



## 1 **Microbial communities inhabiting 600-year-old sediments in the Inka-Coya Lake located in the** 2 **Atacama Desert**

3 Coral Pardo-Esté<sup>1</sup>; Juan Castro-Severyn<sup>2</sup>; Francisco Remonsellez<sup>2,3</sup>; Antonio Maldonado<sup>4,5</sup>; Inger Heine Fuster<sup>6</sup>; Hector  
4 Pizarro<sup>7</sup>; Adriana Aránguiz-Acuña<sup>4,6\*</sup>.

5

6 <sup>1</sup>Departamento de Ciencias Farmacéuticas, Facultad de Ciencias, Universidad Católica del Norte, Antofagasta, Chile.

7 <sup>2</sup>Laboratorio de Microbiología Aplicada y Extremófilos, Departamento de Ingeniería Química, Universidad Católica del Norte,  
8 Antofagasta, Chile.

9 <sup>3</sup>Centro de Investigación Tecnológica del Agua en el Desierto-CEITSAZA, Universidad Católica del Norte, Antofagasta,  
10 Chile.

11 <sup>4</sup>Millennium Nucleus of Andean Peatlands, AndesPeat.

12 <sup>5</sup>Centro de Estudios Avanzados en Zonas Áridas, CEAZA. La Serena, Chile.

13 <sup>6</sup>Laboratorio de Ecología Acuática, Departamento de Recursos Ambientales, Universidad de Tarapacá, Arica, Chile.

14 <sup>7</sup>Departamento de Ciencias Geológicas Universidad Católica del Norte, Antofagasta, Chile.

15 *Correspondence to:* Adriana Aránguiz-Acuña (aaranguiza@academicos.uta.cl)

16 **Abstract.** Lacustrine sediments are natural archives for the surrounding area's biogeochemical dynamics; in particular, the  
17 isolation and extreme conditions in which desert lakes are located make them ideal study models for studying perturbations in  
18 the ecosystem. Specifically, Inka-Coya Lake is in the Atacama Desert, where the presence of metals and metalloids associated  
19 with the active geological activity and local mining industry is a crucial driver for the biological dynamics in this ecosystem,  
20 as have been suggested for macroinvertebrates and plankton communities in the lake. In this study, we aimed to characterize  
21 the microbial communities that inhabit deep lacustrine sediments and their interaction with the surrounding environment. The  
22 results show that the microbial community from lacustrine sediments contains over 70% unclassified organisms, highlighting  
23 this ecosystem's microbial taxonomic novelty. Our results indicate that the microbial communities cluster in three distinct  
24 zones: a superficial community, an intermediate and mixed community, and a more specialized anaerobic community in the  
25 deeper sediments. The microbial composition is dominated by chemoheterotrophic bacteria strongly associated with methane  
26 metabolism. Additionally, there is statistical evidence of strong correlations between particular taxa such as  
27 Sulfurimonadaceae, Metanoregulaceae, and Ktedonobacteroaceae with elements like Cu, As, Fe, Ni, and V, and magnetic  
28 properties of the surrounding environment. Further detailed studies of the metabolic repertoire of these communities are  
29 necessary to understand the complex dynamics between microbial life and geochemical composition in this fragile and extreme  
30 environment.

31

32 **KEYWORDS:** desert lake, deep lacustrine sediment, microbial communities, extremophiles



## 33 1. INTRODUCTION

34 The Atacama Desert is located on the western slopes of the Central Andes Cordillera between 15 and 30°S at elevations  
35 between sea level and 3,500 m a.s.l., in the driest part of the South American dry diagonal, which extends from 5°S on the  
36 west coast to almost 50°S on the east coast, over 4,000 km with less than 200 mm mean annual rainfall (MAR). The Andes  
37 Cordillera represents a physical barrier that directly modulates climatic conditions and water availability in the Atacama Desert  
38 (Garreaud et al., 2003). Particular environmental conditions of the Atacama, such as high solar radiation, low atmospheric  
39 humidity, and other pressures associated with the natural composition of the desert, directly influence life occurring in these  
40 ecosystems (Demergasso et al., 2008; Albarracín et al., 2020; Kurth et al., 2021; Kereszturi et al., 2020; Borsodi et al.,  
41 2022). Thus, the Atacama Desert water bodies are truly natural laboratories for understanding evolutionary processes, not  
42 only of the geomorphology of the landscape but also of different life forms promoted by environmental forces, such as climate  
43 changes at different time scales (Adrian et al., 2009).

44 Aquatic sediments are sources and/or sinks of elements participating in biogeochemical cycles, including both allochthonous  
45 and autochthonous lake processes, influencing biodiversity and trophic dynamics of water bodies (Trolle et al., 2010;  
46 Fernández et al., 2000; Usenko et al., 2007; Bandowe et al., 2018). Aquatic sediments have an advantage over terrestrial  
47 records, and that is that they accumulate at measurable rates because they are often buffered from physical, chemical, and  
48 biotic disturbances, thus allowing the recording of past environmental conditions (Benito, 2020; Picard et al., 2022; Da Costa  
49 et al., 2023; Yan et al., 2024). A wide variety of abiotic (e.g., bulk density, dry mass, radioactive isotopes, mineralogy, chemical  
50 elements) and biotic proxies (e.g., fossils, species abundance, and presence/absence, resting structures, pigments,  
51 environmental DNA) preserved in the sediments are currently analyzed to reconstruct ecosystem change at timescales ranging  
52 from fine-scale (interannual or decadal) to millennial (Cohen, 2003; Korosi et al., 2017).

53 The central Andes water systems have mainly originated after successive glaciations and volcanic and tectonic activity. In the  
54 Atacama Desert, lacustrine sediments are natural archives holding evidence of past precipitations, dust deposition, anthropic  
55 disturbances, and pollution, mainly due to mining activity that occurs in the area (Grosjean and Veit, 2005; Placzek, 2009;  
56 Cerda et al., 2019; Aránguiz-Acuña et al., 2020). Desert lakes are located in extremely arid and isolated areas, making them  
57 susceptible to perturbations (Valero-Garcés et al., 2003; Pueyo et al., 2011; Grosjean and Veit, 2005) and hosting extreme  
58 forms of life, especially microorganisms that have evolved physiological and life-history adaptations allowing them to perform  
59 in challenging conditions (Dib et al., 2009; Ordoñez et al., 2009; Fariás et al., 2013, 2014; Rasuk et al., 2014). Therefore,  
60 lacustrine sediments are expected to contain a great taxonomic diversity, including low-abundance and highly specialized taxa,  
61 directly influenced by small-scale conditions determining local environments (Borsodi et al., 2022).

62 Inka-Coya Lake (22°20'S-68°35'W, 2534 m.a.s.l.) is located at the eastern margin of the Atacama Desert, close to the Salado  
63 River, in the San Francisco de Chiu-Chiu village, northern Chile. It is a karstic sinkhole developed during the Quaternary  
64 period by the dissolution of calcareous layers of the Chiu-Chiu Formation (El Loa Group). The Atacama Desert and its  
65 surroundings have a particular geological history; metals and metalloids found in the Inka-Coya area include Ti, Al, Fe, Ni,  
66 and Cr; also, As and Sb are associated with the local geological activity (Aszalós et al., 2020; Borsdorf and Stadel, 2015;  
67 Pérez-Portilla et al., 2024), that directly influence the chemical composition of the underground water (Vignale et al., 2021).  
68 The central Atacama Desert, specifically the Antofagasta Region, holds large porphyry copper deposits that support the great  
69 metal-mining industry (Dittmar, 2004; Salvarredy-Aranguren et al., 2008). In 2021, Chile was the world's top copper producer,  
70 producing 5,508,084 tons -26.6% of the world's production- (Rodríguez-Luna et al., 2022). The mining industry in the  
71 Antofagasta region has developed extensively since the 19th century (Dittmar, 2004; Salvarredy-Aranguren et al., 2008) with  
72 increasing impacts on the national economy and development, but also on the health of ecosystems, which have triggered



73 social and environmental conflicts, affecting especially relevant groups such as ancient Indigenous communities from Quechua  
74 and Lickan Antay people (see Ramírez et al., 2005; Tapia et al., 2019).

75 Previous studies aimed to assess mining pollution records on environmental matrices have included the sediment records of  
76 different longitudes from the Inka-Coya Lake, showing variation through the geochemistry and magnetic properties (Cerda et  
77 al., 2019; Aránguiz-Acuña et al., 2020; Pérez-Portilla et al., 2024). Analyzed variables have allowed us to identify episodes  
78 associated with changes in water availability, flash flooding, and evidence of perturbations induced by mining activities.  
79 Overall, the lake is polluted at different degrees of severity with Cu, Sb, Mo, and As, and some elements like Cu and Ni have  
80 been enriched in the most recent periods (Cerda et al., 2019; Pérez-Portilla et al., 2024). Additionally, some biological proxies,  
81 such as macroinvertebrates and diatom communities, were found to be directly influenced by the accumulation of metal(loid)  
82 as observed by changes in assemble composition (Aránguiz-Acuña et al., 2020). Surrounding metal-mining exploitation,  
83 which has been maintained and even increased through the last 200 years, in addition to aridity stable conditions, makes Inka-  
84 Coya Lake an excellent site for understanding biological adaptations of aquatic populations to these anthropic pressures  
85 (Aránguiz-Acuña et al., 2018; 2020).

86 While microbial life in arid ecosystems plays a key role in maintaining biogeochemical cycles (Madsen, 2011), there is a high  
87 proportion of unclassified taxa that hold great interest in poly-extreme environments from an ecological, environmental, and  
88 biotechnological point of view (Farias et al., 2014; Castro-Severyn et al., 2021; Dong et al., 2022). Nevertheless, there are  
89 scarce records in which microbial assemblage has been used as a paleolimnological proxy of the possible responses to long-  
90 term sustained anthropogenic metal stress (Da Costa et al., 2023; Yan et al., 2024). The few studies considering this aim have  
91 focused on changes in primary producers' abundance, under-interpreting the impact on other metabolic functional groups  
92 (Benito et al., 2020; Picard et al., 2022).

93 This study aimed to characterize the microbial community along a lacustrine sediment core obtained from Inka-Coya Lake.  
94 To our knowledge, this is a pioneering study in the microbial characterization of a sedimentary core of this length (136 cm)  
95 and date (600 years) from a lake sediment in the Atacama Desert. Results show that the microbial communities have changed  
96 through time, identifying three clear periods in which alpha and beta diversity has been associated with organic matter content,  
97 magnetic susceptibility, and metals and metalloid concentrations. Additional studies of the metabolic functions of the  
98 microorganisms inhabiting these sediments are required to understand the interactions between microbial life and the  
99 geochemical components of the Inka-Coya Lake further.

## 100 1. METHODS

### 101 2.1 Study site and sampling

102 Inka-Coya Lake (San Francisco de Chiu Chiu village, Antofagasta; 22° 20.300' S; 068°35.981' W, Chile) has a surface area of  
103 500 m<sup>2</sup> and a maximum depth of 18 m, is located in the Pre-Andean Depression of the Antofagasta Region at an elevation of



104 2,520 m a.s.l. (Fig. 1 A, B). Around the lake, the predominant vegetation is of vegas, a type of wetland typical of the Andean  
105 pre-Puna zone, strongly associated with the hydric variability of the emerging groundwaters.

## 106 **2.2 Sediment Core Sampling**

107  
108 A fieldwork campaign was driven in August 2021. The topography of the lake bottom was modeled using the Echo-Map Plus  
109 42CV from Garmin. Afterward, three sediment cores from the depocenter of Inka-Coya Lake, where maximum sedimentation  
110 rates are expected, were obtained. The cores were obtained using a 9.0 cm diameter Uwitec gravity corer. This study shows  
111 the most extended core analysis results, measuring 136 cm (labeled LIC-SHC03). X-ray and photography images were  
112 captured before obtaining sections of sediment subsamples from the core. For the geochemical analyses, sediment sub-samples  
113 every 0.5 cm to a depth of 12 cm were obtained. Then, every 1 cm until the end of the core, totaling 146 sediment samples.  
114 Additionally, sub-samples for every 1 cm interval were obtained to develop the magnetic susceptibility analysis. The cores'  
115 detailed treatment, geochemical analysis, and magnetic properties can be reviewed by Pérez-Portilla et al. (2024).

