

1 *Supplement of*

2 **Organic Carbon, Mercury, and Sediment Characteristics along a**
3 **land – shore transect in Arctic Alaska**

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7

8 S1 Supplementary methods

9 S1.1 Grain size analysis

10 The first step of the grain size analysis was the preparation of the samples, by removing the contained organic matter.
11 Therefore, a solution of 100 ml 3 % hydrogen peroxide [H₂O₂] and 4 ml 25 % ammonia [NH₃] were added to the freeze-dried
12 samples. They were then placed on a shaker for approximately four weeks to allow for thorough mixing and reaction of the
13 solution with the samples. During the period of four weeks, 10 ml of 30 % H₂O₂ were added to the samples daily during
14 weekdays, while monitoring the pH to ensure it remained between 6 and 8. If the pH fell outside of this range, it was adjusted
15 accordingly with either ammonia or concentrated acetic acid.

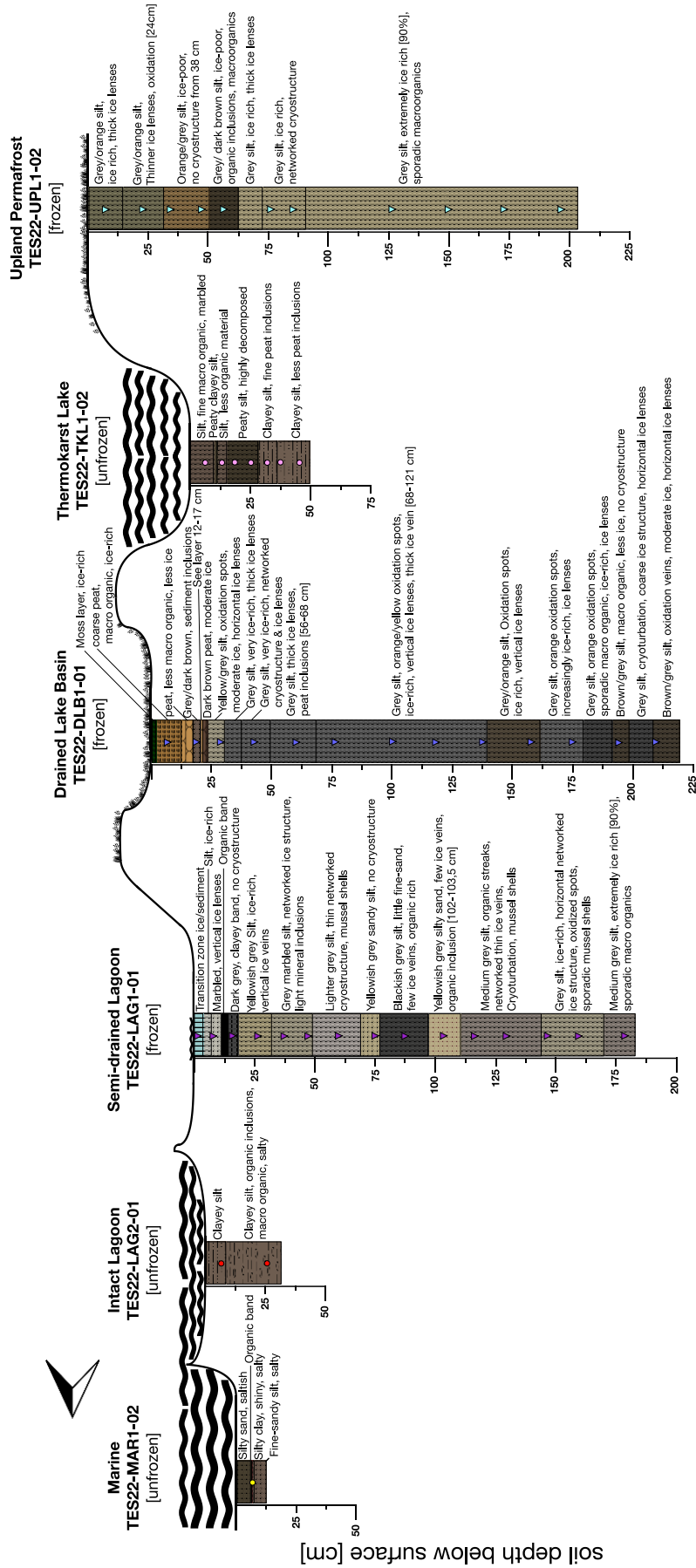
16 The next step was to rinse the samples with purified water, to remove the H₂O₂, followed by centrifugation. The supernatant
17 liquid was then decanted, and the remaining sediment was freeze-dried and manually homogenized. For each sample, 1 g of
18 the homogenized material was placed into a plastic container, to which 0.5 g of tetra-sodium pyrophosphate [Na₄P₂O₇] and
19 0.0001 % ammonia solution [NH₃] were added to disperse the soil particles and prevent them from settling during the
20 subsequent analysis. The sample container was then placed on a "Gerhard Laboshake overhead shaker" for 24 hours to allow
21 for complete dispersion of the added solution. Each sample was then split into eight subsamples with a particle concentration
22 of 5-15 % by a rotary cone sample divider. Simultaneously all particles > 1 mm were sieved out before analysis, weighed, and
23 included in the results at the end. The grain size measurement was carried out using a *Malvern Mastersizer 3000* with a *Malvern*
24 *Hydro LV wet-sample dispersion Unit*.

