

# 1 **Role of nitrogen and iron biogeochemical cycles on the production and export** 2 **of dissolved organic matter in agricultural headwater catchments**

3 Thibault Lambert<sup>1</sup>, Rémi Dupas<sup>1,\*</sup>, Patrick Durand<sup>1</sup>

4 <sup>1</sup> INRAE, UMR SAS 1069, L'Institut Agro, Rennes, France

5 \* Corresponding author

## 6 **Abstract**

7 **To better understand the seasonal variations in environmental conditions regulating**  
8 **dissolved organic matter (DOM) export in headwater catchments**, we combined monitoring of  
9 nitrate, iron, soluble phosphorus and DOM concentration (as dissolved organic carbon; DOC)  
10 and composition (3D fluorescence) in soil and stream waters at regular intervals during one  
11 hydrological year. We installed 17 zero-tension lysimeters in organic-rich top soil horizons  
12 (15 cm below the surface) in the riparian area of a well-monitored agricultural catchment in  
13 French Brittany and collected them at a fortnightly frequency from October 2022 to June  
14 2023. **We observed a large increase in DOC concentrations in soil waters** during the high  
15 flow period linked to the establishment of Fe-reducing conditions and the subsequent release  
16 of DOM. We also noted that the timing and the spatial variability in Fe(II) biodissolution in  
17 soils was regulated by nitrate from agricultural origin and the heterogeneity of water flow  
18 paths at the hillslope scale. Contrary to our current understanding of DOM export in  
19 headwater catchments, these results lead us to consider the winter high flow period as an  
20 active phase of both DOM production and export.

## 21 **1. Introduction**

22 Dissolved organic matter (DOM) is a key component of the ecological and biogeochemical  
23 functioning of aquatic ecosystems (Hanson et al., 2015), affecting for instance light  
24 penetration (Kelly et al., 2001), pollutant transport (Aiken et al., 2011), aquatic microbial  
25 metabolism (Wetzel, 1992), and the treatment of drinking waters (Chow et al., 2005). Aquatic  
26 DOM, which is mainly of terrestrial origin, represents a fundamental link between the  
27 terrestrial, oceanic, and atmospheric compartments of the global carbon cycle (Dean et al.,  
28 2020; Battin et al., 2008). Unravelling the sources and drivers of DOM export has become an  
29 urgent environmental issue in a context of long-term increasing concentrations of dissolved  
30 organic carbon (DOC, a proxy for DOM content) reported in numerous streams in the  
31 northern hemisphere (Monteith et al., 2007; De Wit et al., 2021).

32 **Numerous research carried out in temperate and boreal regions have shown that headwater**  
33 **catchments are the main entry point of DOM into fluvial networks (Ågren et al., 2007; Creed**

34 et al., 2015) and identified riparian areas as the dominant sources of DOM at the catchment  
35 scale owing to their location at the terrestrial-aquatic interface (Sanderman et al., 2009;  
36 Lambert et al., 2014; Laudon et al., 2012; Winterdahl et al., 2014). The flushing of shallow  
37 organic-rich soil layers during storm events (at the daily scale) typically represents the  
38 majority of annual DOC loads (Inamdar et al., 2006), and the DOC *versus* discharge  
39 relationships show that DOC export is transport-limited at the event scale (Buffam et al.,  
40 2001; Zarnetske et al., 2018). Although geomorphological and climatic conditions regulate  
41 DOC loads in aquatic ecosystems (Winterdahl et al., 2014; Laudon et al., 2012), DOC export  
42 at the annual scale is commonly conceptualized as a two-steps process in which DOM is  
43 produced and stored in the catchment during the hot and dry period, and then exported  
44 toward surface waters during the wet and cold period (Boyer et al., 1996). This two-steps  
45 conceptual model often described in temperate catchments (Deirmendjian et al., 2018;  
46 Strohmeier et al., 2020; Wen et al., 2020; Ruckhaus et al., 2023) is also supported by  
47 numerous studies carried out in tropical (Bouillon et al., 2014), boreal (Tiwari et al., 2022),  
48 Mediterranean (Butturini and Sabater, 2000) or Arctic fluvial networks (Neff et al., 2006).  
49 However the processes regulating the size of the riparian DOM pool remain unclear (Tank et  
50 al., 2018 and references below).

51 Antecedent soil conditions of wetness and temperature have been identified as a dominant  
52 control on stream DOC with concentrations typically increasing after dry events (Turgeon and  
53 Courchesne, 2008; Vázquez et al., 2007; Mehring et al., 2013). Periods of drought promote  
54 the production and accumulation of DOM in shallow soil horizons through enhanced soil  
55 organic matter decomposition (Harrison et al., 2008; Fenner and Freeman, 2011; Xu and  
56 Saiers, 2010), resulting in high stream DOC concentrations during the subsequent rewetting  
57 phase of the catchment (Werner et al., 2019; Raymond and Saiers, 2010). In good  
58 agreement with this conceptual model is the observation based on long-term data that the  
59 mean annual DOC concentrations in streams can be related to the intensity and duration of  
60 preceding dry periods (Humbert et al., 2015; Tiwari et al., 2022).

61 However, the establishment of reducing conditions in riparian soils during the winter may  
62 have potential implications on our conceptualization of stream DOC export owing to the  
63 influence of redox conditions on the iron (Fe) cycle in soils. While particulate Fe-hydroxides  
64 absorb organic substances with a high affinity when oxidizing conditions prevail, the  
65 microbially-driven dissolution of Fe oxyhydroxides during reducing conditions leads to the  
66 release of organic molecules previously bounded to surface minerals (Hagedorn et al., 2000;  
67 Blodau et al., 2008). The release of large amounts of DOM in riparian soils during the winter  
68 period – considered as non-productive in our current conceptualisation of stream DOC export  
69 – has been previously reported (Lambert et al., 2013; Lotfi-Kalahroodi et al., 2021), and

70 several studies have suggested that iron redox cycles may play a major role in catchment-  
71 scale DOC export (Knorr, 2013; Selle et al., 2019; Musolff et al., 2017). However, the onset  
72 of Fe reducing conditions and the subsequent DOM release could be limited in agricultural  
73 catchments owing to large inputs of nitrate (an oxidizing specie) from upslope via  
74 groundwater that may prevent Fe reductive biodissolution (Mcmahon and Chapelle, 2008;  
75 Christensen et al., 2000).

76 Because most of the studies investigating DOC export in headwater catchments rely on  
77 stream water monitoring, the processes regulating the size of the mobile DOM pool in  
78 riparian soils and the interaction with other biogeochemical cycles remain largely unknown.  
79 We still lack studies investigating how processes occurring in soil waters reflect our  
80 conceptualization of solutes dynamics based on observations made in surface waters (Knorr,  
81 2013; Dupas et al., 2015; Ledesma et al., 2015; Seibert et al., 2009; Sanderman et al., 2009;  
82 Lambert et al., 2013). In this study, we hypothesized that Fe biodissolution may significantly  
83 affect DOM release in riparian soils during the winter period with consequences on stream  
84 DOC export. We also investigated the potential influence of nitrate from agricultural origin,  
85 which may regulate Fe reduction. To this end, we installed zero-tension lysimeters in the  
86 riparian area of the Kervidy-Naizin catchment, whose stream waters are continuously  
87 monitored for water quality, including DOC at high frequency (Fovet et al., 2018). This  
88 catchment is located in Brittany (France), a region where stream DOC concentrations  
89 exhibited contrasting trends (increasing, decreasing or no trend) over the 2007-2020 period  
90 despite similar geomorphological and climatic conditions (Supplementary Fig. S1). The  
91 Kervidy-Naizin catchment for instance exhibits a weak but significant increase in stream  
92 DOC concentrations over the last two decades (Strohmenger et al., 2020). In this context,  
93 another goal of this study was to explore the hypothesis that long-term regional decrease in  
94 nitrate inputs (Abbott et al., 2018) have impacted long-term trends in DOC through iron  
95 dynamics in riparian soils. We monitored soil water chemistry during the 2022-2023  
96 hydrological year through measurements of DOC, Fe(II) and NO<sub>3</sub> concentrations but also  
97 DOM composition (absorbance and fluorescence properties coupled with parallel factor  
98 analysis) and soluble reactive phosphorus (SRP) as an additional tracer of Fe reductive  
99 dissolution (Gu et al., 2017; Smith et al., 2021). The results allowed us to decipher complex  
100 interactions among C, N, and Fe cycles in agricultural catchments and to highlight the  
101 occurrence of several processes sustaining DOM export during the winter period.

## 102 2. Material and method

### 103 2.1. Study site

104 The Kervidy-Naizin research observatory is a 4.9-km<sup>2</sup> agricultural headwater catchment  
105 located in Brittany (western France, Fig. 1). It belongs to the French Critical Zone  
106 Observatories (OZCAR) network and is instrumented since the 1970s for the long-term  
107 monitoring of the soil-atmosphere-hydrosphere continuum in a context of intensive  
108 agriculture (see Fovet et al., 2018 for a complete presentation of the study site).

109 The site is characterized by gentles slopes (<5%) and low elevation that ranges from 98–140  
110 m above sea level. The bedrock is composed of impermeable Brioverian schists above which  
111 a locally fractured layer of schists is underlain by 1 – 30 m of weathered material and silty  
112 loam soils. Soils are well drained except in riparian zones, where water excess leads to  
113 hydromorphic, poorly drained soil. Soil organic carbon content presents lateral (riparian  
114 versus upland soils) and vertical (surface versus deep soils) gradients, with highest values  
115 about 5.3 – 5.6 % in the uppermost soil horizons (0-20 cm depth) of the riparian area while  
116 soil organic content drop under 1% below 20 cm depth (Lambert et al., 2011).

117 The land use is intensive mixed farming, with 91% of the catchment area under agriculture  
118 that grows crops to feed a high density of dairy cattle, pigs and poultry. Maize (38%), straw  
119 cereals (30%), and grasslands (15%) dominate and wooded areas are mainly confined to  
120 valley bottoms along the stream channel or to some hedgerows (Fig. 1).

121 The climate is temperate oceanic, with mean annual temperature of  $11.2 \pm 0.6^\circ\text{C}$  and mean  
122 annual precipitation of  $810 \pm 180$  mm. Precipitation varies seasonally throughout the year,  
123 with higher precipitation from October to February (mean monthly precipitation of  $92 \pm 31$   
124 mm) and lower precipitation from March to July (mean monthly precipitation of  $50 \pm 14$  mm).  
125 The dynamics of the intermittent stream reflects the seasonal pattern of rainfall and  
126 evapotranspiration with high discharge periods from November to April and completely dry  
127 periods lasting one to three months between July to October depending on the hydrological  
128 year.

129 Groundwater level fluctuations are recorded every 15 min along the Kerolland (K) transect,  
130 rainfall is monitored at hourly intervals using a weather station located ~ 1400 m from the  
131 catchment outlet, and stream discharge is recorded every minute with an automatic gauge  
132 station at the outlet of the catchment. A S::SCAN probe is installed at the outlet of the  
133 catchment for the measurement of DOC and other variables at high-frequency (Fovet et al.,  
134 2018).

