

Ancient clays support contemporary biogeochemical activity in the Critical Zone

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15 Abstract

Late Cretaceous clays exposed at sites located on the north shore of Long Island, New York, USA were sampled to explore questions about how contemporary factors and processes interact with ancient geological materials that are often assumed to not be biologically active. Chemically and biologically catalyzed weathering processes have produced multi-colored clays belonging to the kaolin group with inclusions of hematite, limonite, and pyrite nodules. We sampled exposed clays at three sites to address three questions: 1) Do these exposed clays support significant amounts of microbial biomass and activity, i.e., are they alive? 2) Do these clays support significant ~~amounts of~~ nitrogen (N) cycle activity? 3) Are these clays a potential non-anthropogenic source of reactive N ~~pollution~~ in the contemporary landscape? Samples were analyzed for total carbon (C) and N content, microbial biomass C and N content, microbial respiration, organic matter (OM) content, potential net N mineralization and nitrification, soil nitrate (NO_3^-) and ammonium (NH_4^+) content, and denitrification potential. Results strongly support the idea that ancient geologic materials play a role in contemporary N and C cycling in the Critical Zone. Respiration (average $4.098 \mu\text{g C g}^{-1} \text{d}^{-1}$) was detectable in all samples and was strongly correlated to OM, indicating a living microbial community on the clays. There was evidence of an active N cycle. Higher levels of denitrification potential (average $1.376 \mu\text{g N g}^{-1} \text{d}^{-1}$) compared to both potential net nitrification (average $0.061 \mu\text{g N g}^{-1} \text{d}^{-1}$) and potential net N mineralization (average $0.144 \mu\text{g N g}^{-1} \text{d}^{-1}$) indicate that these clays act more as a sink rather than as a source of ~~reactive N-pollution~~ in the landscape.

1 Introduction

35 The Critical Zone is Earth's constantly evolving boundary layer where rock, soil, water, air, and living organisms interact (Schroeder, 2018). It is comprised of the solid phase matter in which water circulates and is stored, expanding from the top of the vegetation canopy down into the water bearing bedrock. Critical Zone processes are key drivers of chemical transfers between biota and geological materials (Brantley et al., 2006).

40 A major question in Critical Zone science is how contemporary factors and processes interact with ancient geological materials. These materials are often assumed to have little or no biogeochemical activity as they have not been involved in active biological activity for long periods of time and may lack active microbial communities and labile pools of (C) and nitrogen (N) that drive this activity. These interactions are particularly obvious and important in the lower boundary of the Critical Zone, and in places where ancient geologic materials become exposed to contemporary environments (Dass et al., 2021). Major questions center on the ability of ancient materials to support 45 biogeochemical processes related to the cycling of C and N that underlie plant and microbial activity, which underlies environmental and ecosystem "services" of interest to society.

The discovery of significant amounts of N in sedimentary deposits increased interest in the role of geological materials in contemporary N cycling (Morford et al., 2011). Up to 17% of the currently cycling N in some ecosystems may originate from rock materials deep in the Critical Zone (Houlton et al., 2018). Such observations motivate the analysis of surface-exposed ancient rocks in this study. Both NH_4^+ and NO_3^- are highly soluble and the negative charge of NO_3^- makes it highly mobile, thus driving hydrologic losses of N. In excessive amounts, these nutrients lead to ecological stresses such as eutrophication. NH_4^+ is converted to NO_3^- by nitrifying bacteria, and NO_3^- is converted to N_2 by denitrifying bacteria leading to gaseous losses of N back into the atmosphere (Seitzinger et al., 2006).

In this study, we sampled surface exposures of unconsolidated bulk clay units exposed in outcrops that showed no evidence of modern soil development and measured microbial biomass and activity to address three questions: 1) Do these exposed clays support significant amounts of microbial biomass and activity, i.e. are they alive? To address this question we measured microbial biomass C, an index of the living microbial biomass in soil, and microbial respiration, a direct measure of microbial activity (Paul, 2014). Total C and organic matter (OM) content were measured as energy sources for microbial biomass and activity. 2) Do these clays support significant amounts of N cycle activity? N cycle activity was assessed with measurements of microbial biomass N content, potential net N mineralization, and total N content. Microbial biomass N provides an index of the net flux of N through microbial pools. Mineralization results from microbial degradation of N compounds resulting in the production of inorganic, plant-available forms of N. Total N content was measured to quantify the total amount of N potentially available for active cycling. 3) Are these clays a potential non-anthropogenic source of reactive N-pollution in the contemporary landscape? The potential for the clays' microbial activity to be a source or "sink" for reactive N-pollution was evaluated by measuring potential net nitrification, denitrification potential, and pools of ammonium (NH_4^+) and nitrate (NO_3^-). ~~Both NH_4^+ and NO_3^- are highly soluble and the negative charge of NO_3^- makes it highly mobile, thus driving hydrologic losses of N. In excessive amounts, these nutrients lead to ecological stresses such as eutrophication. NH_4^+ is converted to NO_3^- by nitrifying bacteria, and NO_3^- is converted to N_2 by denitrifying bacteria leading to gaseous losses of N back into the atmosphere (Seitzinger et al., 2006).~~

2 Background

2.1 Clay minerals and their role in ecosystem function and nutrient cycling

Clay minerals are hydrous aluminosilicates, more specifically hydrous phyllosilicates, and are some of the most stable products of chemical weathering at surface conditions. These minerals produce a specialized microhabitat and their ability to store and release nutrients make them ecologically important (Kleber et al., 2021). Geochemical composition and pore space of these materials, affect microbial activity and therefore rates of biogeochemical processes in the carbon (C) and nitrogen (N) cycles (Li et al., 2023). Clay minerals have high surface adhesion

capabilities and sorption and desorption of OM in soils varies with mineral assemblage. Sorption of organic carbon (OC) onto phyllosilicates and hydrous iron (Fe) oxides affects accumulation and stabilization of OC in soils (Saidy et al., 2013). The 1:1 layers of hydrated kaolinite clays and 2:1 layers of mixed layer clays, [along with OM](#), serve as nutrient exchange sites between biomass and subsurface weathering horizons. Therefore, the layered structures of clays are key facilitators of seasonal cycling of nutrients including NH_4^+ among others (Eby, 2016; Halama and Bebout, 2021).

Chemical weathering of silicate minerals is a significant mechanism for the availability, uptake, storage, and transport of key nutrients in ecosystems. Variations in mineral weathering and nutrient availability occur due to microorganism and mineral speciation, while intensity of mineral weathering is influenced by a mineral's potential to provide nutrients (P. C. Bennett et al., 2001). The pathways of C and N from the atmosphere to incorporation into the lithosphere progress from fixation into OM to storage in low temperature silicate phases such as clay minerals (Busigny and Bebout, 2013; Halama and Bebout, 2021). This is facilitated by the aqueous solutions and living microbes characteristic of the Critical Zone.

2.2 Geological background

Late Cretaceous clays on the north shore of Long Island, New York, USA offer an opportunity to study the effects of contemporary factors and processes on ancient Critical Zone materials. Long Island is composed of Pleistocene sediments deposited on top of Late Cretaceous formations (Sirkin, 1991) and includes aquifers, confining units, and clay-rich deposits such as the Raritan, Magothy, Gardiners, and Wantagh Clay Formations, that overlie gneissic bedrock. Coastal exposures of the Raritan and Magothy formations present opportunities for biogeochemical analysis of these materials. At these locations clays are exposed in outcrops, offering the chance to investigate their contemporary biogeochemical activity at surface conditions (Fig. 1). Long Island's clay strata have been mainly accessed through core drilling for hydrological studies, and in most instances the clays are a secondary detail rather than the main subject of study. The same is true for palynology studies in which the clay is the assemblage containing the ancient pollen being studied. In contrast, this study focuses on biogeochemical activity and how the clay strata affect and interact with surrounding environments in the Critical Zone. The kaolinitic materials at these sites and their abundance in oxides contribute to long lasting micro-environments (Six et al., 2000).



Figure 1: Map of study sites on Long Island, New York (maps constructed using map data from Esri and Google Earth).

All three study sites are located on the north shore of Long Island, NY. Two of the sites are located along the shoreline - Garvies Point Preserve (GP) on Hempstead Bay and Caumsett State Historic Park Preserve (CSP) along Long Island Sound; Hempstead Harbor Woods (HHW) is located inland from the western side of Hempstead Bay.

Long Island contains numerous clay beds such as Gardiner's Clay, Raritan Confining Unit, Wantagh Clay, and Smithtown Clay, as well as clay lenses in the Magothy Formation (Mills and Wells, 1974). The study area was shaped by the Wisconsin glaciation approximately 75,000 to 11,000 years ago. During this period, Cretaceous strata were sheared off, transported, and re-deposited. Glacially induced thrusting of the strata facilitated bulging of the clay (Mills and Wells, 1974), thus leading to eventual exposure. Evidence of this can be observed in the outcrop sampled for this study at GP, where the clay layers are currently oriented vertically rather than in a near horizontal depositional position. Although there is consensus that the Late Cretaceous deposits of the western north shore of Long Island were formed in a shallow delta or estuary (Fuller, 1914; Swarzenski, 1963) and that they are composed of sand, silt, gravel, and clay, further augmented by eolian action, it is not completely clear to which formation the exposed North Shore clays belong. It is generally accepted that the Late Cretaceous formations exposed on the North Shore tentatively belong to the Magothy formation but may include some younger formations (Isbister, 1966).

It is presumed that Pleistocene deep permafrost formation must have preceded glacial thrusting (Mills and Wells, 1974), which provided the sedimentary cohesion needed to produce the tilted clay strata and layered shale (not sampled for this study) observed today. The three sites of this study also differ in their position on the Manhasset Plateau. The HHW and GP localities are situated on the Upper Manhasset Plateau while CSP is situated on the Lower Manhasset Plateau. The Lower Manhasset Plateau is thought to have sustained a more prolonged grinding by the ice sheet (Fuller, 1914). The most abundant clay species identified from core drills in northwestern Long Island is kaolinite of light gray, brown-yellow, and tan color, along with sparse chlorite, vermiculite, and montmorillonite, with kaolinite ratio to other clays decreasing with depth (Liebling, 1973).

Fe content, among other factors, has contributed to variations in coloring and produced distinctive clay strata, allowing for color based grouping of the samples. The color variations are the result of differences in constituents and geochemical processes that evolved the clays into their modern state. The Fe oxides in hematite are likely sources of the red and brownish coloring and the Fe oxides in goethite are likely sources of the yellow coloring (Davey et al., 1975). This is further supported by portable x-ray fluorescence (pXRF) scan results (sections 4 and 5) which show red, yellow, and brown samples to have some of the highest Fe content. Mn also contributes to brown coloring (Jakobsson et al., 2000), and this is also further supported by pXRF scans that show brown samples to have the highest Mn content. Recent conditions have added sand and water to some clays to yield different textures, providing another observational classification criterion.

