~~Coccolithophorids precipitate carbonate in clumped isotope equilibrium with seawater~~
A well-constrained, coccolith-specific clumped isotope calibration of cultured coccolithophorids for marine temperature reconstructions

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Abstract. Numerous recent studies have tested the clumped isotope (Δ_{47}) thermometer on a variety of biogenic carbonates such as foraminifera and bivalves and showed that ~~all follow a common calibration. While~~most follow a common calibration. However, there may be a difference between biogenic carbonate-based calibrations and the most recent inorganic carbonate calibrations that are assumed to have formed close to isotopic equilibrium. Biogenic calibrations such as those based on foraminifera from seafloor sediments suffer from uncertainties in the determination of the calcification temperatures. Therefore, well-constrained laboratory cultures without temperature uncertainty can help resolve these discrepancies. Although the sample size requirements for a reliable Δ_{47} measurement have decreased over the years, the availability and preservation of many biogenic carbonates is still limited and/or require substantial time to be extracted from sediments in sufficient amounts. Coccoliths, on the other hand, are abundant and often well-preserved in sediments, and are a potential interesting target for palaeoceanography. We thus determined the Δ_{47}-temperature relationship for ~~coccolith carbonate, which is abundant and often well-preserved in sediments~~coccoliths because they are relatively easy to grow in the laboratory. The carbon and oxygen isotopic compositions of coccolith calcite have limited use in palaeoenvironmental reconstructions due to physiological effects that cause variability in the carbon and oxygen isotopic ~~values.~~fractionation during mineralisation. However, the relatively limited data available suggest that clumped isotopes may not be significantly influenced by these effects. We cultured three species of coccolithophores ~~in well constrained~~under controlled carbonate system conditions with ~~a~~ $CO_{2(aq)}$ concentrations between 5 and 45 μM and temperatures between 6°C and 27°C.

Our well-constrained results agree with a previous culture study that there are no species- or genus-specific vital effects on the Δ_{47}-temperature relationship in coccolithophores ~~and~~despite significant deviations from equilibrium in the C and O isotopic composition. We find that varying environmental parameters other than temperature ~~also do~~does not have a significant effect~~.Our~~ on Δ_{47}, changing the parameters yields Δ_{47}-temperature calibrations that agree within 1.2 ppm. Our well-constrained coccolith-specific Δ_{47}-temperature calibration ~~agrees within ±1°C of other biogenic carbonate calibrations yet all biogenic carbonate calibrations show~~shows a ~~consistent~~positive offset of ≥2~~-3°C relative~~ to the inorganic carbonate calibrations. ~~Thus,~~

~~all biogenic~~ , which point to as yet unknown coccolith-specific ~~calibrations can be used interchangeably and must be used for the reconstruction of~~ disequilibrium effects. All published biogenic studies fall within within ±1°C of our coccolith-specific calibration if the original calcification temperatures ~~in~~ are considered, while a cold bias and offsets of 1-2°C are present using the recalculated calcification temperatures of a recent study.

Thus, we suggest the use of our coccolith-specific calibration for further coccolith palaeoceanographic studies and that calibrations derived from laboratory-grown biogenic carbonates, in particular foraminifera, are desirable to reinforce the confidence of clumped isotope-based temperature reconstructions in palaeoceanography.

1 Introduction

Clumped isotope thermometry is a relatively new and increasingly used methodology for the reconstruction of paleotemperatures, palaeohydrology, diagenetic regimes, and as an isotopic tracer (see Huntington and Petersen, 2023 for a recent review). In a carbonate molecule, bonds between the rare heavy isotopes ^{13}C and ^{18}O ~~are thermodynamically preferred, leading to an increase in~~can be formed and their excess abundance ~~–~~relative to a stochastic distribution is denominated as "clumping"~~"~~", which increases with decreasing temperature. To measure this clumping, the carbonate is converted to CO_2 by reaction with phosphoric acid, and the excess abundance of ~~^{13}C and ^{18}O atoms~~$^{13}C^{18}O^{16}O$ (cardinal mass 47) in the released CO_2 relative to a stochastic distribution is measured and reported as Δ_{47} (Schauble et al., 2006; Eiler, 2007). In early studies, before the introduction of carbonate standardisation (Bernasconi et al. 2021), Δ_{47} in different calibration studies of the same carbonate material had widely variable relationships with temperature. ~~However, more~~More recent studies (Petersen et al., 2019; Meinicke et al., 2020; Anderson et al., 2021) have shown that most but not all of these discrepancies were caused by poor interlaboratory comparability caused by the lack of a robust standardisation methodology (Bernasconi et al. 2018, 2021~~;~~ ~~Anderson et al. 2021~~).

