

Supporting Information

S1 Web of Science search criteria

Keywords:

TS=((stem OR root OR branch OR sapwood OR bole OR trunk OR twig OR xylem)
NEAR nitrogen)

AND

TS=(abies OR acer OR alnus OR betula OR carpinus OR carya OR castanea OR cedrus OR chamaecyparis OR cornus OR cryptomeria OR cupressus OR fagus OR fraxinus OR juglans OR juniperus OR larix OR liriodendron OR lithocarpus OR magnolia OR notholithocarpus OR nyssa OR oxydendrum OR phellodendron OR picea OR pinus OR platanus OR populus OR prunus OR pseudotsuga OR quercus OR robinia OR salix OR sequoia OR sequioadendron OR sorbus OR taxodium OR thuja OR tilia OR tsuga OR ulmus)

Date of search:

05.04.2020

S2 Data sources

- Adriaenssens S. 2012. Dry deposition and canopy exchange for temperate tree species under high nitrogen deposition. PhD thesis, Ghent University Ghent, Belgium.
- Akburak S, Oral HV, Ozdemir E, Makineci E. 2013. Temporal variations of biomass, carbon and nitrogen of roots under different tree species. Scandinavian Journal of Forest Research 28(1): 8-16.
- Alban DH, Pastor J. 1993. Decomposition of aspen, spruce, and pine boles on two sites in Minnesota. Canadian Journal of Forest Research 23(9): 1744-1749.
- Albaugh TJ, Allen HL, Fox TR. 2008. Nutrient use and uptake in *Pinus taeda*. Tree Physiology 28(7): 1083-1098.
- Alriksson A, Eriksson HM. 1998. Variations in mineral nutrient and C distribution in the soil and vegetation compartments of five temperate tree species in NE Sweden. Forest Ecology and Management 108(3): 261-273.
- Andre F, Jonard M, Ponette Q. 2010. Biomass and nutrient content of sessile oak (*Quercus petraea* (Matt.) Liebl.) and beech (*Fagus sylvatica* L.) stem and branches in a mixed stand in southern Belgium. Sci Total Environ 408(11): 2285-2294.
- André F, Ponette Q. 2003. Comparison of biomass and nutrient content between oak (*Quercus petraea*) and hornbeam (*Carpinus betulus*) trees in a coppice-with-standards stand in Chimay (Belgium). Annals of Forest Science 60(6): 489-502.
- Aosaar J, Mander Ü, Varik M, Becker H, Morozov G, Maddison M, Uri V. 2016. Biomass production and nitrogen balance of naturally afforested silver birch (*Betula pendula* Roth.) stand in Estonia. Silva Fennica 50(4).
- Aosaar J, Varik M, Becker H, Morozov G, Aun K, Kukumägi M, Padari A, Uri V. 2019. Soil respiration and nitrogen leaching decreased in grey alder (*Alnus incana* (L.) Moench) coppice after clear-cut. Scandinavian Journal of Forest Research 34(6): 445-457.
- Aosaar J, Varik M, Löhmus K, Ostonen I, Becker H, Uri V. 2013. Long-term study of above- and below-ground biomass production in relation to nitrogen and carbon accumulation dynamics in a grey alder (*Alnus incana* (L.) Moench) plantation on former agricultural land. European Journal of Forest Research 132(5-6): 737-749.
- Armolaitis K, Aleinikoviene J, Baniuniene A, Lubyte J, Zekaite V. 2007. Carbon sequestration and nitrogen status in Arenosols following afforestation or following abandonment of arable land. Baltic Forestry 13(2): 169-178.
- Arthur MA, Fahey TJ. 1992. Biomass and nutrients in an Engelmann spruce–subalpine fir forest in north central Colorado: pools, annual production, and internal cycling. Canadian Journal of Forest Research 22(3): 315-325.
- Atkin OK, Bloomfield KJ, Reich PB, Tjoelker MG, Asner GP, Bonal D, Bonisch G, Bradford MG, Cernusak LA, Cosio EG, et al. 2015. Global variability in leaf respiration in relation to climate, plant functional types and leaf traits. New Phytol 206(2): 614-636.
- Bantle A, Borken W, Ellerbrock RH, Schulze ED, Weisser WW, Matzner E. 2014. Quantity and quality of dissolved organic carbon released from coarse woody debris of different tree species in the early phase of decomposition. Forest Ecology and Management 329: 287-294.
- Barbaroux C, Bréda N, Dufrêne E. 2003. Distribution of above-ground and below-ground carbohydrate reserves in adult trees of two contrasting broad-leaved species (*Quercus petraea* and *Fagus sylvatica*). New Phytologist 157(3): 605-615.

- Bauer GA, Persson H, Persson T, Mund M, Hein M, Kummetz E, Matteucci G, van Oene H, Scarascia-Mugnozza G, Schulze E-D 2000. Linking Plant Nutrition and Ecosystem Processes. In: Schulze E-D ed. Carbon and Nitrogen Cycling in European Forest Ecosystems. Berlin: Springer, 63-98.
- Bazilevich NI, Titlyanova AA. 2008. Biotic turnover of five continents: element exchange processes in terrestrial natural ecosystems. Novosibirsk, Russia: SB RAS.
- Belkacem S, Nys C, Gelhaye D. 1992. Effets d'une fertilisation et d'un amendement sur l'immobilisation d'éléments dans la biomasse d'un peuplement adulte d'épicéa commun (*Picea abies* L Karst). Annales des sciences forestières 49(3): 235-252.
- Berg B, Ekbohm G. 1991. Litter mass-loss rates and decomposition patterns in some needle and leaf litter types. Long-term decomposition in a Scots pine forest. VII. Canadian Journal of Botany 69(7): 1449-1456.
- Berg B, Ekbohm G, McClaugherty C. 1984. Lignin and holocellulose relations during long-term decomposition of some forest litters. Long-term decomposition in a Scots pine forest. IV. Canadian Journal of Botany 62(12): 2540-2550.
- Binkley D. 1982. Nitrogen fixation and net primary production in a young Sitka alder stand. Canadian Journal of Botany 60(3): 281-284.
- Binkley D. 1983. Ecosystem production in Douglas-fir plantations: Interaction of red alder and site fertility. Forest Ecology and Management 5(3): 215-227.
- Binkley D, Lousier JD, Cromack K, Jr. 1984. Ecosystem Effects of Sitka Alder in a Douglas-fir Plantation. Forest Science 30(1): 26-35.
- Birk EM, Vitousek PM. 1986. Nitrogen Availability and Nitrogen Use Efficiency in Loblolly Pine Stands. Ecology 67(1): 69-79.
- Bornkamm R, Bennert HW. 1989. Gehalt und Vorrat organischer Inhaltsstoffe in der Baumschicht von Luzulo-Fagetum-Beständen im Solling (BRD) 1) 1)Ergebnisse des „Solling-Projektes“ der Deutschen Forschungsgemeinschaft“ (Internationales Biologisches Programm): Mitteilung Nr. 340. Herrn Prof. Dr. H. Meusel zum 80. Geburtstag gewidmet. Flora 183(1-2): 133-148.
- Brais S, Paré D, Lierman C. 2006. Tree bole mineralization rates of four species of the Canadian eastern boreal forest: implications for nutrient dynamics following stand-replacing disturbances. Canadian Journal of Forest Research 36(9): 2331-2340.
- Bringmark L. 1977. Nutrient cycle in tree stands - Nordic symposium. Silva Fennica 11(3): 201-257.
- Brix H. 1981. Effects of nitrogen fertilizer source and application rates on foliar nitrogen concentration, photosynthesis, and growth of Douglas-fir. Canadian Journal of Forest Research 11(4): 775-780.
- Brozek S. 1990. Effect of soil changes caused by red alder (*Alnus rubra*) on biomass and nutrient status of Douglas-fir (*Pseudotsuga menziesii*) seedlings. Canadian Journal of Forest Research 20(9): 1320-1325.
- Buchmann N, Oren R, Gebauer G, Dietrich P, Schulze E-D. 1992. The use of stable isotopes in ecosystem research. First results of a field study with ^{15}N . Isotopenpraxis Isotopes in Environmental and Health Studies 28(1): 51-59.
- Buchmann N, Schulze E-D, Gebauer G. 1995. ^{15}N -ammonium and ^{15}N -nitrate uptake of a 15-year-old *Picea abies* plantation. Oecologia 102(3): 361-370.
- Calfapietra C, Angelis Pd, Gielen B, Lukac M, Moscatelli MC, Avino G, Lagomarsino A, Polle A, Ceulemans R, Mugnozza GS, et al. 2007. Increased nitrogen-use efficiency of a short-rotation poplar plantation in elevated CO₂ concentration. Tree Physiology 27(8): 1153-1163.

- Campetella G, Botta-Dukát Z, Wellstein C, Canullo R, Gatto S, Chelli S, Mucina L, Bartha S. 2011. Patterns of plant trait–environment relationships along a forest succession chronosequence. *Agriculture, Ecosystems & Environment* 145(1): 38-48.
- Cerasino L, La Porta N. 2014. Allocation of five macroelements and quality of fuels derived from Norway spruce wood obtained by thinning operations. *Biomass and Bioenergy* 70: 553-556.
- Ceschia É, Damesin C, Lebaube S, Pontailler J-Y, Dufrêne É. 2002. Spatial and seasonal variations in stem respiration of beech trees (*Fagus sylvatica*). *Annals of Forest Science* 59(8): 801-812.
- Chen J, Heikkinen J, Hobbie EA, Rinne-Garmston KT, Penttila R, Makipaa R. 2019. Strategies of carbon and nitrogen acquisition by saprotrophic and ectomycorrhizal fungi in Finnish boreal *Picea abies*-dominated forests. *Fungal Biol* 123(6): 456-464.
- Chen Y, Han W, Tang L, Tang Z, Fang J. 2011. Leaf nitrogen and phosphorus concentrations of woody plants differ in responses to climate, soil and plant growth form. *Ecography* 36(2): 178-184.
- Classen AT, Chapman SK, Whitham TG, Hart SC, Koch GW. 2007. Genetic-based plant resistance and susceptibility traits to herbivory influence needle and root litter nutrient dynamics. *Journal of Ecology* 95(6): 1181-1194.
- Cobb WR, Will RE, Daniels RF, Jacobson MA. 2008. Aboveground biomass and nitrogen in four short-rotation woody crop species growing with different water and nutrient availabilities. *Forest Ecology and Management* 255(12): 4032-4039.
- Cole DW, Rapp M, eds. 1980. Elemental cycling in forest ecosystems. Dynamic properties of forest ecosystems. International Biological Programme Synthesis, 23. Cambridge, UK: Cambridge University Press.
- Cong Y, Li M-H, Liu K, Dang Y-C, Han H-D, He HS. 2019. Decreased Temperature with Increasing Elevation Decreases the End-Season Leaf-to-Wood Reallocation of Resources in Deciduous *Betula ermanii* Cham. *Trees. Forests* 10(2).
- Cornelissen JHC, Cerabolini B, Castro-Díez P, Villar-Salvador P, Montserrat-Martí G, Puyravaud JP, Maestro M, Werger MJA, Aerts R. 2003. Functional traits of woody plants: correspondence of species rankings between field adults and laboratory-grown seedlings? *Journal of Vegetation Science* 14(3): 311-322.
- Cotrufo C. 1983. Xylem nitrogen as a possible diagnostic nitrogen test for loblolly pine. *Canadian Journal of Forest Research* 13(2): 355-357.
- Craine JM, Elmore AJ, Aidar MPM, Bustamante M, Dawson TE, Hobbie EA, Kahmen A, Mack MC, McLauchlan KK, Michelsen A, et al. 2009. Global patterns of foliar nitrogen isotopes and their relationships with climate, mycorrhizal fungi, foliar nutrient concentrations, and nitrogen availability. *New Phytol* 183(4): 980-992.
- Dahlin KM, Asner GP, Field CB. 2013. Environmental and community controls on plant canopy chemistry in a Mediterranean-type ecosystem. *Proc Natl Acad Sci U S A* 110(17): 6895-6900.
- Dawson JO, Funk DT. 1981. Seasonal Change in Foliar Nitrogen Concentration of *Alnus Glutinosa*. *Forest Science* 27(2): 239-243.
- Delagrange S, Messier C, Lechowicz MJ, Dizengremel P. 2004. Physiological, morphological and allocational plasticity in understory deciduous trees: importance of plant size and light availability. *Tree Physiology* 24(7): 775-784.
- Denaeyer - De Smet S 1971. Contents in biogenic elements of the plant covers in the deciduous forests of Europe. In: Duvigneaud P ed. Productivity of forest ecosystems. Proceedings of the Brussels symposium

organized by Unesco und the International Biological Programme (27-31 October 1969). Paris: Unesco, 515-525.

Devine WD, Harrington TB, Terry TA, Harrison RB, Slesak RA, Peter DH, Harrington CA, Shilling CJ, Schoenholz SH. 2011. Five-year vegetation control effects on aboveground biomass and nitrogen content and allocation in Douglas-fir plantations on three contrasting sites. *Forest Ecology and Management* 262(12): 2187-2198.

Dewit L, Reid DM. 1992. Branch Abscission in Balsam Poplar (*Populus balsamifera*): Characterization of the Phenomenon and the Influence of Wind. *International Journal of Plant Sciences* 153(4): 556-564.

Doucet A, Savard MM, Begin C, Smirnoff A. 2011. Is wood pre-treatment essential for tree-ring nitrogen concentration and isotope analysis? *Rapid Commun Mass Spectrom* 25(4): 469-475.

Doucet A, Savard MM, Bégin C, Smirnoff A. 2012. Tree-ring $\delta^{15}\text{N}$ values to infer air quality changes at regional scale. *Chemical Geology* 320-321: 9-16.

Du Z, Cai X, Bao W, Chen H, Pan H, Wang X, Zhao Q, Zhu W, Liu X, Jiang Y, et al. 2016. Short-Term vs. Long-Term Effects of Understory Removal on Nitrogen and Mobile Carbohydrates in Overstory Trees. *Forests* 7(12).

Duvigneaud P, Denaeyer - De Smet S 1973. Biological Cycling of Minerals in Temperate Deciduous Forests. In: Reichle DE ed. *Analysis of Temperate Forest Ecosystems*. Berlin, Heidelberg: Springer Berlin Heidelberg, 199-225.

Ebermann R, Stich K. 1985. Distribution and Seasonal Variation of Wood Peroxidase Activity in Oak (*Quercus Robur*) Wood and Fiber Science 17(3): 391-396.

Edmonds RL. 1987. Decomposition rates and nutrient dynamics in small-diameter woody litter in four forest ecosystems in Washington, U.S.A. *Canadian Journal of Forest Research* 17(6): 499-509.