3 Methods

3.1 Study sites

Garvies Point Preserve (GP) is located in Glen Cove, NY, on the eastern shoreline of Hempstead Bay at 40°51'35" N 73°39'07" W. The exposed clays were accessed via trails leading to the beach and samples were collected along approximately 450 m of shoreline, with special focus on the main outcrop which features five types of clays exposed at this site. The most prominent outcrop at GP is approximately 4 m high and yielded samples from five differently colored clays (light gray, dark charcoal gray, white, yellow, and dark red/purplish). The adjacent outcrop is approximately 2 m high containing light and dark gray clay.

Hempstead Harbor Woods (HHW) is located in North Hempstead, NY, at 40°50'11" N 73°39'57" W, on the inland western side of Hempstead Bay, approximately 300 m from the shoreline. Samples were collected from 0 – 1 m from the ground level throughout the wooded area. Although this location does not have direct exposures to the bay, some of the sampled clay materials have indicators of being part of the same formation as those exposed across the bay at GP. Specifically, the red and light gray packed clays collected at HHW share the color and texture attributes of those at GP.

At Caumsett State Historic Park Preserve (CSP) samples were collected from exposures along approximately 1000 m of shoreline. A prominent exposure at CSP located at 40°56'21" N 73°28'13" W has an elevation of

approximately 40 m. Samples were collected starting from the bottom of the cliff, just above the beach floor, and as high as 6 m above the beach floor. Additional samples were collected from exposures at 40°56'08" N 73°28'14" W approximately 4 m above the beach floor, and at 40°56'11" N 73°28'43" W approximately 6 m above the beach floor.

3.2 Sampling

At each site samples were collected from locations that indicated the highest likelihood of yielding multiple clay types. Large outcrops containing multiple variations of clay at GP and CSP were sampled, yielding 14 and 5 samples respectively. The prominent outcrops at these locations feature variety in both color and texture. While our sampling facilitated comparison of these obvious differences, we did not have any *a priori* hypotheses about their causes and biogeochemical effects. HHW featured scattered locations, each featuring clay of a single texture and color, yielding 9 samples. Samples were collected from unconsolidated bulk clay units that showed no evidence of modern soil development, i.e. accumulation of recent organic matter or development of horizons. One sample from HHW (sample 3N-HHW) was collected from a naturally exposed soil horizon (B horizon) underneath topsoil (O horizon).

Samples were collected from the surface of the exposures. Locations where clay had at least a few inches of depth were chosen. The uppermost layer (~1 cm) was scraped off to ensure only clay was collected and to remove any field debris such as loose soil, rocks, sand, and loose plant material. Samples were collected from approximately 5-10 cm of depth. The samples were collected using a trowel, placed in labeled Ziplock bags, and packed loosely with some air remaining in the bag. Rocks, roots, and leaves were removed by hand right after collection. The samples were then refrigerated until laboratory processing. Samples were classified into groups of color (brown, dark grey, light gray, red, white, yellow) and texture (packed clay, sandy clay, watery clay) by visual observation. The packed clay was homogeneous, contained some moisture, and had a putty like texture that easily formed into a ribbon several inches long. The sandy clay was drier and had fine sand mixed in. The watery clay was collected from a small basin of waterlogged clay that was homogeneous, had very fine particles, and a thick viscous texture.

3.3 Laboratory analysis

Samples were analyzed for moisture content by oven drying for 24 hours at 105°C. Moisture content was used to calculate values for all variables on a per g of dry soil basis.

OM content was determined by Loss on Ignition (LOI). Oven dried samples were combusted at 450°C in a muffle furnace and % OM was calculated from weight loss after 8 hours of heating.

For elemental analysis by pXRF, samples were oven dried for 24 hours at 80°C, ground with a mortar and pestle, sieved through a #230 (63µm) sieve, tightly packed into 2.75cm diameter holders, and scanned with a portable Olympus DC-4000 XRF scanner.

Total C, total N, and the C/N ratio were measured using flash combustion / oxidation. Oven dried and ground samples were pressed into 1g pellets. The pellets were analyzed in an Elementar vario_MAX cube elemental analyzer.

For analysis of exchangeable NO_3^- and NH_4^+ , samples were blended with 2M KCl on an orbital shaker at 125 rpm for one hour, followed by filtration (Whatman #42 filter) into scintillation vials that were immediately refrigerated until analysis. The samples were pipetted into microplates and analyzed on a SpectraMax M2 Multi-Mode Microplate Reader from Molecular Devices using wavelengths of 450 nm for NO_3^- , and 650 nm for NH_4^+ (Doane and Horwáth, 2003; Sims et al., 1995). Total inorganic N (TIN) was calculated as the sum of NH_4^+ and NO_3^- .

The chloroform fumigation and incubation method (CFIM) (Jenkinson and Powlson, 1976) was used to determine the C and N content of microbial biomass. Samples (10 g) were fumigated with chloroform for 24 hours, inoculated with 0.2 g of unfumigated clay and incubated for 10 days in 946 ml Mason jars with lids fitted with septa. At the end of the incubation, gas samples were taken by syringe and analyzed for carbon dioxide (CO_2) with a Shimadzu GC-2014 gas chromatograph. After gas sampling, fumigated soils were extracted using KCl as described above after 10 days and the NO_3^- and NH_4^+ produced over the 10 day incubation was taken as an estimate of microbial biomass N. A proportionality constant of 0.41 was used to calculate microbial biomass C from the CO_2 produced over the 10 day incubation (Robertson, 1999).

Unfumigated samples were also incubated for 10 days and provided estimates of microbial respiration and potential net N mineralization and nitrification. These samples were incubated and sampled as described above and production of CO_2 during the 10 day incubation was taken as an estimate of microbial respiration. Production of NO_3^- and NH_4^+ over the 10 day incubation was taken as an estimate of potential net N mineralization and production of NO_3^- was taken as an estimate of potential net nitrification.

A denitrification enzyme assay (DEA) was used to measure the rate of potential denitrification (Groffman et al., 1999; Smith and Tiedje, 1979). 5g of each sample were placed in 125 mL Erlenmeyer flasks and amended with 10mL of DEA media. DEA media was prepared by adding 0.72g KNO_3^- as a substrate for existing enzymes, 0.5g glucose as a source of C / energy, and 0.125g chloramphenicol to block production of new enzymes during incubation, to 1L of nanopure water. The flasks were sealed with rubber stoppers, flushed repeatedly with N_2 gas to create anaerobic conditions, amended with acetylene (C_2H_2) gas, and placed on a shaker at 125 rpm. The headspace of the flasks was sampled (8 mL) by syringe after 30 minutes and 90 minutes of incubation. Samples were analyzed for nitrous oxide (N_2O) with a Shimadzu GC-2014 gas chromatograph.

3.4 Statistical analysis

235 SPSS version 28 was used for all analyses. One-way analysis of variance (ANOVA) with post hoc multiple
comparison tests were run on all response variables using location (GP, HHW, CSP), texture (packed, sandy,
watery), and color (yellow, white, red, light gray, dark gray, brown) as grouping factors. A Sidak adjustment was
applied to the post hoc multiple comparison tests because the number of samples was not equal across groups.
Spearman's rho was used to evaluate linear correlations because the data were not normally distributed and had a
240 heavy positive skew. ANOVA pretest results for homogeneity of variance were reinforced with Welch and Brown –
Forsythe tests. Kruskal Wallis analysis was run to increase confidence in results for groupings by color, texture, and
location. Mann-Whitney analysis with the Monte Carlo option was used to further reinforce ANOVA results
grouped by location. Significance levels were evaluated based on $\alpha < 0.01$ indicating a strong statistically different
significance, $\alpha < 0.05$ indicating a standard statistically different significance, and $\alpha < 0.1$ indicating a marginal
245 statistically different significance. Since ANOVA results yielded the same significance parameters for the majority
of variables as Kruskal Wallis and Mann-Whitney tests, ANOVA values are reported in the results section.

4 Results

4.1 Basic characteristics of clays

250 All samples had detectable amounts of total C, N, and OM (Fig. 2). The dark gray samples had significantly higher
OM, total C, total N, and C/N ratio than several of the other clay color types. The packed clay had the highest
values of all these variables among the texture groups, and significant differences were found for OM between
packed and sandy clays. ANOVA post hoc analysis identified further significant differences in the C/N ratio when
255 comparing dark gray clay to the Fe bearing red, yellow, and brown clays. A number of packed, dark gray and light
gray samples contained concentrations of N exceeding 1000 mg N kg⁻¹ which is considered ecologically significant
for geologic N (Holloway and Dahlgren, 2002).

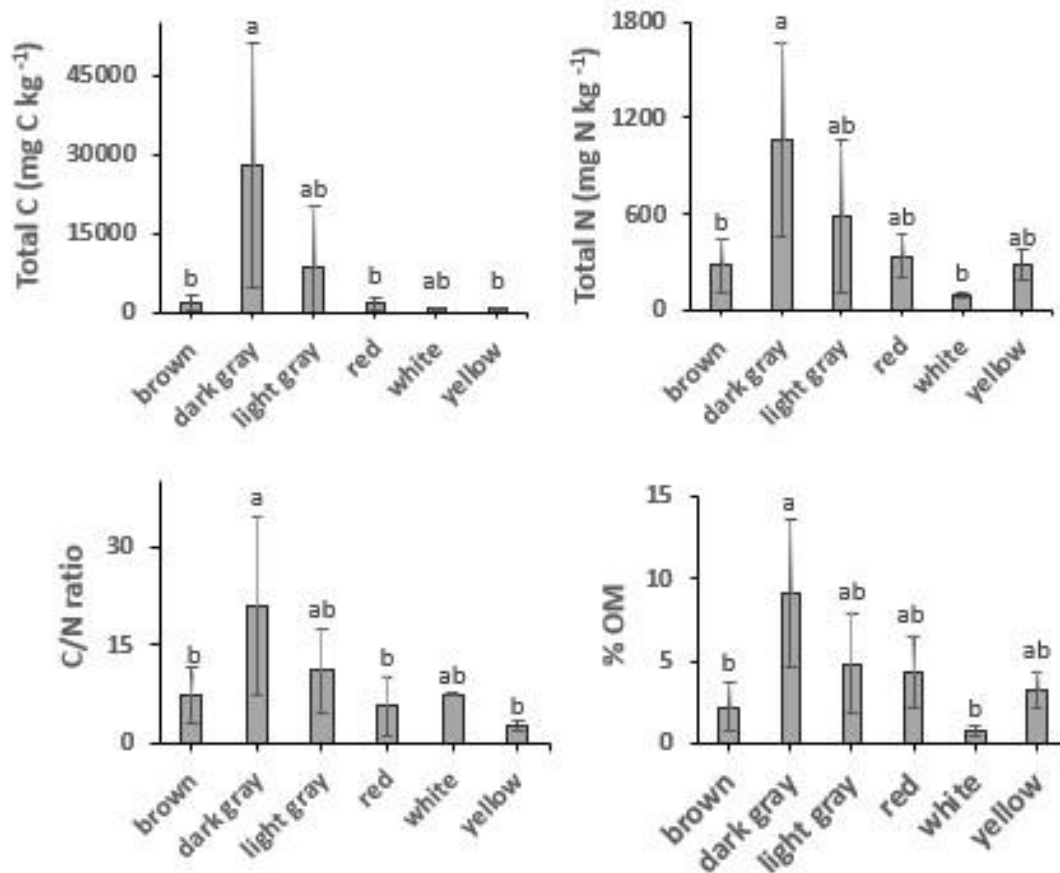


Figure 2: Total C, N, OM content, and C/N ratio of different colored clays. Values are mean $\pm$ SD. Values with different superscripts are significantly different ($p < 0.05$) except: in total C dark gray clay was marginally different ($p < 0.10$) from yellow clay; in total N dark gray clay was marginally different ($p < 0.10$) from brown and white clays; C/N ratio in dark gray clay was marginally different ($p < 0.10$) from brown clay.

