Old Carbon, New Insights: Thermal Reactivity and Bioavailability of Saltmarsh Soils

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Abstract

Saltmarshes are globally important coastal wetlands which can helpstore carbon for millennia, helping to mitigate the impacts of climate change. They accumulate organic carbon from both modern and agedautochthonous sources through in-situ biological(above- and belowground plant production) and the capture of ex-situallochthonous sources which are(terrestrial and marine sediments deposited during tidal inundation...). Previous studies have found that long-term organic carbon storage in saltmarsh soils is driven by the net contribution from the olderpre-aged allochthonous fraction, implying that the inputs of youngautochthonous organic carbon derived from in situ production areis recycled at a faster rate. However, it is also acknowledged that the bioavailability of soil organic carbon depends as much upon environmental conditions as the reactivity of the organic carbon itself. Until now, there has been no empirical evidence linking the reactivity of saltmarsh soil organic carbon with its bioavailability for remineralization.

Using We found that the ¹⁴C age of CO₂ produced during ramped oxidation, we assessed the composition (¹⁴C and ¹³C) of saltmarsh soil carbon pools defined by their thermal reactivity. By relating ¹⁴C measurements of the soil carbon pools to CO₂ respired in aerobic incubations of soils from the same soils, we provide the first empirical evidence linking the thermal reactivity of saltmarsh soil organic carbon with its bioavailability for remineralization.

We found that old (14C) saltmarsh ranged from 201 to 14,875 years BP, and that 14C depleted (older) carbon evolved from higher temperature ramped exidation fractions, indicating that older carbon

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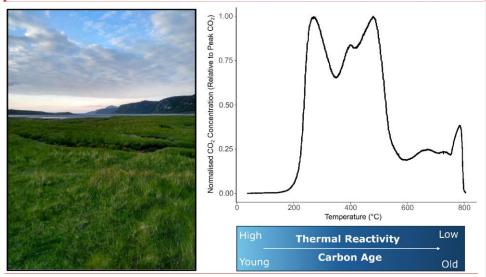
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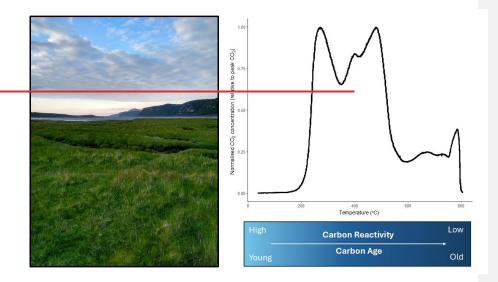
dominates the thermally recalcitrant <u>organic carbon pools</u>, whereas the thermally labile carbon is composed of younger organic carbon sources, fractions. In most cases, the ¹⁴C content of the lowest temperature ramped oxidation fraction (the most thermally labile <u>carbon poolorganic C source)</u> was closest to the previously reported ¹⁴C content of the CO₂ evolved from aerobic incubations of the same soils, implying that the <u>latter was from a thermally labile organic carbon source. This implies that the</u> bioavailability of saltmarsh soil organic carbon to remineralisation in oxic conditions is closely related to its thermal <u>lability</u>.

Our results highlight the importance of saltmarshes as stores of both old, thermally recalcitrant organic carbon, as well as younger, thermally labile organic carbon that is vulnerable to decomposition under oxic conditions, reactivity. Management interventions (e.g. rewetting by tidal inundation) to limit the exposure of saltmarsh soils to elevated oxygen availability may help to protect and conserve these stores of thermallyeld, labile organic carbon and hence limit CO₂ emissions. We also present the first evidence to support the inclusion of thermally labile allochthonous OC stored in saltmarsh soils in additionality assessments, with relevance to international carbon crediting projects and National GHG Inventories.

Graphical Abstract



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1. Introduction

portion of this OC is stored for millennia, providing a climate regulation service, and some is returned to the atmosphere or laterally exported (Komada et al., 2022; Macreadie et al., 2021). Saltmarshes also accumulate and produce inorganic carbon (IC) but the climate regulation service of this is currently under debate and unclear (Granse et al., 2024; Van Dam et al., 2021). To understand the role of saltmarsh soils in carbon cycling and their potential for climate mitigation through targeted management interventions, much research has focussed on determining the autochthonous (in-situ) and allochthonous (ex-situ, trapped during tidal inundation from terrestrial and marine sources) contributions to saltmarsh soils, with the accumulation of autochthonous OC as a direct sequestration of carbon from the atmosphere, reducing the amount of atmospheric greenhouse gases (GHGs) (Macreadie et al., 2019; Saintilan et al., 2013; Van de Broek et al., 2018). The accumulation of allochthonous OC, originally sequestered outside the saltmarsh area, does not directly reduce atmospheric GHGs, but can represent a source of avoided emissions if it remains stored in the saltmarsh soil for longer than in an alternative depositional environment (Howard et al., 2023). Evidence to determine whether this is the case or not, and under what scenarios, has proven challenging to obtain (Geraldi et al.,

2019; Houston et al., 2024a). OC pools with distinct biological turnover times may instead

Saltmarshes accumulate organic carbon (OC) of variable age and reactivity into their soils. A

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provide greater insights(Houston et al., 2024).

21 Another approach is to partition the saltmarsh soil OC pool by reactivity (Luk et al., 2021), which 22 may provide greater insight into the soil carbon residence time and therefore the climate 23 mitigation achieved through targeted management interventions to retain that carbon 24 (Sanderman and Grandy, 2020). Soil OC reactivity can be defined as its availability for 25 remineralisation by soil microbial communities, with different reactivity pools having different 26 turnover times (Plante et al., 2009). 27 Ramped oxidation (RO) and ramped pyrolysis oxidation (RPO) have been used to estimate the 28 thermal reactivity and biological turnover time of soil and sediment OC (Hemingway et al., 2017b; 29 Plante et al., 2011; Rosenheim et al., 2008). RO and RPO involve measuring the quantity of CO2 30 evolved as a sample is increasingly heated at a constant rate in an atmosphere containing oxygen 31 (e.g., Plante et al., 2011; Stoner et al., 2023), or other gases, typically Helium (RPO: e.g., 32 Hemingway et al., 2017a; Rosenheim et al., 2008). The temperature at which CO2 is thermally-33 evolved is related to the activation energy required to thermally decompose C (Hemingway et al., 34 2017b), which is also an estimate of the energy required for biological degradation of OC (Peltre 35 et al., 2013; Plante et al., 2013). CO2 evolved at low temperatures is deemed to be from soil OC 36 pools with a greater thermal lability than CO₂ evolved at higher temperatures (Peltre et al., 2013; 37 Rosenheim et al., 2008). OC thermal reactivity pools can be examined by collecting the evolved 38 CO₂ from set temperature ranges with distinct thermal reactivities and measuring the ¹⁴C (age) 39 and ¹³C content (Rosenheim et al., 2008), which can then be related to the activation energy 40 required to thermally decompose those C sources (Hemingway et al., 2017b). 41 The ¹⁴C content of the thermal reactivity pools provides insight into the turnover time of each pool, 42 with past research showing that the oldest soil organic matter (OM) (most depleted 14C content) 43 tends to dominate the most thermally recalcitrant fractions (Bao et al., 2019b; Plante et al., 2013; 44 Stoner et al., 2023). Similar results have been found for saltmarsh soils (Luk et al., 2021). Young 45 OC, which can be autochthonous or allochthonous (Van de Broek et al., 2018), has been found 46 to turnover at a faster rate than old OC in saltmarsh soils (Komada et al., 2022; Van de Broek et 47 al., 2018), implying that young OC may tend to be more thermally labile than old OC for saltmarsh 48 soils. 49 The ¹³C content of the thermal reactivity pools can also provide insight as to whether the source 50 of OC has an influence on turnover time. Previous work has found that the ¹³C content of evolved 51 CO₂ tends to be more enriched at higher temperatures due to greater contributions from ¹³C-52 enriched, degraded/microbially derived OC (Luk et al., 2021; Sanderman and Grandy, 2020; 53 Stoner et al., 2023). Similarly, comparisons of the isotopic composition of thermally-defined OC

pools to their chemical properties have found that thermally labile OC is derived from mostly lipids and polysaccharides, whereas OC with a higher thermal recalcitrance is derived from a greater proportion of phenolic and aromatic compounds (Sanderman and Grandy, 2020). The thermal reactivity of soil and sediment OC is also influenced by the formation of organo-mineral complexes, which can physically and chemically stabilise OC (Bianchi et al., 2024; Hemingway et al., 2019). Mineral-associations can increase the energy required for decomposition and have been found to increase thermal recalcitrance and to slow turnover times of soil and sediment OC (Hemingway et al., 2019; Stoner et al., 2023). Crucially, the biological availability (bioavailability) of OC for decomposition, and hence its biological turnover time, depends on the prevailing environmental conditions as well as thermal reactivity (Hemingway et al., 2017b; Schmidt et al., 2011). For example, increased hydrodynamic energy can destabilise organo-mineral complexes and increase the bioavailability of previously stable OC (Spivak et al., 2019). Similarly, increased oxygen availability can decrease the energy requirement for microbes to decompose molecularly recalcitrant OC, causing it to be remineralised at a faster rate (Noyce et al., 2023). Houston et al. (2024b) found that young OC stored in saltmarsh soils was preferentially respired as carbon dioxide (CO2) during aerobic incubation experiments, but that a portion of the respired CO₂ was produced from an aged (14C-depleted), allochthonous source. It is possible that this CO2 could have been respired from thermally labile as well as thermally recalcitrant soil OC sources because the increased oxygen availability of the incubations potentially facilitated the degradation of OC which was previously stable in the low-oxygen environment of typical saltmarsh soils (Noyce et al., 2023). The isotopic composition of RO thermal reactivity fractions can be compared to the isotopic composition of the CO₂ that is evolved biologically during incubations of equivalent samples to determine whether or not the age of the most biologically- and thermally-reactive OC pools match. Here, we present the first measurements of the ¹³C and ¹⁴C content of CO₂ derived from saltmarsh soils using RO, and the first comparison of these to the ¹⁴C content of biologically evolved CO₂ from the same soils (Houston et al., 2024b). We hypothesised that the thermally labile C pools would be composed of younger C than the thermally recalcitrant pools, and that the CO₂ evolved from saltmarsh soils exposed to oxic conditions (Houston et al., 2024b) are from a predominantly thermally labile OC pool.

The proportions of autochthonous and allochthonous OC accumulating in saltmarsh soils and

OC reactivity pools are thought to be related, as in-situ processes during burial of saltmarsh soils

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have been suggested to favour the long-term storage of aged, allochthonous OC (Komada et al., 2022; Leorri et al., 2018; Mueller et al., 2019; Van de Broek et al., 2018). Young OC, which can be autochthonous or allochthonous (Van de Broek et al., 2018), is hypothesised to turnover at a faster rate, often resulting in its remineralisation to the atmosphere. It is therefore assumed that old OC is mostly composed of recalcitrant (low reactivity) components, whereas young OC contains a greater proportion of labile (reactive) components (Komada et al., 2022; Van de Broek et al., 2018). Of course, this is not always the case as young OC can contain recalcitrant material, and older OC can be labile if it was stored in a stable environment with low carbon turnover rates, such as a permafrost soil, prior to its mobilisation (Dasari et al., 2024).

Houston et al. (2024) found that a portion of the carbon dioxide (CO_2) evolved during aerobic incubations of saltmarsh soils was from an old, allochthonous source. It is possible that the CO_2 could have been evolved from a labile source, or a physically stabilized source that decomposed due to increased oxygen availability (a thermodynamically favourable terminal electron acceptor facilitating the degradation of OC which was stable in a low-oxygen environment, as saltmarsh soils typically are (Noyce et al., 2023)). To constrain these sources, the ¹⁴C composition of the biologically evolved CO_2 from these experiments can be directly compared to the ¹⁴C composition of the thermally characterized soil CC:

The thermal reactivity of soil OC can usefully be approximated using ramped oxidation (RO), which involves measuring the quantity of CO₂ evolved as a sample is increasingly heated at a constant rate in an atmosphere containing oxygen (Carnett et al., 2023). The energy required to thermally-evolve CO₂ is expected to be related to the energy required for biological degradation of OC, with CO₂ evolved at low temperatures deemed to be from more reactive soil OC pools than CO₂ evolved at higher temperatures (Peltre et al., 2013). The age of the OC reactivity pools can be examined by collecting the evolved CO₂ from set temperature ranges and measuring the ¹⁴C (age) content (Garnett et al., 2023; Plante et al., 2013). These can be compared to the ¹⁴C content of the CO₂ that is evolved biologically during incubations of equivalent samples to determine whether the age of the most biologically- and thermally-reactive OC pools match, or not (Plante et al., 2011).

