



- 1 Enigmatic Fe-Mn-fueled Anaerobic Oxidation of Methane in sulfidic coastal
- 2 sediments of the Eastern Arabian Sea
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# 11 Abstract

12 Anaerobic oxidation of methane (AOM) coupled with Fe-Mn reduction (Fe-Mn-AOM) is considered a globally important biogeochemical process in marine sediments in addition to 13 sulfate-driven AOM ( $SO_4^{2-}AOM$ ) responsible for the consumption of methane, a strong 14 greenhouse gas. Most existing studies have emphasized the significance of Fe-Mn-AOM 15 activities in sediments below the depth of the sulfate methane transition zone (SMTZ) with 16 insignificant dissolved sulfide and sulfate concentrations in the porewaters. Here, we report for 17 18 the first time enigmatic geochemical evidence of focused Fe-Mn-AOM activity across the SMTZ in the presence of high dissolved sulfide concentrations in a sediment core collected 19 20 within the seasonal coastal hypoxic zone of the Eastern Arabian Sea (West coast of India 21 (WCI)). The Fe-Mn-AOM activity is evident from the concurrent decrease in CH<sub>4</sub> concentrations,  $\delta^{13}C_{CH4}$  and  $\delta^{13}C_{DIC}$  values coupled with the enrichment of porewater Fe<sup>2+</sup> and 22





 $Mn^{2+}$  concentrations at multiple depths below the seafloor. Since neither CH<sub>4</sub> nor reactive Fe 23 appears to be the limiting factor controlling the Fe-Mn-AOM activity, we hypothesize that the 24 25 focused Fe-Mn-AOM at multiple depths is likely fueled by the localization of metal-reducing 26 and methanotrophic microbial communities, leading to biogeochemical heterogeneity in a dynamic seasonally hypoxic coastal environment sensitive to climate change. This study 27 28 highlights new insight into CH4-S-Fe-Mn biogeochemical cycling with far-reaching 29 implications in climate studies linked to the estimation of sedimentary methane production and 30 consumption.

# 31 **1 Introduction**

The metal-driven anaerobic oxidation of methane (AOM) is a vital biogeochemical process 32 and may have a significant impact on global biogeochemical cycles, particularly in relation to 33 coupled CH<sub>4</sub>-Fe-Mn-S cycling (Slomp et al., 2013; Riedinger et al., 2014; Leu et al., 2020). 34 Microbial Fe-Mn-driven AOM contributes to the global consumption of CH<sub>4</sub> in marine 35 sediments in addition to sulfate  $(SO_4^{2-})$ , nitrite  $(NO_2^{-})$ , and nitrate  $(NO_3^{-})$ -driven AOM 36 (Raghoebarsing et al., 2006; Beal et al., 2009; Knittel et al., 2009; Ettwig et al., 2010; Riedinger 37 et al., 2014; Xiao et al., 2023). A potential syntrophic coupling of AOM and reduction of Fe-38 Mn-(oxyhydr)oxides in marine sediments was first inferred from incubation experiments using 39 <sup>13</sup>C-labeled CH<sub>4</sub> (Beal et al., 2009). The coupling of AOM and Fe-Mn-reduction is potentially 40 attributed to the activities of methanotrophs (ANME-1, ANME-3) and metal-reducing bacteria 41 (Beal et al., 2009; Oni et al., 2015), while Fe-Mn-(oxyhydr)oxide reduction may also be carried 42 out by ANME-2a, ANME-2c, and ANME-2d using multiheme cytochromes without syntrophic 43 metal-reducing bacterial partners (Ettwig et al., 2016; Scheller et al., 2016; Cai et al., 2018; 44 Leu et al., 2020). Plausible electron transport pathways coupling AOM and metal oxide 45 reduction include (a) direct electron transfer via microbial contact; (b) indirect electron transfer 46 47 by electron shuttling; (c) indirect electron transfer by a metal chelate; and (d) direct electron





transfer by nanowires (Folgosa et al., 2015; Wegener et al., 2015; He et al., 2018). The 48 biogeochemical reactions during Fe-Mn-AOM (Eq. 1 and 2) result in the mutual consumption 49 of CH<sub>4</sub> and Fe-Mn-(oxyhydr)oxides coupled with the enrichment of Fe<sup>2+</sup> and Mn<sup>2+</sup> in the 50 interstitial waters (Beal et al., 2009; Riedinger et al., 2014). Although SO<sub>4</sub><sup>2-</sup>-AOM (Eq. 3) is 51 responsible for > 90% of global methane consumption, Fe-Mn-AOM is thermodynamically 52 more favorable than SO<sub>4</sub><sup>2-</sup>AOM (Beal et al., 2009) as evident from the  $\triangle$ G values (Eq. 1, 2 53 and 3). The global predominance of SO42-AOM may be attributed to its kinetics (Lovley and 54 Phillips, 1987). 55

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$$CH_4 + 8 Fe(OH)_3 + 15H^+ \rightarrow HCO_3^- + 8 Fe^{2+} + 21H_2O \ \triangle G = -270.3 \text{ kJ mol}^{-1} CH_4$$
 (1)