Semi-quantitative results from pXRF scans provide estimates of selected elements of interest (Table 1a). P, a critical nutrient, was highest in brown clay and in watery clay with marginal differences between GP and HHW. Cl, which plays a role in nutrient transport, was highest in yellow, brown, and red clays, and in watery clays with significant differences among color groups and locations. Ti, which plays a role in nutrient cycling and enhances clay adsorption properties, was highest in yellow, red, and brown clays, and in packed clays with significant differences among packed and sandy clay textures and marginal differences between GP and CSP locations. Mn, which plays a role in nitrogen metabolism, was highest in brown clays and watery clays, with significant differences among color groups, textures, and locations. Fe, a critical nutrient, was highest in brown, red, yellow clays, and in watery clays, with significant and marginal differences among color groups and locations. Rb can shed light on clay speciation and therefore nutrient cycling capacity and was highest in dark gray, and in packed clays with significant differences among color groups and locations. Sr can interact with other elements such as Ca and had similar estimated abundances across color groups with the highest concentrations in the light gray, dark gray, brown, and yellow samples. Zr, which can enhance the adsorption properties of clays, was highest in brown and white samples

and in sandy and watery clays, with significant and marginal differences among color groups, textures, and locations. Zn, an essential nutrient, was highest in brown and dark gray, and in watery clays, with significant and marginal differences among color groups and locations. The Rb/Sr ratio can serve as a proxy for environmental changes that affected the clays, and was highest in dark gray clays, nearly the same across textures, with significant differences among color groups and locations (locations had additional marginal differences). There were significant correlations between P and Cl, P and Mn, P and Fe, Cl and Mn, Cl and Fe, Ti and Zr, Mn and Fe, Mn and Zn, Zn and Rb, Zn and Sr, Rb and Sr, Sr and Zr, and marginal correlations between P and Ti, and Ti and Fe.

Garvies Point	P	Cl	Ti	Mn	Fe	Rb	Sr	Zr	Zn	Rb/Sr ratio
dark gray	10991.38	140312.87	4171.20	67.73	6736.80	139.20	73.93	288.47	65.47	1.89
light gray	3372.50	133413.83	3441.33	49.33	4820.00	111.33	70.17	284.33	45.17	1.56
red	30710.67	161559.78	4884.67	52.33	27106.89	98.67	67.67	294.67	48.56	1.46
white	9194.00	126827.67	4351.00	85.67	6296.33	137.00	75.67	335.33	48.33	1.81
yellow	25721.33	160147.78	4566.11	54.00	16648.44	110.33	72.22	340.44	34.00	1.52
Hempstead Harbor Woods										
brown	14576.00	152885.00	3806.00	204.67	14309.00	93.67	80.33	330.00	73.00	1.17
light gray	8068.67	123039.50	3832.17	40.17	6617.50	96.61	76.56	376.56	34.06	1.37
red	5371.50	139207.00	4649.67	23.33	7586.33	42.67	41.00	448.33	33.67	1.04
white	442.00	135305.00	3644.33	32.33	4786.33	79.67	51.00	804.67	23.00	1.56
Caumsett State Park										
brown	31116.83	161317.92	3584.17	352.92	25723.83	98.08	72.08	676.67	74.75	1.39
red	3942.00	142768.00	3741.00	81.00	18065.67	67.33	62.33	387.00	59.67	1.08

Table 1a: Estimated amounts of selected elements of interest detected by pXRF analysis; values are means in ppm.

Garvies Point	SD of P	SD of Cl	SD of Ti	SD of Mn	SD of Fe	SD of Rb	SD of Sr	SD of Zr	SD of Zn	SD of Rb:Sr
dark gray	5524	18068	210	18	1633	8	6	32	21	0.1
light gray	380	1430	647	5	1257	41	9	16	30	0.4
red	23507	12890	906	14	20172	13	6	34	30	0.1
yellow	19354	10021	318	3	11599	18	8	60	2	0.1
Hempstead Harbor Woods										
light gray	5363	7575	560	11	1325	9	28	130	10	0.4
Caumsett State Park										
brown	15314	20994	528	100	4136	9	15	194	18	0.2

Table 1b: Standard deviations of estimated elemental content for groups with more than one sample.

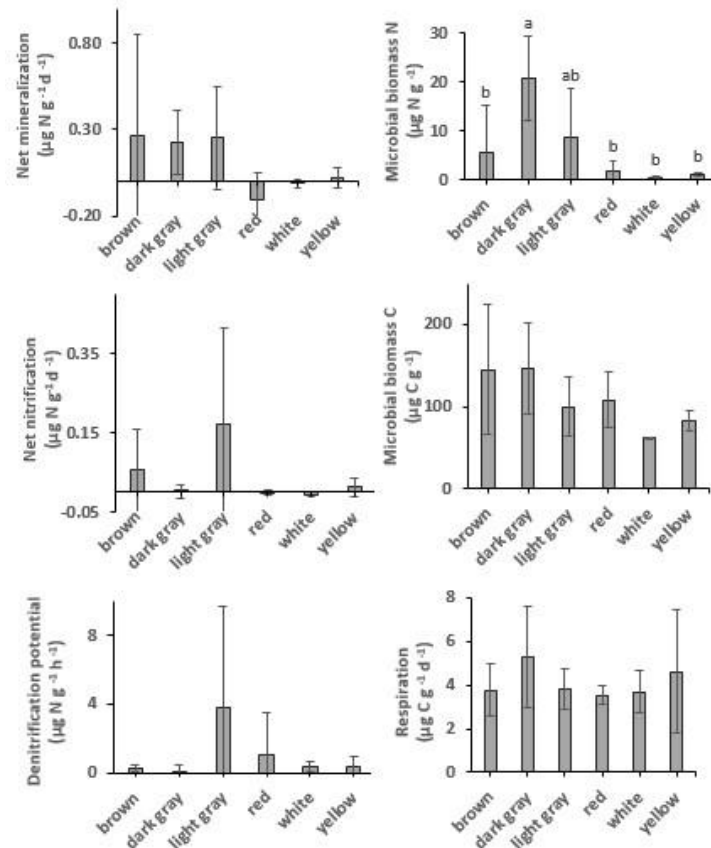


Figure 3: Left column: potential net N mineralization, potential net nitrification, and denitrification potential. Right column top: Microbial biomass N content; values with different superscripts are significantly different at $p < 0.05$ except that dark gray clay was marginally different ($p < 0.10$) from brown and white clays. Right column middle and bottom: Microbial biomass C and respiration. All values are mean $\pm$ SD.

4.2 Do these exposed clays support significant amounts of microbial biomass and activity, i.e., are they alive?

Most clay types exhibited detectable amounts of microbial biomass and activity (Fig. 3). Microbial biomass C was significantly higher (by 135%) in the watery clay than in the packed or sandy clay. A significant difference between watery and sandy clays was identified by ANOVA post hoc analysis. There were no significant differences in microbial biomass C with clay color. Soil respiration was detectable in all materials but there were no significant differences with clay color or texture. There were significant correlations between microbial biomass C and total N, total C (Fig. 4), OM, and respiration. Respiration was significantly correlated with total N (Fig. 4) and OM, and was also marginally correlated with total C and C/N ratio (Fig. 4).

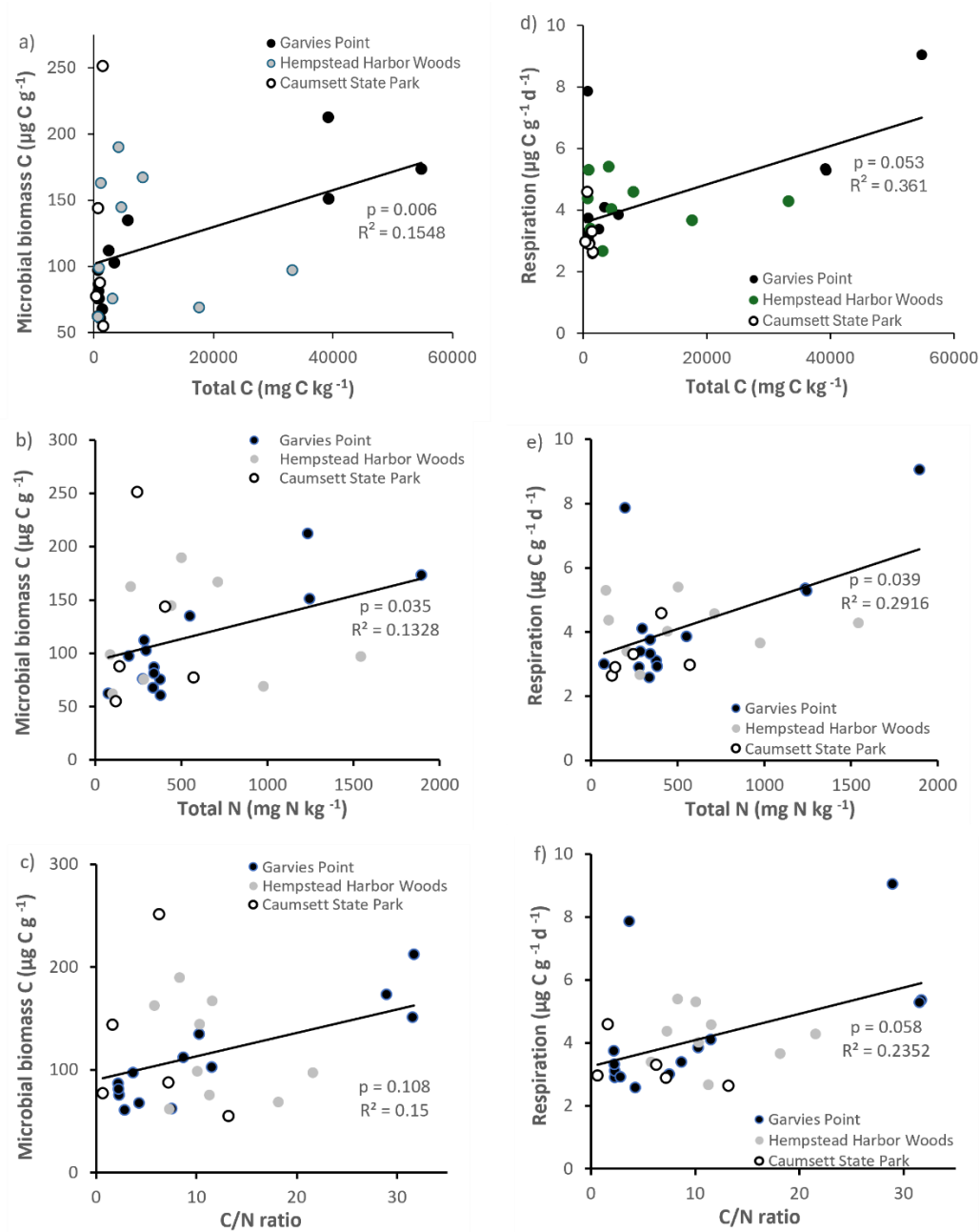


Figure 4: Regression of microbial biomass C and respiration against total C, total N, and C/N ratio. Values are represented for each location; trendline is indicated among all locations. Significant differences are indicated by p value and R^2 .

