



# <sup>1</sup> **Quantifying the soil sink of atmospheric Hydrogen: a full year of field**

# <sup>2</sup> **measurements from grassland and forest soilsin the UK**

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# 9 **Abstract**

10 Emissions of hydrogen (H2) gas from human activities are associated with indirect climate warming effects. As 11 the hydrogen economy expands globally (e.g. the use of  $H_2$  gas as an energy source), the anthropogenic 12 release of  $H_2$  into the atmosphere is expected to rise rapidly as a result of increased leakage. The dominant 13 H<sup>2</sup> removal process is uptake into soils; however, removal mechanisms are poorly understood and the fate 14 and impact of increased H<sub>2</sub> emissions remains highly uncertain. Fluxes of H<sub>2</sub> with soils are rarely measured, 15 and data to inform global models is based on few studies. This study presents soil  $H_2$  fluxes from two field 16 sites in central Scotland, a managed grassland and a planted deciduous woodland, with flux measurements 17 of H<sub>2</sub> covering full seasonal cycles. A bespoke flux chamber measurement protocol was developed to deal 18 with the fast decline in headspace concentrations associated with rapid H<sub>2</sub> fluxes, in which non-linear 19 regression models could be fitted to concentration data over a 7-minute enclosure time. We estimate annual 20 H<sub>2</sub> uptake of -3.1  $\pm$  0.1 and -12.0  $\pm$  0.4 kg H<sub>2</sub> ha<sup>-1</sup> yr<sup>-1</sup> and mean deposition velocities of 0.012  $\pm$  0.002 and 21  $0.088 \pm 0.005$  cm s<sup>-1</sup> for the grassland and woodland sites, respectively. Soil moisture was found to be the 22 primary driver of H<sub>2</sub> uptake at the grassland site, where the high clay content of the soil resulted in anaerobic 23 conditions (near zero H<sub>2</sub> flux) during wet periods of the year. Uptake of H<sub>2</sub> at the forest site was highly variable 24 and did not correlate well with any localised soil properties (soil moisture, temperature, total carbon and 25 nitrogen content). It is likely that the high clay content of the grassland site (55% clay) decreased aeration 26 when soils were wet, resulting in poor aeration and low H<sub>2</sub> uptake. The well-drained forest site (25% clay) was 27 not as restricted by exchange of  $H_2$  between the atmosphere and the soil, showing instead a large variability 28 in H<sub>2</sub> flux that is more likely to be related to heterogeneous factors in the soil that control microbial activity 29 (e.g. labile carbon and microbial densities). The results of this study highlight that there is still much that we





- 30 do not understand regarding the drivers of  $H_2$  uptake in soils and that further field measurements are required
- 31 to improve global models.

# 32 **1. Introduction**

33 Prior to the industrial revolution in the 18<sup>th</sup> century, the atmospheric concentration of Hydrogen gas (H<sub>2</sub>) was 34 relatively stable at approximately 330 ppb (Patterson et al., 2021). Human activity over the past two centuries 35 has resulted in increasing atmospheric H<sub>2</sub> concentrations (546 ppb in 2021, Petron et al. (2023)), partly as a 36 result of increasing industrial leaks (Hitchcock 2019; Cooper et al., 2022), partly due to increases in emissions 37 and concentrations of precursor gases such as methane (CH<sub>4</sub>) and volatile organic compounds (VOCs), and 38 partly due to increasing concentrations of other gases in the atmosphere which extend the natural lifetime 39 of H<sub>2</sub> (Patterson et al., 2021). In the atmosphere, H<sub>2</sub> competes for hydroxyl (OH) radicals with gases such as 40 methane (CH4) and carbon monoxide (CO), thus an increase in concentrations of these gases due to human 41 activities has resulted in increasing competition for OH and extended the lifetimes for each species (Khalil & 42 Rasmussen, 1990; Bertagni et al., 2022). Concentrations of atmospheric H<sub>2</sub> gas are indirectly associated with 43 climate warming effects as a result of extending the atmospheric lifetime of the powerful greenhouse gas  $CH_4$ 44 as well as increasing tropospheric ozone and water vapour, which also have a warming potential (Warwick et 45 al., 2004; Ocko & Hamburg, 2022). The associated indirect global warming potential (GWP) had been 46 estimated to be in the range of 3.3 to 5 over a hundred-year time horizon (Derwent et al., 2020, Field & 47 Derwent, 2021), though recent estimates have been made of up to  $11.6 \pm 2.8$  times that of an equivalent 48 mass of carbon dioxide (Sand et al., 2023). The effective GWP and the atmospheric accumulation of H<sub>2</sub> are 49 highly sensitive to its atmospheric lifetime, which is estimated to be approximately 2 years (Novelli et al., 50 1999).

51 The dominant process for H<sub>2</sub> removal from the atmosphere is uptake by soils, which is estimated to be three 52 times larger than the sink due to atmospheric reaction with OH (Warwick et al., 2004; Derwent et al., 2020; 53 Field & Derwent, 2021; Paulot et al., 2021; Ocko & Hamburg, 2022). Whilst both removal mechanisms are 54 highly uncertain, the fate and impact of increased  $H_2$  emissions depends largely on the soil sink strength 55 (Ehhalt & Rohrer, 2009). The soil H<sup>2</sup> sink is caused by microbial activity, both under aerobic and anaerobic 56 conditions (Piché-Choquette & Constant, 2019). A large spectrum of bacteria and archaea can utilise H<sup>2</sup> as an 57 energy source, via the hydrogenase enzyme. Whilst some investigations have highlighted the importance of 58 high-affinity H2-oxidising bacteria (Saavedra-Lavoie et al., 2020), most studies suggest that this enzyme is 59 widespread across many bacterial and archaeal phyla, and that  $H<sub>2</sub>$  consumption is the norm rather than the 60 exception (Islam et al., 2020; Greening & Grinter, 2022). It has been suggested that the potential soil H<sub>2</sub> sink 61 is very large because of the high H<sub>2</sub> demand of microbes (Smith-Downey et al., 2008). However, specific H<sub>2</sub>





