

Contribution of sulfate to aerobic methane oxidation in upland soils: a mini-review

Rui Su¹, Kexin Li^{2,3}, Nannan Wang², Fenghui Yuan², Ying Zhao¹, Yunjiang Zuo², Ying Sun², Liyuan He⁴, [Lixin Yang⁵](#), Xiaofeng Xu^{4*}, Lihua Zhang^{1,2,5*}

¹College of Life and Environmental Sciences, Minzu University of China, Beijing, 100081, China

²Key Laboratory of Wetland Ecology and Environment, Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences, Changchun, Jilin, 130102, China

³University of Chinese Academy of Sciences, Beijing, 100049, China

⁴Biology Department, San Diego State University, San Diego, CA, 92182, USA

⁵[China Center for Commercial Science, Technology and Quality, Beijing, 100801, China](#)

[State Key Laboratory of Vegetation and Environmental Change, Institute of Botany, Chinese Academy of Sciences, Beijing 100093, China](#)

Corresponding authors: L.Z. (zhanglihua@muc.edu.cn); X.X. (xxu@sdsu.edu)

Abstract. Methane (CH₄) is a potent greenhouse gas, and its global warming potential is 28 times higher than carbon dioxide (CO₂). Various environmental factors influence aerobic CH₄ oxidation in soil. Sulfate (SO₄²⁻) ion is the main component of atmospheric deposition and has been increasing in recent years. It promotes CH₄ production and anaerobic CH₄ oxidation, however, the impact of SO₄²⁻ on aerobic CH₄ oxidation in soils has not yet been comprehensively summarized. We synthesize current research on the effects of SO₄²⁻ on aerobic CH₄ oxidation, examining both its macroscopic manifestations and microscale pathways. Through a literature review, we found that SO₄²⁻ enhances aerobic CH₄ oxidation by 0–42%, moreover, it has been found that various physicochemical properties and processes in the soil are influenced by the addition of SO₄²⁻, which in turn affects aerobic CH₄ oxidation. [This review enhances our understanding of the role of SO₄²⁻ in promoting aerobic CH₄ oxidation. It lays the foundation for future research with two primary goals: \(1\) validating these findings by quantifying CH₄ flux and aerobic oxidation rates, and \(2\) elucidating the underlying](#)

删除了：State Key Laboratory of Vegetation and Environmental Change, Institute of Botany, Chinese Academy of Sciences, Beijing 100093, China

设置了格式：字体：非加粗

删除了：5

删除了：up to

microbial processes through experimental research. Concurrently, the review provides directions for further investigation into the impact of SO_4^{2-} on aerobic CH_4 oxidation.

1 Introduction

CH_4 is an important greenhouse gas, and its atmospheric concentration has increased since pre-industrial times (Place, 2024; Praeg et al., 2016). Its global warming potential is 28 times higher than carbon dioxide (CO_2), owing to its superior heat absorption efficiency (IPCC, 2013). Methanotrophs (aerobic methanotrophs) consume CH_4 under certain conditions (Le Mer and Roger, 2001), reducing CH_4 atmospheric concentration (Singh et al., 2010). Consequently, methanotrophs are crucial microbes that play an indispensable role in regulating and mitigating the CH_4 -related greenhouse effect on Earth. Soil aerobic CH_4 oxidation is the sole known biological sink for atmospheric CH_4 (Ho et al., 2019; Murguia-Flores et al., 2018), contributing to 5%–7% of the global annual atmospheric CH_4 uptake (Saunois et al., 2020). Upland soils are the primary biological CH_4 sink (Bodelier, 2011; Guo et al., 2023), owing to methanotroph-mediated CH_4 consumption (Song et al., 2024). This represents the second-largest atmospheric CH_4 consumption sink, surpassed only by hydroxyl radical depletion (Deng et al., 2019). Aerobic CH_4 oxidation in soils are influenced by many factors, such as soil water content, soil texture, soil type, temperature, soil pH, soil inorganic nitrogen content, metal availability, etc., many of these factors have been extensively reviewed (Shukla et al., 2013; Mishra et al., 2018). However, the effect of SO_4^{2-} , a significant ion component of acid deposition, on aerobic CH_4 oxidation has not yet been reviewed.

Acid rain, involving deposition of SO_4^{2-} and other acidic compounds, remains a globally significant environmental issue (Chen et al., 2020; Qi et al., 2022). The three largest affected regions are Europe, North America, and China (Li et al., 2021). SO_4^{2-} is the major ion in acid rain (Wright and Henriksen, 1978) and has profound impacts on substances and biochemical processes in soils. As a crucial component of terrestrial ecosystems, soils serve as the ultimate receptor of acid deposition. SO_4^{2-} deposition

删除了：This review enhances our understanding of the role of SO_4^{2-} in promoting aerobic CH_4 oxidation and lays the foundation for future studies aimed at validating these findings by quantifying CH_4 flux and oxidation rates, as well as elucidating the underlying microbial processes through experimental research, while also providing directions for further investigation of SO_4^{2-} 's impact on aerobic CH_4 oxidation.

设置了格式：下标

删除了：characterized by the deposition of

删除了：has been

删除了：significant environmental issue

induces soil acidification (Huang et al., 2019), alters soil plant diversity (Li et al., 2022), affects microbial properties (Wang et al., 2018), and limits grass yield potential (Klessa et al., 1989), as well as a reduction in the activities of soil enzymes such as cellulase, invertase, and polyphenol oxidase (Tie et al., 2020). SO_4^{2-} can inhibit CH_4 production (methanogenesis) and promote anaerobic CH_4 oxidation, playing a crucial role in anaerobic CH_4 biogeochemical processes, SO_4^{2-} suppresses methanogenesis primarily due to its thermodynamic and kinetic preference as an electron acceptor (Granberg et al., 2001; Schimel, 2004), leading to decreased CH_4 emissions (Gauci et al., 2004). SO_4^{2-} has been shown to facilitate anaerobic CH_4 oxidation by anaerobic methanotrophic archaea in diverse ecosystems, such as oceans (Boetius et al., 2000), wetlands (La et al., 2022), and paddy fields (Fan et al., 2021), acting as a crucial electron acceptor. Despite these well-documented effects on anaerobic CH_4 biogeochemical processes, the influence of SO_4^{2-} on aerobic CH_4 oxidation, particularly in upland soils, remains underexplored. Given the increasing global deposition of SO_4^{2-} due to industrial activities, understanding its impact on aerobic CH_4 oxidation is essential for predicting future CH_4 dynamics and developing effective climate mitigation strategies.

删除了: the CH_4 cycle

删除了: anaerobic CH_4 cycling

In this review, we have analyzed the literature on the effects of SO_4^{2-} on aerobic CH_4 oxidation. Our analysis not only reveals evidence suggesting that SO_4^{2-} promotes aerobic CH_4 oxidation but also identifies supporting evidence from related studies. In this review, we reviewed references about the influence of SO_4^{2-} on soil properties, substances, or biochemical processes, aiming to elucidate any microscale pathways on aerobic CH_4 oxidation through variations in soil substances or processes. Our analysis reveals that SO_4^{2-} may affect aerobic CH_4 oxidation. Based on the available literature, 3 out of 5 studies that investigated the influence of SO_4^{2-} on aerobic CH_4 oxidation were able to demonstrate a positive effect on aerobic CH_4 oxidation, we infer that SO_4^{2-} favors aerobic CH_4 oxidation. This review summarizes the microscale pathways by which SO_4^{2-} influences aerobic CH_4 oxidation and highlights the importance of future research in this area. By providing a comprehensive synthesis of existing knowledge, this work serves as a valuable reference for future experimental studies. Furthermore,

删除了: Based on the available literature

the findings of this review will contribute to a deeper understanding of global CH₄ cycling, particularly in the context of increasing SO₄²⁻ deposition. Moving forward, we aim to experimentally validate the impact of aerobic CH₄ oxidation following SO₄²⁻ addition and elucidate the underlying microbial mechanisms involved.

