



## 1 What controls planktic foraminiferal calcification?


2 Ruby Barrett<sup>1</sup>, Joost de Vries<sup>2</sup>, Daniela N. Schmidt<sup>1</sup>


3 <sup>1</sup>School of Earth Sciences, University of Bristol, UK

4 <sup>2</sup>BRIDGE, School of Geography, University of Bristol, UK

5

6  RB 0000-0002-6805-1707

7  JDV 0000-0003-3427-6921

8  DNS 0000-0001-8419-2721

9 *Correspondence to:* Ruby Barret, [ruby.barrett@bristol.ac.uk](mailto:ruby.barrett@bristol.ac.uk)

10 **Abstract.** Planktic foraminifera are key producers of pelagic carbonate, and their shell weight is suggested  
11 represent the environment in which they calcify. However, there is debate about the use of size-normalised  
12 weight (SNW) as a proxy, as some authors invoke a carbonate system control on calcification (and by extension  
13 SNW as a  $p\text{CO}_2$  proxy), while others suggest that species optimum conditions, nutrient concentration, or  
14 temperature drive shell weight. To better understand its use as a proxy, we investigate what drives SNW and  
15 whether discrepancies in the proposed control on weight is due to differing data collection methodologies  
16 and/or regionally different drivers. We integrate new and published SNW data with environmental hindcast  
17 data extracted from the CMIP6 modelling suite. Using Bayesian regression modelling, we find that the  
18 environment alone cannot explain the variability in SNW across species. Although physiology likely modulates  
19 the response to the environment, we find little evidence of a unifying driver at the ecogroup-level. Instead, we  
20 identify species-specific responses associated with drivers including (but not limited to) the carbonate system,  
21 which are likely different between ocean basins. We hypothesise that this is partly influenced by cryptic species  
22 and regional phenotypic plasticity in not well understood changes to shell weight, such as the thickness of  
23 calcite deposited during some species' reproductive phase. Consequently, which species to use as a  $p\text{CO}_2$  proxy  
24 or whether multiple species should be used in parallel to reduce uncertainty should be carefully considered.  
25 We strongly encourage the regional testing and calibration of  $p\text{CO}_2$  – SNW relationships.

26

27 **Short summary.** Planktic foraminifera are a plankton whose fossilised shell weight is used to reconstruct past  
28 environmental conditions such as seawater  $\text{CO}_2$ . However, there is debate about whether other environmental  
29 drivers impact shell weight. Here we use a global data compilation and statistics to analyse what controls their  
30 weight. We find that the response varies between species and ocean basin, making it important to use regional  
31 calibrations and consider which species should be used to reconstruct  $\text{CO}_2$ .



32 **1 Introduction**

33 The unprecedented rise in CO<sub>2</sub> and temperature is altering our oceans and impacting marine ecosystems and  
34 **their services** the case of planktic foraminifera (a calcifying zooplankton which lives in the surface ocean),  
35 ocean acidification, sea surface warming and changing nutrient availability are all projected to impact their  
36 calcification (IPCC, 2022; Leung et al., 2022). Currently, these zooplankton contribute approximately a quarter  
37 of modern pelagic carbonate production (Buitenhuis et al., 2019; Langer, 2008) and 23–56% of total carbonate  
38 flux (Neukermans et al., 2023; Schiebel, 2002). The amount of carbonate produced by individual planktic  
39 foraminifers in the first order determines this flux to depth and is a function of their abundance, size and  
40 weight (Barrett et al., 2023). While research generally agrees on what drives foraminiferal size (Schmidt et al.,  
41 2004; c.f. Rillo et al., 2020) and abundance (Bé and Tolderlund, 1971), the controls on the size-normalized  
42 weight (SNW) of planktic foraminifers is debated (e.g. Aldridge et al., 2012; Barker & Elderfield, 2002; de  
43 Villiers, 2004; Lombard et al., 2010; Table 2).

44 As well as resolving what controls SNW to understand how carbonate production could be impacted by  
45 environmental change, it is also important for the interpretation of SNW as a proxy for past ocean conditions.  
46 That is whether SNW should be used to reconstruct carbonate saturation from bottom waters (Lohmann,  
47 1995), and/or as proxy for surface ocean carbonate, and by extension atmospheric pCO<sub>2</sub> (Barker and Elderfield,  
48 2002). The former stipulates that SNW records dissolution post deposition rather than environmental  
49 conditions during life. The latter supports the opposite – that SNW is controlled by carbonate ion concentration  
50 [CO<sub>3</sub><sup>2-</sup>] and records changes in the environment during life and the impact of post depositional processes are  
51 minimal (Russell et al., 2004). If variables other than the carbonate system control SNW, the use of this proxy  
52 should be reassessed.

53 There is **contradicting** evidence of a carbonate system control on foraminiferal calcification, with some studies  
54 showing a positive relationship between SNW and [CO<sub>3</sub><sup>2-</sup>], pH, and calcite saturation (Ω) (Barker & Elderfield,  
55 2002; Beer et al., 2010b; Bijma et al., 2002; Bijma et al., 1999; Broecker & Clark, 2001; Davis et al., 2017; de  
56 Moel et al., 2009; Dong et al., 2022; Lombard et al., 2010; Manno et al., 2012; Moy et al., 2009; Russell et al.,  
57 2004; Weinkauff et al., 2013). However, this response is not uniform between or even within species, with some  
58 studies reporting no response to [CO<sub>3</sub><sup>2-</sup>] (Béjard et al., 2023; Gonzalez-Mora et al., 2008; Henehan et al., 2017;  
59 Mallo et al., 2017; Naik et al., 2011; Pak et al., 2018; Song et al., 2022; Weinkauff et al., 2016). Others suggest  
60 that different environmental parameters are the primary control on SNW, such as temperature (Marr et al.,  
61 2011; Pak et al., 2018; Qin et al., 2020; Song et al., 2022), nutrient concentration (Aldridge et al., 2012), and  
62 optimum growth conditions (de Villiers, 2004). Importantly, many studies identify multivariate environmental  
63 controls on foraminiferal calcification, such as surface ocean carbonate chemistry, temperature, productivity,  
64 nutrient availability, salinity, (Béjard et al., 2023; Mallo et al., 2017; Marshall et al., 2013; Pallacks et al., 2023;  
65 Weinkauff et al., 2016), which can be species-specific and vary between and within ocean basins.

66 Physiology and ecological mechanisms such as biogeography or symbiosis may modulate the environmental  
67 response. Hence different ecogroups (i.e. species grouped by their ecology **which** have functional traits such as



68 spines in common; Table 1) may respond differently to the environment. For example, in symbiont bearing  
69 species the negative impact of low carbonate ion concentration could be reduced due to CO<sub>2</sub> uptake by  
70 symbionts in the foraminifer's microenvironment (Jørgensen et al., 1985; Köhler-Rink and Kühl, 2005; Rink et  
71 al., 1998). Species with spines may better capture food than non-spinose species (Gaskell et al., 2019; Spindler  
72 et al., 1984), providing energy for metabolic processes which support calcification.

73 SNW could additionally be variable between species due to potential differences in biomineralization  
74 pathways. Models suggest different biological controls, such as the intracellular storage of inorganic carbon and  
75 calcium ions (Erez, 2003), pH regulation (Lastam et al., 2023; de Nooijer et al., 2009; Toyofuku et al., 2017), and  
76 active transport of calcium and/or magnesium pumping (Bentov and Erez, 2006; Nehrke et al., 2013). These  
77 different pathways could have different sensitivities to environmental change. Furthermore, SNW  
78 measurements taken at the morphospecies level (i.e. species designated based on morphological features)  
79 could mask differences in the individual genotypes within cryptic species (i.e. organisms that look identical but  
80 represent distinct evolutionary lineages) if these have different environmental preferences (Darling et al., 2000;  
81 Morard et al., 2024).

82 Furthermore, the SNW response may vary spatially. For example, at higher latitudes where carbonate  
83 saturation is close to undersaturation (Mikis et al., 2019), a foraminifera may be at its limit of tolerance and  
84 therefore more vulnerable to small changes in carbonate ion concentration than low latitudes dwellers, akin to  
85 observations of coralline algae species responses to temperature changes at the trailing and leading edges of  
86 their distribution (Kolzenburg et al., 2023).

87 Additionally, the wide range in methodology used to collect weight measurements could also complicate our  
88 understanding of what drives SNW. Results are either generated with a sieved-based approach (SBW), in which  
89 planktic foraminifers are sieved through a narrow size fraction then the average specimen weight is taken, or  
90 through the measurement-based approach (MBW), where the additional step of normalizing to a measured  
91 size parameter (diameter or area) is taken (equation 1). MBW is a more rigorous approach as the use of sieve  
92 fractions (SBW) can be unreliable due to size variability within the sieve fraction itself (Aldridge et al., 2012;  
93 Beer et al., 2010a; Béjard et al., 2023)

$$MBW = \frac{Mean\ SBW_{sample} * Mean\ parameter_{size\ fraction}}{Mean\ parameter_{sample}} \quad (1)$$

94

95 Finally, different sample collection methodologies (i.e. whether results are derived from culture, plankton tow,  
96 core-top, or sediment trap samples) could further complicate our understanding of what drives calcification.  
97 Some authors have analysed foraminiferal SNW from plankton tow samples (Aldridge et al., 2012; Beer et al.,  
98 2010b; Mallo et al., 2017). However, foraminifers living in the water column are likely juvenile and have not  
99 completed calcification, meaning that anomalously light tests could be measured in comparison to the same  
100 size class derived from sediments. The SNW of sediment trap or core-top samples could be impacted by  
101 dissolution as foraminifera fall through the water column, however this can be largely accounted for if samples



102 are derived from above the lysocline. Culture experiments are useful in circumventing these limitations, but  
103 they do not reflect real-world conditions as many are grown in artificial seawater, and the meta-data collected  
104 is variable between publications limiting aggregation of studies.

105 Here, we apply Bayesian regression to statistically infer what drives SNW (measurement-based). We  
106 hypothesise that (1) the environment alone cannot explain variability in foraminiferal SNW across species.  
107 Instead, (2) physiology modulates the foraminiferal SNW response to the environment, hence the SNW  
108 response will be similar within ecogroups. (3) Species-specific SNW sensitivities may overprint the ecogroup  
109 response <sup>^</sup>

## 110 **2 Methods**

111 To infer which environmental variables drive SNW at both a species and group level, we conducted an  
112 exhaustive literature review, pre-processed our data to ensure data quality, and then statistically analysed our  
113 data using Bayesian regression modelling. Details for each step are provided below.

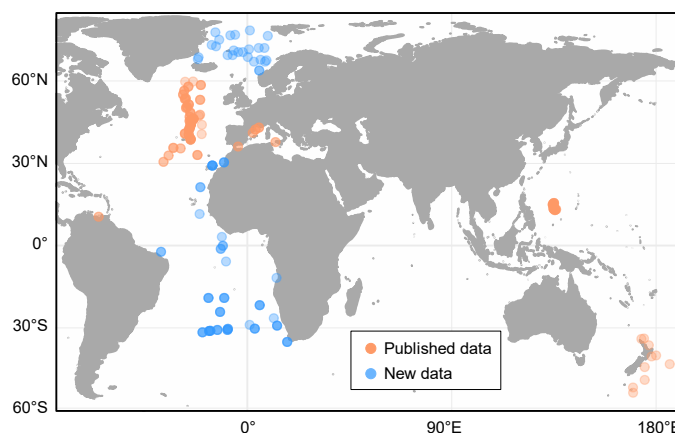
### 114 **2.1 Compilation of planktic foraminiferal SNW data**

115 This study gathers articles on foraminiferal SNW published until the 31<sup>st</sup> October 2023, and includes 790  
116 samples covering 11 species from 7 published datasets and a new dataset ( $n = 229$ ; Fig. 1; Text S1 and S2). A  
117 literature search for planktic foraminiferal SNW was conducted on Google Scholar. Publications with the key  
118 words 'planktic foraminifera' with 'size normalized weight', 'weight', 'calcification' were included. The results  
119 were expanded by exploring citations of key papers and identifying additional studies from the reference list of  
120 review articles. Articles were initially screened considering title relevance, then abstract content, and finally  
121 full-text content. Additionally, we included our own unpublished SNW which significantly increased data  
122 coverage in high latitudes and the subtropical Atlantic (Fig 1, see Text S1 for methodology). The full article list is  
123 available in the supplementary material and the new SNW data can be found in the supplementary data.