25 The international ISO 14688-1:2017 scale was used for the grain size classification. The ranges specified there are ≤ 2 μm for
26 clay, 2 μm to 63 μm for silt and 63 μm to 2 mm for sand (ISO 14688-1:2017, 2017). All statistics of the grain size distribution
27 were calculated using the software GRADISTAT (Blott and Pye, 2001).

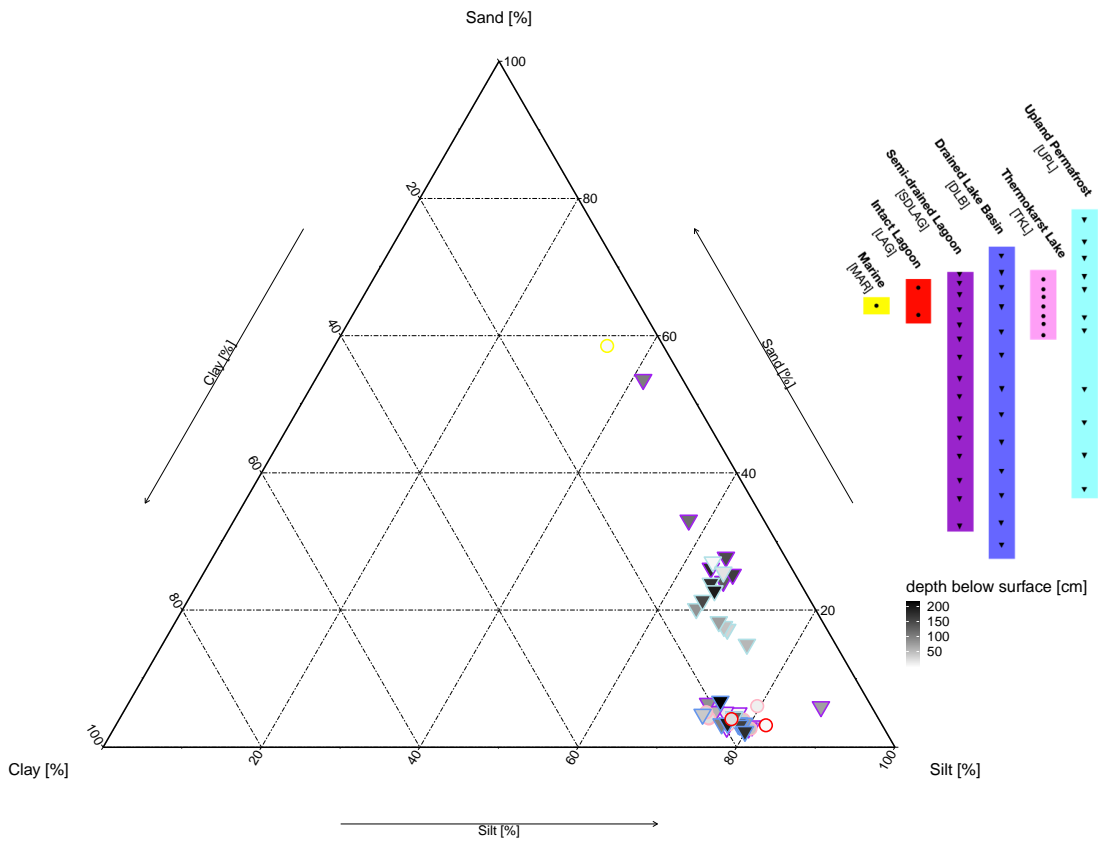
28 S1.2 Hydrochemical analysis

29 The selected samples were thawed at room temperature overnight, to prepare them for the porewater extraction. The porewater
30 was taken the next day, using Rhizons (RHIZONS MOM 5 and 10 cm, Rhizospheres Research Products). Therefore, the
31 Rhizons were inserted into the soil samples and a vacuum was created using plastic syringes. The extracted water was then
32 distributed into different vials for the different hydrochemical measurements (pH value, electrical conductivity (EC), anions).
33 The pH value and EC were measured one day after the pore water extraction with a *WTW Multilab 540* using 4 ml of porewater.
34 The concentration of anions in the porewater can provide different information, like the availability and mobility of nutrients,
35 as well as a potential risk of anion accumulation leading to soil degradation. In this study, the chloride and sulfate concen-
36 trations are of particular interest, as they provide information on the degree of acidification and salinization and can thus
37 indicate the extent to which the sample sites are influenced by the ocean.

38 For the measurement of anions 5 ml of the pore water of every sample was filled into an 8 ml LDPE-Bottle. Samples with a
39 high electrical conductivity were diluted in ratio of 1:25, 1:50 or 1:100, where applied the higher the electrical conductivity
40 the higher the dilution ratio. The Anions were measured using a *Thermo ICS2100*. For the analysis of the DOC content in the