Egnell G, Jurevics A, Peichl M. 2015. Negative effects of stem and stump harvest and deep soil cultivation on the soil carbon and nitrogen pools are mitigated by enhanced tree growth. *Forest Ecology and Management* 338: 57-67.

Ellsworth DS, Reich PB. 1992. Leaf Mass Per Area, Nitrogen Content and Photosynthetic Carbon Gain in *Acer saccharum* Seedlings in Contrasting Forest Light Environments. *Functional Ecology* 6(4): 423-435.

Eriksson HM, Berdén M, Rosén K, Nilsson SI. 1996. Nutrient distribution in a Norway spruce stand after long-term application of ammonium nitrate and superphosphate. *Water, Air, and Soil Pollution* 92(3): 451-467.

Eriksson HM, Rosen K. 1994. Nutrient distribution in a Swedish tree species experiment. *Plant and Soil* 164(1): 51-59.

Fahey TJ, Yavitt JB, Pearson JA, Knight DH. 1985. The nitrogen cycle in lodgepole pine forests, southeastern Wyoming. *Biogeochemistry* 1(3): 257-275.

Feger KH, Raspe S, Schmid M, Zöttl HW. 1991. Verteilung der Elementvorräte in einem schlechtwüchsigen 100jährigen Fichtenbestand auf Buntsandstein. *Forstwissenschaftliches Centralblatt vereinigt mit Tharandter forstliches Jahrbuch* 110: 248-262.

Feng Z, Brumme R, Xu YJ, Lamersdorf N. 2008. Tracing the fate of mineral N compounds under high ambient N deposition in a Norway spruce forest at Solling/Germany. *Forest Ecology and Management* 255(7): 2061-2073.

Finér L. 2008. Nutrient concentrations in *Pinus sylvestris* growing on an ombrotrophic pine bog, and the effects of PK and NPK fertilization. *Scandinavian Journal of Forest Research* 7(1-4): 205-218.

- Fogel R, Hunt G. 1983. Contribution of mycorrhizae and soil fungi to nutrient cycling in a Douglas-fir ecosystem. Canadian Journal of Forest Research 13(2): 219-232.
- Fornes RH, Berglund JV, Leaf AL. 1970. A comparison of the growth and nutrition of *Picea abies* (L.) karst. and *Pinus resinosa* ait. on a K-deficient site subjected to K fertilization. Plant and Soil 33(1): 345-360.
- Freschet GT, Cornelissen JHC, Van Logtestijn RSP, Aerts R. 2010. Evidence of the 'plant economics spectrum' in a subarctic flora. Journal of Ecology 98(2): 362-373.
- Gower ST, Richards JH. 1990. Larches: Deciduous Conifers in an Evergreen World. BioScience 40(11): 818-826.
- Grier CC, Cole DW, Dyrness CT, Fredriksen RL. 1974. Nutrient cycling in 37- and 450-year-old Douglas-fir ecosystems. In: Waring RH, Edmonds RL eds. Integrated research in the coniferous forest biome. Seattle, WA: Coniferous Forest Biome, 21-34.
- Haavik LJ, Ayres MP, Stange EE, Stephen FM. 2012. Phloem and xylem nitrogen variability in *Quercus rubra* attacked by *Enaphalodes rufulus*. The Canadian Entomologist 143(4): 380-383.
- Han W, Chen Y, Zhao FJ, Tang L, Jiang R, Zhang F. 2011. Floral, climatic and soil pH controls on leaf ash content in China's terrestrial plants. Global Ecology and Biogeography 21(3): 376-382.
- Heijari J, Nerg AM, Kainulainen P, Noldt U, Levula T, Raitio H, Holopainen JK. 2008. Effect of long-term forest fertilization on Scots pine xylem quality and wood borer performance. J Chem Ecol 34(1): 26-31.
- Heilman PE, Gessel SP. 1963. The Effect of Nitrogen Fertilization on the Concentration and Weight of Nitrogen, Phosphorus, and Potassium in Douglas-Fir Trees. Soil Science Society of America Journal 27(1): 102-105.
- Hellsten S, Helmisaari H-S, Melin Y, Skovsgaard JP, Kaakinen S, Kukkola M, Saarsalmi A, Petersson H, Akselsson C. 2013. Nutrient concentrations in stumps and coarse roots of Norway spruce, Scots pine and silver birch in Sweden, Finland and Denmark. Forest Ecology and Management 290: 40-48.
- Helmisaari H-S 1995. Nutrient cycling in *Pinus sylvestris* stands in eastern Finland. In: Nilsson LO, Hüttl RF, Johansson UT eds. Nutrient Uptake and Cycling in Forest Ecosystems. Proceedings of the CEC/IUFRO Symposium in Halmstad, Sweden, June, 7-10, 1993: Springer, 327-336.
- Hu B, Zhou M, Bilela S, Simon J, Dannenmann M, Liu X, Alfarraj S, Hou L, Chen H, Zhang S, et al. 2017. Nitrogen nutrition of native and introduced forest tree species in N-limited ecosystems of the Qinling Mountains, China. Trees 31(4): 1189-1202.
- Hunt HW, Ingham ER, Coleman DC, Elliott ET, Reid CPP. 1988. Nitrogen Limitation of Production and Decomposition in Prairie, Mountain Meadow, and Pine Forest. Ecology 69(4): 1009-1016.
- Huss-Danell K, Ohlsson H. 1992. Distribution of biomass and nitrogen among plant parts and soil nitrogen in a young *Alnus incana* stand. Canadian Journal of Botany 70(8): 1545-1549.
- Iivonen S, Kaakinen S, Jolkkonen A, Vapaavuori E, Linder S. 2006. Influence of long-term nutrient optimization on biomass, carbon, and nitrogen acquisition and allocation in Norway spruce. Canadian Journal of Forest Research 36(6): 1563-1571.
- Jach M. 2000. Above- and Below-ground Production of Young Scots Pine (*Pinus sylvestris* L.) Trees after Three Years of Growth in the Field under Elevated CO₂. Annals of Botany 85(6): 789-798.
- Jacobson S, Högbom L, Ring E, Nohrstedt H-Ö. 2016. The distribution of logging residues and its impact on seedling establishment and early plant growth in two Norway spruce stands. Scandinavian Journal of Forest Research 32(2): 134-141.

- Jacquet J-S, Bosc A, O'Grady AP, Jactel H. 2013. Pine growth response to processionary moth defoliation across a 40-year chronosequence. *Forest Ecology and Management* 293: 29-38.
- Jose S, Gillespie AR. 1996. Aboveground production efficiency and canopy nutrient contents of mixed-hardwood forest communities along a moisture gradient in the central United States. *Canadian Journal of Forest Research* 26(12): 2214-2223.
- Jyske T, Kaakinen S, Nilsson U, Saranpää P, Vapaavuori E. 2010. Effects of timing and intensity of thinning on wood structure and chemistry in Norway spruce. *Holzforschung* 64(1).
- Kaakinen S, Piispanen R, Lehto S, Metsometsä J, Nilsson U, Saranpää P, Linder S, Vapaavuori E. 2009. Growth, wood chemistry, and fibre length of Norway spruce in a long-term nutrient optimization experiment. *Canadian Journal of Forest Research* 39(2): 410-419.
- Kaakinen S, Saranpää P, Vapaavuori E. 2006. Effects of growth differences due to geographic location and N-fertilisation on wood chemistry of Norway spruce. *Trees* 21(2): 131-139.
- Kayama M, Makoto K, Nomura M, Satoh F, Koike T. 2009. Nutrient dynamics and carbon partitioning in larch seedlings (*Larix kaempferi*) regenerated on serpentine soil in northern Japan. *Landscape and Ecological Engineering* 5(2): 125-135.
- Kim C. 2019. Carbon and Nitrogen Distribution of Tree Components in *Larix kaempferi* Carriere and *Quercus variabilis* Blume Stands in Gyeongnam Province. *Journal of Korean Society of Forest Science* 108(2): 139-146.
- Kim C, Yoo BO, Jung SY, Lee KS. 2017. Allometric equations to assess biomass, carbon and nitrogen content of black pine and red pine trees in southern Korea. *iForest - Biogeosciences and Forestry* 10(2): 483-490.
- King JS, Giardina CP, Pregitzer KS, Friend AL. 2007. Biomass partitioning in red pine (*Pinus resinosa*) along a chronosequence in the Upper Peninsula of Michigan. *Canadian Journal of Forest Research* 37(1): 93-102.
- Kloepfel BD, Gower ST, Treichel IW, Kharuk S. 1998. Foliar Carbon Isotope Discrimination in *Larix* Species and Sympatric Evergreen Conifers: A Global Comparison. *Oecologia* 114(2): 153-159.
- Kostecki J, Kostecki J, Drab M, Szafraniec M, Stodulski G, Wypych M, Greinert A, Wasylewicz R. 2014. The total content of nitrogen in leaves and wood of trees growing in the area affected by the Głogów Copper Smelter. *Journal of Elementology*(1/2015).
- Kostiainen K, Kaakinen S, Saranpää P, Sigurdsson BD, Linder S, Vapaavuori E. 2004. Effect of elevated [CO₂] on stem wood properties of mature Norway spruce grown at different soil nutrient availability. *Global Change Biology* 10(9): 1526-1538.
- Krutul D, Zielenkiewicz T, Zawadzki J, Radomski A, Antczak A, Drożdżek M, Kłosińska T, Makowski T. 2015. Non-metals accumulation in Scots pine (*Pinus sylvestris* L.) wood and bark affected with environmental pollution. *Wood Research* 60(4): 655–662.
- Kull O, Koppel A, Noormets A. 1998. Seasonal changes in leaf nitrogen pools in two *Salix* species. *Tree Physiology* 18(1): 45-51.
- Kuznetsova T, Lukjanova A, Mandre M, Lõhmus K. 2011. Aboveground biomass and nutrient accumulation dynamics in young black alder, silver birch and Scots pine plantations on reclaimed oil shale mining areas in Estonia. *Forest Ecology and Management* 262(2): 56-64.
- Lahr EC, Krokene P. 2013. Conifer stored resources and resistance to a fungus associated with the spruce bark beetle *Ips typographus*. *PLoS One* 8(8): e72405.

- Lahr EC, Sala A. 2014. Species, elevation, and diameter affect whitebark pine and lodgepole pine stored resources in the sapwood and phloem: implications for bark beetle outbreaks. Canadian Journal of Forest Research 44(11): 1312-1319.
- Lang GE, Reiners WA, Shellito GA. 1982. Tissue chemistry of *Abies balsamea* and *Betula papyrifera* var. *cordifolia* from subalpine forests of northeastern United States. Canadian Journal of Forest Research 12(2): 311-318.
- Larocque G, R. 1999. Performance and morphological response of the hybrid poplar DN-74 (*Populus deltoides* x *nigra*) under different spacings on a 4-year rotation. Ann. For. Sci. 56(4): 275-287.
- Larocque GR. 2000. Performance of young jack pine trees originating from two different branch angle traits under different intensities of competition. Ann. For. Sci. 57(7): 635-649.
- Laughlin DC, Fulé PZ, Huffman DW, Crouse J, Laliberté E. 2011. Climatic constraints on trait-based forest assembly. Journal of Ecology 99(6): 1489-1499.
- Laughlin DC, Leppert JJ, Moore MM, Sieg CH. 2010. A multi-trait test of the leaf-height-seed plant strategy scheme with 133 species from a pine forest flora. Functional Ecology 24(3): 493-501.
- Lee H-S, Park Y-W, Lee H-Y, Choi S-G, Koo C-D. 2018. Changes of nutrient contents in the log of *Quercus acutissima* by cutting period for *Lentinula edodes* log cultivation. Forest Science and Technology 14(1): 33-40.
- Lee WY, Park E-J, Han SU. 2010. Correlation of Growth Performance with Total Nitrogen, Carbon and Nitrogen Isotope Compositions in the Xylem of *Pinus koraiensis*. Jour. Korean For. Soc. 99(3): 353-358.
- Li H, Crabbe MJC, Xu F, Wang W, Ma L, Niu R, Gao X, Li X, Zhang P, Ma X, et al. 2017. Seasonal variations in carbon, nitrogen and phosphorus concentrations and C:N:P stoichiometry in different organs of a *Larix principis-rupprechtii* Mayr. plantation in the Qinling Mountains, China. PLoS One 12(9): e0185163.
- Likens GE, Bormann FH. 1970. Chemical Analyses of Plant Tissues from the Hubbard Brook Ecosystem in New Hampshire. Yale School of Forestry & Environmental Studies Bulletin Series.
- Little SN, Shainsky LJ 1992. Distribution of biomass and nutrients in Lodgepole pine/bitterbrush ecosystems in central Oregon. Res. Pap. PNW-RP-454. Portland, OR: U.S. Department of Agriculture, Forest Service, Pacific Northwest Research Station. 22 p.
- Liu S 1995. Nitrogen cycling and dynamic analysis of man made larch forest ecosystem. In: Nilsson LO, Hüttl RF, Johansson UT eds. Nutrient Uptake and Cycling in Forest Ecosystems. Proceedings of the CEC/IUFRO Symposium in Halmstad, Sweden, June, 7-10, 1993: Springer, 391-397.
- Liu S, Wang H. 2017. N, P, and K characteristics of different age groups of temperate coniferous tree species in northwestern China. Journal of Forestry Research 29(2): 471-478.
- Loewenstein H, Pitkin FH. 1971. Growth responses and nutrient relations of fertilized and unfertilized grand fir. Sta. Pap. 9. Moscow, ID: University of Idaho. Forest, Wildlife and Range Experiment Station.
- Luken JO, Fonda RW. 1983. Nitrogen accumulation in a chronosequence of red alder communities along the Hoh River, Olympic National Park, Washington. Canadian Journal of Forest Research 13(6): 1228-1237.
- Lukeš P, Stenberg P, Rautiainen M, Mõttus M, Vanhatalo KM. 2013. Optical properties of leaves and needles for boreal tree species in Europe. Remote Sensing Letters 4(7): 667-676.
- Maier CA. 2001. Stem growth and respiration in loblolly pine plantations differing in soil resource availability. Tree Physiology 21(16): 1183-1193.