4.3 Do these clays support significant amounts of N cycle activity?

There were significant differences in microbial biomass N among textures, and amounts were highest in dark gray and packed clays. We detected both potential net N mineralization and N immobilization (negative net N mineralization) in our samples (Fig. 3). Potential net N mineralization was highest in brown and in packed clays,

with significant differences among samples from all locations. Net N mineralization was marginally different between the GP and HHW locations.

315 Microbial biomass N was significantly correlated with total C, total N, C/N ratio (Fig. 6), potential net nitrification, OM, and TIN. Total N was significantly correlated with microbial biomass C, and respiration (Fig. 4), OM, TIN (Fig. 5), microbial biomass N, potential net N mineralization (Fig. 6), and total C. Potential net N mineralization was significantly correlated with total C, total N, C/N ratio (Fig. 6), TIN, microbial biomass N, and potential net nitrification. Total N and the C/N ratio were marginally correlated.

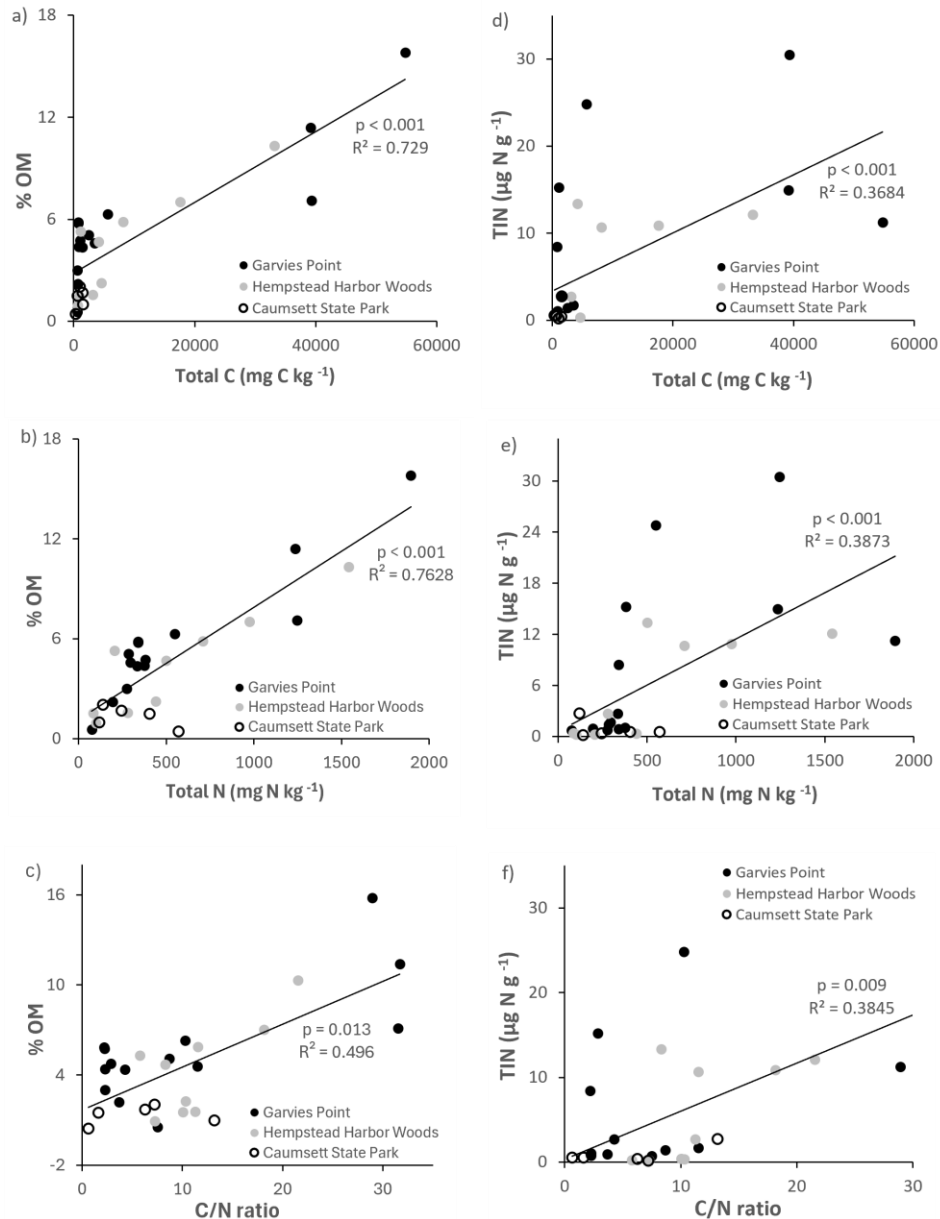


Figure 5: Regression of OM and TIN against total C, total N, and C/N ratio. Values are represented for each location; trendline is indicated among all locations. Significant differences are indicated by p value and R^2 .

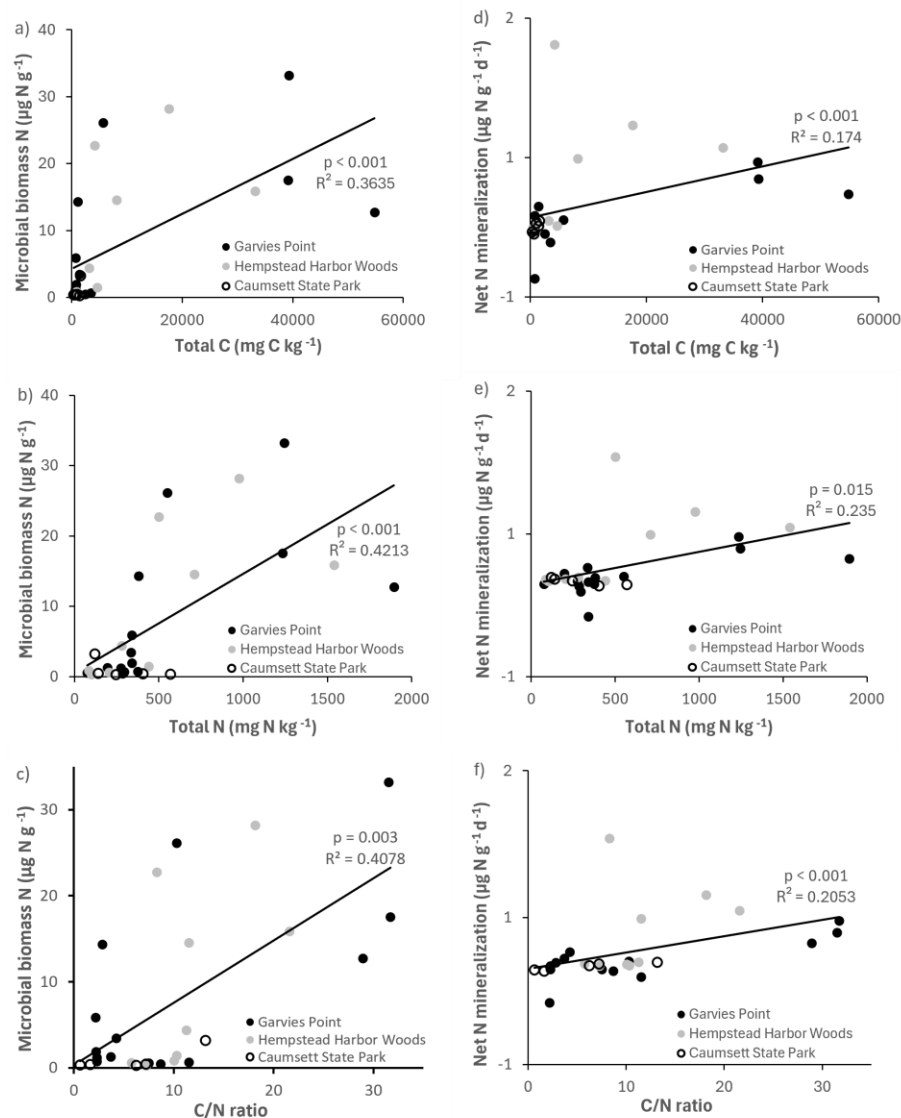


Figure 6: Regression of microbial biomass N and net N mineralization against total C, total N, and C/N ratio. Values are represented for each location; trendline is indicated among all locations. Significant differences are indicated by p value and R^2 .

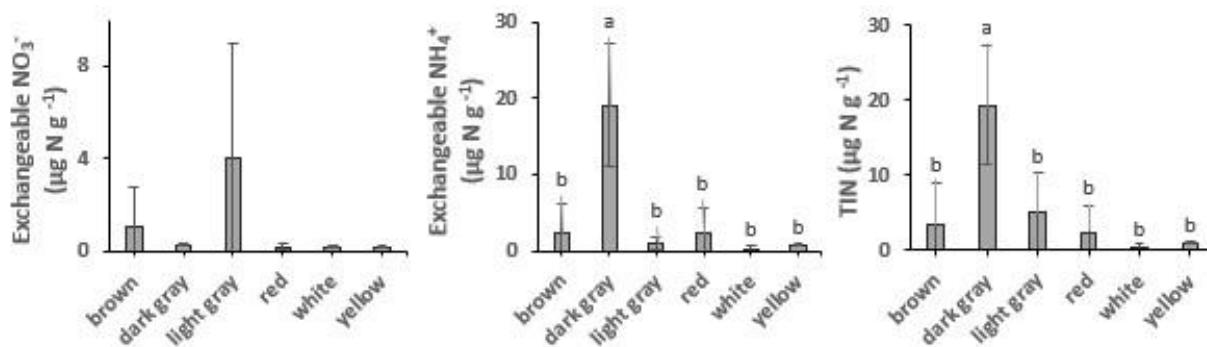


Figure 7: Exchangeable NO_3^- , NH_4^+ , and TIN in clays of different colors. Values are means $\pm$ SD. Values with different superscripts are significantly different at $p < 0.05$.