Based on these recent studies, Δ_{47} has a consistent relationship with temperature~~, independent from the origin of the carbonate,~~ and its temperature dependence is well established empirically at a large range of temperatures (e.g., Anderson et al., 2021; Fiebig et al., 2021). The Δ_{47}-temperature relationship is independent of the carbon and oxygen isotope composition of the fluid from which carbonates precipitate (Ghosh et al., 2006). Empirical calibrations between temperature and Δ_{47} have been established for temperatures between 0°C and 1100°C for inorganic carbonates (Kele et al., 2015; Bonifacie et al., 2017; Kelson et al., 2017; Müller et al., 2019; Swart et al. 2019; Jautzy et al. 2020; Anderson et al., 2021). Further empirical studies on biogenic carbonates, such as for foraminifera, coccoliths, gastropods, and bivalves, have found similar relationships between Δ_{47} and calcification temperature (Katz et al., 2017; Peral et al., 2018; Leutert et al., 2019; Piasecki et al., 2019; de Winter et al., 2022; Huyghe et al., 2022~~). Apart from~~), although specific types of biogenic carbonates such as shallow-water corals (Spooner et al., 2016; Davies et al., 2022), juvenile bivalves (Huyghe et al., 2022), and brachiopods (Bajnai et al., 2018; Davies et al., 2023; Letulle et al., ~~2023), these biogenic~~ 2023) do not. However, there are clear discrepancies between most inorganic calibrations ~~all fall within the same confidence interval (Meinicke et al., 2020,~~(Swart et al. 2019; Jautzy et al. 2020~~;~~

Anderson et al~~.~~, 2021; Fiebig et al., 2021) and an often used, generalised biogenic calibration (Meinicke et al., 2020). One interpretation is that this discrepancy results from uncertainties in the calculation of calcification temperatures for planktonic foraminifera and is resolved with alternate approach to calcification temperature estimation (Daëron and Gray, 2023). With this study using cultured coccoliths we generate biogenic carbonate under well-constrained temperature conditions, so there is little uncertainty in the calcification temperatures.

~~While most calibrations are consistent for inorganic and biogenic calcite, other~~Other methodological problems can also limit the overall use of clumped isotopes. The relative abundance of Δ_{47} to all isotopologues of CO_2 is only ~45 ppm, which requires a sample in the range of 80 to 120 μg with Thermo-Fischer Kiel IV devices coupled to Thermo-Fischer 253 Plus mass spectrometers (Müller et al., 2017) to 2-10 mg for a single measurement with common acid bath systems (e.g. Ghosh et al., 2006; Kelson et al., 2017; Peral et al., 2018; Fiebig et al., 2021). As the analytical error in clumped isotope measurements is large compared to the natural variability, many replicates are needed to achieve an analytical uncertainty allowing meaningful interpretations in palaeoceanography (Bernasconi et al., 2021; Fernandez et al., 2017; Daëron, 2021). Sample size requirements ~~are~~can be a major limiting factor for biogenic calcite ~~with~~. Both limited ~~sample~~abundance and time requirements for picking single species, limits the availability ~~such as~~of planktonic foraminifera ~~from cores and core tops or~~, and the need for sampling precise seasonal increments in slow-growing molluscs can also restrict the mass of carbonate available for analysis (Leutert et al., 2019; de Winter et al., 2022; Huyghe et al., 2022). Coccoliths are a promising alternative as they are often found in greater abundance and have a better preservation potential than foraminifera (Berger, 1973; Subhas et al., 2019). The abundance of coccolith-associated-polysaccharides ~~(CAP)~~ both around and within the coccolith aid in protecting the coccolith calcite from dissolution and overgrowth, and remain in place for millions of years (Henriksen et al., 2004; Sand et al., 2014; Lee et al., 2016). Few anion substitutions and a lack of lattice defects on the coccolith surface further aid in a better preservation relative to foraminifera (Berman et al., 1993; Stoll et al., 2001; Frøhlich et al., 2015; Walker et al., 2019).

