

1 **Biogeochemical Layering and Transformation of**
2 **Particulate Organic Carbon in the Tropical Northwestern**
3 **Pacific Ocean Inferred from $\delta^{13}\text{C}$**

4 **Authors**

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15 **Abstract.** Particulate organic carbon (POC) serves as the main carrier of the biological pump and
16 determines its transmission efficiency, yet the transformation processes of POC remain incompletely
17 understood. This study reports the vertical distribution of POC, dissolved inorganic carbon (DIC), $\delta^{13}\text{C}$ -
18 POC, and $\delta^{13}\text{C}$ -DIC in the tropical Northwestern Pacific Ocean (TNPO). The research identified three
19 distinct biogeochemical layers governing POC transformation: the POC rapid synthesis-degradation
20 layer (RSDL, 0-300 m), the net degradation layer (NDL, 300-1,000 m), and the stable layer (SL, 1,000-
21 2,000 m). From the top to the bottom of the RSDL, $\delta^{13}\text{C}$ -POC values decreased by an average of 2.23‰,
22 while the carbon-to-nitrogen ratios (C:N) increased by an average of 2.3:1, indicating the selective
23 degradation of POC. In the NDL, $\delta^{13}\text{C}$ -POC and $\delta^{13}\text{C}$ -DIC exhibited a significant negative correlation (r
24 = 0.43, $p < 0.05$), indicating a net transformation of POC to DIC. In the SL, POC proved to be resistant
25 to degradation, with POC exhibiting the highest C:N (15:1 on average) and the lowest $\delta^{13}\text{C}$ -POC values
26 (average -27.71‰).

27 **1 Introduction**

28 As the most significant carbon reservoir on the earth's surface, the ocean absorbs about 2.6 billion tons
29 of carbon dioxide (CO₂) from the atmosphere each year, accounting for 25% of global anthropogenic
30 CO₂ emissions (Friedlingstein et al., 2023). After entering the ocean, CO₂ initially dissolves in seawater,
31 forming dissolved inorganic carbon (DIC). Subsequently, phytoplankton and photosynthetic bacteria at
32 the ocean surface convert it into organic carbon through photosynthesis. The majority of carbon in the
33 ocean is in the form of DIC, constituting over 98% of the total carbon content, with the remaining 2%

34 existing as POC and dissolved organic carbon (DOC). Despite being in minimal quantities, POC can be
35 transported to the deep ocean through the biological pump and buried for thousands of years. This process
36 of carbon sequestration aids in the absorption of CO₂ by the ocean, contributing to the regulation of
37 atmospheric CO₂ levels (Longhurst and Glen Harrison, 1989; Turner, 2015). Organic matter produced
38 from the euphotic layer is the primary food source for heterotrophic communities in the dark ocean
39 (Smith et al., 2008); once POC is exported from the euphotic layer, microorganisms rapidly utilize it,
40 releasing DIC (Song, 2010).

41 Some studies have shown that unstable components such as proteins and carbohydrates in POC are
42 preferentially degraded by microorganisms (Eadie and Jeffrey, 1973). However, conducting detailed
43 quantitative analyses of each POC component in actual investigations is challenging, necessitating the
44 use of alternative indicators to demonstrate selective degradation. The One generally accepted indicator
45 is the carbon-to-nitrogen ratios (C:N) due to inherent differences in the C:N of various compounds in
46 POC (Morales et al., 2021). Thus, changes in the C:N during degradation can signify the selective
47 degradation of POC. Nevertheless, the composition of POC is highly complex, and the C:N of its
48 different components are not absolute. For example, lipids typically have a higher C:N than proteins, but
49 the opposite can also occur (Sannigrahi et al., 2005; Hernes and Benner, 2002). Therefore, relying solely
50 on the C:N to reflect the selective degradation process of POC has significant limitations. Although the
51 vital activities of the microbial community in the dark ocean are predominantly driven by heterotrophic
52 respiration (Herndl et al., 2023), many autotrophic organisms use chemical energy to synthesize POC.
53 Compelling evidence indicates that chemoautotrophy plays a substantial role in the fixation of DIC in
54 the minimum oxygen zone (OMZ) (Reinthal et al., 2010) and the deeper ocean (Passos et al., 2022;
55 Walsh et al., 2009). Consequently, there is a continuous conversion of POC and DIC throughout the
56 ocean water column. Exploring the degradation and synthesis of POC in the ocean is imperative to
57 enhance our comprehension of the biological pump processes.

58 The DIC in seawater primarily occurs in four chemical forms: H₂CO₃, HCO₃⁻, CO₃²⁻, and CO₂. In
59 comparison, the composition of POC is more complex. POC comprises various organic compounds
60 originating from living organisms such as phytoplankton, zooplankton, and microorganisms. It also
61 encompasses fecal particles, cell fragments, and diverse organic substances from external sources. Only
62 a small fraction of the POC has been accurately identified in terms of molecular structures (Kharbush et
63 al., 2020). As the depth increases, the readily degradable components in POC are used up, leading to a

64 more intricate structure of the remaining POC through the transformation process. The remaining
65 refractory POC is even more difficult to identify (Lee et al., 2000). Therefore, it becomes challenging to
66 study the chemical characteristics of POC and its transformation process from itself. The $\delta^{13}\text{C}$ is a crucial
67 indicator that can reveal the origin, migration, and transformation of POC, making it significantly
68 important in the investigation of the marine carbon cycle (Ding et al., 2020; Jeffrey et al., 1983).
69 Compared with POC ~~concentrationmolecules~~, $\delta^{13}\text{C}$ -POC provides a more accurate reflection of the
70 chemical properties of the POC pool and the migration and transformation processes of POC (Close and
71 Henderson, 2020). Similarly, $\delta^{13}\text{C}$ -DIC can offer insights into important processes within the ocean
72 carbon cycle. As POC settles, it undergoes a series of biogeochemical processes, including synthesis,
73 degradation, and adsorption. Therefore, the isotope fractionation effect in POC is strong, resulting in
74 significant differences in $\delta^{13}\text{C}$ -POC values at different depths. In contrast, the fractionation of $\delta^{13}\text{C}$ -DIC
75 is subject to fewer influencing factors, and the DIC concentration in the ocean is notably high, thereby
76 engendering minimal variability in $\delta^{13}\text{C}$ -DIC ~~values~~ across the ocean water column (Jeffrey et al., 1983).
77 Therefore, $\delta^{13}\text{C}$ -DIC is more sensitive to the fractionation effect in the ocean carbon cycle. Even slight
78 variations in the $\delta^{13}\text{C}$ -DIC ~~values~~ can reflect significant processes involved in the migration and
79 transformation of POC (Quay and Stutsman, 2003). Through the analysis of $\delta^{13}\text{C}$ -POC and $\delta^{13}\text{C}$ -DIC
80 ~~values~~, we can enhance our comprehension of the intricate composition, transport, and alteration
81 mechanism of POC, providing us with a more profound insight into the dynamic transformations within
82 the ocean biological pump.

83 The tropical Northwestern Pacific Ocean (TNPO) is characterized by intricate current patterns and
84 water mass distributions (Hu et al., 2015; Schönau et al., 2022), and it is also known for the highest
85 surface seawater temperatures globally (Jia et al., 2018). High temperatures facilitate the respiration ~~of~~
86 ~~by~~ heterotrophic organisms, promoting the formation of biological hotspots and ultimately enhancing
87 material circulation and energy flow in the upper ocean (Guo et al., 2023a; Iversen and Ploug, 2013).
88 The air-sea interaction within the TNPO is highly dynamic, exhibiting a shift from being a carbon sink
89 to a carbon source as it extends from higher to lower latitudes (Takahashi et al., 2009; Wu et al., 2005).
90 The complex hydrological characteristics, rapid elemental cycle, and frequent air-sea exchange render
91 the TNPO an ideal laboratory for exploring the ocean carbon cycle. In this research, we collected
92 seawater and particulate matter samples at six stations in the core and boundary regions of the TNPO,

93 and the relationship between DIC, POC, and their stable carbon isotopes was comprehensively analyzed
94 to enhance our understanding of the POC transformation process and the ocean carbon cycle process.