57 
$$CH_4 + 4 MnO_2 + 7H^+ \rightarrow HCO_3^- + 4 Mn^{2+} + 5H_2O$$
  $\Delta G = -556 \text{ kJ mol}^{-1} CH_4$  (2)

58 
$$CH_4 + SO_4^{2-} \rightarrow HS^- + HCO_3^- + H_2O$$
  $\triangle G = -10 \text{ to } -40 \text{ kJ mol}^{-1} \text{ CH}_4$  (3)

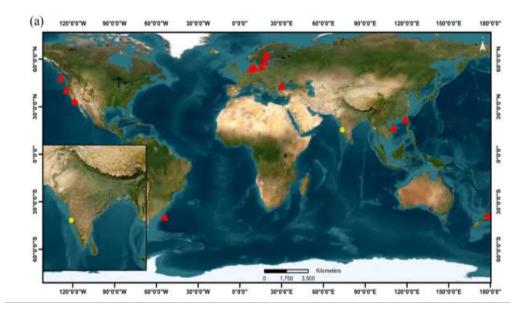
The availability of reactive Fe-Mn-(oxyhydr)oxides and low to undetectable SO42-59 concentrations are considered essential geochemical conditions to drive Fe-Mn-AOM (Beal et 60 61 al., 2009; Oni et al., 2015; Aromokeye et al., 2019), however, a recent report (Li et al., 2019) has shown the feasibility of Fe-Mn-AOM in SO42--rich porewaters. The kinetics of Fe-Mn-62 63 AOM depend on the content and composition of particulate Fe-Mn-(oxyhydr)oxides and 64 soluble ferric-organo complexes (e.g., ferric citrate) (Ettwig et al., 2016). It is observed that the soluble metal complexes and ferrihydrite have higher bioavailability and AOM activity than 65 other solid metal-(oxyhydr)oxides (Lovley and Phillips, 1987; Norði et al., 2013; Ettwig et al., 66 2016; Scheller et al., 2016). However, several incubation experiments and cultures have shown 67 that crystalline goethite, hematite, and magnetite can also serve as significant electron 68 acceptors for Fe-AOM (Bar-Or et al., 2017; Aromokeye et al., 2019; Li et al., 2021). 69

The global distribution of Fe-Mn-AOM is plotted in Figure 1a. Here, we present the first telltale evidence of Fe-Mn-AOM from a seasonally hypoxic shelf zone. The study was carried out



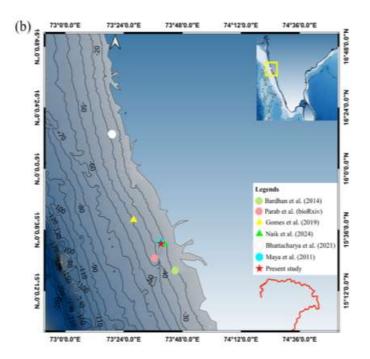


in a sediment core (SSD070/7/GC6) collected off the west coast of India (WCI) at a water 72 depth of 28.5 m (Figure 1b). The seasonal hypoxic zone off the WCI covers an area of 1,80,000 73 km<sup>2</sup> and is the largest of all coastal hypoxic systems (Naqvi et al., 2000). The hypoxia is 74 75 attributed to the upwelling of cold, nutrient-rich waters onto the shelf region during July to September, associated with the southwest monsoon. During the southwest monsoon, the WCI 76 receives substantial terrestrial as well as marine organic matter due to enhanced land runoff 77 78 and upwelling-driven marine productivity respectively. In contrast, during northeast monsoon, 79 the WCI experiences downwelling from November to April resulting in the prevalence of oxic 80 and oligotrophic conditions in the water column (Naqvi et al., 2000; Schott et al., 2001). Consequently, the region exhibits contrasting seasonal biogeochemical conditions including 81 hydrographic and depositional features (Naqvi et al., 2006; Mazumdar et al., 2012). In the 82 83 present study, we have investigated possible driving factors fueling Fe-Mn-AOM activity in a seasonally hypoxic shelf zone characterized by contrasting redox and depositional conditions. 84









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87 Figure 1. a) Global map showing various study locations (red-filled triangles) with reported Fe-Mn-AOM activities. The yellow-filled circle represents the present study location. The references are listed 88 89 in the supplementary material. (b) Map showing locations of the present study (SSD070/7/GC6) along 90 with those collected in previous studies (Maya et al., 2011; Bardhan et al., 2014; Gomes et al., 2019; 91 Bhattacharya et al., 2021; Naik et al., 2023; Parab et al., bioRxiv) from the inner shelf off the West Coast 92 of India (WCI). The red star represents the present study location (SSD070/7/GC6), while other colored symbols represent locations in the WCI where  $\delta^{13}C$  and  $\delta^{15}N$  of suspended particulate organic matter 93 94 and microbiological studies have been conducted.

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## 99 **2 Methodology**

### 100 2.1 Onboard Sampling

The studied gravity core (SSD070/7/GC6, 5.25 m long) was collected onboard ORV *Sindhu Sadhana* in February 2020. The coring operation was carried out off Goa (latitude: 15.50985°N,
longitude: 73.65313°E) from the seasonal hypoxic coastal zone of the Eastern Arabian Sea.
Immediately after recovery, the core was subsampled onboard for hydrocarbon gas analysis
and porewater extraction.

