

1 **Variable contribution of wastewater treatment plant effluents to downstream nitrous**
2 **oxide concentrations and emissions**

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11
12 **Abstract**

13 Nitrous oxide (N₂O), a potent greenhouse gas and ozone-destroying agent, is produced during
14 nitrogen transformations in both natural and human-constructed environments. Wastewater
15 treatment plants (WWTPs) produce and emit N₂O into the atmosphere during the nitrogen removal
16 process. However, the impact of WWTPs on N₂O emissions in downstream aquatic systems
17 remains poorly constrained. By measuring N₂O concentrations at a monthly resolution over a year
18 in the Potomac River Estuary, a tributary of Chesapeake Bay in the eastern United States, we found
19 a strong seasonal variation in N₂O concentrations and fluxes: N₂O concentrations were larger in
20 fall and winter but the flux was larger in summer and fall. Observations at multiple stations across
21 the Potomac River Estuary revealed hotspots of N₂O emissions downstream of WWTPs. N₂O
22 concentrations were higher at stations downstream of WWTPs compared to other stations (median:
23 21.2 nM vs 16.2 nM) despite the similar concentration of dissolved inorganic nitrogen, suggesting
24 the direct discharge of N₂O from WWTPs into the aquatic system or a higher N₂O production yield
25 in waters influenced by WWTPs. Meta-analysis of N₂O measurements associated with WWTPs
26 globally revealed variable influence of WWTPs on downstream N₂O concentrations and
27 emissions. Since wastewater production has increased substantially with the growing population
28 and is projected to continue to rise, accurately accounting for N₂O emissions downstream of the
29 WWTPs is important for constraining and predicting future global N₂O emissions. Efficient N₂O
30 removal, in addition to dissolved nitrogen removal, should be an essential part of water quality
31 control in WWTPs.

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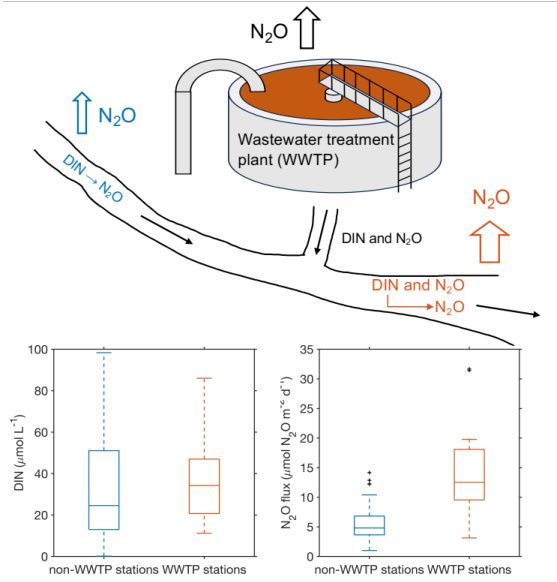
33 Key words: nitrous oxide, greenhouse gas emission, nitrogen pollution, wastewater treatment
34 plants, spatial and seasonal variation

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36 Summary: Wastewater treatment plants (WWTPs) are known to be hotspots of greenhouse gas
37 emissions. However, the impact of WWTPs on the emission of the greenhouse gas N₂O in
38 downstream aquatic environments is less constrained. We found spatially and temporally variable
39 but overall higher N₂O concentrations and fluxes in waters downstream of WWTPs, pointing to
40 the need for efficient N₂O removal in addition to treating nitrogen in WWTPs.

41

42 Graphical abstract



43

44 **Introduction**

45 Nitrogen (N) enters the aquatic environment from agricultural and urban runoff, atmospheric
46 deposition, and wastewater treatment plants (WWTPs), potentially leading to eutrophication,
47 especially in densely populated regions (Galloway et al., 2008; Morée et al., 2013). During
48 microbial transformations of N in aquatic systems (e.g., nitrification and denitrification), a
49 powerful greenhouse gas and ozone depleting agent – N₂O – is produced (Quick et al., 2019).
50 Aquatic systems are large but highly variable sources of N₂O to the atmosphere (Wang et al.,
51 2023). For example, on a global basis, 0.04 - 0.291 Tg N yr⁻¹ and 0.04 - 3.6 Tg N yr⁻¹ of N₂O is
52 estimated to outgas from rivers and estuaries, respectively (Murray et al., 2015; Maavara et al.,
53 2019; Yao et al., 2019; Rosentreter et al., 2023). The high end of the estimates in these inland and
54 coastal waters approaches the scale of the global marine N₂O emissions (2.5 - 4.3 Tg N yr⁻¹ in Tian
55 et al., 2020). The large uncertainty in the estimate of aquatic N₂O emission is partly due to high
56 spatial and temporal variabilities of N₂O flux within/across rivers and estuaries and the lack of
57 observations to capture such variability. Therefore, sampling and measurements of N₂O
58 concentration at high spatial and temporal resolutions would be desirable to constrain aquatic N₂O
59 emission.

60
61 The major factors that appear to correlate with N₂O concentration are dissolved inorganic nitrogen
62 (DIN) and oxygen status (Hu et al., 2016; Zheng et al., 2022). Waste and wastewater release large
63 amounts of DIN into the aquatic environment. In addition, waste and wastewater emit ~0.3 Tg N
64 yr⁻¹ of N₂O (estimated from 2007-2016) into the atmosphere globally, an amount that is
65 continuously increasing at a rate of 0.04±0.01 Tg N yr⁻¹ per decade (Tian et al., 2020). N₂O
66 emission from WWTPs accounts for ~5.2% of total N₂O emission in 2021 in the United States
67 (EPA, 2023). N₂O emissions from different WWTPs are highly variable, and are normally
68 calculated as a function of DIN loading into the WWTPs, using an N₂O emission factor
69 (Kampschreur et al., 2009). N₂O emission factors range from 0.16% to 4.5% (N₂O emitted/DIN
70 loading) (Eggleston et al., 2006; De Haas and Andrews, 2022). In addition to direct emission from
71 the WWTPs, N₂O can be discharged via WWTP effluent and produced due to DIN release from
72 WWTP effluent into the creeks, rivers, and other downstream aquatic systems (McElroy et al.,
73 1978; Beaulieu et al., 2010; Masuda et al., 2018). However, the impact of WWTPs on downstream
74 N₂O concentration is less studied and the downstream N₂O emission remains poorly constrained.

75 Here we specifically compared the N₂O concentration upstream and downstream of the WWTPs
76 in order to assess the impact of WWTPs on N₂O emission, which could help to constrain the
77 emission factor associated with the WWTPs effluents.

78
79 The Potomac River is a major tributary of the Chesapeake Bay – the largest estuary in the United
80 States. The Potomac River Estuary is located in a highly populated area, mainly surrounded by
81 Washington, D.C., and the states of Virginia and Maryland in the eastern United States. The annual
82 mean discharge of Potomac River from 1895 to 2002 measured at Chain Bridge near Washington,
83 DC was 321 m³ s⁻¹ with a large interannual variability (Jaworski et al., 2007). The annual total
84 nitrogen loading was estimated to be around 27.7 ×10⁶ kg N year⁻¹ in 2008-2009 (Bricker et al.,
85 2014). The Potomac River Estuary has experienced ecological degradation for decades partly due
86 to excess nutrient inputs including from the effluents of WWTPs (Bricker et al., 2014; Jaworski et
87 al., 2007). For example, the Blue Plains Advanced WWTP in Washington, D.C. is one of largest
88 WWTPs in the world, treating an average of ~1454 million liters of water per day. Pioneering
89 work in 1978 showed that Blue Plains WWTP was a large source of nitrogen to the Potomac River
90 Estuary, triggering high N₂O production and concentration downstream (McElroy et al., 1978).
91 Thanks to higher standards mandated by governmental agencies (nitrogen concentration in
92 effluents below 7.5 mg L⁻¹) starting in 1980s and the technical improvements in N removal from
93 the wastewater, the nitrogen concentration in effluents of WWTPs in the Potomac River has
94 decreased substantially (Pennino et al., 2016). However, the concurrent effect on N₂O
95 concentration is largely unknown. The Department of Environmental Quality (DEQ) of Virginia
96 maintains an approximately monthly routine monitoring program for water quality (e.g., nitrogen
97 concentration, phosphorus concentration, chlorophyll concentration) and physical properties (e.g.,
98 temperature, salinity, pH, and dissolved oxygen concentration) in the Potomac River Estuary but
99 not for N₂O. Therefore, we collaborated with DEQ of Virginia to measure the spatial and temporal
100 variation of N₂O concentrations in the Potomac River Estuary.

