We sincerely thank Referee 1 for the valuable comments we used to improve the quality of this manuscript. According to these suggestions, we have supplemented several references and corrected some mistakes in our previous draft. These are listed below.

However, I have a major concern on the quality of the data and thus the resulting conclusions. My concern primarily arises from the extremely sparse sampling resolutions: only one time point (late afternoon and high tide) on two days for CO concentration sampling and only one station (Sta. 7) for CO consumption sampling. It is well known that surface water CO concentration changes substantially over diel time scales mainly imposed by daytime photoproduction and bacterial consumption throughout day and night. This diel cycle can be further complicated by tidal variation in coastal areas. It is also well recognized that microbial CO consumption presents large spatiotemporal variations in coastal regions. The data obtained from such poor sampling resolutions can only capture a slim part of the diel cycle and thus cannot support the conclusions reported. For example, if we use the K_{bio} determined at Sta. 7 (0.40 h⁻¹) to estimate the surface water CO concentration in early morning (before dawn, assuming 7 hours of nighttime) at this same locality, we get 0.04 nM, which is below the CO concentration at equilibrium with air (~0.1 nM). Then, the late-afternoon source turns into a nighttime sink. This situation may apply to many other stations, because the reported surface water CO concentrations were mostly quite low.

Reply: Thank you for pointing out this issue. It would have been better if more sampling sites with a higher frequency were sampled to confirm the conclusion of our study. Nevertheless, we think our spatial resolution is adequate to capture the environmental gradients of relevance for CO cycling in the study area. By collecting data at the same specific locations in two days, we can ensure that the data was obtained under similar environmental conditions in the area of influence of low human activities, reducing uncertainties caused by site variations. In the WWTP effluent areas, samples were collected at the same stations at two time periods, both when no effluent was discharged and when effluent was discharged, to better determine its influence on CO. Besides, the sampling time was limited by the tide and the logistical constrain; it was imperative to consider minimizing the interval between sampling and measurement to minimize CO sample uptake within the vial and microbial consumption processes.

The consumption sampling station was located in a representative area for the combined effects of tidal effects and terrestrial contributions, and therefore we considered it to be a sampling site that may feature a good approximation of the rate of microbial consumption in this region.

The second important factor that led to me casting doubt on the quality of the data is the huge correction factor (3.12) for CO concentrations. There is no explanation for what caused the technical problem leading to this correction. Any time- and concentration-dependence of the correction factor? The uncertainty of your atmospheric CO measurement was 14.2%, which alone leads to significant variations in the correction factor. This variability can be larger if the uncertainties of the atmospheric CO measurements at Mace Head and Terceira are taken into account. Please present an uncertainty analysis of this correction.

Reply: Please see the reply to the specific comment on this.

In addition, there are errors in the calculation of CO concentrations (eq. 1, see specific comments on this).

Reply: Please see the reply to the specific comment on this.

The CO budget (eq. 11) is unreliable because the poor sampling resolutions invalidate the steadystate assumption. Moreover, the budgeting misses a potentially important source (terrestrial input including the WWTP effluent) and a potentially important sink (output to the outside sea).

Reply: We disagree. The low resolution might impact the robustness of the results but not necessarily the steady-state assumption. In this study, all the sources and sinks mentioned were considered to be a sort of vertical transport process. The lateral terrestrial input (including WWTP effluent) with high FDOM and nutrients contributed to vertical processes like CO photodegradation, biological production, and microbial consumption. Budgets for the lateral transport processes are indeed very helpful to a complete understanding of their area. However, they were not part of the study presented here.

The interpretation of the data from the incubations of the aquaculture is superficial and often inaccurate or incorrect (see specific comments). In fact, the data allows you to derive the gross production in the "light" treatment.

Reply: Please see the reply to the specific comment on this.

Some suggestions:

Make sure that the calculations of and corrections on CO concentrations are correct.

Base your conclusions on the data. Do not over-extrapolate temporally and/or spatially.

Reply: Many thanks for your suggestions, and we have replied to each specific comment individually.

Specific comments:

Introduction: Put your study in a broad context: why do you choose a human-impacted lagoon system?

