

1 Seasonal and spatial pattern of dissolved organic matter

2 bio- and photodegradation in boreal humic waters

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4 Artem V. Chupakov<sup>1</sup>, Natalia V. Neverova<sup>1</sup>, Anna A. Chupakova<sup>1</sup>, Svetlana A. Zabelina<sup>1</sup>,  
5 Liudmila S. Shirokova<sup>1,2</sup>, Taissia Ya. Vorobyeva<sup>1</sup>, Oleg S. Pokrovsky<sup>2,3\*</sup>

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7 <sup>1</sup> Institute of Ecological Problems of the North, N. Laverov Federal Center for Integrated Arctic  
8 Research, Nab Severnoi Dviny 23, Arkhangelsk 163000, Russia

9 <sup>2</sup> Geoscience and Environment Toulouse, UMR 5563 CNRS, University of Toulouse, 14 Avenue  
10 Edouard Belin, Toulouse 31400, France

11 <sup>3</sup> BIO-GEO-CLIM Laboratory, Tomsk State University, 35 Lenina Pr., Tomsk 634050, Russia

12  
13 \*corresponding author email: [oleg.pokrovsky@get.omp.eu](mailto:oleg.pokrovsky@get.omp.eu)

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16 Synopsis:

17  
18 In boreal humic waters of a forest lake and a bog, the rate of dissolved organic matter  
19 photodegradation is four times higher than that of biodegradation. However, given the shallow  
20 (0.5 m) light-penetrating layer, the biodegradation provides the largest contribution to CO<sub>2</sub>  
21 emission from water surfaces. A few trace metals were partially removed (1-10 %) during photo-  
22 and biodegradation, via precipitation of Fe(III) hydroxides after destabilization of organo-ferric  
23 colloids and organic complexes

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33 **Abstract**

34 Studying competitive effects of microbial and light-induced transformation of dissolved organic  
35 matter (DOM) and trace metals is crucially important for understanding the factors controlling  
36 aquatic carbon (C), micronutrients and toxicants transformation in boreal waters. Here we  
37 determined the bio- and photo-degradability of DOM and its effect on the behavior of dissolved  
38 trace metals in humic surface waters from the European subarctic: an ombrotrophic peatbog  
39 continuum (subsurface water – peatland pool – stream) and a stratified forest lake across seasons.  
40 Along the bog water continuum, biodegradation rate was the highest in subsurface waters  
41 collected via piezometer and the lowest in the acidic peatland pool. Photodegradation was similar  
42 for subsurface waters and the stream, but was not detectable in the peatland pool. The waters of  
43 forest lake exhibited a strong seasonal effect of biodegradation, which was the highest in October  
44 and the lowest in June. Overall, the biodegradation was capable of removing between 1 and 7 %  
45 of initial DOC, being the highest in the forest lake in October and in peatland pool in summer.  
46 The photolysis was capable of degrading a much higher proportion of the initial DOC (10-25 %),  
47 especially in the forest lake during June and the bog stream during July. Only a few trace metals  
48 (TM) were sizably affected by both photo- and biodegradation of DOM (Fe, Al, Ti, Nb and light  
49 REE), whereas V, Mn, Co, Cu and Ba were affected solely by biodegradation. A likely  
50 mechanism of metal removal was their coprecipitation with coagulating Fe(III) hydroxides.  
51 Compared to typical CO<sub>2</sub> emissions from inland waters of the region, biodegradation of DOM  
52 can provide the totality of CO<sub>2</sub> evasion from lake water surfaces whereas bio- and  
53 photodegradation are not sufficient to explain the observed CO<sub>2</sub> fluxes in bog water continuum.  
54 Overall, these results demonstrated strong spatial and seasonal variability in bio- and  
55 photodegradation of DOM and organic TM complexes, and call for the need of a systematic  
56 assessment of both processes across seasons with high spatial resolution.

57

58           **1. Introduction**

59           Organic Carbon (OC) processing via metabolic biological (heterotrophic bacteria uptake  
60 and respiration) and abiotic physico-chemical (photolysis) pathways is considered to be one of  
61 the major source of CO<sub>2</sub> supersaturation in surface waters and related C emissions (Lapierre et  
62 al., 2013; Tranvik et al., 2009), photo vs. biodegradation of Dissolved Organic Matter (DOM)  
63 remains poorly quantified (e.g. Groeneveld et al., 2019; Shirokova et al., 2021; Raudina et al.,  
64 2022). Given sizable CO<sub>2</sub> emissions in boreal and subarctic waters (Karlsson et al., 2021),  
65 together with high concentrations of DOC (Cole et al., 2007; Vonk et al., 2015), and fast ongoing  
66 and predicted environmental changes in high latitude aquatic and terrestrial ecosystems (Wauthy  
67 et al., 2018; Chaudhary et al., 2020; Harris et al., 2022), the surface waters of subarctic regions  
68 are at the forefront of studies on the biogeochemical cycle of C. Although CO<sub>2</sub> emissions from  
69 these waters are significantly lower than those in the 10 °S – 10 °N equatorial belt (e.g., Borges  
70 et al., 2015), the magnitude of possible changes in C flux from northern waters to the atmosphere  
71 remains much less known. Further, there are still important geographical biases linked to  
72 insufficient knowledge of rates and mechanisms of DOC transformation in certain regions. An  
73 example is wetland-dominated northern aquatic settings, where high concentrations of soil  
74 organic C surrounding the bogs provide high concentrations of DOC but also some related trace  
75 metals, whose concentration and migration can be strongly controlled by processes of DOM  
76 transformation.

77           Thorough laboratory and field work on DOM bio- and photolability conducted over the  
78 past decades have demonstrated both phenomena are important, and, depending on  
79 environmental setting (nutrient regime, photic layer depth, nature of DOM, etc.), one or the other  
80 may dominate overall DOM removal in surface waters (Vachon et al., 2016, 2017; Vähätalo and  
81 Wetzel, 2008; Obernosterer and Benner, 2004). In addition to DOM, trace elements (limiting  
82 micronutrients, toxicants and geochemical tracers) present in the form of organic and organo-

83 mineral (Fe, Al) colloids and metal – organic complexes may be subjected to strong  
84 transformations during microbiological and photolytic degradations of DOM. This in turn may  
85 impact the bioavailability, toxicity and export fluxes of trace metals from terrestrial to continental  
86 aquatic and finally coastal environments.

87         Recently, specific attention was devoted to the aquatic systems of permafrost peatlands  
88 given their high vulnerability to climate warming and huge potential for release of soil organic  
89 C to surface waters (Vonk et al., 2015; Shirokova et al., 2019; Payandi-Rolland et al., 2020;  
90 Prijac et al., 2022; Rosset et al., 2022; Taillardet et al., 2022). These studies provided a range of  
91 DOM susceptibility to biotic degradation. Thus, between 10 and 40 % of the DOC in lakes, rivers  
92 and soil waters of the boreal zone may be available for bacterial uptake over a time frame of  
93 several weeks (Berggren et al., 2010; Roehm et al., 2009). This range is consistent with 14-16%  
94 of biodegradable DOC (BDOC) assessed globally (Begum et al. 2022). The necessity for further  
95 studies was also indicated, most notably with regard to *i*) seasonal aspects, given that the  
96 overwhelming majority of available studies were performed during Arctic summer (see  
97 discussions in Vonk et al., 2015; Laurion et al., 2021), and *ii*) increased spatial resolution, given  
98 that sizable variations of BDOC can be observed within quite short distances of a hydrological  
99 continuum (Payandi-Rolland et al., 2020; Raudina et al., 2022). Another poorly known aspect is  
100 DOM photo- and biolability across the depth of the water column, especially in seasonally  
101 stratified lakes which are subject to spring and autumn overturn.

102         Based on a compilation of available studies on BDOC and their own research, Vonk et  
103 al. (2015) argued there is a negligible amount of biodegradable DOC in aquatic systems without  
104 permafrost. This is, however, contradictory to available assessments on biodegradation of aquatic  
105 DOM as major driver of CO<sub>2</sub> emission in general (Amaral et al., 2021; Liu and Wang, 2022) and  
106 in boreal waters in particular (Ask et al., 2012; Lapierre et al., 2013). Furthermore, among all  
107 Arctic rivers, the highest annual (20%) and winter (ca. 45%) BDOC was reported for the Ob

108 River, which drains through peatlands with minimal permafrost influence (Wickland et al.,  
109 2012). These non-exhaustive examples illustrate certain inconsistency in current estimations of  
110 DOC biodegradability in surface organic-rich waters of high latitudes, which precludes  
111 quantitative modeling of future C fluxes between land, water and atmosphere in these  
112 environmentally important regions. Towards addressing these inconsistencies, in this study, we  
113 chose a typical hydrological continuum in a boreal ombrotrophic bog in a glacial lake-ridge  
114 complex that includes subsurface water, a small peatland pool in the central part of the bog and  
115 an outlet stream. Further, we selected a well-studied deep stratified humic lake in the same region  
116 (Lake Temnoe; Chupakov et al., 2017) where we sampled surface and deep horizons for the  
117 incubation experiments. The chosen waters represent subarctic non-permafrost regions that  
118 exhibit sizable organic C pool in their soils and high concentrations of DOC in their surface  
119 waters. In contrast to previous studies of permafrost peatlands (Shirokova et al., 2019; Laurion  
120 et al., 2021; Payandi-Rolland et al., 2020; Mazoyer et al., 2022) where the main source of DOM  
121 is peat or ground vegetation like mosses and lichens, in this highly productive southern taiga  
122 region, DOC may be more vulnerable to microbial activity due to the presence of forest leachates  
123 (i.e., Don and Kalbitz, 2005; Kalbitz et al., 2003; Kawahigashi et al., 2004; Kiikkilä et al., 2013)  
124 and much higher bioproductivity for both the terrestrial and aquatic parts of the lake-river  
125 ecosystems.

126         The first working hypothesis behind our study design is that the DOC-rich subsurface  
127 water and deep horizons of the humic lake are mostly sensitive to sunlight impact (Stubbins et  
128 al., 2010), and that maximal impact of photodegradation is expected during allochthonous  
129 aromatic DOM input (high surface inflow to lakes and bogs in June and October). In contrast,  
130 maximal biodegradation of DOM is expected during periods of possible phytoplankton bloom in  
131 August, when autochthonous organic material is generated in the water column. A broad  
132 importance of DOM bio- and photodegradation dynamics is that these processes can contribute

133 to CO<sub>2</sub> emissions from water surfaces thereby directly controlling the C cycling between the land  
134 and the atmosphere. Therefore, the first aim of this study was to relate the measured rates of  
135 DOM photo and biodegradation to CO<sub>2</sub> emissions observed in the studied water bodies.

136 A novelty of the present study is addressing trace metal (TM) partitioning during bio- and  
137 photodegradation. The link between DOM and TE is straightforward: in humic waters of  
138 peatlands, most TE (except probably some alkalis and oxyanions) are strongly (> 80%)  
139 associated to DOM in the form of organic and organo-mineral (Fe, Al) colloids (Pokrovsky et  
140 al., 2005, 2012, 2016). As a result, any DOM transformation processes may directly control the  
141 pattern of TE. From the other hand, some TE may be photosensitive (Mn, Fe), toxic (Al, Cu, As,  
142 Cd, Pb), or act as limiting micronutrients (Zn, Co, Ni, Mo) for the bacteria. Our second working  
143 hypothesis here is that removal of DOM via photo- or bio-degradation will change the  
144 partitioning of trace elements which are either *i*) strongly bound to DOM, such as divalent  
145 transition metals, or *ii*) incorporated into organo-mineral (Fe, Al) colloids, such as trivalent and  
146 tetravalent hydrolyses. The TE of 1<sup>st</sup> group might either remain in solution (during  
147 photodegradation), hence not modifying their total dissolved concentration, or being taken up by  
148 growing bacteria during bio-degradation of TE-bound organic matter (Shirokova et al., 2017a,  
149 c). The elements of the second group are capable of co-precipitating with Fe and Al hydroxides  
150 hence being scavenged from the aqueous solution. (e.g., Kopacek et al., 2005, 2006). To test  
151 these hypotheses, we examined DOM and related trace metals bio- and photodegradability  
152 aiming to assess 1) spatial variations along a hydrological continuum of non-permafrost peatland  
153 and different horizons of a neighboring deep stratified lake located in the forest, and 2) temporal  
154 variability during 3 main hydrological seasons (high flow in June, baseflow in August and  
155 autumn rain season in October) in the forest lake. Achieving these objectives should allow  
156 quantifying the relative share of bio- and photodegradation on overall DOC and TM removal  
157 from surface waters via biotic and physico-chemical mechanisms.