2 The microbial aerobic CH₄ oxidation processes

2.1 Aerobic CH₄ oxidation processes

Aerobic CH₄ oxidation is mediated by methanotrophs, a group of specialized microorganisms (Chistoserdova et al., 2005). In soils, aerobic CH₄ oxidation can be classified into two distinct forms based on the concentration of CH₄ (Walsh et al., 2009). The first form, known as high-affinity oxidation, occurs at CH₄ concentrations close to atmospheric levels (<2ppm) and is carried out by high-affinity methanotrophs (Chowdhury and Dick, 2013). This process is commonly observed in upland soils, particularly in environments with high NH₄⁺ concentrations (Ho et al., 2019; Le Mer and Rob  rt, 2001). The second form, referred to as low-affinity oxidation, occurs at CH₄ concentrations exceeding 40 ppm and is mediated by low-affinity methanotrophs (Chowdhury and Dick, 2013). This form is typically found in wetland environments, where CH₄ concentrations are significantly higher than atmospheric levels (Bechtold et al., 2025). Aerobic CH₄ oxidation converts CH₄ to CO₂ in four steps:   MMO oxidizes CH₄ to methanol (CH₃OH),   methanol dehydrogenase (MDH) oxidizes CH₃OH to formaldehyde (HCHO),   FADH oxidizes HCHO to formate (HCOOH),   formate dehydrogenase (FDH) oxidizes HCOOH to CO₂ (Fig. 1, paths   1  4) (Mancinelli, 1995).

2.2 Methanotrophs

Methanotrophs constitute a distinct subset of methylotrophs, primarily dependent on the one-carbon compound CH₄ as their sole source of carbon and energy (Hanson and Hanson, 1996). In the traditional classification system, Proteobacterial methanotrophs were categorized into type I (*Methylococcaceae* and *Crenotrichaceae*), type II

设置了格式: 字体: 加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 下标

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 下标

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 下标

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 下标

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

删除了: Aerobic CH₄ oxidation processes can be further

classified into assimilatory and dissimilatory pathways. In the dissimilatory pathways, CH₄ is sequentially oxidized to CO₂ by multiple enzymes (Fig. 1  2). (Mancinelli, 1995). In the assimilation pathways, methanotrophs convert formaldehyde, an intermediate product of aerobic CH₄ oxidation, into biomass and other organic compounds mainly through the ribulose monophosphate pathway (RuMP pathway) (Fig. 1  3), serine pathway (Fig. 1  4), and xylulose monophosphate pathway (XyMP pathway) (Fig. 1  1)(Yang et al., 2023).

(*Methylocystaceae* and *Beijerinckiaceae*), and type X (*Methylococcaceae*) (Li et al., 2020) based on their cell membrane arrangement, chemotaxonomic properties, physiological characteristics, and phylogenetic locations. However, due to the discovery of non-canonical methanotrophs, the traditional classification system has become outdated. Consequently, methanotrophs are now classified into seven categories based on phylogenetic analysis: Type I-A (*Methylomonadaceae*), I-B (*Methylococcaceae*), I-C (*Methylothermaceae*), I-D (*Crenotrichaceae*), II-A (*Methylocystaceae*), II-B (*Beijerinckiaceae*), III (*Methylacidiphilaceae*), and NC10 (Fenibo et al., 2023). *Methylomonadaceae*, *Methylococcaceae*, *Methylothermaceae*, and *Crenotrichaceae* belong to the class *Gammaproteobacteria*, while *Methylocystaceae* and *Beijerinckiaceae* are classified under *Alphaproteobacteria*. *Methylacidiphilaceae* belongs to the phylum *Verrucomicrobia*. The composition of different types of methanotrophs is shown in Figure 1 (Fenibo et al., 2023). Notably, only four genera—*Methylocella*, *Methyacidimicrobium*, *Methylacidiphilum*, and *Methanomirabilis*—are capable of carbon fixation via the Calvin-Benson-Bassham (CBB) cycle (Fenibo et al., 2023; Op den Camp et al., 2009). Among Actinobacterial methanotrophs, *Candidatus Mycobacterium methanotrophicum* is classified with the *Mycobacterium* genus (van Spanning et al., 2022). Methanotrophs utilize two forms of methane monooxygenase (MMOs): soluble cytoplasmic monooxygenase (sMMO) and particulate membrane-bound monooxygenase (pMMO). Except for *Methylocella silvestris* and *Methyloferula stellata*, all methanotrophs possess pMMO. sMMO has only been detected in a few specific genera, namely *Methylomonas* sp., *Methylomicrobium* sp., *Methylosinus* sp., and *Methylococcus capsulatus* (DiSpirito et al., 2016). Copper (Cu) concentration differentially regulates MMO expression (Fig. 1⑤): high Cu concentrations induces pMMO (Fig. 1⑥), whereas low Cu concentrations triggers sMMO (Fig. 1⑦) (Hakemian & Rosenzweig, 2007).

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

删除了: The expression of these enzymes is regulated by copper (Cu) concentration (Hakemian and Rosenzweig, 2007)....

3 Soil CH₄ oxidation in response to SO₄²⁻ addition

Sulfates, including SO₄²⁻ and sulfuric acid (H₂SO₄), enhance aerobic CH₄ oxidation

within a range of 0–42% (Table 1), ~~so~~ we hypothesize that SO_4^{2-} may stimulate aerobic
185 CH_4 oxidation. For example, in a temperate mixed deciduous woodland, the cumulative
uptake of aerobic CH_4 oxidation was 25% higher in the experimental group with H_2SO_4
addition compared to the control group during the final quarter of the study period
(Bradford et al., 2001b). Similar results were reported by Sitaula et al. (1995). In
190 another study, King and Schell (1998) found that adding SO_4^{2-} (Na_2SO_4) increased
aerobic CH_4 oxidation by 3% at a CH_4 concentration of 250 ppm compared to the
control group, although this result was not statistically significant. The lack of
significance may be attributed to the insufficient concentration gradient of Na_2SO_4 in
the experimental setup, which limited the ability to fully assess the effects of SO_4^{2-} on
aerobic CH_4 oxidation. Therefore, we propose that the observed enhancement of
195 aerobic CH_4 oxidation following H_2SO_4 addition is primarily due to the increase in
 SO_4^{2-} concentration.

删除了: thus,

The promotional effect of SO_4^{2-} on aerobic CH_4 oxidation is further supported by
comparisons with other anions under similar cationic conditions. Benstead and King
200 (2001) observed that HNO_3 exerted a stronger inhibitory effect on aerobic CH_4
oxidation under equivalent soil acidic conditions than H_2SO_4 . This finding is consistent
with the results of Bradford et al. (2001a), who experimentally confirmed the inhibitory
effect of nitrate (NO_3^-) on aerobic CH_4 oxidation (Dunfield and Knowles, 1995; Wang
and Ineson, 2003). When H_2SO_4 and HNO_3 were added to the soil to achieve H^+
205 concentrations of 10 and 1 $\mu\text{mol H}^+$ per gram of fresh weight (gfw), respectively, both
acids inhibited aerobic CH_4 oxidation to a similar extent. However, H_2SO_4 exhibited a
lesser inhibitory effect than HNO_3 . We hypothesize that SO_4^{2-} may promote aerobic
 CH_4 oxidation, as evidenced by the findings of Benstead and King (2001) and Bradford
et al. (2001a).

设置了格式: 下标

210 However, not all studies support the hypothesis that SO_4^{2-} promotes aerobic CH_4
oxidation. For instance, Bradford et al. (2001a) observed no significant difference in
aerobic CH_4 oxidation between low (564 μM) and high (1408 μM) concentrations of

删除了: Consequently, when H_2SO_4 and HNO_3 are added to the soil, resulting in equivalent acidic conditions, the inhibitory effect of H_2SO_4 is less pronounced than that of HNO_3 .

H₂SO₄ compared to the control group. This discrepancy may be due to differences in H₂SO₄ concentration across studies. Similarly, Hu et al. (2018) reported no significant effect of SO₄²⁻ on aerobic CH₄ oxidation. Based on the available evidence, SO₄²⁻ promotes aerobic CH₄ oxidation within a range of 0–42%. Although the mechanisms by which SO₄²⁻ influences aerobic CH₄ oxidation are not yet fully understood, we have identified potential microscopic pathways through which SO₄²⁻ may affect this aerobic process by reviewing relevant literature.

4 Microscale pathways by which SO₄²⁻ addition influences aerobic CH₄ oxidation

SO₄²⁻'s impact on aerobic CH₄ oxidation—particularly its mechanisms for enhancement—remains unclear. Our literature review reveals two promotion pathways: Shifts in methanotroph activity and community structure (Fig. 2 path d) (Bradford et al., 2001b; Sitaula et al., 1995). Alterations to soil physicochemical properties (Fan et al., 2017), substrate availability (Bjorneras et al., 2019; Palmer et al., 2013; Xu et al., 2017), and nutrient dynamics (Islam, 2012) (Fig. 2).