95 **2 Sampling and Methods**

96 The samples were collected in the TNPO during an expedition on R/V *Kexue* from March to April 2022.
97 A total of 6 stations were set up: EQ-6 (150.99° E, 0.00° N, 1944 m), E142-3 (140.99° E, 12.01° N, 4091
98 m), E142-7 (140.99° E, 15.99° N, 4725 m), E142-11 (140.99° E, 20.00° N, 462 4m), E142-13 (142.04°
99 E, 0.00° N, 3382 m) and E142-19 (141.99° E, 6.01° N, 2580 m) (Fig. 1). The 12-L Niskin bottles (KC-
100 Denmark, Denmark) mounted on a Conductivity-Temperature-Depth (CTD, Sea-bird SBE911, United
101 States) rosette ~~was-were~~ used to obtain water samples from the vertical profile of 0-2,000 m at each
102 station for analysis of temperature, salinity, dissolved oxygen (DO), POC, $\delta^{13}\text{C}$ -POC, particulate nitrogen
103 (PN), DIC, $\delta^{13}\text{C}$ -DIC, and chlorophyll a (Chl-*a*). The specific sampling and analysis methods are as
104 follows.

105 **Temperature and salinity:** The temperature and salinity were measured by CTD (Sea-bird SBE911,
106 United States) in situ during sampling.

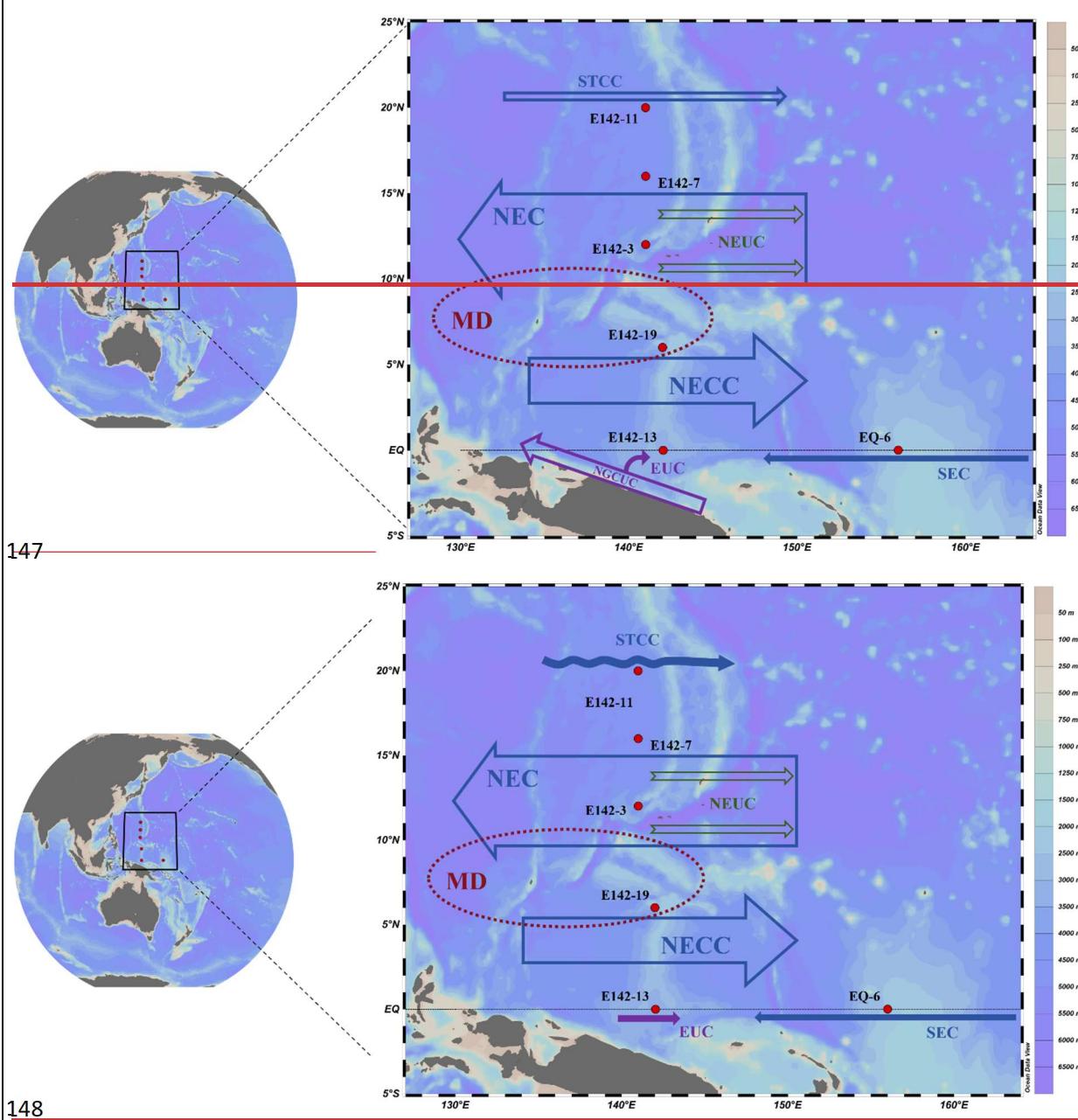
107 **DO:** ~~DO was determined in situ using the Winkler method with a measurement precision of 2.2×10^{-3}
108 $\mu\text{mol/L}$. At each depth, we collected samples in 50 mL brown bottles, added manganese sulfate and
109 alkaline potassium iodide to fix the oxygen, then titrated the released iodine with sodium thiosulfate to
110 calculate DO concentrations~~ Water samples were collected, fixed, and titrated according to the classic
111 Winkler method, the precision of which was $2.2 \times 10^{-3} \mu\text{mol/L}$ (Bryan et al., 1976; Zuo et al., 2018).
112 ~~The discrete DO samples were used to calibrate the DO concentration data obtained by the CTD sensor.~~

113 **POC, $\delta^{13}\text{C}$ -POC, and PN:** Particle samples were obtained by filtering 2-5 L of seawater onto a GF/F
114 glass filter (0.7 μm , Whatman) that had been combusted in a muffle furnace (450°C, 4 h) and acid-soaked
115 (0.5 M hydrochloric acid (HCl), 24 h). The filter was treated with HCl to remove inorganic carbonates
116 and oven-dried at 60°C. ~~After collection, samples were stored below -20 °C until laboratory analysis.~~
117 Afterward, POC, PN concentration, and $\delta^{13}\text{C}$ -POC ~~values~~ were analyzed using an elemental analyzer
118 and an isotope mass spectrometer (Thermo Fisher Scientific Flash EA 1112 HT-Delta V Advantages,
119 United States) with an accuracy of $\pm 0.8\%$ and $\pm 0.2\%$, respectively. ~~Standard reference materials~~
120 ~~were used to calibrate $\delta^{13}\text{C}$ and POC, PN measurements, including USGS64 ($\delta^{13}\text{C} = -40.8 \pm 0.04\%$, C%~~

121 = 31.97%, N% = 18.65%, Indiana University), USGS40 ($\delta^{13}\text{C} = -26.39 \pm 0.04\text{\textperthousand}$, C% = 40.8%, N% =
122 9.52%, Geological Survey, United States), and Urea #2a ($\delta^{13}\text{C} = -9.14 \pm 0.02\text{\textperthousand}$, C% = 20%, N% =
123 46.67%, Indiana University). We implemented a quality control protocol by randomly inserting a
124 certified reference material after every 10 samples. The measured values of these reference materials
125 were subsequently plotted against the calibration curve to monitor and verify instrument stability
126 throughout the analytical process (Ma et al., 2021).

127 **DIC and $\delta^{13}\text{C-DIC}$:** Sampling was performed using a 50 ml glass bottle. After the water sample
128 overflowed, 1 ml of the sample was taken out with a pipette and then fixed with saturated mercuric
129 chloride solution to remove the influence of biological activity. After collection, samples were stored in
130 refrigerator at 4°C for later laboratory measurement of DIC concentration using a total DIC analyzer
131 The DIC concentration was measured using a DIC analyzer (Apollo SciTech AS-C3, United States) with an
132 accuracy of $\pm 0.1\%$ (Ma et al., 2020). For calibration, certified reference material (Batch 144, 2031.53
133 $\pm 0.62 \mu\text{mol/kg}$ provided by the Scripps Institution of Oceanography (University of California, San
134 Diego) was used. $\delta^{13}\text{C-DIC}$ values automatic: Automatic analysis was performed using a Thermo Delta-
135 V isotope ratio mass spectrometer (ThermoFisher Scientific MAT 253Plus, United States). For
136 calibration, certified reference materials for $\delta^{13}\text{C-DIC}$ were used, including GBW04498 ($\delta^{13}\text{C} = -27.28$
137 $\pm 0.10\text{\textperthousand}$), GBW04499 ($\delta^{13}\text{C} = -19.58 \pm 0.10\text{\textperthousand}$), and GBW04500 ($\delta^{13}\text{C} = -4.58 \pm 0.12\text{\textperthousand}$), all provided
138 by the Institute of Geophysical and Geochemical Exploration (Chinese Academy of Geological Sciences).
139 We inserted a reference standard every 10 samples, using its measured values to verify instrument
140 stability via the calibration curve.