101

102 **Materials and Methods**

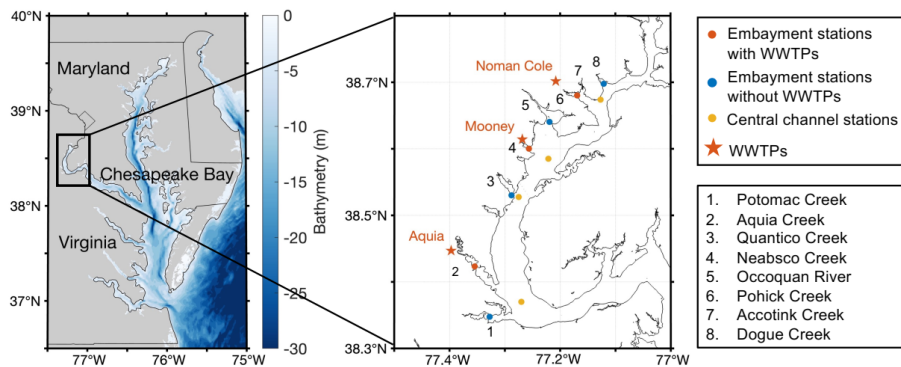
103 **Sample collection for N₂O and nutrients**

104 Surface waters at ~0.5 m depth at eleven stations in the tidal Potomac River Estuary were sampled
105 monthly or bimonthly (depending on the weather) on a vessel (Grady White 208) for the analysis

106 of DIN concentration, and both concentration and nitrogen isotopes of N₂O from April 2022 to
107 May 2023 (Figure 1). The eleven stations are characterized into 3 groups: embayment downstream
108 of WWTPs, embayment not associated with WWTPs, and the central channel of the Potomac
109 River. Three embayment stations downstream of WWTPs are associated with three different
110 WWTPs: Noman Cole, Mooney and Aquia, all of which implement tertiary treatment of the
111 wastewater. We obtained the volume discharge and total N in treated water of each WWTP from
112 Discharge Monitoring Reporting required by Virginia Pollutant Discharge Elimination System
113 permit. Noman Cole WWTP discharges ~140.8 million liters of water and 370 kg N per day into
114 Pohick Creek. Mooney WWTP discharges ~54.9 million liters of water and 147 kg N per day into
115 the Neabsco Creek. Aquia WWTP discharges much less water and N into the Aquia Creek (~21.2
116 million liters per day and 35 kg N per day). The distances from the embayment stations
117 downstream of WWTPs to Noman Cole, Mooney, Aquia WWTPs were approximately 4, 1.8 and
118 5.8 km, respectively.

119
120 The embayment stations were 2-3 meters deep while the average depth of central channel stations
121 was around 8 meters. The embayment stations have been routinely sampled for water quality
122 analyses by the DEQ of Virginia since the early 1970's. The central channel stations were added
123 for this study. The purposes of this sampling design are to evaluate the impact of WWTPs on
124 downstream distribution of DIN and N₂O, and to compare DIN and N₂O concentrations between
125 edge and central channel of the river. The central channel is likely affected both by the Potomac
126 mainstem flow and by the input from tributaries, while the embayment stations may be mainly
127 affected by water flow from tributaries but also influenced by the tidal cycle (see the salinity
128 change in Supplementary Figure 1b). While estuarine N₂O concentrations could be affected by
129 tides (Gonçalves et al., 2015), sampling was not always conducted at the same tidal state due to
130 logistic difficulties. Triplicate water samples for N₂O concentrations and isotopes were collected
131 via a submersible pump into 60 mL serum bottles after overflowing three times the bottle's volume.
132 After removing 3 mL water to create a 3 mL air headspace via a syringe, the serum bottles were
133 immediately sealed with butyl stoppers and aluminum crimps and preserved with 0.5 mL of 10 M
134 NaOH solution to stop biological activities. NaOH has been shown to be an effective and less
135 environmentally hazardous preservative for N₂O and nutrient analysis (Frame et al., 2016; Wong
136 et al., 2017).

137



138

139 Figure 1. Sampling stations in the Potomac River Estuary including embayment stations associated with WWTPs (red circles) and without WWTPs (blue circles), and central channel stations (yellow circles). Locations of WWTPs (Noman Cole, Mooney and Aquia) are shown in red stars. Creeks/ivers with sampling stations are numbered in the map with names shown in the legend. Stream sampling sites upstream and downstream of WWTPs in creeks 4 – 7 are shown in Figure 4 below.

145

146 In addition to the routine sampling in the Potomac River Estuary, we also sampled its tributaries, some of which were associated with the WWTPs, on May 18, 2023 (Figure 1) to specifically evaluate the impact of WWTPs on downstream N₂O concentrations. Four creeks/ivers were sampled including Neabsco Creek (5 stations: 2 stations upstream and 3 stations downstream of Mooney WWTP), Occoquan River (3 stations, no WWTP), Pohick Creek (4 stations: 2 stations upstream and 2 stations downstream of Noman Cole WWTP), and Accotink Creek (2 stations, no WWTP). Because Aquia WWTP discharges much less water and N into the Aquia Creek, its impact was not specifically investigated. Since water depths of these creeks/ivers were shallow, the water samples were collected by directly submerging 60 mL serum bottles into the surface water (~0.1 m) and preserving them as described above.

156
157 Besides N₂O sampling, temperature, salinity, and dissolved O₂ concentrations were recorded via a
158 YSI EXO1 sonde. Chlorophyll-a samples (300 mL) were filtered onto GF/F filters and kept on ice
159 in a cooler. The filters were then kept frozen at -20°C in the lab until analysis within 3 months
160 (Arar and Collins, 1997). One additional sample for total nitrogen and phosphorus (both particulate
161 and dissolved) was collected into 250 mL HDPE bottles and kept in ice in a cooler until analysis
162 within 48 hours on land (Rice et al., 2012; EPA, 1983). Total nitrogen is the sum of total Kjeldahl
163 nitrogen and nitrite plus nitrate.

164 Measurement of N₂O and nutrient concentrations

166 N₂O in the serum bottles was stripped by helium carrier gas into a Delta V Plus mass spectrometer
167 (Thermo) for the analyses of N₂O concentration and isotope ratio (m/z = 44, 45, 46) (Tang et al.,
168 2022). The total amount of N₂O in the serum bottles was determined using a standard curve of
169 N₂O peak area with N₂O standards containing a known amount of N₂O reference gas (0, 0.207,
170 0.415, 0.623, 0.831, 1.247 nmol N₂O). The total amount of N₂O dissolved in the water was
171 calculated after subtracting the amount of N₂O in 3 mL air headspace. The amount of N₂O in 3
172 mL air headspace was generally less than 4% of the amount of N₂O dissolved in the 57 mL water
173 samples. The N₂O concentration in samples was then calculated from the total amount of N₂O
174 dissolved in the water divided by the volume of water in the serum bottles. The detection limit and
175 precision of N₂O concentration measurement were 1.29 and 0.33 nM, respectively. We used N₂O
176 produced from nitrate isotope standards (USGS34 = -1.8‰ and IAEA = 4.7‰) to calibrate for
177 δ¹⁵N of N₂O samples. We then estimated N₂O saturation (%): $\frac{N_2O_{measured}}{N_2O_{equilibrium}} \times 100$. The
178 equilibrium N₂O concentration ($N_2O_{equilibrium}$) was calculated based on the solubility of N₂O and
179 atmospheric N₂O concentrations (Weiss and Price, 1980). The monthly atmospheric N₂O
180 concentrations were obtained from the nearby atmospheric station in Brentwood, Maryland
181 (<https://gml.noaa.gov/>) (Andrews et al., 2023).

183 After analyzing N₂O concentration, samples were neutralized to pH ~7 by adding 10%
184 hydrochloric acid. NO₂⁻ + NO₃⁻ (NO_x⁻) concentration in these samples was measured using the
185 vanadium (III) reduction method by converting NO_x⁻ to NO, which was then quantified by
186 chemiluminescence analyzer (Braman and Hendrix, 1989). The detection limit of NO_x⁻

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203 concentration was 0.15 μM . NH_4^+ and NO_2^- concentrations were measured at a few selected
204 stations using the fluorometric orthophthalaldehyde method (Holmes et al., 1999) and the
205 colorimetric method (Hansen and Koroleff, 1999), respectively. Their concentrations were much
206 smaller than NO_3^- alone, mostly accounting for less than 10% of the DIN concentration. Therefore,
207 we only present NO_x^- data in this study.

208

209 **N₂O flux calculation**

210 Surface N₂O flux was calculated using the following equation: $Flux = k \times (N_2O_{measured} -$
211 $N_2O_{equilibrium})$. The gas transfer velocity (k) was estimated based on three different
212 parameterizations: $k = 1.91 \times e^{0.35 \times U} \times (\frac{Sc}{600})^{-0.5}$ (Raymond and Cole, 2001); $k = (0.314 \times$
213 $U^2 - 0.436 \times U + 3.99) \times (\frac{Sc}{600})^{-0.5}$ (Jiang et al., 2008); $k = 0.251 \times U^2 \times (\frac{Sc}{660})^{-0.5}$
214 (Wanninkhof, 2014). U is the wind speed at the 10 m height obtained from the National Centers
215 for Environmental Prediction (NCEP) reanalysis (Kalnay et al., 1996;
216 <https://psl.noaa.gov/data/gridded/data.ncep.reanalysis.html>). Sc is the Schmidt number that could
217 be estimated as a function of temperature (Wanninkhof, 2014). Since our samples have salinity
218 close to 0, we used the parameterization of Sc for freshwater. Average values of the three N₂O flux
219 estimates are presented in the paper and N₂O fluxes estimated by different parameterizations are
220 provided in the associated dataset. We acknowledge large variations in estimating k values in the
221 riverine and estuarine systems by using different empirical models (Raymond and Cole, 2001;
222 Borges et al., 2004; Rosentreter et al., 2021). For instance, the effect of water velocity and water
223 depth on gas transfer velocity was not considered in the parameterizations above. Therefore, we
224 focus on evaluating the spatiotemporal variations in N₂O fluxes and their driving factors instead
225 of their absolute magnitude.