First, the addition of SO₄²⁻ alters soil physicochemical properties (Fig. 2 path a), i.e., particularly by reducing soil pH (Fig. 2 ①). Soil acidification increases due to enhanced base cation leaching associated with SO₄²⁻ addition (Hu et al., 2013), leading to a decrease in the pH of forest soils (Fasth et al., 1991; Tie et al., 2020). The addition of H₂SO₄ has been shown to promote aerobic CH₄ oxidation by altering the activity or community structure of methanotrophs (Bradford et al., 2001b; Sitaula et al., 1995). However, in experiments involving H₂SO₄ addition, it remains unclear whether the observed enhancement in aerobic CH₄ oxidation is primarily due to the decreased pH (Fig. 2 path e) or the increase in SO₄²⁻ concentration (Fig. 2 path d). Generally, CH₄ consumption is greater at higher pH conditions in forest soils (Brumme and Borken,

删除了：At present, the impact of SO₄²⁻ on aerobic CH₄ oxidation is not fully understood, and its role in this process remains unclear, particularly regarding its potential enhancing effects. To address this knowledge gap, we conducted a literature review focusing on studies involving the addition of SO₄²⁻. Through this review, we have identified several potential microscopic pathways through which SO₄²⁻ may promote aerobic CH₄ oxidation. One possible pathway involves changes in methanotroph activity and community structure in response to SO₄²⁻ (Fig. 2 path d) (Bradford et al., 2001b; Sitaula et al., 1995). Alternatively, SO₄²⁻ may affect aerobic CH₄ oxidation by altering soil physical properties (Fan et al., 2017), microbial substrates availability (Bjorneras et al., 2019; Palmer et al., 2013; Xu et al., 2017), and soil nutrition content (Islam, 2012) (Fig. 2).

设置了格式：字体：非加粗

设置了格式：字体：非加粗

设置了格式：字体：非加粗

删除了：al

1999; Silver et al., 1999), therefore, the reduction in soil pH may lead to a decrease in aerobic CH₄ oxidation. However, in acidic soils, a decrease in pH has been shown to increase aerobic CH₄ oxidation (Sitaula et al., 1995). Consequently, when evaluating the impact of SO₄²⁻ addition on aerobic CH₄ oxidation, it is essential to consider the initial soil pH (Fig. 2 path e), as methanotrophs exhibit different pH preferences in acidic and alkaline environments (Shukla et al., 2013).

Second, SO₄²⁻ addition can alter the soil microbial substrate (Fig. 2 path b), particularly by decreasing soil di-O-alkyl C content (Fig. 2 ②) (Xu et al., 2017). In a subtropical forest, SO₄²⁻ addition has been shown to increase the activity of gram-negative bacteria in soil by reducing the litter di-O-alkyl carbon (di-O-alkyl C) (Fig. 2 ② and path g) (Xu et al., 2017). Di-O-alkyl C is a component of soil organic carbon (SOC). SOC degradation is accelerated when the percentage of di-O-alkyl C is high (Huang et al., 2021). Conversely, when the content of di-O-alkyl C is low, SOC degradation slows down, leading to a greater availability of substrates for microorganisms, including methanotrophs. Methanotrophs, which are gram-negative bacteria (Schimel and Gullledge, 1998), may exhibit increased activity in response to SO₄²⁻ addition. This enhancement of methanotrophs activity (Fig. 2 path h) can ultimately promote aerobic CH₄ oxidation (Fig. 2 path o).

Third, SO₄²⁻ can alter soil nutrition content (Fig. 2 path c), specifically increasing soil Cu availability (Fig. 2 ③) (Islam, 2012), phosphorus (P) content (Fig. 2 ④) by enhancing acid phosphatase activity (Lv et al., 2014; Veraart et al., 2015), (aluminum ion) Al³⁺ toxicity (Fig. 2 ⑤) (Hu et al., 2013; Sogn and Abrahamsen, 1998), and NH₄⁺ absorption (Bradford et al., 2001b; Gullledge and Schimel, 1998; King and Schnell, 1998) (Fig. 2 ⑥). Cu is a crucial component in aerobic CH₄ oxidation processes, with its critical role stemming from its high abundance in catalytically active pMMO complexes—where it directly participates in CH₄ oxidation and facilitates electron transfer from endogenous reductants to molecular oxygen (Balasubramanian &

删除了: .

删除了: T

删除了: caused by SO₄²⁻ addition

批注 [芮苏 1]: Literature shows that H₂SO₄ addition increases CH₄ oxidation rates, but it remains unclear whether this result is caused by pH decrease or SO₄²⁻ increase. Generally, low pH inhibits CH₄ oxidation rates, yet the study sites mentioned in this review where H₂SO₄ enhanced soil CH₄ oxidation were acidic, a decrease in pH may stimulate the activity of methanotrophs and increase the CH₄ oxidation rate.

Thus, methanotrophs in these acidic soils might exhibit different pH preferences compared to those in non-acidic soils. Since SO₄²⁻ addition decreases soil pH, the impact of pH decline on CH₄ oxidation rates must be considered in the context of initial soil pH. Therefore, we have included this part of the content.

删除了: Cu is a crucial component in aerobic CH₄ oxidation processes, as it is utilized by methanotrophs in their molecular machinery, synthesized from metabolized CH₄ through the secretion of methanobactin into the environment. This process facilitates the oxidation of CH₄ to methanol (Dassama et al., 2016).

设置了格式: 字体: 非加粗

设置了格式: 下标

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

Rosenzweig, 2007; Semrau et al., 2010). This process drives the conversion of CH₄ to methanol (Dassama et al., 2016). It was anticipated that methanobactin secreted by methanotrophs during aerobic CH₄ oxidation would facilitate Cu uptake (Knapp et al., 2007); however, the specific mechanism by which methanobactin affects Cu uptake remains unclear (Fig. 2 path j). For methanotrophs capable of expressing both sMMO and pMMO, the expression of these enzymes is regulated by the availability of Cu, a phenomenon known as the classic “copper switch” (Stanley et al., 1983). Under Cu-deficient conditions, these methanotrophs express sMMO. However, as the ratio of Cu to biomass increases, the expression of sMMO significantly decreases, while the expression of pMMO increases (Semrau et al., 2018). Notably, nearly all methanotrophs possess pMMO (Koo and Rosenzweig, 2021); therefore, increased Cu availability can enhance the expression of pMMO. Research indicates that Cu can serve as a promoter of aerobic CH₄ oxidation (Ho et al., 2013). Therefore, SO₄²⁻ addition may promote aerobic CH₄ oxidation by increasing the availability of soil Cu, thereby enhancing the expression of pMMO (Fig. 2 path i and k).

设置了格式: 下标

设置了格式: 字体: 非加粗

删除了: "

删除了: "

删除了:

A positive correlation has been found between P and aerobic CH₄ oxidation in soils (Veraart et al., 2015; Zhang et al., 2020). P can potentially enhance the activity of soil methanotrophs (Fig. 2 path n) (Zhang et al., 2011), with an increase in soil P content achieved through the hydrolysis of organic compounds, including nucleic acids, phospholipids, and phosphate esters, by acid and alkaline phosphatases (Veraart et al., 2015). The addition of SO₄²⁻ accelerated acid phosphatase activity, thereby increasing soil P content (Lv et al., 2014). Therefore, we hypothesize that SO₄²⁻ may indirectly enhance aerobic CH₄ oxidation through the augmentation of soil P content, subsequently promoting the activity of methanotrophs in the soil (Fig. 2 path n and o). It is well-established that Al³⁺ inhibits aerobic CH₄ oxidation (Tamai et al., 2007; Tamai et al., 2003). Additionally, soil acidification resulting from SO₄²⁻ addition has been shown to intensify the toxicity of Al³⁺ in forest soils (Fig. 2 ⑤) (Hu et al., 2013; Sogn and Abrahamsen, 1998). The increase in Al³⁺ can inhibit the activity of methanotrophs (Nanba and King, 2000; Shukla et al., 2013) (Fig. 2 path l), thereby inhibiting aerobic

