

# Covariation of redox potential profiles and water table level in peatland sites representing different drainage regimes: implications for ecological modelling

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**Abstract.** Reduction-oxidation (redox) reactions are ubiquitous in nature, responsible for the energy acquisition of all organisms. Redox reactions are electron transfer reactions that necessarily involve two participants: one being oxidised (electron donor) and one being reduced (electron acceptor).

Availability of terminal electron acceptors (TEAs) is a major determinant of the extent to which carbon in organic matter can be oxidised in an ecosystem. This is most important under waterlogged conditions, such as peatlands, where diffusion of O<sub>2</sub>, the most effective common TEA, into soil is blocked by water. Under these conditions, alternative TEAs can be used by microbiota to continue organic matter oxidation.

Decomposition processes in soil can be characterised by its redox state, i.e. which TEA is responsible for organic matter oxidation at a given time. This can in principle be measured as a voltage between the soil solution and a known reference electrode, known as the redox potential.

Current soil ecosystem models do not depict the use of alternative TEAs well. This limits their applicability for predicting soil carbon loss under different drainage regimes, and thus their usefulness for assessing best management practices for soil carbon preservation and water course protection. The most common determinant of mode of decomposition presently used in ecosystem models is water table level (WTL), which relies on the assumption that the redox state of a peatland ecosystem responds predictably to changes in WTL.

We conducted a two-year redox monitoring experiment in a boreal mesotrophic peatland under three drainage regimes: undrained, short-term drainage and long-term drainage. In addition, an ombrotrophic long-term drained plot was monitored. Snapshot assessments of the activity of three major metabolic enzymes, arginine deaminase, protease and urease, were also undertaken at the mesotrophic plots as an indicator of differences in microbial activity between drainage regimes.

20 We found that WTL was a poor temporal predictor of redox potential but that the position of major transition zones between  
oxic and anoxic states, as well as enzymatic activities within the peat profile, were somewhat determined by the dominant  
WTL depth. In the undrained plots especially, redox potential values reflecting oxic or suboxic conditions were often found  
below the WTL, whereas on the drained plots anoxia was present above the WTL. Preceding redox potential was found to  
affect activities of protease and urease, but not arginine, in all plots.

## 25 1 Introduction

Oxidation-reduction (redox) reactions are central to the energy acquisition processes of all life. All redox reactions consist of  
an atom donating an electron (being oxidised) and an atom accepting an electron (being reduced). In soils, there is usually  
no shortage of electron donors, the most usual source being organic matter; thus, energetically feasible redox reactions are  
generally limited by which electron acceptors are available (Green and Paget, 2004).

30 Many elements found in soils alter their behaviour according to their redox state. As an important example, iron (Fe) is  
mainly found in non-water soluble compounds in its oxidised ferric  $\text{Fe}^{3+}$  (Fe(III)) state, whereas in its reduced ferrous  $\text{Fe}^{2+}$   
(Fe(II)) state it is soluble. This has implications for the movement of elements in soil solution. For example, phosphorus (P)  
forms complexes with Fe(III) compounds that may then 'unravel' when Fe is reduced, making the P available to soil solution  
(e.g. Zak et al., 2004).

35 The strongest common oxidising element in soil systems is oxygen ( $\text{O}_2$ ). This is consumed whenever available, and such  
reactions release the most energy. When availability of  $\text{O}_2$  is limited, e.g. under waterlogged conditions, other terminal electron  
acceptors (TEAs) will be used. Common TEAs in soils include, in descending order of available energy, nitrate ( $\text{NO}_3^-$ ),  
manganese(III) ( $\text{Mn}^{3+}$ ), Fe(III), sulphate ( $\text{SO}_4^{2-}$ ) and carbon dioxide ( $\text{CO}_2$ ), each of which require increasing electron activity  
(pe) in the soil solution to be energetically feasible. This can be measured against a redox pair of known activity to provide the  
40 redox potential ( $E_h$ , V).

Redox processes in soils, whether mineral or organic, are an interplay between microbes, their enzymes, and purely chemical  
reactions, i.e. both biotic and abiotic drivers. Which process is dominant will depend on the relative availability of oxidants  
and the activity of  $\text{H}^+$ , i.e. the environment's pH. For example, Fe(III) may be reduced to Fe(II) either biotically, by microbes  
oxidising organic carbon (C), or chemically, by reduced (i.e. electron-rich) humic substances (Melton et al., 2014). In more  
45 extreme circumstances, Fe(III) may also be reduced by archaea in the anoxic oxidation of  $\text{CH}_4$  (Ettwig et al., 2016). As  
microbe activity is often the dominant factor affecting redox status, it is conceivable that changes in  $E_h$  could be more rapid  
under conditions where microbes are more active, reflected by microbial enzyme activity in the soil.

In peatlands, the activity of oxidative and hydrolytic enzymes produced by microorganisms will be regulated by the site's  
vegetation cover, soil water regime, temperature and nutrient availability, along with interactions between physicochemical  
50 factors, such as changing pH or redox  $E_h$  (Freeman et al., 1996; Bonnett et al., 2006). Interactions between redox condi-  
tions, extracellular enzyme activity and persistence of phenolic substances are complex. For example, Freeman et al. (2001)  
hypothesised that a process termed "enzymatic capture" occurs due to the accumulation of phenolic substances (phenolic dis-

solved organic matter, or phenolic DOM) and their persistence in the soil. This condition occurs when phenoloxidase activity is suppressed under anoxic conditions. The accumulation of these phenolic substances leads to inhibition of hydrolytic enzyme activity, e.g. of protease or urease (Kane et al., 2019), and has been suggested as one mechanism leading to stabilisation of peat C (Freeman et al., 2004).

Humic substances, prevalent in organic soils such as peat, have been found to act as both TEAs and donors, potentially reducing CH<sub>4</sub> emissions from boreal peatlands by a large factor (Klöpfer et al., 2014). The E<sub>h</sub> of humic acid reduction reactions, for example, has been shown to range between +150 – -300 mV, thus overlapping with several non-organic electron acceptors (Aeschbacher et al., 2011). Furthermore, the electron accepting capacity of organic matter has been found to be greater in ombrotrophic than minerotrophic peats (Keller and Takagi, 2013). Finally, other non-O<sub>2</sub> electron acceptors have been found to inhibit production and/or promote oxidation of CH<sub>4</sub> (Kumaraswamy et al., 2001).

Generally speaking, current ecological models used for predicting ecosystem C and nutrient fluxes either only include an oxic-anoxic state controlled by soil permeability and water table level (WTL) (Palviainen et al., 2024), ignore redox altogether and base their estimates on WTL and/or soil moisture and temperature only (e.g. Gong et al., 2013) or use only one electron acceptor other than O<sub>2</sub>, e.g. NO<sub>3</sub><sup>-</sup> (Wriedt and Rode, 2006) or Fe(III) (Tang et al., 2016). To improve such models, reactions of TEAs other than O<sub>2</sub> need to be modelled. A first step in this direction would be to model the redox state of a peatland empirically, using easily measurable (and computationally cheap) variables and site properties to predict the momentary E<sub>h</sub>.

Northern peatland ecosystems generally have high WTLs, which has encouraged the sequestration of huge amounts of C under anoxic conditions (e.g. Yu et al., 2010). At such sites, any variation in soil redox E<sub>h</sub> due to changes in the WTL could unlock the stored C into water-dissoluble and microbially degradable forms (Freeman et al., 2001). Indeed, the ecological characteristics of such wet and relatively nutrient-rich fens, as well as the C stored in the peat, have been shown to be particularly sensitive to any decrease in WTL (Straková et al., 2012; Jaatinen et al., 2007; Gong et al., 2013; Kokkonen et al., 2019).

It is generally believed that redox conditions in peatlands are regulated by the WTL through its effect on soil O<sub>2</sub> concentrations, i.e. that in water-logged soil, the water blocks gas movement between the atmosphere and the soil matrix (e.g. Belyea, 1999; Blodau et al., 2004; Kiuru et al., 2022). However, covariation of WTL and soil redox E<sub>h</sub> at different depths in peat soils has not been examined in detail. In dense peat especially, the depth distribution of redox processes may be insensitive to WTL fluctuations (Knorr and Blodau, 2009). On freshwater tidal wetlands, for example, redox E<sub>h</sub> has been shown to fluctuate at 20 cm depth, but not at 50 cm, despite the WTL fluctuating between -40 and +18 cm (Seybold et al., 2002).

In recent years, there has been growing interest in defining the best approach for describing redox phenomena in ecosystems. For example, it has been suggested that only assessing the E<sub>h</sub> state of an ecosystem or soil profile should be amended by examining the resistance of that system to change under redox conditions, i.e. its redox buffering capacity (Burgin and Loecke, 2023). In such cases, it could be postulated that:

1. TEAs other than O<sub>2</sub> and CO<sub>2</sub> may play a significant role in an ecosystem's redox palette; and
2. the readings given by E<sub>h</sub> measurement devices reflect the dominant redox pair in an ecosystem at a given time; then
3. under increasingly anoxic conditions, E<sub>h</sub> should "pause" its descent at values reflecting the available TEAs; and

4. this would cause the probability distribution of  $E_h$  values measured in such an ecosystem to express multi-modality rather than bi-modality, which would be indicative of an  $O_2$ - $CO_2$ -dominated system.

From a modelling perspective, this would indicate that current peatland models that do not recognise TEAs other than  $O_2$  would work better on ombrotrophic than minerotrophic sites as more TEAs are present in minerotrophic peatlands.

In this study, we examine the effects of WTL fluctuation on soil solution redox  $E_h$  at different depths in a mesotrophic sedge fen site (ME) situated in a minerotrophic peatland in southern Finland. Four plots were examined, comprising three ME plots with different soil WTL (drainage) regimes, i.e. wet (undrained control), short-term water-level drawdown (15 years), and long-term drawdown (55 years), along with a long-term drained bog plot at an ombrotrophic site (OM) in the same peatland massif, used for comparison. In each case, WTL drawdown has had marked impacts on local peat properties (e.g. chemical composition, density, pH), vegetation cover and soil microbial communities (Straková et al., 2011; Peltoniemi et al., 2012). Consequently, we hypothesize that the drainage regime, and the ensuing depth range of WTL fluctuation, will be reflected in the depth at which redox conditions are at their most dynamic, i.e. at ME plots, with the wet undrained plot having WTL fluctuations and redox dynamics occurring close to the peat surface, the long-term drained plot having fluctuations at the greatest depth, and the short-term drained plot lying somewhere between.

We also tested for relationships between WTL and  $E_h$  under the different drainage regimes using analysis of wavelet coherence between WTL and  $E_h$  conditions, using both  $E_h$  values from individual sensors at different depths and the depth isopotential of 0mV  $E_h$ , indicative of Fe reduction.

Our second hypothesis is, therefore, that  $E_h$  from sensors in the most active peat layer in terms of alternating redox conditions will follow fluctuations in the WTL, and that the 0 mV  $E_h$  isopotential will consistently follow the WTL.

Third, we hypothesise that the lower levels of alternative TEAs in the OM plot will cause  $E_h$  conditions to become reducing more quickly after a rise in WTL than in the ME plots. This will be revealed as a lower phase difference, i.e. lag time, in a wavelet coherence analysis at the OM plot compared to the ME plots, and as a more extreme distribution of  $\delta E_h$  in the long-term drained plot compared to the wet, undrained plot.

Fourth, we hypothesise that the lack of alternative TEAs at the OM plot will be expressed as bi-modality in the probability density distribution of  $E_h$  measurements, and that the presence of alternative TEAs at the ME plots will be expressed as multi-modality.

Finally, we test whether 'snapshots' of soil enzyme activity at the ME plots correlate with preceding redox conditions and drainage status. For this study we selected hydrolytic enzymes, i.e. proteases, ureases and arginine deaminases, knowing that their activity can be affected by phenolic substances. We did not investigate the activity of oxidative enzymes because this issue has already been thoroughly investigated by other authors. Thus, our fifth hypothesis is that the more aerobic conditions at the drained plots will be reflected in increased enzyme activity compared to the wet, undrained plot.

## 2 Material and methods

### 2.1 Measurement sites

120 This study took place at the Lakkasuo mire, a raised bog complex with a large minerotrophic lagg, in southern Finland (61.797 °N, 24.309 °E). In 1961, half of the mire complex was drained to improve forest growth, which created conditions where the impact of persistent WTL drawdown could be studied on plots either side of a border ditch, both of which shared similar hydrological conditions, vegetation cover and soil properties prior to drainage (e.g. Minkkinen et al., 1999). The surface layer of the undrained minerotrophic fen receives ground water from the Vatiharju esker bordering the mire complex in the west, 125 potentially bringing in electron acceptors such as Fe to the ME plots (Sallantausta and Kaipainen, 1996).

In 2001, several new experimental WTL drawdown plots were established, with conditions allowing the study of shorter-term impacts (e.g. Straková et al., 2012; Kokkonen et al., 2019). These measurement plots were positioned to represent three WTL regimes, i.e. an undrained plot (ME-UD), a short-term (ca. 15 yrs at the time of study) persistent WTL drawdown plot (ME-STD) and a long-term (ca. 55 yrs) persistent WTL drawdown plot (ME-LTD) (Fig. 1). Conditions at the STD plot were 130 maintained via narrow, ca. 30-cm deep ditches that conducted surface water into the border ditch. An initially ca. 1m deep ditch network within the LTD area had not been intensively reconditioned for some years, thus the drainage effect was less effective than would be the case at more intensively managed forest sites. In addition, a comparative ombrotrophic (OM) plot under an LTD regime (OM-LTD) was located on the same peatland massif, ca. 500 metres from the ME-LTD plot.

The different WTL regimes being examined were reflected in the composition of plant communities on the plots. For example, the ME-UD plot supported typical boreal mesotrophic open mire vegetation, being dominated by sedges, such as *Carex lasiocarpa* (woollyfruit sedge) and *C. rostrata* (beaded sedge), with some forbs, such as *Menyanthes trifoliata* (bog bean) and *Comarum palustre* (swamp cinquefoil), shrubs *Betula nana* (dwarf birch) and *Salix* sp. (willows), and several moss species, the most common being the peat mosses *Sphagnum fallax* and *S. flexuosum*. On the ME-STD plot, the lowered WTL had already had a significant effect on the vegetation (Kokkonen et al., 2019), the original sedges and forbs largely having been re- 140 placed by species tolerating drier conditions, such as *C. echinata* (star sedge), *C. canescens* (silvery sedge), *Calamagrostis* spp. (reed grass), *Cirsium palustre* (European swamp thistle) and *Trientalis europaea* (arctic starflower). A dense spread of *Pinus sylvestris* (Scots pine) and *Betula pubescens* (Downy birch) seedlings had started to form an overstorey layer and, while the moss layer was sparse, it included several species. The ME-LTD plot could be classified as a mesotrophic peatland forest (*Vaccinium myrtillus* type II in the local classification), with surface vegetation characterised by shrubs, such as *Empetrum nigrum* (black crowberry), *V. myrtillus* (bilberry) and *V. vitis-idaea* (lingonberry), with *Eriophorum vaginatum* (cottongrass) as the most common sedge. The tree stand comprised *P. sylvestris* and *B. pubescens*, with some *Picea abies* (Norway spruce), while the moss layer was dominated by *Pleurozium schreberi* (feathermoss), other moss species including the peat mosses *S. russowii*, *S. medium* and some *Dicranum* (fork mosses) and *Polytrichum* (haircap moss) species. In comparison, the OM-LTD plot was 145 classified as a nutrient-poor peatland forest, with *Sphagnum* mosses intermixed with forest mosses, and peatland shrubs such as *Andromeda polifolia* (bog rosemary) and *Vaccinium uliginosum* (bog bilberry) sparsely present among *E. vaginatum* tillers.

