Effects of photosymbiosis and related processes on planktic foraminifera-bound nitrogen isotopes in South Atlantic sediments

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Abstract. Foraminifera often form symbiotic relationships with photosynthetic algae, providing a host environment and inorganic nutrients in exchange for photosynthetic organic matter from the algal symbiont. To date, the history of this relationship has been studied in paleoceanographic records using the oxygen and carbon stable isotopes of foraminiferal calcite. More recently, photosymbiotic activity has been observed to impact the nitrogen isotope ratio ($\delta^{15}N$) of foraminiferal tissue and the organic matter incorporated into foraminiferal tests. Dinoflagellate symbiont-bearing species appear to be lower in $\delta^{15}N$ than symbiont-barren species and more similar to their feeding sources, likely due to their retention of low-$\delta^{15}N$ metabolic ammonium and thus a weaker amplitude for the "trophic enrichment factor", the $\delta^{15}N$ increase per trophic level that is widely observed in food webs. We report new glacial/interglacial foraminifera-bound $\delta^{15}N$ (FB-$\delta^{15}N$) data from Deep Sea Drilling Program Site 516, located in the subtropical South Atlantic gyre, which contains multiple foraminifera species at adequately high abundance for inter-species comparison of foraminiferal nitrogen, carbon, and oxygen isotopes over a full glacial cycle. Our data show a conserved $\delta^{15}N$ difference of 3-5 ‰ between dinoflagellate-bearing species and the other species, qualitatively consistent with, but greater in amplitude than, the $\delta^{15}N$ difference observed in previous modern ocean and core-top studies. We propose that this greater amplitude at Site 516 is the result of lateral transport of symbiont-barren species into the South Atlantic subtropical gyre, which appears to represent a small region of low thermocline nitrate $\delta^{15}N$ surrounded by regions with higher thermocline nitrate $\delta^{15}N$.

We demonstrate that FB-$\delta^{15}N$ provides a clear signal of dinoflagellate endosymbiosis, and that it may be able to identify other, weaker endosymbioses (e.g., with chrysophytes or pelagophytes). However, the data also suggest caution in regions with strong gradients, where species from contrasting environments may occur in a single sediment sample.

1 Introduction

Biologically available nitrogen (or "fixed N") is an essential nutrient and thus central to ocean productivity and biogeochemical cycling (Sarmiento and Gruber, 2006), with links to atmospheric CO_2 and thus climate (Broecker, 1982; Broecker and Henderson, 1998; Falkowski, 1997; McElroy, 1983). The ocean's fixed N reservoir size is dominantly controlled by the balance between N_2 fixation and denitrification (Gruber and Sarmiento, 1997), which can be reconstructed by N isotopes ($^{15}N/^{14}N$, or $\delta^{15}N$) measured on organic matter in marine sediments (Brandes and Devol, 2002; Deutsch et al., 2004; Galbraith et al., 2013). In addition, variation in the degree of nitrate consumption in ocean surface waters can also be investigated with N isotopes in marine sediments (Altabet and Francois, 1994).

The isotopic composition of bulk N in marine sediments is variably influenced by diagenesis on the seabed and exogenous N inputs (e.g., (Meckler et al., 2011; Robinson et al., 2012; Möbius, 2013; Schubert and Calvert, 2001). Largely to address these concerns, over the past decades, fossil-bound nitrogen isotope methods have been developed (Kast et al., 2022; Lueders-Dumont et al., 2018; Sigman et al., 1999; Wang et al., 2015; Wang et al., 2014; Brunelle et al., 2007; Robinson et al., 2004), including foraminifera-bound (FB-) $\delta^{15}N$ (Ren et al., 2009). In contrast to bulk sediment $\delta^{15}N$, fossil-bound organic matter represents a specific N pool in the sediment archive that is physically protected from bacterial/chemical diagenesis, exogenous N contamination and thermal stress by the biomineral matrix, and appears to be minimally affected by partial calcite dissolution (Martínez-García et al., 2022). The first applications of the FB-$\delta^{15}N$ proxy focused on reconstructing past changes in the N cycle in the North Atlantic (Ren et al., 2009; Straub et al., 2013), Pacific (Ren et al., 2012a; Ren et al., 2017; Ren et al., 2015) and Southern Ocean (Martínez-García et al., 2014) over Pleistocene glacial/interglacial cycles. More recent studies have shown that FB-$\delta^{15}N$ can provide reliable information about changes in the N cycle through the Cenozoic (Auderset et al., 2022; Hess et al., 2023; Kast et al., 2019; Wang et al., 2022).

Studies in the oligotrophic ocean, where surface-water nitrate is fully consumed, have established that FB-$\delta^{15}N$ in surface sediments reflect the $\delta^{15}N$ of shallow subsurface nitrate (i.e., the nitrate that is supplied to and consumed in the euphotic zone) (Ren et al., 2012b; Ren et al., 2009; Schiebel et al., 2018). Water column studies indicate seasonal changes in FB-$\delta^{15}N$ that parallel the changes in the $\delta^{15}N$ of bulk filtered particles (>0.7 μm) and/or net-tow collected material (>150 μm) in surface waters (Smart et al., 2020; Smart et al., 2018), as is consistent with the role of planktic foraminifera as heterotrophic zooplankton that feed variously on phytoplankton, other heterotrophs, and other types of particulate N (PN) (e.g., (Anderson et al., 1979; LeKieffre et al., 2020; Spindler et al., 1984; Takagi et al., 2019). Phytoplankton assimilate the nitrate supplied from below and provide the food to zooplankton such as foraminifera. The metabolism of zooplankton and other heterotrophs releases low-$\delta^{15}N$ regenerated N (e.g., ammonium), which is avidly reassimilated by phytoplankton. The net result is that phytoplankton biomass, on average, is lower in $\delta^{15}N$ than the nitrate supply (Altabet, 1988; Fawcett et al., 2011; Knapp et al., 2005), while herbivorous zooplankton are similar in $\delta^{15}N$ to the nitrate supply (Montoya et al., 2002). Foraminifera, which feed on both zooplankton and phytoplankton, thus have a $\delta^{15}N$ that varies with the $\delta^{15}N$ of the nitrate supplied and consumed in the euphotic zone (Ren et al., 2009; Ren et al., 2012b; Smart et al., 2018; Schiebel et al., 2018).

Aside from the δ^{15}N of their diet, some foraminifera species have a special aspect of their physiology that influences their δ^{15}N: most extant spinose planktic foraminifera species host algal symbionts (Hemleben et al., 1989). A typical heterotrophic organism puts most of its dietary organic carbon toward energy generation, releasing the metabolized carbon as CO_2. If the C:N:P of the organism is not vastly different from that of its food, this will leave the organism with an excess of N (as well as P). The N must be removed, typically being lost by deamination and excretion of the resulting ammonium (or of urea or uric acid that are produced from the ammonium). Foraminifera, like other symbiont-bearing organisms (e.g., scleractinian corals), take advantage of their metabolic N and P waste to obtain additional organic carbon: they provide these nutrients to symbiotic algae, which use them to fix additional organic carbon, with a portion of this organic matter then being returned to the host for its nutrition. This strategy is particularly prominent in low-nutrient marine systems, where environmental nutrient concentrations are low and light is abundant, such as in the oligotrophic subtropical gyres.

Because decisive steps in N catabolism break a N-containing bond, which involves substantial isotopic fractionation, the catabolically produced ammonium is low in δ^{15}N (Silfer et al., 1992). In normal heterotrophic organisms, the loss of this low-δ^{15}N ammonium raises the δ^{15}N of the organism, paraphrased by the isotope ecologist's rule of thumb: "You are what you eat plus a few (permille)" (Deniro, 1976; Deniro and Epstein, 1981). In dinoflagellate-bearing foraminifera, however, the dinoflagellates are thought to assimilate the low-δ^{15}N ammonium, recycling it back into the host-symbiont system and thus reducing the δ^{15}N elevation of the foraminifera relative to its food (Ren et al., 2009; Smart et al., 2018). In summary, because N excretion to the environment can be greatly reduced by photosynthetic endosymbiosis, foraminifera that host symbionts are expected to have a δ^{15}N closer to that of their food source and lower than "pure" heterotrophs with the same food source.

The role of symbiosis in ancient organisms and ecosystems is of great interest. In low- to mid-latitude surface waters of the ocean, limitation of autotrophic biomass by the availability of nutrients (especially N and P) is common, and changes in ocean nutrient conditions through time may have played a major role in evolution (Hohmann-Marriott and Blankenship, 2011). Photosynthetic endosymbiosis – hosted by organisms such as scleractinian corals and planktic foraminifera – has been described as an adaptation to settings of N- and P-limited photosynthetic growth, with the heterotrophic host sharing its metabolic N and P with the autotrophic symbionts in exchange for organic carbon (C) (e.g., (Hallock, 1981)). Thus, the origination and history of these symbioses may reflect changes in the ocean's nutrient inventories and cycling. In turn, these symbioses may affect nutrient cycling and the availability of nutrients, especially in systems such as shallow reefs.

Photosymbiosis in modern foraminifera species can be investigated by microscopic observations of intracellular algae (Anderson and Be, 1976; Gastrich, 1987; Lee et al., 1965), pulse-chase experiments with isotope labels (Bird et al., 2020; LeKieffre et al., 2020), active chlorophyll fluorometry (Takagi et al., 2019), or detection of algal DNA (Gast and Caron, 1996; Gast et al., 2000). However, these methods may be complicated by foraminiferal feeding on algal cells, making it difficult to distinguish endosymbionts from prey. More to the point, identifying and/or quantifying photosymbiotic activity from fossil foraminifera is even more challenging, particularly for extinct species. The carbon isotopic composition (δ^{13}C) of the $CaCO_3$ of planktic foraminifera tests may be affected by photosymbiosis. Symbiotic photosynthetic organisms have a preference for taking up ^{12}C, resulting in local dissolved inorganic carbon (DIC) becoming enriched in ^{13}C, which is subsequently integrated into the calcium carbonate tests of foraminifera (Spero and Deniro, 1987). In larger foraminifera specimens, which host a greater number of photosymbionts, there is a relatively higher enrichment of ^{13}C than in smaller individuals (Spero et al., 1991). Thus, the relationship between the $\delta^{13}C$ and test size of symbiont-bearing foraminifera has been proposed and applied as a metric by which to identify symbiosis in ancient foraminifera (D'Hondt et al., 1994; Edgar et al., 2013; Ezard et al., 2015; Norris, 1998). The oxygen isotopic composition ($\delta^{18}O$) of the $CaCO_3$ can be used to infer the depth habitat of different foraminifera species, with higher $\delta^{18}O$ indicating deeper and colder water masses (e.g., Mulitza et al. (1997), Rashid and Boyle (2007) and references therein). Thus, it has also been proposed that, due to persistent shallower depth habitats throughout ontogeny of symbiont-bearing foraminifera, their size-specific $\delta^{18}O$ relationship should be minimal in comparison to symbiont-barren foraminifera species, which have a larger potential range of depth habitats. However, there are potential pitfalls to these approaches: The foraminiferal test size can be influenced by environmental conditions such as surface water stratification, with smaller test sizes reported during periods of upwelling (Schmidt et al., 2004). Furthermore, sediment mixing preferentially affects finer fractions and thus impacts the size fraction *vs.* $\delta^{13}C$ and $\delta^{18}O$ relationships (Hupp et al., 2019). Both proxies, their biological mechanisms, and their pitfalls require further investigation.