124 Data were only included if SNW was normalized by the measurement based weight (MBW) method as in  
125 equation 1 (Barker & Elderfield, 2002) using diameter or silhouette area. (Aldridge et al., 2012; Beer et al.,  
126 2010a; Béjard et al., 2023). Because the count of foraminifera collected can be low in sediment traps, selecting  
127 narrow size classes was not always possible for this data type as restricting sieve size would have resulted in a  
128 very small number of specimens. Data from plankton tows were removed from analysis as these may contain  
129 juvenile foraminifers. Given typical sedimentation rates in the open ocean and bioturbation, core-top data  
130 were considered preindustrial (unless the publication stated otherwise). Core samples were considered  
131 preindustrial if dated between 1000 AD and 1900 AD as CO<sub>2</sub> remained fairly stable over the Holocene (IPCC,  
132 2021). *G. ruber* white and *G. ruber* pink are combined to increase sample size.



133 Samples were omitted if dissolution of foraminifera specimens was reported, or if the water depth was more  
 134 than 4000m thereby approaching the CCD (Carbonate compensation depth; Broecker & Clark, 2009). Due to  
 135 sampling effort and preservation (i.e. the CCD being shallower in the Pacific), data are focused in the Atlantic  
 136 with only some Pacific data. Measurements span a wide latitudinal gradient (54°S to 78°N; Fig. 1). Planktic  
 137 foraminifers were assigned to one of three ecogroups following Aze et al. (2011) (Table 1).



138 **Figure 1** Location of SNW data. See Fig. S1 for a breakdown of species by location.  $n_{samples} = 790$

139

140

141 **Table 1** Planktic foraminifera species and their features which determine their ecogroup. The number in  
 142 brackets indicate the genotype counts from Morard et al. (2024).

| Species                     | Ecogroup                         | Habitat depth   | Cryptic diversification |
|-----------------------------|----------------------------------|-----------------|-------------------------|
| <i>G. bulloides</i>         | ymbiont-barren, spinose          | mixed layer     | High (10)               |
| <i>G. inflata</i>           | ymbiont-barren, non-spinose      | thermocline     | Low (2)                 |
| <i>N. pachyderma</i>        | ymbiont-barren, non-spinose      | mixed layer     | High (8)                |
| <i>G. truncatulinooides</i> | ymbiont-barren, non-spinose      | sub-thermocline | Moderate (5)            |
| <i>N. incompta</i>          | ymbiont-barren, non-spinose      | mixed layer     | Low (2)                 |
| <i>G. ruber</i>             | ymbiont-obligate, spinose        | mixed layer     | Moderate (4)            |
| <i>O. universa</i>          | ymbiont-obligate, spinose        | mixed layer     | Low (2)                 |
| <i>T. sacculifer</i>        | ymbiont-obligate, spinose        | mixed layer     | None (1)                |
| <i>G. elongatus</i>         | ymbiont-obligate, spinose        | mixed layer     | None (1)                |
| <i>N. dutertrei</i>         | ymbiont-facultative, non-spinose | thermocline     | None (1)                |
| <i>P. obliquiloculata</i>   | ymbiont-facultative, non-spinose | thermocline     | Low (2)                 |

143



144 **2.2 CMIP6 data extraction: compilation of environmental data**

145 For all SNW data, corresponding surface ocean environmental data were extracted from models in the CMIP6  
146 ensemble for the modern and preindustrial. Environmental data includes sea surface temperature, phosphate  
147 concentration, nitrate concentration, salinity, chlorophyll *a* concentration, net primary productivity (NPP),  
148 alkalinity, CO<sub>3</sub><sup>2-</sup>, DIC, Calcite Ω and pH.

149 Carbonate system, salinity and temperature data were derived from Jiang et al. (2023), in which 14 CMIP6  
150 ESMs were corrected for bias and model drift (see Table S1 and Jiang et al. 2023). Environmental data for the  
151 Mediterranean was not available from the Jiang et al. (2023). For this region, sea surface temperature (SST),  
152 sea surface salinity (SSS), dissolved inorganic carbon (DIC) and total alkalinity (TA) were extracted from CESM2  
153 (Danabasoglu et al., 2020) (Fig. S2) as the carbonate system output from CESM2 was closest to the median of  
154 the global average for the 14 ESMs (see Table S4 and S5 in Jiang et al. 2023).

155 The CESM2 data used in this manuscript were manipulated the same as other ESMs in Jiang et al. (2023). For  
156 consistency with other models, CESM2 outputs were converted from mol m<sup>-3</sup> to μmol kg<sup>-1</sup> using a density  
157 function calculated from the Thermodynamic Equation of Seawater (TEOS-10; IOC et al., 2010; McDougall &  
158 Barker, 2011). Interannual variability was reduced by calculating a 10 year average for each decade. Model bias  
159 was removed by correcting to DIVA gridded (Troupin et al., 2012) GLODAP (Lauvset et al., 2022) observational  
160 data and model drift was removed using the relevant CESM2 preindustrial control (piControl). The adjusted SST,  
161 SSS, DIC and TA were then used to calculate the rest of the OA indicators (CO<sub>3</sub><sup>2-</sup>, Calcite Ω and pH) using  
162 CO2System (van Heuven et al., 2011; Lewis and Wallace, 1998). Ice core-based atmospheric CO<sub>2</sub> data  
163 (Etheridge et al., 1996; MacFarling Meure et al., 2006) were used to approximate the oceanic fCO<sub>2</sub> change from  
164 1750 to 1850, thereby enabling estimation of the carbonate system for the preindustrial (1750) assuming that  
165 all locations are in equilibrium with the atmosphere (Takahashi et al., 2014).

166 Five Earth System Models (ESMs) were used to extract phosphate concentration, nitrate concentration,  
167 chlorophyll *a* concentration and net primary productivity (NPP) data to determine 'optimum conditions' (Table  
168 S1; Fig.S3). NPP and chlorophyll are indicators of the algal biomass concentration, which is a large part of some  
169 foraminifera species' diet (Schiebel and Hemleben, 2017). Nutrient concentration is a step detached from this,  
170 and represents the food available for their prey. There is some evidence that phosphate can inhibits  
171 calcification in some other calcifiers. Decadal averages were calculated for these variables. For comparison to  
172 existing data and to improve data readability phosphate and nitrate were converted from mol m<sup>-3</sup> to μmol kg<sup>-1</sup>,  
173 and chlorophyll *a* from kg m<sup>-3</sup> to mg m<sup>-3</sup>. The median of the non-corrected environmental outputs were  
174 calculated and the preindustrial (1750) values were assumed the same as in 1850. These data were not  
175 corrected to observational data as the data coverage is insufficient. Although species' abundance is also often  
176 used to inform optimum conditions, these data were not available for the same locations.

177



178 **2.3 Statistical modelling**

179 **2.3.1 Data cleaning: addressing size fraction bias and collinearity in environmental data**

180 All statistical analyses were carried out using R version 4.2.1 (R Core Team, 2018). To remove size fraction bias  
181 in SNW, the size fractions 250-300 and 300-350 were merged into one size fraction and this used. These size  
182 fractions were chosen because of their large sample number, they are in the middle of the size range, and  
183 allow us cover a wide environmental gradient (Fig. 2). This resulted in statistical analysis of 512 samples  
184 covering seven species from four published datasets and our data (Text S2).

185 Four of the initial ten environmental parameters were analysed: phosphate concentration, salinity, NPP, and  
186  $\text{CO}_3^{2-}$ . We were unable to analyse the impact of sea surface temperature due to collinearity, which would  
187 inflate the variance and standard error of coefficient estimates (Dormann et al., 2013). Nitrate was excluded as  
188 phosphate and nitrate concentration are highly correlated ( $\rho = 0.83$ ,  $p = <.000$ ). We chose to keep phosphate  
189 as it is more commonly assessed in the literature. Similarly, the carbonate system parameters are highly  
190 correlated (Fig. S4), but as carbonate ion concentration is often used in the literature we use this to represent  
191 the carbonate system. Because NPP is more directly linked with plankton biomass than chlorophyll  $a$   
192 concentration, the former is analysed here. Due to this data cleaning, it is important to note that while in the  
193 following we emphasise the parameter we analysed, the impacts on SNW could also be driven by the highly  
194 correlated driver.

195 **2.3.2 Model Specification**

196 All models were fitted using the Bayesian regression model package, brms (Bürkner, 2017) which uses the  
197 probabilistic programming language Stan (Carpenter et al., 2017). The models were specified to be Gamma  
198 distributed and were fitted using the NUTS (Hoffman and Gelman, 2014) sampler with 4 chains and 2000  
199 iterations, each of which the first 1000 are warmup to calibrate the sampler, thus leading to 4000 posterior  
200 samples.

201 All models were checked with appropriate tests before interpretation to ensure model assumptions were not  
202 violated. Variables were centred and standardised to reduce structural collinearity, and a QR decomposition  
203 term added to models to reduce correlation between variables. To check for any remaining collinearity pairs  
204 plots were visually assessed, and variance inflation factors (VIF) were verified using the package 'performance'  
205 which passes the brms model to its frequentist counterpart. A VIF of ten or less indicates that collinearity is not  
206 problematic (Marcoulides & Raykov, 2019). Outliers were detected using Pareto's  $k$ , for which a value of 0.7 or  
207 higher indicated an unduly influential observation. Visual posterior predictive checks were carried out to assess  
208 model fit and chain mixing (Fig. S5). An Rhat value close to 1 (i.e. less than 1.1) indicates the chains have  
209 converged (Bürkner, 2017).



210 **2.3.3 Modelling: Can the environment explain foraminiferal SNW across species?**

211 To assess whether there is a universal driver and how much variability in SNW across all foraminifers can be  
212 explained by the environment, a “group-level” (i.e. foraminifera species pooled together;  $n_{samples} = 512$ )  
213 Bayesian multi-level model was fitted (Bürkner, 2018). The full model included carbonate ion concentration  
214 ( $CO_3^{2-}$ ), salinity, phosphate concentration, and net primary productivity (NPP) as fixed environmental effects  
215 and species as a random effect (intercept only). Data type (i.e. sediment trap, sediment core and core-top) was  
216 added as a fixed effect (not a random effect because data type had less than five levels (Harrison et al., 2018).  
217 Because the range of variance was unequal (“heteroscedastic”) between species (Fig. S6), we add a shape term  
218 to the model which allows the variance between each species to vary.

219 The full model was compared to a ‘null’ model which did not consider species and included fixed  
220 environmental effects only (the impact of data type as a fixed effect was removed from bayes R2 values to  
221 ensure it was environmental effect only that was measured). Both models were compared using leave-one-out  
222 cross-validation (‘LOO’; Vehtari et al., 2017), a measure which informs which model is performing best.

223 LOO indicated that adding species as a random effect improved model fit ( $\widehat{elpd}_{loo}$  improved by  $261.3 \pm 18.6$ ,  
224 see details in results). As such, we fit models for individual species to assess their association with the  
225 environment.

226 **2.3.4 Modelling: Is the SNW response to the environment similar between ecogroups or species specific?**

227 The size fraction restriction imposed for analysis of SNW across species (250-350  $\mu m$  only) was relaxed (Text S2)  
228 as it is less relevant at the species-level which recognises the size ranges of taxa. Only sieve size fractions that  
229 are 50  $\mu m$  in range were used (unless data were from sediment traps). Similar to the group-level model, data  
230 type was added as a fixed effect for each species-level model. (data, *T. sacculifer*, *N. dutertrei*, *P.*  
231 *obliquiloculata* and *O. universa* were not modelled because of their low number of observations ( $n < 30$ ). *N.*  
232 *incompta* was excluded from analysis because of significant multi-collinearity that prevented meaningful  
233 inference of environmental effects. Bayesian models were fitted to the remaining five species. To assess how  
234 much of the variability in foraminiferal SNW for different species can be explained by the environment, the  
235 effect size and credible interval (i.e. Bayesian confidence interval) of coefficients (environmental variables)  
236 were extracted from each model. Results were clustered by ecogroup to assess whether there were differences  
237 in the SNW response to the environment between ecogroups (Fig. 4).