~~Kinetic and disequilibrium effects due to the biological origin of the calcite, collectively known as vital effects, are known to affect the carbon and oxygen isotopic composition of~~Biogenic carbonates often feature carbon and oxygen isotopic compositions that differ from those expected for abiogenic carbonates near equilibrium, offsets informally called "vital effects". Such offsets have been described for coccolith calcite (Ziveri et al., 2003; Rickaby et al., 2010; Ziveri et al., 2012; Candelier et al., 2013; Hermoso et al., 2014; Stevenson et al., 2014; Hermoso et al., 2016, Katz et al., 2017) and the contributing processes simulated in models (Langer et al., 2012; Ziveri et al., 2012; Bolton and Stoll, 2013; Holtz et al., 2017; McClelland et al., 2017). ~~).~~ ~~Initial~~The vital effects limit the use of stable isotope signatures of coccolith calcite in palaeoceanography. However, initial studies ~~find~~of clumped isotopes in coccolithophorids found that ~~while~~ multiple coccolithophore species ~~with different~~display clear carbon and oxygen vital effects ~~do,~~ they follow previous Δ_{47}-temperature calibrations (Drury and John, 2016; Katz et al., 2017~~).~~), making them potentially useful for palaeoceanography. However, because these studies and calibrations were performed before carbonate standardisation (Bernasconi et al., 2021) and have a limited number of replicate analyses, ~~limiting their~~robustness and interpretation of their conclusions need to be verified with new studies. Other aspects

such as the dissolved inorganic carbon chemistry in the cultures and growth conditions as possible influences on Δ_{47} also still need to be examined.

To this end, three species of calcifying coccolithophores were cultured under controlled temperature and carbonate chemistry conditions. A temperature range of 21°C and $CO_{2(aq)}$ range of 40 μM was covered. Coccolithophores from the *Gephyrocapsa* genus were cultured between 6°C and 27°C, using the warm-adapted *G. oceanica* and the cold-adapted *G. muellerae*. Inter-genus vital effects were tested through comparison with *Calcidiscus leptoporus*, which features distinct carbon and oxygen isotopic vital effects compared to *Gephyrocapsa* in previous studies (Ziveri et al., 2003; Hermoso et al., 2014; Katz et al., 2017). Finally, through comparison with a previous coccolith culture (Katz et al., 2017), inorganic (Anderson et al., 2021; Daëron and Vermeesch, 2024), biogenic (Peral et al., 2018; Meinicke et al., 2020; de Winter et al., 2022), and recalculated (Daëron and Gray, 2023) Δ_{47}-temperature calibrations, the potential need for a coccolith-specific calibration is assessed.

2 Materials and methods

2.1 Batch and continuous culture setup

Three monoclonal coccolithophore strains, *Gephyrocapsa oceanica* (RCC 1303), *Gephyrocapsa muellerae* (RCC 3370), and *Calcidiscus leptoporus* (RCC 1130) obtained from the Roscoff Culture Collection were cultured at ETH Zürich. *G. oceanica* is an abundant cosmopolitan coccolithophore species that has been found at temperatures between 12-27°C but favours temperatures above 20°C (Sett et al., 2014; von Dassow et al., 2021). *G. muellerae* is from the same genus as *G. oceanica*, favours colder and less saline conditions, and can grow at 6°C (von Dassow et al., 2021). *C. leptoporus* is a large coccolithophore, is especially abundant in low latitude upwelling settings, is resistant to dissolution and thus a major contributor to coccolith carbonate, and is known to have large isotopic vital effects (Thierstein and Young, 2004; Ziveri et al., 2007; Langer et al., 2012; Hermoso, 2014). Thus, the three strains were cultured to provide a temperature range of 21°C while also highlighting potential isotopic vital effects. *G. oceanica* was cultured in a turbidostat or continuous culture setup as seen in Fig. 1a, between a $CO_{2(aq)}$ of 5 and 45 μM for at least two temperatures (Table 1). All three strains were grown in at least two temperatures in batch culture setup as seen in Fig. 1b, and provide a wide range of temperatures (Table 1).

All cultures were carried out in ~~Artificial Seawater (ASW),~~artificial seawater, following Kester et al. (1967), as the basis of a K/2 medium (Keller et al., 1987). The final pH and DIC were adjusted through addition of HCl and Na_2CO_3 and covered a range of 40 μM of $CO_{2(aq)}$ values. A Tris buffer was not included for all experiments as this would interfere with the carbonate chemistry control. Prior to inoculation, the K/2 media was sterilized through an 0.2 µm Millipore Stericup filter.

