
283 of carbonate chemistry conditions.

284 The TA:DIC threshold was computed to indicate the point at which the current calcification rate
285 (i.e. the calcification rate at the baseline) is reduced by a half for linear negative, threshold negative
286 and parabolic responders. The thresholds and the amount of NaOH and Na₂CO₃ required (starting
287 point at 10 μmol/kg in the addition of 50 μmol/kg) to reach this threshold were determined. For
288 parabolic responders, the inflection points that tell us when the rate is predicted to change slope
289 are also included in the Supplementary Table 1. Once the species' responses were determined, an
290 attempt was made to group them based on functional groups. However, since species within the
291 same functional group had varying responses, grouping them together meant these responses were
292 no longer visible due to a wide spread of data. Therefore, most of the analysis remained on the
293 species level (Table 1).



294

295 **Figure 2:** Overview of the categories of responses between carbonate chemistry parameters
 296 (TA:DIC) and calcification rate: linear positive (calcification increase at higher TA:DIC), linear
 297 negative (calcification decrease at higher TA:DIC), exponential for the threshold positive
 298 response (calcification increase with plateauing at higher TA:DIC), exponential for the threshold
 299 negative response (calcification decline with plateauing at lower TA:DIC) and parabolic
 300 (calcification increase followed by a decrease at higher TA:DIC). Responses were only considered
 301 significant when $p < 0.05$, otherwise they were categorized as neutral.

302 **2.5 Conceptual framework to evaluate increases in TA:DIC**

303 The regression models applied to each species could be used to predict calcification rates at higher

304 TA:DIC ratio. We conceptually added alkalinity from the current calcification rate baseline. This
305 baseline was computed for each species using CO2SYS with $p\text{CO}_2 = 425$ ppm and $\text{pH}_{\text{tot}} = 8.1$, for
306 a temperature of 20°C and the average salinity for each species, based on their respective OA
307 dataset(s) (see Supplement Table 2). All CO2SYS calculations in this study were carried out with
308 the Python version of CO2SYS (Humphreys et al., 2022) using the stoichiometric dissociation
309 constants for carbonic acid from Sulpis et al. (2020), for sulfuric acid by Dickson et al. (1990) and
310 for total boron from Uppström (1974). From this baseline, TA was added in the form of both NaOH
311 and Na_2CO_3 . These two compounds were chosen as they differentially change the carbonate
312 chemistry settings, with NaOH changing TA:DIC in the 1:1 ratio, and Na_2CO_3 inducing a 2:1
313 TA:DIC change. For example, $10 \mu\text{mol/kg}$ of NaOH addition will increase TA by $10 \mu\text{mol/kg}$ and
314 not affect DIC. For Na_2CO_3 , $10 \mu\text{mol/kg}$ addition will increase TA by $10 \mu\text{mol/kg}$ and increase
315 DIC by $5 \mu\text{mol/kg}$. Figure 1 demonstrates the usefulness of this approach. For both NaOH and
316 Na_2CO_3 , $10 \mu\text{mol/kg}$ was conceptually added using the principles of mass balance approach for
317 the carbonate system via CO2SYS. This was repeated for increments of $100 \mu\text{mol/kg}$ up until a
318 total of $500 \mu\text{mol/kg}$. The new TA:DIC ratios were estimated using the ratio of the new TA and
319 DIC by adding the direct effect of ΔTA and ΔDIC due to chemical additions of NaOH (assume
320 $\Delta\text{DIC}=0$) or Na_2CO_3 (assume $\Delta\text{DIC}=0.5*\Delta\text{TA}$). A maximum of $500 \mu\text{mol/kg}$ was chosen to have
321 more realistic additions of TA that resemble those appropriate within the OAE field trials (e.g.
322 Wang et al., 2023). With the new TA:DIC ratios after TA addition, the species' regression models
323 based on the fitted OA response data were used to compute respective calcification rates (note that
324 added points with NaOH or Na_2CO_3 were not calculated as part of the regression). These data
325 points were all plotted along with the experimental data, regression model and prediction intervals
326 as shown in Fig. 3.

327 We also determine the amount of NaOH needed to reach pH 9 for each study. This was computed
328 for each species using CO2SYS starting from $p\text{CO}_2 = 425$ ppm and $\text{pH}_{\text{tot}} = 8.1$, for a temperature
329 of 20°C and the average salinity in the increments of $500 \mu\text{mol/kg}$ until desired pH was not
330 reached. We have conducted this step for all the studies involving negative responders, as it does
331 not negatively impact positive or neutral responders.

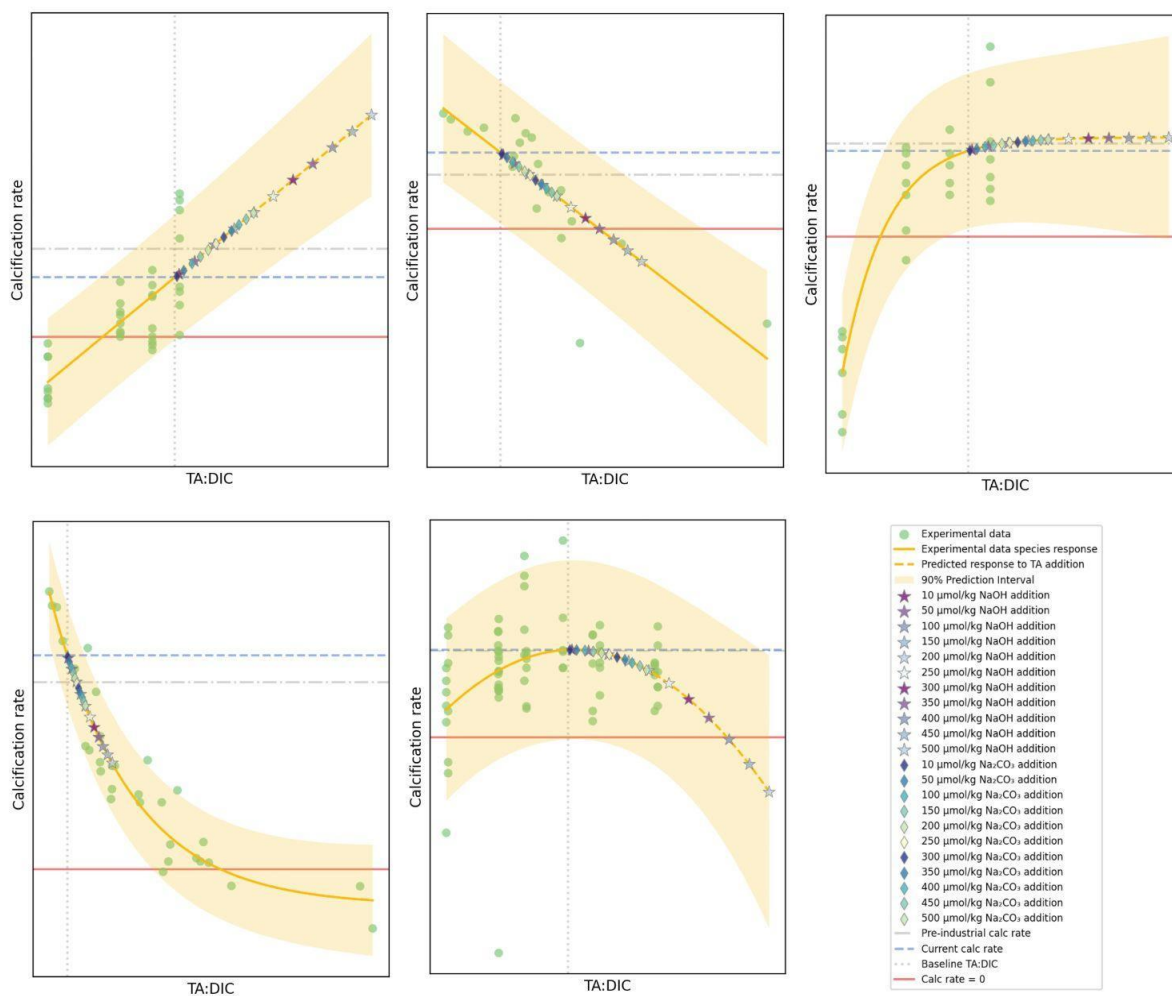
332 **2.6 Evaluation of the biological responses based on the OAE addition**

333 The species with significant correlations were grouped visually based on their best-fit regression
334 models and are classified into positive, negative, and neutral as the following:

335 1) *Positive responders*: species with predicted linear positive and threshold positive calcification
336 rate response corresponding with increased TA addition.

337 2) *Negative responders*: species with predicted *linear negative, parabolic and threshold negative*
338 *response* in calcification rate upon (a certain amount of) TA addition. For the parabolic and
339 threshold negative response, a concentration of NaOH was determined that indicates the threshold
340 in TA:DIC ratio beyond which the response becomes negative. Additionally, NaOH concentration
341 was determined to reduce the calcification rate to a half, with the threshold at the corresponding

342 TA:DIC.
 343 3) *Neutral responders*: species with *no significant correlation* ($p < 0.05$) in calcification rate upon
 344 TA addition.



345
 346 **Figure 3:** *Conceptual diagrams for five types of responses (linear positive, negative,*
 347 *exponential positive, negative and parabolic) plotted with experimental data from OA studies*
 348 *(green dots), predicted values at various additions of alkalinity (stars), the regression line and*
 349 *prediction error margins fitted for a given species. The red line indicates zero net dissolution*
 350 *(calcification rate is equal to 0; dissolution rate = calcification rate). The dotted lines indicate*
 351 *the pre-industrial (TA:DIC = 1.16) and current calcification rate (TA:DIC ≈ 1.12), in gray and*
 352 *blue, respectively.*

353 **2.7 Determining threshold values indicative of negative biological response to OAE**

354 The metrics to evaluate the sensitivity of calcification rate of the negative responders in this study
355 were based on the amount of NaOH or Na₂CO₃ addition required to reduce the current calcification
356 rate by a half. The greater the TA:DIC ratio value to trigger half calcification rate reduction, the
357 less sensitive species was to NaOH addition. We refer to this TA:DIC ratio as the biological
358 threshold, which we also report along with corresponding pH and Ω_{ar} and the associated
359 uncertainty. TA:DIC thresholds were converted to their respective pH and Ω_{ar} , which are affected
360 by temperature and salinity. To calculate threshold pH and Ω_{ar} we normalized for a temperature of
361 20°C and used the average salinity per species. This is because most experiments were done at
362 constant salinities, but at varying temperatures.

