

1 **Locally Produced Sedimentary-Leaf Wax Biomarkers in the High-Altitude Catchments**
2 **Areguni Mountains Outweigh Upstream-Downstream River-TTtransport in Sedimentary**
3 **Archives**

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19 **Abstract:** ~~Sedimentary records of lipid biomarkers such as leaf wax *n*-alkanes are not only~~
20 ~~influenced by ecosystem turnover and physiological changes in plants, they are also influenced by~~
21 ~~earth surface processes integrating these signals into the sedimentary record, though the effect of~~
22 ~~these integration processes are not fully understood. Sedimentary records of lipid biomarkers such~~
23 ~~as leaf wax *n*-alkanes are not only influenced by ecosystem turnover and physiological changes in~~
24 ~~plants, they are also influenced by earth surface processes integrating these signals. The integration~~
25 ~~of biomarkers into the sedimentary record and the effects of integration processes on recorded~~
26 ~~environmental signals are complex and not fully understood.~~ To determine the depositional
27 constraints on biomarker records in a high-altitude small catchment system, we collected both soil
28 and stream sediments along a 1000 m altitude transect (1500 – 2500 masl) in the Areguni
29 Mountains, a subrange of the Lesser Caucasus Mountains in Armenia. We utilize ~~the existence of~~
30 a treeline at ~ 2000 masl, which separates alpine meadow above from deciduous forest below, to
31 assess the relative contribution of upstream biomarker transport to local vegetation input in the
32 stream. We find that average chain length (ACL), hydrogen isotope (δD) and carbon isotope ($\delta^{13}\text{C}$)
33 values of *n*-alkanes are significantly different in soils collected above and below the treeline.
34 However, samples collected from the stream sediments do not integrate these signals quantitatively.
35 As the stream drops below the treeline, the ACL, δD and $\delta^{13}\text{C}$ values of *n*-alkanes preserved in
36 streambed sediments reflect a bias toward *n*-alkanes sourced from trees. This suggests that there
37 is either 1) minimal transportation of organic matter from the more open vegetation in higher
38 elevations, or 2) greater production of target biomarkers by trees and ~~shrubs~~ shrubs found at lower
39 elevations results in overprinting of stream signals by local vegetation. Though ~~these is latter~~
40 observations may preclude using *n*-alkanes to measure past treeline movement in these mountains,
41 δD values of biomarkers in fluvial deposits in these settings are more likely to record local

42 hydrological changes rather than reflect fractionation changes due to turnover in upstream
43 vegetation structure. ~~changes in upstream fractionation differences associated with vegetation~~
44 ~~turnover.~~

45

46 1. Introduction

47 Mountain regions are important hubs for biodiversity and can provide refuge for a number
48 of endemic species of flora and fauna (Antonelli et al., 2018). However, these high-altitude
49 environments are often particularly vulnerable to climate change (Guisan and Theurillat, 2000).
50 Therefore, gaining an understanding of sensitivity of these regions to past climate change is
51 important for projecting the effects of future climate change on fragile ecosystems. The ~~so-called~~
52 Caucasus Region in particular has been identified as a biodiversity hotspot covering the Republics
53 of Armenia, Georgia, Azerbaijan, and parts of the Russian Federation, Türkiye, and Iran, ~~supports~~
54 that supports a wide variety of plant and animal species (Zazanashvili, 2009; Gasparyan and
55 Glauberman, 2022). To better understand climate and environmental change in both the past and
56 the present, it is necessary to refine our understanding and interpretation of paleoclimate records
57 in this region. Plant wax biomarkers have been used in this region in both geological and
58 archaeological contexts to reconstruct past climates, therefore understanding modern variability
59 and transport processes will help refine these interpretations (Brittingham et al., 2019; Glauberman
60 et al., 2020; Malinsky-Buller et al., 2021, 2024; Trigui et al., 2019). Specifically, we are interested
61 in understanding the sedimentary processes involved in the formation, transport, recycling, and
62 accumulation of organic biomarkers in sedimentary archives and assessing whether these archives
63 record a local environmental signal or are a mix of local and transported organic material.

64 Normal alkanes (*n*-alkanes) are an important component of the epicuticular wax in
65 terrestrial plants. This waxy coating on plants protects against ultraviolet damage, water loss and
66 predation (Jetter et al., 2006). Specific compounds in this wax, such as *n*-alkanes, are a useful tool
67 for reconstructing past environmental changes through the analysis of the distribution of alkane
68 homologues as well as their stable hydrogen (δD) and carbon ($\delta^{13}C$) isotope values. Previous

69 research in the Greater and Lesser Caucasus Mountains has documented the applicability of the
70 average chain length (ACL) of leaf wax biomarkers as a tool for differentiating between grassy
71 and deciduous vegetation (Bliedtner et al., 2018; Trigui et al., 2019a), although on a global scale
72 ACL does not differentiate well between vegetation types (Bush and McInerney, 2013a).

73 ~~The biggest driver of~~ The carbon isotope ($\delta^{13}\text{C}$) values of plant tissue is primarily
74 determined by the photosynthetic pathway of the plant (Diefendorf and Freimuth, 2017). C_3 plants,
75 which thrive in areas with cooler growing season temperatures, have more negative $\delta^{13}\text{C}$ values
76 than do C_4 plants, which thrive in warmer growing season temperatures (Ehleringer et al., 1977).
77 C_3 vegetation is further influenced by water use efficiency, as water stress influences the c_i/c_a ratio
78 of plants (Farquhar et al., 1982). $\delta^{13}\text{C}$ values in lipids generally follow the same trends, and C_3
79 plants have more negative $\delta^{13}\text{C}$ lipid values than C_4 plants (Diefendorf and Freimuth, 2017).
80 However, carbon fractionation of lipids is not consistent in different classes of plants (Diefendorf
81 et al., 2011; Pedentchouk et al., 2008; Sikes et al., 2013). Currently, C_4 vegetation makes up around
82 3% of plant species in Armenia (Rudov et al., 2020), and was present in the Kalavan region during
83 the Holocene (Tornero et al., 2016).