Reply: According to the comment, we integrated the last paragraph of the introduction:

Ria Formosa is a major lagoon system in southern Portugal ($36^{\circ}58$ ' N, $8^{\circ}02$ ' W to $37^{\circ}03$ ' N, $7^{\circ}32$ ' W) (Newton and Mudge, 2003; Cravo et al., 2014). It is a mesotidal coastal lagoon with an average water depth of ~2 m. Water temperatures typically range from about 12 °C in the winter to about 27 °C in the summer (Newton and Mudge, 2003). Salinities range from 13 to 36.5 (Newton and Mudge, 2003). The lagoon is vertically well-mixed due to limited freshwater inputs and the predominance of a tidal-forced circulation pattern (Cravo et al., 2014). Being a typical coastal lagoon system, the water exchange with the adjacent shelf is restricted (Tett et al., 2003). In addition

to nutrient inputs from aquaculture facilities, the Ria Formosa Lagoon receives significant nutrient inputs from agricultural runoff and sewage, particularly from the cities of Faro, Olhão, and Tavira (Newton and Mudge, 2005). A part of the Ria Formosa Lagoon system is a natural park established in 1987 (Aníbal et al., 2019). Ria Formosa also plays a vital role in the region's tourism industry, as well as supporting a fishery and aquaculture industry of national significance. Due to its restricted (estuarine-type) circulation and high nutrient and organic matter inputs, the Ria Formosa Lagoon system could be a potential source of atmospheric CO. The unique setting at the Ria Formosa lagoon system provides an excellent opportunity to investigate the dynamics of carbon monoxide in an anthropogenically-influenced coastal system.

Additionally, eutrophication and algal bloom often occur in aquaculture ponds in Ria Formosa, which should be a non-negligible component of the RF-CO cycle process. How these environmental factors will affect CO production and emissions from the lagoon system (accounting for approximately 13% of the coastal areas worldwide) is unknown due to limited measurements and knowledge gaps regarding its sources and sinks. In this study, we present the first in situ observations of CO in the region, which we combined with incubation experiments in waters directly influenced by anthropogenic activities. The main objectives of this study were to i) elucidate the CO distribution and sea–air flux densities in the coastal lagoon system, ii) estimate the different CO sources and sinks in the lagoon system during summer, and iii) determine the potential impact of aquaculture activities on CO cycling in this region.

L32: Ossola et al. (2022) used model compounds not natural CDOM and their experiments were conducted at pH 5.8-6.5, conditions atypical of coastal and open oceans. Replace it with a more appropriate reference.

Reply: Ossola et al. (2022) was replaced with Stubbins et al. (2006).

Integrate "Study site description" into the last paragraph of the "Introduction". Elaborate more about the human activities (including aquaculture) in the region.

Reply: See our first reply to the specific comment, and we added more details about human activities (including aquaculture) in the region: In addition to nutrient inputs from aquaculture facilities, the Ria Formosa Lagoon receives significant nutrient inputs from agricultural runoff and sewage, particularly from the cities of Faro, Olhão, and Tavira (Newton and Mudge, 2005). A part of the Ria Formosa Lagoon system is a natural park established in 1987 (Aníbal et al., 2019). Ria Formosa also plays a vital role in the region's tourism industry, as well as supporting a fishery and aquaculture industry of national significance.

L63-64: Give more details about how seawater was transferred from the Niskin bottles to the quartz glass bottles (e.g., the material of the tube used for sample transfer; if the bottles were overflowed; if sunlight was avoided during the transfer; what caps were used to close the bottles, etc.).

Reply: Here we added more details about the sample transfer process: Bubble-free water samples were collected as triplicates for the determination of dissolved CO and were transferred to 100 mL quartz glass bottles by using Tygon® tubing to avoid contamination by silicone rubber (Xie et al., 2002) from the Niskin bottle, taking care to keep the tube at the bottom of the vail and control the

speed of water flow to avoid bubbles and swirls. Our samples were immediately sealed with rubber stoppers and aluminum caps, stored between 0 and 4 °C in the dark to suppress further CO photoproduction and uptake within the vial at ambient conditions (Day and Faloona, 2009), and transported within 1.5 hours to the Centro de Ciências do Mar (CCMAR) for analysis.

L72: Samples for dark incubation were collected only on one day? Then was it May 25 or 26?

Reply: The samples for the dark incubation (microbial CO consumption experiment) were collected only on 26 May. For more clarity, we adapted the text as follows: Water duplicate samples from Station 7 (the representative area for the combined effects of tidal effects and terrestrial contributions) for determining CO consumption on May 26th were introduced into 100 mL quartz glass bottles from the Niskin bottle and then covered with aluminum foil and stored in a cooling box ($0 \sim 4 \,^{\circ}$ C) after sampling.

L73: How many replicate incubations?