238 **3 Results**

239 **3.1 Qualitative assessment of existing data**

240 Assessing the available SNW data and their suggested drivers in the literature, there is no single environmental  
241 control on foraminiferal size normalised weight across species (Table 2). Although this summary suggests that a  
242 low carbonate ion concentration does not reduce foraminiferal SNW, it is inconclusive as to whether an  
243 increase in carbonate ion concentration has no impact on shell weight or increases it. For other environmental  
244 variables, it is either a mixed response or there is too little information to determine a direction of response.  
245 However, it is important to note that where no significant effect is reported in Table 2, this could possibly  
246 reflect the lack of statistical power rather than no response. Using environmental data from earth system  
247 models allows us to reanalyse the data and determine whether any environmental drivers emerge for SNW  
248 across all species.



249 **Table 2** Compilation of results from previous studies assessing the relationship between planktonic  
 250 foraminiferal size-normalized weight (SNW) and the environment. + = positive correlation, - = negative  
 251 correlation, ~ = no response. This table summarizes information from measurement based SNW (i.e. silhouette  
 252 area, or diameter normalised) studies only and omits those which only normalised to size by sieving (i.e. sieve-  
 253 based weights; SBW) or use plankton tow data. See supplementary Table S2 for detail on SNW measurement  
 254 method. [1] Barker & Elderfield (2002); [2] Bédard et al. (2023); [3] Marr et al. (2011); [4] Marshall et al. (2013);  
 255 [5] Osborne et al. (2016); [6] Pallacks et al. (2023); [7] Weinkauff et al. (2016).

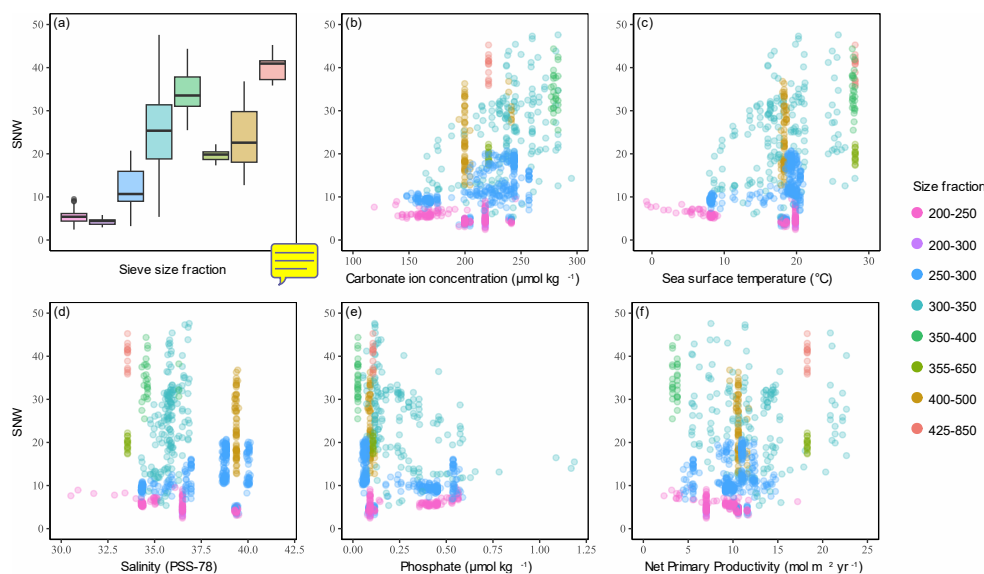
256

| Species                             | Data type | Biogeozone  | Carbonate ion | pH | CO <sub>2</sub> | Temperature | Productivity | Phosphate | Nitrate | Salinity | Optimum conditions |
|-------------------------------------|-----------|-------------|---------------|----|-----------------|-------------|--------------|-----------|---------|----------|--------------------|
| <i>symbiont-barren, spinose</i>     |           |             |               |    |                 |             |              |           |         |          |                    |
| <i>G. bulloides</i> <sup>6</sup>    | Core      | Subtropical | +             |    | -               | -           |              |           |         |          |                    |
| <i>G. bulloides</i> <sup>1</sup>    | Core-top  | Temperate   | +             |    |                 | ~           |              |           |         |          |                    |
| <i>G. bulloides</i> <sup>3</sup>    | Core-top  | Subtropical |               |    |                 | -           |              |           |         |          |                    |
| <i>G. bulloides</i> <sup>5</sup>    | Trap/Core | Tropical    | +             |    |                 | ~           |              | ~         |         |          |                    |
| <i>G. bulloides</i> <sup>2</sup>    | Trap      | Subtropical | ~             | ~  | ~               | ~           | ~            | ~         | ~       | ~        | ~                  |
| <i>G. bulloides</i> <sup>7</sup>    | Trap      | Subtropical | ~             |    |                 | ~           | ~            |           |         |          | -                  |
| <i>symbiont-obligate, spinose</i>   |           |             |               |    |                 |             |              |           |         |          |                    |
| <i>G. elongatus</i> <sup>6</sup>    | Core      | Subtropical | +             |    | -               | -           |              |           |         |          |                    |
| <i>G. elongatus</i> <sup>7</sup>    | Trap      | Subtropical | ~             |    |                 | +           | -            |           |         |          | +                  |
| <i>G. ruber</i> <sup>7</sup>        | Trap      | Subtropical | ~             |    |                 | +           | -            |           |         |          | ~                  |
| <i>G. ruber</i> <sup>4</sup>        | Trap      | Tropical    | +             |    |                 | +           |              |           |         |          |                    |
| <i>G. sacculifer</i> <sup>4</sup>   | Trap      | Tropical    | +             |    |                 | +           |              |           |         |          |                    |
| <i>symbiont-barren, non-spinose</i> |           |             |               |    |                 |             |              |           |         |          |                    |
| <i>G. inflata</i> <sup>1</sup>      | Core-top  | Temperate   | +             |    |                 | ~           |              |           |         |          |                    |
| <i>G. trunc</i> <sup>1</sup>        | Core-top  | Temperate   | +             |    |                 | ~           |              |           |         |          |                    |
| <i>G. trunc</i> <sup>2</sup>        | Trap      | Subtropical | +             | ~  | ~               | +           | -            | ~         | ~       | ~        | -                  |
| <i>N. incompta</i> <sup>2</sup>     | Trap      | Subtropical | ~             | ~  | ~               | +           | ~            | ~         | ~       | ~        | ~                  |
| <i>N. incompta</i> <sup>1</sup>     | Core-top  | Temperate   | +             |    |                 | ~           |              |           |         |          |                    |



257 **3.2 Qualitative assessment of reanalysed data**

258 Here we qualitatively assess the integrated published SNW and new SNW dataset alongside the environmental  
 259 output from the CMIP6 modelling suite. Generally, larger foraminifers (e.g. 425-850  $\mu\text{m}$ ) have heavier tests  
 260 (average 40.14  $\mu\text{g}$ ) and smaller foraminifers (e.g. 200-250  $\mu\text{m}$ ) have lighter tests (average 5.49  $\mu\text{g}$ ; (Fig. 2a). The  
 261 300-350  $\mu\text{m}$  size fraction shows greatest variability in weight (standard deviation [ $\sigma$ ] 9.32; Fig. 2a), likely as it  
 262 has a higher species diversity ( $n = 5$ ) compared to other size fractions ( $n = 1$  to 4). Interestingly, the second  
 263 highest variability in weight is for the 400-500  $\mu\text{m}$  size fraction ( $\sigma$  6.77; Fig. 2a) and is linked to only one  
 264 species, *G. truncatulinoides*, from one publication (Béjard et al., 2023; Fig. S7). The species is atypical as a very  
 265 large proportion of the weight is in the gametogenic calcite covering the entire test (Schmidt et al., 2008)  
 266 whose thickness might be driven by environmental parameters as well. Furthermore, the species has a year-  
 267 long life cycle (whilst other species analysed here have lunar cycles and peak in a specific season), meaning  
 268 that this species is exposed to greater environmental variability throughout the year. The lack of environmental  
 269 variability shown here for these samples likely reflects averaging of the seasons in this annual environmental  
 270 record.



271

272 **Figure 2** (a) Boxplot showing SNW distribution across sieve size fractions. (b-f) Planktic foraminiferal size-  
 273 normalised weight (MBW) against environmental variables extracted from the CMIP6 modelling suite (see  
 274 methods). Colour indicates the size-fraction foraminifers were initially sieved at before being normalised to  
 275 their length or area. See Fig. S7 for planktic foraminiferal SNW separated by species, with sieve size fraction  
 276 information.

277

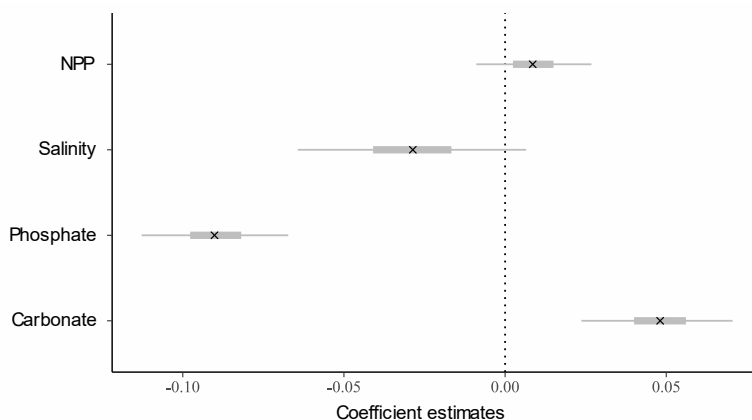
278 The smallest size fractions must be interpreted with caution (Fig. 2) as they have not been systematically  
 279 assessed in warm regions (where carbonate ion concentration is higher) due to a preference for using larger  
 280 sieve size fractions in these regions. Although the smaller size fractions are meaningful in polar and subpolar



281 areas (as foraminifers are smaller at the poles), there are data missing for small sizes in warm, high calcite  
282 environments. The absence of heavy foraminifer in low carbonate ion saturation (Fig. 2b) and cool (Fig. 2c)  
283 environments suggest that these environments limit foraminiferal weight. To remove size fraction bias, the size  
284 fractions 250-300 and 300-350 have been merged to create a 250-350 size fraction and (unless stated  
285 otherwise) the following statistics has been performed on this reduced dataset.

### 286 3.3 Is there an environmental control on SNW at the group-level?

287 We use Bayesian regression to determine whether there is an environmental control on SNW across  
288 species. An 'environment only' model explains 23% of the variability in SNW (Bayes R<sup>2</sup>; Gelman et al., 2019),  
289 whilst a model which additionally includes species as a random effect explains 86% of the variability in SNW,  
290 indicating that species-specific differences are more important than environmental effects for SNW at the  
291 group-level. Higher SNWs are associated with a higher carbonate ion concentration (0.05 [0.02, 0.07]; effect  
292 size and 95% credible interval [lower, upper]; Fig. 3) and lower phosphate concentration (-0.09 [-0.11, -0.07];  
293 Fig. 3; Table S3), though the effect size is small. To dive deeper into the link between SNW and the  
294 environment, Bayesian models were fitted at the species level.