## 135 **2.2. Monitoring and manual sampling**

136 We investigated the seasonal variability in riparian DOM concentration and composition  
137 using zero-tension lysimeters designed to collect free soil waters (Supplementary Fig. S2)  
138 and installed in September 2022 in topsoil horizons (15 cm depth) in the Kerolland riparian

139 zone, an area known to be a major contributor to stream DOC export in this catchment  
140 (Lambert et al., 2014). We placed the lysimeters along three lines parallel to the stream  
141 channel, about 10-20 m apart from each other and from the stream, with the aim to capture  
142 the heterogeneity of water flow paths and nitrate concentration coming from the upslope  
143 cultivated fields. Lysimeters were all located in the hydromorphic soils unit according to the  
144 soil map (Fig. 1). We installed 29 zero-tension lysimeters, but some were lost during the  
145 study period because of damage by rodents. We kept lysimeters for which at least seven  
146 consecutive dates were available, resulting in 17 lysimeters used for the study. We collected  
147 soil waters from November 2022 to June 2023 at a weekly to fortnightly frequency depending  
148 on the hydro-climatic conditions (Fig. 2). The end of sampling was imposed by the lack of  
149 water in lysimeters owing to the gradual drawdown of the water table in the riparian zone  
150 during the spring period. We sampled soil waters with a vacuum pump and filtered them at  
151 0.2  $\mu\text{m}$  with acetate cellulose syringe encapsulated filters directly on site for all analyses  
152 including DOC,  $\text{NO}_3^-$ , SRP, Fe(II), and DOM composition (absorbance and fluorescence). We  
153 used unfiltered water samples to measure physico-chemistry variables including temperature  
154 and pH with an ODEON probe. In addition, we collected surface waters right next to the  
155 riparian area where lysimeters were located and at the outlet of the catchment. The  
156 laboratory analyses were identical for soil and surface waters.

### 157 **2.3. Analytical procedures**

158 With the exception of Fe(II) measurements that were performed the same day as sampling,  
159 all analyses were done within two weeks after sampling. Samples were stored in a 4°C cold  
160 room in the dark. Fe(II) analyses were determined using the 1,10-phenanthroline colorimetric  
161 method (Lambert et al., 2013): dissolved iron was trapped on site and the optical density of  
162 the complex formed with phenanthroline was measured the same day once back to the  
163 laboratory at 510 nm with an UV-vis spectrophotometer. DOC concentrations were measured  
164 using a total carbon analyzer (SHIMADZU TOC-V) with a precision estimated at  $\pm 5\%$  using  
165 a standard potassium hydrogen phthalate solution (SIGMA ALDRICH). Nitrate as  $\text{N-NO}_3^-$  and  
166 SRP were determined by spectrometry with an automatic sequential analyzer (SmartChem  
167 200, AMS Alliance, France).

168 Absorbance for colored DOM (CDOM) was measured with a Lambda 365 UV/vis  
169 spectrophotometer (Perkin Elmer) from 200 to 700 nm (1 nm increment) using a 1 cm quartz  
170 cuvette. Samples were diluted in most case due the DOM-rich nature of soil waters. The only  
171 purpose of CDOM spectra was to correct excitation-emission matrices (EEMs) for inner filter  
172 effects (Ohno, 2002). The dilution factor used for fluorescence measurements were applied  
173 to CDOM spectra. Fluorescence DOM (FDOM) was collected as EEMs with a Lambda LS45

174 (Perkin Elmer) using a 1 cm quartz cuvette across excitation wavelengths of 270 – 450 nm (5  
175 nm increment) and emission wavelengths of 290 – 600 nm (0.5 nm increment). Samples  
176 were diluted so absorbance at 254 nm was below 0.3 to reduce inner filter effects (Ohno,  
177 2002).

178 In our study, the Fe(II):DOC ratio was  $0.30 \pm 0.24$ , implying that significant interferences on  
179 DOM fluorescence from iron can be expected (Poulin et al., 2014). The degree of iron  
180 quenching, however, varies greatly between samples depending on the iron:DOC ratio  
181 (Pullin et al., 2007) but also on DOM composition (Jia et al., 2021; Poulin et al., 2014) and  
182 Fe(III) concentrations (Ohno et al., 2008), making difficult to predict the influence of Fe on  
183 EEMs. That being said, quenching was clearly apparent in some samples ( $n < 10$ ) that  
184 showed the fluorescence intensity to increase with dilution factor, reflecting the influence of  
185 high level of Fe that reduces DOM fluorescence (Pullin et al., 2007). The quenching  
186 impacted EEMs at low ( $< 270$  nm) and moderate to high (420 – 490 nm) excitation and  
187 emission wavelengths, respectively, which is consistent with previous studies concluding that  
188 Fe mainly impacts fluorescence intensity in EEM locations associated with humic-like  
189 fluorophores, namely A and C peaks (Jia et al., 2021; Poulin et al., 2014). Thus, although we  
190 cannot rule out an effect of iron on EEMs, this would have impacted the relative contribution  
191 of humic-like fluorophores associated with C1 and C2 components of our model (see below)  
192 who behaved similarly between clusters and across seasons.

#### 193 2.4. PARAFAC modelling

194 EEMs preprocessing (Raman scattering removal and standardization to Raman units) was  
195 performed prior to the PARAFAC modeling. Normalization was done using a Milli-Q water  
196 sample run the same day as the sample. A five-component PARAFAC model was obtained  
197 using the drEEM 0.3.0 Toolbox (Murphy et al., 2013) for MATLAB (MathWorks, Natick, MA,  
198 USA). Split-half analysis, random initialization, and visualization of residuals EEMs were  
199 used to test and validate the model. The positions of maximum peaks of the PARAFAC  
200 components were compared to previous studies carried out in similar context of human-  
201 impacted catchments with the open fluorescence database OpenFluor using the OpenFluor  
202 add-on for the open-source chromatography software OpenChrom (Murphy et al., 2014). The  
203 maximum fluorescence  $F_{\text{Max}}$  values of each component for a particular sample provided by  
204 the model were summed to calculate the total fluorescence signal  $F_{\text{Tot}}$  of the sample in  
205 Raman units. The relative abundance of any particular PARAFAC component X was then  
206 calculated as  $\%C_X = F_{\text{Max}}(X)/F_{\text{Tot}}$ .

#### 207 2.5 Statistical Analyses

208 A principal component analysis (PCA) coupled to a clustering analysis was used to  
209 discriminate and group lysimeters based on the presence or absence of iron biodissolution in  
210 soil waters. The aim was to help visualize temporal pattern for each of the two clusters rather  
211 than 17 time series if data were plotted for each lysimeter. For this reason, data (DOC, NO<sub>3</sub>,  
212 SRP and Fe(II) concentrations and the relative contribution of PARAFAC components) were  
213 averaged for each lysimeters then normalized. The PCA was performed using the *prcomp*  
214 function in the R software, and the *factoextra* package was used to identify the variables that  
215 contribute the most to the first two dimensions of the PCA. The cluster analysis, based on the  
216 results from the PCA and called Hierarchical Clustering on Principal Components (Josse,  
217 2010), was performed with the *FactoMineR* package for R (Lê et al., 2008). Relationships  
218 between variables were investigated either through Pearson or Spearman correlations  
219 depending of the nature (linear or not) of the correlations.

### 220 3. Results

#### 221 3.1. Hydro-climatic context

222 The hydrological regime of the study site is characterized by a succession of three distinct  
223 periods determined by water table fluctuations along the hillslope, corresponding to different  
224 hydrological regimes for the riparian soils (Fig. 2; Lambert et al., 2013): (i) a period of  
225 progressive rewetting of riparian soils after the dry season and of low groundwater flow and  
226 low stream discharge (01/09/2022 – 18/12/2022, mean and cumulated precipitation =  $5.1 \pm 5.3$   
227 mm d<sup>-1</sup> and 338.5 mm, respectively); (ii) a period of prolonged waterlogging of riparian soils  
228 induced by the rise of the water table in the upland domain, corresponding to high values of  
229 hillslope groundwater flow and stream discharge (18/12/2022 – 9/05/2023, mean and  
230 cumulated precipitation =  $6.8 \pm 7.9$  mm d<sup>-1</sup> and 573 mm, respectively); and (iii) a period of  
231 drainage and progressive drying of the riparian soils induced by the drawdown first in the  
232 upland domain then in the bottomland domain and corresponding to the decrease of both the  
233 hillslope groundwater flow and stream discharge (09/05/2023 – 01/07/2023, mean and  
234 cumulated precipitation =  $4.3 \pm 4.4$  mm d<sup>-1</sup> and 42.5 mm, respectively). Air temperature (Fig.  
235 2C) showed a smoothed seasonal variability with decreasing values from September to  
236 December (from ~20°C to -2°C) followed by a rise in temperature from 0°C to 20°C from  
237 February to July. This pattern was only interrupted by a relatively short episode of higher  
238 temperature (close to 10°C) during the winter, coinciding with the first intense rainfall period  
239 of the year.

#### 240 3.2. Fluorescence properties of DOM

241 Five PARAFAC components were identified in soil waters (Supplementary Fig. S3), all of  
242 which already described in previous studies. All five components had humic-like fluorescence

243 properties (Fellman et al., 2010). Components C1 (excitation/emission peaks = 350 nm /444  
244 nm), C2 (<270/450), and C5 (410/488) predominantly cover the regions of EEMs associated  
245 with peaks A and C and are common tracers of terrestrially-derived DOM in surface waters  
246 (Kothawala et al., 2015; Stedmon and Markager, 2005; Logozzo et al., 2023; Lambert et al.,  
247 2017) while C3 (330/406) and C4 (295/410) are both located near the classical peak M,  
248 indicating a microbial transformation of terrestrial DOM (Williams et al., 2010; Lambert et al.,  
249 2022; Yamashita et al., 2010). The maximum fluorescence intensity of all components were  
250 strongly related to DOC concentrations (not shown) and the relative contribution of each  
251 component decreased from as C1 ( $29.7 \pm 3.1$  %) > C2 ( $28.3 \pm 3.6$  %) > C3 ( $19.5 \pm 2.5$  %) > C4  
252 ( $12.9 \pm 6.6$  %) > C5 ( $9.7 \pm 2.1$  %).

### 253 3.3. Seasonal variations in soil and stream waters

254 Temperature in soil waters (Fig. 3A) followed the same pattern as air temperature: values  
255 oscillated between 5°C and 15°C during November – January, reached minimums between 4  
256 and 7 °C in January – March and then increased gradually during the end of the study period  
257 up to 18 – 20 °C in June. pH varied between 6.2 and 7.4 (mean  $6.9 \pm 0.3$ ) across lysimeters  
258 and didn't exhibit significant trends over the study period (Fig. 3B). Solutes, however,  
259 exhibited complex patterns with a high variability across lysimeters and time, especially  
260 during the high flow period (Fig. 3C-F). **Despite the fact that lysimeters were installed along**  
261 **three lines ranging 10-30 m from the stream, no spatial pattern was identified.** Overall, these  
262 elements were strongly linked to each other (Fig. 4). DOC concentrations ranged from 2.3 to  
263  $87.4 \text{ mg L}^{-1}$  (mean =  $30.2 \pm 12.8 \text{ mg L}^{-1}$ ) over the study period and were linearly and positively  
264 **(Pearson  $r = 0.73$ ,  $p$  value < 0.0001)** associated with Fe(II) that ranged from 0 to  $45.8 \text{ mg L}^{-1}$   
265 (mean =  $9.8 \pm 7.6 \text{ mg L}^{-1}$ ). Fe(II) was negatively **(Spearman  $r = -0.56$ ,  $p$  value < 0.0001)**  
266 correlated with  $\text{NO}_3$  (from 0 to  $16.4 \text{ mg L}^{-1}$ , mean =  $0.9 \pm 1.1 \text{ mg L}^{-1}$ ), and SRP (from 0 to  $0.5$   
267  $\text{mg L}^{-1}$ , mean =  $0.1 \pm 0.1 \text{ mg L}^{-1}$ ) was also positively **(Pearson  $r = 0.21$ ,  $p$  value = 0.0005)**  
268 related to Fe(II), but not as strongly as for DOC.