Reply: We did the microbial CO consumption experiment once for duplicate water samples on May 26th. For more clarity, we adapted the text as follows: Water duplicate samples from Station 7 (the representative area for the combined effects of tidal effects and terrestrial contributions) for determining CO consumption on May 26th were introduced into 100 mL quartz glass bottles from the Niskin bottle and then covered with aluminum foil and stored in a cooling box (0~4 °C) after sampling.

L74: "cooling box"—what was the temperature inside the box, well below the in situ temperature? If so, could significantly affect bacterial activity.

Reply: The temperature of the cooling box was between 0 and 4 °C. For more clarity, we adapted the text as follows: From Fig. S1, it is clear that after sampling and transporting the samples to the CCMAR for analysis within 0.5 h, the CO loss is less than 5%.

L75-78: Was temperature stable during the incubation? At what temperature? At or close to in situ temperature?

Reply: During the incubation experiment, all samples were stored at an in-situ temperature maintained at ± 1 °C. For more clarity, we adapted the text as follows: All samples for the consumption experiment were transported back to the laboratory within half an hour and stored at an in-situ temperature maintained at ± 1 °C in the lab.

CO measurements: What was the precision and detection limit of the method.

Reply: The detection limit of our measurements of dissolved CO was 10 ppb, and the analytical precision for the method was \pm 0.011 nmol L⁻¹. We added an explanation to the revised manuscript as follows: CO in the seawater and atmosphere was detected using a CO Gas Analyzer (ta3000R; Ametek, USA) with a lower detection limit of 10 ppb, and the analytical precision for the method was \pm 0.011 nmol L⁻¹.

L81: Did the 113.9 ppb standard cover the range of CO concentrations in the headspace of your samples? If not, what was the linear response range of the CO analyzer and was the range large enough to cover your samples' headspace concentrations?

Reply: The system was calibrated with the CO standard every half hour, or before and after related sample blocks, by replicating injections. For our datasets, it is convenient to correct data by interpolating standard peak areas over time periods between calibrations, like other studies using a similar method (e.g., Xie et al. 2002). Besides, the linear response range of the analyzer is 0~3 ppm CO. The standard gas was chosen as it lies in the expected range of the CO mole fraction equilibrated with the study area.

L85-89: Provide more details about how the headspace was created and how the headspace gas was subsampled (i.e., creation of headspace: how water was removed from and how gas was injected into the bottle? Subsampling the headspace gas: how the headspace gas was sampled? During both the creation and subsampling of the headspace gas, how the headspace was maintained at the atmospheric pressure?)

Reply: A syringe with a short needle was used as a conduit to introduce CO-free gas (12 mL) into a vial. The inflow of CO-free gas expels the water sample through the longer needle, creating a headspace within the vial (Fig. 1). The vial was equilibrated at room temperature and atmospheric pressure for 5 minutes at 120 rpm on a KS-90 shaking table (Edmund Bühler, Germany), followed by an additional equilibration period of 3 min before analysis. Then, using a gas-tight Luer-lock syringe, we extracted the equilibrium gas as the gaseous subsample of the headspace with the CO analyzer. This information was added to the revised manuscript.



Figure 1. A sketch of the headspace introduction apparatus.

Equations 1-5: List units for all parameters. Is Vw the volume before making the headspace or after making the headspace? Beta in Wiesenburg and Guinasso (1979) is in mL gas (mL H_2O)⁻¹ atm⁻¹. Moreover, the units of R are atm L mol⁻¹ K⁻¹. How did you get the units of nmol L⁻¹ for CO_{surf} (L94)?

Reply: We apologize for the confusion we caused. Units for all parameters have been added to the manuscript (see e.g., β (mL CO (mL H₂O)⁻¹ atm⁻¹)). V_w is the volume of the water sample after making the headspace (88 mL), and V_a is the volume of the headspace air (12 mL). In the second term of equation 1, the unit is in mol L⁻¹. According to Equation 7 and Table 4 (Wiesenburg and Guinasso, 1979), constants for the calculation of solubilities in nmol/L (CO) from moist air at 1 atm total pressure (first term of equation 1). Then, we could get the CO_{surf} by the unit of nmol L⁻¹.

Equation 1: The equation seems incorrect. The first term should be multiplied "P" and "Vw (volume of remaining water after the creation of headspace) then divided by the volume of the water before the creation of the headspace.

Reply: We disagree. We simplified Eq. 1: the first term should be multiplied by 'P' and 'V_w' indeed, then divided by 'V_w', not the volume of the water before the creation of the headspace. 'P' was 1 atm.

L104: "in equilibrium with the headspace". Note COeq is the dissolved CO concentration in the water remaining after the creation of the headspace.