- Maire V, Wright IJ, Prentice IC, Batjes NH, Bhaskar R, van Bodegom PM, Cornwell WK, Ellsworth D, Niinemets Ü, Ordonez A, et al. 2015. Global effects of soil and climate on leaf photosynthetic traits and rates. *Global Ecology and Biogeography* 24(6): 706-717.
- Major JE, Johnsen KH, Barsi DC, Campbell M, Malcolm JW. 2013. Stem biomass, C and N partitioning and growth efficiency of mature pedigreed black spruce on both a wet and a dry site. *Forest Ecology and Management* 310: 495-507.
- Mälkönen E. 1974. Annual primary production and nutrient cycle in some Scots pine stands. *Communicationes Instituti Forestalis Fenniae* 84(5): 1-87.
- Mälkönen E. 1977. Annual primary production and nutrient cycle in a birch stand. *Communicationes Instituti Forestalis Fenniae* 91(5): 1-35.
- Mandre M, Klöško J, Lukjanova A, Tullus H. 2011. Variation of carbohydrates and lignin in hybrid aspen (*Populus tremula* x *P. tremuloides*) on alkaline soil. *Cellulose Chemistry and Technology* 45(5-6): 299-311.
- Mandre M, Korsjukov R, Ots K. 2004. Effect of wood ash application on the biomass distribution and physiological state of Norway spruce seedlings on sandy soils. *Plant and Soil* 265(1/2): 301-314.
- Martin AR, Gezahegn S, Thomas SC. 2015. Variation in carbon and nitrogen concentration among major woody tissue types in temperate trees. *Canadian Journal of Forest Research* 45(6): 744-757.
- Martin JG, Bolstad PV. 2005. Annual soil respiration in broadleaf forests of northern Wisconsin: influence of moisture and site biological, chemical, and physical characteristics. *Biogeochemistry* 73(1): 149-182.
- Martin JG, Kloepel BD, Schaefer TL, Kimbler DL, McNulty SG. 1998. Aboveground biomass and nitrogen allocation of ten deciduous southern Appalachian tree species. *Canadian Journal of Forest Research* 28(11): 1648-1659.
- Matyssek R, Schulze E-D. 1987. Heterosis in hybrid larch (*Larix decidua* x *leptolepis*). I. The role of leaf characteristics. *Trees* 1(4): 219-224.
- Mei L, Xiong Y, Gu J, Wang Z, Guo D. 2015. Whole-tree dynamics of non-structural carbohydrate and nitrogen pools across different seasons and in response to girdling in two temperate trees. *Oecologia* 177(2): 333-344.
- Meier CE, Grier CC, Cole DW. 1985. Below- and Aboveground N and P Use by *Abies amabilis* Stands. *Ecology* 66(6): 1928-1942.
- Meir P, Kruijt B, Broadmeadow M, Barbosa E, Kull O, Carswell F, Nobre A, Jarvis PG. 2002. Acclimation of photosynthetic capacity to irradiance in tree canopies in relation to leaf nitrogen concentration and leaf mass per unit area. *Plant, Cell & Environment* 25(3): 343-357.
- Merilä P, Mustajärvi K, Helmisaari H-S, Hilli S, Lindroos A-J, Nieminen TM, Nöjd P, Rautio P, Salemaa M, Ukonmaanaho L. 2014. Above- and below-ground N stocks in coniferous boreal forests in Finland: Implications for sustainability of more intensive biomass utilization. *Forest Ecology and Management* 311: 17-28.
- Merrill W, Cowling EB. 1966. Role of nitrogen in wood deterioration: amounts and distribution of nitrogen in tree stems. *Canadian Journal of Botany* 44(11): 1555-1580.
- Mfarrej MFB, Sharaf NS. 2011. Host Selection of Peach Rootborer *Capnodis tenebrionis* L. (Coleoptera: Buprestidae) to Stone-Fruit Trees in Jordan. *Jordan Journal of Agricultural Sciences* 7(4): 682-689.
- Milla R, Reich PB. 2011. Multi-trait interactions, not phylogeny, fine-tune leaf size reduction with increasing altitude. *Ann Bot* 107(3): 455-465.

- Mitchell AK, Barclay HJ, Brix H, Pollard DFW, Benton R, deJong R. 1996. Biomass and nutrient element dynamics in Douglas-fir: effects of thinning and nitrogen fertilization over 18 years. Canadian Journal of Forest Research 26(3): 376-388.
- Mori AS, Shiono T, Haraguchi TF, Ota AT, Koide D, Ohgue T, Kitagawa R, Maeshiro R, Aung TT, Nakamori T, et al. 2015. Functional redundancy of multiple forest taxa along an elevational gradient: predicting the consequences of non-random species loss. Journal of Biogeography 42(8): 1383-1396.
- Morrison IK. 1974. Dry-matter and Element Content of Roots of Several Natural Stands of *Pinus banksiana* Lamb. in Northern Ontario. Canadian Journal of Forest Research 4(1): 61-64.
- Mussche S, Bussche B, De Schrijver A, Neirynck J, Nachtergale L, Lust N. 1998. Nutrient uptake of a mixed oak/beech forest in Flanders (Belgium). *Silva Gandavensis* 63: 120-133.
- Naidu SL, DeLucia EH, Thomas RB. 1998. Contrasting patterns of biomass allocation in dominant and suppressed loblolly pine. Canadian Journal of Forest Research 28(8): 1116-1124.
- Ne'eman G, Goubitz S, Werger MJ, Shmida A. 2011. Relationships between tree size, crown shape, gender segregation and sex allocation in *Pinus halepensis*, a Mediterranean pine tree. Ann Bot 108(1): 197-206.
- Nihlgård B. 1972. Plant Biomass, Primary Production and Distribution of Chemical Elements in a Beech and a Planted Spruce Forest in South Sweden. Oikos 23(1): 69-81.
- Nihlgård B, Lindgren L. 1977. Plant Biomass, Primary Production and Bioelements of Three Mature Beech Forests in South Sweden. Oikos 28(1): 95-104.
- Niinemets Ü. 2001. Global-Scale Climatic Controls of Leaf Dry Mass per Area, Density, and Thickness in Trees and Shrubs. Ecology 82(2): 453-469.
- Noh NJ, Son Y, Kim RH, Seo KW, Koo JW, Park IH, Lee YJ, Lee KH, Son YM. 2007. Biomass accumulations and the distribution of nitrogen and phosphorus within three *Quercus acutissima* stands in central Korea. Journal of Plant Biology 50(4): 461-466.
- Nordborg F, Nilsson U, Gemmel P, Örlander G. 2007. Carbon and nitrogen stocks in soil, trees and field vegetation in conifer plantations 10 years after deep soil cultivation and patch scarification. Scandinavian Journal of Forest Research 21(5): 356-363.
- Nykqvist N. 1971. The Effect of Clear Felling on the Distribution of Biomass and Nutrients. Bulletins from the Ecological Research Committee 14: 166-178.
- Nys C, Ranger D, Ranger J. 1983. Etude comparative de deux écosystèmes forestiers feuillus et résineux des Ardennes primaires françaises. III. - Minéralomasse et cycle biologique. Annales des sciences forestières 40(1): 41-66.
- Ordonez JC, van Bodegom PM, Witte JP, Bartholomeus RP, van Hal JR, Aerts R. 2010. Plant strategies in relation to resource supply in mesic to wet environments: does theory mirror nature? Am Nat 175(2): 225-239.
- Oren R, Werk KS, Schulze ED, Meyer J, Schneider BU, Schramel P. 1988. Performance of Two *Picea abies* (L.) Karst. Stands at Different Stages of Decline. VI. Nutrient Concentration. Oecologia 77(2): 151-162.
- Ovington JD, Madgwick HAI. 1959. Distribution of Organic Matter and Plant Nutrients in a Plantation of Scots Pine. Forest Science 5(4): 344-355.
- Park BB, Yanai RD, Sahm JM, Lee DK, Abrahamson LP. 2005. Wood ash effects on plant and soil in a willow bioenergy plantation. Biomass and Bioenergy 28(4): 355-365.
- Perala DA, Alban DH. 1982. Biomass, nutrient distribution and litterfall in *Populus*, *Pinus* and *Picea* stands on two different soils in Minnesota. Plant and Soil 64(2): 177-192.

- Phillips T, Watmough SA. 2012. A nutrient budget for a selection harvest: implications for long-term sustainability. *Canadian Journal of Forest Research* 42(12): 2064-2077.
- Pierce S, Brusa G, Vagge I, Cerabolini BEL, Thompson K. 2013. Allocating CSR plant functional types: the use of leaf economics and size traits to classify woody and herbaceous vascular plants. *Functional Ecology* 27(4): 1002-1010.
- Ponette Q, Ranger J, Ottorini J-M, Ulrich E. 2001. Aboveground biomass and nutrient content of five Douglas-fir stands in France. *Forest Ecology and Management* 142(1): 109-127.
- Portsmouth A, Niinemets Ü, Truu L, Pensa M. 2005. Biomass allocation and growth rates in *Pinus sylvestris* are interactively modified by nitrogen and phosphorus availabilities and by tree size and age. *Canadian Journal of Forest Research* 35(10): 2346-2359.
- Pozdnyakov LK, Protopopov VV, Gorbatenko VM. 1969. Biological productivity of forests in Middle Siberia and Yakutia. Krasnoyarsk, Russia.
- Prentice IC, Meng T, Wang H, Harrison SP, Ni J, Wang G. 2011. Evidence of a universal scaling relationship for leaf CO₂ drawdown along an aridity gradient. *New Phytol* 190(1): 169-180.
- Preston KA, Cornwell WK, Denoyer JL. 2006. Wood density and vessel traits as distinct correlates of ecological strategy in 51 California coast range angiosperms. *New Phytol* 170(4): 807-818.
- Prokushkin A, Hagedorn F, Pokrovsky O, Viers J, Kirdyanov A, Masyagina O, Prokushkina M, McDowell W. 2018. Permafrost Regime Affects the Nutritional Status and Productivity of Larches in Central Siberia. *Forests* 9(6).
- Prokushkin AS unpublished. Nitrogen concentration of *Larix gmelinii* near Tura, Russia.
- Pyttel PL, Köhn M, Bauhus J. 2015. Effects of different harvesting intensities on the macro nutrient pools in aged oak coppice forests. *Forest Ecology and Management* 349: 94-105.
- Quested HM, Cornelissen JHC, Press MC, Callaghan TV, Aerts R, Trosien F, Riemann P, Gwynn-Jones D, Kondratchuk A, Jonasson SE. 2003. Decomposition of Sub-Arctic Plants with Differing Nitrogen Economies: A Functional Role for Hemiparasites. *Ecology* 84(12): 3209-3221.
- Ranger J, Cuirin G, Bouchon J, Colin M, Gelhaye D, Ahamed DM. 1992. Biomasse et minéralomasse d'une plantation d'épicéa commun (*Picea abies* Karst) de forte production dans les Vosges (France). *Annales des sciences forestières* 49(6): 651-668.
- Ranger J, Gelhaye D. 2001. Belowground biomass and nutrient content in a 47-year-old Douglas-fir plantation. *Annales des sciences forestières* 58(4): 423-430.
- Ranger J, Marques R, Colin-Belgrand M, Flammang N, Gelhaye D. 1995. The dynamics of biomass and nutrient accumulation in a Douglas-fir (*Pseudotsuga menziesii* Franco) stand studied using a chronosequence approach. *Forest Ecology and Management* 72(2): 167-183.
- Redmon LA, Rouquette FM, Smith GR, Florence MJ, Stuth JW. 1997. Interseeded legumes with loblolly pine. I. Effect of phosphorus and legume variety on pine seedling establishment and mortality. *Journal of Plant Nutrition* 20(12): 1755-1764.
- Reich PB, Kloepel BD, Ellsworth DS, Walters MB. 1995. Different Photosynthesis-Nitrogen Relations in Deciduous Hardwood and Evergreen Coniferous Tree Species. *Oecologia* 104(1): 24-30.
- Reich PB, Oleksyn J, Wright IJ. 2009. Leaf phosphorus influences the photosynthesis-nitrogen relation: a cross-biome analysis of 314 species. *Oecologia* 160(2): 207-212.

- Reich PB, Tjoelker MG, Pregitzer KS, Wright JJ, Oleksyn J, Machado JL. 2008. Scaling of respiration to nitrogen in leaves, stems and roots of higher land plants. *Ecol Lett* 11(8): 793-801.
- Rodriguez-Calcerrada J, Lopez R, Salomon R, Gordaliza GG, Valbuena-Carabana M, Oleksyn J, Gil L. 2015. Stem CO₂ efflux in six co-occurring tree species: underlying factors and ecological implications. *Plant Cell Environ* 38(6): 1104-1115.
- Røsberg I, Frank J, Stuanes AO. 2006. Effects of liming and fertilization on tree growth and nutrient cycling in a Scots pine ecosystem in Norway. *Forest Ecology and Management* 237(1-3): 191-207.
- Rose AK, Nicholas NS. 2008. Coarse Woody Debris in a Southern Appalachian Spruce-fir Forest of the Great Smoky Mountains National Park. *Natural Areas Journal* 28(4): 342-355.
- Royer-Tardif S, Delagrange S, Nolet P, Rivest D. 2017. Using Macronutrient Distributions within Trees to Define a Branch Diameter Threshold for Biomass Harvest in Sugar Maple-Dominated Stands. *Forests* 8(2).
- Sala A, Hopping K, McIntire EJB, Delzon S, Crone EE. 2012. Masting in whitebark pine (*Pinus albicaulis*) depletes stored nutrients. *New Phytol* 196(1): 189-199.
- Santa Regina I. 2000. Organic matter distribution and nutrient fluxes within a sweet chestnut (*Castanea sativa* Mill.) stand of the Sierra de Gata, Spain. *Ann. For. Sci.* 57(7): 691-700.
- Santa Regina I, Tarazona T, Calvo R. 1997. Aboveground biomass in a beech forest and a Scots pine plantation in the Sierra de la Demanda area of northern Spain. *Ann. For. Sci.* 54(3): 261-269.
- Saurer M, Cherubini P, Ammann M, De Cinti B, Siegwolf R. 2004. First detection of nitrogen from NO_x in tree rings: a ¹⁵N/¹⁴N study near a motorway. *Atmospheric Environment* 38(18): 2779-2787.
- Scherer-Lorenzen M, Schulze E-D, Don A, Schumacher J, Weller E. 2007. Exploring the functional significance of forest diversity: A new long-term experiment with temperate tree species (BIOTREE). *Perspectives in Plant Ecology, Evolution and Systematics* 9(2): 53-70.
- Schowalter TD, Morrell JJ. 2002. Nutritional Quality of Douglas-Fir Wood: Effect of Vertical and Horizontal Position on Nutrient Levels. *Wood and Fiber Science* 34(1): 158-164.
- Schröder J, Klinner S, Körner M. 2017. A new set of biomass functions for *Quercus petraea* in Western Pomerania. *Baltic Forestry* 23(2): 449-462.
- Schulze E-D, Schulze W, Koch H, Arneth A, Bauer G, Kelliher FM, Hollinger DY, Vygodskaya NN, Kusnetsova WA, Sogatchev A, et al. 1995. Aboveground biomass and nitrogen nutrition in a chronosequence of pristine Dahurian Larix stands in eastern Siberia. *Canadian Journal of Forest Research* 25(6): 943-960.
- Seidel F, Lopez C ML, Celi L, Bonifacio E, Oikawa A, Yamanaka T. 2019b. N Isotope Fractionation in Tree Tissues During N Reabsorption and Remobilization in *Fagus crenata* Blume. *Forests* 10(4).
- Seidel F, Lopez C ML, Oikawa A, Yamanaka T. 2019a. Seasonal nitrogen partitioning in Japanese cedar (*Cryptomeria japonica*, D. Don) tissues. *Plant and Soil* 442(1-2): 511-529.
- Sicard C, Saint-Andre L, Gelhaye D, Ranger J. 2006. Effect of initial fertilisation on biomass and nutrient content of Norway spruce and Douglas-fir plantations at the same site. *Trees* 20(2): 229-246.
- Silvestri N, Giannini V, Antichi D. 2018. Intercropping cover crops with a poplar short rotation coppice: Effects on nutrient uptake and biomass production. *Italian Journal of Agronomy*: 126-133.
- Son Y, Gower ST. 1992. Nitrogen and phosphorus distribution for five plantation species in southwestern Wisconsin. *Forest Ecology and Management* 53(1): 175-193.
- Sprugel DG. 1984. Density, Biomass, Productivity, and Nutrient-Cycling Changes During Stand Development in Wave-Regenerated Balsam Fir Forests. *Ecological Monographs* 54(2): 165-186.