269 **DOC concentrations in stream waters varied from  $2.9$  to  $36.8 \text{ mg L}^{-1}$  during the study period**  
270 **(Fig. 5). Maximum concentrations were reached during storm events due to a rapid response**  
271 **to rainfall and the mobilisation of riparian wetland waters (Durand and Juan Torres, 1996).**  
272 **There was a tendency for minimum (at base flow) and maximum (at peak discharge)**  
273 **concentrations to decrease from November to March. From March to July, however, minimal**  
274 **concentrations remained stable while maximum values showed a slight increasing trend.**

### 275 3.4. Clustering of soil waters

276 The first two components of the PCA explained 69.4 % of the total variance of the data and  
277 discriminated lysimeters depending on the presence or absence of Fe(II) biodissolution in



278 soil waters of the riparian area (Fig. 6). The first principal component (PC1, 54% of the total  
279 variance) was mainly related to NO<sub>3</sub> concentrations and terrestrial humic-like components  
280 (C1, C2, and C5) on positive scores, and to DOC and Fe(II) concentrations and the microbial  
281 humic-like component C4 on negative scores. The second component (PC2, 15.4% of the  
282 total variance) was related to SRP (positive score) and the component C3 (negative score).  
283 PARAFAC components had similar or even higher scores than DOC, Fe(II), and NO<sub>3</sub>  
284 concentrations on the two first dimensions of the PCA (Supplementary Fig. S4), illustrating  
285 the importance of DOM composition as an important factor contributing to explain the spatial  
286 variability across lysimeters. The distribution of PARAFAC components along the first  
287 dimension reflects the relationships between their relative contribution and Fe(II),  
288 concentrations (not shown). More specifically, %C4 was strongly and positively correlated  
289 with Fe(II) ( $R^2 = 0.38$ , Pearson  $r = 0.62$ ) compared to other components that exhibited  
290 weakest and negative relationships with Fe(II) ( $R^2$  from 0.09 to 0.19, Pearson  $r$  from -0.30 to  
291 -0.43). In other words, lysimeters capturing Fe biodissolution in the riparian area were  
292 associated with high DOC and a greater proportion of the microbial C4 component compared  
293 to lysimeters enriched in nitrate where no Fe(II) was measured.

294 The hierarchical clustering based on the PCA results grouped the lysimeters in two distinct  
295 clusters based on the presence (cluster 1) or absence (cluster 2) of Fe(II) (Fig. 6). This  
296 approach allowed us to gain insight into the temporal evolution of solutes in soil waters since  
297 clear patterns appeared once the data were grouped by cluster (Fig. 7). In cluster 1, DOC, N-  
298 NO<sub>3</sub> and SRP decreased from  $39.8 \pm 13.3$  to  $23.4 \pm 8.4$  mg L<sup>-1</sup>, from  $2.6 \pm 3.6$  to  $1.2 \pm 1.8$  mg L<sup>-1</sup>,  
299 and from  $0.18 \pm 0.18$  to  $0.08 \pm 0.15$  mg L<sup>-1</sup>, respectively, during the rewetting phase of the  
300 catchment while Fe(II) was not measured at significant levels. During the high flow period,  
301 however, Fe(II) increased gradually from  $3.7 \pm 3.2$  to  $26.5 \pm 7.8$  mg L<sup>-1</sup>, and both DOC and SRP  
302 followed a similar trend with concentrations raising from  $27.3 \pm 9.5$  to  $54.9 \pm 25.0$  mg L<sup>-1</sup> and  
303 from  $0.07 \pm 0.13$  to  $0.18 \pm 0.11$  mg L<sup>-1</sup>, respectively. During this period and until the end of the  
304 hydrological cycle, N-NO<sub>3</sub> were very low, decreasing from  $0.54 \pm 0.66$  mg L<sup>-1</sup> at the beginning  
305 of the high flow period to values below 0.15 mg L<sup>-1</sup> the rest of the survey. The start of the  
306 third hydrological period corresponding to the drawdown of the water table and the  
307 consecutive aeration of riparian soils was marked by the rapid drop of Fe(II) at  $8.1 \pm 7.4$  mg L<sup>-1</sup>,  
308 DOC at  $17.5 \pm 10.9$  mg L<sup>-1</sup>, and SRP at  $0.02 \pm 0.02$  mg L<sup>-1</sup>.

309 Similarly to cluster 1, soil waters from the cluster 2 exhibited a decline in DOC and SRP  
310 concentrations during the rewetting phase of the catchment but these trends continued  
311 during the high flow period, with minimal values reached in the middle of February. Thus,  
312 DOC dropped from  $34.5 \pm 7.1$  to  $9.4 \pm 3.1$  mg L<sup>-1</sup> and SRP from  $0.19 \pm 0.08$  to  $0.02 \pm 0.01$  mg L<sup>-1</sup>  
313 during this period, before showing an increasing trend to reach concentrations about

314 21.0±6.1 mg L<sup>-1</sup> for DOC and 0.16±0.13 mg L<sup>-1</sup> for SRP at the end of the high flow period.  
315 DOC remained elevated (24.1±3.1 mg L<sup>-1</sup>) at the start of the dry period, but SRP dropped  
316 close to depletion. In contrast, N-NO<sub>3</sub> first increased from 0.57±0.81 mg L<sup>-1</sup> in November to  
317 maximum values of 6.5±5.9 mg L<sup>-1</sup> in the middle of March, and then exhibited decreasing  
318 concentrations until a complete depletion at the beginning of the third hydrological period.  
319 Contrary to cluster 1, Fe(II) was not measured at significant concentrations in cluster 2 (*i.e.*  
320 below 0.5 mg L<sup>-1</sup>) except in March, during which Fe(II) increased from 1.2±1.9 to 4.1±0.2 mg  
321 L<sup>-1</sup>.

## 322 4. Discussion

### 323 4.1. The buffering effect of nitrate on iron reductive dissolution

324 The reductive biodissolution of iron during the high-water winter period is a recurrent process  
325 in riparian soils of headwater catchments (Smolders et al., 2017; Knorr, 2013; Selle et al.,  
326 2019). The magnitude of variations in Fe(II) and associated DOC and SRP dynamics  
327 reported in this study are in line with previous works conducted in the same research  
328 catchment (Lambert et al., 2013; Lotfi-Kalahroodi et al., 2021; Gu et al., 2017). In addition,  
329 our results evidenced a marked variability in the intensity of iron dissolution across lysimeters  
330 that we attributed to the spatial distribution of NO<sub>3</sub>-rich water flow paths that can inhibit and  
331 delay the release of Fe(II) and DOC in soil waters.

332 A fundamental condition for the establishment of reductive conditions is the prolonged  
333 waterlogging of riparian soils. As shown earlier for this and other lowland catchments on  
334 impervious bedrock, the increase of the hydraulic gradient induced by the rise of  
335 groundwater in the upland domain during the high flow period maintains a strong hydrologic  
336 connection between upland and riparian domains (Pacific et al., 2010; Molenat et al., 2008).  
337 Under these conditions, riparian soils remain waterlogged owing to a high and continuous  
338 hillslope groundwater flow, leading to the gradual establishment of reductive conditions and  
339 the subsequent triggering of Fe-biodissolution as long as inputs of oxidizing species  
340 remained limited and/or counterbalanced by higher rate of consumption through microbial  
341 activity (Lotfi-Kalahroodi et al., 2021; Lambert et al., 2013). This pattern is well illustrated by  
342 data from lysimeters of the first cluster (Fig. 7). After a quick depletion of an initial stock of  
343 nitrate accumulated during the previous summer, reductive conditions were rapidly  
344 established at the beginning of the high flow period and increasing Fe(II) concentrations in  
345 soil waters lead to the onset of the reductive Fe biodissolution in riparian soils. The gradual  
346 increase in Fe(II) during all the high flow period despite variations in temperature or rainfall  
347 patterns (with some intense precipitation events > 20 mm d<sup>-1</sup>) suggests a limited impact of  
348 these climatic episodes, except during a period of low precipitation during which both Fe(II)

349 and DOC exhibited a slight decrease in February/March. We attributed this small drop to the  
350 drawdown of the water table in upland groundwater flow following a prolonged absence of  
351 precipitations (see PK3 fluctuations, Fig. 2) that may have re-oxygenated soil waters (as no  
352 changes in N-NO<sub>3</sub> occurred).

353 Therefore, large release of DOC occurred in soils of the first cluster. Iron biodissolution also  
354 affected SRP, but the relationships was weaker suggesting that the reductive dissolution of  
355 soil Fe was not the primary driver of SRP concentrations in soils. For instance, soil  
356 properties, and more specifically soil phosphorus content and speciation, have been shown  
357 to strongly regulate SRP in soil waters of the Kervidy-Naizin catchment (Gu et al., 2017).  
358 Regarding DOC, the mean DOC:Fe(II) molar ratio was 142.4±285.5. This was higher than  
359 the DOC:Fe(II) ratio measured in experimental conditions (74.5±74.6) but similar to value  
360 measured on the field (134.4±25.6) by Lotfi-Kalahroodi et al. (2021) who aimed to investigate  
361 Fe reduction in the riparian area of our study catchment. Fe(III) concentrations in soil waters  
362 were not measured, but, based on the work of Lotfi-Kalahroodi et al. (2021), we can estimate  
363 a ratio between total Fe and Fe(II) of 4.8. Keeping in mind that this is a rough estimation, our  
364 mean DOC:Fe ratio would be about 29.3±58.8, which is consistent with previous studies (e.g.  
365 Selle et al., 2019; Musolff et al., 2017; Grybos et al., 2009; Cabezas et al., 2013). The nature  
366 of processes releasing DOC upon the reduction of soil-Fe oxyhydroxides in riparian soils of  
367 our study site has been studied in laboratory conditions (Grybos et al., 2009). Results have  
368 shown that up to 60% of the release is due to DOC desorption caused by the pH increase  
369 that accompanies the reduction of Feoxyhydroxides in these soils, the remaining 40% being  
370 due to the dissolution of Fe-oxyhydroxides that strongly adsorb organic compounds  
371 previously bounded to surface minerals (e.g. Hagedorn et al., 2000). In good agreement with  
372 these results, soil DOC was positively related to pH (Supplementary Fig. S5). The abrupt  
373 decrease in DOC in June illustrates the restoration of aerobic conditions owing to the  
374 drawdown of the water table in the bottomland domain led to the formation of Fe-minerals  
375 and the subsequent retention of DOC and SRP (Gu et al., 2017).

376 Lysimeters from the second cluster showed a very different pattern. Although some of them  
377 were located close (3-4 m) to lysimeters in which reducing conditions prevailed, there was no  
378 evidence of Fe(II) release, arguably because of the presence of nitrate. Indeed, and in  
379 agreement with studies carried out in wetland (Lucassen et al., 2004) and lacustrine  
380 (Andersen, 1982) sediments, we argue that the Fe-biodissolution biodissolution was inhibited  
381 as long as long as NO<sub>3</sub> remained in sufficient quantity in soil waters. In the absence of such  
382 production or regeneration process, both DOC and SRP showed a net depletion pattern from  
383 November to March. The influence of nitrate as a buffer of Fe-biodissolution was furthermore  
384 supported by the observation of a slight release of Fe(II) in May, at a moment when nitrate

385 became depleted from soil waters, probably because of plant uptake. Interestingly, we found  
386 that the threshold value of nitrate above which the process is activated (based on the  $\text{NO}_3$   
387 *versus* Fe(II) relationship (Fig. 4) as well as timing of Fe-biodissolution identified in cluster 1  
388 and cluster 2) ranged between 1.2 and 1.8 N- $\text{NO}_3$  ( $4.1 - 6.2 \text{ mg L}^{-1}$ ), which is close to the  
389 threshold value of  $6 \text{ mg L}^{-1}$  established at the catchment scale by Musolff et al. (2017) in  
390 German streams.