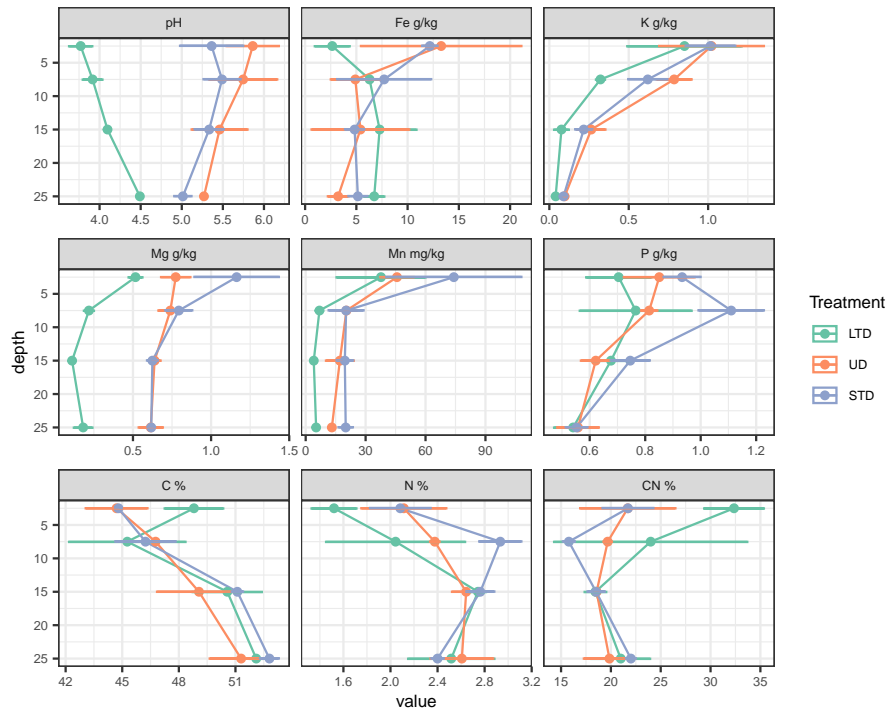


**Figure 1.** Left: location of the measurement site (made with Natural Earth); right: map of the Lakkasuo mire (north at top), with the measurement plots marked (background map: National land survey of Finland, topographic map, 4/2024. Green hatched rectangle is a feature of the topographic map defining the perimeter of a nature conservation area.)

Soil chemical properties (Laiho et al., 2024) at the ME plots reflected the typical effects of drainage on fen peats, *i.e.* a relative increase in C content and a decrease in nitrogen (N) content, along with a decrease in pH due to decomposition increasing the amount of organic acids in the soil, and loss of soluble earth metals such as magnesium (Mg) and potassium (K) (Fig. 2). From a redox perspective, the most important elements analysed were Fe and manganese (Mn), with Fe content being significantly lower in the surface layer of LTD plots than STD and UD plots, Mn content showing no significant differences. While a soil chemical analysis was not available for the OM-LTD plot, concentrations of the most important elements for redox couples (*i.e.* Fe and Mn) on drained ombro-oligotrophic peatlands are known to be around one-fifth those present on drained mesotrophic sites (e.g. Laiho and Laine, 1995).

## 2.2 Redox sensors

Three redox probes (Vorenhout consulting, Netherlands) fitted with platinum (Pt) electrodes were installed at five depths (*i.e.* 5, 15, 25, 35 and 45 cm below the surface) at each measurement plot in early Autumn 2014. A silver-silver chloride (Ag-AgCl) reference electrode was also installed in a groundwater well for each trio of probes. A Hypnos III logger system (Vorenhout Consulting, Netherlands) (Vorenhout et al., 2011) was then used to record data from the probes at 15 minute intervals continuously over two years, *i.e.* from Autumn 2014 to Autumn 2016. The redox measurement systems were calibrated beforehand according to Vorenhout et al. (2004) using a 220 mV (pH 7) reference solution.



**Figure 2.** Chemical properties of peat soils at the three mesotrophic study sites (means and standard deviation of three samples; element concentrations in mg or g/kg dry weight unless stated otherwise in the panel headers; depth in cm. LTD (green) = long-term drawdown, UD (red) = undrained, STD (blue) = short-term drawdown).

**Table 1.** Common redox pairs in soils and their corresponding field-observable  $E_h$  values (mV). All values reported in pH 7, according to Borch et al. (2010) and Vorenhout et al. (2004)

$E_h$ range	Redox pair	
	Oxidised species	Reduced species
> 400	$O_2$	$H_2O$
300	$NO_3^-$	$N_2$
150	$Mn^{4+}$	$Mn^{3+}$
-50 ... 100	$Fe^{3+}$	$Fe^{2+}$
-150	$SO_4^{2-}$	$H_2S$
<-200	$CO_2$	$CH_4$

## 2.3 Ancillary measurements

The WTL at each measurement plot was continuously monitored using TruTrack WT-HR1000 probes (Intech Instruments, New Zealand), while soil temperatures were monitored using i-Button DS1921G temperature loggers (MaximIntegrated Products, USA) at the same depths as  $E_h$  (i.e. 5, 15, 25, 35 and 45 cm) at one point per treatment.

170 Soil pH was determined for each depth at each ME and OM plot on two occasions, October 2014 and May 2015, from peat samples obtained from the plots. The pH was measured under laboratory conditions using a slurry of one part peat to three parts water.

Activity of the enzymes arginine deaminase, protease and urease were determined at ME plots as indicators of soil microbial activity at the same depths as redox measurements on peat samples obtained in October 2014, May 2015 and July 2015. Briefly, 175 arginine deaminase activity was determined from soil samples incubated with arginine to produce ammonium ( $\text{NH}_4$ ). This was then extracted with a potassium chloride (KCl) solution and the amount of extracted  $\text{NH}_4$  determined colourimetrically (Alef and Kleiner, 1986). Determination of protease activity was based on the decomposition of added casein during incubation, determined colourimetrically by measuring the amount of L-tyrosine produced (Rejsek et al., 2008). Urease activity was determined by incubating the samples with added urea and colourimetrically determining the amount of  $\text{NH}_4$  released (Kandeler 180 and Gerber, 1988).

Air temperature was measured continuously at 2m above the soil's surface near the measurement plots throughout the measurement campaign. Precipitation and snow depth were measured hourly at the SMEAR II measurement station, ca. 6 km distant (Aalto et al., 2023).

## 2.4 Data processing

185 The potential given by the Pt-Ag/AgCl-pairs was converted to  $E_h$  by adding +200mV to the reading to account for the difference in electrical potential between the Ag/AgCl reference electrode used in our setup and the standard hydrogen electrode against which  $E_h$  values are reported. The readings were then corrected for  $\text{H}^+$  activity by applying the Nernst equation, i.e.  $(+59\text{mV} \times (\text{pH} - 7))$ , normalising them to pH 7. To reduce noise, the readings were averaged over an hour or over a day, depending on the analysis being undertaken. The data were further corrected for drift by fitting a linear model of  $E_h$  over time to the maximum 190 ( $> 400 \text{ mV}$ ) and minimum ( $< -400 \text{ mV}$ ) readings, as grouped by the logger. Drift was determined at  $-0.16\text{mVd}^{-1}$  for the STD and LTD plots and  $-0.08\text{mVd}^{-1}$  for the UD and OM-LTD sites.

Basic data processing and statistical analyses were undertaken using the R statistical package (R Core Team, 2023).

## 2.5 Linear and non-linear modelling

Linear and non-linear models, using hourly data, were fitted to assess differences in  $E_h$  response to WTL between the ME 195 measurement plots. Both the models and the results are described in more detail in the supplementary materials.



## 2.6 Wavelet analysis

The frequency spectra of hourly averaged redox potentials, soil temperatures and WTL measurements were analysed using wavelet decomposition (Torrence and Compo, 1998) to identify patterns that would inform future directions for  $E_h$  modelling. Briefly, wavelet decomposition consists of fitting a wavelet function with different frequencies at each time point in a time series and estimating how well each of the frequencies fits the data, enabling identification of time patterns with above- and below-average values.

The coefficients were then compared against soil temperature and WTL changes *via* a cross-correlation spectrum, using a *Mexican hat* style mother wavelet to achieve sufficient temporal resolution (Wang, 2015). Owing to large differences between the probe  $E_h$  readings, wavelet fitting was undertaken separately for each probe and depth profile. All hourly wavelet analyses were performed using the PyWavelets package in Python (Lee et al., 2019). The full wavelet coherence figures are presented in Supplement 1.

In addition, wavelet coherence between daily average WTL and the depth isopotential of  $E_h$  0 mV indicative of Fe reduction was estimated based on the mean  $E_h$  of all three probes on all measurement plots using the WaveletComp package in R (Schmidbauer, 2018).

In this case, the depth isopotential was calculated by linearly interpolating daily average  $E_h$  depth profiles from 5 to 45 cm depth, using a random time series with the same Fourier transform properties as a surrogate time series to analyse the significance of coherence. Only those days where the  $E_h$  0 isopotential was present (i.e. the whole  $E_h$  profile was not above or below 0 mV) were included in the analysis.

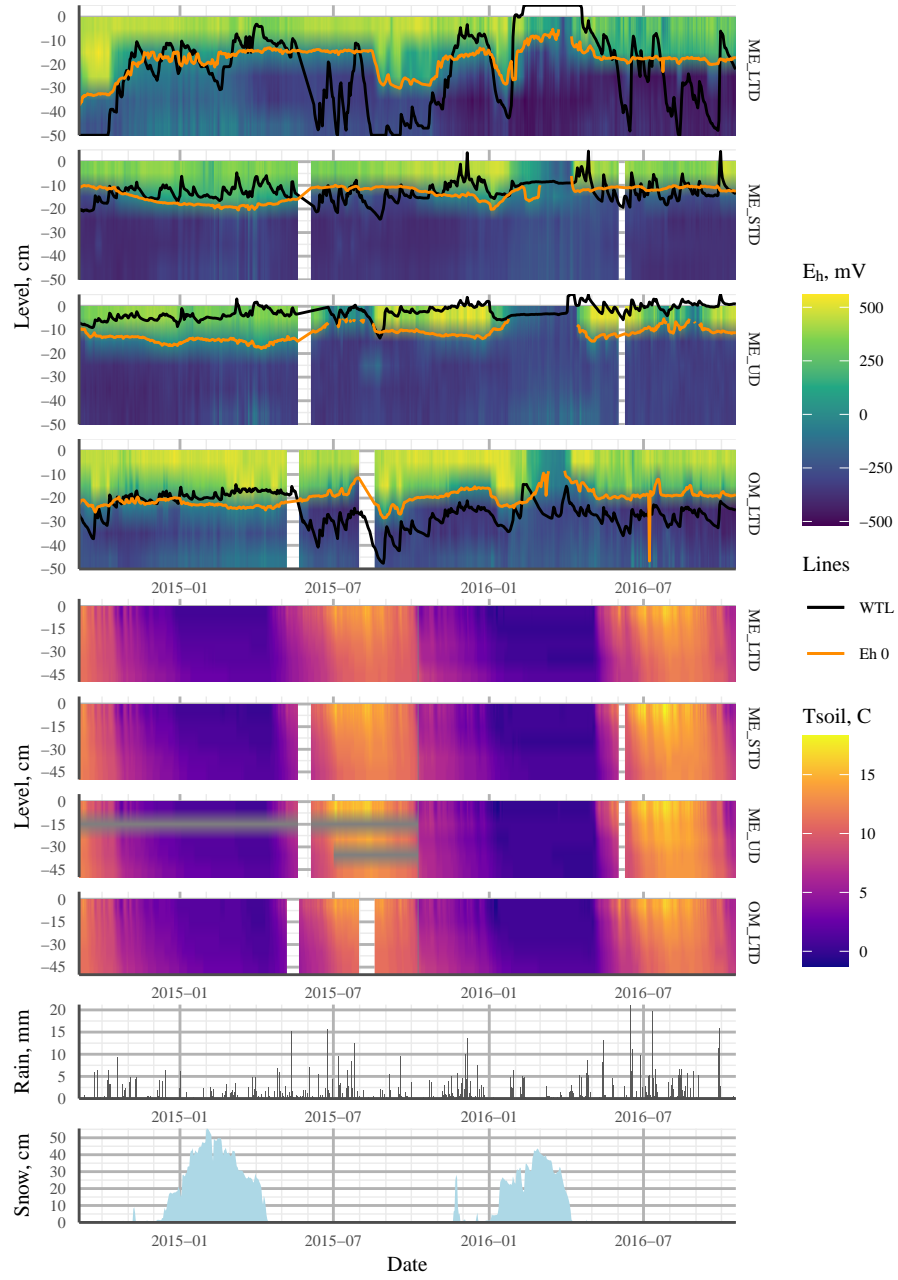
Although there are differing views on how Fe(III)/Fe(II)-related redox potential should be calculated, based on electrochemical theory and the number of protons involved in the processes (e.g. see Boonman et al., 2024), we chose 0 mV as the pH 7-normalised potential representing Fe reduction based on the literature (e.g. Borch et al., 2010; Vorenhout et al., 2004) and our own experimental observations in the laboratory (Marttunen, 2024), where Fe addition to anaerobic peat incubations stabilised the  $E_h$  to around 0 mV. This is also supported by a study published by Pyzola et al. (2025), where Fe(II) started to appear in incubated slurries at close to 0 mV  $E_h$ .