84 The hydrogen isotope (δD) values of *n*-alkanes in terrestrial plants record the δD values of
85 environmental water (Sachse et al., 2012). This is typically reflective of δD values in precipitation,
86 though precipitation δD values can also undergo positive shifts due to soil evaporation. The δD
87 values of plant waxes are also influenced by fractionation during biological synthesis of lipids,
88 which imparts a strong negative fractionation on δD values, as well as transpiration of leaf water
89 (Gamarra et al., 2016). The fractionation between meteoric water and lipids is typically larger in
90 gymnosperms than in angiosperms (Oakes and Hren, 2016; Pedentchouk et al., 2008).

91 ———Despite the benefits in measuring δD and $\delta^{13}C$ values in *n*-alkanes for
92 understanding environmental and hydrological processes, not all the processes modifying isotope
93 values from plant to *n*-alkane deposition are well understood. Sedimentary integration is one of
94 the most poorly understood aspects of this process (Sachse et al., 2012). A number of studies on
95 the integration of leaf waxes in catchments have been published in recent years which help clarify
96 these processes (Alewell et al., 2016; Feakins et al., 2018; Häggi et al., 2016; Hemingway et al.,
97 2016; Ponton et al., 2014; Suh et al., 2019). Previous studies on the integration of organic
98 biomarkers have produced mix results, with some demonstrating spatial integration of catchment
99 signals (Alewell et al., 2016; Feakins et al., 2018; Hemingway et al., 2016), whereas others did not
100 observe this (Häggi et al., 2016; Ponton et al., 2014). However, these previous studies typically
101 focused on very large river systems, which will undergo different transport processes than the first-
102 order streams analyzed in this study. A number of these studies (Alewell et al., 2016; Feakins et
103 al., 2018; Hemingway et al., 2016; Ponton et al., 2014) also observed seasonal differences in
104 biomarker load in river sediments. Collecting seasonal samples in the Areguni Mountains, as well
105 as testing these processes in other first order streams, could further help clarify the transport
106 processes measured in this setting.

107 ~~However, most of these studies have focused on large river systems rather than first order~~
108 ~~streams.~~ Thus, the sedimentary processes involved in the formation, transport, recycling, and
109 accumulation of organic biomarkers in first and second order streams are not well understood. One
110 challenge in assessing these processes in small streams is that the environment and plant
111 communities are often homogenous, and thus it is not possible to differentiate between local and
112 upstream transported organic material. To better understand transport processes affecting organic
113 material in small catchments, we studied a set of streams in the Dany River, a tributary of the

114 Barepat River, located in the Areguni Mountains in the Lesser Caucasus Range. This stream
115 system is divided into two distinct ecological regions by the treeline (at ~ 2000 masl), which
116 separates alpine meadow above the tree line (2000 – 2500 masl) from deciduous forest below
117 (1500 – 2000 masl). To evaluate the input of *n*-alkanes from upstream transported organic material
118 relative to vegetation near the stream, we collected soil samples on the slopes of the mountains
119 from both above- and below the treeline throughout the watershed and sediments deposited in the
120 streambed along an elevation transect. ~~Comparing the hillslope sedimentary biomarkers and the~~
121 ~~streambed sedimentary~~ Comparison of the hillside and streambed sedimentary *n*-alkanes
122 ~~biomarkers~~ allows assessment of the input of *n*-alkanes locally produced by vegetation compared
123 to those transported in stream sediments within the catchment.

124 An additional motivation of this research is that treelines are a vulnerable feature of higher
125 altitude environments. Previous research in the Areguni Mountains study area has assessed the
126 relationship between treeline dynamics and climate forcing in the past (Ghukasyan et al., 2010;
127 Malinsky-Buller et al., 2021; Montoya et al., 2013; Tornero et al., 2016), ~~and the~~ Pleistocene
128 sediments uncovered at archaeological sites at Kalavan village within this area have the potential
129 to reconstruct this relationship through the analysis of plant wax biomarkers deposited in fluvial
130 sediments. However, in order to reconstruct these systems in the past it is important to understand
131 modern biomarkers integration processes in the first and second order streams and their potential
132 effects on the sedimentary archives of the Areguni Mountains.

133 **2. Methods**

134 **2.1 Sample Collection and Extraction**

135 Hillslope soil samples were collected in September 2018 along an altitude transect (1500 – 2500
136 masl) above the Dany River watershed, a first order tributary of the Barepat River in the Areguni

137 Mountains, Armenia (Fig 1), which traverses the treeline at ~2000 masl. Forest vegetation is
138 predominantly oak (*Quercus macranthera*), beech (*Fagus orientalis*) and hornbeam (*Carpinus*
139 *orientalis*), while above treeline alpine meadow is comprised of *Heracleum sp.* and *Senecio sp.*
140 (Joannin et al., 2022; Volodicheva, 2002). Soil samples were collected by first clearing the top ~10
141 cm of soil to remove roots. Stream bed sediment samples were collected from the Dany River
142 throughout the altitude transect at intervals of ~100 m in altitude. In all cases, roughly 100 g of
143 sediment were collected for extraction of *n*-alkanes. ~~In order to extract *n*-alkanes, s~~ Samples were
144 extracted using a Soxhlet apparatus with 2:1 dichloromethane:methanol for 48 hours. Following
145 lipid extraction, *n*-alkanes were separated from total liquid extract by passing samples through a
146 column of activated silica gel (1.25 g) in baked Pasteur pipettes with 2 mL hexane (non-polar
147 fraction), 4 mL dichloromethane (slightly polar fraction) and 4 mL methanol (polar fraction). *n*-
148 alkanes were quantified through the analysis of the hexane fraction. We quantified *n*-alkanes using
149 a BP-5 column (30 m × 0.25 mm i.d., 0.25 μm film thickness) with He as the carrier (1.5 ml/min).
150 Oven temperature was set at 50 °C for 1 min, ramped to 180 °C at 12 °C/min, then ramped to
151 320 °C at 6 °C/min and held for 4 min. (Brittingham et al., 2017; Smolen and Hren, 2023). We
152 measured a standard mixture of *n*-alkanes (C₂₀-C₃₃) of known concentration to correct for mass
153 dependent response decreases in longer chain *n*-alkanes. ~~*n*-alkanes were separated with silica gel~~
154 column chromatography and quantified on a Thermo Scientific Trace GC Ultra (Manufacturer)
155 fitted with a split-splitless (SSL) injector and flame ionization detector (FID) using a BP-5 column
156 (30 m × 0.25 mm i.d., 0.25 μm film thickness) with He as the carrier (1.5 ml/min). Odd over even
157 predominance (OEP) (Eq. 1) and average chain length (ACL) (Eq. 2) were used to evaluate
158 distributions of *n*-alkanes- (Bush and McInerney, 2013b) ~~(REF)~~. We also calculated P_{aq}, an *n*-