158 **2. Materials and Methods**

159 *2.1. Natural settings of subarctic bog and stratified lake*

160 The study site is in the NE part of the European boreal zone (Arkhangelsk region), **Fig.**  
161 **1**. The mean annual air temperature is 0 °C and average annual precipitation is 700 ± 50 mm.  
162 The pristine ombrotrophic Ilasskoe Bog is located 30 km SE of Arkhangelsk, and is a typical  
163 lake-ridge complex formed from the last glaciation approximately 10,000 years ago. Its total  
164 surface area is 89 km<sup>2</sup>, with an average peat thickness of 3 m. The hydrological continuum of the  
165 Ilasskoe Bog includes subsurface water collected via piezometer (2-2.5 m depth), a small lake  
166 (Severnoe) and a stream outlet (**Fig. 1**). Lake Severnoe, located in the central part of the bog, is  
167 a typical peatland pool with an average depth of 1.5 m and a surface area of 0.013 km<sup>2</sup>. The  
168 Chernyi Stream is an outlet for the eastern part of the bog. The stream is 0.7-2.0 m wide, 10 km  
169 long and it flows in a forested (taiga) zone in the shade of tree canopy. The waters of the Ilasskoe  
170 Bog are acidic (pH ranges from 3.9-4.0 in piezometer and peatland pool to 5.7 in stream Chernyi),  
171 organic-rich (DOC is equal to 88, 13 and 38 mg L<sup>-1</sup> in the piezometer, lake and stream,  
172 accordingly) and low mineralized (Electrical Conductivity is 17-46 μS cm<sup>-1</sup>), as listed in **Table**  
173 **1**.

174 Lake Temnoe is located in a pristine taiga forest 100 km NNE of the town of Arkhangelsk,  
175 an area that does not receive any direct anthropogenic impact (**Fig. 1**). The watershed area is 3.08  
176 km<sup>2</sup> and the lake surface area is 0.091 km<sup>2</sup>, with a maximum depth of 37 m and a Secchi disk  
177 depth of 3.5±0.5 m. The water residence time in the lake is 394 days. Bogs constitute 31% of  
178 lake's watershed area, which is represented by carbonate-free loamy moraine atop the peat,  
179 podzol and gley soils. The lake water is slightly acidic (pH = 5.1 to 6.0), humic (DOC = 13-20  
180 mg L<sup>-1</sup>) and dominated by allochthonous DOM with a low concentration of total dissolved ions  
181 (Electrical Conductivity of 20 μS cm<sup>-1</sup>). Similar to other deep boreal and subarctic lakes, the lake  
182 exhibits 2 main periods of pronounced stratification (November to April and June to September)

183 and two periods of lake overturn (October and May). Maximal winter stratification occurs in  
184 March; the highest water temperature typically occurs in July (see Chupakov et al., 2017 for  
185 details). The upper 0 – 10 m water layer (epilimnion) is not stratified in the course of the year.

186 The surface waters were collected from the shore (peatland pool and stream) or a PVC  
187 boat (Lake Temnoe). Surface (30-50 cm depth) waters were sampled in the Ilasskoe bog and 3  
188 water horizons (0.5, 5 and 10 m) were sampled in the Temnoe Lake using a pre-cleaned  
189 polycarbonate horizontal water sampler (Aquatic Research Co, ID, USA). The water samples  
190 were placed into 2-L Milli-Q pre-cleaned PVC jars and kept refrigerated (4 °C) until arrival at  
191 the laboratory within 2-3 hours of collection.

192

## 193 *2.2. Experiments*

### 194 2.2.1. Biodegradation

195 For DOM biodegradation assessments, we followed the recommended protocol and used  
196 the appropriate type of labware for assessing biodegradable DOC of Arctic waters without  
197 external nutrient addition (Vonk et al., 2015; Payandi-Rolland et al., 2020) and applied a slight  
198 modification from Shirokova et al. (2019) to assess maximal possible biodegradation. Initial  
199 water samples brought to the laboratory within 2-3 hours after sampling were filtered through 3  
200 µm sterilized Nylon Sartorius membranes (47 mm diameter); these were used because  
201 ‘conventional’ 0.8-1.2 µm (GF/F) filtration membranes might remove too many microbial cells  
202 (Dean et al., 2018).

203 Duplicate 30 mL aliquots of 3 µm-filtered water were placed into pre-combusted (4.5  
204 hours at 450°C) dark borosilicate 40 mL glass bottles wrapped in Al foil to prevent any  
205 photolysis, without nutrient amendment and incubated at 22±1°C in the dark. The bottles were  
206 closed with loosened sterilized PVC caps to allow air exchange. The bottles were shaken  
207 manually once a day avoiding the liquid touching the cap. The entire reactor was used for



208 sampling after 0, 2, 5, 8, 12, and 21 days of exposure. Sampled solutions were filtered through  
209 sterile, MilliQ-cleaned Sartorius 0.22  $\mu\text{m}$  filters. The DOC blanks for these filters did not exceed  
210 1% of DOC concentrations in experimental samples. Sterilized control reactors were filled with  
211 natural water that was filtered through a 0.22  $\mu\text{m}$  sterile filter and incubated together with  
212 experimental reactors following the approach of Köhler et al (2002). They were re-filtered  
213 through 0.22  $\mu\text{m}$  filters the day of sampling. All handling and sampling of bottles was performed  
214 in the laminar hood box (class A100) in a sterilized workspace. Filtered samples were acidified  
215 with 30  $\mu\text{L}$  of concentrated (8.1 M) double distilled HCl, tightly capped and stored in the  
216 refrigerator before DOC analyses. The non-acidified portion of filtrate was used for pH, Specific  
217 Conductivity, DIC and UV<sub>254 nm</sub> and optical spectra measurement.

218

### 219 2.2.2. Photodegradation

220 For photodegradation incubations, water samples were collected in Al-foil covered pre-  
221 cleaned polypropylene jars and sterile filtered (0.22  $\mu\text{m}$  Nalgene Rapid-Flow Sterile Systems)  
222 within 2 hours of sampling and refrigerated. The filtrates were transferred under laminar hood  
223 box into sterilized, acid-washed quartz tubes (150 mL volume, 20% air headspace) with silicate  
224 stoppers and placed at  $3 \pm 2$  cm depth into an outdoor pool which was filled by river water having  
225 the light transparency similar to that of the Ilaskoe and Temnoe lakes. The outdoor pools were  
226 placed in an unshaded area with a latitude similar to the sampling sites (< 30 km from Ilaskoe  
227 Bog and Temnoe Lake). Slight wind movement and regular manual shaking allowed for  
228 sufficient mixing of reactor interiors during exposure. All photodegradation experiments were  
229 run in duplicates. The water temperature (EBRO EBI 20) and light intensity (Luxmeter Testo  
230 545) were continuously recorded every 3 hours.

231 For DOM photodegradation experiments, we followed conventional methods requiring  
232 exposure of 0.2  $\mu\text{m}$ -sterile filtered samples in quartz reactors in the outdoor pool (Vähätalo et al.,

233 2003; Chupakova et al., 2018; Gareis and Lesack, 2018), solar simulator (Lou and Xie, 2006;  
234 Amado et al., 2014) or directly in the lake water (Laurion and Mladenov, 2013; Groeneveld et  
235 al., 2016). Note that the 0.22  $\mu\text{m}$  sterile filtration is the only way of conducting photodegradation  
236 experiments, given that autoclave sterilization of DOM-rich natural waters would coagulate  
237 humic material and thereby would not be suitable (Andersson et al., 2018). Filtration through a  
238 smaller pore size (i.e.,  $< 0.025 \mu\text{m}$ ), however, would decrease the concentration of DOC and  
239 trace metals (i.e., Ilina et al., 2014; Vasyukova et al., 2010). We have chosen a 16 day exposure  
240 time for logistical constraints, which is consistent with biodegradation experiments described  
241 above and with the duration used in previous studies on photodegradation under sunlight, from  
242 15 to 70 days (Moran et al., 2000; Vähätalo and Wetzel, 2004; Mostofa et al., 2007; Chupakova  
243 et al., 2018). Dark control experiments were conducted also in duplicates, using sterilized glass  
244 tubes filled with sterile 0.22  $\mu\text{m}$ -filtered water, wrapped in Al foil and placed in the same outdoor  
245 pool as the experiments. The headspace (approx. 20% of total reaction volume) was similar in  
246 experimental and control reactors. The individual reactors were sterile sampled at the beginning  
247 and after the 0, 2, 5, 8, 12, and 16 days of exposure. The Milli-Q blanks were collected and  
248 processed to monitor for any potential sample contamination introduced by our filtration,  
249 incubation, handling and sampling procedures. The organic carbon blanks of the filtrates did not  
250 exceed 0.2 mg/L.

251

### 252 2.3. Analyses

253 The temperature, pH,  $\text{O}_2$  and specific conductivity in surface waters were measured in  
254 the field. The dissolved  $\text{CO}_2$  concentration in the studied bodies of water was measured in-situ  
255 using submersible Vaissala Carbocap® GM70 handheld carbon dioxide meter with GMP222  
256 probes (accuracy 1.5%; see Serikova et al. (2018, 2019) for methodological details). The  
257 diffusional  $\text{CO}_2$  flux was calculated using a wind-based model (Cole and Caraco, 1998) with  $k_{600}$

258  $=2.07+0.215 \times u_{10}^{1.7}$ , where  $u_{10}$  is the wind speed at 10 m height, following the approaches  
259 developed for surface waters of peatlands (Zabelina et al., 2021). The DOC was analyzed by  
260 high-temperature catalytic oxidation using a Shimadzu® TOC-VCSN (uncertainty  $\pm 2\%$ , 0.1 mg  
261  $L^{-1}$  detection limit) in acidified samples after sparging it with C-free air for 3 min at 100 mL  $min^{-1}$   
262 as non-purgable organic carbon (NPOC). Internationally certified water samples  
263 (MISSISSIPPI-03 and Pérade-20) were used to check validity and reproducibility of the analysis.

264 The UV- and visual absorbance of water samples was measured using a 10 mm quartz  
265 cuvette on a CARY-50 UV-vis spectrophotometer to assess the aromaticity of pore fluids via  
266 specific UV absorbance ( $SUVA_{254}$ ). In the filtrates, we measured optical density at 254 nm and  
267 at selected wavelengths (365, 436, 470, and 665 nm) as well as the entire UV-visible spectrum.  
268 The specific UV-absorbency ( $SUVA_{254}$ ,  $L\ mg^{-1}\ m^{-1}$ ) and  $E_{470}:E_{665}$  ratios are used as a proxy for  
269 degree of condensation of aromatic groups of DOM, or humification (Chin et al., 1994; Weishaar  
270 et al., 2003; Hur et al., 2006; Peacock et al., 2013). The ratio  $E_{254}:E_{436}$  is useful for evaluation  
271 of contributions of autochthonous (aquatic) DOM compared to terrestrial (soil) C (Hur et al., 2006;  
272 Ilina et al., 2014). The ratio  $E_{254}:E_{365}$  also allows approximating the mean molecular weight of  
273 DOM (Hiriart-Baer et al., 2008; Berggren et al., 2007). For better visualization of the differences  
274 in spectral parameters between experimental and control reactors, we calculated the difference  
275 ( $\Delta A$ ) between the absorbance of the photo- or bio-reactor and that of the control reactor at each  
276 sampling time.

277 Filtered samples collected from photodegradation experiments were acidified with  
278 ultrapure nitric acid and analyzed for major and TE following the procedures employed by  
279 Geoscience and Environment Toulouse Laboratory for analyses of boreal humic waters  
280 (Oleinikova et al., 2017, 2018). Major cations, Si, P and ~40 TE were measured with a  
281 quadrupole ICP-MS (Agilent 7500 ce) using In and Re as internal standards. The international  
282 geo-standard SLRS-6 (Riverine Water Reference Material for Trace Metals) was used to check

283 validity and reproducibility of analyses. Note that for both bio- and photodegradation  
284 experiments, ICP MS analyses were performed over 16 days of incubation time.

285 To check for possible microbial development in biodegradation experiments, we  
286 performed oligotrophic and eutrophic bacteria counts over the course of incubation, following  
287 the standard methodology used in biodegradation experiments of peat waters (Stutter et al., 2013)  
288 and also described previously (Shirokova et al., 2017b; Chupakova et al., 2018). Specifically,  
289 active bacteria number count (colony forming units, CFU mL<sup>-1</sup>) was performed using Petri dishes  
290 inoculation (0.1 to 1.0 mL of lake water in three replicates) performed in a laminar hood box  
291 immediately prior the experimental incubation start and upon each sampling. Samples were  
292 inoculated on Nutrient Agar (5 g L<sup>-1</sup> beef extract, 5 g L<sup>-1</sup> gelatine peptone, 15 g L<sup>-1</sup> bacteriological  
293 agar, pH=6.8±0.2 at 25 °C) to determine the total number of heterotrophic bacteria. Difco® agar  
294 (granulated powder, Lot No 6290083) inoculation was used to assess the number of oligotrophic  
295 bacteria. Inoculation of blanks was routinely performed to assure the absence of contamination  
296 from external environments.