CH₄ oxidation (Fig. 2 path m). Therefore, SO₄²⁻ addition may directly affect
methanotrophs by enhancing the toxicity of Al³⁺ in the soil, thereby inhibiting aerobic
CH₄ oxidation (Fig. 2 path o). When NH₄Cl and (NH₄)₂SO₄ were added to the soil at
the same molar concentration of NH₄⁺, the inhibitory effect of (NH₄)₂SO₄ on aerobic
CH₄ oxidation was weaker than that of NH₄Cl (Adamsen and King, 1993; Bradford et
al., 2001a; King and Schnell, 1998). NH₄⁺ has been found to inhibit aerobic CH₄
oxidation (Bronson and Mosier, 1994; Dunfield and Knowles, 1995), and the key
mechanism is the competition between CH₄ and NH₄⁺ for the same MMO enzyme
(Gulledge et al., 2004). Due to the similar molecular structures of CH₄ and NH₄⁺, MMO
can oxidize both CH₄ (to CH₃OH) and NH₄⁺ (to NO₂⁻). The inhibitory effect of NH₄Cl
is greater than that of (NH₄)₂SO₄, as SO₄²⁻ may enhance the adsorption of NH₄⁺ onto
cation exchange sites in the soil (Bradford et al., 2001b; Gulledge and Schimel, 1998;
King and Schnell, 1998) (Fig. 2 ⑥). This reduced availability of NH₄⁺ limits its ability
to compete with methanotrophs for MMO enzymes, thereby increasing the availability
of MMO (Fig. 2 path p), promoting aerobic CH₄ oxidation (Fig. 2 path k), and further
intensifying the inhibitory effect of NH₄Cl compared to (NH₄)₂SO₄. In conclusion,
SO₄²⁻ served as a facilitator of aerobic CH₄ oxidation, mitigating the inhibitory effects
of NH₄⁺ on this process.

5 Conclusions

This review synthesizes the double-scale mechanisms by which SO₄²⁻ influences aerobic CH₄ oxidation. Macroscopically, SO₄²⁻ enhances aerobic CH₄ oxidation rates by 0–42%. Mechanistic studies demonstrate that this regulation occurs through SO₄²⁻-driven alteration of environmental factors (e.g., pH, Cu/P availability, Al³⁺ toxicity, NH₄⁺ absorption), which subsequently modulate methanotroph physiology and MMO activity. Based on synthesized evidence, we hypothesize a net stimulatory effect of SO₄²⁻ on aerobic CH₄ oxidation. Validating this hypothesis requires deeper mechanistic insights; therefore, future research should prioritize quantifying aerobic CH₄ oxidation

删除了: SO₄²⁻ plays a pivotal role in global acid deposition, with annual deposition rates ranging from 141.64 ± 120.04 TgS a⁻¹ year⁻¹ (Gao et al., 2022). By synthesizing the available literature and exploring both its macroscopic effects and microscopic mechanisms, we investigated how SO₄²⁻ affects aerobic CH₄ oxidation. We observed that SO₄²⁻ enhances aerobic CH₄ oxidation by up to 0–42% on a macro scale. At the microscopic mechanism level, SO₄²⁻ can influence methanotrophs or MMO by modulating pH, di-O-alkyl C content, Cu availability, P content, Al³⁺ toxicity, and NH₄⁺ absorption, thereby promoting or inhibiting aerobic CH₄ oxidation. Based on these findings, we hypothesize that SO₄²⁻ would promote aerobic CH₄ oxidation.

设置了格式: 上标

设置了格式: 字体: 非加粗

responses to SO_4^{2-} exposure while elucidating underlying microbial mechanisms. This integrated approach is projected to advance CH_4 mitigation strategies amid rising global SO_4^{2-} deposition.

删除了: If this hypothesis is validated in the future, it could provide significant benefits for CH_4 mitigation, particularly in the context of increasing global sulfur deposition. Therefore, future research in this field should focus on investigating the response of aerobic CH_4 oxidation and its influencing factors under increasing SO_4^{2-} conditions, as well as clarifying the underlying microbial mechanisms.

380 Data availability

All raw data can be provided by the corresponding authors upon request.

Author contribution

385 Rui Su finished writing; Kexin Li, Nannan Wang, Fenghui Yuan, Yunjiang Zuo, Ying Sun, Ying Zhao, Lixin Yang, and Liyuan He gave constructive comments and revised the structure and content of the article; Xiaofeng Xu and Lihua Zhang reviewed and edited the manuscript.

Competing interests

390 The authors declare that they have no conflict of interest.

Acknowledgments

This study was partially supported by the National Natural Science Foundation of China (32471777; 32271681) and the Joint Funds of China's National Natural Science Foundation (U2006215), and by Academic Team Leadership Program (2024XSYL01), Minzu University of China, and by the Major Program of the National Natural Science Foundation of China (42494823), the National Key Research and Development Program of China (2024YFF0808703), the Young Scientists Innovation Funds of State Key Laboratory of Black Soils Conservation and Utilization (2023HTDGZ-QN-03).
400 X.X. acknowledged the financial assistance provided by the National Science Foundation (2145130) and SPRUCE and NGEE Arctic projects, supported by the Office of Biological and Environmental Research in the Department of Energy Office of Science.

带格式的: 行距: 1.5 倍行距

删除了: Key Laboratory of Ecology and Environment in Minority Areas (Minzu University of China), National Ethnic Affairs Commission (KLEEMA202206)

删除了: .

删除了:

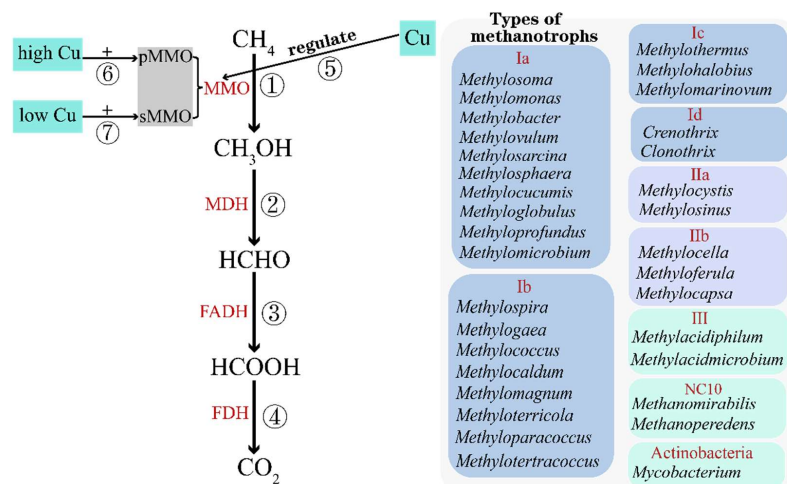
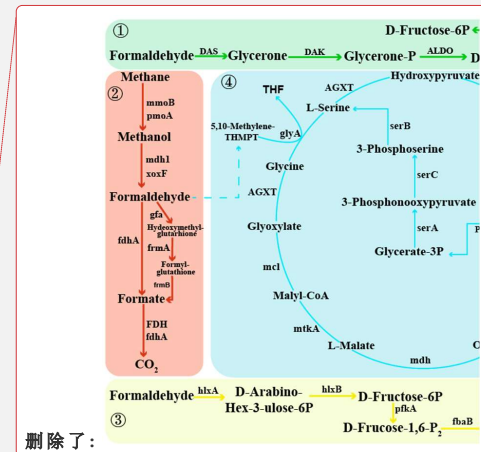


Figure 1: Diagram of the aerobic methane oxidation process and classification of methanotrophs.

① CH_4 is oxidized to methanol (CH_3OH) by MMO; ② CH_3OH is oxidized to formaldehyde (HCHO) by methanol dehydrogenase (MDH); ③ HCHO is oxidized to formate (HCOOH) by formaldehyde dehydrogenase (FADH); ④ HCOOH is oxidized to CO_2 by formate dehydrogenase (FDH); ⑤ Cu controls two MMOs expression; ⑥ High Cu concentration regulates pMMO expression in soil; ⑦ Low Cu concentration regulates sMMO expression in soil.



删除了:

删除了: complete

删除了: including methane assimilation and dissimilation,

删除了: ① Xylulose monophosphate pathway (XyMP pathway) of formaldehyde assimilation; ② CH_4 is oxidized to CO_2 under the sequential action of multiple enzymes; ③ Ribulose monophosphate pathway (RuMP pathway) of formaldehyde assimilation; ④ Serine pathway of formaldehyde assimilation.