The ¹⁴C content of the thermal reactivity pools also provides insight into the turnover time of each pool, with past research showing that the oldest soil organic matter (OM) (most depleted ¹⁴C content) tends to dominate the most thermally stable fractions (Bao et al., 2019; Plante et al., 2013; Stoner et al., 2023). Similar results have been found for saltmarsh soils (Luk et al., 2021). The ¹⁶C content of the thermal reactivity pools can also provide insight as to whether the source

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123 2023).

Here, we present the first measurements of the ¹⁴C and ¹⁴C content of CO₂ derived from saltmarsh soils using ramped oxidation, and the first comparison of these to the ¹⁴C content of biologically

of OC has an influence on turnover time. Previous work shows that the ¹³C content of evolved CO₂

tends to be more enriched at higher temperatures due to greater contributions from ¹³C-enriched,

degraded/microbially derived C (Luk et al., 2021; Sanderman and Grandy, 2020; Stoner et al.,

evolved CO₂ from the same soils (Houston et al., 2024). We hypothesised that the pre-aged,

allochthonous CO2 respired from saltmarsh soils in Houston et al. (2024) was from a thermally

128 labile source, and that the thermally recalcitrant OC pools would be predominantly composed of

129 older OC.

2. Methods

2.1. Field site and sample collection

Three saltmarsh soil cores (T1-3) were retrieved ca. 30 m apart from the lower marsh zone from Skinflats (SK), an estuarine saltmarsh in Scotland (56° 3'34.04"N, 3°43'59.16"W), as detailed in Houston et al. (2024b2024). Field methods and laboratory sub-sampling procedures are described in detail in Houston et al. (2024b2024). Briefly, the cores were split into 1 cm thick slices as follows: core T1 (0-1 cm, 5-6 cm, and 18-19 cm); T2 (0-1 cm, 5-6 cm, and 15-16 cm), and T3 (0-1 cm, 5-6 cm, and 19-20 cm) (with the deepest sample from each core being the deepest retrieved sample from the 20 cm length of the corer. On the occasions when a full core was not retrieved, the deepest retrieved soil was used).). Each slice was subsequently divided to provide sample material for the RO procedure, and for aerobic laboratory incubations from which the biologically evolved CO₂ was collected for ¹³C and ¹⁴C analysis (Houston et al., 2024b2024).

2.2. Ramped oxidation

÷The RO sub-samples were individually dried to constant mass before milling to a fine powder to homogenise and limit potential shielding effects from aggregates. <u>Unlike most RO and RPO</u> studies (e.g., Hemingway et al., 2017b), we did not remove carbonates from our samples. Acid treatment, which is required to remove carbonates from samples has been demonstrated to result in losses from the labile OC fraction (Bao et al., 2019a). A loss of labile OC for our samples could seriously impact the interpretations in our study, and our ability to compare the ¹⁴C content of the CO₂ respired from bulk (untreated) soils in the incubation experiments (Houston et al., 2024b) to the ¹⁴C content of the RO thermal fractions.

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The samples were then sent to the NEIF Radiocarbon Laboratory for the RO procedure, which is described in Garnett et al. (2023). The RO procedure involved two stages, a first combustion to determine the relationship between the rate of CO2 evolution and temperature (thermogram), and a second combustion where evolved sample gases were collected across defined temperature ranges, for subsequent isotope analysis. For the first combustion, ca. 200 mg of dried and homogenized sample material was weighed into a quartz vial which was inset into a quartz combustion tube, which was subsequently placed into a furnace set initially to room temperature. The furnace was(2023). In brief, the samples were progressively heated at a constant rate of 5°C per minute from ambient room temperature to 800°C in a stream of high purity oxygen (N5.5, BOC, UK). Heating caused combustion of the sample and the evolution of gas which was passed into a second quartz combustion tube containing platinised wool in a furnace set to a constant temperature of 950°C. The platinised wool acted as a catalyst to ensure complete combustion of the evolved gases. Upon exiting the secondary combustion chamber the sample passed through a glass tube containing magnesium perchlorate desiccant to remove moisture and subsequently the CO2 concentration of the gas was and the evolved CO2 measured using a non-dispersive infrared CO2 sensor (SprintIR®-WF-5, Gas Sensing Solutions, UK). The sample was then passed out of the sensor unit and vented to the atmosphere.

The measured CO₂ concentration (normalised for sample mass) was plotted against temperature to produce thermograms which were used to identify temperature ranges, which defined C thermal. Temperature ranges, which defined OC reactivity pools for this study: , were identified from the resulting thermograms:150-325°C_{*}(t1), 325-425°C_{*}(t2), 425-500°C_{*}(t3), 500-650°C_{*}(t4), and 650-800°C.

For each sample, the required mass of material to evolve sufficient CO_2 (> 3 mL) for ^{14}C measurement was calculated based on the thermogram. A new sub-set from the original dried and homogenised sample (t5). The RO procedure was then re-run following the RO procedure outlined above, but instead of venting to atmosphere, after its measurement the evolved CO_2 was collected into foil gas bags based on the defined temperature ranges. CO_2 was with new sample material and the thermally evolved CO_2 collected for ^{13}C analysis from 650-800 °C, but sufficient CO_2 was evolved for ^{14}C analysis from this thermal fraction for only one sample (T1 0.5 cm, Table A1) and we do not consider this fraction further because it is likely dominated by carbonates and not relevant to the purpose of this study.

The foil gas bags (5 L Spout Pouch, https://www.pouchshop.co.uk/) used for sample collection were sealed with one-hole rubber bungs into which a 0.6 cm diameter x 5 cm length stainless

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steel tube was inserted. Isoversinic tubing (Saint Gobain, France) was fitted over the stainless steel to connect it to a quick coupling (Colder Products Company, USA), which allowed connection to the RO kit.

Prior to the RO CO₂ collection, all equipment was cleaned using a standardised procedure

Prior to the RO CO₂ collection, all equipment was cleaned using a standardised procedure (Garnett et al., 2023). All glassware was combusted at 900°C for a minimum of two hours, and all couplings and connectors were washed in carbon-free detergent (Decon) and rinsed in Milli-Q water. The foil gas bags were cleaned by repeatedly (3 times) filling with ca. 1 L high purity nitrogen gas (Research Grade 99.9995% purity, BOC, UK) and evacuating with an air pump, over a period of at least 24 hours (to aid out-gassing of CO₂). The final evacuation, immediately before connecting to the RO rig, involved pumping out the bags with an SBA-5 CO₂ analyser (PPsystems, USA) to ensure that the bags did not contain significant contamination. Before commencing a sample combustion, the entire RO rig was checked for leaks and other potential sources of contamination by measuring the CO₂ concentration in the oxygen carrier gas exiting the kit, using the SBA-5 CO₂ analyser.

Within 3 days of combusting a sample, the evolved gas in each foil bag was connected to a vacuum rig for and 14C analyses from the pre-defined temperature increments. The 13C and 14C analyses of these CO2 samples followed the same methodology at the same laboratory as in Houston et al. (2024). Briefly, following cryogenic recovery of pure sample CO2 by passing it through slush (-78°C; dry ice and industrial methylated spirits) and then liquid nitrogen (-196°C) traps, under high vacuum (ca. 3 x 10⁻³ millibars). The sample CO₂ was then split into three aliquots: One for δ^{13} C analysis using isotope ratio mass spectrometry (IRMS; Delta V, Thermo-Fisher, Germany), one for graphitisation and subsequent AMS ¹⁴C analysis, and one for an archive back-up. The purification, the recovered sample CO2 was graphitised AMS samples were measuredand analysed for 14C content at the SUERC AMSScottish Universities Environmental Research Centre Accelerator Mass Spectrometry (AMS) Laboratory (see Ascough et al., 2024). $\underline{\text{The}^{13}\text{C}}. \text{A sub-sample of the recovered CO}_{2} \\ \underline{\text{was analysed for}^{13}\text{C}} \\ \text{content (} \\ \delta^{13}\text{C-VPDB)} \\ \underline{\text{was using}} \\ \underline{\text{was analysed for}^{13}\text{C}} \\ \underline{\text{content (}} \\ \delta^{13}\text{C-VPDB)} \\ \underline{\text{was using}} \\ \underline{\text{content (}} \\ \delta^{13}\text{C-VPDB)} \\ \underline{\text{was analysed for}^{13}\text{C}} \\ \underline{\text{content (}} \\ \delta^{13}\text{C-VPDB)} \\ \underline{\text{was analysed for}^{13}\text{C-VPDB)}} \\ \underline{\text{content (}} \\ \underline{\text{content (}} \\ \delta^{13}\text{C-VPDB)} \\ \underline{\text{content (}} \\ \underline{$ isotope ratio mass spectrometry (Thermo-Fisher Delta V, Germany) and used to normalise the 14C results to a δ^{13} C of -25 ‰ to correct for isotopic fractionation. Following convention, ¹⁴C results are presented as %Modern (fraction modern x 100) and conventional radiocarbon ages (years BP, where 0 BP = AD 1950 and age = $-8033 \times Ln \text{ (}\%\text{Modern/100)}\text{)}.$

2.3. Data Analysis

Continuous activation energy distributions were modelled from thermograms using the 'rampedpyrox' package in Python V3.8 (Hemingway, 2016; Hemingway et al., 2017b). The

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rampedpyrox model calculates mean activation energies (μΕ) and the standard deviation of activation energy (σΕ), which is a measure of the heterogeneity of bond strength, for each temperature fraction which CO₂ was collected from. Mean μΕ, σΕ and activation energy distribution (p (o,E)) are also calculated for each sample using the *rampedpyrox* model. We do not use the *rampedpyrox* model for calculation of isotope values as it applies a blank correction to ¹⁴C (Hemingway et al., 2017a, b) which is not relevant to the analytical set-up for this study (Garnett et al., 2023), and the ¹³C values generated varied significantly from our IRMS measured values (Table A2). Further data analysis and visualisation of thermograms and isotopic data was undertaken using RStudio V4.2.2 (R Core Team, 2022).

3. Results

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3.1. Radiocarbon

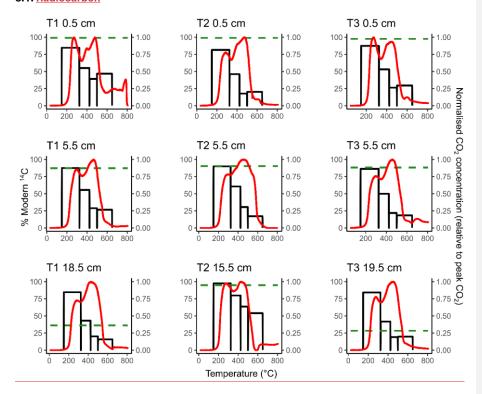


Figure 1. Thermograms

The CO₂ evolved from the RO analysis had bimodal distributions for most samples, with the major peaks occurring at approximately 250°C and 450°C (Fig. 1). These peaks were within t1 (150°C) and 450°C (Fig. 1).

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325°C) and t3 (425-500°C), respectively. We calculated the proportion of CO_2 evolved from the lower temperature CO_2 peak (150-425°C; t1 and t2 combined) and the proportion of CO_2 evolved from the higher temperature CO_2 peak (425-650°C; t3 and t4 combined) (Table A1). There were no significant trends with depth for either T1, T2, or T3 (spearman's rho, p > 0.05). Visually, for both T1 and T3 the size of the second major peak relative to the lower temperature peak increased with depth, whereas for T2 the opposite was the case (Fig. 1).

Most of the samples showed similar trends for CO₂ produced during the ramped combustion procedure, with a lesser CO₂ peak (t2, 325-425°C) between the two larger peaks of t1 and t3 (Fig. 1, see graphical abstract for magnified example). Following the t3 peak (425-500°C) there was another smaller peak in CO₂ evolution between 500-650°C (t4) (Fig. 1). T1 0.5 cm and T3 5.5 cm had high temperature CO₂ peaks between 650-800°C, but this peak was not present for most of the samples (Fig. 1).

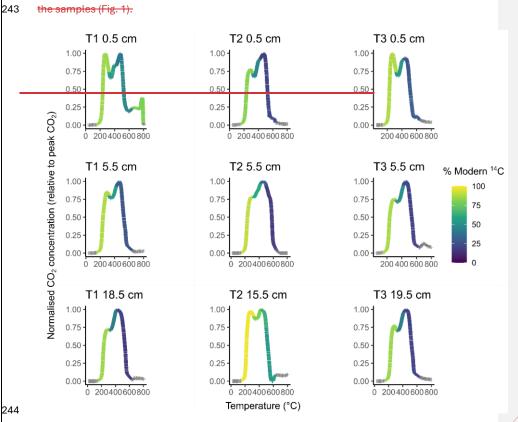


Figure 1- (red lines, right-hand y-axis) overlaying the Thermograms for each of the soil samples. The shading colour indicates the % Modern 14C content of the CO2 evolved throughout the ramped oxidation from

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temperature fractions (black bars, left-hand y-axis) for each sample. The horizontal green dashed

<u>lines represent the 150-325°C, 325-425°C, 425-500°, 500-650°C (and 650-800°C for T1 0.5 cm only).</u> Crey shading

indicates values outside of the CO₂ collection range (150-800°C).