 uptake rates for different soil types and conditions are lacking. In addition to microbial activity, diffusion into the soil is a further important rate limiting step. Gases penetrate the soil by passive diffusion and diffusion rates are mainly influenced by porosity, which is affected by soil structure, texture, organic matter contents, vegetation types (roots) and moisture content. Thus, for the same microbial activity, porous soils can be 66 expected to be much larger H<sub>2</sub> sinks than compacted and/or waterlogged soils due to increased gas exchange rates with the atmosphere. At the larger scale, diffusion rates will depend on the changing climate: a wetter 68 climate may lower the H<sub>2</sub> diffusion rates (Paulot et al., 2021). Temperature is another important factor as it determines the rate of microbial enzyme reactions, and a carbon source is required for heterotrophic microbial activity (Islam et al., 2020; Meredith et al., 2016; Baril et al., 2022). In addition, soil H<sup>2</sup> 71 concentrations will be competing with CH<sub>4</sub> as the energy source for soil microbes, hence the H<sub>2</sub> sink strength 72 may in turn affect the CH<sub>4</sub> sink strength and vice versa (Conrad, 1999). The biological sink of atmospheric H<sub>2</sub> has been suggested to be more sensitive to spatial variations of drivers compared to the fluxes of other gases 74 with high variability such as nitrous oxide (N<sub>2</sub>O); however, H<sub>2</sub> measurement data are limited (Baril et al., 2022).

76 Historically, the processes that control  $H_2$  uptake in soils have been severely understudied due to the logistical 77 difficulties and technical constraints on measuring H<sub>2</sub> fluxes. This study presents measurements of H<sub>2</sub> fluxes between the soil and the atmosphere at two field sites in central Scotland, a managed grassland and a planted 79 deciduous woodland. These are the first reported flux measurements of  $H_2$  covering a full annual cycle in the 80 UK. It has previously been reported that forest ecosystems exhibit higher H<sub>2</sub> uptake rates than agroecosystems (Ehhalt and Rohrer, 2009); however, the generality of this and exact mechanisms are still unclear. This study 82 aims to investigate the response of microbial H<sub>2</sub> uptake at a grassland and a forest site to environmental drivers, and to identify differences between the sites. We also describe a dedicated flux chamber 84 methodology which has been developed to best address the challenges of measuring H<sub>2</sub> flux using gas chromatography (GC) analysers.

## **2. Methods**

#### *2.1. Field Sites*

 Measurements of trace gas fluxes and environmental variables were made at two field sites within the Midlothian region in central Scotland (UK, approximately 6 miles south of Edinburgh, Table 1). The first of these was the long-term environmental monitoring site at Easter Bush Farm (grassland). The grassland site (55.8653 °N, -3.206 °W) is an intensively managed, improved grassland (South field in Cowan et al., 2020 and Drewer et al., 2016) that since 2001 has been used predominantly to graze sheep, with a species composition







 **Table 1** Field site environmental properties as reported in previous studies and ongoing research. Mean annual values taken from 10+ years of site data. Rainfall represents throughfall (e.g. rain that reaches the

soil).



#### *2.2. Meteorological and soil measurements*

Continuous environmental measurements were made at both field sites. Air temperature, soil temperature,

soil volumetric water content (VWC) at three depths (5, 10 and 20 cm at the grassland site; 5, 10 and 15 cm





 at the woodland site), relative humidity (RH) and rainfall were measured at both sites throughout the flux measurement campaign (Table S1). For each flux chamber measurement, soil temperature and soil VWC were also measured next to the chamber (<0.5 m distance) at the time of the flux measurement. Soil temperature was measured at 10 cm depth using a handheld probe (ETI Ltd., Worthing, UK), and soil VWC was measured at 12 cm depth using an HS2 HydroSense II handheld soil moisture sensor (Campbell Scientific, Utah, USA), with 4 replicates for each chamber. Soil samples were collected for total carbon (C) and total nitrogen (N) analysis from the top 10 cm of soil at the woodland site in March 2021, September 2021, May 2022, August 2022, November 2022, and March 2023. Subsamples were dried at 105 °C until constant weight, milled using a ball mill (MM200 ball mill, Retsch, Haan, Germany) and analysed using an elemental analyser (Flash SMART, Thermo Fisher Scientific, MA, USA).

#### *2.3. Flux measurements*

130 Fluxes of hydrogen (H<sub>2</sub>), methane (CH<sub>4</sub>) and nitrous oxide (N<sub>2</sub>O) were measured using the static chamber method (e.g. Drewer et al., 2016). Chambers (diameter = 40 cm, height = 30 cm) consisting of opaque polypropylene open-ended cylinders, were installed at each field site: 20 at Easter Bush (grassland) and 36 at Glencorse (woodland). The chambers were inserted into the ground to a depth of approximately 10 cm for the entire study period. The depth to the surface in each chamber was measured at 5 points on the sides of the chamber base using a ruler, from which the average was used to calculate the volume of air within. During measurement periods, aluminium lids were fastened onto the bases using four strong clips; a strip of draft excluder glued onto the lid provided a gas tight seal between chamber and lid. A three-way tap was used for gas sample removal using a 100 ml syringe. 20 ml glass vials were filled with a double needle system to flush the vials with five times their volume. Storage tests using gas standards revealed that gases stored in the vials 140 were stable for up to 24 hours, after which  $H_2$  leakage could be observed in the data. Hence all analyses of  $H_2$  gas samples from the chambers were carried out within 24 hours of measurement in the field (typically within 142 6 hours). Measurements of  $H_2$  and GHGs were made approximately monthly.

143 Two separate measurement protocols were employed to measure greenhouse gases (GHGs) and  $H_2$  fluxes, due to the differences in how the gases behaved within the chamber over a given timespan. For GHG measurements, the standard practice of extracting four gas samples (100 ml) at regular intervals over one 146 hour (0, 20, 40, 60 min) was used (Drewer et al. 2017). However, due to the rapid uptake of H<sub>2</sub> observed in 147 trial measurements (H<sub>2</sub> in the chamber headspace could reach zero ppb in under 10 mins), the time-evolution 148 of H<sub>2</sub> in the chamber was non-linear and therefore a separate measurement protocol was developed for H<sub>2</sub> fluxes. Fluxes of H<sup>2</sup> were measured during entirely separate enclosure periods to the GHGs (albeit on the same day) using an enclosure period with 6 samples taken over 7 minutes (0, 1, 2, 3, 5 & 7 mins). Chambers