391 The PARAFAC components identified in the model suggest a dominance of highly aromatic  
392 and conjugated molecules across all lysimeters and dates, which is typical of DOM derived  
393 from soil organic matter and found in poorly drained soils in riparian or wetland areas  
394 (Sanderman et al., 2009; Lambert et al., 2013; Yamashita et al., 2010). The larger proportion  
395 of C4 in the first cluster however indicates that the Fe oxyhydroxides reduction leads to  
396 greater proportion of microbially-derived compounds within the DOM pool. In agreement with  
397 previous studies showing that the Fe(III) reduction could enhance the decomposition of  
398 organic matter in soils (Chen et al., 2020; Kappler et al., 2021), the close link between Fe(II)  
399 and C4 likely reflects an indirect effect of Fe biodissolution promoting the degradation of soil  
400 OM and the subsequent incorporation of microbially-derived compounds into the DOM pool  
401 (Dong et al., 2023). This hypothesis is well consistent with previous experimental studies  
402 performed with soils from the Kervidy-Naizin riparian area, which showed that bacterial  
403 reduction of Fe(III)-oxides to Fe(II) was concomitant with the release of large biological  
404 organic by-products upon the growth of bacterial communities (Lotfi-Kalahroodi et al., 2021).

405 Our study evidences a strong spatial heterogeneity of the establishment of reducing  
406 conditions in the riparian area of the Kervidy-Naizin catchment, associated with differences in  
407 the composition of DOM released in soil waters. It remain to be determined, however, the  
408 reason for such variability in biogeochemical processes in riparian soils. A first explanation  
409 can be related to the heterogeneity in water flowpaths in soils. In intensive agricultural  
410 catchments such as our study site, inflow of  $\text{NO}_3$ -rich water may arise from the rise of  
411 contaminated groundwater in valley bottom s and/or from subsurface flow paths that connect  
412 upland soils to riparian soils (Molenat et al., 2008). It is likely that lysimeters from the second  
413 cluster captured preferential flow paths of  $\text{NO}_3$ -rich waters while lysimeters from the first  
414 cluster were disconnected from those preferential water circulations. Alternatively, the  
415 absence of nitrate in soil waters may arise from a higher rate of denitrification that  
416 counterbalanced  $\text{NO}_3$  inputs. Research based on field observation remained limited to  
417 decipher the respective role of hydrology *versus* biogeochemistry in controlling Fe(II)  
418 biodissolution in riparian soils, and experimental studies would be required to provide more  
419 quantitative values on these potential drivers and their interactions.

#### 4.2. Implication for stream DOM export at the catchment scale

The current understanding of DOM export in headwater catchments is based on a two-steps conceptual model, in which a pool of mobile DOM is built in soils during the dry season and then flushed towards surface waters during the following wet season (e.g. Tiwari et al., 2022; Ruckhaus et al., 2023; Strohmenger et al., 2020; Raymond and Saiers, 2010). However, the high-frequency measurements of DOC in the stream do not fully support this statement. The establishment of a hydrological connection between riparian soils and the stream during the winter period showed the stream DOC to gradually decrease both at peak discharge during successive storm events and at base flow during inter-storm periods (Figure 5). This pattern, which repeats every year in this catchment (Strohmenger et al., 2020), is well consistent with the hypothesis of the mobilisation and exhaustion of a DOM pool limited in size built during the summer period (Humbert et al., 2015). However, stream DOC were found to increase slightly in March/April after the low-flow period that showed the hydrological connection between soils and the stream to decrease. It is unlikely that the mobilisation of an additional pool of DOM from upland soils may explain this small raises in stream DOC because this pool is 1) relatively small in terms of size, and 2) quickly exhausted at the beginning of the winter period (Lambert et al., 2014). Therefore, the seasonal pattern of stream DOC likely reflects the regeneration of the riparian DOM pool during the winter period as shown by our data collected in soil waters of riparian wetlands.

Stable carbon isotopes have indeed demonstrated that riparian soils of the Kervidy-Naizin catchment – and more particularly the DOM-rich uppermost soil horizons – are the dominant source of stream DOC at the catchment scale (Lambert et al., 2014), a feature commonly shared by headwater catchments (e.g. Sanderman et al., 2009). Thus, the decline in DOC and SRP observed in soil waters, particularly in the second cluster whereby these elements became almost depleted (Fig. 7), was consistent with the general flushing behaviour of the catchment shown by stream DOC from November to February. Similarly, the large two to three fold increase in DOC concentrations in riparian soils (in cluster 1 and 2, respectively) denotes a large mobilisation of DOM between March and May despite wet and low temperature conditions, that could explain in turn the pattern observed in stream DOC at the same time. While part of this regeneration can be attributed to iron biodissolution, the release of large amount of DOC the cluster 2 where the reductive biodissolution of Fe(III) was limited implies that another production mechanisms contributed to release DOM in riparian soils. It is unlikely that agricultural inputs (crop residues, manure application, etc) main may explain the increases in the riparian area, as these sources are episodic and/or size-limited (Lambert et al., 2014; Humbert et al., 2015; Pacific et al., 2010). This observation echoes previous works on the Kervidy-Naizin catchment showing effective inter-annual regeneration mechanisms of

456 the pool of soluble phosphorus in soils unrelated to iron dynamics (Gu et al., 2017), a  
457 statement supported here by the fact that SRP concentrations followed a similar pattern as  
458 DOC in soils grouped in the second cluster (Fig. 7).

459 The PARAFAC results suggest that DOM mobilized from soil to streams is only composed by  
460 aromatic molecules of high molecular weight. Although complex organic molecules indeed  
461 dominate stream DOM export (Fellman et al., 2009), it should be noted however that protein-  
462 like components are commonly found in stream waters (Inamdar et al., 2012), including in  
463 our study site (Humbert et al., 2020). The lack of such components in our model results from  
464 our sampling approach and not from their absence in catchment soils. Indeed, the production  
465 of protein-like components in catchment soils is restricted to the summer hot and dry period  
466 during which a pool made of low-aromatic and microbially-derived compounds built up in  
467 riparian soils (Lambert et al., 2013). However, this DOM pool is quickly flushed and  
468 exhausted during the rewetting phase in October-November, and soil DOM during the winter  
469 period is mainly composed by highly-aromatic molecules originating from soil organic  
470 material (Lambert et al., 2014). Agricultural practices such as fertilizer applications can  
471 represent another source of protein-like DOM in the catchment (Humbert et al., 2020), but  
472 these inputs remain episodic with a low impact on DOM at the catchment scale (Humbert et  
473 al., 2015; Lambert et al., 2014). For instance, a recent one-year of monitoring of soil waters  
474 at different locations in the catchment has shown that protein-like components represent only  
475  $3.44 \pm 2.8\%$  of the total fluorescence signal in catchment soils, this contribution being  
476 particularly low in riparian areas (Humbert et al., 2020). Therefore, the absence of protein-  
477 like components in our PARAFAC model is the consequence of our sampling design that  
478 focused on DOM production mechanisms in riparian soils (distant from agricultural inputs)  
479 during the winter period (period of production of highly aromatic compounds in soils).

480 Taking together, our results have two important implications regarding our conceptualisation  
481 of DOM export in headwater catchments. First, it challenges the idea that the wet period acts  
482 solely as a passive export period for DOC, with no or little DOC production (Strohmenger et  
483 al., 2020; Ruckhaus et al., 2023; Wen et al., 2020). Second, it emphasizes that stream DOC  
484 dynamics at the outlet is an integrative signal, potentially masking the high spatial  
485 heterogeneity of the system owing to complex interactions between biogeochemical cycles in  
486 soils, nutrient transfer at the soil/stream interface and hydrological functioning of catchments.  
487 While the patterns of stream DOC were consistent with that observed in soils, our study  
488 remains however limited in its capacity to quantify the relative contribution of the cluster  
489 identified to stream DOC export. Additionally, we do not have the necessary data such as  
490 isotopes or molecular markers to elucidate the precise origin and DOM (and SRP) release in

491 soils unrelated to iron biodissolution, and this should be the focus of future work combining  
492 experimental and field studies.

### 493 **Conclusion**

494 The combined monitoring of soil and stream waters in a temperate headwater catchment  
495 allowed us to evidence the dual role of high flow period as both an active phase of DOC  
496 production and export. In agreement with previous studies (e.g. Selle et al., 2019; Knorr,  
497 2013), the establishment of Fe-reducing conditions in riparian areas was identified as a major  
498 mechanism for the release of large amount of DOM in soil waters. In agricultural catchments,  
499 however, we found that this process can be buffered by nitrate, leading to a strong spatial  
500 heterogeneity in the magnitude of iron biodissolution and its consequences on soil DOC  
501 dynamics. Our study also evidenced that another production mechanisms unrelated to Fe  
502 dynamics contributed to release DOM in riparian soils during the winter period, pointing to  
503 the need to further investigate stream DOC export at the soil/stream interface.

504 The interactions between the N and Fe biogeochemical cycles may have potential  
505 implications regarding long-term increases in DOC in streams of Brittany. Indeed, stream  
506 DOC in the Kervidy-Naizin catchment has been slowly but significantly increasing in the last  
507 two decades, and this trend is mirrored by a decline in NO<sub>3</sub> concentrations (Strohmenger et  
508 al., 2020). While part of the DOC trend can be related to changes in climatic conditions as  
509 winters tend to wetter over the years (Strohmenger et al., 2020), the long-term decline in N  
510 inputs from agriculture may have favoured the increase in stream DOC by enhancing Fe(II)  
511 biodissolution in riparian soils. This hypothesis could partly explain why catchments having  
512 similar geomorphological and climatic properties present contrasting long-term trends at the  
513 scale of the Brittany region (Supplementary Fig. S1). Indeed, nitrate concentrations have  
514 largely decreased during the last decades, but the rate of recovery is not uniform across the  
515 region (Abbott et al., 2018). Studies carried out at the regional scale aiming to decipher the  
516 interactions between local (agricultural practices) and global (climatic conditions) and the  
517 consequences on stream DOC export would be critical considering the influence of DOM on  
518 water quality and on the ecological and biogeochemical functioning of surface waters.

### 519 **Data availability**

520 Data on soil waters will be published on Zenodo.org upon the reservation that the paper will  
521 be accepted for publication. Hydrological and climatic data from the Kervidy-Naizin site are  
522 available here: [https://geosass.fr/web/?page\\_id=103](https://geosass.fr/web/?page_id=103).

### 523 **Acknowledgements**

524 We thank Militza G., Harald F., P. Petitjean and Celine B. for their assistance in field and lab  
525 work.

## 526 **Financial support**

527 This study has received funding from the H2020 European Research Council under the Marie  
528 Skłodowska-Curie grant agreement COSTREAM No 101064945.

## 529 **Author contribution**

530 TL conceived the study. TL defined protocols with contribution from RD and PD. TL collected  
531 field samples with help from RD. TL made laboratory analysis. TL analysed the data and  
532 drafted the manuscript with inputs from RD and PD. All authors contributed and approved to  
533 the manuscript.