106 For headspace methane analysis, the sediment was extracted using 50 ml cut syringes at an 107 interval of 10 cm and transferred into 20 ml headspace vials filled with 3 ml of KOH and 3 ml of NaN<sub>3</sub> to trap CO<sub>2</sub> and arrest microbial activities respectively. The vials were flushed with 108 helium, homogenized, and stored at 4°C after sealing with butyl rubber septa. The core was 109 110 subsampled onboard for porewater extraction at 10 cm intervals and transferred into 50 ml tarsons centrifuge vials using a 50 ml cut syringe under a stream of argon gas to avoid oxidation 111 of dissolved sulfide. The samples were centrifuged at 7000 rpm for 20 minutes in a Remi C-30 112 113 centrifuge. The supernatant porewater was filtered through a 0.2 µm Whatman syringe filter. The filtered porewater was then stored in crimp vials under a helium head and subsequently 114 preserved at 4°C for measurement of various constituents. For dissolved sulfide ( $\Sigma HS^- = H_2S$ 115 116 +  $HS^-$  +  $S^{2-}$ ) analysis, 1 ml of 1 M CdNO<sub>3</sub> was added to fix all sulfides present as CdS. 117 Porewater samples for trace metal analysis were acidified with a known amount of concentrated supra-pure HNO<sub>3</sub>. All porewater samples were immediately crimp-sealed under a helium head 118 119 and stored at 4 °C until shore-based analysis. The core was subsampled for solid phase analysis at 1 cm resolution and stored at 4 °C for further analysis. 120

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## 123 2.2 Porewater and headspace gas analysis

The headspace hydrocarbon gas concentrations were determined using a trace GC equipped 124 125 with a flame ionization detector. Methane concentration measurements were carried out using an external calibration line prepared from six different gas mixtures (CH4, C2H6, C3H8, C4H10, 126 iso- $C_4H_{10}$  and  $CO_2$ ) standards with concentrations varying from 10-1000 ppm. The carbon 127 128 isotopic composition of methane was determined using an isotope ratio mass spectrometer 129 (Thermo-Delta V plus) coupled with a gas chromatograph (Thermo-Trace GC Ultra). The carbon isotope ratios of methane ( $\delta^{13}C_{CH4}$ ) are reported in delta notation as permil deviation 130 from Vienna Pee Dee Belemnite (VPDB) standard, and the external precision calculated for 131 the measurement is typically 0.07–0.09‰. The concentration of dissolved inorganic carbon 132 (DIC) was measured using a carbon coulometer (UIC-CM5130) with ultrapure CaCO<sub>3</sub> as 133 134 standard. The measurement yielded a sample reproducibility of  $\pm 0.3$  %. The carbon isotope ratios of DIC ( $\delta^{13}C_{DIC}$ ) were analyzed using a Thermo Delta V continuous flow isotope ratio 135 mass spectrometer coupled with a GASBENCH II equipped with a PAL auto-sampler, 136 following Torres et al. (2005) and Peketi et al. (2020). Reference materials NBS-18, NBS-19, 137 Carrara marble standards, and in-house laboratory standards (MERCK high purity CaCO<sub>3</sub> and 138 Carbonate reference standard) were measured multiple times during the analysis with each 139 batch. The carbon isotope composition of DIC is reported in delta notation as permil deviation 140 141 from the VPDB standard. The measurement yielded a sample reproducibility of +0.02 %. Porewater samples were diluted 40-fold for the determination of trace metals using high-142 resolution inductively coupled plasma-mass spectrometry (Nu-ATTOM ES). NASS-5 143 144 (Seawater-certified reference material) and CASS-6 (Near Shore Seawater reference material) 145 were analyzed to check the accuracy and reproducibility of the sample analysis. The analytical 146 precision was monitored by repeated measurements of sample/standard and the calculated RSD was less than 3%. 147





148	The porewater sulfate concentration was measured from the $\Sigma HS^-$ free supernatant solution
149	using a Metrohm ion chromatograph (Basic IC plus 883) equipped with a suppressed
150	conductivity detector (Metrohm, IC detector 1.850.9010). A mixed solution of 1 mM NaHCO <sub>3</sub>
151	and 3.2 mM $\mathrm{Na_2CO_3}$ was used as the eluent, and 0.2 N $\mathrm{H_2SO_4}$ was used as the suppressor
152	regeneration fluid. The samples were diluted 1000-fold with 18 M $\Omega$ ultrapure water prior to
153	analysis. The calibration line was prepared using a standard IC sulfate solution from a 100-
154	ppm mixed anion standard (Fluka), and the sample reproducibility was $\pm$ 0.1%. Quantification
155	of dissolved sulfide ( $\Sigma$ HS <sup>-</sup> ) was carried out following the methylene blue method (Cline et al.,
156	1969). Absorbance was measured at 670 nm on a spectrophotometer (Chemito Spectrascan
157	UV-2700). Sodium sulfide nonahydrate (≥99.99% purity, Merck) was used for the preparation
158	of calibration standards. The analytical error based on replicate standard measurements was $<$
159	3%.