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3.2. 14C content of the CO2 respired from the aerobic incubation experiments of Houston et al.

252 <u>(2024b).ramped oxidation fractions.</u>

253 The ¹⁴C content of the CO₂ evolved during the ramped oxidation (¹⁴C-RO<u>fractions () decreased</u>

exponentially from 150-650°C (t1-t4) (p<0.001, Fig. 12, Table 1) were statistically similar between the 0.5 cm, 5.5 cm, and deepest sample (T1 18.5 cm, T2 15.5 cm, T3 19.5 cm) depth increments

for). This regression was calculated using the mid-point of each of the temperature fractions

(Kruskal-Wallis; p = 0.83, 0.38, 0.66, 0.99, for 150-325°C, 325- 425°C, 425-500°C, 500-650°C,

respectively). There were, however, clear differences range (e.g., t1 mid-point is 237.5°C):

Sufficient CO₂ for ¹⁴C analysis for 650-800°C (t5) was only recovered from one sample (T1 0.5 cm,

Fig. 1, Table 1). This sample did not follow the same decreasing trend in ¹⁴C contents between the

content with increasing temperature fractions, with ranges, as it contained a greater amount of

81.50-97.54^{†4}C (79.75 ± 0.37) % Modern for 150-325 °C, 41.67-79.80) than t2, t3 and t4 (39.18 –

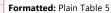
55.02 % Modern for 325-425 °C, 17.67-63.56 % Modern for 425-500 °C, and 15.69-53.96 %

Modern for 500-650 °C (Fig. 1, Table 1).

) (Fig. 1, Table 1).

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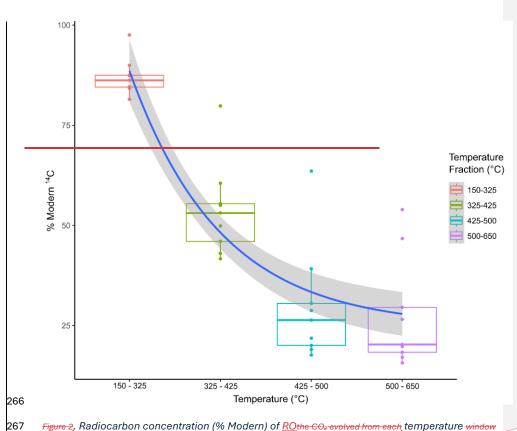


Figure 2, Radiocarbon concentration (% Modern) of ROthe CO₂ evolved from each temperature window during RO (150-325°C, 325-425°C, 425-500°, 500-650°C). The blue line is the exponential regression between temperature and ¹⁴C content ($y = 215.76e^{-0.004n}$, p < 0.001, $R_2 = 0.77$, SE = 1.86, n = 36). The grey shading is the 95 % confidence interval of the regression.

For the entire sample set, the 14 C content of the evolved CO_2 ranged from 97.53 ± 0.50 % Modern for T2 15.5 cm (201 ± 41 years BP) to 15.70 ± 0.12 % Modern for T1 18.5 cm (14,875 ± 61 years BP) (Fig. 2, Table 1, Table A2). There were no significant trends in 14 C content with depth, for example the 14 C contents of the CO_2 evolved from the 150-325°C fraction for the deepest layer (T1 18.5 cm, T2 15.5 cm, T3 19.5 cm), ranged from 84-97 % Modern (1,347-201 years BP), whereas in the surface (0.5 cm) samples it ranged from 81-87 % Modern (1,643-1,085 years BP) (Table 1, Table A2). The CO_2 evolved from the 500-650°C fraction for the deepest layer (T1 18.5 cm, T2 15.5 cm, T3 19.5 cm), ranged from 15-54 % Modern (14,875-4,956 years BP), whereas in the surface (0.5 cm) samples it ranged from 20-47 % Modern (12,826-6,108 years BP) (Table 1, Table A2).

Table 1. fractions and the CO₂ produced in soil Radiocarbon concentration (% Modern) of the CO₂ evolved from each RO temperature fraction and the incubation experiments in Houston et al. (2024b). Errors are

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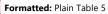
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	o one standard deviat reported in Table A1. (2		n. A sole "C meas	surement for 110.	<u>5 cm 650-</u>	////	Formatted	
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T1 0.5 cm	n 84.62 ± 0.44	55.02 ± 0.29	39.18 ± 0.21	46.75 ± 0.26	79.75 ± ₄	0.37	Formatted	
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T1 5.5 cm	n 87.51 ± 0.43	55.43 ± 0.28	28.76 ± 0.17	26.56 ± 0.16	. I ◆	h	Formatted	
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T1 18.5 cr	m 84.56 ± 0.44	43.06 ± 0.23	20.07 ± 0.13	15.70 ± 0.12		1	Formatted	
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T2 0.5 cm	n 81.50 ± 0.43	46.04 ± 0.24	17.67 ± 0. <u>13<mark>013</mark></u>	20.26 ± 0.14	•	1		
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T2 5.5 cm	n 89.95 ± 0.42	60.55 ± 0.30	30.54 ± 0.17	17.11 ± 0.12			Formatted	
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T2 15.5 cr	m 97.53 ± 0.50	79.80 ± 0.41	63.56 ± 0.31	53.96 ± 0.27			Formatted	
T2 0 5 CH	07 27 + 0 <i>1</i> 5	E2 00 + 0 28	26 27 + 0 15	20 55 + 0 17	Į.		Formatted	
T3 0.5 cm	n 87.37 ± 0.45	53.09 ± 0.28	26.37 ± 0.15	29.55 ± 0.17			Formatted	
T3 5.5 cm	n 86.23 ± 0.42	49.86 ± <u>0.</u> 25	21.87 ± 0.14	18.36 ± 0.12	•		Formatted Table	
TO CIC C	1 00.20 - 0	40.00	Z1.07 - 0	10.00 - 0		ne comme	Formatted	
T3 19.5 cr	m 84.23 ± 0.41	41.67 ± 0.22	19.04 ± 0.13	19.76 ± 0.14	· I	h	Formatted	
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3. <u>2. δ¹³C</u>					Į!	William Control	Formatted	
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There w	vere no significa	<u>ant difference</u>	es in thes. 13C	content of the	<u>.e</u>		Formatted Table	
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ramped	l oxidation fracti	ions <u>(Fig. 2, T</u>	able 2)				Formatted	
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There was	a positive linear relat	ı tionship between	the <u>depth increm</u>	<u>nents (Kruskal-Wal</u>	llis; p = 0.66,		Formatted	
0.63, 0.63	3, 0.44, 0.17, ¹³ C cont	tent of the evolve	ed CO₂ (¹³C-RO) ≀	and temperature (from the RO		Formatted	
	· · etween 150-650°C (p						Formatted	
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the mid-po	oint of each temperat	ture range (e.g., tı	mid-point is 237.	. 5°C).	1		Formatted	
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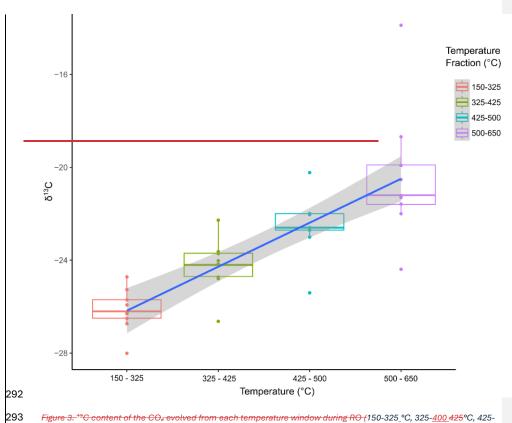
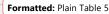


Figure 3. ⁺³C content of the CO₂ evolved from each temperature window during RO (150-325_°C, 325-400_425°C, 425-500_°C, $^{\circ}$ C, 500-650_°C, 650-800_°C respectively). ¹³C contents followed the opposite trend to ¹⁴C contents with temperature, with ranges of °C). The blue line is the linear regression between temperature and ¹³C content (y = 0.017x = 30.39, p<0.001, R_2 = 0.62, SE = 1.72, n = 45). The grey shading is the 95 % confidence interval of the regression.

^{†3}C values of the RO temperature fractions ranged from -28.0 ‰ to -24.7 ‰4.0 ‰ (Table 2). Values for 150-325 °C, -26.6 to -22.3 ‰650°C (t1-t4), the range at which ¹⁴C was also measured (except for 325-425 one ¹⁴C measurement from the 650-800°C, -25.4 to -20.2 ‰ fraction for 425-500 °C, -24.4T1 0.5 cm) ranged from -28.0 ‰ to -13.9 ‰ for 500-650 °C, (Fig. 3, Table 2), and -21.1 to -4.0 ‰values for 650-800 °C (Fig. 2, Table 2).

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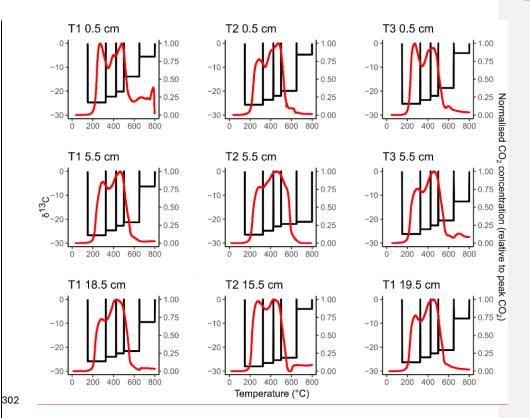


Figure 2. Thermograms (red lines, right-hand y-axis) overlaying the ¹³C content of the RO temperature fractions (black bars, left-hand y-axis) for each sample. Unlike Fig. fraction (t5) ranged from -21-1, we did not attempt % to relate the ¹³C-RO to the ¹³C content of the CO₂ respired in the incubation experiments, due to the potential for microbial fractionation during the incubation experiments.

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-4.0 % (Table 2).

Fable 2, δ¹³C-VPDB‰ signature offer the CO₂ evolved from each RO temperature fractions fraction and the incubation experiments in Houston et al. (2024b). Errors are reported to one standard deviation from the mean. (2024).

δ¹³C-

	*	7	VPDB‰		*	
						Incubations
	150-325°C	325-425°C	425-500°C	500-650°C	650-800°C	(Houston et
1						al., 2024b)
T1 0.5 cm	-24.7 ± 0.1	-22.3 <u>± 0.1</u>	-20.2 <u>± 0.1</u>	-13.9 <u>± 0.1</u>	-5.6 <u> ± 0.1</u>	-23.3 <u>± 0.4</u>
T1 <u>50</u> .5 cm	-26.7 <u> ± 0.1</u>	-24.7 <u> ± 0.1</u>	-22.7 <u>± 0.1</u>	-21.3 <u>± 0.1</u>	-6.3 <u>± 0.1</u>	-23.6 <u>± 0.</u> €
T1 18.5 cm	-25.9 <u> ± 0.1</u>	-24.0 <u>± 0.1</u>	-22.6 <u> ± 0.1</u>	-21.6 <u> ± 0.1</u>	-9.5 <u> ± 0.1</u>	-6.1 <u>± 0.1</u>
T2 0.5 cm	-25.7 <u> ± 0.1</u>	-23.6 <u> ± 0.1</u>	-22.0 <u>± 0.1</u>	-19.9 <u> ± 0.1</u>	-4.7 <u> ± 0.1</u>	-22.9 <u>± 0.</u> ◀
T2 5.5 cm	-26.5 <u> ± 0.1</u>	-24.8 <u>± 0.1</u>	-23.0 <u>± 0.1</u>	-22.0 <u>± 0.1</u>	-21.1 <u>± 0.1</u>	-23.1 <u>± 0.</u> €
T2 15.5 cm	-28.0 <u> ± 0.1</u>	-26.6 <u>± 0.1</u>	-25.4 <u>± 0.1</u>	-24.4 ± 0.1	-4.0 <u>± 0.1</u>	-20.2 <u>± 0.</u> €
T3 0.5 cm	-25.3 <u> ± 0.1</u>	-23.7 <u>± 0.1</u>	-22.0 <u>± 0.1</u>	-18.7 <u>± 0.1</u>	-4.1 <u> ± 0.1</u>	-20.6 <u>± 0.</u> €
T3 5.5 cm	-26.2 <u>± 0.1</u>	-24.2 ± 0.1	-22.6 <u>± 0.1</u>	-20.5 <u>± 0.1</u>	-12.6 <u>± 0.1</u>	-23.4 <u>± 0.</u> €
T3 19.5 cm	-26.3 ± 0.1	-24.2 ± 0.1	-22.6 ± 0.1	-21.2 ± 0.1	-8.0 ± 0.1	-3.7 ± 0.1