- Stewart CM, Van Deelen TR, Dawson JO. 2008. Autumn Herbivory by White-Tailed Deer and Nutrient Loss in Planted Seedlings. *The American Midland Naturalist* 160(2): 342-349.
- Stockfors J, Linder S. 1998. Effect of nitrogen on the seasonal course of growth and maintenance respiration in stems of Norway spruce trees. *Tree Physiology* 18(3): 155-166.
- Svoboda M, Matějka K, Kopáček J. 2006. Biomass and element pools of selected spruce trees in the catchments of Plešné and Čertovo Lakes in the Šumava Mts. *Journal of Forest Science* 52(10): 482-495.
- Tamminen P, Saarsalmi A, Kukkola M. 2012. Amount of boron in Norway spruce stands in eastern Finland. *Forest Ecology and Management* 269: 92-98.
- Terziev N, Boutelje J, Larsson K. 2008. Seasonal fluctuations of low-molecular-weight sugars, starch and nitrogen in sapwood of *Pinus sylvestris* L. *Scandinavian Journal of Forest Research* 12(2): 216-224.
- Thiébeau P, Bertrand I. 2020. Production de biomasse et immobilisation de carbone et d'azote sur des sols marginaux: cas de taillis à très courte rotation conduits sans fertilisation. *Biotechnologie, Agronomie, Société et Environnement / Biotechnology, Agronomy, Society and Environment* 24(1): 1-13.
- Tognetti R, Johnson JD, Michelozzi M, Raschi A. 1998. Response of foliar metabolism in mature trees of *Quercus pubescens* and *Quercus ilex* to long-term elevated CO₂. *Environmental and Experimental Botany* 39(3): 233-245.
- Tomlinson G, Siegwolf RT, Buchmann N, Schleppi P, Waldner P, Weber P. 2014. The mobility of nitrogen across tree-rings of Norway spruce (*Picea abies* L.) and the effect of extraction method on tree-ring delta(1)(5)N and delta(1)(3)C values. *Rapid Commun Mass Spectrom* 28(11): 1258-1264.
- Tsutsumi T 1971. Accumulation and circulation of nutrient elements in forest ecosystems. In: Duvigneaud P ed. Productivity of forest ecosystems. Proceedings of the Brussels symposium organized by Unesco und the International Biological Programme (27-31 October 1969). Paris: Unesco, 543-552.
- Turner J, Cole DW, Gessel SP. 1976. Mineral Nutrient Accumulation and Cycling in a Stand of Red Alder (*Alnus Rubra*). *Journal of Ecology* 64(3): 965-974.
- Turner J, Singer MJ. 1976. Nutrient Distribution and Cycling in a Sub-Alpine Coniferous Forest Ecosystem. *Journal of Applied Ecology* 13(1): 295-301.
- Uchytlova T, Krejza J, Vesela B, Holub P, Urban O, Horacek P, Klem K. 2019. Ultraviolet radiation modulates C:N stoichiometry and biomass allocation in *Fagus sylvatica* saplings cultivated under elevated CO₂ concentration. *Plant Physiol Biochem* 134: 103-112.
- Uri V, Lõhmus K, Ostonen I, Tullus H, Lastik R, Villo M. 2007. Biomass production, foliar and root characteristics and nutrient accumulation in young silver birch (*Betula pendula* Roth.) stand growing on abandoned agricultural land. *European Journal of Forest Research* 126(4): 495-506.
- Varik M, Aosaar J, Ostonen I, Lõhmus K, Uri V. 2013. Carbon and nitrogen accumulation in belowground tree biomass in a chronosequence of silver birch stands. *Forest Ecology and Management* 302: 62-70.
- Vergutz L, Manzoni S, Porporato A, Novais RF, Jackson RB. 2012. Global resorption efficiencies and concentrations of carbon and nutrients in leaves of terrestrial plants. *Ecological Monographs* 82(2): 205-220.
- Vogt KA, Grier CC, Meier CE, Edmonds RL. 1982. Mycorrhizal Role in Net Primary Production and Nutrient Cycling in *Abies Amabilis* Ecosystems in Western Washington. *Ecology* 63(2): 370-380.
- Voigt GK, Steucek GL. 1969. Nitrogen Distribution and Accretion in an Alder Ecosystem. *Soil Science Society of America Journal* 33(6): 946-949.

- Voronin PY, Mukhin VA, Velivetskaya TA, Ignat'ev AV, Kuznetsov VV. 2017. Isotope composition of carbon and nitrogen in tissues and organs of *Betula pendula*. Russian Journal of Plant Physiology 64(2): 184-189.
- Wang H, Chen D, Sun X. 2019. Nutrient Allocation to Different Compartments of Age-Sequence Larch Plantations in China. Forests 10(9).
- Węgiel A, Bielinis E, Polowy K. 2018. Macronutrient Stocks in Scots Pine Stands of Different Densities. Forests 9(10).
- Wei H, Xu C, Ma L, Duan J, Jiang L, Ren J. 2014. Effect of Late-Season Fertilization on Nutrient Reserves and Carbohydrate Accumulation in Barerootlarix Olgensisseedlings. Journal of Plant Nutrition 37(2): 279-293.
- Weigt RB, Haberle KH, Millard P, Metzger U, Ritter W, Blaschke H, Gottlein A, Matyssek R. 2012. Ground-level ozone differentially affects nitrogen acquisition and allocation in mature European beech (*Fagus sylvatica*) and Norway spruce (*Picea abies*) trees. Tree Physiol 32(10): 1259-1273.
- Wielgolaski FE, Kjelvik S, Kallio P 1975. Mineral Content of Tundra and Forest Tundra Plants in Fennoscandia. In: Wielgolaski FE ed. Fennoscandian Tundra Ecosystems: Part 1 Plants and Microorganisms. Berlin, Heidelberg: Springer Berlin Heidelberg, 316-332.
- Willis CG, Halina M, Lehman C, Reich PB, Keen A, McCarthy S, Cavender-Bares J. 2010. Phylogenetic community structure in Minnesota oak savanna is influenced by spatial extent and environmental variation. Ecography 33(3): 565-577.
- Wilson KB, Baldocchi DD, Hanson PJ. 2000. Spatial and seasonal variability of photosynthetic parameters and their relationship to leaf nitrogen in a deciduous forest. Tree Physiology 20(9): 565-578.
- Wirth C, Schulze E-D, Lüker B, Grigoriev S, Siry M, Hardes G, Ziegler W, Backor M, Bauer G, Vygodskaya NN. 2002. Fire and site type effects on the long-term carbon and nitrogen balance in pristine Siberian Scots pine forests. Plant and Soil 242(1): 41-63.
- Wittwer RF, Stringer JW. 1985. Biomass production and nutrient accumulation in seedling and coppice hardwood plantations. Forest Ecology and Management 13(3): 223-233.
- Woodwell GM, Whittaker RH, Houghton RA. 1975. Nutrient Concentrations in Plants in the Brookhaven Oak-Pine Forest. Ecology 56(2): 318-332.
- Wright TW, Will GM. 1958. The nutrient content of Scots and Corsican pines growing on sand dunes. Forestry: An International Journal of Forest Research 31(1): 13-25.
- Wullschleger SD, Norby RJ, Love JC, Runck C. 1997. Energetic Costs of Tissue Construction in Yellow-poplar and White Oak Trees Exposed to Long-term CO₂ Enrichment. Annals of Botany 80(3): 289-297.
- Xiao Y, Peng F, Dang Z, Jiang X, Zhang J, Zhang Y, Shu H. 2015. Influence of rhizosphere ventilation on soil nutrient status, root architecture and the growth of young peach trees. Soil Science and Plant Nutrition 61(5): 775-787.
- Xiao Y, Peng Y, Peng F, Zhang Y, Yu W, Sun M, Gao X. 2018. Effects of concentrated application of soil conditioners on soil-air permeability and absorption of nitrogen by young peach trees. Soil Science and Plant Nutrition 64(3): 423-432.
- Xu L, Baldocchi DD. 2003. Seasonal trends in photosynthetic parameters and stomatal conductance of blue oak (*Quercus douglasii*) under prolonged summer drought and high temperature. Tree Physiology 23(13): 865-877.
- Yan CF, Gessler A, Rigling A, Dobbertin M, Han XG, Li MH. 2016. Effects of mistletoe removal on growth, N and C reserves, and carbon and oxygen isotope composition in Scots pine hosts. Tree Physiol 36(5): 562-575.

- Yao X, Liu Q. 2009. Responses in Some Growth and Mineral Elements of Mono Maple Seedlings to Enhanced Ultraviolet-B and to Nitrogen Supply. *Journal of Plant Nutrition* 32(5): 772-784.
- Yeo J-K, Lee W-W, Koo Y-B, Woo K-S, Byun J-K. 2010. Nitrogen Storage Potential in Aboveground Biomass of Three-year-old Poplar Clones in a Riparian Area. *Journal of Agriculture & Life Science* 44(3): 15-21.
- Yguel B, Bailey R, Tosh ND, Vialatte A, Vasseur C, Vitrac X, Jean F, Prinzing A. 2011. Phytophagy on phylogenetically isolated trees: why hosts should escape their relatives. *Ecol Lett* 14(11): 1117-1124.
- Youssefi F, Brown P, Weinbaum S. 2015. Relationship between tree nitrogen status, xylem and phloem sap amino acid concentrations, and apparent soil nitrogen uptake by almond trees(*Prunus dulcis*). *The Journal of Horticultural Science and Biotechnology* 75(1): 62-68.
- Zhao K, Zheng M, Fahey TJ, Jia Z, Ma L. 2018. Vertical gradients and seasonal variations in the stem CO₂ efflux of *Larix principis-rupprechtii* Mayr. *Agricultural and Forest Meteorology* 262: 71-80.
- Zhao Q, Liu X-y, Zeng D-h. 2011. Aboveground biomass and nutrient allocation in an age-sequence of *Larix olgensis* plantations. *Journal of Forestry Research* 22(1): 71-76.

S3 Classification of tree species into Growth / Leaf type classes

Broadleaf deciduous, fast-growing

Acer campestre, Acer davidii, Acer macrophyllum, Acer negundo, Acer negundo subsp. californicum, Acer pictum, Acer platanoides, Acer pseudoplatanus, Acer rubrum, Aesculus hippocastanum, Alnus cordata, Alnus glutinosa, Alnus hirsuta, Alnus incana, Alnus incana subsp. rugosa, Alnus rhombifolia, Alnus rubra, Alnus rugosa, Betula alleghaniensis, Betula ermanii, Betula lenta, Betula nigra, Betula papyrifera, Betula pendula, Betula platyphylloides, Betula platyphylloides subsp. mandshurica, Betula populifolia, Betula pubescens, Betula tortuosa, Betula utilis, Betula verrucosa, Cornus alternifolia, Cornus controversa, Cornus walteri, Fraxinus americana, Fraxinus angustifolia, Fraxinus angustifolia subsp. oxycarpa, Fraxinus excelsior, Fraxinus pennsylvanica, Juglans mandshurica, Juglans nigra, Liquidambar styraciflua, Platanus occidentalis, Populus alba, Populus balsamifera, Populus cathayana, Populus deltoides, Populus deltoides subsp. monilifera, Populus euphratica, Populus fremontii, Populus gileadensis, Populus grandidentata, Populus hopeiensis, Populus koreana, Populus nigra, Populus robusta, Populus simonii, Populus suaveolens, Populus tremula, Populus tremula var. Davidiana, Populus tremula x tremuloides, Populus tremuloides, Populus trichocarpa, Prunus avium, Prunus pensylvanica, Prunus persica, Prunus serotina, Prunus virginiana, Quercus cerris, Quercus dentata, Quercus lobata, Quercus nigra, Quercus rubra, Quercus texana, Robinia pseudoacacia, Robinia pseudoacacia var. Inermis, Salix amygdaloides, Salix atrocinerea, Salix babylonica, Salix bebbiana, Salix caprea, Salix cinerea, Salix dunnii, Salix eleagnos subsp. eleagnos, Salix fragilis, Salix hultenii, Salix lasiolepis, Salix lucida, Salix lucida ssp. laetiandra, Salix pentandra, Salix rorida, Salix triandra, Salix udensis, Salix viminalis, Sorbus alnifolia, Sorbus torminalis, Ulmus glabra, Ulmus minor, Ulmus pumila

Broadleaf deciduous, slow-/medium-growing

Acer buergerianum, Acer caudatum subsp. ukurundense, Acer davidii subsp. grosseri, Acer japonicum, Acer mandshuricum, Acer monspessulanum, Acer opalus, Acer opalus subsp. obtusatum, Acer palmatum, Acer pensylvanicum, Acer pseudosieboldianum, Acer saccharum, Acer saccharum subsp. floridanum, Acer spicatum, Acer tataricum subsp. ginnala, Acer tegmentosum, Acer truncatum, Aesculus californica, Betula albosericea, Betula costata, Betula dahurica, Carpinus betulus, Carya alba, Carya glabra, Carya ovata, Castanea dentata, Castanea mollissima, Castanea sativa, Cornus florida, Cornus kousa subsp. chinensis, Cornus kousa subsp. kousa, Cornus macrophylla, Cornus mas, Cornus officinalis, Fagus crenata, Fagus grandifolia, Fagus sylvatica, Fraxinus chinensis, Fraxinus chinensis subsp. rhynchophylla, Fraxinus mandshurica, Fraxinus nigra, Ilex verticillata, Juglans ailanthifolia, Juglans californica, Juglans cinerea, Juglans hindsii, Juglans regia, Liquidambar formosana, Liriodendron tulipifera, Magnolia fraseri, Magnolia kobus, Magnolia obovata, Magnolia officinalis, Malus baccata, Malus sylvestris, Nyssa sylvatica, Oxydendrum arboreum, Phellodendron amurense, Prunus armeniaca, Prunus cerasifera, Prunus davidiana, Prunus domestica, Prunus padus, Quercus acutissima, Quercus alba, Quercus aliena var. acutiserrata, Quercus bicolor, Quercus chenii, Quercus coccinea, Quercus douglasii, Quercus ellipsoidalis, Quercus faginea, Quercus falcata, Quercus gambelii, Quercus ilicifolia, Quercus kelloggii, Quercus laevis, Quercus macrocarpa, Quercus michauxii, Quercus mongolica, Quercus mongolica subsp. crispula, Quercus petraea, Quercus prinus, Quercus pubescens, Quercus pyrenaica, Quercus robur, Quercus serrata subsp. serrata, Quercus variabilis, Quercus velutina, Robinia neomexicana, Sorbus americana, Sorbus aria, Sorbus aucuparia, Sorbus commixta, Tilia americana, Tilia amurensis, Tilia cordata, Tilia japonica, Tilia mandshurica, Tilia mongolica, Tilia platyphyllos, Ulmus americana, Ulmus davidiana, Ulmus davidiana var. japonica, Ulmus laciniata, Ulmus parvifolia