141 **Chl -a:** 2 L of water sample after zooplankton removal was filtered onto pre-combusted (450°C for 5 h)
142 GF/F filters (0.7 μm , Whatman); and placed in the refrigerator at -20°C before measurement. In the
143 laboratory, the filters were extracted with 90% propanol for 12-24 h, and the concentration was measured
144 using a fluorescence photometer (Turner Designs, United States) For calibration, Chlorophyll a analytical
145 standard (purity $\geq 95.0\%$) provided by Sigma-Aldrich (SIAL, St. Louis, MO, United States) were used.
146 (Ma et al., 2020).



149 **Figure 1. TPWO sampling stations (red dots in the figure) and ocean current distribution. In the figure, blue**
150 **represents the ocean currents from the surface to the bottom of the thermocline, mainly STCC, NEC, NECC,**
151 **and SEC; green represents the ocean currents in the subthermocline, mainly NEUC; purple represents the**
152 **ocean currents from the bottom of the thermocline to the subthermocline, mainly EUC.**

153 **3 Results and Discussion**

154 **3.1 Hydrological Characteristics**

155 Except for station E142-11, the remaining five stations are all located at the Western Pacific Warm Pool
156 (WPWP). The SST of the five stations in the warm pool area was higher, averaging 29.01 ± 0.67 °C,
157 while station E142-11 had a lower SST of 25.02 °C. The strong seawater stratification in the study area

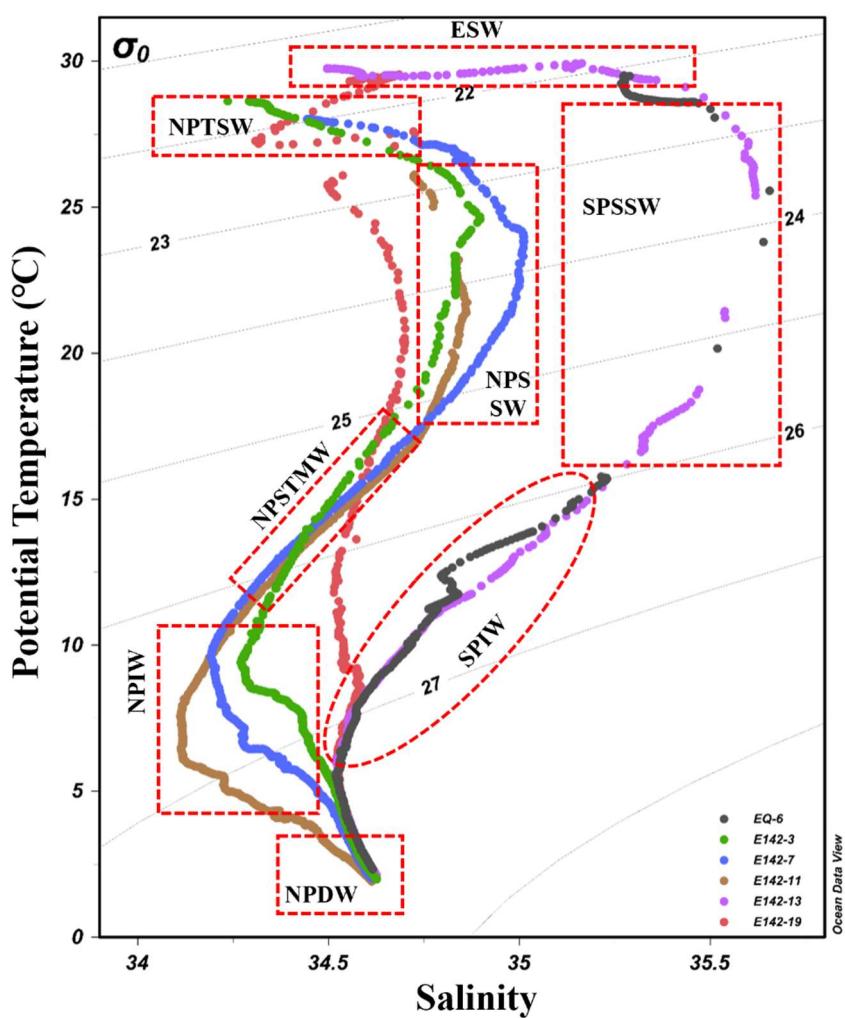
158 restricted the movement of nutrient-rich water from the deep to the upper ocean, resulting in the region
 159 showing oligotrophic characteristics (Radenac et al., 2013). Therefore, the Chl-*a* concentration in-at the
 160 deep chlorophyll maximum layer depth (DCMD) was notably low, with an average of only 0.24 ± 0.04
 161 $\mu\text{g/L}$. Based on the fluorescence intensity measured by the CTD in-situ fluorescence sensor, we
 162 calculated the Primary Production Zone Depth (PPZD), which is the depth where the fluorescence
 163 intensity drops to 10% of its maximum value above this depth (Owens et al., 2015). Additionally, the
 164 Mixed Layer Depth (MLD) at each station was determined using the temperature threshold method
 165 (Table 1) (Thompson, 1976). The results indicate that the PPZD at each station is deeper than the MLD,
 166 suggesting that the POC generated at these stations does not undergo particularly complex physical
 167 mixing after its formation (Buesseler et al., 2020).

168 **Table 1. The water depth (WD), the PPZD, the MLD, the deep chlorophyll maximum layer depth**
 169 **(DCMD) and the Chl-*a* at DCMD for each station.**

<u>Station</u>	<u>Longitude</u>	<u>Latitude</u>	<u>WD</u>	<u>PPZD</u>	<u>MLD</u>	<u>DCMD</u>	<u>Chl-<i>a</i></u>
	<u>°E</u>	<u>°N</u>	<u>m</u>	<u>m</u>	<u>m</u>	<u>m</u>	<u>$\mu\text{g/L}$</u>
<u>EQ-6</u>	<u>155.99</u>	<u>0.00</u>	<u>1944</u>	<u>129</u>	<u>65</u>	<u>50</u>	<u>0.31</u>
<u>E142-3</u>	<u>141.00</u>	<u>12.01</u>	<u>4091</u>	<u>216</u>	<u>102</u>	<u>140</u>	<u>0.19</u>
<u>E142-7</u>	<u>141.00</u>	<u>16.00</u>	<u>4725</u>	<u>204</u>	<u>68</u>	<u>150</u>	<u>0.25</u>
<u>E142-11</u>	<u>140.99</u>	<u>20.00</u>	<u>4624</u>	<u>203</u>	<u>42</u>	<u>90</u>	<u>0.21</u>
<u>E142-13</u>	<u>142.04</u>	<u>0.00</u>	<u>3382</u>	<u>165</u>	<u>45</u>	<u>90</u>	<u>0.25</u>
<u>E142-19</u>	<u>142.00</u>	<u>6.01</u>	<u>2580</u>	<u>170</u>	<u>109</u>	<u>100</u>	<u>0.21</u>

170
 171 Based on the relationship between potential temperature and salinity (θ-S) (Fig. 2), eight water masses
 172 in the study area were identified: North Pacific Tropical Surface Water (NPTSW), North Pacific
 173 Subsurface Water (NPSSW), North Pacific Subtropical Mode Water (NPSTMW), North Pacific
 174 Intermediate Water (NPIW), North Pacific Deep Water (NPDW), as well as Equatorial Surface Water
 175 (ESW), South Pacific Subsurface Water (SPSSW) and South Pacific Intermediate Water (SPIW). In the
 176 upper ocean (0-300 m), we found that both NPTSSW and SPSSW exhibited high salinity characteristics.
 177 The salinity of NPTSSW was distributed between 34.66 and 35.01, while the salinity of SPSSW was
 178 distributed between 35.15 and 35.65. In addition, as the water depth increased, the temperature of
 179 NPTSSW and SPSSW decreased significantly, with NPTSSW dropping from 27.18°C to 16.21°C and
 180 SPSSW dropping from 29.23°C to 14.81°C . The representative water mass in the middle ocean (300-

181 1000 m) is NPIW, which is characterized by a rapid decrease in temperature (11.44-5.57°C) and a slight
 182 increase in salinity (~0.3) with increasing water depth. The representative water mass in the deep ocean
 183 (1000-2000 m) is NPDW, which has stable properties and slight changes in salinity and temperature.
 184 Notably, the water mass distribution at station E142-19 is quite special. Ranging from the subsurface to
 185 the deep layer, the water mass properties of this station are relatively stable, showing low-salinity and
 186 low-temperature characteristics. This is attributed to the intrusion of both North Pacific Intermediate
 187 Water (NPIW) and South Pacific Intermediate Water (SPIW) into the station in the mid-ocean region.
 188 Additionally, the station is situated within the MD upwelling area, where strong upwelling transports
 189 low-temperature, low-salinity North Pacific Deep Water (NPDW) from the bottom to the upper layer,
 190 enhancing seawater exchange. Consequently, the water at station E142-19 comprises a mixture of diverse
 191 water masses (MW).