159 alkane proxy to evaluate the possible biomarker contribution of aquatic and emergent plants (Eq.
160 3) (Ficken et al., 2000).

$$161 \text{ OEP} = \frac{C_{25} + C_{27} + C_{29} + C_{31} + C_{33}}{C_{24} + C_{26} + C_{28} + C_{30} + C_{32}}$$

$$162 \text{ ACL} = \frac{25 * C_{25} + 27 * C_{27} + 29 * C_{29} + 31 * C_{31} + 33 * C_{33}}{C_{25} + C_{27} + C_{29} + C_{31} + C_{33}}$$

$$163 \text{ Paq} = \frac{C_{23} + C_{25}}{C_{23} + C_{25} + C_{29} + C_{31}}$$

164 2.3 Stable Isotope Analysis

165 δD and $\delta^{13}\text{C}$ values of individual *n*-alkanes were measured with a Thermo GC-Isolink coupled
166 with a Thermo Scientific MAT 253 (manufacturer) isotope ratio mass spectrometer with a BP-5
167 column (30 m \times 0.25 mm i.d., 0.25 μm film thickness). Oven temperature was set at 50°C for 1
168 min, ramped to 180°C at 12°C/min, then ramped to 320°C at 6°C/min and held for 4 min. Internal
169 standards (Mix A5 from A. Schimmelman) were run every four samples across a range of
170 concentrations (5-30 V/s) to correct for size effects. Standard ~~deviations-errors~~ were 0.54‰ for
171 $\delta^{13}\text{C}$ and 43‰ for δD . Isotope ratios (R) were converted to δX ($\delta^{13}\text{C}$ and δD) values (Eq. 3) and
172 are expressed in permill (‰).

$$173 \delta\text{X} = \left(\frac{R_{\text{Sample}}}{R_{\text{Standard}}} - 1 \right) * 1000$$

174

175 3. Results

176 3.1. Alkane abundances

177 The most abundant alkane homolog in samples collected in the Areguni Mountains is the
178 C₂₉ or C₃₁ alkane, which is typical for terrestrial plants ([see S1-S4 for illustrative chromatograms](#)).
179 Odd numbered alkanes are significantly more abundant than even numbered alkanes, and the OEP
180 of all samples ~~averages is~~ 11.2, with a range from 7.4-18.4. There is no significant difference
181 between the mean OEP of soil (11.1) and stream (11.3) samples in the watershed. These values
182 are similar to those previously measured in the Greater and Lesser Caucasus Mountains (Bliedtner
183 et al., 2018; Trigui et al., 2019).

184 The mean average chain length (ACL) of all samples averages 29.7, with a range from 28.4
185 to 31.8 (Fig 3). In soils above the treeline, the ~~average-mean~~ ACL value is 30.6 (range of 29.8-
186 31.8). In soils below the treeline, the ~~average-mean~~ ACL value is 29.5 (range of 28.4-30.4). There
187 is a significant difference between the average ACL values of the [n-alkanes in](#) above treeline and
188 below treeline soils ([Student's t-test, p<0.001, n=30](#)). Stream sediment above the treeline ~~has~~ an
189 average ACL value of 29.7 (range of 29.1-30.2) and stream sediments below the treeline have an
190 average ACL value of 29.3 (range of 28.6 ~~to~~ 30.0). The stream sediments from below the treeline
191 have a significantly ([Student's t-test, p<0.001, n=21](#)~~p<0.001~~) lower average ACL value than those
192 above the treeline.

193 The P_{aq} values of *n*-alkanes in these samples suggests a mostly terrestrial origin of the
194 organic matter. Higher P_{aq} values indicate contributions of floating and emergent macrophytes.
195 However, we do not find a ~~significant~~ difference between the P_{aq} values in the stream sediments
196 when compared to the soil samples, indicating that the organic load of the stream sediments is
197 mostly of terrestrial origin. Terrestrial plants have average P_{aq} values of 0.09, with emergent plants
198 averaging 0.25 (Ficken et al., 2000). Only eight of the 51 samples in this study had P_{aq} values above
199 0.20, four stream and four soil samples. This indicates that there was not a significant contribution

200 of aquatic plants in the Dany stream sediments, and the biomarker load is primarily terrestrial in
201 origin.