This study investigates foraminifera-bound nitrogen isotopes (FB-$\delta^{15}N$) as a new tool for the reconstruction of past symbiosis in planktic foraminifera. We measure shell-bound $\delta^{15}N$ in five foraminifera species from a South Atlantic sediment core over a full glacial cycle. We also analyse the same samples for test $\delta^{18}O$ and $\delta^{13}C$. In addition, in a subset of samples, we measure $\delta^{18}O$ and $\delta^{13}C$ in different test size fractions.

2 Material and methods

2.1 Core site and local hydrography

We analyzed samples from sediment core DSDP 516, located on the Rio Grande Rise (30°16'S, 35°17'W, 1313 m water depth) near the core of the oligotrophic subtropical gyre in the South Atlantic Ocean (Fig.1). The local hydrography is dominated by the wind-driven anticyclonic South Atlantic gyre. The subtropical gyre is associated with net subduction, deepened isopycnals, slow nutrient supply from the subsurface to the euphotic zone, and N and P impoverishment in the euphotic zone (Cullen et al., 2002). The nutrient supply to the euphotic zone at Site 516 can be augmented by eddy mixing with the nutrient-bearing waters of the Antarctic Circumpolar Current, which flow east-northeastward along the southern boundary of the gyre (Reid et al., 1977; Schmid et al., 2000). The Brazil Current to the west may also contribute nutrients. However, Site 516 is near the core of the gyre, and relatively isolated from nutrient inputs at the margins. Accordingly, the annual average nitrate concentration ranges from 0.2 μmol/kg to below the limit of detection and chlorophyll-*a* concentrations are <0.7 mg/m^3 (Siccha et al., 2018). These conditions are comparable to those observed at the oligotrophic Sargasso Sea in the North Atlantic, as established for the Bermuda Atlantic Time-series Station (Lomas et al., 2013; Steinberg et al., 2001). The eastern boundary of the subtropical gyre is marked by the Benguela Current, where the persistent southerly and southeasterly winds drive surface water offshore and facilitate upwelling of cold and nutrient-rich subsurface water (Cole and Villacastin, 2000). The Benguela

Current flows northward and turns into the southern branch of the South Equatorial Current, which forms the northern edge of the subtropical gyre (Stramma, 1991).

2.2 Nitrogen isotope analysis

2.2.1 Foraminifer-bound nitrogen isotopes

Sediment samples were wet sieved through a 63 µm mesh and dried overnight in a clean oven at 40°C. Between 600-800 individual foraminifera tests were manually picked from size fraction 250-400 µm. We analyzed dinoflagellate-bearing *Trilobatus sacculifer*, the two chromotypes *Globigerionoides ruber ruber* and *G. ruber albus* (pink and white, respectively), chrysophyte-bearing *Globigerinella siphonifera (*syn. *G. aequilateralis*), as well as symbiont-barren *Globigerina bulloides* and *Globorotalia truncatulinoides.* The detailed protocol used here for measuring FB-$\delta^{15}N$ can be found in (Auderset et al., 2022) and (Moretti et al., 2024). In short, about 5-7 mg of foraminifera tests were gently crushed and chemically treated to remove external organic matter, clay, and manganese coatings. Subsequently, 3-5 mg of the cleaned material was weighed out per sample for N content determination and ultimately N isotope analysis. The test fragments were first dissolved in hydrochloric acid (50 µl of 3N HCl), and the organic N was converted into nitrate (NO_3^-) by persulfate oxidation following a slightly modified protocol first used for foraminifera by (Ren et al., 2009). The NO_3^- was converted to nitrous oxide (N_2O) using denitrifying bacteria *Pseudomonas chlororaphis* and its $\delta^{15}N$ was measured by an automated, custom-built N_2O extraction system and inlet by continuous helium carrier flow to an isotope ratio mass spectrometer (Thermo MAT 253) (Casciotti et al., 2002; Sigman et al., 2001; Weigand et al., 2016). To quantify the precision and accuracy of the corrected isotope values, for each series of 30 samples, a total of three different in-house (Max Planck Institute for Chemistry, MPIC) calcite and aragonite laboratory standards were analyzed in triplicate, namely a coral standard from the taxon *Porites* (PO-1) with $\delta^{15}N$ of 6.2 ± 0.3 ‰, a coral standard from the taxon *Lophelia* (LO-1) with $\delta^{15}N$ of 10.1 ± 0.4 ‰, and a mixed foraminifera standard (MF-1) (63-315 µm size fraction) from the North Atlantic (MSM58-17-1, Repschläger et al. (2018)) with $\delta^{15}N$ of 5.92 ± 0.28 ‰ (Moretti et al., 2024). After correcting for blank isotope composition, the precision of the multi-analysis average from each run is better than 0.2 ‰ (1SD).

2.2.2 Seawater nitrate nitrogen isotopes

Seawater samples from the South Atlantic A13.5 section were collected from full water column hydrocasts, spanning 32.0 °S to 41.5 °S. Acid-washed 60 mL HDPE bottles (Nalgene) were rinsed at least three times with sample water prior to filling and immediately freezing at −20 °C until analysis for NO_3^- concentration and $\delta^{15}N$. For comparison, we also measured seawater from the upper 1000 m of the water column in the subtropical North Atlantic gyre, collected during the CLIMODE (CLIvar MOde water Dynamics Experiment) campaign in 2006 (Hutto et al., 2006). Sampling stations are located across the northern boundary of the subtropical gyre between roughly 36.45 °N and 38.6 °N, which covers the sharp transition in water properties associated with the northern edge of the subtropical gyre (Fig. 1).

As with FB-$\delta^{15}N$ analysis, the $\delta^{15}N$ (*vs.* air) and $\delta^{18}O$ (*vs.* VSMOW) of NO_3^- were determined using the denitrifier method (Casciotti et al., 2002; Sigman et al., 2001). For seawater, denitrifying bacteria *Pseudomonas chlororaphis ssp. aureofaciens* (ATCC 13985, Manassas, VA, USA) were used to quantitatively convert NO_3^- and nitrite (NO_2^-) in samples to N_2O. The product N_2O was measured by a gas chromatography-isotope ratio mass spectrometer (Thermo MAT 253) (Weigand et al., 2016). For the shallow water samples, where NO_2^- constitutes over 10% of the total $NO_2^- + NO_3^-$ pool, mostly in the upper 200 meters, sulfamic acid was added to remove NO_2^- following the protocol described by (Granger and Sigman, 2009). The pooled standard deviation of all NO_3^--only and $NO_2^- + NO_3^-$ $\delta^{15}N$ measurements, respectively, were 0.24‰ (for shallow samples, <200 m) and 0.08‰ (for deeper samples). Two international potassium nitrate (KNO_3) reference materials, IAEA-N3 ($\delta^{15}N$ of 4.7‰, $\delta^{18}O$ of 25.6‰) and USGS-34 ($\delta^{15}N$ of −1.8‰, $\delta^{18}O$ of −27.9‰) were used for standardization, and a lab N_2O standard in helium was run in parallel to monitor the consistency of mass spectrometry.

2.3 Stable oxygen and carbon isotope analysis

For the down-core records of $\delta^{18}O$ and $\delta^{13}C$, we picked samples of all five species from the 250-400 µm size fraction throughout the core. For reference, the largest test sizes for *T. sacculifer* and *G. siphonifera* were observed in the 400-630 µm size fraction, and *G. truncatulinoides* in the >630 µm size fraction. To investigate the size fraction-specific $\delta^{13}C$ and $\delta^{18}O$, we chose specimens with sizes of 200-250 µm, 250-315 µm, 315-400 µm and > 400 µm from two climatically distinct (end-member) settings: an interglacial (129 ka, Marine Isotope Stage (MIS) 5) and glacial (27 ka, Last Glacial Maximum (LGM)) time period. There were not enough *G. bulloides* tests within the two largest size fractions (315-400 µm and > 400 µm); thus, size-specific $\delta^{13}C$ and $\delta^{18}O$ could not be evaluated for this species. For each sample measured, 30 specimens were gently cracked open with a watch glass, submerged in aliquots of ethanol (pure grade) and put in an ultrasonic bath for 20 seconds. Subsequently, the ethanol was decanted and the samples were dried at 60°C in a clean oven. Clean sample fragments were placed in 4.5 ml exetainer vials, flushed with helium and digested by anhydrous orthophosphoric acid (H_3PO_4) at 70°C. The liberated CO_2-He mixture was transported to the Thermo Finnigan GasBench II preparation device with He as carrier gas and analyzed using a Thermo Finnigan Delta V mass spectrometer (Breitenbach and Bernasconi, 2011). For each series of 60 samples, four different calcite laboratory standards were analyzed in quintuplicate. $CaCO_3$ standard weights were chosen so that they span the entire range of sample weights. VU-Internal-Carbonate Standard (VICS) was used to correct for sample size-based fractionation effects. VICS, IAEA and two in-house carbonate standards (Travertine, Carrara Marble) were used to detect mass spectrometer drift during the run and to quantify the precision and accuracy of the corrected isotope data. After correction for these effects, the reproducibility of the standards was typically better than 0.1 ‰ (1SD) for both $\delta^{18}O$ and $\delta^{13}C$.