## 534 **Competing interests**

535 The authors declare that they have no conflict of interest.

## 536 **References**

- 537 Abbott, B. W., Moatar, F., Gauthier, O., Fovet, O., Antoine, V., and Ragueneau, O.: Trends and  
538 seasonality of river nutrients in agricultural catchments: 18years of weekly citizen science in France,  
539 *Science of The Total Environment*, 624, 845-858, <https://doi.org/10.1016/j.scitotenv.2017.12.176>,  
540 2018.
- 541 Ågren, A., Buffam, I., Jansson, M., and Laudon, H.: Importance of seasonality and small streams for the  
542 landscape regulation of dissolved organic carbon export, *Journal of Geophysical Research:*  
543 *Biogeosciences*, 112, <https://doi.org/10.1029/2006JG000381>, 2007.
- 544 Aiken, G. R., Hsu-Kim, H., and Ryan, J. N.: Influence of Dissolved Organic Matter on the Environmental  
545 Fate of Metals, Nanoparticles, and Colloids, *Environmental Science & Technology*, 45, 3196-3201,  
546 10.1021/es103992s, 2011.
- 547 Andersen, J. M.: Effect of nitrate concentration in lake water on phosphate release from the sediment,  
548 *Water Research*, 16, 1119-1126, [https://doi.org/10.1016/0043-1354\(82\)90128-2](https://doi.org/10.1016/0043-1354(82)90128-2), 1982.
- 549 Battin, T. J., Kaplan, L. A., Findlay, S., Hopkinson, C. S., Marti, E., Packman, A. I., Newbold, J. D., and  
550 Sabater, F.: Biophysical controls on organic carbon fluxes in fluvial networks, *Nature Geoscience*,  
551 1, 95-100, 10.1038/ngeo101, 2008.
- 552 Blodau, C., Fulda, B., Bauer, M., and Knorr, K.-H.: Arsenic speciation and turnover in intact organic soil  
553 mesocosms during experimental drought and rewetting, *Geochimica et Cosmochimica Acta*, 72,  
554 3991-4007, <https://doi.org/10.1016/j.gca.2008.04.040>, 2008.
- 555 Bouillon, S., Yambele, A., Gillikin, D. P., Teodoru, C., Darchambeau, F., Lambert, T., and Borges, A. V.:  
556 Contrasting biogeochemical characteristics of the Oubangui River and tributaries (Congo River  
557 basin), *Sci Rep*, 4, 5402, 10.1038/srep05402, 2014.
- 558 Boyer, E. W., Hornberger, G. M., Bencala, K. E., and McKnight, D.: Overview of a simple model  
559 describing variation of dissolved organic carbon in an upland catchment, *Ecological Modelling*, 86,  
560 183-188, [https://doi.org/10.1016/0304-3800\(95\)00049-6](https://doi.org/10.1016/0304-3800(95)00049-6), 1996.
- 561 Buffam, I., Galloway, J. N., Blum, L. K., and McGlathery, K. J.: A stormflow/baseflow comparison of  
562 dissolved organic matter concentrations and bioavailability in an Appalachian stream,  
563 *Biogeochemistry*, 53, 269-306, 10.1023/A:1010643432253, 2001.
- 564 Butturini, A. and Sabater, F.: Seasonal variability of dissolved organic carbon in a Mediterranean stream,  
565 *Biogeochemistry*, 51, 303-321, 10.1023/A:1006420229411, 2000.
- 566 Chen, C., Hall, S. J., Coward, E., and Thompson, A.: Iron-mediated organic matter decomposition in  
567 humid soils can counteract protection, *Nature Communications*, 11, 2255, 10.1038/s41467-020-  
568 16071-5, 2020.



569 Chow, A. T., Gao, S., and Dahlgren, R. A.: Physical and chemical fractionation of dissolved organic  
570 matter and trihalomethane precursors: A review, *Journal of Water Supply: Research and*  
571 *Technology-Aqua*, 54, 475-507, [10.2166/aqua.2005.0044](https://doi.org/10.2166/aqua.2005.0044), 2005.

572 Christensen, T. H., Bjerg, P. L., Banwart, S. A., Jakobsen, R., Heron, G., and Albrechtsen, H.-J.:  
573 Characterization of redox conditions in groundwater contaminant plumes, *Journal of Contaminant*  
574 *Hydrology*, 45, 165-241, [https://doi.org/10.1016/S0169-7722\(00\)00109-1](https://doi.org/10.1016/S0169-7722(00)00109-1), 2000.

575 Creed, I. F., McKnight, D. M., Pellerin, B. A., Green, M. B., Bergamaschi, B. A., Aiken, G. R., Burns, D.  
576 A., Findlay, S. E. G., Shanley, J. B., Striegl, R. G., Aulenbach, B. T., Clow, D. W., Laudon, H.,  
577 McGlynn, B. L., McGuire, K. J., Smith, R. A., and Stackpoole, S. M.: The river as a chemostat: fresh  
578 perspectives on dissolved organic matter flowing down the river continuum, *Canadian Journal of*  
579 *Fisheries and Aquatic Sciences*, 72, 1272-1285, [10.1139/cjfas-2014-0400](https://doi.org/10.1139/cjfas-2014-0400), 2015.

580 de Wit, H. A., Stoddard, J. L., Monteith, D. T., Sample, J. E., Austnes, K., Couture, S., Fölster, J.,  
581 Higgins, S. N., Houle, D., Hruška, J., Krám, P., Kopáček, J., Paterson, A. M., Valinia, S., Van Dam,  
582 H., Vuorenmaa, J., and Evans, C. D.: Cleaner air reveals growing influence of climate on dissolved  
583 organic carbon trends in northern headwaters, *Environmental Research Letters*, 16, 104009,  
584 [10.1088/1748-9326/ac2526](https://doi.org/10.1088/1748-9326/ac2526), 2021.

585 Dean, J. F., Meisel, O. H., Martyn Rosco, M., Marchesini, L. B., Garnett, M. H., Lenderink, H., van  
586 Logtestijn, R., Borges, A. V., Bouillon, S., Lambert, T., Röckmann, T., Maximov, T., Petrov, R.,  
587 Karsanaev, S., Aerts, R., van Huissteden, J., Vonk, J. E., and Dolman, A. J.: East Siberian Arctic  
588 inland waters emit mostly contemporary carbon, *Nature Communications*, 11, 1627,  
589 [10.1038/s41467-020-15511-6](https://doi.org/10.1038/s41467-020-15511-6), 2020.

590 Deirmendjian, L., Loustau, D., Augusto, L., Lafont, S., Chipeaux, C., Poirier, D., and Abril, G.: Hydro-  
591 ecological controls on dissolved carbon dynamics in groundwater and export to streams in a  
592 temperate pine forest, *Biogeosciences*, 15, 669-691, [10.5194/bg-15-669-2018](https://doi.org/10.5194/bg-15-669-2018), 2018.

593 Dong, H., Zeng, Q., Sheng, Y., Chen, C., Yu, G., and Kappler, A.: Coupled iron cycling and organic  
594 matter transformation across redox interfaces, *Nature Reviews Earth & Environment*, 4, 659-673,  
595 [10.1038/s43017-023-00470-5](https://doi.org/10.1038/s43017-023-00470-5), 2023.

596 Dupas, R., Gruau, G., Gu, S., Humbert, G., Jaffrézic, A., and Gascuel-Oudou, C.: Groundwater control  
597 of biogeochemical processes causing phosphorus release from riparian wetlands, *Water Research*,  
598 84, 307-314, <https://doi.org/10.1016/j.watres.2015.07.048>, 2015.

599 Durand, P. and Juan Torres, J. L.: Solute transfer in agricultural catchments: the interest and limits of  
600 mixing models, *Journal of Hydrology*, 181, 1-22, [https://doi.org/10.1016/0022-1694\(95\)02922-2](https://doi.org/10.1016/0022-1694(95)02922-2),  
601 1996.

602 Fellman, J. B., Hood, E., and Spencer, R. G. M.: Fluorescence spectroscopy opens new windows into  
603 dissolved organic matter dynamics in freshwater ecosystems: A review, *Limnology and*  
604 *Oceanography*, 55, 2452-2462, <https://doi.org/10.4319/lo.2010.55.6.2452>, 2010.

605 Fellman, J. B., Hood, E., D'Amore, D. V., Edwards, R. T., and White, D.: Seasonal changes in the  
606 chemical quality and biodegradability of dissolved organic matter exported from soils to streams in  
607 coastal temperate rainforest watersheds, *Biogeochemistry*, 95, 277-293, [10.1007/s10533-009-9336-6](https://doi.org/10.1007/s10533-009-9336-6), 2009.

609 Fenner, N. and Freeman, C.: Drought-induced carbon loss in peatlands, *Nature Geoscience*, 4, 895-  
610 900, [10.1038/ngeo1323](https://doi.org/10.1038/ngeo1323), 2011.

611 Fovet, O., Ruiz, L., Gruau, G., Akkal, N., Aquilina, L., Busnot, S., Dupas, R., Durand, P., Fauchoux, M.,  
612 Fauvel, Y., Fléchar, C., Gilliet, N., Grimaldi, C., Hamon, Y., Jaffrezic, A., Jeanneau, L., Labasque,  
613 T., Le Henaff, G., Mérot, P., Molénat, J., Petitjean, P., Pierson-Wickmann, A.-C., Squidant, H.,  
614 Viaud, V., Walter, C., and Gascuel-Oudou, C.: AgrHyS: An Observatory of Response Times in  
615 Agro-Hydro Systems, *Vadose Zone Journal*, 17, 180066, <https://doi.org/10.2136/vzj2018.04.0066>,  
616 2018.

617 Grybos, M., Davranche, M., Gruau, G., Petitjean, P., and Pédrot, M.: Increasing pH drives organic matter  
618 solubilization from wetland soils under reducing conditions, *Geoderma*, 154, 13-19,  
619 <https://doi.org/10.1016/j.geoderma.2009.09.001>, 2009.

620 Gu, S., Gruau, G., Dupas, R., Rumpel, C., Crème, A., Fovet, O., Gascuel-Oudou, C., Jeanneau, L.,  
621 Humbert, G., and Petitjean, P.: Release of dissolved phosphorus from riparian wetlands: Evidence  
622 for complex interactions among hydroclimate variability, topography and soil properties, *Science of*  
623 *The Total Environment*, 598, 421-431, <https://doi.org/10.1016/j.scitotenv.2017.04.028>, 2017.

624 Hagedorn, F., Kaiser, K., Feyen, H., and Schleppei, P.: Effects of Redox Conditions and Flow Processes  
625 on the Mobility of Dissolved Organic Carbon and Nitrogen in a Forest Soil, *Journal of Environmental*  
626 *Quality*, 29, 288-297, <https://doi.org/10.2134/jeq2000.00472425002900010036x>, 2000.

627 Hanson, P. C., Pace, M. L., Carpenter, S. R., Cole, J. J., and Stanley, E. H.: Integrating Landscape  
628 Carbon Cycling: Research Needs for Resolving Organic Carbon Budgets of Lakes, Ecosystems,  
629 18, 363-375, 10.1007/s10021-014-9826-9, 2015.

630 Harrison, A. F., Taylor, K., Scott, A., Poskitt, J., Benham, D., Grace, J., Chaplow, J., and Rowland, P.:  
631 Potential effects of climate change on DOC release from three different soil types on the Northern  
632 Pennines UK: examination using field manipulation experiments, *Global Change Biology*, 14, 687-  
633 702, <https://doi.org/10.1111/j.1365-2486.2007.01504.x>, 2008.

634 Humbert, G., Jaffrezic, A., Fovet, O., Gruau, G., and Durand, P.: Dry-season length and runoff control  
635 annual variability in stream DOC dynamics in a small, shallow groundwater-dominated agricultural  
636 watershed, *Water Resources Research*, 51, 7860-7877, <https://doi.org/10.1002/2015WR017336>,  
637 2015.

638 Humbert, G., Parr, T. B., Jeanneau, L., Dupas, R., Petitjean, P., Akkal-Corfini, N., Viaud, V., Pierson-  
639 Wickmann, A.-C., Denis, M., Inamdar, S., Gruau, G., Durand, P., and Jaffrézic, A.: Agricultural  
640 Practices and Hydrologic Conditions Shape the Temporal Pattern of Soil and Stream Water  
641 Dissolved Organic Matter, *Ecosystems*, 23, 1325-1343, 10.1007/s10021-019-00471-w, 2020.

642 Inamdar, S., Finger, N., Singh, S., Mitchell, M., Levia, D., Bais, H., Scott, D., and McHale, P.: Dissolved  
643 organic matter (DOM) concentration and quality in a forested mid-Atlantic watershed, USA,  
644 *Biogeochemistry*, 108, 55-76, 10.1007/s10533-011-9572-4, 2012.