297

#### 298 *2.4. Data treatment*

299 The bio- and photodegradable DOC and trace metals were calculated as percent loss  
300 relative to control similar to other studies (Vonk et al., 2015; Chupakova et al., 2018; Shirokova  
301 et al., 2017b, 2019). However, previous works in similar environmental contexts of high-DOC  
302 humic waters demonstrated that the effects of DOC and element decrease are rather low and often  
303 comparable to uncertainties of duplicates (Shirokova et al., 2019). To assess the net effect of bio-  
304 or photodestruction during the experiment, we used the integral values of concentration change,  
305 estimated as the difference between the experiment and the control, while taking into account the  
306 standard deviation of replicates. For this, we first calculated the mean of replicates at the *i*-th  
307 time of sampling for the experiment and the control of *X* component ( $^{mean}X_i$  and  $^{control}X_i$ ,

308 respectively). We next calculated the sum of mean concentration of replicates and its standard  
 309 deviation ( $^{mean}X_i+SD_i$ ). Thus, we obtained 3 values characterizing the bio- or photo-degradation  
 310 process: 1) the change of concentration in the experimental reactor ( $^{mean}S$ ), 2) the change of  
 311 concentration not linked to the studied process ( $^{control}S$ ), and 3) the maximal uncertainty of the  
 312 concentration change in the reactor ( $^{mean+SD}S$ ). This allowed calculating, in percentages, the  
 313 efficiency of bio or photodegradation of  $X$  component relative to the control, taken into account  
 314 relevant uncertainties as following:

$$315 \quad X (\%) = 100 \times (|^{mean}X| - |^{control}X|) / |^{control}X| \quad (1)$$

$$316 \quad SD (\%) = 100 \times (|^{mean+SD}X| - |^{mean}X|) / |^{control}X| \quad (2)$$

317 where  $X$  is biodegradable DOC or trace element (BDOC and BTE, respectively) or  
 318 photodegradable DOC and trace element (PDOC and PTE, respectively). The sign of  $X$   
 319 designates either a decrease (« $\leftarrow$ ») or an increase (« $\rightarrow$ ») of solute concentration during the  
 320 experiment. We considered the decrease of concentration significant when  $X (\%) > SD (\%)$ . In  
 321 other cases, the change was non-systematic over the course of experiment or non-measurable  
 322 using the experimental technique employed in the present study.

323 The mean rate of bio- or photodegradation of  $X$  component ( $V_X$ ) was calculated based on  
 324 the overall change ( $\Delta X$ , in %) between the initial ( $X_0$ ) and final value normalized to overall  
 325 duration of the experiment  $t$  (22 and 16 days for bio- and photodegradation, respectively):

$$326 \quad V_X = ((\Delta X / X_0) / t) \quad (3)$$

327 The SD for rates of component change were calculated in a similar way.

328 The spectral differences between experimental and control reactors were presented as X-  
 329 Y-Z diagrams where X is elapsed time, Y is wavelength, and Z is  $\Delta A$ . The data were plotted in  
 330 a Surfer software package using triangulation with a linear interpolation method. Statistical  
 331 treatment included the least squares method and the Pearson correlation, as the data were

332 normally distributed. All calculations were performed in STATISTICA ver. 10 (StatSoft  
333 Inc.,Tulsa) at  $p = 0.05$ ).

334

### 335 **3. Results**

#### 336 *3.1. Field measured C concentration and calculated CO<sub>2</sub> fluxes*

337 The DOC concentration ([DOC]) ranged from 13 to 21 mg L<sup>-1</sup> in Lake Temnoe,  
338 depending on depth and season. The CO<sub>2</sub> concentrations and fluxes increased from June to  
339 October and varied from 99 to 337 μmol L<sup>-1</sup> and 32 to 71 mmol CO<sub>2</sub> m<sup>-2</sup> d<sup>-1</sup>, respectively (**Table**  
340 **1**). In Ilasskoe Bog hydrological continuum, the DOC decreased from 88 mg L<sup>-1</sup> in the peat soil  
341 water to 38 mg L<sup>-1</sup> in the outlet stream. The DOC concentration was generally similar (within ±  
342 5 %) between 3, 0.8 (GFF), 0.45 and 0.22 μm pore size filtration of the initial sample, which is  
343 in agreement with former size fractionation measurements for Arctic and subarctic systems  
344 (Vasyukova et al., 2010; Pokrovsky et al., 2012, 2016, Shirokova et al., 2019). The waters of  
345 Ilasskoe Bog continuum exhibited CO<sub>2</sub> supersaturation with respect to atmosphere (from 55 to  
346 3300 μmol L<sup>-1</sup>) and calculated CO<sub>2</sub> emission (diffusion) flux ranging from 22 mmol CO<sub>2</sub> m<sup>-2</sup> d<sup>-1</sup>  
347 in the peatland pool to 1600 mmol CO<sub>2</sub> m<sup>-2</sup> d<sup>-1</sup> in the piezometer (**Table 1**).

348

#### 349 *3.2. Biodegradation of DOM*

##### 350 3.2.1. DOC concentration evolution in the experiments

351 In the Temnoe Lake, the range of [DOC] change during 2-3 week incubation in the  
352 experimental reactors did not exceed 2 mg L<sup>-1</sup> and remained within +0.5 to -1.5 mg L<sup>-1</sup>, which is  
353 less than 10% of the initial DOC amount (**Fig. 2 A and Fig. S1** of the Supplement). The  
354 biodegradable DOC was both season and depth dependent and ranged from 2 to 6 % (**Table 2**).  
355 The integral 2-week rates of biodegradation (**Table 3, Fig. 3 A**) demonstrated the highest values  
356 during autumn at depths of 0.5 m and 10 m and the lowest values during June at all depths. The

357 final 0-10 m water column- and season-averaged biodegradation rate in Lake Temnoe ranged  
358 from 0.02 to 0.04 mg DOC L<sup>-1</sup> d<sup>-1</sup>. Rates of bio-degradation in the 0-10 m layer demonstrated an  
359 increase from May to October, over the entire open-water period (**Fig. 4 A**).

360 For Ilaskoe Bog, the BDOC was highest in the peatland pool ( $4.9 \pm 1.4$  %) and lowest  
361 in the outlet stream ( $3.1 \pm 2.4$  %; **Fig. 2 A** and **Fig. S1**). The integral rate of DOC biodegradation  
362 followed the order 'piezometer >> stream > peatland pool' and ranged from 0.03 to 0.17 mg C  
363 L<sup>-1</sup> d<sup>-1</sup> (**Table 3, Fig. 3 A**).

364

### 365 3.2.2. Optical parameters of DOM

366 In Lake Temnoe, the SUVA<sub>254</sub> remained relatively constant (4.2 to 4.6 L mg C<sup>-1</sup> m<sup>-1</sup>)  
367 across seasons and depths (**Table 1 B**). Over the course of biodegradation, the SUVA<sub>254</sub> did not  
368 change significantly (i.e., less than 0.2 units, which is comparable to the variability of duplicates;  
369 **Fig. S2**). The ratio E<sub>254</sub>:E<sub>436</sub>, which is an indicator of humification, increased with incubation  
370 time in Lake Temnoe waters; the magnitude of this increase across depth followed the order "0.5  
371 m > 5 m > 10 m" (**Fig. S3**). The ratio E<sub>254</sub>/E<sub>365</sub> also increased over the course of biodegradation,  
372 corresponding to an increase of mean molecular weight of DOM (Hiriart-Baer et al., 2008;  
373 Berggren et al., 2007). The ratio E<sub>365</sub>/E<sub>470</sub> also demonstrated the strongest increase in surface  
374 horizons and virtually no change in the deepest horizon (**Fig. S3**). An increase in the ratio  
375 E<sub>470</sub>:E<sub>665</sub> corresponds to a decrease in the degree of aromaticity (humification). An increase in  
376 the ratio E<sub>254</sub>:E<sub>436</sub> signifies a decrease in contribution of autochthonous (aquatic) DOM compared  
377 to terrestrial (soil) C, whereas an increase in the E<sub>254</sub>:E<sub>365</sub> ratio characterizes removal of low  
378 molecular weights compounds.

379 In Ilaskoe Bog samples, the highest SUVA was observed in the water of the piezometer  
380 and the lowest in the stream, but the evolution of this parameter in the course of biodegradation  
381 was rather weak (**Fig. S3**). The E<sub>254</sub>:E<sub>365</sub> and E<sub>254</sub>:E<sub>436</sub> ratios increased with incubation time in

382 the piezometer and decreased with time in the stream (**Fig. S3**). The optical ratios ( $E_{254}:E_{436}$ ,  
383  $E_{365}:E_{470}$ ,  $E_{470}:E_{665}$ ) increased in the peatland pool, suggesting an increase in the molecular  
384 weight and an increase in the ratio of aromatic to aliphatic compounds.

385 Complete spectral differences between the experimental and control samples  
386 demonstrated rather weak ( $\Delta A \leq 0.04$ ) changes of spectral parameters, mostly detectable after  
387 10-12 days of incubation (**Fig. S4**). These results were generally consistent with the discrete  
388 spectral parameters presented above and demonstrated maximal effects in the piezometer and  
389 bog outlet stream. In Lake Temnoe, the maximal impact of biodegradation on spectral parameters  
390 was observed in June, at 0.5 m depth.

391

### 392 3.2.3. Bacterial number evolution during biodegradation experiments

393 The number of cultivable eutrophic bacteria (EB) sizably (ca., 2 orders of magnitude)  
394 increased during biodegradation of Lake Temnoe waters. However, this evolution was not  
395 systematic in the course of incubation; there was a pronounced decrease after 2 weeks of  
396 exposure in June and August and rather stable concentration in waters of all horizons sampled in  
397 October (**Fig. S5**). Such maxima in June and August might be linked to consumption of  
398 substrate/nutrient limitations on bacterial growth. In Ilasskoe Bog continuum, the number of  
399 eutrophic bacteria decreased by an order of magnitude in the peatland pool and piezometer while  
400 it remained constant in the stream. The number of oligotrophic bacteria (OB) increased in waters  
401 of all Lake Temnoe horizons by ca. 2 orders of magnitude in August and October and 1 order of  
402 magnitude in June. In contrast, the OB number did not change or slightly decreased during  
403 incubations of waters from Ilasskoe Bog continuum (**Fig. S5**).

404

405

406



#### 407 3.2.4 Trace element behavior

408           During biodegradation experiments, a number of trace metals [Group 1] demonstrated a  
409 significant ( $X > SD$ , Eqn. 1) decrease in concentration across the incubation period (**Table 2**):  
410 Al, Ti, Fe, Co, Cu, Ba, Nb, light REE (LREE) and Pb (as illustrated for Fe in **Fig. 5**) as well as  
411 Mn, V, and La (**Figs. S6, S7 and S8**, respectively). The most significant effects were observed  
412 for Fe in the 0-5 m horizon of Lake Temnoe (9 to 18 % in June, 6 to 13.5 % in August and 8 to  
413 9.5 % in October) and 14% in the peatland pool of Ilaskoe Bog. Overall, for most elements  
414 except Fe and Mn, this decrease was less pronounced than that of DOC; maximal effects were  
415 achieved for Lake Temnoe in August and October (V, Mn, Co, Cu, Ni, Nb, Hf, Pb and Th) and  
416 in June (Al and Ti). These elements are typically linked to DOM and Fe and present in the form  
417 of organic- and organo-mineral colloids. Second group of major and trace elements did not  
418 appreciably change their concentration ( $< 2$  % decrease): Li, B, Na, Mg, K, Ca, Si, Ge, As, Rb,  
419 Sr, Mo, Sb, Mo and Ba. These elements are not linked to colloids of Fe(III) hydroxide and organic  
420 matter. Finally, some elements [Group 3] exhibited unstable behavior without systematic change  
421 in concentration during the exposure ( $X < SD$ , Eqns. 1-2): Cr, Zn, Cu, Sr, Cd, (Y, Zr), Cs, Tl and  
422 U. These elements cannot be considered as significantly impacted by the biodegradation process  
423 in Lake Temnoe water.

424           In the Ilaskoe Bog hydrological continuum, the most significant changes during  
425 biodegradation were observed in the peatland pool and outlet stream. Elements strongly ( $> 5-10$   
426 %;  $X > S.D.$  in Eqn. 1) affected by biodegradation were organically / colloidally complexed V,  
427 Fe, Ni, Ga, Y, LREEs and Pb.

428

429

430

431

### 432 3.3. Photodegradation of DOM

#### 433 3.3.1. DOC concentration evolution

434 Compared to biodegradation, photodegradation demonstrated much higher values of  
435 PDOC and rates of reaction as well as higher variability among seasons and sites. In Lake  
436 Temnoe, the PDOC was the highest in June and the lowest in October (**Fig. 2 B** and **Table 2**).  
437 The maximal range of concentration change during 2-week period achieved 6-8 mg L<sup>-1</sup> (**Fig. S9**)  
438 which was 10 to 20 % of the initial [DOC] values. The rates strongly decreased from May-June  
439 to the end of summer – autumn. This is consistent with much higher solar radiation in June  
440 compared to August and October as measured on site (mean maximal daytime light intensity of  
441 5170±2760, 3220±2160, and 419±176 Lux, respectively). The depth integrated (0 to 10 m) rate  
442 of DOM photodegradation in Lake Temnoe ranged from 0 in October to 0.2 mg C L<sup>-1</sup> d<sup>-1</sup> in June  
443 (**Table 3; Fig. 4 B**).

444 In the Ilasskoe Bog hydrological continuum during July, the photodegradation rate  
445 followed the order “outlet stream > piezometer >> peatland pool” (**Fig. 3 B**), where integral rates  
446 equaled to 0.27±0.04, 0.33±0.07, and 0±0.05 mg C L<sup>-1</sup> d<sup>-1</sup>, respectively (**Table 3**).