41 porewater 10-20 ml of each sample was filled into a glass vial. To preserve the samples until further analysis they were
42 acidified with 50 μl of 30 % hydrochloric acid (HCL) and stored at 4 °C. The measurement was carried out using a *Shimadzu*
43 *Total Organic Carbon Analyzer (TOC-VCPH)*. The results of three to five injections were used as an average to determine the
44 total DOC content. The detection limit was 0.25 mg/L and the uncertainty ranged between ±10 % for measured values higher
45 than 1.5 mg/L and ±15-20 % for values lower than 1.5 mg/L.



47 **Figure S1: Stratigraphic core description of the soil profiles. Samples marked as points (unfrozen sediments) and triangles (frozen**
 48 **sediments). Core abbreviations: UPL: upland permafrost, TKL: thermokarst lake, DLB: drained lake basin, SLAG: semi-drained**
 49 **lagoon, LAG: lagoon, MAR: marine.**



50 **Figure S2: Soil triangle showing the grain size distribution of the soil profiles, with circles for unfrozen sediments and triangles for**
 51 **frozen sediments and a colour gradient over depth below surface [cm]. Core abbreviations: UPL: upland permafrost, TKL:**
 52 **thermokarst lake, DLB: drained lake basin, SDLAG: semi-drained lagoon, LAG: lagoon, MAR: marine.**

53 **S3 Supplementary tables**

54 **Table S1: Results of the Hydrochemical analysis. Core abbreviations: UPL: upland permafrost; TKL: thermokarst lake; DLB:**
 55 **drained lake basin; SDLAG: semi-drained lagoon; LAG: lagoon; MAR: marine.**
 56

Hydrochemical Analysis	Mean depth [cm]	pH value	Electrical conductivity [mS cm ⁻¹]	DOC as NPOC [mg l ⁻¹]
MAR	6.25	7.67	39.5	19.2
LAG	5	7.64	54.6	NA
SDLAG	7.75	7.49	39.0	80.8
SDLAG	87	5.94	61.5	NA
SDLAG	179	7.58	17.85	NA
DLB	6.75	4.52	0.26	52.3
DLB	60.5	6.99	0.67	187
DLB	215	6.61	0.33	378
TKL	6	7.45	0.83	NA
UPL	7	6.44	0.31	72.3
UPL	56	7.32	0.36	NA
UPL	195	7.25	0.12	228