- 151 used to measure H<sub>2</sub> were fitted with a small 5 cm diameter PC fan which ran from a 9 V battery during chamber 152 enclosure times to ensure rapid air mixing over the shorter measurement period.
- 153 Concentrations of H<sub>2</sub> were measured using an Agilent 8890 gas chromatograph fitted with a pulsed discharge 154 helium ionization detector (GC-PDHID) equipped with a 7697A headspace autosampler, with capacity for 108 155 vials (Agilent, Santa Clara, California, USA). Concentrations of CH<sub>4</sub> and N<sub>2</sub>O were measured using a gas 156 chromatograph (Agilent 7890B with headspace autosampler 7697A with capacity for 108 vials; Agilent, Santa 157 Clara, California, USA) with a micro-electron capture detector (μECD) for N2O analysis and flame ionization 158 detector (FID) for CH<sub>4</sub> analysis run in parallel. Each analytical run of H<sub>2</sub> and GHG samples included at least 159 three sets of four certified standard concentrations for calibration purposes (certified to  $\pm$  5%). The 160 instrumental noise (σ) of the instruments were 40, 5, and 15 ppb for CH<sub>4</sub>, N<sub>2</sub>O and H<sub>2</sub>, respectively. Based on 161 the methods used, the analytical uncertainty in flux estimates were 0.38, 0.047 and 1.08 nmol m<sup>-2</sup> sec<sup>-1</sup> for 162 CH<sub>4</sub>, N<sub>2</sub>O and H<sub>2</sub>, respectively.
- 163 Fluxes were calculated using linear and non-linear regression methods using the HMR package for the 164 statistical software R (Pedersen *et al.*, 2010). By convention, positive fluxes represent emission from the soil, 165 and negative fluxes indicate that the soil acts as a sink. Fluxes of GHGs were all calculated using linear 166 regression, where *dC/dt* is calculated using the standard line of best fit through the concentration data. As 167 concentrations of H<sub>2</sub> fall exponentially during chamber measurements when soil uptake of H<sub>2</sub> is high, linear 168 regression is not always appropriate. To account for this, fluxes of  $H_2$  were calculated using both linear 169 regression and the HM model, depending on the magnitude of the rate of change observed in each chamber 170 measurement. The HM model is a commonly used non-linear model derived by Hutchinson & Mosier (1981) 171 with a negative exponential form of curvature which calculates the rate of change of a gas concentration at 172  $t = 0$ . The concentration *C* at time *t* is given by Equation 1, where  $C_0$  is the initial concentration,  $C_{max}$  is the 173 value at equilibrium and *k* is a constant. *dC/dt* is is the initial rate of change in concentration at *t* = 0 in nmol 174 mol<sup>-1</sup> s<sup>-1</sup>, calculated using Equation 2.

$$
C_t = C_{max} - (C_{max} - C_o) \exp(-kt)
$$
 (Equation 1)

$$
\frac{dC}{dt} = k(C_{max} - C_o)
$$
 (Equation 2)

177 The initial *dC/dt* is used to calculate the flux using Equation 3, where *F* is gas flux from the soil (nmol m<sup>-2</sup> s<sup>-1</sup>), 178 p is the density of air in mol m<sup>-3</sup>, V is the volume of the chamber in m<sup>3</sup> and A is the ground area enclosed by 179 the chamber in  $m<sup>2</sup>$ .

180 
$$
F = \frac{dC}{dt} \times \rho \times \frac{V}{A}
$$
 (Equation 3)





- At low concentrations near the limit of detection of the analyser, a clear exponential decline was hard to discern from the measurement noise and could give rise to spurious fits to Equation 1. (Examples 1 and 2 in Figure 1 and Table 2). The criteria for using the HM model for each individual flux calculation was based on i) *k* is not unrealistically large in Equation 2 (as defined and limited by the HMR package in R), ii) the flux 185 estimated by linear regression is larger than the analytical uncertainty of the method (1.08 nmol m<sup>-2</sup> s<sup>-1</sup> for H2) and iii) the 95 % confidence interval (95% C.I.) of the HM model fit is less than 5 times the magnitude of the flux estimated using linear regression (removes poor-fitting outliers). In Figure 1 and Table 3, six examples are given in which three selections of linear regression fitting and three selections of the HMR model fitting are used to determine flux. For large uptake fluxes (Examples 4, 5 and 6) the HMR model provides a more suitable fit to the non-linearity in *dC*/*dt*, which linear regression does not accurately represent. Deposition 191 velocity of  $H_2$  was calculated by dividing the calculated flux by the ambient concentration at the site (mean 192 of  $t = 0$  measurements on day of measurement in mol m<sup>-3</sup>).
- 



 **Figure 1.** Examples of concentration data collected during H<sup>2</sup> flux chamber sampling. Linear regression (grey) and HM model (brown) are used to determine *dC*/*dt* for each chamber measurement. Error bars represent the analytical uncertainty of H<sup>2</sup> measurements by GC analysis (15 ppb in this study). Comparisons of flux data presented in Table 2.









205

#### 206 **3. Results**

#### 207 *3.1. Hydrogen Flux measurements*

208 Fluxes of H<sub>2</sub> measured from the grassland site ranged from -15.5 to +5.3 nmol m<sup>-2</sup> s<sup>-1</sup> (deposition velocity (Vd) 209 ranged from 0.070 to -0.026 cm s<sup>-1</sup>) (Figures 2 and S1) over the period of September 2023 to September 2024. 210 More than 90% of the H<sub>2</sub> fluxes measured at the grassland site were negative (soil uptake) and only 2 of 251 211 chamber measurements showed emissions from the soil which exceed the analytical uncertainty of the 212 method. Fluxes of H<sub>2</sub> at the grassland site changed seasonally, with greater uptake in the spring and summer 213 compared with winter, where the flux was close to zero. Fluxes at the grassland site had a median of -1.2 nmol 214  $\text{m}^2$  s<sup>-1</sup> and 95% percentiles of -9.9 to 0.2 nmol  $\text{m}^2$  s<sup>-1</sup>. Fluxes measured from the woodland site ranged from 215  $-40.7$  to  $-1.1$  nmol m<sup>-2</sup> s<sup>-1</sup> (Vd ranged from 0.191 to 0.005 cm s<sup>-1</sup>) (Figures 2 and S1). All fluxes measured at the 216 woodland site showed H<sub>2</sub> uptake in the soil. Spatial variability of H<sub>2</sub> flux at the woodland site was an order of 217 magnitude larger than those observed at the grassland site. Fluxes at the woodland site had a median of - 218 18.7 nmol m<sup>-2</sup> s<sup>-1</sup> and 95% percentiles of -32.4 to -4.3 nmol m<sup>-2</sup> s<sup>-1</sup>. Ambient concentrations of H<sub>2</sub> at the sites 219 ranged from 424.8 to 566.5 ppb. Mean ambient concentrations at the woodland site (484.4 ppb) were on 220 average 21.7 ppb (4.3 %) lower than the grassland site (506.5 ppb) which could be considered statistically 221 insignificant (t-test,  $p > 0.1$ ), but differences were fairly consistent throughout the year (summary statistics 222 presented in Table S2).