226

227 **Results and discussion**

228 **Spatial and temporal variations of N₂O concentrations in the Potomac River Estuary**

229 Along the roughly 50 km sampling transect in the Potomac River Estuary, NO_x^- concentration
230 decreased from 98 to $<1 \mu\text{M}$ from upstream to downstream (Figure 2a). NO_x^- concentration
231 showed a clear seasonal pattern: higher in winter and spring while lower in summer and fall. The
232 spatial and temporal patterns were likely attributable to the distribution of nutrient sources into the

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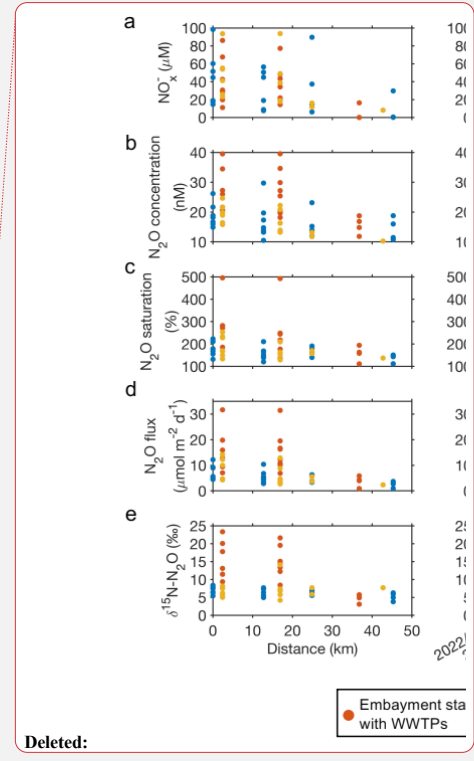
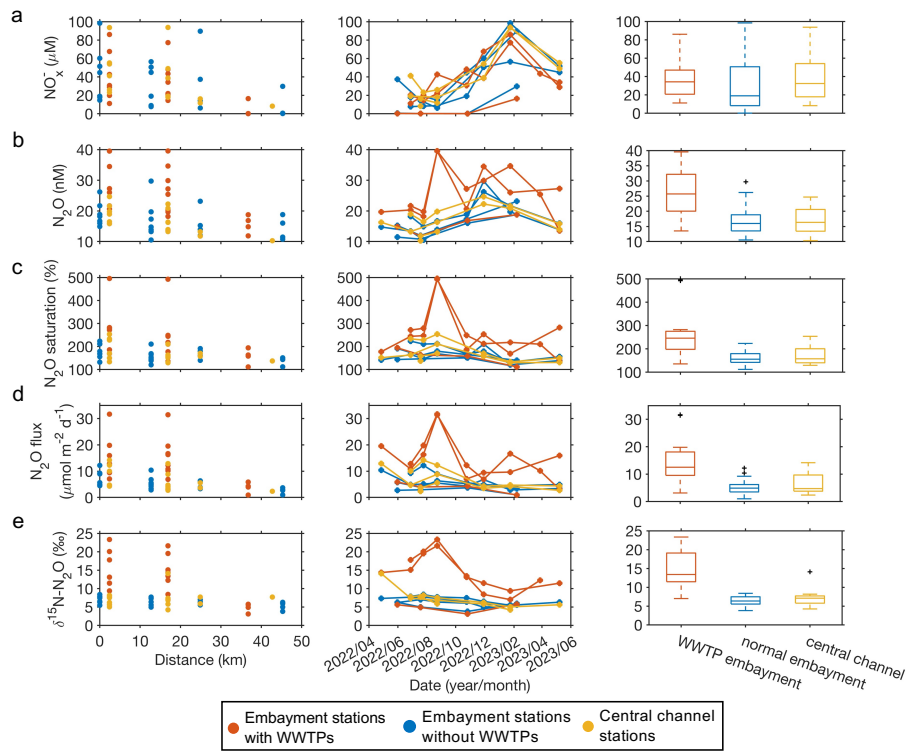
241 Potomac River, DIN uptake and other removal processes along the river (Glibert et al., 1995;
242 Carstensen et al., 2015). For example, the maximum N loading into the Chesapeake Bay occurs in
243 winter and spring (Da et al., 2018). Meanwhile, N₂O concentration decreased from approximately
244 40 to 10 nM along the sampling transect and was higher in the fall and winter (Figure 2b). Since
245 temperature decreased from ~31°C in summer to 4°C in winter (Supplementary Figure 1a), the
246 increase in N₂O solubility in colder water during winter partly explained the seasonal change. In
247 contrast, N₂O saturation had higher values in summer and fall (Figure 2c), suggesting a higher
248 N₂O production in summer and fall. It is worth noting that N₂O saturation was above 100% at all
249 sampling stations with a maximum reaching 500%, indicating the Potomac River Estuary was a
250 consistent and strong source of N₂O to the atmosphere. N₂O fluxes ranged from 1 to 31.7 μmol
251 N₂O m⁻² d⁻¹, generally decreasing from upstream to downstream (Figure 2d). N₂O fluxes showed
252 a similar seasonal pattern to N₂O saturation: higher in summer and fall. N₂O concentrations
253 (median: 18.2 nM) and fluxes (median: 5.6 μmol N₂O m⁻² d⁻¹) in the Potomac River Estuary were
254 substantially higher than in the mainstem of the Chesapeake Bay (2.6 to 20.9 nM N₂O with a
255 median value at 10.6 nM and -0.3 to 4.3 μmol N₂O m⁻² d⁻¹ with a median at 0.5 μmol N₂O m⁻² d⁻¹
256 (Tang et al., 2022; Laperriere et al., 2019)). Therefore, the tributaries (i.e., Potomac River) are
257 more intense sources of N₂O to the atmosphere than mainstem of the bay.

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262 Figure 2. Spatial and temporal variations of NO_x^- concentration (a), N_2O concentration (b), N_2O
 263 saturation (c), N_2O flux (d) and $\delta^{15}\text{N}$ of N_2O (e). The distance shows from upstream to downstream
 264 stations in the Potomac River. Embayment stations associated with WWTPs (red circles and lines)
 265 and without WWTPs (blue circles and lines), and central channel stations (yellow circles and
 266 lines). For the boxplots, the red line in each box is the median. The bottom and top of each box are
 267 the 25th and 75th percentiles of the observations, respectively. The error bars represent 1.5 times
 268 the interquartile range away from the bottom or top of the box, with black + signs showing outliers
 269 beyond that range. Embayment stations associated with WWTPs had significantly higher N_2O
 270 concentration, N_2O saturation, N_2O flux and $\delta^{15}\text{N}$ values compared to other stations ($p < 0.01$, t -
 271 test) but not significantly different NO_x^- concentration.

272

274 Stations close to each other had similar NO_x^- concentrations (e.g., upstream stations > downstream
275 stations), regardless of station category (i.e., with WWTP, without WWTP, central channel of the
276 Potomac River). In contrast, N_2O concentrations and fluxes varied within locations according to
277 the station category: N_2O concentrations and fluxes were substantially higher at stations
278 downstream of WWTPs ($p < 0.01$, *t*-test). N_2O concentrations and fluxes were similar between
279 stations in embayments without WWTPs and the central channel (Figure 2). This suggests these
280 WWTPs are efficient in removing DIN from sewage and other sources but WWTPs may discharge
281 N_2O directly into the effluent or enhance downstream N_2O production (e.g., higher N_2O production
282 yield from the same amount of DIN). This effect extended to our sampling stations ~1.8-4 km
283 downstream of the WWTPs. However, the effect of WWTPs on downstream N_2O varied among
284 stations. For example, elevated N_2O concentrations were observed downstream from Noman Cole
285 and Mooney WWTPs but not downstream from Aquia WWTP. This difference may be related to
286 the different N removal processes of WWTPs that produce N_2O at different yields (de Haas and
287 Andrews, 2022; Zhao et al., 2024). However, we don't have detailed information about the three
288 WWTPs other than that they all implement tertiary treatment. In addition, the different dilution
289 factors by riverine discharges also matter. For example, the volume of effluent from Mooney
290 WWTP was higher than the discharge of Neabsco Creek while the volume of effluent from Aquia
291 WWTP were generally lower than the discharge of Aquia Creek (Supplementary Figure 2a-b).
292 Particularly, the highest N_2O concentration of up to 40 nM was found at two stations downstream
293 of the Noman Cole and Mooney WWTPs on August 23, 2022 when the river discharge was low
294 (Supplementary Figure 2). Thus, the effect of WWTPs on downstream N_2O concentrations also
295 varies seasonally (Schult et al., 2023; Murray et al., 2020), with a relatively more important role
296 in the dry season. Repeated spatial and temporal sampling allowed us to capture these N_2O
297 hotspots. Previous studies have shown the impact of WWTPs on downstream N_2O concentrations
298 and emissions in aquatic environments. For example, the highest N_2O concentration ~675 nM in
299 the Potomac River was measured near the discharge of the Blue Plains WWTP in 1977 (McElroy
300 et al., 1978). Highest N_2O emissions in the Ohio River near Cincinnati were attributed to direct
301 input of N_2O from WWTPs' effluents (Beaulieu et al., 2010).