363 **2.8 Extraction of the carbonate chemistry data from the GLODAP dataset**

364 We extracted total alkalinity, dissolved inorganic carbon, Ω_{ar} , and pH_{tot} from the Global Ocean
365 Data Analysis Project GLODAPv2.2023 dataset (<https://glodap.info>). We used regression in
366 MATLAB with a second-order polynomial equation to predict Ω_{ar} from the TA:DIC. The
367 regression analysis was performed using data from various depth intervals (0–10m, 0–30m, 0–
368 50m, 0–100m, 0–200m) regionally and globally. The regional analysis divided the global oceans
369 into the following groupings: Arctic (north of 65°N), Southern (south of 40°S), North Pacific
370 (north of 40°N), Central Pacific (40°S to 40°N), North Atlantic (North of 40°N), Central Atlantic
371 (40°S to 40°N), and Indian Ocean (north of 40°S).

372 **2.9. Calculating calcification in the pre-industrial times**

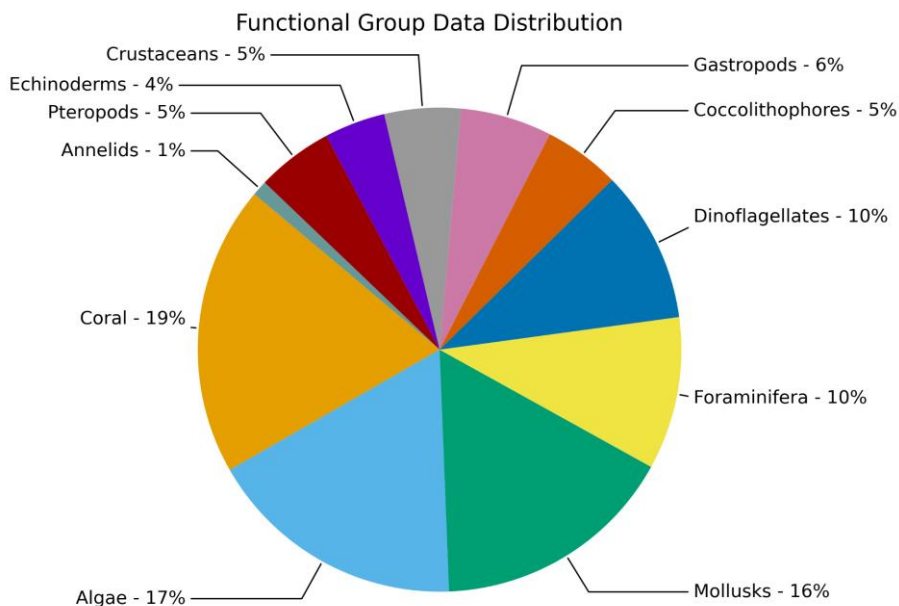
373 With respect to the changes since the pre-industrial times, the aim was to examine the difference
374 in calcification between current and pre-industrial, and to what extent NaOH addition is required
375 to compensate for this difference. This was done by first inferring the industrial TA:DIC ratio of
376 1.16 (Feely et al., 2004) vs. a current TA:DIC of 1.12 (derived based on the current pCO₂ and
377 global pH surface ocean value) and using the regression lines of TA:DIC vs. calcification rate to
378 calculate the corresponding calcification rates (Fig. 3). In the next step, calcification rate was
379 calculated due to the addition of NaOH and Na₂CO₃ from the species-specific baselines (see
380 Method Section 2.5 for detailed explanation), specifically for the positive responders. This was
381 done using the principles of mass balance approach for the carbonate system via CO2SYS, where
382 the carbonate system is calculated for each increment of NaOH or Na₂CO₃ added. The difference
383 between the pre-industrial calcification and current, increased by the NaOH was calculated and
384 compared on the species level.

385 **3. Results**

386 **3.1 Data collection for the calcification rate responses of different biological groups**

387 We examined 70 datasets, which covered 84 different species that were divided into 11 different
388 groups (Fig. 4). These functional groups were corals (19% of datasets), calcifying algae (17%),

389 mollusks (16%), foraminifera (10%), dinoflagellates (10%), coccolithophores (5%), gastropods
 390 (6%), crustaceans (5%), echinoderms (4%), pteropods (5%), and annelids (1%). In the mollusks
 391 group, we have separated out the gastropod and pteropod because of a higher number of studies
 392 that explicitly cover these two groups. The group of gastropods refers to all gastropods that are not
 393 pteropods. If all three groups were combined (mollusks, gastropods, pteropods), this group would
 394 be the largest.



395
 396 **Figure 4:** Percent of studies for multiple groups (N=11) with available data for the calcification
 397 rate responses as part of data compilation of 70 studies covering 84 species).

398 3.2 Species-specific responses to NaOH/Na₂CO₃ addition

399 Calcification rate responses of species from different groups were correlated to TA:DIC and
 400 summarized to obtain calcification rate response. The calcification rate responses encompassed
 401 linear (positive and negative), threshold (positive and negative), parabolic, and neutral responses,
 402 with the slope and the intercept of the response determining the type and the magnitude of the
 403 response. We present fitted responses of calcification rate per TA:DIC ratio for each examined
 404 species (Table 1; Supplement Fig. 3). When possible, we fit a regression to multiple datasets of
 405 the same species that used the same units. We also present the response with the additions of NaOH
 406 and Na₂CO₃ for each species per examined study and corresponding rate unit and their biological
 407 TA:DIC thresholds (Table 2; Supplement Table 3).

408 **Table 1:** The summary of all the OA studies from which the chemical and biological data was
 409 collected, including the name of the species and group and the accompanying calcification rate
 410 unit. The response for each species and rate unit was determined by the regression with the lowest
 411 p-value, where the p-value was smaller than 0.05. These responses include p-value, goodness of
 412 fit (R^2) and Root Mean Square Error (RMSE) in this table. Non-significant responses are

413 categorized as having a ‘neutral’ response. The type of regression (linear positive or negative,
 414 threshold positive or negative, parabolic, and neutral) as well as an overall response (positive,
 415 negative, neutral) per species is indicated.