Reply: We see that there has been a misunderstanding. We corrected and revised the equation and text as follows: The dissolved CO concentration in equilibrium with surface water (CO_{eq} in mol L⁻¹) was calculated using Eq 6 (Wiesenburg and Guinasso, 1979), x'_{atm} is the atmospheric CO dry mole fraction:

$$CO_{eq} = x'_{atm} \times \beta \tag{6}$$

Equation 4 is the first term of equation 1. If Ceq here refers to the CO concentration in surface water in equilibrium with the atmosphere (see equation 6), then x' here refers to the dry mole fraction of CO in the air samples. Do not mix this x' with that in equation 1.

Reply: We are sorry for the confusion. We have corrected and revised the equations:

$$CO_{eq} = x'_{atm} \times \beta$$

L107: Please explain what was the technical problem? The correction fact f = 3.12. This was huge!

Reply: We acknowledged that the issue stems from a malfunction in the calibration system – only errors in the calibration algorithm, not issues with the detector or lamp: The CO analyzer did not report any error during the warm-up progress including self-test after power-on, indicating that the detector, carrier gas pressure, and lamp were working normally. We repeated calibrations over time, using the same standard gases, to ensure that the detector consistently responds to the point calibration. The results were consistent and reproducible. However, the values obtained from the analyzer during the calibration were only about one-third of those given for the standard gases.

Further, we compared the average atmospheric CO mole fraction for the same month (May 2021) from atmospheric monitoring stations in Mace Head and Teixeira. A correction factor f (3.12) was determined. This was the fundamental reason for using the correction factor f for the CO analyzer measurements.

L111-113: "nearly uniform atmospheric background CO mole fractions". This is usually true over open oceans but not near the coast. The relative standard deviation of your measurements (5/35.1=14.2%) indicates this.

Reply: Thank you for pointing out this issue. The data obtained from the nearshore atmosphere were variable. But from the aspect of the monthly mean atmospheric background CO mole fractions across the Northwestern Atlantic Ocean during our measurement period were nearly uniform.

Equation 6. See comment on equation 4.

Reply: We have corrected and revised the equations.

L124-129: Did you measure u2?

Reply: Yes, instantaneous wind speeds at 2 m above sea level were measured onboard during atmospheric sampling using a vane anemometer (Testo, Germany). This information will be added to the revised manuscript.

Equation 7: Flux densities were calculated using spot windspeeds? If not, how windspeeds were processed? (if averaged, give the relevant spatial and temporal scales).

Reply: Flux densities were calculated using spot windspeeds (see Table 1).

Figure 3: Temperature, pH, and nutrients are shown in Figure 3 but not cited in the text.

Reply: Temperature, pH, and nutrients were the important basis for distinguishing the WWTP thermal effluent plume zone. We discuss them in lines 180 to 197.

Figure 2: What is "Stations 114" in the figure caption?

Reply: Thanks for spotting this mistake. We have corrected the caption of Fig.2. 'Figure 2: Mean CO surface concentrations (\pm standard error estimate) at Stations 1–14 measured on 25 May 2021 (blue bars) and on 26 May 2021 (orange bars).'

L180: Justify using one station (Sta. 18) to represent a region (Praia de Faro zone).

Reply: In the paper, we intended to point out that the chosen station 8 is close to the "Praia de Faro" zone. We did not use one station (Sta. 8) to represent a region (Praia de Faro zone). Notably, during the high tide, the water of the Faro channel from the region of Montenegro through Esteiro Largo and to the West region through the Ramalhete. Station 8 was a very representative sampling location.

L188: CO concentration at Sta. 13 was among the lowest (Figures 2 and 4).

Reply: Thanks for pointing this out. We have corrected station 13 to station 14.

L188-189: "biological CO production". This is purely speculative. FDOM at Sta. was high as well.

Reply: We agree that this idea is very speculative. However, we would like to keep the small paragraph as it seems justified by the results presented by Gros et al. (2009) and McLeod et al. (2021). The production and distribution of CO were affected by multiple factors; CO was also produced by phytoplankton, albeit to a minor extent.

L189-191: "WWTP thermal effluent plume". What was your purpose of mentioning this plume?

Reply: Here, we would like to illustrate that at station 8, CO concentrations were also affected by anthropogenic (e.g., high FDOM) influence.

L196-197: n = 14 for the relationship between CO and phosphate, but n = 28 for the relationship between CO and FDOM. Why? Any implication for the significant correlation between CO and phosphate?