## 116 **2.3 Sediment Core Dating**

117 The geochronology of the sediment core from Inka-Coya Lake was determined through radiocarbon dating ( $^{14}\text{C}$ ) on the  
118 remaining macroscopic carbon along the record. The measurements were done using accelerator mass spectrometry (AMS),  
119 and the results were corrected for isotopic fractionation with an unreported  $\delta^{13}\text{C}$  value. Subsequently, the age-depth model  
120 for this sedimentary core was established using the Bayesian radiocarbon chronology package Bchron in R, using the 'shcal20'  
121 as the calibration curve (Hogg et al., 2020; Haslett and Parnell, 2008). A detailed description of the procedure is available in  
122 Pérez-Portilla et al. (2024).

## 123 **2.4 Magnetic and Geochemical Properties Analysis**

124 Five grams of each sediment sub-sample were placed into paleomagnetic boxes of  $8\text{ cm}^3$  to measure the mass magnetic  
125 susceptibility ( $\chi$ ) using a Kappabridge MFK1\_FA instrument (AGICO Co) under environmental conditions ( $22\text{--}24^\circ\text{C}$ ) and a  
126 magnetic field of 200 A/m. The samples were measured at a low frequency of 976 Hz ( $\chi_{lf}$  or simply  $\chi$ ) and a high frequency  
127 of 15,616 Hz ( $\chi_{hf}$ ). The magnetic susceptibility dependent on the frequency was calculated using both measurements, as  
128 described by Pérez-Portilla et al. (2024). The  $\chi_{fd}\%$  parameter is used to indicate the presence of magnetic particles near the  
129 limit of the superparamagnetic/single domain (SP/SD) magnetic size (Verosub and Roberts, 1995), which can be linked to the  
130 presence of magnetic particles of authigenic origin (Dearing et al., 1996). Additionally, sub-samples of each 1 cm slice were  
131 dried in an oven at  $50^\circ\text{C}$ . Afterwards, they were homogenized using an agate mortar in the Geochemistry Laboratory of  
132 Universidad Católica del Norte (UCN), Antofagasta, Chile. The sediments were then digested using reverse aqua regia (4 mL  
133 HCl + 12 mL of  $\text{HNO}_3$  + 300 mg of sediment sample) and a microwave digester (Perkin Elmer MPS 320; EPA 3052 method),  
134 following Tapia et al. (2022) in Centro de Investigación Tecnológica del Agua en el Desierto (CEITSAZA-UCN). The  
135 elements aluminum (Al), titanium (Ti), vanadium (V), manganese (Mn), iron (Fe), nickel (Ni), Cu, zinc (Zn), arsenic (As),  
136 molybdenum (Mo), and antimony (Sb) were measured by the inductively coupled plasma atomic emission spectroscopy (ICP-  
137 OES) Perkin Elmer Optima 7000 in the digested residue at CEITSAZA. The organic, inorganic matter, and carbonate contents  
138 were estimated using the loss on ignition (LOI) method, which was assessed at a contiguous 1 cm interval following Heiri et  
139 al. (2001). This procedure involved drying  $1\text{ cm}^3$  of each sediment sample in crucibles at  $105^\circ\text{C}$  for two hours and weighing  
140 them. The dry samples were weighed before heating to  $550^\circ\text{C}$  in a flask over 1.5 h, left at  $550^\circ\text{C}$  for two h, then allowed to



141 cool. The samples were weighed, then the crucibles were transferred to the flask and burned at 925°C. Finally, the crucibles  
142 were weighed again once they cooled.

## 143 2.5 Sediment sample processing and DNA extraction

144 Sediment samples were obtained every 1 cm from the top to the bottom of the core. According to the manufacturer's  
145 instructions, total DNA was extracted from the 250 mg of sediment samples using the DNeasy PowerSoil kit (Qiagen Inc.,  
146 Hilden, Germany). DNA integrity, quality, and quantity were verified by 1% agarose gel electrophoresis and fluorescence  
147 using a Qubit 3.0 fluorometer and the Qubit dsDNA HS assay kit (Thermo Fisher Scientific, MA, USA). Following, DNA  
148 samples were sent to AustralOmics, Chile, for amplification of the bacterial 16S rRNA gene V4 region (~250 bp) using the  
149 515F and 806R primers (Caporaso et al., 2011), construction of 250 bp paired-end libraries and sequencing on a MiSeq  
150 (Illumina) platform.

## 151 2.6 Taxonomic Composition Analysis

152 This analysis was conducted in R v4.0.3 and RStudio v1.3.1093 following the DADA2 v1.16.0 R package pipeline (Callahan  
153 et al., 2016) to infer amplicon sequence variants (ASVs) for each sub-sample. Briefly, the reads were evaluated for quality  
154 control and subsequently trimmed ( $N_s = 0$ ,  $\text{length} \geq 150$  bp,  $\text{expected errors} \leq 2$ ), followed by dereplication, denoising, and  
155 merging of paired reads. Following, an ASV table was built to allow a maximum of two expected errors, removing chimeras  
156 and assigning taxonomy using the Silva v138 database (Quast et al., 2012). Also, all ASVs identified as Eukarya, Chloroplast,  
157 and Mitochondria were removed. A multi-sequence alignment was created to infer phylogeny using FastTree v2.1.10 (Price et  
158 al., 2009), and phyloseq-object (containing the ASVs, taxonomy assignment, phylogenetic tree, and the samples meta-data)  
159 was created using the R package Phyloseq v1.34.0 (McMurdie et al., 2013). Finally, taxa relative abundance and taxonomic  
160 composition at different ranks were visualized using the ggplot2 v3.3.3 (Wickham, 2016), Fantaxtic v0.2.0 (Teunisse, 2022),  
161 and ampvis2 v2.7.4 (Andersen et al., 2018) R packages.

## 162 2.7 Diversity Analysis

163 Alpha diversity metrics (Shannon, Chao, phylogenetic diversity, and Simpson indexes) were calculated for each segment along  
164 the core using the microbiome v1.24.0 (Lahti et al., 2017) and btools v0.0.1 R packages. Also, Wilcoxon statistical tests to  
165 compare means between the identified zones were carried out and visualized using the DESeq2 v1.42.0 (Love et al., 2014) and  
166 ggpubr v0.6.0 (Kassambara, 2017). Moreover, beta diversity was evaluated by principal coordinates analysis using Hellinger  
167 transformed Bray Curtis distances based on the ASV abundance matrix were calculated using Phyloseq v1.34.0 (McMurdie et  
168 al., 2013) and ampvis2 v2.4.5 (Andersen et al., 2018) R package. Also, redundancy analysis (RDAs) was calculated using



169 depth gradient and zone parameters to constrain the multivariate space, and ANOVA tested the statistical significance of the  
170 selected geochemical variables.

171

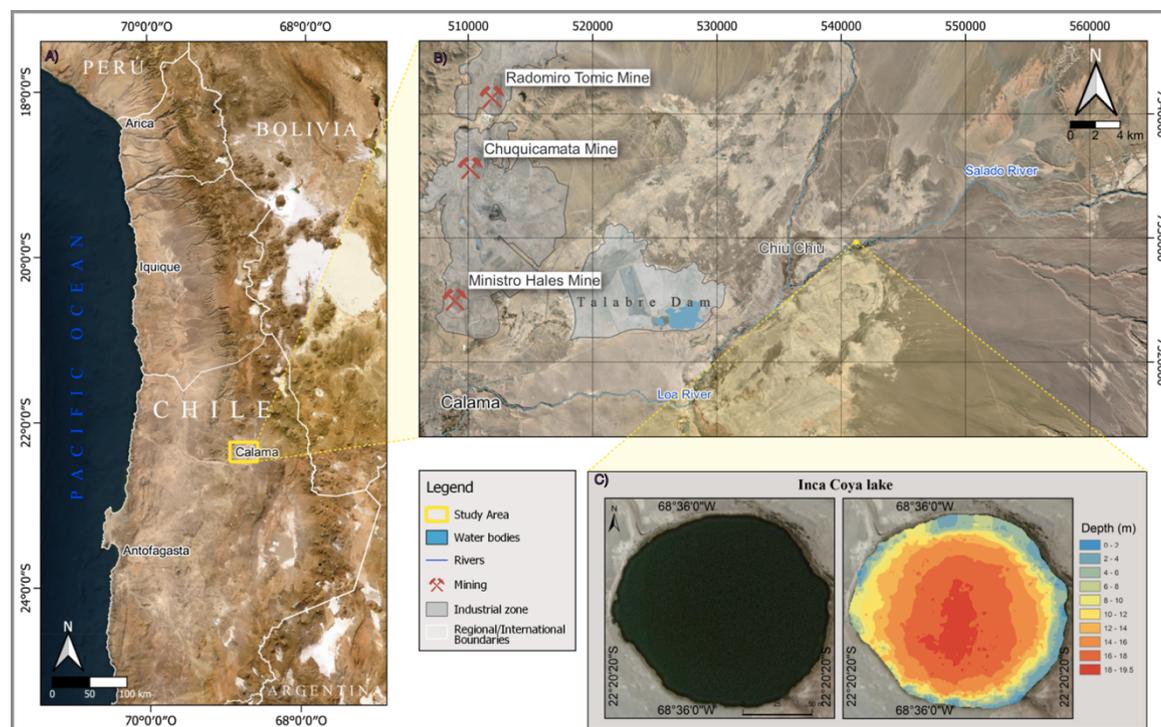
## 172 **2.8 Functional Predictions**

173 Functional potential signatures and metabolic pathways abundances were predicted based on the ASV abundance and  
174 taxonomy matrices using PICRUSt2 v2.4.1 software (Douglas et al., 2020) through the Kyoto Encyclopedia of Genes and  
175 Genomes (KEGG) (Kanehisa et al., 2012) and MetaCyc (Caspi et al., 2018) pathway databases. The analysis of Differential  
176 Abundance represented pathways was calculated using the Kruskal-Wallis test (confidence interval = 0.95) and the Benjamini-  
177 Hochberg correction false-discovery rate using ggpicrust2 v1.7.2 R package (Yang et al., 2023). Also, we use the Functional  
178 Annotation of Prokaryotic Taxa (FAPROTAX) database v1.2.7 (Louca et al., 2016) to map the identified ASVs and quantify  
179 changes in established ecologically relevant functions.

## 180 **1. RESULTS**

181 The samples analyzed correspond to a sediment core from Lake Inka-Coya, located in the Atacama Desert, with a water depth  
182 of 18.5 meters (Fig. 1). The sediment core age-depth model was constructed based on six charcoal sample dates, where the  
183 more superficial at 41 cm corresponds to  $75 \pm 32$  cal years BP, and the deepest found at 94 cm corresponds to an age value of  
184  $505 \pm 22$  cal years BP. Based on the age-depth model constructed, the sediment core of Inka-Coya Lake analyzed had 630  
185 years of age (Fig. 2).