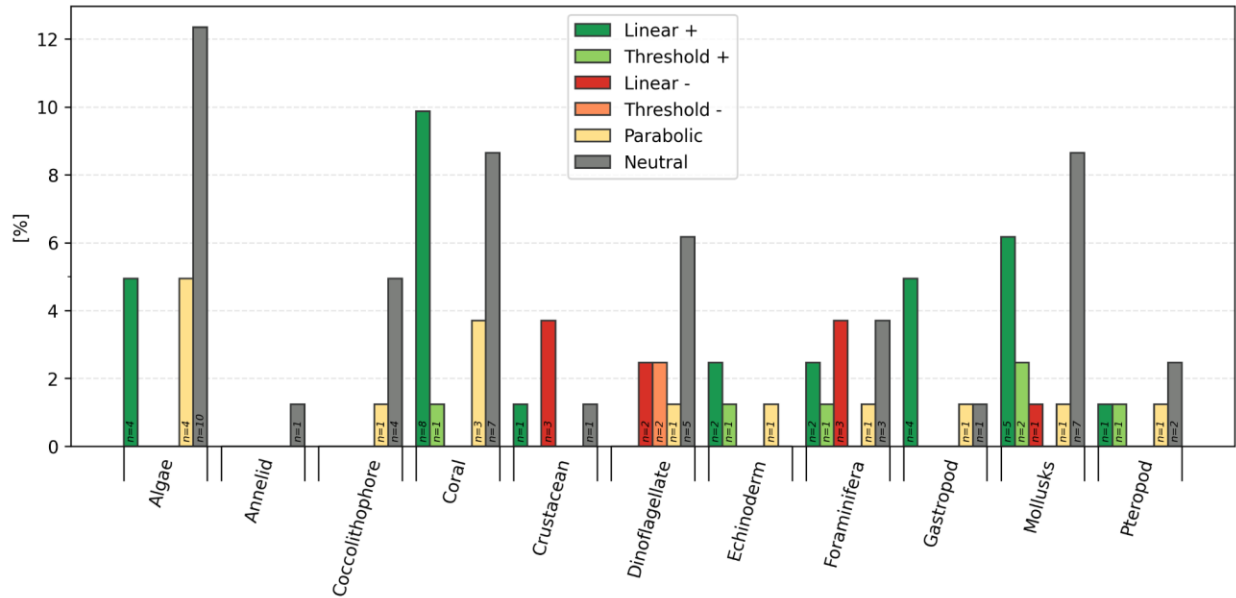
Studies	n	Group	Species	Rate unit	Response	Pos/Neg/ Neut	p- value	R2	RMSE
Vasquez-Elizondo et al. (2016)	4	Algae	<i>Amphiroa tribulus</i>	mmol/m ² /hr	neutral	Neutral			
Sinutok et al. (2011)	16	Algae	<i>Halimeda cylindracea</i>	mmol/hr	neutral	Neutral			
Comeau et al. (2013)	71	Algae	<i>Halimeda macroloba</i>	mmol/g/hr	parabolic	Negative	0.0127	0.1200	0.0028
Meyer et al. (2016)	24	Algae	<i>Halimeda macroloba</i>	mmol/m ² /hr	neutral	Neutral			
Sinutok et al. (2011)	16	Algae	<i>Halimeda macroloba</i>	mmol/hr	parabolic	Negative	0.0108	0.5000	0.0001
Comeau et al. (2013)	62	Algae	<i>Halimeda minima</i>	mmol/g/hr	neutral	Neutral			
Meyer et al. (2016)	24	Algae	<i>Halimeda opuntia</i>	mmol/m ² /hr	linear +	Positive	0.0080	0.2800	0.0222
Comeau et al. (2013)	72	Algae	<i>Hydrolithon reinboldii</i>	mmol/g/hr	linear +	Positive	0.0053	0.1100	0.0026
Cornwall et al. (2018)	23	Algae	<i>Hydrolithon reinboldii</i>	mmol/m ² /hr	neutral	Neutral			
Comeau et al. (2013)	72	Algae	<i>Lithophyllum flavescens</i>	mmol/g/hr	neutral	Neutral			
Johnson et al. (2021)	420	Algae	<i>Lithophyllum sp.</i>	mmol/g/hr	linear +	Positive	0.0000	0.1000	0.1136
Vasquez-Elizondo et al. (2016)	4	Algae	<i>Lithothamnion sp.</i>	mmol/m ² /hr	neutral	Neutral			
Monserrat et al. (2022)	62	Algae	<i>Neogoniolithon brassica- florida</i>	mmol/m ² /hr	neutral	Neutral			
Ries et al. (2009)	42	Algae	<i>Neogoniolithon sp.</i>	mmol/g/hr	parabolic	Negative	0.0000	0.4100	0.0003
Vasquez-Elizondo et al. (2016), Comeau et al. (2018)	26	Algae	<i>Neogoniolithon sp.</i>	mmol/m ² /hr	neutral	Neutral			
Briggs-Carpenter et al. (2019)	425	Algae	<i>Porolithon onkodes</i>	mmol/m ² /hr	linear +	Positive	0.0010	0.0300	0.8093
Comeau et al. (2018, 2019)	64	Algae	<i>Sporolithon durum</i>	mmol/m ² /hr	parabolic	Negative	0.0012	0.2000	0.1704
Ries et al. (2009)	41	Annelid	<i>Hydroides crucigera</i>	mmol/g/hr	neutral	Neutral			
Fiorini et al. (2011), Langer et al. (2006, 2011)	14	Cocco.	<i>Calcidiscus leptoporus</i>	mmol/#/hr	neutral	Neutral			
Langer et al. (2006)	6	Cocco.	<i>Calcidiscus leptoporus</i>	mmol/#	neutral	Neutral			
*	233	Cocco.	<i>Emiliana huxleyi</i>	mmol/#/hr	parabolic	Negative	0.0000	0.1600	0.0000
Casareto et al. (2009)	14	Cocco.	<i>Pleurochrysis carterae</i>	mmol/m ³ /hr	neutral	Neutral			
White et al. (2018)	118	Cocco.	<i>Pleurochrysis carterae</i>	mmol/#	neutral	Neutral			
Meyer et al. (2016)	24	Coral	<i>Acropora millepora</i>	mmol/m ² /hr	neutral	Neutral			
Camp et al. (2017), Comeau et al. (2013)	74	Coral	<i>Acropora pulchra</i>	mmol/m ² /hr	parabolic	Negative	0.0000	0.2900	1.3257
Agostini et al. (2021)	18	Coral	<i>Acropora solitaryensis</i>	mmol/m ² /hr	neutral	Neutral			
Comeau et al. (2018), Comeau et al. (2019)	81	Coral	<i>Acropora yongei</i>	mmol/m ² /hr	linear +	Positive	0.0000	0.2900	1.9447
Bove et al. (2020)	27	Coral	<i>Duncanopsammia axifuga</i>	mmol/m ² /hr	linear +	Positive	0.0016	0.3300	5.0785
Cornwall et al. (2018)	44	Coral	<i>Goniopora sp.</i>	mmol/m ² /hr	neutral	Neutral			
Maier et al. (2009)	237	Coral	<i>Lophelia pertusa</i>	mmol/g/hr	linear +	Positive	0.0030	0.0400	0.0002
Bove et al. (2020)	65	Coral	<i>Montastraea cavernosa</i>	mmol/m ² /hr	linear +	Positive	0.0154	0.0900	0.5047
Ries et al. (2009)	54	Coral	<i>Oculina arbuscula</i>	mmol/g/hr	parabolic	Negative	0.0000	0.8600	0.0001
Comeau et al. (2013)	72	Coral	<i>Pavona cactus</i>	mmol/m ² /hr	parabolic	Negative	0.0002	0.2200	0.9093
Comeau et al. (2019)	49	Coral	<i>Plesiastrea versipora</i>	mmol/m ² /hr	linear +	Positive	0.0069	0.1500	0.6003
Brown et al. (2022)	4	Coral	<i>Pocillopora damicornis</i>	mmol/g/hr	neutral	Neutral			
Comeau et al. (2013, 2018), Putnam-Gates et al. (2015)	117	Coral	<i>Pocillopora damicornis</i>	mmol/m ² /hr	neutral	Neutral			
Evensen-Edmunds et al. (2016)	60	Coral	<i>Pocillopora verrucosa</i>	mmol/m ² /hr	linear +	Positive	0.0132	0.1000	0.8297
Agostini et al. (2021)	18	Coral	<i>Porites heronensis</i>	mmol/m ² /hr	neutral	Neutral			
Comeau et al. (2013)	72	Coral	<i>Porites rus</i>	mmol/m ² /hr	linear +	Positive	0.0020	0.1300	2.0281
Okazaki et al. (2013)	75	Coral	<i>Siderastrea radians</i>	mmol/m ² /hr	linear +	Positive	0.0004	0.1600	2.7886
Okazaki et al. (2013)	64	Coral	<i>Solenastrea hyades</i>	mmol/m ² /hr	threshold +	Positive	0.0004	0.2300	2.0385
Krueger et al. (2017)	36	Coral	<i>Stylophora pistillata</i>	mmol/m ² /hr	neutral	Neutral			
Pansch et al. (2014)	36	Crust.	<i>Amphibalanus improvisus</i>	mmol/g/hr	linear +	Positive	0.0000	0.4300	0.0004
Ries et al. (2009)	36	Crust.	<i>Callinectes sapidus</i>	mmol/g/hr	linear -	Negative	0.0000	0.4000	0.0082
Ries et al. (2009)	18	Crust.	<i>Homarus americanus</i>	mmol/g/hr	linear -	Negative	0.0014	0.4800	0.0079
Ries et al. (2009)	12	Crust.	<i>Penaeus plebejus</i>	mmol/g/hr	linear -	Negative	0.0124	0.4800	0.0006
Findlay et al. (2010)	6	Crust.	<i>Semibalanus balanoides</i>	mmol/g/hr	neutral	Neutral			
Tatters et al. (2013)	45	Dino.	<i>Alexandrium sp.</i>	1/hr	neutral	Neutral			
Hansen et al. (2007)	19	Dino.	<i>Caratium lineatum</i>	#/hr	linear -	Negative	0.0000	0.6700	0.0043
Tatters et al. (2013)	45	Dino.	<i>Gonyaulax sp.</i>	1/hr	neutral	Neutral			
Hansen et al. (2007)	31	Dino.	<i>Heterocapsa triquetra</i>	#/hr	threshold -	Negative	0.0000	0.9100	0.0027
Wang et al. (2019)	4	Dino.	<i>Karenia mikimotoi</i>	1/hr	neutral	Neutral			
Tatters et al. (2013)	45	Dino.	<i>Lingulodinium polyedrum</i>	1/hr	neutral	Neutral			
Tatters et al. (2013)	45	Dino.	<i>Prorocentrum micans</i>	1/hr	neutral	Neutral			
Hansen et al. (2007)	21	Dino.	<i>Prorocentrum minimum</i>	#/hr	threshold -	Negative	0.0000	0.8800	0.0019
Brading et al. (2011)	175	Dino.	<i>Symbiodinium sp.</i>	#/hr	linear -	Negative	0.0010	0.0600	0.0066
Van, de, Waal et al. (2013)	12	Dino.	<i>Thracosphaera heimii</i>	mmol/hr	parabolic	Negative	0.0002	0.8500	0.0000
Ries et al. (2009)	17	Echino.	<i>Arbacia punctulata</i>	mmol/g/hr	parabolic	Negative	0.0000	0.8900	0.0003
Courtney et al. (2015)	28	Echino.	<i>Echinometra viridis</i>	%	linear +	Positive	0.0009	0.3500	13.0388
Courtney et al. (2021)	4	Echino.	<i>Echinometra viridis</i>	1/hr	linear +	Positive	0.0244	0.9500	2.3854

Ries et al. (2009)	18	Echino.	<i>Eucidaris tribuloides</i>	mmol/g/hr	threshold +	Positive	0.0000	0.8400	0.0004
Keul et al. (2013)	205	Foram.	<i>Ammonia sp.</i>	mmol/#/hr	linear -	Negative	0.0277	0.0200	0.0000
Prazeres et al. (2015)	32	Foram.	<i>Amphistegina lessonii</i>	%/hr	parabolic	Negative	0.0008	0.3900	0.0010
Kisakurek et al. (2011)	16	Foram.	<i>Globigerinella siphonifera</i>	mmol/hr	neutral	Neutral			
Kisakurek et al. (2011)	14	Foram.	<i>Globigerinoides ruber</i>	mmol/hr	neutral	Neutral			
Reymond et al. (2013)	179	Foram.	<i>Marginopora rossi</i>	%/hr	linear +	Positive	0.0000	0.1900	0.0090
Uthicke-Fabricsius et al. (2012)	47	Foram.	<i>Marginopora vertebralis</i>	mmol/g/hr	threshold +	Positive	0.0000	0.4000	0.0004
Sinutok et al. (2011)	16	Foram.	<i>Marginopora vertebralis</i>	mmol/hr	neutral	Neutral			
Prazeres et al. (2015)	32	Foram.	<i>Marginopora vertebralis</i>	%/hr	linear -	Negative	0.0006	0.3300	0.0005
Manno et al. (2012)	192	Foram.	<i>Neogloboquadrina pachyderma</i>	mmol/#/hr	linear +	Positive	0.0000	0.7100	0.0000
Oron et al. (2020)	96	Foram.	<i>Operculina ammonoides</i>	mmol/g/hr	linear -	Negative	0.0031	0.0900	0.0017
Manriquez et al. (2016)	74	Gastropod	<i>Concholepas concholepas</i>	mmol/g/hr	linear +	Positive	0.0000	0.2400	0.0009
Noisette et al. (2016), Ries et al. (2009)	173	Gastropod	<i>Crepidula fornicata</i>	mmol/g/hr	parabolic	Negative	0.0000	0.2100	0.0028
Ries et al. (2009)	42	Gastropod	<i>Littorina littorea</i>	mmol/g/hr	linear +	Positive	0.0001	0.3400	0.0002
Bibby et al. (2007)	4	Gastropod	<i>Littorina littorea</i>	µm (shell thickness)	neutral	Neutral			
Ries et al. (2009)	21	Gastropod	<i>Strombus alatus</i>	mmol/g/hr	linear +	Positive	0.0000	0.6400	0.0001
Ries et al. (2009)	33	Gastropod	<i>Urosalpinx cinerea</i>	mmol/g/hr	linear +	Positive	0.0000	0.5700	0.0001
Ries et al. (2009)	18	Mollusks	<i>Argopecten irradians</i>	mmol/g/hr	linear +	Positive	0.0097	0.3500	0.0002
Ramajo et al. (2016)	6	Mollusks	<i>Argopecten purpuratus</i>	mmol/g/hr	neutral	Neutral			
Zhang et al. (2011)	5	Mollusks	<i>Azumapecten farreri</i>	mmol/g/hr	linear +	Positive	0.0106	0.9200	0.0001
Ong et al. (2017)	24	Mollusks	<i>Cerastoderma edule</i>	mmol/g/hr	neutral	Neutral			
Sordo et al. (2021)	27	Mollusks	<i>Chamelea gallina</i>	mmol/g/hr	neutral	Neutral			
Gazeau et al. (2007)	20	Mollusks	<i>Crassostrea gigas</i>	mmol/g/hr	linear +	Positive	0.0001	0.6100	0.0000
Ries et al. (2009), Waldbusser et al. (2011)	28	Mollusks	<i>Crassostrea virginica</i>	mmol/g/hr	threshold +	Positive	0.0000	0.5600	0.0003
Garilli et al. (2015)	68	Mollusks	<i>Cyclope neritea</i>	mmol/g/hr	linear -	Negative	0.0020	0.1400	0.0037
Ries et al. (2009)	25	Mollusks	<i>Mercenaria mercenaria</i>	mmol/g/hr	threshold +	Positive	0.0000	0.8300	0.0000
Ries et al. (2009)	14	Mollusks	<i>Mya arenaria</i>	mmol/g/hr	linear +	Positive	0.0001	0.7300	0.0003
Ninokawa et al. (2020)	13	Mollusks	<i>Mytilus californianus</i>	mmol/m ² /hr	neutral	Neutral			
Ries et al. (2009), Gazeau et al. (2007)	86	Mollusks	<i>Mytilus edulis</i>	mmol/g/hr	linear +	Positive	0.0119	0.0700	0.0002
Gazeau et al. (2014)	11	Mollusks	<i>Mytilus galloprovincialis</i>	mmol/g/hr	neutral	Neutral			
Gazeau et al. (2014)	5	Mollusks	<i>Mytilus galloprovincialis</i>	mmol/m ³ /hr	neutral	Neutral			
Garilli et al. (2015)	315	Mollusks	<i>Nassarius corniculus</i>	mmol/g/hr	parabolic	Negative	0.0000	0.2500	0.0064
Cameron et al. (2019)	30	Mollusks	<i>Pecten maximus</i>	mmol/g/hr	neutral	Neutral			
Comeau et al. (2010)b	5	Pteropod	<i>Cavolinia inflexa</i>	mm (shell length)	neutral	Neutral			
Comeau et al. (2009, 2010)	12	Pteropod	<i>Limacina helicina</i>	mmol/g/hr	linear +	Positive	0.0000	0.8500	0.0001
Lischka et al. (2011, 2012)	119	Pteropod	<i>Limacina helicina</i>	mm (shell length)	threshold +	Positive	0.0003	0.1300	0.1303
Bednarsek (2021), Mekkes et al. (2021)	117	Pteropod	<i>Limacina helicina</i>	µm (shell thickness)	parabolic	Negative	0.0000	0.1800	0.0038
Lischka et al. (2012)	28	Pteropod	<i>Limacina retroversa</i>	mm (shell length)	neutral	Neutral			