447

#### 448 3.3.2. Optical parameters of DOM

449 Similar to the DOC concentration, the optical parameters of DOM more strongly evolved  
450 over the course of photodegradation compared to the biodegradation experiments. In the Temnoe  
451 Lake, the strongest decrease in SUVA<sub>254</sub> was observed in the waters of all horizons in June. This  
452 decrease was less pronounced in October (**Fig. S10**). The E<sub>254</sub>:E<sub>365</sub> ratio demonstrated a sizable  
453 increase in June, with much weaker increase in October. The E<sub>254</sub>:E<sub>436</sub> ratio strongly decreased  
454 with exposure time throughout all seasons (10 m depth) and only in June in the surface horizons  
455 (**Fig. S11**). An increase in the ratio E<sub>254</sub>:E<sub>365</sub> over the course of photodegradation corresponded  
456 to an increase in mean molecular weight of DOM. The ratios E<sub>365</sub>:E<sub>470</sub> and E<sub>470</sub>:E<sub>665</sub> decreased

457 in all experiments with the Temnoe Lake waters (**Fig. S11**), suggesting a decrease in the degree  
458 of humification (Battin, 1998) and a decrease in the ratio of aromatic to aliphatic moieties.

459 The  $SUVA_{254}$  in Ilasskoe Bog waters remained stable during photodegradation of stream  
460 waters and piezometer and strongly decreased in the peatland pool (**Fig. S10**). The  $E_{254}:E_{436}$  ratio  
461 strongly increased in the peatland pool and exhibited a decrease in stream waters and piezometer,  
462 whereas the  $E_{365}:E_{470}$  ratio systematically decreased in all photodegradation experiments with the  
463 Ilasskoe Bog continuum (**Fig. S11**). Finally, the  $E_{470}:E_{665}$  ratio exhibited sizable decrease, in the  
464 order 'stream >> pool  $\geq$  piezometer'. The total spectral differences between experimental and  
465 control reactors were mostly pronounced in stratified forest lake waters in June ( $\Delta A = -0.4$  to -  
466 0.4) and in the bog continuum in July, where effects were strongest in the piezometer and outlet  
467 stream waters ( $\Delta A$  value of  $-0.4$  (**Fig. S12**)).

468

### 469 3.3.3. TE in photodegradation experiments

470 The elements affected by photodegradation also formed three groups similar to those  
471 impacted by biodegradation. Concentrations of Al, Fe, trivalent and tetravalent hydrolysates (Ti,  
472 Ga, Zr, Y, LREE and Th) and Nb of [Group 1] significantly ( $> 2\%$ ;  $p < 0.05$ ) decreased during  
473 photolysis as illustrated for Fe in **Fig. 6**, and for Ti and Zr in **Figs. S13** and **S14**, respectively.  
474 The decrease of Fe was mostly pronounced in Lake Temnoe water from 10 m depth, whereas  
475 that of Ti and Zr was detectable for all horizons and seasons except in October. For the Ilasskoe  
476 Bog continuum, there was no systematic change in Fe concentration, whereas concentrations of  
477 Ti and Zr systematically decreased over the course of sunlight exposure (**Figs. S13, S14**). Alkali  
478 (Li, Rb), alkaline-earth metals (Mg, Ca, Sr, Ba), Si and oxyanions (As, Mo, Sb) of [Group 2]  
479 were weakly ( $< 2\%$ ) affected by photolysis. Finally, the remaining trace elements of [Group 3]  
480 did not exhibit any systematic evolution of concentration during exposure to sunlight, or these  
481 changes were inferior to the uncertainties of replicates ( $X < S.D.$  in Eqn. 1).

482 We found that, unlike for DOC, the magnitude of trace element concentration decrease  
483 during photodegradation was generally lower than that of biodegradation experiments. Overall,  
484 the strongest effects were observed for Ti (3 to 9% in Lake Temnoe; 20% in Ilasskoe Bog), Ga  
485 (6 to 14%), Zr (14-17% in Lake Temnoe), Nb (8 to 13%) and Th (8 to 19% in the Temnoe Lake  
486 and up to 50% in the Ilasskoe Bog). These effects were mostly pronounced in the Temnoe Lake  
487 in June and August and in peatland pool of the Ilasskoe Bog (July).

488

## 489 **4. Discussion**

### 490 *4.1. Comparison between biodegradation and photolysis*

491 The impact of season on the biodegradable DOC could be tested only for Lake Temnoe  
492 because it was sampled during the 3 main hydrological periods. The maximal biodegradation of  
493 the lake water was observed during autumn, when large amount of labile fresh soil OM and plant  
494 litter were delivered to the lake from the watershed via surface runoff. The water temperature  
495 seems to be of secondary importance for the intensity and rate of DOM biodegradation. This is  
496 also confirmed by lack of statistically significant (at  $p < 0.05$ ) correlation between water  
497 temperature and BDOC parameters (overall magnitude and rate). It is worth noting that the  
498 seasonal pattern of BDOC in the humic lake quantified in this study (**Fig. 4 A**) contrasted with  
499 previous works on biodegradation of large Arctic streams and rivers whose BDOC decreased as  
500 the Arctic summer progressed (Vonk et al., 2015). Presumably, the input of fresh plant litter from  
501 the forested watershed of Lake Temnoe provided elevated biodegradation in the water column at  
502 the end of the open water season. Another reason could be due to lake overturn in October and  
503 exposure of deep, partially autochthonous, and thus biodegradable, DOM to the surface horizons.  
504 A supply of limiting nutrients (N and P) to the upper 0-10 m layer during lake overturn could  
505 also promote such biodegradation in October.

506           The highest biodegradation rates in the uppermost sections of the bog hydrological  
507 continuum (piezometer, **Fig. 3 A**) are consistent with recent findings on organic-rich waters of  
508 permafrost peatlands (Shirokova et al., 2019; Payandi-Rolland et al., 2020) and earlier results on  
509 headwaters, small streams and soil leachates (Roehm et al., 2009; Ilina et al., 2014; Mann et al.,  
510 2014, 2015; Larouche et al., 2015; Spencer et al., 2015; Vonk et al., 2015; Moody et al., 2013;  
511 Pickard et al., 2017; Dean et al., 2019). This could be due to the very short water residence time  
512 and freshly leached DOM in these water objects (i.e., Mann et al., 2012; Abbott et al., 2014;  
513 Payandi-Rolland et al., 2020), given that bioavailable DOM components leached from plant litter  
514 are rapidly utilized (Textor et al., 2018). At the same time, overly low BDOC (2-8 %) values,  
515 regardless of depth and season in humic lake and across the hydrological continuum of the bog  
516 (**Fig. 2 A**), are supportive of previous results for permafrost peatlands from the neighboring  
517 region (Shirokova et al., 2019). A general path for DOM spectral properties modification over  
518 the course of biodegradation consisted of an increase in aromaticity of DOM due to preferential  
519 uptake of non-humic low molecular weight (LMW) compounds. However, this was not  
520 accompanied by a sizable increase in SUVA (**Fig. S2**). Presumably, the proportion of these  
521 compounds in the overall DOC level was quite low and could not impact SUVA evolution.  
522 Globally, the evolution of optical ratios was consistent with bacterial consumption of aliphatic  
523 LMW compounds and an increase in the overall aromaticity of DOM.

524           Concerning the seasonal variation of photodegradation in the deep humic lake, maximal  
525 effects were observed in June, at the highest solar radiation. These effects likely occurred due to  
526 fresh terrestrial organic matter that was leached from the watershed and then efficiently processed  
527 during Arctic summer. It should be noted that labile phenolic, carbohydrates, N-containing bases  
528 and smaller molecular weight compounds are abundant in litter leachates produced during initial  
529 decay stages (Kiikkilä et al., 2011, 2012, 2013; Hensgens et al., 2021). By July, most of the  
530 photodegradable DOM was already removed, and in October, the effects were much lower. This

531 was consistent with drastic decrease of sunlight intensity: 5170, 3200 and 420 Lux in June,  
532 August and October, as measured in this study. Therefore, photolabile DOM is delivered from  
533 the forested watershed to the lake essentially during surface flux, at high water flow. It is then  
534 quickly removed from the water column, which was especially seen in the 0.5 and 5 m horizons  
535 of Lake Temnoe. Although labile organic matter from litter fall was also delivered during autumn  
536 rain season, presumably, during this period, the conditions for photolysis (low temperature, short  
537 daytime period and insufficient light) were not as favorable as those in June or August.

538         Photodegradation of waters from the Ilasskoe Bog continuum demonstrated maximal  
539 rates in the piezometer (**Fig. 3 B**). During photolysis of humic water, a decrease in optical ratios  
540 ( $E_{365}:E_{470}$ ;  $E_{470}:E_{665}$ ) clearly indicated preferential degradation of humic aromatic compounds.  
541 The strong effect of photodegradation on DOM optical properties in the 500-650 nm region may  
542 be linked to decomposition of complex DOM into smaller molecules, whereas a decrease of  
543 absorbance in the 230-400 nm region (**Fig. S12**) indicates degradation of aromatic compounds,  
544 progressively increasing over insolation time. A recent study of DOM photolysis in humic-rich  
545 forested streams demonstrated that high aromatic material was photochemically converted into  
546 smaller non-fluorescent molecules (Wilske et al., 2020).

547         Results obtained on the more important role of photodegradation over biodegradation are  
548 generally consistent with earlier reports on the dominance of photolysis for DOM processing in  
549 Arctic waters within North America (Cory et al., 2014; Ward et al., 2017), the Canadian  
550 temperate zone (Winter et al., 2007; Porcal et al., 2013, 2014, 2015), and Swedish headwater  
551 catchments (Köhler et al., 2002). It is known that DOM photolysis mainly decreases the  
552 proportion of aromatic (colored) DOC and produces rather small ( $\leq 10\%$ ) change in bulk DOC  
553 concentration (Laurion and Mladenov, 2013; Koehler et al., 2014; Groeneveld et al., 2016;  
554 Oleinikova et al., 2017; Chupakova et al., 2018; Gareis and Lesack, 2018). The present study

555 corroborates these former findings across a much larger seasonal scale and spatial resolution of  
556 boreal surface waters.

557 As a further perspective of this work, one has to consider biodegradation of photolytically  
558 altered DOM given that photo-oxidation is known to transform molecular structures into more  
559 bioavailable forms (e.g., Cory and Kling, 2018; Sulzberger et al., 2019) thereby stimulating  
560 microbial growth under sunlight, as is known for other Arctic and subarctic settings (i.e.,  
561 Drozdova et al., 2020; Laurion et al., 2020).

562

#### 563 *4.2. Possible impact of microbial and photolytic processing on CO<sub>2</sub> emissions from* 564 *water surfaces*

565 A broad importance of DOM bio- and photodegradation dynamics is that these processes  
566 can contribute to CO<sub>2</sub> emissions from water surfaces thereby directly controlling the C cycling  
567 between the land and the atmosphere (Lapierre et al., 2013; Tranvik et al., 2009; Cory et al.,  
568 2014, 2018). In this study, we attempted to relate, for the first time for several diverse aquatic  
569 systems across seasons, experimentally measured rates of DOM degradation to in-situ measured  
570 CO<sub>2</sub> emissions. The integral rates of DOM bioprocessing in the water column of Lake Temnoe  
571 (**Table 3, Fig. 4 A**) allow quantifying the potential contribution of biodegradation to CO<sub>2</sub>  
572 production and emission. Assuming all biodegraded DOM is transformed into CO<sub>2</sub> and there is  
573 no biomass increase or sedimentation, a 1 m water layer of the lake can emit 1.7 mmol CO<sub>2</sub> m<sup>-2</sup>  
574 d<sup>-1</sup> in June and 3.3 mmol CO<sub>2</sub> m<sup>-2</sup> d<sup>-1</sup> in October. Note that across seasons, Lake Temnoe is not  
575 chemically stratified in the first 0 – 10 m water layer, which does not mix up with anoxic  
576 hypolimnion and is not subjected to the influence of sediment respiration (Chupakov et al., 2017).  
577 Therefore, integral flux from 10 m deep water layer amounts to 17 – 33 mmol CO<sub>2</sub> m<sup>-2</sup> d<sup>-1</sup> across  
578 the seasons. These values are comparable to typical values of CO<sub>2</sub> evasion from the surface of  
579 this lake during different seasons (30-70 mmol CO<sub>2</sub> m<sup>-2</sup> d<sup>-1</sup>; **Table 1 B**). For surface waters of

580 Ilasskoe Bog, maximal CO<sub>2</sub> production due to DOM biomineralization alone (**Table 3**) ranged  
581 from 5.0 mmol CO<sub>2</sub> m<sup>-2</sup> d<sup>-1</sup> for the peatland pool (2 m deep) to 2.5 mmol CO<sub>2</sub> m<sup>-2</sup> d<sup>-1</sup> for the  
582 outlet stream (0.5 m deep). However, in summer, the peatland pool and stream emitted 23 and  
583 150 mmol CO<sub>2</sub> m<sup>-2</sup> d<sup>-1</sup> (**Table 1 A**) which could not be sustained by DOM biodegradation.