## 3 Results

### 3.1 Water table level

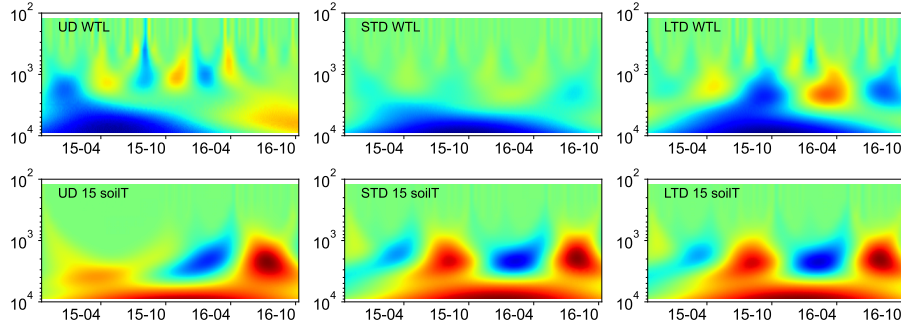
Differences in WTL were significant ( $p < 0.05$ ) between all treatments, with mean WTLs over the measurement campaign being -1.5 cm, -13.8 cm and -26.8 cm for the ME-UD, ME-STD and ME-LTD plots, respectively (Fig. 3). Fluctuations in WTL were greatest at the LTD plot, while the STD and UD plots showed roughly the same magnitude of WTL fluctuation, i.e. between 0 and -15 cm.

Wavelet analysis indicated clear seasonal patterns in WTL fluctuation for the UD and LTD plots, but not for the STD plot. This pattern was especially strong in the LTD plot, which showed below average values from July to November and above



**Figure 3.** Heat maps showing mean values (mean of three profiles per treatment) interpolated from the 5-45 cm depth profiles of  $E_h$  (**four topmost plots:** ME LTD, ME STD, ME UD, OM LTD; WTL and the  $E_h$  0 isopotential shown as solid black and orange lines, respectively) and soil temperature (**four middle plots:** ME LTD, ME STD, ME UD, OM LTD). **Bottom two plots:** daily rainfall (mm) and snow pack thickness (cm) at the peatland over the study period. ME = mesotrophic sedge fen; OM = ombrotrophic bog; UD = undrained; STD = short-term ( $\sim 15$  years) drainage; LTD = long-term ( $\sim 55$  years) drainage.

average values from January to May (Fig. 4). In comparison, temperature plots at all sites showed the opposite pattern. At the UD plot, WTL fluctuated over a shorter period than at the LTD plot (UD 1000-hour period vs. LTD 2000-hour period) (Fig. 4).



**Figure 4.** Mexican hat wavelet coherence: **top, left to right** : WTL data from the 1) ME-UD plot, 2) ME-STD plot, and 3) ME-LTD plot, and **bottom, left to right**: soilT data at 15 cm from the 4) ME-UD plot, 5) ME-STD plot, and 6) ME-LTD plot. Note inverted Y-axes (wavelet period, hours).

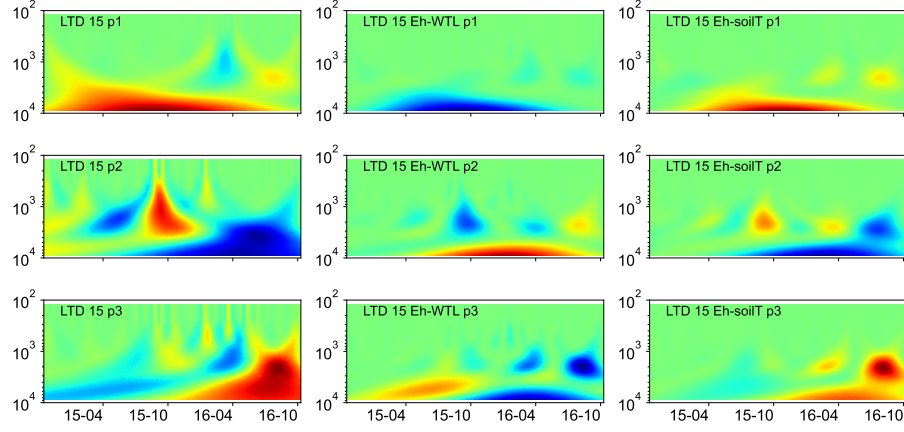
### 230 3.2 Redox

Generally speaking,  $E_h$  varied between oxic and completely anoxic in all profiles, with the 5 cm surface layer being over +400 mV and layers below 25 cm being < 0 mV for all three treatments. The main difference between treatments was at 15 cm depth, where  $E_h$  was almost permanently below -200 mV after June 2015 at the UD plot, but varied between +250 mV and -400 mV in the STD and LTD plots (Fig. 3).

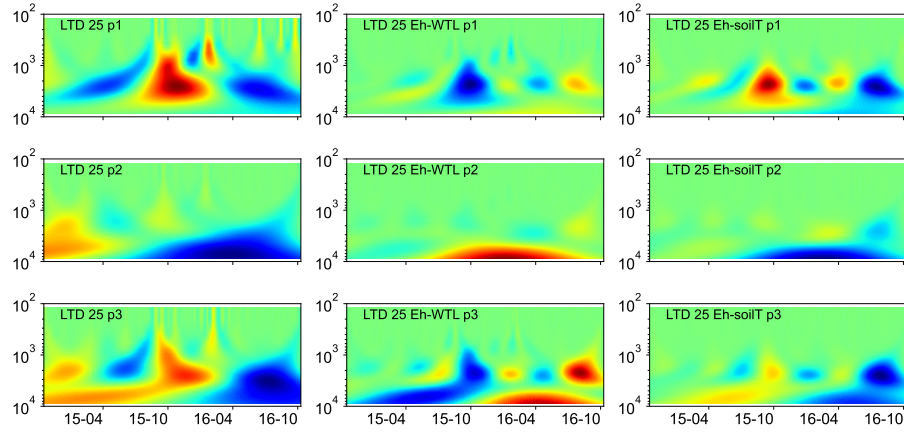
235 While  $E_h$  values varied greatly between the three profiles for all treatments, especially at 15cm depth (Fig. 3), sensitivity to change in WTL differed between profiles, with some sensors showing no sensitivity to WTL and others consistently reacting to WTL change.

Wavelet analysis revealed a periodic pattern of roughly 100 days (2000-3000 hours) at differing depths on all plots. At the ME drained plots (STD, LTD), for example, clearest patterns were observed at 15 and 25 cm depth (Figs. 5, 6, 7), whereas  
 240 strongest patterns were observed at 5 cm depth at the UD plot (Fig. 8). There were also additional shorter-period patterns at the UD plot ranging from 200 to 500 hours. At the STD plot, interactions between  $E_h$  and WTL in the most active layers were mostly in synphase, i.e. a lowering WTL (increasing WTL depth) coinciding with rising  $E_h$  values, compared with an alternating counter- and synphase interaction at the UD and LTD plots.

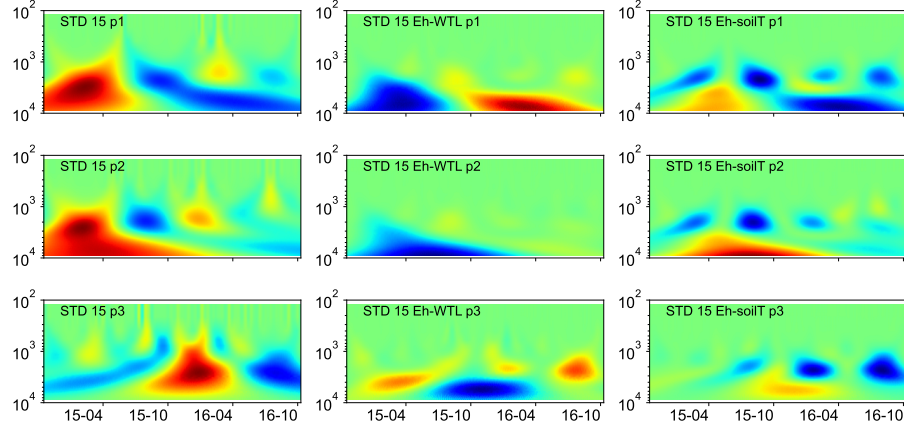
Analysis of coherence between the Fe-reducing depth isopotential (-100 ... +50 mV (Table 1), represented by the  $E_h$  0  
 245 mV isopotential) and WTL (Fig. 9) indicated 128-day and 64-day periods of significant ( $p < 0.05$ ) coherence at all plots, but



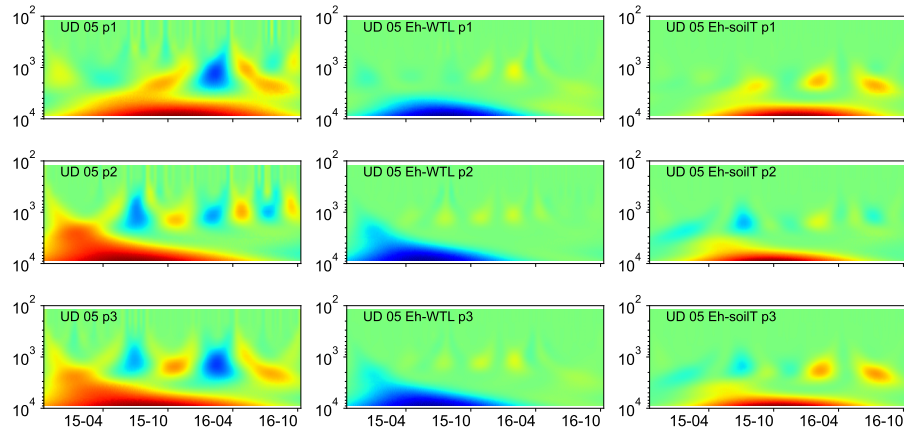
**Figure 5. Left:** Wavelet coherence between Mexican hat wave and  $E_h$  at the ME-LTD plot (15 cm depth); **centre:** interaction between  $E_h$  and WTL wavelets; **right:** interaction between  $E_h$  and soil temperature wavelets at the LTD plot (15 cm depth). **Top to bottom:** three probes plotted separately (p1 ... p3). Note logarithmic, inverted scale (hours) on y-axis, dates as YY-MM on x-axis.



**Figure 6. Left:** Wavelet coherence between Mexican hat wave and  $E_h$  at the ME-LTD plot (25 cm depth); **centre:** interaction between  $E_h$  and WTL wavelets; **right:** interaction between  $E_h$  and soil temperature wavelets at the ME-LTD plot (25 cm depth). **Top to bottom:** three probes plotted separately (p1 ... p3). Note logarithmic, inverted scale (hours) on y-axis, dates as YY-MM on x-axis.

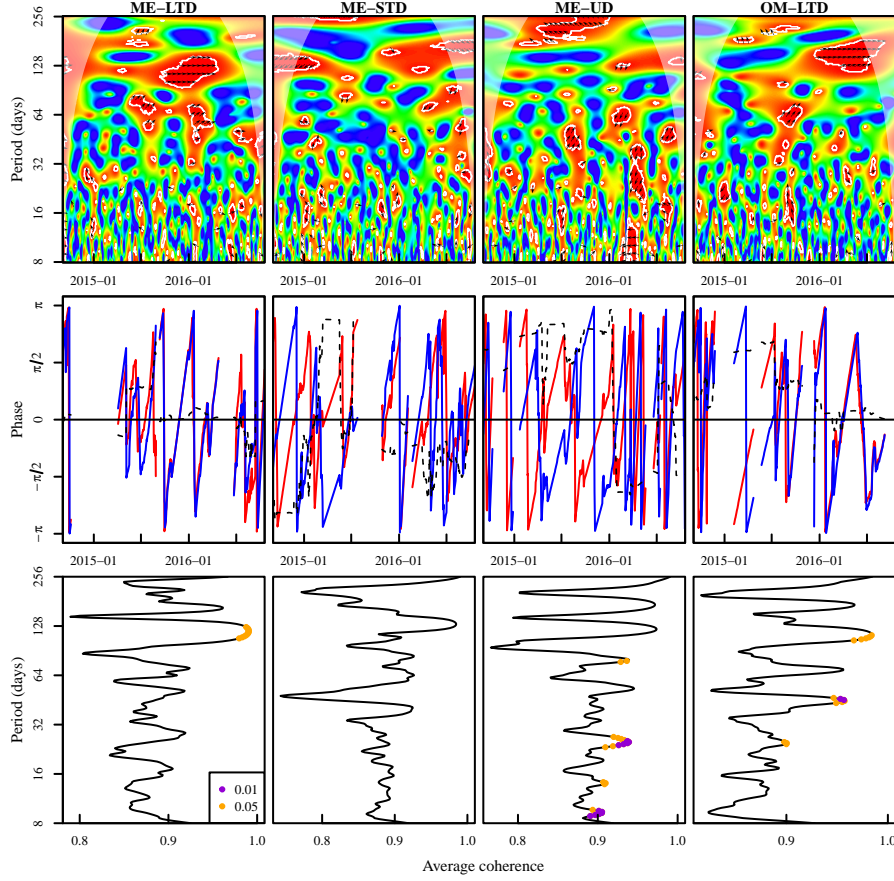


**Figure 7. Left:** Wavelet coherence between Mexican hat wave and  $E_h$  at the ME-STD plot (15 cm depth); **centre:** interaction between  $E_h$  and WTL wavelets; **right:** interaction between  $E_h$  and soil temperature wavelets at the ME-STD plot (15 cm depth). **Top to bottom:** three probes plotted separately (p1 ... p3). Note logarithmic, inverted scale (hours) on y-axis, dates as YY-MM on x-axis.



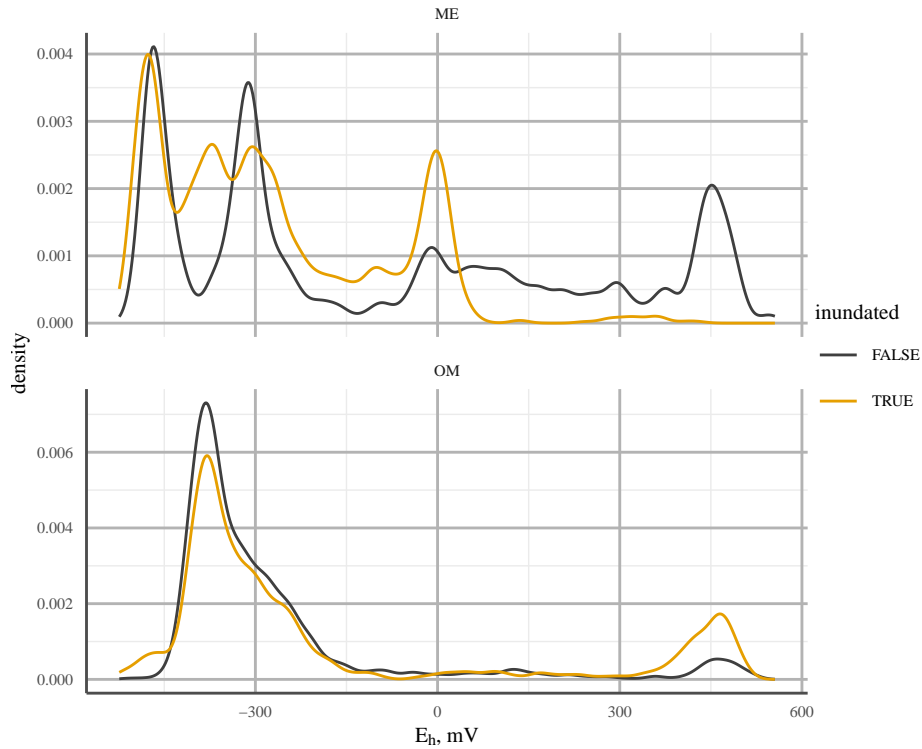
**Figure 8. Left:** Wavelet coherence between Mexican hat wave and  $E_h$  at the the UD plot (5 cm depth); **centre:** interaction between  $E_h$  and WTL wavelets; **right:** interaction between  $E_h$  and soil temperature wavelets at the ME-UD plot (5 cm depth). **Top to bottom:** three probes plotted separately (p1 ... p3). Note logarithmic, inverted scale (hours) on y-axis, dates as YY-MM on x-axis.

particularly so at the LTD plot, where WTL led the change during these periods. At the ME-STD and ME-UD plots, there were fewer periods of significant coherence and phase differences alternated between WTL and the  $E_h$  0 isopotential leading.