202 3.2. δD and $\delta^{13}C$ values

203 The $\delta^{13}C$ values in soils and stream sediments collected from the Areguni Mountains reflect
204 a C_3 landscape, which is typical in Armenia. $\delta^{13}C$ values in all samples ranged from -36.0 to -
205 32.3‰ (Fig 4). The range is similar for both soil samples (-35.9 to -32.3‰) and stream samples (-
206 36.0 to -32.5‰). However, there is a significant difference in the $\delta^{13}C$ values of above and below
207 treeline samples, both in the stream and soil samples collected. Above the treeline, $\delta^{13}C$ values in
208 soils averages -34.9‰, and below the treeline soil alkanes average -33.3‰ ($p < 0.0001$, student's t-
209 test, $n=30$). Stream sediment $\delta^{13}C$ values average -35.0‰ above the treeline and -33.6‰ below
210 the treeline ($p < 0.0001$, student's t-test, $n=21$). ~~In both cases, these values are significant~~
211 ~~($p < 0.0001$, student's t test).~~ $\delta^{13}C$ values in stream samples exhibit a step-like behavior, with ~2‰
212 shift to more negative values as the stream drops below the treeline.

213 The δD values measured in soil samples collected in the catchment ranged from -144 to -
214 185‰ (Fig 5). These values were significantly ($p < 0.001$, student's t test). ~~More~~ more negative in
215 above treeline sediments (-175‰) than in below treeline sediments (-156‰) ($p < 0.001$, student's
216 t-test, $n=30$). δD values were also more negative in stream sediment samples collected above ~~This~~
217 ~~is also true in sediment collected from stream samples, which are significantly more negative~~
218 ~~above~~ the treeline (-175‰) than below the treeline (-158‰) ($p < 0.001$, student's t-test, $n=21$). As
219 with the $\delta^{13}C$ values, the δD values of stream sediment samples show sudden change as the stream
220 drops below the treeline.

221 4. Discussion

222 4.1 Integration of local and upstream soil n -alkanes into the river sediments

223 The hillslope soil leaf wax ACL (Fig 3), δD , $\delta^{13}C$ (Fig. 4) and δD ACL (Fig. 5) show a step-
224 like change at the treeline, indicating a significant separation between upstream (above treeline)
225 and downstream (below treeline) soils. Using this separation, it is possible to assess the
226 contributions and integration of upstream vs. downstream soils to the streambed sediments along
227 the altitude transect. The step-like transition in streambed δD and $\delta^{13}C$ values indicates an over-
228 printing of upstream alkane isotope values by input from deciduous vegetation. Thus, local
229 production largely outweighs upstream transport in this setting. However, to firmly evaluate the
230 upstream and downstream hillslope soil contribution to streambed sediments, there is a need to
231 quantitatively evaluate the area-weighted production of *n*-alkanes above and below the treeline.

232 **4.2. Modeling *n*-alkane production and estimating upstream transport and integration**

233 To further evaluate the integration of *n*-alkanes above and below the treeline, we created a
234 mixing model that calculates the expected δD , $\delta^{13}C$ and ACL values at each one of the sampling
235 locations based on the *n*-alkane production of hillslope sediments above each streambed sampling
236 point (Fig. 6). This-Our mixing model assumes that the *n*-alkanes in the river are a function of the
237 weighted *n*-alkane production above the sampling location.

238 The parameters we used for this-our mixing model are: 1. Satellite images (Google Earth)
239 to map the areas of tree and grass sediment covered by alpine meadow and forest vegetation
240 throughout the Dany River catchment. 2. An estimate of net primary productivity of organic
241 material production in grasses and trees (grams per area) (Brun et al., 2022). 3. Estimates of *n*-
242 alkane production in grasses and trees in the Greater and Lesser Caucasus Mountains (grams of *n*-
243 alkane per gram of organic material) (Bliedtner et al., 2018; Trigui et al., 2019). 4. End member
244 values of δD , $\delta^{13}C$ and ACL derived from the average hillslope soils above and below the treeline.
245 At each sample point within the catchment, we first calculated the upstream area covered by the

246 two dominant vegetation types within the catchment (deciduous forest and alpine meadow) (Figure
247 6). This area was then multiplied by the previously mentioned constants (Table 1). By multiplying
248 these terms (area x organic mass production x *n*-alkane production x end member soils value), we
249 created an *n*-alkane production map for the Dany River catchment. ~~Using this map, we~~Using this
250 method, we calculated, ~~for each riverbed sampling location~~, the amount of grass and tree *n*-alkanes
251 produced on the hillslopes above the sampling locations and the expected δD , $\delta^{13}C$ and ACL values
252 for each stream sampling location (Figure 7a, 7c, 7e).

253 We compared the results of ~~this our~~ mixing model with the measured δD , $\delta^{13}C$ and ACL
254 in the streams. Stream sediment samples collected above the treeline (from ~2000-2600 masl) fall
255 within the range of expected values, however, samples below the treeline consistently over-sample
256 ~~deciduous-sourced~~ *n*-alkanes sourced from below treeline vegetation. Measured δD , $\delta^{13}C$ and ACL
257 values do not have a linear relationship with the expected values based on vegetation area (Fig 7b,
258 7d, 7f). These measured values would produce under-estimates of the upstream area of alpine
259 grasses, yielding incorrect reconstructions of paleo-vegetation in sedimentary records. Comparing
260 the mixing model with the observations indicates that an area-weighted mixing process is not an
261 adequate model for explaining the *n*-alkanes signal in the streambed sediments. A simple and
262 straightforward way to interpret this discrepancy is that an area-weighted quantitative integration
263 of *n*-alkanes is not a good model for describing this catchment system, and that local production
264 is much larger than transported organic material.

265 However, there are still other factors that may be driving this process that our mixing model
266 does not account for. First, the average slope of forested areas in the Dany watershed is higher than
267 those in grassy areas. These steeper slopes would cause more sediment transport into the stream
268 bed. Second, ~~though~~although production of *n*-alkanes in grasses and trees is not significantly

269 different in the Greater and Lesser Caucasus Mountains, concentrations are higher in soils in
270 deciduous areas (Bliedtner et al., 2018; Trigui et al., 2019). This retention of more biomarkers in
271 forest soils would also increase the contribution of deciduous alkanes into the stream bed. Third,
272 stream downcutting into older sediments has the potential to re-mobilize stored organic carbon,
273 which may contain a greater load of deciduous *n*-alkanes. However, analysis of pollen from a
274 ~~nearby~~-lake core nearby (~ 5km from the Dany catchment) -in the Areguni Mountains shows a
275 gradual shift over the last 4000 years from a grass-dominated landscape to the deciduous forest
276 present today (Joannin et al., 2022b). Therefore, stored biomarkers are more likely to be grass-
277 dominant, and this is unlikely to explain the measured bias to deciduous alkanes.