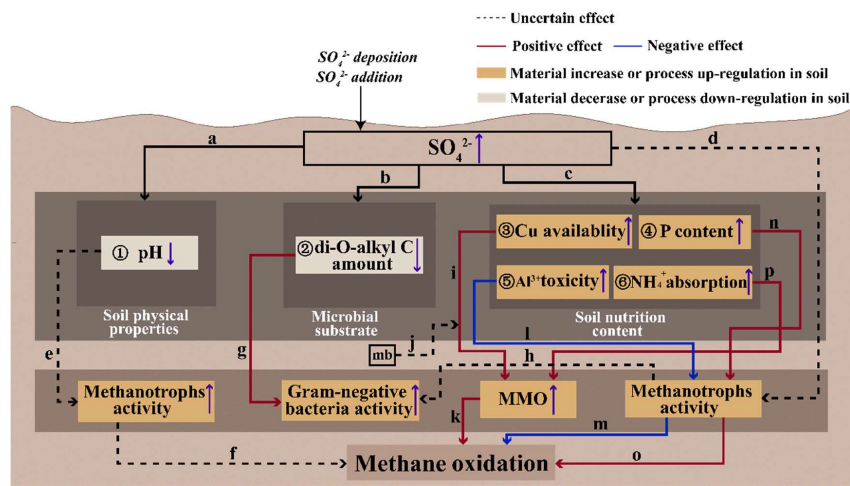
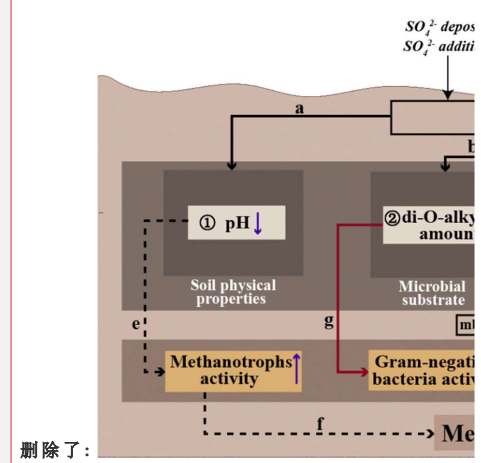


Figure 2. Conceptual diagram of the potential microscopic mechanisms by which sulfate influences aerobic methane oxidation in upland soil.

① SO_4^{2-} decreases soil pH (Fasth et al., 1991; Tie et al., 2020); ② SO_4^{2-} decreases soil di-O-alkyl C amount (Xu et al., 2017); ③ SO_4^{2-} increases soil Cu availability (Islam, 2012); ④ SO_4^{2-} increases soil P content by increasing soil acid phosphatase activity (Lv et al., 2014; Veraart et al., 2015); ⑤ SO_4^{2-} increases soil Al^{3+} toxicity (Hu et al., 2013; Sogn and Abrahamsen, 1998); ⑥ SO_4^{2-} increases NH_4^+ absorption (Bradford et al., 2001b; Gullledge and Schimel, 1998; King and Schnell, 1998); a. Changes in soil physical properties due to increased soil SO_4^{2-} content; b. Changes in soil microbial substrate due to increased soil SO_4^{2-} content; c. SO_4^{2-} may promote CH_4 oxidation; d. SO_4^{2-} affects the activity or community size of methanotrophs in soils (Bradford et al., 2001b; Sitaula et al., 1995); e. Decreased pH may inhibit or stimulate soil CH_4 oxidation (Sitaula et al., 1995); f. Decreased pH may inhibit or stimulate soil CH_4 oxidation (Sitaula et al., 1995); g. Decreased di-O-alkyl C amount increases soil gram-negative bacteria activity (Xu et al., 2017); h. The increased activity of gram-negative bacteria may stem from the enhanced activity of methanotrophs.; i. Elevated Cu availability stimulates soil aerobic CH_4 oxidation (Ho et al., 2013); j. mb (methanobactin) is expected to accelerate Cu uptake (Knapp et al., 2007); k. Enhanced MMO activity facilitates aerobic CH_4 oxidation. l. Elevated Al^{3+} toxicity inhibits soil methanotrophs activity (Nanba and King, 2000; Shukla et al., 2013); m. Decreased methanotrophs



删除了:

activity inhibits soil CH₄ oxidation. n. Elevated P content increases soil methanotrophs activity (Zhang et al., 2011); o. Elevated methanotrophs activity stimulates soil CH₄ oxidation (Bradford et al., 2001b; Sitaula et al., 1995); p. The increased adsorption of NH₄⁺ enhances the availability of MMO to soil methanotrophs.

465

Study site	Sulfate concentration	CH ₄ concentration	Effect	Reference
Perridge Forest	H ₂ SO ₄ (50 Kg S ha ⁻¹)	Ambient air	25 % increased	Bradford et al., 2001b
Perridge Forest	H ₂ SO ₄ (5mM)	Ambient air	no effect	Bradford et al., 2001a
	(NH ₄) ₂ SO ₄ (5mM)	Ambient air	no effect	
Maine forest	Na ₂ SO ₄ 0.5µg S g ⁻¹ soil	250ppm	3% increased	King and Schell, 1998
Norway Scots Pine forest	H ₂ SO ₄ pH3	Ambient air	42% increased	Sitaula et al., 1995
Birch taiga	Na ₂ SO ₄ 2.8 µmol S g ⁻¹ soil	4ppm	no effect	Gulledge and Schimel, 1998
	K ₂ SO ₄ 2.8 µmol S g ⁻¹ soil	4ppm	no effect	

Table 1. Promotion effect of sulfates on methane oxidation in diverse upland soils.

Reference

- 470 Adamsen, A.P.S., King, G.M., Methane consumption in temperate and subarctic forest soils rates, vertical zonation, and responses to water and nitrogen. *Applied and Environmental Microbiology* 59, 485-490. <http://doi.org/10.1128/aem.59.2.485-490.1993>, 1993.
- 475 [Balasubramanian, R., Rosenzweig, A.C., Structural and mechanistic insights into methane oxidation by particulate methane monooxygenase. *Accounts of Chemical Research* 40:573-580. <http://doi.org/10.1021/ar700004s>, 2007.](http://doi.org/10.1021/ar700004s)
- 480 Bechtold, E.K., Ellenbogen, J.B., Villa, J.A., Ferreira, D.K.d.M., Oliverio, A.M., Kostka, J.E., Rich, V.I., Varner, R.K., Bansal, S., Ward, E.J., Bohrer, G., Borton, M.A., Wrighton, K.C., Wilkins, M.J., Metabolic interactions underpinning high methane fluxes across terrestrial freshwater wetlands. *Nature Communications* 16. <http://doi.org/10.1038/s41467-025-56133-0>, 2025.
- Bjorneras, C., Skerlep, M., Floudas, D., Persson, P., Kritzberg, E.S., High sulfate concentration enhances iron mobilization from organic soil to water.