Needleleaf deciduous, fast-growing

Larix decidua, Larix decidua x leptolepis, Larix kaempferi, Larix leptolepis, Larix occidentalis, Larix x eurolepis, Taxodium distichum

Needleleaf deciduous, slow-/medium-growing

Larix dahurica, Larix gmelinii, Larix gmelinii var. olgensis, Larix laricina, Larix lyallii, Larix olgensis, Larix principis-rupprechtii, Larix sibirica

Needleleaf evergreen, fast-growing

Abies alba, *Abies bornmulleriana*, *Abies fraseri*, *Abies grandis*, *Cryptomeria fortunei*, *Cryptomeria japonica*, *Picea abies*, *Picea crassifolia*, *Picea glauca*, *Picea omorika*, *Picea rubens*, *Picea sitchensis*, *Pinus banksiana*, *Pinus brutia*, *Pinus contorta*, *Pinus monticola*, *Pinus muricata*, *Pinus nigra*, *Pinus nigra* var. *calabrica*, *Pinus palustris*, *Pinus pinaster*, *Pinus radiata*, *Pinus strobus*, *Pinus sylvestris*, *Pseudotsuga menziesii*, *Sequoia sempervirens*, *Thuja plicata*, *Tsuga heterophylla*

Needleleaf evergreen, slow-/medium-growing

Abies amabilis, *Abies balsamea*, *Abies concolor*, *Abies firma*, *Abies lasiocarpa*, *Abies mayriana*, *Abies nephrolepis*, *Abies sachalinensis*, *Abies sibirica*, *Cedrus deodara*, *Chamaecyparis obtusa*, *Juniperus communis*, *Juniperus deppeana*, *Juniperus monosperma*, *Juniperus osteosperma*, *Juniperus oxycedrus*, *Juniperus oxycedrus* var. *oxycedrus*, *Juniperus scopulorum*, *Juniperus virginiana*, *Juniperus virginiana*, *Picea engelmannii*, *Picea jezoensis*, *Picea mariana*, *Picea mariana* x *rubens*, *Picea meyeri*, *Picea obovata*, *Picea orientalis*, *Picea wilsonii*, *Pinus albicaulis*, *Pinus aristata*, *Pinus armandii*, *Pinus bungeana*, *Pinus cembra*, *Pinus densiflora*, *Pinus echinata*, *Pinus edulis*, *Pinus flexilis*, *Pinus halepensis*, *Pinus jeffreyi*, *Pinus koraiensis*, *Pinus massoniana*, *Pinus monophylla*, *Pinus pinea*, *Pinus ponderosa*, *Pinus resinosa*, *Pinus rigida*, *Pinus serotina*, *Pinus sibirica*, *Pinus strobiiformis*, *Pinus tabulaeformis*, *Pinus tabuliformis*, *Pinus taeda*, *Pinus thunbergii*, *Pinus uncinata*, *Thuja occidentalis*, *Tsuga canadensis*, *Tsuga chinensis*, *Tsuga mertensiana*, *Tsuga sieboldii*

S4 Generalized additive models

A total of 17 generalized additive models (GAMs) are implemented for each tree tissue N concentration, using different combinations of explanatory variables. While GAMs (1) – (9) consider plant trait variables, GAMs (10) – (12) consider environmental condition variables. GAMs (13) – (17) incorporate plant traits and environmental conditions:

- (1) Leaf types (LT; broadleaf deciduous, needleleaf deciduous, needleleaf evergreen)
- (2) Growth rate (GR) classes (slow-/medium growing, fast-growing)
- (3) Leaf type / growth rate (LT/GR) classes
- (4) Tree age
- (5) Tree height
- (6) Compartment biomass
- (7) LT/GR + Age
- (8) LT/GR + Height
- (9) LT/GR + Biomass
- (10) MAT + MAP
- (11) Soil N concentration
- (12) MAT + MAP + Soil N
- (13) LT/GR + Soil N
- (14) LT + MAT + MAP + Soil N
- (15) LT + Age + MAT + MAP
- (16) LT + Height + MAT + MAP
- (17) LT + Biomass + MAT + MAP

Table S1: Modelling efficiencies (MEFs) of all the 17 applied generalized additive models (GAMs) for modelling leaf N concentration using different combinations of explanatory variables. n indicates the number of available measurements for each GAM.

GAM	Variables	Formula	n	MEF
(1)	Leaf Type (LT)	Leaf_N ~ factor(LT)	5944	0.51
(2)	Growth Rate (GR)	Leaf_N ~ factor(GR)	5944	0.032
(3)	Leaf Type / Growth Rate (LT/GR)	Leaf_N ~ factor(LTGR)	5944	0.524
(4)	Age	Leaf_N ~ s(Age)	428	0.07
(5)	Height	Leaf_N ~ s(Height)	416	0.336

(6)	Biomass	$\text{Leaf_N} \sim s(\text{Biomass})$	73	0.368
(7)	LT/GR + Age	$\text{Leaf_N} \sim s(\text{Age}) + \text{factor(LTGR)}$	428	0.454
(8)	LT/GR + Height	$\text{Leaf_N} \sim s(\text{Height}, \text{by} = \text{LTGR}) + \text{factor(LTGR)}$	416	0.743
(9)	LT/GR + Biomass	$\text{Leaf_N} \sim s(\text{Biomass}, \text{by} = \text{LTGR})$	73	0.772
(10)	MAT + MAP	$\text{Leaf_N} \sim s(\text{MAT}) + s(\text{MAP}) + \text{te}(\text{MAT}, \text{MAP})$	5944	0.134
(11)	Soil N	$\text{Leaf_N} \sim s(\text{Soil_N})$	624	0.27
(12)	MAT + MAP + Soil N	$\text{Leaf_N} \sim s(\text{MAT}) + s(\text{MAP}) + s(\text{Soil_N}) + \text{te}(\text{MAT}, \text{MAP}) + \text{te}(\text{MAT}, \text{Soil_N}) + \text{te}(\text{MAP}, \text{Soil_N})$	624	0.516
(13)	LT/GR + Soil N	$\text{Leaf_N} \sim s(\text{Soil_N}, \text{by} = \text{LTGR})$	624	0.616
(14)	LT + MAT + MAP + Soil N	$\text{Leaf_N} \sim s(\text{MAT}, \text{by} = \text{LT}) + s(\text{MAP}, \text{by} = \text{LT}) + s(\text{Soil_N}, \text{by} = \text{LT}) + \text{factor(LT)} + \text{te}(\text{MAT}, \text{Soil_N}) + \text{te}(\text{MAP}, \text{Soil_N})$	624	0.698
(15)	LT + Age + MAT + MAP	$\text{Leaf_N} \sim s(\text{MAT}, \text{by} = \text{LT}) + s(\text{MAP}, \text{by} = \text{LT}) + s(\text{Age}, \text{by} = \text{LT}) + \text{factor(LT)} + \text{te}(\text{MAT}, \text{Age})$	428	0.618
(16)	LT + Height + MAT + MAP	$\text{Leaf_N} \sim s(\text{MAT}, \text{by} = \text{LT}) + s(\text{MAP}, \text{by} = \text{LT}) + s(\text{Height}, \text{by} = \text{LT}) + \text{factor(LT)} + \text{te}(\text{MAP}, \text{Height})$	416	0.761
(17)	LT + Biomass + MAT + MAP	$\text{Leaf_N} \sim s(\text{MAT}) + s(\text{MAP}) + s(\text{Biomass}) + \text{factor(LT)} + \text{te}(\text{MAT}, \text{Biomass})$	73	0.779

Table S2: Modelling efficiencies (MEFs) of all the 17 applied generalized additive models (GAMs) for modelling branch N concentration using different combinations of explanatory variables. n indicates the number of available measurements for each GAM.

GAM	Variables	Formula	n	MEF
(1)	Leaf Type (LT)	$\text{Branch_N} \sim \text{factor(LT)}$	599	0.078
(2)	Growth Rate (GR)	$\text{Branch_N} \sim \text{factor(GR)}$	599	0.019
(3)	Leaf Type / Growth Rate (LT/GR)	$\text{Branch_N} \sim \text{factor(LTGR)}$	599	0.146
(4)	Age	$\text{Branch_N} \sim s(\text{Age})$	437	0.248
(5)	Height	$\text{Branch_N} \sim s(\text{Height})$	312	0.041
(6)	Biomass	$\text{Branch_N} \sim s(\text{Biomass})$	300	0.022
(7)	LT/GR + Age	$\text{Branch_N} \sim s(\text{Age}, \text{by} = \text{LTGR})$	437	0.402
(8)	LT/GR + Height	$\text{Branch_N} \sim s(\text{Height}, \text{by} = \text{LTGR})$	312	0.348
(9)	LT/GR + Biomass	$\text{Branch_N} \sim s(\text{Biomass}, \text{by} = \text{LTGR})$	300	0.379
(10)	MAT + MAP	$\text{Branch_N} \sim s(\text{MAT}) + s(\text{MAP}) + \text{te}(\text{MAT}, \text{MAP})$	599	0.428
(11)	Soil N	$\text{Branch_N} \sim s(\text{Soil_N})$	201	0.087
(12)	MAT + MAP + Soil	$\text{Branch_N} \sim s(\text{MAT}) + s(\text{MAP}) + s(\text{Soil_N}) + \text{te}(\text{MAT}, \text{MAP}) + \text{te}(\text{MAT}, \text{Soil})$	201	0.692

	N	Soil_N)		
(13)	LT/GR + Soil N	Branch_N ~ s(Soil_N, by = LTGR)	201	0.55
(14)	LT + MAT + MAP + Soil N	Branch_N ~ s(MAT) + s(MAP) + s(Soil_N) + factor(LT) + te(MAT, MAP) + te(MAT, Soil_N)	201	0.701
(15)	LT + Age + MAT + MAP	Branch_N ~ s(MAT, by = LT) + s(MAP, by = LT) + s(Age, by = LT) + factor(LT) + te(MAT, MAP) + te(MAP, Age)	437	0.599
(16)	LT + Height + MAT + MAP	Branch_N ~ s(MAT, by = LT) + s(MAP, by = LT) + s(Height, by = LT) + te(MAT, Height) + te(MAP, Height)	312	0.573
(17)	LT + Biomass + MAT + MAP	Branch_N ~ s(MAT, by = LT) + s(MAP, by = LT) + s(Biomass, by = LT) + te(MAT, MAP) + te(MAT, Biomass) + te(MAP, Biomass)	300	0.702

Table S3: Modelling efficiencies (MEFs) of all the 17 applied generalized additive models (GAMs) for modelling stem N concentration using different combinations of explanatory variables. n indicates the number of available measurements for each GAM.

GAM	Variables	Formula	n	MEF
(1)	Leaf Type (LT)	Stem_N ~ factor(LT)	1048	0.119
(2)	Growth Rate (GR)	Stem_N ~ factor(GR)	1048	0
(3)	Leaf Type / Growth Rate (LT/GR)	Stem_N ~ factor(LTGR)	1048	0.122
(4)	Age	Stem_N ~ s(Age)	823	0.366
(5)	Height	Stem_N ~ s(Height)	515	0.315
(6)	Biomass	Stem_N ~ s(Biomass)	320	0.228
(7)	LT/GR + Age	Stem_N ~ s(Age, by = LTGR)	823	0.605
(8)	LT/GR + Height	Stem_N ~ s(Height, by = LTGR) + factor(LTGR)	515	0.555
(9)	LT/GR + Biomass	Stem_N ~ s(Biomass, by = LTGR) + factor(LTGR)	320	0.416
(10)	MAT + MAP	Stem_N ~ s(MAT) + s(MAP) + te(MAT, MAP)	1048	0.151
(11)	Soil N	Stem_N ~ s(Soil_N)	323	0.002
(12)	MAT + MAP + Soil N	Stem_N ~ s(MAT) + s(MAP) + s(Soil_N) + te(MAT, MAP) + te(MAT, Soil_N)	323	0.488
(13)	LT/GR + Soil N	Stem_N ~ s(Soil_N, by = LTGR) + factor(LTGR)	323	0.724
(14)	LT + MAT + MAP + Soil N	Stem_N ~ s(MAT, by = LT) + s(MAP, by = LT) + s(Soil_N, by = LT) + te(MAT, MAP) + te(MAT, Soil_N) + te(MAP, Soil_N)	323	0.922
(15)	LT + Age + MAT + MAP	Stem_N ~ s(MAT, by = LT) + s(MAP, by = LT) + s(Age, by = LT) + te(MAT, Age)	823	0.682
(16)	LT + Height + MAT + MAP	Stem_N ~ s(MAT, by = LT) + s(MAP, by = LT) + s(Height, by = LT) + te(MAT, MAP) + te(MAT, Height) + te(MAP, Height)	515	0.669
(17)	LT + Biomass +	Stem_N ~ s(MAT, by = LT) + s(MAP, by = LT) + s(Biomass, by = LT) +	320	0.657

	MAT + MAP	factor(LT) + te(MAT, MAP) + te(MAT, Biomass) + te(MAP, Biomass)		
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Table S4: Modelling efficiencies (MEFs) of all the 17 applied generalized additive models (GAMs) for modelling root N concentration using different combinations of explanatory variables. n indicates the number of available measurements for each GAM.