278 Since *n*-alkanes in ~~this~~-the first order stream in this study do not quantitatively integrate *n*-
279 alkanes based on the upstream area of different vegetation types, this likely precludes the use of
280 *n*-alkanes as a tool to reconstruct vertical treeline movement in this setting. However, this is a
281 benefit for attempts to reconstruct hydrological changes through the analysis of δD values in *n*-
282 alkanes. Given the ~20‰ difference in apparent fractionation (ϵ) values for above and below
283 treeline sediments, changes in upstream vegetation cover would alter measured δD values in *n*-
284 alkanes in sedimentary archives. Without this quantitative integration, *n*-alkanes measured in the
285 Pleistocene sediments found in this watershed are more likely to reflect changes in δD values of
286 precipitation, and therefore would serve to reconstruct hydrological cycles, rather than changes in
287 upstream vegetation cover. Since $\delta^{13}C$ and ACL of *n*-alkanes are also different in above and below
288 treeline sediments, these other analyses would also be useful to identify periods with large changes
289 in treeline that might complicate interpretation of δD values.

290 In order to illustrate this point, we present hypothetical records of biomarker δD values
291 from three points in the Dany watershed (Fig. 7) documenting 20‰ and 30‰ shifts in precipitation

292 δD values. We use δD values of precipitation from water samples collected at the nearest
293 meteorological station with isotope data in Armenia (Dilijan, (Brittingham et al., 2019b)) Given
294 the lack of quantitative integration in the catchment, a paleoclimate record from either above (A)
295 or below (C) treeline would record the shift in precipitation δD values. Below treeline sedimentary
296 records, with stream organic biomarker load overprinted by local vegetation production, would
297 likely provide a means to reconstruct the δD precipitation values. However, records near the
298 treeline (B) would be ~~heavily affected~~influenced by changes in apparent fractionation values
299 associated with changes in vegetation around the stream. Co-occurring climate forcing of shifts in
300 δD values of precipitation and changes in treeline altitude would cause paleoclimate records in this
301 zone to over-estimate the magnitude of precipitation δD value shifts.

~~302 Previous studies on the integration of organic biomarkers has produced mix results, with
303 some demonstrating spatial integration of catchment signals, whereas others did not observe this
304 However, these previous studies typically focused on very large river systems, which will
305 undergo different transport processes than the first order streams analyzed in this study. A number
306 of these studies also observed seasonal differences in biomarker load in river sediments.
307 Collecting seasonal samples in the Areguni Mountains, as well as testing these processes in other
308 first order streams, could further help clarify the transport processes measured in this setting.~~

309
310

311 5. Conclusion

312 Sediment and stream samples from the Areguni Mountains, a subrange of the Lesser Caucasus
313 Mountains in Armenia, demonstrate that there is a significant difference in hillslope soil δD , $\delta^{13}C$
314 and ACL values above and below treeline. *n*-alkanes in sediments in the Areguni Mountains can

315 be used to differentiate between the above and below treeline sediments. However, *n*-alkanes
316 extracted from stream sediments reflect their local area, rather than demonstrating transport from
317 the higher-altitude alpine meadow. These results provide a complication for attempts to reconstruct
318 changes in past treeline in this mountain range, given that the biomarker load in stream does not
319 reflect the relative area of different upstream vegetation types. However, these results simplify
320 interpretation of past *n*-alkane δD values, as apparent fractionation differences between grasses
321 and trees are less likely to impart a significant influence on δD values in stream bed *n*-alkanes.

322 **6. Competing interests**

323 The contact author has declared that none of the authors has any competing interests

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339

340 **Works Cited/References**

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	<u>Conc</u> ¹	<u>NPP</u> ²	<u>ACL</u> ³	$\delta^{13}\text{C}$ ³	δD ³
<u>Forest</u>	<u>7.69</u>	<u>1099.4</u>	<u>29.5</u>	<u>-33.3</u>	<u>-156</u>
<u>Alpine Meadow</u>	<u>3.03</u>	<u>719.1</u>	<u>30.6</u>	<u>-34.9</u>	<u>-175</u>

Table 1: Constants used for mixing model. 1: *n*-alkane concentration in grasses and trees in the Caucasus Mountains (grams of *n*-alkane/gram of organic material) (Bliedtner et al., 2018; Trigui et al., 2019). 2: NPP for forest and grassland areas (grams per area) (Brun et al., 2022). 3: mean values for ACL, $\delta^{13}\text{C}$ and δD (this study)

555 Figure Captions

556 Figure 1: (Left) Topographic map (ASTER Global DEM) of Armenia with inset map of
557 sampling location (black box) (Right) Inset map of soil (yellow circles) and stream (blue circles)
558 samples collected in the Areguni Mountains, along with the limit of the Barepat (dashed line)
559 and Dany watersheds (solid line)

560 Figure 2: The δD and $\delta^{13}C$ values of *n*-alkanes extracted from above treeline (green squares) and
561 below treeline (red triangles) sediments

562 Figure 3: The average chain length (ACL) values of *n*-alkanes extracted from above treeline
563 (~~green-yellow~~ squares) and below treeline (~~red-green~~ squares) sediment and stream (~~blue~~
564 ~~triangles~~) ~~sediments~~ across the sampling elevation gradient

565 Figure 4: The $\delta^{13}C$ values of *n*-alkanes extracted from above treeline (yellow squares) and below
566 treeline (green squares) sediment ~~above treeline (green squares) and below treeline (red squares)~~
567 ~~and stream (blue triangles) sediments~~ across the sampling elevation gradient

568 Figure 5: The δD values of *n*-alkanes extracted from above treeline (yellow squares) and below
569 treeline (green squares) sediment ~~above treeline (green squares) and below treeline (red squares)~~
570 ~~and stream (blue triangles) sediments~~ across the sampling elevation gradient

571 Figure 6: Mixing model used to calculate expected values of stream sample points. Upstream
572 watershed area covered by deciduous forest (green) and alpine meadow (yellow) was calculated
573 at each sample location (blue dots). Point watershed for sample 24 (dashed line) is shown here as
574 an example.