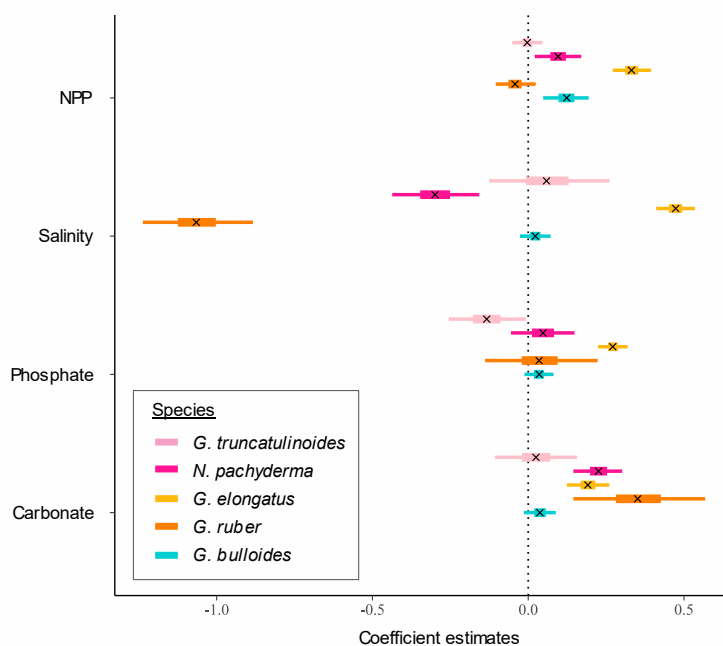
295 **Figure 3** Effect size and credible intervals for the association between SNW and the environment for the group-  
296 level model. A cross [x] represents the median value, the thicker line the 50% interval (i.e. where 50% of the  
297 posterior probability lies) and the thinner line the 95% interval. If the 95% interval does not cross zero, then  
298 there is a 95% probability there is an effect of the environmental variable. A negative value represents a  
299 negative correlation between SNW and the coefficient. Note that the modelled dataset is slightly different to  
300 the species-level dataset. The group-level model dataset includes species which were omitted from species-  
301 level models due to their low sample size, and the size fraction ranges are more restricted for the group-level  
302 model due to a bias against larger size fractions in cooler environments (see methods).

303



304 **3.4 Is there a species specific or an ecogroup response?**

305



306 **Figure 4** Effect size and credible intervals for the association between SNW and the environment for the species-  
 307 level Bayesian modelling. A cross [x] represents the median value, the thicker line the 50% interval (i.e. where  
 308 50% of the posterior probability lies) and the thinner line the 95% interval. If the 95% interval does not cross zero  
 309 then there is a 95% probability there is an effect of the environmental variable. A negative value represents a  
 310 negative correlation between SNW and the environmental variable. Ecogroups are grouped by colour. *G.*  
 311 *bulloides* is a symbiont barren, spinose species. *G. ruber* and *G. elongatus* are symbiont-obligate, spinose species.  
 312 *N. pachyderma* and *G. truncatulinoides* are symbiont barren, non-spinose species.

313

314 In agreement with published literature (Aldridge et al., 2012; Barker and Elderfield, 2002; Béjard et al., 2023;  
 315 Marshall et al., 2013; Osborne et al., 2016; Pallacks et al., 2023), an increase in carbonate ion concentration  
 316 does not negatively impact SNW (Fig. 4; Table S3). The relationship is not always positive though, with *G.*  
 317 *bulloides* (0.04 [-0.02, 0.09]) and *G. truncatulinoides* (0.03 [-0.11, 0.16]) exhibiting no notable response to a  
 318 change in carbonate ion concentration (i.e. 95% interval crosses zero).

319 It remains up for debate which part of the carbonate system exerts control on calcification. It has been  
 320 suggested that the  $\text{HCO}_3^- / \text{H}^+$  ratio (where  $\text{HCO}_3^-$  [bicarbonate ions] are the inorganic carbon substrate and  $\text{H}^+$   
 321 [protons] are a calcification inhibitor) controls calcification and that  $\text{CO}_3^{2-}$  correlates because of a  
 322 proportionality between  $\text{CO}_3^{2-}$  and this ratio (Bach, 2015). Yet even if this is the case, this implies that  $\text{CO}_3^{2-}$  can  
 323 be proxy for the  $\text{HCO}_3^- / \text{H}^+$  ratio hence it is still important for calcification.



324 An increase in phosphate concentration is unlikely (<95% probability and <50% probability for *G. ruber*) to  
325 impact SNW other than for *G. truncatulinoides* (−0.13 [−0.26, −0.01]), and *G. elongatus* (0.27 [0.22, 0.32]). For  
326 the former, increased phosphate may reduce SNW and for the latter, SNW increases with phosphate  
327 concentration (Fig. 4; Table S3). Given the evidence for calcification inhibition in high phosphate conditions (Lin  
328 and Singer, 2006) for other calcifiers, such as corals (Kinsey and Davies, 1979), coccolithophores (Paasche and  
329 Brubak, 1994), and calcifying green algae (Demes et al., 2009), it is interesting that we do not observe stronger  
330 detrimental effect of phosphate on these foraminiferal species. However, this disparity could be explained by  
331 the different calcification mechanisms. For example, foraminifers biomineralize extracellularly by engulfing  
332 calcite-forming materials through seawater vacuolisation (potentially assisted by transmembrane ion transport;  
333 Bentov et al., 2009; de Nooijer et al., 2014; Erez, 2003; Nehrke et al., 2013). In contrast, coccolithophores  
334 biomineralize by forming coccoliths in intracellular organelles called ‘coccolith forming vesicles’ (Brownlee and  
335 Taylor, 2004).

336 There is no consensus on the impact of phosphate on calcification even within a taxon, with a recent study on  
337 coccolithophores not showing calcification inhibition but instead showing decreased calcification with  
338 phosphate limitation (Gerecht et al., 2018). Hence pointing to other taxa exhibiting similar response to our  
339 species-level modelling. Our *G. bulloides* result conflicts with a study of North Atlantic *G. bulloides*, in which a  
340 decrease in SNW with increased phosphate was recorded (Aldridge et al., 2012), though Bédard (2023) and  
341 Mallo et al. (2017) did not observe this in the Mediterranean. This disparity could be due to the use of shallow  
342 plankton tows in Aldridge et al. (2012) which is likely to complicate the SNW signal as juveniles which had not  
343 completed their development may have been measured. Additionally, *G. bulloides* has several cryptic species  
344 (Morard et al., 2024) which have their own ecological adaptation and spatial variability. Hence the geographic  
345 difference might further complicate the interpretation of data in these studies (Fig. S2). In our group-level  
346 model though we observe a negative impact of phosphate on SNW across species (−0.09 [−0.11, −0.07]; Fig. 3;  
347 Table S3). This is unlikely an effect of sampling bias toward the Atlantic as the Atlantic has near-even sampling  
348 ( $n = 263$ ) to the Mediterranean ( $n = 239$ ). Instead, as the group-level model contains some different species  
349 than the species-level modelling, we suggest that this difference reflects that certain species of foraminifera  
350 are sensitive to phosphate, while others are not.

351 Salinity has a mixed impact on foraminiferal SNW. For *G. ruber* SNW is lighter at high salinity (−1.06 [−1.24,  
352 −0.88]), and *N. pachyderma* has a similar but weaker response (−0.30 [−0.44, −0.16]; Fig. 4; Table S3).  
353 Meanwhile, the SNW of *G. elongatus*, closely related to *G. ruber* and by some assumed to be an ecotype,  
354 increases with salinity (0.47 [0.41, 0.54]). Laboratory experiments which exposed foraminifers to a wider  
355 salinity range than observed under normal ocean conditions concluded that *G. ruber* was most tolerant to  
356 changes in salinity out of the seven species analysed (Bijma et al., 1990). For other foraminiferal species, they  
357 found that under low salinity growth rate reduced and final size was smaller. This difference could be because  
358 salinity values reported by Bijma et al. (1990) were more extreme than normal ocean conditions, or that  
359 growth rate and size are impacted differently to weight, i.e. foraminifers could be smaller but have a thicker  
360 test. Unfortunately, weight was not recorded in the study so this cannot be tested.



361 Similar to carbonate ion concentration, it is unlikely (<95% probability) that an increase in NPP decreases SNW.  
362 Instead, for *N. pachyderma*, *G. elongatus* and *G. bulloides*, increasing NPP (food availability) results in a heavier  
363 SNW (Fig. 4; Table S3). For *G. elongatus*, their symbionts should make the species less dependent on  
364 productivity due to cross transfer of sugars (LeKieffre et al., 2018), and for *G. bulloides* the presence of spines  
365 should make it easier for them to capture prey therefore should similarly be less associated with NPP. Yet, both  
366 SNWs increase with food availability (*G. bulloides*: 0.12 [0.05, 0.19]; *G. elongatus*: 0.33 [0.27, 0.40]). Even in the  
367 asymbiotic, non-spinose ecogroup, *N. pachyderma* and *G. truncatulinoides* there is no clear pattern, with the  
368 former's SNW increasing with productivity (0.09 [0.02, 0.17]) and the latter showing no response (-0.00 [-0.05,  
369 0.05]). It is interesting that despite constructing a secondary calcite crust (which could overprint the primary  
370 SNW signal), *N. pachyderma* (Kohfeld et al., 1996) still exhibits a response to the environment.

371 Due to collinearity we are unable to assess the impact of SST on SNW for the species-level models. However,  
372 we could expect an increase in SNW with warming as warmer water decreases the solubility of atmospheric  
373 CO<sub>2</sub>, which elevates surface water carbonate ion concentration, and also increases enzymatic activity which  
374 promotes growth and calcification rate (Lombard et al., 2009; Spero et al., 1991). Although some past research  
375 has identified an increase in SNW with warming (Béjard et al., 2023; Davis et al., 2013; Gonzalez-Mora et al.,  
376 2008; Marshall et al., 2013; Osborne et al., 2016; Qin et al., 2020; Song et al., 2022; Weinkauff et al., 2016),  
377 there is also evidence for the reverse (Mallo et al., 2017; Naik et al., 2010, 2011; Pallacks et al., 2023). This  
378 dichotomy has been attributed to overriding effect of decreasing carbonate ion concentration on SNW due to  
379 ocean carbon input (Naik et al., 2010; Pallacks et al., 2023), temperature induced sea surface stratification and  
380 lower food availability (Mallo et al., 2017).

381 Due to limited shell flux data, we were unable to investigate how optimum growth conditions (OGC) impacted  
382 SNW. Although NPP may facilitate OGC by making food available for growth, we cannot assume that high NPP  
383 results in optimum conditions as it also hinders photosynthesis and excludes species (Ortiz et al., 1995). There  
384 is some evidence of SNW increasing where a species is at its OGC (i.e. where shell flux for that species is high;  
385 de Villiers, 2004), but there is no consensus in the data (Table 2) with some observing a negative correlation  
386 between OGC and SNW (Béjard et al., 2023; Weinkauff et al., 2016).

387 The SNW response to the environment is largely species specific and shows little evidence of an overriding  
388 ecological driven response. For some taxa, similar responses can be found, e.g. the symbiont-obligate, spinose  
389 species *G. ruber* and *G. elongatus* show the same direction of response to carbonate, though the strength of  
390 response is variable (0.35 [0.14, 0.56] and 0.19 [0.12, 0.26], respectively; Fig. 4; Table S3). The symbiont barren,  
391 non-spinose species (*N. pachyderma*, and *G. truncatulinoides*) lack a unifying driver linked to their ecology.  
392 Though it is important to note that the SNWs of these species are likely to be more heavily impacted the  
393 production of a secondary calcite crust than other species analysed here (Kohfeld et al., 1996; Schmidt et al.,  
394 2008).



395 **3.5 Should SNW be used as proxy for CO<sub>2</sub>?**

396 Disentangling the controls on SNW is important for understanding the use of SNW as a proxy for interpreting  
397 past ocean conditions. This paper cautions the use of planktic foraminiferal SNW as a reliable proxy for the  
398 surface ocean carbonate system and palaeo  $p\text{CO}_2$ .

399 Although there is a small but likely (i.e. >95% probability) effect of carbonate on a group level (i.e. across  
400 species; 0.05 [0.02, 0.07]), phosphate is also likely associated with SNW (-0.09 [-0.11, -0.07]; Fig. S3; Table  
401 S3). Hence, unless the impact of phosphate on SNW can be quantified and disentangled from the carbonate  
402 effect, SNW is not a reliable predictor for  $p\text{CO}_2$ . As SNW is variable on a species level, there is a need to  
403 consider which species to use for paleo proxies, or a need to consider multiple species in parallel to reduce  
404 uncertainty from species-specific differences.

405 Although the use of SNW to inform past CO<sub>2</sub> has been shown to work regionally with certain species, e.g. *G.*  
406 *bulloides* in the North Atlantic (Barker and Elderfield, 2002), the relationship between SNW and carbonate ion  
407 concentration seems to break down when taken out of its calibration region. When expanding the *G. bulloides*  
408 dataset to include Pacific, Mediterranean and higher latitude North Atlantic samples (Fig. S1) we find no  
409 correlation between SNW and carbonate ion concentration. Hence we advocate for the regional calibration of  
410  $p\text{CO}_2$  – SNW relationships, and caution against the extrapolation and global application of SNW as proxy for  
411  $p\text{CO}_2$ .