302
303 In addition, a higher nitrogen isotopic signature ($\delta^{15}\text{N}$) of N_2O associated with WWTPs (median
304 $\delta^{15}\text{N}$ at 13‰) also suggests the distinct sources or cycling processes of N_2O compared to stations

305 of the central channel and without the influence of WWTPs (median $\delta^{15}\text{N}$ of N_2O at 6‰, Figure
306 2e) in the Potomac River Estuary. In comparison, the average $\delta^{15}\text{N}$ of N_2O in the tropospheric air
307 is around 6.55‰ (Snider et al., 2015). $\delta^{15}\text{N}$ of N_2O for stations with the influence of WWTPs
308 showed a clear seasonal variation: higher in summer than winter (Figure 2e). This seasonal
309 difference may be related to the seasonal change in the relative importance of WWTPs' effluents
310 versus riverine discharge (Supplementary Figure 2c). For example, a relatively larger WWTPs'
311 effluent volume compared to the riverine discharge led to a larger $\delta^{15}\text{N}$ of N_2O in summer.
312 However, no clear seasonal pattern of $\delta^{15}\text{N}$ of N_2O was seen for stations without the influence of
313 WWTPs. $\delta^{15}\text{N}$ of N_2O produced in WWTPs depends on the treatment stages and aeration
314 conditions (Toyoda et al., 2011; Tumendelger et al., 2014). For example, the average $\delta^{15}\text{N}$ values
315 were reported to be -24.5‰ and 0‰ respectively for N_2O produced from nitrification during oxic
316 treatment versus N_2O produced from anaerobic denitrification in a California WWTP (Townsend-
317 Small et al., 2011). ~~The $\delta^{15}\text{N}$ values of N_2O in these urban WWTPs were lower than those found~~
318 ~~in waters downstream of WWTPs in the Potomac River (median $\delta^{15}\text{N}$ at 13‰).~~ One of the reasons
319 for the increased $\delta^{15}\text{N}$ of N_2O may be partial N_2O reduction via denitrification in the WWTPs, in
320 downstream creeks, or in sediments; this denitrification effect has been seen in the marine oxygen
321 minimum zones (Kelly et al., 2021). Denitrification as the cause of the elevated $\delta^{15}\text{N}$ is partly
322 supported by the higher $\delta^{15}\text{N}$ of N_2O when NO_x^- was reduced to less than 40 μM , suggesting the
323 occurrence of N_2O reduction when the concentration of other denitrification substrates became
324 low (Supplementary Figure 3). However, we do not know the exact locations where denitrification
325 occurred (e.g., WWTPs, anoxic niches in suspended particles, sediments), which deserves further
326 investigations. The influence of denitrification on unique isotopic signatures of N_2O produced
327 from WWTPs has also been observed in Tama River in Japan (Toyoda et al., 2009).

328

329 Environmental controls on N_2O concentrations

330 N_2O concentrations showed positive correlations with total N ($r=0.62$, $p<0.01$) and NO_x^-
331 concentrations ($r=0.51$, $p<0.01$) (Figure 3a). Correlation analyses done separately for stations with
332 or without WWTPs had similar patterns (Supplementary Figure 4). A better correlation between
333 the N_2O concentration and total N may indicate the contribution of other N sources besides NO_x^-
334 to N_2O production. N_2O could be produced from nitrification in the process of oxidizing NH_4^+ to
335 NO_x^- in the oxic environment as previously shown in the oxygenated mainstem of the Chesapeake

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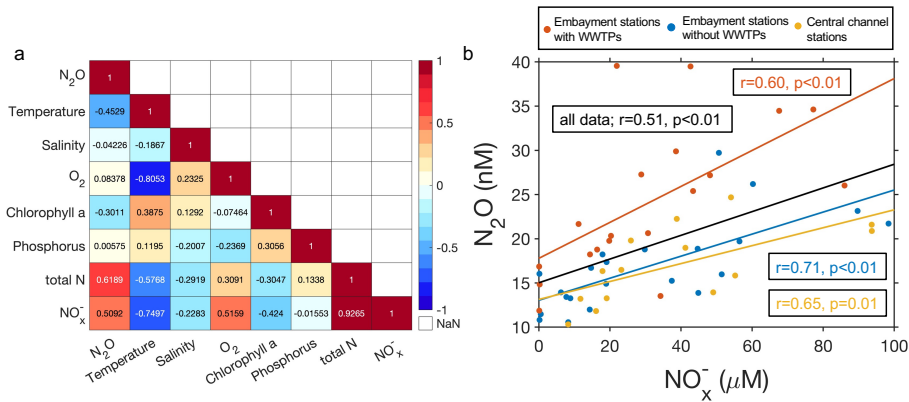
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341 Bay (Tang et al., 2022). However, we can't exclude the possibility of N₂O production from
 342 denitrification associated with anaerobic microsites in particles or in sediment (Beaulieu et al.,
 343 2011; Wan et al., 2023). Future investigations with ¹⁵N tracers should be conducted to differentiate
 344 N₂O production pathways around the WWTPs. Furthermore, N₂O concentration was negatively
 345 correlated with temperature since higher temperature reduced the N₂O solubility. Although
 346 previous studies have showed dissolved oxygen to be an important driver of N₂O concentrations
 347 or fluxes in rivers and estuaries (Rosamond et al., 2012; Wang et al., 2015; Zheng et al., 2022), we
 348 did not find a strong dependence of N₂O on oxygen concentrations in the Potomac River Estuary
 349 (Figure 3a). This lack of strong dependence is probably because of the overall oxygenated
 350 conditions (Supplementary Figure 1c), and opposite correlations found in stations without WWTPs
 351 (positive) versus in stations with WWTPs (negative) (Supplementary Figure 4), which may be
 352 influenced by the different N₂O production pathways.

353

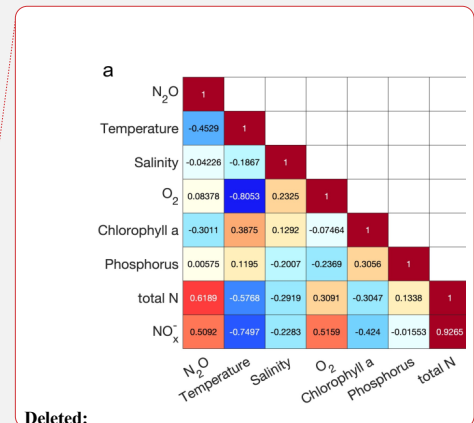


354

355 Figure 3. (a) Correlation coefficients among different environmental factors and N₂O
 356 concentrations. (b) Relationship between N₂O and NO_x⁻ concentrations at different categories of
 357 sampling stations.

358

359 The significant positive relationship between N₂O and NO_x⁻ concentration existed for samples
 360 collected at stations from all three different categories (Figure 3b). N₂O concentrations at stations
 361 downstream of WWTPs were notably higher than at other stations not associated with WWTPs
 362 even under the similar range of NO_x⁻ concentration. The larger slope of N₂O concentration versus



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364 NO_x^- concentration at stations downstream of WWTPs may be related to the direct input of N_2O
365 from WWTPs into the downstream waters or different N_2O production pathways and production
366 yields that deserve further investigations. The DIN concentration has been found to be a good
367 predictor of N_2O concentration and emission in many other rivers and estuaries (Murray et al.,
368 2015; Reading et al., 2020; Zheng et al., 2022;). However, the correlation varied spatially, which
369 may be affected by the variable N_2O emission factors from DIN cycling. The emission factors are
370 affected by temperature, concentration and forms of N, oxygen, organic carbon concentration and
371 many other factors (Hu et al., 2016). The external N_2O input (e.g., input from WWTPs) could also
372 affect the relationship between N_2O and DIN concentrations (Dong et al., 2023). Compared to DIN
373 (~28 to 71 μM) and N_2O concentrations (~16 to 61 nM) measured approximately 45 years ago in
374 the same section of the Potomac River (McElroy et al., 1978), current DIN and N_2O concentrations
375 have slightly decreased. Thus, an additional benefit of nutrient regulation is the reduction of
376 greenhouse gas - N_2O - emissions, beyond improving water quality.