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576 Figure 76: Comparison of the measured ACL, δD and $\delta^{13}C$ values against expected values of
577 stream sediments. Dashed line represents the range of expected values from stream sediments if
578 vegetation was integrated equally by area

579 Figure 87: A photograph of the Dany watershed with hypothetical paleoclimate record from
580 three locations: (A, dashed line) Below treeline, (B, solid line) near treeline with fluctuations in
581 treeline altitude, and (C, dotted line) above treeline

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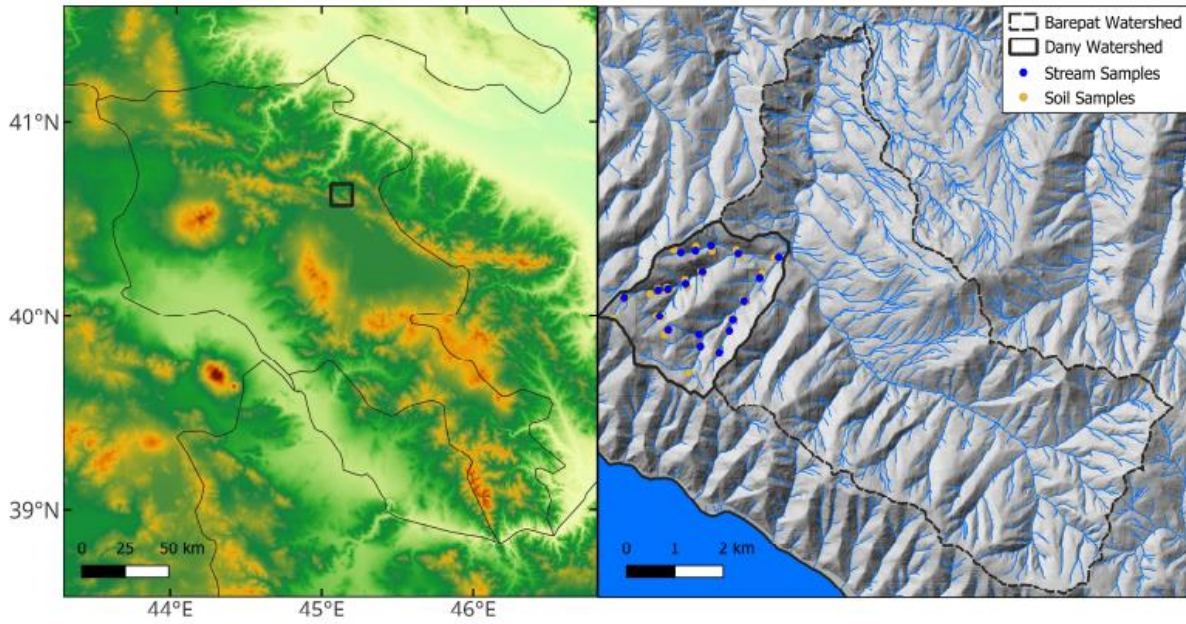
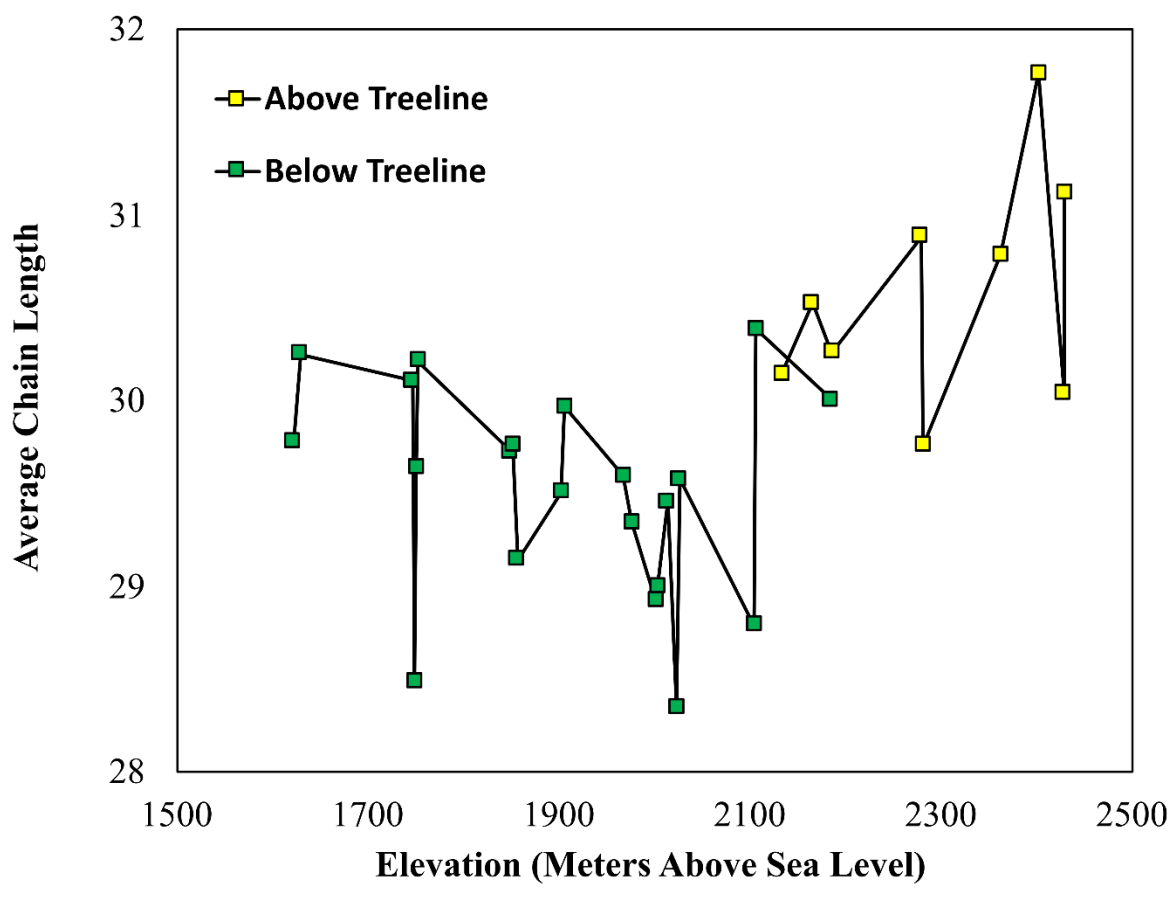
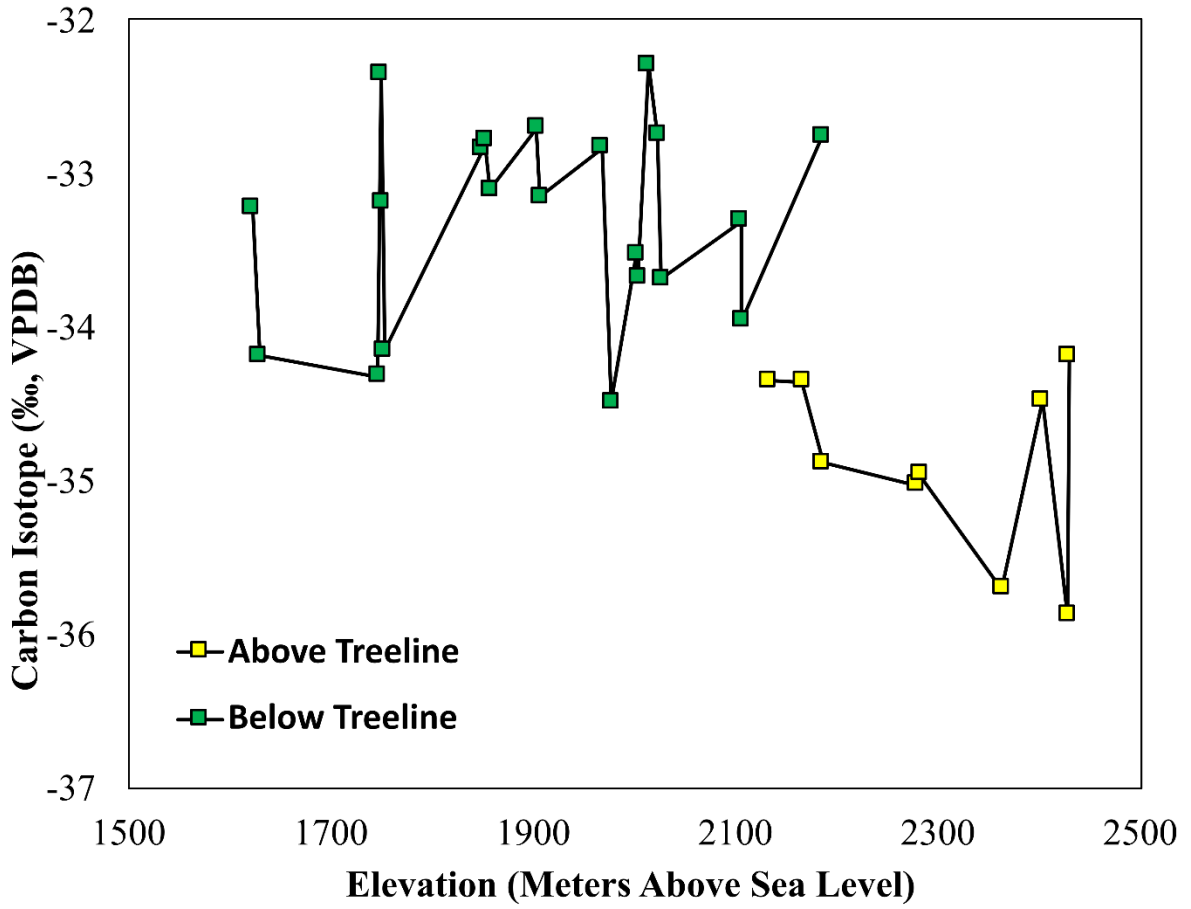