192
 193 [Figure 2. Relationship between potential temperature \(\$\sigma_0\$ \) and salinity \(S\) at each sampling station. The](#)
 194 [water mass distribution is marked with a dotted line.—](#)

195 The study area is traversed by six major ocean currents: the South Equatorial Current (SEC), the North
196 Equatorial Current (NEC), the North Equatorial Undercurrent (NEUC), the Subtropical Countercurrent
197 (STCC), the Equatorial Undercurrent (EUC) and the North Equatorial Countercurrent (NECC). Among
198 them, the SEC flows from east to west along the equator and is characterized by high temperature and
199 low salinity, notably impacting station EQ-6. The NEC is a major westward current in the study area,
200 accompanied by a series of eastward undercurrents of NEUC in its lower part; stations E142-3 and E142-
201 7 are mainly affected by them. The STCC is characterized by a multi-eddy structure that flows eastward
202 in the subtropical region of the North Pacific and notably impacts station E142-11. The EUC is a strong
203 eastward current rich in oxygen and nutrients, which are present in the subsurface layer of the equatorial
204 Pacific, forming the main body of the thermocline of this area; station E142-13 is deeply affected by it.
205 The NECC is an important current in the tropical Pacific equatorial current system, transporting warm
206 pool water from the western Pacific to the eastern Pacific; Station E142-19 is mainly affected by it.
207 Furthermore, the area features a substantial upwelling system known as the Mindanao Dome (MD),
208 greatly impacting Station E142-19, situated southeast of the MD.

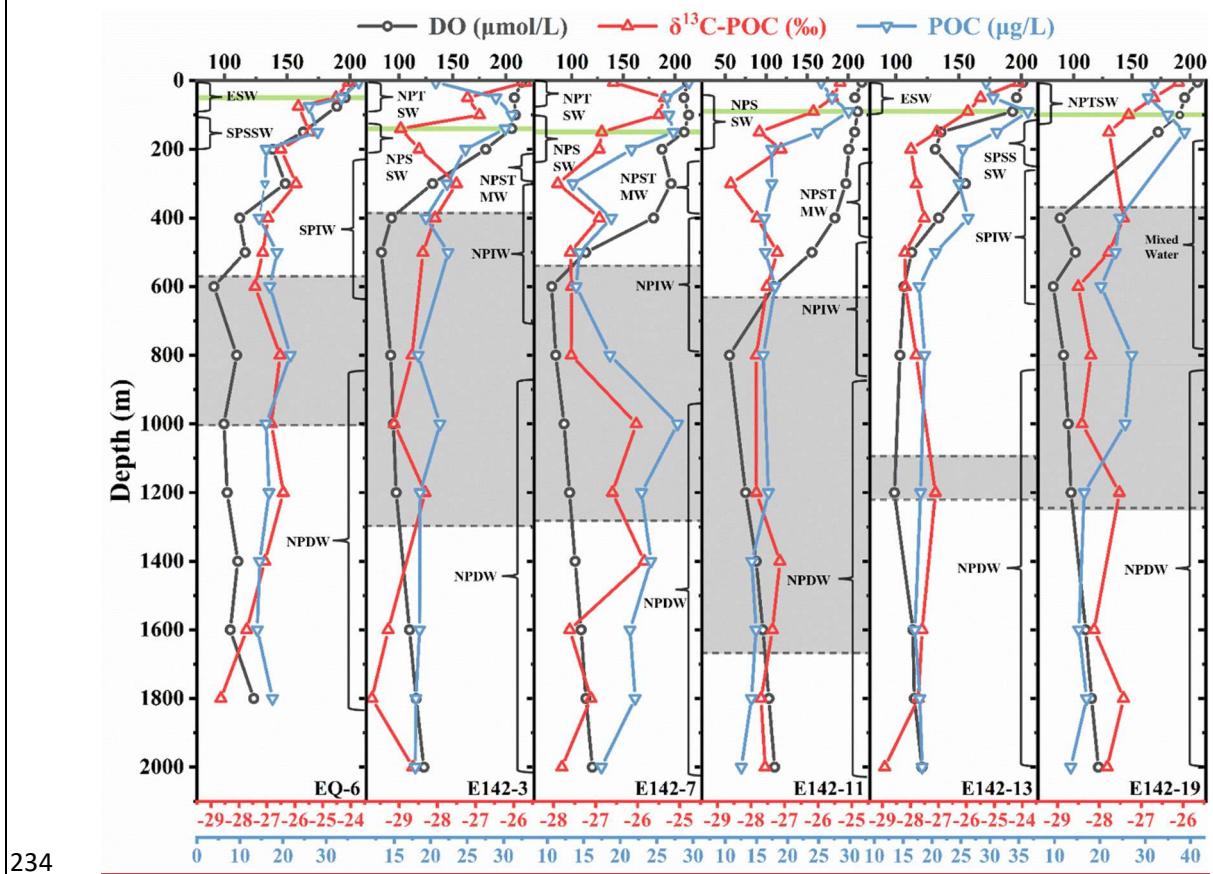
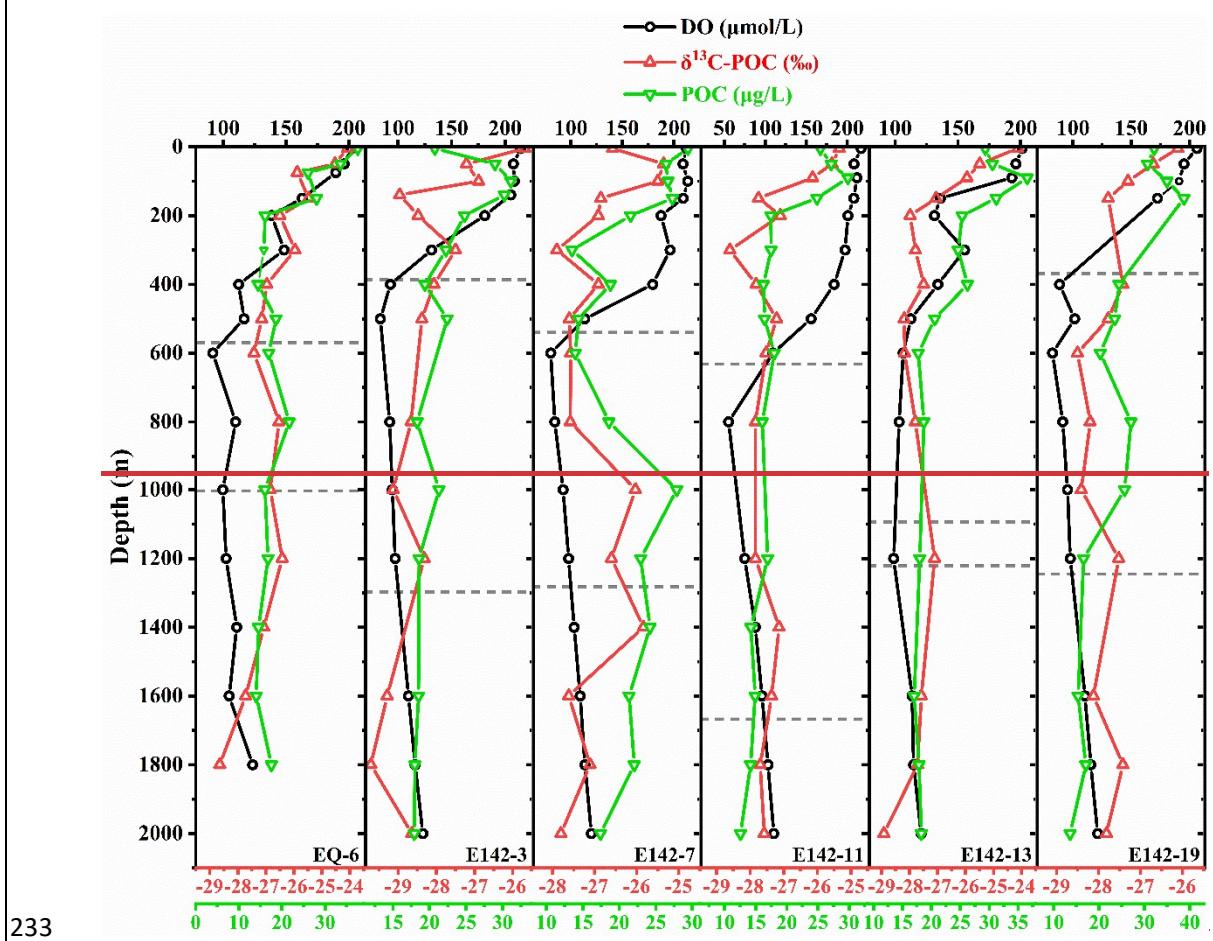
209 **3.2 Vertical distribution characteristics of POC and $\delta^{13}\text{C}$ -POC**