377
378 Since N_2O concentrations had the strongest correlation with total N concentrations (reflecting the
379 N_2O production potential) and temperature (affecting N_2O solubility), we developed a predictive
380 model of N_2O concentration based on total N and temperature. Predictions were performed
381 separately for stations with WWTPs ($\text{N}_2\text{O concentration} = 0.115 \times \text{total N} - 0.241 \times$
382 $\text{temperature} + 17.185$, $n=18$, $r=0.78$; $p<0.01$) and without WWTPs including central channel
383 stations ($\text{N}_2\text{O concentration} = 0.049 \times \text{total N} - 0.298 \times \text{temperature} + 18.888$, $n=23$,
384 $r=0.81$, $p<0.01$). The observed N_2O variability was generally captured by these simple linear
385 models (Supplementary Figure 5) but there were variabilities in the observations remaining to be
386 explained. Addition of other predictors did not significantly improve the model performance, so
387 we chose the simple predictive model that is mechanistically understandable. We then applied the
388 two predictive models separately to estimate N_2O concentrations at the embayment station in the
389 Pohick Bay (with WWTP) and the embayment station in the Occoquan Bay (without WWTP)
390 using total N concentration and temperature that were measured since 2008 by the DEQ of Virginia
391 monitoring program (Supplementary Figures 6 and 7). Predicted N_2O concentrations showed a
392 clear seasonality: higher in winter and lower in summer. N_2O concentrations in the Pohick Bay
393 decreased substantially (-0.9 nM/year) possibly due to the nutrient reduction (total N concentration
394 decreasing at 8.8 $\mu\text{M}/\text{year}$) over the last 14 years (Supplementary Figure 6). However, N_2O

395 concentrations in the Occoquan Bay only decreased slightly (-0.1 nM /year, not statistically
396 significant) along with the minor nutrient reduction (total N concentration decreasing at non-
397 statistically significant rate of 0.5 $\mu\text{M}/\text{year}$) (Supplementary Figure 7). Continuation of
398 environmental monitoring in the Potomac River (e.g., N nutrients and temperature), which is much
399 easier than sampling and measuring N_2O gas, could be used to indirectly estimate the changes in
400 N_2O concentrations in the future. These predictors are likely to be important in other estuaries, but
401 the weighting would vary among locations.

402

403 **Impact of wastewater treatment plants on N_2O concentrations and emissions**

404 To further evaluate how WWTPs affect the N_2O distribution in the Potomac River, we measured
405 N_2O concentrations upstream and downstream of the two WWTP effluents (Mooney and Noman
406 Cole in Neabsco Creek and Pohick Creek, respectively) and compared them to N_2O concentrations
407 measured in two creeks that do not have WWTPs (Figure 4a). Interestingly, the N_2O concentration
408 and flux at the station downstream of Mooney WWTP in Neabsco Creek were lower than the N_2O
409 concentration and flux at the station upstream of Mooney WWTP (15.0 nM vs 20.1 nM; 14.6 μmol
410 $\text{m}^{-2} \text{d}^{-1}$ vs 24.7 $\mu\text{mol m}^{-2} \text{d}^{-1}$). The exact mechanisms were not clear but one of the potential reasons
411 could be the influence by tidal cycles: high tide during the sampling time (salinity was 0.17 instead
412 of 0) may have reversed the water flow and diluted the WWTP effluent with low N_2O
413 concentration Potomac water (12.1 nM at the outflow of Neabsco Creek into the Potomac River
414 Estuary). In contrast, we found substantially a higher N_2O concentration and flux downstream of
415 the Noman Cole WWTP than the upstream station (30.8 nM vs 16.7 nM; 55 $\mu\text{mol m}^{-2} \text{d}^{-1}$ vs 17.6
416 $\mu\text{mol m}^{-2} \text{d}^{-1}$) in the Pohick Creek, which is less affected by the tidal cycle due to its semi-closed
417 geography (salinity was 0.12). The high downstream N_2O concentration and flux may suggest the
418 direct addition of N_2O from WWTP effluent to the downstream environment. Furthermore, $\delta^{15}\text{N}$
419 of N_2O in stations downstream of WWTPs were generally higher than the other two creeks that do
420 not have WWTPs (Figure 4b), confirming the distinct source of N_2O production by WWTPs found
421 in the Potomac River Estuary. Overall, the influence of WWTP effluents on downstream
422 distribution of N_2O is variable, and could be affected by the physical movement of water.

423

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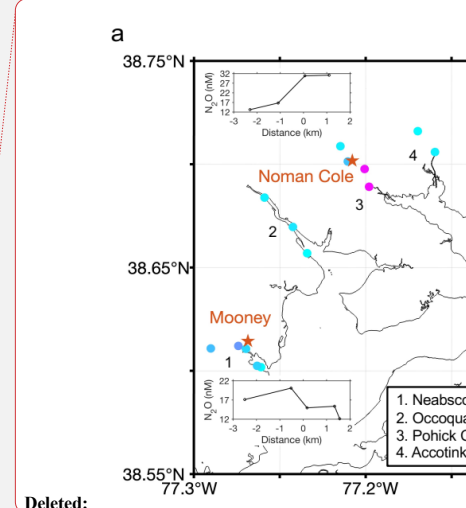
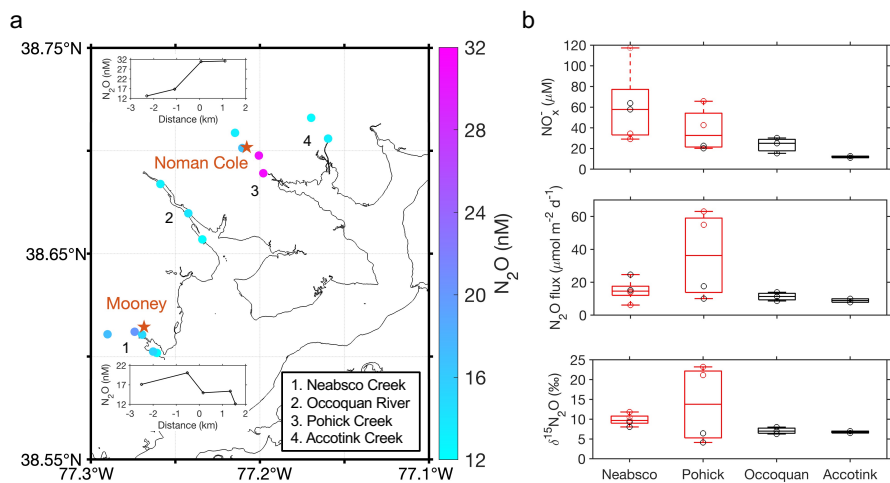
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433
 434 Figure 4. (a) Color-coded N₂O concentration at creek sampling stations on May 18, 2023. WWTPs
 435 (Mooney and Noman Cole) are shown in red stars. The insert figures show the change in N₂O
 436 concentrations as a function of distance up or down stream from the WWTPs. Creeks/ivers with
 437 sampling stations are numbered in the map with names shown in the legend. (b) Box plots of NO₃⁻
 438 , N₂O flux and δ¹⁵N of N₂O comparing four creeks. Neabsco and Pohick Creeks with WWTPs are
 439 displayed with red color boxes. Red and black circles in the boxplots show the data points of
 440 stations downstream and upstream/or without WWTPs, respectively. NO₃⁻, N₂O flux and δ¹⁵N of
 441 N₂O were clearly higher at stations downstream from the WWTP in Pohick Creek.

442
 443 Dong et al. (2023) evaluated the potential impact of wastewater nitrogen discharge on estuarine
 444 N₂O emissions globally. Here we compiled data from previous studies with direct N₂O
 445 measurements in aquatic systems associated with WWTPs (not included in Dong et al., 2023) to
 446 assess the global impact of WWTPs on aquatic N₂O concentrations or emissions (McElroy et al.,
 447 1978; Hemond and Duran, 1989; Toyoda et al., 2009; Beaulieu et al., 2010; Rosamond et al., 2012;
 448 Chun et al., 2020; Masuda et al., 2021; Masuda et al., 2018; Dylla, 2019). WWTP effluents or
 449 water downstream of the WWTPs contain some of the highest N₂O concentrations and fluxes
 450 observed in the aquatic system (Table 1 and Supplementary Figure 8). For example, up to
 451 12,411.4% saturation of N₂O was measured in the effluent of WWTPs in the Tama River in Japan
 452 (Toyoda et al., 2009). In addition, N₂O flux up to 40,800 µmol N₂O-N m⁻² d⁻¹ was found

454 downstream of the Regina WWTP in the Wascana Creek in Canada (Dylla, 2019). The
 455 downstream N₂O flux was >300 times higher than the N₂O flux upstream of the Regina WWTP.
 456 In comparison, the maximum N₂O saturation and flux previously reported in a global riverine N₂O
 457 dataset were around 2,500% and 12,754 $\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$ (Hu et al., 2016). Across the sites
 458 listed in Table 1, N₂O concentration/saturation/flux downstream of the WWTPs was 1.45 to 374-
 459 fold of the upstream waters. The only exception was our observed decrease in N₂O concentrations
 460 downstream of Mooney WWTP on May 18, 2023, which was likely influenced by the tidal cycle.
 461 The wide range of apparent WWTP effect is related to many factors including the variable N₂O
 462 emission factors in the WWTPs, the ratio of WWTP effluent volume to riverine discharge, the
 463 distance from the WWTPs where measurements were conducted, and the direction of water flow
 464 (e.g., tidal cycle). In addition, the estuarine type, mixing regime, and stratification are also
 465 important factors controlling N₂O emissions (Brown et al., 2022). Overall, failing to account for
 466 N₂O emissions downstream of the WWTPs and their variability would substantially bias estimates
 467 of aquatic N₂O emissions. This uncertainty is increased by the fact that only a few observations
 468 are available (all in the northern hemisphere) (Supplementary Figure 8) compared to >58 000
 469 WWTPs present globally (Ehalt Macedo et al., 2022). It is also important to restrict the N₂O
 470 emission via efficient N₂O reduction in the WWTPs considering the projected increase in future
 471 wastewater production (Qadir et al., 2020).