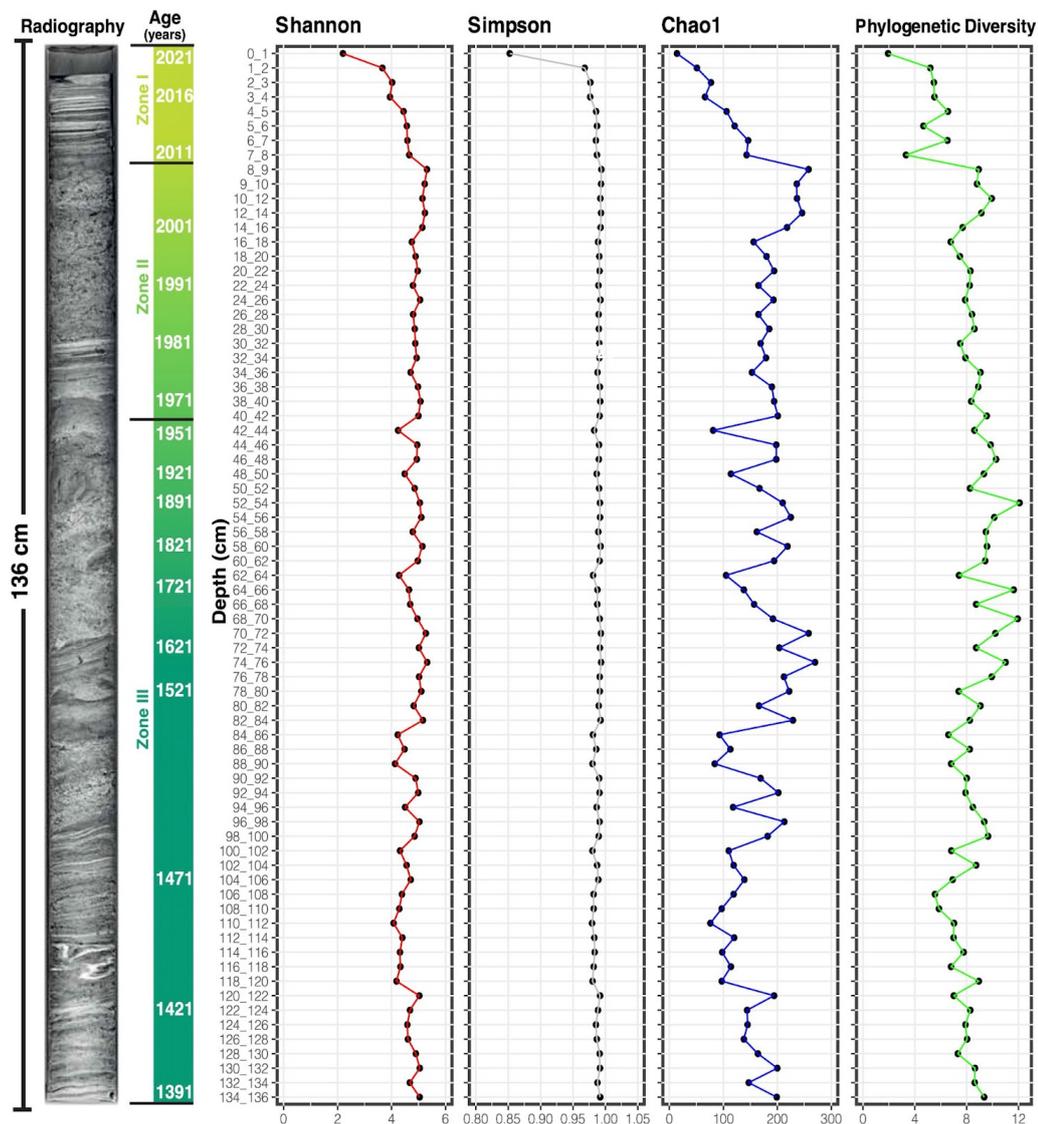
186



187

188 **Figure 1. Sampling site location in Northern Chile (A), Inka-Coya Lake, and important surrounding mining and urban**  
189 **centers (B) and the bathymetry of the lake (C).**

190



191

192 **Figure 2. Diversity of the microbial community of the Inka-Coya sediment core. The core sedimentary radiography,**  
193 **dating, and alpha diversity indices variation along the core depth are displayed.**

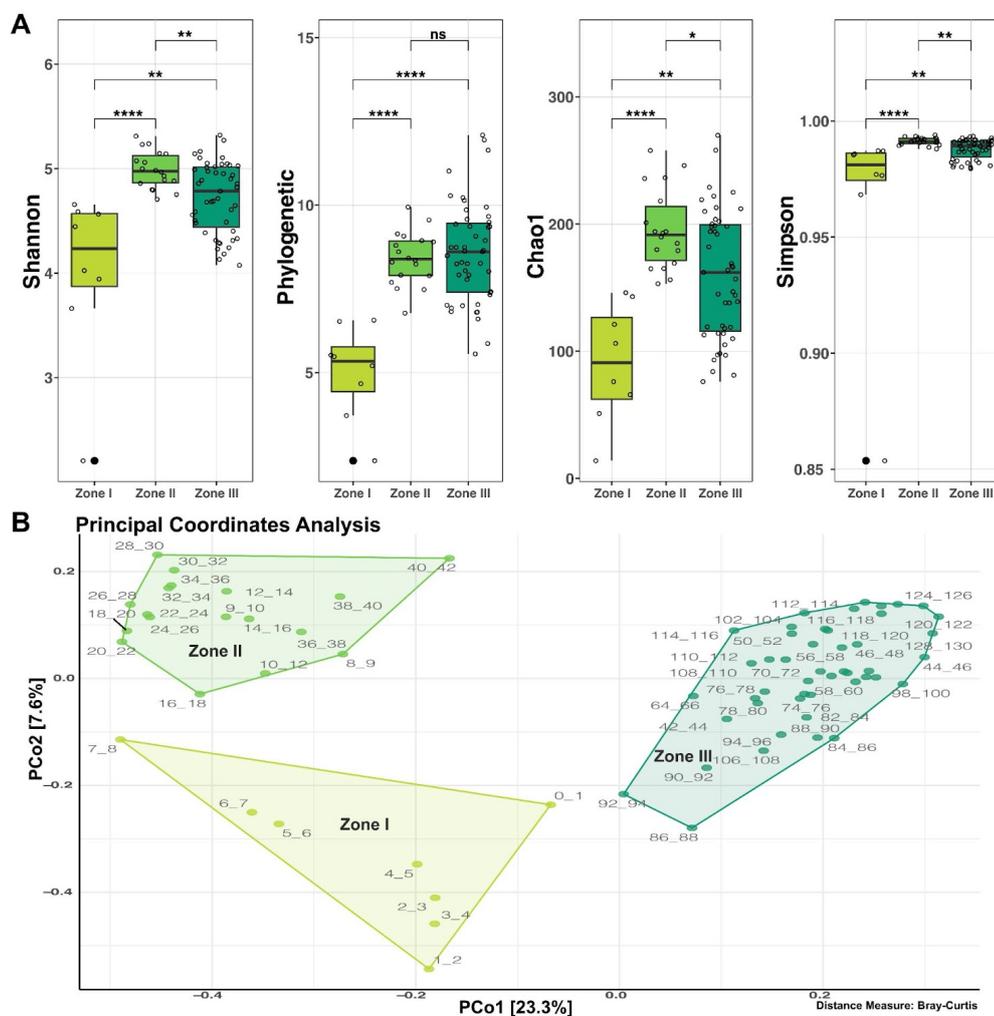
194 The variation in diversity within the sample was measured to determine the changes in the microbial community along the  
195 sediment core by calculating the Shannon, Simpson, Chao, and Phylogenetic indices as standard measures of the taxonomic  
196 diversity within a sample (Thukral, 2017). The microbial community observed along the core was diverse, based on the DNA  
197 samples analyzed. Quantification of diversity showed that it increased with depth, and the Simpson index remained stable after  
198 the 2 cm surface layer. At the same time, Chao1 considered low-abundance taxa, and the phylogenetic index based on the



199 phylogenetic history of the species (Fig. 2). The diversity increases in all cases at two cm long, and maximum values were  
200 observed between 8 and 86 cm, with minor peaks at 96, 120, and 130 cm near the bottom samples.

201 Three distinct zones in terms of microbial taxonomy could be identified and are statistically different for the four evaluated  
202 diversity indices, except between Zone II and III in the context of phylogenetic diversity (Fig. 3A). These three disjunct  
203 clustering zones of microbial community diversity along the sediment core were also identified in the beta diversity analysis,  
204 where zone I includes the less diverse upper layer (0-8 cm), zone II, the middle zone of the core (9-42 cm) with significant  
205 greater diversity, and zone III the deepest are sampled in this study, depicted in three shades of green going from lighter (zone  
206 I) to darker (zone III) in the PCoA clustering analysis (Fig. 3B).

207



208

209 **Figure 3. Clustering of the microbial communities inhabiting the deep sediment of Inka-Coya Lake** A) Statistical  
 210 **differences in alpha diversity between depth zones.** B) **Principal Coordinates Analysis (PCoA) with the ASVs relative**  
 211 **abundance using Bray-Curtis as distance metric; each point corresponds to a community, tagged by depth and colored**  
 212 **by zone.**

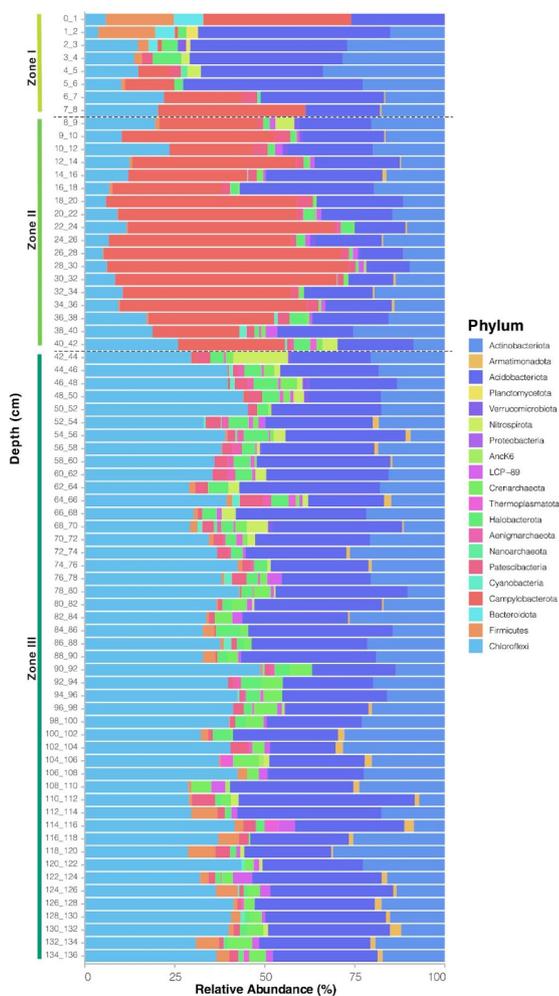
213 The taxonomic composition and abundance along the communities at phylum rank also reflect the clustering on three distinct  
 214 zones, where zone I is dominated by Actinobacteriota and includes a great abundance of Firmicutes in the top layers, and  
 215 Campylobacterota, there is also the presence of Bacteroidota, Halobacteriota, and Plantomycetota. Zone II is more diverse and  
 216 composed mainly of Campylobacterota, Chloroflexi, Acidobacteriota, and Actinobacteriota. While zone III is the largest and  
 217 more homogeneous, composed of several low-abundance taxa, dominated by Chloroflexi, Acidobacteriota, and



218 Actinobacteriota, there is also a higher representation of Crenarchaeota, Nitrospira, Aenigmarchaeota, and Armatimonadota  
219 that in the rest of the zones (Fig. 4).

220 Notably, 76.6% of the taxa could not be identified at the genus level (0% matched any known species). Thus, Figure 5 shows  
221 the abundance at the “best hit,” where Campylobacterota (*Sulfuricurvum*, *Sulfurimonas*), *Mycobacterium*, and *Methanolinea*  
222 dominated the overall community. While in each zone, there are particular taxa associated; for instance, species belonging to  
223 the Aminicenantes Phyla are very common in Zone I; *Pseudarcobacter* is prevalent in Zone II as *Pelolinea* is in Zone III.

224

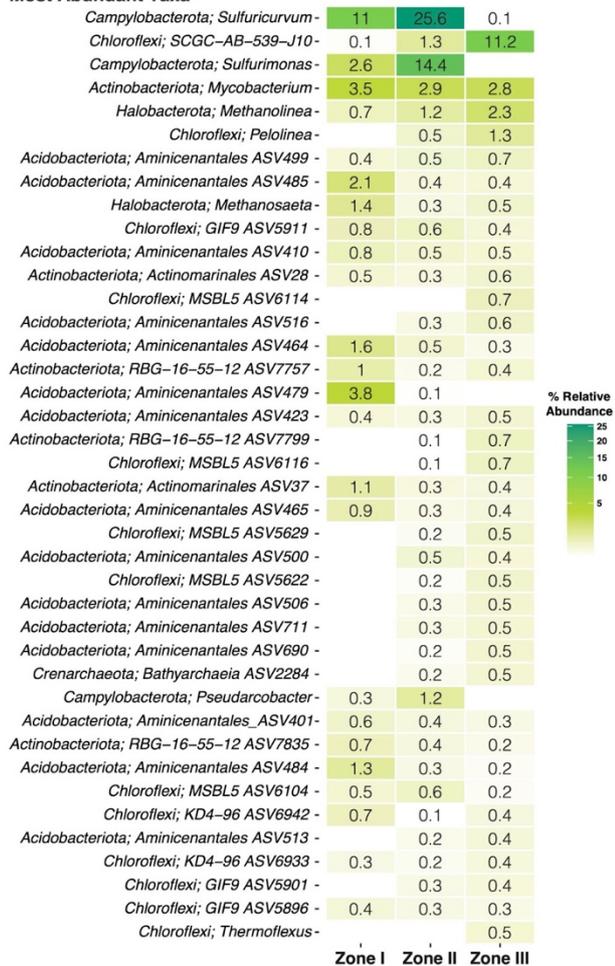


225

226 **Figure 4. Taxonomic composition of the microbial community in the deep sediments of Inka-Coya Lake. Stacked bar**  
227 **of the taxonomic composition at the phylum level.**