**Figure 9. Top row:** Wavelet coherence between the  $E_h$  0 depth isopotential and WTL (blue areas = low coherence, red areas = high coherence, areas of significant coherence ( $p < 0.05$  vs random time series with identical fourier transform characteristics) outlined in white; black arrows = phase difference: pointing right = synphase interaction, pointing left = counterphase interaction; right-up or left-down = WTL leads, left-up or right-down =  $E_h$  0 depth isopotential leads; areas shaded with white = cone of influence). **Middle row:** phases and phase differences during periods of significant coherence (blue line = phase of  $E_h$  0 depth isopotential, red line = WTL phase, dashed black line = phase difference:  $-\pi \dots -\pi/2$  and  $0 \dots \pi/2$  = WTL leads,  $-\pi/2 \dots 0$  and  $\pi/2 \dots \pi$  =  $E_h$  0 isopotential leads). **Bottom row:** average coherence (x-axis) per period length (y-axis), with period length of significant coherence denoted with orange and dark violet points ( $p < 0.05$  and  $p < 0.01$ , respectively). Measurement plots indicated in top row panel titles; x-axes aligned between panels in first and second rows.

During the first winter,  $E_h$  remained above Fe-reducing levels in soil  $> 15$  cm depth at all plots, regardless of WTL fluctuation and snow pack height. During the second winter, however,  $E_h$  values indicative of  $CO_2$  reduction were achieved at all plots, even at 5 cm depth (Fig. 3).

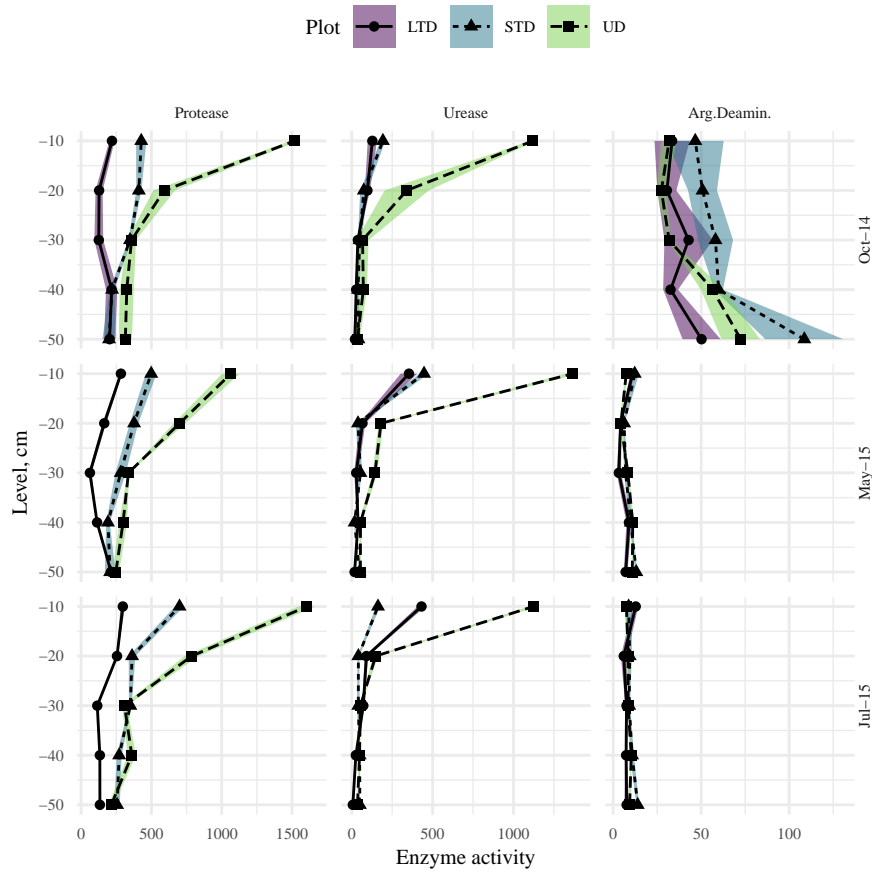


**Figure 10.** Probability density of  $E_h$  values at 25 cm depth for mesotrophic (ME; top) and ombrotrophic (OM; bottom) long-term drainage (LTD) plots. Line colour indicates whether WTL was above (orange) or below (black) the sensor. Note: different y-axis scales used due to the more continuous distribution at the ME site.

A comparison of  $E_h$  probability densities at 25 cm depth on the ME-LTD and OM-LTD plots (where most  $E_h$  dynamics occurred) indicated bi-modality between  $O_2$  reduction and  $CO_2$  reduction at the OM-LTD plot, and multi-modality, with peaks at  $E_h$  levels corresponding to  $O_2$ ,  $Fe^{3+}$  and  $CO_2$  reduction, at the ME-LTD plot (Fig. 10). Inundation of the sensor had a more pronounced effect at the ME plot, shifting the distribution toward lower  $E_h$  values.

### 255 3.3 Enzyme activity

Monitoring at the ME site in Autumn 2014 revealed a significant decrease in protease and urease activity with depth at all plots (LTD, STD, UD); however, the rate at which activities decreased differed between plots (Fig. 11). At the control plot (UD), both protease and urease were found in high levels in the topsoil, decreasing rapidly down to 20–30 cm, after which the rate of decline slowed. At the STD plot, subsoil (40–50 cm depth) protease levels were ca. 4x, and urease ca. 6x, lower than those  
 260 in the topsoil (0–10 cm depth), and the rate of decline in both cases was less steep than that at UD plot, tending to level off at around 30–40 cm. While 40–50 cm depth urease levels were similar to those at UD plots, protease levels at the same depth were significantly lower. At LTD plots, while urease showed a gradual but significant decline with depth, with levels around



**Figure 11.** Protease, urease and arginine deaminase activity depth profiles at the ME-LTD (solid line, round point, purple shaded area), ME-STD (short-dashed line, triangle point, blue shaded area) and ME-UD (long-dashed line, square point, pale green shaded area) plots, measured in October 2014, May 2015 and July 2015. Points and lines respectively represent observed and interpolated means, shaded areas represent standard errors of the means (N=3). Activities presented as  $\mu\text{g}$  L-tyrosine for Protease and  $\text{NH}_4\text{-N}$  for Urease and Arg. Deaminase,  $\text{h}^{-1} \text{g}^{-1}$  d.w. soil, respectively. Note the different x-axis scales between columns.



half those at UD plots at 40–50 cm, protease levels first declined but then began increasing again at around 20 cm, reaching similar levels to topsoil at 30–40 cm. After remaining relatively stable down to 10–20 cm, levels of arginine deaminase showed increasing trends with depth at all plots, with significantly higher levels in deeper soils at UD (30–40 cm) and STD (40–50 cm) plots, but not at LTD plot (Fig. 11). Arginine levels at the STD plot were higher than those at the UD and LTD plots at all depths, with especially high levels at 40–50 cm. Overall, the rate of increase was steepest at the UD plot, with levels at 40–50 cm 2.5x higher than those in topsoil, followed closely by the STD plot, with a significant increase at 40–50 cm ca. 2.1x higher than that in the topsoil. In comparison, the LTD plot showed a much shallower rate of increase, with levels at 40–50 cm just ca. 0.5x higher than those in topsoil.

In Spring 2015, both the levels and activity patterns for protease were similar to those in Autumn 2014 (Fig. 11). In comparison, topsoil urease levels were noticeably higher than in 2014 at all three plots, and especially so at the STD (ca. 2x) and LTD (ca. 3x) plots. However, levels dropped rapidly with depth at all plots, levelling off at 30–40 cm at plot UD at higher levels than 2014; dropping but peaking significantly at 40–50 cm at the STD plot at levels slightly higher than 2014; and dropping but peaking significantly at 30–40 cm at the LTD plot, with final levels slightly lower than in 2014. The greatest change between Spring 2015 and Autumn 2014 was observed in arginine deaminase levels, with levels significantly lower in 2015 at all plots (UD topsoil ca. 4.5 x lower, STD ca.3.5x lower, LTD ca. 4x lower). All three plots showed a further decline to 10–20 cm (LTD = 20–30 cm) followed by a significant increase up to 40–50 cm (LTD = 30–40 cm followed by a further drop). Levels at 40–50 cm were between 7 and 8x lower than in 2014.

Overall, enzyme activity in Summer 2015 was similar to that in Spring (Fig. 11), with the difference that topsoil protease levels were slightly higher at UD and STD plots, and slightly lower at the LTD plot; urease topsoil levels were slightly lower at UD and STD, and slightly higher at LTD; and topsoil arginine levels were relatively similar at UD and LTD but slightly lower at STD (Fig. 11). In all cases, levels at 40–50 cm depth were slightly lower than Spring at UD and LTD plots, and slightly higher at STD plots. While the rates of decline were similar between years, protease showed a continuous decline with depth (levelling at 30–40 cm at LTD), rather than the u-shaped pattern typically seen in Autumn 2014 and Spring 2015 (Fig. 11); urease declines were similar between years at UD and LTD, but at STD dropped rapidly to 10–20 cm and rose slowly from 30–50 cm; while arginine levels at depth remained very similar to those in Spring 2015, except for slightly lower topsoil levels and slightly higher subsoil levels (10–30 cm) at the STD plot.

Correlations between protease and urease activity and the average  $E_h$  of the preceding two weeks were frequently statistically significant (Table 2), especially in Summer 2015 samples. This positive correlation between  $E_h$  and enzyme activity points to higher microbial activity under less reducing conditions.

**Table 2.** Within-profile correlation between  $E_h$  (mean of preceding two weeks) and enzyme activities measured from peat samples (N=5 for each sampling) taken from Lakkasuo in Autumn 2014 (LTD and STD only), Spring 2015 and Summer 2015. Numbers in cells indicate how often a significant ( $p < 0.05$ ) correlation was observed between each variable pair.

Variable	$E_h$	Protease	Urease	Arg.deam.	$E_h$	Protease	Urease	Arg.deam.	$E_h$	Protease	Urease	Arg.deam.
Treatment	LTD	LTD	LTD	LTD	STD	STD	STD	STD	UD	UD	UD	UD
$E_h$ -LTD	1											
Protease-LTD	1/3	1										
Urease-LTD	2/3	0/3	1									
Arg.deam.-LTD	0/3	0/3	0/3	1								
$E_h$ -STD	2/3	0/3	1/3	1/3	1							
Protease-STD	3/3	0/3	2/3	1/3	2/3	1						
Urease-STD	2/3	0/3	3/3	1/3	2/3	1/3	1					
Arg.deam.-STD	0/3	0/3	0/3	0/3	0/3	0/3	0/3	1				
$E_h$ -UD	2/2	0/2	2/2	1/2	2/2	2/2	2/2	0/2	1			
Protease-UD	2/2	1/2	2/2	0/2	2/2	2/2	1/2	0/2	2/2	1		
Urease-UD	2/2	0/2	2/2	1/2	1/2	1/2	2/2	0/2	2/2	2/2	1	
Arg.deam-UD	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	0/2	1

### 3.4 Statistical modelling of redox potential

The results of modelling proved inconclusive. It was not possible to predict momentary  $E_h$ , or even the momentary change or direction of change in  $E_h$  based on momentary WTL, time elapsed since WTL had risen above a given level, or the direction of WTL change, even when combined with soil temperature and precipitation.

While it was possible to fit the non-linear model to some inundation periods, mostly on the ME-LTD plot at 25 cm depth, more often than not  $E_h$  showed no consistent behaviour during inundation. Furthermore, the predictive power of the linear models proved extremely poor when autocorrelation of  $E_h$  was taken into account.

For further description of the models, please see Appendix A.

## 4 Discussion

### 4.1 WTL and redox

Though predicting  $E_h$  using WTL proved impossible based on the models we applied, our results show a clear effect of WTL regime on  $E_h$  in terms of depth distribution. At the ME-LTD, ME-STD and OM-LTD plots,  $E_h$  varied widely between 5 and 15 cm depth (and also at 25 cm depth in the case of the LTD plots), while at the ME-UD plot, variability was mostly limited to 5 cm depth (Fig. 3).

Generally speaking, differences between sensors were more pronounced when average  $E_h$  was between the extremes, i.e.  $O_2$ -reducing and  $CO_2$ -reducing conditions, and when  $E_h$  was in a state of change (Fig 3). This emphasises the highly localised nature of redox conditions and the fact that the values at each Pt sensor only represent the conditions prevalent in a small volume of soil.

The overall  $E_h$  values at different depths could be interpreted as indicating different TEAs being used at different depths in the peat matrix. In this case,  $E_h$  values over 400mV, mostly present at 5 cm in all treatments, would be indicative of oxic conditions, values of +100...+400 at 15cm depth would indicate  $NO_3^-$  reduction, and  $E_h$  values at 25 cm depth and deeper would be more indicative of  $Fe^{3+}$ ,  $SO_4^{2-}$  or  $CO_2$  reduction (Table 1). Just such a connection between dynamic WTL and  $Fe^{2+}$  and  $SO_4^{2+}$  concentrations in porewater (indicating  $E_h$  values below +50 and -100 mV, respectively) has previously been observed in a degraded fen in Central Europe (Estop-Aragonés et al., 2013), confirming that WTL does indeed have an effect on  $E_h$  conditions, even if current measurements and analysis methods fail to record the connection. A more recent example of the effect of WTL on the dominant redox pair and  $E_h$  is in Boonman et al. (2024), where subsoil irrigation increased the average WTL on drained, nutrient-rich agricultural peatlands, which corresponded to lower  $E_h$  closer to the surface. Also the dominant redox process was affected by WTL, with the dominant WTL corresponding to  $E_h$  values implying iron reduction. Note, however, that this connection broke down at times, mostly when WTL was below -40 cm. Our results were more ambiguous, however, as there appeared to be no clear WTL that would make or break the connection between WTL and apparent Fe-reduction. Rather, low WTL during summer did not cause the 0 mV isopotential to drop in June or August 2015 or 2016 on the

ME-LTD plot, whereas in the Autumn of 2015 a drop and consequent rise in WTL was reflected as a corresponding drop and rise in the 0 mV depth isotherm (Fig. 3).