2.4 Age model

The age model for DSDP 516 was produced by graphically aligning benthic foraminifera oxygen isotopes (Auderset et al., 2024) to global benthic stack LR04 by Lisiecki and Raymo (2005) using the software AnalySeries (Paillard et al., 1996)

3 Results

Downcore changes in FB-δ^{15}N and δ^{13}C are small relative to inter-species differences. The weak changes in δ^{15}N across the last 160 ka are discussed elsewhere (Auderset et al., 2024). Here we focus on the inter-species differences (Fig. 2, Fig. 3). FB-δ^{15}N data show two distinct groups: *T. sacculifer*, *G. ruber albus* and *G. ruber ruber* of ca. 4.2 ‰ (± 0.42), 4.1 ‰ (± 0.43), and 4.3 ‰ (± 0.37), respectively, *versus G. truncatulinoides*, *G. bulloides* and *G. siphonifera,* which show more elevated δ^{15}N values of 8.0 ‰ (± 0.67), 9.3 ‰ (± 0.45), and 8.8 ‰ (± 0.46), respectively (Fig. 2a, 3a). The N contents range between 3.8 nmol/mg and 4.5 nmol/mg (Fig. 2b, 3b). We observe similar N contents throughout the record and among the species. The studied foraminifera are well preserved, and the stability of the N content through the record implies little to no diagenetic loss of FB-N after burial through the sediment mixed layer (Fig. S1).

The δ^{18}O data show a fairly even spread between the different species (Fig. 2c, 3c). *Trilobatus sacculifer* and *G. ruber* (*albus* and *ruber*) are more depleted in ^{18}O (with δ^{18}O values of 0.3-0.5 ‰) compared to *G. truncatulinoides*, *G. bulloides,* and *G. siphonifera* (1.1-1.9 ‰). Over the last 160 ka, *T. sacculifer, G. ruber* (*albus* and *ruber*), and *G. siphonifera* generally follow the same trend. These species show a temporal variation of ~0.7-1 ‰ with a maximum at 40 ka and minimum at 90 ka (Fig. 2c, 3c). *Globorotalia truncatulinoides* and *G. bulloides* show a smaller amplitude of variation of ~0.5 ‰ and no clear δ^{18}O maximum/minimum (Fig 2c, 3c). Apart from *G. bulloides* with a δ^{13}C range between −1.6 ‰ and −0.3 ‰*,* all species range between 0.6 ‰ and 1.7 ‰. *Globigerina bulloides* δ^{13}C shows a substantial negative offset of ca. 1.5 ‰ from the other species and is more variable than all other species analyzed here (Fig. 2d, 3d).

All species investigated appear to show a positive relationship between test carbonate δ^{13}C and test size, although some reversals occur in the largest (400 μm) size fraction (data for *G. bulloides* are for the two smallest size fractions only) (Fig. S2). We record different slopes of the linear fit in LGM and MIS 5 samples (the two end-member settings) for *T. sacculifer* and *G. ruber* (*albus* and *ruber*) (Table S1). The Pearson correlation coefficient (r-value) for the LGM ranges between 0.60 and 0.98, and for MIS 5 between 0.68 and 0.96, with *G. siphonifera* and *G. truncatulinoides* having the lowest r-values among the studied species. The slopes for δ^{13}C *vs.* test size remain relatively constant for *G. siphonifera* (0.0013 *vs.* 0.0024) and *G. truncatulinoides* (0.0043 *vs.* 0.0038) between the LGM and MIS 5. However, the strength of the relationship differs between the two intervals, with r-values for *G. siphonifera* increasing from 0.60 during the LGM to 0.86 in MIS 5, and for *G. truncatulinoides* decreasing from 0.91 to 0.68. *T. sacculifer* exhibits the highest average slope for both time slices (0.053), while *G. siphonifera* records the lowest (0.018). Notably, significant relationships ($p < 0.05$) for δ^{13}C are observed in *G. ruber albus* ($p = 0.02$) and *G. ruber ruber* ($p = 0.03$) during the LGM and *T. sacculifer* ($p = 0.04$) during MIS 5.

In contrast to δ^{13}C, the test carbonate δ^{18}O shows only a weak correlation with test size (Table S1) and the relationship varies downcore and between species (Fig. S3). The only significant relationship is observed for *G.ruber albus* in the combined time slice ($p=0.02$).

Seawater nitrate nitrogen isotopes and nitrate concentration profiles from CLIMODE and A13.5 stations are shown in Fig. S4 and discussed in the Supplementary Material.

4 Discussion

4.1 FB-$\delta^{15}N$ at DSDP Site 516

In planktic foraminifera species *T.sacculifer* and *G.ruber*, dinoflagellate symbionts may recycle low-$\delta^{15}N$ ammonium and thus keep FB-$\delta^{15}N$ low, preventing the full ca. 3 ‰ elevation characteristic of a trophic level increase (Minagawa and Wada, 1984; Ren et al., 2012b; Smart et al., 2018). Therefore, the $\delta^{15}N$ of *T. sacculifer* and *G. ruber* should more closely match the $\delta^{15}N$ of their food source than in species without dinoflagellate symbionts. In oligotrophic areas, such as the subtropical South Atlantic gyre, surface nitrate is fully consumed by phytoplankton. With the role of euphotic zone N recycling described in Section 1 (Altabet, 1988; Fawcett et al., 2011), this leads to herbivorous zooplankton close to or slightly higher than the $\delta^{15}N$ of this consumed nitrate (e.g., 3.6 ± 1.0 ‰ (n = 107) for zooplankton *vs.* 2.6 ‰ for thermocline nitrate in the Sargasso Sea (Montoya et al., 2002; Smart et al., 2018). With foraminifera feeding on a mixture of small and large phytoplankton and zooplankton, this yields a $\delta^{15}N$ for the foraminifera diet that is close to the $\delta^{15}N$ of the subsurface nitrate supply (Fawcett et al., 2011). At DSDP Site 516, the FB-$\delta^{15}N$ of the dinoflagellate-bearing species *T. sacculifer* and *G. ruber* (Gastrich, 1987; Hemleben et al., 1989) ranges between 4 ‰ and 5 ‰. This value is lower than global mean pycnocline nitrate (of 6.25‰; Fripiat et al. (2021)), suggesting that regional subsurface nitrate $\delta^{15}N$ has been lowered by N_2 fixation, as has been observed in other subtropical gyres (Harms et al., 2019; Knapp et al., 2008; Marshall et al., 2022; Casciotti et al., 2008; Liu et al., 1996; Yoshikawa et al., 2015) especially the North Atlantic (Marconi et al., 2015; Marconi et al., 2017). For comparison, in the Sargasso Sea – an ocean region with persistent N_2 fixation – shallow thermocline nitrate $\delta^{15}N$ is 2.6‰ (Fawcett et al., 2015) and surface sediment FB-$\delta^{15}N$ are 2.4‰ (*T. sacculifer*) and 2.6‰ (*G. ruber*) (Smart et al., 2018).

In *G. truncatulinoides*, the bulk of the shell derives from calcification below 100 m, indicating that they spend much of their lives below the euphotic zone, where light conditions are unsuitable for photosymbionts (Reynolds et al., 2018; Schiebel and Hemleben, 2017). This is consistent with their high $\delta^{15}N$ compared to dinoflagellate symbiont bearers. *G. bulloides* and *G. siphonifera* are more enriched in ^{15}N than the other analyzed species at DSDP Site 516. This may indicate minimal nitrogen recycling associated with non-dinoflagellate symbionts, e.g. chrysophytes for *G. siphonifera*, such that this species groups with the symbiont-barren species (Faber et al., 1989; Faber et al., 1988; Ren et al., 2012b; Smart et al., 2018; Granger et al., 2024). As for the higher $\delta^{15}N$ of *G. bulloides* and *G. siphonifera* relative to *G. truncatulinoides,* this might be explained by a higher $\delta^{15}N$ for their food source. This possibility is pursued in Section 4.4.

4.2 Comparison with carbonate oxygen and carbon isotopes at DSDP Site 516

To gain additional insight into the controls on FB-$\delta^{15}N$, we compare the data with our measurements of carbonate $\delta^{18}O$ and $\delta^{13}C$ of the foraminiferal shells. The $\delta^{18}O$ differences among species have been used to reconstruct depth habitat . Minimum $\delta^{18}O$ values in foraminiferal assemblages point to warmer waters and shallower depth habitats, which are likely to be required or at least best-suited for photosymbiont-bearing foraminifera (Fig. 4a) (see also Supplementary Material). By cross-plotting FB-δ^{15}N and δ^{18}O from Site 516, we see two distinct groups: shallow-dwellers with dinoflagellate symbionts with lighter FB-δ^{15}N and δ^{18}O, and deeper-dwellers with non-dinoflagellate symbionts/ symbiont-barren recording heavier FB-δ^{15}N and δ^{18}O (Fig. 4b). Thus, unsurprisingly, the measurements are consistent with a shallow depth habitat for photosymbiosis.

The controls on the δ^{13}C of planktic foraminiferal calcite are a matter of debate; however, size-specific δ^{13}C measurements have been proposed as a proxy for photosymbiosis. Foraminiferal δ^{13}C is likely sensitive to both environmental and organism-scale δ^{13}C gradients (induced by photosymbiont activity and foraminifera respiration), with a ^{13}C enrichment for increased photosynthetic activity (Spero et al., 1997; Spero and Deniro, 1987). However, the δ^{13}C of deepest dwelling, symbiont barren species *G. truncatulinoides* at DSDP Site 516*,* does not distinguish from even the most photosymbiotically active species (Fig. 2d, Fig. 3d). This is because depth habitat, variation in dissolved inorganic carbon, remineralization and other environmental factors also influence carbonate δ^{13}C, such that it alone is unlikely to be a reliable proxy for photosymbiosis. Surface mixed layer DIC is typically enriched in ^{13}C due to photosynthesis, whereas subsurface water DIC δ^{13}C is lower due to bacterial respiration and remineralization of the low-δ^{13}C organic matter arriving from the euphotic zone (Kroopnick, 1985). In a stratified ocean, such as the South Atlantic subtropical gyre, shallow-dwelling foraminifera should thus incorporate a higher δ^{13}C in contrast to thermocline-dwellers with lower δ^{13}C. Yet at DSDP Site 516, modern surface ocean $\delta^{13}C_{DIC}$ seems to be only marginally higher (~ 1.7 ‰) than the $\delta^{13}C_{DIC}$ in the upper thermocline (~ 1.4 ‰) (Fig. S7), which could explain why calcite δ^{13}C of *T. sacculifer* and *G. truncatulinoides* at Site 516 are so similar. In our measurements, shallow subsurface/ thermocline-dweller *G. bulloides* is the only species to be clearly lower in δ^{13}C than the other foraminifera. It has been speculated that the presence of metabolically active cyanobacterial endobionts (*Synechococcus*) affects the carbon isotopes measured in the host *G. bulloides* through respiration, instead of photosynthesis from the endobiont, leading to ^{13}C depletion instead of ^{13}C enrichment in the calcifying microenvironment (Bird et al., 2017; Febvre-Chevalier, 1971; Spero and Lea, 1996). Alternatively, we suspect that this offset could be due to other effects associated with seasonality (Section 4.4.2) or a different environmental baseline for foraminifera originating from outside the South Atlantic gyre (Section 4.4.3).