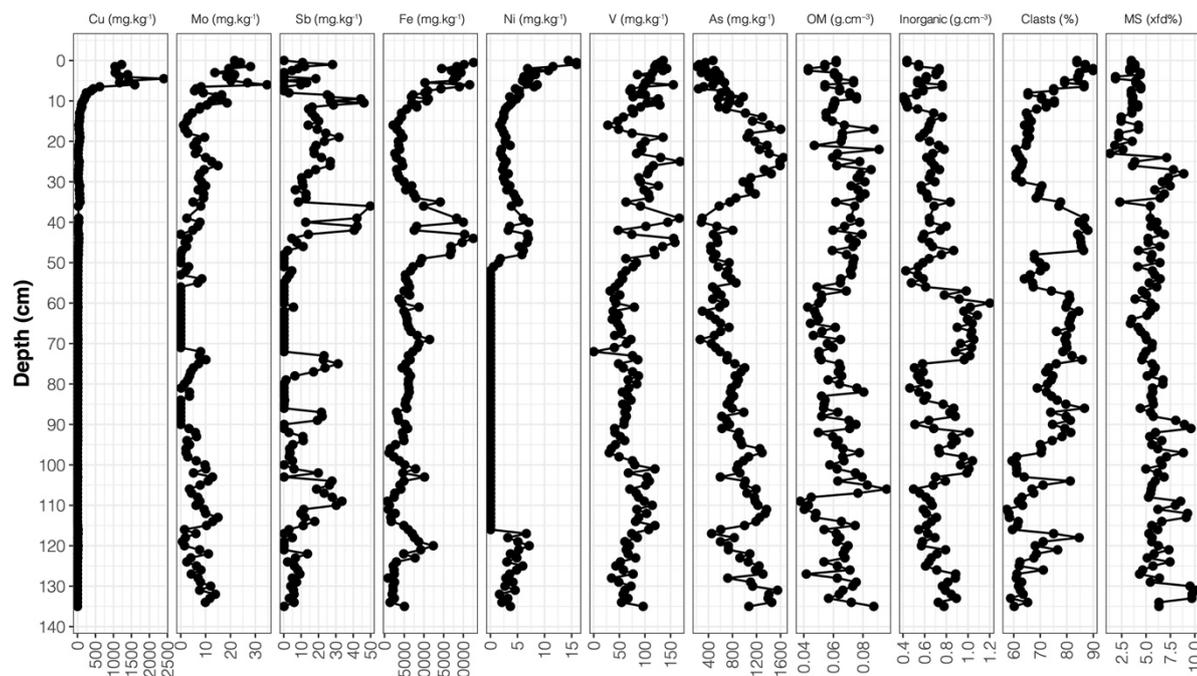


**Most Abundant Taxa**



228

229 **Figure 5. Heatmap of the abundance of the microbial community at the family level. The color gradient indicates the**  
 230 **abundance of the specific taxa**



231

232 **Figure 6. Depth variation of metals and metalloids concentrations ( $\text{mg}\cdot\text{kg}^{-1}$ ), organic matter (OM) and inorganic**  
233 **concentrations ( $\text{g}\cdot\text{cm}^{-3}$ ), clasts percent, and magnetic susceptibility (MS) measured in Inka-Coya Lake sediment core**  
234 **(modified from Pérez-Portilla et al. 2024).**

235 Variations in analyzed sediment properties along the core, such as magnetic susceptibility, organic matter, and  
236 carbonates/clasts composition, are shown in Figure 6. Copper (Cu) and nickel (Ni) were in lower concentration and variability  
237 at greater depths and showed concentration peaks in surface sediments. Elements such as iron (Fe), molybdenum (Mo), and  
238 vanadium (V) also showed top sediment peaks. Still, overall, they had more variable behavior than previously mentioned  
239 elements in the middle and bottom sediments ( $> 40$  cm). Metalloids, arsenic (As), and antimony (Sb) exhibited the highest  
240 concentrations between 10-45 cm depth.

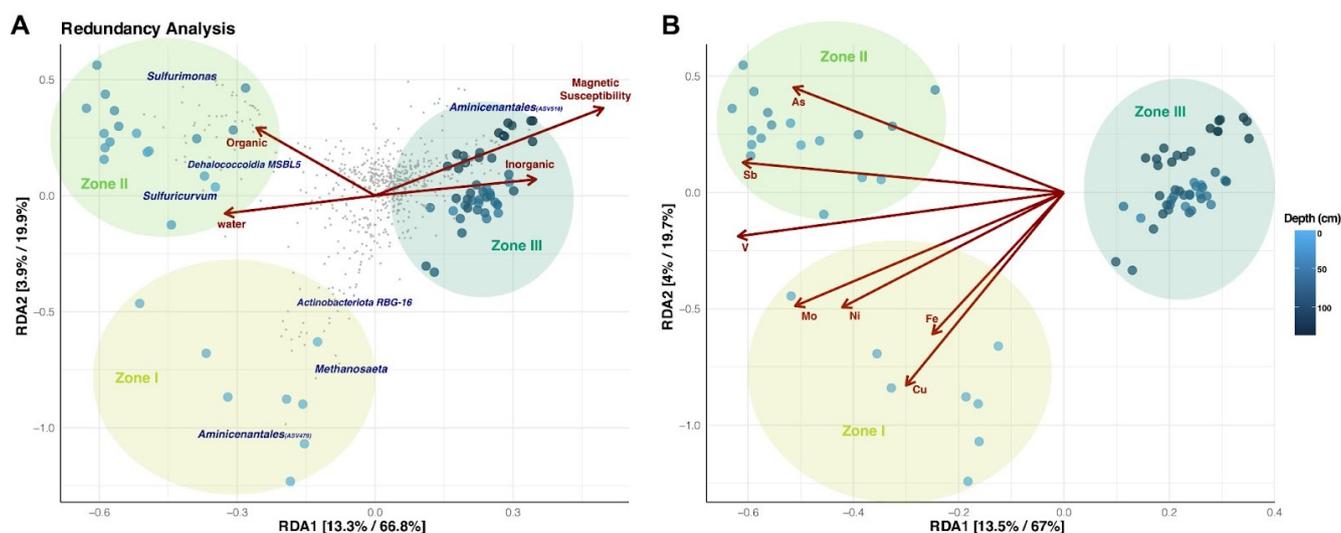
241 The mean organic matter and carbonate contents are around 8.7% and 19.5%, respectively, while the inorganic density (91%)  
242 showed the highest averaged values ( $0.71 \text{ g}\cdot\text{cm}^{-3}$ ). The sediment composition shifts to clay and silt from 28 cm to the top of  
243 the core. The content of clasts was predominant in the inorganic fraction, with 71.8% along the core. The carbonate peaks were  
244 observed at 10-36 cm, 46-56, and more significant and variable proportions below 96 cm, which did not exceed 40%.

245 Magnetic susceptibility (MS,  $\chi$ ) values range from  $-6.09\times 10^{-9}$  to  $8.13\times 10^{-7} \text{ m}^3\cdot\text{kg}^{-1}$ , with an average value of  $2.77\times 10^{-7} \text{ m}^3\cdot\text{kg}^{-1}$ .  
246 Frequency-dependent susceptibility ( $\chi_{fd}\%$ ) values range between 1.31 and 10.17%, with an average of 5.62%. Zone I has



247 the highest values of  $\chi$  and the lowest values of  $\chi_{fd}\%$ , while Zone II shows the lowest  $\chi$  values and intermediate  $\chi_{fd}\%$  values.  
 248 Zone III presents intermediate  $\chi$  values and the highest  $\chi_{fd}\%$  values.

249 Geochemical and magnetic variables are associated with microbial diversity found in the sediment of Inka-Coya Lakes,  
 250 differentiated into zones (I, II, and III). Deeper and older fractions of sediments (dark blue), especially Aminicenantaes, are  
 251 positively influenced by the magnetic susceptibility and inorganic elements in the sediments. Microbial assemblage found in  
 252 the middle sediments (Zone II) of the core is driven by organic matter content and water availability, where taxa like  
 253 *Sulfurimonas*, *Sulfuricurvum*, and *Dehalococcoidia* were the most represented. The middle zone is associated with a significant  
 254 As peak, which suggests that the presence of metal(loid)s positively affects the microorganisms assemblage inhabiting since  
 255 middle-to-superficial layers. In Zone II, microbial diversity is mainly associated with low but stable concentrations of organic  
 256 matter, a more significant proportion of clasts, and the higher peak of As and Sb in the sediments, where  $\chi$  values decreased.  
 257 Upper Zone I is mainly characterized by metal enrichment, with elevated concentrations of Cu, Zn, Ni, Fe, and Mo, among  
 258 other elements. These peaks correlate with high  $\chi$  values (Fig. 7).



259

260 **Figure 7. Redundancy analysis on Hellinger transformed Bray-Curtis distances (corrected by unobserved species) for**  
 261 **the microbial communities along the core distance. A) Influence of physicochemical parameters and B) Elemental**  
 262 **composition. Depth gradient and Zone parameters were chosen to constrain the multivariate space in a supervised**  
 263 **approach. Each axis in the graph shows the percentage of variance explained in an unsupervised and supervised**  
 264 **analysis.**

265 There is statistical significance between some key taxa and the physicochemical and elemental composition along the sediment  
 266 gradient in Inca-Coya Lake; for instance, *Methanoregulaceae*, *Ktedonobacteriaceae*, and *Sulfurimonadaceae* are some of the  
 267 taxa with the strongest correlation to Cu, Fe, Ni and V presence while zones II and III are the most influenced by these dynamics  
 268 (Supplementary Fig. 1).

269 Regarding metabolic approximation, chemoheterotrophy is the most abundant function in all three zones, while aerobic  
 270 chemoheterotrophs are the most prevalent in zone II. Still, chemoheterotrophs (including several electron acceptors) thrive in



271 zones II and III (Supplementary Fig. 2). Other functions, such as Methanogenesis, were abundant only in deep sediments (zone  
272 III). Moreover, as expected, phototrophy and photoautotrophy were present only in low abundance in zone I, where little light  
273 could reach the community (Supplementary Fig. 2). Methane metabolism is very relevant in all three zones, especially in zone  
274 III, where acetate is the primary source for this pathway (Supplementary Fig. 2). Other relevant forms of energy transformation  
275 are Nitrate reduction VI (assimilation) and starch degradation in zone II. Nitrotoluene degradation and biosynthesis of  
276 unsaturated fatty acids are also crucial in the benthic microbial community as a whole (Supplementary Fig. 2). A somewhat  
277 homogeneous prediction for metabolic ability regarding energy production is found along the communities inhabiting over a  
278 meter deep in the Inka-Coya Lake sediments, where geochemical and magnetic dynamics directly influence microbial  
279 activities.

#### 280 4. DISCUSSION

281 Microbial dynamics along the length of the sediment of Inka-Coya Lake are tightly associated with sediment attributes, such  
282 as metal(loid)s concentrations and  $\chi_{fd}\%$ , organic compounds- and water- availability. The geochemical characteristics of the  
283 area surrounding Inka-Coya Lake, where active volcanic activity results in the enrichment of elements such as arsenic, sulfur,  
284 copper, and others (Romero et al., 2003; Tapia et al., 2018), suggests that microorganisms assemblage inhabiting the Atacama  
285 area have developed broad tolerance range to this potential toxic compounds. Microbial biomarkers serve as criteria to assess  
286 anthropogenic impact (Yan et al., 2024), and microorganisms can alter the speciation and bioavailability of meta(oids) in an  
287 ecosystem (Niu et al., 2020).

288 Five stratigraphic zones in the sedimentary core in Inka-Coya were defined from the sediment core here analyzed by Pérez-  
289 Portilla et al. (2024), where the concentration of rock-forming elements such as Cr, Zn, and V are found in concentrations as  
290 expected for the Atacama Desert; while Cu, Mo, Sb and As are higher than expected, suggesting influence from nearby mining  
291 activities (Pérez-Portilla et al., 2024). Copper production in the region generates by-products such as Mo, As, and, to a lesser  
292 extent, Zn (Ramírez et al., 2005; Tapia et al., 2019). Also, mining wastes contain high concentrations of chemical products  
293 such as Pb, Cr, Cd, Cu, Zn, Hg, and Ni, and metalloids, which are often stored in dams or reservoirs (Csavina et al., 2012) or  
294 passed through lotic systems, making them an important source of contamination of inorganic chemical elements for the  
295 aquatic biological communities (Keller et al., 1992; Pollard et al., 2003; Pigati et al., 2011; Hamilton et al., 2017; Ritter et al.,  
296 2019). Previous studies have shown impacts and metal enrichment from the mining industry in the Antofagasta region  
297 surrounding Calama City and the Loa River basin near the Inka-Coya Lake. Cerda et al. (2019), Vargas-Machuca et al. (2021),  
298 Aránguiz-Acuña et al. (2020), and Zanetta-Colombo et al. (2022, 2024), using both abiotic and biological proxies, have  
299 evidenced an increase in the concentration of metals in different environmental matrices during post-industrial time, attributing  
300 this difference to the mining activities in the area. Additionally, changes in the composition of the zooplankton community  
301 (inferred by diapausing egg banks) and benthic diatoms could be attributed to the increase in Cu concentration evidenced in  
302 the sedimentary cores obtained in this lake.