584         The addition of photodegradation (assuming a photic layer depth of 3.5 m) to DOM  
585 bioprocessing in the water column of the Temnoe Lake during open water season can further  
586 increase potential CO<sub>2</sub> production in the water column. For the case of Ilasskoe Bog waters, the  
587 addition of photolytic degradation increases projected CO<sub>2</sub> emission from the outlet stream by a  
588 factor of 5, which is still below the actual CO<sub>2</sub> flux, whereas DOM photolysis has no impact on  
589 CO<sub>2</sub> emissions from the peatland pool. Note that, although the depth of sunlight processing in  
590 boreal waters is typically 0.8-1.0 m (Vähätalo et al., 2000; Koehler et al., 2014), a more recent  
591 study concluded that direct photomineralization of DOM in Arctic humic ponds could be limited  
592 to the first centimeters of the water column (Mazoyer et al., 2022). Furthermore, in typical DOM-  
593 rich Arctic waters, only half of sunlight-associated DOC losses is converted into CO<sub>2</sub> and the  
594 rest may be turned into particles through photoflocculation (e.g., Mazoyer et al., 2022).  
595 Therefore, despite a faster photodegradation rate compared to biodegradation, due to the shallow  
596 photic layer in humic waters, the biodegradation may provide the largest impact on CO<sub>2</sub> emission  
597 from the water column of boreal waters.

598         At the same time, our assumption that all CO<sub>2</sub> in lake water is produced by bio- or  
599 photodegradation of DOM might not be warranted because there are multiple sources of CO<sub>2</sub> in  
600 the lake waters, which were not assessed in the present study. These including but not limited to:  
601 particulate organic matter bio- and photodegradation, whose importance can strongly exceed that  
602 of DOC (e.g., Attermeyer et al., 2018; Lau et al., 2021; Keskitalo et al., 2022), sediment  
603 respiration, plankton and periphyton diel photosynthetic cycle, underground water discharge at  
604 the lake bottom, and delivery of DOC and CO<sub>2</sub>-rich waters via lateral surface and shallow



605 subsurface influx. Given that the contribution of each CO<sub>2</sub> source can vary among different water  
606 bodies and across seasons, the assessment of DOM bio- and photodegradation contribution to  
607 overall CO<sub>2</sub> flux in this study should be considered as highly conservative.

608

#### 609 *4.3. Impact of DOM bio- and photo transformation on trace element pattern*

610 In this study we hypothesized the following link between DOC and TE: in humic surface  
611 waters of peatlands, most TE, which include divalent transition metals (Cu, Ni, Co, Zn, Mn),  
612 toxicants (Be, Cr, Cd, Pb), trivalent and tetravalent hydrolysates (Al, Ga, Y, REE, Ti, Zr, Hf,  
613 Th), with an exception of some alkalis and oxyanions, are strongly (> 80%) associated to DOM  
614 in the form of organic, organo-ferric and organo-aluminium colloids (Pokrovsky et al., 2012,  
615 2016). As a result, any DOM transformation processes, be it bio- or photo-degradation, may  
616 directly affect the concentration pattern of TE. Specifically, the DOM removal via photo- or bio-  
617 degradation should change the speciation of those elements, that are strongly bound to DOM  
618 such as divalent transition metals, or incorporated into organo-mineral (Fe, Al) colloids, such as  
619 trivalent and tetravalent hydrolysates (TE<sup>3+</sup>, TE<sup>4+</sup>). The former might either remain in solution  
620 (during photodegradation), hence not modifying their total dissolved concentration, or being  
621 taken up by growing bacteria during bio-degradation. The latter (TE<sup>3+</sup>, TE<sup>4+</sup>) are capable of co-  
622 precipitating with Fe and Al hydroxides, especially during photodegradation (i.e., Kopacek et al.,  
623 2005, 2006), hence being sizably removed from the aqueous solution. From the other hand, some  
624 TE are known to be photosensitive (Mn, Fe), toxic (Al, Cu, As, Cd, Pb), or potentially limiting  
625 micronutrients (Zn, Co, Ni, Mo) for the bacteria and therefore they are capable affecting the  
626 overall rate of photo- or bio-degradation.

627 However, contrary to our expectations, among all major and trace elements measured in  
628 the experiments, only trivalent and tetravalent hydrolysates (TE<sup>3+</sup>, TE<sup>4+</sup>) were sizably impacted  
629 by both photo- and biodegradation. It is known that these elements are essentially present in the

630 form of large molecular size, highly polymerized and presumably aromatic, organo-Fe/Al  
631 colloids in humic boreal/subarctic lakes (Pokrovsky et al., 2012, 2016), rivers (Krickov et al.,  
632 2019; Pokrovsky et al., 2010), and soil porewaters (Pokrovsky et al., 2005; Raudina et al., 2021).  
633 Therefore, insoluble  $TE^{3+}$  and  $TE^{4+}$  generally followed the removal of Fe(III) in the form of  
634 particulate Fe hydroxides, after breaking the Fe-DOM bonds that stabilized colloidal Fe(III)  
635 hydroxides. This destabilization and Fe hydroxide particle formation is known to occur either via  
636 biodegradation (i.e., Oleinikova et al., 2018) or photolysis (Kopacek et al., 2005, 2006;  
637 Oleinikova et al., 2017; Chupakova et al., 2018). At the same time, some micronutrients (V, Mn,  
638 Co, Cu and Ba) were affected solely by biodegradation. This can reflect uptake of these metals  
639 by growing bacterial cells, as is known from laboratory experiments with pure cultures of  
640 heterotrophic bacteria (Shirokova et al., 2017a).

641 Note that the effects of bio- and photodegradation were more pronounced for light REE  
642 (LREE) compared to heavy REE (HREE). This result is consistent with the fact that LREE have  
643 stronger association with Fe hydroxide compared to organic complexes, as known from general  
644 chemical considerations and laboratory experiments (i.e., Bau, 1999) and evidenced in various  
645 boreal and subarctic settings (Pokrovsky et al., 2016; Krickov et al., 2019). Given that the main  
646 effect of both photolysis and biodegradation of DOM in humic Fe(III)-rich surface waters is  
647 coagulation of dissolved Fe(III) in the form of Fe oxy(hydr)oxides, the LREE are removed from  
648 solution. This removal occurs in the form of adsorbed complexes or coprecipitated with Fe  
649 oxy(hydr)oxides, while HREE remain in the form of strong aqueous complexes.

650

## 651 **Conclusions**

652 Seasonally resolved bio- and photo-degradability of DOM in a deep stratified lake and  
653 summer measurements from a peat bog's hydrological continuum within the boreal zone  
654 demonstrated that the subsurface and deep horizons of these stratified waters are mostly sensitive

655 to sunlight impact, and that maximal effects of photodegradation occurred in June, during  
656 strongest insolation. In contrast, the biodegradation of DOM from the humic lake was mostly  
657 pronounced during October, when fresh leachates of forest litter were exported from the  
658 watershed. Insoluble, low-mobility trace metals such as trivalent and tetravalent hydrolysates  
659 were affected by both bio- and photodegradation, as they are associated with coagulating Fe(III)  
660 oxyhydroxides.

661 A broad implication of obtained results is that, although DOM photodegradation rates  
662 were sizably higher compared to those of biodegradation, the rather thin photic layer in humic  
663 waters does not allow for significant contribution of photolysis in overall CO<sub>2</sub> emission from  
664 lake and bog surfaces. Further work is needed on biodegradation of photolytically altered DOM  
665 given that photo-oxidation is known to transform molecular structures into more bioavailable  
666 forms. The high seasonal dynamics and spatial variability in both photo- and biodegradability of  
667 DOM and related trace elements of humic surface waters in the boreal zone encountered in this  
668 study suggest the need for studying these processes during “shoulder seasons” (early spring and  
669 late autumn), the periods of maximal photo- and biodegradation, respectively. These efforts  
670 should be focused on the most dynamic components such as small streams and subsurface waters,  
671 which demonstrated the highest rates of both photo- and biodegradation.

672

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678

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680

681 **Authors contribution.**

682 AVC and OP designed the study and wrote the paper; AC, NN and SB performed sampling,  
683 analysis and their interpretation; LS performed bacterial number assessment and DOC results  
684 interpretation; AVC, TV and OP provided analyses of literature data.

#### 685 **Competing interests.**

686 The authors declare that they have no conflict of interest.

687

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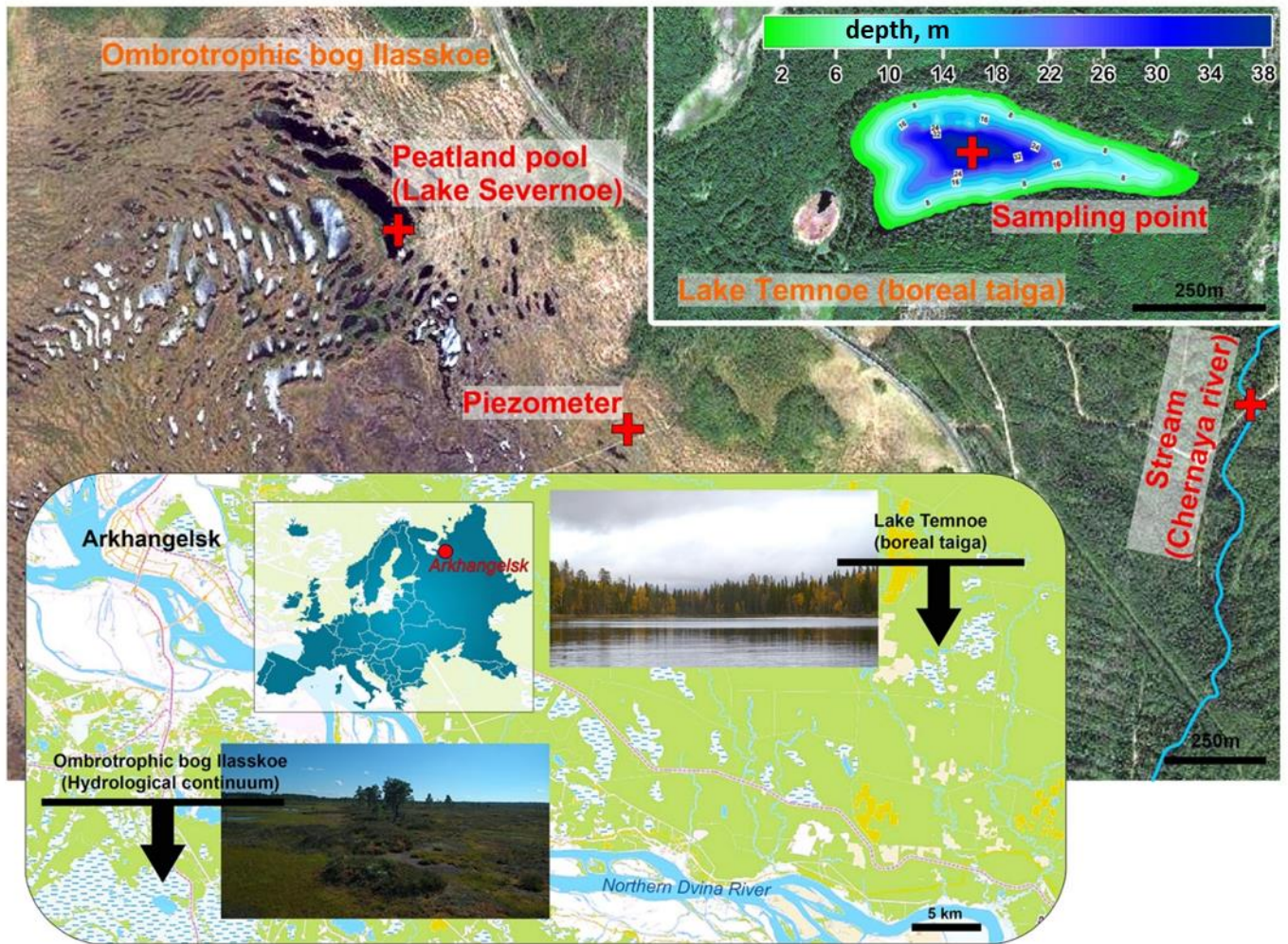
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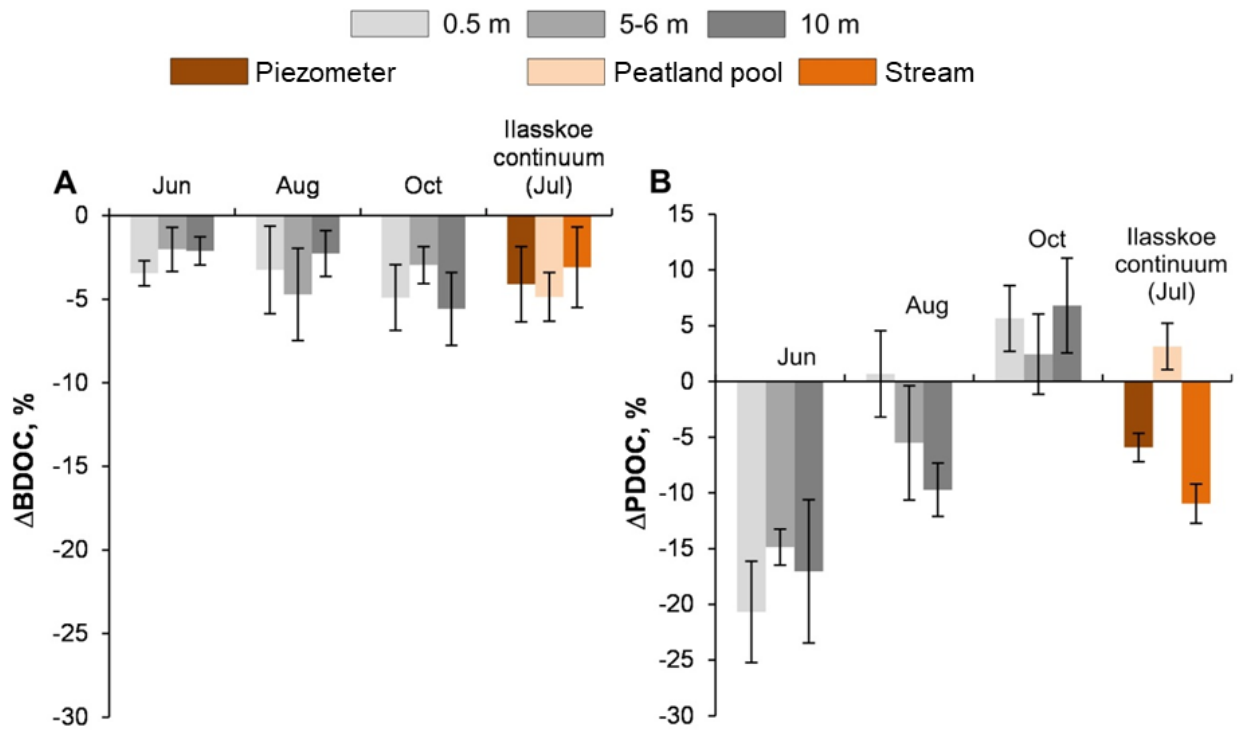
1062 **Fig. 1.** Geographical location of studied hydrological continuum for Ilasskoe Bog waters and  
1063 deep stratified Lake Temnoe in the boreal forest. Photo and map credits of Chupakov A.V.