412 One of the challenges in assessing a unifying calcification response is unequal methodologies and data  
413 reporting. In this paper 57 publications were screened for their SNW data, but only 7 publications (and our  
414 data) could be used for the species-level modelling. Around half were omitted as they were older than  
415 preindustrial and therefore could not be used to determine drivers. Otherwise, data were often not freely  
416 available (or at all available) and if deposited, only provided processed data with different methods of  
417 normalising weight to size. We strongly encourage the community to deposit raw data to make the legacy of  
418 data longer. 28 publications were omitted because shell weights were reported using the sieve-based weight  
419 (SBW) methodology and not normalised to size or area (MBW). Although there is some debate as to whether  
420 this additional step of normalising weight to measurement-based size is necessary, some publications  
421 (Aldridge et al., 2012; Beer et al., 2010a; Bédard et al., 2023) indicate that MBW SNW is more robust than SBW.  
422 It would be a step forward for the community to derive protocols for SNW akin to trace element analysis e.g.  
423 Hathorne et al. (2013) and Rosenthal et al. (2004). Additionally, it is important to acknowledge the different  
424 developmental stages in plankton tow samples compared to sediment trap and core-top samples. Post-  
425 depositional dissolution will reduce weights, while infilling and diagenesis increase weight and both need to be  
426 carefully monitored (Bassinot et al., 1994; Broecker & Clark, 2001). Additionally, we still have important gaps in  
427 our understanding of foraminiferal ecology, for example the dynamics of the habitat throughout the year, the  
428 peak times of biomass production in different regions and the drivers of thickness of gametogenic calcite. All  
429 of these factors limit the use of the proxy.





430 Importantly, our analyses lack data from the Indian Ocean, Southern high latitudes and large parts of the  
431 Pacific - highlighting challenges of preservation in deep sea sediments, logistics of reaching remote areas, and  
432 bias due to the traditional areas of sampling of sea going nations. As analyses expand to ocean regions below  
433 the lysocline, authors should provide a measure of dissolution and/or high resolution images of specimens  
434 which can help assess the impact of post-diagenetic alteration. Although such images can also support  
435 morphological assessment of cryptic species, these images are still not systematically implemented in  
436 palaeoceanographic studies.

#### 437 **4 Conclusions**

438 Although higher carbonate ion concentration and lower phosphate concentration are associated with heavier  
439 SNWs at the group-level (i.e. across species), the environment alone explains relatively little of the variability in  
440 SNW at the group-level. Instead, we identify species-specific SNW responses that better explain variability in  
441 weight. Although physiology is likely to modulate the foraminiferal response to the environment, we find  
442 limited evidence of an ecogroup-level response.

443 The species-specific SNW response to the environment is complex, with each species responding to a different  
444 combination of environmental drivers. We hypothesise that this is in part influenced by cryptic species and our  
445 limited understanding of what drives the thickness of gametogenic calcite. The SNW response being species-  
446 specific and responding to drivers other than carbonate implies there is a need to consider which species to  
447 use as a  $p\text{CO}_2$  proxy, or a need to consider multiple species in parallel to reduce uncertainty from species-  
448 specific differences. Furthermore, due to differences in the published response of *G. bulloides* in the North  
449 Atlantic and our more global dataset of *G. bulloides* SNW, we advocate for the regional calibration of  $p\text{CO}_2$ –  
450 SNW relationships.

451 Our understanding of SNW as a proxy would be greatly improved with some community efforts to solve some  
452 of the above questions including (1) making raw SNW data freely available, (2) community agreed protocols,  
453 i.e. whether SBW or MBW should be used in such analyses, (3) improving our understanding of the calcification  
454 process itself and how the environment drives the thickness of gametogenic calcite, and (4) resolving the  
455 impact that cryptic species have on SNW measurements.



456 **Code availability**

457 The code (R script) supporting this article has been uploaded as part of the supplement and is available at

458 DOIXXXX

459 **Data availability**

460 All data used in this study are available at DOIXXXX

461 **Supplement**

462 The supplement related to this article is available at: DOIXXXX

463 **Author contribution**

464 R.B. and D.N.S. conceptualised the study. R.B. collated existing SNW data and processed CMIP6 model data,  
465 and conducted analysis of these data. J.V. contributed to the methodological design and statistical analysis. R.B.  
466 prepared the manuscript with contributions from all co-authors.

467 **Competing interests**

468 The authors declare that they have no conflict of interest.

469

470

471 **Acknowledgements**

472 This work would not have been possible without the efforts of the ODP and IODP programs. We acknowledge  
473 the University of Bremen, Dr Barbara Donner and Dr KH Baumann for sharing samples. We would like to thank  
474 Dale Thompson and Dr Maricel Williams for processing samples to produce these new SNW data. Thank you to  
475 Dr Li-Qing Jiang for providing support early on in the study design, and for sharing their processed CMIP6 data.

476 **Financial support**

477 R.B. was funded by NERC GW4+ DTP grant NE/S007504/1

478 D.N.S. was funded by NERC grant NE/P019439/1

479 J.V. was funded by NERC grant NE/X001261/1

480

481



482 **References**

- 483 Aldridge, D., Beer, C. J., and Purdie, D. A.: Calcification in the planktonic foraminifera *Globigerina bulloides*  
484 linked to phosphate concentrations in surface waters of the North Atlantic Ocean, *Biogeosciences*, 9, 1725–  
485 1739, <https://doi.org/10.5194/bg-9-1725-2012>, 2012.
- 486 Aze, T., Ezard, T. H. G., Purvis, A., Coxall, H. K., Stewart, D. R. M., Wade, B. S., and Pearson, P. N.: A phylogeny  
487 of Cenozoic macroperforate planktonic foraminifera from fossil data, *Biol. Rev. Camb. Philos. Soc.*, 86, 900–  
488 927, <https://doi.org/10.1111/j.1469-185X.2011.00178.X>, 2011.
- 489 Bach, L. T.: Reconsidering the role of carbonate ion concentration in calcification by marine organisms,  
490 *Biogeosciences*, 12, 4939–4951, <https://doi.org/10.5194/BG-12-4939-2015>, 2015.
- 491 Barker, S. and Elderfield, H.: Foraminiferal calcification response to glacial-interglacial changes in atmospheric  
492 CO<sub>2</sub>, *Science* (80-. ), 297, 833–836, <https://doi.org/10.1126/science.1072815>, 2002.
- 493 Barrett, R., Adebowale, M., Birch, H., Wilson, J. D., and Schmidt, D. N.: Planktic Foraminiferal Resilience to  
494 Environmental Change Associated With the PETM, *Paleoceanogr. Paleoclimatology*, 38,  
495 <https://doi.org/10.1029/2022PA004534>, 2023.
- 496 Bassinot, F. C., Beaufort, L., Vincent, E., Labeyrie, L. D., Rostek, F., Müller, P. J., Quidelleur, X., and Lancelot, Y.:  
497 Coarse fraction fluctuations in pelagic carbonate sediments from the tropical Indian Ocean: A 1500-kyr record  
498 of carbonate dissolution, *Paleoceanography*, 9, 579–600, <https://doi.org/10.1029/94PA00860>, 1994.
- 499 Bé, A. W. H. and Tolderlund, D. S.: Distribution and ecology of living planktonic foraminifera in surface waters  
500 of the Atlantic and Indian Oceans, in: *The Micropalaeontology of Oceans*, Cambridge University Press, 105–  
501 149, 1971.
- 502 Beer, C. J., Schiebel, R., and Wilson, P. A.: On methodologies for determining the size-normalised weight of  
503 planktic foraminifera, *Biogeosciences*, 7, 2193–2198, <https://doi.org/10.5194/bg-7-2193-2010>, 2010a.
- 504 Beer, C. J., Schiebel, R., and Wilson, P. A.: Testing planktic foraminiferal shell weight as a surface water [CO<sub>2</sub>-3]  
505 proxy using plankton net samples, *Geology*, 38, 103–106, <https://doi.org/10.1130/G30150.1>, 2010b.
- 506 Béjard, T. M., Rigual-Hernández, A. S., Flores, J. A., Tarruella, J. P., Durrieu De Madron, X., Cacho, I., Haghipour,  
507 N., Hunter, A., and Sierro, F. J.: Calcification response of planktic foraminifera to environmental change in the  
508 western Mediterranean Sea during the industrial era, *Biogeosciences*, 20, 1505–1528,  
509 <https://doi.org/10.5194/BG-20-1505-2023>, 2023.
- 510 Bentov, S. and Erez, J.: Impact of biomineralization processes on the Mg content of foraminiferal shells: A  
511 biological perspective, *Geochemistry, Geophys. Geosystems*, 7, 8, <https://doi.org/10.1029/2005GC001015>,  
512 2006.
- 513 Bentov, S., Brownlee, C., and Erez, J.: The role of seawater endocytosis in the biomineralization process in  
514 calcareous foraminifera, *Proc. Natl. Acad. Sci.*, 106, 21500–21504, <https://doi.org/10.1073/PNAS.0906636106>,



- 515 2009.
- 516 Bijma, J., Faber, W. W., and Hemleben, C.: Temperature and salinity limits for growth and survival of some  
517 planktonic foraminifers in laboratory cultures, *J. Foraminifer. Res.*, 20, 95–116, 1990.
- 518 Bijma, J., Spero, H. J., and Lea, D. W.: Reassessing Foraminiferal Stable Isotope Geochemistry: Impact of the  
519 Oceanic Carbonate System (Experimental Results), *Use Proxies Paleocyanogr.*, 489–512,  
520 [https://doi.org/10.1007/978-3-642-58646-0\\_20](https://doi.org/10.1007/978-3-642-58646-0_20), 1999.
- 521 Bijma, J., Hönisch, B., and Zeebe, R. E.: Impact of the ocean carbonate chemistry on living foraminiferal shell  
522 weight: Comment on “Carbonate ion concentration in glacial-age deep waters of the Caribbean Sea” by W. S.  
523 Broecker and E. Clark, *Geochemistry, Geophys. Geosystems*, 3, 1–7, <https://doi.org/10.1029/2002GC000388>,  
524 2002.
- 525 Broecker, W. and Clark, E.: An evaluation of Lohmann’s foraminifera weight dissolution index,  
526 *Paleoceanography*, 16, 531–534, <https://doi.org/10.1029/2000PA000600>, 2001a.
- 527 Broecker, W. and Clark, E.: Reevaluation of the CaCO<sub>3</sub> size index paleocarbonate ion proxy, *Paleoceanography*,  
528 16, 669–671, <https://doi.org/10.1029/2001PA000660>, 2001b.
- 529 Broecker, W. and Clark, E.: Ratio of coccolith CaCO<sub>3</sub> to foraminifera CaCO<sub>3</sub> in late Holocene deep sea  
530 sediments, *Paleoceanography*, 24, <https://doi.org/10.1029/2009PA001731>, 2009.
- 531 Brownlee, C. and Taylor, A.: Calcification in coccolithophores: A cellular perspective, in: *Coccolithophores*,  
532 Springer, Berlin, 31–49, [https://doi.org/10.1007/978-3-662-06278-4\\_2](https://doi.org/10.1007/978-3-662-06278-4_2), 2004.
- 533 Buitenhuis, E. T., Le Quéré, C., Bednaršek, N., and Schiebel, R.: Large Contribution of Pteropods to Shallow  
534 CaCO<sub>3</sub> Export, *Global Biogeochem. Cycles*, 33, 458–468, <https://doi.org/10.1029/2018GB006110>, 2019.
- 535 Bürkner, P. C.: brms: An R Package for Bayesian Multilevel Models Using Stan, *J. Stat. Softw.*, 80, 1–28,  
536 <https://doi.org/10.18637/JSS.V080.I01>, 2017.
- 537 Bürkner, P. C.: Advanced Bayesian multilevel modeling with the R package brms, *R J.*, 10, 395–411,  
538 <https://doi.org/10.32614/RJ-2018-017>, 2018.
- 539 Carpenter, B., Gelman, A., Hoffman, M. D., Lee, D., Goodrich, B., Betancourt, M., Brubaker, M. A., Guo, J., Li, P.,  
540 and Riddell, A.: Stan: A Probabilistic Programming Language, *J. Stat. Softw.*, 76,  
541 <https://doi.org/10.18637/JSS.V076.I01>, 2017.
- 542 Danabasoglu, G., Lamarque, J. F., Bacmeister, J., Bailey, D. A., DuVivier, A. K., Edwards, J., Emmons, L. K.,  
543 Fasullo, J., Garcia, R., Gettelman, A., Hannay, C., Holland, M. M., Large, W. G., Lauritzen, P. H., Lawrence, D. M.,  
544 Lenaerts, J. T. M., Lindsay, K., Lipscomb, W. H., Mills, M. J., Neale, R., Oleson, K. W., Otto-Bliessner, B., Phillips,  
545 A. S., Sacks, W., Tilmes, S., van Kampenhout, L., Vertenstein, M., Bertini, A., Dennis, J., Deser, C., Fischer, C.,  
546 Fox-Kemper, B., Kay, J. E., Kinnison, D., Kushner, P. J., Larson, V. E., Long, M. C., Mickelson, S., Moore, J. K.,  
547 Nienhouse, E., Polvani, L., Rasch, P. J., and Strand, W. G.: The Community Earth System Model Version 2