303 A strong correlation between mineral composition and microbial diversity in other arid region water bodies, such as salt flats  
304 and brines, is expected (Fariás et al., 2014; Castro-Severyn et al., 2021; Dong et al., 2022), as demonstrated in pre-Puna salt  
305 lakes, such as Tebenchique and La Brava (Fariás et al., 2014; Ramos-Tapia et al., 2023). The diversity of microbial life in  
306 these shallow salty lakes is dominated by Bacteroidetes, Proteobacteria, and Euryarchaeota (Fariás et al., 2014; Fernandez et  
307 al., 2016; Kurth et al., 2021) and hypersaline lakes are mainly composed by Bacteroidetes, Chloroflexi, Cyanobacteria and  
308 Proteobacteria (Dorador et al., 2018). Inka-Coya, one of the few brackish water lakes located in the Antofagasta Region below



309 3,000 m a.s.l., and its sediment communities are dominated by Phylum Acidobacteriota, Chloroflexi, Actinobacteriota, and  
310 Campylobacterota, sharing some similarities and taking into account changes in taxonomy (Oren and Garrity, 2021).

311 At the lower taxonomic rank, the community is dominated by microorganisms with a broad repertoire for mineral interactions,  
312 e.g., there is experimental evidence of organomineralization in extracellular S<sup>0</sup> formation by a species of the sulfur-oxidizing  
313 bacteria *Sulfuricurvum* (Cron et al., 2019). Another remarkable microorganism found along the lacustrine sediment is the  
314 cosmopolitan and highly diverse *Sulfurimonas*, which can grow using sulfur, hydrogen, nitrogen, oxygen, and organic  
315 compounds, suggesting it is critical in maintaining trophic dynamics (Han and Perner, 2015). Additionally, *Mycobacterium* is  
316 a saprophytic bacterium commonly found in lakes, rivers, and other water sources (Falkinham et al., 2015), there are some  
317 species representatives of this genus that have bioremediation potential for polycyclic aromatic hydrocarbons (Deng et al.,  
318 2023), suggesting endurance and a broad range of metabolic capacities. There are extensive studies on Mycobacteriaceae  
319 representatives that cause human disease, as reviewed by Falkinham (2009). However, environmental representatives with the  
320 capacity to inhabit poly-extreme environments have yet to be determined; Inka-Coya sediment is an uncharacterized species  
321 of this genus.

322 Furthermore, several representatives of the candidate phylum Aminicenantales are common in current sediments from Zone I,  
323 i.e., recently deposited sediments during the last ten years. So far, these microorganisms are associated with a fermentative  
324 saccharolytic lifestyle that does not have an isolated representative yet (Kadnikov et al., 2019). While in Zone II, comprising  
325 the period between 10 and 50 years ago approximately, is mainly represented by microorganisms classified as the  
326 *Pseudarcobacter* genus, that was recently separated from the *Arcobacter* genus (Pérez-Cataluña et al., 2018) and are  
327 characterized as mesophilic bacteria that can grow in microaerophilic conditions (Collado et al., 2011). Finally, in the deeper  
328 and older sediments, where anaerobic (or facultative anaerobes) microorganisms can thrive, there is a particular abundance of  
329 an ASV from the *Pelolinea* genus that has only one described species that was isolated from the subseafloor sediment (Imachi  
330 et al., 2014), and an unknown Chloroflexi species that is associated with Dehalococcoidia class a common sub-seafloor  
331 bacterium (Wasmund et al., 2014). This finding suggests past conditions of high salinities for the lake, close to marine  
332 conditions and much higher than the lake's current salinity of around 5 g·L<sup>-1</sup> (Aránguiz-Acuña et al., 2020). Archaea  
333 representatives found inhabiting the lacustrine sediments include the hydrogenotrophic methanogen *Methanolinea* (Imachi et  
334 al., 2008; Rainey et al., 2015), that are very abundant, especially towards the deeper and more anoxic environment, where  
335 methanogenesis is the central metabolism at play. Overall, there is great taxonomic and metabolic diversity associated with  
336 the microbial community from this lacustrine sediment.

337 The microbial community in Inka-Coya Lake is primarily heterotrophic with a special enrichment in methanogenic organisms  
338 in the oldest deposits previous to 1950 (zone III), where oxygen levels are lower as evidenced by the metabolic approximation  
339 done in this study, and another kind of metabolism depending of CO<sub>2</sub> concentrations, could be dominating. Furthermore, there  
340 is a vast taxonomic novelty harbored in Inka-Coya Lake sediment; over 70% of taxa cannot be identified to the genus level,  
341 indicating there is a significant amount of “microbial dark matter,” a term associated with unknown microbial representatives  
342 that can potentially harbor novel bioactive compounds with numerous applications (Zha et al., 2022; Jiao et al., 2021).

343 The main drivers for microbial community composition in the sinks-Coya sediments were As, Sb, V, Mo, Mi, Fe, and Cu,  
344 which suggest that there are numerous strategies that microorganisms use to resist high concentrations of metal(loid)s that  
345 thrive in this ecosystem (Rahman, 2020; Mathivanan et al., 2021), as observed in the Atacama and Altiplano area (Orellana et  
346 al., 2018; Donati et al., 2019; Aszalós et al., 2020; Castro-Severyn et al., 2019). Additionally, microorganisms can use oxido-  
347 reduction processes to obtain energy from metal ions (Raab and Feldman, 2003; Staicu and Stolz, 2021), and given the known  
348 geochemical characteristics of the area it is expected to find strong relationships between the microbial life and inorganic  
349 compounds, as they can dissolve and precipitate ores and influence metal(loid)s transformations (Raab et al., 2003; Zhou et  
350 al., 2022). In this extreme environment where competition is strong and abiotic pressures are constant, organic matter and



351 water availability -both parameters critical for most life forms- govern community abundance and composition, suggesting a  
352 delicate dynamic balance reached between abiotic and biotic entities at play. It is important to remark that with the number of  
353 unclassified taxa, a significant number of novel resistant or usage mechanisms remain to be characterized.

354 Observed trends in magnetic susceptibility in Inka-Coya sediment could be mainly attributed to variations in the concentration  
355 of ferromagnetic minerals, such as titanomagnetite, and authigenic origin minerals, such as sulfide (greigite), as is broadly  
356 explained in Pérez-Portilla et al. (2024) for this sediment core. In this case, the high X values would result from a high  
357 concentration of ferromagnetic minerals of detrital origin, primarily Fe oxides, while elevated xfd% values would be linked to  
358 the presence of greigite of authigenic origin (Pérez-Portilla et al., 2014). Thus, greigite formation typically occurs through the  
359 dissolution of titanomagnetite or other detrital minerals containing Fe (e.g., Chan et al., 2001; Fialová et al., 2006; Versteeg et  
360 al., 1995).

361 Elevated magnetic susceptibility values in the upper layers of sediments may be attributed to a recent deposition of fine Fe  
362 oxide grains, which could be originating and transported from industrial and urban sources (e.g., Chan et al., 2001); this concurs  
363 with the Fe top sediment peak, and it could be related to a diverse superficial community, while in zone II the lowest values  
364 of X are observed, which could be associated with a mixed community. All these processes occur under a high production of  
365 Fe minerals of authigenic origin (xfd%>3%; Dearing et al., 1996). Moreover, lower levels of Xfd% could be involved in the  
366 production and assimilation of iron sulfides such as greigite (Bazylinski et al., 2001; Lins et al., 2007), promoting a microbial  
367 specialization and increasing resistance of the anaerobic community found in the deeper sediments. Microbial metabolic  
368 responses could satisfactorily support several processes associated with greigite formation. The presence of greigite in the lake  
369 sediments could be associated with reducing or low-oxygen environments (e.g., Benning et al., 2000), where additionally  
370 magnetotactic bacteria would contribute to sedimentary greigite formation through the biomineralization of magnetosomes in  
371 anoxic aqueous environments (Moskowitz et al., 2008).

372 There are statistically significant correlations between some particular taxa and the geochemical composition along the  
373 sediment gradient in Inca-Coya Lake suggesting that each element directly influences the metabolic capabilities of the  
374 microorganisms and shapes the community selecting taxa that can resist metal(loid)s toxicity (Yao and Gao, 2007; Laplante  
375 et al., 2013; Stankevica et al., 2020; Kostka and Leśniak, 2021; Yan et al., 2020). Further studies that elucidate the functional  
376 properties that these microbial communities have will enhance our understanding in terms of metal(loid)s resistance and the  
377 use of different electron acceptors for energy production.

378 Finally, increasing metal(loid)s exploitation in the region during the last 100 years has directly influenced their mobility and  
379 the local geochemistry. In this context, microorganisms from these extreme environments are known to be highly adaptive and  
380 have developed several resistance mechanisms and the ability to use these compounds to their benefit. Hence, some bacteria  
381 (and their genes) can be used as biomarkers for the bioavailability of such metals and contamination of soils (Li and Wong,  
382 2010; Roosa et al., 2014). In Inka-Coya Lake, a selective process could be evidenced along the sediment record, in which the  
383 recent period is characterized by a marked increase in chemical elements and microbial composition, which could be associated  
384 with the increasing mining activity and other anthropological activities, as water extraction or aridity increase by climatic  
385 changes, that also would increase the disturbance of this relevant area in the core of the most arid non-polar Desert.

|386

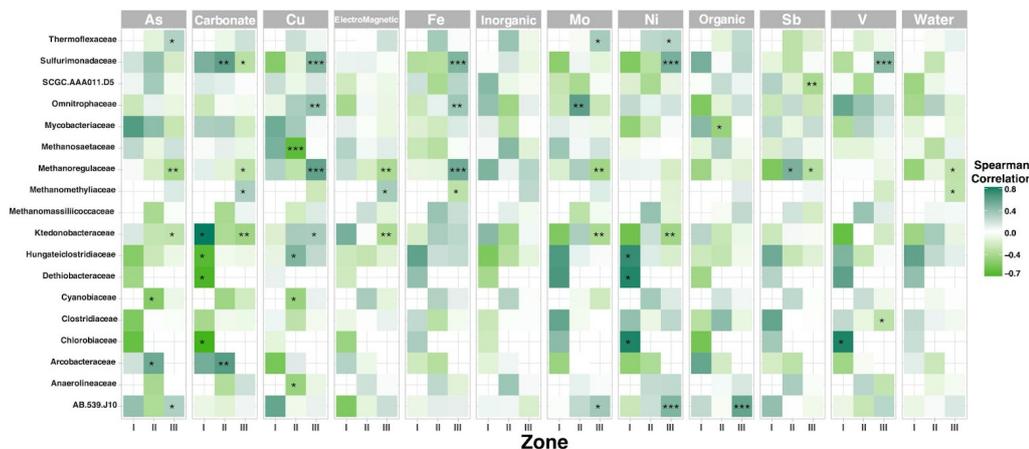


387 **5. CONCLUSIONS**

388 This study represents the first to encompass a deep gradient of microbial life in a desert lake in the Atacama area, proposing a  
 389 biological clustering of taxa and function in three periods that stratified for over 600 years, since the pre-mining period, the  
 390 mining development and the most industrialized mega mining observed nowadays. A great taxonomic novelty exists among  
 391 the microbial community inhabiting lacustrine sediments of Inka-Coya that potentially holds an abundant novel repertoire of  
 392 bioactive compounds of biotechnological interest. Mineralogical enrichment, water, organic matter availability, and magnetic  
 393 susceptibility are also variables that explain the changes within the microbial community. There are strong relationships  
 394 between geochemical composition and microbial diversity, especially in Cu, Fe, Ni, and V. The first zone is less diverse and  
 395 dominated by Actinobacteria; the second zone has a high abundance of Chloroflexi, Acidobacteriota, and Actinobacterota.  
 396 The third zone shows more rare taxa with lower abundance and clusters the higher area of the studied sediments, including  
 397 archaea. Overall, chemoheterotrophy is the prevalent energy production mechanism along the sediment core. This unique and  
 398 fragile ecosystem depends on biogeochemical dynamics vulnerable to anthropogenic activities and climate change.