645 Inamdar, S. P., O'Leary, N., Mitchell, M. J., and Riley, J. T.: The impact of storm events on solute exports  
646 from a glaciated forested watershed in western New York, USA, *Hydrological Processes*, 20, 3423-  
647 3439, <https://doi.org/10.1002/hyp.6141>, 2006.

648 Jia, K., Manning, C. C. M., Jollymore, A., and Beckie, R. D.: Technical note: Effects of iron(II) on  
649 fluorescence properties of dissolved organic matter at circumneutral pH, *Hydrol. Earth Syst. Sci.*,  
650 25, 4983-4993, 10.5194/hess-25-4983-2021, 2021.

651 Josse, J.: Principal component methods - hierarchical clustering - partitional clustering: why would we  
652 need to choose for visualizing data?,

653 Kappler, A., Bryce, C., Mansor, M., Lueder, U., Byrne, J. M., and Swanner, E. D.: An evolving view on  
654 biogeochemical cycling of iron, *Nature Reviews Microbiology*, 19, 360-374, 10.1038/s41579-020-  
655 00502-7, 2021.

656 Kelly, D. J., Clare, J. J., and Bothwell, M. L.: Attenuation of solar ultraviolet radiation by dissolved organic  
657 matter alters benthic colonization patterns in streams, *Journal of the North American Benthological  
658 Society*, 20, 96-108, 10.2307/1468191, 2001.

659 Knorr, K. H.: DOC-dynamics in a small headwater catchment as driven by redox fluctuations and  
660 hydrological flow paths – are DOC exports mediated by iron reduction/oxidation cycles?,  
661 *Biogeosciences*, 10, 891-904, 10.5194/bg-10-891-2013, 2013.

662 Kothawala, D. N., Ji, X., Laudon, H., Ågren, A. M., Futter, M. N., Köhler, S. J., and Tranvik, L. J.: The  
663 relative influence of land cover, hydrology, and in-stream processing on the composition of  
664 dissolved organic matter in boreal streams, *Journal of Geophysical Research: Biogeosciences*,  
665 120, 1491-1505, <https://doi.org/10.1002/2015JG002946>, 2015.

666 Lambert, T., Perolo, P., Escoffier, N., and Perga, M. E.: Enhanced bioavailability of dissolved organic  
667 matter (DOM) in human-disturbed streams in Alpine fluvial networks, *Biogeosciences*, 19, 187-200,  
668 10.5194/bg-19-187-2022, 2022.

669 Lambert, T., Pierson-Wickmann, A.-C., Gruau, G., Thibault, J.-N., and Jaffrezic, A.: Carbon isotopes as  
670 tracers of dissolved organic carbon sources and water pathways in headwater catchments, *Journal  
671 of Hydrology*, 402, 228-238, <https://doi.org/10.1016/j.jhydrol.2011.03.014>, 2011.

672 Lambert, T., Bouillon, S., Darchambeau, F., Morana, C., Roland, F. A. E., Descy, J.-P., and Borges, A.  
673 V.: Effects of human land use on the terrestrial and aquatic sources of fluvial organic matter in a  
674 temperate river basin (The Meuse River, Belgium), *Biogeochemistry*, 136, 191-211,  
675 10.1007/s10533-017-0387-9, 2017.

676 Lambert, T., Pierson-Wickmann, A.-C., Gruau, G., Jaffrezic, A., Petitjean, P., Thibault, J.-N., and  
677 Jeanneau, L.: Hydrologically driven seasonal changes in the sources and production mechanisms  
678 of dissolved organic carbon in a small lowland catchment, *Water Resources Research*, 49, 5792-  
679 5803, <https://doi.org/10.1002/wrcr.20466>, 2013.

680 Lambert, T., Pierson-Wickmann, A. C., Gruau, G., Jaffrezic, A., Petitjean, P., Thibault, J. N., and  
681 Jeanneau, L.: DOC sources and DOC transport pathways in a small headwater catchment as  
682 revealed by carbon isotope fluctuation during storm events, *Biogeosciences*, 11, 3043-3056,  
683 10.5194/bg-11-3043-2014, 2014.

684 Laudon, H., Buttle, J., Carey, S. K., McDonnell, J., McGuire, K., Seibert, J., Shanley, J., Soulsby, C.,  
685 and Tetzlaff, D.: Cross-regional prediction of long-term trajectory of stream water DOC response

686 to climate change, *Geophysical Research Letters*, 39, <https://doi.org/10.1029/2012GL053033>,  
687 2012.

688 Lê, S., Josse, J., and Husson, F.: FactoMineR: An R Package for Multivariate Analysis, *Journal of*  
689 *Statistical Software*, 25, 1 - 18, 10.18637/jss.v025.i01, 2008.

690 Ledesma, J. L. J., Grabs, T., Bishop, K. H., Schiff, S. L., and Köhler, S. J.: Potential for long-term transfer  
691 of dissolved organic carbon from riparian zones to streams in boreal catchments, *Global Change*  
692 *Biology*, 21, 2963-2979, <https://doi.org/10.1111/gcb.12872>, 2015.

693 Logozzo, L. A., Hosen, J. D., McArthur, J., and Raymond, P. A.: Distinct drivers of two size fractions of  
694 operationally dissolved iron in a temperate river, *Limnology and Oceanography*, 68, 1185-1200,  
695 <https://doi.org/10.1002/lno.12338>, 2023.

696 Lotfi-Kalahroodi, E., Pierson-Wickmann, A.-C., Rouxel, O., Marsac, R., Bouhnik-Le Coz, M., Hanna, K.,  
697 and Davranche, M.: More than redox, biological organic ligands control iron isotope fractionation in  
698 the riparian wetland, *Scientific Reports*, 11, 1933, 10.1038/s41598-021-81494-z, 2021.

699 Lucassen, E. C. H. E. T., Smolders, A. J. P., van der Salm, A. L., and Roelofs, J. G. M.: High groundwater  
700 nitrate concentrations inhibit eutrophication of sulphate-rich freshwater wetlands, *Biogeochemistry*,  
701 67, 249-267, 10.1023/B:BIOG.0000015342.40992.cb, 2004.

702 McMahon, P. B. and Chapelle, F. H.: Redox Processes and Water Quality of Selected Principal Aquifer  
703 Systems, *Groundwater*, 46, 259-271, <https://doi.org/10.1111/j.1745-6584.2007.00385.x>, 2008.

704 Mehring, A. S., Lowrance, R. R., Helton, A. M., Pringle, C. M., Thompson, A., Bosch, D. D., and Vellidis,  
705 G.: Interannual drought length governs dissolved organic carbon dynamics in blackwater rivers of  
706 the western upper Suwannee River basin, *Journal of Geophysical Research: Biogeosciences*, 118,  
707 1636-1645, <https://doi.org/10.1002/2013JG002415>, 2013.

708 Molenat, J., Gascuel-Oudou, C., Ruiz, L., and Gruau, G.: Role of water table dynamics on stream nitrate  
709 export and concentration in agricultural headwater catchment (France), *Journal of Hydrology*, 348,  
710 363-378, <https://doi.org/10.1016/j.jhydrol.2007.10.005>, 2008.

711 Monteith, D. T., Stoddard, J. L., Evans, C. D., de Wit, H. A., Forsius, M., Høgåsen, T., Wilander, A.,  
712 Skjelkvåle, B. L., Jeffries, D. S., Vuorenmaa, J., Keller, B., Kopáček, J., and Vesely, J.: Dissolved  
713 organic carbon trends resulting from changes in atmospheric deposition chemistry, *Nature*, 450,  
714 537-540, 10.1038/nature06316, 2007.

715 Murphy, K. R., Stedmon, C. A., Graeber, D., and Bro, R.: Fluorescence spectroscopy and multi-way  
716 techniques. PARAFAC, *Analytical Methods*, 5, 6557-6566, 10.1039/C3AY41160E, 2013.

717 Murphy, K. R., Stedmon, C. A., Wenig, P., and Bro, R.: OpenFluor— an online spectral library of auto-  
718 fluorescence by organic compounds in the environment, *Analytical Methods*, 6, 658-661,  
719 10.1039/C3AY41935E, 2014.

720 Musolff, A., Selle, B., Büttner, O., Opitz, M., and Tittel, J.: Unexpected release of phosphate and organic  
721 carbon to streams linked to declining nitrogen depositions, *Global Change Biology*, 23, 1891-1901,  
722 <https://doi.org/10.1111/gcb.13498>, 2017.

723 Neff, J. C., Finlay, J. C., Zimov, S. A., Davydov, S. P., Carrasco, J. J., Schuur, E. A. G., and Davydova,  
724 A. I.: Seasonal changes in the age and structure of dissolved organic carbon in Siberian rivers and  
725 streams, *Geophysical Research Letters*, 33, <https://doi.org/10.1029/2006GL028222>, 2006.

726 Ohno, T.: Fluorescence Inner-Filtering Correction for Determining the Humification Index of Dissolved  
727 Organic Matter, *Environmental Science & Technology*, 36, 742-746, 10.1021/es0155276, 2002.

728 Ohno, T., Amirbahman, A., and Bro, R.: Parallel Factor Analysis of Excitation–Emission Matrix  
729 Fluorescence Spectra of Water Soluble Soil Organic Matter as Basis for the Determination of  
730 Conditional Metal Binding Parameters, *Environmental Science & Technology*, 42, 186-192,  
731 10.1021/es071855f, 2008.

732 Pacific, V. J., Jencso, K. G., and McGlynn, B. L.: Variable flushing mechanisms and landscape structure  
733 control stream DOC export during snowmelt in a set of nested catchments, *Biogeochemistry*, 99,  
734 193-211, 10.1007/s10533-009-9401-1, 2010.

735 Poulin, B. A., Ryan, J. N., and Aiken, G. R.: Effects of Iron on Optical Properties of Dissolved Organic  
736 Matter, *Environmental Science & Technology*, 48, 10098-10106, 10.1021/es502670r, 2014.

737 Pullin, M. J., Anthony, C., and Maurice, P. A.: Effects of Iron on the Molecular Weight Distribution, Light  
738 Absorption, and Fluorescence Properties of Natural Organic Matter, *Environmental Engineering*  
739 *Science*, 24, 987-997, 10.1089/ees.2006.0040, 2007.

740 Raymond, P. A. and Saiers, J. E.: Event controlled DOC export from forested watersheds,  
741 *Biogeochemistry*, 100, 197-209, 10.1007/s10533-010-9416-7, 2010.

742 Ruckhaus, M., Seybold, E. C., Underwood, K. L., Stewart, B., Kincaid, D. W., Shanley, J. B., Li, L., and  
743 Perdrial, J. N.: Disentangling the responses of dissolved organic carbon and nitrogen  
744 concentrations to overlapping drivers in a northeastern United States forested watershed, *Frontiers*  
745 *in Water*, 5, 10.3389/frwa.2023.1065300, 2023.

746 Sanderman, J., Lohse, K. A., Baldock, J. A., and Amundson, R.: Linking soils and streams: Sources and  
747 chemistry of dissolved organic matter in a small coastal watershed, *Water Resources Research*,  
748 45, <https://doi.org/10.1029/2008WR006977>, 2009.

749 Seibert, J., Grabs, T., Köhler, S., Laudon, H., Winterdahl, M., and Bishop, K.: Linking soil- and stream-  
750 water chemistry based on a Riparian Flow-Concentration Integration Model, *Hydrol. Earth Syst.*  
751 *Sci.*, 13, 2287-2297, 10.5194/hess-13-2287-2009, 2009.

