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High seasonal and spatial dynamics of bio- and photodegradation in boreal humic waters

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Synopsis:

In boreal (non-permafrost) humic (>15 mg DOC/L) waters of a stratified lake and an ombrotrophic bog, the experimentally measured rate of DOM photodegradation is 4 times higher than that of biodegradation. However, given the shallow (0.5 m) photic layer versus the full depth of water column (2 - 10 m), the biodegradation may provide the largest contribution to aerial CO_2 emission.

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

Abstract

Studying competitive effects of microbial and light-induced degradation of dissolved organic matter (DOM) is crucially important for understanding the factors controlling aquatic carbon (C) transformation in boreal waters. However, studies addressing both DOM and trace element (TE) behavior are limited, which does not allow assessment of coupled C – TE (including macro- and micronutrients and toxicants) biogeochemical cycles in these environmentally important settings. Here we conducted a seasonally-resolved assessment on the degree of DOM and related major and TE transformation under biotic activity and sunlight using conventional incubations of humic surface waters from the European subarctic. We studied bio- and photodegradation over 2 - 3 weeks in an ombrotrophic peatbog continuum (subsurface water from piezometer - small peatland pool - outlet stream) during July, and in three horizons (0.5, 5 and 10 m) of deep (30 m), a stratified forest lake from the same region during June, August and September.

Along the bog water continuum in July, biodegradation rate was the highest in subsurface waters collected via piezometer and the lowest in the acidic peatland pool (0.17 to 0.03 mg C L^{-1} d^{-1}, respectively). Photodegradation was similar for piezometrically collected subsurface waters and the stream (about 0.3 mg C L^{-1} d^{-1}), but was not detectable in the peatland pool. The waters of forest lake exhibited a strong seasonal effect of biodegradation, which was the highest in October and the lowest in June (0.04 and 0.02 mg C L^{-1} d^{-1}, respectively). The photodegradation of DOM from the forest lake was observed only in June and August (0.19 and 0.07 mg C L^{-1} d^{-1}, respectively). Biodegradation was capable of removing between 1 and 7 % of initial DOC, being the highest in the forest lake in October and in peatland pool in summer. The photolysis was capable of degrading a much higher proportion of the initial DOC (10-25 %), especially in the forest lake during June and the bog stream during July. The change of optical parameters confirmed the highest photodegradation occurs in June (Arctic summer) and demonstrates a

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

decrease of chromophoric (aromatic) compounds during incubation, whereas biodegradation acted preferentially on aliphatic, low molecular weight compounds. Only a few trace metals were sizably affected by both photo- and biodegradation of DOM (Fe, Al, Ti, Nb and light REE), whereas V, Mn, Co, Cu and Ba were affected solely by biodegradation. Typical values of TE removal over a 2-week period of incubation ranged from 1 to 10 %. These effects were mostly pronounced in the less acidic forest lake compared to the bog waters. A likely mechanism of TE removal was their coprecipitation with coagulating Fe(III) hydroxides.

When averaged across sites and seasons, DOM biodegradation and photodegradation processes could remove 5.3 and 10.8 mg C L^{-1} y^{-1}, respectively. Compared to typical CO_2 emissions from inland waters of the region, biodegradation of DOM can provide the totality of C-CO_2 evasion from lake water surfaces whereas bio- and photodegradation are not sufficient to explain the observed fluxes in bog water continuum. Overall, these results demonstrated strong spatial and seasonal variability in DOM and TE complexes bio- and photodegradation, which was poorly accessed until now, and call for the need of a systematic assessment of both processes across seasons with high spatial resolution.

1. Introduction

Organic Carbon (OC) processing via metabolic biological (heterotrophic bacteria uptake and respiration) and inorganic physico-chemical (photolysis) pathways is considered to be one of the major source of CO_2 supersaturation in surface waters and related C emissions (Lapierre et al., 2013; Tranvik et al., 2009), although the relative role of dissolved vs particulate organic carbon (POC) remains poorly quantified (e.g. Attermeyer et al., 2018; Lau et al., 2021; Shirokova et al., 2021; Raudina et al., 2022). Given sizable C emissions in boreal and subarctic waters (Karlsson et al., 2021), together with high concentrations of DOC (Cole et al., 2007; Vonk et al., 2015), and fast ongoing and predicted environmental changes in high latitude aquatic and

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

terrestrial ecosystems (Wauthy et al., 2018; Chaudhary et al., 2020; Harris et al., 2022), the surface waters of subarctic regions are at the forefront of studies on the biogeochemical cycle of C. Although emissions from these waters are significantly lower than those in the 10 °S – 10 °N equatorial belt (e.g., Borges et al., 2015), the magnitude of possible changes in C flux from northern waters to the atmosphere remains much less known. Further, there are still important geographical biases linked to insufficient knowledge of rates and mechanisms of DOC transformation in certain regions. An example is peatland dominant northern aquatic settings, where high concentrations of soil organic C surrounding the bogs provide elevated concentrations of DOC. These soils and their organic C content become highly vulnerable to biological and physico-chemical impact depending on local environmental context, permafrost presence and season (Vonk et al., 2015).

Thorough laboratory and field work on DOM bio- and photolability conducted over the past decades have demonstrated both phenomena are important, and, depending on environmental setting (nutrient regime, photic layer depth, nature of DOM, etc.), one or another may dominate overall DOM removal in surface waters (Vachon et al., 2016, 2017; Vähätalo and Wetzel, 2008; Obernosterer and Benner, 2004). Recently, specific attention was devoted to the aquatic systems of permafrost peatlands given their high vulnerability to climate warming and huge potential for release of soil organic C to surfaces waters (Vonk et al., 2015; Shirokova et al., 2019; Payandi-Rolland et al., 2020; Prijac et al., 2022; Rosset et al., 2022; Taillardet et al., 2022). These former studies provided a range of DOM susceptibility to biotic degradation. Thus, between 10 and 40 % of the DOC in lakes, rivers and soil waters of the boreal zone may be available for bacterial uptake over a time frame of several weeks (Berggren et al., 2010; Roehm et al., 2009). This range is consistent with 14-16% of biodegradable DOC (BDOC) assessed globally (Begum et al. 2022). The necessity for further studies was also indicated, most notably with regard to *i*) seasonal aspects, given that the overwhelming majority of available studies were

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

performed during Arctic summer (see discussions in Vonk et al., 2015; Laurion et al., 2021), and *ii*) increased spatial resolution, given that sizable variations of BDOC can be observed within quite short distances of a hydrological continuum (Payandi-Rolland et al., 2020; Raudina et al., 2022). Another poorly studied aspect is DOM photo- and biolability across the depth of the water column, especially in seasonally stratified lakes which are subject to spring and autumn overturn.

Based on a compilation of available studies on BDOC and their own research, Vonk et al. (2015) argued there is a negligible amount of biodegradable DOC in aquatic systems without permafrost. This is, however, contradictory to available assessments on biodegradation of aquatic DOM as major driver of CO_2 emission in general (Amaral et al., 2021; Liu and Wang, 2022) and in boreal waters in particular (Ask et al., 2012; Lapierre et al., 2013). Furthermore, among all Arctic rivers, the highest annual (20%) and winter (ca. 45%) biodegradable DOC (BDOC) was reported for the Ob River, which drains through peatlands with minimal permafrost influence (Wickland et al., 2012). These non-exhaustive examples illustrate certain inconsistency in current estimations of DOC biodegradability in surface organic-rich waters of high latitudes, which precludes quantitative modeling of future C fluxes between land, water and atmosphere in these environmentally important regions. Towards addressing these inconsistences, in this study, we chose a typical hydrological continuum in a boreal ombrotrophic bog in a glacial lake-ridge complex that includes subsurface water, a small peatland pool in the central part of the bog and an outlet stream. Further, we selected a well-studied deep stratified humic lake in the same region (Lake Temnoe; Chupakov et al., 2017) where we sampled surface and deep horizons for the incubation experiments. The chosen waters represent subarctic non-permafrost regions that exhibit sizable organic C pool in their soils and high concentrations of DOC in their surface waters. In contrast to previous studies of permafrost peatlands (Shirokova et al., 2019; Laurion et al., 2021; Payandi-Rolland et al., 2020; Mazoyer et al., 2022) where the main source of DOM is peat or ground vegetation like mosses and lichens, in this highly productive southern taiga

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

region, DOC may be more vulnerable to microbial activity due to the presence of forest leachates (i.e., Don and Kalbitz, 2005; Kalbitz et al., 2003; Kawahigashi et al., 2004; Kiikkilä et al., 2013) and much higher bioproductivity for both the terrestrial and aquatic parts of the lake-river ecosystems.

The working hypothesis behind our study design is that the DOC-rich subsurface water and deep horizons of the humic lake are mostly sensitive to sunlight impact (Stubbins et al., 2010), and that maximal impact of photodegradation is expected during allochthonous aromatic DOM input (high surface inflow to lakes and bogs in June and October). In contrast, maximal biodegradation of DOM is expected during periods of possible phytoplankton bloom in August, when autochthonous organic material is generated in the water column. Trace metals (TM) are supposed to either be taken up by biotic processing of DOM (e.g., micronutrients), or follow the transformation of colloidal organically-bound Fe/Al into particulate Fe, Al hydroxides (e.g., Kopacek et al., 2005, 2006), capable of scavenging TM. To test these hypotheses, we examined DOM and related trace metals bio- and photodegradability aiming to assess 1) spatial variations along a hydrological continuum of non-permafrost peatland and different horizons of a neighboring deep stratified lake located in the forest, and 2) temporal variability during 3 main hydrological seasons (high flow in June, baseflow in August and autumn rain season in October) in the forest lake. Achieving these objectives should allow quantifying the relative share of bio- and photodegradation on overall DOC and TM removal from surface waters via biotic and physico-chemical mechanisms.

2. Materials and Methods

2.1. Natural settings of subarctic bog and stratified lake

The study site is in the NE part of the European boreal zone (Arkhangelsk region), **Fig. 1**. The mean annual air temperature is 0 °C and average annual precipitation is 700 ± 50 mm.