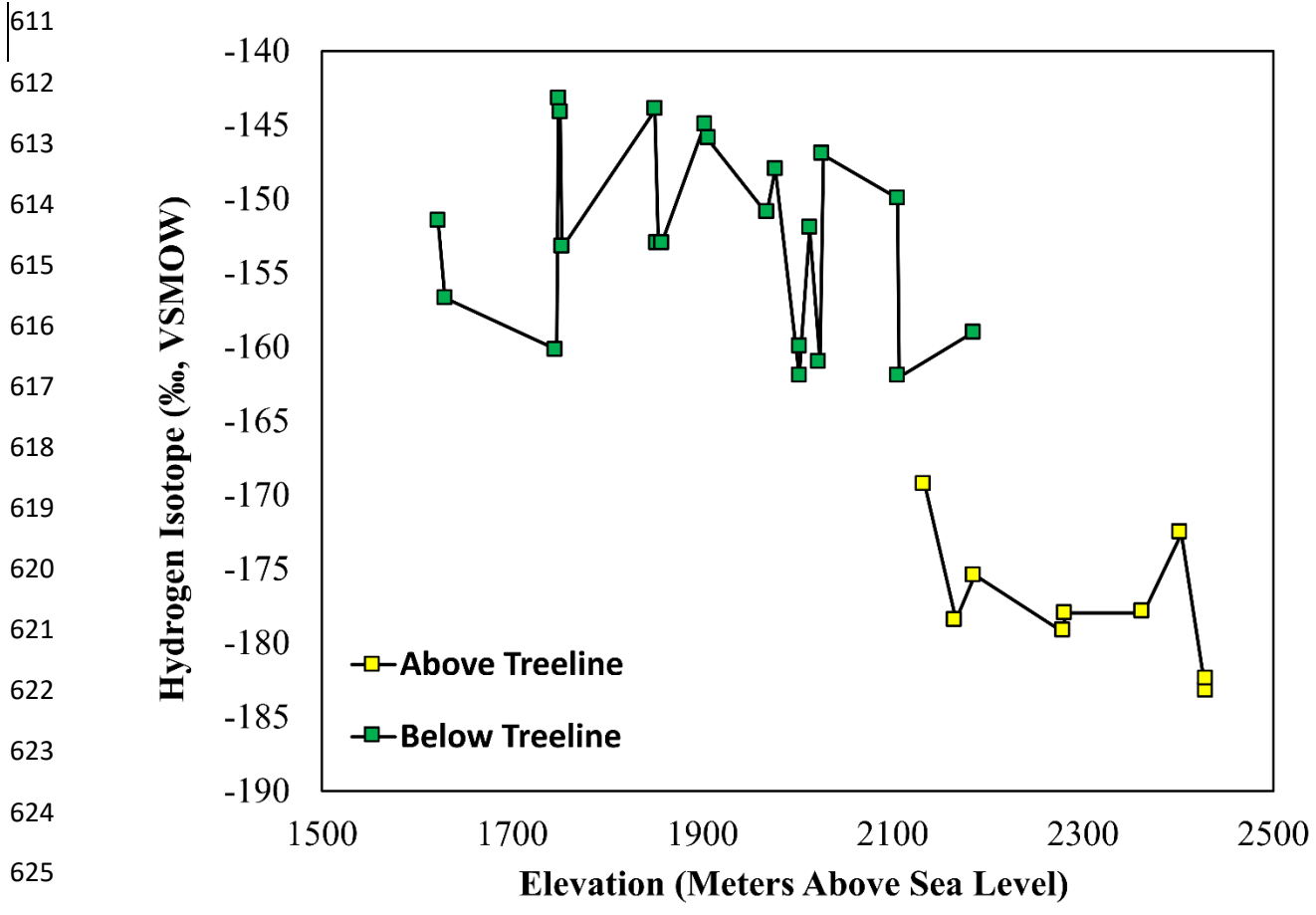
Figure 1: (Left) Topographic map of Armenia with inset map of sampling location (black box) (Right) Inset map of soil (yellow circles) and stream (blue circles) samples collected in the Areguni Mountains, along with the limit of the Barepat (dashed line) and Dany watersheds (solid line)