#### 2.3 Solid phase analysis 160

161 Iron extractions were conducted under anaerobic conditions using frozen sediment samples following ascorbate and sodium dithionite steps. The leaching solutions were nitrogen-flushed 162 before extraction. The iron concentrations in ascorbate leached fraction (FeAsc) were 163 164 determined following previous studies (Hyacinthe et al., 2006; Raiswell et al., 2010) using 165 Atomic Absorption Spectrometer (Agilent-240AA) using air-acetylene flame. The sample reproducibility for replicate analysis was within 5 %. The FeAse free sediment residue was 166 leached for 2 hrs using sodium dithionite buffer for determination of dithionite extractable iron 167 (Fe<sub>D</sub>) content following previous works (Mehra and Jackson, 1960; Canfield et al., 1989; 168 Volvaikar et al., 2020). The Fe<sub>D</sub> content was determined using ferrozine-complexometry 169 technique on a Chemito spectrophotometer (Spectroscan UV 2700). The calibration line was 170 prepared from pure ferrous ammonium sulfate (Merck) and absorbance was measured at 515 171 nm. The replicate for Fe<sub>D</sub> analysis yielded sample reproducibility within 6 %. The total 172





173	inorganic carbon content was measured using a UIC carbon coulometer (CM 5130). Ultrapure
174	CaCO3 from Sigma-Aldrich was used as a standard reference material for measurement
175	(Carbon content: $12.0 \pm 0.25\%$ ). The measurement of total carbon (TC) content was carried out
176	on freeze-dried and desalinated samples using an EA1112 elemental analyzer (Thermo Fisher
177	Scientific, Germany). The total organic carbon (TOC) content (Figure S2; Table S2) was
178	calculated by subtracting TIC from TC. NC soil was used as a calibration standard for TC. The
179	reproducibility for TC in soil standards B2184 and B2152 was found to be 2.11 $\pm$ 0.1 % and
180	$1.53$ + 0.07 % respectively. The carbon and nitrogen isotope ratio of TOC ( $\delta^{13}C_{TOC})$ and
181	nitrogen ( $\delta^{15}N$ ) were measured on decarbonated samples using a Delta-V-plus isotope ratio
182	mass spectrometer coupled with an elemental analyzer (Thermo Flash EA 2000). The carbon
183	and nitrogen isotope ratios are reported as per mil (‰) deviations from the isotopic composition
184	of VPDB and atmospheric N <sub>2</sub> with reproducibility better than $\pm 0.3$ %.

## 185 **3 Results and Discussion**

# 186 **3.1 Evidence for SO4<sup>2-</sup> and Fe-Mn-AOM**

Based on the porewater SO<sub>4</sub><sup>2-</sup> and Fe-Mn concentration profiles (Figures 2a-2e; Table S1), the 187 sediment core can be divided into three distinctive diagenetic regimes (zone-i, zone-ii, and 188 zone-iii). Zone-i (63 to 303 cmbsf) is characterized by Fe<sup>2+</sup> and Mn<sup>2+</sup> enrichment (Fe<sup>2+</sup>: 10.75 189 to 361.3  $\mu$ M and Mn<sup>2+</sup>: 0.28 to 7.39  $\mu$ M) at high SO<sub>4</sub><sup>2-</sup> concentrations (0.5 to 18.2 mM); zone-190 ii (303 to 463 cmbsf) is characterized by  $SO_4^{2-}$  depletion (0.4 to 1.45  $\mu$ M) coupled with Fe<sup>2+</sup> 191 and Mn<sup>2+</sup> enrichment (Fe<sup>2+</sup>: 14.9 to 387.5 µM and Mn<sup>2+</sup>: 0.41 to 10.17 µM); and zone-iii (463-192 523 cmbsf) shows  $SO_4^{2-}$  depletion (below detection limit to 0.8  $\mu$ M) coupled with depleted  $Fe^{2+}$ 193 and  $Mn^{2+}$  concentrations (Fe<sup>2+</sup>: 11.85 to 58.2  $\mu$ M and  $Mn^{2+}$ : 1.13 to 2.68  $\mu$ M). The  $\Sigma$ HS<sup>-</sup> 194 concentrations remain consistently high (163 to 10,385 µM) below 63 cm from the seabed. 195