In any case, the cross-plot between FB-δ^{15}N and δ^{13}C strongly discriminates between *G. bulloides versus* the other foraminifera analyzed at Site 516 (Fig. 4c). Interestingly, *G. siphonifera* clusters in the same group as *G. truncatulinoides*, although both species are very different in terms of depth habitat and symbiotic relationship. Altogether, the δ^{15}N-δ^{13}C comparison might be interpreted as indicating that the non-dinoflagellate symbionts have a significant but variable effect on carbonate δ^{13}C while having no appreciable effect on FB-δ^{15}N (Fig. 4c). However, this view is complicated for δ^{13}C by the observation that *G. truncatulinoides,* which is symbiont-barren, has a similar δ^{13}C to species with a range of symbionts.

An alternative method proposed for the detection of photosymbiosis in fossil foraminifera is through the relationship of foraminiferal carbonate δ^{13}C with test size (Spero and Deniro, 1987). The argument is that the preferential uptake of $^{12}CO_2$ by the photosynthesizing symbionts raises the δ^{13}C of the internal DIC pool from which the calcification proceeds (Spero et al., 1997). Larger individual foraminifera can host more symbionts per surface area of the individual, resulting in δ^{13}C elevation compared to smaller foraminifera (Spero et al., 1991). Thus, a steeper rise in δ^{13}C with increasing specimen size (steeper positive slope) indicates potentially higher photosymbiotic activity (Edgar et al., 2013; Norris, 1996; Shaw et al., 2021). According to this method, dinoflagellate-bearing *G. ruber* and *T. sacculifer* and potentially chrysophyte-hosting *G. siphonifera* would be predicted to have a steeper rise in $\delta^{13}C$ with size than *G. truncatulinoides* or *G. bulloides*. However, for all investigated species except *G. siphonifera,* our measurements indicate similar slopes in $\delta^{13}C$ *vs*. size, with dinoflagellate bearing species *T. sacculifer*, and *G. ruber* (*albus* and *ruber*) showing no statistically significant difference in slopes compared to non-symbiotic species *G. truncatulinoides* (Fig. S2, Table S1). The weak/ near-zero slope for *G. siphonifera* $\delta^{13}C$ (Fig. S2e) could be the result of the size fraction between 125-400 µm used in this study not capturing the larger, adult life stages of this species; we also observed a small number of *G. siphonifera* in the 400-630 µm fraction but did not measure them. Nevertheless, the anomalously weak/near-zero slope for *G. siphonifera* $\delta^{13}C$ has been observed before. Bijma et al. (1998) showed a steeper slope for *G. siphonifera* type II than type I. They proposed that *G. siphonifera* type I symbionts are less efficient at removing low-$\delta^{13}C$ host-respired CO_2, allowing respiration to partially counteract the photosynthesis-induced $\delta^{13}C$ rise in the microenvironment and shells during ontogeny. The significant contribution of host-respired CO_2 to symbionts in *G. siphonifera* type II is supported by the recent experimental study by Takagi et al. (2022).

That *G. truncatulinoides* falls into the same range of slopes (as well as absolute $\delta^{13}C$ values) as *T. sacculifer* and *G. ruber* and has a steeper slope than *G. siphonifera* indicates that either the size fraction was too low and we observe the difference between juveniles and adults in *G. truncatulinoides* or that the previously proposed symbiosis metric of $\delta^{13}C$ *vs*. size may not be sensitive enough to capture all photosymbiotic relationships.

However, using FB-$\delta^{15}N$ as a symbiosis proxy may also have its complexities. In particular, the FB-$\delta^{15}N$ differences between foraminifera with and without dinoflagellate symbionts is notably greater at DSDP Site 516 than reported at other sites across the global ocean (Costa et al., 2016; Martínez-García et al., 2014; Ren et al., 2012b; Schiebel et al., 2018; Smart et al., 2020; Smart et al., 2018). Interspecies FB-$\delta^{15}N$ relationships might vary due to processes in the water column, such as seasonal changes in the supply of N affecting the $\delta^{15}N$ of the food source (Smart et al., 2020; Smart et al., 2018) or lateral transport (Granger et al., 2024). In the next section we compare DSDP Site 516 with core top samples from geographically distinct regions to examine the variability of FB-$\delta^{15}N$ differences amongst foraminifera species relative to hydrologic environment.

4.3 FB-$\delta^{15}N$ as a new proxy for photosymbiosis?

To further advance our understanding of FB-$\delta^{15}N$ and its suitability as a metric of photosymbiosis, we now broaden our view beyond DSDP Site 516. We compile all available and published core top FB-$\delta^{15}N$ data that contain dinoflagellate bearing and non-dinoflagellate bearing foraminifera (Fig. S5, Table S2). The calculated offsets between the two categories across all core sites show that non-dinoflagellate bearing foraminifera (*Globorotalia menardii, Neogloboquadrina dutertrei, G. siphonifera, Globorotalia tumida, Globorotalia hirsuta, Globorotalia inflata, G. truncatulinoides, G. bulloides*) are consistently enriched in ^{15}N compared to dinoflagellate bearing foraminifera (*G. ruber, T. sacculifer, O. universa*) (Fig. 5a). For the most part, FB-$\delta^{15}N$ offsets are consistent across wide geographic ranges of the core sites over different latitudes and biogeochemical regimes.

With this compilation, we first consider the average FB-δ^{15}N values that arise for different species and their implications for FB-δ^{15}N differences among different symbioses (Fig. 5b, Fig. 6).

The calculated mean FB-δ^{15}N offsets between dinoflagellate bearing and non-dinoflagellate/symbiont-barren foraminifera from the global compilation correlate with chlorophyll-*a* (Chl-*a*) measurements by Takagi et al. (2019). They analyzed living foraminifera from the central and western Pacific Ocean and the tropical eastern Atlantic Ocean across different seasons and estimated the intensity of photosymbiosis (Fig. 7). Based on Takagi et al. (2019), *O. universa*, *T. sacculifer, G. ruber ruber* and *G. ruber albus* have a higher percentage of intracellular Chl-*a* and thus photosymbiont activity. This is in contrast to *G. truncatulinoides, G. bulloides* and *G. inflata* with little/no Chl-*a* inside of the tests, which record the highest FB-δ^{15}N compared to dinoflagellate bearing foraminifera (*G. ruber, T. sacculifer, O. universa*) (Fig.5b). *Neogloboquadrina dutertrei* and *G. menardii* are consistently higher in Chl-*a*/biomass and lower in their FB-δ^{15}N offset than photosymbiont-barren *G. bulloides* and *G. truncatulinoides*. However, the strength of the Chl-*a* /biomass *vs.* FB-δ^{15}N relationship is not extraordinarily strong (r = 0.73), suggesting that Chl-*a* content is not the only factor driving FB-δ^{15}N (Fig. 7).

Globigerinella siphonifera stands out in the correlation of Chl-*a* /biomass in the global compilation (Fig. 7), as it records similar levels of Chl-*a*/biomass compared to some dinoflagellate symbiont-bearing foraminifera *G. ruber (ruber and albus)*, despite their high FB-δ^{15}N offset from dinoflagellate-hosting foraminifera reported at DSDP Site 516 (Fig. 3a) and in previous studies (Li et al., 2019; Ren et al., 2012b; Smart et al., 2018; Granger et al., 2024). Takagi et al. (2016) find that the chlorophyll content of *G. siphonifera* peaks before the final chamber formation, resulting in a minimum chlorophyll content during formation of the largest-second-largest chamber, which likely dominate calcite mass (and thus geochemistry). This may reflect digestion of its symbionts prior to gametogenesis(Faber et al., 1988), making *G. siphonifera* symbiont-barren in its later stages of calcification. In any case, this ontogenetic change could explain why FB-δ^{15}N would identify *G. siphonifera* as having a weaker host-symbiont N cycle. As an alternative or additional explanation, the high FB-δ^{15}N and relatively high Chl-*a* of *G. siphonifera,* may indicate a less active symbiotic relationship, whether due to less efficient internal ammonium recycling (Smart et al., 2018), low chrysophyte growth rates, and/or higher harvesting rates.

All such interpretations, however, are currently uncertain. Across species of foraminifera, symbiont physiology, photosynthetic rates, and host-symbiont interactions may vary, with unforeseen impacts on the N isotopes. Moreover, fixed N may enter foraminifera through more than just feeding. Depending on the physiology of the endosymbionts, their host, and environmental conditions, direct nitrate uptake by the foraminifera host and/or its symbionts is a possibility (Piña-Ochoa et al., 2010; Uhle et al., 1999).

Regardless, the uniquely low FB-δ^{15}N of the dinoflagellate-bearing species supports higher photosynthetic rates (averaged over their lifetime) in dinoflagellate symbionts and, thus, a generally more important role for the symbiosis in the dinoflagellate-bearing species (e.g., as in *O. universa*; LeKieffre et al. (2020)). The shallow depth (i.e. high-light) habitats of the dinoflagellate species is fully consistent with this interpretation, as is the dominance of dinoflagellate symbioses among modern symbiotic corals (Davy et al., 2012).

4.4 Deviations from the FB-δ^{15}N photosymbiosis paradigm at Site 516

Despite the overall consistency observed across sites in the global FB-δ^{15}N compilation, there are notable differences in inter-species offsets between the global compilation and DSDP Site 516 (Fig. 5a). In particular, among the foraminiferal species analyzed, the FB-δ^{15}N differences between the three dinoflagellate-bearing species and the three other species (*G. truncatulinoides, G. bulloides, G. siphonifera*) are unusually great at DSDP Site 516 (Fig. 8). These differences warrant investigation here, with a focus on their cause and thus their implications for interspecies FB-δ^{15}N differences as a paleoproxy for foraminiferal photosymbiosis.