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

The pristine ombrotrophic Ilasskoe Bog is located 30 km SE of Arkhangelsk, and is a typical lake-ridge complex formed from the last glaciation approximately 10,000 years ago. Its total surface area is 89 km², with an average peat thickness of 3 m. The hydrological continuum of the Ilasskoe Bog includes subsurface water collected via piezometer (2-2.5 m depth), a small lake (Severnoe) and a stream outlet (**Fig. 1**). Lake Severnoe, located in the central part of the bog, is a typical peatland pool with an average depth of 1.5 m and a surface area of 0.013 km². The Chernyi Stream is an outlet for the eastern part of the bog. The stream is 0.7-2.0 m wide, 10 km long and it flows in a forested (taiga) zone in the shade of tree canopy. The waters of the Ilasskoe Bog are acidic (pH ranges from 3.9-4.0 in piezometer and peatland pool to 5.7 in stream Chernyi), organic-rich (DOC is equal to 88, 13 and 38 mg L^{-1} in the piezometer, lake and stream, accordingly) and lesser mineralized (Electrical Conductivity is 17-46 µS cm^{-1}), as listed in **Table 1**.

Lake Temnoe is located in a pristine forest 100 km NNE of the town of Arkhangelsk, an area that does not receive any direct anthropogenic impact (**Fig. 1**). The watershed area is 3.08 km² and the lake surface area is 0.091 km², with a maximum depth is 37 m and a Secchi disk depth of 3.5±0.5 m. The water residence time in the lake is 394 days. Bogs constitute 31% of lake's watershed area, which is represented by carbonate-free loamy moraine atop the peat, podzol and gley soils. The lake water is slightly acidic (pH = 5.1 to 6.0) and humic (DOC = 13-20 mg L^{-1}) and dominated by allochthonous DOM with a low concentration of total dissolved ions (Electrical Conductivity of 20 µS cm^{-1}). Similar to other deep boreal and subarctic lakes, the lake exhibits 2 main periods of pronounced stratification (November to April and June to September) and two periods of lake overturn (October and May). Maximal winter stratification occurs in March; the highest water temperature typically occurs in July (see Chupakov et al., 2017 for details).

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

The surface waters were collected from the shore (peatland pool and stream) or a PVC boat (Lake Temnoe). Surface (30-50 cm depth) waters were sampled in the Ilasskoe bog and 3 water horizons (0.5, 5 and 10 m) were sampled in the Temnoe Lake using a pre-cleaned polycarbonate horizontal water sampler (Aquatic Research Co, ID, USA). The water samples were placed into 2-L Milli-Q pre-cleaned PVC jars and kept refrigerated until arrival at the laboratory within 2-3 hours of collection.

2.2. Experiments

2.2.1. Biodegradation

For biodegradation assessments we followed the recommended protocol and used the appropriate type of labware for assessing biodegradable DOC of Arctic waters without external nutrient addition (Vonk et al., 2015; Payandi-Rolland et al., 2020) and applied a slight modification from Shirokova et al. (2019) to assess maximal possible biodegradation. Initial water samples brought to the laboratory within 2-3 hours after sampling were filtered through 3 µm sterilized Nylon Sartorius membranes (47 mm diameter); these were used as 'conventional' 0.7 µm (GF/F) filtration membranes might remove too many microbial cells (Dean et al., 2018).

Duplicate 30 mL aliquots of 3 µm-filtered water were placed into pre-combusted (4.5 hours at 450°C) dark borosilicate 40 mL glass bottles wrapped in Al foil to prevent any photolysis, without nutrient amendment and incubated at 22±1°C in the dark. The bottles were closed with loosened sterilized PVC caps. The bottles were shaken manually once a day avoiding the liquid touching the cap. The entire reactor was used for sampling after 0, 2, 5, 8, 12, and 21 days of exposure. Sampled solutions were filtered through sterile, MilliQ-cleaned Sartorius 0.22 µm filters. The DOC blanks for these filters did not exceed 1% of DOC concentrations in experimental samples. Sterilized control reactors were filled with natural water that was filtered

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

through a 0.22 µm sterile filter and incubated together with experimental reactors following the approach of Köhler et al (2002).

All handling and sampling of bottles was performed in the laminar hood box in a sterilized workspace. Filtered samples were acidified with 30 µL of concentrated (8.1 M) double distilled HCl, tightly capped and stored in the refrigerator before DOC analyses. The non-acidified portion of filtrate was used for pH, Specific Conductivity, DIC and $UV_{254\ nm}$ and optical spectra measurement. Control runs were 0.22 µm sterile-filtered water which was incubated in parallel with experiments and re-filtered through 0.22 µm filters the day of sampling. To ensure minimized release from sterilized Nylon membrane, we ran blank (Milli-Q) filtrations through both 0.7 µm GF/F and 0.22 µm Nylon filters; in both cases the DOC blank was below 0.1-0.2 mg/L which is less than 1% of DOC concentration in our samples. The glass bottles were incubated in duplicates at 22±1°C and agitated manually at least once a day over the 16 days of exposure.

2.2.2. Photodegradation

For photodegradation incubations, water samples were collected in Al-foil covered pre-cleaned polypropylene jars and sterile filtered (0.22 µm Nalgene Rapid-Flow Sterile Systems) within 2 hours of sampling and refrigerated. The filtrates were transferred under laminar hood box into sterilized, acid-washed quartz tubes (150 mL volume, 20% air headspace) with silicate stoppers and placed at 3 ± 2 cm depth into an outdoor pool which was filled by river water having the light transparency similar to that of the Ilasskoe and Temnoe lakes. In-situ measurements of sunlight intensity were conducted using a submersible sunlight sensor. The outdoor pools were placed in an unshaded area with a latitude similar to the sampling sites (< 30 km from Ilasskoe Bog and Temnoe Lake). Slight wind movement and regular manual shaking allowed for sufficient mixing of reactor interiors during exposure. All photodegradation experiments were

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

run in duplicates. The water temperature (EBRO EBI 20) and light intensity (Luxmeter Testo 545) were continuously recorded every 3 hours.

For photodegradation experiments, we followed conventional methods requiring exposure of 0.2 µm-sterile filtered samples in quartz reactors in the outdoor pool (Vähätalo et al., 2003; Chupakova et al., 2018; Gareis and Lesack, 2018), solar simulator (Lou and Xie, 2006; Amado et al., 2014) or directly in the lake water (Laurion and Mladenov, 2013; Groeneveld et al., 2016). Note that the 0.22 µm sterile filtration is the only way of conducting photodegradation experiments, given that autoclave sterilization of DOM-rich natural waters would coagulate humic material and thereby would not be suitable (Andersson et al., 2018). Filtration through a smaller pore size, however, would decrease the concentration of DOC and trace metals (i.e., Ilina et al., 2014; Vasyukova et al., 2010). We have chosen a 16 day exposure time for consistency with biodegradation experiments described above and following the previous studies on photodegradation under sunlight, which typically ranges from 15 to 70 days (Moran et al., 2000; Vähätalo and Wetzel, 2004; Mostofa et al., 2007; Chupakova et al., 2018). Dark control experiments were conducted also in duplicates, using sterilized glass tubes filled with sterile 0.22 µm-filtered water, wrapped in Al foil and placed in the same outdoor pool as the experiments. The headspace (approx. 20% of total reaction volume) was similar in experimental and control reactors. The individual reactors were sterile sampled at the beginning and after the 0, 2, 5, 8, 12, and 16 days of exposure. Each sampling sacrificed the entire reactor. The Milli-Q blanks were collected and processed to monitor for any potential sample contamination introduced by our filtration, incubation, handling and sampling procedures. The organic carbon blanks of the filtrates did not exceed 0.2 mg/L.

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

2.3. Analyses

The temperature, pH, O_2 and specific conductivity in surface waters were measured in field. The dissolved CO_2 concentration in the studied bodies of water was measured in-situ using submersible Vaissala Carbocap® GM70 handheld carbon dioxide meter with GMP222 probes (accuracy 1.5%; see Serikova et al. (2018, 2019) for methodological details). The diffusional CO_2 flux was calculated using a wind-based model (Cole and Caraco, 1998) with $k_{600} = 2.07 + 0.215 \times u_{10}^{1.7}$, where u_{10} is the wind speed at 10 m height, following the approaches developed for surface waters of peatlands (Zabelina et al., 2021).

The DOC and DIC were analyzed by high-temperature catalytic oxidation using a Shimadzu® TOC-VCSN (uncertainty ± 2%, 0.1 mg L^{-1} detection limit). DIC was measured after sample acidification with HCl and DOC was analyzed in acidified samples after sparging it with C-free air for 3 min at 100 mL min^{-1} as non-purgable organic carbon (NPOC). Internationally certified water samples (MISSISSIPPI-03 and Pérade-20) were used to check validity and reproducibility of the analysis. Filtered sampled collected from photodegradation experiments were acidified with ultrapure nitric acid and analyzed for major and TE following the procedures employed by GET (Toulouse) for analyses of boreal humic waters (Oleinikova et al., 2017, 2018).

The UV- and visual absorbance of water samples was measured using a 10 mm quartz cuvette on a CARY-50 UV-vis spectrophotometer to assess the aromaticity of pore fluids via specific UV absorbance ($SUVA_{254}$). In the filtrates, we measured optical density at 254 nm and at select wavelengths (365, 436, 470, and 665 nm) as well as the entire UV-visible spectrum. The specific UV-absorbency ($SUVA_{254}$, L mg^{-1} m^{-1}) and E_{470}:E_{665} ratios are used as a proxy for degree of condensation of aromatic groups of DOM, or humification (Chin et al., 1994; Weishaar et al., 2003; Hur et al., 2006; Peacock et al., 2013). The ratio E_{254}:E_{436} is useful for evaluation of contributions of autochtonous (aquatic) DOM compared to terrestrial (soil) C (Hur et al., 2006;

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

Ilina et al., 2014). The ratio E_{254}:E_{365} also allows approximating the mean molecular weight of DOM (Hiriart-Baer et al., 2008; Berggren et al., 2007). For better visualization of the differences in spectral parameters between experimental and control reactors, we calculated the difference (ΔA) between the absorbance of the photo- or bio-reactor and that of the control reactor at each sampling time.

Major cations, Si, P and ~40 TE were measured with a quadrupole ICP-MS (Agilent 7500 ce) using In and Re as internal standards. The international geo-standard SLRS-5 (Riverine Water Reference Material for Trace Metals) was used to check validity and reproducibility of analyses. Note that for both bio- and photodegradation experiments, ICP MS analyses were performed over 16 days of incubation time.

To check for possible microbial development in biodegradation experiments, we performed oligotrophic and eutrophic bacteria counts over the course of incubation, following the standard methodology used in biodegradation experiments of peat waters (Stutter et al., 2013) and also described previously (Shirokova et al., 2017b; Chupakova et al., 2018).