416 *Barcelos-Ramos et al. (2010), Fiorini et al. (2011), Iglesias-Rodriguez et al. (2008), Richier et al. (2011), Sciandra et al. (2003), Stoll et al. (2012),
417 Gafar et al. (2018), Bach et al. (2011), Sett et al. (2014).

418 Within each of the 11 groups, several categories of calcification response occur within each
419 functional group, with the most varied being the group of dinoflagellates, foraminifera and
420 mollusks, with each of them showing 4 or 5 different categories of calcification responses (Fig. 5).
421 Of the six types of responses of calcification rate vs. TA:DIC, 27% were linear positive (N=27),
422 9% linear negative (N=9), 6% threshold positive (N=6), 2% threshold negative (N=2), 14%
423 parabolic (N=14) and 41% neutral (N=41).

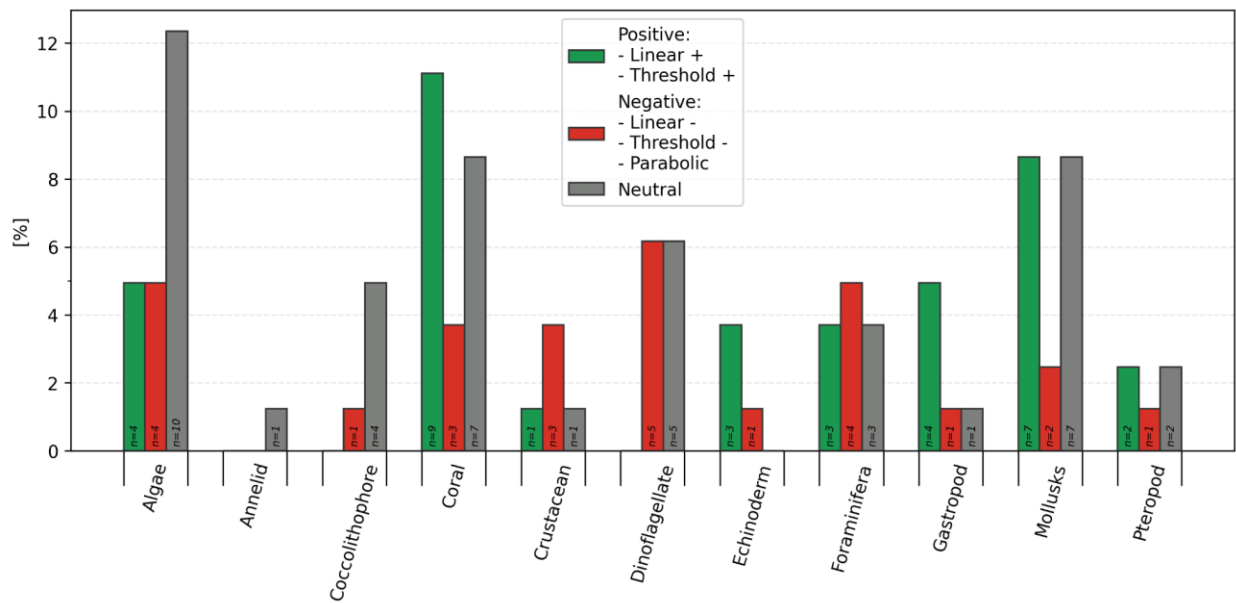
424 Such responses could be further summed up into positive (linear and threshold positive), negative
425 (linear and threshold negative, parabolic) and neutral responses (Fig. 6) when generalized across
426 the calcification rate against the TA:DIC ratio. A summary of responses includes 33.3% positive
427 (N=33), 25.3% negative (N=25), while 41.4% show a neutral response (N=41).



428

429 **Figure 5:** Categories of calcification rate responses and percentage (%) response across eleven
 430 groups (calcifying coralline algae, annelids, coccolithophores, corals, crustaceans,
 431 dinoflagellate, echinoderms, foraminifera, gastropods, mollusks, pteropods). The number on the
 432 bar indicates the number of studies of species included.

433



434

435 **Figure 6:** Summary of percentage (%) responses in calcification rates as positive (linear and
 436 threshold positive), negative (linear and threshold negative, parabolic) and neutral across
 437 eleven groups (calcifying algae, annelids, coccolithophores, corals, crustaceans, dinoflagellate,
 438 echinoderms, foraminifera, gastropods, mollusks, pteropods). The number on the
 439 bar indicates the number of studies with species included.

440

441 3.3 Evaluation of the responses to NaOH/Na₂CO₃ additions

442 Upon added TA, the calcification rate in positive responders will increase, either in a linear or
443 threshold positive response, where calcification plateaus, with the concentration being dependent
444 on the species-specific rate of response (Fig. 2; Supplement Fig. 3). The negative responders
445 (linear or threshold negative and parabolic) will be negatively impacted as follows: first, for the
446 linear negative responders, addition of the Na₂CO₃ will linearly decrease calcification rate, but
447 there is no associated threshold to it; second, for the threshold negative responders, calcification
448 rate will decline in an exponential way until reaching a DIC:TA value, where the response plateaus;
449 and third, for the parabolic responders, the calcification rate will initially increase until reaching
450 certain TA:DIC threshold upon which calcification starts declining. We conclude that all the
451 TA:DIC thresholds for negative responders will be species-specific (Table 2; Supplement Table
452 3).

453 3.4 Threshold values indicative of negative biological response to OAE

454 The TA:DIC biological thresholds in Table 2 are determined by the amount of NaOH addition
455 required to reduce calcification rate by a half (see Supplement Table 3 for Na₂CO₃ thresholds).
456 These thresholds demonstrate the range of carbonate chemistry conditions over which the negative
457 biological effects of OAE deployment might occur and are shown alongside the corresponding pH
458 and Ω_{ar} . Uncertainties are highest for the experimental studies where the experimental temperature
459 was much lower or higher than the 20°C we assume to calculate the thresholds. Experiments done
460 at temperatures below 10°C include mainly pteropods and crustaceans, whereas experiments done
461 at temperatures above 30°C were mainly for algae.

462 For the negative responders, TA:DIC thresholds range from 1.13 to 1.74. The majority of species
463 have reached their threshold below the addition of 500 $\mu\text{mol/kg}$ NaOH, though for 6 species a
464 NaOH addition of more than 500 $\mu\text{mol/kg}$ is required to cross the thresholds in the TA:DIC range
465 of 1.40 to 1.74. The most sensitive species include *Arbacia punctulata* (echinoderm), *Limacina*
466 *helicina* (pteropod), *Halimeda macroloba* (algae) and *Nassarius corniculus* (mollusk). These are
467 all parabolic responders but vary in functional groups. Crustaceans and algae generally require
468 between 50 and 300 $\mu\text{mol/kg}$ to reach their thresholds. Overall, dinoflagellates, corals and
469 foraminifera are the least sensitive groups, with the linear negative responder *Ammonia sp.* of the
470 foraminifera group requiring the highest NaOH addition of 1500 $\mu\text{mol/kg}$ to reduce calcification
471 rate in half. The other least sensitive species include mostly parabolic responders, as well as one
472 threshold negative.