223

224 **Figure 2.** Fluxes of H<sup>2</sup> measured using the flux chamber method at grassland (Easter Bush, grassland; grey) 225 and forest (Glencorse Forest, woodland; red) sites in Midlothian, Scotland. Boxplots (a) represent the median, 226 and 25<sup>th</sup> and 75<sup>th</sup> percentiles of flux data of 20 chambers, respectively (whiskers represent the 95<sup>th</sup> 227 percentiles). (b) Frequency distributions of the flux data for both sites (Figure replicated for Vd in Figure S1).

# 228 *3.2. Greenhouse gas fluxes*

229 Fluxes of CH<sub>4</sub> at both sites were close to zero, with mostly small negative fluxes observed at both sites (Figure 230 S3). Soil uptake of CH<sub>4</sub> was observed during the summer months at both sites but during colder months, only 231 the woodland site continued to observe consistent negative CH<sub>4</sub> fluxes. Fluxes of CH<sub>4</sub> measured from the 232 grassland site ranged from -1.2 to 1.0 nmol m<sup>-2</sup> s<sup>-1</sup> with a median of -0.14 nmol m<sup>-2</sup> s<sup>-1</sup>. Fluxes of CH<sub>4</sub> measured 233 from the woodland site ranged from -1.3 to 2.3 nmol  $m^2 s^1$  with a median of -0.32 nmol  $m^2 s^1$ . Only 40% of 234 all CH<sub>4</sub> flux measurements exceeded the analytical uncertainty of the chamber method deployed, highlighting 235 the magnitude of observed fluxes were near the limit of detection of the methodology. Fluxes of N<sub>2</sub>O 236 measured at both sites were relatively low for all measurement dates (58% of all data below the analytical





- 237 uncertainty) with the exception of measurements made in April at the grassland site. Nitrogen fertiliser was
- 238 applied to the field on the 28<sup>th</sup> of March, resulting in increased N<sub>2</sub>O emissions for several weeks (Figure S3).
- 239

# 240 *3.3. Drivers of H<sup>2</sup> flux*

241 Correlations of H<sub>2</sub> flux with soil moisture and soil temperature can be observed at both sites (Figures 4a & 242 4b); however, each site responds differently. Fluxes of H<sub>2</sub> at the grassland site were close to zero when water 243 filled pore space (WFPS) was high (>45%), then tended towards uptake as WFPS decreased. The correlation 244 between H<sub>2</sub> flux and WFPS is weaker at the woodland site and flux data are widely scattered. Fluxes of H<sub>2</sub> at 245 both the grassland and woodland site tended towards higher uptake as temperature increased, though 246 scatter increased toward higher uptake at both sites (>12 °C). A simplistic multiple regression fit between H<sub>2</sub> 247 Ilux (y) with soil moisture (x) and soil temperature (z) ( $y = a_1x^2 + a_2x + b_1z^2 + b_2z + c$ ) accounts for more than 248 half of the variance in the observed fluxes at the grassland site  $(R^2 = 0.60)$  with a significant contribution from 249 soil moisture, but the same approach does not adequately represent the large flux variability at the woodland 250 site ( $R^2$  = 0.14) for which neither soil moisture or soil temperature was found to correlate significantly (Table 251 S3). Fluxes of CH<sub>4</sub> at the sites followed the same trends as H<sub>2</sub> flux in terms of emission/uptake and follow 252 similar correlations with soil moisture and soil temperature as H<sub>2</sub> flux (Figures 4c & 4d). Fluxes of CH<sub>4</sub> at both 253 sites were close to zero (or emission) when soils were wet (>45 % WFPS) and cold (<6 °C). Uptake of CH<sub>4</sub> was 254 greatest when soils were drier and warm.

255 Total carbon (C) and total nitrogen (N) from the woodland site provided comparisons of H<sub>2</sub> flux with soil C and N at the chamber level (Figure S4). Variability in C and N in the replicated cores per chamber was similar to the magnitude of spatial variability at the plot scale, suggesting that localised soil samples were not adequately representative of the soil within a chamber (high spatial variability of C and N in the soil at the  $\leq$  1 m<sup>2</sup> scale). No correlation between H<sub>2</sub> flux with measured total soil C or N in the top 10 cm was found at 260 the woodland site  $(R^2 < 0.01$  for each).

261 By combining continuous soil measurement data collected at each site (soil moisture and temperature at 10 262 cm depth), with the multiple regression model with soil moisture and soil temperature (Figures 4b & 4c) as 263 described in Table S1, continuous H<sub>2</sub> flux predictions were made for a full year (Figure 4a). This model predicts 264 that  $H_2$  flux at the grassland site remains close to zero for most of the time, except when soil moisture drops 265 (e.g. warm months in spring and summer). The model predicts that  $H_2$  flux at the grassland site is strongly 266 dependent on the soil moisture content, with relatively strong periods of  $H_2$  uptake during drier periods 267 (warm periods between rainfall events). H2 flux estimates at the woodland site are more variable, and less 268 susceptible to changes in meteorology or soil conditions. The model predicts a slowdown in H<sub>2</sub> uptake in the





- 269 forest soils during the colder months in winter but is not significantly impacted by changing soil moisture. 270 Total annual estimates of H<sub>2</sub> flux predicted by the model are -3.1  $\pm$  0.1 and -12.0  $\pm$  0.4 kg H<sub>2</sub> ha<sup>-1</sup> yr<sup>-1</sup> for the 271 grassland and woodland sites, respectively. By comparison, a straight average of the measurements, without 272 using models to gap-fill the data, suggests mean fluxes (with 95% C.I.s) of -2.6  $\pm$  0.4 and -18.7  $\pm$  1.0 nmol m<sup>-2</sup> 273 s<sup>-1</sup> which would translate to annual cumulative fluxes of approximately -1.6 ± 0.2 and -11.7 ± 0.6 kg H<sub>2</sub> ha<sup>-1</sup> yr<sup>-</sup> 274 <sup>1</sup> for the grassland and GC sites, respectively. The two estimates agree well at the woodland site, but the gap 275 filling increases the estimated annual  $H_2$  uptake at the grassland site by 56%.
- 276



279 **Figure 3.** Correlations between H<sub>2</sub> flux and (a) water filled pore space (WFPS) and (b) Soil Temperature. 280 Correlations between CH<sup>4</sup> flux and (c) water filled pore space (WFPS) and (d) Soil Temperature. WFPS and soil





- temperature measured at 10 cm depth via sampling probe. A 2nd order polynomial fit (black dashed line) is
- 282 *included as a visual aid* ( $y = a_1x^2 + a_2x + c$ ) (Figure replicated for Vd in Figure S2).