57 **S4 Supplementary R-scripts**

58 **S4.1 R-script Kruskal-Wallis rank sum test**

```
59 ### required packages
60 library("psych")
61 library("rstatix")
62 library("dplyr")
63 library("readxl")
64 ### load data
65 kw_data <- read_excel("../kw_data.xlsx")
66 # descriptive statistic by group
67 describeBy(kw_data$Parameter, kw_data$Prefix)
68 ### using Kruskal-Wallis-Test
69 kruskal.test(kw_data$Parameter ~ kw_data$Prefix)
70 # example result: Kruskal-Wallis chi-squared = 11.131, df = 3, p-value = 0.01104
71 ### post-hoc analyse (dunn's test)
72 dunn_test(Parameter ~ Prefix, data = kw_data, p.adjust.method = "bonferroni")
```

73 **S4.2 R-script Mann-Whitney-Wilcoxon test**

```
74 ### required packages
75 library("psych")
76 library("readxl")
77 ### Mann-Whitney-U-Test pairwise site comparison
78 ### load data pairwise
79 data_two_cores <- read_excel("../core_1-core_2.xlsx")
80 ### calculate Mann-Whitney-U-Test
81 wilcox.test(parameter ~ Prefix, data = data_two_cores, exact = TRUE, correct = FALSE, conf.int = FALSE)
82 # example results: p-value = 0.01154
83 ### effect size
84 z_parameter <- qnorm(p-value/2)
85 # print z-value
86 z_parameter
87 # example result: z = -2.525907
88 r_parameter <- z_parameter/sqrt(49)
89 # print r-value
90 r_parameter
91 # example result: 0.3608439
92 ### Mann-Whitney-U-Test comparison frozen vs. unfrozen
93 ### load data
94 fn_data <- read_excel("../fro_unfro.xlsx")
95 ### descriptive statistic by groups
96 describeBy(fn_data$parameter, fn_data$Prefix)
97 # example result: frozen: mean = 4.76 wt%, median = 5.1 ; unfrozen: mean = 4.75 wt%, median 3.17
```

```

98 ### calculate Mann-Whitney-U-Test
99 wilcox.test(parameter~Prefix, data = fn_data, exact = FALSE, correct = FALSE, conf.int = FALSE)
100 #example result: p-value = 0.007946
101 ### effect size
102 z_parameter <- qnorm(p-value/2)
103 # print z-value
104 z_paramter
105 # example result: z = -2.654356
106 r_parameter <- z_parameter/sqrt(49)
107 # pront r-value
108 r_parameter
109 # example result: 0.3791937
110 ### same procedure for saline/non-saline comparison
111
112 S4.3 R-script Correlation matrix
113 ### required packages
114 library("corrplot")
115 library("readxl")
116
117 ### load data
118 cm_data <- read_excel("../...correlation_matrix.xlsx")
119 ### create correlation matrix
120 Correlationm <- cor(cm_data)
121 ### create correlation matrix with p-value
122 Correlationp <- cor.mtest(cm_data)
123 ### start the plot
124 pdf(file = "../... /correlations_matrix.pdf",
125     width = 20,
126     height = 20)
127 ### plot with numbers
128 corrplot(Correlationm, method = "number", p.mat=Correlationp$p, insig = "blank",
129         tl.col = "black", tl.srt = 59, tl.cex = 3,
130         number.cex = 2, label.srt = 50)
131 ### add the second half of the plot as dots
132 corrplot(Correlationm, p.mat=Correlationp$p, insig = "blank", type = "upper", tl.pos = "n", tl.cex =1.5, add = TRUE)
133 ### save the plot
134 dev.off()

```

135 **Supplementary References**

136 Blott, S. J. and Pye, K.: GRADISTAT: a grain size distribution and statistics package for the analysis of unconsolidated
137 sediments, Earth Surf Processes Landf, 26, 1237–1248, <https://doi.org/10.1002/esp.261>, 2001.