- Biogeochemistry 144, 245-259. <http://doi.org/10.1007/s10533-019-00581-6>, 2019.
- 485 Bodelier, P.L.E., Interactions between nitrogenous fertilizers and methane cycling in wetland and upland soils. *Current Opinion in Environmental Sustainability* 3, 379-388. <http://doi.org/10.1016/j.cosust.2011.06.002>, 2011.
- Boetius, A., Ravensschlag, K., Schubert, C., Rickert, D., Widdel, F., Gieseke, A., Amann, R., Jorgensen, B., Witte, U., Pfannkuche, O., A marine microbial consortium
- 490 apparently mediating anaerobic oxidation of methane. *Nature* 407, 623-626. <http://doi.org/10.1038/35036572>, 2000.
- Bradford, M.A., Ineson, P., Wookey, P.A., Lappin-Scott, H.M., The effects of acid nitrogen and acid sulphur deposition on CH₄ oxidation in a forest soil: a laboratory study. *Soil Biology and Biochemistry* 33, 1695-1702.
- 495 [http://doi.org/10.1016/S0038-0717\(01\)00091-8](http://doi.org/10.1016/S0038-0717(01)00091-8), 2001a.
- Bradford, M.A., Wookey, P.A., Ineson, P., Lappin-Scott, H.M., Controlling factors and effects of chronic nitrogen and sulphur deposition on methane oxidation in a temperate forest soil. *Soil Biology and Biochemistry* 33, 93-102. [http://doi.org/10.1016/S0038-0717\(00\)00118-8](http://doi.org/10.1016/S0038-0717(00)00118-8), 2001b.
- 500 Bronson, K.F., Mosier, A.R., Suppression of methane oxidation in aerobic soil by nitrogen fertilizers, nitrification inhibitors, and urease inhibitors. *Biology and Fertility of Soils* 17, 263-268. <http://doi.org/10.1007/bf00383979>, 1994.
- Brumme, R., Borken, W., Site variation in methane oxidation as affected by atmospheric deposition and type of temperate forest ecosystem. *Global Biogeochemical Cycles* 13, 493-501. <http://doi.org/10.1029/1998gb900017>, 1999.
- 505 Chistoserdova, L., Vorholt, J.A., Lidstrom, M.E., A genomic view of methane oxidation by aerobic bacteria and anaerobic archaea. *Genome Biology* 6, 1-6. <http://doi.org/10.1186/gb-2005-6-2-208>, 2005.
- Chowdhury, T.R., Dick, R.P., Ecology of aerobic methanotrophs in controlling methane fluxes from wetlands. *Applied Soil Ecology* 65, 8-22. <http://doi.org/10.1016/j.apsoil.2012.12.014>, 2013.
- 510 Dassama, L.M., Kenney, G.E., Ro, S.Y., Zielazinski, E.L., Rosenzweig, A.C., Methanobactin transport machinery. *Proceedings of the National Academy of Sciences of the United States of America* 113, 13027-13032. <http://doi.org/10.1073/pnas.1603578113>, 2016.
- 515 Deng, Y., Che, R., Wang, F., Conrad, R., Dumont, M., Yun, J., Wu, Y., Hu, A., Fang, J., Xu, Z., Cui, X., Wang, Y., Upland Soil Cluster Gamma dominates methanotrophic communities in upland grassland soils. *Science of the Total Environment* 670, 826-836. <http://doi.org/10.1016/j.scitotenv.2019.03.299>, 2019.
- 520 DiSpirito, A.A., Semrau, J.D., Murrell, J.C., Gallagher, W.H., Dennison, C., Vuilleumier, S., Methanobactin and the link between copper and bacterial methane oxidation. *Microbiology and Molecular Biology Reviews* 80:387-409. <http://doi.org/10.1128/mmb.00058-15>, 2016.
- 525 Dunfield, P., Knowles, R., Kinetics of inhibition of methane oxidation by nitrate, nitrite, and ammonium in a humisol. *Applied and Environmental Microbiology* 61, 3129-3135. <http://doi.org/10.1128/aem.61.8.3129-3135.1995>, 1995.
- Fan, J.L., Xu, Y.H., Chen, Z.M., Xiao, J., Liu, D.Y., Luo, J.F., Bolan, N., Ding, W.X.,

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

设置了格式: 字体: 非加粗

- Sulfur deposition suppressed nitrogen-induced soil N₂O emission from a subtropical forestland in southeastern China. *Agricultural and Forest Meteorology* 233, 163-170. <http://doi.org/10.1016/j.agrformet.2016.11.017>, 2017.
- 530 Fan, L.C., Schneider, D., Dippold, M.A., Poehlein, A., Wu, W.C., Gui, H., Ge, T.D., Wu, J.S., Thiel, V., Kuzyakov, Y., Dorodnikov, M., Active metabolic pathways of anaerobic methane oxidation in paddy soils. *Soil Biology and Biochemistry* 156, 108215. <http://doi.org/10.1016/j.soilbio.2021.108215>, 2021.
- 535 Fasth, W.J., David, M.B., Vance, G.F., Sulfate retention and cation leaching of forest soils in response to acid additions. *Canadian Journal of Forest Research* 21, 32-41. <http://doi.org/10.1139/x91-005>, 1991.
- Fenibo, E.O., Selvarajan, R., Wang, H.Q., Wang, Y., Abia, A.L.K., Untapped talents: insight into the ecological significance of methanotrophs and its prospects. *Science of the Total Environment* 903, 166145. <http://doi.org/10.1016/j.scitotenv.2023.166145>, 2023.
- 540 Gauci, V., Matthews, E., Dise, N., Walter, B., Koch, D., Granberg, G., Vile, M., Sulfur pollution suppression of the wetland methane source in the 20th and 21st centuries. *Proceedings of the National Academy of Sciences of the United States of America* 101, 12583-12587. <http://doi.org/10.1073/pnas.0404412101>, 2004.
- 545 Granberg, G., Sund, I., Svensson, B.H., Nilsson, M., Effect of temperature, and nitrogen and sulfur deposition, on methane emission from a boreal mire. *Ecology* 82, 1982-1998. 2001.
- Gulledge, J., Hrywna, Y., Cavanaugh, C., Steudler, P.A., Effects of long-term nitrogen fertilization on the uptake kinetics of atmospheric methane in temperate forest soils. *FEMS Microbiology Ecology* 49, 389-400. <http://doi.org/10.1016/j.femsec.2004.04.013>, 2004.
- 550 Gulledge, J., Schimel, J., Low-concentration kinetics of atmospheric CH₄ oxidation in soil and mechanism of NH₄⁺ inhibition. *Applied and Environmental Microbiology* 64, 4291-4298. 1998.
- 555 Guo, J.H., Feng, H.L., Peng, C.H., Chen, H., Xu, X., Ma, X.H., Li, L., Kneeshaw, D., Ruan, H.H., Yang, H.Q., Wang, W.F., Global climate change increases terrestrial soil CH₄ emissions. *Global Biogeochemical Cycles* 37, e2021GB007255. <http://doi.org/10.1029/2021gb007255>, 2023.
- 560 Hakemian, A.S., Rosenzweig, A.C., The biochemistry of methane oxidation. *Annual Review of Biochemistry* 76, 223-241. <http://doi.org/10.1146/annurev.biochem.76.061505.175355>, 2007.
- Hanson, R.S., Hanson, T.E., Methanotrophic bacteria. *Microbiological Reviews* 60, 439-471. <http://doi.org/10.1128/mmbr.60.2.439-471.1996>, 1996.
- 565 Ho, A., Lee, H.J., Reumer, M., Meima-Franke, M., Raaijmakers, C., Zweers, H., de Boer, W., Van der Putten, W.H., Bodelier, P.L.E., Unexpected role of canonical aerobic methanotrophs in upland agricultural soils. *Soil Biology and Biochemistry* 131, 1-8. <http://doi.org/10.1016/j.soilbio.2018.12.020>, 2019.
- 570 Ho, A., Lüke, C., Reim, A., Frenzel, P., Selective stimulation in a natural community of methane oxidizing bacteria: effects of copper on *pmoA* transcription and activity. *Soil Biology and Biochemistry* 65, 211-216.

删除了: Gao, J., Zhou, W., Liu, Y., Sha, L., Song, Q., Lin, Y., Yu, G., Zhang, J., Zheng, X., Fang, Y., Grace, J., Zhao, J., Xu, J., Gui, H., Sinclair, F., Zhang, Y., Litter-derived nitrogen reduces methane uptake in tropical rainforest soils. *Science of the Total Environment* 849, 157891. <http://doi.org/10.1016/j.scitotenv.2022.157891>, 2022.

<http://doi.org/10.1016/j.soilbio.2013.05.027>, 2013.

580 Hu, M.J., Wilson, B.J., Sun, Z.G., Huang, J.F., Tong, C., Effects of nitrogen and sulphate addition on methane oxidation in the marsh soil of a typical subtropical estuary (Min River) in China. *Chemistry and Ecology* 34, 610-623. <http://doi.org/10.1080/02757540.2018.1464153>, 2018.

585 Hu, Y.L., Jung, K., Zeng, D.H., Chang, S.X., Nitrogen- and sulfur-deposition-altered soil microbial community functions and enzyme activities in a boreal mixedwood forest in western Canada. *Canadian Journal of Forest Research* 43, 777-784. <http://doi.org/10.1139/cjfr-2013-0049>, 2013.

590 Huang, J., Zhou, K.J., Zhang, W., Liu, J.X., Ding, X., Cai, X.A., Mo, J.M., Sulfur deposition still contributes to forest soil acidification in the Pearl River Delta, South China, despite the control of sulfur dioxide emission since 2001. *Environmental Science and Pollution Research* 26, 12928-12939. <http://doi.org/10.1007/s11356-019-04831-w>, 2019.