## 



 **Figure 4.** (a) H<sup>2</sup> flux measurements and model predictions for both field sites using a multiple regression fit 286 with soil moisture (x) and soil temperature (z) ( $y = a_1x^2 + a_2x + b_1z^2 + b_2z + c$ ). (b) Continuous water filled pore space (WFPS) at measurements made at 10 cm depth (average of 60 mins). (c) Continuous soil temperature at measurements made at 10 cm depth (average of 60 mins).





#### 290 **4. Discussion**

#### 291 *4.1. Quantification of H<sup>2</sup> flux*

292 Fluxes of H<sub>2</sub> measured in this study range from -40.7 to 5.3 nmol m<sup>-2</sup> s<sup>-1</sup> with mean fluxes of -2.6 ± 0.4 and -293 18.7  $\pm$  1.0 nmol m<sup>-2</sup> s<sup>-1</sup> for the grassland and woodland sites, respectively. Using regression to model (gap-fill) 294 flux data, we estimate annual H<sub>2</sub> uptake of 3.1  $\pm$  0.1 and 12.0  $\pm$  0.4kg H<sub>2</sub> ha<sup>-1</sup> yr<sup>-1</sup> for the grassland and 295 woodland sites, respectively, which increases the expected mean uptake at the grassland site to  $4.3 \pm 0.2$ 296 nmol m<sup>-2</sup> s<sup>-1</sup> while the expected mean uptake at the woodland site remains near 18 nmol m<sup>-2</sup> s<sup>-1</sup>. Predicted 297 uptake is higher at the grassland site due to the expectation in the model that uptake will increase during 298 periods of drier soils that were not measured directly. Predicted uptake estimated by the model and the 299 extrapolation of the mean flux are not significantly different at the woodland site due to the lack of correlation 300 with soil drivers in the model. However, the model does predict that uptake will slow down during the coldest 301 months when fewer measurements were made at the site.

302 Mean measured uptake of H<sub>2</sub> at the grassland site is at the lower end of uptake reported in other studies that 303 directly measured H<sub>2</sub> flux from soils, which range from -1.5 to >20 nmol m<sup>-2</sup> s<sup>-1</sup> (Table 3). The mean soil uptake 304 of  $H_2$  at the woodland site is at the higher end in terms of uptake magnitude, close in magnitude to high 305 deposition velocities reported for peatlands in Simmonds et al., (2011). While uptake at this site seems high, 306 we are confident that the flux measurements are accurate based on the consistency of flux observations and 307 the quality controls put in place. Concentrations of  $H_2$  in the chambers consistently fell exponentially, reaching 308 near zero within 5 minutes (often within 3 mins) of enclosure. At the time of chamber closure (*t*o), a volume 309 of 0.025 m<sup>3</sup> of ambient air at the woodland site contains approximately 400-500 nmol of H<sub>2</sub> gas. To reach zero 310 within 5 mins would require fluxes approximately 10-12 nmol  $m^2 s^1$  in magnitude. While dealing with the 311 exponential non-linearity of the rate of change of the concentration (*dC*/*dt*) does introduce an element of 312 uncertainty in the flux calculations, we are confident the method used in this study (HMR fitting) accurately 313 captures the flux at  $t_0$  and thus a realistic magnitude of soil  $H_2$  uptake.

314 Only two of the measured  $H_2$  fluxes were both positive and larger than the analytical noise of the 315 measurement method. However, these measurements from separate chambers on separate dates (from the 316 grassland site) both showed 7 consecutive concentration measurements, all clearly increasing with time, 317 highlighting that it is possible for  $H_2$  emissions to occur in soils, even where uptake is the predominant 318 direction of flux. It has been observed that legumes produce  $H_2$  during the nitrogen fixation process (e.g. 319 Schubert and Evans 1976; Flynn et al., 2014); however, no legume plants were present in any of the chamber 320 locations during the study. The source of these  $H_2$  emissions remains unknown and at no point did either of 321 the field sites become a source of  $H_2$ , but our observations do highlight that there remain unknown microbial 322 processes at the sub-field scale.





323 **Table 4.** A summary of H<sup>2</sup> fluxes and deposition velocity (Vd) measurements reported in literature, compared

324 with measured and modelled values in this study. Mean values and reported uncertainties. Where only flux

325 or Vd is reported, missing values are estimated using an ambient  $H_2$  concentration of 500 ppb.



326

#### 327 *4.2. Drivers of H<sup>2</sup> flux*

328 This study provides evidence of large variability in  $H_2$  flux behaviour across two different soil types and the 329 importance of environmental factors such as soil temperature and moisture content. At the grassland site, 330 soil moisture (WFPS) dominated the H<sub>2</sub> flux behaviour in the soils. The relationship between H<sub>2</sub> uptake and 331 soil moisture was statistically significant (p <0.001) and explained 60% of the variance observed in the 332 grassland H<sub>2</sub> fluxes observed. While H<sub>2</sub> flux does appear to correlate with soil temperature at the grassland 333 site when compared directly, this is almost entirely due to the strong correlation between soil moisture and 334 soil temperature ( $R^2$  = 0.68). Multiple regression finds soil temperature to be an insignificant variable once 335 the effect of soil moisture is accounted for at the grassland site. Spatial variability in  $H_2$  fluxes at the woodland 336 site were an order of magnitude higher than those at the grassland site. This spatial variability could not be 337 explained by soil moisture, temperature or the total carbon content of the soil. While there do appear to be 338 weak relationships between the flux data and soil moisture and soil temperature, neither is found to be 339 statistically significant (maximum p-value of 0.15 for soil temperature).