210 The average POC concentration from the surface to the deep chlorophyll maximum layer (DCM, 0-150
211 m) of the six stations was: E142-19 ($34.12 \pm 3.53 \text{ }\mu\text{g/L}$) > E142-13 ($31.90 \pm 3.19 \text{ }\mu\text{g/L}$) > EQ-6 (31.32
212 $\pm 5.27 \text{ }\mu\text{g/L}$) > E142-3 ($27.77 \pm 4.78 \text{ }\mu\text{g/L}$) > E142-7 ($27.43 \pm 1.35 \text{ }\mu\text{g/L}$) > E142-11 ($26.81 \pm 2.25 \text{ }\mu\text{g/L}$).

213 Since the nutrient concentration in ESW and SPSSW is higher than that in NPTSW and NPTSSW, The the
214 surface POC concentrations at stations E142-13 and EQ-6 were slightly higher than those at other stations.
215 However, the surface POC concentration at station E142-19 was the highest among the six stations
216 because the robust upwelling of MD brought rich nutrients to the surface seawater, alleviating the
217 nitrogen nutrient limitation of the surface water at this station (Gao et al., 2021).

218 The POC concentration of each station demonstrated a decreasing trend with increasing water depth and
219 tended to remain stable in the deep ocean ($> 1,000 \text{ m}$) (Fig. 2). The most significant drop in POC
220 concentration occurred between the DCM and 600 m (Fig. 3). The seawater within this depth range was
221 abundant in POC and also exhibited relatively high temperature and DO concentration, which likely
222 enhanced the metabolic activities of heterotrophic organisms, thereby accelerating their utilization of
223 POC (Iversen and Ploug, 2013; Sun et al., 2021). The aerobic degradation of POC led to a significant

224 consumption of DO. Therefore, the change in DO in this water layer was consistent with the change of
225 POC concentration (Fig. 2Fig. 3). It could be inferred that the rapid degradation of POC contributes to
226 the accelerated formation of the oxygen cline. Since the microbial life activities below the oxygen cline
227 were still active, leading to the continued consumption of DO through POC degradation, the DO could
228 not be replenished in time. As a result, the low oxygen zone (where DO < 100 $\mu\text{mol/L}$) emerged in the
229 middle ocean at all stations (Fig. 2Fig. 3). However, the hypoxic conditions observed at station E142-13
230 were comparatively less pronounced than those observed at other stations (Fig. 2Fig. 3). This can be
231 attributed to the consistent transport of oxygen and nutrient-rich seawater by the EUC to this station,
232 facilitating oxygen replenishment and mitigating deoxygenation (Brandt et al., 2021).



235 **Figure 2Figure 3.** Vertical distribution of DO concentration, $\delta^{13}\text{C}$ -POC values, and POC concentration at
236 each sampling station. The gray area dotted line marks the hypoxic zone with DO = 100 $\mu\text{mol/L}$ as the
237 boundary. The green line represents the DCM depth.

238 The vertical distribution of $\delta^{13}\text{C}$ -POC values closely resembles that of POC concentration (Figs. 2, 3a).

239 This similarity suggests that specific ^{13}C -enriched components may be preferentially degraded during
240 POC degradation. Although the molecular composition of oceanic POC cannot be fully identified, it is
241 generally understood to primarily consist of lipids, amino acids, carbohydrates, nucleic acids, and a small
242 number of heterogeneous components (Kharbush et al., 2020). The metabolic activity of amino acids and
243 carbohydrates is higher than lipids, leading microorganisms to preferentially use these compounds as
244 energy sources, enriching lipids in POC (Hwang et al., 2006; Jeffrey et al., 1983). Previous studies have

245 reported that during the degradation of POC, the carbon isotope fractionation characteristics of amino
246 sugar monomers closely align with changes in $\delta^{13}\text{C}$ -POC values (Guo et al., 2023b). Moreover, several

247 studies have highlighted that the carbon isotopic composition of lipid monomers does not exhibit
248 significant depletion during POC degradation; in fact, it may even show a trend of enrichment (Close et

249 al., 2014; Häggi et al., 2021). These observations further indicates the preferential degradation of amino
250 acids and carbohydrates in POC. On the other hand, compared with lipids, amino acids and carbohydrates

251 exhibit higher $\delta^{13}\text{C}$ values (Hayes, 1993; Hwang and Druffel, 2003; Schouten et al., 1998). When large
252 quantities of amino acids and carbohydrates undergo selective degradation, the residual POC will show

253 low $\delta^{13}\text{C}$ value characteristics. Therefore, as POC is continuously consumed in the water column, the
254 $\delta^{13}\text{C}$ -POC values will gradually decrease. In addition, lipids have a low nitrogen content in comparison

255 to amino acids and carbohydrates, leading to a relatively high C:N (Morales et al., 2021). Our findings
256 demonstrated a strong negative correlation between $\delta^{13}\text{C}$ -POC values and C:N (Fig. 3Fig. 4b), which

257 implied that as the water depth increases, $\delta^{13}\text{C}$ -POC values decreases while the C:N in the remaining
258 POC increases. This suggests that selective degradation of POC occurs in our study, during which amino

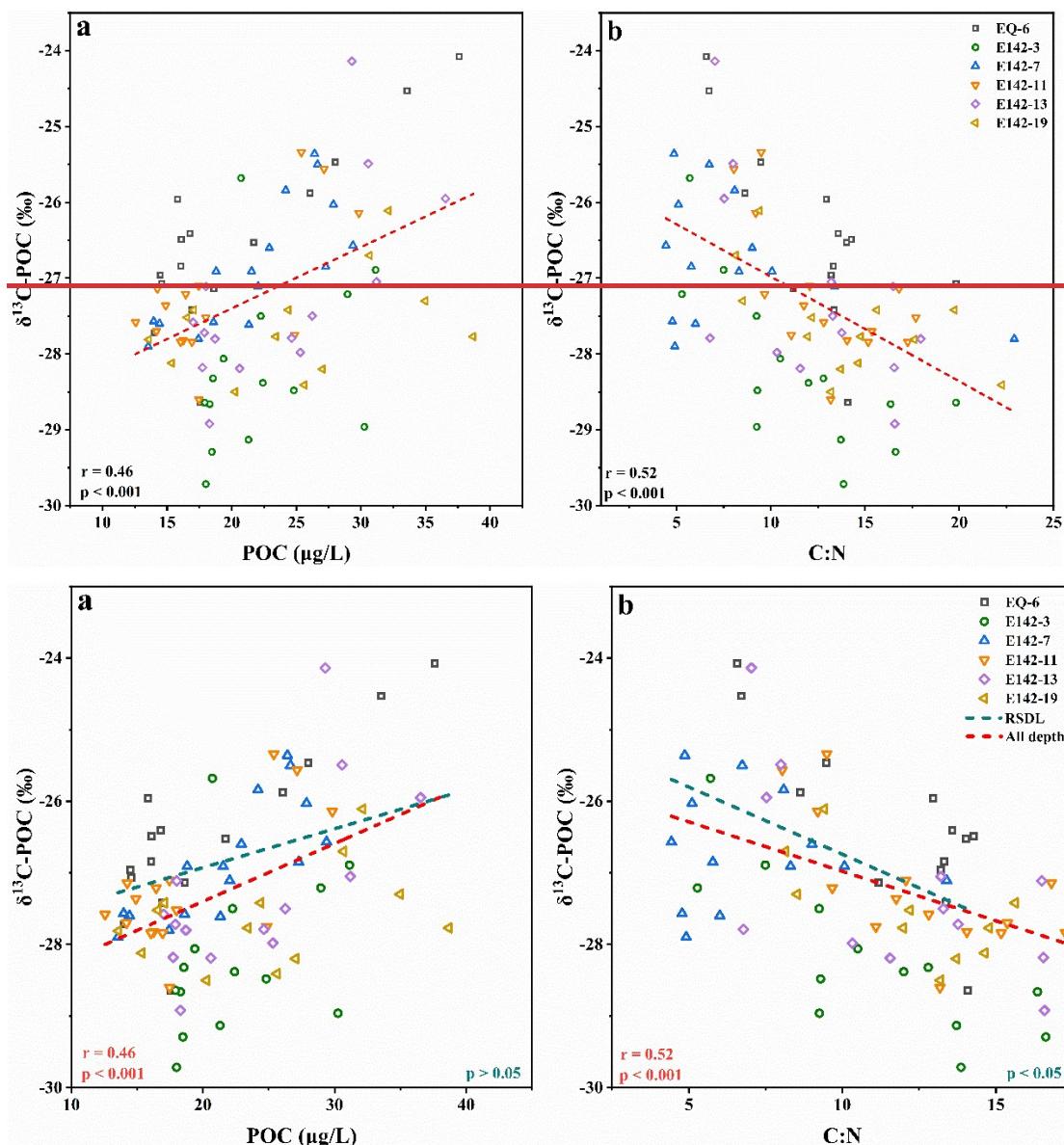
259 acids and carbohydrates in the POC were preferentially removed, resulting in a relative increase in the
260 proportion of lipids in the remaining POC (Druffel et al., 2003; Guo et al., 2023a). However, it is