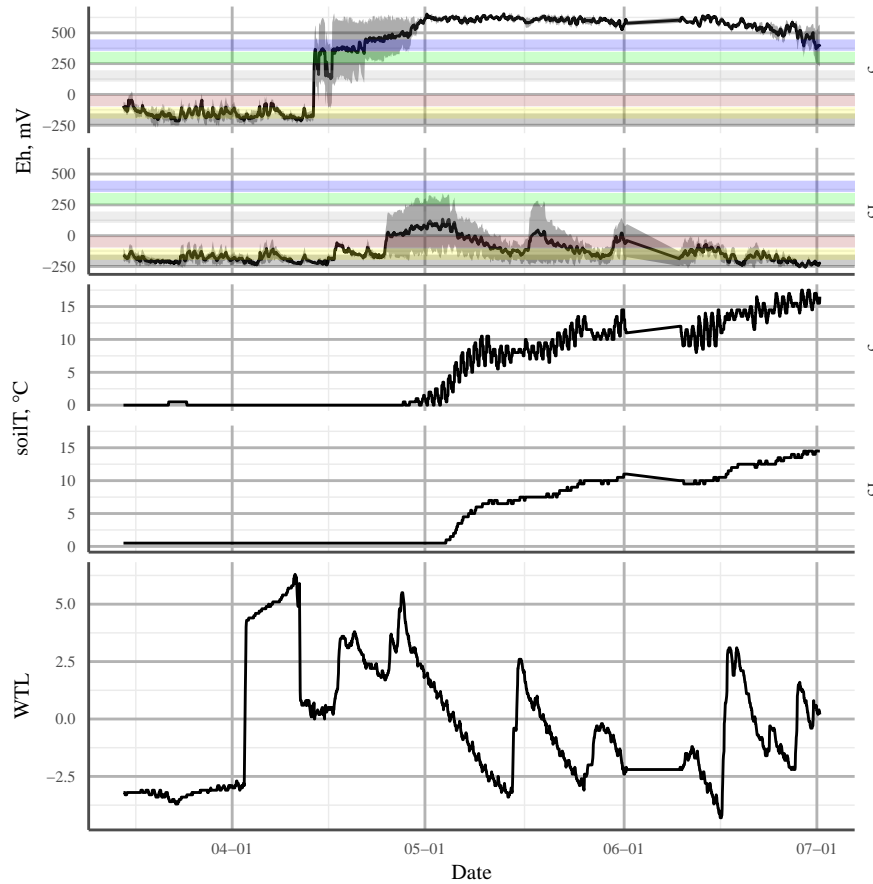
325 The effect of WTL on  $E_h$  was complex. One might expect a pattern where WTL rising over a certain level would, in all cases, cause conditions to become more reducing as water blocks movement of air into the soil matrix. This type of relationship was previously observed by Mchergui et al. (2014), under laboratory conditions, on silt and sandy mineral soils under both permanent and intermittent flooding, with the settling time for  $E_h$  varying between 0 and 14 days, depending on soil type. On peatlands, de Mars and Wassen (1999) reported a clear relationship between WTL and  $E_h$  values at 15 cm depth, especially  
330 on moderately rich fens. Note, however, that in their study, measurements were taken at many sites with only 2 – 4 time points measured per site; hence, variation in  $E_h$  values at a given WTL was in the range of 200 – 400 mV throughout the WTL range.

Our continuous measurements suggested that factors in addition to WTL were affecting the  $E_h$ , and that the relationship between WTL and  $E_h$  was not always simple. For example, in the Summer of 2015 at the LTD plot,  $E_h$  at 25 cm depth, where most changes in  $E_h$  were observed, responded directly to changes in WTL, even though WTL at the time was below -30 cm.  
335 At the same time, there was no change in  $E_h$  below 25 cm depth, even when the WTL dropped below -50 cm. At the UD plot, there were several occasions where a rising WTL caused a rise in  $E_h$ , e.g. after snow melt in Spring and Summer 2016 (Fig. 12). This could be an indication of incoming water bringing in new TEAs from the surrounding area. A similar trend of  $E_h$  increasing with rising WTL was observed by Yu et al. (2008) in a swamp environment. In this case, the major difference to our site was that the WTL was, on average, much higher than the soil surface. It was assumed by the authors that the rise in  $E_h$  was  
340 caused by  $O_2$  transport to the rooting zone by plants. As the UD plot in our study was dominated by aerenchymous vegetation, such as sedges and wetland herbs (see section 2.1), this could also be the case in the current experiment. That the effect was limited to 5 cm depth is consistent with the maximal  $O_2$  release depth observed for *Carex rostrata* (bottle sedge) (Mainiero and Kazda, 2005).

Over the same period, rain events caused a rapid drop in  $E_h$  at 15 and 25 cm depth in the ME-LTD plot, despite the WTL  
345 never rising above 40 cm. This suggests that infiltration of rain water down to the WTL following heavy rainfall causes enough blockage of gas transport to prevent reoxidisation of the TEAs used by the microbial population.

Mitchell and Branfireun (2005) found that late summer precipitation events of > 30 mm sharply increased the  $E_h$  close to surface on an upland-peatland interface. The WTL was close to surface in their study, comparable to our ME-UD plot; they also observed transport of  $SO_4^{2-}$  and Fe(II), and to a lesser extent Fe(III), from the upland to the edge of the peatland during  
350 stormflow conditions. They interpreted the  $E_h$  rise as due to transport of dissolved  $O_2$  in the rainwater along the flowpath from the watershed to the peatland. Our observations outlined above indicate the same processes, i.e. on the undrained plot, hydrological connection to the watershed is unbroken and oxygen can be transported along the water, whereas on the drained plots, water from the uplands is diverted by the drainage network and only the oxygen delivered in the rainwater falling on the plot itself affected the  $E_h$ . Undrained ombrotrophic peatlands, which do not receive water from a mineral soil watershed,  
355 could be expected to exhibit similar response to precipitation events as the ME-LTD plot in this study. Season could also be expected to play a role in how precipitation affects redox conditions: during peak growing season (late June, July) the soil is warmer than during early and late growing season, as is generally the precipitation itself. This causes  $O_2$  availability after a

precipitation event to drop in two ways: first,  $O_2$  solubility in water decreases along rising temperature; and second, higher biological activity causes the  $O_2$  in the water to be used up more quickly. Thus, precipitation during peak growing season could  
 360 cause a drop in  $E_h$  below soil surface, while off-season precipitation could cause a rise in  $E_h$  instead.



**Figure 12.** Redox potential ( $E_h$ ) and soil temperature (soiT) at 5 and 15 cm depth, and water table level (WTL) at the undrained (UD) plot in Spring 2016. Solid black line in  $E_h$ : mean of three profiles, shaded area: Standard Error. Horizontal coloured bars represent approximate theoretical  $E_h$  values of transition between different TEAs, from top to bottom:  $O_2$ ,  $NO_3^-$ ,  $Mn^{4+}$ ,  $Fe^{3+}$ ,  $SO_4^{2-}$ ,  $CO_2$

The difference in  $E_h$  dynamics at the ME plots between winters appeared to be related to differences in WTL dynamics (Fig. 3). During the winter of 2014-2015, for example, the WTL fluctuated between -30 and -10 cm at the LTD plot, and between -20 and -5 at the STD and UD plots; whereas the WTL remained constant during the winter of 2015-2016, lying either close to the surface or above it at all sites. This may have been due to a brief period of frost early in the winter, when up to 20 cm  
 365 of snow accumulated, followed by thawing over about a month, followed by freezing conditions once again. This could have formed a layer of ice on the surface of the peat, preventing flow of  $O_2$  into the system. The fact that the deeper snow pack during the winter of 2014-2015 failed to cause reducing conditions in the top layers suggests that  $O_2$  movement into peat is

not necessarily prevented by snow. As redox conditions are intrinsically linked with water chemistry (e.g. Zak et al., 2004; Kjaergaard et al., 2012; Riedel et al., 2013; Kaila et al., 2016), this suggests that there may be differences in Spring melt runoff  
370 water quality and CH<sub>4</sub> emissions between years, depending on whether gas transport between the atmosphere and soil has been blocked by ice or not. Surface, as opposed to deep, peat through-flow, indicative of frozen top soil, has previously been associated with a higher CH<sub>4</sub> content (Dinsmore et al., 2011), as well as lower dissolved O<sub>2</sub> concentrations in Spring runoff water (Eskelinen et al., 2016).

The behaviour of E<sub>h</sub> on the ME-LTD and OM-LTD plots revealed bimodality in E<sub>h</sub> value distribution on the OM-LTD plot  
375 and multimodality on the ME-LTD plot (Fig. 10). This suggests that the presence of alternative TEAs at the ME plot caused the drop in E<sub>h</sub> to "pause" at characteristic levels during inundated conditions. Both plots had peaks around +450 mV and below -300 mV, suggesting O<sub>2</sub>-driven respiration and CO<sub>2</sub> reduction through methanogenesis, respectively (Table 1). The additional density peak observed at the ME-LTD plot was around 0 mV, suggesting that the Fe<sup>3+</sup>–Fe<sup>2+</sup> redox pair was responsible. A possible complication regarding this interpretation is that the WTL at the OM-LTD plot was more stable than at the ME-LTD  
380 plot (Fig. 3). However, the sensors at 25 cm depth on the OM-LTD plot experienced more transitions between non-inundated and inundated conditions than the corresponding sensors on the ME-LTD plot (26 vs 14 transitions, respectively). As alternative TEAs are important during transition phases, this suggests that the OM-LTD plot should be more, not less, prone to multimodal E<sub>h</sub> behaviour than the ME-LTD plot if alternative TEAs are present.

To the best of our knowledge, multi-year studies of Spring melt-water quality and greenhouse gas emissions in relation to  
385 soil frost conditions have yet to be conducted. Nevertheless, the dynamics between soil frost and E<sub>h</sub> observed in this study (Fig 3) suggest that pore-water chemical quality under the snow pack and its relationship to E<sub>h</sub> measurements could be an interesting subject for future study.

The complexity of the WTL-E<sub>h</sub> relationship revealed by wavelet analysis most likely explains why the non-dynamic linear and non-linear models failed to predict changes in E<sub>h</sub> using momentary WTL or duration of inundation. Furthermore, there  
390 was no sign of E<sub>h</sub> reacting more rapidly to changes in WTL at the OM-LTD plot than the ME plot, whether using 0 mV depth isopotential-WTL wavelet analysis or a comparisons of daily  $\delta E_h$  value distribution. This, combined with our disappointing modelling results, raises the question of whether non-dynamic linear or nonlinear models are the best way of modelling a complex phenomenon such as the redox state of a soil system.

A possible non-dynamic approach could be to use a multinomial or ordinal model (e.g. Liang et al., 2020), which could better  
395 reflect the multimodal nature of redox conditions without the need for extensive estimation of available electron acceptors, which a dynamic model such as the decomposition model of DNDC (Haas et al., 2013) would require. This approach would produce the probability of each electron acceptor being the dominant one at a given moment, reflected in observations by a redox potential range, rather than a specific redox potential.

Applying a dynamic approach to peat redox state modelling would require extensive laboratory-, microcosm- and mesocosm  
400 studies to parametrise the links between peat physico-chemical properties such as pore size distribution, elemental and organic carbon composition, pore- and runoff water quality and the mass and species diversity of the microbiota using the TEAs.

The difficulties faced in characterising  $E_h$  behaviour based on WTL could, at least in part, be understood by defining exactly what WTL measured from a WTL well represents. It is easy to mistake the clear interface between air and the water surface in a well as representing a sudden switch from dry to wet conditions in the peat. In fact, the water level in the well represents the depth at which water content in the peat reaches field capacity, ie. the degree of saturation where all pores small enough to allow capillary action to hold on to water against the force of gravity are filled. Depending on the peat's pore size distribution, there will still be varying amounts of non-saturated pores present in the peat at field capacity, and the proportion of saturated pores will decrease linearly above the water level (Zhang and Furman, 2021). Thus, not only can the actual soil water content at the WTL differ between sites and treatments, it may actually still increase below the WTL. As saturation of pores with water is the main obstacle preventing  $O_2$  from penetrating the soil profile, soil water content and pore size distribution at different depths could prove a better means of predicting  $E_h$  behaviour in peatlands, and soils in general, than simply measuring the WTL. Indeed, Estop-Aragonés et al. (2012) reported that concentration of dissolved oxygen in peat porewater did not reliably drop to zero when WTL reached the peat layer; instead, significant amounts of  $O_2$  were occasionally available in the porewater even when WTL was more than 10 cm above the monitored peat layer. Correspondingly, the air-filled pore space depth profile did not linearly follow the WTL.

Our observation of no clear-cut correlation between  $E_h$  and WTL could, in part, explain the often poor predictive power of WTL in momentary  $CH_4$  dynamics (e.g. Koskinen et al., 2016; Korkiakoski et al., 2017) as  $CH_4$  production and oxidation are intrinsically connected with  $E_h$  conditions.

## 4.2 Enzyme activity

Studies reporting hydrolytic enzyme activity in peatlands are rare. However, compared to the available research, (e.g. Wojciech and Styła, 2011; Lloyd and Sheaffe, 1973; Singh and Kumar, 2008), the activities observed at the ME plots were within an order of magnitude of previously reported values, whether they were measured in peat or mineral soil.

The variations in enzyme activity observed at the ME plots between Autumn 2014 and Summer 2015 were potentially the result of local differences in organic substrate quality and, more specifically, differences in the activity of hydrolytic enzymes at different points and depths at each site.

Generally speaking, in more oxidative environments, i.e. those with higher  $E_h$ , enzyme activity increased (Table 2). This was almost universally true for protease and urease activity, though not for arginine deaminase. This connection between hydrolytic enzymes and  $E_h$  indicates that degradation of complex organic substances is inhibited under reducing conditions, which would promote carbon accumulation in the soil. Interestingly, this general increase in enzyme activity under more oxic conditions was also confirmed by Alef and Kleiner (1987a) for arginine deaminases, the authors going on to state that aerobic and anaerobic ammonification of arginine (including their ratio) was most likely strongly influenced by the redox potential. Consequently, the non-correlation between arginine deaminase and redox potential observed in our study may have been due to the inhibition of arginine deaminase, perhaps by an increased amount of readily available substrate for microorganisms and a high amount of  $NH_4$  in the soil (Alef and Kleiner, 1987b; Lin and Brookes, 1999). The potential for ammonification by microorganisms could also have been limited by a higher C/N ratio in the soil environment (Fujii et al., 2019). While we are unable to deduce the

exact cause of this unexpected arginine deaminase inhibition from our results, redox potential could prove to be an indicator for this condition.

