1 Classification accuracy and compatibility across devices of a new

2 Rapid-E+ flow cytometer

Branko Sikoparija¹, Predrag Matavulj², Isidora Simovic¹, Predrag Radisic¹, Sanja Brdar¹, Vladan Minic¹,
 Danijela Tesendic³, Evgeny Kadantsev⁴, Julia Palamarchuk⁴ and Mikhail Sofiev⁴

¹BioSense Institute Research Institute for Information Technologies in Biosystems, University of Novi Sad, Novi Sad, 2100,
 Serbia

7 ²Institute for Data Science, University of Applied Sciences North Western Switzerland, Windish, 5210, Switzerland

8 ³Department of mathematics and informatics, Faculty of Sciences, University of Novi Sad, Novi Sad, 21000, Serbia

9 ⁴Finnish Meteorological Institute, Helsinki, Erik Palmenin Aukio 1, FI-00560, Finland

- 10 Correspondence to: Mikhail Sofiev (Mikhail.Sofiev@fmi.fi)
- 11 Matt Smith #1 (Citation: https://doi.org/10.5194/egusphere-2024-187-CC1)

The authors present a very interesting and robust study examining the classification accuracy and compatibility across devices of a new Rapid-E+ flow cytometer for examining airborne pollen. The paper is generally well written, although it could do with thorough editing with specific focus on the use of articles. I have listed some minor comments below that I hope will help.

16 **Reply:** The authors would like to thank Matt Smith for interest in the study and for his helpful comments, which we have used 17 to improve our manuscript. Below we answer the questions and indicate the changes we have made to the revised manuscript.

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My one comment about the methods relates to the use of the Hirst type trap (Lines 161 to 165). When calibrating such sensitive instruments as the Rapid-E and Rapid-E+, it is important to remove as much uncertainty as possible. The authors might therefore consider counting whole slides from the Hirst type trap to reduce error. Obviously, this is not always feasible when examining whole seasons, but even examining a small subset of slides in this way might provide some interesting insights. Although I note that correlations were only conducted for or days when average pollen concentrations measured by the manual method exceeded 10 pollen m⁻³ in order to reduce uncertainty.

25 Reply: The reviewer is correct that the standard method (EN16868) has large uncertainty that originates from different critical 26 points (i.e. flow measurements, pollen identification, subsampling during analysing collected samples). Recent study by Mimic 27 and Sikoparija (2021) confirmed that analysing 100% of samples coming from Hirst type traps is expected to improve comparison of time series obtained from different devices especially for low concentrations. However, as the reviewer correctly 28 29 pointed out, analysing an entire sample under microscope for the entire season is nor realistic, the effect is quite small, and all 30 measurement critical points exist in an automatic approach as well (Tummon et al. 2022), but still are not precisely quantified. 31 This is why we followed the recommendations of the EN16868 norm: to limit the influence of measurement uncertainty when 32 comparing results from different methods. We focused on daily values and only considered cases where a sufficient amount 33 of pollen was detected. To clearly address this aspect, we have added the following sentence to section 3.3:

34 "Limited improvement in correlations could be expected if the measurement uncertainty of the standard Hirst volumetric 35 method (EN16868), inherited from the subsampling during analysing collected samples, is eliminated by counting 100% of 36 microscopic slides (Mimic and Sikoparija 2021). However, such analysis for the entire season is extremely difficult and even 37 if done so, the effect is presumed to be small."