4.4.1 Depth habitat

The isotopic composition of suspended PN varies systematically with depth in the water column (Altabet, 1988; Altabet et al., 1991). Suspended PN has its lowest δ^{15}N in the euphotic layer, increasing with depth below the euphotic zone as a consequence of the preferential release of ^{14}N-ammonium during microbial decomposition. The PN δ^{15}N increase from the surface to ~500 m depth can be up to 6 ‰ (Altabet, 1988; Altabet et al., 1991; Hannides et al., 2013). Therefore, planktic foraminifera living below the euphotic zone might be expected to have a higher δ^{15}N for their feeding source than surface-dwellers.

At Site 516, our N isotope results show clear differences between two groups of species. The group of *G. truncatulinoides*, *G. bulloides,* and *G. siphonifera* have a δ^{15}N composition that is 3 to 5 ‰ higher than the group of *T. sacculifer* and *G. ruber*. These observations are in good agreement with previous studies (Ren et al., 2012b; Smart et al., 2018), including samples from upper-ocean net tows (surface 200 m), moored sediment traps, core-tops, and down-core sediments. Planktic foraminifera living below the euphotic zone, when feeding on suspended PN, such as *G. truncatulinoides*, could incorporate food that is enriched in ^{15}N (Mintenbeck et al., 2007). To date, there are not yet any published depth profile data for PN or nitrate isotopes close to core Site 516. Based on data from the North Atlantic (Altabet, 1988; Altabet et al., 1991) and South Atlantic surface particles (Mino et al., 2002), PN δ^{15}N may be up to 9 ‰ higher at 1000 m water depth.

However, several lines of evidence suggest that depth habitat is not the main cause of the difference in δ^{15}N between the two groups. The two morpho- and geno-types of *G. siphonifera* (Types I and II) are surface-to-thermocline dwellers (Bijma et al., 1990; Schiebel and Hemleben, 2017), and yet have a higher δ^{15}N than the deeper dwelling *G. truncatulinoides* (Hemleben et al., 1985) (Fig. 2a). Other differences in depth habitat inferred from foraminiferal δ^{18}O and δ^{13}C (Supplementary Material, Figs. S6, S7) are also not consistent with the inter-species δ^{15}N offsets that would be expected if depth were the main driver of FB-δ^{15}N. For example, of the species we measured at Site 516, *G. truncatulinoides* is the deepest dweller according to its δ^{18}O values, but it has a δ^{15}N that is lower than that of *G. siphonifera* and *G. bulloides*. In addition, at DSDP Site 516, *G. siphonifera* δ^{15}N is more similar to *G. truncatulinoides* during interglacials but more similar to *G. bulloides* during glacials. Yet the δ^{18}O of *G. siphonifera* maintains its position in δ^{18}O space relative to *T. sacculifer* and *G. ruber*, which would indicate no major change in the relative depth habitat of *G. siphonifera* through the glacial cycle. These observations suggest that depth habitat *per se* is not the main driver of the grouping we see in the FB-δ^{15}N data. Depth also cannot explain the larger than average FB-δ^{15}N offset at Site 516, given the higher FB-δ^{15}N values for *G. siphonifera* and *G. bulloides* than for *G. truncatulinoides*.

4.4.2 Seasonality

Seasonality can also play a role in determining which nitrogen isotope signals are incorporated by foraminifera. While total foraminifera production often follows the seasonality of phytoplankton productivity in a given environment, there are major differences among species (Schiebel et al., 2001). In temperate latitudes and parts of the subtropical gyres, most nitrate supply from below occurs during the winter, with a nitrate-fueled phytoplankton bloom in the spring giving way to intensive ammonium recycling between phytoplankton and zooplankton in the summer-autumn (Dugdale and Goering, 1967; Eppley and Peterson, 1979). Depending on the most productive season of a given species of planktic foraminifera, it may feed within a nitrate-based or ammonium-based ecosystem, with the latter being associated with a lower δ^{15}N for suspended PN, zooplankton, and sinking PN (Fawcett et al., 2011). This distinction is apparent in the seasonality of foraminifera δ^{15}N (Smart et al., 2020; Smart et al., 2018). Thus, if we assume the seasonality of production is similar in the South Atlantic compared to the North Atlantic gyre, then we would expect Site 516 summer-bloomers *T. sacculifer* and *G. ruber* to be lower in δ^{15}N than *G. truncatulinoides*, which subsist in deep waters year-round but exhibit peak fluxes to the seafloor in winter-spring (Deuser, 1987; Hemleben et al., 1985; Salmon et al., 2015) when they reproduce in surface waters around the time of peak phytoplankton production (Schiebel et al., 2002; Reynolds et al., 2018). However, the offset that we observe between *T. sacculifer* and *G. ruber versus G. truncatulinoides* is 4-4.5 ‰, which is substantially higher than typical seasonal offsets between summer and winter PN δ^{15}N in oligotrophic regions (e.g., Smart et al. (2018)). Thus, if seasonality is the explanation for the particularly large FB-δ^{15}N inter-species offsets at DSDP Site 516, it would appear that some special source of seasonality is required for the South Atlantic subtropical gyre.

4.4.3 Lateral transport

In austral winter-spring, between July and September, the oligotrophic South Atlantic gyre is affected by wind-driven mixing and an Ekman transport-driven incursion of nutrient-rich waters with high-δ^{15}N nitrate from the Southern Ocean (Figs. 1, S8). *Globigerina bulloides,* perhaps the most opportunistic among the species discussed here, is abundant at the southern margin of the Atlantic subtropical gyre around 35ºS (Boltovskoy, 1962), thus recording the higher δ^{15}N associated with high degrees of nitrate assimilation from a nitrate source with a δ^{15}N that is similar to that of Subantarctic Mode Water (Smart et al., 2015). Indeed, it has been shown that *G. bulloides* reaches highest abundances when food supply is increased (Kretschmer et al., 2018; Mortyn and Charles, 2003; Schiebel et al., 1995). *Globigerinella siphonifera* although it is a subtropical species, similarly yields highest shell fluxes at times of highest bioproductivity (Jonkers and Kučera, 2015), which would be the winter-spring season in the South Atlantic. For both *G. bulloides* and *G. siphonifera*, we observed highly variable specimen counts across the last 160 ka (Fig. S9), in strong contrast to the uniformly high abundances of *T. sacculifer*, *G. ruber albus* and *G. truncatulinoides* throughout the record. Therefore, the large FB-δ^{15}N offset between summer-blooming species (*T. sacculifer* and *G. ruber*) and *G. bulloides* and *G. siphonifera* at Site 516 could reflect increased nutrient influx from the Southern Ocean with an elevated δ^{15}N signature in austral winter-spring during northward movement of the subtropical front. As previously documented, not only nutrients but also planktic foraminifera and/or their particulate food sources (PN) can be transported from other oceanic regions to the South Atlantic (Peeters et al., 2004; Smart et al., 2020; Granger et al., 2024).

Here we compare the nitrate profiles for "inside-gyre" (BATS, Fawcett et al. (2015)) *vs.* "outside-gyre" (CLIMODE, this study) stations in the North Atlantic (Fig. 9a) to evaluate baseline (nitrate δ^{15}N) differences between the two regions, providing a reference point for the baseline (nitrate δ^{15}N) differences we could expect in the South Atlantic. Subsurface nitrate at 200 m inside the North Atlantic gyre is ca. 3 ‰ lower than subsurface nitrate outside the gyre, which is due to gyre-related N_2-fixation processes keeping thermocline nitrate δ^{15}N low. A similar upper-ocean nitrate δ^{15}N gradient is observed in other subtropical gyres in the Indian and Pacific Ocean with ~2.5‰ lower δ^{15}N inside the gyre *vs*. outside (Harms et al., 2019; Yoshikawa et al., 2018; Yoshikawa et al., 2015; Marconi et al., 2024; Marshall et al., 2023). While no similar data exists from inside the South Atlantic gyre, a hypothetical "inside gyre" δ^{15}N profile based on the southern "outside gyre" would put the subsurface nitrate δ^{15}N at 4 ‰ (inside gyre) *versus* 7 ‰ (outside gyre) (Fig. 9b), closely matching the FB-δ^{15}N of *T. sacculifer* and *G. ruber* at Site 516. Symbiont-barren *G. truncatulinoides* has the expected ~ 3‰ δ^{15}N offset above "inside-gyre" nitrate, i.e., the full trophic enrichment typically observed for marine zooplankton (Minagawa and Wada, 1984). *G. bulloides* and *G. siphonifera,* in contrast, appear to be elevated by an additional 1-2 ‰. This additional offset could be due to a Southern Ocean influence, where subsurface nitrate derived from Subantarctic Mode Water is enriched in ^{15}N due to incomplete nutrient consumption (Sigman et al., 2000; Smart et al., 2015). Indeed, the distinct δ^{13}C of *G. bulloides* has been interpreted as reflecting its Southern Ocean origin (Ninnemann and Charles, 1997) (Supplementary Material, Fig. S7). In our interpretation, both the relatively high δ^{15}N of Subantarctic-sourced nitrate and the greater potential for incursion of extra-subtropical water into the small South Atlantic gyre contribute to the abnormally large FB-δ^{15}N difference, at DSDP Site 516, between *G. bulloides/G. siphonifera* and the dinoflagellate-bearing species. The South Atlantic may be more prone to these effects than the North Atlantic gyre or other sites from the core top compilation due to the smaller geographic extent of the gyre and/or its proximity to the Southern Ocean.

5 Conclusions

FB-δ^{15}N is a promising new geochemical tool to identify fossil foraminiferal photosymbiosis, in particular, the strong symbiosis with dinoflagellates, and also has the potential to identify symbiosis with non-dinoflagellate symbionts (e.g., chrysophytes and pelagophytes) (Figs. 5, 6, 10). The FB-δ^{15}N range of the non-dinoflagellate symbiont group may indicate that non-dinoflagellate symbioses in foraminifera are relatively weak in terms of the internal N demand they generate, which could indicate that they generate substantially less photosynthetic energy for the host than do dinoflagellate endosymbionts. This possibility warrants testing with other approaches.