There are two other possible explanations for the differences observed, both related to water regime. Phenol oxidases are one of the few enzyme groups capable of breaking down phenolic compounds, and any reduction in their activity, i.e. their ability to degrade phenolic compounds, may explain the reduced activity of hydrolytic enzymes (Bonnett et al., 2006). Phenol oxidases are a group of extracellular enzymes that catalyse the oxidation of lignin and phenolic compounds in soil organic matter (Sanchez-Julia and Turner, 2021). The influence of such oxidising enzymes on the activity of hydrolytic enzymes has been described by Freeman et al. (2001) using the 'enzyme latch' hypothesis, which states that "under conditions of full saturation with water, there is a lack of O<sub>2</sub> in the environment, which suppresses the activity of oxidising enzymes". Furthermore, the low levels of O<sub>2</sub> naturally found in peat bogs and fens leads to the accumulation of polyphenolic substances, which bind to hydrolytic enzymes and thereby inhibit their functioning (Fenner and Freeman, 2011). However, Romanowicz et al. (2015) found that fluctuations in the WTL can cause changes in the accumulation of polyphenolic substances (i.e. phenol oxidase activity) in peat soils, as well as redox potential, with both redox potential and phenol oxidase activity decreasing with increasing WTL. Up until now, however, studies on the impact of WTL on extracellular enzymes have produced conflicting results (e.g. Xiang and Freeman, 2009; Sun et al., 2010; Fenner et al., 2011), and further research will be needed to clarify the processes involved.

A second possibility is the 'iron gate' paradigm described by Wang et al. (2017), whereby the effect of phenolic substances on the oxidation, and subsequent mineralisation, of soil organic carbon (SOC) following a drop in water levels (water balance) is attributed to Fe transformation, which is redox induced (Li et al., 2012). In this case, Fe(II) is oxidised to Fe(III) through abiotic or biotic reactions (Hall and Silver, 2013). As Fe(II) positively affects the oxidation of phenols (Sinsabaugh, 2010; Hall and Silver, 2013), Fe oxidation may serve to protect against C loss as the WTL drops (Wang et al., 2017). Under conditions of reduced moisture, oxidation of Fe(II) will reduce the oxidation of phenols and thereby affect the mineralisation rate of SOC (Wang et al., 2017), thus indirectly affecting the activity of hydrolytic enzymes. The Fe oxidation process also lowers the pH, which will also affect phenol oxidase activity as its optimal pH is close to neutral (Sinsabaugh, 2010; Toberman et al., 2010). Furthermore, the products of Fe(II) oxidation can immobilise P and dissolved organic C, thereby reducing C and P availability, and thus enzyme synthesis (Lalonde et al., 2012; Riedel et al., 2013). Extracellular enzymes can also bind to amorphous ferric hydroxide (Fe(OH)<sub>3</sub>) formed during Fe(II) oxidation, further limiting the catalytic activity of enzymes in peat soil (Allison, 2006; Wen et al., 2019).

## 5 Conclusions

To the best of our knowledge, this is the first study to undertake long-term continuous monitoring of redox conditions at multiple depths in a boreal peatland while also monitoring soil WTL and temperature. Furthermore, the side-by-side existence of different drainage histories on plots with similar ecohydrological development before drainage makes our dataset unique.



Our results show that enzyme activity changed at different depths and in different seasons, most likely due to differences in substrate quality at the different sites, particularly in relation to water saturation. Such effects were particularly pronounced where water saturation was directly affected by the drainage regime.

Our first hypothesis, that drainage regime through dominant WTL level would be reflected in the depth at which most  $E_h$  dynamics occur, was supported by the data, with  $E_h$  at its most dynamic at 25 cm depth at the ME-LTD plot, 15 cm at the ME-STD plot and 5 cm at the ME-UD plot.

Our second hypothesis, that  $E_h$  and Fe reduction isopotential fluctuations would closely follow WTL fluctuations, was only partially supported by the data. The connection between WTL and  $E_h$  was intermittently in syn- and counter-phase, suggesting that different hydrological phenomena may affect redox conditions in opposite ways, even where the effect on WTL is constant in all cases.

Our third hypothesis, that reduced levels of alternative TEAs in the OM-LTD plot would cause Eh conditions to become reducing more quickly after a rise in the WTL, was not supported by our data, which indicated that alternative TEAs were utilised in drained peatlands with more decomposed peat. It is possible that, in this case, the greater organic matter-associated electron accepting capacity of decomposed *Sphagnum* peat, as opposed to sedge peat, compensated for the lack of mineral elements.

Our fourth hypothesis, that  $E_h$  measurements at the OM drained plot would display a bi-modal distribution, and those in the ME drained plot a multi-modal distribution due to the greater availability of mineral TEAs, was supported. This suggests that, if the electron accepting capacity of organic matter is slowing down the transition between oxic and CO<sub>2</sub>-reducing conditions, its  $E_h$  may not be picked up by the sensor's Pt electrodes.

Our fifth hypothesis, that the ME-LTD plot, with its generally higher  $E_h$  values, would display higher enzyme activity than the ME-UD plot, was partially supported by the data. While a higher  $E_h$  correlated with higher enzyme activity within profiles, the opposite was true between plots.

Finally, the relationship between WTL fluctuation and redox conditions proved too complex to model with non-dynamic linear or non-linear modelling, suggesting that characterisation of redox fluctuations between treatments will require more complex models.

We suggest that future field  $E_h$  measurement campaigns should be paired with pore water chemistry and greenhouse gas monitoring, and that the role of OM and changes in input water quality in the yearly redox cycle should be clarified. Smaller-scale laboratory and other *ex-situ* studies should be made in order to tease apart the contributions of different factors affecting redox dynamics in peatlands.

*Data availability.* The original ( $E_h$ , WTL, soilT, enzymatic activity, pH) data is available in Zenodo: 10.5281/zenodo.12544806. Other data and scripts are available from corresponding author upon reasonable request.

## Appendix A: Modelling results

### 500 A1 Methods

For the nonlinear models (Eq. A1), it was assumed that different sites and depths in the peat profile would have characteristically low  $E_h$  values toward which the system would asymptotically move under inundated conditions, i.e. when movement of  $O_2$  into the peat matrix was limited or prevented. It was also assumed that  $E_h$  values would rise rapidly once inundation ended as oxygen penetration into the soil profile oxidised the reduced TEAs. Consequently, an exponential asymptotic model was fitted  
 505 separately to the mean measurements from each peat depth on all treatments using the time inundated (i.e. the time elapsed since WTL had risen above each Pt sensor in the peat profile) as the explanatory variable. Each period of inundation was fit separately, with a lower limit of 240 hours for inundation length to exclude short-term WTL fluctuations that may not be long enough for the  $\delta E_h$  to show signs of saturation. In this way, characteristic asymptote and rate values were obtained for each depth and treatment.

$$510 \quad E_h = A_0 + (A_{ref} - A_0) * (1 - e^{-t_{in}/\tau}) \quad (A1)$$

where  $E_h$  is the redox potential measured,  $A_0$  is  $E_h$  at the beginning of the inundation period,  $A_{ref}$  is the asymptotic term of  $E_h$ ,  $t_{in}$  is the time inundated (hours) and  $\tau$  is the rate term. Model fitting was undertaken in the R statistical environment (R Core Team, 2023) using the Levenburg-Marquardt-algorithm and the minpack.lm library (Elzhov et al., 2022).

A multiple linear model was also fit to each treatment and sensor depth using available ancillary measurements as parameters  
 515 (Table A1). Autocorrelation between consecutive  $E_h$  measurements was then estimated using the ar function in R, with a lag value of 1. The autocorrelation was then entered into the linear model fitting procedure.

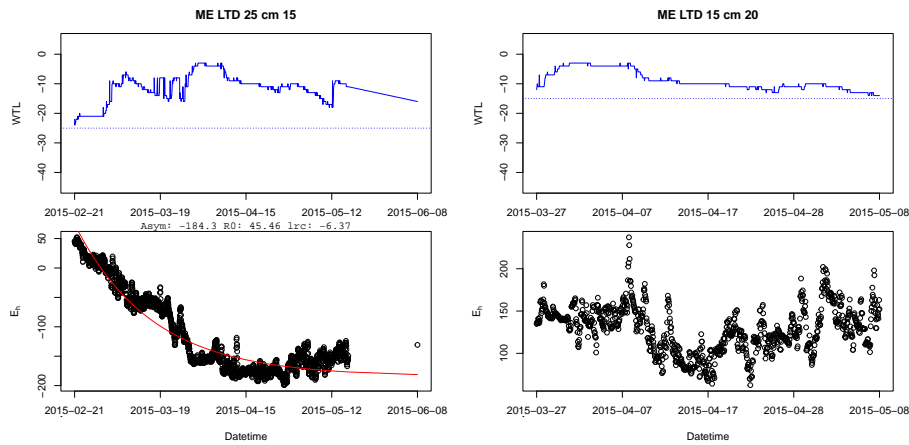
**Table A1.** Parameters used in the linear models

Name	explanation	unit
$D_o$	depth of WTL over the sensor	cm
$Pr_d$	running sum of precipitation, 24h	mm
$Pr_w$	running sum of precipitation, 7 days	mm
$T_{soil}$	temperature of soil at sensor depth	°C
$T_{air}$	temperature of air at 4.2m height (at SMEAR II station)	°C
$I_{in}$	time since WTL rose above sensor depth	h

## A2 Results

It was only possible to fit the asymptotic model to 36% of inundation periods with a continuous duration of 240 hours or more. Grouped by treatment and depth, fitting success rate varied between 0 and 56% of inundation periods for those groups where the number of viable periods available was  $> 1$ .

More often than not, the  $E_h$  value did not react to inundation in a monotonic fashion, which was the main assumption behind use of an asymptotic function to model the connection between time inundated and  $E_h$ . In practice,  $E_h$  values often fluctuated around the Fe reduction level or higher, reaching levels indicative of  $O_2$  reduction, even during months-long inundation periods.

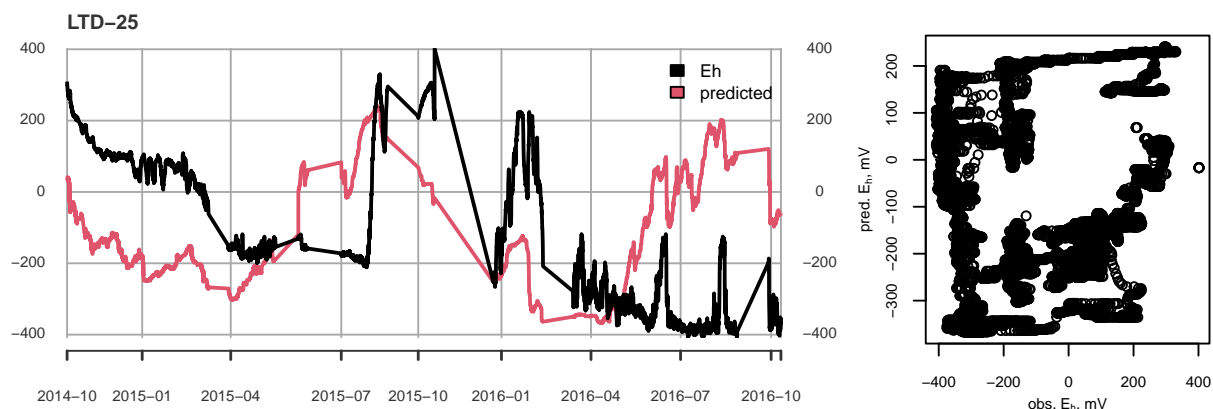


**Figure A1.** Example of a successful (left) and failed (right) fit of the asymptotic model to  $E_h$  during periods of inundation. Top panels, solid blue line: water table level, dashed blue line: position of  $E_h$  sensor in the depth profile. Left Figures: Asym is the value of the asymptote, R0 the  $E_h$  value when inundation began and lrc the natural logarithm of the rate constant ( $\text{mV s}^{-1}$ )

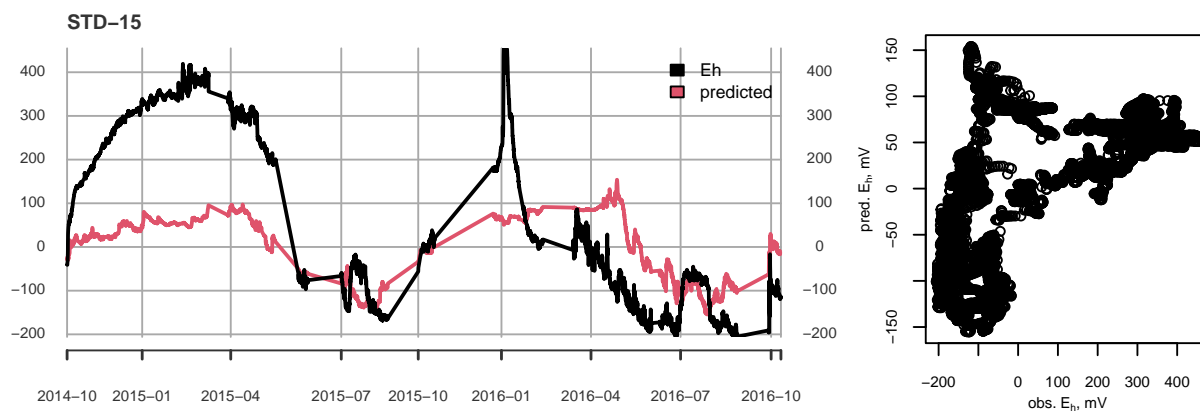
Even though the linear models were fitted for each depth at each treatment separately, for the most part they completely failed to reproduce the temporal dynamics of  $E_h$  (Figs. A2, A3). Only on the UD plot, at 35 cm depth, was the average  $E_h$  value predicted well and even then any shorter term temporal dynamics were completely missing (Fig. A4).

## A3 Conclusions

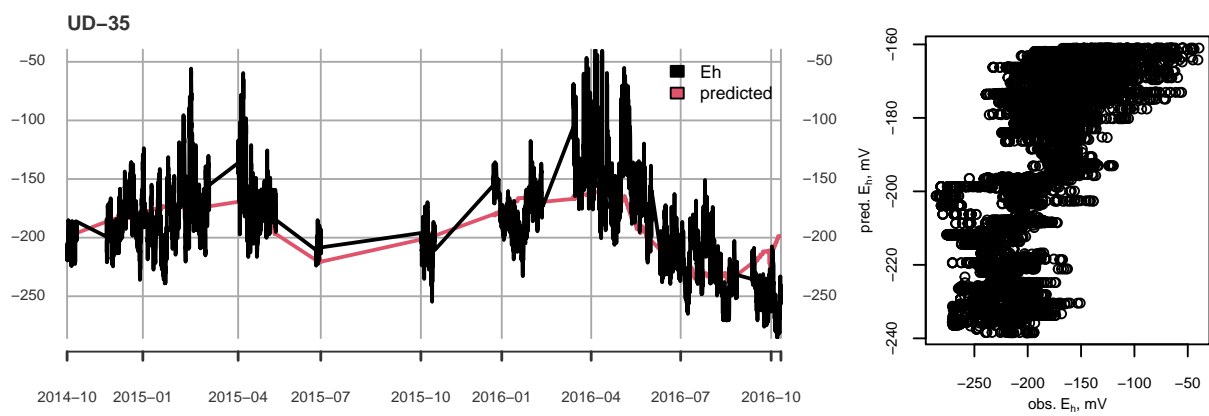
In the data presented here, momentary WTL, soil and air temperature and historical precipitation were not sufficient to reliably explain the temporal changes in  $E_h$ . Possibly some sort of dynamic model could work better in predicting how  $E_h$  is affected by environmental variables over time.