- 548 (CESM2), *J. Adv. Model. Earth Syst.*, 12, <https://doi.org/10.1029/2019MS001916>, 2020.
- 549 Darling, K. F., Wade, C. M., Stewart, I. A., Kroon, D., Dingle, R., and Leigh Brown, A. J.: Molecular evidence for  
550 genetic mixing of Arctic and Antarctic subpolar populations of planktonic foraminifers, *Nature*, 405, 43–47,  
551 <https://doi.org/10.1038/35011002>, 2000.
- 552 Davis, C. V., Badger, M. P. S., Bown, P. R., and Schmidt, D. N.: The response of calcifying plankton to climate  
553 change in the Pliocene, *Biogeosciences*, 10, 6131–6139, <https://doi.org/10.5194/bg-10-6131-2013>, 2013.
- 554 Davis, C. V., Rivest, E. B., Hill, T. M., Gaylord, B., Russell, A. D., and Sanford, E.: Ocean acidification  
555 compromises a planktic calcifier with implications for global carbon cycling, *Sci. Rep.*, 7, 1–8,  
556 <https://doi.org/10.1038/s41598-017-01530-9>, 2017.
- 557 Demes, K. W., Bell, S. S., and Dawes, C. J.: The effects of phosphate on the biomineralization of the green alga,  
558 *Halimeda incrassata* (Ellis) Lam., *J. Exp. Mar. Bio. Ecol.*, 374, 123–127,  
559 <https://doi.org/10.1016/J.JEMBE.2009.04.013>, 2009.
- 560 Dong, S., Lei, Y., Bi, H., Xu, K., Li, T., and Jian, Z.: Biological Response of Planktic Foraminifera to Decline in  
561 Seawater pH, *Biology (Basel)*, 11, <https://doi.org/10.3390/BIOLOGY11010098/S1>, 2022.
- 562 Dormann, C. F., Elith, J., Bacher, S., Buchmann, C., Carl, G., Carré, G., Marquéz, J. R. G., Gruber, B., Lafourcade,  
563 B., Leitão, P. J., Münkemüller, T., McClean, C., Osborne, P. E., Reineking, B., Schröder, B., Skidmore, A. K., Zurell,  
564 D., and Lautenbach, S.: Collinearity: a review of methods to deal with it and a simulation study evaluating their  
565 performance, *Ecography (Cop.)*, 36, 27–46, <https://doi.org/10.1111/J.1600-0587.2012.07348.X>, 2013.
- 566 Erez, J.: The Source of Ions for Biomineralization in Foraminifera and Their Implications for Paleoceanographic  
567 Proxies, *Rev. Mineral. Geochemistry*, 54, 115–149, <https://doi.org/10.2113/0540115>, 2003.
- 568 Etheridge, D. M., Steele, L. P., Langenfelds, R. L., Francey, R. J., Barnola, J. M., and Morgan, V. I.: Natural and  
569 anthropogenic changes in atmospheric CO<sub>2</sub> over the last 1000 years from air in Antarctic ice and firn, *J.*  
570 *Geophys. Res. Atmos.*, 101, 4115–4128, <https://doi.org/10.1029/95JD03410>, 1996.
- 571 Gaskell, D. E., Ohman, M. D., and Hull, P. M.: Zooglider-Based Measurements of Planktonic Foraminifera in the  
572 California Current System, *J. Foraminifer. Res.*, 49, 390–404, <https://doi.org/10.2113/GSJFR.49.4.390>, 2019.
- 573 Gelman, A., Goodrich, B., Gabry, J., and Vehtari, A.: R-squared for Bayesian Regression Models, *Am. Stat.*, 73,  
574 307–309, <https://doi.org/10.1080/00031305.2018.1549100>, 2019.
- 575 Gerecht, A. C., Šupraha, L., Langer, G., and Henderiks, J.: Phosphorus limitation and heat stress decrease  
576 calcification in *Emiliania huxleyi*, *Biogeosciences*, 15, 833–845, <https://doi.org/10.5194/BG-15-833-2018>, 2018.
- 577 Gonzalez-Mora, B., Sierro, F. J., and Flores, J. A.: Controls of shell calcification in planktonic foraminifers, *Quat.*  
578 *Sci. Rev.*, 27, 956–961, <https://doi.org/10.1016/j.quascirev.2008.01.008>, 2008.
- 579 Harrison, X. A., Donaldson, L., Correa-Cano, M. E., Evans, J., Fisher, D. N., Goodwin, C. E. D., Robinson, B. S.,



- 580 Hodgson, D. J., and Inger, R.: A brief introduction to mixed effects modelling and multi-model inference in  
581 ecology, *PeerJ*, 2018, <https://doi.org/10.7717/PEERJ.4794/FIG-3>, 2018.
- 582 Hathorne, E. C., Gagnon, A., Felis, T., Adkins, J., Asami, R., Boer, W., Caillon, N., Case, D., Cobb, K. M., Douville,  
583 E., Demenocal, P., Eisenhauer, A., Garbe-Schönberg, D., Geibert, W., Goldstein, S., Huguen, K., Inoue, M.,  
584 Kawahata, H., Kölling, M., Cornec, F. L., Linsley, B. K., McGregor, H. V., Montagna, P., Nurhati, I. S., Quinn, T.  
585 M., Raddatz, J., Rebaubier, H., Robinson, L., Sadekov, A., Sherrell, R., Sinclair, D., Tudhope, A. W., Wei, G.,  
586 Wong, H., Wu, H. C., and You, C. F.: Interlaboratory study for coral Sr/Ca and other element/Ca ratio  
587 measurements, *Geochemistry, Geophys. Geosystems*, 14, 3730–3750, <https://doi.org/10.1002/GGGE.20230>,  
588 2013.
- 589 Henehan, M. J., Evans, D., Shankle, M., Burke, J. E., Foster, G. L., Anagnostou, E., Chalk, T. B., Stewart, J. A., Alt,  
590 C. H. S., Durrant, J., and Hull, P. M.: Size-dependent response of foraminiferal calcification to seawater  
591 carbonate chemistry, *Biogeosciences*, 14, 3287–3308, <https://doi.org/10.5194/bg-14-3287-2017>, 2017.
- 592 van Heuven, S., Pierrot, D., Rae, J. W. B., Lewis, E., and Wallace, D. W. R.: MATLAB Program Developed for CO2  
593 System Calculations, [https://doi.org/https://doi.org/10.3334/CDIAC/otg.CO2SYS\\_MATLAB\\_v1.1](https://doi.org/https://doi.org/10.3334/CDIAC/otg.CO2SYS_MATLAB_v1.1), 2011.
- 594 Hoffman, M. D. and Gelman, A.: The No-U-Turn Sampler: Adaptively Setting Path Lengths in Hamiltonian  
595 Monte Carlo, *J. Mach. Learn. Res.*, 15, 1593–1623, 2014.
- 596 IOC, SCOR, and IAPSO: The International Thermodynamic Equation of Seawater - 2010: Calculation and use of  
597 thermodynamic properties, ntergovernmental Oceanogr. Comm. Manuals Guid. No. 56. UNESCO.196., 2010.
- 598 IPCC: Summary for Policymakers. In: *Climate Change 2021: The Physical Science Basis. Contribution of Working*  
599 *Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*, edited by: Masson-  
600 Delmotte, V., Zhai, P., Pirani, A., Connors, S. L., Péan, C., Berger, S., Caud, N., Chen, Y., Goldfarb, L., Gomis, M.  
601 I., Huang, M., Leitzell, K., Lonnoy, E., Matthews, J. B. R., Maycock, T. K., Waterfield, T., Yelekçi, O., Yu, R., and  
602 Zhou, B., Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA,  
603 <https://doi.org/10.1017/9781009157896>, 2021.
- 604 IPCC: *Climate Change 2022: Impacts, Adaptation, and Vulnerability. Contribution of Working Group II to the*  
605 *Sixth Assessment Report of the Intergovernmental Panel on Climate Change*, edited by: Pörtner, H. O., Roberts,  
606 D. C., Tignor, M., Poloczanska, E. S., Mintenbeck, K., Alegría, A., Craig, M., Langsdorf, S., Lösschke, S., Möller, V.,  
607 Okem, A., and Rama, B., Cambridge, UK and New York, NY, USA, <https://doi.org/10.1017/9781009325844>,  
608 2022.
- 609 Jiang, L. Q., Dunne, J., Carter, B. R., Tjiputra, J. F., Terhaar, J., Sharp, J. D., Olsen, A., Alin, S., Bakker, D. C. E.,  
610 Feely, R. A., Gattuso, J. P., Hogan, P., Ilyina, T., Lange, N., Lauvset, S. K., Lewis, E. R., Lovato, T., Palmieri, J.,  
611 Santana-Falcón, Y., Schwinger, J., Séférian, R., Strand, G., Swart, N., Tanhua, T., Tsujino, H., Wanninkhof, R.,  
612 Watanabe, M., Yamamoto, A., and Ziehn, T.: Global Surface Ocean Acidification Indicators From 1750 to 2100,  
613 *J. Adv. Model. Earth Syst.*, 15, <https://doi.org/10.1029/2022MS003563>, 2023.