2.4. Data treatment

The bio- and photodegradable DOC and trace metals were calculated as percent loss relative to control in similar fashion with other studies (Vonk et al., 2015; Chupakova et al., 2018; Shirokova et al., 2017b, 2019). However, previous works in similar environmental contexts of high-DOC humic waters demonstrated that the effects of DOC and element decrease are rather low and often comparable to uncertainties of duplicates (Shirokova et al., 2019). To assess the net effect of bio- or photodestruction during the experiment, we used the integral values of concentration change, estimated as the difference between the experiment and the control, while taking into account the standard deviation of replicates. For this, we first calculated the mean of replicates at the *i*-th time of sampling for the experiment and the control of *X* component ($^{mean}X_i$

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

and ${}^{control}X_i$, respectively). We next calculated the sum of mean concentration of replicates and its standard deviation (${}^{mean}X_i+SD_i$). Thus, we obtained 3 values characterizing the bio- or photo-degradation process: 1) the change of concentration in the experimental reactor (${}^{mean}S$), 2) the change of concentration not linked to the studied process (${}^{control}S$), and 3) the maximal uncertainty of the concentration change in the reactor (${}^{mean+SD}S$). This allowed calculating, in percentages, the efficiency of bio or photodegradation of *X* component relative to the control, taken into account relevant uncertainties as following:

$$X\ (\%) = 100 \times (|{}^{mean}X| - |{}^{control}X|)/|{}^{control}X| \quad (1)$$

$$SD\ (\%) = 100 \times (|{}^{mean+SD}X| - |{}^{mean}X|)/|{}^{control}X| \quad (2)$$

where *X* is biodegradable DOC or trace element (BDOC and BTE, respectively) or photodegradable DOC and trace element (PDOC and PTE, respectively). The sign of *X* designates either a decrease («–») or an increase (« + ») of solute concentration during the experiment. We considered the decrease of concentration significant when *X* (%) > SD (%). In other cases, the change was non-systematic over the course of experiment or non-measurable using the experimental technique employed in the present study.

The mean rate of bio- or photodegradation of *X* component (V_X) was calculated based on the overall change (ΔX, in %) between the initial (X_0) and final value normalized to overall duration of the experiment *t* (22 and 16 days for bio- and photodegradation, respectively):

$$V_X = ((\Delta X / X_0) / t \quad (3)$$

The SD for rates of component change were calculated in a similar way.

The spectral differences between experimental and control reactors were presented as X-Y-Z diagrams where X is elapsed time, Y is wavelength, and Z is ΔA. The data were plotted in a Surfer software package using triangulation with a linear interpolation method. Statistical treatment included the least squares method and the Pearson correlation, as the data were normally distributed. The ANOVA method was used to test the differences in the average DOC

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

and metal concentration versus time in incubation experiments and in the controls and to assess the difference between the light experiments and the dark control for photodegradation experiments. All calculations were performed in STATISTICA ver. 10 (StatSoft Inc.,Tulsa) at p = 0.05).

3. Results

3.1. Field measured C concentration and calculated CO_2 fluxes

The DOC concentration ranged from 13 to 21 mg L^{-1} in Lake Temnoe, depending on depth and season. The CO_2 concentrations and fluxes increased from June to October and varied from 99 to 220 µmol L^{-1} and 32 to 71 mmol CO_2 m^{-2} d^{-1}, respectively (**Table 1**). In Ilasskoe Bog hydrological continuum, the DOC decreased from 88 mg L^{-1} in the peat soil water to 38 mg L^{-1} in the outlet stream. The DOC concentration was generally similar (within ± 5 %) between 3, 0.8 (GFF), 0.45 and 0.22 µm pore size filtration of the initial sample, which is in agreement with former size fractionation measurements for Arctic and subarctic systems (Vasyukova et al., 2010; Pokrovsky et al., 2012, 2016, Shirokova et al., 2019). The waters of Ilasskoe Bog continuum exhibited CO_2 supersaturation with respect to atmosphere (from 55 to 3300 µmol L^{-1}) and calculated CO_2 emission (diffusion) flux ranging from 22 mmol CO_2 m^{-2} d^{-1} in the peatland pool to 1600 mmol CO_2 m^{-2} d^{-1} in the piezometer (**Table 1**).

3.2. DOC concentration evolution in the experiments

3.2.1. Biodegradation

In the Temnoe Lake, the range of DOC concentration change during 2-3 week incubation in the experimental reactors did not exceed 2 mg/L and typically remained within +0.5 to -1.5 mg/L, which is typically less than 10% of the initial DOC amount (**Fig. 2 and Fig. S1** of the Supplement). The biodegradable DOC was both season and depth dependent and ranged from 2

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

to 6 % (**Table 2**). The integral 2-week rates of biodegradation (**Table 3, Fig. 3 A**) demonstrated the highest values during autumn at depths of 0.5 m and 10 m and the lowest values during June at all depths. The final 0-10 m water column- and season-averaged biodegradation rate in Lake Temnoe ranged from 0.02 to 0.04 mg DOC L^{-1} d^{-1}. Integral rates of bio-degradation in the 0-10 m layer of the lake demonstrated a general increase over the entire open-water period (May to October; **Fig. 4**).

For Ilasskoe Bog, in the hydrological continuum studied during July, the BDOC was highest in the peatland pool (4.9 ± 1.4 %) and lowest in the outlet stream (3.1 ± 2.4 %; **Fig. 2** and **Fig. S1**). The integral rate of DOC biodegradation followed the order 'piezometer >> stream > lake' and ranged from 0.03 to 0.17 mg C L^{-1} d^{-1} (**Table 3, Fig. 3 A**).

3.2.2. Photodegradation

Compared to biodegradation, photodegradation demonstrated much higher values of PDOC and rates of reaction as well as higher variability among seasons and sites. In Lake Temnoe, the PDOC was the highest in June and the lowest in October (**Fig. 2 B** and **Table 2**). The maximal range of concentration change during 2-week period achieved 6-8 mg L^{-1} (**Fig. S2**) which was 10 to 20 % of the initial [DOC] values. The rates strongly decreased from May-June to the end of summer – autumn. The depth integrated (0 to 10 m) rate of DOM photodegradation in Lake Temnoe ranged from 0 in October to 0.2 mg C L^{-1} d^{-1} in June (**Table 3**; **Fig. 4 B**).

In the Ilasskoe Bog hydrological continuum during July, the photodegradation rate followed the order "outlet stream > piezometer >> peatland pool" (**Fig. 3 B**), where integral values of photodegradation equaled to 0.27±0.04, 0.33±0.07, and 0±0.05 mg C L^{-1} d^{-1}, respectively (**Table 3**).

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

3.3. Optical parameters of DOM

3.3.1 Biodegradation

In Lake Temnoe, the $SUVA_{254}$ remained relatively constant (4.2 to 4.6 L mg C^{-1} m^{-1}) across seasons and depths (**Table 1 B**). Over the course of biodegradation, the $SUVA_{254}$ did not change significantly (i.e., less than 0.2 units, which is comparable to the variability of duplicates; **Fig. S3**). The ratio E_{254}:E_{436}, which is an indicator of humification, increased with incubation time in Lake Temnoe waters; the magnitude of this increase followed the order "0.5 m > 5 m > 10 m" (**Fig. S4**). The ratio E_{254}/E_{365} also increased over the course of biodegradation, corresponding to an increase of mean molecular weight of DOM (Hiriart-Baer et al., 2008; Berggren et al., 2007). The ratio E_{365}/E_{470} also demonstrated the strongest increase in surface horizons and virtually no change in the deepest horizon (**Fig. S4**). An increase in the ratio E_{470}:E_{665} corresponds to a decrease in the degree of aromaticity (humification). An increase in the ratio E_{254}:E_{436} signifies a decrease in contribution of autochthonous (aquatic) DOM compared to terrestrial (soil) C, whereas an increase in the E_{254}:E_{365} ratio characterizes removal of low molecular weights compounds.

In Ilasskoe Bog samples, the highest SUVA was observed in the water of the piezometer and the lowest in the stream, but the evolution of this parameter in the course of biodegradation was rather weak (**Fig. S4**). The E_{254}:E_{365} and E_{254}:E_{436} ratios increased with incubation time in the piezometer and decreased with time in the stream (**Fig. S4**). The optical ratios (E_{254}:E_{436}, E_{365}:E_{470}, E_{470}:E_{665}) increased in the peatland pool, suggesting an increase in the molecular weight and an increase in the ratio of aromatic to aliphatic compounds.

Complete spectral differences between the experimental and control samples demonstrated rather weak ($\Delta A \leq 0.04$) changes of spectral parameters, mostly detectable after 10-12 days of incubation (**Fig. S5**). These results were generally consistent with the discrete spectral parameters presented above and demonstrated maximal effects in the piezometer and

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

bog outlet stream. In Lake Temnoe, the maximal impact of biodegradation on spectral parameters was observed in June, at 0.5 m depth.

3.3.2. Photodegradation

Similar to the DOC concentration, the optical parameters of DOM more strongly evolved over the course of photodegradation compared to the biodegradation experiments. In the Temnoe Lake, the strongest decrease in $SUVA_{254}$ was observed in the waters of all horizons in June. This decrease was less strong in October (**Fig. S6**). The E_{254}:E_{365} ratio demonstrated a sizable increase, mostly pronounced in June, with the lowest but still measurable increase in October. The E_{254}:E_{436} ratio strongly decreased with exposure time throughout all seasons (10 m depth) and only in June in the surface horizons (**Fig. S7**). An increase in the ratio E_{254}:E_{365} over the course of photodegradation corresponded to an increase in mean molecular weight of DOM, likely due to coagulation. The ratios E_{365}:E_{470} and E_{470}:E_{665} decreased in all experiments with the Temnoe Lake waters (**Fig. S7**), suggesting a decrease in the degree of humification (Battin, 1998) and a decrease in the ratio of aromatic to aliphatic moieties.

The SUVA in Ilasskoe Bog waters remained stable during photodegradation of stream waters and piezometer and strongly decreased in the peatland pool (**Fig. S6)**. The E_{254}:E_{436} ratio strongly increased in the peatland pool and exhibited measurable decrease in stream waters and piezometer, whereas the E_{365}:E_{470} ratio systematically decreased in all photodegradation experiments with the Ilasskoe Bog continuum (**Fig. S7**). Finally, the E_{470}:E_{665} ratio exhibited sizable decrease, in the order 'stream >> pool ≥ piezometer'. The total spectral differences between experimental and control reactors were mostly pronounced in stratified forest lake waters in June (ΔA = -0.4 to -0.4) and in the bog continuum in July, where effects were strongest in the piezometer and outlet stream waters (ΔA parameter as high as – 0.4 (**Fig. S8**).