473 **Table 2:** *Studies with negative responders (linear and threshold negative, parabolic) with*
474 *demonstrated TA:DIC thresholds, indicating the amount of NaOH needed to halve the current*
475 *calcification rate (i.e. at the baseline). The value for TA:DIC threshold is used to determine the*
476 *pH and Ω_{ar} (at $T=20^\circ\text{C}$ and the average salinity per species). See Supplement Table 3 for Na₂CO₃*
477 *thresholds.*

Species	Group	Rate unit	NaOH TA:DIC threshold	NaOH addition [$\mu\text{mol/kg}$]	NaOH addition [mg/L]	pH at threshold	Ω_{ar} at threshold
<i>Arbacia punctulata</i>	Echino.	mmol/g/hr	1.128	10	0.39	8.11	3.73
<i>Halimeda macroloba</i>	Algae	mmol/hr	1.132	10	0.39	8.11	3.84
<i>Limacina helicina</i>	Ptero	μm (shell thickness)	1.13	10	0.39	8.11	3.78
<i>Nassarius corniculus</i>	Mollusks	mmol/g/hr	1.149	10	0.39	8.11	4.20
<i>Crepidula fornicata</i>	Gastropod	mmol/g/hr	1.152	50	1.95	8.17	4.40
<i>Penaeus plebejus</i>	Crust.	mmol/g/hr	1.145	50	1.95	8.17	4.21
<i>Neogoniolithon sp.</i>	Algae	mmol/g/hr	1.164	100	3.90	8.23	4.76
<i>Amphistegina lessonii</i>	Foram.	%/hr	1.19	150	5.85	8.28	5.48
<i>Caratium lineatum</i>	Dino.	#/hr	1.18	150	5.85	8.30	5.21
<i>Homarus americanus</i>	Crust.	mmol/g/hr	1.186	150	5.85	8.29	5.37
<i>Oculina arbuscula</i>	Coral	mmol/g/hr	1.185	150	5.85	8.29	5.35
<i>Sporolithon durum</i>	Algae	mmol/m ² /hr	1.218	200	7.80	8.32	6.31
<i>Cyclope neritea</i>	Mollusks	mmol/g/hr	1.224	200	7.80	8.32	6.22
<i>Callinectes sapidus</i>	Crust.	mmol/g/hr	1.227	250	9.76	8.39	6.57
<i>Thoracosphaera heimii</i>	Dino.	mmol/hr	1.232	250	9.76	8.38	6.70
<i>Halimeda macroloba</i>	Algae	mmol/g/hr	1.259	300	11.71	8.41	7.40
<i>Operculina ammonoides</i>	Foram.	mmol/g/hr	1.321	450	17.56	8.53	9.20
<i>Symbiodinium sp.</i>	Dino.	#/hr	1.316	450	17.56	8.54	9.15
<i>Heterocapsa triquetra</i>	Dino.	#/hr	1.325	500	19.51	8.62	9.56
<i>Marginopora vertebralis</i>	Foram.	%/hr	1.332	500	19.51	8.59	9.71
<i>Acropora pulchra</i>	Coral	mmol/m ² /hr	1.379	600	23.41	8.64	11.00
<i>Pavona cactus</i>	Coral	mmol/m ² /hr	1.379	600	23.41	8.64	11.00
<i>Prorocentrum minimum</i>	Dino.	#/hr	1.388	650	25.37	8.73	11.48
<i>Emiliana huxleyi</i>	Cocco.	mmol/#!/hr	1.457	800	31.22	8.78	13.48
<i>Ammonia sp.</i>	Foram.	mmol/#!/hr	1.738	1500	58.54	9.22	22.50

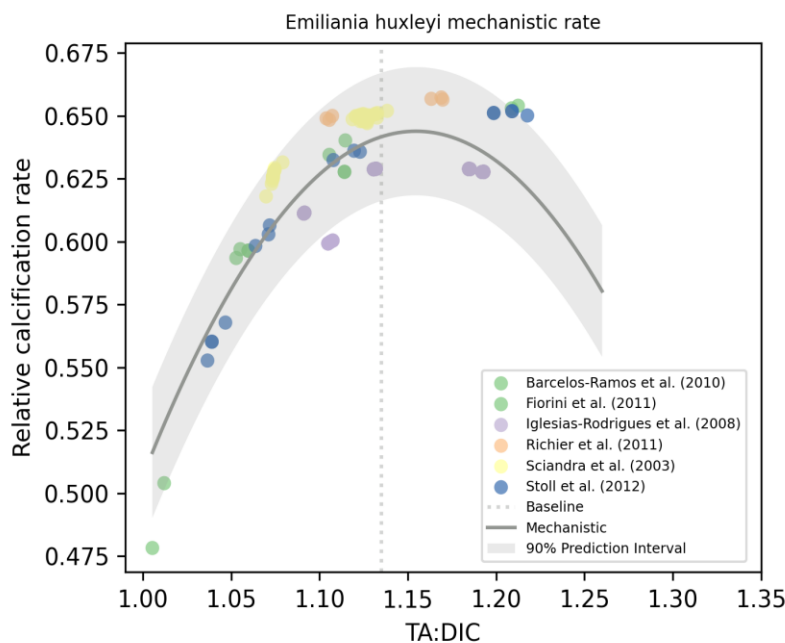
478 3.5 Comparison of current vs. pre-industrial calcification rates

479 To understand the extent can OAE offset the negative effects induced by OA, we have focused on
480 positive (linear and exponential) responders from this study, which were negatively impacted by
481 OA. Comparing their average pre-industrial calcification rate to the rates induced by OAE, we
482 note that the change depends on the category of response (Supplement Fig. 3): in the species with
483 the threshold positive rate, calcification is similar for the pre-industrial and current conditions.
484 This is likely because this type of response retains maximum calcification rate across greater
485 TA:DIC range, and as such these species have not been compromised in their calcification by the
486 changes that occurred from the pre-industrial onwards. On the other hand, linear positive calcifiers
487 seem to be more severely impacted and current calcification rates are substantially lower compared
488 to the pre-industrial calcification rates. It follows that for the positive linear responders, an increase
489 using NaOH would compensate for the calcification rate loss since the pre-industrial times. For
490 most of the investigated linear responders, there would have to be NaOH addition of 50 to 100
491 $\mu\text{mol/kg}$ to fully compensate for the difference between the pre-industrial and current conditions.
492 However, two species *Amphibalanus improvisus* and *Azumapekten farreri* require up to 200
493 $\mu\text{mol/kg}$ NaOH, while some species, i.e. coral *Siderastrea radians* might return to the levels of
494 the pre-industrial calcification by a much smaller NaOH amount, less than 50 $\mu\text{mol/kg}$ NaOH.

495 3.6 Comparing calcification mechanisms with the empirical studies

496 This study establishes the predictions of responses that ultimately need to be validated in the field
497 or experimental studies, this being the most pertinent for highly sensitive species. A good
498 alternative to validating the predicted responses is to use species-specific mechanistic responses,
499 which should be inherently more accurate than just empirical responses. Here, we compared the
500 obtained results of this study with the predicted mechanistic relationship to determine to what
501 extent can mechanistic relationships contribute to improved, i.e. more accurate and less
502 uncertainty, OAE predictions.

503 For *Emiliana huxleyi*, we used the experimental TA and DIC data to calculate the $[\text{HCO}_3^-]$, $[\text{H}^+]$
504 and $[\text{CO}_2]$ concentrations. Using the mechanistic rate equation from Bach et al. (2015) and their
505 computed sensitivity parameters, we calculated and plotted the rate derived via mechanistic
506 approach. We applied linear, polynomial (second-order) and exponential regressions and chose the
507 best fit based on the lowest p-value, using the same method as for our experimental calcification
508 rate data regressions. Like the mechanistic rate regression, our experimental calcification rate also
509 shows a significant parabolic relationship for *Emiliana huxleyi* (see Fig. 9). However, when using
510 the same approach for another coccolithophore species *Calcidiscus leptoporus* (Bach et al., 2015),
511 our best fit did not align with the proposed mechanistic response; instead, a non-significant
512 relationship was obtained using experimental data (Supplement Fig. 5). Such comparisons reveal
513 species-specific relationships that are likely dependent on a lot of parameters, with one equation
514 alone not being operable among different species from different regional settings.



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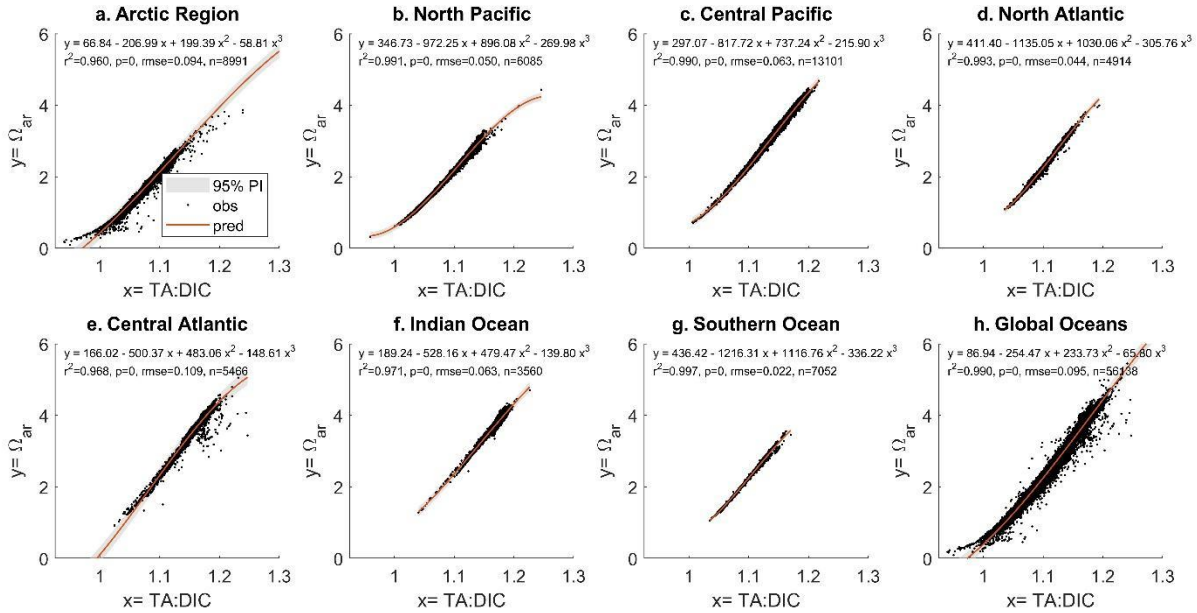
517 **Figure 9:** Mechanistic rate equation and parameters ($a = 9.56e-1$, $b = 7.04e-4$ mol/kg, $c = 2.1e6$
518 kg/mol, $d = 8.27e6$ kg/mol) taken from Bach et al. (2015) and fitted using experimental data for
519 *E. huxleyi* (used data from the studies indicated in legend).