399 **Appendix A.**

400

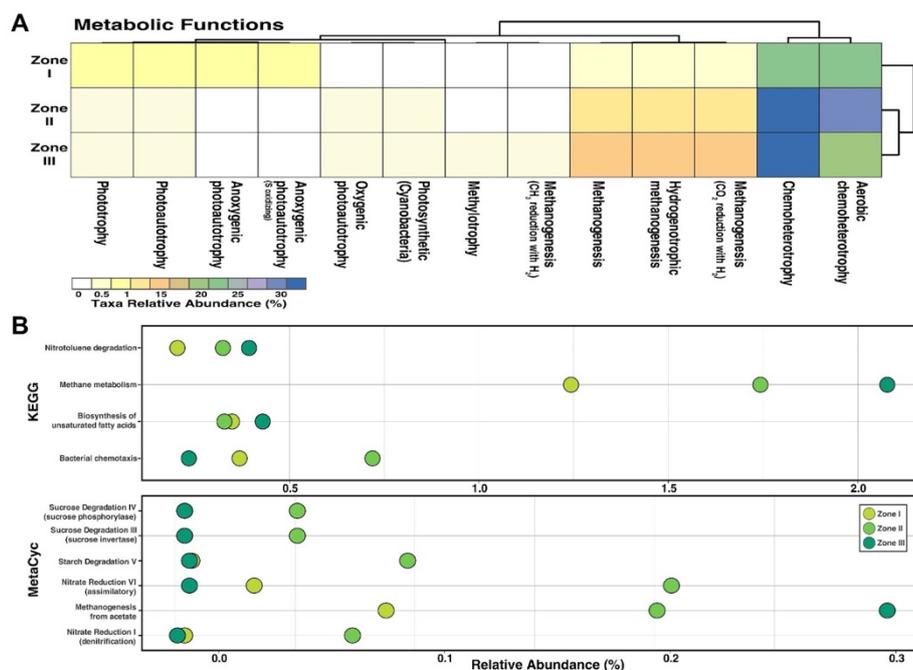


401

402 **Figure A1.** Pearson correlation between the top microbial families and geochemical parameters. Asterisks show the level of  
 403 significance (\**p* value < 0.05, \*\**p* value < 0.01, Pearson correlation). Purple, blue, and white indicate positive, negative, and  
 404 no correlation, respectively.

405

406 **Appendix B**



407

408 **Figure B1.** Metabolic pathways prediction of the microbial community inhabiting sediments along a depth gradient in Inka-  
 409 Coya Lake. The predictions are based on the identified taxonomic composition according to the comparison with different  
 410 databases, such as: A) FAPROTAX; B) KEGG and C) MetaCyc. Categories with significant differences ( $p < 0.05$ ) according  
 411 to Kruskal-Wallis test are displayed.

412

#### 413 DATA AVAILABILITY STATEMENT

414 The raw sequencing data presented in this study have been deposited in the DDBJ/ENA/GenBank SRA database under the  
 415 BioProject: PRJNA1067596.

416

#### 417 AUTHOR CONTRIBUTIONS

418 Conceptualization: AAA; FR, JCS, CPE. Data curation: CPE, JCS, FR, IHF, AAA. Formal analysis: CPE, JCS, FR, AAA,  
 419 AM, HP. Funding acquisition: FR, AAA Methodology: CPE, JCS, AM, HP, FR, AAA. Supervision: AAA, FR Writing original  
 420 draft: CPE, JCS. Writing review and editing: AAA, AM, HP, JCS, CPE. All authors have read and agreed to the published  
 421 version of the manuscript.

422

#### 423 FUNDING

424 ANID 2020 FONDECYT Regular 1200423 (AA-A), UTA-Mayor 9738-24 (AA-A)

425 ANID 2022 FONDECYT Regular 1220902 (FR)

426 ANID 2023 FONDECYT postdoctoral 3230189 (CPE)



427 ANID 2021 FONDECYT postdoctoral 3210156 (JCS)

428

#### 429 **COMPETING INTERESTS**

430 The contact author has declared that none of the authors has any competing interests.

431

#### 432 **ACKNOWLEDGMENTS**

433 Likan antay Community of San Francisco de Chiu-Chiu.

434 MAINI-UCN (Kappabridge)

#### 435 **References**

- 436 1. Adrian R, O'Reilly CM, Zagarese H, Baines SB, Hessen DO, Keller W, Livingstone DM, Sommaruga R, Straile D,  
437 van Donk E, Weyhenmeyer GA, Winder M.: Lakes as sentinels of climate change. *Limnol Oceanogr.*, 54:2283–2297,  
438 2009
- 439 2. Albarracín, V. H., Galván, F. S., and Farías, M. E.: Extreme microbiology at Laguna Socompa: A high-altitude  
440 Andean lake (3570 m asl) in Salta, Argentina. *Microbial Ecosystems in Central Andes Extreme Environments:*  
441 *Biofilms, Microbial Mats, Microbialites and Endoevaporites*, 205-220, 2020
- 442 3. Andersen KS, Kirkegaard RH, Karst SM, Albertsen M.: ampvis2: an R package to analyse and visualise 16S rRNA  
443 amplicon data. *BioRxiv*. 2018
- 444 4. Aránguiz-Acuña, A., Luque, J. A., Pizarro, H., Cerda, M., Heine-Fuster, I., Valdés, J., ... and Wennrich, V.: Aquatic  
445 community structure as sentinel of recent environmental changes unraveled from lake sedimentary records from the  
446 Atacama Desert, Chile. *PLoS One*, 15(2), e0229453, 2020
- 447 5. Aránguiz-Acuña, A., Pérez-Portilla, P., De la Fuente, A., and Fontaneto, D.: Life-history strategies in zooplankton  
448 promote coexistence of competitors in extreme environments with high metal content. *Sci Rep*, 8(1), 11060, 2018
- 449 6. Aszalós, J. M., Szabó, A., Felföldi, T., Jurecska, L., Nagy, B., and Borsodi, A. K.: Effects of active volcanism on  
450 bacterial communities in the highest-altitude crater lake of Ojos del Salado (Dry Andes, Altiplano-Atacama Region).  
451 *Astrobiology*, 20(6), 741-753, 2020
- 452 7. Bandowe FL, Grosjean M, Tylmann W, Mosquera P, Hampel H, Schneider T.: A 150-year record of polycyclic  
453 aromatic compound (PAC) deposition from high Andean Cajas National Park, southern Ecuador. *STOTEN*,  
454 621:1652–1663, 2018
- 455 8. Bazylinski, D. A.: Bacterial mineralization. *Encyclopedia of materials: Science and technology*, 441-447, 2001
- 456 9. Benito, X. Benthic Foraminifera and diatoms as ecological indicators. *Modern Trends in Diatom Identification:*  
457 *Fundamentals and Applications*, 257-280, 2020



- 458 10. Benning, L. G., Wilkin, R. T., and Barnes, H. L.: Reaction pathways in the Fe–S system below 100 C. *Chem Geol.*, 167(1-  
459 2), 25-51, 2000
- 460 11. Borsdorf, A., and Stadel, C.: *The Andes: A geographical portrait*. Springer, 2015
- 461 12. Borsodi, A. K., Aszalós, J. M., Megyes, M., and Nagy, B.: Benthic Bacterial Diversity of High-Altitude  
462 Athalassohaline Lakes of the Puna de Atacama (Central Andes). *Geomicrobiol J*, 39(1), 28-38, 2022
- 463 13. Callahan, B.J., McMurdie, P.J., Rosen, M.J., Han, A.W., Johnson, A., Holmes, S.P.: DADA2: high-resolution sample  
464 inference from Illumina amplicon data. *Nat Methods*. 13:581–583. doi: 10.1038/nmeth.3869, 2016
- 465 14. Caporaso, J. G., Lauber, C. L., Walters, W. A., Berg-Lyons, D., Lozupone, C. A., Turnbaugh, P. J., ... and Knight,  
466 R.: Global patterns of 16S rRNA diversity at a depth of millions of sequences per sample. *PNAS*. 108(Supplement 1):4516–  
467 4522, 2011
- 468 15. Caspi, R., Billington, R., Fulcher, C. A., Keseler, I. M., Kothari, A., Krummenacker, M., ... and Karp, P. D.: The  
469 MetaCyc database of metabolic pathways and enzymes. *Nuc acids Res*, 46(D1), D633-D639, 2018
- 470 16. 16. Castro-Severyn, J., Pardo-Esté, C., Mendez, K. N., Fortt, J., Marquez, S., Molina, F., ... and Saavedra, C. P.:  
471 Living to the high extreme: unraveling the composition, structure, and functional insights of bacterial communities thriving in  
472 the arsenic-rich Salar de Huasco altiplanic ecosystem. *mSpectrum*, 9(1), 10-1128, 2021
- 473 17. Castro-Severyn, J., Pardo-Esté, C., Sulbaran, Y., Cabezas, C., Gariazzo, V., Briones, A., ... and Saavedra, C. P.:  
474 Arsenic response of three altiplanic *Exiguobacterium* strains with different tolerance levels against the metalloid species: a  
475 proteomics study. *Front Microbiol*, 10, 2161, 2019
- 476 18. Cerda, M., Evangelista, H., Valdés, J., Siffedine, A., Boucher, H., Nogueira, J., ... and Ortlieb, L.: A new 20th century  
477 lake sedimentary record from the Atacama Desert/Chile reveals persistent PDO (Pacific Decadal Oscillation) impact. *J South  
478 Am Earth Sci*. 95: 102302, 2019
- 479 19. Chang, L., Heslop, D., Roberts, A. P., Rey, D., and Mohamed, K. J.: Discrimination of biogenic and detrital magnetite  
480 through a double Verwey transition temperature. *J Geophys.*, 121(1), 3-14, 2016
- 481 20. Cohen, A. S., Manobianco, J., Dettman, D. L., Black, B. A., Beck, C., Feibel, C. S., ... and Vonhof, H. Seasonality  
482 and lake water temperature inferred from the geochemistry and sclerochronology of quaternary freshwater bivalves from the  
483 Turkana Basin, Ethiopia and Kenya. *Quaternary Sci Rev*, 317, 108284, 2023
- 484 21. Collado, L., Levican, A., Perez, J., and Figueras, M. J.: *Arcobacter defluvii* sp. nov., isolated from sewage samples.  
485 *IJSE*, 61(9), 2155-2161, 2011
- 486 22. Cron, B., Henri, P., Chan, C. S., Macalady, J. L., and Cosmidis, J.: Elemental sulfur formation by *Sulfuricurvum*  
487 *kujjense* is mediated by extracellular organic compounds. *Front Microbiol*, 10, 2710, 2019
- 488 23. Csavina, J., Field, J., Taylor, M. P., Gao, S., Landázuri, A., Betterton, E. A., and Sáez, A. E.: A review on the importance  
489 of metals and metalloids in atmospheric dust and aerosol from mining operations. *STOTEN*, 433, 58-73, 2012
- 490 24. Da Costa, C., Colin, Y., Debret, M., Copard, Y., Gardes, T., Jacq, K., ... and Berthe, T. Shifts in sediment bacterial  
491 communities reflect changes in depositional environments in a fluvial context. *STOTEN*, 885, 163890, 2023