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

3.4. Bacterial number evolution during biodegradation experiments

The number of cultivable eutrophic bacteria (EB) sizably (ca., 2 orders of magnitude) increased during biodegradation of Lake Temnoe waters. However, this evolution was not systematic in the course of incubation; there was a pronounced decrease after 2 weeks of exposure in June and August and rather stable concentration in waters of all horizons sampled in October (**Fig. S9**). Such maxima in June and August might be linked to consumption of substrate/nutrient limitations on bacterial growth. In Ilasskoe Bog continuum, the number of eutrophic bacteria decreased by an order of magnitude in the peatland pool and piezometer while remaining constant in the stream. The number of oligotrophic bacteria (OB) increased in waters of all Lake Temnoe horizons by ca. 2 orders of magnitude in August and October and 1 order of magnitude in June. In contrast, the OB number did not change or slightly decreased during incubations of waters from Ilasskoe Bog continuum (**Fig. S9**).

3.5. Trace element patterns

3.5.1 TE in biodegradation experiments

During biodegradation experiments, a number of components [Group 1] demonstrated a statistically significant (X > SD, Eqn. 1) decrease in concentration across the incubation period (**Table 2**): Al, Ti, Fe, Co, Cu, Ba, Nb, light REE (LREE) and Pb (as illustrated for Fe in **Fig. 5)** as well as Mn, V, and La (**Figs. S10, S11 and S12**, respectively). The most significant effects were observed for Fe in the 0-5 m horizon of Lake Temnoe (9 to 18 % in June, 6 to 13.5 % in August and 8 to 9.5 % in October) and 14% in the peatland pool of Ilasskoe Bog. Overall, for most elements except Fe and Mn, this increase was less pronounced than that of DOC; maximal effects were achieved for Lake Temnoe in August and October (V, Mn, Co, Cu, Ni, Nb, Hf, Pb and Th) and in June (Al and Ti). These elements are typically linked to DOM and Fe and present in the form of organic- and organo-mineral colloids. Certain elements [Group 2] did not

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

appreciably change their concentration (< 2 % decrease): Li, B, Na, Mg, K, Ca, Si, Ge, As, Rb, Sr, Mo, Sb, Mo and Ba. These elements are not linked to colloids of Fe(III) hydroxide and organic matter. Finally, some elements [Group 3] exhibited unstable behavior without systematic change in concentration during the exposure (X < SD, Eqns. 1-2): Cr, Zn, Cu, Sr, Cd, (Y, Zr), Cs, Tl and U. These elements cannot be considered as significantly impacted by the biodegradation process in Lake Temnoe water.

In the Ilasskoe Bog hydrological continuum, the most significant changes during biodegradation were observed in the peatland pool and outlet stream. Elements strongly (> 5-10 %; X > S.D. in Eqn. 1) affected by biodegradation were V, Fe, Ni, Ga, Y, LREEs and Pb.

3.5.2. TE in photodegradation experiments

The elements affected by photodegradation also formed three groups similar to those impacted by biodegradation. Aluminum, Fe, trivalent and tetravalent hydrolysates (Ti, Ga, Zr, Y, LREE and Th) and Nb of [Group 1] significantly (> 2 %; $p < 0.05$) decreased their concentration during photolysis as illustrated for Fe in **Fig. 6,** and for Ti and Zr in **Figs. S13** and **S14,** respectively. The decrease of Fe was mostly pronounced in Lake Temnoe water from 10 m depth, whereas that of Ti and Zr was detectable for all horizons and seasons except in October. For the Ilasskoe Bog continuum, there was no systematic change in Fe concentration, whereas concentrations of Ti and Zr systematically decreased over the course of sunlight exposure (**Figs. S13, S14**). Alkali (Li, Rb), alkaline-earth metals (Mg, Ca, Sr, Ba), Si and oxyanions (As, Mo, Sb) of [Group 2] were weakly (< 2 %) affected by photolysis. Finally, the remaining trace elements of [Group 3] did not exhibit any systematic evolution of concentration during exposure to sunlight, or these changes were inferior to the uncertainties of replicates (X < S.D. in Eqn. 1).

We found that, unlike for DOC, the magnitude of trace element concentration decrease during photodegradation was generally lower than that of biodegradation experiments. Overall,

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

the strongest effects were observed for Ti (3 to 9% in Lake Temnoe; 20% in Ilasskoe Bog), Ga (6 to 14%), Zr (14-17% in Lake Temnoe), Nb (8 to 13%) and Th (8 to 19% in the Temnoe Lake and up to 50% in the Ilasskoe Bog). These effects were mostly pronounced in the Temnoe Lake in June and August and in peatland pool of the Ilasskoe Bog (July).

4. Discussion

4.1. Comparison between biodegradation and photolysis

The impact of season on the biodegradable DOC could be tested only for Lake Temnoe because it was sampled during the 3 main hydrological periods. The maximal biodegradation of the lake water was observed during autumn, when large amount of labile fresh soil OM and plant litter were delivered to the lake from the watershed via surface runoff. The water temperature seems to be of secondary importance for the intensity and rate of DOM biodegradation. It is worth noting that the seasonal pattern of BDOC in the humic lake quantified in this study (Fig. 4 A) contrasted with previous works on biodegradation of large Arctic streams and rivers whose BDOC decreased as the Arctic summer progressed (Vonk et al., 2015). Presumably, the input of fresh material (plant litter at the end of summer-autumn from the forested watershed of Lake Temnoe) provided elevated biodegradation in the water column at the end of the open water season. Another reason could be due to lake overturn in October and exposure of deep partially autochthonous, and thus biodegradable, DOM to the surface biota. This biodegradable DOM originated from leaching of organic detritus accumulated during summer months along the bottom lake horizons (Chupakov et al., 2017). A supply of limiting nutrients (N and P) to the upper 0-10 m layer during lake overturn could also promote such biodegradation in October.

The highest biodegradation rates in the uppermost sections of the bog hydrological continuum (piezometer, **Fig. 3 A**) are consistent with recent findings on organic-rich waters of permafrost peatlands (Shirokova et al., 2019; Payandi-Rolland et al., 2020) and earlier results on

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

headwaters, small streams and soil leachates (Roehm et al., 2009; Ilina et al., 2014; Mann et al., 2014, 2015; Larouche et al., 2015; Spencer et al., 2015; Vonk et al., 2015; Moody et al., 2013; Pickard et al., 2017; Dean et al., 2019). This could be due to the very short water residence time and freshly leached DOM in these water objects (i.e., Mann et al., 2012; Abbott et al., 2014; Payandi-Rolland et al., 2020), given that bioavailable DOM components leached from plant litter are rapidly utilized (Textor et al., 2018). At the same time, overly low BDOC (2-8 %) values, regardless of depth and season in humic lake and across the hydrological continuum of the bog (**Fig. 2 A**), are supportive of previous results for permafrost peatlands from the neighboring region (Shirokova et al., 2019). A general path for DOM spectral properties modification over the course of biodegradation consisted of an increase in aromaticity of DOM due to preferential uptake of non-humic low molecular weight (LMW) compounds. However, this was not accompanied by an increase in SUVA (**Fig. S3**). Presumably, the proportion of these compounds in the overall DOC level was quite low and could not impact SUVA evolution. Globally, the evolution of optical ratios was consistent with bacterial consumption of aliphatic LMW compounds and an increase in the overall aromaticity of DOM.

Concerning the seasonal variation of photodegradation in the deep humic lake, maximal effects were observed in June. These maximal effects likely occurred due to fresh terrestrial organic matter (plant litter) leached from the watershed and then efficiently processed during Arctic summer. By July, the majority of DOM in lake surface layers was already degraded; this was observed for both the deep stratified Lake Temnoe and shallow peatland pool samples. In the end of summer, photodegradable DOM occurred solely in the deep horizon (10 m) of the lake. However, in October, even this deep horizon did not exhibit photodegradable DOM, presumably due to low insolation and unfavorable temperatures. It should be noted that labile phenolic, carbohydrates, N-containing bases and smaller molecular weight compounds are abundant in litter leachates produced during initial decay stages (Kiikkilä et al., 2011, 2012,

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

2013; Hensgens et al., 2021). By July, most of the biodegradable DOM was already removed, and in October, the effects were much lower. Therefore, photolabile DOM is delivered from the forested watershed to the lake essentially during surface flux, at high water flow. It is then quickly removed from the water column, which was especially seen in the 0.5 and 5 m horizons of Lake Temnoe. Although labile organic matter from litter fall was also delivered during autumn high flow, presumably, during this period, the conditions for photolysis (low temperature, short daytime period and insufficient light) were not as favorable as those in June or August.

Photodegradation of waters from the Ilasskoe Bog continuum demonstrated maximal rates in soil waters from the piezometer (**Fig. 3 B**). During photolysis of humic water, a decrease in optical ratios (E_{365}:E_{470}; E_{470}:E_{665}) clearly indicated preferential degradation of humic aromatic compounds. The strong effect of photodegradation on DOM optical properties in the 650-500 nm region may be linked to decomposition of complex DOM into smaller molecules, whereas a decrease of absorbance in the 230-400 nm region **(Fig. S8)** indicates destruction of aromatic compounds, progressively increasing over insolation time. A recent study of DOM photolysis in humic-rich forested streams demonstrated that high aromatic material was photochemically converted into smaller non-fluorescent molecules (Wilske et al., 2020).

Results obtained on the more important role of photodegradation over biodegradation are generally consistent with earlier reports on the dominance of photolysis for DOM processing in Arctic waters within North America (Cory et al., 2014; Ward et al., 2017), the Canadian temperate zone (Winter et al., 2007; Porcal et al., 2013, 2014, 2015), and Swedish headwater catchments (Köhler et al., 2002). According to former results for Scandinavian surface waters, the main impact of DOM photolysis is reflected by a decrease in the proportion of aromatic (colored) DOC and a rather small (≤ 10 %) change in bulk DOC concentration (Groeneveld et al., 2016; Koehler et al., 2014), Canada (Laurion and Mladenov, 2013; Gareis and Lesack, 2018) and NW Russia (Oleinikova et al., 2017; Chupakova et al., 2018).