- 614 Jørgensen, B. B., Erez, J., Revsbech, P., and Cohen, Y.: Symbiotic photosynthesis in a planktonic foraminiferan,  
615 Globigerinoides sacculifer (Brady), studied with microelectrodes<sup>1</sup>, *Limnol. Oceanogr.*, 30, 1253–1267,  
616 <https://doi.org/10.4319/LO.1985.30.6.1253>, 1985.
- 617 Kinsey, D. W. and Davies, P. J.: Effects of elevated nitrogen and phosphorus on coral reef growth, *Limnol.*  
618 *Oceanogr.*, 24, 935–940, <https://doi.org/10.4319/LO.1979.24.5.0935>, 1979.
- 619 Kohfeld, K. E., Fairbanks, R. G., Smith, S. L., and Walsh, I. D.: Neogloboquadrina pachyderma (sinistral coiling)  
620 as paleoceanographic tracers in polar oceans: Evidence from northeast water polynya plankton tows, sediment  
621 traps, and surface sediments, *Paleoceanography*, 11, 679–699, <https://doi.org/10.1029/96PA02617>, 1996.
- 622 Köhler-Rink, S. and Köhl, M.: The chemical microenvironment of the symbiotic planktonic foraminifer *Orbulina*  
623 *universa*, *Mar. Biol. Res.*, 1, 68–78, <https://doi.org/10.1080/17451000510019015>, 2005.
- 624 Kolzenburg, R., Moreira, H., Storey, C., and Ragazzola, F.: Structural integrity and skeletal trace elements in  
625 intertidal coralline algae across the Northeast Atlantic reveal a distinct separation of the leading and the  
626 trailing edge populations, *Mar. Environ. Res.*, 190, <https://doi.org/10.1016/J.MARENRES.2023.106086>, 2023.
- 627 Langer, M. R.: Assessing the contribution of foraminiferan protists to global ocean carbonate production, *J.*  
628 *Eukaryot. Microbiol.*, 55, 163–169, <https://doi.org/10.1111/J.1550-7408.2008.00321.X>, 2008.
- 629 Lastam, J., Griesshaber, E., Yin, X., Rupp, U., Sánchez-Almazo, I., Heß, M., Walther, P., Checa, A., and Schmahl,  
630 W. W.: The unique fibrillar to platy nano- and microstructure of twinned rotaliid foraminiferal shell calcite, *Sci.*  
631 *Reports* 2023 131, 13, 1–19, <https://doi.org/10.1038/s41598-022-25082-9>, 2023.
- 632 Lauvset, S. K., Lange, N., Tanhua, T., Bittig, H. C., Olsen, A., Kozyr, A., Alin, S., Álvarez, M., Azetsu-Scott, K.,  
633 Barbero, L., Becker, S., Brown, P. J., Carter, B. R., Da Cunha, L. C., Feely, R. A., Hoppema, M., Humphreys, M. P.,  
634 Ishii, M., Jeansson, E., Jiang, L. Q., Jones, S. D., Lo Monaco, C., Murata, A., Müller, J. D., Pérez, F. F., Pfeil, B.,  
635 Schirnick, C., Steinfeldt, R., Suzuki, T., Tilbrook, B., Ulfsbo, A., Velo, A., Woosley, R. J., and Key, R. M.:  
636 GLODAPv2.2022: the latest version of the global interior ocean biogeochemical data product, *Earth Syst. Sci.*  
637 *Data*, 14, 5543–5572, <https://doi.org/10.5194/ESDD-14-5543-2022>, 2022.
- 638 LeKieffre, C., Spero, H. J., Russell, A. D., Fehrenbacher, J. S., Geslin, E., and Meibom, A.: Assimilation,  
639 translocation, and utilization of carbon between photosynthetic symbiotic dinoflagellates and their planktic  
640 foraminifera host, *Mar. Biol.*, 165, 1–15, <https://doi.org/10.1007/S00227-018-3362-7/FIGURES/9>, 2018.
- 641 Leung, J. Y. S., Zhang, S., Connell, S. D., Leung, J. Y. S., Zhang, S., and Connell, S. D.: Is Ocean Acidification Really  
642 a Threat to Marine Calcifiers? A Systematic Review and Meta-Analysis of 980+ Studies Spanning Two Decades,  
643 *Small*, 18, <https://doi.org/10.1002/SMLL.202107407>, 2022.
- 644 Lewis, E. and Wallace, D. W. R.: Program Developed for CO<sub>2</sub> System Calculations, 1998.
- 645 Lin, Y. P. and Singer, P. C.: Inhibition of calcite precipitation by orthophosphate: Speciation and thermodynamic  
646 considerations, *Geochim. Cosmochim. Acta*, 70, 2530–2539, <https://doi.org/10.1016/J.GCA.2006.03.002>, 2006.



- 647 Lohmann, G. P.: A model for variation in the chemistry of planktonic foraminifera due to secondary  
648 calcification and selective dissolution, *Paleoceanography*, 10, 445–457, <https://doi.org/10.1029/95PA00059>,  
649 1995.
- 650 Lombard, F., Labeyrie, L., Michel, E., Spero, H. J., and Lea, D. W.: Modelling the temperature dependent growth  
651 rates of planktic foraminifera, *Mar. Micropaleontol.*, 70, 1–7, <https://doi.org/10.1016/j.marmicro.2008.09.004>,  
652 2009.
- 653 Lombard, F., da Rocha, R. E., Bijma, J., and Gattuso, J. P.: Effect of carbonate ion concentration and irradiance  
654 on calcification in planktonic foraminifera, *Biogeosciences*, 7, 247–255, [https://doi.org/10.5194/BG-7-247-](https://doi.org/10.5194/BG-7-247-2010)  
655 2010, 2010.
- 656 MacFarling Meure, C., Etheridge, D., Trudinger, C., Steele, P., Langenfelds, R., Van Ommen, T., Smith, A., and  
657 Elkins, J.: Law Dome CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O ice core records extended to 2000 years BP, *Geophys. Res. Lett.*, 33,  
658 <https://doi.org/10.1029/2006GL026152>, 2006.
- 659 Mallo, M., Ziveri, P., Graham Mortyn, P., Schiebel, R., and Grelaud, M.: Low planktic foraminiferal diversity and  
660 abundance observed in a spring 2013 west-east Mediterranean Sea plankton tow transect, *Biogeosciences*, 14,  
661 2245–2266, <https://doi.org/10.5194/BG-14-2245-2017>, 2017.
- 662 Manno, C., Morata, N., and Bellerby, R.: Effect of ocean acidification and temperature increase on the  
663 planktonic foraminifer *Neogloboquadrina pachyderma* (sinistral), *Polar Biol.*, 35, 1311–1319,  
664 <https://doi.org/10.1007/s00300-012-1174-7>, 2012.
- 665 Marcoulides, K. M. and Raykov, T.: Evaluation of Variance Inflation Factors in Regression Models Using Latent  
666 Variable Modeling Methods, *Educ. Psychol. Meas.*, 79, 874, <https://doi.org/10.1177/0013164418817803>,  
667 2019.
- 668 Marr, J. P., Baker, J. A., Carter, L., Allan, A. S. R., Dunbar, G. B., and Bostock, H. C.: Ecological and temperature  
669 controls on Mg/Ca ratios of *Globigerina bulloides* from the southwest Pacific Ocean, *Paleoceanography*, 26,  
670 <https://doi.org/10.1029/2010PA002059>, 2011.
- 671 Marshall, B. J., Thunell, R. C., Henehan, M. J., Astor, Y., and Wejnert, K. E.: Planktonic foraminiferal area density  
672 as a proxy for carbonate ion concentration: A calibration study using the Cariaco Basin ocean time series,  
673 *Paleoceanography*, 28, 363–376, <https://doi.org/10.1002/palo.20034>, 2013.
- 674 Getting started with TEOS-10 and the Gibbs seawater (GSW) oceanographic toolbox: [https://www.teos-](https://www.teos-10.org/pubs/Getting_Started.pdf)  
675 10.org/pubs/Getting\_Started.pdf.
- 676 Mikis, A., Hendry, K. R., Pike, J., Schmidt, D. N., Edgar, K. M., Peck, V., Peeters, F. J. C., Leng, M. J., Meredith, M.  
677 P., Jones, C. L. C., Stammerjohn, S., and Ducklow, H.: Temporal variability in foraminiferal morphology and  
678 geochemistry at the West Antarctic Peninsula: A sediment trap study, *Biogeosciences*, 16, 3267–3282,  
679 <https://doi.org/10.5194/BG-16-3267-2019>, 2019.





- 680 de Moel, H., Ganssen, G. M., Peeters, F. J. C., Jung, S. J. A., Kroon, D., Brummer, G. J. A., and Zeebe, R. E. :  
681 Planktic foraminiferal shell thinning in the arabian sea due to anthropogenic ocean acidification?,  
682 Biogeosciences, 6, 1917–1925, <https://doi.org/10.5194/bg-6-1917-2009>, 2009.
- 683 Morard, R., Darling, K. F., Weiner, A. K. M., Hassenrück, C., Vanni, C., Cordier, T., Henry, N., Greco, M., Vollmar,  
684 N. M., Milivojevic, T., Rahman, S. N., Siccha, M., Meilland, J., Jonkers, L., Quillévéré, F., Escarguel, G., Douady,  
685 C. J., Garidel-Thoron, T. de, Vargas, C. de, and Kucera, M.: The global genetic diversity of planktonic  
686 foraminifera reveals the structure of cryptic speciation in plankton, *Biol. Rev.*,  
687 <https://doi.org/10.1111/BRV.13065>, 2024.
- 688 Moy, A. D., Howard, W. R., Bray, S. G., and Trull, T. W.: Reduced calcification in modern Southern Ocean  
689 planktonic foraminifera, *Nat. Geosci.*, 2, 276–280, <https://doi.org/10.1038/ngeo460>, 2009.
- 690 Naik, S. S., Naidu, P. D., Govil, P., and Godad, S.: Relationship between weights of planktonic foraminifer shell  
691 and surface water CO<sub>3</sub><sup>=</sup> concentration during the Holocene and Last Glacial Period, *Mar. Geol.*, 275, 278–282,  
692 <https://doi.org/10.1016/j.margeo.2010.05.004>, 2010.
- 693 Naik, S. S., Godad, S. P., and Naidu, D. P.: Does carbonate ion control planktonic foraminifera shell calcification  
694 in upwelling regions?, *Curr. Sci.*, 101, 1370–1375, 2011.
- 695 Nehrke, G., Keul, N., Langer, G., de Nooijer, L. J., Bijma, J., and Meibom, A.: A new model for biomineralization  
696 and trace-element signatures of Foraminifera tests, *Biogeosciences*, 10, 6759–6767,  
697 <https://doi.org/10.5194/bg-10-6759-2013>, 2013.
- 698 Neukermans, G., Bach, L. T., Butterley, A., Sun, Q., Claustre, H., and Fournier, G. R.: Quantitative and  
699 mechanistic understanding of the open ocean carbonate pump - perspectives for remote sensing and  
700 autonomous in situ observation, *Earth-Science Rev.*, 239, 104359,  
701 <https://doi.org/10.1016/J.EARSCIREV.2023.104359>, 2023.
- 702 de Nooijer, L. J., Toyofuku, T., Kitazato, H., and Stanley, S. M.: Foraminifera promote calcification by elevating  
703 their intracellular pH, *PNAS*, 106, 15374–15378, 2009.
- 704 de Nooijer, L. J., Spero, H. J., Erez, J., Bijma, J., and Reichart, G. J.: Biomineralization in perforate foraminifera,  
705 <https://doi.org/10.1016/j.earscirev.2014.03.013>, 1 August 2014.
- 706 Ortiz, J. D., Mix, A. C., and Collier, R. W.: Environmental control of living symbiotic and asymbiotic foraminifera  
707 of the California Current, *Paleoceanography*, 10, 987–1009, <https://doi.org/10.1029/95PA02088>, 1995.
- 708 Osborne, E. B., Thunell, R. C., Marshall, B. J., Holm, J. A., Tappa, E. J., Benitez-Nelson, C., Cai, W. J., and Chen,  
709 B.: Calcification of the planktonic foraminifera *Globigerina bulloides* and carbonate ion concentration: Results  
710 from the Santa Barbara Basin, *Paleoceanography*, 31, 1083–1102, <https://doi.org/10.1002/2016PA002933>,  
711 2016.
- 712 Paasche, E. and Brubak, S.: Enhanced calcification in the coccolithophorid *Emiliania huxleyi* (Haptophyceae)