261 noteworthy that in the upper ocean (0-300 m), although there is a significant negative correlation between
262 $\delta^{13}\text{C}$ -POC values and C:N ratios ($p < 0.05$), no significant correlation is observed between $\delta^{13}\text{C}$ -POC

263 values and POC concentration ($p > 0.05$) (Fig. 4a). This suggests that the fractionation of $\delta^{13}\text{C}$ -POC at

264 this depth layer is not entirely controlled by selective degradation. Photosynthesis exerts a certain

265 influence on the fractionation of $\delta^{13}\text{C}$ -POC within this depth range, primarily manifested as an increase
 266 in photosynthetic carbon isotope fractionation with depth, leading to a decrease in $\delta^{13}\text{C}$ -POC values.
 267 In a study conducted in the subtropical North Atlantic, the photosynthetic carbon isotope fractionation
 268 increased by 5.6‰ from the upper to the lower euphotic zone, while the $\delta^{13}\text{C}$ values of the
 269 photosynthetic product, phytol, decreased by 6.3‰ (Henderson et al., 2024). Therefore, although the
 270 process of selective degradation significantly affects the fractionation of $\delta^{13}\text{C}$ -POC, it is still necessary
 271 to consider the regulatory effects of other processes in certain unique marine environments.



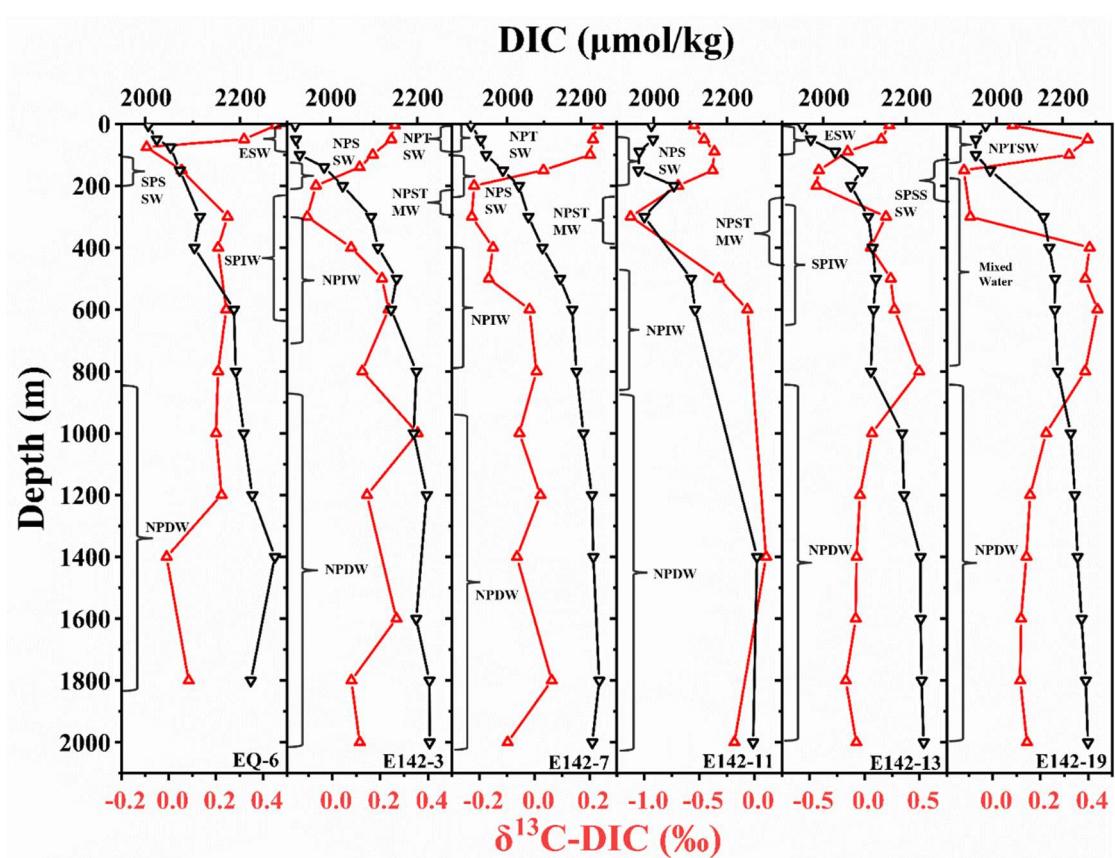
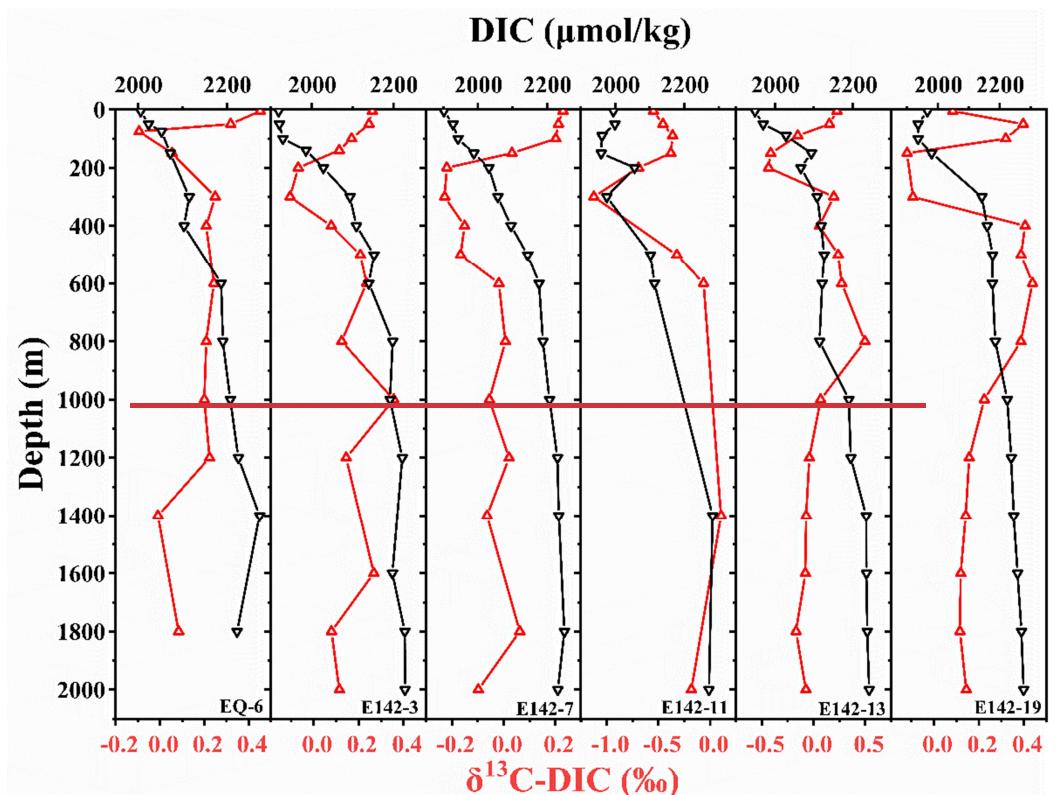
272
 273 **Figure 3Figure 4.** a. Relationship between $\delta^{13}\text{C}$ -POC values and POC concentration; b. Relationship between
 274 $\delta^{13}\text{C}$ -POC values and C:N
 275

276 **3.3 Vertical distribution characteristics of DIC and $\delta^{13}\text{C}$ -DIC**