GAM	Variables	Formula	n	MEF
(1)	Leaf Type (LT)	Root_N ~ factor(LT)	267	0.118
(2)	Growth Rate (GR)	Root_N ~ factor(GR)	267	0
(3)	Leaf Type / Growth Rate (LT/GR)	Root_N ~ factor(LTGR)	267	0.193
(4)	Age	Root_N ~ s(Age)	245	0.232
(5)	Height	Root_N ~ s(Height)	98	0.046
(6)	Biomass	Root_N ~ s(Biomass)	111	0.147
(7)	LT/GR + Age	Root_N ~ s(Age, by = LTGR) + factor(LTGR)	245	0.455
(8)	LT/GR + Height	Root_N ~ s(Height, by = LTGR) + factor(LTGR)	98	0.568
(9)	LT/GR + Biomass	Root_N ~ s(Biomass, by = LTGR)	111	0.403
(10)	MAT + MAP	Root_N ~ s(MAT) + s(MAP) + te(MAT, MAP)	267	0.352
(11)	Soil N	Root_N ~ s(Soil_N)	136	0.552
(12)	MAT + MAP + Soil N	Root_N ~ s(MAT) + s(MAP) + s(Soil_N) + te(MAT, Soil_N) + te(MAP, Soil_N)	136	0.862
(13)	LT/GR + Soil N	Root_N ~ s(Soil_N, by = LTGR) + factor(LTGR)	136	0.826
(14)	LT + MAT + MAP + Soil N	Root_N ~ s(MAT, by = LT) + s(MAP, by = LT) + s(Soil_N, by = LT)	136	0.871
(15)	LT + Age + MAT + MAP	Root_N ~ s(MAT, by = LT) + s(MAP, by = LT) + s(Age, by = LT) + te(MAT, MAP) + te(MAP, Age)	245	0.775
(16)	LT + Height + MAT + MAP	Root_N ~ s(MAT) + s(MAP) + s(Height) + factor(LT) + te(MAT, MAP)	98	0.757
(17)	LT + Biomass + MAT + MAP	Root_N ~ s(MAT, by = LT) + s(MAP, by = LT) + s(Biomass, by = LT) + te(MAT, MAP)	111	0.928

S5 N concentration summary statistics

Table S5: Leaf N Concentration [gN g^{-1}] summary statistics for all growth / leaf type classes.

Leaf N Concentration [gN g^{-1}]	1 st quartile	Median	Mean	3 rd quartile
Broadleaf Deciduous Fast-growing	0.0194	0.0241	0.0238	0.0282
Broadleaf Deciduous Slow-/Medium-growing	0.0183	0.0215	0.0216	0.0247
Needleleaf Deciduous Fast-growing	0.0167	0.0194	0.0193	0.0213
Needleleaf Deciduous Slow-/Medium-growing	0.0133	0.0172	0.0172	0.0211
Needleleaf Evergreen Fast-growing	0.0107	0.0125	0.0126	0.0143
Needleleaf Evergreen Slow-/Medium-growing	0.0099	0.0118	0.0122	0.0148
All Broadleaf Deciduous	0.0185	0.0222	0.0224	0.0260
All Needleleaf Deciduous	0.0155	0.0185	0.0183	0.0213
All Needleleaf Evergreen	0.0105	0.0124	0.0125	0.0144
All Fast-growing	0.0118	0.0144	0.0168	0.0209
All Slow-/Medium-growing	0.0147	0.0194	0.0192	0.0234
All	0.0124	0.0167	0.0179	0.0226

Table S6: Branch N Concentration [gN g^{-1}] summary statistics for all growth / leaf type classes.

Branch N Concentration [gN g^{-1}]	1 st quartile	Median	Mean	3 rd quartile
Broadleaf Deciduous Fast-growing	0.0035	0.0053	0.0063	0.0076
Broadleaf Deciduous Slow-/Medium-growing	0.0026	0.0038	0.0041	0.0053
Needleleaf Deciduous Fast-growing	0.0047	0.0050	0.0053	0.0055
Needleleaf Deciduous Slow-/Medium-growing	0.0038	0.0048	0.0044	0.0053
Needleleaf Evergreen Fast-growing	0.0024	0.0033	0.0036	0.0049
Needleleaf Evergreen Slow-/Medium-growing	0.0020	0.0027	0.0032	0.0038
All Broadleaf Deciduous	0.0029	0.0042	0.0050	0.0059
All Needleleaf Deciduous	0.0040	0.0049	0.0046	0.0053
All Needleleaf Evergreen	0.0022	0.0030	0.0035	0.0045
All Fast-growing	0.0025	0.0039	0.0044	0.0054
All Slow-/Medium-growing	0.0022	0.0032	0.0037	0.0049
All	0.0024	0.0035	0.0040	0.0051

Table S7: Stem N Concentration [gN g^{-1}] summary statistics for all growth / leaf type classes.

Stem N Concentration [gN g^{-1}]	1 st quartile	Median	Mean	3 rd quartile
Broadleaf Deciduous Fast-growing	0.0010	0.0016	0.0023	0.0026
Broadleaf Deciduous Slow-/Medium-growing	0.0013	0.0018	0.0021	0.0025
Needleleaf Deciduous Fast-growing	0.0006	0.0007	0.0008	0.0009
Needleleaf Deciduous Slow-/Medium-growing	0.0009	0.0010	0.0013	0.0013
Needleleaf Evergreen Fast-growing	0.0006	0.0008	0.0011	0.0012
Needleleaf Evergreen Slow-/Medium-growing	0.0003	0.0006	0.0009	0.0010
All Broadleaf Deciduous	0.0012	0.0017	0.0022	0.0025
All Needleleaf Deciduous	0.0008	0.0010	0.0013	0.0013
All Needleleaf Evergreen	0.0005	0.0008	0.0010	0.0011
All Fast-growing	0.0007	0.0010	0.0014	0.0015
All Slow-/Medium-growing	0.0007	0.0010	0.0014	0.0017
All	0.0007	0.0010	0.0014	0.0016

Table S8: Root N Concentration [gN g^{-1}] summary statistics for all growth / leaf type classes.

Root N Concentration [gN g^{-1}]	1 st quartile	Median	Mean	3 rd quartile
Broadleaf Deciduous Fast-growing	0.0052	0.0078	0.0094	0.0111
Broadleaf Deciduous Slow-/Medium-growing	0.0035	0.0045	0.0056	0.0068
Needleleaf Deciduous Fast-growing	0.0026	0.0032	0.0032	0.0033
Needleleaf Deciduous Slow-/Medium-growing	0.0058	0.0074	0.0073	0.0093
Needleleaf Evergreen Fast-growing	0.0015	0.0033	0.0045	0.0075
Needleleaf Evergreen Slow-/Medium-growing	0.0016	0.0046	0.0045	0.0062
All Broadleaf Deciduous	0.0039	0.0064	0.0077	0.0089
All Needleleaf Deciduous	0.0051	0.0071	0.0070	0.0091
All Needleleaf Evergreen	0.0015	0.0038	0.0045	0.0070
All Fast-growing	0.0024	0.0052	0.0060	0.0079
All Slow-/Medium-growing	0.0038	0.0061	0.0062	0.0085
All	0.0033	0.0060	0.0061	0.0083

S6 Significance of differences between leaf, branch, stem, and root N concentration

The significance of differences between leaf, branch, stem, and root N concentration is quantified by the p-values of pairwise t-tests (Table S9).

Table S9: P-values of pairwise t-tests of N concentration in all compartments.

p-value	Leaf N Concentration	Branch N Concentration	Stem N Concentration
Branch N Concentration	$< 2*10^{-16}$	—	—
Stem N Concentration	$< 2*10^{-16}$	$< 2*10^{-16}$	—
Root N Concentration	$< 2*10^{-16}$	$3.2*10^{-6}$	$< 2*10^{-16}$

S7 Differences between tree species

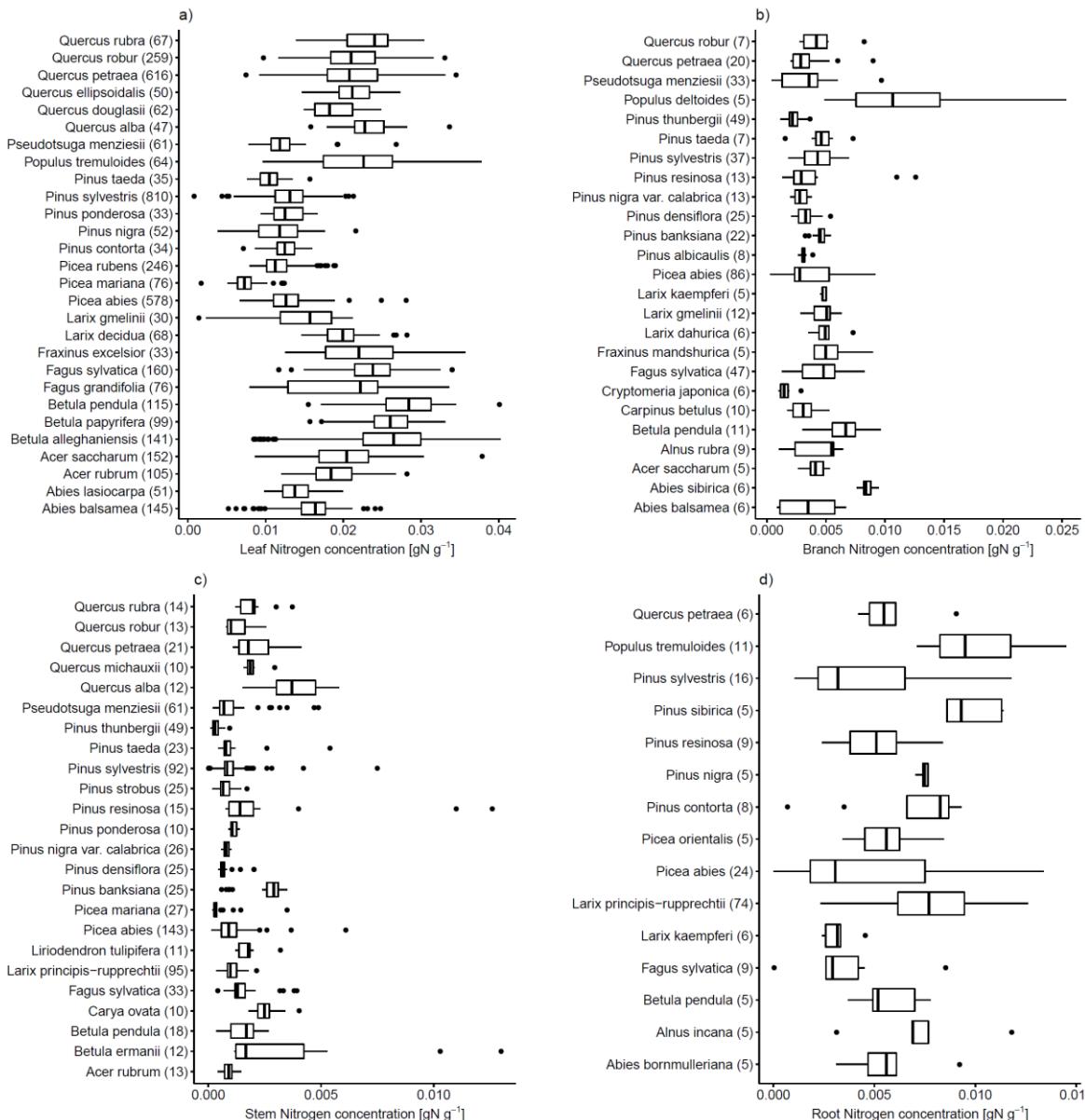


Fig. S1: a) Leaf, b) branch, c) stem and d) root N concentration [gN g⁻¹] for the most common tree species in the database. Only tree species with at least 30 in case of leaf, 10 in case of stem, and 5 in case of branch and root N concentration measurements are included in this figure.

Table S10: Leaf N Concentration [gN g⁻¹] summary statistics for all species with at least 30 measurements.

Leaf N Concentration [gN g ⁻¹]	1 st quartile	Median	Mean	3 rd quartile
Abies balsamea	0.0146	0.0164	0.0159	0.0177
Abies lasiocarpa	0.0122	0.0138	0.0139	0.0155
Acer rubrum	0.0165	0.0184	0.0189	0.0211
Acer saccharum	0.0169	0.0204	0.0197	0.0232

Betula alleghaniensis	0.0225	0.0265	0.0248	0.03
Betula papyrifera	0.024	0.026	0.0258	0.0283
Betula pendula	0.0255	0.0284	0.0281	0.0313
Fagus grandifolia	0.0129	0.0222	0.02	0.0244
Fagus sylvatica	0.0215	0.0238	0.0235	0.026
Fraxinus excelsior	0.0177	0.022	0.0225	0.0264
Larix decidua	0.018	0.02	0.0198	0.0214
Larix gmelinii	0.0119	0.0157	0.0148	0.0185
Picea abies	0.011	0.0126	0.0127	0.0142
Picea mariana	0.0064	0.0072	0.0075	0.0081
Picea rubens	0.0101	0.0113	0.0116	0.0127
Pinus contorta	0.0115	0.0125	0.0124	0.0137
Pinus nigra	0.0091	0.0118	0.0115	0.0141
Pinus ponderosa	0.0111	0.0125	0.0128	0.0148
Pinus sylvestris	0.0112	0.0132	0.0131	0.0148
Pinus taeda	0.0093	0.0105	0.0105	0.0115
Populus tremuloides	0.0174	0.0226	0.0223	0.0263
Pseudotsuga menziesii	0.0107	0.0118	0.0122	0.0131
Quercus alba	0.0215	0.0228	0.0231	0.0252
Quercus douglasii	0.0163	0.0182	0.0189	0.0212
Quercus ellipsoidalis	0.0194	0.0212	0.0214	0.0233
Quercus petraea	0.0179	0.0208	0.0211	0.0244
Quercus robur	0.0184	0.021	0.0213	0.0241
Quercus rubra	0.0205	0.024	0.0232	0.0257

Table S11: Branch N Concentration [gN g^{-1}] summary statistics for all species with at least 5 measurements.

Branch N Concentration [gN g^{-1}]	1 st quartile	Median	Mean	3 rd quartile
Abies balsamea	0.0011	0.0035	0.0036	0.0058
Abies sibirica	0.0082	0.0084	0.0085	0.0088
Acer saccharum	0.0037	0.0041	0.0041	0.0048
Alnus rubra	0.0024	0.0055	0.0044	0.0057

Betula pendula	0.0055	0.0067	0.0066	0.0075
Carpinus betulus	0.0022	0.0031	0.0032	0.0038
Cryptomeria japonica	0.0012	0.0015	0.0016	0.0018
Fagus sylvatica	0.003	0.0048	0.0045	0.0058
Fraxinus mandshurica	0.004	0.005	0.0056	0.006
Larix dahurica	0.0044	0.0049	0.005	0.0052
Larix gmelinii	0.004	0.0051	0.0048	0.0054
Larix kaempferi	0.0047	0.0047	0.0048	0.005
Picea abies	0.0024	0.0028	0.0036	0.0053
Pinus albicaulis	0.003	0.003	0.0031	0.0032
Pinus banksiana	0.0044	0.0046	0.0046	0.0049
Pinus densiflora	0.0027	0.0033	0.0033	0.0037
Pinus nigra var. calabrica	0.0024	0.0028	0.0029	0.0034
Pinus resinosa	0.0023	0.0029	0.0043	0.0041
Pinus sylvestris	0.0032	0.0043	0.0043	0.0053
Pinus taeda	0.0042	0.0046	0.0046	0.0052
Pinus thunbergii	0.0019	0.0022	0.0023	0.0026
Populus deltoides	0.0076	0.0107	0.0126	0.0147
Pseudotsuga menziesii	0.0013	0.0036	0.0034	0.0043
Quercus petraea	0.0022	0.0029	0.0034	0.0036
Quercus robur	0.0031	0.0042	0.0045	0.0051

Table S12: Stem N Concentration [gN g^{-1}] summary statistics for all species with at least 10 measurements.