- 713 under phosphorus limitation, *Phycologia*, 33, 324–330, <https://doi.org/10.2216/I0031-8884-33-5-324.1>, 1994.
- 714 Pak, D. K., Hendy, I. L., Weaver, J. C., Schimmelmann, A., and Clayman, L.: Foraminiferal proxy response to  
715 ocean temperature variability and acidification over the last 150 years in the Santa Barbara Basin (California),  
716 *Quat. Int.*, 469, 141–150, <https://doi.org/10.1016/J.QUAINT.2016.07.049>, 2018.
- 717 Pallacks, S., Ziveri, P., Schiebel, R., Vonhof, H., Rae, J. W. B., Littley, E., Garcia-Orellana, J., Langer, G., Grelaud,  
718 M., and Martrat, B.: Anthropogenic acidification of surface waters drives decreased biogenic calcification in the  
719 Mediterranean Sea, *Commun. Earth Environ.*, 4, 1–10, <https://doi.org/10.1038/s43247-023-00947-7>, 2023.
- 720 Qin, B., Li, T., Xiong, Z., Algeo, T. J., and Jia, Q.: Calcification of planktonic foraminifer *Pulleniatina*  
721 *obliquiloculata* controlled by seawater temperature rather than ocean acidification, *Glob. Planet. Change*, 193,  
722 103256, <https://doi.org/10.1016/J.GLOPLACHA.2020.103256>, 2020.
- 723 R Core Team: R: A language and environment for statistical computing. R Foundation for Statistical Computing.,  
724 <https://www.r-project.org/>, 2018.
- 725 Rillo, M. C., Miller, C. G., Kučera, M., and Ezard, T. H. G.: Intraspecific size variation in planktonic foraminifera  
726 cannot be consistently predicted by the environment, *Ecol. Evol.*, 10, 11579–11590,  
727 <https://doi.org/10.1002/ECE3.6792>, 2020.
- 728 Rink, S., Kühl, M., Bijma, J., and Spero, H. J.: Microsensor studies of photosynthesis and respiration in the  
729 symbiotic foraminifer *Orbulina universa*, *Mar. Biol.*, 131, 583–595, <https://doi.org/10.1007/s002270050350>,  
730 1998.
- 731 Rosenthal, Y., Perron-Cashman, S., Lear, C. H., Bard, E., Barker, S., Billups, K., Bryan, M., Delaney, M. L.,  
732 DeMenocal, P. B., Dwyer, G. S., Elderfield, H., German, C. R., Greaves, M., Lea, D. W., Marchitto, T. M., Pak, D.  
733 K., Paradis, G. L., Russell, A. D., Schneider, R. R., Scheiderich, K., Stott, L., Tachikawa, K., Tappa, E., Thunell, R.,  
734 Wara, M., Weldeab, S., and Wilson, P. A.: Interlaboratory comparison study of Mg/Ca and Sr/Ca measurements  
735 in planktonic foraminifera for paleoceanographic research, *Geochemistry, Geophys. Geosystems*, 5, 4–09,  
736 <https://doi.org/10.1029/2003GC000650>, 2004.
- 737 Russell, A. D., Hönisch, B., Spero, H. J., and Lea, D. W.: Effects of seawater carbonate ion concentration and  
738 temperature on shell U, Mg, and Sr in cultured planktonic foraminifera, *Geochim. Cosmochim. Acta*, 68, 4347–  
739 4361, <https://doi.org/10.1016/j.gca.2004.03.013>, 2004.
- 740 Schiebel, R.: Planktic foraminiferal sedimentation and the marine calcite budget, *Global Biogeochem. Cycles*,  
741 16, <https://doi.org/10.1029/2001gb001459>, 2002.
- 742 Schiebel, R. and Hemleben, C.: *Planktic Foraminifers in the Modern Ocean*, Springer, Berlin, 154 pp.,  
743 [https://doi.org/10.1007/978-3-662-50297-6\\_1](https://doi.org/10.1007/978-3-662-50297-6_1), 2017.
- 744 Schmidt, D. N., Renaud, S., Bollmann, J., Schiebel, R., and Thierstein, H. R.: Size distribution of Holocene  
745 planktic foraminifer assemblages: Biogeography, ecology and adaptation, *Mar. Micropaleontol.*, 50, 319–338,



- 746 [https://doi.org/10.1016/S0377-8398\(03\)00098-7](https://doi.org/10.1016/S0377-8398(03)00098-7), 2004.
- 747 Schmidt, D. N., Elliott, T., and Kasemann, S. A.: The influences of growth rates on planktic foraminifers as  
748 proxies for palaeostudies - A review, *Geol. Soc. Spec. Publ.*, 303, 73–85, <https://doi.org/10.1144/SP303.6>,  
749 2008.
- 750 Song, Q., Qin, B., Tang, Z., Liu, Y., Chen, Z., Guo, J., Xiong, Z., and Li, T.: Calcification of planktonic foraminifer  
751 *Neogloboquadrina pachyderma* (sinistral) controlled by seawater temperature rather than ocean acidification  
752 in the Antarctic Zone of modern Southern Ocean, *Sci. China Earth Sci.*, 65, 1824–1836,  
753 <https://doi.org/10.1007/S11430-021-9924-7/METRICS>, 2022.
- 754 Spero, H. J., Lerche, I., and Williams, D. F.: Opening the carbon isotope “vital effect” black box, 2, Quantitative  
755 model for interpreting foraminiferal carbon isotope data, *Paleoceanography*, 6, 639–655,  
756 <https://doi.org/10.1029/91PA02022>, 1991.
- 757 Spindler, M., Hemleben, C., Salomons, J. B., and Smit, L. P.: Feeding behavior of some planktonic foraminifers  
758 in laboratory cultures, *J. Foraminifer. Res.*, 14, 237–249, <https://doi.org/10.2113/GSJFR.14.4.237>, 1984.
- 759 Takahashi, T., Sutherland, S. C., Chipman, D. W., Goddard, J. G., and Ho, C.: Climatological distributions of pH,  
760 pCO<sub>2</sub>, total CO<sub>2</sub>, alkalinity, and CaCO<sub>3</sub> saturation in the global surface ocean, and temporal changes at  
761 selected locations, *Mar. Chem.*, 164, 95–125, <https://doi.org/10.1016/J.MARCHEM.2014.06.004>, 2014.
- 762 Toyofuku, T., Matsuo, M. Y., De Nooijer, L. J., Nagai, Y., Kawada, S., Fujita, K., Reichert, G. J., Nomaki, H.,  
763 Tsuchiya, M., Sakaguchi, H., and Kitazato, H.: Proton pumping accompanies calcification in foraminifera, *Nat.*  
764 *Commun.*, 8, 1–6, <https://doi.org/10.1038/ncomms14145>, 2017.
- 765 Troupin, C., Barth, A., Sirjacobs, D., Ouberdous, M., Brankart, J. M., Brasseur, P., Rixen, M., Alvera-Azcárate, A.,  
766 Belounis, M., Capet, A., Lenartz, F., Toussaint, M. E., and Beckers, J. M.: Generation of analysis and consistent  
767 error fields using the Data Interpolating Variational Analysis (DIVA), *Ocean Model.*, 52–53, 90–101,  
768 <https://doi.org/10.1016/J.OCEMOD.2012.05.002>, 2012.
- 769 Vehtari, A., Gelman, A., and Gabry, J.: loo: Efficient Leave-One-Out Cross-Validation and WAIC for Bayesian  
770 Models. R Package Version 1.1.0, <https://github.com/stan-dev/loo>, 2017.
- 771 de Villiers, S.: Optimum growth conditions as opposed to calcite saturation as a control on the calcification rate  
772 and shell-weight of marine foraminifera, *Mar. Biol.*, 144, 45–49, <https://doi.org/10.1007/s00227-003-1183-8>,  
773 2004.
- 774 Weinkauff, M. F. G., Moller, T., Koch, M. C., and Kučera, M.: Calcification intensity in planktonic Foraminifera  
775 reflects ambient conditions irrespective of environmental stress, *Biogeosciences*, 10, 6639–6655,  
776 <https://doi.org/10.5194/bg-10-6639-2013>, 2013.
- 777 Weinkauff, M. F. G., Kunze, J. G., Waniek, J. J., and Kučera, M.: Seasonal Variation in Shell Calcification of  
778 Planktonic Foraminifera in the NE Atlantic Reveals Species-Specific Response to Temperature, Productivity, and

<https://doi.org/10.5194/egusphere-2024-2405>

Preprint. Discussion started: 5 August 2024

© Author(s) 2024. CC BY 4.0 License.



779 Optimum Growth Conditions, PLoS One, 11, e0148363, <https://doi.org/10.1371/journal.pone.0148363>, 2016.

780

781



782 [Supplementary reference list](#)

- 783 Barker, S.: Planktonic foraminiferal proxies for temperature and pCO<sub>2</sub>, University of Cambridge, 2002.
- 784 Barker, S. and Elderfield, H.: Foraminiferal calcification response to glacial-interglacial changes in atmospheric  
785 CO<sub>2</sub>, *Science* (80-. ), 297, 833–836, <https://doi.org/10.1126/science.1072815>, 2002.
- 786 Bédard, T. M., Rigual-Hernández, A. S., Flores, J. A., Tarruella, J. P., Durrieu De Madron, X., Cacho, I., Haghpor,  
787 N., Hunter, A., and Sierro, F. J.: Calcification response of planktic foraminifera to environmental change in the  
788 western Mediterranean Sea during the industrial era, *Biogeosciences*, 20, 1505–1528,  
789 <https://doi.org/10.5194/BG-20-1505-2023>, 2023.
- 790 van Heuven, S., Pierrot, D., Rae, J. W. B., Lewis, E., and Wallace, D. W. R.: MATLAB Program Developed for CO<sub>2</sub>  
791 System Calculations, [https://doi.org/https://doi.org/10.3334/CDIAC/otg.CO2SYS\\_MATLAB\\_v1.1](https://doi.org/https://doi.org/10.3334/CDIAC/otg.CO2SYS_MATLAB_v1.1), 2011.
- 792 Jiang, L. Q., Dunne, J., Carter, B. R., Tjiputra, J. F., Terhaar, J., Sharp, J. D., Olsen, A., Alin, S., Bakker, D. C. E.,  
793 Feely, R. A., Gattuso, J. P., Hogan, P., Ilyina, T., Lange, N., Lauvset, S. K., Lewis, E. R., Lovato, T., Palmieri, J.,  
794 Santana-Falcón, Y., Schwinger, J., Séférian, R., Strand, G., Swart, N., Tanhua, T., Tsujino, H., Wanninkhof, R.,  
795 Watanabe, M., Yamamoto, A., and Ziehn, T.: Global Surface Ocean Acidification Indicators From 1750 to 2100, *J.*  
796 *Adv. Model. Earth Syst.*, 15, <https://doi.org/10.1029/2022MS003563>, 2023.
- 797 Marr, J. P., Baker, J. A., Carter, L., Allan, A. S. R., Dunbar, G. B., and Bostock, H. C.: Ecological and temperature  
798 controls on Mg/Ca ratios of *Globigerina bulloides* from the southwest Pacific Ocean, *Paleoceanography*, 26,  
799 <https://doi.org/10.1029/2010PA002059>, 2011.
- 800 Marshall, B. J., Thunell, R. C., Henehan, M. J., Astor, Y., and Wejnert, K. E.: Planktonic foraminiferal area density  
801 as a proxy for carbonate ion concentration: A calibration study using the Cariaco Basin ocean time series,  
802 *Paleoceanography*, 28, 363–376, <https://doi.org/10.1002/palo.20034>, 2013.
- 803 Getting started with TEOS-10 and the Gibbs seawater (GSW) oceanographic toolbox: [https://www.teos-10.org/pubs/Getting\\_Started.pdf](https://www.teos-10.org/pubs/Getting_Started.pdf).  
804
- 805 Osborne, E. B., Thunell, R. C., Marshall, B. J., Holm, J. A., Tappa, E. J., Benitez-Nelson, C., Cai, W. J., and Chen, B.:  
806 Calcification of the planktonic foraminifera *Globigerina bulloides* and carbonate ion concentration: Results from  
807 the Santa Barbara Basin, *Paleoceanography*, 31, 1083–1102, <https://doi.org/10.1002/2016PA002933>, 2016.
- 808 Pallacks, S., Ziveri, P., Schiebel, R., Vonhof, H., Rae, J. W. B., Little, E., Garcia-Orellana, J., Langer, G., Grelaud,  
809 M., and Martrat, B.: Anthropogenic acidification of surface waters drives decreased biogenic calcification in the  
810 Mediterranean Sea, *Commun. Earth Environ.*, 4, 1–10, <https://doi.org/10.1038/s43247-023-00947-7>, 2023.
- 811 Qin, B., Li, T., Xiong, Z., Algeo, T. J., and Chang, F.: Deepwater carbonate ion concentrations in the western  
812 tropical Pacific since 250 ka: Evidence for oceanic carbon storage and global climate influence,  
813 *Paleoceanography*, 32, 351–370, <https://doi.org/10.1002/2016PA003039>, 2017.
- 814 Weinkauff, M. F. G., Kunze, J. G., Wanek, J. J., and Kučera, M.: Seasonal Variation in Shell Calcification of  
815 Planktonic Foraminifera in the NE Atlantic Reveals Species-Specific Response to Temperature, Productivity, and  
816 Optimum Growth Conditions, *PLoS One*, 11, e0148363, <https://doi.org/10.1371/journal.pone.0148363>, 2016.
- 817