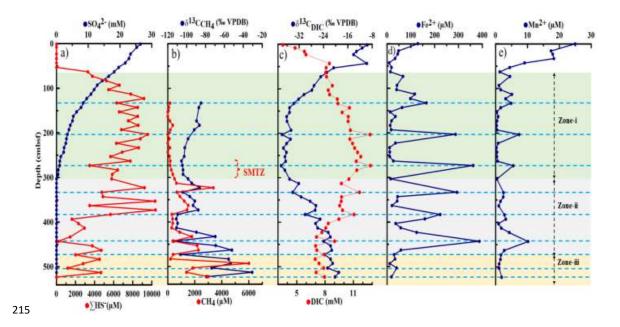




The SMTZ is identified within an approximate depth window of 263 to 303 cmbsf (Figures 2a 196 197 and 2b). Within the SMTZ, AOM activity is identified by simultaneous depletion in CH4 concentrations (188.9 to 503.2  $\mu$ M) and C-isotope ratios of CH<sub>4</sub> ( $\delta^{13}C_{CH4}$ : -105.6 to -98.6 ‰) 198 and DIC ( $\delta^{13}C_{DIC}$ : -38 to 35.2 %). The AOM-driven depletion in  $\delta^{13}C_{CH4}$  and  $\delta^{13}C_{DIC}$  values 199 within the SMTZ may be attributed to (i) DIC back flux during AOM (Yoshinaga et al., 2014) 200 (ii) concurrent methanotrophic and methanogenic activity (Borowski et al., 1997), (iii) the 201 202 ability of ANMEs to perform as facultative methanogens (Pohlman et al., 2008), and (iv) intracellular reaction reversibilities along enzymatic AOM pathway (Wegener et al., 2021). 203 Above SMTZ, the upward increasing trend in  $\delta^{13}C_{CH4}$  values may be attributed to  $^{13}C$ 204 enrichment (Figure 2b; Alperin et al., 1988; Whiticar et al., 1996; Martens et al., 1999; 205 Whiticar, 1999) in the residual methane (kinetic isotope effect) due to methane oxidation and 206 lack of carbon recycling at high SO4<sup>2-</sup> concentrations (SO4<sup>2</sup>>0.5 mM; Yoshinaga et al., 2014; 207 Wegener et al., 2021). Below the SMTZ, the methanogenesis trend is superimposed by depths 208 of intermittent AOM activity evident from depletion in CH<sub>4</sub> concentrations (1.04 to 4.19 mM) 209 coupled with a sharp decrease in  $\delta^{13}C_{CH4}$  (-17.95 to -61 ‰) and  $\delta^{13}C_{DIC}$  values (-1.8 to - 4.1 210 ‰) and increase in DIC concentrations (1 to 2.7 mM). The cross plot of DIC concentrations vs 211 212  $\delta^{13}C_{DIC}$  values for the overall core and the Fe-Mn-AOM specific points show negative correlations (Figure 3). The intermittent increase in DIC concentrations and decrease in  $\delta^{13}C_{DIC}$ 213 214 values can also be identified within the sulfatic zone-i.



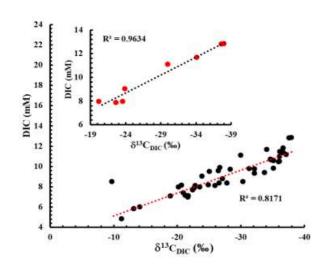




**Figure 2.** Figure showing Fe-Mn-AOM and  $SO_4^{2-}$ -AOM activities by means of pore fluid chemistry in seasonally hypoxic coastal sediment of the Eastern Arabian Sea. (a) Porewater  $SO_4^{2-}$ (mM) and  $\Sigma HS^-$  ( $\mu M$ ) concentrations. (b) Dissolved CH<sub>4</sub> concentrations ( $\mu M$ ) and  $\delta^{13}C_{CH4}$  values. (c) Dissolved DIC concentrations (mM) and  $\delta^{13}C_{DIC}$  values (d, e) Dissolved Fe and Mn ( $\mu M$ ) concentrations. The blue dashed lines represent the depth layers with AOM activities. The red curly bracket depicts SMTZ. The different color shades represent zone-ii, zone-ii, and zone-iii.







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Figure 3. Cross plot of DIC concentration (mM) vs  $\delta^{13}C_{DIC}$  (‰) values in the studied sediment core (represented by black dots with an R<sup>2</sup> value of 0.82). The inset cross plot of DIC vs.  $\delta^{13}C_{DIC}$  (represented by red dots with an R<sup>2</sup> value of 0.96) corresponds to values specific to Fe-Mn-AOM (marked by blue dashed lines in Figure 2).

227 The most intriguing observation of the present study is the intermittent increase in porewater Fe<sup>2+</sup> and Mn<sup>2+</sup> concentrations (Figures 2d & 2e). Within zone-i and ii, the marked increase in 228 porewater Fe<sup>2+</sup> and Mn<sup>2+</sup> concentrations are associated with AOM activities. However, in zone-229 iii, the AOM activities are not associated with significant porewater Fe<sup>2+</sup> and Mn<sup>2+</sup> enrichment. 230 The porewater SO<sub>4</sub><sup>2-</sup>, Fe<sup>2+</sup>, Mn<sup>2+</sup>, CH<sub>4</sub>, and DIC concentrations along with  $\delta^{13}C_{CH4}$  and  $\delta^{13}C_{DIC}$ 231 depth profiles provide tell-tale evidence of both SO42-AOM and Fe-Mn-AOM in the present 232 study. Zonation of the geochemical profiles shows co-occurrence of SO42-AOM and Fe-Mn-233 AOM in zone-i, and Fe-Mn-AOM in zone-ii and iii. The occurrence of Fe-Mn-AOM in SO42-234 rich zone has also been reported in an earlier study from the Dongsha area, South China Sea 235 (Li et al., 2019). These observations are further supported by an incubation study (Segarra et 236 al., 2013) where notable rates of Fe-Mn-AOM coincide with comparatively elevated levels of 237