520 For most species, we must still rely on empirical, single-parameter relationships, including
521 saturation state, bicarbonate ion concentration and the substrate-to-inhibitor ratio (SIR) (i.e. the
522 bicarbonate ion to hydrogen ion concentration ratio). When comparing empirical data from the
523 experiments involving the mollusk, coral and coccolithophore groups against the SIR ratio, we
524 found large discrepancies between this and SIR-proposed mechanisms (Supplement Fig. 6). For
525 most of the coccolithophore group, the experimental rate regressions cannot be explained using
526 SIR mechanisms. Only in the case of *Calcidiscus leptoporus*, the experimental and mechanistic
527 responses remain the same. Reasons for these discrepancies could potentially be that SIR might
528 insufficiently include the multitude of biological processes involved in the calcification (e.g. how
529 carbon is provisioned or the ability to regulate calcifying fluid pH), as well as other environmental
530 parameter variations. For mollusks, a quarter of the mechanistic rate regressions based on the SIR
531 agreed with the experimental calcification rate regressions. The other 75% did not, especially for
532 the studies with experimental conditions of $\Omega_{ar} > 1$. For corals, the majority of coral species ($n =$
533 14) were classified as having a linear positive mechanistic relationship when using SIR
534 relationships. When comparing this to our experimental rate regressions, we only found
535 agreements with the mechanistic regressions in 6 out of 18 species.

536 **3.7 Global and regional carbonate chemistry data coverage based on GLODAP datasets**

537 The compilation of chemical observational data (pH, Ω_{ar} , TA, DIC) was done for the GLODAP
538 data across the regional ocean and global scales to determine the range of Ω_{ar} , TA and DIC (as
539 represented by the TA:DIC ratio) and TA:DIC vs Ω_{ar} correlation down to the depths averaged over
540 200 m. This allows us to apply the thresholds even for the regions for which we do not have
541 sufficient or reliable data or experimental coverage, making the inferences about the OAE impact
542 even in those regions.

543 Here, we focus on showing the results ranging over the 0–50m because this covers most of the
544 biological habitat for examined species and it is where the OAE enhancement would induce the
545 greatest changes. Over the 0–50 m depth, Ω_{ar} ranges from 0.2 to 5 and TA:DIC ranges from 0.1 to
546 1.25 and both parameters are correlated across all the regions, as demonstrated by the fitted second-
547 order polynomial regressions, with R^2 of 0.96 or higher, and all the correlations being significant
548 (Fig. 7), with regional specific relationships not impacting the fit. All the correlation parameters
549 are presented in Supplement Table 4. Similar fits were found at different depths. The conditions
550 in the higher latitude regions are located at the lower range of Ω_{ar} vs TA:DIC, while the conditions
551 in the low latitudes and temperate regions are at the upper range, with the highest values present
552 in the central Atlantic and Pacific region. Such strong correlation as observed for Ω_{ar} vs TA:DIC
553 does not exist with pH, regardless of the depth interval examined. While the correlations are still
554 significant, they are broadly distributed and represented over a shorter TA:DIC range, with
555 significantly lower goodness of fit (Supplement Fig. 4), with the correlations being highly
556 regionally dependent due to pH and temperature co-linearity. Because of this, all further biological
557 analyses are only done using the Ω_{ar} vs TA:DIC ratio.



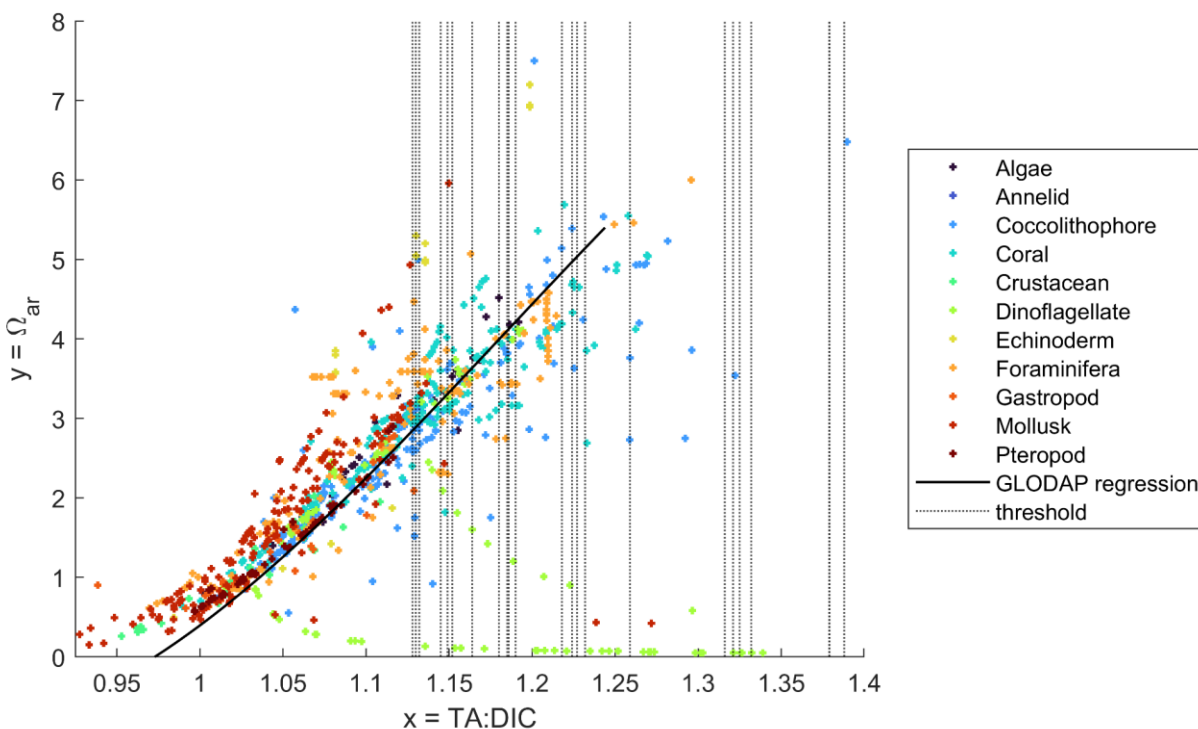
558

559 **Figure 7:** The range of observed Ω_{ar} and TA and DIC values (as represented by the TA:DIC ratio)
 560 values and the relationship with the best fitted curve between Ω_{ar} vs TA:DIC across regional (a-g)
 561 and global (h) scales based on the observational GLODAP dataset averaged over 0-50 m depth.

562 **3.8 TA:DIC vs Ω_{ar} for experimental data and GLODAP**

563 We compared the ranges and TA:DIC and Ω_{ar} of biological experimental data with field
 564 biogeochemical data (GLODAP) to examine if similar range of conditions and TA:DIC
 565 correlations are applicable over a broader, global dataset. For this, we plotted Ω_{ar} vs TA:DIC along
 566 with the GLODAP regression line for Ω_{ar} vs TA:DIC (Fig. 8). For each TA and DIC datapoint, the
 567 salinity and temperature specific to that data point were used to compute Ω_{ar} . We show the
 568 similarity in the conditions, which gives the validity of our experimentally derived thresholds to
 569 be extrapolated within the global GLODAP dataset.

570 Figure 8 also shows that various biological groups are clustered around specific TA:DIC ratios,
 571 for example, mollusks, coral and coccolithophores are represented on the lower, mid, and higher
 572 TA:DIC spectra, respectively, while dinoflagellates are randomly scattered off the TA:DIC line.
 573 This indicates that there is a general lack of data distribution in the upper ranges of TA:DIC ratio,
 574 especially for the groups that are lying at the lower and mid end of the TA:DIC ratio spectra.
 575 Plotting biological data from the OA datasets against the regional and global TA:DIC gradient
 576 derived from GLODAP (Fig. 7), we also observed that experimental data ranges were not always
 577 consistent with natural conditions, for example, having a lower Ω_{ar} at a higher TA:DIC ratio.



578

579 **Figure 8:** Experimental TA:DIC from experimental biological studies for eleven investigated
 580 functional groups (see legend) plotted against Ω_{ar} , with the latter being computed using
 581 experimental TA and DIC. The black line represents the regression line of TA:DIC and Ω_{ar} data
 582 from the GLODAP dataset (covering 0-50m depth). See Supplement Fig. 5 for GLODAP Ω_{ar} vs
 583 TA:DIC, from which the black regression line shown here is derived.

584

585 4. Discussion

586 OAE is a quickly developing strategy that is already in the field-testing phase despite extremely
 587 limited understanding of biological implications and environmental concerns. Hence, gaining
 588 insights of potential risks for the biological species and communities is essential and timely. In
 589 retrospect, it took decades for the OA research community to get a more accurate and
 590 comprehensive understanding leading to predictions of biological responses to OA (Gattuso and
 591 Riebesell, 2015). Without a very clear conceptual strategy for the OAE testing, research
 592 community might also need years to decades before we comprehensively understand the OAE-
 593 related implications. Consequently, there is an essential need to develop an assessment framework
 594 of predictive responses and testing appropriate to OAE, which will assist in OAE scaling while
 595 avoiding the risks. This paper aims at developing such an assessment, where responses are
 596 categorized per species responses against TA:DIC. We propose to use the TA:DIC ratio in the
 597 biological studies reporting OAE results, as we believe it simplifies the system and makes it easier
 598 to use and translate the carbonate chemistry in the experimental setting. Such a TA:DIC ratio
 599 allows to ultimately standardize the biogeochemical and biological data and is useful for easier
 600 comparisons among the conducted experiments.

601 **4.1. Identified strengths and limitations of the synthesis approach based on OA studies**

602 Prior to conducting this study, several drawbacks were identified that could potentially limit such
603 a synthesis work: first, insufficient amount of data at the upper range of carbonate chemistry
604 conditions (high pH, high Ω_{ar}); second, experimental data under conditions with no relevance to
605 natural settings (Fig. 8); and third, an insufficient number of validation studies under high TA
606 conditions to validate the results of this synthesis. To overcome the first two limitations, the
607 decision was made to combine multiple OA datasets for a single species with the aim to achieve a
608 greater range in carbonate chemistry conditions, including higher pH, Ω_{ar} experimental values,
609 which should reduce the uncertainty of the predictions. However, combining raw data on species
610 calcification rate proved to be more challenging because even across the same species the reporting
611 of the calcification rates was highly variable. The use of different measuring approaches of
612 calcification rates while conducting OA studies generated data with divergent units that do not
613 allow for the intercomparison of data and results. As different studies for a single species could
614 not be combined, we chose to increase the number of studies and thus, the number of examined
615 species. Based on the response categories from the OA studies (Ries et al., 2009), our hypothesis
616 was that OAE will elucidate the same categories of responses, i.e. positive, negative and neutral.
617 Within each of the groups examined, multiple categories of predicted calcification response were
618 found. In this way, we demonstrated that it was possible to develop a useful framework for
619 assessing and predicting species-specific OAE responses that can delineate different responders,
620 identify species with greater OAE sensitivity and determine the thresholds where such negative
621 responses could happen.