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1073 **Fig. 2.** Percentage of bio- (A) and photo- (B) degradable DOC presented as relative decrease in  
 1074 DOC concentration between the initial and final value for the Temnoe Lake (June, August and  
 1075 October) and Ilasskoe Bog surface waters (July). Error bars are 1 s.d. of duplicates relative to  
 1076 the control (see Eqn. 1-2 in the text). In accord with unified protocol of biodegradation  
 1077 experiments (Vonk et al., 2015), positive values signify nil photodegradation (experimental  
 1078 artifacts of DOC production).

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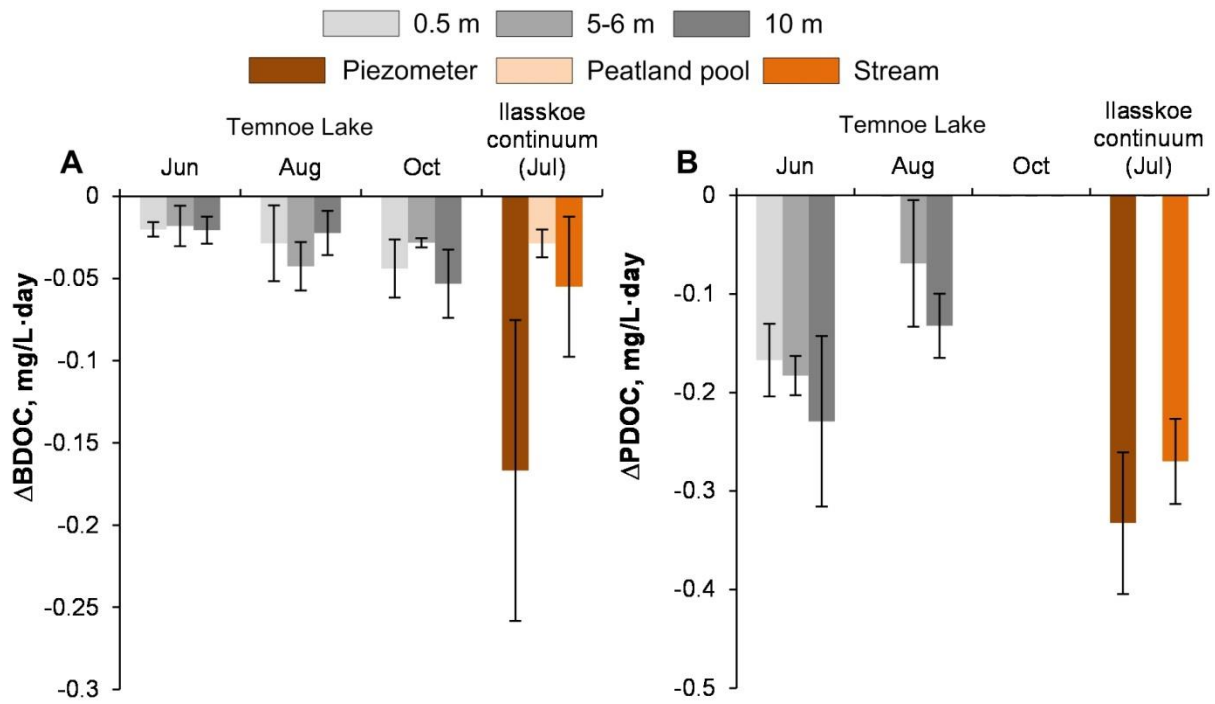
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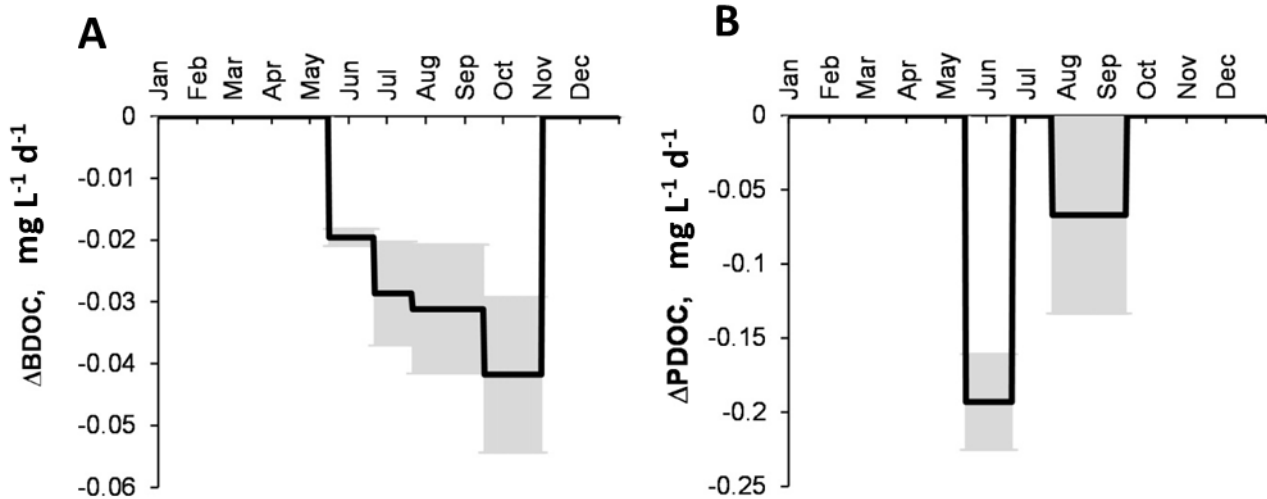
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1097 **Fig. 3.** Rates of DOC bio- (A) and photo- (B) degradation. The values are negative because  
1098 they represent a decrease in DOC concentration over the course of the experiment.

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1106 **Fig. 4.** Integral rates of bio- ( $\Delta\text{BDOC}$ , **A**) and photo- ( $\Delta\text{PDOC}$ , **B**) degradation in the 0-10 m  
 1107 layer of Lake Temnoe across the entire open-water period (May to October). Rate values are  
 1108 negative because they signify a decrease in DOC concentration. Note that there was no  
 1109 sampling from December to April and the photodegradation was not studied in July.  
 1110 Uncertainties are represented by gray shaded rectangles.

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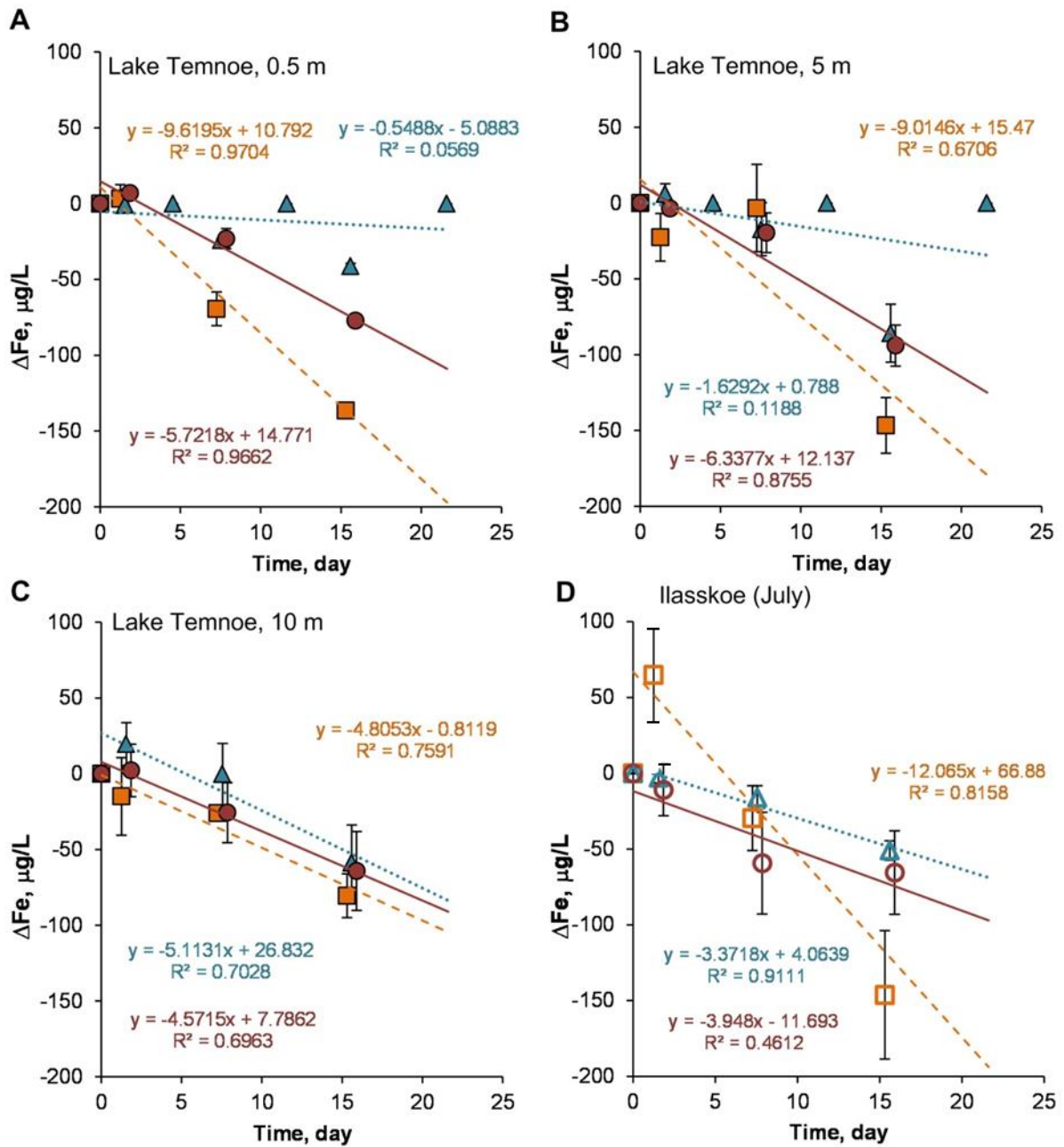
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1121 ■ June ▲ August ● October □ Piezometer △ Peatland pool ○ Stream

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1123 **Fig. 5.** Change in Fe concentration (relative to control) over time in biodegradation  
 1124 experiments. Error bars are 1 s.d. of duplicates. Temnoe Lake 0.5 m (A), 5 m (B) and 10 m (C)  
 1125 in June (squares), August (triangles) and October (circles). Ilasskoe Bog continuum in July (D)  
 1126 including piezometer (squares), Severnoe peatland pool (triangles) and stream Chernyi  
 1127 (circles).