Stem N Concentration [gN g^{-1}]	1 st quartile	Median	Mean	3 rd quartile
Acer rubrum	0.0007	0.0009	0.0009	0.001
Betula ermanii	0.0012	0.0017	0.0037	0.0042
Betula pendula	0.001	0.0017	0.0015	0.002
Carya ovata	0.0022	0.0025	0.0026	0.0027
Fagus sylvatica	0.0012	0.0013	0.0016	0.0016
Larix principis-rupprechtii	0.0009	0.001	0.0011	0.0012
Liriodendron tulipifera	0.0014	0.0018	0.0018	0.0018

Picea abies	0.0006	0.0009	0.001	0.0013
Picea mariana	0.0003	0.0003	0.0005	0.0004
Pinus banksiana	0.0026	0.0029	0.0025	0.0031
Pinus densiflora	0.0006	0.0006	0.0008	0.0007
Pinus nigra var. calabrica	0.0007	0.0008	0.0008	0.0009
Pinus ponderosa	0.001	0.0011	0.0011	0.0012
Pinus resinosa	0.0009	0.0014	0.0029	0.002
Pinus strobus	0.0005	0.0007	0.0008	0.0009
Pinus sylvestris	0.0007	0.0008	0.001	0.0011
Pinus taeda	0.0007	0.0008	0.0011	0.001
Pinus thunbergii	0.0002	0.0003	0.0003	0.0004
Pseudotsuga menziesii	0.0005	0.0007	0.001	0.0011
Quercus alba	0.003	0.0037	0.0039	0.0048
Quercus michauxii	0.0017	0.0019	0.0019	0.002
Quercus petraea	0.0014	0.0018	0.0022	0.0027
Quercus robur	0.0009	0.001	0.0013	0.0016
Quercus rubra	0.0014	0.002	0.002	0.0021

Table S13: Root N Concentration [gN g^{-1}] summary statistics for all species with at least 5 measurements.

Root N Concentration [gN g^{-1}]	1 st quartile	Median	Mean	3 rd quartile
Abies bornmulleriana	0.0047	0.0056	0.0057	0.0061
Alnus incana	0.0069	0.0069	0.0073	0.0077
Betula pendula	0.0049	0.0052	0.0057	0.007
Fagus sylvatica	0.0026	0.0029	0.0035	0.0042
Larix kaempferi	0.0026	0.0032	0.0032	0.0033
Larix principis-rupprechtii	0.0062	0.0077	0.0077	0.0095
Picea abies	0.0018	0.0031	0.0046	0.0075
Picea orientalis	0.0045	0.0056	0.0057	0.0062
Pinus contorta	0.0066	0.0083	0.0069	0.0087
Pinus nigra	0.0074	0.0075	0.0074	0.0077
Pinus resinosa	0.0038	0.0051	0.0051	0.0061

<i>Pinus sibirica</i>	0.0086	0.0093	0.0098	0.0113
<i>Pinus sylvestris</i>	0.0022	0.0032	0.0043	0.0065
<i>Populus tremuloides</i>	0.0082	0.0095	0.0102	0.0118
<i>Quercus petraea</i>	0.0048	0.0055	0.0058	0.0061

S8 Partial correlations

Table S14: Partial correlation (in brackets: respective p-values) between leaf, branch, stem, and root N and tree age, mean annual temperature (MAT), mean annual precipitation sum (MAP), and soil N concentration, controlled for the other respective explanatory variables, and for leaf types (BD: broadleaf deciduous, ND: needleleaf deciduous, NE: needleleaf evergreen) separately. In some cases, not enough measurements are available (-).

Partial correlation (p-value)	Leaf N	Branch N	Stem N	Root N
Age (controlled for MAT); BD	0.056 (0.487)	-0.327 (0.004)	-0.380 (0.001)	-0.522 (0.005)
Age (controlled for MAT); ND	-0.322 (0.335)	-0.175 (0.392)	-0.091 (0.316)	-0.014 (0.897)
Age (controlled for MAT); NE	0.313 (0.092)	-0.160 (0.030)	-0.151 (0.013)	-0.063 (0.615)
Age (controlled for MAP); BD	-0.036 (0.576)	-0.322 (0.003)	0.072 (0.364)	-0.465 (0.015)
Age (controlled for MAP); ND	-0.351 (0.219)	-0.041 (0.841)	-0.069 (0.448)	0.082 (0.456)
Age (controlled for MAP); NE	-0.255 (0.174)	0.109 (0.121)	-0.022 (0.703)	-0.134 (0.254)
Age (controlled for Soil N); BD	-0.160 (0.151)	-0.389 (0.004)	-0.438 (2.7×10^{-4})	-0.579 (0.002)
Age (controlled for Soil N); ND	-	-0.187 (0.444)	-0.441 (1.2×10^{-5})	0.289 (0.009)
Age (controlled for Soil N); NE	-	-0.015 (0.914)	-0.146 (0.148)	0.113 (0.636)
MAT (controlled for Age); BD	-0.286 (2.8×10^{-4})	0.201 (0.086)	0.273 (0.024)	0.289 (0.143)
MAT (controlled for Age); ND	0.120 (0.726)	0.033 (0.871)	-0.143 (0.117)	0.278 (0.010)
MAT (controlled for Age); NE	0.552 (0.002)	-0.469 (1.6×10^{-11})	-0.385 (5.6×10^{-11})	0.101 (0.418)
MAT (controlled for MAP); BD	-0.185 (2.8×10^{-13})	0.316 (3.3×10^{-4})	0.236 (0.013)	0.448 (0.005)
MAT (controlled for MAP); ND	0.040 (0.753)	-0.289 (0.136)	-0.018 (0.847)	-0.089 (0.419)
MAT (controlled for MAP); NE	0.121 (1.6×10^{-5})	-0.142 (0.048)	-0.247 (1.2×10^{-5})	0.157 (0.180)
MAT (controlled for Soil N); BD	-0.182 (3.2×10^{-4})	0.367 (0.005)	0.146 (0.326)	0.749 (0.020)
MAT (controlled for Soil N); ND	-0.044 (0.876)	-0.380 (0.120)	-0.556 (1.3×10^{-8})	0.159 (0.159)
MAT (controlled for Soil N); NE	0.044 (0.592)	-0.034 (0.834)	-0.207 (0.101)	0.170 (0.598)
MAP (controlled for Age); BD	0.072 (0.268)	-0.221 (0.043)	-0.038 (0.629)	-0.255 (0.199)
MAP (controlled for Age); ND	0.753 (0.002)	0.319 (0.112)	-0.127 (0.163)	0.479 (3.5×10^{-6})
MAP (controlled for Age); NE	0.234 (0.214)	-0.465 (2.7×10^{-12})	-0.179 (0.002)	-0.116 (0.326)
MAP (controlled for MAT); BD	-0.067 (0.008)	-0.311 (4.2×10^{-4})	-0.215 (0.023)	-0.504 (0.001)
MAP (controlled for MAT); ND	0.298 (0.018)	0.437 (0.020)	-0.026 (0.773)	0.408 (1.1×10^{-4})
MAP (controlled for MAT); NE	0.317 (< 2×10^{-16})	-0.269 (1.4×10^{-4})	-0.057 (0.320)	-0.125 (0.287)
MAP (controlled for Soil N); BD	-0.236 (2.7×10^{-6})	-0.192 (0.123)	-0.073 (0.584)	-0.838 (0.005)
MAP (controlled for Soil N); ND	-0.082 (0.771)	0.274 (0.271)	-0.342 (9.8×10^{-4})	0.226 (0.044)
MAP (controlled for Soil N); NE	0.071 (0.390)	-0.491 (0.001)	0.053 (0.677)	-0.055 (0.858)
Soil N (controlled for Age); BD	-0.120 (0.282)	-0.200 (0.156)	-0.235 (0.059)	0.540 (0.004)
Soil N (controlled for Age); ND	-	-0.198 (0.417)	-0.022 (0.836)	-0.466 (1.3×10^{-5})
Soil N (controlled for Age); NE	-	0.118 (0.394)	0.239 (0.017)	0.186 (0.432)
Soil N (controlled for MAT); BD	0.006 (0.905)	-0.066 (0.628)	-0.059 (0.694)	0.168 (0.666)

Soil N (controlled for MAT); ND	0.062 (0.825)	-0.344 (0.162)	-0.476 (2.2×10^{-6})	-0.421 (1.0×10^{-4})
Soil N (controlled for MAT); NE	0.061 (0.461)	0.054 (0.739)	-0.024 (0.853)	-0.261 (0.413)
Soil N (controlled for MAP); BD	0.058 (0.253)	-0.207 (0.095)	-0.181 (0.170)	0.825 (0.006)
Soil N (controlled for MAP); ND	0.076 (0.788)	-0.079 (0.757)	-0.201 (0.058)	-0.134 (0.238)
Soil N (controlled for MAP); NE	0.054 (0.517)	0.109 (0.504)	-0.034 (0.791)	-0.036 (0.906)

S9 Aridity Index

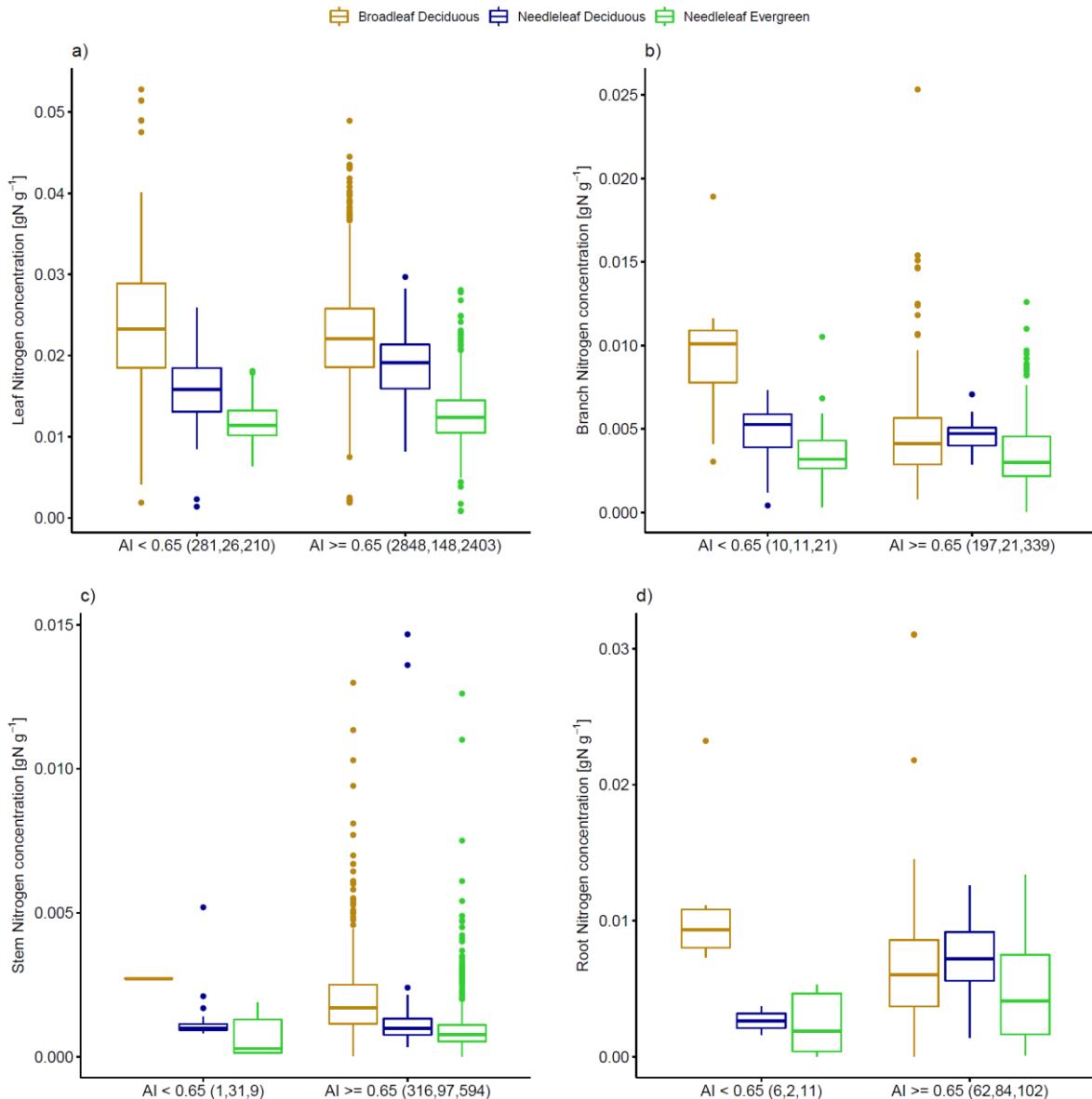


Fig. S2: The variation in a) leaf, b) branch, c) stem, and d) root N concentration for aridity index (AI) classes (AI < 0.65 vs. AI ≥ 0.65) and for leaf types (broadleaf deciduous, needleleaf deciduous, needleleaf evergreen) separately. The number of observations in each climatic class and for each leaf type is stated in brackets. The box-whisker plots show the median and the interquartile range of values. The whiskers extend up to the most extreme data point which is no more than 1.5 times the interquartile range away from the box. Outliers are drawn as points.