238	$SO_4{}^2\mbox{-}AOM.$ Within zone-ii, where $SO_4{}^2\mbox{-}$ concentrations are reduced to 0.4 to 1.45 $\mu M,$ the
239	$Fe^{2\scriptscriptstyle +}$ and $Mn^{2\scriptscriptstyle +}$ concentration spikes may be attributed primarily to Fe-Mn-(oxyhydr)oxides
240	fueling AOM activity (Riedinger et al., 2014; Egger et al., 2015; Egger et al., 2017).

In most of the previous studies (Crowe et al., 2009; Sivan et al., 2011; Slomp et al., 2013; 241 Riedinger et al., 2014; Egger et al., 2015; Egger et al., 2016a; Egger et al., 2016b; Egger et al., 242 243 2017; Aromokeve et al., 2019; Vigderovich et al., 2019; Luo et al., 2020; Xiao et al., 2023), AOM-driven  $Fe^{2+}$ -Mn<sup>2+</sup> enrichment has been reported at low to negligible  $\Sigma HS^-$  concentrations 244 (Figure S1). However, the present study demonstrates tell-tale signatures of substantially high 245 porewater  $Fe^{2+}$ -Mn<sup>2+</sup> concentrations despite high  $\Sigma$ HS<sup>-</sup> concentrations. A similar observation 246 was also reported in previous studies (Ramírez-Pérez et al., 2015; Li et al., 2019). Equation-1 247 shows that for every mole of DIC produced via Fe-AOM, 8 moles of Fe<sup>2+</sup> are produced. The 248 DIC concentration perturbations (along the blue dashed lines) may be converted to equivalent 249  $\mu$ M of Fe<sup>2+</sup> produced via Fe<sup>3+</sup> reduction in the pore waters. The stoichiometrically calculated 250 amount of Fe<sup>2+</sup> produced ranges from 11673  $\mu$ M to 17163  $\mu$ M in zone-i-ii and 6192 to 8259 251  $\mu$ M in zone-iii. The measured porewater Fe<sup>2+</sup> concentrations are approximately 1.65-3% of the 252 calculated Fe<sup>2+</sup> in zone-i-ii and 0.2-0.6 % in zone-iii. Thus, the measured Fe<sup>2+</sup> concentration is 253 the residual Fe<sup>2+</sup> left after sulfidization (FeS/FeS<sub>2</sub>: Hensen et al., 2003; Treude et al., 2014; 254 Peketi et al., 2015). Therefore, the lack of significant Fe<sup>2+</sup> and Mn<sup>2+</sup> enrichment in zone-iii may 255 indicate complete consumption of Fe<sup>2+</sup> and Mn<sup>2+</sup> via sulfidization. 256

Additional processes that may enhance the metal concentrations in the sediment pore waters is the presence of metal (Fe-Mn) sulfide collides (Rickard et al., 2007) as nanoparticles. Dissolution of porewater Fe-Mn sulfide nanoparticles (Morse et al., 1999; Olson et al., 2017) in acid during sample preparation results in the simultaneous occurrence of  $Fe^{2+}$  and  $Mn^{2+}$ spikes in porewater. However, if the observed  $Fe^{2+}-Mn^{2+}$  concentration spikes in this study are





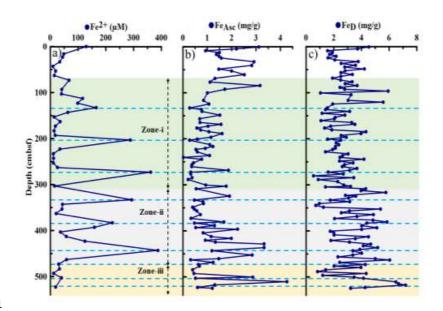
merely an analytical artifact, then under the presence of such high dissolved sulfide concentrations,  $Fe^{2+}$ -Mn<sup>2+</sup> spikes would be expected at all depths. The absence of Fe-Mn spikes corroborated with distinct increase in DIC concentration and decrease in  $\delta^{13}C_{DIC}$  values rules out such artifact generation.