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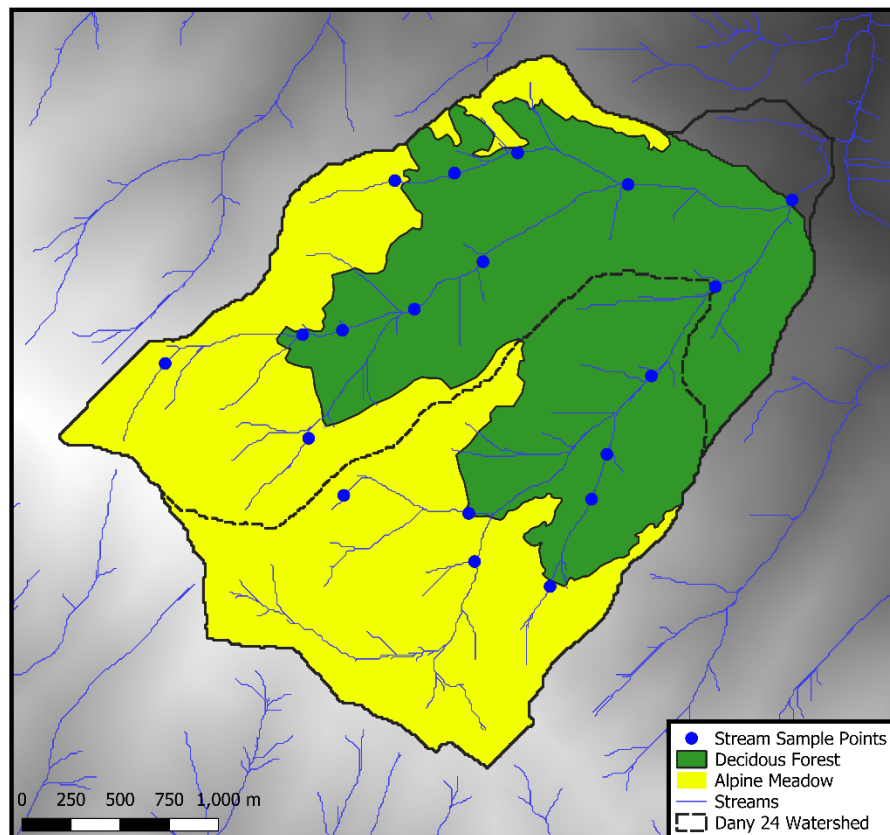


Table 1: Constants used for mixing model. 1: *n*-alkane concentration in grasses and trees in the Caucasus Mountains (grams of *n*-alkane/gram of organic material) (Bliedtner et al., 2018; Trigui et al., 2019). 2: NPP for forest and grassland areas (grams per area) (Brun et al., 2022). 3: mean values for ACL, $\delta^{13}\text{C}$ and δ (this study)**Figure 6:** Mixing model used to calculate expected values of stream sample points. Upstream watershed area covered by deciduous forest (green) and alpine meadow (yellow) was calculated at each sample location (blue dots). Point watershed for sample 24 (dashed line) is shown here as an example.