622 **4.2 Synthesizing biological response under OAE additions identifies positive and negative** 623 **responders**

624 The greatest variability in calcification rate response upon NaOH addition was evident in
625 foraminifera, mollusks, corals, dinoflagellates and pteropods, where four to five different
626 categories of responses were found. Such variability confirms that the responses to OAE will be
627 species-specific and is related to various calcification mechanisms across the observed groups.
628 Despite such specificity, the responses were summarized across three emerging groups of
629 responses: positive, negative, and neutral (Fig. 6), which we discuss in the context of possible
630 mechanisms of calcification or available OAE experimental studies used for validation.

631 Positive responders (33%) show an increased calcification rate to OAE additions, observed within
632 all functional groups besides annelids, coccolithophores and dinoflagellates. Corals mostly have
633 positive and neutral responses, suggesting that coral species would not be negatively impacted
634 during OAE field trials. This mostly positive response is validated by increased coral calcification,
635 shown for two coral species of *Acropora* and *Siderastre* in experiments conducted by Palmer et
636 al. (2022).

637 The metrics to evaluate the sensitivity of calcification rate for the negative responders (negative
638 linear and threshold) to OAE addition was based on the amount of alkalinity addition required to
639 halve the current calcification rate (Fig. 3; Tables 1, 2). The most negative responses are expected

640 in dinoflagellates (6% of all species), crustaceans and foraminifera (both 5% of all species). As
641 such, these groups are one of the priorities for the future OAE experimental work to determine at
642 which TA: DIC negative response happens. The worst out of these are dinoflagellates that
643 demonstrate negative response in 5 cases, 5 neutral responses and 0 positive (see Table 1;
644 Supplement Fig. 3). The reason for negative response to OAE in this group is related to the fact
645 that their growth gets limited at higher pH, with further carbon limitation playing a role at very
646 high pH levels and low DIC concentration (Hansen et al., 2002; 2007). On the other hand,
647 crustaceans only demonstrated positive response in one study (Pansch et al., 2014), while
648 remaining results predict either negative or neutral response. While crustaceans are effective in
649 retaining homeostasis at lower pH, they might be less so at higher pH, which was shown in the
650 OA experiments by Ries et al. (2009) for three crustacean species (*Callinectes sapidus*, *Homarus*
651 *americanus*, *Penaeus plebejus*), confirmed in the OAE study by Cripps et al. (2013) in *Carcinus*
652 *meanas*. While studies are still lacking, physiological acid-base regulation at higher pH is
653 associated with higher costs (Cripps et al., 2013). Crustaceans show a disrupted acid–base balance,
654 evident through the increase in hemolymph pH, K, Na ions and osmolality, coupled with a decrease
655 in extracellular pCO₂ and HCO₃⁻, which is indicative of respiratory alkalosis (Truchot, 1984;1986).
656 This is often associated with hyperventilation, the aim of which is to flush out the hemolymph CO₂
657 and increase the affinity of oxygen uptake. However, while this temporarily creates a
658 physiologically favorable condition, it also represents the physiological costs and potential
659 metabolic composition of other processes, likely also for the calcification.

660 For the neutral responders or groups with no significant correlation between TA:DIC and
661 calcification rates, it is somewhat uncertain to predict if such responses will be retained under
662 OAE. While parabolic responders show a physiologically understandable parabolic type of dose-
663 response, positioning the TA:DIC values where the threshold occurs is also highly species-specific
664 and potentially uncertain, meaning that it might depend on other environmental factors.

665 With respect to the coccolithophores, we note that this was the only group where data compilation
666 on calcification rate across the groups was possible because the OA studies were conducted in a
667 more uniform way, using similar approaches, and reporting the result in the same units. When data
668 for *E. huxleyi* across the comparable studies was compiled (Barcelos-Ramos et al., 2010; Fiorini
669 et al., 2011; Iglesias-Rodrigues et al., 2008; Sciandra et al., 2003; Stoll et al., 2012; Richier et al.,
670 2011), a significant parabolic response was obtained (Table 1), although the goodness of fit was
671 fairly low ($R^2=0.16$). Despite lower R^2 , we decided to use the compiled dataset because of the
672 increased statistical power. The parabolic response obtained aligns with Langer et al. (2006) and
673 also with the parabolic type responses found in the synthesis study by Paul and Bach (2020) and
674 Bach et al. (2015). The threshold indicates the mechanisms of coccolithophore growth that are
675 driven by CO₂, which is shown to decline with the OAE addition. The threshold based on all
676 studies for *E. huxleyi* combined was positioned at TA:DIC of 1.457 ($\Omega_{ar} = 13.48$, see Table 2),
677 which would be triggered at 800 $\mu\text{mol/kg}$ of added NaOH and at 60 μatm . Comparatively with the
678 phytoplanktonic diatoms, such growth limitation is predicted at a pCO₂ amount at 100 μatm
679 (Riebesell et al., 1993). It is important to note that when these studies were analyzed individually,
680 a mixture of different responses was observed. We emphasize the variability within the

681 coccolithophore responses, which are species-specific and inherently related to the strain
682 adaptation to their innate regional settings and dependent on a variety of other factors (Bach et al.,
683 2015; Gafar and Schultz, 2018), including the longevity of the species, the experimental settings
684 used in the study, for example nutrient-replete vs nutrient deficient conditions, and the presence or
685 absence of (un)suitable light conditions. Interestingly, for all the coccolithophore species other
686 than *E. huxleyi*, responses were neutral. For validation purposes, the results of our study could not
687 be compared to Gately et al. (2023) because calcification rates were not studied, and it is urgent
688 that more validation studies for coccolithophores are conducted.

689 **4.3 TA:DIC thresholds related to biological sensitivity and their implementation**

690 Lastly, and most importantly, a set of species-specific thresholds was developed in this study, with
691 demonstrated application across the global Ω_{ar} vs TA:DIC conditions (Table 2; Fig. 8). The range
692 of OAE additions to exceed the thresholds vary significantly between the species and the type of
693 response, with the parabolic responders generally having the lowest threshold compared to the
694 linear or exponential negative responders. The TA:DIC thresholds upon TA application ranged
695 between 10–500 $\mu\text{mol/kg}$ of NaOH for all but 5 of negatively responding species, pointing to the
696 most and least species-specific OAE-related sensitivities, respectively. The lowest TA:DIC
697 threshold is predicted for echinoderm *Arbacia punctulata*, with the lowest pH value of 8.1, which
698 is at least an order of magnitude lower compared to what the lab experimental OAE trials use in
699 their treatment levels.

700 In this study, we have calculated the thresholds for all negatively impacted species irrespective of
701 the quantity that needs to be added, but we note that high threshold value exceedance is likely not
702 realistic. Although biogeochemical model outputs show high OAE-related concentration at the
703 injection site for a short-time, realistic field dosing upon dilution might be low. Wang et al. (2023)
704 reported that the nearfield maxima in the respective investigation area of the Bering Sea is to
705 increase TA by about 10 $\mu\text{mol/kg}$ in the nearfield and by about 1 $\mu\text{mol/kg}$ of NaOH in the farfield
706 region.

707 Similarly, a lot of laboratory-based OAE experiments are conducted with the aim to gain a wide-
708 ranging empirical response, applying very high treatment levels that are not realistic with respect
709 to potential level of OAE dosing in the field. Here, we explicitly emphasize the importance of
710 including much lower additions of TA as the experimental treatment levels to better support
711 biological understanding in the field. In addition, prior to the lab experiments it would be important
712 to identify if the selected species for the experiments might be negative responders with a threshold
713 at lower TA:DIC range. This is especially pertinent for the groups for which OA experimental
714 data is limited and only distributed at the lowest and the mid TA:DIC ratio, such as mollusks and
715 dinoflagellates (Fig. 8; Supplement Fig. 3).

716 We note that the calculations of the thresholds in this study were taken for the global surface ocean
717 conditions assuming current $p\text{CO}_2$ conditions of 425 and pH of 8.1 as a control from where the
718 OAE compound was added. However, this does not take into consideration the conditions of the
719 local ecosystems in which OAE would be implemented and as such, the thresholds might be

720 different depending on the local surface pH conditions where organisms are residing. As such, to
721 get the most accurate and regionally applicable threshold value for the species of interest, the
722 recommendation is that the baseline for the OAE addition should be determined based on the local
723 settings.

724 **4.4 Bringing realism of OAE experiments to the field trials**

725 OAE-related biological responses and risks are not going to depend solely on the concentration of
726 OAE compound used but also on the baseline carbonate chemistry conditions at the site of
727 deployment, such as baseline TA:DIC ($\text{pH}/\Omega_{\text{ar}}$) and variability of carbonate chemistry parameters
728 across horizontal and vertical depths. Physical parameters of importance are related to the dilution
729 effect, mixing, retention capacity, as well as the rate of the equilibration effects of the air-sea CO_2
730 uptake (Ferderer et al., 2022; He and Tyka, 2023; Schulz et al., 2023; Wang et al., 2023).
731 Variability on the seasonal and annual scales of the air-sea CO_2 uptake can have impacts not only
732 on the chemical processes related to the variable OAE efficiency, but also for the biological
733 implications related to the crossing of biologically sensitive thresholds. It is the combination of all
734 these factors that creates baseline conditions to which biota would ultimately be exposed in their
735 natural environment upon OAE deployment (Wang et al., 2022).

736 If similar conditions as induced by the OAE field trial are present as part of the natural variability
737 within the species' habitat, it is more likely that the species might be adjusted to it. On the contrary,
738 rapidly induced novel conditions might be the most detrimental. As such, it is worth considering
739 if OAE deployments could be, when possible, carried out not as a single high dosage deployment,
740 but rather as a more continuous, lower dosage application that would eliminate the swings and
741 maxima in conditions, while also allowing more time for species acclimation or migration during
742 the initial injection of the OAE deployment.