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39 Minor comments

40 Line 47 - "which is a new model stemming from the PA-300 (Crouzy et al., 2016) and Rapid-E (Sauliene et al., 2019)". 41 Reply: Corrected as suggested. 42 43 Line 49 – "In particular, Rapid-E+ samples at a flow rate of 51 min-1" 44 Reply: Corrected as suggested. 45 Line 53 – "Like its predecessor" 46 47 **Reply:** Corrected as suggested. 48 49 Line 73 – "was trained in the Novi Sad laboratory" 50 Reply: Corrected as suggested. 51 52 Line 74 - "owned by the City of Osijek in Croatia" 53 Reply: Corrected as suggested. 54 55 Line 74 - "and the Finnish Meteorological Institute" 56 **Reply:** Corrected as suggested. 57 58 Lines 79/80 – "for the Pannonian Plain" Lines 85/86 – "or capturing the main features" 59 Reply: Corrected as suggested. 60 Line 91 – "Scientific names should be italics" (review throughout including figures and tables). 61 **Reply:** The scientific names of the plant species from which pollen was used in the model training were written in italics 62 (Table A2). For classes of pollen identified from aerobiological samples (automatic and manual) we did not use the taxonomic 63 nomenclature because the pollen classes do not fully represent taxonomic categories. For example, in real time detections class 64 65 Artemisia is trained on pollen from Artemisia absintium L., Artemisia vulgaris L., thus it cannot be fully representative for 66 genus Artemisia. Similarly, in manual analysis the class Artemisia recorded in the given day could consist of pollen coming either from one or several species thus never being representative for the entire genus Artemisia. To address this, we have 67 added the following info in the Table A2: 68 69 "* does not fully represent taxonomic rank (i.e. pollen in reference data coming only from one or several species of the 70 respective taxonomic category) thus not written in italics" 71 Lines 98/99 - "To ensure identification" 72 73 Reply: Corrected as suggested 74 75 Line 102 - by using a Swisens Atomizer 76 Reply: Corrected as suggested 77 78 Line 193 – "It is interesting to note that after the start of rainfall the coarse particles" 79 **Reply:** Corrected as suggested. 80 81 Line 196 - The following lacks clarity and should be rewritten "However, quite low flow rate" 82 **Reply:** The sentence is rewritten and now reads: "However, following the equations given in Tummon et al. (2022), the flow rate of the Rapid-E+ (5 l min⁻¹) is not sufficient 83 84 to measure all relevant concentrations at sub hour temporal resolution with reasonably low uncertainty." 85 86 Line 208 – "despite these pollen grains being morphologically similar" (note that the plural of pollen is pollen) 87 Reply: Corrected as suggested. 88 89 Line 245 – "There was a clear tendency towards confusion of different pollen occurring"

- 90 **Reply:** Corrected as suggested.91
- 92 Table 1 It would be interesting to see the correlation coefficients for Taxaceae/Cupressaceae combined and for the Urticaceae
- family, as many pollen monitoring networks do not separate these into different genera due to the difficulty in identification.
- 94 **Reply:** We have calculated correlations for Taxaceae/Cupressaceae (sum of Taxus and Juniperus in Rapid-E+ data), Urticaceae
- 95 (sum of Urtica and Parietaria in Rapid-E+ data) and Cannabaceae (sum of Cannabis and Humulus in Rapid-E+ data), and
- 96 added coefficients into Table 1. Also, we added the following sentence to the results section:
- 97 "Merging Rapid-E+ measurements for classes that are difficult to identify by manual method (i.e. Taxus and Juniperus, Urtica
 98 and Parietaria, Cannabis and Humulus) did not improve the correlations (Table 1)."
- 99
- 100 Line 256 "Repeating it for each device in a network is unfeasible"
- 101 **Reply:** Corrected as suggested.
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Lines 261/262 – The following text lacks clarity and needs reworking, perhaps linked to another sentence "Demonstrated for
 Rapid-E, the problem also existed for Rapid-E+ (Fig. 4)".

- 105 **Reply:** The text is now rewritten and reads:
- "As a result, classification performance falls when a model trained on a reference dataset from one device is tested on a reference dataset from another one, which was demonstrated for Rapid-E (Matavulj et al. 2021). The same problem exists in Rapid-E+ (Fig. 4)."
- 109

110 Line 263 - pollen not pollens

- 111 **Reply:** Corrected as suggested.112
- 113 Line 274 pollen not pollens
- 114 **Reply:** Corrected as suggested.
- 116 Line 277 "Although this was not seen for all pollen types, there are pollen classes with comparable"
- 117 **Reply:** Corrected as suggested.
- 118119 Line 317 "datasets, the creation of which is a highly demanding process".
- 120 **Reply:** Corrected as suggested.
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122123 References

- Mimic, G., Sikoparija, B.: Analysis of airborne pollen time series originating from Hirst-type volumetric samplers—
 comparison between mobile sampling head oriented toward wind direction and fixed sampling head with two-layered
 inlet, Aerobiologia 37:321-331, https://doi.org/10.1007/s10453-021-09695-7, 2021.
- Tummon, F., Bruffaerts, N., Celenk, S., Choël, M., Clot, B., Crouzy, B., Galán, C., Gilge, S., Hajkova, L., Mokin, V.,
 O'Connor, D., Rodinkova, V., Sauliene, I., Sikoparija, B., Sofiev, M., Sozinova, O., Tesendic, D. and Vasilatou, K.:
 Towards standardisation of automatic pollen and fungal spore monitoring: best practises and guidelines, Aerobiologia.
 https://doi.org/10.1007/s10453-022-09755-6, 2022.