3.34. Ramped oxidation and incubation comparison

Figure 14 presents a comparison of the 14 C content of the CO_2 evolved from RO temperature fractions (this study) and respired CO_2 from the same soils during aerobic laboratory incubations (Houston et al. 2024b_2024). These comparisons show that for each of the 0.5 cm depth samples, the 14 C content of the respired CO_2 was greater than the 14 C content of the CO_2 evolved from the same soils in any of the RO temperature fractions in the same soils (Fig. 14). For the 5.5 cm depth samples, the 14 C content of the CO_2 evolved from the 150-325°C RO temperature fraction (Fig. 14). For T2 15.5 cm, the 14 C content of the respired CO_2 was also closest to the 150-325°C RO temperature fraction (Fig. 14). For the T1 18.5 cm and T3 19.5 cm samples, the 14 C contents of the incubation CO_2 were depleted relative to the 150-325°C RO temperature fraction for both samples, and instead, were closest to the 325-425°C and 425-500°C RO temperature fractions, respectively (Fig. 1).4)

3.4. Activation Energy

Mean activation energy (μ E) ranged from 157.50-170.97 kJ/mol for the 0.5 cm depth samples, 159.97-165.32 kJ/mol for the 5.5 cm depth samples, and 154.38-160.44 kJ/mol for the deepest samples (T118.5 cm, T215.5 cm, T319.5 cm. Table 3). The standard deviation of activation energy (σ E) ranged from 23.16-35.83 kJ/mol for the 0.5 cm depth samples, 22.16-25.25 kJ/mol for the 5.5

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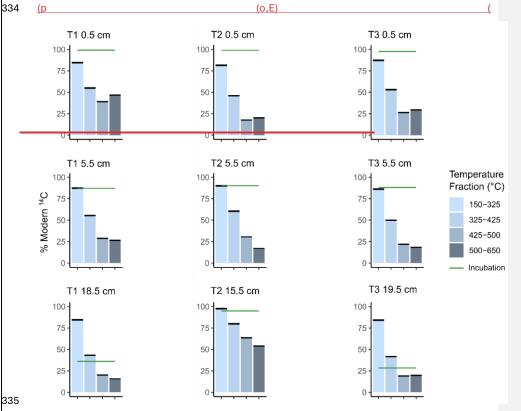


Figure 4. Radiocarbon content (% Modern) of the CO₂ evolved from each temperature window during RO ((150-325°C, 325-425°C, 425-500°, 500-650°C. Blue bars) and incubation experiments (green lines, from Houston et al. (2024)).

<u>Table 1.</u> ANOVA; p = 0.47, 0.37, and 0.14, respectively).

Table 3. Mean activation energy (μE), standard deviation of activation energy (σE), and activation energy distribution for each sample.

	μΕ (kJ/mol)	σE (kJ/mol)	<u>p (oE)</u>
T1 0.5 cm	<u>170.97</u>	<u>35.83</u>	0.02
T1 5.5 cm	159.97	22.16	0.02
T1 18.5 cm	160.44	22.72	0.02
T2 0.5 cm	160.47	23.16	0.02
T2 5.5 cm	165.32	25.25	<u>0.01</u>
T2 15.5 cm	<u>154.38</u>	21.43	0.02
T3 0.5 cm	<u>157.50</u>	<u>24.01</u>	0.02

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T3 5.5 cm	162.31	24.44	0.02
T3 19.5 cm	160.13	23.51	0.02

Table 4 shows μ E and the associated σ E for each thermal fraction. μ E ranged from 131.04-133.23 kJ/mol for 150-325 °C, 156.83-157.78 kJ/mol for 325-425 °C, 176.14-177.79 kJ/mol for 425-500 °C, 185.44-199.19 kJ/mol for 500-650 °C, and 213.06-247.75 kJ/mol for 650-800 °C (Table 4). σ E ranged from 7.33-8.71 kJ/mol for 150-325 °C, 9.83-10.23 kJ/mol for 325-425 °C, 6.88-8.83 kJ/mol for 425-500 °C, 3.68-16.04 kJ/mol for 500-650 °C, and 1.83-10.94 kJ/mol for 650-800 °C (Table 4). μ E and σ E both varied significantly between the thermal fractions, increasing sequentially (Kruskal-Wallis, ρ = 0.001 and 0.001, respectively). We therefore infer that the thermal recalcitrance of RO fractions is greater at higher temperatures and use temperature as a proxy for thermal reactivity herein.

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	_	_	μΕ (σΕ) (kJ/mol)	_	_
	150-325°C	325-425°C	425-500°C	500-650°C	650-800°C
T1 0.5 cm	132.43 (7.33)	157.52 (10.17)	177.79 (7.32)	199.19 (16.04)	242.42 (10.94)
T1 5.5 cm	133.23 (8.07)	157.00 (10.13)	177.07 (7.58)	191.06 (7.87)	213.06 (2.12)
T1 18.5 cm	131.90 (8.62)	157.60 (9.83)	176.93 (7.83)	189.53 (6.70)	239.73 (6.73)
T2 0.5 cm	132.10 (8.21)	157.42 (10.08)	177.23 (7.38)	191.39 (10.62)	226.84 (6.81)
T2 5.5 cm	132.15 (8.71)	157.11 10.01)	177.68 (8.83)	195.80 (7.95)	224.56 (4.55)
T2 15.5 cm	131.04 (8.46)	156.83 (10.23)	176.69 (6.88)	185.44 (3.68)	247.74 (1.83)
T3 0.5 cm	131.59 (7.48)	157.33 (10.07)	176.14 (7.07)	193.65 (12.46)	231.21 (10.04)
T3 5.5 cm	133.05 (8.28)	157.67 (10.14)	177.19 (7.70)	191.13 (9.11)	236.57 (4.29)
T3 19.5 cm	131.73 (8.38)	157.78 (10.00)	176.71 (7.34)	191.7 (10.53)	232.23 (9.69)

4. Discussion

Soils are complex mixtures of many different OC sources and ages, with different vulnerabilities 4.1. to decomposition and turnover. In this study, we aimed to improve our understanding of the carbon cycling of saltmarsh soils by measuring the ¹³C and ¹⁴C content of thermally-fractionated soil carbon pools, and comparing these results to the ¹⁴C content of biologically evolved CO₂ from the same soils (Houston et al., 2024b).

Thermograms

During ramped combustion, CO₂ evolved at low temperatures is deemed to be more energetically favourable for decomposition than CO₂ evolved at higher temperatures, implying that the reactivity of soil OC decreases with increasing temperature (Peltre et al., 2013; Williams and Plante, 2018). Due to the approximately bimodal thermogram distribution (Fig. 1), we can therefore define the low-temperature CO₂ peak as relatively 'labile' and the higher temperature CO₂ peak as 'recalcitrant' OC pools (Capel et al., 2006). There were no significant trends between

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the proportion of CO₂ evolved from the labile (150-425°C) and recalcitrant (425-650°C) RO fractions for either T1, T2, or T3 (Spearman's rho, p>0.05), limiting what we can infer from the distributions of the thermograms. Despite the lack of statistical significance, for both T1 and T3 the height of the higher temperature (recalcitrant) peak relative to the lower temperature (labile) peak increases with increasing depth. This may be caused by OM degradation throughout the burial process during which microbes degrade soil OM, and preferentially deplete the labile OM pool as more favourable for decomposition, which in turn can result in deeper soils having greater proportions of recalcitrant, microbially derived OC (Luk et al., 2021; Soldatova et al., 2024).

Conversely, for T2 the height of the labile peak increased relative to the recalcitrant peak with increasing depth (Fig. 1). This is likely to be because, as well as the soil burial process and the degradation of OM, all samples were within the soil rooting zone (0-40 cm), which can facilitate the transfer of labile OM from the surface to deep soils (Bernal et al., 2017; Rumpel and Kögel-Knabner, 2011). This process may highlight heterogeneity in these saltmarsh soils and in this case may have introduced new labile OM to the deeper soil samples, which was subsequently captured by the RO procedure for the T2 15.5 cm sample, resulting in the increased labile fraction with depth in core T2 (15.5 cm). Due to the low-oxygen conditions in waterlogged saltmarsh soils, decomposition rates are slow, and labile OM can persist for extended time periods compared to aerobic soils (Chapman et al., 2019; McTigue et al., 2021). Hence, it is also feasible that the greater proportion of labile OG at depth for T2 is due to preservation of larger inputs of labile OM compounds at the soil surface during the burial process. The difference in the thermogram distributions with depth between T2, and T1/T3, may highlight heterogeneity across the Skinflats saltmarsh soils, even within the same lower marsh zone.

4.2. 14 C content of ramped oxidation CO₂ fractions

The changes with depth between the thermogram CO₂ peaks follow similar trends to the RO-¹⁴C content of the corresponding temperature fractions (Fig. 1, 4.1. Carbon provenance of ramped oxidation CO₂ fractions

The first three RO temperature fractions (150-325°C, 325-425°C, 425-500°C) were derived solely from OC sources, as IC begins to breakdown from ca. 550°C (Hemingway et al., 2017b). CO_2 from the 500-650°C and 650-800°C fractions may, however, have been evolved from a mix of OC and IC sources. The IC contents of the studied soils (0.11-0.48%) were low relative to OC contents (4.18-7.71%), and IC makes only 1.95-10.48% of the total soil C pool for these samples (Table A3). Wider μ E ranges (mean activation energy of each thermal fraction) and increased bond strength

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diversity (σE) compared to the first three RO fractions (Table 4) may have been caused by non-first order decomposition of carbonates (a form of IC) from 550 °C, as first order decomposition kinetics are a requirement for the *rampedpyrox* model (Hemingway et al., 2017b). Hemingway (pers. comm. 16/01/2025) confirmed that due to the low amounts of carbonates in these samples (Table A3) that it would be appropriate to calculate activation energies using the *rampedpyrox* model

IC could have been removed from our saltmarsh soil samples to allow complete analysis of the soil OC pool, and many R(P)O studies have taken this approach (Bao et al., 2019b; Hemingway et al., 2017b; Luk et al., 2021; Stoner et al., 2023; Williams and Rosenheim, 2015). However, our samples have low IC contents (Table A3), and acid-treatment, which is required to remove IC from samples, can cause losses of labile OC (Bao et al., 2019a). Indeed, in Hemingway et al. (2017), acid treatment of samples prior to RO resulted in a shift of 4 % Modern ¹⁴C, which could change one of our samples from having a pre-bomb ¹⁴C content to a post-bomb ¹⁴C content, or vice-versa. A similar shift in ¹⁴C content for our samples could seriously impact the interpretations in our study, and our ability to compare the ¹⁴C content of the CO₂ respired from bulk (untreated) soils in the incubation experiments (Houston et al., 2024) to the ¹⁴C content of the RO fractions. The soils in the incubation experiments were also not decarbonated as the acid-treatment would have affected soil respiration processes and made the results incomparable to in-situ soil degradation processes (Houston et al., 2024b).

4.2. ¹⁴C content of ramped oxidation CO₂ fractions

Table 1). For T1 and T3, the increasing proportion of evolved CO₂ associated with the recalcitrant (high temperature) peak corresponds to a decrease in RO-¹⁴C content (Fig. 1), i.e., the increase in the amount of CO₂ evolved from a recalcitrant pool corresponds to an increase in its age. For T2, the opposite is observed; the decreasing proportion of the evolved CO₂ associated with the recalcitrant (high temperature) peak corresponds to an increase in RO-¹⁴C content (Fig. 1), i.e., the increase in the amount of CO₂ evolved from the recalcitrant pool corresponds to a decrease in its age. This may suggest the input of a different source of younger but recalcitrant (allochthonous) material in T2. As saltmarsh soil accumulates, and OC is buried and degraded, our data show that the proportion of aged, recalcitrant OC tends to increase with depth, while comparatively younger, labile OC pools also persist.