Before starting the experiments, the strains were maintained at a constant temperature of 18°C and slowly acclimatized to the experimental temperature for at least 4-6 generations. Cultures were inoculated at cell densities of ~5000 cells mL^{-1} for *G. oceanica* and *G. muellerae* and ~2000 cells mL^{-1} for *C. leptoporus*.

Batch cultures were carried out in 1 or 2 L Nalgene polycarbonate sterile flasks with 50- and 100-mL headspace respectively and kept in a lit incubator set to the experimental temperature ($\pm$ 0.1°C). For the duration of the experiments, the flasks were rotated on a roller set at 10 rpm to reduce settling and allow for uniform light exposure, see Fig. 1b. *C. leptoporus* did not grow under turbulent conditions and was manually shaken every day (Houdan et al., 2006). The cultures were illuminated by LED
130 lights on a sinusoidal 14/10-hour light/dark cycle and a maximum luminosity of 120 µmol photons m^{-2} s^{-1}. Harvesting took place in a semi-continuous manner, where 80-90% of the total volume was harvested and then refreshed when the culture reached a set cell density (100-150 x 10^3 cells mL^{-1}). In general, we used a higher cell density compared to previous culturing studies due to the larger sample material requirement for clumped isotope measurements. Yet each strain was maintained in their early- to mid-exponential phase to prevent a reservoir effect and an enriching of the $\delta^{13}C$ of the media ($\delta^{13}C_{DIC}$) relative
135 to the initial conditions (Barry et al., 2012; Hermoso, 2014). Certain batch and continuous cultures had a higher cell density and significant drift in the DIC, pH, or $CO_{2(aq)}$ over the course of the experiment and serve to test the sensitivity of vital effects to a varying carbon system.

Continuous cultures were performed in 1L (FMT 150/1000) or 3L (FMT 150/3000) photobioreactor (PBR) connected to a PP600 peristaltic pump and a GMS 150 gas mixing system (Photon Systems Instruments, Drásov, Czech Republic), see Fig.
140 1a. For a more detailed description of the experimental setup see Zhang et al. (2022). The strains were illuminated with LEDs on a 16/8-hour light/dark cycle with a maximum luminosity of 200 µmol photons m^{-2}s^{1}. The optical density (OD), which is proportional to the cell density, was calibrated to a zero point before inoculation. With continuous OD monitoring and (sub-)daily measurements of the cell density, the OD could be set to a cell density threshold between 100 and 200 x 10^3 cells mL^{-1}. If the OD exceeded this threshold, the peristaltic pump pumped fresh media into the PBR equivalent to 10% of the culture
145 volume, and media from inside the PBR was pushed into the outflow bottle. Ambient air was first scrubbed of CO_2 and then passed through a gas mixing system with a set amount of CO_2 with a constant composition ($\delta^{13}C$ = -14.36 $\pm$ 0.06‰ VPDB; $\delta^{18}O$ = 19.85 $\pm$ 0.14‰ VSMOW). Thus, the pCO$_2$ was controlled and maintained at a constant level, which was checked at the beginning and end of each experiment. Temperatures were continuously controlled with a pH-temperature probe in the vessel, which kept a constant temperature with <0.1°C deviation through Peltier-elements at the bottom of the vessel.
150 Cell density and size were measured using a Z2 Coulter Particle Counter and Size Analyzer with an aperture size of 100 µm (Beckman Coulter, Inc., Brea, California, United States), which were used to determine growth rates. Cell counts were taken systematically every 24 hours with an extra measurement taken before and after harvest.

The net growth rate for batch cultures were calculated as:

$$\mu = \frac{\ln(N_t) - \ln(N_0)}{t - t_0} \tag{1}$$

155 where N_t is the cell count at time t (in days) and N_0 is the initial cell count.

For continuous cultures, the dilution rate, which equates to the volume of the output (V_{out}) over the volume of the PBR over time t also needs to be considered:

$$\mu = \frac{\ln(N_t) - \ln(N_0) + \frac{V_{out}}{V_{PBR}}}{t - t_0} \tag{2}$$

Cells were harvested by centrifugation at 4000 rpm for 5 minutes. The pellet containing coccoliths was rinsed with deionized
160   water to remove traces of salts by 4 cycles of centrifugation and removal of the supernatant. Subsequently, the wet pellet of
organic matter and coccoliths was transferred to a 2 mL vial, which was then stored at -20°C for later analysis.