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

As a further perspective of this work, one has to consider biodegradation of photolytically altered DOM given that photo-oxidation is known to transform molecular structures into more bioavailable forms (e.g., Cory and Kling, 2018; Sulzberger et al., 2019) thereby stimulating microbial growth under sunlight, as is known for other Arctic and subarctic settings (i.e., Drozodova et al., 2020; Laurion et al., 2020).

4.2. Possible impact of microbial and photolytic processing on CO_2 emissions from water surfaces

The integral rates of DOM bioprocessing in the water column of Lake Temnoe (**Table 3, Fig. 4 A**) allow quantifying the potential contribution of biodegradation to CO_2 production and emission. Assuming all biodegraded DOM is transformed into CO_2 and there is no biomass increase or sedimentation, a 1 m water layer of the lake can emit 0.02 g C-CO_2 m^{-2} d^{-1} in June and 0.04 g C-CO_2 m^{-2} d^{-1} in October. Assuming the entire water column studied in this work (10 m depth of Lake Temnoe) participates in DOM biodegradation and CO_2 emission, the integral flux amounts to 0.2-0.4 g C-CO_2 m^{-2} d^{-1} across the seasons. These values are comparable to typical values of CO_2 evasion from the surface of this lake during different seasons (30-70 mmol CO_2 m^{-2} d^{-1}, or 0.36-0.84 g C-CO_2 m^{-2} d^{-1}; **Table 1 B**).

For surface waters of Ilasskoe Bog, maximal CO_2 production due to DOM biomineralization alone (**Table 3**) ranged from 0.06 g C-CO_2 m^{-2} d^{-1} for the peatland pool (2 m deep) to 0.03 g C-CO_2 m^{-2} d^{-1} for the outlet stream (0.5 m deep). However, in summer, the peatland pool and stream emitted 0.27 and 1.8 g C-CO_2 m^{-2} d^{-1} (**Table 1 A**) which could not be sustained by DOM biodegradation.

The addition of photodegradation (assuming a photic layer depth of 3.5 m) to DOM bioprocessing in the water column of the Temnoe Lake during open water season can further increase potential CO_2 production in the water column thus making it possible to provide entire

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

observed CO_2 evasions. For the case of Ilasskoe Bog waters, the addition of photolytic degradation increases projected CO_2 emission from the outlet stream by a factor of 5, which is still below the actual CO_2 flux, whereas DOM photolysis has no impact on CO_2 emissions from the peatland pool. Note that, although the depth of sunlight processing in boreal waters is typically 1-0.8 m (Vähätalo et al., 2000; Koehler et al., 2014), a more recent study concluded that direct photomineralization of DOM in Artic humic ponds is limited to the first centimeters of the water column (Mazoyer et al., 2022). Furthermore, in typical DOM-rich Arctic waters, only half of sunlight-associated DOC losses is converted into CO_2 and the rest may be turned into particles through photoflocculation (e.g., Mazoyer et al., 2022). Therefore, despite a faster photodegradation rate compared to biodegradation, due to the shallow photic layer in humic waters, the biodegradation may provide the largest impact on CO_2 emission from the water column of boreal waters.

4.3. Impact of DOM bio- and photo transformation on trace element cycling

Among all major and trace elements measured in the experiments, only a few (trivalent and tetravalent hydrolysates, TE^{3+}, TE^{4+}) were impacted by both photo- and biodegradation. It is known that these elements are essentially present in the form of large molecular size, highly polymerized and presumbaly aromatic, organo-Fe/Al colloids in humic boreal/subacrtic lakes (Pokrovsky et al., 2012, 2016), rivers (Krickov et al., 2019; Pokrovsky et al., 2010), and soil porewaters (Pokrovsky et al., 2005; Raudina et al., 2021). Therefore, insoluble TE^{3+} and TE^{4+} generally followed the removal of Fe(III) in the form of particulate Fe hydroxides, after breaking the Fe-DOM bonds that stabilized colloidal Fe(III) hydroxides. This destabilization and Fe hydroxide particle formation is known to occur either via biodegradation (i.e., Oleinikova et al., 2018) or photolysis (Kopacek et al., 2005, 2006; Oleinikova et al., 2017; Chupakova et al., 2018). At the same time, some micronutrients (V, Mn, Co, Cu and Ba) were affected solely by

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

biodegradation. This can reflect uptake of these metals by growing bacterial cells, as is known from laboratory experiments with pure cultures of heterotrophic bacteria (Shirokova et al., 2017a).

Note that the effects of bio- and photodegradation were more pronounced for light REE (LREE) compared to heavy REE (HREE). This result is consistent with the fact that LREE have stronger association with Fe hydroxide compared to organic complexes, as known from general chemical considerations and laboratory experiments (i.e., Bau, 1999) and evidenced in various boreal and subacrtic settings (Pokrovsky et al., 2016; Krickov et al., 2019). Given that the main effect of both photolysis and biodegradation of DOM in humic Fe(III)-rich surface waters is coagulation of dissolved Fe(III) in the form of Fe oxy(hydr)oxides, the LREE are removed from solution [either in the form of adsorbed complexes or coprecipitated with Fe oxy(hydr)oxides] while HREE remain [in the form of strong aqueous complexes].

In former studies of photo- and biodegradation of surface waters from permafrost peatlands, only a few nutrients (P, Fe, Zn and V) and insoluble low mobility trace metals (Ti, Zr, Nb and Th) demonstrated a decrease in concentration (Shirokova et al., 2019). This list of elements is generally consistent with that established in the present study of humic subarctic lakes of the non-permafrost zone, except P and Zn which did not exhibit sizable removal in our experiments. It is possible that a high proportion of low molecular weight $LMW_{<1\ kDa}$ (and thus, potentially bioavailable) forms of macro- and micronutrients, such as P and Zn, in the permafrost ice (i.e., Kuzmina et al., 2023) can be delivered to the lake and river via suprapermafrost flow (Raudina et al., 2018, 2021), which is eventually responsible for elevated bioavailability of these elements in permafrost surface waters, as reported in former experiments.

Conclusions

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

Seasonally resolved bio- and photo-degradability of DOM in a deep stratified lake and summer measurements from a peat bog's hydrological continuum within the boreal zone confirmed the initial hypothesis that the subsurface and deep horizons of these stratified waters are mostly sensitive to sunlight impact, and that maximal effects of photodegradation occurred in the month of June during strong insolation. In contrast, the biodegradation of DOM from the humic lake was mostly pronounced during October, when fresh leachates of forest litter were exported from the watershed. The evolution of optical parameters of DOM demonstrated removal of aliphatic, presumably autochthonous, organic ligands during biodegradation and photolysis of aromatic humic molecules. Insoluble low-mobility trace metals such as trivalent and tetravalent hydrolysates were affected by both bio- and photodegradation, as they are associated with coagulating Fe(III) oxyhydroxides. A few micronutrients (V, Mn, Co, Cu and Ba) were, however, removed during biodegradation experiments, thus reflecting their possible uptake by microorganisms.

Although DOM photodegradation rates were sizably higher compared to those of biodegradation, the rather thin photic layer in humic waters does not allow for significant contribution of photolysis in overall CO_2 emission from lake and bog surfaces. In the deep stratified lake, the biodegradation alone was capable explaining observed CO_2 emissions, while in the shallow bog continuum, the sum of bio- and photodegradation were not sufficient to provide CO_2 flux. The high seasonal dynamics and spatial variability in both photo- and biodegradability of DOM and related trace elements of humic surface waters in the boreal zone encountered in this study suggest the need for further assessment of rates of these processes with focus on early spring and late autumn, the periods of maximal photo- and biodegradation, respectively. Considering the strong spatial variations of DOM processing in the aquatic continuum, focus should be centered on the most dynamic components such as small streams and subsurface waters, which demonstrated the highest rates of both photo- and biodegradation.

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

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Authors contribution.

AVC and OP designed the study and wrote the paper; AC and SB performed sampling, analysis and their interpretation; LS performed bacterial number assessment and DOC results interpretation; AVC and OP provided analyses of literature data

Competing interests.

The authors declare that they have no conflict of interest.

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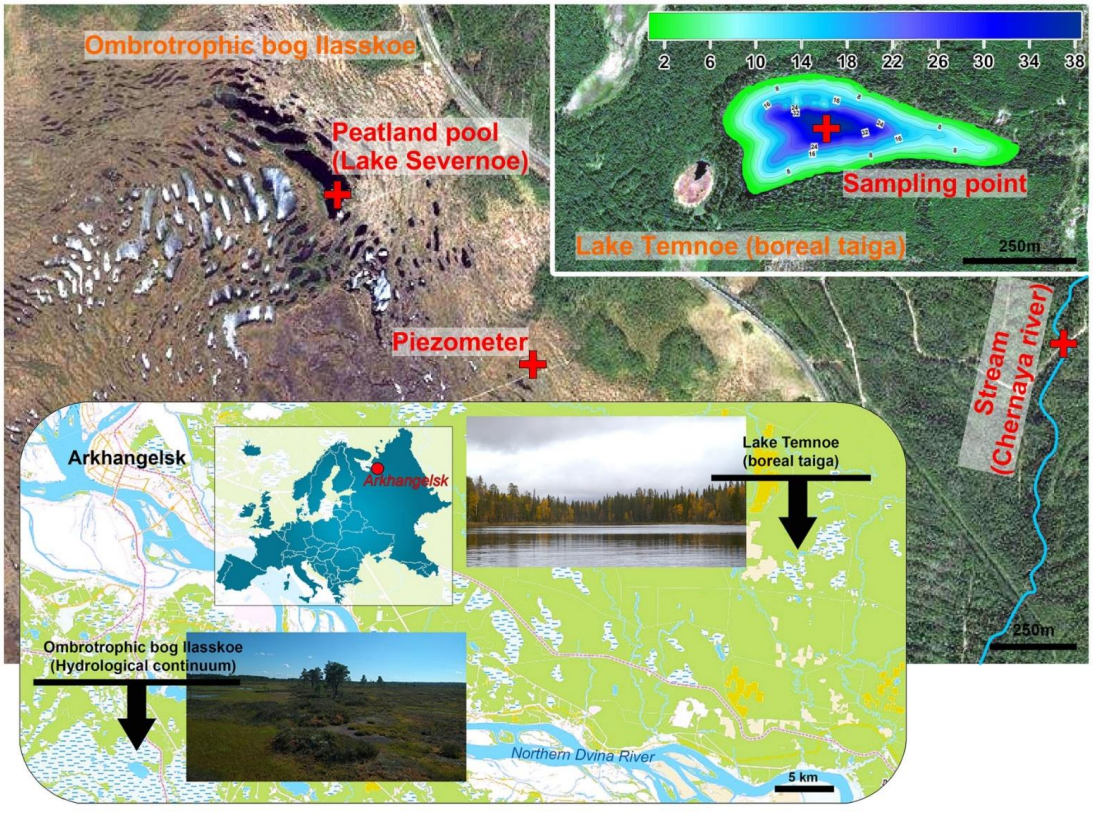

Fig. 1. Geographical location of studied hydrological continuum for Ilasskoe Bog waters and deep stratified Lake Temnoe in the boreal forest. Photo and map credits of Chupakov A.V.