472
 473 Table 1. Global N₂O observations in aquatic systems associated with wastewater treatment plants.
 474 N₂O data are presented in concentration (nM), saturation (%) or flux ($\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$)
 475 according to how they are reported in different studies. ~~*downstream vs upstream.~~

River/location	WWTP	N ₂ O upstream or in tributaries without WWTP §	N ₂ O in WWTP effluent §	N ₂ O downstream or in tributaries with WWTP §	§ Average fold change	Reference
Potomac River/ Washington, D.C., USA	Blue Plains WWTP	11-34 nM		147-318 nM	9.3	McElroy et al., 1978
Assabet River/ Massachusetts, USA	Westborough WWTP	~10 nM	1045 nM	163 nM	16.3	Hemond and Duran, 1989
Tama River/ Tokyo, Japan	Plant 1 Plant 2	350.7% ¶ 219.3%	12411.4% ¶ 3326.2%	3454.8% ¶ 1029.6%	9.8 4.7	Toyoda et al., 2009
Ohio River/ Cincinnati, USA		27.9 $\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$		1068 $\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$	38.2	Beaulieu et al., 2010

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Grand River/ Ontario, Canada	e.g., Kitchener WWTP	4-12 $\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$		9-113 $\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$	9.4	Rosamond et al., 2012
Wascana Creek/ Saskatchewan, Canada	Regina WWTP	-32.5 to 109 $\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$	227 to 72800 $\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$	398 to 40800 $\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$	374	Dylla. 2019
Han River/ Seoul, Korea	JNW	39.7 nM	602.1 nM	441.6 nM	11.1	Chun et al., 2020
A-river	A-WWTP	61 nM	493 nM	180 nM	3	Masuda et al., 2021
B-river	B-WWTP	95 nM	246 nM	286 nM	3	Masuda et al., 2018
C-river/Miyagi, Japan	C-WWTP	100 nM	319 nM	145 nM	1.45	
Potomac River Estuary /Virginia, USA	Noman Cole Mooney Aquia	10.8-29.7 nM <u>1-12.2</u> <u>$\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$</u>		11.87-39.5 nM <u>0.95-31.7</u> <u>$\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$</u>	1.6 <u>2.2</u>	This study
Neabsco Creek/ Virginia, USA	Mooney	20.1 nM <u>24.7</u> <u>$\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$</u>		15.0 nM <u>14.6</u> <u>$\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$</u>	0.75 <u>0.59</u>	This study
Pohick Creek/ Virginia, USA	Noman Cole	16.7 nM <u>17.6</u> <u>$\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$</u>		30.8 nM <u>55</u> <u>$\mu\text{mol N}_2\text{O-N m}^{-2} \text{ d}^{-1}$</u>	1.84 <u>3.12</u>	This study

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481

482 Conclusion

483 Taking advantage of the routine water monitoring program by the DEQ of Virginia, we detected
484 strong spatial and temporal variabilities of N_2O concentrations and emissions in the Potomac River
485 Estuary, a major tributary of Chesapeake Bay. Observations across the Potomac River Estuary also
486 allowed us to identify hotspots of N_2O emissions associated with WWTPs effluents. Higher N_2O
487 concentrations downstream of WWTPs compared to regions with similar nitrogen nutrient
488 concentrations suggested the direct discharge of dissolved N_2O from WWTPs and/or intense N_2O
489 production. The influence of WWTPs on downstream N_2O concentrations and emissions is largely
490 affected by volumes of river discharges versus WWTPs effluents. A survey of globally available
491 data shows N_2O concentrations or emissions are consistently elevated in waters downstream from
492 WWTPs. Future ^{15}N tracer incubations would help to explain the high N_2O concentration
493 downstream of WWTPs by disentangling the N_2O production pathways. In addition, concurrent
494 measurements of the N flux and N_2O concentration downstream of WWTPs will help to constrain
495 overall N_2O emission factors associated with WWTPs. Our work could encourage potential
496 collaborations between scientific community and governmental agencies/the public to better
497 observe the environmental pollution or quality, e.g., increasing the frequency and resolution of
498 observations for N_2O and other greenhouse gases along with many regularly monitored

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499 environmental factors like temperature and nutrients. Such efforts may identify previously
500 overlooked sources of N₂O emission and help to better estimate N₂O emissions from aquatic
501 systems.

502

503 **Data availability**

504 Data presented in this study has been deposited in Zenodo repository:
505 <https://doi.org/10.5281/zenodo.10775250>.

506

507 **Author contribution**

508 W.T. conceived the study. J.T., T.J., and W.T. collected N₂O samples from the Potomac River
509 Estuary. W.T. analyzed samples and interpreted data with other coauthors. W.T. wrote the first
510 draft of the manuscript with input from B.B.W. All coauthors contributed to the result discussion
511 and manuscript writing.

512

513 **Competing interests**

514 The authors declare that they have no conflict of interest.

515

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520 sampling and for providing the opportunity to collect N₂O samples in the Potomac River Estuary.
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522 quality data of wastewater treatment plants. This study is supported by Princeton University.

523

524 **References**

- 525 Andrews, A., Crotwell, A., Crotwell, M., Handley, P., Higgs, J., Kofler, J., Lan, X., Legard, T.,
526 Madronich, M., McKain, K., Miller, J., Moglia, E., Mund, J., Neff, D., Newberger, T., Petron, G.,
527 Turnbull, J., Vimont, I., Wolter, S., and NOAA Global Monitoring Laboratory.: NOAA Global
528 Greenhouse Gas Reference Network Flask-Air PFP Sample Measurements of N₂O at Tall Tower
529 and other Continental Sites, 2005-Present [Data set]. NOAA GML. 10.15138/C11N-KD82
530 Version: 2023-08-23.
- 531 Arar, E. J. and Collins, G. B.: Method 445.0: In vitro determination of chlorophyll a and
532 pheophytin a in marine and freshwater algae by fluorescence, United States Environmental
533 Protection Agency, Office of Research and Development, 1997.
- 534 Beaulieu, J. J., Shuster, W. D., and Rebolz, J. A.: Nitrous Oxide Emissions from a Large,
535 Impounded River: The Ohio River, *Environmental Science & Technology*, 44, 7527-7533,
536 10.1021/es1016735, 2010.
- 537 Beaulieu, J. J., Tank, J. L., Hamilton, S. K., Wollheim, W. M., Hall, R. O., Jr., Mulholland, P. J.,
538 Peterson, B. J., Ashkenas, L. R., Cooper, L. W., Dahm, C. N., Dodds, W. K., Grimm, N. B.,
539 Johnson, S. L., McDowell, W. H., Poole, G. C., Valett, H. M., Arango, C. P., Bernot, M. J., Burgin,
540 A. J., Crenshaw, C. L., Helton, A. M., Johnson, L. T., O'Brien, J. M., Potter, J. D., Sheibley, R.
541 W., Sobota, D. J., and Thomas, S. M.: Nitrous oxide emission from denitrification in stream and
542 river networks, *Proceedings of the National Academy of Sciences of the United States of America*,
543 108, 214-219, 10.1073/pnas.1011464108, 2011.
- 544 Borges, A. V., Vanderborght, J.-P., Schiettecatte, L.-S., Gazeau, F., Ferrón-Smith, S., Delille, B.,
545 and Frankignoulle, M.: Variability of the gas transfer velocity of CO₂ in a macrotidal estuary (the
546 Scheldt), *Estuaries*, 27, 593-603, 10.1007/BF02907647, 2004.
- 547 Braman, R. S. and Hendrix, S. A.: Nanogram nitrite and nitrate determination in environmental
548 and biological materials by vanadium (III) reduction with chemiluminescence detection,
549 *Analytical Chemistry*, 61, 2715-2718, 1989.
- 550 Bricker, S. B., Rice, K. C., and Bricker, O. P.: From Headwaters to Coast: Influence of Human
551 Activities on Water Quality of the Potomac River Estuary, *Aquatic Geochemistry*, 20, 291-323,
552 10.1007/s10498-014-9226-y, 2014.

553 Brown, A. M., Bass, A. M., and Pickard, A. E.: Anthropogenic-estuarine interactions cause
554 disproportionate greenhouse gas production: A review of the evidence base, *Marine Pollution*
555 *Bulletin*, 174, 113240, 10.1016/j.marpolbul.2021.113240, 2022.

556 Carstensen, J., Klais, R., and Cloern, J. E.: Phytoplankton blooms in estuarine and coastal waters:
557 Seasonal patterns and key species, *Estuarine, Coastal and Shelf Science*, 162, 98-109,
558 10.1016/j.ecss.2015.05.005, 2015.