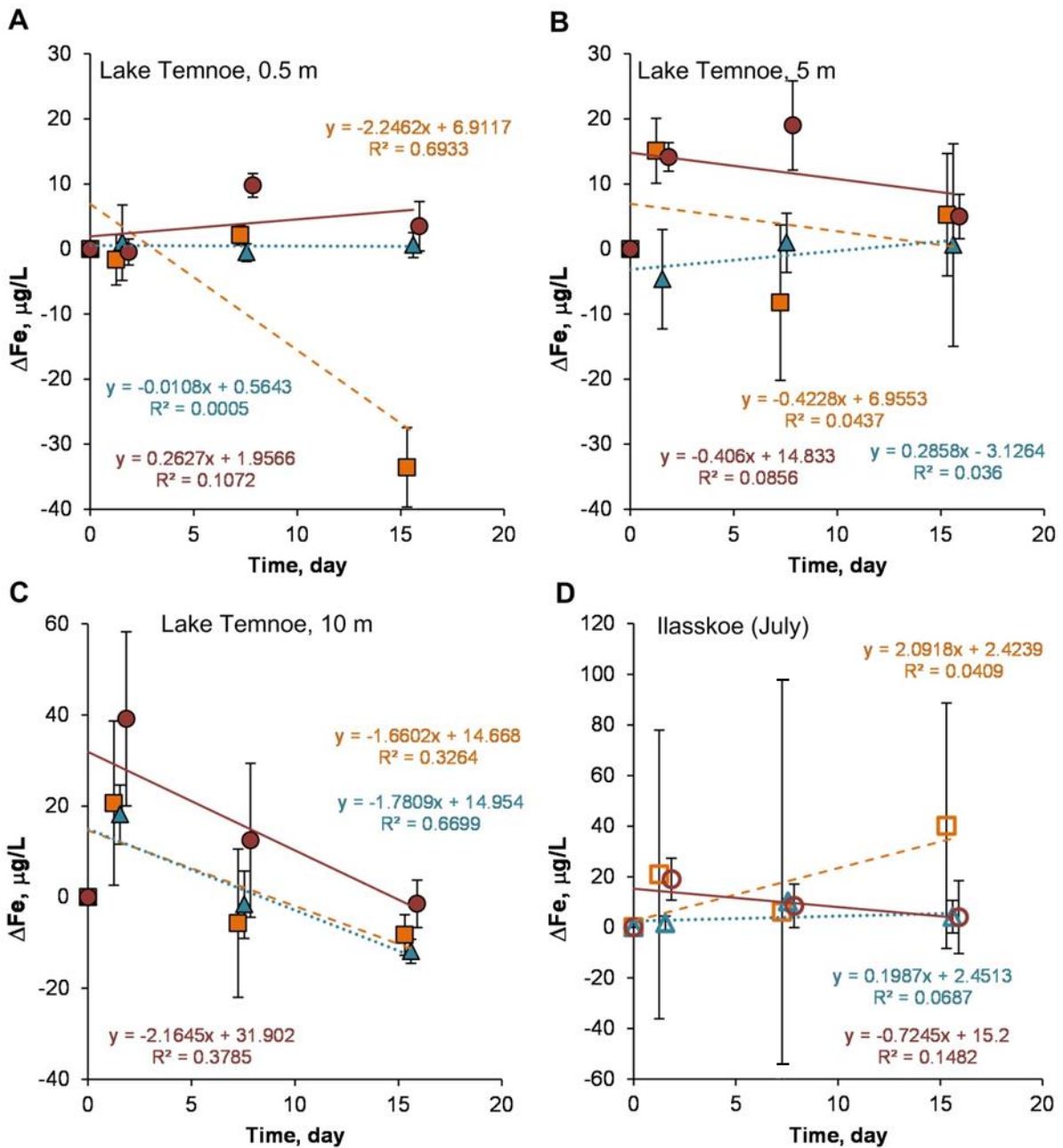
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■ June   
 ▲ August   
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  Piezometer   
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**Fig. 6.** Change in Fe concentration (relative to the control) over time in photo-degradation experiments. The error bars are 1 s.d. of duplicates. Lake Temnoe 0.5 m (A), 5 m (B) and 10 m (C) in June (squares), August (triangles) and October (circles). Ilasskoe continuum in July (D) includes piezometer (squares), peatland pool Severnoe (triangles) and stream Chernyi (circles)

1143 **Table 1.** Landscape setting, hydrochemical characteristics and CO<sub>2</sub> concentration and emission  
 1144 flux of studied waters. S.C. is specific conductivity and EB and OB is eutrophic and  
 1145 oligotrophic bacteria count, respectively.

1146 **1A.** Ilasskoe bog continuum in July.

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	Piezometer	Lake Severnoe	Stream Chernyi
<b>GPS coordinates</b>	N64.328694° E40.612556°	N64.334361° E40.609667°	N64.330982° E40.653352°
<b>Description</b>	Shallow groundwater	Peatland pool	Outlet stream
<b>T, °C</b>	11.4	19.4	13
<b>O<sub>2</sub>, mg/L</b>	0.6	8.6	7.5
<b>pH</b>	3.9	4.0	5.7
<b>S.C., μS cm<sup>-1</sup></b>	46	17	26
<b>DOC, mg L<sup>-1</sup></b>	87.6	12.7	38.4
<b>SUVA<sub>254</sub></b>	4.13	3.80	4.85
<b>P-PO<sub>4</sub>, μg L<sup>-1</sup></b>	8.6	3.0	1.7
<b>P<sub>total</sub>, μg L<sup>-1</sup></b>	153	10	20
<b>N-NO<sub>3</sub>, μg L<sup>-1</sup></b>	111	70	98
<b>N-NH<sub>4</sub>, μg L<sup>-1</sup></b>	85.4	16.1	12.6
<b>N<sub>total</sub>, μg L<sup>-1</sup></b>	1180	222	399
<b>Si, μg L<sup>-1</sup></b>	1808	47	2076
<b>CO<sub>2</sub>, μmol/L</b>	3360	55	318
<b>CO<sub>2</sub> flux, mmol m<sup>-2</sup> d<sup>-1</sup></b>	1600	22	151
<b>EB, CFU mL<sup>-1</sup></b>	49360	56600	9000
<b>OB, CFU mL<sup>-1</sup></b>	54560	37900	21600

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1149 **1B.** Lake Temnoe across seasons and depths.

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<b>Month</b>	Jun	Jun	Jun	Aug	Aug	Aug	Oct	Oct	Oct
	0.5	5	10	0.5	5	10	0.5	6	10
<b>GPS</b>	N64.47683°			E041.74533°					
<b>Description</b>	Lake in the northern taiga								
<b>T, °C</b>	12.7	4,9	4,5	18.4	5.5	4.3	9.0	5.8	4.4
<b>O<sub>2</sub>, mg/L</b>	8,45	4,8	4,5	7.78	4.93	2.63	8.90	4.46	2.14
<b>pH</b>	5.2	5.2	5.3	6.0	5.5	5.7	5.2	5.2	5.1
<b>S.C., μS cm<sup>-1</sup></b>	17	17	19	17	17	19	18	18	20
<b>DOC, mg L<sup>-1</sup></b>	12.6	19.2	21	19	19.5	21.2	19.4	20.6	20.6
<b>SUVA<sub>254</sub></b>	4.6	4.7	4.6	4.2	4.5	4.5	4.3	4.3	4.7
<b>P-PO<sub>4</sub>, μg L<sup>-1</sup></b>	2.9	3.3	6.4	0.9	3.6	9.4	3.8	4.6	4.2
<b>P<sub>total</sub>, μg L<sup>-1</sup></b>	19	17	19	20	16	20	18	19	20
<b>N-NO<sub>3</sub>, μg L<sup>-1</sup></b>	119	150	137	86	152	254	88	85	100
<b>N-NH<sub>4</sub>, μg L<sup>-1</sup></b>	7.1	8.0	10.0	9.1	17.5	13.8	16.4	14.1	15.5
<b>N<sub>total</sub>, μg L<sup>-1</sup></b>	305	420	408	355	315	337	425	416	396
<b>Si, μg L<sup>-1</sup></b>	1940	2268	2354	1183	2208	2714	2269	2380	2380
<b>CO<sub>2</sub>, μmol/L</b>	99	309	329	110	256	337	223	232	253
<b>CO<sub>2</sub> flux, mmol m<sup>-2</sup> d<sup>-1</sup></b>	32	-	-	46	-	-	71	-	-
<b>EB, CFU mL<sup>-1</sup></b>	-	36	50	259	92	270	780	220	105
<b>OB, CFU mL<sup>-1</sup></b>	50	570	420	-	190	-	680	150	66

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**Table 2.** The % bio- and photodegradable solutes (mean  $\pm$  s.d.) whose relative change (concentration decrease) in the course of experiment was superior to that of SD. Prefix  $\Delta B$  and  $\Delta P$  represents the effect of bio- and photodegradation, respectively. Duration of biodegradation and photodegradation is  $21.6 \pm 0.1$  and  $15.6 \pm 0.1$  days, respectively.  $W$  represents the probability of measurable effect, significantly different from changes in the control reactors. Only the components with  $W \geq 33\%$  are presented. Temnoe Lake is deep stratified lake in the forest. Peizometer, peatland pool and outlet stream represent the hydrological continuum of the Ilasskoe Bog.

Index	Temnoe Lake		Temnoe Lake		Temnoe Lake		Temnoe Lake		Temnoe Lake		Temnoe Lake		Piezo-meter (Jul)		Peatland pool (Jul)		Outlet stream (Jul)	
	0.5 m (Jun)	5 m (Jun)	10 m (Jun)	0.5 m (Aug)	5 m (Aug)	10 m (Aug)	0.5 m (Oct)	5 m (Oct)	10 m (Oct)	0.5 m (Oct)	6 m (Oct)	10 m (Oct)	10 m (Oct)					
$\bar{\alpha}$ , $\mu S/cm$	17	17	19	17	17	17	18	18	19	18	18	20	20	46	17	26		
$\Delta B(\bar{\alpha} \pm SD)$	-24 $\pm$ 4	-26 $\pm$ 7	-30 $\pm$ 5	-23 $\pm$ 3	-27 $\pm$ 3	-27 $\pm$ 3	-24 $\pm$ 7	-23 $\pm$ 4	-33 $\pm$ 16	-24 $\pm$ 7	-23 $\pm$ 4	-17 $\pm$ 5	-17 $\pm$ 5	0	-18 $\pm$ 10	-29 $\pm$ 4		
DOC, mg/L	12.6	19.2	21.0	19.0	19.5	19.5	19.4	20.6	21.2	19.4	20.6	20.6	20.6	87.6	12.7	38.4		
$\Delta B(DOC \pm SD)$	3.4 $\pm$ 0.8	2.0 $\pm$ 1.4	2.1 $\pm$ 0.8	3.2 $\pm$ 2.6	4.7 $\pm$ 1.6	4.7 $\pm$ 1.6	4.9 $\pm$ 2.0	3.0 $\pm$ 0.3	2.3 $\pm$ 1.4	4.9 $\pm$ 2.0	3.0 $\pm$ 0.3	5.6 $\pm$ 2.2	5.6 $\pm$ 2.2	4.1 $\pm$ 2.3	4.9 $\pm$ 1.4	3.1 $\pm$ 2.4		
$\Delta P(DOC \pm SD)$	20.7 $\pm$ 4.6	14.9 $\pm$ 1.6	17.0 $\pm$ 6.4	0	5.5 $\pm$ 5.1	0	9.7 $\pm$ 2.4	0	9.7 $\pm$ 2.4	0	0	0	0	5.9 $\pm$ 1.3	0	11.0 $\pm$ 1.8		
Al, $\mu g/L$	275	298	329	254	296	296	275	288	335	275	288	323	323	276	59	388		
$\Delta B(Al \pm SD)$	3.5 $\pm$ 1.4	1.8 $\pm$ 0.9	0	2.0 $\pm$ 1.3	0	1.4 $\pm$ 1.5	2.0 $\pm$ 1.9	0	1.4 $\pm$ 1.5	2.0 $\pm$ 1.9	0	0	0	0.9 $\pm$ 2.2	0	1.3 $\pm$ 1.8		
$\Delta P(Al \pm SD)$	1.9 $\pm$ 1.1	2.7 $\pm$ 0.9	3.6 $\pm$ 1.3	0	2.5 $\pm$ 1.3	0	1.7 $\pm$ 2.0	0.7 $\pm$ 0.9	1.7 $\pm$ 2.0	0.7 $\pm$ 0.9	0	0	0	0	0	0.8 $\pm$ 0.9		
Ti, $\mu g/L$	1.5	2.1	2.6	1.1	2.0	2.0	1.7	1.9	2.6	1.7	1.9	2.5	2.5	3.7	0.6	5.0		
$\Delta B(Ti \pm SD)$	-9.2 $\pm$ 1.6	-9.9 $\pm$ 7.4	-2.6 $\pm$ 2.7	-4.8 $\pm$ 3.4	-1.8 $\pm$ 2.7	0	-3.6 $\pm$ 1.7	-1.0 $\pm$ 3.1	0	-3.6 $\pm$ 1.7	-1.0 $\pm$ 3.1	-1.0 $\pm$ 3.9	-1.0 $\pm$ 3.9	-2.3 $\pm$ 3.6	-2.2 $\pm$ 1.7	-1.4 $\pm$ 2.2		
$\Delta P(Ti \pm SD)$	-0.1 $\pm$ 3	-3 $\pm$ 3	-8 $\pm$ 3	0 $\pm$ 0	-9 $\pm$ 1	-9 $\pm$ 1	-2 $\pm$ 4	0	-3 $\pm$ 2	-2 $\pm$ 4	0	0	0	0	-20 $\pm$ 4	-3.3 $\pm$ 0.5		
V, $\mu g/L$	0.5	0.6	0.7	0.4	0.5	0.5	0.4	0.5	0.7	0.4	0.5	0.7	0.7	1.1	0.5	1.3		
$\Delta B(V \pm SD)$	-8.3 $\pm$ 16.2	-5.4 $\pm$ 3.2	-4.9 $\pm$ 2.3	-6.8 $\pm$ 7.5	-10.0 $\pm$ 4.6	-10.0 $\pm$ 4.6	-14.7 $\pm$ 11	-13.9 $\pm$ 4.3	-1.7 $\pm$ 1.6	-14.7 $\pm$ 11	-13.9 $\pm$ 4.3	-16.1 $\pm$ 1.7	-16.1 $\pm$ 1.7	-3.2 $\pm$ 2.6	-0.2 $\pm$ 3.4	-17.9 $\pm$ 5.0		
Mn, $\mu g/L$	39	55	79	17	48	48	30	47	93	30	47	105	105	78	9	47		
$\Delta B(Mn \pm SD)$	0	0	-0.3 $\pm$ 2.2	-31.8 $\pm$ 1.3	-3.2 $\pm$ 1.6	-3.2 $\pm$ 1.6	-4.8 $\pm$ 2.2	-3.2 $\pm$ 1.7	-0.6 $\pm$ 2.2	-4.8 $\pm$ 2.2	-3.2 $\pm$ 1.7	-0.4 $\pm$ 0.1	-0.4 $\pm$ 0.1	0	0	-1.6 $\pm$ 2.8		
Fe, $\mu g/L$	358	527	710	165	460	460	317	448	795	317	448	820	820	4402	157	1006		
$\Delta B(Fe \pm SD)$	-18.1 $\pm$ 2.5	-9.1 $\pm$ 2.6	-5.4 $\pm$ 1.6	-13.5 $\pm$ 1.0	-6.3 $\pm$ 2.6	-6.3 $\pm$ 2.6	-9.5 $\pm$ 1.4	-7.8 $\pm$ 1.9	-1.4 $\pm$ 1.9	-9.5 $\pm$ 1.4	-7.8 $\pm$ 1.9	-3.3 $\pm$ 1.8	-3.3 $\pm$ 1.8	-0.8 $\pm$ 0.8	-13.6 $\pm$ 4.3	-4.5 $\pm$ 2.4		
$\Delta P(Fe \pm SD)$	-3.9 $\pm$ 0.6	-2.0 $\pm$ 1.9	-4.0 $\pm$ 1.3	0	-2.9 $\pm$ 1.5	-2.9 $\pm$ 1.5	-1.2 $\pm$ 0.4	0	-0.2 $\pm$ 0.6	-1.2 $\pm$ 0.4	0	0	0	0	0	0		