340 Meteorological conditions were almost identical at the local scale (sites are less than 3 km apart) and soil at 341 both sites was of a similar pH and had similar total carbon and nitrogen contents. A small difference in





 ambient H<sup>2</sup> concentrations was observed between the sites which may be caused by the large soil uptake and 343 poorer circulation of air at the woodland site, resulting in lower near surface H<sub>2</sub> concentrations. The reason 344 for the large difference in flux of  $H_2$  measured between the two sites is not entirely clear from the measured data, but it is likely that the physical properties of the soils played a role. While rooting systems and carbon structure within the surface layers of the soils will be different at the sites, one large and obvious disparity is the clay content of the soils which is approximately 55% and 25% at the grassland and woodland sites, respectively. The higher density clay soil of the grassland site results in the soil becoming highly anaerobic 349 when moisture levels increase, as can be seen in the switching from CH<sub>4</sub> uptake to CH<sub>4</sub> emission when WFPS exceeded 40%. At the woodland site, a thin layer of organic materials (forest litter) lies on top of a sandy, well-351 drained soil, which may provide ideal conditions for H<sub>2</sub> uptake. Uptake of CH<sub>4</sub> is generally greater than at the grassland site, and WFPS remains lower throughout the year, showing that drainage is significantly faster at 353 the site and suggests that the soils are more aerobic than at the grassland site (e.g. better penetration of  $H_2$  to active regions within the soil). While the differences in soil texture may partly explain the large magnitude 355 of difference in H<sub>2</sub> uptake between the sites, it does not account for the large spatial variability of H<sub>2</sub> flux at the woodland site. While the flux at the grassland site is largely dependent on physical factors at the field scale such as the moisture content (aeration) of the soil, the woodland site showed large variations between plots. This variation may be due to microbial factors that are highly spatial in a forest floor, such as available nutrients (labile carbon from rotting plant litter), canopy shading and varying microbial densities.

#### *4.3. Considerations for future research*

 Chamber flux methods are commonplace in the field of GHG flux measurements, but there are several 363 important factors that need to be considered when carrying out  $H_2$  flux measurements in the field. One of the most important - when using gas chromatography analysis - is the lifetime of samples stored in vials. While it is possible to keep GHG samples in these vials for weeks or even months without significant storage loss, H<sub>2</sub> concentrations were found to change relatively quickly, and should be analysed as soon as is possible (within 24 h of measurement). This severely limits the reach of a particular field experiment to within travel 368 distances of a working H<sub>2</sub> gas chromatography instrument (e.g. not suitable for international shipment of 369 samples). Almost all published H<sub>2</sub> flux measurements to date are within the temperate region of the northern hemisphere (USA and Europe), which limits the available data for models to predict soil/atmosphere 371 interactions at the global scale. Building  $H_2$  flux datasets at a global level would require either investment in localised infrastructure that allowsfor samples to be analysed in-country, or for the deployment of temporary roving measurement methodology which travels between sites. We emphasise that unless particular care and





374 attention is applied to the transportation of gas samples (e.g. tests and quality control checks), the H<sub>2</sub> flux 375 cannot be analysed over a large distance due to leakage of samples.

376 Field measurements of  $H_2$  are beneficial due to realistic environmental conditions. However, the manual aspects of chamber sampling create logistical issues (extensive fieldwork) and the overlap of many 378 environmental and soil variables can make it difficult to identify the driving forces behind  $H_2$  flux (e.g. the soil moisture/temperature comparison). With this setup, the GC-PDHID is limited to one gas sample every 4 minutes, thus auto-chambers (chambers that open/close and measure gas samples automatically) are limited 381 in capability. New faster instruments able to measure H<sub>2</sub> gas via infra-red spectroscopy (by converting H<sub>2</sub> to H2O) are becoming more commercially available (see aerodyne.com/laser-analyzers), but there are no studies 383 using these analysers to date. Previously gas chromatography instrumentation has been used to measure  $H_2$  flux via the aerodynamic gradient method (Meredith et al., 2017), which allows half hourly fluxes to be measured at the field scale. While micrometeorological methods such as the aerodynamic gradient method allow for a greater temporal and spatial coverage of soil fluxes, they also require certain field conditions, such as flat open terrain and large (mains) power supply. In the case of the woodland site in this study, 388 micrometeorological methods are not feasible. With current available  $H_2$  measurement methods, care must be given when planning measurement activities to ensure efficiency in data collection.

390 Lab-based incubation studies of  $H<sub>2</sub>$  flux in literature are similar in number to those measured in the field. Incubation studies allow for better control of soil conditions such as moisture, temperature and nutrient content, environmental conditions (air temperature) and also for consistency in microbial populations (via replicates of well mixed/homogenised soils). For example, in this study, it was difficult to determine the impact of soil temperature due to the correlation with soil moisture. Due to the climate in the region, there were no periods when the soils were cold and also dry, preventing observations of different extremes of the 396 driving forces behind H<sub>2</sub> flux. Incubation studies would be able to provide more information on these drivers which may help modelling efforts; however, field measurements are still required to validate flux models as incubation studies inevitably come with the caveat that flux measurements are not representative of true soil conditions due to soil cores being repacked and creating therefore artificial conditions.

# 400 **5. Conclusions**

401 This study reports that the soil sink (uptake) of  $H_2$  for a grassland and a forest site in close proximity is -3.1  $\pm$ 402 0.1 and -12.0  $\pm$  0.4 kg H<sub>2</sub> ha<sup>-1</sup> yr<sup>-1</sup>, respectively (with mean Vds of 0.012  $\pm$  0.002 and 0.088  $\pm$  0.005 cm s<sup>-1</sup> for 403 grassland and forest soils, respectively). Soil moisture was found to be the primary driver of H<sub>2</sub> uptake at the 404 grassland site, where the high clay content of the soil resulted in anaerobic conditions (near zero H<sub>2</sub> flux) 405 during wet periods of the year. Uptake of  $H<sub>2</sub>$  at the forest site was highly variable and did not correlate well





 with any localised soil properties. Both sites were exposed to similar meteorological conditions (3 km apart) and had similar basic soil properties (such as pH and carbon content), thus we conclude that the large difference in uptake between the soils was dependent on soil aeration. It is likely that the high clay content of the grassland site (55%) resulted in a lack of aeration when soils were wet, while the well-drained forest 410 site (25% clay) was not restricted by exchange of H<sub>2</sub> between the atmosphere and the soil, showing instead a 411 large variability in H<sub>2</sub> flux that could be related to heterogeneous factors that control microbial activity (e.g. labile carbon and microbial densities). In order to account for the large magnitude of site-scale differences 413 like those observed in this study, further field sites should be studied over a range of soil and land cover types 414 and management activities to improve global models of the soil  $H_2$  sink. In addition, laboratory incubations are needed to measure H<sup>2</sup> fluxes under controlled environmental conditions to refine the main driving 416 parameters of  $H_2$  fluxes further.