277 Among the six stations, only the equatorial stations E143-13 and EQ-6 exhibited average upper DIC
278 concentrations exceeding 2000 $\mu\text{mol/kg}$, with values of 2036 and 2054 $\mu\text{mol/kg}$, respectively. This
279 phenomenon can be attributed to the fact that the surface water masses at these stations are composed of
280 high-temperature and high-salinity ESW (Fig. 2). Although high temperatures generally hinder the
281 dissolution of CO_2 , they can accelerate the rate of CO_2 release by heterotrophic organisms. Meanwhile,
282 high salinity increases the ionic strength and buffering capacity of seawater, promoting DIC
283 accumulation (Zeebe and Wolf-Gladrow, 2001). These factors collectively contribute to the high DIC
284 concentrations observed in the surface layers of these two stations. The average upper DIC concentration
285 at station E142-19 was the next highest, reaching 1992 $\mu\text{mol/kg}$. This is due to upwelling at this station,
286 which transports deep, high-DIC seawater to the middle ocean. Consequently, this station also recorded
287 the highest average mid-layer DIC concentration among the six stations, at 2184 $\mu\text{mol/kg}$. Furthermore,
288 since stations E142-3, E142-7, and E142-11 are predominantly influenced by the same water mass across
289 all depths, their DIC concentrations are relatively similar at each depth (Fig. 5). The average DIC
290 concentrations of all six stations in the upper ocean, middle ocean, and deep ocean were 2004 ± 65 , 2147
291 ± 35 , and $2234 \pm 26 \mu\text{mol/kg}$, respectively. There was a significant increase in DIC concentration from
292 the upper to the deep ocean (Fig. 4Fig. 5). Affected by photosynthesis, DIC increases gradually in the
293 upper ocean. In contrast, in the middle ocean, the rapid decomposition of POC released a large amount
294 of inorganic carbon, causing a rapid increase in DIC throughout the water column. Then, in the deep
295 ocean, a small amount of POC continues to degrade, while the release of DIC due to decreasing carbonate
296 saturation with depth contributes to a gradual increase in DIC concentration within this layer.~~in deeper~~
297 ~~layers, only a tiny amount of POC continued to degrade, so the DIC concentration of this layer increased~~
298 ~~slowly.~~

299 Moreover, we observed surface $\delta^{13}\text{C}$ -DIC values ranging from -0.55 to 0.45‰ (average 0.12‰) in the
300 research region, which is significantly lower than those reported in studies conducted in the Pacific region
301 in the 1990s (Quay et al., 2017; Quay and Stutsman, 2003). This suggests that the ocean has absorbed
302 more anthropogenic CO_2 as atmospheric CO_2 concentrations have increased over the years. The surface
303 $\delta^{13}\text{C}$ -DIC value of station E142-11 was the lowest among the six stations, only -0.55‰ , while the surface
304 $\delta^{13}\text{C}$ -DIC value of station EQ-6 was the highest among the six stations, reaching 0.45‰ . This is because

station E142-11 was located at the strongest atmospheric CO₂ net sink area, while station EQ-6 was located at the atmospheric CO₂ net source area (Zhong et al., 2022). The sea-air exchange at station E142-11 was sufficient, leading to a lower $\delta^{13}\text{C}$ -DIC value in its surface water, as it was more likely to reach isotopic equilibrium with atmospheric CO₂. In contrast, the surface water of station EQ-6 was more susceptible to seawater mixing and biological primary production influences. The higher $\delta^{13}\text{C}$ -DIC values observed in the surface water of station EQ-6 can be attributed to the isotope fractionation caused by the consumption of a substantial amount of CO₂ by biological primary production (Quay et al., 2003). In analyzing the vertical distribution of $\delta^{13}\text{C}$ -DIC, the findings revealed a rapid decrease in $\delta^{13}\text{C}$ -DIC values at each station, mirroring the decline seen in $\delta^{13}\text{C}$ -POC values in the upper ocean (0-300 m) (Figs. 4, 5d). Within this depth range, the average decrease in $\delta^{13}\text{C}$ -POC values was 2.23‰, while the average decrease of $\delta^{13}\text{C}$ -DIC values was 0.30‰, with $\delta^{13}\text{C}$ -DIC reaching its minimum value in the subsurface. However, in the middle ocean layer (300-1,000 m), unlike $\delta^{13}\text{C}$ -POC, $\delta^{13}\text{C}$ -DIC values increased first and then stabilized (Fig. 4Fig. 5). Therefore, distinct differences exist in the overall change trends of $\delta^{13}\text{C}$ -DIC values and $\delta^{13}\text{C}$ -POC values in the ocean water column. Since the mutual conversion between POC and DIC was ongoing, this conversion process will inevitably cause changes in $\delta^{13}\text{C}$ -POC and $\delta^{13}\text{C}$ -DIC. Generally, the variation range of $\delta^{13}\text{C}$ -POC values was more significant than that of $\delta^{13}\text{C}$ -DIC, indicating the more complex biogeochemical processes experienced by POC (Meyer et al., 2016; Schmittner et al., 2013). This difference is also partly due to the much larger size of the DIC pool compared to the POC pool (Jeffrey et al., 1983). The high DIC concentration in the ocean buffers its isotopic variability, resulting in minimal changes in $\delta^{13}\text{C}$ -DIC values across the water column, whereas the smaller POC pool is more sensitive to localized biogeochemical processes, leading to greater variability in $\delta^{13}\text{C}$ -POC values.

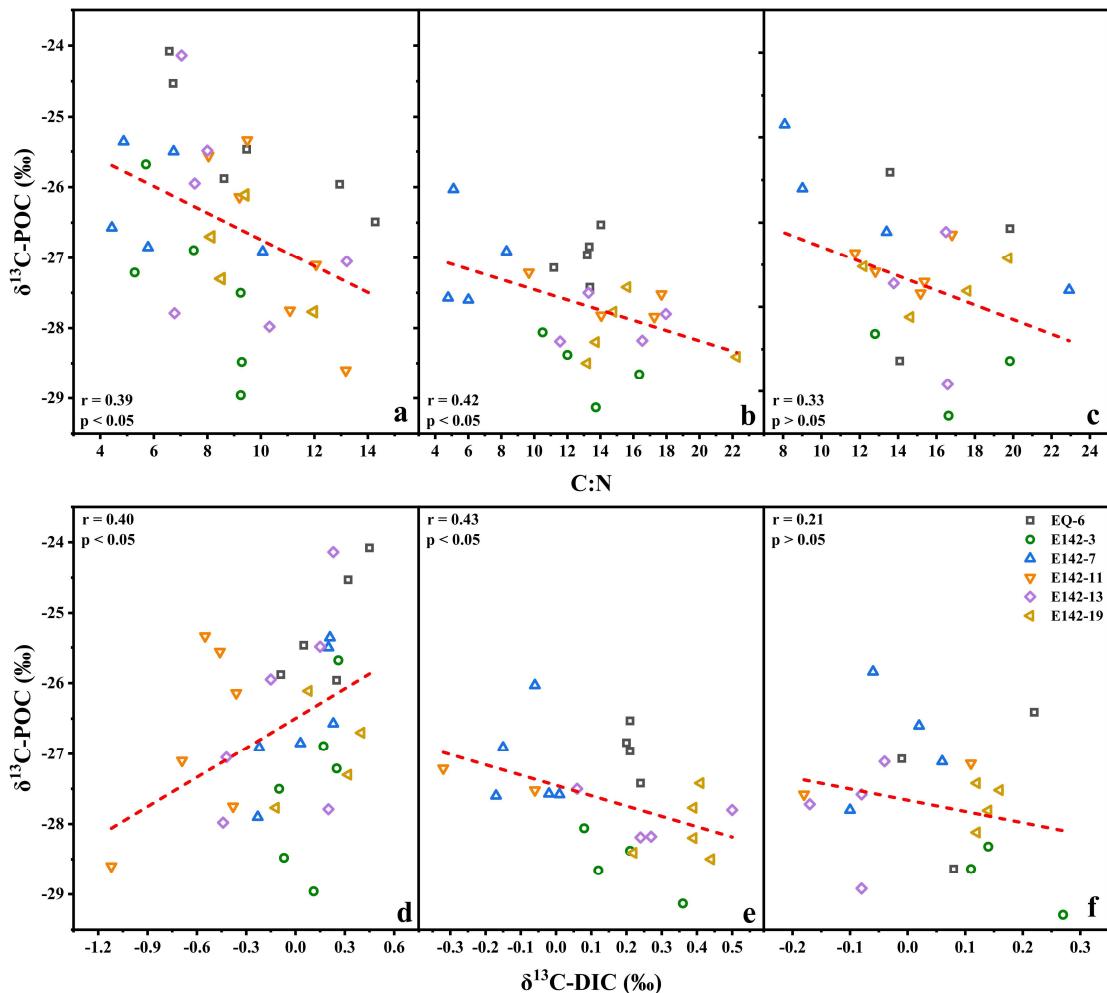


329 **Figure 4****Figure 5**. Vertical distribution of DIC concentration and $\delta^{13}\text{C-DIC}$ values at each sampling station.
330 The black line represents DIC, and the red line represents $\delta^{13}\text{C-DIC}$ values.