- 492 25. Dearing, J. A., Hay, K. S., Baban, A. S., Huddleston, E. M., Wellington H., and Loveland P. J.: Magnetic susceptibility  
493 of topsoils: a test of conflicting theories using a national database. *Geophys. J. Int.* 127: 728–734, 1996
- 494 26. Demergasso, C., Escudero, L., Casamayor, E. O., Chong, G., Balagué, V., and Pedrós-Alió, C.: Novelty and spatio-  
495 temporal heterogeneity in the bacterial diversity of hypersaline Lake Tebenquiche (Salar de Atacama). *Extremophiles*, 12,  
496 491-504, 2008
- 497 27. Deng, Y., Mou, T., Wang, J., Su, J., Yan, Y., and Zhang, Y. Q.: Characterization of three rapidly growing novel  
498 *Mycobacterium* species with significant polycyclic aromatic hydrocarbon bioremediation potential. *Front Microbiol*, 14, 2023
- 499 28. Dib, J., Motok, J., Zenoff, V. F., Ordoñez, O., and Fariás, M. E.: Occurrence of resistance to antibiotics, UV-B, and  
500 arsenic in bacteria isolated from extreme environments in high-altitude (above 4400 m) Andean wetlands. *Curr. Microbiol.*  
501 56, 510–517, 2008
- 502 29. Dittmar T.: Hydrochemical process controlling arsenic and heavy metal contamination in the Elqui river system  
503 (Chile). *STOTEN*. 325: 193–207, 2004
- 504 30. Donati, E. R., Sani, R. K., Goh, K. M., and Chan, K. G.: Recent advances in bioremediation/biodegradation by  
505 extreme microorganisms. *Front Microbiol*, 10, 1851, 2019
- 506 31. Dong, H., Huang, L., Zhao, L., Zeng, Q., Liu, X., Sheng, Y., ... and Chen, H.: A critical review of mineral–microbe  
507 interaction and co-evolution: mechanisms and applications. *National Sci Rev*, 9(10), nwac128, 2022
- 508 32. Dorador, C., Fink, P., Hengst, M., Icaza, G., Villalobos, A. S., Vejar, D., ... and Harrod, C.: Microbial community  
509 composition and trophic role along a marked salinity gradient in Laguna Puilar, Salar de Atacama, Chile. *Antonie Van*  
510 *Leeuwenhoek*, 111, 1361-1374, 2018
- 511 33. Douglas, G. M., Maffei, V. J., Zaneveld, J. R., Yurgel, S. N., Brown, J. R., Taylor, C. M., ... and Langille, M. G.:  
512 PICRUSt2 for prediction of metagenome functions. *Nat biotechnol*, 38(6), 685-688, 2020
- 513 34. Falkinham, J. O.: Environmental sources of nontuberculous mycobacteria. *Clin chest med*, 36(1), 35-41, 2015
- 514 35. Fariás, M. E., Contreras, M., Rasuk, M. C., Kurth, D., Flores, M. R., Poire, D. G., ... and Visscher, P. T.:  
515 Characterization of bacterial diversity associated with microbial mats, gypsum evaporites and carbonate microbialites in  
516 thalassic wetlands: Tebenquiche and La Brava, Salar de Atacama, Chile. *Extremophiles* 18, 311–329, 2014.
- 517 36. Fernandez, A. B., Rasuk, M. C., Visscher, P. T., Contreras, M., Novoa, F., Poire, D. G., ... and Fariás, M. E.: Microbial  
518 diversity in sediment ecosystems (evaporites domes, microbial mats, and crusts) of hypersaline Laguna Tebenquiche, Salar de  
519 Atacama, Chile. *Front Microbiol*, 7, 1284, 2016
- 520 37. Fialová, H., Maier, G., Petrovský, E., Kapička, A., Boyko, T., Scholger, R., and MAGPROX Team.: Magnetic  
521 properties of soils from sites with different geological and environmental settings. *J Appl Geophys.*, 59(4), 273-283, 2006
- 522 38. Garreaud, R., Vuille, M., and Clement, A. C.: The climate of the Altiplano: observed current conditions and  
523 mechanisms of past changes. *Palaeogeogr.*, 194(1-3), 5-22, 2003
- 524 39. Falkinham III, J. O.: The biology of environmental mycobacteria. *Env Microbiol Rep*, 1(6), 477-487, 2009



- 525 40. Grosjean, M., and Veit, H. Water resources in the arid mountains of the Atacama Desert (northern Chile): past climate  
526 changes and modern conflicts. *Global change and mountain regions: an overview of current knowledge*, 93-104, 2005
- 527 41. Hamilton, P. B., Rolshausen, G., Uren Webster, T. M., and Tyler, C. R.: Adaptive capabilities and fitness  
528 consequences associated with pollution exposure in fish. *Proc. R. Soc. B*, 372(1712), 20160042, 2017
- 529 42. Han, Y., and Perner, M.: The globally widespread genus *Sulfurimonas*: versatile energy metabolisms and adaptations  
530 to redox clines. *Front Microbiol*, 6, 989, 2015
- 531 43. Haslett, J., and Parnell, A.: A simple monotone process with application to radiocarbon-dated depth chronologies. *J*  
532 *R Stats*, 57(4), 399-418, 2008
- 533 44. Heiri, O., Lotter, A. F., and Lemcke, G.: Loss on ignition as a method for estimating organic and carbonate content  
534 in sediments: reproducibility and comparability of results. *J. Paleolimnol*, 25, 101-110, 2001
- 535 45. Hernández, A., Bao, R., Giralt, S., Barker, P. A., Leng, M. J., Sloane, H. J., and Sáez, A.: Biogeochemical processes  
536 controlling oxygen and carbon isotopes of diatom silica in Late Glacial to Holocene lacustrine rhythmites. *Palaeogeogr.*  
537 *Palaeoclimatol. Palaeoecol.* 299(3–4), 413–425, 2012.
- 538 46. Hogg, A. G., Heaton, T. J., Hua, Q., Palmer, J. G., Turney, C. S. M., Southon, J., Bayliss, A., Blackwell, P. G.,  
539 Boswijk, G., Bronk Ramsey, C., Pearson, C., Petchey, F., Reimer, P., Reimer, R. and Wacker, L.: “SHCal20 SOUTHERN  
540 HEMISPHERE CALIBRATION, 0–55,000 YEARS CAL BP,” *Radiocarbon*. Cambridge University Press, pp. 1–20, 2020
- 541 47. Imachi, H., Sakai, S., Lipp, J. S., Miyazaki, M., Saito, Y., Yamanaka, Y., ... and Takai, K. *Pelolinea submarina* gen.  
542 nov., sp. nov., an anaerobic, filamentous bacterium of the phylum Chloroflexi isolated from seafloor sediment. *International*  
543 *journal of systematic and evolutionary microbiology*, 64(Pt\_3), 812-818, 2014
- 544 48. Imachi, H., Sakai, S., Sekiguchi, Y., Hanada, S., Kamagata, Y., Ohashi, A., and Harada, H. *Methanolinea tarda* gen.  
545 nov., sp. nov., a methane-producing archaeon isolated from a methanogenic digester sludge. *IJSEM*, 58(1), 294-301, 2008
- 546 49. Jiao, J. Y., Liu, L., Hua, Z. S., Fang, B. Z., Zhou, E. M., Salam, N., ... and Li, W. J.: Microbial dark matter coming to  
547 light: challenges and opportunities. *Nat Sci Rev*, 8(3), nwaa280, 2021
- 548 50. Kadnikov, V. V., Mardanov, A. V., Beletsky, A. V., Karnachuk, O. V., and Ravin, N. V. Genome of the candidate  
549 phylum Aminicenantes bacterium from a deep subsurface thermal aquifer revealed its fermentative saccharolytic lifestyle.  
550 *Extremophiles*, 23, 189-200, 2019
- 551 51. Kanehisa, M., Goto, S., Sato, Y., Furumichi, M., and Tanabe, M.: KEGG for integration and interpretation of large-  
552 scale molecular data sets. *Nucleic acids Res*, 40(D1), D109-D114, 2012
- 553 52. Kassambara, A.: *R Graphics Essentials for Great Data Visualization: 200 Practical Examples You Want to Know for*  
554 *Data Science*. STHDA, 2017
- 555 53. Keller W, Yan ND, Holtze K, Pitblado JR. Chemical response of acid lakes in the Sudbury, Ontario area to reduced  
556 smelter emissions, 1981–1989. *Can J Fish Aquat Sci.* 49 (Suppl.1): 25–32, 1992



- 557 54. Kereszturi, Á., Aszalós, J. M., Heiling, Z., Ignéczi, Á., Kapui, Z., Király, C., ... and Szalai, Z. Cold, dry, windy, and  
558 UV irradiated: surveying Mars-relevant conditions in Ojos del Salado Volcano (Andes Mountains, Chile). *Astrobiology*, 20(6),  
559 677-683, 2020
- 560 55. Korosi, J. B., Thienpont, J. R., Smol, J. P., and Blais, J. M. Paleo-ecotoxicology: what can lake sediments tell us about  
561 ecosystem responses to environmental pollutants?. *Environ. Sci. Technol.*, 51(17), 9446-9457, 2017
- 562 56. Kostka, A., and Leśniak, A. Natural and anthropogenic origin of metals in lacustrine sediments; assessment and  
563 consequences—A case study of Wigry lake (Poland). *Minerals*, 11(2), 158, 2021
- 564 57. Kurth, D., Elias, D., Rasuk, M. C., Contreras, M., and Farias, M. E. Carbon fixation and rhodopsin systems in  
565 microbial mats from hypersaline lakes Brava and Tebenquiche, Salar de Atacama, Chile. *PLoS One*, 16(2), e0246656, 2021
- 566 58. Lahti, L., Shetty, S., Blake, T., and Salojarvi, J. Tools for microbiome analysis in R Version 2.1. 26, 2017
- 567 59. Laplante K, Sébastien B, Derome N. Parallel changes of taxonomic interaction networks in lacustrine bacterial  
568 communities induced by a polymetallic perturbation. *Evol Appl.*, 6(4):643-59, 2013
- 569 60. Li, W. C., and Wong, M. H. Effects of bacteria on metal bioavailability, speciation, and mobility in different metal  
570 mine soils: a column study. *J soils sediments*, 10, 313-325, 2010
- 571 61. Lins, U., Keim, C. N., Evans, F. F., Farina, M., and Buseck, P. R.: Magnetite (Fe<sub>3</sub>O<sub>4</sub>) and greigite (Fe<sub>3</sub>S<sub>4</sub>) crystals  
572 in multicellular magnetotactic prokaryotes. *Geomicrobiology J*, 24(1), 43-50, 2007
- 573 62. Louca, S., Parfrey, L. W., and Doebeli, M.: Decoupling function and taxonomy in the global ocean microbiome.  
574 *Science*, 353(6305), 1272-1277, 2016
- 575 63. Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2.  
576 *Genome Biol.* 2014;15(12):1–21
- 577 64. Madsen, E. L.: Microorganisms and their roles in fundamental biogeochemical cycles. *Curr opinion Biotech.*, 22(3),  
578 456-464, 2011
- 579 65. Mathivanan, K., Chandirika, J. U., Vinothkanna, A., Yin, H., Liu, X., and Meng, D. Bacterial adaptive strategies to  
580 cope with metal toxicity in the contaminated environment—A review. *Ecotox Env Safety*, 226, 112863, 2021
- 581 66. McMurdie P, Holmes S. phyloseq: an R package for reproducible interactive analysis and graphics of microbiome  
582 census data. *PLoS ONE*. 8(4):e61217, 2013
- 583 67. Moskowitz, B. M., Bazylinski, D. A., Egli, R., Frankel, R. B., and Edwards, K. J.: Magnetic properties of marine  
584 magnetotactic bacteria in a seasonally stratified coastal pond (Salt Pond, MA, USA). *GJI* 174(1), 75-92, 2008
- 585 68. Niu, Z. S., Yang, Y., Tou, F. Y., Guo, X. P., Huang, R., Xu, J., ... and Hochella, M. F. Sulfate-reducing bacteria (SRB)  
586 can enhance the uptake of silver-containing nanoparticles by a wetland plant. *Env Sci: Nano*, 7(3), 912-925, 2020.
- 587 69. Ordoñez, O. F., Flores, M. R., Dib, J. R., Paz, A., and Fariás, M. E. Extremophile culture collection from Andean  
588 lakes: extreme pristine environments that host a wide diversity of microorganisms with tolerance to UV radiation. *Microb.*  
589 *Ecol.* 58, 461–473, 2009.