While FB-δ^{15}N offsets are remarkably consistent in our global compilation, DSDP Site 516 stands out with its larger than expected FB-δ^{15}N offsets between dinoflagellate-bearing foraminifera (*G. ruber* and *T. sacculifer*) and the non-dinoflagellate bearing *G. siphonifera* and *G. bulloides*. Site 516 sits near the core of the uniquely small South Atlantic subtropical gyre. As a subtropical gyre, it hosts convergence, fed by inflow from the surrounding surface ocean. Accordingly, we propose that DSDP Site 516 is influenced by the input to the region of *G. bulloides* and *G. siphonifera* specimens and/or their N sources that were sourced from the higher-δ^{15}N nitrate of the Subantarctic Zone to the South and/or the more proximal temperate ocean waters surrounding the gyre. With this caveat regarding regions with strong environmental gradients, our findings support the use of inter-species FB-δ^{15}N offsets to investigate the emergence and evolution of photosymbiosis in foraminifera and potentially other animals.

Data availability. The data is available in Supplementary Data and in the PANGAEA database.

Competing interests. The authors declare that they have no conflict of interest.

Author contribution. A.A. and A.M.-G. designed the study. A.A. measured foraminifera-bound nitrogen isotopes in the lab of A.M.-G.. Y.R. and D.M. measured seawater nitrate nitrogen isotopes. A.A., S.M.S, D.M.S. and A.M.-G. wrote the manuscript. All authors contributed to the interpretation of the data and provided input to the final manuscript at different stages of the project.

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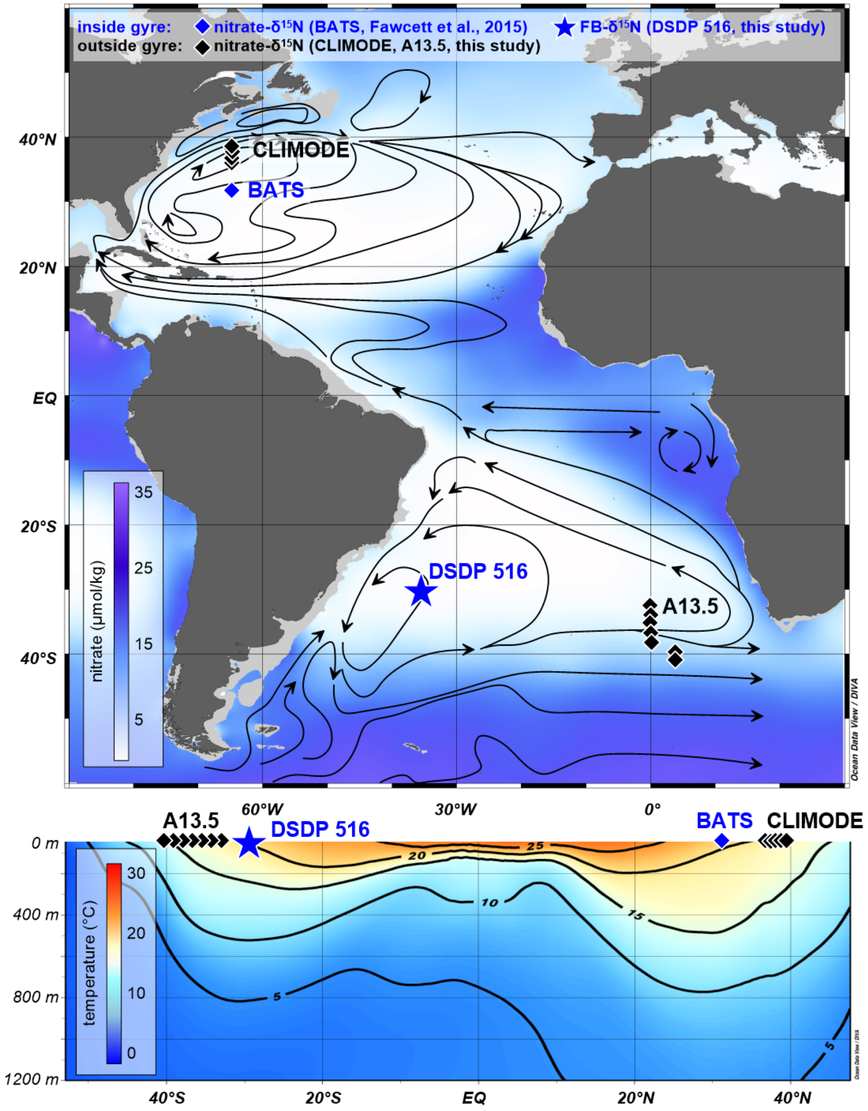
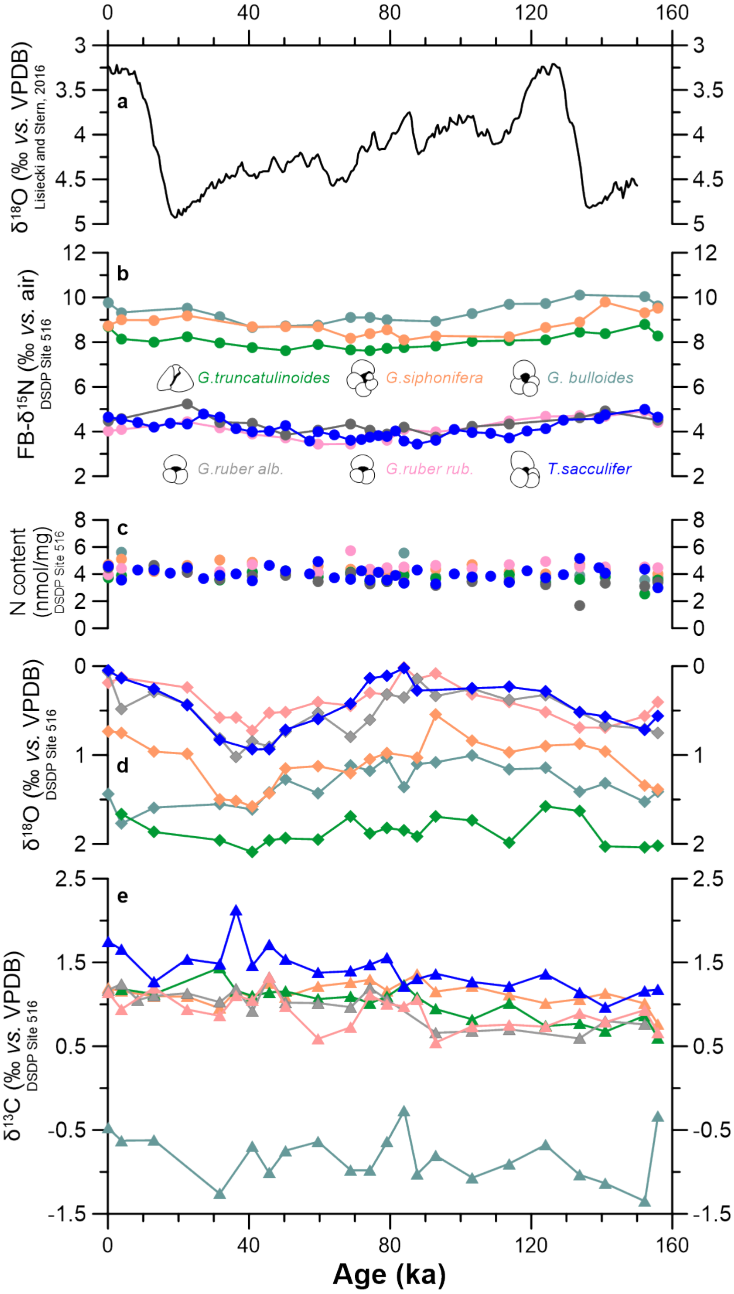
Figures



480 **Figure 1. Oceanographic context for DSDP Site 516.** (**a**) Nitrate map (in µmol/kg) at 100 m water depth with sample location DSDP Site 516 for FB-δ15N (this study) and nitrate δ15N profiles from CLIMODE (this study), BATS (Fawcett et al., 2015) and A13.5 (this study). Main surface currents and oceanographic fronts modified from (Peterson and Stramma, 1991; Wefer et al., 1996) and (Schmitz Jr. and McCartney, 1993) (**b**) Mean annual temperature cross section. Map and cross section generated with Ocean Data View and data set from World Ocean Atlas (Garcia et al., 2013; Schlitzer, 2015).



485 **Figure 2. 160 kyr time series of species-specific nitrogen, carbon and oxygen isotopes at DSDP Site 516. (a)** benthic foraminifera δ^{18}O stack (Lisiecki and Stern, 2016). MIS = Marine Isotope Stages. **(b)** Species-specific FB-δ^{15}N measured at DSDP 516 for *G. bulloides* (turquoise), *G. siphonifera* (orange), *G. truncatulinoides* (green), *T. sacculifer* (blue), *G.ruber ruber* (pink), and *G. ruber albus* (grey). **(c)** N content for each species. **(d)** Species-specific calcite-δ^{18}O and **(d)** calcite-δ^{13}C.