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## **7. Competing interests**

The authors declare that they have no conflict of interest.

#### **8. Data availability**

Data currently undergoing preparation for submission to the Environmental Information Data Centre (EIDC).

https://eidc.ac.uk/





#### **9. Author contributions**

- N. Cowan was the primary author of the manuscript and carried out all data analysis presented. The field
- team that developed measurement methodology protocols, carried out measurements, maintained field
- instrumentation and performed lab analysis consisted of T. Roberts, M. Hanlon, A. Bezanger, G. Toteva, A.
- Tweedie, K. Yeung and A. Deshpande. The project management and significant contributors to the
- manuscript text consisted of P. Levy, U. Skiba, E. Nemitz and J. Drewer. All coauthors contributed to the
- writing of the manuscript before submission.

#### **10.References**

- Baril, X., Durand, A.-A., Srei, N., Lamothe, S., Provost, C., Martineau, C., Dunfield, K., Constant, P., 2022. The 441 biological sink of atmospheric H2 is more sensitive to spatial variation of microbial diversity than  $N_2O$  and CO<sup>2</sup> emissions in a winter cover crop field trial. Science of The Total Environment. https://doi.org/10.1016/j.scitotenv.2022.153420
- Bertagni, M.B., Pacala, S.W., Paulot, F., Porporato, A., 2022. Risk of the hydrogen economy for atmospheric methane. Nat Commun. https://doi.org/10.1038/s41467-022-35419-7
- Billington, H.L. and Pelham, J., 1991. Genetic Variation in the Date of Budburst in Scottish Birch Populations: Implications for Climate Change. Functional Ecology, 5(3), pp. 403–409. https://doi.org/10.2307/2389812.
- Buzzard, V., Thorne, D., Gil-Loaiza, J., Cueva, A., Meredith, L.K., 2022. Sensitivity of soil hydrogen uptake to natural and managed moisture dynamics in a semiarid urban ecosystem. PeerJ. https://doi.org/10.7717/peerj.12966
- Conrad, R., 1999. Contribution of hydrogen to methane production and control of hydrogen concentrations in methanogenic soils and sediments. FEMS Microbiology Ecology. https://doi.org/10.1111/j.1574- 6941.1999.tb00575.x
- Cooper, J., Dubey, L., Bakkaloglu, S., Hawkes, A., 2022. Hydrogen emissions from the hydrogen value chain- emissions profile and impact to global warming. Science of The Total Environment. https://doi.org/10.1016/j.scitotenv.2022.154624
- Cowan, N., Levy, P., Maire, J., Coyle, M., Leeson, S.R., Famulari, D., Carozzi, M., Nemitz, E., Skiba, U., 2020. An evaluation of four years of nitrous oxide fluxes after application of ammonium nitrate and urea fertilisers measured using the eddy covariance method. Agricultural and Forest Meteorology. https://doi.org/10.1016/j.agrformet.2019.107812





- Derwent, R.G., Stevenson, D.S., Utembe, S.R., Jenkin, M.E., Khan, A.H., Shallcross, D.E., 2020. Global modelling studies of hydrogen and its isotopomers using STOCHEM-CRI: Likely radiative forcing consequences of a future hydrogen economy. International Journal of Hydrogen Energy. https://doi.org/10.1016/j.ijhydene.2020.01.125
- Deshpande, A.G., Jones, M.R., van Dijk, N., Mullinger, N.J., Harvey, D., Nicoll, R., Toteva, G., Weerakoon, G., Nissanka, S., Weerakoon, B., Grenier, M., Iwanicka, A., Duarte, F., Stephens, A., Ellis, C.J., Vieno, M., Drewer,
- J., Wolseley, P.A., Nanayakkara, S., Prabhashwara, T., Bealey, W.J., Nemitz, E., Sutton, M.A., 2024. Estimation of ammonia deposition to forest ecosystems in Scotland and Sri Lanka using wind-controlled
- 469 NH<sub>3</sub> enhancement experiments. Atmospheric Environment.
- https://doi.org/10.1016/j.atmosenv.2023.120325
- Drewer, J., Anderson, M., Levy, P.E., Scholtes, B., Helfter, C., Parker, J., Rees, R.M., Skiba, U.M., 2016. The impact of ploughing intensively managed temperate grasslands on N2O, CH4 and CO2 fluxes. Plant Soil. https://doi.org/10.1007/s11104-016-3023-x
- Drewer, J., Anderson, M., Levy, P.E., Scholtes, B., Helfter, C., Parker, J., Rees, R.M., Skiba, U.M., 2016. The impact of ploughing intensively managed temperate grasslands on N2O, CH4 and CO2 fluxes. Plant Soil. https://doi.org/10.1007/s11104-016-3023-x
- Ehhalt, D.H., Rohrer, F., 2009. The tropospheric cycle of H2: a critical review. Tellus B: Chemical and Physical Meteorology. https://doi.org/10.1111/j.1600-0889.2009.00416.x
- Field, R.A., Derwent, R.G., 2021. Global warming consequences of replacing natural gas with hydrogen in the

domestic energy sectors of future low-carbon economies in the United Kingdom and the United States of

- America. International Journal of Hydrogen Energy. https://doi.org/10.1016/j.ijhydene.2021.06.120
- Flynn, B., Graham, A., Scott, N., Layzell, D.B., Dong, Z., 2014. Nitrogen fixation, hydrogen production and N2O emissions. Can. J. Plant Sci. https://doi.org/10.4141/cjps2013-210
- Greening, C., Grinter, R., 2022. Microbial oxidation of atmospheric trace gases. Nat Rev Microbiol. https://doi.org/10.1038/s41579-022-00724-x
- Hammer, S., Levin, I., 2009. Seasonal variation of the molecular hydrogen uptake by soils inferred from continuous atmospheric observations in Heidelberg, southwest Germany. Tellus B: Chemical and Physical Meteorology. https://doi.org/10.1111/j.1600-0889.2009.00417.x
- Hitchcock, W. K., Beamish, B. B. and Cliff D., 2019. A Study of the Formation of Hydrogen Produced During the Oxidation of Bulk Coal Under Laboratory Conditions, in Naj Aziz and Bob Kininmonth (eds.), Proceedings
- of the 2008 Coal Operators' Conference, Mining Engineering, University of Wollongong