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

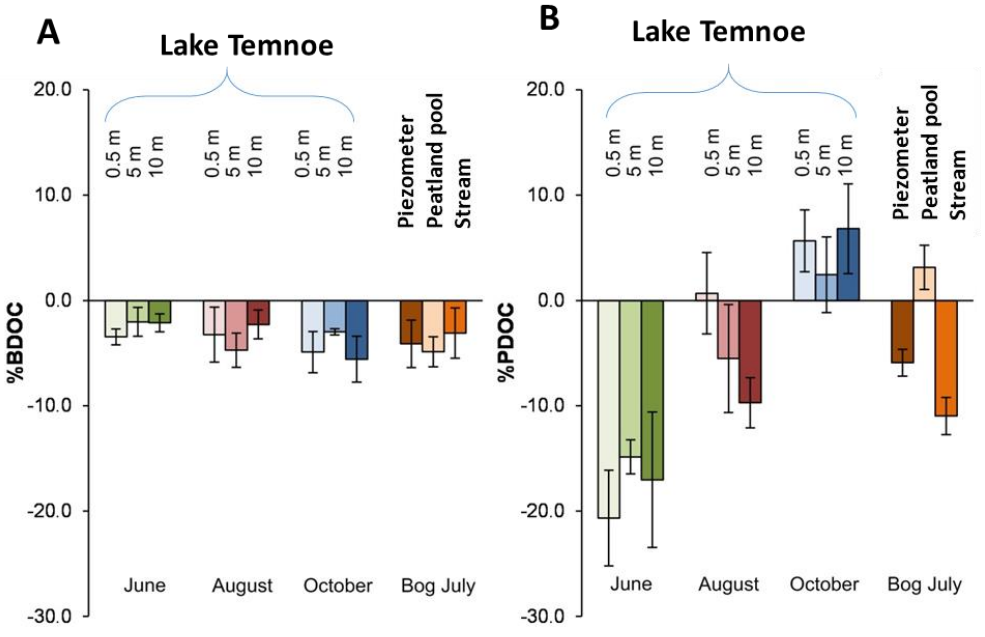

Fig. 2. Percentage of bio- (A) and photo- (B) degradable DOC presented as relative decrease in DOC concentration between the initial and final value for the Temnoe Lake (June, August and October) and Ilasskoe Bog surface waters (July). Error bars are 1 s.d. of duplicates relative to the control (see Eqn. 1-2 in the text). Positive values signify nil photodegradation (experimental artifacts of DOC production).

https://doi.org/10.5194/egusphere-2024-233
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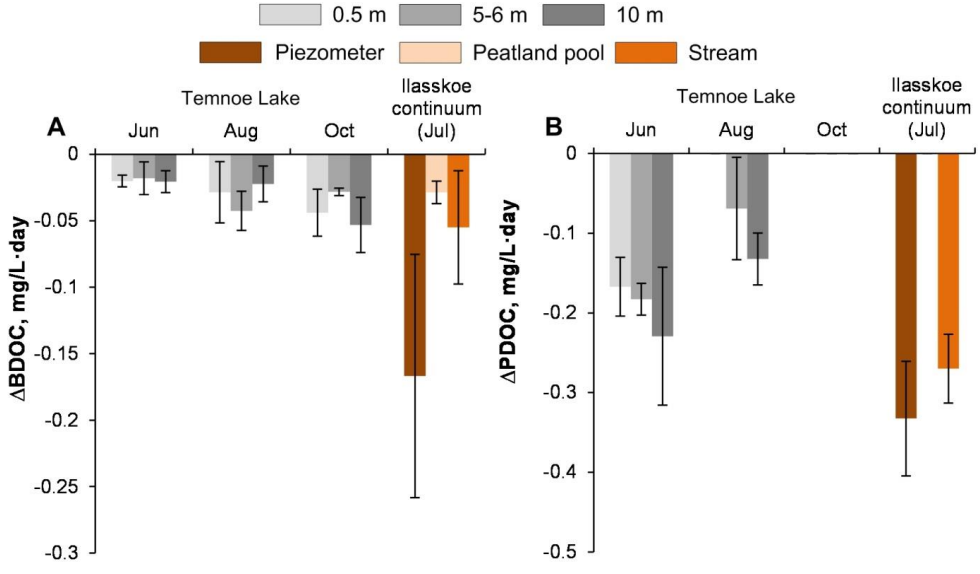

Fig. 3. Rates of DOC bio- (A) and photo- (B) degradation. The values are negative because they represent a decrease in DOC concentration over the course of the experiment.

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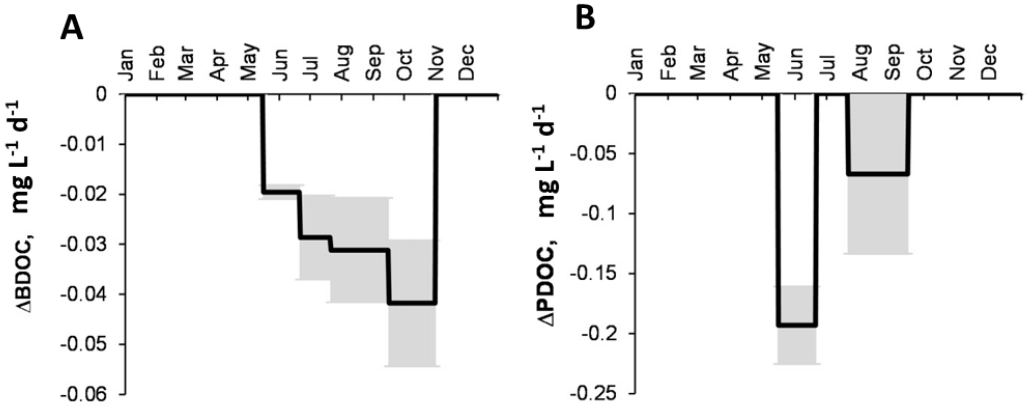

Fig. 4. Integral rates of bio- (ΔBDOC, **A**) and photo- (ΔPDOC, **B**) degradation in the 0-10 m layer of Lake Temnoe across the entire open-water period (May to October). Rate values are negative because they signify a decrease in DOC concentration. Uncertainties are represented by gray shaded rectangles.

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Preprint. Discussion started: 13 February 2024

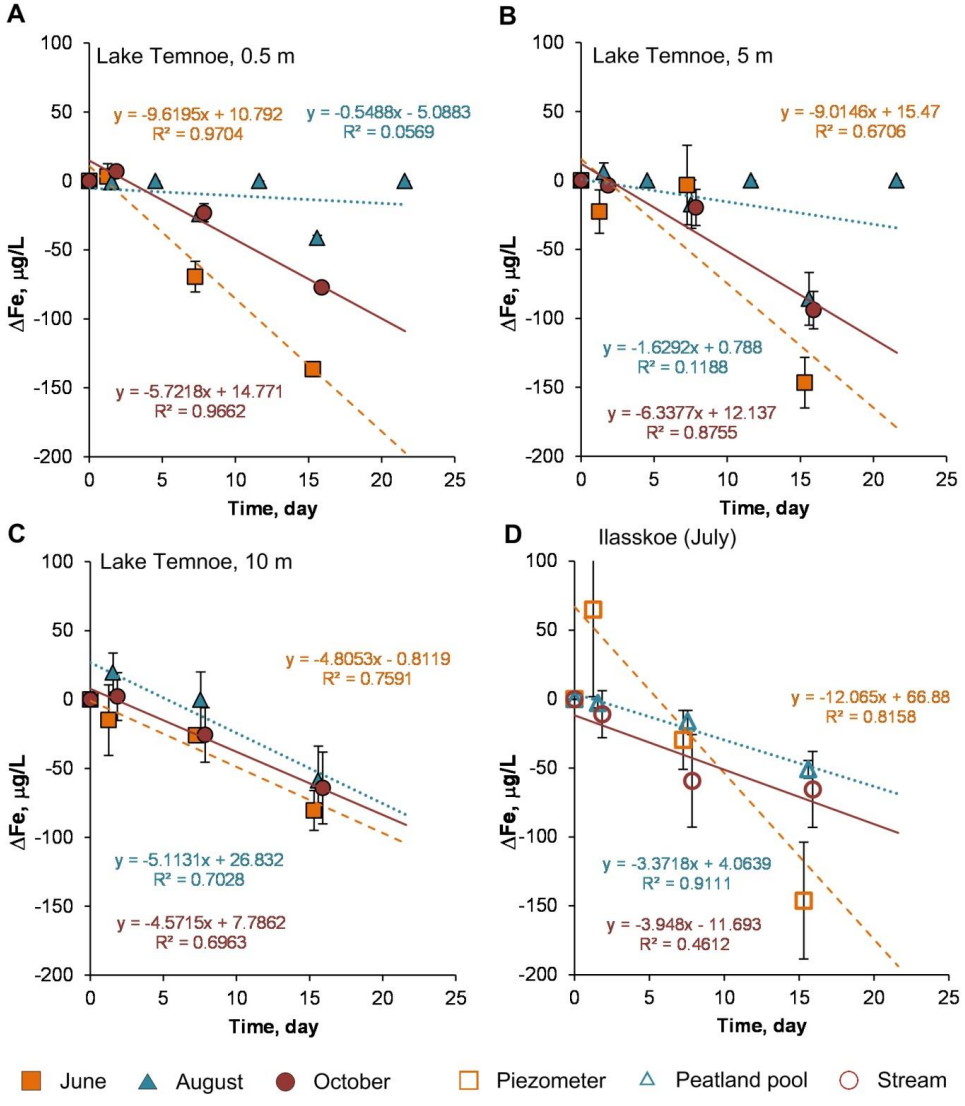

Fig. 5. Change in Fe concentration (relative to control) over time in biodegradation experiments. Error bars are 1 s.d. of duplicates. Temnoe Lake 0.5 m (A), 5 m (B) and 10 m (C) in June (squares), August (triangles) and October (circles). Ilasskoe Bog continuum in July (D) including piezometer (squares), Severnoe peatland pool (triangles) and stream Chernyi (circles).