4.4 Are these clays a potential non-anthropogenic source of reactive N-pollution in the contemporary landscape?

The highest amounts of potential net nitrification were found in light gray (Fig. 3) and packed clays, with significant differences between the GP and HHW locations identified by ANOVA post hoc analysis. (light gray clays at HHW had 100% higher values than the same color group at GP). The highest amounts of NO_3^- were found in light gray (Fig. 7) and packed clays (light gray clays at HHW had 6447% higher values than the same color group at GP), with significant differences between GP and HHW, and between HHW and CSP identified by ANOVA post hoc analysis. The highest amounts of NH_4^+ were found in dark gray (Fig. 7), packed clays at GP, with significant differences among color groups (763% higher amount in dark gray than red clays at GP). The highest amounts of TIN were found in dark gray and packed clays (280% higher than light gray, and 1057% higher than sandy clay, respectively), with significant differences among color and texture groups. Kruskal Wallis analysis identified a marginally significant difference for TIN by location. There were no significant differences in denitrification potential.

Potential net nitrification was significantly correlated with total C, C/N ratio (Fig. 8), microbial biomass N, potential net N mineralization, and marginally correlated with TIN. Denitrification potential was significantly correlated with the C/N ratio (Fig. 8), suggesting joint control by C and N availability. There were significant correlations between NH_4^+ pools and the following variables: total C, total N, C/N ratio (Fig. 9), microbial biomass N, potential net N mineralization, TIN, and OM. NO_3^- pools were significantly correlated with total N (Fig. 9), microbial biomass N, and potential net N mineralization. There was a marginal correlation between NO_3^- and total C (Fig. 9).

5 Discussion

Our study sought to explore organic-mineral interactions by investigating the functional performance of clays through measurements of biogeochemical processes occurring within their structures. This research aimed to expand knowledge beyond the traditional approach of investigating C and N cycle dynamics based on single predictor values such as phyllosilicate clay, abundance of a certain mineral species, and specific surface area and adsorption values (Kleber et al., 2021) by making a suite of measurements of microbial biomass and activity relevant to C and N cycles. This approach allowed us to determine that ancient clays are contributing to contemporary Critical Zone biogeochemical processes at ecosystem and landscape scales.

5.1 N, C, and OM in geological materials

Over geological time scales, N and C cycle from the Earth's surface to depth through subduction zones and are returned to the surface through arc magmatism. Geological cycling of N is greatly facilitated by storage of NH_4^+ in silicate minerals. NH_4^+ in minerals is a major influence on fluxes between reservoirs (Halama and Bebout, 2021), thus giving geological materials such as the clays we have analyzed a potential role in ecosystem function.

For contemporary ecosystem processes, it is generally assumed that geologic materials are not an important source of N (Schlesinger, 2013), as the dominant global pool of N is in the atmosphere. The focus of much N cycle research is on the energetically expensive movement of atmospheric N into biological pools (Galloway et al., 2004). The largest pools of N in ecosystems are in particulate and dissolved OM pools in soils, sediments, and the ocean (Groffman et al., 2021). While it is reasonable to assume that some of the N in our samples could be of modern biological origin and is adsorbed onto the surface of the clay particles, our sampling approach that targeted areas with no evidence of modern OM or soil development suggest that our analysis was of older N associated with geological materials. Increasingly, recognition of the fact that abundant organic N becomes incorporated into geological materials has fueled interest in the role of these materials in contemporary ecosystem processes. Relevant analyses show that 10^{21} g of global fixed N were incorporated into meta-sedimentary mica-rich rocks by the burial of OM in marine and freshwater sediments during the Cretaceous (Morford et al., 2011). These analyses have stimulated studies of the movement of deeply buried organic N into actively cycling pools of N in soils and vegetation (Houlton et al., 2018) and interest in the exposure of ancient materials at the soil surface, such as the clays collected for this study.

At our study sites, burial of OM likely occurred in a Late Cretaceous shallow delta (Fuller, 1914). The N content of these samples is within the range of previous reports of sedimentary and metasedimentary rock N content of 200 – 1200 mg N kg⁻¹ (Holloway and Dahlgren, 2002; Morford et al., 2011). Organic-rich marine sediments commonly exceed 1000 mg N kg⁻¹ and some of our samples fit this criterion (Li, 1991; Morford et al., 2011). As these bedrock materials weather, N is released in plant available forms that stimulate ecosystem productivity and C storage (Dahlgren, 1994; Morford et al., 2011). Bedrock is also a source of N to aquifers, which is relevant for our samples which have a hydrogeologic origin as part of the clay confining units of Long Island’s aquifer system. The samples were collected at exposed aquifer margins in this region with great concern about groundwater ~~reactive N-pollution~~ (Karamouz et al., 2020). The limited literature on the N content of geological materials does not include any studies of clay materials similar to those studied here.

The surface exposure of the clays in our study allowed us to directly measure microbial biomass and activities that are central to biogeochemical cycling of C and N. These measurements shed light on the role that these secondary minerals, that were produced by the weathering of primary silicate minerals which originated at depth, may be playing in the contemporary N cycle on Long Island and in the Critical Zone elsewhere on Earth. These measurements also allowed for comparison of these geological materials with surface and subsurface soils in the region that have been assayed with the same methods (Groffman et al., 2009; Morse et al., 2014). Our analysis showed significant microbial biomass and activity in many samples, with much of the variation in activity driven by the total C and N content of the samples. The results strongly support the idea that ancient geologic materials play a role in contemporary N and C cycling in the Critical Zone.

Biogeochemical processes are influenced by microbial-mineral associations that influence the rates and magnitudes at which biogeochemical reactions occur. The structure of the mineral space influences the extent of biologically

mediated reactions occurring within this habitat. Clays are expected to support less of these processes due to their tightly bound layered structure and fine particle size (Kleber et al., 2021). This is consistent with the lower amounts of activity detected in our samples compared to other soils, discussed in section 5.2. The tight structure of the clay offers a micro-environment that is more constant over time, as compared to the more dynamic conditions found within soil (Kleber et al., 2021). The longevity of a micro-environment is affected by both OM and mineralogy, with kaolinitic 1:1 clays and oxides producing longer lasting environments (Six et al., 2000). The smaller pore size of clay facilitates anaerobic conditions leading to higher denitrification rates in clay rich materials than in sandy ones (Li et al., 2023; Pihlatie et al., 2004). This relationship between particle size and microbial activity was evident in the analysis results obtained from our samples.

What then are the sources of the OC supporting microbial biomass and activity in the samples in this study, which varied with clay color and texture? Plant litter and soil debris from the overlaying deposits are certain contributors of OC to the exposed clays at these locations. Ample C in coarse clay (0.2 – 2.0 μm) has been documented to be in the form of charcoal or black carbon (BC) (Laird et al., 2008). We have observed BC within the clay layers at the GP site, both distinctly layered with white clay and as small inclusions in some of the gray clay, which contains the highest amount of OM at that site (up to 15.8%). This presence indicates that BC is one of the contributing sources of OC and has become incorporated into silt and clay fraction minerals in the geological unit that is exposed at the GP site, through processes such as adsorption of dissolved biogenic compounds onto the clay particle surfaces (Laird et al., 2008).

Clay color is also affected by Fe, which likely contributed to the pigmentation of the red, yellow, and brown clays in our study. Our pXRF scans detected Fe content as high as 4.55 % in some samples. Possible sources of Fe include oxidative reactions and organic ligand bonding, both of which can be catalyzed by bacteria. The Fe content of Long Island's aquifers is driven by processes that include oxidative dissolution of minerals such as pyrite (FeS_2) (Brown and Schoonen, 2004). Pyrite inclusions occur in the clays at these study sites, and our pXRF scans detected the presence of S in addition to the Fe in our samples. Chelation of ions and organic acids is common in the Critical Zone. In the presence of water, Fe is one of the metal ions that can chelate with organic acids and become mobilized through the subsurface (Schroeder, 2018). Furthermore, Fe, Ti, and Mn, all of which were detected in our samples, play a key role in mineral and organic oxidation reactions. Fe(II), FeO_2 , and TiO_2 bearing clays produce the highest amounts of reactive O species, which in turn react with organic C to transform soil and sediment OM, and to produce CO_2 as the end-product of organic C oxidation (Kleber et al., 2021). Mn oxides are the strongest naturally occurring oxidants and play an important role in organic C transformation (Remucal and Ginder-Vogel, 2014). These color variations indicating variation in oxidative or reducing conditions may reflect biogeochemical conditions and activity during their formation that have legacy effects on contemporary activity.

The much higher occurrence of significant differences among the clays when grouped by color and texture, as opposed to when grouped by location, indicates that land use history and other local characteristics have less or no

effect, and that the main differentiator for microbial activity is the clay material itself and variations therein, such as Fe oxides, trace elements, and variations in clay speciation. It also indicates that the clays from the different locations are likely part of the same larger formation (presumably Raritan or Magothy).

5.2 Are these ancient clays contributing to contemporary biogeochemical processes?

Our study was driven by three questions, the first of which was the most fundamental: do these materials support living microbial biomass? Our analysis detected a living microbial community on most of the clay samples. There were strong positive relationships between total C content, total N content, and microbial biomass and activity. The dark gray clays had the highest total N, total C, and microbial biomass N, and microbial biomass C was highest in both dark gray and brown clays. White clays had the lowest contents of these variables, indicating that inclusions of Fe and C (discussed in section 5.1) in the clays play a role in their capacity to sequester C and N and support microbial biomass and activity.

While our clay samples had significant amounts of microbial biomass and activity, it is important to compare our results with other materials and soils in our region (the northeastern U.S.) to evaluate just how important this activity might be. Both surface organic and subsurface mineral horizons of fully developed soils often have substantial C and N pools (Bohlen et al., 2001), with variations in distribution. We compared the data of our clay samples to two studies that used the same analysis methods (Groffman et al., 2009; Morse et al., 2014) (Table 2), to elucidate the differences in microbial biomass and activity between different soils and the clays. In a comparison with forest, agricultural, and grassland soils in the Baltimore, Maryland metropolitan area, respiration and microbial biomass C of the clays (respiration $4.1 \mu\text{g C g}^{-1} \text{ d}^{-1}$; microbial biomass C $114 \mu\text{g C g}^{-1}$; ratio 28) was closest to the agricultural soil (respiration $6.7 \text{ mg C kg}^{-1} \text{ d}^{-1}$; microbial biomass C 224 mg C kg^{-1} ; ratio 33) (Groffman et al., 2009) and lower than the forest or grassland soils.