The ¹⁴C-RO content decreased over the four thermal fractions (150-325 °C, 325-425 °C, 425-500 °C, 500-650 °C. Fig. 1exponentially with increasing temperature (Fig. 2), implying that ¹⁴C-depleted the CO₂ evolved from labile (low temperature) OC had a greater thermal recalcitrance

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than ¹⁴C-enriched OC for these saltmarsh soil samples ¹⁴C content than the CO₂ evolved from recalcitrant (high temperature) OC. Since the ¹⁴C content of eachall RO fractionfractions was <100 % Modern (Fig. 2-Table 1), each of the OC reactivity pools were likely to be predominantly composed of carbon sequestered from the atmosphere before the 1963 ¹⁴C bomb-spike caused by atmospheric nuclear weapons testing, although we cannot completely discount some contributions from post-bomb carbon (Hajdas et al., 2021). Nevertheless, using ¹⁴C content as an estimate of the age of the OC we can infer that the older (¹⁴C-depleted) OC has a greater thermal recalcitrance than young OCcarbon reactivity decreases with increasing age for these samples, which. This finding is consistent with previous studies on the thermal reactivity of carbon stored in soils and sediments (e.g., Bao et al., 2019b; Luk et al., 2021; Plante et al., 2023) (Bao et al., 2019; Luk et al., 2021; Plante et al., 2013; Stoner et al., 2023)

The results suggest inhomogeneity within at least one of the temperature fractions for each sample as, although there were no post-bomb ¹⁴C contents for the incubation or RO samples (Table 1), there is likely to be a fraction of post-bomb (post-AD1955) OC in at least one of the temperature fractions. AutochthonousThis is due to autochthonous OC sequestration (post-bomb) at this accreting saltmarsh (Hajdas et al., 2021; Smeaton et al., 2024) which may become obscured by contributions from pre-bomb (pre-AD1955) OC. Observing the exponential decline in ¹⁴C content with increasing temperature (Fig. 12), we hypothesise that, if present, this mixing of pre- and post-bomb C most likely occurred in the 150-325°C fraction. As ¹⁴C content decreases with increasing temperature for the RO fractions, CO₂ with ¹⁴C content greater than any of the measured RO fractions would be expected to have been evolved within the lowest temperature fraction (150-325°C).

As the oldest (most ¹⁴C-depleted) C had the greatest thermal recalcitrance (Fig. 1), this emphasises that saltmarshes accumulating greater amounts of older (¹⁴C-depleted) OC will likely provide the most thermally recalcitrant OC stores, and saltmarshes accumulating greater proportions of contemporary OC, either through in-situ production or young allochthonous components, contain soil OC stores which are of greater thermal lability (Komada et al., 2022; Van de Broek et al., 2018). However, the ¹⁴C contents of the lowest temperature RO fraction (81-98 % Modern; Table 1) highlight that although the thermal reactivity of OC decreases with ¹⁴C content (Fig. 1), thermally labile OC can still be aged (at least hundreds of years old) for these soils. Due to the often anaerobic and non-eroding conditions of buried sediments, saltmarshes can therefore be stores of old, but thermally labile carbon. Of course, the thermal recalcitrance of OC is not necessarily related to biological turnover time, as this is also dependent on the prevailing environmental conditions (Schmidt et al., 2011; Spivak et al., 2019).

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The 14C-RO contents ranged from 97.53 ± 0.50 % Modern for T2 15.5 cm in the 150-325°C fraction to 15.70 ± 0.12 % Modern for T1 18.5 cm (Table 1), highlighting the role of saltmarshes both as stores of contemporary and highly aged carbon as these 14C contents correspond to conventional radiocarbon years BP (relative to AD 1950) of 201 ± 41 years BP and 14,875 ± 61 years BP, respectively (Table A2). While regional deglaciation of this part of Scotland is likely to postdate the age of 14,875 ± 61 years BP (Ballantyne and Small, 2019), we note that the catchment geology contains sources of petrogenic carbon (Miller et al., 2023), which would be ¹⁴C-dead and may have diluted the ¹⁴C content of the highest temperature RO-CO₂ fractions. As the oldest carbon was stored in the lowest reactivity fraction (Fig. 2), this emphasises that saltmarshes accumulating greater amounts of pre-aged OC will likely provide the most stable OC stores, and saltmarshes accumulating greater proportions of contemporary OC, either through in-situ production or young allochthonous components, contain soil OC stores which are of greater vulnerability for remineralisation and loss to the atmosphere (Komada et al., 2022; Van de Brock et al., 2018), However, the ¹⁴C contents of the 150-325°C fraction (81-98 % Modern) correspond to uncalibrated 14C ages of 201 - 1643 years BP (SI), highlighting that although OC reactivity decreases with age (Fig. 2), the labile OC fraction can still be centuries to millennia in age for these soils and that, due to the often anaerobic and non-eroding conditions of buried sediments, saltmarshes can be stores of old, but reactive, carbon.

4.3. ¹³C content of ramped oxidation CO₂ fractions

¹³C-RO increased sequentially with the thermal fractions (Fig. 2), due to greater contributions from relatively ¹³C-enriched C sources from the higher temperature thermal fractions. The ¹³C-RO contents of the 150-650 °C fractions were each typical of OC sources (Leng and Lewis, 2017), whereas the ¹³C-RO contents of the 650-800 °C fraction were mostly typical of at least a partial contribution from an IC source, with the exception of T2 5.5 cm and T3 5.5 cm (Table 2) (Brand et al., 2014; Ramnarine et al., 2012). As IC can begin to evolve from 550 °C, it is possible that a mix of OC and IC sources was present in the 500-650 °C thermal fractions.

As ¹³C-RO increased with temperature (Fig. 2, Table 2), ¹³C-enriched OC had a greater thermal recalcitrance than ¹³C-depleted OC for these samples. Previous work has demonstrated that >80 % of the OC accumulating at Skinflats saltmarsh is autochthonous/terrestrial in origin (Miller et al., 2023), with limited contributions from marine OC. The thermally recalcitrant OC was potentially composed of a greater amount of OC which has undergone microbial decomposition as this process tends to enrich the degraded OC in ¹³C (Boström et al., 2007; Etcheverría et al., 2009; Luk et al., 2021; Sanderman and Grandy, 2020; Soldatova et al., 2024; Stoner et al., 2023).

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The thermally recalcitrant OC may instead/also have been composed of more different OM compounds (e.g., lignins, aromatics) than the more thermally labile OC (e.g., carbohydrates, lipids) (Sanderman and Grandy, 2020). It is also possible that methodological artefacts, such as kinetic fractionation, influenced the 13 C-RO contents. Kinetic fractionation is explained by different carbon isotopes evolving as CO_2 from the soil sample at different rates during the ramped heating (Hemingway et al., 2017a). Kinetic fractionation would cause the 13 C content of the evolved CO_2 to increase linearly with temperature (Hemingway et al., 2017a), and we cannot rule out this artefact. Hemingway et al. (2017a) determined that kinetic fractionation was not an important factor in their RPO procedure, but we used a different set-up (described in Garnett et al., 2023).

thermally recalcitrant OC (oxidised at high temperatures) tended to be ¹³C enriched compared to labile OC. This implies that the recalcitrant OC was likely to be composed of a greater amount of OC which has undergone microbial decomposition as this process tends to enrich the degraded OC in ¹³C (Boström et al., 2007; Etcheverría et al., 2009; Luk et al., 2021; Sanderman & Grandy, 2020; Soldatova et al., 2024; Stoner et al., 2023). It is also possible that methodological artefacts, such as kinetic fractionation, influenced the ¹³C-RO contents. Kinetic fractionation is explained by different carbon isotopes evolving as CO₂ from the soil sample at different rates during the ramped heating (Hemingway et al., 2017). Kinetic fractionation would cause the ¹³C content of the evolved CO₂ to increase linearly with temperature (Hemingway et al., 2017), as we observed in Fig. 3, so we cannot rule out this artefact.

4.4. Changes in the isotopic content of ramped oxidation CO2 fractions with depth

The isotopic composition of the evolved CO_2 did not vary significantly with depth for any of the temperature fractions. The lack of an increase in the age (14 C-depletion) of soil C with sample depth is unusual, as typically C undergoes a burial process, and previous work has shown diagenetic ageing of saltmarsh soils with depth as young OC is turned over faster than old OC (Komada et al., 2022; Van de Broek et al., 2018).

Compared to other UK saltmarshes, Skinflats has relatively high C accumulation rates (Miller et al., 2023; Smeaton et al., 2024). Depleted ¹⁴C contents of the OC accumulating at the Skinflats saltmarsh (Houston et al., 2024b)The range in ¹⁹C-RO contents between 150-800°C shows a clear distinction between 150-650°C and 650-800°C (Table 2). There was a strong positive linear relationship between ¹⁹C content and temperature for the 150-650°C range (Fig. 3), which was not the case for the 650-800°C range as the ¹⁹C contents of the CO₂ evolved from the 500-650°C

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to the 650-800°C range increases in a non-linear manner (Table 2). This is likely to be because the C pools from 150-650°C are mostly composed of OC, whereas the 650-800°C pool is mostly composed of IC. We deemed this to be the case as the ¹³C contents of the CO₂ evolved between 150-650°C were typical of OC sources (Leng et al., 2006; Leng & Lewis, 2017), whereas the ¹³C contents of the CO₂ evolved between 650-800°C were mostly typical of at least a partial contribution from an IC source, with the exception of T2 5.5 cm and T3 5.5 cm (Table 2) (Brand et al., 2014; Ramnarine et al., 2012).

The CO₂ evolved from 650-800°C for T1 0.5 cm had a-¹³C content of -5.6 ‰, indicating that it was from a predominantly IC source. Given the-¹⁴C content was 79.75 ± 0.37 % Modern, we determined that this was likely to be from a biogenic IC source, potentially shell fragments.

4.4. Comparison of biologically and thermally evolved CO₂

For these saltmarsh soils, thermally recalcitrant OC was—¹³C-enriched and—¹⁴C-depleted compared to thermally labile OC. This implies that labile OC tends to be relatively young compared to recalcitrant OC which tends to be composed of more degraded and aged OC. These findings are consistent with a previous study on the thermal reactivity (using ramped pyrolysis oxidation) of saltmarsh soil OM (Luk et al., 2021) and other soil and sediment OM studies (Sanderman and Grandy, 2020; Stoner et al., 2023).

We did not attempt to relate the ¹³C-RO to the ¹³C content of the CO_2 -respired in the incubation experiments, due to the potential for microbial fractionation during the incubation experiments. Microbial alteration can change the ¹³C content of the respired CO_2 -and the resulting soil OC (Soldatova et al., 2024; Werth and Kuzyakov, 2010), so it is possible that the CO_2 -collected for isotopic analysis in Houston et al. (2024) did not reflect the ¹³C content of the OC pool it was respired from. Therefore, the ¹³C content of the biologically evolved CO_2 and the ¹³C-RO measured in this study are not comparable. We focus the remainder of our discussion on comparing the ¹⁴C content of the biologically evolved CO_2 (Houston et al., 2024) to ¹⁴C-RO measured in this study, which would not be affected by microbial fractionation during the incubation period (¹⁴C results are normalised using the measured S^{13} C values and therefore corrected for isotopic fractionation).

Fig. 4 shows that for each of the 0.5 cm depth samples, the ¹⁴C content of the CO₂ respired in the aerobic laboratory experiments was ¹⁴C-enriched relative to any of the RO temperature fractions, which was also the case for the T3 5.5 cm sample (Table 1). This was likely to be caused by inhomogeneity in the OC reactivity pools, as each defined thermal reactivity pool may be composed of multiple OC sources of variable age and composition. Due to the negative

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exponential relationship between **4CO₂-RO and temperature (Fig. 2), we hypothesise that for soil samples producing respired CO₂ that was **14C-enriched relative to any of the RO fractions (T1 0.5 cm, T3 0.5 cm, T3 5.5 cm; Table 1, Fig. 4), that this CO₂ was biologically-produced from an OC pool within the most thermally labile RO fraction (150-325°C). Thus, we suggest that even within the 150-325 °C RO fraction there are pools of even younger OM, but that they are masked by older, **14C-depleted OM. This implies that RO-**14C analysis of finer temperature fractions could provide further insights into the turnover of young carbon in these soils.