Table 2, continued.

Index	Temnoe Lake 0.5 m (Jun)		Temnoe Lake 5 m (Jun)		Temnoe Lake 10 m (Jun)		Temnoe Lake 0.5 m (Aug)		Temnoe Lake 5 m (Aug)		Temnoe Lake 10 m (Aug)		Temnoe Lake 0.5 m (Oct)		Temnoe Lake 6 m (Oct)		Temnoe Lake 10 m (Oct)		Piezo-meter (Jul)	Peatland pool (Jul)	Outlet stream (Jul)
	Value	SD	Value	SD	Value	SD	Value	SD	Value	SD	Value	SD	Value	SD	Value	SD	Value	SD			
Co, µg/L	0.28		0.39		0.68		0.07		0.30		0.65		0.18		0.31		0.74		0.45	0.06	0.30
ΔB(Co±SD)	-2.2±5.1		-1.2±2.1		-3.7±4.6		-32.7±2.6		-8.1±5.6		-2.7±3.3		-11.0±4.4		-9.1±5.1		-1.6±0.4		0	0	-20.6±27.8
Cu, µg/L	0.5		0.6		0.7		0.6		0.6		0.7		0.7		0.7		0.5		1.5	0.3	0.8
ΔB(Cu±SD)	0		0		0		-14.3±1.4		-6.8±4.0		-17.9±11.0		-5.3±4.8		-4.1±8.0		-1.4±12.3		0	-7.7±9.9	0
Ga, µg/L	0.017		0.022		0.026		0.012		0.016		0.023		0.017		0.015		0.024		0.126	0.016	0.066
ΔP(Ga±SD)	-14±6		-13±5		-10±4		0		-1±8		0		-10±4		0		0		-7±5	-5±8	-6±3
Y, µg/L	0.22		0.25		0.28		0.20		0.24		0.28		0.22		0.23		0.28		0.10	0.01	0.21
ΔP(Y±SD)	-1.3±4.6		-6.7±0.9		-5.3±2.5		0		-2.3±0.7		-1.0±1.6		-1.4±0.2		0		0		0	-5.8±2.7	0
Zr, µg/L	0.4		0.4		0.5		0.4		0.5		0.5		0.4		0.4		0.5		0.3	0.1	0.4
ΔP(Zr±SD)	-15±4		-14±0		-13±2		-9±20		-17±1		-14±3		-4±4		0		0		0	-32±3	-8±1
Nb, µg/L	0.016		0.020		0.025		0.012		0.020		0.024		0.017		0.018		0.025		0.033	0.005	0.042
ΔB(Nb±SD)	-3.6±10.2		-1.7±7.0		0		-7.7±4.8		-1.1±2.6		0		-7.3±2.3		-1.5±6.3		-5.0±4.0		-2.4±1.5	0	0
ΔP(Nb±SD)	-9±3		-8±3		-9±1		-6±23		-13±2		-10±5		-8±4		0		-3±3		0	-13±10	-10±4
Ba, µg/L	4.8		5.1		5.8		4.6		5.0		5.7		4.9		4.8		5.6		54.4	1.5	56.8
ΔB(Ba±SD)	-2.2±0.7		-2.8±1.7		-1.0±2.7		0		0		0		-1.9±0.5		-1.7±3.7		-5.9±1.6		0	0	-1.3±3.4
La, µg/L	0.23		0.26		0.30		0.21		0.26		0.32		0.24		0.27		0.31		0.07	0.01	0.22
ΔB(La±SD)	-4.9±6.5		0		0		-3.9±0.9		-0.3±1.6		-2.6±1.5		-1.1±3.6		-2.4±1.2		-4.0±2.8		-0.8±10.4	-29.7±10.0	-2.0±2.8
ΔP(La±SD)	-3.8±3.6		-1.2±5.9		-2.0±2.7		0		-3.6±1.0		-3.2±1.6		-1.8±0.9		-2.6±1.0		0		0	-13.5±3.0	0
Ce, µg/L	0.58		0.65		0.71		0.50		0.62		0.78		0.59		0.63		0.78		0.21	0.03	0.56
ΔB(Ce±SD)	-5.2±4.2		0		0		-4.4±0.7		-0.1±1.4		-0.8±1.0		-0.9±2.9		-0.8±1.1		-2.2±2.2		0	-9.3±5.9	-2.0±1.9
ΔP(Ce±SD)	-4.9±1.9		-6.2±1.2		-1.1±1.7		0		-1.5±0.5		-3.2±1.7		-1.9±1.2		0		-0.047±0.46		-3.7±1.5	-2.4±1.6	-3.6±1.1

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Table 2, continued.

Index	Temnoe Lake 0.5 m (Jun)		Temnoe Lake 5 m (Jun)		Temnoe Lake 10 m (Jun)		Temnoe Lake 0.5 m (Aug)		Temnoe Lake 5 m (Aug)		Temnoe Lake 10 m (Aug)		Temnoe Lake 0.5 m (Oct)		Temnoe Lake 6 m (Oct)		Temnoe Lake 10 m (Oct)		Piezometer (Jul)	Peatland pool (Jul)	Outlet stream (Jul)	
	Value	SD	Value	SD	Value	SD	Value	SD	Value	SD	Value	SD	Value	SD	Value	SD	Value	SD	Value	Value	Value	
Pr, µg/L	0.075	0.085	0.094	0.069	0.082	0.105	0.077	0.084	0.102	0.027	0.005	0.070	0.070	0.070	0.005	0.070	0.070	0.070	0.027	0.005	0.070	
ΔB(Pr±SD)	-1.9±5.2	0	-4.0±1.1	0	-0.7±1.3	0	-0.9±1.6	0	-3.2±2.3	-3.0±1.7	-10.8±8.4	-1.7±2.3	-1.7±2.3	-1.7±2.3	-10.8±8.4	-1.7±2.3	-1.7±2.3	-1.7±2.3	-3.0±1.7	-10.8±8.4	-1.7±2.3	
ΔP(Pr±SD)	-3.0±3.9	-5.5±0.9	-0.4±2.9	13.7±20.8	0	-6.1±2.8	0	-1.3±1.8	0	0	0	0	0	0	0	0	0	0	-0.01±2.4	-16.9±3.0	-2.4±2.1	
Nd, µg/L	0.33	0.34	0.39	0.29	0.33	0.42	0.33	0.32	0.41	0.11	0.02	0.27	0.27	0.27	0.11	0.02	0.27	0.27	0.11	0.02	0.27	
ΔP(Nd±SD)	-7.8±2.4	-4.5±2.5	-0.8±2.1	0	-3.8±2.5	-2.0±3.3	0	0	0	0	0	0	0	0	0	0	0	0	-2.5±2.8	-24.7±6.2	0	
Eu, µg/L	0.015	0.017	0.016	0.012	0.016	0.020	0.014	0.018	0.021	0.011	0.001	0.017	0.017	0.017	0.011	0.001	0.017	0.017	0.011	0.001	0.017	
ΔP(Eu±SD)	0	-10.9±3.3	0	0	-0.8±4.7	-3.0±4.0	0	-8.7±8.4	-6.9±4.2	-23.7±8.6	-58.2±15.2	-1.0±1.0	-1.0±1.0	-1.0±1.0	-23.7±8.6	-58.2±15.2	-1.0±1.0	-1.0±1.0	-23.7±8.6	-58.2±15.2	-1.0±1.0	
Gd, µg/L	0.06	0.07	0.08	0.05	0.07	0.08	0.06	0.07	0.09	0.02	0.00	0.06	0.06	0.06	0.02	0.00	0.06	0.06	0.02	0.00	0.06	
ΔP(Gd±SD)	-0.2±4.4	-6.5±2.9	-3.5±3.5	0	-6.0±3.2	-7.6±1.9	-2.3±2.0	0	-6.7±3.2	-0.2±4.4	-6.5±2.9	-3.5±3.5	-0.2±4.4	-6.5±2.9	-3.5±3.5	-7.6±1.9	-2.3±2.0	-6.7±3.2	0	0	0	-3.2±3.5
Ho, µg/L	0.009	0.009	0.011	0.007	0.009	0.011	0.009	0.009	0.011	0.007	0.009	0.009	0.011	0.009	0.004	0.0004	0.009	0.011	0.004	0.0004	0.009	
ΔP(Ho±SD)	-1.9±1.5	-0.3±1.6	-4.6±6.5	0	-0.1±3.6	-3.4±3.2	-11.0±4.9	0	0	0	0	0	0	0	-21.5±11.3	0	-10.9±5.5	-10.9±5.5	-21.5±11.3	0	-10.9±5.5	
Er, µg/L	0.023	0.025	0.033	0.022	0.026	0.030	0.022	0.023	0.031	0.011	0.001	0.023	0.023	0.023	0.011	0.001	0.023	0.023	0.011	0.001	0.023	
ΔB(Er±SD)	0	0	0	-5.2±3.0	-2.1±1.8	0	-2.1±3.5	0	-2.0±5.3	0	0	0	0	0	-15.6±4.9	-22.9±19.5	0	0	-15.6±4.9	-22.9±19.5	0	
Pb, µg/L	0.23	0.24	0.23	0.16	0.23	0.39	0.28	0.28	0.32	11	0.35	0.65	0.65	0.65	11	0.35	0.65	0.65	11	0.35	0.65	
ΔB(Pb±SD)	0	0	0	-21.3±2.5	-2.0±7.6	-2.4±1.6	-8.2±3.3	-7.2±9.9	-0.8±2.7	0	0	0	0	0	0	0	0	0	0	0	0	-8.5±10.9
Th, µg/L	0.046	0.052	0.066	0.058	0.054	0.064	0.053	0.054	0.061	0.019	0.005	0.050	0.050	0.050	0.019	0.005	0.050	0.050	0.019	0.005	0.050	
ΔP(Th±SD)	0	0	-11.6±2.6	-12.2±22.5	-7.8±3.2	-18.1±5.6	0	0	-2.0±1.9	0	0	0	0	0	0	0	0	0	0	0	0	-10.6±0.8

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**Table 3.** Mean ( $\pm$ SD), depth-integrated rates of bio- and photodegradation ( $\text{mg C L}^{-1} \text{d}^{-1}$ )

Object	$V_{\text{Biodegradation}}$	$V_{\text{Photodegradation}}$
Lake Temnoe		
Forest Lake (Jun)	$-0.02 \pm 0.0014$	$-0.19 \pm 0.03$
Forest Lake (Aug)	$-0.031 \pm 0.010$	$-0.067 \pm 0.066$
Forest Lake (Oct)	$-0.042 \pm 0.013$	0
Ilaskoe Bog continuum (July)		
Piezometer water	$-0.17 \pm 0.09$	$-0.33 \pm 0.07$
Peatland pool	$-0.029 \pm 0.008$	0
Outlet stream (Chernyi)	$-0.055 \pm 0.043$	$-0.27 \pm 0.043$

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