S10 Q-Q plots

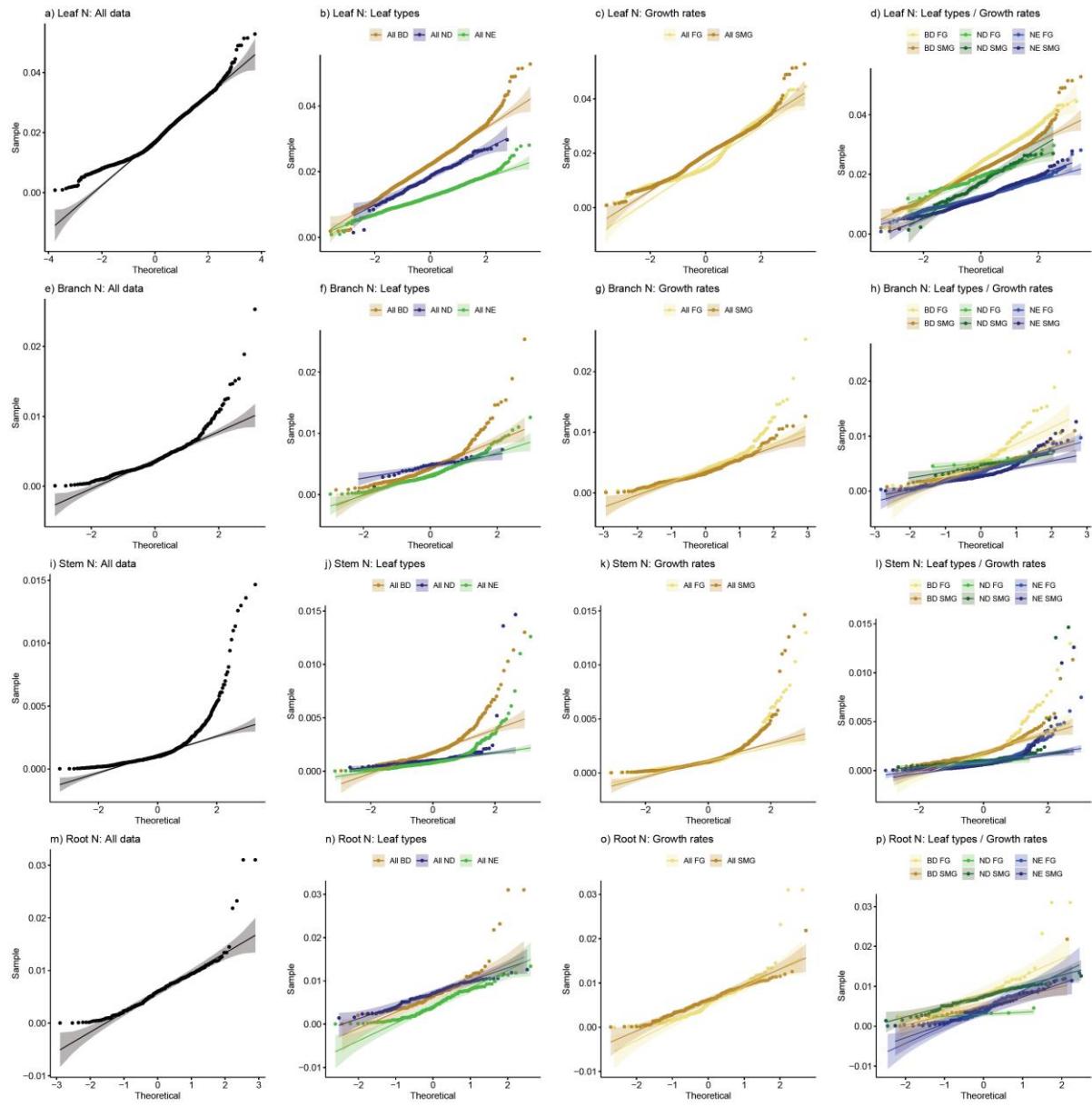


Fig. S3: Q-Q plots comparing the distributions of a-d) leaf, e-h) branch, i-l) stem, and m-p) root N concentration measurements with a standard normal distribution for leaf types and growth classes (BD = broadleaf deciduous, ND = needleleaf deciduous, NE = needleleaf evergreen, SMG = slow-/medium-growing, FG = fast-growing) separately. The straight line visualizes perfect normality, the shaded area shows 95 % confidence intervals.

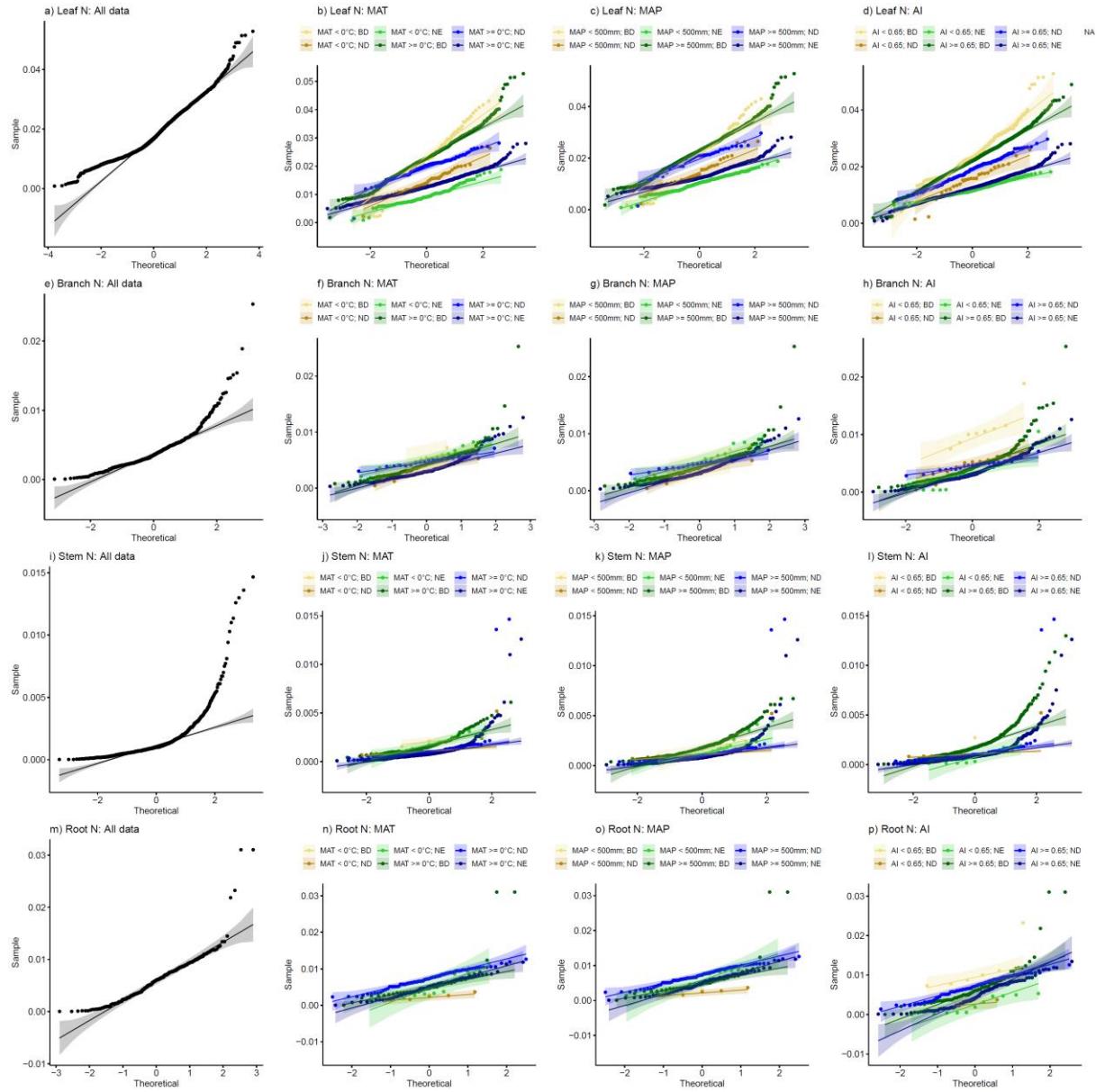


Fig. S4: Q-Q plots comparing the distributions of a-d) leaf, e-h) branch, i-l) stem, and m-p) root N concentration measurements with a standard normal distribution for mean annual temperature (MAT) classes (MAT < 0°C vs. MAT >= 0°C), mean annual precipitation sum (MAP) classes (MAP < 500mm vs. MAP >= 500mm), aridity index (AI) classes (AI < 0.65 vs. AI >= 0.65) and for leaf types (BD = broadleaf deciduous, ND = needleleaf deciduous, NE = needleleaf evergreen) separately. The straight line visualizes perfect normality, the shaded area shows 95 % confidence intervals.

S11 Residual plots

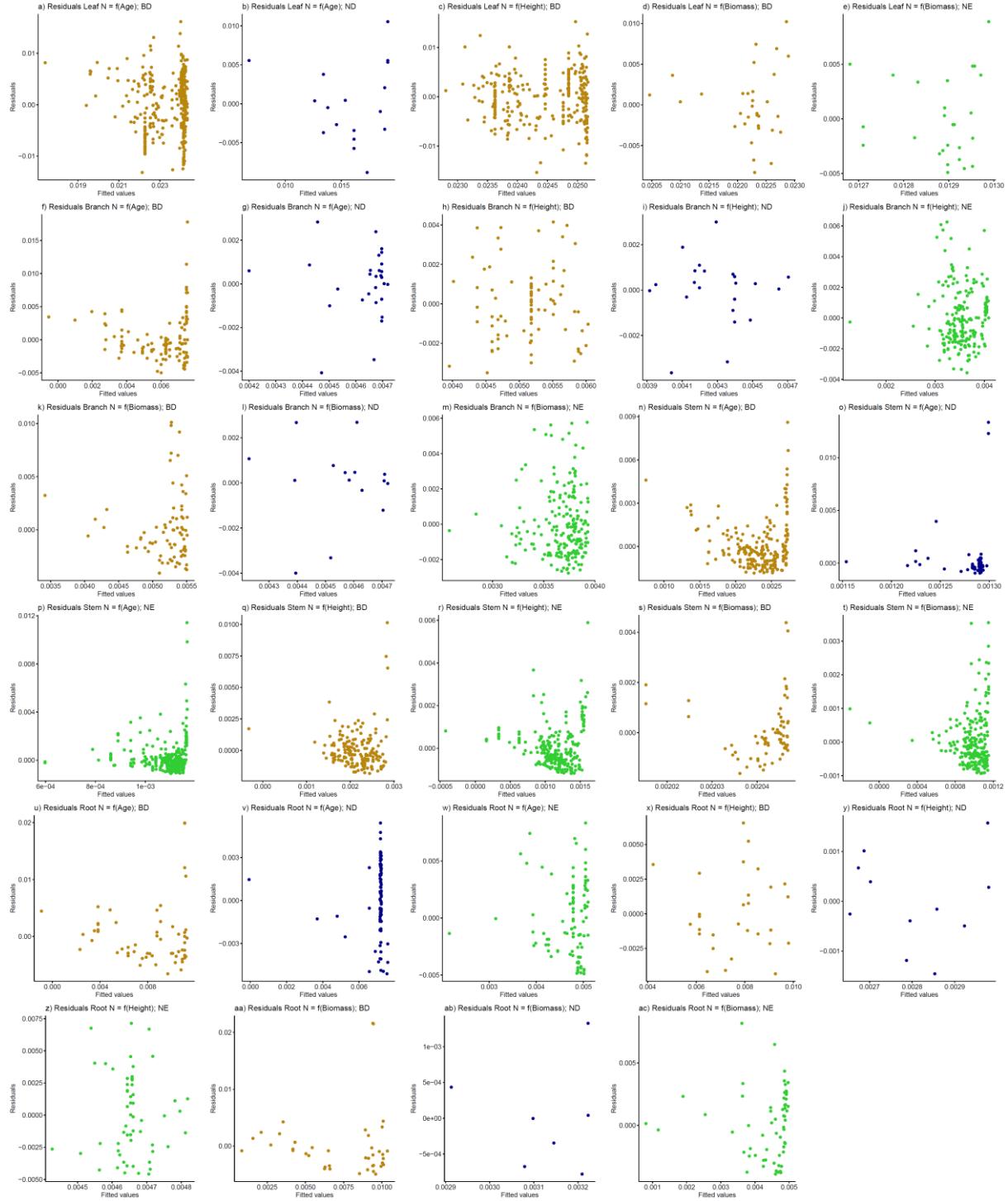


Fig. S5: Residuals from the fitted linear models (shown in Fig. 2) between a-e) leaf, f-m) branch, n-t) stem, and u-ac) root N concentration and tree age, tree height, and compartment biomass for leaf types (BD = broadleaf deciduous, ND = needleleaf deciduous, NE = needleleaf evergreen) separately.

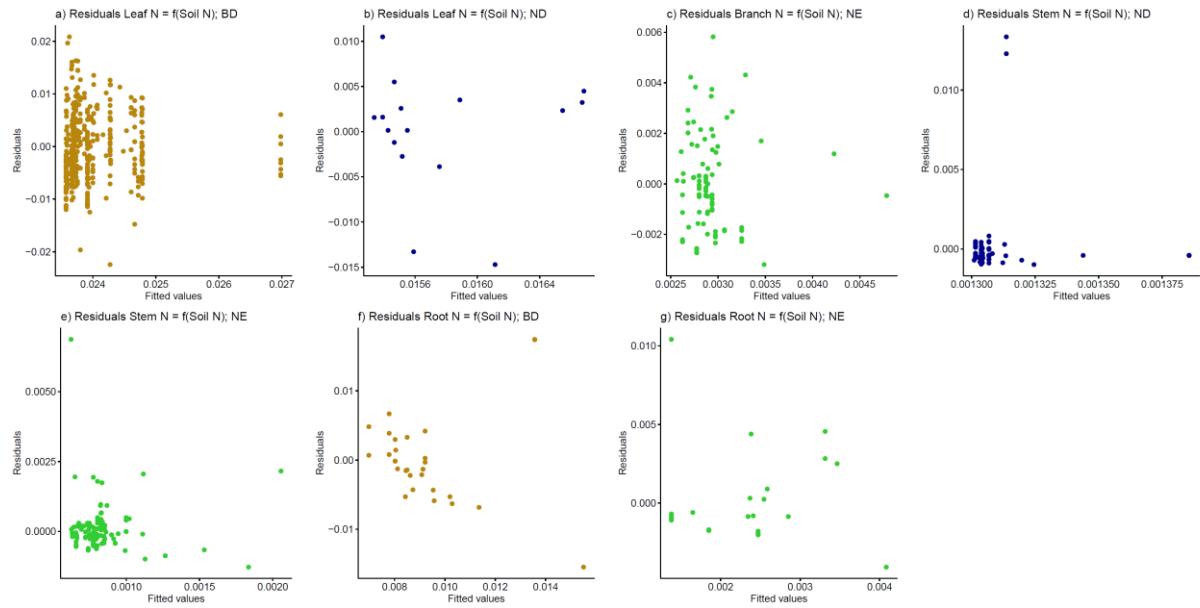


Fig. S6: Residuals from the fitted linear models (shown in Fig. 5) between a-b) leaf, c) branch, d-e) stem, and f-g) root N concentration and soil N concentration for leaf types (BD = broadleaf deciduous, ND = needleleaf deciduous, NE = needleleaf evergreen) separately.