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The **G content of respired CO₂ from the 5.5 cm depth samples tended to be closer to the **G content of the lowest temperature (150-925°G) RO fraction (Fig. 4), implying that for these samples the biologically evolved CO₂ was from a thermally labile OC pool. The T2 15.5 cm respired CO₂ sample was also similar in **G content to the lowest temperature RO fraction, whereas respired CO₂ from the slightly deeper T1 10.5 cm and T3 19.5 cm samples was **G depleted relative to the 150-325°C RO fraction, instead aligning closer to the higher temperature RO fractions (Fig. 4). This implies that the biologically evolved CO₂ from T1 18.5 cm and T3 19.5 cm was not from a thermally labile OC pool. The **G content of the CO₂ evolved from the aerobic incubations of T1 18.5 cm and T3 19.5 cm was hypothesized to have been evolved from an inorganic C source due to the enriched **G contents of -6.1**w and -3.7**w, respectively (Houston et al., 2024). As IC reactivity is controlled by different factors than OC reactivity (Van Dam et al., 2021), and the remainder of the samples were determined to evolve from OC substrates, this is likely to explain why the ***G content of the CO₂ evolved from the aerobic incubation experiments for T1 18.5 cm and T3 19.5 cm did not align with the lowest temperature (most thermally tabile) RO fraction (Fig. 4). Therefore, there was a clear depth trend in the relationship between the ***G

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The depleted ¹⁴C contents of some of the OC accumulating at the Skinflats saltmarsh (201 ± 41 years BP to 14,875 ± 61 years BP; Table A2) imply that a proportion of the OC being buried may already have been pre-aged at the time of deposition on the marsh surface, as the marsh formed in the 1930's (Miller et al., 2023). The combination of high carbon accumulation rates and depleted soil ¹⁴C contents implies that the Skinflats saltmarsh accumulates a high proportion of old, most likely allochthonous OC. Some of the aged, allochthonous OC may have undergone significant microbial processing and degradation prior to its accumulation in the saltmarsh soil. As the OM is degraded, and the energetically favourable components are consumed, the resulting OM becomes increasingly thermally. This means that some of the OC may have undergone significant microbial processing and degradation prior to its accumulation in the

content of CO2 respired in the aerobic incubation experiments and the 14C content of the CO2

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evolved during RO of the same bulk soils.

saltmarsh soil. It is also possible that highly aged OC could remain labile if it had been stored in an environment with low rates of microbial decomposition, e.g., a peatland (Dean et al., 2023). Regardless of the age and degradation state of the OC deposited onto the marsh surface, as it gets buried it will undergo a degree of microbial processing and degradation in the saltmarsh soil (Luk et al., 2021). As the OM is degraded, and the energetically favourable components are consumed, the resulting OM becomes increasingly recalcitrant (Luk et al., 2021; Sanderman and Grandy, 2020; Soldatova et al., 2024). The accumulation of a high proportion of degraded OC on the Skinflats saltmarsh may therefore explain the lack of observed change in the isotopic composition of the soil OC pools with depth. Therefore, at depth we would expect to see an increase in the relative proportion of recalcitrant OC, which we do for T1 and T3 (Fig. 1). The insitu degradation of soil OM may reduce these inhomogeneities with depth as the labile OM components either get consumed or degraded to a recalcitrant (higher RO temperature) state. We did not measure fine-scale changes in RO-14C content within the 150-325°C temperature window for any of the samples, but if this is the case, there would be less of a range of the Contents within the 150-325°C RO fraction for the 5.5 cm and deeper samples than for the 0.5 cm samples,

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Not all old OC is degraded or thermally recalcitrant, and our results show that the Skinflats saltmarsh is also a store of old (14C-depleted), thermally labile OC (Fig. 1). Old OC can be thermally labile if it 'ages' (is stored) in an environment with low decomposition rates, e.g., a peatland (Dean et al., 2023), prior to transport and accumulation into the saltmarsh. There are extensive peatlands in the Skinflats catchment, many of which are degrading (Lilly et al., 2012). Regardless of the age and degradation state of the OC deposited onto the marsh surface, as it gets buried it will undergo a degree of microbial processing and degradation in the saltmarsh soil (Luk et al., 2021), but that process is potentially less prevalent at Skinflats than saltmarshes accumulating younger, less degraded OC.

Through isotopic analysis of saltmarsh soils partitioned using ramped oxidation, we have determined that increased thermal recalcitrance is related to older (14C-depleted; Fig. 1), more degraded/microbially derived (13C-enriched; Fig. 2) soil C. These findings are consistent with previous research on the thermal reactivity of soil and sediment C, that more energy is required (higher temperature/µE) to decompose older (14C-depleted), degraded/microbially derived (13C-enriched) C than younger (14C-enriched), less processed (13C-depleted) C (e.g., Bao et al., 2019b;

Plante et al., 2013; Stoner et al., 2023), including one saltmarsh study (Luk et al., 2021).

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4.5. Comparison of biologically and thermally evolved CO₂

¹⁴C-depleted OC.

As the biological turnover time of OC depends on the prevailing environmental conditions as well as thermal reactivity (Schmidt et al., 2011), the isotopic composition of the most biologically- and thermally-reactive saltmarsh soil OC pools may not be the same. To determine if this is the case, or not, we compared the isotopic composition of the RO thermal reactivity fractions to the isotopic composition of the CO_2 that was evolved biologically during incubations of equivalent samples (Houston et al., 2024b) (Fig. 1).

Figure 1 shows that for each of the 0.5 cm depth samples, the 14 C content of the CO $_2$ respired in the aerobic laboratory experiments was 14 C-enriched relative to any of the RO temperature fractions, which was also the case for the T3 5.5 cm sample (Table 3). The relative 14 C-enrichment of the biologically respired CO $_2$ compared to the thermally evolved CO $_2$ was likely caused by inhomogeneity in the OC thermal reactivity pools, as each defined thermal reactivity pool may be composed of multiple OC sources of variable age and composition. As thermal recalcitrance is related to 14 C-depletion for these samples (Fig. 1), we hypothesise that for saltmarsh soil samples producing respired CO $_2$ that was 14 C-enriched relative to any of the RO fractions (T1 0.5 cm, T2 0.5 cm, T3 0.5 cm, T3 5.5 cm; Table 1, Fig. 1), that this CO $_2$ was biologically-produced from an OC pool within the most thermally labile RO fraction (150-325°C). Thus, we suggest that even within the 150-325 °C RO fraction there are pools of even younger OC, but that they are masked by older.

The ¹⁴C content of respired CO₂ from the 5.5 cm depth samples tended to be closer to the ¹⁴C content of the lowest temperature (150-325°C) RO fraction (Fig. 1), implying that for these samples the biologically evolved CO₂ was from a thermally labile OC pool. The T2 15.5 cm respired CO₂ sample was also similar in ¹⁴C content to the lowest temperature RO fraction, whereas respired CO₂ from the slightly deeper T1 18.5 cm and T3 19.5 cm samples was ¹⁴C-depleted relative to the 150-325°C RO fraction, instead aligning closer to the higher temperature RO fractions (Fig. 1). The biologically evolved CO₂ from T1 18.5 cm and T3 19.5 cm was therefore not from a thermally labile OC pool. The ¹⁴C content of the CO₂ evolved from the aerobic incubations of T1 18.5 cm and T3 19.5 cm was hypothesized to have been derived from an inorganic C source due to the enriched ¹³C contents of -6.1‰ and -3.7‰, respectively (Houston et al., 2024b). As IC biological turnover times are controlled by different factors than OC (Van Dam et al., 2021), and the remainder of the samples were determined to evolve from OC substrates, this is likely to explain why the ¹⁴C content of the CO₂ evolved from the aerobic incubation experiments for T1 18.5 cm and T3 19.5 cm did not align with the lowest temperature

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(most thermally labile) RO fraction (Fig. 1). Therefore, there was a clear depth trend in the relationship between the ¹⁴C content of CO₂ respired in the aerobic incubation experiments and the ¹⁴C content of RO fractions of the same bulk soils. Degradation of the thermally labile OM components to a more thermally recalcitrant state during burial may reduce the inhomogeneities within the most thermally labile RO fraction for this study.

For seven out of nine samples (T1 18.5 cm and T3 19.5 cm being the outliers), the 14 C content of the CO_2 evolved from the aerobic laboratory incubations was closest to the 14 C content of the 150-325°C RO temperature fraction. Therefore, even though the CO_2 evolved from the aerobic incubation experiments was determined to be from a predominantly aged, allochthonous OC source (Houston et al., 2024b)(Houston et al., 2024), it can now also be shown to be derived from a predominantly thermally labile OC pool (Fig. 1).

We did not attempt to relate the 13 C-RO to the 13 C content of the CO_2 respired in the incubation experiments, due to the potential for microbial fractionation during the incubation experiments 4). The results from our study suggest that saltmarshes can be stores of old, labile OC which can change the 13 C content of the respired CO_2 and the resulting soil OC (Soldatova et al., 2024; Werth and Kuzyakov, 2010). In contrast, 14 C results are normalised using the measured δ^{13} C values and are therefore immune to such isotopic fractionation effects.

4.6. Implications

Our results show that aged (presumed allochthonous), thermally labile OC stored in saltmarsh soils remains vulnerable to remineralisation and loss to the atmosphere upon habitat drainage. Saltmarsh soils usually exist in low-oxygen, tidally-inundated conditions which slow decomposition of OC (Chapman et al., 2019), but many saltmarshes globally have been in oxic conditions, e.g. when a saltmarsh is drained (and their soils subsequently oxidised) to convert them for land uses such as housing developments and agriculture (Bromberg and Bertness, 2005; Campbell et al., 2022; Morris et al., 2012). In the Forth Estuary, where the Skinflats saltmarsh is located, as much as 50% of the intertidal area has been converted to agricultural land since 1600, often involving the drainage of saltmarshes (Hansom and McGlashan, 2008). or the soils disturbed:

Protecting saltmarshes from degradation following drainage is listed as an eligible activity for generating carbon credits for blue carbon ecosystem (BCE) projects (VERRA, 2023) and there is significant potential for climate mitigation by avoided emissions from protecting vulnerable stocks of soil OC in BCEs (Goldstein et al., 2020; Griscom et al., 2017; Kwan et al., 2025; Sasmito et al., 2025). Similarly, the re-creation of saltmarsh habitat through managed realignment (rewetting by tidal inundation) of historic saltmarsh habitats which were previously reclaimed for

697 land use purposes (e.g., agriculture) could reduce (and possible reverse) the emissions of aged 698 OC to the atmosphere, both locally to Skinflats, and globally. 699 The evidence for the respiration of thermally labile, allochthonous OC from saltmarsh soils in a 700 drainage degradation scenario demonstrates that at least this fraction of allochthonous OC 701 should be counted as additional in carbon crediting projects and National GHG Inventories. 702 Because allochthonous OC can account for up to 90 % of saltmarsh soil carbon (Komada et al., 703 2022), the inclusion of allochthonous OC (or even a fraction of it) would significantly increase the 704 climate mitigation awarded to blue carbon projects (as carbon credits, or contributions to 705 National GHG Inventories) (Houston et al., 2024a). 706 As the bioavailable OC respired in the experiments of Houston et al. (2024b) was (in most cases) 707 from a predominantly thermally labile OC pool, and ¹⁴C-RO decreased (C became older) with 708 increasing temperature (thermal recalcitrance), RO measurements could be useful for 709 characterising the turnover times of OC pools for saltmarsh soils exposed to oxic conditions 710 (drainage degradation scenario). The use of thermally defined OC pools to characterize OC 711 turnover times for saltmarsh soils would require a modelling advancement to constrain 712 degradation rates and residence times. Such efforts are not within the scope of this study but 713 could inform additionality/permanence in these saltmarsh systems. Experimentally defined 714 turnover times of OC thermal reactivity pools could, for example, provide a more robust approach 715 than inclusion/exclusion of allochthonous OC from saltmarsh 'blue carbon' projects (Houston et 716 al., 2024a). 717 Further research is needed to determine if the relationship between biological and thermal 718 lability exists for different degradation scenarios such as nutrient enrichment, as OC turnover 719 time depends on the environmental conditions as well as the thermal lability of the OC pools. 720 Similarly, these experiments would need to be replicated for a wider range of saltmarshes (high 721 and low latitude saltmarshes, different typologies), as there are likely to be differences in OC 722 turnover in different systems. 723 The samples used for this study were from the low marsh zone only, but it is likely that the thermal 724 reactivity of the Skinflats saltmarsh soil C will vary spatially across the marsh, as the proportion 725 of OC sources has been shown to be variable across saltmarshes (Middelburg et al., 1997). Given 726 our findings that old (14C-depleted) OC has greater thermal recalcitrance than young (14C-727 enriched) OC (Fig. 1), we anticipate that higher marsh zones, which typically have greater 728 proportions of autochthonous OC than lower marsh zones (Spohn et al., 2013), would contain a 729 greater proportion of thermally labile OC. However, it is important to recognise that some of the

young (14C-enriched), autochthonous OC in saltmarsh soils can also be thermally recalcitrant. As well as marsh zonation, we expect that the proportion of OC sources (and associated mix of thermal reactivities) would also vary with proximity to marsh creeks which redistribute autochthonous and allochthonous C across the saltmarsh habitat (Middelburg et al., 1997; Reed et al., 1999). In previously published work we showed that Skinflats accumulates OC of a much greater 'age' (depleted soil ¹⁴C contents) than two other saltmarshes in Scotland (Houston et al., 2024b).