595 Huang, R.L., Crowther, T.W., Sui, Y.Y., Sun, B., Liang, Y.T., High stability and metabolic capacity of bacterial community promote the rapid reduction of easily decomposing carbon in soil. *Communications Biology* 4, 1376. <http://doi.org/10.1038/s42003-021-02907-3>, 2021.

600 IPCC, 2013. Climate change 2013: the physical science basis. In: Stocker, T.F., Qin, D., Plattner, G.-K., Tignor, M., Allen, S.K., Boschung, J., Nauels, A., Xia, Y., Bex, V., Midgley, P.M. (Eds.), Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.

Islam, M., The effect of different rates and forms of sulfur on seed yield and micronutrient uptake by chickpea. *Plant, Soil and Environment* 58, 399-404. <http://doi.org/10.17221/145/2012-pse>, 2012.

605 King, G.M., Schnell, S., Effects of ammonium and non-ammonium salt additions on methane oxidation by *Methylosinus trichosporium* OB3b and Maine forest soils. *Applied and Environmental Microbiology* 64, 253-257. 1998.

610 Klessa, D.A., Frame, J., Golightly, R.D., Harkess, R.D., THE EFFECT OF FERTILIZER SULFUR ON GRASS PRODUCTION FOR SILAGE. *Grass and Forage Science* 44, 277-281. <http://doi.org/10.1111/j.1365-2494.1989.tb02165.x>, 1989.

615 Knapp, C.W., Fowle, D.A., Kulczycki, E., Roberts, J.A., Graham, D.W., Methane monooxygenase gene expression mediated by methanobactin in the presence of mineral copper sources. *Proceedings of the National Academy of Sciences of the United States of America* 104, 12040-12045. <http://doi.org/10.1073/pnas.0702879104>, 2007.

Koo, C.W., Rosenzweig, A.C., Biochemistry of aerobic biological methane oxidation. *Chemical Society Review* 50, 3424-3436. <http://doi.org/10.1039/d0cs01291b>, 2021.

620 La, W., Han, X., Liu, C.Q., Ding, H., Liu, M., Sun, F., Li, S., Lang, Y., Sulfate concentrations affect sulfate reduction pathways and methane consumption in coastal wetlands. *Water Research* 217, 118441.

<http://doi.org/10.1016/j.watres.2022.118441>, 2022.

Le Mer, J., Roger, P., Production, oxidation, emission and consumption of methane by soil: a review. *European Journal of Soil Biology* 37, 25-50.

[http://doi.org/10.1016/S1164-5563\(01\)01067-6](http://doi.org/10.1016/S1164-5563(01)01067-6), 2001.

Li, C., Liu, X., Meng, M.J., Zhai, L., Zhang, B., Jia, Z.H., Gu, Z.Y., Liu, Q.Q., Zhang, Y.L., Zhang, J.C., The use of Biolog Eco microplates to compare the effects of sulfuric and nitric acid rain on the metabolic functions of soil microbial communities in a subtropical plantation within the Yangtze River Delta region. *Catena* 198, 105039. <http://doi.org/10.1016/j.catena.2020.105039>, 2021.

Li, C.H., Wang, B., Fang, Z., Yu, H.L., Huang, J.Y., Plant species diversity is driven by soil base cations under acid deposition in desert coal-mining region in northwestern China. *Ecological Indicators* 145, 109682. <http://doi.org/10.1016/j.ecolind.2022.109682>, 2022.

Li, Y., Liu, Y.W., Pan, H., Hernández, M., Guan, X.M., Wang, W., Zhang, Q.C., Luo, Y., Di, H.J., Xu, J.M., Impact of grazing on shaping abundance and composition of active methanotrophs and methane oxidation activity in a grassland soil. *Biology and Fertility of Soils* 56, 799-810. <http://doi.org/10.1007/s00374-020-01461-0>, 2020.

Lv, Y., Wang, C.Y., Jia, Y.Y., Wang, W.W., Ma, X., Du, J.J., Pu, G.Z., Tian, X.J., Effects of sulfuric, nitric, and mixed acid rain on litter decomposition, soil microbial biomass, and enzyme activities in subtropical forests of China. *Applied Soil Ecology* 79, 1-9. <http://doi.org/10.1016/j.apsoil.2013.12.002>, 2014.

Mancinelli, R., The regulation of methane oxidation in soil. *Annual Review of Microbiology* 49, 581-605. 1995.

Mishra, V.K., Shukla, R., Shukla, P.N., Inhibition of Soil Methane Oxidation by Fertilizer Application: an Intriguing but Persistent Paradigm. *Environmental Pollution and Protection* 3, 57-69. <http://doi.org/10.22606/epp.2018.32001>, 2018.

Murguia-Flores, F., Arndt, S., Ganesan, A.L., Murray-Tortarolo, G., Hornibrook, E.R.C., Soil Methanotrophy Model (MeMo v1.0): a process-based model to quantify global uptake of atmospheric methane by soil. *Geoscientific Model Development* 11, 2009-2032. <http://doi.org/10.5194/gmd-11-2009-2018>, 2018.

Nanba, K., King, G., Response of atmospheric methane consumption by maine forest soils to exogenous aluminum salts. *Applied and Environmental Microbiology* 66, 3674-3679. <http://doi.org/10.1128/AEM.66.9.3674-3679.2000>, 2000.

Op den Camp, H.J., Islam, T., Stott, M.B., Harhangi, H.R., Hynes, A., Schouten, S., Jetten, M.S., Birkeland, N.K., Pol, A., Dunfield, P.F., Environmental, genomic and taxonomic perspectives on methanotrophic *Verrucomicrobia*. *Environmental Microbiology Reports* 1, 293-306. <http://doi.org/10.1111/j.1758-2229.2009.00022.x>, 2009.

Palmer, S.M., Clark, J.M., Chapman, P.J., van der Heijden, G.M.F., Bottrell, S.H., Effects of acid sulphate on DOC release in mineral soils: the influence of SO_4^{2-} retention and Al release. *European Journal of Soil Science* 64, 537-544. <http://doi.org/10.1111/ejss.12048>, 2013.

Place, S.E., Examining the role of ruminants in sustainable food systems. *Grass and*

Forage Science 79, 135-143. <http://doi.org/10.1111/gfs.12673>, 2024.

Praeg, N., Wagner, A.O., Illmer, P., Plant species, temperature, and bedrock affect net methane flux out of grassland and forest soils. *Plant and Soil* 410, 193-206. <http://doi.org/10.1007/s11104-016-2993-z>, 2016.

670 Saunois, M., Stavert, A.R., Poulter, B., Bousquet, P., Canadell, J.G., Jackson, R.B.,
Raymond, P.A., Dlugokencky, E.J., Houweling, S., Patra, P.K., Ciais, P., Arora,
V.K., Bastviken, D., Bergamaschi, P., Blake, D.R., Brailsford, G., Bruhwiler, L.,
Carlson, K.M., Carrol, M., Castaldi, S., Chandra, N., Crevoisier, C., Crill, P.M.,
Covey, K., Curry, C.L., Etiope, G., Frankenberg, C., Gedney, N., Hegglin, M.I.,
675 Hoglund-Isaksson, L., Hugelius, G., Ishizawa, M., Ito, A., Janssens-Maenhout, G.,
Jensen, K.M., Joos, F., Kleinen, T., Krummel, P.B., Langenfelds, R.L., Laruelle,
G.G., Liu, L., Machida, T., Maksyutov, S., McDonald, K.C., McNorton, J., Miller,
P.A., Melton, J.R., Morino, I., Muller, J., Murguia-Flores, F., Naik, V., Niwa, Y.,
Noce, S., Doherty, S.O., Parker, R.J., Peng, C., Peng, S., Peters, G.P., Prigent, C.,
680 Prinn, R., Ramonet, M., Regnier, P., Riley, W.J., Rosentreter, J.A., Segers, A.,
Simpson, I.J., Shi, H., Smith, S.J., Steele, L.P., Thornton, B.F., Tian, H., Tohjima,
Y., Tubiello, F.N., Tsuruta, A., Viovy, N., Voulgarakis, A., Weber, T.S., van Weele,
M., van der Werf, G.R., Weiss, R.F., Worthy, D., Wunch, D., Yin, Y., Yoshida, Y.,
Zhang, W., Zhang, Z., Zhao, Y., Zheng, B., Zhu, Q., Zhu, Q., Zhuang, Q., The
685 Global Methane Budget 2000-2017. *Earth System Science Data* 12, 1561-1623.
<http://doi.org/10.5194/essd-12-1561-2020>, 2020.