743 What is needed urgently for the community performing biological field trials is a best practice
744 guide for evaluating the biological responses as part of the field environmental risk monitoring
745 approach. Along with the guide for conducting the lab (Iglesias-Rodrigues et al., 2023) and field
746 OAE studies (Cyronak et al., 2023), the recommendations in the risk assessment guide would
747 address the topics of initial baseline conditions, identify suitable risk analyses, determine
748 thresholds, and propose the development of regionally specific indicators for monitoring, while
749 also identifying the guidance for the regulators. Ideally, such biological and environmental risk
750 monitoring and assessment would be accompanied by the application of the physical mixing
751 models with site-specific biogeochemical processes (Ho et al., 2023; Fennel et al., 2023) that can
752 predict the maximum expected TA increase in the nearfield and farfield regions of the study site,
753 representing a more realistic exposure and better informing further experimental work.

754 **4.5 Comparison of calcification rate with the pre-industrial conditions**

755 Positive calcifying responders identified in this study were the most impacted through reduced
756 calcification due to anthropogenically driven OA since the pre-industrial changes. For these
757 species, retentive addition of NaOH in the range of 10 to 200 $\mu\text{mol}/\text{kg}$ NaOH would allow these

758 species to fully bounce back to the pre-industrial calcification. Achieving such long-lasting
759 increases of NaOH is currently not feasible in the field but shows additional mitigating benefits of
760 OAE against OA. With continuous OAE implementation, the calcification of positive responders
761 would be reversed back to their pre-industrial capacity, indicating that such co-benefiting effects
762 could be considered within the context of OA mitigating effects and could be part of the ecosystem
763 restoration and protection strategy. In addition, it is probably less likely that such reversal of the
764 processes might lead to concern for the species as ecological winners and potential shift on the
765 community level. However, current modeling efforts show that temporal and spatial extent of OAE
766 as a mitigation capacity for OA is variable and might induce significant biogeochemical changes
767 that further exacerbate ecological risks (González and Ilyina, 2018; González et al., 2016; Mongin
768 et al., 2021). Nevertheless, with 40% positive responders, there is substantial opportunity for
769 species-specific variations and an indication that some species that could indeed benefit beyond
770 just improved calcification.

771 **4.6 Unknowns about ecological and biogeochemical implications call for the precautionary** 772 **approach**

773 The value of calcification as the proxy is indicative of organismal fitness which directly relates to
774 OAE effects as harmful or beneficial for the species. From an ecological perspective, 31% of
775 negative responders demonstrate a potential for negative implications and ecological shifts. In
776 addition, we note that this study did not include diatoms in the analyses, which are predicted to be
777 negatively impacted by the carbonate-based OAE (Ferderer et al., 2022), leader to possible
778 community shifts (Bach et al., 2019). The possibility of the ecological shifts should not be
779 neglected given the variety of the positive responders, understudied effects of OAE in non-
780 calcifiers and their relationship with the calcifiers through the grazing impact, and lastly, unknown
781 and highly unpredictable indirect effects. In addition, the inferences on the neutral responders
782 should also remain cautious.

783 From a biogeochemical perspective, it is reasonable to infer that OAE will introduce changes in
784 calcification rate across species, potentially resulting in changing carbon export or carbonate
785 counter pump. Species-specific responses in major carbonate producers, i.e. coccolithophores,
786 foraminifera and pteropods show both, negative and positive response, which could have strong
787 effects on biogeochemical fluxes (Riebesell et al., 2017; Bach et al., 2019). Increased calcification
788 could result in thicker and denser shells, contributing to faster sinking and increased carbonate
789 fluxes, while the negative calcification has an opposite effect. This could ultimately, over
790 centennial time scales (Oschlies et al., 2023), induce changes on the subsurface total alkalinity at
791 intermediate and deeper depths in the water column, and dissolution at or near the seafloor (Gehlen
792 et al., 2011) with potential feedback that results in increased CO₂ flux to the atmosphere (Gattuso
793 et al., 2021). Full scale of ecological and biogeochemical shifts remains a high priority topic for
794 future investigations and until these huge uncertainties are resolved, we should exercise a
795 precautionary principle in considering the next steps of OAE field implementations.

796 **4.7 Potential confounding effects and the validation issues**

797 This study only considered the changes in carbonate chemistry due to the addition of NaOH and
798 Na₂CO₃. However, other OAE feedstocks contain compounds that could induce biological toxicity
799 due to the presence of the trace metals (Ni, Cu, Ca, Si; Bach et al., 2019), as well as potential
800 negative environmental impacts due to the secondary precipitation (Hartmann et al., 2022; Moras
801 et al., 2022). This study also did not focus on the sensitivity across different life stages, even
802 though stage-stage sensitivities to OAE are expected based on previous OA results. Furthermore,
803 we did include data from the experimental lab or field studies that involve multiple stressors in
804 their experimental designs. As such, an additional impact of warming, dissolved oxygen, light
805 intensity on the OAE-induced responses was not determined, although they could elicit different
806 biological pathways than OAE alone or have additional confounding effects.

807 The synthesis of the experimental studies always includes implicit biases that are based on the
808 published experimental studies, the range and species used, regional coverage and heterogeneity.
809 Important consideration is the adaptation of the species used in the experimental studies because
810 their calcification optimums optimum might be pre-determined based on their local habitat
811 conditions. Given that the baseline for the OAE-compound addition was chosen at the global
812 current surface pH value, some of the thresholds might actually be lower than expected.

813 An additional parameter of importance, which has not been addressed in this study because it was
814 not possible to do it on such a diverse group of species with varied generations, is the duration of
815 the experimental exposure. Duration can impact physiology with respect to the exposure until the
816 OAE-related effects become evident.

817 The predictive results of these studies need to be validated with subsequent studies, but suitable
818 approach to evaluate such predictions could be done by comparing mechanistic studies with the
819 experimental data. One of the problems is the lack of a known mechanistic relationship of the
820 calcifying species. However, a lot of mechanistic relationships are based on one parameter only.
821 Ninokawa et al. (2024) and Li et al. (2023) emphasized that using only one parameter to describe
822 calcification process is insufficient and strongly recommend using at least two parameters for more
823 accurate calcification predictions. Our findings agree with Ninokawa et al. (2024), for examples,
824 we observe that using SIR relationships to successfully describe calcification was limited to only
825 a few species and there are no generalizable patterns that could be applicable across multiple
826 groups. This clearly delineates a major gap in the mechanistic understanding of calcification so
827 far, the lack of which significantly limits our ability of ecological and biogeochemical predictions
828 to OAE. As such, more research is urgently needed on broader mechanistic understanding of
829 calcification across different species, and additionally, one parameter calcification processes
830 should be replaced with more accurate and comprehensive using two or three parameters.

831 **4.8 Applications within the existing governmental regulations and the guiding principle**

832 Our results, especially related to the use of biological thresholds or NaOH dosing, have
833 applications outside the academic realm, most notably with policy-management governmental
834 regulations. Here, we provide one example how current regulations could be compared against the

835 results obtained from this study, focusing on the US Environmental Protection Agency’s rule for
836 waste water not exceeding a pH of 9 when entering the coastal ocean (see NPDES manual, 2010).
837 We have added NaOH study until reaching the threshold 9 and analyzed which species could be
838 compromised because of added OAE. For most of the studies, NaOH concentration of 1100 to
839 1250 $\mu\text{mol/kg}$ needs to be added before exceeding a pH of 9. This represents a very high
840 concentration and the thresholds for most of the negative responders with identified thresholds
841 (Table 2), will be exceeded far below the regulatory standards of pH 9 (Table 2), especially if the
842 exposure occurred over a duration period that matters for calcification and for the organism’s
843 physiological status. This case demonstrates discrepancy of the current chemical pH regulation
844 and associated biological effects, where the safety biological limits are violated and biological
845 harm is not prevented, thus likely induced. Despite the fact that achieving such high pH through
846 continuous NaOH implementation is unlikely to occur in the field, such regulations are of
847 particular concern and regulations to assure safety space for marine biota need to be urgently
848 addressed and determined.

849 **5. Conclusions and next steps**

850 Sufficient certainty in predicting biological responses reduces the risks and supports safe operating
851 space for OAE implementation and scaling up. Overall, given that almost 70% of examined species
852 showed non-neutral response (either positive or negative), this calls for care in OAE field
853 implementation until the temporal and spatial scales of safe operations are determined and OA
854 mitigation established. The goal of this study is to serve as a baseline for prioritizing experimental
855 and field OAE research and assess environmental risks. Such prioritization identifies those species
856 for which experimental work needs to be conducted first. This would involve species with the
857 greatest OAE-related sensitivity (negative responders), species with the greatest uncertainty in
858 response, as well as the species with very strong predicted positive response that could potentially
859 introduce a shift on the community level. In addition, it would also recognize the species for which
860 the existing knowledge is sufficient and there is less immediate need for the OAE experiments.
861 We hope that all presented tools provide guidance for the practicing and regulatory community
862 considering OAE field application within the safe limits.

863 It is important to emphasize that this study represents the first synthesis of OAE effects. It would
864 benefit from continuous update and data additions, while cross-validated with further experimental
865 work. Furthermore, we underline that our predictions were developed as calcification responses of
866 the pelagic organisms only. Similar OA datasets are available for the benthic calcifiers and non-
867 calcifiers, and also for other biological responses, including growth, survival, with much greater
868 uniformity of data and units, which mean they could be easily transformed into an OAE predictive
869 risk assessment as conducted in this study.

870

871 **Data availability**

872 No additional data were generated as part of this study, they were all collected from the already
873 published studies. The compiled data is currently available on request. The Python code used for

874 computing baselines per species, conceptually adding alkalinity in the form of NaOH and
875 Na₂CO₃, predicting calcification rate response, visualizing data and computing thresholds is
876 available in the GitHub repository at https://github.com/hannavdmortel/OAE_calc_responses (last
877 access: 24 July 2024) and is archived on Zenodo at <https://doi.org/10.5281/zenodo.12806137> (van
878 de Mortel, 2024). PyCO₂SYS v1.8.0 (Humphreys et al., 2022) was used to solve for the carbonate
879 system, with software available at <https://doi.org/10.5281/zenodo.3744275> (Humphreys et al.,
880 2023).

881 The code is stored at the following link: <https://doi.org/10.5281/zenodo.12806137>

882 **Author contributions**

883 NB designed and conceptualized the research and wrote the first draft of the paper. GP provided
884 the analyses using GLODAP data, and also provided visualizations and formal analyses. HvdM
885 collected and curated data, conducted formal analyses and provided visualization. MGR has
886 provided formal statistical analyses and visuals. RAF and AD have provided insights, suggestions,
887 and generated discussion about specific parts of the paper. All have contributed to the writing of
888 this draft.

889

890 **Competing interests**

891 The contact author has declared that none of the authors has any competing interests.

892

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