- 590 70. Orellana, R., Macaya, C., Bravo, G., Dorochesi, F., Cumsille, A., Valencia, R., ... and Seeger, M. Living at the  
591 frontiers of life: extremophiles in Chile and their potential for bioremediation. *Front Microbiol*, 9, 2309, 2018
- 592 71. Oren, A. (2014). The Family *Methanoregulaceae*. In: Rosenberg, E., DeLong, E.F., Lory, S., Stackebrandt, E.,  
593 Thompson, F. (eds) *The Prokaryotes*. Springer, Berlin, Heidelberg.
- 594 72. Oren, A., and Garrity, G. M. Valid publication of the names of forty-two phyla of prokaryotes. *International journal*  
595 *of systematic and evolutionary microbiology*, 71(10), 005056, 2021
- 596 73. Pérez-Cataluña, A., Salas-Massó, N., Diéguez, A. L., Balboa, S., Lema, A., Romalde, J. L., and Figueras, M. J.  
597 Revisiting the taxonomy of the genus *Arcobacter*: getting order from the chaos. *Front Microbiol*, 9, 2077, 2018
- 598 74. Pérez-Portilla, P., Aránguiz-Acuña, A., Pizarro, H., and Herrera, J.: Assessing the Effects of Long-Term Mining  
599 Exploitation on a Lacustrine System from the Arid Region of the Atacama Desert, Chile. *STOTEN*, 949:  
600 <https://doi.org/10.1016/j.scitotenv.2024.174771>, 2024
- 601 75. Picard, M., Wood, S. A., Pochon, X., Vandergoes, M. J., Reyes, L., Howarth, J. D., ... and Puddick, J. Molecular and  
602 pigment analyses provide comparative results when reconstructing historic cyanobacterial abundances from lake sediment  
603 cores. *Microorganisms*, 10(2), 279, 2022
- 604 76. Pigati JS, Miller DM, Bright J, Mahan SA, Nekola JC, Paces JB. Chronology, sedimentology, and micro-fauna of  
605 ground-water discharge deposits in the central Mojave Desert, Valley Wells, California. *Geol Soc Am Bull.*, 123: 2224–2239,  
606 2011
- 607 77. Placzek, C., Quade, J., Betancourt, J. L., Patchett, P. J., Rech, J. A., Latorre, C., ... and English, N. B. Climate in the  
608 dry central Andes over Geologic, millennial, and interannual timescales. *Ann. Mo. Bot. Gard.*, 96(3), 386–397, 2009.
- 609 78. Pollard HG, Colbourne JK, Keller W. Reconstruction of Centuries-old *Daphnia* Communities in a Lake Recovering  
610 from Acidification and Metal Contamination. *Ambio.*, 32: 214–218, 2003
- 611 79. Price, M.N., Dehal, P.S., Arkin, A.P.: FastTree: computing large minimum evolution trees with profiles instead of a  
612 distance matrix. *Mol Biol Evol.*, 26(7):1641–1650, 2009
- 613 80. Pueyo, J. J., Sáez, A., Giralt, S., Valero-Garcés, B. L., Moreno, A., Bao, R., Schwalb, A., Herrera, C., Klosowska, B.,  
614 and Taberner, C.: Carbonate and organic matter sedimentation and isotopic signatures in Lake Chungará, Chilean Altiplano,  
615 during the last 12.3 kyr, *Palaeogeogr. Palaeoclimatol.*, 307, 339–355, 2011
- 616 81. Quast C, Pruesse E, Yilmaz P, Gerken J, Schweer T, Yarza P, et al. The SILVA ribosomal RNA gene database project:  
617 improved data processing and web-based tools. *Nucl Acids Res.*, 41(D1):D590–D596, 2012.
- 618 82. Raab, A., and Feldmann, J. Microbial transformation of metals and metalloids. *Science Progress*, 86(3), 179-202,  
619 2003
- 620 83. Rahman, Z. An overview on heavy metal resistant microorganisms for simultaneous treatment of multiple chemical  
621 pollutants at co-contaminated sites, and their multipurpose application. *J. Haz. Mat.*, 396, 122682, 2020
- 622 84. Rainey, F., Kämpfer, P., Trujillo, M., Chun, J., DeVos, P., Hedlund, B., and Dedysh, S. (2015). *Bergey's manual of*  
623 *systematics of Archaea and Bacteria* (Vol. 410). W. B. Whitman (Ed.). Hoboken, NJ: Wiley.



- 624 85. Ramírez M, Massolo S, Frache R, Correa JA. Metal speciation and environmental impact on sandy beaches due to El  
625 Salvador copper mine, Chile. *Mar Pollut Bull.*, 50, 62–72, 2005
- 626 86. Ramos-Tapia I, Salinas P, Núñez R, Cortez D, Soto J, Paneque M. Compositional Changes in Sediment Microbiota  
627 Are Associated with Seasonal Variation of the Water Column in High-Altitude Hyperarid Andean Lake Systems. *Microbiol*  
628 *Spectr.* 15;11(3):e0520022, 2023
- 629 87. Rasuk, M. C., Kurth, D., Flores, M. R., Contreras, M., Novoa, F., Poire, D., and Farias, M. E.: Microbial  
630 characterization of microbial ecosystems associated to evaporites domes of gypsum in Salar de Llamara in Atacama Desert.  
631 *Microb. Ecol.* 71, 44–56, 2014
- 632 88. Ritter, B., Wennrich, V., Medialdea, A., Brill, D., King, G., Schneiderwind, S., ... and Dunai, T. J.: Climatic  
633 fluctuations in the hyperarid core of the Atacama Desert during the past 215 ka. *Sci Rep.*, 9: 5270, 2019
- 634 89. Rodríguez-Luna, D., Encina-Montoya, F., Alcalá, F. J., and Vela, N.: An Overview of the Environmental Impact  
635 Assessment of Mining Projects in Chile. *Land*, 11(12), 2278, 2022
- 636 90. Romero, H., Méndez, M., Smith, P.: Mining development and environmental in-justice in the Atacama desert of  
637 northern Chile. *Environ. Justice* 5, 70–76, 2012
- 638 91. Romero, L., Alonso, H., Campano, P., Fanfani, L., Cidu, R., Dadea, C., ... and Farago, M. Arsenic enrichment in  
639 waters and sediments of the Rio Loa (Second Region, Chile). *App Geochem*, 18(9), 1399-1416, 2003
- 640 92. Roosa, S., Wattiez, R., Prygiel, E., Lesven, L., Billon, G., and Gillan, D. C. Bacterial metal resistance genes and metal  
641 bioavailability in contaminated sediments. *Env pollution*, 189, 143-151, 2014
- 642 93. Salvarredy-Aranguren MM, Probst A, Roulet M, Isaure M-P.: Contamination of surface waters by mining wastes in  
643 the Milluni Valley (Cordillera Real, Bolivia): Mineralogical and hydrological influences. *Appl Geochem.*, 23: 1299–1324,  
644 2008
- 645 94. Staicu, L. C., and Stolz, J. F.: Microbes vs. metals: Harvest and recycle. *FEMS microbiol ecology*, 97(5), fiab056,  
646 2021
- 647 95. Stankevica, K., Vincevica-Gaile, Z., Klavins, M., Kalnina, L., Stivrins, N., Grudzinska, I., and Kaup, E.:  
648 Accumulation of metals and changes in composition of freshwater lake organic sediments during the Holocene. *Chem Geol*,  
649 539, 119502, 2020
- 650 96. Tapia, J., Davenport, J., Townley, B., Dorador, C., Schneider, B., Tolorza, V., and von Tümpling, W.: Sources,  
651 enrichment, and redistribution of As, Cd, Cu, Li, Mo, and Sb in the Northern Atacama Region, Chile: implications for arid  
652 watersheds affected by mining. *J Geochem Exploration*, 185, 33-51, 2018
- 653 97. Tapia, J., Mukherjee, A., Rodríguez, M. P., Murray, J., and Bhattacharya, P.: Role of tectonics and climate on elevated  
654 arsenic in fluvial systems: Insights from surface water and sediments along regional transects of Chile. *Env Pollution*, 314,  
655 120151, 2022
- 656 98. Tapia, J., Murray, J., Ormachea, M., Tirado, N., and Nordstrom, D. K. Origin, distribution, and geochemistry of  
657 arsenic in the Altiplano-Puna plateau of Argentina, Bolivia, Chile, and Perú. *STOTEN*, 678, 309-325, 2019



- 658 99. Teunisse, G. M.: Fantaxtic - Nested Bar Plots for Phyloseq Data (Version 2.0.1) [Computer software].  
659 <https://github.com/gmteunisse/Fantaxtic>, 2022
- 660 100. Thukral, A. K. A review on measurement of Alpha diversity in biology. *Agri Res J*, 54(1), 2017
- 661 101. Tomazini Jr, A., Lal, S., Munir, R., Stott, M., Henrissat, B., Polikarpov, I., ... and Levin, D. B.: Analysis of  
662 carbohydrate-active enzymes in *Thermogemmatospira* sp. strain T81 reveals carbohydrate degradation ability. *Canadian*  
663 *journal of microbiology*, 64(12), 992-1003, 2018
- 664 102. Trolle, D., Hamilton, D.P., Pilditch, C.A.: Evaluating the influence of lake morphology, trophic status and diagenesis  
665 on geochemical profiles in lake sediments. *Appl Geochem.*, 25:621–632, 2010
- 666 103. Usenko, S., Landers, D.H., Appleby, P.G., Simonich, S.L.: Current and historical deposition of PBDEs, pesticides,  
667 PCBs, and PAHs to Rocky Mountain National Park. *Environ Sci Technol.*, 41:7235–7241, 2007
- 668 104. Valero-Garcés, B. L., Delgado-Huertas, A., Navas, A., Edwards, L., Schwalb, A., and Ratto, N.: Patterns of regional  
669 hydrological variability in central-southern Altiplano (18–26°S) lakes during the last 500 years, *Palaeogeogr. Palaeoclimatol.*, 194,  
670 319–338, 2003.
- 671 105. Vargas-Machuca, B. D., Zanetta-Colombo, N., De Pol-Holz, R., and Latorre, C. Variations in local heavy metal  
672 concentrations over the last 16,000 years in the central Atacama Desert (22° S) measured in rodent middens. *STOTEN* 775,  
673 145849, 2021
- 674 106. Verosub, K. L., and Roberts, A. P.: Environmental magnetism: Past, present, and future. *J. Geophys.*, 100(B2), 2175-  
675 2192, 1995
- 676 107. Versteeg, J. K., Morris, W. A., and Rukavina, N. A.: The utility of magnetic properties as a proxy for mapping  
677 contamination in Hamilton Harbour sediment. *J Great Lakes Res.*, 21(1), 71-83, 1995
- 678 108. Vignale, F. A., Lencina, A. I., Stepanenko, T. M., Soria, M. N., Saona, L. A., Kurth, D., ... and Farías, M. E.: Lithifying  
679 and non-lithifying microbial ecosystems in the wetlands and salt flats of the Central Andes. *Microb Ecol.*, 1-17, 2021
- 680 109. Wasmund, K., Schreiber, L., Lloyd, K. G., Petersen, D. G., Schramm, A., Stepanauskas, R., ... and Adrian, L.:  
681 Genome sequencing of a single cell of the widely distributed marine subsurface *Dehalococcoidia*, phylum Chloroflexi. *ISME*  
682 *J.* 8, 383–397. doi: 10.1038/ismej.2013.143, 2014
- 683 110. Wickham, H.: *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York. ISBN 978-3-319-24277-4,  
684 <https://ggplot2.tidyverse.org>, 2016
- 685 111. Yan, C., Wang, F., Liu, H., Liu, H., Pu, S., Lin, F., ... and Yuan, R.: Deciphering the toxic effects of metals in gold  
686 mining area: microbial community tolerance mechanism and change of antibiotic resistance genes. *Env Res.*, 189, 109869,  
687 2020
- 688 112. Yan, Y., He, A., Dilek, Y., Zhu, Z., and Zhao, Q. Landscape inversion episodes in SE China during the Mesozoic–  
689 early Cenozoic: Constrained by trace-element contents, Nd isotope geochemistry, and detrital zircon U-Pb geochronology of  
690 sedimentary basins. *GSA Bulletin.*, 136(7-8), 2978-2998, 2024



- 691 113. Yang, C., Mai, J., Cao, X., Burberry, A., Cominelli, F., and Zhang, L.: ggpicrust2: an R package for PICRUSt2  
692 predicted functional profile analysis and visualization. *Bioinformatics*, 39(8), btad470, 2023
- 693 114. Yao, Z., Gao, P.: Heavy metal research in lacustrine sediment: a review. *Chin. J. Ocean. Limnol.* 25, 444–454, 2007.
- 694 115. Zanetta-Colombo, N. C., Fleming, Z. L., Gayo, E. M., Manzano, C. A., Panagi, M., Valdés, J., and Siegmund, A.:  
695 Impact of mining on the metal content of dust in indigenous villages of northern Chile. *Env Intl*, 169, 107490, 2022
- 696 116. Zanetta-Colombo, N. C., Scharnweber, T., Christie, D. A., Manzano, C. A., Blersch, M., Gayo, E. M., ... and Nüsser,  
697 M. When another one bites the dust: Environmental impact of global copper demand on local communities in the Atacama  
698 mining hotspot as registered by tree rings. *STOTEN*, 920, 170954, 2024
- 699 117. Zha, Y., Chong, H., Yang, P., and Ning, K. (2022). Microbial dark matter: from discovery to applications. *GPB*,  
700 20(5), 867-881.
- 701 117. Zhao, Y.N., Li, X.F., Ren, Y.P., Wang, X.H.: Effect of static magnetic field on the performances of and anode biofilms  
702 in microbial fuel cells. *RSC Adv* 6(85):82301–82308, 2016
- 703 118. Zhou, H., and van Hullebusch, E. D.: Microbial interaction and transformation of metals and metalloids. *Letters*  
704 *Applied Microbiol*, 75(5), 1074-1075, 2022
- 705
- 706