752 Selle, B., Knorr, K.-H., and Lischeid, G.: Mobilisation and transport of dissolved organic carbon and iron  
753 in peat catchments—Insights from the Lehstenbach stream in Germany using generalised additive  
754 models, *Hydrological Processes*, 33, 3213-3225, <https://doi.org/10.1002/hyp.13552>, 2019.

755 Smith, G. J., McDowell, R. W., Condon, L. M., Daly, K., Ó hUallacháin, D., and Fenton, O.: Reductive  
756 dissolution of phosphorus associated with iron-oxides during saturation in agricultural soil profiles,  
757 *Journal of Environmental Quality*, 50, 1207-1219, <https://doi.org/10.1002/jeq2.20256>, 2021.

758 Smolders, E., Baetens, E., Verbeeck, M., Nawara, S., Diels, J., Verdier, M., Peeters, B., De Cooman,  
759 W., and Baken, S.: Internal Loading and Redox Cycling of Sediment Iron Explain Reactive  
760 Phosphorus Concentrations in Lowland Rivers, *Environmental Science & Technology*, 51, 2584-  
761 2592, 10.1021/acs.est.6b04337, 2017.

762 Stedmon, C. A. and Markager, S.: Resolving the variability in dissolved organic matter fluorescence in  
763 a temperate estuary and its catchment using PARAFAC analysis, *Limnology and Oceanography*,  
764 50, 686-697, <https://doi.org/10.4319/lo.2005.50.2.0686>, 2005.

765 Strohmenger, L., Fovet, O., Akkal-Corfini, N., Dupas, R., Durand, P., Faucheux, M., Gruau, G., Hamon,  
766 Y., Jaffrezic, A., Minaudo, C., Petitjean, P., and Gascuel-Oudou, C.: Multitemporal Relationships  
767 Between the Hydroclimate and Exports of Carbon, Nitrogen, and Phosphorus in a Small Agricultural  
768 Watershed, *Water Resources Research*, 56, e2019WR026323,  
769 <https://doi.org/10.1029/2019WR026323>, 2020.

770 Tank, S. E., Fellman, J. B., Hood, E., and Kritzberg, E. S.: Beyond respiration: Controls on lateral carbon  
771 fluxes across the terrestrial-aquatic interface, *Limnology and Oceanography Letters*, 3, 76-88,  
772 <https://doi.org/10.1002/lo.10065>, 2018.

773 Tiwari, T., Sponseller, R. A., and Laudon, H.: The emerging role of drought as a regulator of dissolved  
774 organic carbon in boreal landscapes, *Nature Communications*, 13, 5125, 10.1038/s41467-022-  
775 32839-3, 2022.

776 Turgeon, J. M. L. and Courchesne, F.: Hydrochemical behaviour of dissolved nitrogen and carbon in a  
777 headwater stream of the Canadian Shield: relevance of antecedent soil moisture conditions,  
778 *Hydrological Processes*, 22, 327-339, <https://doi.org/10.1002/hyp.6613>, 2008.

779 Vázquez, E., Romaní, A. M., Sabater, F., and Butturini, A.: Effects of the Dry–Wet Hydrological Shift on  
780 Dissolved Organic Carbon Dynamics and Fate Across Stream–Riparian Interface in a  
781 Mediterranean Catchment, *Ecosystems*, 10, 239-251, 10.1007/s10021-007-9016-0, 2007.

782 Wen, H., Perdrial, J., Abbott, B. W., Bernal, S., Dupas, R., Godsey, S. E., Harpold, A., Rizzo, D.,  
783 Underwood, K., Adler, T., Sterle, G., and Li, L.: Temperature controls production but hydrology  
784 regulates export of dissolved organic carbon at the catchment scale, *Hydrol. Earth Syst. Sci.*, 24,  
785 945-966, 10.5194/hess-24-945-2020, 2020.

786 Werner, B. J., Musolff, A., Lechtenfeld, O. J., de Rooij, G. H., Oosterwoud, M. R., and Fleckenstein, J.  
787 H.: High-frequency measurements explain quantity and quality of dissolved organic carbon  
788 mobilization in a headwater catchment, *Biogeosciences*, 16, 4497-4516, 10.5194/bg-16-4497-  
789 2019, 2019.

790 Wetzel, R. G.: Gradient-dominated ecosystems: sources and regulatory functions of dissolved organic  
791 matter in freshwater ecosystems, *Hydrobiologia*, 229, 181-198, 10.1007/BF00007000, 1992.

792 Williams, C. J., Yamashita, Y., Wilson, H. F., Jaffé, R., and Xenopoulos, M. A.: Unraveling the role of  
793 land use and microbial activity in shaping dissolved organic matter characteristics in stream  
794 ecosystems, *Limnology and Oceanography*, 55, 1159-1171,  
795 <https://doi.org/10.4319/lo.2010.55.3.1159>, 2010.

796 Winterdahl, M., Erlandsson, M., Futter, M. N., Weyhenmeyer, G. A., and Bishop, K.: Intra-annual  
797 variability of organic carbon concentrations in running waters: Drivers along a climatic gradient,  
798 *Global Biogeochemical Cycles*, 28, 451-464, <https://doi.org/10.1002/2013GB004770>, 2014.

799 Xu, N. and Saiers, J. E.: Temperature and Hydrologic Controls on Dissolved Organic Matter Mobilization  
800 and Transport within a Forest Topsoil, *Environmental Science & Technology*, 44, 5423-5429,  
801 10.1021/es1002296, 2010.

802 Yamashita, Y., Scinto, L. J., Maie, N., and Jaffé, R.: Dissolved Organic Matter Characteristics Across a  
803 Subtropical Wetland's Landscape: Application of Optical Properties in the Assessment of  
804 Environmental Dynamics, *Ecosystems*, 13, 1006-1019, 10.1007/s10021-010-9370-1, 2010.

805 Zarnetske, J. P., Bouda, M., Abbott, B. W., Saiers, J., and Raymond, P. A.: Generality of Hydrologic  
806 Transport Limitation of Watershed Organic Carbon Flux Across Ecoregions of the United States,  
807 Geophysical Research Letters, 45, 11,702-711,711, <https://doi.org/10.1029/2018GL080005>, 2018.

808

## 809 **Figure Caption**

810 **Figure 1** – Location map of the Kervidy-Naizin experimental catchment showing land uses.  
811 Hatched areas located along the stream channel network indicate the extent of hydromorphic  
812 soils commonly waterlogged during the winter period. Lysimeters were located downslope the  
813 piezometer PK1.

814 **Figure 2** – (A) Record of hourly discharge and daily rainfall, (B) record of hourly piezometric  
815 levels in wetland (PK1) and upland (PK3) domains, and (C) record of daily air temperature.  
816 Black triangles in panel A indicate fieldwork for manual sampling of soil and stream waters.  
817 Vertical black dashed lines delimit the different hydrologic periods, namely the rewetting, high  
818 flow, and recession phases. See text for details.

819 **Figure 3** – Evolution of (A) air temperature and (B) pH, (C) DOC, (D) NO<sub>3</sub>, (E) Fe(II), and (F)  
820 SRP in soil waters during the study period. Vertical black dashed lines delimit the different  
821 hydrologic periods, namely the rewetting, high flow, and recession phases. See text for details.

822 **Figure 4** – Relationships between (A) DOC and Fe(II), (B) Fe(II) and NO<sub>3</sub>, and (C) SRP and  
823 Fe(II) in soil waters during the study period.

824 **Figure 5** – Variations in stream DOC measured at high frequency at the outlet of the  
825 catchment. Vertical black dashed lines delimit the different hydrologic periods, namely the  
826 rewetting, high flow, and recession phases. See text for details.

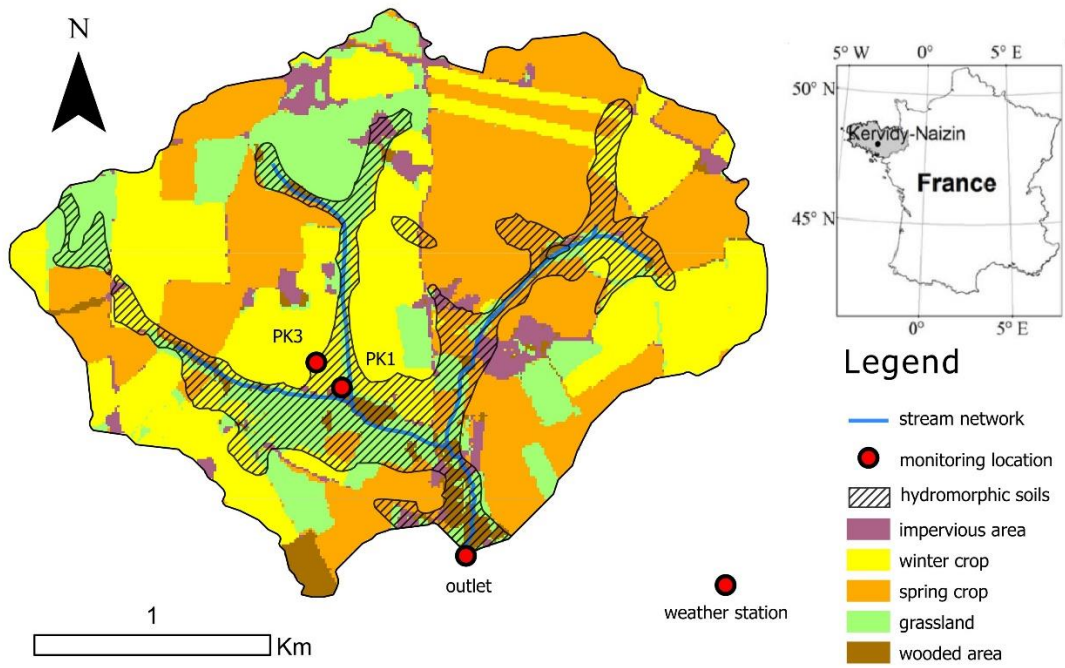
827 **Figure 6** – PCA biplot, including loadings plot for the input variables and scores plot for  
828 lysimeters. One point represents one lysimeters, PCA being based on average values  
829 calculated over the study period. Markers are coloured according to the cluster identified by  
830 the Hierarchical Clustering on Principal Components (see material and methods).

831 **Figure 7** – Evolution of (A) DOC, (B) Fe(II), (C) NO<sub>3</sub>, and (D) SRP in soil waters for each  
832 cluster. Lysimeters are grouped according the Hierarchical Clustering on Principal  
833 Components (see text for details and Fig. 6). Vertical black dashed lines delimit the different  
834 hydrologic periods, namely the rewetting, high flow, and recession phases. See text for details.