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

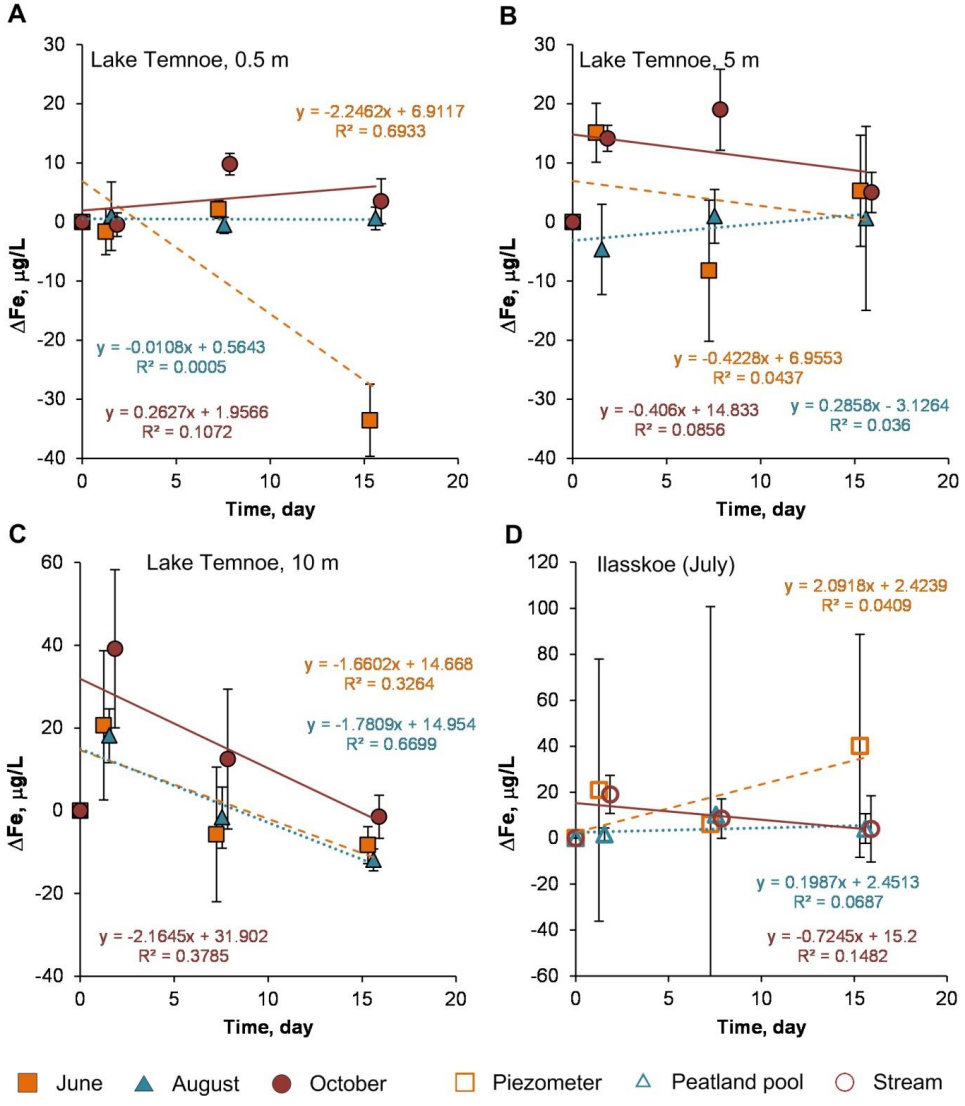

Fig. 6. Change in Fe concentration (relative to the control) over time in photo-degradation experiments. The error bars are 1 s.d. of duplicates. Lake Temnoe 0.5 m (A), 5 m (B) and 10 m (C) in June (squares), August (triangles) and October (circles). Ilasskoe continuum in July (D) includes piezometer (squares), peatland pool Severnoe (triangles) and stream Chernyi (circles)

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

Table 1. Landscape setting, hydrochemical characteristics and CO_2 concentration and emission flux of studied waters. S.C. is specific conductivity and EB and OB is eutrophic and oligotrophic bacteria count, respectively.

1A. Ilasskoe bog continuum in July.

	Piezometer	Lake Severnoe	Stream Chernyi
GPS coordinates	N64.328694° E40.612556°	N64.334361° E40.609667°	N64.330982° E40.653352°
Description	Shallow groundwater	Peatland pool	Outlet stream
T,°C	11.4	19.4	13
O2, mg/L	0.6	8.6	7.5
pH	3.9	4.0	5.7
S.C., µS cm^{-1}	46	17	26
DOC, mg L^{-1}	87.6	12.7	38.4
DIC, mg L^{-1}	0.32	0.40	0.38
$SUVA_{254}$	4.13	3.80	4.85
P-PO_4, µg L^{-1}	8.6	3.0	1.7
P_{total}, µg L^{-1}	153	10	20
N-NO_3, µg L-1	111	70	98
N-NH_4, µg L^{-1}	85.4	16.1	12.6
N_{total}, µg L^{-1}	1180	222	399
Si, µg L^{-1}	1808	47	2076
CO_2, µmol/L	3360	55	318
CO_2 flux, mmol m^{-2} d^{-1}	1600	22	151
EB, CFU mL^{-1}	49360	56600	9000
OB, CFU mL^{-1}	54560	37900	21600

1B. Lake Temnoe across seasons and depths.

Month	Jun	Jun	Jun	Aug	Aug	Aug	Oct	Oct	Oct
	0.5	5	10	0.5	5	10	0.5	6	10
GPS	N64.47683° E041.74533°								
Description	Lake in the northern taiga								
T,°C	12.7	4,9	4,5	18.4	5.5	4.3	9.0	5.8	4.4
O2, mg/L	8,45	4,8	4,5	7.78	4.93	2.63	8.90	4.46	2.14
pH	5.2	5.2	5.3	6.0	5.5	5.7	5.2	5.2	5.1
S.C., µS cm^{-1}	17	17	19	17	17	19	18	18	20
DOC, mg L^{-1}	12.6	19.2	21	19	19.5	21.2	19.4	20.6	20.6
DIC, mg L^{-1}	0.55	0.53	0.49	0.70	0.71	0.66	0.67	0.67	0.64
$SUVA_{254}$	4.6	4.7	4.6	4.2	4.5	4.5	4.3	4.3	4.7
P-PO_4, µg L^{-1}	2.9	3.3	6.4	0.9	3.6	9.4	3.8	4.6	4.2
P_{total}, µg L^{-1}	19	17	19	20	16	20	18	19	20
N-NO_3, µg L-1	119	150	137	86	152	254	88	85	100
N-NH_4, µg L^{-1}	7.1	8.0	10.0	9.1	17.5	13.8	16.4	14.1	15.5
N_{total}, µg L^{-1}	305	420	408	355	315	337	425	416	396
Si, µg L^{-1}	1940	2268	2354	1183	2208	2714	2269	2380	2380
CO_2, µmol/L	99	309	329	110	256	337	223	232	253
CO_2 flux, mmol m^{-2} d^{-1}	32	-	-	46	-	-	71	-	-
EB, CFU mL^{-1}	-	36	50	259	92	270	780	220	105
OB, CFU mL^{-1}	50	570	420	-	190	-	680	150	66

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

Table 2. The % bio- and photodegradable solutes (mean ± s.d.) whose relative change (concentration decrease) in the course of experiment was superior to that of SD. Prefix ΔB and ΔP represents the effect of bio- and photodegradation, respectively. Duration of biodegradation and photodegradation is 21.6±0.1 and 15.6±0.1 days, respectively. W represents the probability of measurable effect, significantly different from changes in the control reactors. Only the components with W ≥ 33% are presented. Temnoe Lake is deep stratified lake in the forest. Peizometer, peatland pool and outlet stream represent the hydrological continuum of the Ilasskoe Bog.

Index	Temnoe Lake 0.5 m (Jun)	Temnoe Lake 5 m (Jun)	Temnoe Lake 10 m (Jun)	Temnoe Lake 0.5 m (Aug)	Temnoe Lake 5 m (Aug)	Temnoe Lake 10 m (Aug)	Temnoe Lake 0.5 m (Oct)	Temnoe Lake 6 m (Oct)	Temnoe Lake 10 m (Oct)	Piezo-meter (Jul)	Peatland pool (Jul)	Outlet stream (Jul)
æ, µS/cm	17	17	19	17	17	19	18	18	20	46	17	26
ΔB(æ ±SD)	-24±4	-26±7	-30±5	-23±3	-27±3	-33±16	-24±7	-23±4	-17±5	0	-18±10	-29±4
DOC, mg/L	12.6	19.2	21.0	19.0	19.5	21.2	19.4	20.6	20.6	87.6	12.7	38.4
ΔB(DOC±SD)	3.4±0.8	2.0±1.4	2.1±0.8	3.2±2.6	4.7±1.6	2.3±1.4	4.9±2.0	3.0±0.3	5.6±2.2	4.1±2.3	4.9±1.4	3.1±2.4
ΔP(DOC±SD)	20.7±4.6	14.9±1.6	17.0±6.4	0	5.5±5.1	9.7±2.4	0	0	0	5.9±1.3	0	11.0±1.8
Al, µg/L	275	298	329	254	296	335	275	288	323	276	59	388
ΔB(Al±SD)	3.5±1.4	1.8±0.9	0	2.0±1.3	0	1.4±1.5	2.0±1.9	0	0	0.9±2.2	0	1.3±1.8
ΔP(Al±SD)	1.9±1.1	2.7±0.9	3.6±1.3	0	2.5±1.3	1.7±2.0	0.7±0.9	0	0	0	0	0.8±0.9
Ti, µg/L	1.5	2.1	2.6	1.1	2.0	2.6	1.7	1.9	2.5	3.7	0.6	5.0
ΔB(Ti±SD)	-9.2±1.6	-9.9±7.4	-2.6±2.7	-4.8±3.4	-1.8±2.7	0	-3.6±1.7	-1.0±3.1	-1.0±3.9	-2.3±3.6	-2.2±1.7	-1.4±2.2
ΔP(Ti±SD)	-0.1±3	-3±3	-8±3	0±0	-9±1	-3±2	-2±4	0	0	0	-20±4	-3.3±0.5
V, µg/L	0.5	0.6	0.7	0.4	0.5	0.7	0.4	0.5	0.7	1.1	0.5	1.3
ΔB(V±SD)	-8.3±16.2	-5.4±3.2	-4.9±2.3	-6.8±7.5	-10.0±4.6	-1.7±1.6	-14.7±11	-13.9±4.3	-16.1±1.7	-3.2±2.6	-0.2±3.4	-17.9±5.0
Mn, µg/L	39	55	79	17	48	93	30	47	105	78	9	47
ΔB(Mn±SD)	0	0	-0.3±2.2	-31.8±1.3	-3.2±1.6	-0.6±2.2	-4.8±2.2	-3.2±1.7	-0.4±0.1	0	0	-1.6±2.8
Fe, µg/L	358	527	710	165	460	795	317	448	820	4402	157	1006
ΔB(Fe±SD)	-18.1±2.5	-9.1±2.6	-5.4±1.6	-13.5±1.0	-6.3±2.6	-1.4±1.9	-9.5±1.4	-7.8±1.9	-3.3±1.8	-0.8±0.8	-13.6±4.3	-4.5±2.4
ΔP(Fe±SD)	-3.9±0.6	-2.0±1.9	-4.0±1.3	0	-2.9±1.5	-0.2±0.6	-1.2±0.4	0	0	0	0	0