**Figure A2. Left:** Measured (black) and predicted (red) redox potential, **right:** observed vs. predicted  $E_h$ , at ME-LTD plot, 25 cm depth.



**Figure A3. Left:** Measured (black) and predicted (red) redox potential, **right:** observed vs. predicted  $E_h$ , at ME-STD plot, 15 cm depth.



**Figure A4.** Left: Measured (black) and predicted (red) redox potential, right: observed vs. predicted  $E_h$ , at ME-UD plot, 35 cm depth.

*Author contributions.* MK processed and analysed the data and did all calculations and model fitting unless otherwise noted, wrote the first draft and revised the manuscript with input from all co-authors; JA did the wavelet analysis of hourly data; VV and LH performed the field work, analysis and interpretation of the enzyme activity assessment and wrote the subsections concerning enzymatic activity; KR performed English proofreading; MV designed, built and installed the field redox potential measurement system; RL conceived the original research  
545 idea, planned the field experiment and supervised the project.

*Competing interests.* The authors declare no competing interests

*Acknowledgements.* The authors wish to thank Petra Strakova for help with peat sampling and chemical analysis. This work was supported by the Finnish Research Council (Grant no's 289116 and 339489) and the Maj and Tor Nessling Foundation.

## References

- 550 Aalto, J., Aalto, P., Keronen, P., Kolari, P., Rantala, P., Taipale, R., Kajos, M., Patokoski, J., Rinne, J., Ruuskanen, T., et al.: SMEAR II Hyytiälä forest meteorology, greenhouse gases, air quality and soil, <https://doi.org/10.23729/23dd00b2-b9d7-467a-9cee-b4a122486039>, university of Helsinki, Institute for Atmospheric and Earth System Research, 2023.
- Aeschbacher, M., Vergari, D., Schwarzenbach, R. P., and Sander, M.: Electrochemical Analysis of Proton and Electron Transfer Equilibria of the Reducible Moieties in Humic Acids, *Environmental Science & Technology*, 45, 8385–8394, <https://doi.org/10.1021/es201981g>, 2011.
- 555 Alef, K. and Kleiner, D.: Arginine Ammonification, a Simple Method to Estimate Microbial Activity Potentials in Soils, *Soil Biology and Biochemistry*, 18, 233–235, [https://doi.org/10.1016/0038-0717\(86\)90033-7](https://doi.org/10.1016/0038-0717(86)90033-7), 1986.
- Alef, K. and Kleiner, D.: Applicability of Arginine Ammonification as Indicator of Microbial Activity in Different Soils, *Biology and Fertility of Soils*, 5, 148–151, <https://doi.org/10.1007/BF00257650>, 1987a.
- Alef, K. and Kleiner, D.: Estimation of Anaerobic Microbial Activities in Soils by Arginine Ammonification and Glucose-Dependent CO<sub>2</sub>-production, *Soil Biology and Biochemistry*, 19, 683–686, [https://doi.org/10.1016/0038-0717\(87\)90048-4](https://doi.org/10.1016/0038-0717(87)90048-4), 1987b.
- 560 Allison, S. D.: Soil Minerals and Humic Acids Alter Enzyme Stability: Implications for Ecosystem Processes, *Biogeochemistry*, 81, 361–373, <https://doi.org/10.1007/s10533-006-9046-2>, 2006.
- Belyea, L. R.: A Novel Indicator of Reducing Conditions and Water-Table Depth in Mires, *Functional Ecology*, 13, 431–434, <https://doi.org/10.1046/j.1365-2435.1999.00333.x>, 1999.
- 565 Blodau, C., Basiliko, N., and Moore, T. R.: Carbon Turnover in Peatland Mesocosms Exposed to Different Water Table Levels, *Biogeochemistry*, 67, 331–351, <https://doi.org/10.1023/B:BIOG.0000015788.30164.e2>, 2004.
- Bonnett, S. A. F., Ostle, N., and Freeman, C.: Seasonal Variations in Decomposition Processes in a Valley-Bottom Riparian Peatland, *Science of The Total Environment*, 370, 561–573, <https://doi.org/10.1016/j.scitotenv.2006.08.032>, 2006.
- Boonman, J., Harpenslager, S. F., van Dijk, G., Smolders, A. J. P., Hefting, M. M., van de Riet, B., and van der Velde, Y.: Redox Potential Is a Robust Indicator for Decomposition Processes in Drained Agricultural Peat Soils: A Valuable Tool in Monitoring Peatland Wetting Efforts, *Geoderma*, 441, 116 728, <https://doi.org/10.1016/j.geoderma.2023.116728>, 2024.
- 570 Borch, T., Kretzschmar, R., Kappler, A., Cappellen, P. V., Ginder-Vogel, M., Voegelin, A., and Campbell, K.: Biogeochemical Redox Processes and Their Impact on Contaminant Dynamics, *Environmental Science & Technology*, 44, 15–23, <https://doi.org/10.1021/es9026248>, 2010.
- 575 Burgin, A. J. and Loecke, T. D.: The Biogeochemical Redox Paradox: How Can We Make a Foundational Concept More Predictive of Biogeochemical State Changes?, *Biogeochemistry*, <https://doi.org/10.1007/s10533-023-01036-9>, 2023.
- de Mars, H. and Wassen, M. J.: Redox Potentials in Relation to Water Levels in Different Mire Types in the Netherlands and Poland, *Plant Ecology*, 140, 41–51, <https://doi.org/10.1023/A:1009733113927>, 1999.
- Dinsmore, K. J., Billett, M. F., Dyson, K. E., Harvey, F., Thomson, A. M., Piirainen, S., and Kortelainen, P.: Stream Water Hydrochemistry as an Indicator of Carbon Flow Paths in Finnish Peatland Catchments during a Spring Snowmelt Event, *Science of The Total Environment*, 409, 4858–4867, <https://doi.org/10.1016/j.scitotenv.2011.07.063>, 2011.
- 580 Elzhov, T. V., Mullen, K. M., Spiess, A.-N., and Bolker, B.: minpack.lm: R Interface to the Levenberg-Marquardt Nonlinear Least-Squares Algorithm Found in MINPACK, Plus Support for Bounds, <https://CRAN.R-project.org/package=minpack.lm>, r package version 1.2-2, 2022.

- 585 Eskelinen, R., Ronkanen, A.-K., Marttila, H., Isokangas, E., and Kløve, B.: Effect of Soil Frost on Snow-melt Runoff Generation : Stable Isotope Study in Drained Peatlands, *Boreal Environment Research*, 21, 2743, 2016.
- Estop-Aragonés, C., Knorr, K.-H., and Blodau, C.: Controls on in Situ Oxygen and Dissolved Inorganic Carbon Dynamics in Peats of a Temperate Fen, *Journal of Geophysical Research: Biogeosciences*, 117, <https://doi.org/10.1029/2011JG001888>, 2012.
- Estop-Aragonés, C., Knorr, K.-H., and Blodau, C.: Belowground in Situ Redox Dynamics and Methanogenesis Recovery in a Degraded Fen during Dry-Wet Cycles and Flooding, *Biogeosciences*, 10, 421–436, <https://doi.org/10.5194/bg-10-421-2013>, 2013.
- 590 Ettwig, K. F., Zhu, B., Speth, D., Keltjens, J. T., Jetten, M. S. M., and Kartal, B.: Archaea Catalyze Iron-Dependent Anaerobic Oxidation of Methane, *Proceedings of the National Academy of Sciences*, 113, 12 792–12 796, <https://doi.org/10.1073/pnas.1609534113>, 2016.
- Fenner, N. and Freeman, C.: Drought-Induced Carbon Loss in Peatlands, *Nature Geoscience*, 4, 895–900, <https://doi.org/10.1038/ngeo1323>, 2011.
- 595 Fenner, N., Williams, R., Toberman, H., Hughes, S., Reynolds, B., and Freeman, C.: Decomposition ‘hotspots’ in a Rewetted Peatland: Implications for Water Quality and Carbon Cycling, *Hydrobiologia*, 674, 51–66, <https://doi.org/10.1007/s10750-011-0733-1>, 2011.
- Freeman, C., Liska, G., Ostle, N. J., Lock, M. A., Reynolds, B., and Hudson, J.: Microbial Activity and Enzymic Decomposition Processes Following Peatland Water Table Drawdown, *Plant and Soil*, 180, 121–127, <https://doi.org/10.1007/BF00015418>, 1996.
- Freeman, C., Ostle, N., and Kang, H.: An Enzymic ‘latch’ on a Global Carbon Store., *Nature*, 409, 149, <https://doi.org/10.1038/35051650>, 2001.
- 600 Freeman, C., Ostle, N. J., Fenner, N., and Kang, H.: A Regulatory Role for Phenol Oxidase during Decomposition in Peatlands, *Soil Biology and Biochemistry*, 36, 1663–1667, <https://doi.org/10.1016/j.soilbio.2004.07.012>, 2004.
- Fujii, K., Yamada, T., Hayakawa, C., Nakanishi, A., and Funakawa, S.: Kinetics of Arginine Ammonification to Estimate Microbial Activity of N Mineralization in Forest and Cropland Soils, *European Journal of Soil Biology*, 92, 1–7, <https://doi.org/10.1016/j.ejsobi.2019.03.001>, 2019.
- 605 Gong, J., Kellomäki, S., Wang, K., Zhang, C., Shurpali, N., and Martikainen, P. J.: Modeling CO<sub>2</sub> and CH<sub>4</sub> Flux Changes in Pristine Peatlands of Finland under Changing Climate Conditions, *Ecological Modelling*, 263, 64–80, <https://doi.org/10.1016/j.ecolmodel.2013.04.018>, 2013.
- Green, J. and Paget, M. S.: Bacterial Redox Sensors, *Nature Reviews Microbiology*, 2, 954–966, <https://doi.org/10.1038/nrmicro1022>, 2004.
- 610 Haas, E., Klatt, S., Fröhlich, A., Kraft, P., Werner, C., Kiese, R., Grote, R., Breuer, L., and Butterbach-Bahl, K.: LandscapeDNDC: A Process Model for Simulation of Biosphere–Atmosphere–Hydrosphere Exchange Processes at Site and Regional Scale, *Landscape Ecology*, 28, 615–636, <https://doi.org/10.1007/s10980-012-9772-x>, 2013.
- Hall, S. J. and Silver, W. L.: Iron Oxidation Stimulates Organic Matter Decomposition in Humid Tropical Forest Soils, *Global Change Biology*, 19, 2804–2813, <https://doi.org/10.1111/gcb.12229>, 2013.
- 615 Jaatinen, K., Fritze, H., Laine, J., and Laiho, R.: Effects of Short- and Long-Term Water-Level Drawdown on the Populations and Activity of Aerobic Decomposers in a Boreal Peatland, *Global Change Biology*, 13, 491–510, <https://doi.org/10.1111/j.1365-2486.2006.01312.x>, 2007.
- Kaila, A., Asam, Z., Koskinen, M., Uusitalo, R., Smolander, A., Kiikkilä, O., Sarkkola, S., O’Driscoll, C., Kitunen, V., Fritze, H., Nou-siainen, H., Tervahauta, A., Xiao, L., and Nieminen, M.: Impact of Re-wetting of Forestry-Drained Peatlands on Water Quality—a Laboratory Approach Assessing the Release of P, N, Fe, and Dissolved Organic Carbon, *Water, Air, & Soil Pollution*, 227, 292, <https://doi.org/10.1007/s11270-016-2994-9>, 2016.
- 620



- Kandeler, E. and Gerber, H.: Short-Term Assay of Soil Urease Activity Using Colorimetric Determination of Ammonium, *Biology and Fertility of Soils*, 6, 68–72, <https://doi.org/10.1007/BF00257924>, 1988.
- Kane, E. S., Veverica, T. J., Tfaily, M. M., Lilleskov, E. A., Meingast, K. M., Kolka, R. K., Daniels, A. L., and Chimner, R. A.: Reduction-  
625      Oxidation Potential and Dissolved Organic Matter Composition in Northern Peat Soil: Interactive Controls of Water Table Position and Plant Functional Groups, *Journal of Geophysical Research: Biogeosciences*, 124, 3600–3617, <https://doi.org/10.1029/2019JG005339>, 2019.
- Keller, J. K. and Takagi, K. K.: Solid-Phase Organic Matter Reduction Regulates Anaerobic Decomposition in Bog Soil, *Ecosphere*, 4, art54, <https://doi.org/10.1890/ES12-00382.1>, 2013.
- 630      Kiuru, P., Palviainen, M., Marchionne, A., Grönholm, T., Raivonen, M., Kohl, L., and Laurén, A.: Pore Network Modeling as a New Tool for Determining Gas Diffusivity in Peat, *Biogeosciences*, 19, 5041–5058, <https://doi.org/10.5194/bg-19-5041-2022>, 2022.
- Kjaergaard, C., Heiberg, L., Jensen, H. S., and Hansen, H. C. B.: Phosphorus Mobilization in Rewetted Peat and Sand at Variable Flow Rate and Redox Regimes, *Geoderma*, 173–174, 311–321, <https://doi.org/10.1016/j.geoderma.2011.12.029>, 2012.
- Klüpfel, L., Piepenbrock, A., Kappler, A., and Sander, M.: Humic Substances as Fully Regenerable Electron Acceptors in Recurrently Anoxic  
635      Environments, *Nature Geoscience*, 7, 195–200, <https://doi.org/10.1038/ngeo2084>, 2014.
- Knorr, K. H. and Blodau, C.: Impact of Experimental Drought and Rewetting on Redox Transformations and Methanogenesis in Mesocosms of a Northern Fen Soil, *Soil Biology and Biochemistry*, 41, 1187–1198, <https://doi.org/10.1016/j.soilbio.2009.02.030>, 2009.
- Kokkonen, N. A. K., Laine, A. M., Laine, J., Vasander, H., Kurki, K., Gong, J., and Tuittila, E.-S.: Responses of Peatland Vegetation to 15-Year Water Level Drawdown as Mediated by Fertility Level, *Journal of Vegetation Science*, 30, 1206–1216,  
640      <https://doi.org/10.1111/jvs.12794>, 2019.
- Korkiakoski, M., Tuovinen, J. P., Aurela, M., Koskinen, M., Minkkinen, K., Ojanen, P., Penttilä, T., Rainne, J., Laurila, T., and Lohila, A.: Methane Exchange at the Peatland Forest Floor - Automatic Chamber System Exposes the Dynamics of Small Fluxes, *Biogeosciences*, 14, 1947–1967, <https://doi.org/10.5194/bg-14-1947-2017>, 2017.
- Koskinen, M., Maanavilja, L., Nieminen, M., Minkkinen, K., and Tuittila, E.-S.: High Methane Emissions from Restored Norway Spruce  
645      Swamps in Southern Finland over One Growing Season, *Mires and Peat*, 17, 1–13, 2016.
- Kumaraswamy, S., Ramakrishnan, B., and Sethunathan, N.: Methane Production and Oxidation in an Anoxic Rice Soil as Influenced by Inorganic Redox Species, *Journal of Environment Quality*, 30, 2195, <https://doi.org/10.2134/jeq2001.2195>, 2001.
- Laiho, R. and Laine, J.: Changes in Mineral Element Concentrations in Peat Soils Drained for Forestry in Finland, *Scandinavian Journal of Forest Research*, 10, 218–224, <https://doi.org/10.1080/02827589509382887>, 1995.
- 650      Laiho, R., Peltoniemi, K., and Fritze, H.: Peat Characteristics, Microbial PLFA, and Fungal and Actinobacterial Sequences from Lakkasuo Peatland Drainage Experiment, Year 2004, <https://doi.org/10.5281/zenodo.12566743>, 2024.
- Lalonde, K., Mucci, A., Ouellet, A., and Gélinas, Y.: Preservation of Organic Matter in Sediments Promoted by Iron, *Nature*, 483, 198–200, <https://doi.org/10.1038/nature10855>, 2012.
- Lee, G. R., Gommers, R., Waselewski, F., Wohlfahrt, K., and O’Leary, A.: PyWavelets: A Python Package for Wavelet Analysis, *Journal of*  
655      *Open Source Software*, 4, 1237, <https://doi.org/10.21105/joss.01237>, 2019.
- Li, Y., Yu, S., Strong, J., and Wang, H.: Are the Biogeochemical Cycles of Carbon, Nitrogen, Sulfur, and Phosphorus Driven by the “FeIII–FeII Redox Wheel” in Dynamic Redox Environments?, *Journal of Soils and Sediments*, 12, 683–693, <https://doi.org/10.1007/s11368-012-0507-z>, 2012.