331 **3.4 Transformation characteristics of POC in different water layers**

332 According to the distribution characteristics of $\delta^{13}\text{C}$ -POC and $\delta^{13}\text{C}$ -DIC values, we divided the ocean
333 water column into three biogeochemical layers: the POC rapid synthesis-degradation layer (RSDL, 0-
334 300 m), the net degradation layer (NDL, 300-1,000 m) and the stable layer (SL, 1,000-2,000 m). Within
335 the RSDL, POC was rapidly degraded while being synthesized. The synthesis of POC likely exceeded
336 its degradation from the surface to the DCM layer, while the degradation of POC appeared to dominate
337 below the DCM. The synthesis rate was greater than the degradation rate from the surface to the DCM
338 layer, while the degradation rate was greater than the synthesis rate below the DCM, reflecting the rapid
339 decrease in photosynthetic rate with depth. In this layer, In addition, the $\delta^{13}\text{C}$ -POC values and C:N in this
340 layer exhibited a pronounced negative correlation, while no significant correlation is observed between
341 $\delta^{13}\text{C}$ -POC values and POC concentration ($p > 0.05$) (Fig. 5Figs. 4a, 6a). Therefore, the rapid decrease of
342 $\delta^{13}\text{C}$ -POC values in this layer was dominated by the selective degradation of POC and
343 photosynthesis amino acids and carbohydrates. Both $\delta^{13}\text{C}$ -POC and $\delta^{13}\text{C}$ -DIC values decrease with
344 increasing depth in the RSDL (Figs. 3, 5), and they exhibit a significant positive correlation within this
345 layer (Fig. 6d). Although the degradation of POC typically lowers the $\delta^{13}\text{C}$ value of DIC, as the $\delta^{13}\text{C}$
346 value of POC is lower than that of DIC, the significant decline in $\delta^{13}\text{C}$ -DIC values observed in the RSDL,
347 when considering the substantial difference in magnitude between the POC pool and the DIC pool,
348 suggests the influence of additional processes. However, at the same time, $\delta^{13}\text{C}$ -POC and $\delta^{13}\text{C}$ -DIC
349 showed a significant positive correlation in this layer (Fig. 5d). Supposing that POC underwent selective
350 degradation, the resulting DIC should exhibit an enrichment in $\delta^{13}\text{C}$. However, contrary to expectations,
351 our findings indicate a decline in $\delta^{13}\text{C}$ DIC within the RSDL. This perplexing occurrence can be
352 attributed to two primary reasons. Specifically, On the one hand, the phytoplankton and photosynthetic
353 bacteria in the upper ocean tended to use the light $^{12}\text{CO}_2$ in the seawater for photosynthesis; thus the
354 $\delta^{13}\text{C}$ -DIC values of the surface ocean at all stations was relatively high. However, light intensity
355 diminishes with increasing depth, which is unfavorable for photosynthesis. This leads to the
356 accumulation of $^{12}\text{CO}_2$ produced by the respiration of heterotrophic communities. However, light
357 intensity diminished with depth increases, causing the photosynthesis rate to slow. Meanwhile, the
358 respiration rate of the biological community was still very fast, resulting in the accumulation of light
359 $^{12}\text{CO}_2$. Consequently, the $\delta^{13}\text{C}$ -DIC values in this layer steadily declined (Ge et al., 2022). In the NDL,

360 sunlight was extremely weak, and photosynthesis was nearly absent. Heterotrophic communities
361 dominate, leading to a continuous decrease in POC concentration and a corresponding increase in DIC
362 concentration (Figs. 3, 5). Generally, the degradation of POC would be expected to lower the $\delta^{13}\text{C}$ value
363 of DIC. However, in this layer, $\delta^{13}\text{C}$ -POC values showed a significant negative correlation with both
364 C:N and $\delta^{13}\text{C}$ -DIC values (Fig. 6b, e), indicating the influence of additional processes on $\delta^{13}\text{C}$ -DIC
365 fractionation. The NDL often encompasses low-oxygen zones (Fig. 3), which are known to favor the
366 activity of chemoautotrophic microorganisms. Compared to aerobic environments, the energy required
367 for microorganisms to fix inorganic carbon into organic carbon is lower under low-oxygen condition
368 (Hugler and Sievert, 2011; Mccollom and Amend, 2005). During this process, chemoautotrophic
369 microorganisms preferentially utilize lighter ^{12}C isotopes, leading to the enrichment of ^{13}C in the
370 remaining DIC pool. This microbial activity explains the observed increase in $\delta^{13}\text{C}$ -DIC values in the
371 NDL. In the NDL, the sunlight was pretty weak, and there was almost no photosynthesis. The rate of
372 chemosynthesis of organic carbon was lower than the degradation rate of POC, causing the concentration
373 of POC to continue decreasing. Additionally, the $\delta^{13}\text{C}$ -POC in this layer showed a significant negative
374 correlation with both C:N and $\delta^{13}\text{C}$ -DIC (Fig. 5b, e), suggesting a very active mutual conversion process
375 between POC and DIC. The large amount of selective degradation of amino acids and carbohydrate POC
376 caused the $\delta^{13}\text{C}$ -DIC in this layer to continue to increase. In the SL, the POC concentration remained
377 consistently low. $\delta^{13}\text{C}$ -POC values did not correlate significantly with either C:N or $\delta^{13}\text{C}$ -DIC (Fig. 5Fig.
378 6c, f). This was because the easily degradable components in POC had been completely consumed in the
379 RSDL and NDL, and the remaining components were relatively refractory. As a result, the conversion
380 of POC to DIC was rare in SL, leading to an absence of a clear link between $\delta^{13}\text{C}$ -POC and $\delta^{13}\text{C}$ -DIC.



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Figure 5 **Figure 6.** Relationships between $\delta^{13}\text{C-POC}$ and C:N at different depths: (a) 0-300 m, (b) 300-1,000 m, (c) 1,000-2,000 m, and between $\delta^{13}\text{C-POC}$ and $\delta^{13}\text{C-DIC}$ at different depths: (d) 0-300 m, (e) 300-1,000 m, (f) 1,000-2,000 m.

385 4 Conclusions

386 In general, this study investigated the transformation characteristics of POC in the tropical northwest
 387 Pacific Ocean based on the $\delta^{13}\text{C}$ perspective. Our findings revealed three distinct stages of POC behavior
 388 in the ocean: rapid synthesis-degradation, net degradation, and stable existence. Below the RSDL, the
 389 selective degradation of POC dominated the changes in $\delta^{13}\text{C-POC}$. The C:N ratio data in RSDL and NDL
 390 indicate an increase in the proportion of refractory lipids in POC, relative to more labile components
 391 such as amino acids and carbohydrates. Following vigorous selective degradation in the RSDL and NDL,
 392 an increase in the proportion of refractory lipids in POC was observed. Consequently, in the SL, POC
 393 was found to be stable with a slow degradation rate. The fractionation of $\delta^{13}\text{C-DIC}$ in the ocean is
 394 influenced by both the production and degradation processes of POC. Within the RSDL, $\delta^{13}\text{C-DIC}$

395 fractionation is predominantly governed by primary production, whereas within the NDL and SL, it is
396 primarily influenced by the degradation process of POC.

397 Although we utilized $\delta^{13}\text{C}$ -POC and $\delta^{13}\text{C}$ -DIC to assess the overall transformation characteristics of POC,
398 the specific synthesis and decomposition ratios of POC are still challenging to determine. Further
399 research is needed on the monomer carbon isotopic composition of POC (lipids, amino acids, etc.) to
400 enhance our understanding of the transformation process of POC.

401 **Data Availability.** The data files used in this paper are available at (Tian et al., 2024).

402 **Competing interest.** The authors declare that they have no conflict of interest.

403 **Author contribution.** Detong Tian: Investigation, Data Curation, Writing-original draft. Xuegang Li
404 and Jinming Song: Conceptualization, Funding acquisition, Writing-review & editing. Jun Ma, Funding
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