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

Table 2, continued.

Index	Temnoe Lake 0.5 m (Jun)	Temnoe Lake 5 m (Jun)	Temnoe Lake 10 m (Jun)	Temnoe Lake 0.5 m (Aug)	Temnoe Lake 5 m (Aug)	Temnoe Lake 10 m (Aug)	Temnoe Lake 0.5 m (Oct)	Temnoe Lake 6 m (Oct)	Temnoe Lake 10 m (Oct)	Piezo-meter (Jul)	Peatland pool (Jul)	Outlet stream (Jul)
Co, µg/L	0.28	0.39	0.68	0.07	0.30	0.65	0.18	0.31	0.74	0.45	0.06	0.30
ΔB(Co±SD)	-2.2±5.1	-1.2±2.1	-3.7±4.6	-32.7±2.6	-8.1±5.6	-2.7±3.3	-11.0±4.4	-9.1±5.1	-1.6±0.4	0	0	-20.6±27.8
Cu, µg/L	0.5	0.6	0.7	0.6	0.6	0.7	0.7	0.7	0.5	1.5	0.3	0.8
ΔB(Cu±SD)	0	0	0	-14.3±1.4	-6.8±4.0	-17.9±11.0	-5.3±4.8	-4.1±8.0	-1.4±12.3	0	-7.7±9.9	0
Ga, µg/L	0.017	0.022	0.026	0.012	0.016	0.023	0.017	0.015	0.024	0.126	0.016	0.066
ΔP(Ga±SD)	-14±6	-13±5	-10±4	0	-1±8	0	-10±4	0	0	-7±5	-5±8	-6±3
Y, µg/L	0.22	0.25	0.28	0.20	0.24	0.28	0.22	0.23	0.28	0.10	0.01	0.21
ΔP(Y±SD)	-1.3±4.6	-6.7±0.9	-5.3±2.5	0	-2.3±0.7	-1.0±1.6	-1.4±0.2	0	0	0	-5.8±2.7	0
Zr, µg/L	0.4	0.4	0.5	0.4	0.5	0.5	0.4	0.4	0.5	0.3	0.1	0.4
ΔP(Zr±SD)	-15±4	-14±0	-13±2	-9±20	-17±1	-14±3	-4±4	0	0	0	-32±3	-8±1
Nb, µg/L	0.016	0.020	0.025	0.012	0.020	0.024	0.017	0.018	0.025	0.033	0.005	0.042
ΔB(Nb±SD)	-3.6±10.2	-1.7±7.0	0	-7.7±4.8	-1.1±2.6	0	-7.3±2.3	-1.5±6.3	-5.0±4.0	-2.4±1.5	0	0
ΔP(Nb±SD)	-9±3	-8±3	-9±1	-6±23	-13±2	-10±5	-8±4	0	-3±3	0	-13±10	-10±4
Ba, µg/L	4.8	5.1	5.8	4.6	5.0	5.7	4.9	4.8	5.6	54.4	1.5	56.8
ΔB(Ba±SD)	-2.2±0.7	-2.8±1.7	-1.0±2.7	0	0	0	-1.9±0.5	-1.7±3.7	-5.9±1.6	0	0	-1.3±3.4
La, µg/L	0.23	0.26	0.30	0.21	0.26	0.32	0.24	0.27	0.31	0.07	0.01	0.22
ΔB(La±SD)	-4.9±6.5	0	0	-3.9±0.9	-0.3±1.6	-2.6±1.5	-1.1±3.6	-2.4±1.2	-4.0±2.8	-0.8±10.4	-29.7±10.0	-2.0±2.8
ΔP(La±SD)	-3.8±3.6	-1.2±5.9	-2.0±2.7	0	-3.6±1.0	-3.2±1.6	-1.8±0.9	-2.6±1.0	0	0	-13.5±3.0	0
Ce, µg/L	0.58	0.65	0.71	0.50	0.62	0.78	0.59	0.63	0.78	0.21	0.03	0.56
ΔB(Ce±SD)	-5.2±4.2	0	0	-4.4±0.7	-0.1±1.4	-0.8±1.0	-0.9±2.9	-0.8±1.1	-2.2±2.2	0	-9.3±5.9	-2.0±1.9
ΔP(Ce±SD)	-4.9±1.9	-6.2±1.2	-1.1±1.7	0	-1.5±0.5	-3.2±1.7	-1.9±1.2	0	-0.047±0.46	-3.7±1.5	-2.4±1.6	-3.6±1.1

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

1097 **Table 2**, continued.

Index	Temnoe Lake 0.5 m (Jun)	Temnoe Lake 5 m (Jun)	Temnoe Lake 10 m (Jun)	Temnoe Lake 0.5 m (Aug)	Temnoe Lake 5 m (Aug)	Temnoe Lake 10 m (Aug)	Temnoe Lake 0.5 m (Oct)	Temnoe Lake 6 m (Oct)	Temnoe Lake 10 m (Oct)	Piezo-meter (Jul)	Peatland pool (Jul)	Outlet stream (Jul)
Pr, µg/L	0.075	0.085	0.094	0.069	0.082	0.105	0.077	0.084	0.102	0.027	0.005	0.070
ΔB(Pr±SD)	-1.9±5.2	0	0	-4.0±1.1	0	-0.7±1.3	0	-0.9±1.6	-3.2±2.3	-3.0±1.7	-10.8±8.4	-1.7±2.3
ΔP(Pr±SD)	-3.0±3.9	-5.5±0.9	-0.4±2.9	13.7±20.8	0	-6.1±2.8	0	-1.3±1.8	0	-0.01±2.4	-16.9±3.0	-2.4±2.1
Nd, µg/L	0.33	0.34	0.39	0.29	0.33	0.42	0.33	0.32	0.41	0.11	0.02	0.27
ΔP(Nd±SD)	-7.8±2.4	-4.5±2.5	-0.8±2.1	0	0	-3.8±2.5	-2.0±3.3	0	0	-2.5±2.8	-24.7±6.2	0
Eu, µg/L	0.015	0.017	0.016	0.012	0.016	0.020	0.014	0.018	0.021	0.011	0.001	0.017
ΔP(Eu±SD)	0	-10.9±3.3	0	0	-0.8±4.7	-3.0±4.0	0	-8.7±8.4	-6.9±4.2	-23.7±8.6	-58.2±15.2	-1.0±1.0
Gd, µg/L	0.06	0.07	0.08	0.05	0.07	0.08	0.06	0.07	0.09	0.02	0.00	0.06
ΔP(Gd±SD)	-0.2±4.4	-6.5±2.9	-3.5±3.5	0	-6.0±3.2	-7.6±1.9	-2.3±2.0	0	-6.7±3.2	0	0	-3.2±3.5
Ho, µg/L	0.009	0.009	0.011	0.007	0.009	0.011	0.009	0.009	0.011	0.004	0.0004	0.009
ΔP(Ho±SD)	-1.9±1.5	-0.3±1.6	-4.6±6.5	0	-0.1±3.6	-3.4±3.2	-11.0±4.9	0	0	-21.5±11.3	0	-10.9±5.5
Er, µg/L	0.023	0.025	0.033	0.022	0.026	0.030	0.022	0.023	0.031	0.011	0.001	0.023
ΔB(Er±SD)	0	0	0	-5.2±3.0	-2.1±1.8	0	-2.1±3.5	0	-2.0±5.3	-15.6±4.9	-22.9±19.5	0
Pb, µg/L	0.23	0.24	0.23	0.16	0.23	0.39	0.28	0.28	0.32	11	0.35	0.65
ΔB(Pb±SD)	0	0	0	-21.3±2.5	-2.0±7.6	-2.4±1.6	-8.2±3.3	-7.2±9.9	-0.8±2.7	0	-17.4±1.0	-8.5±10.9
Th, µg/L	0.046	0.052	0.066	0.058	0.054	0.064	0.053	0.054	0.061	0.019	0.005	0.050
ΔP(Th±SD)	0	0	-11.6±2.6	-12.2±22.5	-7.8±3.2	-18.1±5.6	0	0	-2.0±1.9	0	-49.5±1.3	-10.6±0.8

1098

https://doi.org/10.5194/egusphere-2024-233
Preprint. Discussion started: 13 February 2024

Table 3. Mean (±SD), depth-integrated rates of bio- and photodegradation (mg C $L^{-1}d^{-1}$)

Object	$V_{Biodegradation}$	$V_{Photodegradation}$
Lake Temnoe		
Forest Lake (Jun)	-0.02±0.0014	-0.19±0.03
Forest Lake (Aug)	-0.031±0.010	-0.067±0.066
Forest Lake (Oct)	-0.042±0.013	0
Ilasskoe Bog continuum (July)		
Piezometer water	-0.17±0.09	-0.33±0.07
Peatland pool	-0.029±0.008	0
Outlet stream (Chernyi)	-0.055±0.043	-0.27±0.043