## 266 **3.2** Possible limiting factors and focused Fe-Mn-AOM

An essential prerequisite for Fe-Mn-AOM is the presence of appreciable quantities of Fe-Mn-267 (oxyhydroxides) to react with CH<sub>4</sub> (Beal et al., 2009) as evident from the stoichiometric ratios 268 269 (Eq. 1). Results from iron extractions indicate the presence of significant amount of iron in 270 highly reactive/bioavailable (Fe<sub>Asc</sub>) (Figure 4b) and dithionite extractable phases (Fe<sub>D</sub>) (Figure 271 4c, Table S2). The FeAsc and FeD contents in the sediment range from 0.01 to 4.27 mg/g (Avg: 1.2 $\pm$ 0.86 mg/g) and 0.54 to 7.2 mg/g (Avg:3.03 $\pm$ 1.36 mg/g) respectively. The porewater Fe<sup>2+</sup> 272 concentration spikes are associated with marked depletion in FeAsc content, indicating 273 274 consumption of ferrihydrite and other bioavailable iron through Fe-AOM (Figure 4). However, the presence of substantial content of both FeD and FeAsc throughout the core length suggests 275 276 Fe-AOM is not limited by reactive iron content. In the present study, the high reactive iron flux 277 in the shelf zone may be attributed to rapid sedimentation rates (150 to 1500 cm/ky) and fluvial 278 input from Fe-Mn-rich provenances (Fernandes et al., 2020). Additionally, the presence of CH4 throughout the Fe-Mn-AOM zones suggests that neither CH<sub>4</sub> nor reactive Fe are limiting 279 280 factors responsible for the focused occurrences of Fe-AOM in the sediment core.







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Figure 4. Depth profiles of (a) Porewater Fe<sup>2+</sup> concentrations (µM). (b) solid phase ascorbic acid
leachable iron (Fe<sub>Asc</sub>) and (c) sodium dithionite leachable iron (Fe<sub>D</sub>) contents (in mg/g). The blue dashed
lines represent the depth layers with AOM activities. The different color shades represent zone-i,
zone-ii, and zone-iii.

286 One of the plausible factors responsible for the occurrence of focused Fe-Mn-AOM in distinct sediment layers could be the spatiotemporal enrichment of the microbial population driving 287 288 Fe-Mn-AOM. The latter in turn could be the result of the interplay between several biogeochemical factors. For instance, the shallow shelf off WCI is highly dynamic, 289 experiencing drastic changes in water column redox conditions, marine productivity, fluvial 290 291 fluxes of organic matter, sediment load, and extensive denitrification (Naqvi et al., 2000; Schott et al., 2001; Naqvi et al., 2006; Maya et al., 2011; Mazumdar et al., 2012; Fernandes et al., 292 2020). A large range in TOC contents (1.45 to 31.3 mg/g), (TOC/TN)<sub>molar</sub> ratios (3.47 to 27.32), 293  $\delta^{13}C_{TOC}$  (-20.69 to -25.93 ‰), and  $\delta^{15}N$  values (-3.6 to 7.7 ‰) in the present and previous 294 studies (Mazumdar et al., 2012; Fernandes et al., 2020) indicate marked temporal variation in 295 the fluvial and marine organic matter fluxes and denitrification conditions in WCI (Figure S2, 296 15





297	Table S2). Previous studies (near SSD070/7/GC6; Figure 1b) investigating carbon and nitrogen
298	stable isotopes of suspended particulate organic matter (SPOM) in the estuary (Bardhan et al.,
299	2014) and shelf zone (Maya et al., 2011) of WCI revealed significant intra-annual variations in
300	$\delta^{15}N$ (estuary: 0.69 to 7.26 ‰; shelf: -4.17 to 10.43 ‰) and $\delta^{13}C$ (estuary: -30.14 to -19.52 ‰,
301	shelf: -17.64 to -26.74 ‰) throughout the year. These variations reflect the complex and
302	dynamic nature of biogeochemical processes and organic matter sources in the coastal waters
303	of the WCI. Corroboratively, significant variations in the diversity, abundance, and activity of
304	microorganisms, attributable to seasonal differences in nutrient availability, have been recorded
305	in the water column between monsoon and non-monsoon seasons (Gomes et al., 2019; Naik et
306	al., 2024; Parab et al., bioRxiv). Spatiotemporally contrasting biogeochemical conditions of
307	shallow coastal waters have profound influence on the structure and function of underlying
308	sedimentary microbiomes (Bhattacharya et al., 2021). An earlier study (Orsi et al., 2017) from
309	the North Eastern Arabian Sea oxygen minimum zone reported stratigraphic microbial
310	distribution attributed to palaeoceanographic conditions. It was also proposed that focused
311	abundances of microbial communities in sediment layers are expected to be most pronounced
312	in the coastal region due to drastic variations in marine redox conditions and terrestrial fluxes.
313	In the context of the biogeochemical phenomenon revealed here, we hypothesize a dominant
314	role of the localized abundance of metal-reducing bacterial/archaeal communities in restricting
315	Fe-Mn-AOM activities into specific sedimentary strata. The focusing of microbial activity in
316	different sediment layers may be attributed to factors such as past environmental conditions
317	and depositional processes (Parkes et al., 2000; Orsi et al., 2017; Hoshino et al., 2020). Seasonal
318	coastal hypoxia coupled with a wide range of organic matter (marine to terrestrial) fluxes may
319	play important roles in stratigraphic microbial distributions and heterogeneity in
320	biogeochemical processes.