559 Chun, Y., Kim, D., Hattori, S., Toyoda, S., Yoshida, N., Huh, J., Lim, J. H., and Park, J. H.:
560 Temperature control on wastewater and downstream nitrous oxide emissions in an urbanized river
561 system, *Water Res*, 187, 116417, 10.1016/j.watres.2020.116417, 2020.

562 Da, F., Friedrichs, M. A. M., and St-Laurent, P.: Impacts of Atmospheric Nitrogen Deposition and
563 Coastal Nitrogen Fluxes on Oxygen Concentrations in Chesapeake Bay, *Journal of Geophysical*
564 *Research: Oceans*, 123, 5004-5025, 10.1029/2018jc014009, 2018.

565 de Haas, D. and Andrews, J.: Nitrous oxide emissions from wastewater treatment - Revisiting the
566 IPCC 2019 refinement guidelines, *Environmental Challenges*, 8, 10.1016/j.envc.2022.100557,
567 2022.

568 Dong, Y., Liu, J., Cheng, X., Fan, F., Lin, W., Zhou, C., Wang, S., Xiao, S., Wang, C., Li, Y., and
569 Li, C.: Wastewater-influenced estuaries are characterized by disproportionately high nitrous oxide
570 emissions but overestimated IPCC emission factor, *Communications Earth & Environment*, 4,
571 10.1038/s43247-023-01051-6, 2023.

572 Dylla, N. P.: Downstream effects on denitrification and nitrous oxide from an advanced
573 wastewater treatment plant upgrade, University of Saskatchewan, 2019.

574 Eggleston, H., Buendia, L., Miwa, K., Ngara, T., and Tanabe, K.: 2006 IPCC guidelines for
575 national greenhouse gas inventories, 2006.

576 Ehalt Macedo, H., Lehner, B., Nicell, J., Grill, G., Li, J., Limtong, A., and Shakya, R.: Distribution
577 and characteristics of wastewater treatment plants within the global river network, *Earth System*
578 *Science Data*, 14, 559-577, 10.5194/essd-14-559-2022, 2022.

579 EPA, U.: Method 365.4: Phosphorous, total (Colorimetric, automated, block digester AA II),
580 1983.

581 EPA, U.: Inventory of US Greenhouse gas emissions and sinks: 1990-2021, United States
582 Environmental Protection Agency, 2023.

583 Frame, C. H., Lau, E., Nolan, E. J. t., Goepfert, T. J., and Lehmann, M. F.: Acidification Enhances
584 Hybrid N₂O Production Associated with Aquatic Ammonia-Oxidizing Microorganisms, *Front*
585 *Microbiol*, 7, 2104, 10.3389/fmicb.2016.02104, 2016.

586 Galloway, J. N., Townsend, A. R., Erisman, J. W., Bekunda, M., Cai, Z., Freney, J. R., Martinelli,
587 L. A., Seitzinger, S. P., and Sutton, M. A.: Transformation of the nitrogen cycle: recent trends,
588 questions, and potential solutions, *Science*, 320, 889-892, 10.1126/science.1136674, 2008.

589 Gonçalves, C., Brogueira, M. J., and Nogueira, M.: Tidal and spatial variability of nitrous oxide
590 (N₂O) in Sado estuary (Portugal), *Estuarine, Coastal and Shelf Science*, 167, 466-474,
591 10.1016/j.ecss.2015.10.028, 2015.

592 Glibert, P. M., Conley, D. J., Fisher, T. R., Harding, L. W., and Malone, T. C.: Dynamics of the
593 1990 winter/spring bloom in Chesapeake Bay, *Marine Ecology Progress Series*, 122, 27-43, 1995.

594 Hansen, H. P. and Koroleff, F.: Determination of nutrients, in: *Methods of Seawater Analysis*,
595 159-228, 10.1002/9783527613984.ch10, 1999.

596 Hemond, H. F. and Duran, A. P.: Fluxes of N₂O at the sediment-water and water-atmosphere
597 boundaries of a nitrogen-rich river, *Water Resources Research*, 25, 839-846,
598 10.1029/WR025i005p00839, 1989.

599 Holmes, R. M., Aminot, A., Kérouel, R., Hooker, B. A., and Peterson, B. J.: A simple and precise
600 method for measuring ammonium in marine and freshwater ecosystems, *Canadian Journal of*
601 *Fisheries and Aquatic Sciences*, 56, 1801-1808, 10.1139/f99-128, 1999.

602 Hu, M., Chen, D., and Dahlgren, R. A.: Modeling nitrous oxide emission from rivers: a global
603 assessment, *Global Change Biology*, 22, 3566-3582, 10.1111/gcb.13351, 2016.

604 Jaworski, N. A., Romano, B., Buchanan, C., and Jaworski, C.: *The Potomac River Basin and its*
605 *Estuary: landscape loadings and water quality trends, 1895–2005, Report, Interstate Commission*
606 *on the Potomac River Basin, Rockville, Maryland, USA, 2007.*

607 Jiang, L. Q., Cai, W. J., and Wang, Y.: A comparative study of carbon dioxide degassing in river-
608 and marine-dominated estuaries, *Limnology and Oceanography*, 53, 2603-2615,
609 10.4319/lo.2008.53.6.2603, 2008.

610 Kalnay, E., Kanamitsu, M., Kistler, R., Collins, W., Deaven, D., Gandin, L., Iredell, M., Saha, S.,
611 White, G., and Woollen, J.: The NCEP/NCAR 40-year reanalysis project, *Bulletin of the American*
612 *meteorological Society*, 77, 437-471, 1996.

613 Kampschreur, M. J., Temmink, H., Kleerebezem, R., Jetten, M. S., and van Loosdrecht, M. C.:
614 Nitrous oxide emission during wastewater treatment, *Water Res*, 43, 4093-4103,
615 10.1016/j.watres.2009.03.001, 2009.

616 Kelly, C. L., Travis, N. M., Baya, P. A., and Casciotti, K. L.: Quantifying Nitrous Oxide Cycling
617 Regimes in the Eastern Tropical North Pacific Ocean With Isotopomer Analysis, *Global*
618 *Biogeochemical Cycles*, 35, 10.1029/2020gb006637, 2021.

619 Laperriere, S. M., Nidzicko, N. J., Fox, R. J., Fisher, A. W., and Santoro, A. E.: Observations of
620 Variable Ammonia Oxidation and Nitrous Oxide Flux in a Eutrophic Estuary, *Estuaries and*
621 *Coasts*, 42, 33-44, 10.1007/s12237-018-0441-4, 2019.

622 Maavara, T., Lauerwald, R., Laruelle, G. G., Akbarzadeh, Z., Bouskill, N. J., Van Cappellen, P.,
623 and Regnier, P.: Nitrous oxide emissions from inland waters: Are IPCC estimates too high?, *Global*
624 *Change Biology*, 25, 473-488, 10.1111/gcb.14504, 2019.

625 Masuda, S., Otomo, S., Maruo, C., and Nishimura, O.: Contribution of dissolved N₂O in total N₂O
626 emission from sewage treatment plant, *Chemosphere*, 212, 821-827,
627 10.1016/j.chemosphere.2018.08.089, 2018.

628 Masuda, S., Sato, T., Mishima, I., Maruo, C., Yamazaki, H., and Nishimura, O.: Impact of nitrogen
629 compound variability of sewage treated water on N₂O production in riverbeds, *J Environ Manage*,
630 290, 112621, 10.1016/j.jenvman.2021.112621, 2021.

631 McElroy, M. B., Elkins, J. W., Wofsy, S. C., Kolb, C. E., Durán, A. P., and Kaplan, W. A.:
632 Production and release of N₂O from the Potomac Estuary 1, *Limnology and Oceanography*, 23,
633 1168-1182, 10.4319/lo.1978.23.6.1168, 1978.

634 Morée, A. L., Beusen, A. H. W., Bouwman, A. F., and Willems, W. J.: Exploring global nitrogen
635 and phosphorus flows in urban wastes during the twentieth century, *Global Biogeochemical*
636 *Cycles*, 27, 836-846, 10.1002/gbc.20072, 2013.

637 Murray, R. H., Erler, D. V., and Eyre, B. D.: Nitrous oxide fluxes in estuarine environments:
638 response to global change, *Global Change Biology*, 21, 3219-3245, 10.1111/gcb.12923, 2015.

639 Murray, R., Erler, D. V., Rosentreter, J., Wells, N. S., and Eyre, B. D.: Seasonal and spatial controls
640 on N₂O concentrations and emissions in low-nitrogen estuaries: Evidence from three tropical
641 systems, *Marine Chemistry*, 221, 103779, 10.1016/j.marchem.2020.103779, 2020.

642 Pennino, M. J., Kaushal, S. S., Murthy, S. N., Blomquist, J. D., Cornwell, J. C., and Harris, L. A.:
643 Sources and transformations of anthropogenic nitrogen along an urban river–estuarine continuum,
644 *Biogeosciences*, 13, 6211-6228, 10.5194/bg-13-6211-2016, 2016.

645 Qadir, M., Drechsel, P., Jiménez Cisneros, B., Kim, Y., Pramanik, A., Mehta, P., and Olaniyan,
646 O.: Global and regional potential of wastewater as a water, nutrient and energy source, *Natural*
647 *Resources Forum*, 44, 40-51, 10.1111/1477-8947.12187, 2020.