We further compared our results to soils in various spodic hydopedologic settings in a northern hardwood forest at the Hubbard Brook Experimental Forest, New Hampshire (Morse et al., 2014). These settings included typical podzols (T), bimodal podzols (Bi), Bh podzols (Bh), and seeps, and three depths (Oi/Oe, Oa/A; B horizons) were sampled. Common features of spodosols are complexation with Al and Fe (often organic complexation) and enrichment with Fe and Mn (Van Ranst et al., 2018), thus sharing some elemental features with our clay samples since Al is a major constituent of clay and pXRF scans confirmed the presence of Mn and Fe in our samples.

		Total C	Respiration	MBC	MBN	NH ₄ ⁺	NO ₃ ⁻	TIN	ratio of MBC/Total C	ratio of MBC/Respiration
	Groffman et al., 2009		mg C kg ⁻¹ d ⁻¹	MBC mg C kg ⁻¹		NH ₄ ⁺ mg N kg ⁻¹	NO ₃ ⁻ mg N kg ⁻¹	mg N kg ⁻¹		
	Forest mean		10.3	346		2.1	0.4	2.5		33.592
	Agriculture mean		6.7	224		0.9	8.7	9.6		33.433
	Grassland mean		8.2	306		0.5	1.2	1.7		37.317
	Values source: Table 3									
Morse et al., 2014										
Hydropedologic setting	Horizon	Total C mg C kg ⁻¹		MBC ug C g ⁻¹	MBN ug N g ⁻¹	NH ₄ ⁺ ug N g ⁻¹	NO ₃ ⁻ ug N g ⁻¹	TIN ug N g ⁻¹	ratio of MBC/Total C	
T: typical podzols	Oi/Oe	506000		4780	774	152	16.8	168.8	0.009	
	Oa/A	329000		2320	320	30.4	18.3	48.7	0.007	
	mean of surface Oi/Oe, Oa/A	417500		3550	547	91.2	17.55	108.75	0.008	
	B > 10 cm	74000		481	32.4	2.4	3.33	5.73	0.007	
Bi: bimodal podzols	Oi/Oe	378000		6210	511	83.7	9.6	93.3	0.016	
	Oa/A	158000		1300	226	6.7	9.97	16.67	0.008	
	mean of surface Oi/Oe, Oa/A	268000		3755	368.5	45.2	9.785	54.985	0.012	
	B > 10 cm	56000		275	21.6	2.12	0.53	2.65	0.005	
Bh: Bh podzols	Oi/Oe	443000		8230	710	144	17.8	161.8	0.019	
	Oa/A	234000		3310	333	4.93	15.9	20.83	0.014	
	mean of surface Oi/Oe, Oa/A	338500		5770	521.5	74.465	16.85	91.315	0.016	
	B > 10 cm	60000		569	36	1.46	2.67	4.13	0.009	
Seep	Oa/A	232000		5060	238	9.18	1.02	10.2	0.022	
Values source: Table 1										
This study										
	Total C mg C kg ⁻¹	ug C g ⁻¹ d ⁻¹	MBC ug C g ⁻¹	MBN ug N g ⁻¹	NH ₄ ⁺ ug N g ⁻¹	NO ₃ ⁻ ug N g ⁻¹	TIN ug N g ⁻¹	ratio of MBC/Total C	ratio of MBC/Respiration	
	All locations, surface, mean	8220.18	4.1	113.53	7.64	4.63	1.47	6.11	0.014	27.690
HPS: hydropedologic setting, T: typical podzols, Bi: bimodal podzols, and Bh: Bh podzols; MBC: microbial biomass C, MBN: microbial biomass N.										

Table 2: comparison of response variables from this study to data from Groffman et al., 2009 and Morse et al., 2014.

Our samples, all of which were collected from the surface, were most comparable to the B horizons and to the Bi setting from Morse et al. (2014) (Table 2). B horizons in the Bi setting soils had the lowest total C and microbial biomass C and N contents of the Hubbard Brook soils but were still much higher than our clay values. The clay had a higher microbial biomass C / total C ratio (0.014) than the surface (0.012) and B (0.005) horizons at Hubbard Brook suggesting relatively high OM quality in the clays.

Morse et al., (2014), found that differences in total C and N content were more notable across soil horizons than across hydropedologic setting, where the B horizon, which accumulated Al and Fe, had less C and N and microbial biomass and activity. The dominance of chemical (OM quality) versus location controls is consistent with the stronger differences that we observed with color than with location. The composition of OM in clay fractions differs from that in sand and silt fractions, and there is usually a decrease in C/N ratio as particle size decreases from coarse silt to fine clay (Laird et al., 2008). In clay, the packed textures dominated by smaller particle size contain more OM than larger particle sandy samples, but the C/N ratio increases with smaller particle size. In our samples, dark gray clays had the highest total N, total C, and microbial biomass N; microbial biomass C was highest in both dark gray and brown clays. White clays had the lowest contents of these variables, indicating that inclusions (such as Fe, BC, and others) in the clays play a role in increasing their capacity to accommodate C and N rich contents.

The second question our study addressed is whether the clays support an active N cycle. We measured several indices of N cycling activity that showed that the exposed clays do indeed support and active N cycle and have potential to supply N to support plant growth. Microbial biomass N is an index of the size of the actively cycling labile N pool in soil. Mineralization, the production of simple, soluble, inorganic forms of N that are a dominant source of N for plant growth, is strongly tied to the C cycle (Hart et al., 1994). When microbes degrade N containing compounds during mineralization, their N is converted to proteins which release ammonia (NH₃) that converts to NH₄⁺, and their C is converted to biomass or CO₂ (Groffman et al., 2021).

495 Our findings have general similarities to those of Morse et al. (2014), who found that clay rich B horizons have lower rates of biogeochemical activity (lower net N mineralization and net nitrification potentials) as well as smaller C and N pools than surface soils with less clay and Al. Clay minerals play a role in the C and N adsorption and stabilization in soil. The storage potential of C is influenced by the size of the silicate mineral's surface area and the amount of cations adhering to these minerals (Kahle et al., 2002). Adsorption rates and amounts of OM on mineral

500 surfaces are influenced by variations in aqueous solution dynamics, mineralogy, and OM chemistry, while OM affects mineral growth, transformation, and dissolution (Kleber et al., 2021).

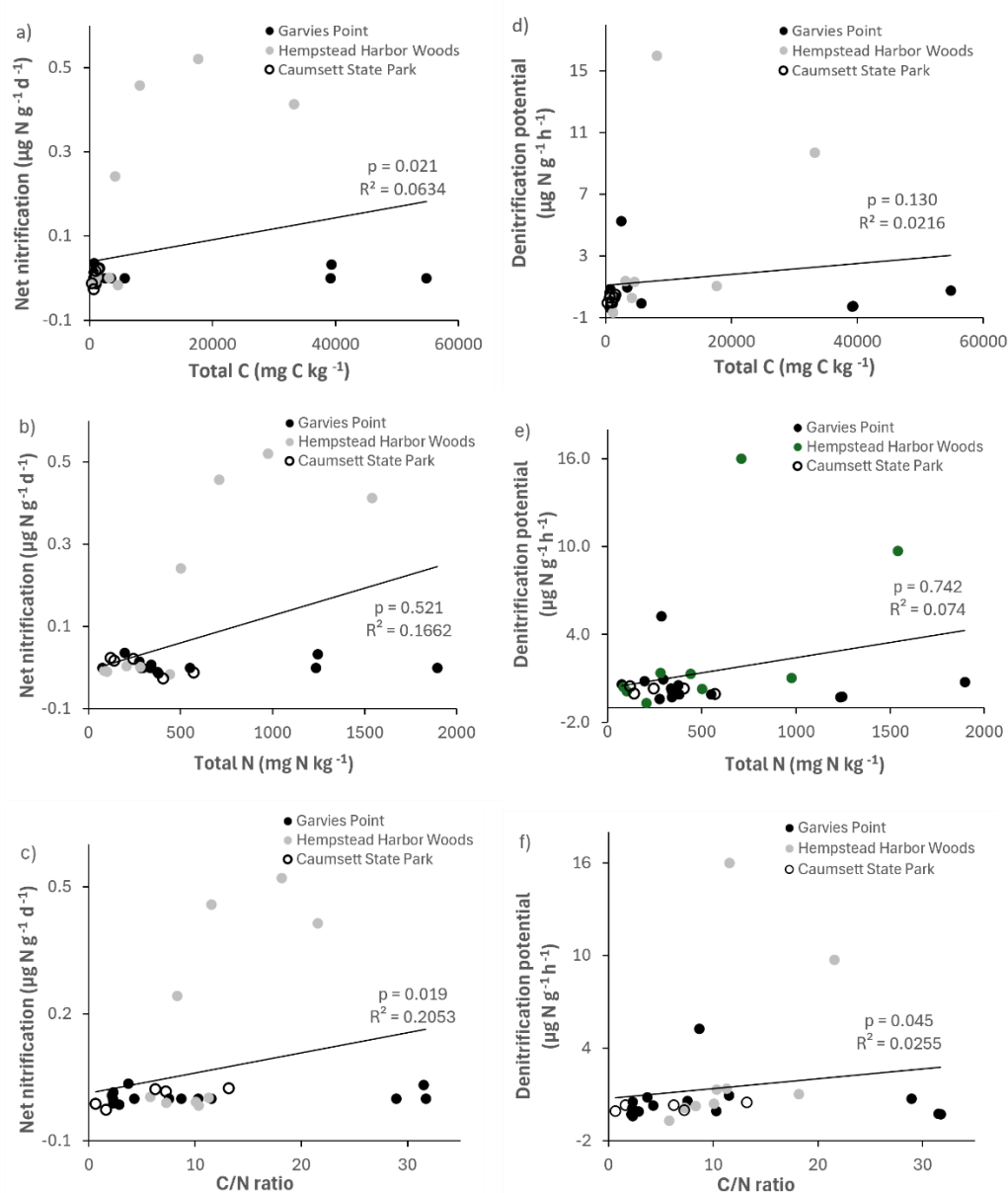


Figure 8: Regression of net nitrification and denitrification potential against total C, total N, and C/N ratio. Values are represented for each location; trendline is indicated among all locations. Significant differences are indicated by p value and R².

The third question we asked is whether the clays are a non-anthropogenic source of reactive N-pollution. There is great concern about N pollution of groundwater and coastal waters in our region of study (Karamouz et al., 2020).

There is particular concern about NO₃⁻, the most highly mobile form of reactive N that is a drinking water pollutant and a prime cause of eutrophication in coastal waters (Conley et al., 2009). We therefore assessed the potential of these clays to contribute to high levels of NO₃⁻ in the environment by measuring both NO₃⁻ pools as well as processes that produce (nitrification) and consume (denitrification) NO₃⁻.

Nitrification is carried out by chemoautotrophic bacteria that oxidize NH₄⁺ into NO₂⁻ which is further oxidized into NO₃⁻. When this process is stimulated by the addition of N through application of fertilizers, atmospheric deposition, groundwater and runoff sources, it can lead to excessive production of NO₃⁻ (Groffman et al., 2021).