Schimel, J., Playing scales in the methane cycle: from microbial ecology to the globe.
Proceedings of the National Academy of Sciences of the United States of America
101, 12400-12401. <http://doi.org/10.1073/pnas.0405075101>, 2004.

690 Schimel, J., Gullledge, J., Microbial community structure and global trace gases. *Global
Change Biology* 4, 745-758. <http://doi.org/10.1046/j.1365-2486.1998.00195.x>,
1998.

Semrau, J.D., DiSpirito, A.A., Gu, W., Yoon, S., Metals and Methanotrophy. *Applied
and Environmental Microbiology* 84. <http://doi.org/10.1128/aem.02289-17>,
2018. [c](https://doi.org/10.1128/aem.02289-17)

695 [Semrau, J.D., DiSpirito, A.A., Yoon, S., Methanotrophs and copper. *FEMS
Microbiology Reviews* 34:496-531. \[http://doi.org/10.1111/j.1574-
6976.2010.00212.x\]\(http://doi.org/10.1111/j.1574-
6976.2010.00212.x\), 2010.](https://doi.org/10.1128/aem.02289-17)

700 Shukla, P.N., Pandey, K.D., Mishra, V.K., Environmental determinants of soil methane
oxidation and methanotrophs. *Critical Reviews in Environmental Science and
Technology* 43, 1945-2011. <http://doi.org/10.1080/10643389.2012.672053>, 2013.

Silver, W.L., Lugo, A.E., Keller, M., Soil oxygen availability and biogeochemistry
along rainfall and topographic gradients in upland wet tropical forest soils.
Biogeochemistry 44, 301-328. <http://doi.org/10.1007/bf00996995>, 1999.

705 Singh, B.K., Bardgett, R.D., Smith, P., Reay, D.S., Microorganisms and climate change:
terrestrial feedbacks and mitigation options. *Nature Reviews Microbiology* 8, 779-
790. <http://doi.org/10.1038/nrmicro2439>, 2010.

Sitaula, B.K., Bakken, L.R., Abrahamsen, G., CH₄ uptake by temperate forest soil:
effect of N input and soil acidification. *Soil Biology and Biochemistry* 27, 870-

- 710 880. [http://doi.org/10.1016/0038-0717\(95\)00017-9](http://doi.org/10.1016/0038-0717(95)00017-9), 1995.
- Sogn, T., Abrahamsen, G., Effects of N and S deposition on leaching from an acid forest soil and growth of Scots pine (*Pinus sylvestris* L.) after 5 years of treatment. *Forest Ecology and Management* 103, 177-190. [http://doi.org/10.1016/S0378-1127\(97\)00188-6](http://doi.org/10.1016/S0378-1127(97)00188-6), 1998.
- 715 Song, H., Peng, C., Zhu, Q., Chen, Z., Blanchet, J.-P., Liu, Q., Li, T., Li, P., Liu, Z., Quantification and uncertainty of global upland soil methane sinks: Processes, controls, model limitations, and improvements. *Earth-Science Reviews* 252. <http://doi.org/10.1016/j.earscirev.2024.104758>, 2024.
- 720 Stanley, S.H., Prior, S.D., Leak, D.J., Dalton, H., COPPER STRESS UNDERLIES THE FUNDAMENTAL CHANGE IN INTRACELLULAR LOCATION OF METHANE MONO-OXYGENASE IN METHANE-OXIDIZING ORGANISMS - STUDIES IN BATCH AND CONTINUOUS CULTURES. *Biotechnology Letters* 5, 487-492. <http://doi.org/10.1007/bf00132233>, 1983.
- 725 Tamai, N., Takenaka, C., Ishizuka, S., Water-soluble Al inhibits methane oxidation at atmospheric concentration levels in Japanese forest soil. *Soil Biology and Biochemistry* 39, 1730-1736. <http://doi.org/10.1016/j.soilbio.2007.01.029>, 2007.
- Tamai, N., Takenaka, C., Ishizuka, S., Tezuka, T., Methane flux and regulatory variables in soils of three equal-aged Japanese cypress (*Chamaecyparis obtusa*) forests in central Japan. *Soil Biology and Biochemistry* 35, 633-641. [http://doi.org/10.1016/s0038-0717\(03\)00010-5](http://doi.org/10.1016/s0038-0717(03)00010-5), 2003.
- 730 Tie, L.H., Fu, R., Penuelas, J., Sardans, J., Zhang, S.B., Zhou, S.X., Hu, J.X., Huang, C.D., The additions of nitrogen and sulfur synergistically decrease the release of carbon and nitrogen from litter in a subtropical forest. *Forests* 11, 1280. <http://doi.org/10.3390/f11121280>, 2020.
- 735 van Spanning, R.J.M., Guan, Q., Melkonian, C., Gallant, J., Polerecky, L., Flot, J.F., Brandt, B.W., Braster, M., Iturbe Espinoza, P., Aerts, J.W., Meima-Franke, M.M., Piersma, S.R., Bunduc, C.M., Ummels, R., Pain, A., Fleming, E.J., van der Wel, N.N., Gherman, V.D., Sarbu, S.M., Bodelier, P.L.E., Bitter, W., Methanotrophy by a *Mycobacterium* species that dominates a cave microbial ecosystem. *Nature Microbiology* 7, 2089-2100. <http://doi.org/10.1038/s41564-022-01252-3>, 2022.
- 740 Veraart, A.J., Steenbergh, A.K., Ho, A., Kim, S.Y., Bodelier, P.L.E., Beyond nitrogen: the importance of phosphorus for CH₄ oxidation in soils and sediments. *Geoderma* 259-260, 337-346. <http://doi.org/10.1016/j.geoderma.2015.03.025>, 2015.
- 745 Walsh, C., O'Regan, B., Moles, R., Incorporating methane into ecological footprint analysis: A case study of Ireland. *Ecological Economics* 68, 1952-1962. <http://doi.org/10.1016/j.ecolecon.2008.07.008>, 2009.
- Wang, Q., Kwak, J.-H., Choi, W.-J., Chang, S.X., Decomposition of trembling aspen leaf litter under long-term nitrogen and sulfur deposition: effects of litter chemistry and forest floor microbial properties. *Forest Ecology and Management* 412, 53-61. <http://doi.org/10.1016/j.foreco.2018.01.042>, 2018.
- 750 Wang, Z.P., Ineson, P., Methane oxidation in a temperate coniferous forest soil: effects of inorganic N. *Soil Biology and Biochemistry* 35, 427-433. [http://doi.org/10.1016/s0038-0717\(02\)00294-8](http://doi.org/10.1016/s0038-0717(02)00294-8), 2003.

- 755 Xu, Y.H., Fan, J.L., Ding, W.X., Gunina, A., Chen, Z.M., Bol, R., Luo, J.F., Bolan, N.,
Characterization of organic carbon in decomposing litter exposed to nitrogen and
sulfur additions: links to microbial community composition and activity.
Geoderma 286, 116-124. <http://doi.org/10.1016/j.geoderma.2016.10.032>, 2017.
- 760 | Zhang, L.H., Yuan, F.H., Bai, J.H., Duan, H.T., Gu, X.Y., Hou, L.Y., Huang, Y., Yang,
M.G., He, J.S., Zhang, Z.H., Yu, L.J., Song, C.C., Lipson, D.A., Zona, D., Oechel,
W., Janssens, I.A., Xu, X.F., Phosphorus alleviation of nitrogen-suppressed
methane sink in global grasslands. *Ecology Letters* 23, 821-830.
<http://doi.org/10.1111/ele.13480>, 2020.
- 765 Zhang, T., Zhu, W., Mo, J., Liu, L., Dong, S., Increased phosphorus availability
mitigates the inhibition of nitrogen deposition on CH₄ uptake in an old-growth
tropical forest, southern China. *Biogeosciences* 8, 2805-2813.
<http://doi.org/10.5194/bg-8-2805-2011>, 2011.

删除了：Yang, C., Chen, Y.T., Zhang, Q., Qie, X.H.,
Chen, J.X., Che, Y.J., Lv, D.T., Xu, X.Y., Gao, Y.X., Wang,
Z.Y., Sun, J., Mechanism of microbial regulation on
methane metabolism in saline-alkali soils based on
metagenomics analysis. *Journal of Environmental
Management* 345, 118771.
<http://doi.org/10.1016/j.jenvman.2023.118771>, 2023.