In this paper we have determined that age (14C-content) is related to the thermal recalcitrance of saltmarsh soil OC. We therefore speculate that sites accumulating younger OC would have more thermally labile soil OC than sites accumulating older OC, like Skinflats, with wider implications for the risks to these vulnerable stores of soil carbon from human disturbances.

5. Conclusions

This is the first study on saltmarsh soils to employ the ramped oxidation method. We show that old (14C-depleted (up to 14,875 years BP) carbon dominates the thermally recalcitrant OC pools. The thermally , whereas the labile OC pools are also aged (14C-depleted) compared to the contemporary atmosphere but are composed of younger than the thermally recalcitrant OC pools. (201-1843 years BP) carbon. These results highlight the role of saltmarshes as mixed stores of both old, thermally recalcitrant OC, as well as younger, thermallyold, labile OC.

We present the first comparison of the bioavailability (CO_2 evolved from incubation experiments; Houston et al., 2024)) and the thermal reactivity (RO) of saltmarsh soil OC. We show that preaged, allochthonous CO_2 evolved from saltmarsh soils exposed to oxic conditions (Houston et al., 2024b)(Houston et al., 2024) are from a predominantly thermally labile OC pool. As saltmarsh soils exist mostly in low oxygen, waterlogged conditions, management interventions to limit their exposure to elevated oxygen availability may protect and conserve these stores of thermally labile OC and provide a climate abatement service. Therefore, we recommend that thermally labile allochthonous OC stored in saltmarsh soils should be counted as additional in some carbon crediting projects and National GHG Inventories.

Appendix A

Table A1. Additional ¹⁴C measurement from the 650-800 °C. ¹⁴C was measured at the Scottish Universities Environmental Research Centre Accelerator Mass Spectrometer (AMS) Laboratory. δ^{13} C (relative to Vienna PDB standard) was measured using isotope ratio mass spectrometry on a Delta V (Thermo, Germany) and used to normalize the ¹⁴C results to a δ^{13} C = -25‰, which

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762 763 were reported as %Modern ¹⁴C (i.e., Fraction modern × 100). Errors are reported to one standard deviation from the mean.

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 $\label{eq:table A1. Percentage of CO_second during ramped oxidation of each sample from the labile (150-425^{\circ}\text{C}) and recalcitrant 388 (425-650^{\circ}\text{C}) temperature ranges.}$

Sample ID	% Modern 14C Labile (150-	Recalcitrant (425-650°C)	•
-	425°C)		
Skin T1 0.5 cm 650-800 °C	79.75 ± 0.5059.54	40.46	•
T1 5.5 cm	62.99	37.01	
T1 18.5 cm	60.12	39.88	
T2 0.5 cm	57.53	42.47	
T2 5.5 cm	50.00	50.00	
T2 15.5 cm	64.07	35.93	
T3 0.5 cm	64.97	35.03	
T3 5.5 cm	59.61	4 0.39	
T3 19.5 cm	64.86	35.1 4	

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770 771 772 Table A2. Isotopic compositions measured by IRMS (δ^{13} C) compared to values estimated by the rampedpyrox model (Hemingway, 2016). Modelled and measured δ^{13} C values are significantly different (Mann-Whittney-U test, p = 0.04).

Table A2. Conventional radiocarbon age (years BP, where 0 BP = AD 1950 and age = -8033 x Ln (%Modern/100)) for each of the samples. Measurement errors are reported to one standard deviation (1 σ). We also report the amount of CO₂ evolved from each temperature fraction from which CO₂ was collected for ¹⁴C measurement, reported as %Carbon.

	δ ¹³ C	δ ¹³ C	
	(measured)Radio	(modelled)Radi	
	carbon Age (years	ocarbon Age 1 σ	%Carbon
Sample ID	BP)	uncertainty	4
		42	<u>-30.</u> 1 <u>±</u> ◀
Skin T1 0-1cm.5 cm 150-325 °C	<u>-24.7 ± 0.</u> 1 ,341		<u>0.2</u> . 42 ◆
			1.17
Skin T1 0-1cm.5 cm 325-425 °C	<u>-22.3 ± 0.1</u> 4,800	<u>-27.8 ± 0.2</u> 43	
			1.07
Skin T1 0-1cm.5 cm 425-500 °C	<u>-20.2 ± 0.17,527</u>	<u>-25.6 ± 0.2</u> 42	
		44	/
Skin T1 0-1cm.5 cm 500-650 °C	<u>-13.9 ± 0.1</u> 6,108		<u>-19.5 ±</u> 0.2 69 ◀
		37	
Skin_T1 0 <u>-1cm.5 cm</u> _650-800_°C	<u>-5.6 ± 0.1,818</u>		<u>-11.1 ± 0.253 </u>
			1.44
Skin T1 5 <u>-6cm.5 cm</u> 150-325 °C	<u>-26.7 ± 0.</u> 1 ,072	-27.7 ± 0.240	<i>\</i>
			1.59
Skin T1 5 <u>-6cm.5 cm</u> 325-425°C	<u>-24.7 ± 0.1</u> 4,740	<u>-25.8 ± 0.2</u> 41	
			1.11
Skin T1 5 <u>-6cm.5 cm</u> 425-500 °C	<u>-22.7 ± 0.1</u> 10,011	<u>-23.8 ± 0.2</u> 46	
			0.67
Skin T1 5 <u>-6cm.5 cm</u> 500-650 °C	<u>-21.3 ± 0.1</u> 10,650	<u>-22.6 ± 0.2</u> 48	
Skin T1 5-6cm 650-800			
<u>°C</u> <u>-6.3</u>	± 0.1		

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Skin_T1 18-19cm.5 cm	1.48	- //
150-325°C -25.9 ± 0.1,347 -27.3 ± 0.242		///
		1.43
Skin_T1 18 <u>-19cm</u> .5 cm 325-425 °C -24.0 ± 0.16,769	-25.4 ± 0.2 43	4
	53	-24.1± ◆
Skin_T1 18-19cm-5 cm 425-500 °C -22.6 ± 0.112,900		0.2 .31
<u> </u>		0.62
Skin_T1 18-19cm-5 cm 500-650 °C -21.6 ± 0.114,875	-23.3 ± 0.2 61	4
Skin T1 18-19cm 650-	20.0 - 0.201	
800 °C -9.5 ± 0.1 -11.0 ± 0.2		
Skin T2 0-1cm-5 cm	1.45	4
150-325°C -25.7 ± 0.1,643 -27.9 ± 0.242		
130-025 ₁ 0 <u>-23.7 - 0.1,040 -27.3 - 0.242</u>		1.30
Skin_T2 0-1cm-5 cm 325-425 °C -23.6 ± 0.1,230	-25.9 ± 0.2 42	1.00
<u>-23.0 ± 0.1,230</u>	<u>-23.9 ± 0.2</u> 42	1.44
Skin T2 0-1cm-F-cm 425-500 °C 22.0 ± 0.142-024	-24 2 + 0 2 50	1.44
Skin T2 0-1cm.5 cm 425-500 °C -22.0 ± 0.113,924	-24.3 ± 0.2 58	0.59
Skin T2 0 10m F cm F00 6F0 9C 10.0 ± 0.110 000	22 4 + 0 255	0.08
Skin T2 0-1cm -5 cm 500-650 °C -19.9 ± 0.112,826 Skin T2 0-1cm 650-800	-22.4 ± 0.2 55	
°C -4.7 ± 0.1 -7.2 ± 0.2		
	1.09	
Skin T2 5-6cm.5 cm 150-325 °C -26.5 ± 0.1851 -27.4 ± 0.238	7.00	
150-325°C -26.5 ± 0.1851 -27.4 ± 0.238		1.13
Skin T0 F Com F om 20F 40F 90	25.0 + 0.240	1.10
Skin T2 5-6cm. 325-425°C -24.8 ± 0.14,030	<u>-25.8 ± 0.2</u> 40	1.12
Older TO F Oness Frame 40F F00 00	04.0 + 0.040	1.12
Skin_T2 5-6cm.5 cm 425-500 °C -23.0 ± 0.19,528	<u>-24.0 ± 0.2</u> 43	◆ \ \
		1.10
A		1.10
Skin,T2 5-6cm-5 cm,500-650,°C -22.0 ± 0.114,184	-23.1 ± 0.2 56	1.10
Skin T2 5-6cm-5 cm, 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800		1.10
Skin T2 5-6cm-5 cm 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800 -21.1 ± 0.1 -22.3 ± 0.2	-23.1 ± 0.2 56	1.10
Skin_T2 5-6cm-5 cm, 500-650, °C -22.0 ± 0.114,184 Skin_T2 5-6cm 650-800 °C -21.1 ± 0.1 -22.3 ± 0.2 Skin_T2 15-16cm.5 cm,		1.10
Skin T2 5-6cm-5 cm 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800 -21.1 ± 0.1 -22.3 ± 0.2	-23.1 ± 0.2 56	
Skin_T2 5-6cm-5 cm_500-650 °C -22.0 ± 0.114,184 Skin_T2 5-6cm 650-800 -21.1 ± 0.1 -22.3 ± 0.2 Skin_T2 15-16cm-5 cm_1 -28.0 ± 0.1201 -26.9 ± 0.241	-23.1 ± 0.256 2.66	1.10 2.60
Skin_T2 5-6cm:5 cm_500-650 °C -22.0 ± 0.114,184 Skin_T2 5-6cm 650-800 °C -21.1 ± 0.1 -22.3 ± 0.2 Skin_T2 15-16cm:5 cm_1 -28.0 ± 0.1201 -26.9 ± 0.241 Skin_T2 15-16cm:5 cm_325 °C -26.6 ± 0.1,813	-23.1 ± 0.2 56	
Skin_T2 5-6cm:5 cm_500-650 °C -22.0 ± 0.114,184 Skin_T2 5-6cm 650-800 °C -21.1 ± 0.1 -22.3 ± 0.2 Skin_T2 15-16cm:5 cm_1 -28.0 ± 0.1201 -26.9 ± 0.241 Skin_T2 15-16cm:5 cm_325-425 °C -26.6 ± 0.1,813 Skin_T2 7272_15-	-23.1 ± 0.256 2.66 -25.5 ± 0.241	2.60
Skin_T2 5-6cm:5 cm_500-650 °C -22.0 ± 0.114,184 Skin_T2 5-6cm 650-800 °C -21.1 ± 0.1 -22.3 ± 0.2 Skin_T2 15-16cm:5 cm_1 -28.0 ± 0.1201 -26.9 ± 0.241 Skin_T2 15-16cm:5 cm_325-425 °C -26.6 ± 0.1,813 Skin_T2 72/15-16cm:5 cm_425-	-23.1 ± 0.256 2.66	
Skin T2 5-6cm:5 cm 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800 °C -21.1 ± 0.1 -22.3 ± 0.2 Skin T2 15-16cm:5 cm 325-425 °C -26.6 ± 0.1;813 Skin T2 15-16cm:5 cm 325-425 °C -26.6 ± 0.1;813 Skin Tr2 72, 15-16cm:5 cm 425-500 °C -24.3 ± 0.2;640	-23.1 ± 0.256 2.66 -25.5 ± 0.241	2.60 2.12
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	-23.1 ± 0.256 2.66 -25.5 ± 0.241 40	2.60
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	-23.1 ± 0.256 2.66 -25.5 ± 0.241	2.60 2.12
Skin T2 5-6cm 5 cm 500-650 °C	-23.1 ± 0.256 2.66 -25.5 ± 0.241 40	2.60 2.12
Skin T2 5-6cm:5 cm 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800 °C -21.1 ± 0.1 -22.3 ± 0.2 Skin T2 15-16cm:5 cm 150-325 °C -28.0 ± 0.1201 -26.9 ± 0.241 Skin T2 15-16cm:5 cm 325-425 °C -26.6 ± 0.1;813 Skin T2 ₹2 15-16cm:5 cm 425-500 °C -25.4 ± 0.1 -24.3 ± 0.2;640 Skin T2 ₹2 15-16cm:5 cm 500-650 °C -24.4 ± 0.1;956 Skin T2 15-16cm 650-800 °C -2.9 ± 0.2	-23.1 ± 0.256 2.66 -25.5 ± 0.241 40 -23.6 ± 0.240	2.60 2.12
Skin T2 5-6cm.5 cm 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800 -21.1 ± 0.1 -22.3 ± 0.2 Skin T2 15-16cm.5 cm 150-325 °C -28.0 ± 0.1201 -26.9 ± 0.241 Skin T2 15-16cm.5 cm 325-425 °C -26.6 ± 0.1,813 Skin T2 72 15-16cm.5 cm 425-500 °C -25.4 ± 0.1 -24.3 ± 0.2,640 Skin T2 72 15-16cm.5 cm 500-650 °C -24.4 ± 0.1,956 Skin T2 15-16cm 650-800 °C -4.0 ± 0.1 -2.9 ± 0.2 Skin T3 0-1cm.5 cm	-23.1 ± 0.256 2.66 -25.5 ± 0.241 40	2.60 2.12
Skin T2 5-6cm:5 cm 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800 °C -21.1 ± 0.1 -22.3 ± 0.2 Skin T2 15-16cm:5 cm 150-325 °C -28.0 ± 0.1201 -26.9 ± 0.241 Skin T2 15-16cm:5 cm 325-425 °C -26.6 ± 0.1;813 Skin T2 ₹2 15-16cm:5 cm 425-500 °C -25.4 ± 0.1 -24.3 ± 0.2;640 Skin T2 ₹2 15-16cm:5 cm 500-650 °C -24.4 ± 0.1;956 Skin T2 15-16cm 650-800 °C -2.9 ± 0.2	-23.1 ± 0.256 2.66 -25.5 ± 0.241 40 -23.6 ± 0.240	2.60 2.12 0.83
Skin T2 5-6cm 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800 °C -21.1 ± 0.1 -22.3 ± 0.2 Skin T2 15-16cm.5 cm 150-325 °C -28.0 ± 0.1201 -26.9 ± 0.241 Skin T2 15-16cm.5 cm 325-425 °C -26.6 ± 0,1,813 Skin T2 72 15- 16cm.5 cm 425- 500 °C -25.4 ± 0.1 -24.3 ± 0.2,640 Skin T2 72 15-16cm.5 cm 500-650 °C -24.4 ± 0.1,956 Skin T2 15-16cm 650- 800 °C -4.0 ± 0.1 -2.9 ± 0.2 Skin T3 0-1cm.5 cm 150-325 °C -25.3 ± 0,1,085 -27.3 ± 0.241	-23.1 ± 0.256 2.66 -25.5 ± 0.241 40 -23.6 ± 0.240 1.98	2.60 2.12
Skin T2 5-6cm.5 cm 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800 -21.1 ± 0.1 -22.3 ± 0.2 Skin T2 15-16cm.5 cm 150-325 °C -28.0 ± 0.1201 -26.9 ± 0.241 Skin T2 15-16cm.5 cm 325-425 °C -26.6 ± 0.1,813 Skin T2 72 15-16cm.5 cm 425-500 °C -25.4 ± 0.1 -24.3 ± 0.2,640 Skin T2 72 15-16cm.5 cm 500-650 °C -24.4 ± 0.1,956 Skin T2 15-16cm 650-800 °C -4.0 ± 0.1 -2.9 ± 0.2 Skin T3 0-1cm.5 cm	-23.1 ± 0.256 2.66 -25.5 ± 0.241 40 -23.6 ± 0.240	2.60 2.12 0.83
Skin T2 5-6cm.5 cm, 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800 °C -21.1 ± 0.1 -22.3 ± 0.2 Skin T2 15-16cm.5 cm 150-325 °C -28.0 ± 0.1201 -26.9 ± 0.241 Skin T2 15-16cm.5 cm, 325-425 °C -26.6 ± 0.1,813 Skin Tr272 15- 16cm.5 cm, 425- 500 °C -25.4 ± 0.1 -24.3 ± 0.2,640 Skin T2 T2-16cm.5 cm, 500-650 °C -24.4 ± 0.1,956 Skin T2 15-16cm 650- 800 °C -4.0 ± 0.1 -2.9 ± 0.2 Skin T3 0-1cm.5 cm 150-325 °C -25.3 ± 0.1,085 -27.3 ± 0.241 Skin T3 0-1cm.5 cm, 325-425 °C	-23.1 ± 0.256 2.66 -25.5 ± 0.241 40 -23.6 ± 0.240 1.98 -25.7 ± 0.242	2.60 2.12 0.83
Skin T2 5-6cm 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800 °C -21.1 ± 0.1 -22.3 ± 0.2 Skin T2 15-16cm.5 cm 150-325 °C -28.0 ± 0.1201 -26.9 ± 0.241 Skin T2 15-16cm.5 cm 325-425 °C -26.6 ± 0,1,813 Skin T2 72 15- 16cm.5 cm 425- 500 °C -25.4 ± 0.1 -24.3 ± 0.2,640 Skin T2 72 15-16cm.5 cm 500-650 °C -24.4 ± 0.1,956 Skin T2 15-16cm 650- 800 °C -4.0 ± 0.1 -2.9 ± 0.2 Skin T3 0-1cm.5 cm 150-325 °C -25.3 ± 0,1,085 -27.3 ± 0.241	-23.1 ± 0.256 2.66 -25.5 ± 0.241 40 -23.6 ± 0.240 1.98	2.60 2.12 0.83
Skin T2 5-6cm 5 cm 500-650 °C	-23.1 ± 0.256 2.66 -25.5 ± 0.241 40 -23.6 ± 0.240 1.98 -25.7 ± 0.242 -24.1 ± 0.247	2.60 2.12 0.83
Skin T2 5-6cm.5 cm, 500-650 °C -22.0 ± 0.114,184 Skin T2 5-6cm 650-800 °C -21.1 ± 0.1 -22.3 ± 0.2 Skin T2 15-16cm.5 cm 150-325 °C -28.0 ± 0.1201 -26.9 ± 0.241 Skin T2 15-16cm.5 cm, 325-425 °C -26.6 ± 0.1,813 Skin Tr272 15- 16cm.5 cm, 425- 500 °C -25.4 ± 0.1 -24.3 ± 0.2,640 Skin T2 T2-16cm.5 cm, 500-650 °C -24.4 ± 0.1,956 Skin T2 15-16cm 650- 800 °C -4.0 ± 0.1 -2.9 ± 0.2 Skin T3 0-1cm.5 cm 150-325 °C -25.3 ± 0.1,085 -27.3 ± 0.241 Skin T3 0-1cm.5 cm, 325-425 °C	-23.1 ± 0.256 2.66 -25.5 ± 0.241 40 -23.6 ± 0.240 1.98 -25.7 ± 0.242	2.60 2.12 0.83