Denitrification is an anaerobic process that converts NO₃⁻ to gaseous forms NO, N₂O, N₂ (Robertson and Groffman, 2015), removing reactive N from the soil and facilitating the cycling of N between the biosphere and lithosphere to the atmosphere.

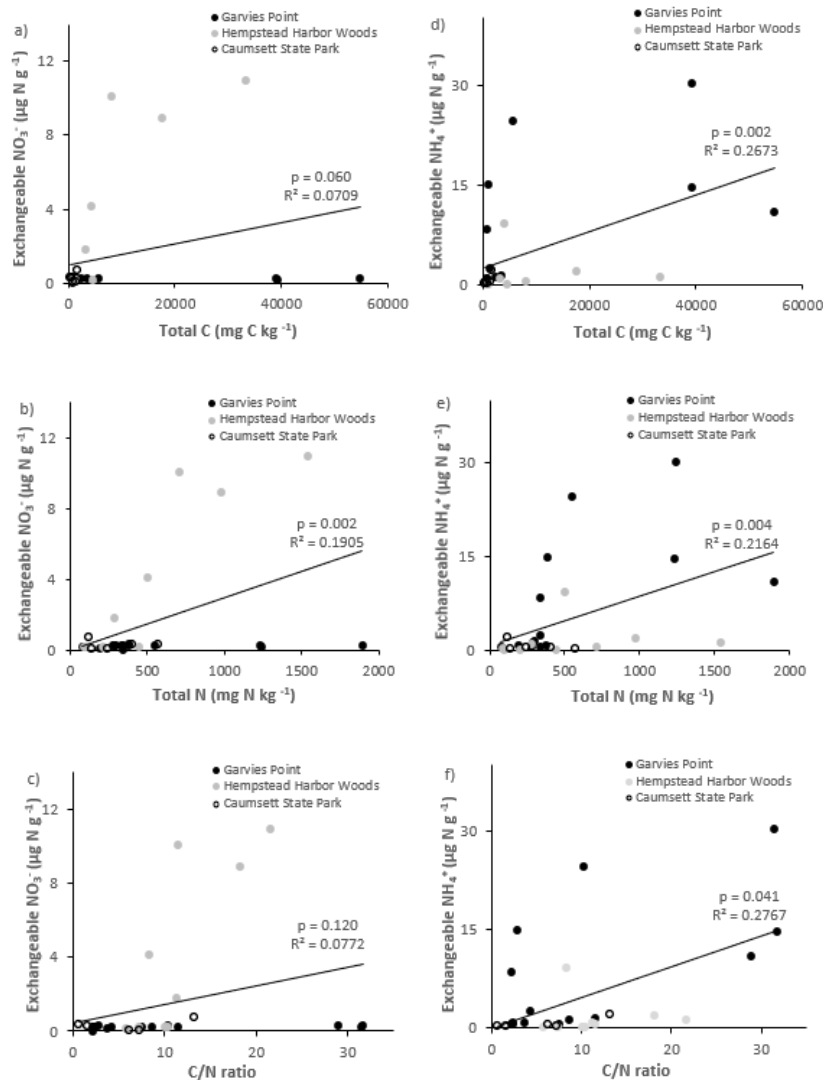


Figure 9: Regression of exchangeable NO_3^- and NH_4^+ against total C, total N, and C/N ratio. Values are represented for each location; trendline is indicated among all locations. Significant differences are indicated by p value and R^2 .

5.3 N Pollution due to reactive N species

N pollution occurs in the air, soil, and water. In the air it is the addition of volatile forms of N gasses, such as NO_y caused by industrial and commercial activity; in soil there is often an overload of nutrients from overuse of fertilizers and deposition from the atmosphere; in water a major cause is sewage system drainage and agricultural runoff, causing eutrophication which leads to dead zones in the oceans, in the most extreme scenario. The main source for N pollution (in the form of NO_3^-) to Long Island's aquifers is from septic systems, with additional inputs from agriculture, lawn care, and atmospheric deposition (Szymczycha et al., 2017). NO_3^- fluxes in Long Island's aquifers can negatively impact coastal ecosystems (Karamouz et al., 2020) and drinking water quality. The clay

units included in Long Island's aquifer and aquitard strata can act as confining units, and as ~~sinks or~~ non-anthropogenic ~~sinks or~~ sources of reactive N ~~pollution~~.

The clays examined in this study could act as a minor source or a potentially significant sink for reactive N ~~pollution~~ in an aquifer system. Clays with significant rates of potential net nitrification (especially the light gray clay) could be a source of NO_3^- if they were in contact with the aquifer. However, rates of denitrification potential were generally much higher than potential net nitrification, suggesting that these clay materials are more likely to act as NO_3^- sinks. The light gray clays have the potential to act as both NO_3^- sources and sinks depending on environmental conditions, while the white and Fe-bearing (red, yellow) clays have the potential to act as NO_3^- sinks.

6 Conclusions

This investigation shed light on the amount and type of microbial activity that occurs in geological microhabitats in the Critical Zone at a coastal exposure of temperate northern latitude and allowed us to evaluate their potential local and regional impact.

Our analyses have taken a step towards better understanding the nature and extent of the biogeochemical activity that these types of microhabitats support. The approach of using laboratory measurements of microbial biomass and activity in ancient materials was successful in characterizing the biogeochemical potential of these materials, even at low levels, and could be applied in other Critical Zone studies.

The results from this study provide quantitative data showing microbial activity of silt and clay fraction materials of hydrogeologic origin, and confirm that these materials contain ecologically significant concentrations of geologic N exceeding $1000 \text{ mg N kg}^{-1}$ (Holloway and Dahlgren, 2002). Their surface exposure allowed us to explore the interaction between ancient geological stratigraphic components and modern day environmental conditions. This type of interaction is illustrative of the conceptual reach of Critical Zone science and the cohesive understanding of multiple differing factors that it provides. The unity of various disciplines and their individual approaches to investigation allow for greater understanding of the ensuing conditions that occur in the Critical Zone, where the resulting processes work together to support all living organisms.

Our results advance the emerging science of the geological N cycle and clearly show that ancient geological materials are contributing to contemporary biogeochemical processes in the Critical Zone of our study region. Further, we have shown that these materials support a wide range of N cycle processes encompassing mineralization, immobilization, nitrification, and denitrification. There is a clear need for further research on how clay physical and chemical characteristics influence the flows of N to and from clays and biogeochemical processes in the Critical Zone, and to see how these materials and processes are contributing to the growth of vegetation and the dynamics of pollutants in the Critical Zone of this dynamic, densely populated, and environmentally sensitive region.

Author contributions

570 VMA conceptualized the study, performed field work and sample collection, laboratory analyses, data analysis, and wrote the manuscript; PMG designed the study, provided laboratory methodology, access, and supplies, interpreted results, co-wrote, and revised the manuscript; ZC provided laboratory equipment and access, interpreted results, and revised and edited the manuscript; DES interpreted results, revised and edited the manuscript.

575 Competing interests

None of the authors have any competing interests.

Acknowledgements

The authors thank Clare Kohler and Kaitlin McLaughlin for help with laboratory analyses; and Veronica Natale and
580 Dr. Herbert Mills for help obtaining samples.

References

- Bohlen, P. J., Groffman, P. M., Driscoll, C. T., Fahey, T. J., and Siccama, T. G.: PLANT–SOIL–MICROBIAL
585 INTERACTIONS IN A NORTHERN HARDWOOD FOREST, *Ecology*, 82, 965–978, [https://doi.org/10.1890/0012-9658\(2001\)082\[0965:PSMIIA\]2.0.CO;2](https://doi.org/10.1890/0012-9658(2001)082[0965:PSMIIA]2.0.CO;2), 2001.
- Brantley, S. L., White, T. S., White, A. F., Sparks, D. L., Richter, D., Pregitzer, K. S., Derry, L. A., Chorover, J., Chadwick, O. A., April, R., Anderson, S. P., and Amundson, R. C.: Frontiers in exploration of the critical zone. Report of a Workshop Sponsored by the National Science Foundation(NSF), Newark, DE, 30, 2006.
- 590 Brown, C. J. and Schoonen, M. A. A.: The origin of high sulfate concentrations in a coastal plain aquifer, Long Island, New York, *Appl. Geochem.*, 19, 343–358, [https://doi.org/10.1016/S0883-2927\(03\)00154-9](https://doi.org/10.1016/S0883-2927(03)00154-9), 2004.
- Busigny, V. and Bebout, G. E.: Nitrogen in the Silicate Earth: Speciation and Isotopic Behavior during Mineral-Fluid Interactions, *Elements*, 9, 353–358, <https://doi.org/10.2113/gselements.9.5.353>, 2013.
- Conley, D. J., Paerl, H. W., Howarth, R. W., Boesch, D. F., Seitzinger, S. P., Havens, K. E., Lancelot, C., and
595 Likens, G. E.: Controlling Eutrophication: Nitrogen and Phosphorus, *Science*, 323, 1014–1015, <https://doi.org/10.1126/science.1167755>, 2009.
- Dahlgren, R. A.: Soil acidification and nitrogen saturation from weathering of ammonium-bearing rock, *Nature*, 368, 838–841, <https://doi.org/10.1038/368838a0>, 1994.
- 600 Dass, P., Houlton, B. Z., Wang, Y., Wårlind, D., and Morford, S.: Bedrock Weathering Controls on Terrestrial Carbon-Nitrogen-Climate Interactions, *Glob. Biogeochem. Cycles*, 35, e2020GB006933, <https://doi.org/10.1029/2020GB006933>, 2021.