- Liang, J., Bi, G., and Zhan, C.: Multinomial and Ordinal Logistic Regression Analyses with Multi-Categorical Variables Using R, *Annals of Translational Medicine*, 8, 982, <https://doi.org/10.21037/atm-2020-57>, 2020.
- Lin, Q. and Brookes, P. C.: Arginine Ammonification as a Method to Estimate Soil Microbial Biomass and Microbial Community Structure, *Soil Biology and Biochemistry*, 31, 1985–1997, [https://doi.org/10.1016/S0038-0717\(99\)00121-2](https://doi.org/10.1016/S0038-0717(99)00121-2), 1999.
- Lloyd, A. B. and Sheaffe, M. J.: Urease Activity in Soils, *Plant and Soil*, 39, 71–80, <https://doi.org/10.1007/BF00018046>, 1973.
- Mainiero, R. and Kazda, M.: Effects of *Carex Rostrata* on Soil Oxygen in Relation to Soil Moisture, *Plant and Soil*, 270, 311–320, <https://doi.org/10.1007/s11104-004-1724-z>, 2005.
- Marttunen, S.: Impacts of Controlled Redox Conditions on Greenhouse Gas Dynamics from Peat, Master's thesis, University of Helsinki, Faculty of Biological and Environmental Sciences, Helsinki, Finland, 2024.
- Mchergui, C., Besaury, L., Langlois, E., Aubert, M., Akpa-Vinceslas, M., Buatois, B., Quillet, L., and Bureau, F.: A Comparison of Permanent and Fluctuating Flooding on Microbial Properties in an Ex-Situ Estuarine Riparian System, *Applied Soil Ecology*, 78, 1–10, <https://doi.org/10.1016/j.apsoil.2014.01.012>, 2014.
- Melton, E. D., Swanner, E. D., Behrens, S., Schmidt, C., and Kappler, A.: The Interplay of Microbially Mediated and Abiotic Reactions in the Biogeochemical Fe Cycle, *Nature Reviews Microbiology*, 12, 797–808, <https://doi.org/10.1038/nrmicro3347>, 2014.
- Minkinen, K., Vasander, H., Jauhiainen, S., Karsisto, M., and Laine, J.: Post-Drainage Changes in Vegetation Composition and Carbon Balance in Lakkasuo Mire, Central Finland, *Plant and Soil*, 207, 107–120, <https://doi.org/10.1023/a:1004466330076>, 1999.
- Mitchell, C. P. J. and Branfireun, B. A.: Hydrogeomorphic Controls on Reduction–Oxidation Conditions across Boreal Upland–Peatland Interfaces, *Ecosystems*, 8, 731–747, <https://doi.org/10.1007/s10021-005-1792-9>, 2005.
- Palviainen, M., Pumpanen, J., Mosquera, V., Hasselquist, E. M., Laudon, H., Ostonen, I., Kull, A., Wilson, F. R., Peltomaa, E., Könönen, M., Launiainen, S., Peltola, H., Ojala, A., and Laurén, A.: Extending the SUSI Peatland Simulator to Include Dissolved Organic Carbon Formation, Transport and Biodegradation - Proper Water Management Reduces Lateral Carbon Fluxes and Improves Carbon Balance, *Science of The Total Environment*, 950, 175 173, <https://doi.org/10.1016/j.scitotenv.2024.175173>, 2024.
- Peltoniemi, K., Straková, P., Fritze, H., Iráizoz, P. A., Pennanen, T., and Laiho, R.: How Water-Level Drawdown Modifies Litter-Decomposing Fungal and Actinobacterial Communities in Boreal Peatlands, *Soil Biology and Biochemistry*, 51, 20–34, <https://doi.org/10.1016/j.soilbio.2012.04.013>, 2012.
- Pyzola, S. M., Dhakal, P., Coyne, M. S., Grove, J. H., Vandiviere, M. M., and Matocha, C. J.: Transformation of Organic Matter under Anoxic Conditions in Soils, *Science of The Total Environment*, 970, 178 899, <https://doi.org/10.1016/j.scitotenv.2025.178899>, 2025.
- R Core Team: R: A Language and Environment for Statistical Computing, R Foundation for Statistical Computing, 2023.
- Rejsek, K., Formanek, P., and Pavelka, M.: Estimation of Protease Activity in Soils at Low Temperatures by Casein Amendment and with Substitution of Buffer by Demineralized Water, *Amino Acids*, 35, 411–417, <https://doi.org/10.1007/s00726-007-0601-5>, 2008.
- Riedel, T., Zak, D., Biester, H., and Dittmar, T.: Iron Traps Terrestrially Derived Dissolved Organic Matter at Redox Interfaces, *Proceedings of the National Academy of Sciences*, 110, 10 101–10 105, <https://doi.org/10.1073/pnas.1221487110>, 2013.
- Romanowicz, K. J., Kane, E. S., Potvin, L. R., Daniels, A. L., Kolka, R. K., and Lilleskov, E. A.: Understanding Drivers of Peatland Extracellular Enzyme Activity in the PEATcosm Experiment: Mixed Evidence for Enzymic Latch Hypothesis, *Plant and Soil*, 397, 371–386, <https://doi.org/10.1007/s11104-015-2746-4>, 2015.
- Sallantausta, T. and Kaipainen, H.: Water-Carried Element Balances of Peatlands, *Northern peatlands in global climatic change*, edited by: Laiho, R., Laine, J., and Vasander, H., Publications of the Academy of Finland, Edita, Helsinki, pp. 197–203, 1996.

Sanchez-Julia, M. and Turner, B. L.: Abiotic Contribution to Phenol Oxidase Activity across a Manganese Gradient in Tropical Forest Soils, *Biogeochemistry*, 153, 33–45, <https://doi.org/10.1007/s10533-021-00764-0>, 2021.

Schmidbauer, A. R. a. H.: *WaveletComp: Computational Wavelet Analysis*, 2018.

Seybold, C. A., Mersie, W., Huang, J., and McNamee, C.: Soil Redox, pH, Temperature, and Water-Table Patterns of a Freshwater Tidal  
700 Wetland, *Wetlands*, 22, 149–158, [https://doi.org/10.1672/0277-5212\(2002\)022\[0149:SRPTAW\]2.0.CO;2](https://doi.org/10.1672/0277-5212(2002)022[0149:SRPTAW]2.0.CO;2), 2002.

Singh, D. K. and Kumar, S.: Nitrate Reductase, Arginine Deaminase, Urease and Dehydrogenase Activities in Natural Soil (Ridges with Forest) and in Cotton Soil after Acetamiprid Treatments, *Chemosphere*, 71, 412–418, <https://doi.org/10.1016/j.chemosphere.2007.11.005>, 2008.

Sinsabaugh, R. L.: Phenol Oxidase, Peroxidase and Organic Matter Dynamics of Soil, *Soil Biology and Biochemistry*, 42, 391–404,  
705 <https://doi.org/10.1016/j.soilbio.2009.10.014>, 2010.

Straková, P., Niemi, R. M., Freeman, C., Peltoniemi, K., Toberman, H., Heiskanen, I., Fritze, H., and Laiho, R.: Litter Type Affects the Activity of Aerobic Decomposers in a Boreal Peatland More than Site Nutrient and Water Table Regimes, *Biogeosciences*, 8, 2741–2755, <https://doi.org/10.5194/bg-8-2741-2011>, 2011.

Straková, P., Penttilä, T., Laine, J., and Laiho, R.: Disentangling Direct and Indirect Effects of Water Table Drawdown on Above- and Be-  
710 lowground Plant Litter Decomposition: Consequences for Accumulation of Organic Matter in Boreal Peatlands, *Global Change Biology*, 18, 322–335, <https://doi.org/10.1111/j.1365-2486.2011.02503.x>, 2012.

Sun, X., Xiang, W., He, L., and Zhao, Y.: Impacts of Hydrological Conditions on Enzyme Activities and Phenolic Concentrations in Peatland Soil: An Experimental Simulation, *Frontiers of Earth Science in China*, 4, 463–470, <https://doi.org/10.1007/s11707-010-0140-3>, 2010.

Tang, G., Zheng, J., Xu, X., Yang, Z., Graham, D. E., Gu, B., Painter, S. L., and Thornton, P. E.: Biogeochemical Modeling of CO<sub>2</sub> and CH<sub>4</sub>  
715 Production in Anoxic Arctic Soil Microcosms, *Biogeosciences*, 13, 5021–5041, <https://doi.org/10.5194/bg-13-5021-2016>, 2016.

Toberman, H., Laiho, R., Evans, C. D., Artz, R. R. E., Fenner, N., Straková, P., and Freeman, C.: Long-Term Drainage for Forestry Inhibits Extracellular Phenol Oxidase Activity in Finnish Boreal Mire Peat, *European Journal of Soil Science*, 61, 950–957, <https://doi.org/10.1111/j.1365-2389.2010.01292.x>, 2010.

Torrence, C. and Compo, G. P.: A Practical Guide to Wavelet Analysis, *Bulletin of the American Meteorological Society*, 79, 61–78,  
720 [https://doi.org/10.1175/1520-0477\(1998\)079<0061:APGTWA>2.0.CO;2](https://doi.org/10.1175/1520-0477(1998)079<0061:APGTWA>2.0.CO;2), 1998.

Vorenhout, M., van der Geest, H. G., van Marum, D., Wattel, K., Eijssackers, H. J. P., and July, P.: Automated and Continuous Redox Potential Measurements in Soil, *Journal of Environmental Quality*, 33, 1562–1567, <https://doi.org/10.2134/jeq2004.1562>, 2004.

Vorenhout, M., van der Geest, H. G., and Hunting, E. R.: An Improved Datalogger and Novel Probes for Continuous Redox Measurements in Wetlands, *International Journal of Environmental Analytical Chemistry*, 91, 801–810, <https://doi.org/10.1080/03067319.2010.535123>,  
725 2011.

Wang, Y.: Frequencies of the Ricker Wavelet, *GEOPHYSICS*, 80, A31–A37, <https://doi.org/10.1190/geo2014-0441.1>, 2015.

Wang, Y., Wang, H., He, J.-S., and Feng, X.: Iron-Mediated Soil Carbon Response to Water-Table Decline in an Alpine Wetland, *Nature Communications*, 8, 15 972, <https://doi.org/10.1038/ncomms15972>, 2017.

Wen, Y., Zang, H., Ma, Q., Evans, C. D., Chadwick, D. R., and Jones, D. L.: Is the 'enzyme Latch' or 'Iron Gate' the Key to Protecting Soil  
730 Organic Carbon in Peatlands?, *Geoderma*, 349, 107–113, <https://doi.org/10.1016/j.geoderma.2019.04.023>, 2019.

Wojciech, S. L. and Styła, K.: Changes of Urease Activity in Peat Profile of Peatland By Nierybno Lake in "Bory Tucholskie" National Park, p. 4, 2011.

- Wriedt, G. and Rode, M.: Modelling Nitrate Transport and Turnover in a Lowland Catchment System, *Journal of Hydrology*, 328, 157–176, <https://doi.org/10.1016/j.jhydrol.2005.12.017>, 2006.
- 735 Xiang, W. and Freeman, C.: Annual Variation of Temperature Sensitivity of Soil Organic Carbon Decomposition in North Peatlands: Implications for Thermal Responses of Carbon Cycling to Global Warming, *Environmental Geology*, 58, 499–508, <https://doi.org/10.1007/s00254-008-1523-6>, 2009.
- Yu, K., Faulkner, S. P., and Baldwin, M. J.: Effect of Hydrological Conditions on Nitrous Oxide, Methane, and Carbon Dioxide Dynamics in a Bottomland Hardwood Forest and Its Implication for Soil Carbon Sequestration, *Global Change Biology*, 14, 798–812, <https://doi.org/10.1111/j.1365-2486.2008.01545.x>, 2008.
- 740 Yu, Z., Loisel, J., Brosseau, D. P., Beilman, D. W., and Hunt, S. J.: Global Peatland Dynamics since the Last Glacial Maximum, *Geophysical Research Letters*, 37, <https://doi.org/10.1029/2010GL043584>, 2010.
- Zak, D., Gelbrecht, J., and Steinberg, C. E. W.: Phosphorus Retention at the Redox Interface of Peatlands Adjacent to Surface Waters in Northeast Germany, *Biogeochemistry*, 70, 357–368, <https://doi.org/10.1007/s10533-003-0895-7>, 2004.
- 745 Zhang, Z. and Furman, A.: Soil Redox Dynamics under Dynamic Hydrologic Regimes - A Review, *Science of The Total Environment*, 763, 143 026, <https://doi.org/10.1016/j.scitotenv.2020.143026>, 2021.