648 Quick, A. M., Reeder, W. J., Farrell, T. B., Tonina, D., Feris, K. P., and Benner, S. G.: Nitrous
649 oxide from streams and rivers: A review of primary biogeochemical pathways and environmental
650 variables, *Earth-Science Reviews*, 191, 224-262, 10.1016/j.earscirev.2019.02.021, 2019.

651 Raymond, P. A. and Cole, J. J.: Gas exchange in rivers and estuaries: Choosing a gas transfer
652 velocity, *Estuaries*, 24, 312-317, 10.2307/1352954, 2001.

653 Reading, M. J., Tait, D. R., Maher, D. T., Jeffrey, L. C., Looman, A., Holloway, C., Shishaye, H.
654 A., Barron, S., and Santos, I. R.: Land use drives nitrous oxide dynamics in estuaries on regional
655 and global scales, *Limnology and Oceanography*, 10.1002/lno.11426, 2020.

656 Rice, E. W., Bridgewater, L., and Association, A. P. H.: Standard methods for the examination of
657 water and wastewater, American public health association Washington, DC2012.

658 Rosamond, M. S., Thuss, S. J., and Schiff, S. L.: Dependence of riverine nitrous oxide emissions
659 on dissolved oxygen levels, *Nature Geoscience*, 5, 715-718, 10.1038/ngeo1556, 2012.

660 Rosentreter, J. A., Wells, N. S., Ulseth, A. J., and Eyre, B. D.: Divergent Gas Transfer Velocities
661 of CO₂, CH₄, and N₂O Over Spatial and Temporal Gradients in a Subtropical Estuary, *Journal of*
662 *Geophysical Research: Biogeosciences*, 126, 10.1029/2021jg006270, 2021.

663 Rosentreter, J. A., Laruelle, G. G., Bange, H. W., Bianchi, T. S., Busecke, J. J., Cai, W. J., Eyre,
664 B. D., Forbrich, I., Kwon, E. Y., Maavara, T. and Moosdorf, N.: Coastal vegetation and estuaries
665 are collectively a greenhouse gas sink, *Nature Climate Change*, 13, 579-587, 10.1038/s41558-023-
666 01682-9, 2023.

667 Schulz, G., Sanders, T., Voynova, Y. G., Bange, H. W., and Dähnke, K.: Seasonal variability of
668 nitrous oxide concentrations and emissions in a temperate estuary, *Biogeosciences*, 20, 3229-
669 3247, 10.5194/bg-20-3229-2023, 2023.

670 Snider, D. M., Venkiteswaran, J. J., Schiff, S. L., and Spoelstra, J.: From the ground up: global
671 nitrous oxide sources are constrained by stable isotope values, *PloS one*, 10, e0118954,
672 10.1371/journal.pone.0118954, 2015.

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Moved up [1]: Rosentreter, J. A., Wells, N. S., Ulseth, A. J., and Eyre, B. D.: Divergent Gas Transfer Velocities of CO₂, CH₄, and N₂O Over Spatial and Temporal Gradients in a Subtropical Estuary, *Journal of Geophysical Research: Biogeosciences*, 126, 10.1029/2021jg006270, 2021. ¶

679 Tang, W., Tracey, J. C., Carroll, J., Wallace, E., Lee, J. A., Nathan, L., Sun, X., Jayakumar, A.,
680 and Ward, B. B.: Nitrous oxide production in the Chesapeake Bay, *Limnology and Oceanography*,
681 10.1002/lno.12191, 2022.

682 Tian, H., Xu, R., Canadell, J. G., Thompson, R. L., Winiwarter, W., Suntharalingam, P., Davidson,
683 E. A., Ciais, P., Jackson, R. B., Janssens-Maenhout, G., Prather, M. J., Regnier, P., Pan, N., Pan,
684 S., Peters, G. P., Shi, H., Tubiello, F. N., Zaehle, S., Zhou, F., Arneth, A., Battaglia, G., Berthet,
685 S., Bopp, L., Bouwman, A. F., Buitenhuis, E. T., Chang, J., Chipperfield, M. P., Dangal, S. R. S.,
686 Dlugokencky, E., Elkins, J. W., Eyre, B. D., Fu, B., Hall, B., Ito, A., Joos, F., Krummel, P. B.,
687 Landolfi, A., Laruelle, G. G., Lauerwald, R., Li, W., Lienert, S., Maavara, T., MacLeod, M., Millet,
688 D. B., Olin, S., Patra, P. K., Prinn, R. G., Raymond, P. A., Ruiz, D. J., van der Werf, G. R.,
689 Vuichard, N., Wang, J., Weiss, R. F., Wells, K. C., Wilson, C., Yang, J., and Yao, Y.: A
690 comprehensive quantification of global nitrous oxide sources and sinks, *Nature*, 586, 248-256,
691 10.1038/s41586-020-2780-0, 2020.

692 Townsend-Small, A., Pataki, D. E., Tseng, L. Y., Tsai, C. Y., and Rosso, D.: Nitrous oxide
693 emissions from wastewater treatment and water reclamation plants in southern California, *J*
694 *Environ Qual*, 40, 1542-1550, 10.2134/jeq2011.0059, 2011.

695 Toyoda, S., Iwai, H., Koba, K., and Yoshida, N.: Isotopomeric analysis of N₂O dissolved in a river
696 in the Tokyo metropolitan area, *Rapid Communications in Mass Spectrometry*, 23, 809-821,
697 10.1002/rcm.3945, 2009.

698 Toyoda, S., Suzuki, Y., Hattori, S., Yamada, K., Fujii, A., Yoshida, N., Kouno, R., Murayama, K.,
699 and Shiomi, H.: Isotopomer Analysis of Production and Consumption Mechanisms of N₂O and
700 CH₄ in an Advanced Wastewater Treatment System, *Environmental Science & Technology*, 45,
701 917-922, 10.1021/es102985u, 2011.

702 Tumendelger, A., Toyoda, S., and Yoshida, N.: Isotopic analysis of N₂O produced in a
703 conventional wastewater treatment system operated under different aeration conditions, *Rapid*
704 *Commun Mass Spectrom*, 28, 1883-1892, 10.1002/rcm.6973, 2014.

705 Wan, X. S., Sheng, H. X., Liu, L., Shen, H., Tang, W., Zou, W., Xu, M. N., Zheng, Z., Tan, E.,
706 Chen, M., Zhang, Y., Ward, B. B., and Kao, S. J.: Particle-associated denitrification is the primary
707 source of N₂O in oxic coastal waters, *Nat Commun*, 14, 8280, 10.1038/s41467-023-43997-3, 2023.

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710 Wang, J., Chen, N., Yan, W., Wang, B., and Yang, L.: Effect of dissolved oxygen and nitrogen on
711 emission of N₂O from rivers in China, *Atmospheric Environment*, 103, 347-356,
712 10.1016/j.atmosenv.2014.12.054, 2015.

713 Wang, J., Vilmin, L., Mogollon, J. M., Beusen, A. H. W., van Hoek, W. J., Liu, X., Pika, P. A.,
714 Middelburg, J. J., and Bouwman, A. F.: Inland Waters Increasingly Produce and Emit Nitrous
715 Oxide, *Environmental science & technology*, 57, 13506-13519, 10.1021/acs.est.3c04230, 2023.

716 Wanninkhof, R.: Relationship between wind speed and gas exchange over the ocean revisited,
717 *Limnol. Oceanogr. Methods*, 12, 351-362, 2014.

718 Weiss, R. F. and Price, B. A.: Nitrous oxide solubility in water and seawater, *Marine Chemistry*,
719 8, 347-359, 10.1016/0304-4203(80)90024-9, 1980.

720 Wong, G. T. F., Li-Tzu Hou, L., and Li, K. Y.: Preservation of seawater samples for soluble
721 reactive phosphate, nitrite, and nitrate plus nitrite analyses by the addition of sodium hydroxide,
722 *Limnology and Oceanography: Methods*, 15, 320-327, 10.1002/lom3.10160, 2017.

723 Yao, Y., Tian, H., Shi, H., Pan, S., Xu, R., Pan, N., and Canadell, J. G.: Increased global nitrous
724 oxide emissions from streams and rivers in the Anthropocene, *Nature Climate Change*,
725 10.1038/s41558-019-0665-8, 2019.

726 Zhao, Y. W., Du, L. L., Hu, B., Lin, H. Y., Liang, B., Song, Y. P., Wang, Y. Q., Wang, H. W., Li,
727 P. F., Wang, A. J. and Wang, H. C.: Impact of influent characteristics and operational parameters
728 on nitrous oxide emissions in wastewater treatment: Strategies for mitigation and microbial
729 insights, *Current Research in Biotechnology*, 7, 100207, 10.1016/j.crbiot.2024.100207, 2024.

730 Zheng, Y., Wu, S., Xiao, S., Yu, K., Fang, X., Xia, L., Wang, J., Liu, S., Freeman, C., and Zou, J.:
731 Global methane and nitrous oxide emissions from inland waters and estuaries, *Glob Chang Biol*,
732 28, 4713-4725, 10.1111/gcb.16233, 2022.

733