- Davey, B. G., Russell, J. D., and Wilson, M. J.: Iron oxide and clay minerals and their relation to colours of red and yellow podzolic soils near Sydney, Australia, *Geoderma*, 14, 125–138, [https://doi.org/10.1016/0016-7061\(75\)90071-3](https://doi.org/10.1016/0016-7061(75)90071-3), 1975.
- 605 Doane, T. A. and Horwath, W. R.: Spectrophotometric Determination of Nitrate with a Single Reagent, *Anal. Lett.*, 36, 2713–2722, <https://doi.org/10.1081/AL-120024647>, 2003.
- Downey, A. E., Groffman, P. M., Mejía, G. A., Cook, E. M., Sritairat, S., Karty, R., Palmer, M. I., and McPhearson, T.: Soil carbon sequestration in urban afforestation sites in New York City, *Urban For. Urban Green.*, 65, 127342, <https://doi.org/10.1016/j.ufug.2021.127342>, 2021.
- 610 Eby, N.: Principles of environmental geochemistry, Waveland Press, Inc, Long Grove, Illinois, 514 pp., 2016.
- Fuller, M.: *The Geology of Long Island, New York*. Myron L. Fuller, *J. Geol.*, 24, 303–304, <https://doi.org/10.1086/622335>, 1914.
- Galloway, J. N., Dentener, F. J., Capone, D. G., Boyer, E. W., Howarth, R. W., Seitzinger, S. P., Asner, G. P., Cleveland, C. C., Green, P. A., Holland, E. A., Karl, D. M., Michaels, A. F., Porter, J. H., Townsend, A. R., and Vorosmarty, C. J.: Nitrogen Cycles: Past, Present, and Future, *Biogeochemistry*, 70, 153–226, <https://doi.org/10.1007/s10533-004-0370-0>, 2004.
- 615 Groffman, P. M., Holland, E. A., Myrold, D. D., Robertson, G. P., and Zou, X.: “Denitrification.” In *Standard soil methods for long-term ecological research*, Oxford University Press, New York, 272–288 pp., 1999.
- Groffman, P. M., Williams, C. O., Pouyat, R. V., Band, L. E., and Yesilonis, I. D.: Nitrate Leaching and Nitrous Oxide Flux in Urban Forests and Grasslands, *J. Environ. Qual.*, 38, 1848–1860, <https://doi.org/10.2134/jeq2008.0521>, 2009.
- 620 Groffman, P. M., Rosi, E. J., and Fulweiler, R. W.: The nitrogen cycle. In K. C. Weathers, D. L. Strayer, & G. E. Likens (Eds.), in: *Fundamentals of Ecosystem Science*, Academic Press, 161–188, 2021.
- Halama, R. and Bebout, G.: Earth’s Nitrogen and Carbon Cycles, *Space Sci. Rev.*, 217, 45, <https://doi.org/10.1007/s11214-021-00826-7>, 2021.
- 625 Hart, S. C., Nason, G. E., Myrold, D. D., and Perry, D. A.: Dynamics of Gross Nitrogen Transformations in an Old-Growth Forest: The Carbon Connection, *Ecology*, 75, 880–891, <https://doi.org/10.2307/1939413>, 1994.
- Holloway, J. M. and Dahlgren, R. A.: Nitrogen in rock: Occurrences and biogeochemical implications: BIOGEOCHEMICAL IMPLICATIONS OF N IN ROCK, *Glob. Biogeochem. Cycles*, 16, 65-1-65–17, <https://doi.org/10.1029/2002GB001862>, 2002.
- 630 Houlton, B. Z., Morford, S. L., and Dahlgren, R. A.: Convergent evidence for widespread rock nitrogen sources in Earth’s surface environment, *Science*, 360, 58–62, <https://doi.org/10.1126/science.aan4399>, 2018.
- Jakobsson, M., Løvlie, R., Al-Hanbali, H., Arnold, E., Backman, J., and Mörtz, M.: Manganese and color cycles in Arctic Ocean sediments constrain Pleistocene chronology, *Geology*, 28, 23, [https://doi.org/10.1130/0091-7613\(2000\)28<23:MACCIA>2.0.CO;2](https://doi.org/10.1130/0091-7613(2000)28<23:MACCIA>2.0.CO;2), 2000.
- 635 Jenkinson, D. S. and Powlson, D. S.: The effects of biocidal treatments on metabolism in soil—V, *Soil Biol. Biochem.*, 8, 209–213, [https://doi.org/10.1016/0038-0717\(76\)90005-5](https://doi.org/10.1016/0038-0717(76)90005-5), 1976.
- Kahle, M., Kleber, M., and Jahn, R.: Review of XRD-based quantitative analyses of clay minerals in soils: the suitability of mineral intensity factors, *Geoderma*, 109, 191–205, [https://doi.org/10.1016/S0016-7061\(02\)00175-1](https://doi.org/10.1016/S0016-7061(02)00175-1), 2002.
- 640

- Karamouz, M., Mahmoodzadeh, D., and Oude Essink, G. H. P.: A risk-based groundwater modeling framework in coastal aquifers: a case study on Long Island, New York, USA, *Hydrogeol. J.*, 28, 2519–2541, <https://doi.org/10.1007/s10040-020-02197-9>, 2020.
- 645 Kleber, M., Bourg, I. C., Coward, E. K., Hansel, C. M., Myneni, S. C. B., and Nunan, N.: Dynamic interactions at the mineral–organic matter interface, *Nat. Rev. Earth Environ.*, 2, 402–421, <https://doi.org/10.1038/s43017-021-00162-y>, 2021.
- Laird, D. A., Chappell, M. A., Martens, D. A., Wershaw, R. L., and Thompson, M.: Distinguishing black carbon from biogenic humic substances in soil clay fractions, *Geoderma*, 143, 115–122, <https://doi.org/10.1016/j.geoderma.2007.10.025>, 2008.
- 650 Li, L., Shields, J., Snow, D. D., Kaiser, M., and Malakar, A.: Labile carbon and soil texture control nitrogen transformation in deep vadose zone, *Sci. Total Environ.*, 878, 163075, <https://doi.org/10.1016/j.scitotenv.2023.163075>, 2023.
- Li, Y. H.: Distribution patterns of the elements in the ocean: A synthesis, *Geochim. Cosmochim. Acta*, 55, 3223–3240, [https://doi.org/10.1016/0016-7037\(91\)90485-N](https://doi.org/10.1016/0016-7037(91)90485-N), 1991.
- 655 Liebling, R. S.: Clay Minerals of the Weathered Bedrock Underlying Coastal New York, *Geol. Soc. Am. Bull.*, 84, 1813, [https://doi.org/10.1130/0016-7606\(1973\)84<1813:CMOTWB>2.0.CO;2](https://doi.org/10.1130/0016-7606(1973)84<1813:CMOTWB>2.0.CO;2), 1973.
- Mills, H. C. and Wells, P. D.: Ice-Shove Deformation and Glacial Stratigraphy of Port Washington, Long Island, New York, *Geol. Soc. Am. Bull.*, 85, 357, [https://doi.org/10.1130/0016-7606\(1974\)85<357:IDAGSO>2.0.CO;2](https://doi.org/10.1130/0016-7606(1974)85<357:IDAGSO>2.0.CO;2), 1974.
- 660 Morford, S. L., Houlton, B. Z., and Dahlgren, R. A.: Increased forest ecosystem carbon and nitrogen storage from nitrogen rich bedrock, *Nature*, 477, 78–81, <https://doi.org/10.1038/nature10415>, 2011.
- Morse, J. L., Werner, S. F., Gillin, C. P., Goodale, C. L., Bailey, S. W., McGuire, K. J., and Groffman, P. M.: Searching for biogeochemical hot spots in three dimensions: Soil C and N cycling in hydropedologic settings in a northern hardwood forest: Biogeochemical hotspots in soil profiles, *J. Geophys. Res. Biogeosciences*, 119, 1596–1607, <https://doi.org/10.1002/2013JG002589>, 2014.
- 665 P. C. Bennett, Rogers, J. R., Chol, W. J., and Hiebert, F. K.: Silicates, Silicate Weathering, and Microbial Ecology, *Geomicrobiol. J.*, 18, 3–19, <https://doi.org/10.1080/01490450151079734>, 2001.
- Paul, E. A.: Soil microbiology, ecology, and biochemistry, Fourth edition., Academic Press, Amsterdam, 2014.
- 670 Pihlatie, M., Syväsalö, E., Simojoki, A., Esala, M., and Regina, K.: Contribution of nitrification and denitrification to N₂O production in peat, clay and loamy sand soils under different soil moisture conditions, *Nutr. Cycl. Agroecosystems*, 70, 135–141, <https://doi.org/10.1023/B:FRES.0000048475.81211.3c>, 2004.
- Remucal, C. K. and Ginder-Vogel, M.: A critical review of the reactivity of manganese oxides with organic contaminants, *Environ. Sci. Process. Impacts*, 16, 1247, <https://doi.org/10.1039/c3em00703k>, 2014.
- 675 Robertson, G. P. (Ed.): Standard soil methods for long-term ecological research, Oxford University Press, New York, 462 pp., 1999.
- Robertson, G. P. and Groffman, P. M.: Nitrogen transformations., in: *Soil Microbiology, Ecology, and Biogeochemistry*, Academic Press, Burlington, MA, 421–446, 2015.
- 680 Saidy, A. R., Smernik, R. J., Baldock, J. A., Kaiser, K., and Sanderman, J.: The sorption of organic carbon onto differing clay minerals in the presence and absence of hydrous iron oxide, *Geoderma*, 209–210, 15–21, <https://doi.org/10.1016/j.geoderma.2013.05.026>, 2013.

Schlesinger, W. H.: An estimate of the global sink for nitrous oxide in soils, *Glob. Change Biol.*, 19, 2929–2931, <https://doi.org/10.1111/gcb.12239>, 2013.

Schroeder, P. A.: *Clays in the Critical Zone*, Cambridge university press, Cambridge New York, 2018.

685 Seitzinger, S., Harrison, J. A., Böhlke, J. K., Bouwman, A. F., Lowrance, R., Peterson, B., Tobias, C., and Drecht, G. V.: DENITRIFICATION ACROSS LANDSCAPES AND WATERSCAPES: A SYNTHESIS, *Ecol. Appl.*, 16, 2064–2090, [https://doi.org/10.1890/1051-0761\(2006\)016\[2064:DALAWA\]2.0.CO;2](https://doi.org/10.1890/1051-0761(2006)016[2064:DALAWA]2.0.CO;2), 2006.

Sims, G. K., Ellsworth, T. R., and Mulvaney, R. L.: Microscale determination of inorganic nitrogen in water and soil extracts, *Commun. Soil Sci. Plant Anal.*, 26, 303–316, <https://doi.org/10.1080/00103629509369298>, 1995.

Sirkin, L.: Stratigraphy of the Long Island Platform, *J. Coast. Res.*, 217–227, 1991.

690 Six, J., Elliott, E. T., and Paustian, K.: Soil Structure and Soil Organic Matter II. A Normalized Stability Index and the Effect of Mineralogy, *Soil Sci. Soc. Am. J.*, 64, 1042–1049, <https://doi.org/10.2136/sssaj2000.6431042x>, 2000.

Smith, M. S. and Tiedje, J. M.: Phases of denitrification following oxygen depletion in soil, *Soil Biol. Biochem.*, 11, 261–267, [https://doi.org/10.1016/0038-0717\(79\)90071-3](https://doi.org/10.1016/0038-0717(79)90071-3), 1979.

695 Swarzenski, W. V.: *Hydrogeology of Northwestern Nassau and Northeastern Queens counties, Long Island, New York*, USGS Geological Survey Water Supply Paper 1657, 88 pp., 1963.

Szymczycha, B., Kroeger, K. D., Crusius, J., and Bratton, J. F.: Depth of the vadose zone controls aquifer biogeochemical conditions and extent of anthropogenic nitrogen removal, *Water Res.*, 123, 794–801, <https://doi.org/10.1016/j.watres.2017.06.048>, 2017.

700 Van Ranst, E., Wilson, M. A., and Righi, D.: Spodic Materials, in: *Interpretation of Micromorphological Features of Soils and Regoliths*, Elsevier, 633–662, <https://doi.org/10.1016/B978-0-444-63522-8.00022-X>, 2018.