835

836

837 **Figure 1**



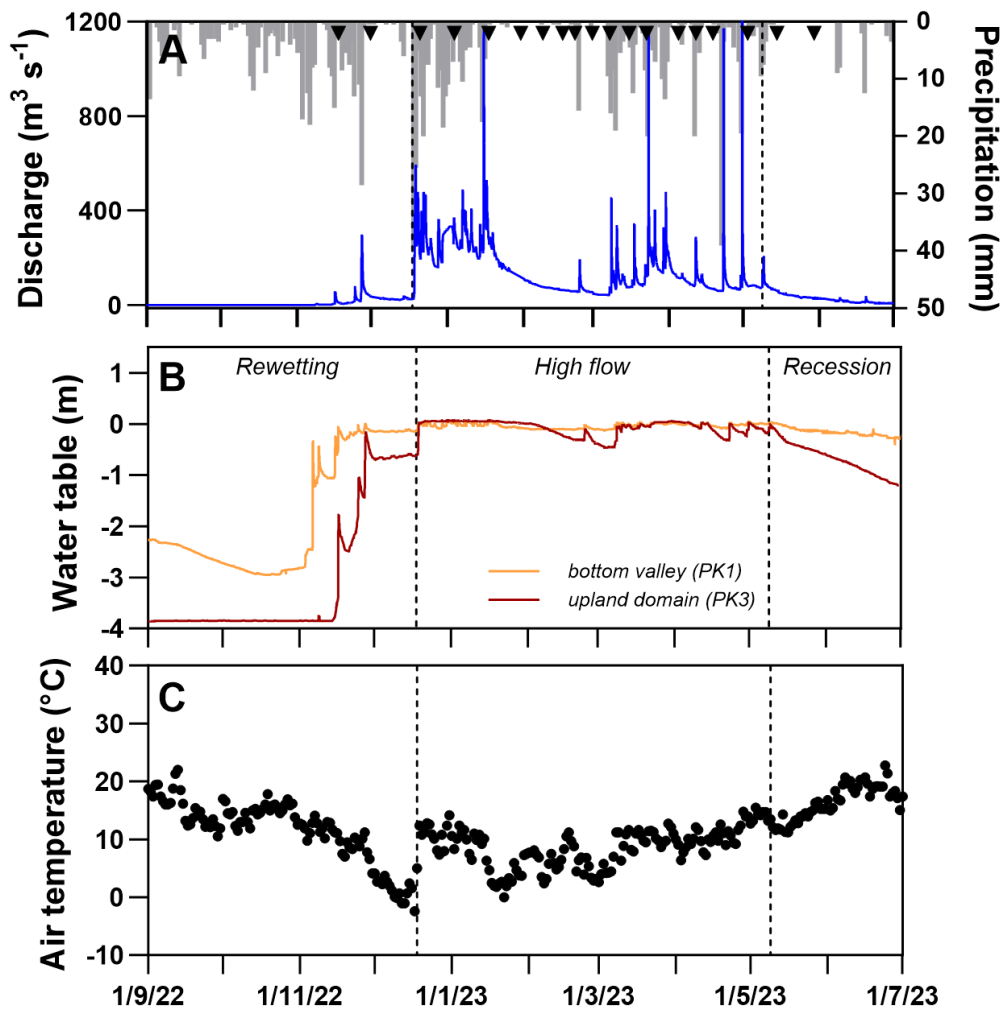
838

839

840

841

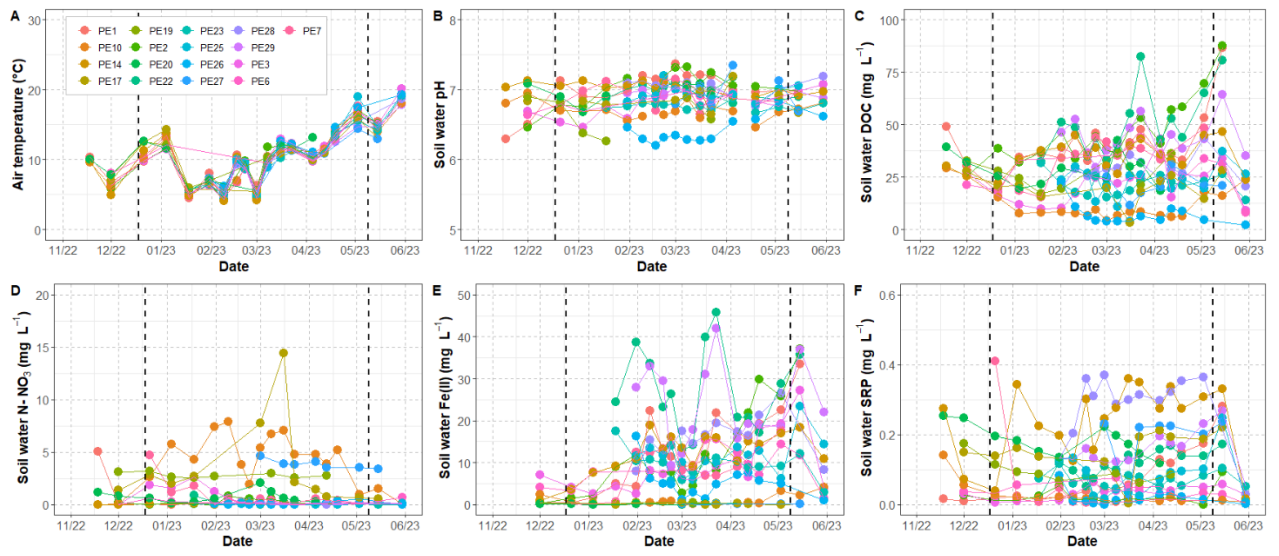
842 **Figure 2**



843

844

845 **Figure 3**

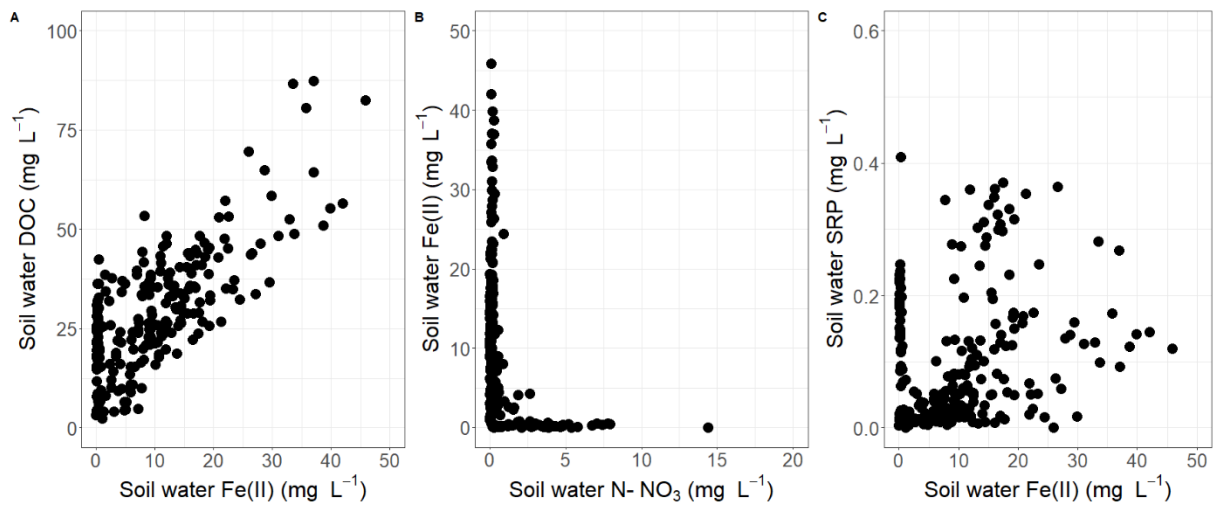


846

847



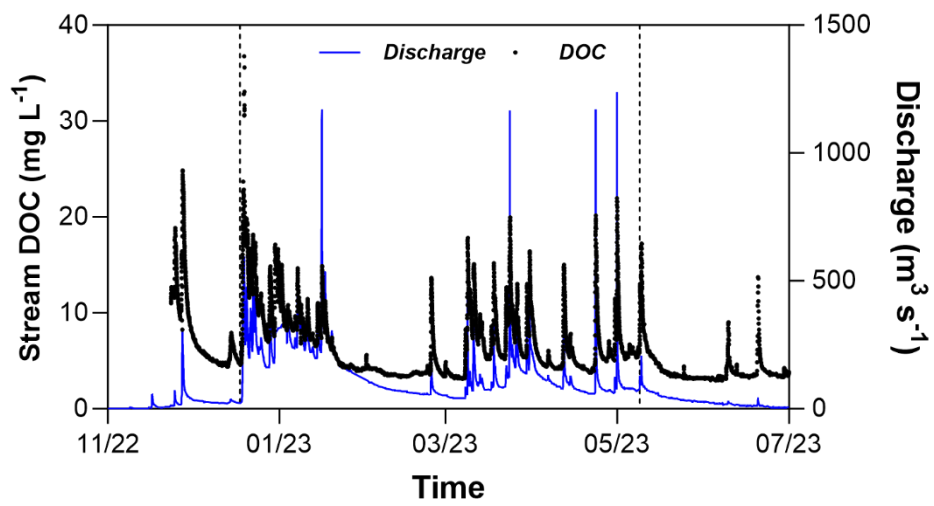
848 **Figure 4**



849

850

851 **Figure 5**

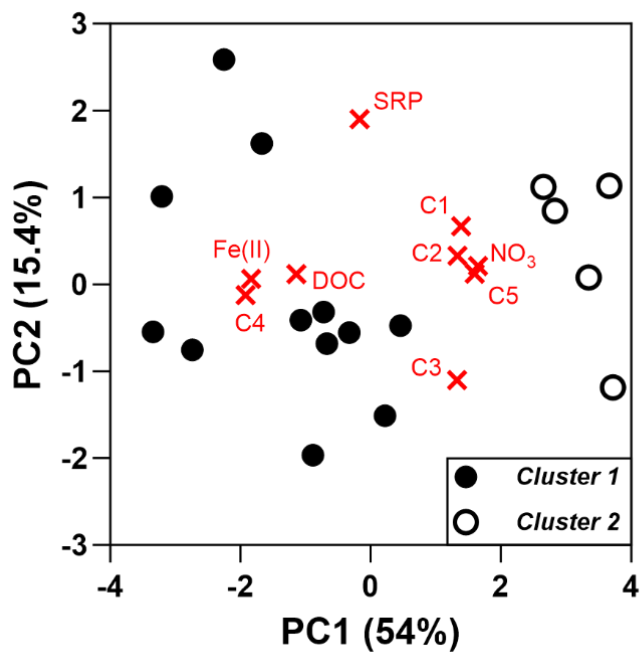


852

853

854

855 **Figure 6**

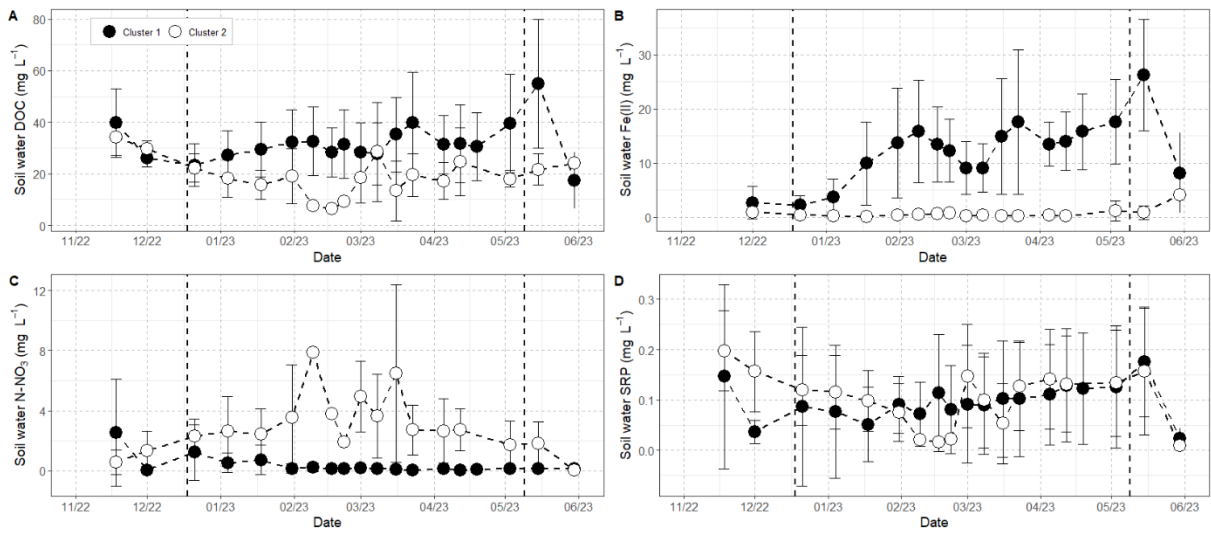


856

857

858

859 **Figure 7**



860