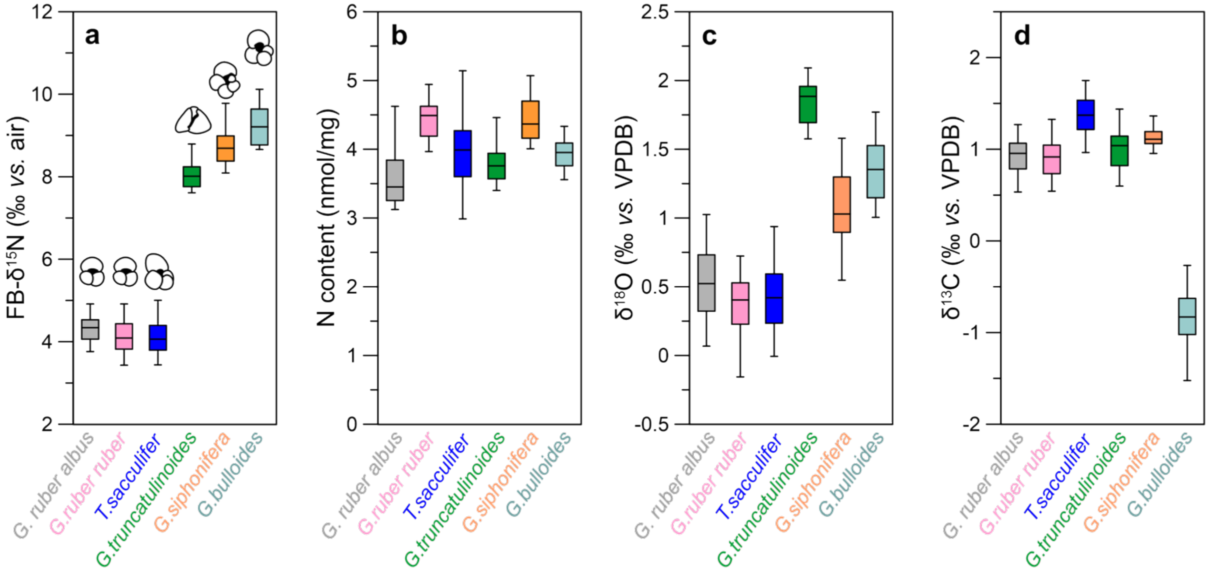


Figure 3. Box plots of species-specific nitrogen, carbon and oxygen isotopes at DSDP Site 516. (a) Species-specific FB-δ^{15}N values averaged over the last 160 ka for *G. bulloides* (turquoise), *G. siphonifera* (orange), *G. truncatulinoides* (green), *T. sacculifer* (blue), *G.ruber ruber* (pink) and *G.ruber albus* (grey). **(b)** N content for each species. **(c & d)** Species-specific δ^{18}O and δ^{13}C, respectively.

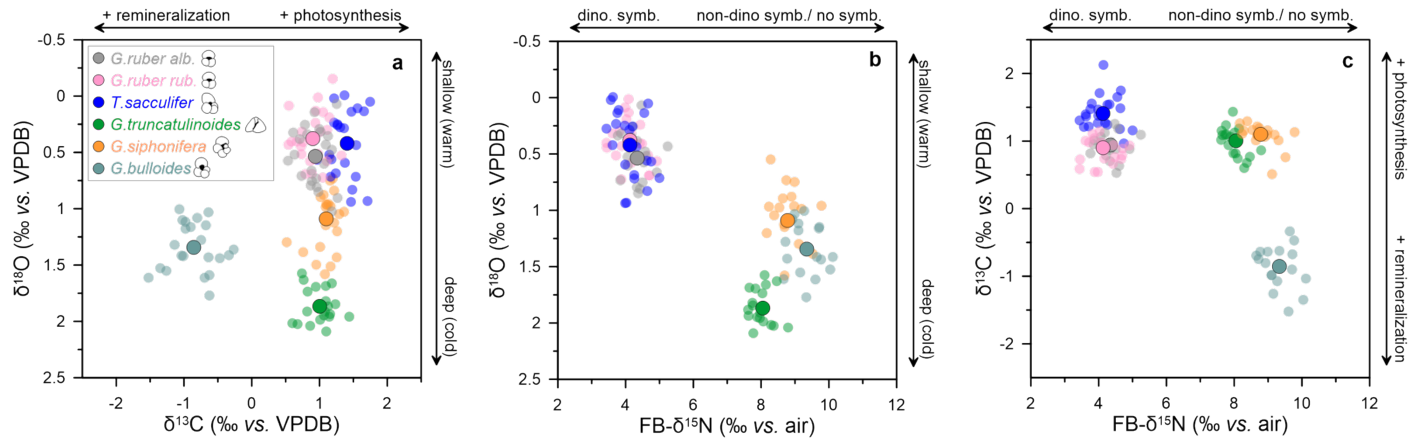


Figure 4. Relationships between oxygen, carbon and nitrogen isotopes. Interspecies comparison between *G. ruber* (*albus* and *ruber*), *T. sacculifer, G. truncatulinoides*, *G. siphonifera* and *G. bulloides* at DSDP Site 516. Arrows indicate distinction between shallow (warm) *vs.* deep (cold) dweller, remineralization *vs.* photosynthesis in the water column and non-dinoflagellate bearing/non-symbiotic *vs.* dinoflagellate bearing foraminifera.

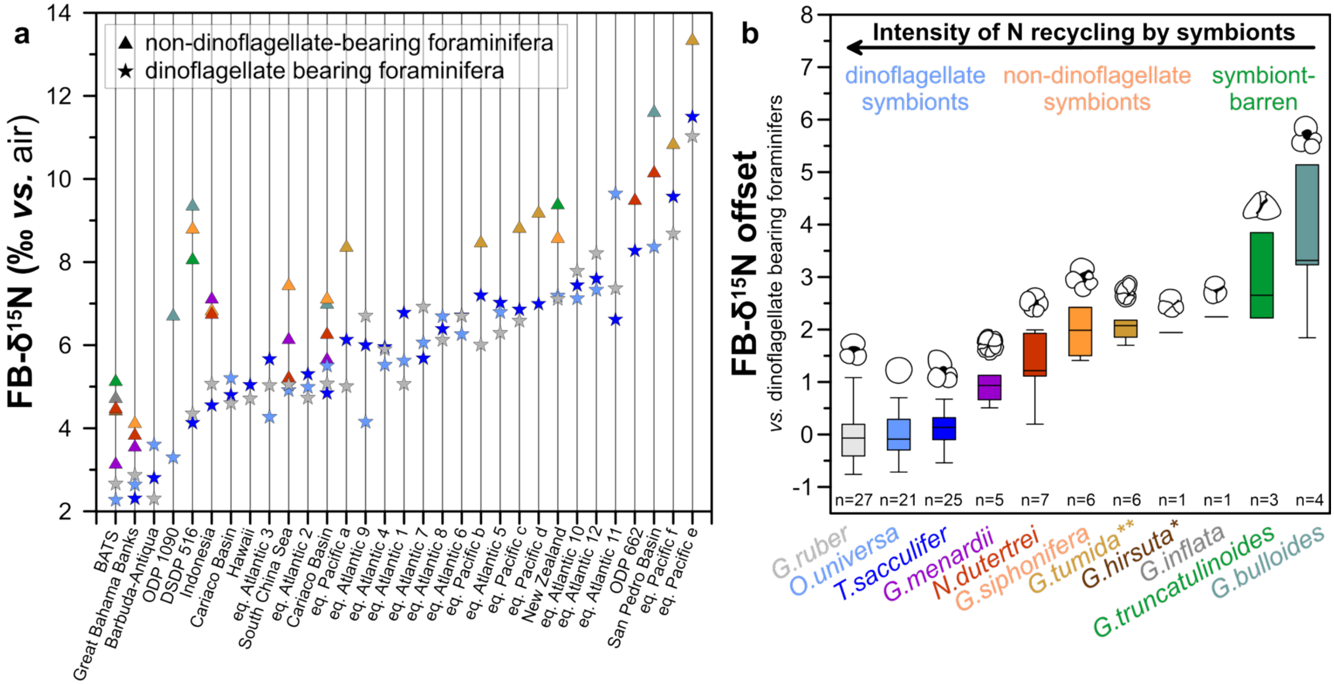


Figure 5. Compilation of FB-δ^{15}N values and offsets between dinoflagellate-bearing and non-dinoflagellate-bearing foraminifera from different locations. (**a**) Foraminifera from core tops in the Sargasso Sea (BATS) (Smart et al., 2018), Great Bahama Banks, Indonesia, Hawaii, South China Sea, New Zealand (Ren et al., 2012b), equatorial Atlantic 1-12 (Schiebel et al., 2018) as well as Holocene samples from the equatorial Pacific a-f (Costa et al., 2016), ODP 1090 (Martínez-García et al., 2014) and ODP 662 (Auderset et al., 2024) (See Table S2 and Fig. S5 for more information about core sites). **(b)** Across all sites in (a), the average FB-δ^{15}N offset between and dinoflagellate-barren and dinoflagellate-bearing foraminifera, categorized by symbiont relationship and proposed intensity of internal ammonium recycling. * = suspected chrysophyte symbionts/ barren, ** = unknown symbiotic status.

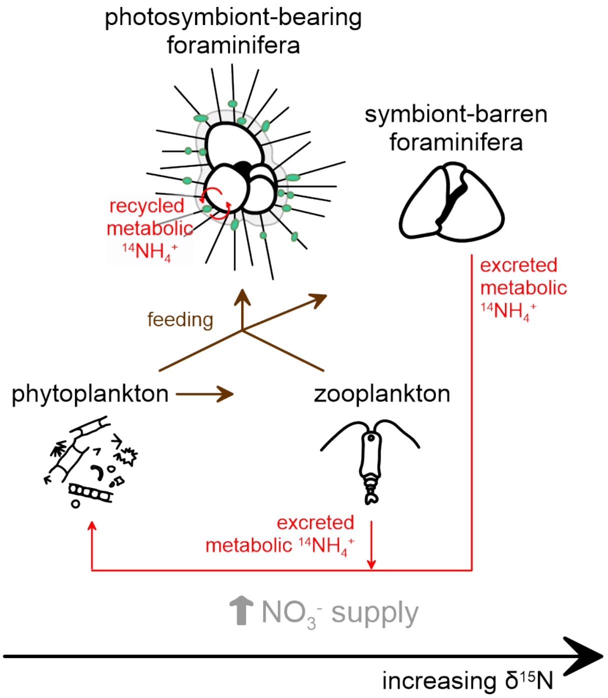


Figure 6. Proposed interactions and processes influencing planktic foraminifera-bound nitrogen isotopes (FB-δ^{15}N) in relation to symbiotic status and food sources. Symbiont-barren foraminifera, or those hosting symbionts other than dinoflagellates, record elevated FB-δ^{15}N relative to their food source, as with typical zooplankton. In contrast, symbiont-bearing foraminifera exhibit lower FB-δ^{15}N values due to their association with photosymbionts, which internally reassimilate the low-δ^{15}N metabolic ammonium (NH_4^+) from the foraminiferal host through photosynthesis, preventing or reducing the δ^{15}N elevation that typically results from heterotrophy.