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Skin T3 0-1cm 650-800					
<u>°C</u>	-4.1 ± 0.1	-6.2 ± 0.2			
Skin T3 5-6cm.5 cm			1.13		•
150-325°C	<u>-26.2 ± 0.</u> 1 ,19	0 <u>-27.8 ± 0.2</u>39			
				1.32	
Skin T3 5-6cm.5 cm 325-	-425 <u>°</u> C	<u>-24.2 ± 0.15,591</u>	<u>-25.9 ± 0.2</u> 41		•
				1.08	
Skin T3 5-6cm.5 cm 425-	-500 <u>°</u> C	22.6 ± 0.1 12,211	<u>-24.3 ± 0.252</u>		•
		<u>-20.50 ±</u>		0.58	
Skin T3 5-6cm.5 cm 500-	-650 <u>°</u> C	<u>0.1</u> 13,617	-22.32 ± 0.2 54		4
Skin T3 5-6cm 650-					
800 °C	-12.60 ± 0.1	-14.29 ± 0.2			1
Skin T3 19-20cm.5	<u>-26.30 ±</u>	39			•
cm 150-325°C	<u>0.</u> 1 ,379		<u>-27</u> 1.4	48 <u>± 0.2</u>	4
				1.75	\\\
Skin T3 19-20cm.5 cm 32	25-425°C	24.20 ± 0.1 7,033	-25.40 ± 0.243		4
		-22.60 ±		1.11	11
Skin T3 19-20cm.5 cm 42	25-500 <u>°</u> C	<u>0.1</u> 13,322	-23.78 ± 0.2 55		4
		<u>-21.20 ±</u>		0.64	
Skin T3 19-20cm.5 cm 5	00-650 <u>°</u> C	<u>0.1</u> 13,026	<u>-22.57 ± 0.255</u>		4
Skin T3 19-20cm 650-80	<u>0</u>				
<u>°C</u>	$-8.00 \pm 0.$	<u>-9.30 ± 0.3</u>	<u>2</u>		- 1

Table A3. Soil carbon properties measured on equivalent sub-samples prior to the RO procedure, as reported in Houston et al. (2024). Total organic carbon (TCC), Total carbon (TC) for the soil samples were measured by a SoliTOC analyser (Elementar Analysensysteme, Hanau, Germany). 14 C was measured at the Scottish Universities Environmental Research Centre Accelerator Mass Spectrometer (AMS) Laboratory. δ^{13} C (relative to Vienna PDB standard) was measured using isotope ratio mass spectrometry on a Delta V (Thermo, Germany) and used to normalize the 14 C results to a δ^{13} C = $^{-25}$ ‰, which were reported as %Modern 14 C (i.e., Fraction modern × 100). Errors are reported to one standard deviation from the mean.

Sample ID	TOC (%)	TIC (%)	TC (%)	% Modern ¹⁴ C	<u>δ¹³C</u>
SK T1 0.5 cm	<u>4.1</u>	0.48	4.58	47.49 ± 0.23	-23.5 ± 0.1
SK T1 5.5 cm	4.96	0.11	<u>5.06</u>	45.03 ± 0.20	-24.5 ± 0.1
SK T1 18.5 cm	4.8	0.39	<u>5.18</u>	41.36 ± 0.19	-23.8 ± 0.1
SK T2 0.5 cm	<u>4.71</u>	0.16	4.87	31.47 ± 0.15	-22.2 ± 0.1
SK T2 5.5 cm	4.23	0.13	4.36	43.69 ± 0.21	-24.1 ± 0.1
SK T2 15.5 cm	7.56	0.15	<u>7.71</u>	50.93 ± 0.24	-25.1 ± 0.1
SK T3 0.5 cm	5.37	0.12	5.49	47.03 ± 0.22	-23.7 ± 0.1
SK T3 5.5 cm	4.06	0.11	<u>4.18</u>	44.15 ± 0.21	24.0 ± 0.1
SK T3 19.5 cm	<u>5.23</u>	0.12	<u>5.35</u>	44.48 ± 0.21	-24.1 ± 0.1

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Formatted: Plain Table 5 **Data Availability** 783 784 All data presented in this manuscript is available in the main text and, appendices, and 785 supporting information. **Author Contribution Statement** 786 787 A.H. undertook the study, fieldwork, sample processing, data acquisition, and wrote the first draft 788 of the manuscript. M.G. conducted the laboratory procedures with the help of A.H. A.H., W.A., 789 and M.G. contributed to designing the study, fieldwork, and laboratory analyses. W.A., M.G., and J.S. oversaw the study and contributed to writing and revision of the manuscript. 790 791 **Competing Interests** 792 The authors declare that they have no conflict of interest. 793 Acknowledgements Formatted: Font: +Body (Aptos), 11 pt, Font color: Auto 794 We thank Jo Smith (University of Aberdeen) for her comments and edits on the first draft of this Formatted: Heading 2, Left, Line spacing: single 795 manuscript. We thank the NERC SUPER DTP for funding the PhD through which this research Formatted: Left 796 was undertaken (NE/S007342/1). We acknowledge support from the National Environmental 797 Isotope Facility in funding the_-14C measurements for this study under grant NE/S011587/1 798 (allocation numbers 2594.1022, 2709.1023), WENAWA also acknowledges support provided by Formatted: Font: (Default) +Body (Aptos), Font color: Auto 799 the HORIZON-CL5-2023-D1-02-02 grant C-BLUES, Innovation to advance the evidence base for 800 reporting of Blue Carbon inventories and greenhouse gas fluxes in coastal wetlands. 801 Thanks Finally, thanks are extended to Chloe Bates for assisting with sample collection. Finally, 802 we thank the editor and both reviewers for their comments which have improved this 803 manuscript. 804 **Reference List** 805 References 806 Ballantyne, C. K. and Small, D.: The Last Scottish Ice Sheet, Earth and Environmental Science Formatted: Font: Not Bold 807 Transactions of The Royal Society of Edinburgh, 110, 93-131, https://doi.org/10.1017/S1755691018000038, 2019. 808 809 Bao, R., Zhao, M., McNichol, A., Wu, Y., Guo, X., Haghipour, N., and Eglinton, T. I.: On the Origin 810 of Aged Sedimentary Organic Matter Along a River-Shelf-Deep Ocean Transect, Journal of Geophysical Research: Biogeosciences, 124, 2582-2594, 811 https://doi.org/10.1029/2019JG005107, 2019. 812 Formatted: Header

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