Reply: Because of logistical constraints in the number of phosphate samples we could take. There was no valid evidence for the phosphate to directly affect the photodegradation of CDOM. Still, it was also an important factor affecting the distribution of CO at the lagoon system. More studies are needed to explain the positive correlation between these two better.

L201-202: "The lagoon was a source of atmospheric CO in May 2021". This is a premature conclusion since you only sampled on two days, each at a specific time point and tidal level (see general comment). Better to say "The lagoon was source of atmospheric CO at the time of sampling".

Reply: Thank you for pointing out this issue, and we adapted the text as follows: This indicates that the lagoon was a source of atmospheric CO at the time of sampling, which is in line with the observations from other coastal waters.

Figures 3, 4, and 5(b) and Table 1: Samples were collected on two days. Are the data shown in these figures the average of the two samples? If averaged, what are the variabilities of the two sampling times? Justify the use of average values.

Reply: Figures 3, 4, and Table 1 were the averages of two days' data, and Fig.5(b) was the flux densities of CO and wind speeds at Stations 8-12 and 14 on day two (we have corrected the caption). One aspect of our averaging was because some of the data were obtained from CCMAR (nutrients). More important was the point that CO concentrations in this study area were strongly affected by environmental conditions with anthropogenic activities, especially the WWTP. Thus, averaging the samples from the two days with and without WWTP emissions can better evaluate the CO distribution and potential impact of other activities on CO cycling in this region.

Figure 8: What do you mean by "CDOM absorption factor"?

Reply: We have replaced the 'CDOM absorption factor' with 'CDOM absorption coefficient'.

L266-267: Why CO concentration did not decrease during night? Hard to understand. Bacteria were killed by solar UV during daytime?

Reply: There seems to be a misunderstanding. Microbial consumption was always persisting, both during the day and at night. The experiments with sampling from the incubation bottles were conducted at time points 0, 24, and 48 h. In this 48-hour incubation experiment, the total photogeneration of CO was greater than microbial consumption, so it reflected continuous growth.

L267-269: In fact, the dark incubation roughly followed an exponential decay trend. No evidence suggests that a steady state reached at the end of the incubation.

Reply: To clarify our statement, we modified it. It reads now, 'This indicates that microbial CO consumption, probably restricted by CO dark production, led to a lower consumption state at the end of the dark incubation.'

L284-289: Much of the discussion is purely speculative.

Reply: We are sorry, but we disagree. We would like to keep the small paragraph as it seems to be justified by the results presented in Ossola et al. (2022) and Xu et al. (2023):

The ratio of the spectral slope (S_R) of the shorter waveband (280–295 nm) to that of the longer waveband (350–400 nm) was used to identify the origin of the CDOM: the smaller the S_R , the higher the proportion of terrestrial CDOM, and vice versa (Helms et al., 2013; John et al., 2009; Zhu et al., 2017). Besides, Xu et al. (2023) pointed out that there was a significant negative correlation between the apparent efficiency of CO photoproduction and S_R . Ossola et al. (2022) demonstrated for the first time a precise mechanism for CO production: the aromatic methoxy groups can produce CO. Summarizing, using S_R , the CO photoproduction capacity can be more accurately evaluated.

L291-293: Such a comparison does not mean much because you measured net production while the others reported photoproduction.

Reply: Thank you for pointing to this issue. We changed the comparison about the photoproduction in different areas of the manuscript.

Technical corrections:

L16: remove "of the Ria Formosa Lagoon".

Reply: Done.

L21: replace "it" with "this reaction".

Reply: Done.

L27: remove "may".

Reply: Done.

L28: "early" à "earlier"; "indicate" à "reported".

Reply: Done.

L125: at 10 m.

Reply: Done.

Equation 9: define u2.

Reply: Done.

L161: in the Ria...

Reply: Done.

L175: Show the Ramalhete and Faro–Olhão inlets in the maps of Figure 3.

Reply: Done.

L176: replace first "." with ",".

Reply: Done.

Figure 4: Caption is incorrect (now the same as that of Figure 3). Chl-a is missing.

Reply: Many thanks. We have corrected the caption: 'Figure 4: (A) mean salinity (filled light blue squares) vs. mean temperature (filled orange dots) and (B) mean concentration of dissolved CO (filled blue dots) and mean FDOM (filled red squares) at all stations in the Ria Formosa Lagoon on 25 May (Day1) and 26 May 2021 (Day2).'

L197: add "," after "r = 0.860".

Reply: Done.

Figure 8: Absence of symbol legends. What do you mean by "CDOM absorption factor"?

Reply: Done, and we have replaced the 'CDOM absorption factor' with 'CDOM absorption coefficient'.

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