- Hutchinson, G.L., Mosier, A.R., 1981. Improved Soil Cover Method for Field Measurement of Nitrous Oxide Fluxes. Soil Science Society of America Journal. https://doi.org/10.2136/sssaj1981.03615995004500020017x
- Islam, Z.F., Welsh, C., Bayly, K., Grinter, R., Southam, G., Gagen, E.J., Greening, C., 2020. A widely distributed hydrogenase oxidises atmospheric H2 during bacterial growth. The ISME Journal. https://doi.org/10.1038/s41396-020-0713-4
- Khalil, M.A.K., Rasmussen, R.A., 1990. The global cycle of carbon monoxide: Trends and mass balance. Chemosphere. https://doi.org/10.1016/0045-6535(90)90098-e
- Field, R.A., Derwent, R.G., 2021. Global warming consequences of replacing natural gas with hydrogen in the
- domestic energy sectors of future low-carbon economies in the United Kingdom and the United States of
- America. International Journal of Hydrogen Energy. https://doi.org/10.1016/j.ijhydene.2021.06.120
- Lallo, M., Aalto, T., Hatakka, J., Laurila, T., 2009. Hydrogen soil deposition at an urban site in Finland. Atmos. Chem. Phys. https://doi.org/10.5194/acp-9-8559-2009
- Meredith, L.K., Commane, R., Keenan, T.F., Klosterman, S.T., Munger, J.W., Templer, P.H., Tang, J., Wofsy, S.C.,
- Prinn, R.G., 2016. Ecosystem fluxes of hydrogen in a mid-latitude forest driven by soil microorganisms and plants. Global Change Biology. https://doi.org/10.1111/gcb.13463
- Nagai, M., Kakiuchi, H., Masuda, T., 2024. Measurements of hydrogen deposition velocities by farmland soil using D2 gas. Radiation Protection Dosimetry. https://doi.org/10.1093/rpd/ncae055
- Novelli, P.C., Lang, P.M., Masarie, K.A., Hurst, D.F., Myers, R., Elkins, J.W., 1999. Molecular hydrogen in the
- troposphere: Global distribution and budget. J. Geophys. Res. https://doi.org/10.1029/1999jd900788
- Ocko, I.B., Hamburg, S.P., 2022. Climate consequences of hydrogen emissions. Atmos. Chem. Phys. https://doi.org/10.5194/acp-22-9349-2022
- Paulot, F., Paynter, D., Naik, V., Malyshev, S., Menzel, R., Horowitz, L.W., 2021. Global modeling of hydrogen
- using GFDL-AM4.1: Sensitivity of soil removal and radiative forcing. International Journal of Hydrogen Energy. https://doi.org/10.1016/j.ijhydene.2021.01.088
- Paulot, F., Pétron, G., Crotwell, A.M., Bertagni, M.B., 2024. Reanalysis of NOAA H2 observations: implications for the H2 budget. Atmos. Chem. Phys. https://doi.org/10.5194/acp-24-4217-2024
- Patterson, J.D., Aydin, M., Crotwell, A.M., Pétron, G., Severinghaus, J.P., Krummel, P.B., Langenfelds, R.L.,
- Saltzman, E.S., 2021. H 2 in Antarctic firn air: Atmospheric reconstructions and implications for
- anthropogenic emissions. Proc. Natl. Acad. Sci. U.S.A. https://doi.org/10.1073/pnas.2103335118





- Pedersen, A.R., Petersen, S.O., Schelde, K., 2010. A comprehensive approach to soil-atmosphere trace-gas flux estimation with static chambers. European J Soil Science. https://doi.org/10.1111/j.1365- 2389.2010.01291.x
- Petron, G. B., Crotwell, A. M., Mund, J., Crotwell, M., Mefford, T., Thoning, K., Hall, B. D., Kitzis, D. R., Madronich, M., Moglia, E., Neff, D., Wolter, S., Jordan, A., Krummel, P., Langenfelds, R., and Patterson, J.
- D.: Atmospheric H2 observations from the NOAA Global Cooperative Air Sampling Network, Atmos. Meas.
- Tech. Discuss. [preprint], https://doi.org/10.5194/amt-2024-4, in review, 2024.
- Piché-Choquette, S., Constant, P., 2019. Molecular Hydrogen, a Neglected Key Driver of Soil Biogeochemical Processes. Appl Environ Microbiol. https://doi.org/10.1128/aem.02418-18
- Saavedra-Lavoie, J., de la Porte, A., Piché-Choquette, S., Guertin, C., Constant, P., 2020. Biological H2 and CO
- oxidation activities are sensitive to compositional change of soil microbial communities. Can. J. Microbiol. https://doi.org/10.1139/cjm-2019-0412
- Sand, M., Skeie, R.B., Sandstad, M., Krishnan, S., Myhre, G., Bryant, H., Derwent, R., Hauglustaine, D., Paulot,
- F., Prather, M., Stevenson, D., 2023. A multi-model assessment of the Global Warming Potential of hydrogen. Commun Earth Environ. https://doi.org/10.1038/s43247-023-00857-8
- Schubert, K.R., Evans, H.J., 1976. Hydrogen evolution: A major factor affecting the efficiency of nitrogen fixation in nodulated symbionts. Proc. Natl. Acad. Sci. U.S.A. https://doi.org/10.1073/pnas.73.4.1207
- Simmonds, P.G., Derwent, R.G., Manning, A.J., Grant, A., O'doherty, S., Spain, T.G., 2011. Estimation of hydrogen deposition velocities from 1995–2008 at Mace Head, Ireland using a simple box model and concurrent ozone depositions. Tellus B: Chemical and Physical Meteorology. https://doi.org/10.1111/j.1600-0889.2010.00518.x
- Smith-Downey, N.V., Randerson, J.T., Eiler, J.M., 2008. Molecular hydrogen uptake by soils in forest, desert, and marsh ecosystems in California. J. Geophys. Res. https://doi.org/10.1029/2008jg000701
- Warwick, N.J., Bekki, S., Nisbet, E.G., Pyle, J.A., 2004. Impact of a hydrogen economy on the stratosphere and
- troposphere studied in a 2-D model. Geophysical Research Letters. https://doi.org/10.1029/2003gl019224