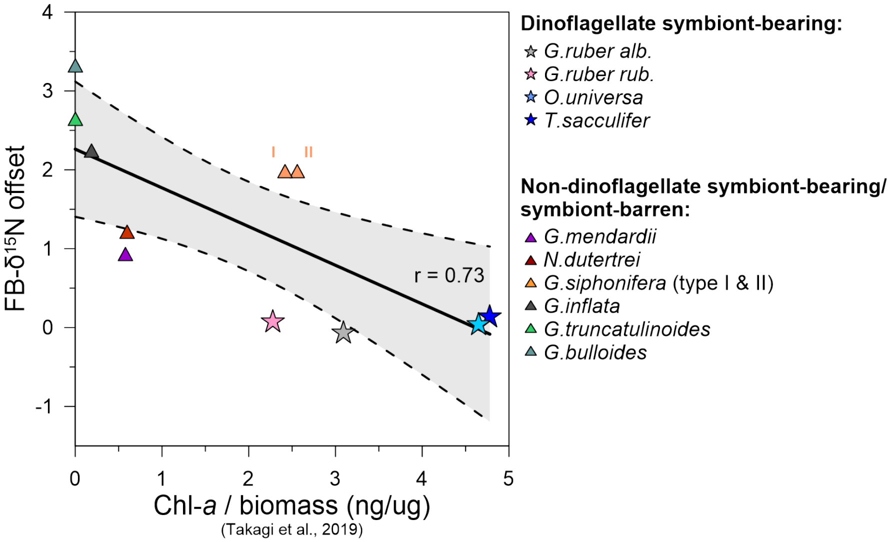


Figure 7. Chlorophyll-*a* concentrations in living planktic foraminifera *vs.* FB-δ^{15}N offsets between dinoflagellate-bearing and other non-dinoflagellate-bearing foraminifera. Chlorophyll-*a*/biomass values were measured in foraminifera collected in the central and western Pacific Ocean and the tropical eastern Atlantic Ocean across different seasons (Takagi et al., 2019) and compared to the median of the FB-δ^{15}N offsets in our core top compilation (Fig. 5, S5, Table S2).

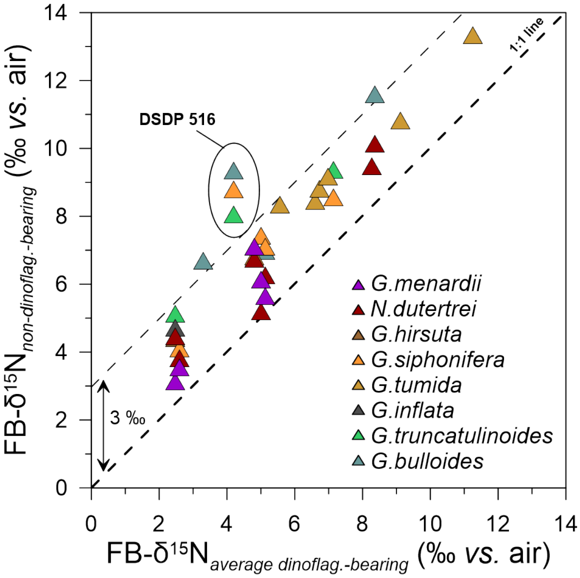
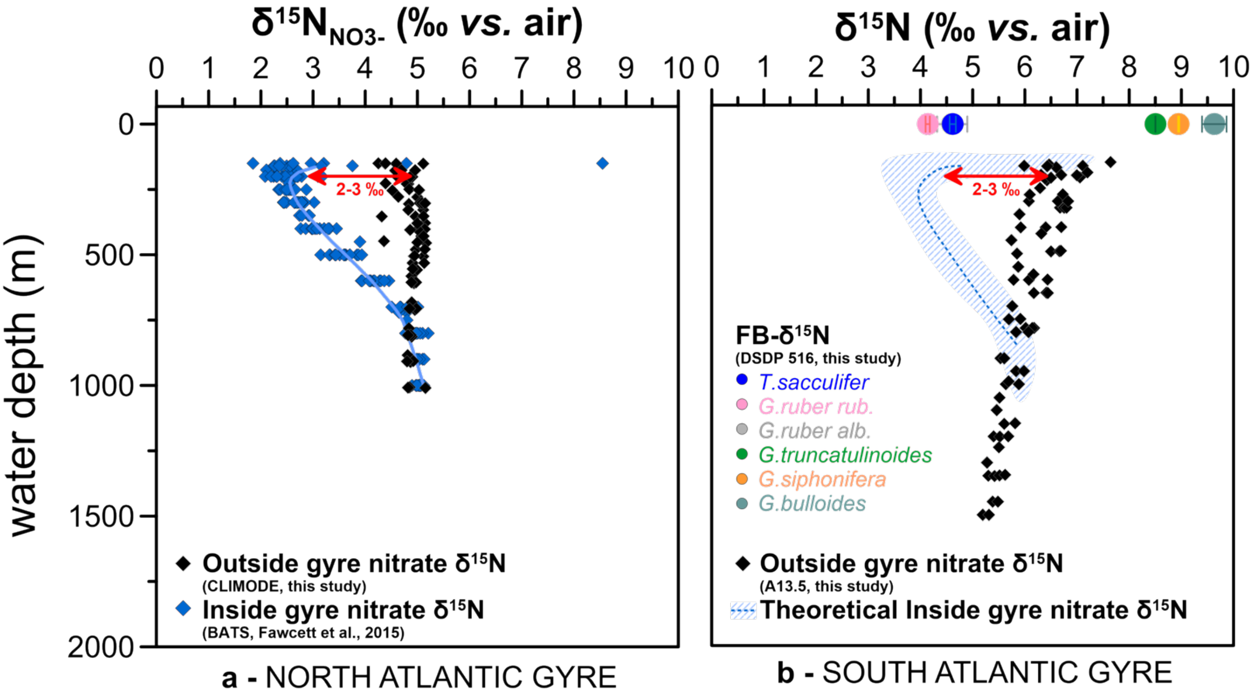


Figure 8. FB-δ^{15}N offsets between non-dinoflagellate-bearing foraminifera *vs* dinoflagellate-bearing foraminifera (the average of *G. ruber albus*, *T. sacculifer* and *O. universa*), with DSDP Site 516 indicated having uniquely high FB-δ^{15}N offsets. DSDP Site 516 exhibits stronger ^{15}N enrichment for non-dinoflagellate-bearing foraminifera than other sites from the global core top compilation (see Table S1 and Figure S5).



a - NORTH ATLANTIC GYRE

b - SOUTH ATLANTIC GYRE

Figure 9. Proposal for the N isotopes of the South Atlantic gyre based on North Atlantic gyre nitrate isotopic measurements. (a) Seawater nitrate $\delta^{15}N$ for the North Atlantic gyre system with distinct isotopic signature for outside (black diamonds, CLIMODE, this study) and inside (blue diamonds, BATS, Fawcett et al. (2015)) the gyre system, **(b)** Seawater nitrate $\delta^{15}N$ outside the South Atlantic gyre system (black diamonds, profile A13.5, this study). Hashed area with blue line indicates inferred possible seawater nitrate $\delta^{15}N$ for a location within the South Atlantic gyre based on the difference between inside and outside the North Atlantic gyre and nitrate isotopic data from the South Atlantic. Coloured circles indicate FB-$\delta^{15}N$ from Site 516. Locations for seawater $\delta^{15}N$ profiles are shown in Fig. 1.

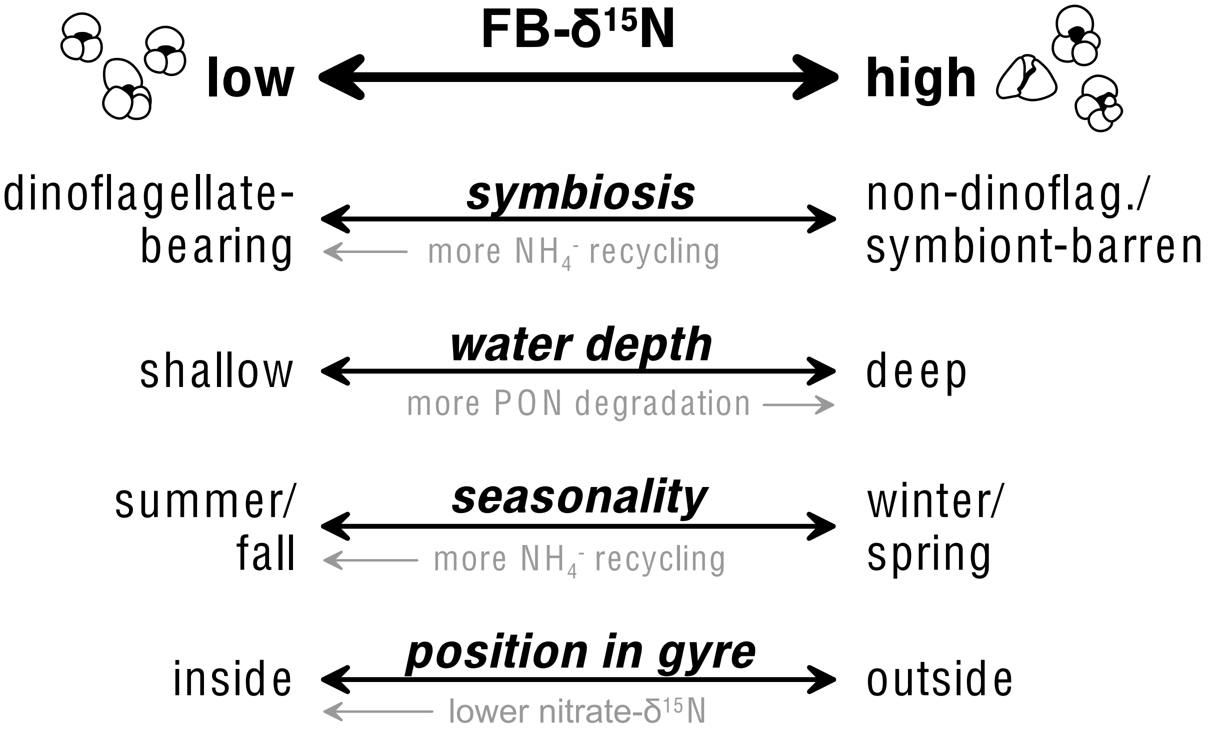


Figure 10. Proposed different processes affecting FB-δ^{15}N at DSDP Site 516. Shallow-dwelling, summer-blooming, dinoflagellate-bearing foraminifera located within the oligotrophic gyre record generally lower FB-δ^{15}N values than symbiont-barren or foraminifera hosting symbionts other than dinoflagellates that live generally in a deeper depth habitat, bloom during winter/spring season and/or live outside oligotrophic gyres.

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