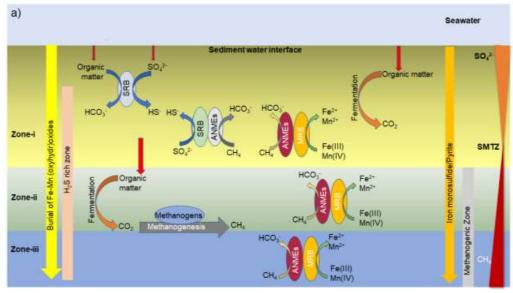
321	Our hypothesis is supported by metagenomic sequencing (Sarkar et al., in prep; Table S3)
322	carried out in a ~3 m long, sulfide-rich, sediment core (SSK42/9; water depth: 31 m; Figure
323	1b), collected from a nearby shallow coastal site (Bhattacharya et al., 2021) which shows the
324	presence of two species-level entities affiliated to the uncultivated archaeon
325	Candidatus Methanoperedens (GenBank assembly numbers GCA_003104905.1 and
326	GCA_001317315.1) reported thus far for potential abilities to harness anaerobic methane
327	oxidation to the reduction of Fe (III) (Ettwig et al., 2016; Cai et al., 2018). However, the final
328	microbiological corroboration of the focused Fe-Mn-AOM phenomenon hinges on the future
329	enrichment of the genome sequence database for microorganisms having definite and complete
330	Fe-Mn-AOM attributes.
331	Being the world's largest coastal hypoxic zone covering an area of 0.18 million sq. km, the net
332	influence of Fe-Mn-AOM could be significant in sedimentary methane consumption. A

 $\label{eq:schematic representation of CH_4-S-Fe-Mn cycling in coastal hypoxic sediments of the Eastern$ 

Arabian Sea is presented in Figures 5a, 5b and 5c respectively.









SRB: Sulfate reducing bacteria; MRB: Metal reducing bacteria; ANME: Anaerobic methanotrophic archaea

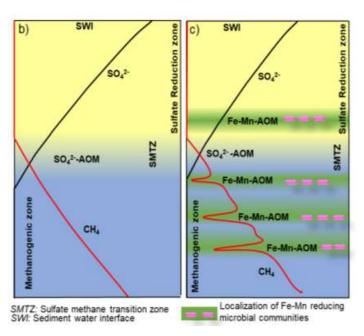




Figure 5. (a) Schematic representation of Fe-Mn-AOM and SO<sub>4</sub><sup>2-</sup>-AOM activities within the sediment
column. The figure depicts different zonation within the sediment column, showing the co-occurrence
of SO<sub>4</sub><sup>2-</sup>-AOM and Fe-Mn-AOM in zone-i and Fe-Mn-AOM in zone-ii-iii. Schematic representation of





pore fluid vertical profiles in the absence (b) and presence (c) of Fe-Mn-AOM activity. The greenshaded regions in the methanogenic zone represent potential Fe-Mn-AOM depth layers, controlled by
the localization of metal-reducing microbial communities.

## 343 4 Conclusions

The comprehensive geochemical investigation of the sediment core (5.25 m) retrieved from the 344 seasonally hypoxic shelf sediment off the eastern Arabian Sea (west coast of India) has 345 unveiled compelling evidence of focused Fe-Mn-AOM occurring at multiple depths beneath 346 the seafloor. The accumulation of porewater Fe<sup>2+</sup> and Mn<sup>2+</sup> despite elevated concentrations of 347  $\Sigma$ HS<sup>-</sup>, suggests the predominant production of Fe<sup>2+</sup> and Mn<sup>2+</sup> rather than consumption through 348 349 metal sulfidization processes. This study demonstrates that Fe-Mn-AOM remains a significant biogeochemical phenomenon even within  $SO_4^{2-}$  and  $\Sigma HS^{-}$ -rich sediments. Our findings are also 350 351 relevant in assessing the consumption of CH<sub>4</sub>, a greenhouse gas in organic-rich coastal sediments, thus having implications for climate change research. This study presents a new 352 perspective by documenting the biogeochemical heterogeneity in the occurrence of Fe-Mn-353 354 AOM, possibly attributed to deposition-controlled Fe-Mn reducing microbial population 355 distribution in a highly dynamic coastal environment sensitive to climate change. The findings of the present study may have a far-reaching influence on coupled CH4-Mn-Fe-S cycling in 356 357 expanding hypoxic coastal regions of the global ocean. Our findings lead to a hypothesis that necessitates future examination of microbial communities in seasonally hypoxic sediments and 358 SPOM at high depth and temporal resolution which may uncover the complex interactions 359 360 between microbial life and their environment.

361 Appendices: No appendix is there in this manuscript.

362 Code Availability: No code was used in this manuscript.

363 Data availability: All the data used in this study is incorporated in the supplementary text as
364 table S1, table S2 and table S3.





- **Sample availability:** Samples are available with the author.
- 366 Video supplement: No video component is there in this manuscript.
- 367 Author Contributions: K.S., A.P., and A.M. designed the research; A.M. obtained the funding;
- 368 K.S., A.Z., and S.P.K.P. analyzed the data; A.G., M.S., and J.M. carried out sampling; and K.S.,
- 369 A.P., and A.M. wrote the paper.
- 370 **Competing interests:** The authors declare that they have no conflict of interest.

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678	Supplementary Data. For figures (Figures S1 and S2), tables (Tables S1, S2 and S3), and
679	supporting text, please see supplementary data file.
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