At the beginning, at each harvest, and at the end of an experiment, 12 mL of water were sampled for pH and cell density
measurement. Each aliquot was centrifuged once at 4000 rpm for 5 minutes, and the supernatant was used for DIC, $\delta^{13}C_{DIC}$,
and $\delta^{18}O_{SW}$ measurements. Some batch and continuous cultures were kept until higher cell densities were reached. In these
165   cases, pH, DIC, and $\delta^{13}C_{DIC}$ were measured more often to track the evolutions from initial conditions as a result of cell growth
and cell density increase.

2.2 Sample cleaning

Contaminants from organic matter or sulphur compounds contained within carbonates are known to interfere with the clumped
isotopic measurements for carbonates (Eiler and Schauble, 2004; Dennis and Schrag, 2010). In this study we used three
170   methodologies for the removal of possible contaminants from the coccolith pellet. Initially the aliquots were dried in a 60°C
oven for at least 4 hours to remove any remaining water that could dilute the cleaning reagents. Then one of three cleaning
methodologies was performed.

One method used oxidation of the organic matter through reaction with a 18% H_2O_2 solution neutralized with NaOH to pH 8-
9 following the protocol of Falster et al. (2018). The second methodology used a solution of 10% H_2O_2 neutralized with NH_3
175   to pH 8-9. For both solutions the pellets were reacted overnight at room temperature and then rinsed 4-5 times with milli-Q
until a neutral pH of 7 was obtained. The third method used is Total Lipid Extraction (TLE), following Matyash et al. (2008).
The pellets were first ~~reacted with~~suspended in 1 mL of isopropanol and homogenized through vortexing and sonication until
there were no visible clumps. 3 mL of methyl-tert-butyl ether ~~(MTBE)~~ were added and the total solution was again vortexed.
1 mL of DI water was added and the sample was centrifuged at 4000 rpm for 5 minutes. This method removes the polar and
180   apolar organic compounds from the pellet into the upper 4 mL of the solution. This supernatant is subsequently discarded,
while maintaining a 1 mL of aqueous phase with the cleaned coccolith pellet at the bottom. When abundant organic matter
was still visible after one cycle of cleaning, the procedure was repeated. After all three cleaning methodologies, the samples
were dried at 60°C for at least 4 hours.

In order to confirm the effectiveness of the different cleaning methodologies and their effect on the coccolith carbonate, high
185   resolution scanning electron images were taken using a JSM-7100F JEOL Scanning electron Microscope (SEM) at ScopeM,
ETH Zürich, see Fig. S1. As harvested from the cultures, all coccoliths exhibited regular morphology with no evidence of
coccolith malformation. The cleaning protocol causes slight dissolution, fragmentation, and breakage of some of the coccoliths.

2.3 Isotope measurements

2.3.1 Coccolith carbon, oxygen, and clumped isotopes

Carbon, oxygen, and clumped isotopes from the cultured coccoliths were measured on two ThermoFisher Scientific mass spectrometers, a MAT253 and a 253Plus, coupled to Kiel IV preparation devices as described in Müller et al. (2017). Aliquots of 110-180 μg of sample and carbonate standards (ETH-1, 2, 3, and IAEA C2) were measured with a Kiel device at 70°C. The samples were reacted for 300 seconds with three drops of 104% phosphoric acid, and the released CO_2 was immediately frozen in a liquid nitrogen (LN_2) trap kept at -190°C. After the sample was reacted fully, the first LN_2 trap is heated to -100°C, and the CO_2 gas was transferred through a 10 mm PorapakQ (50-80 mesh) to a second LN_2 trap kept at -190°C. The PorapakQ tubing was coated with Sulfinert 2000, stuffed with silver wool, and kept at -40°C to capture and eliminate possible organic contaminants. The second LN_2 trap was then heated to room temperature and measured using the Long Integration Dual Inlet (LIDI) protocol (Hu et al., 2014, Müller et al., 2017). The initial gas intensity of the sample gas is recorded and then immediately measured for 400 seconds, 40 cycles of 10 seconds. Subsequently, the reference gas is measured for the same duration starting at the same initial gas intensity. Before measuring, background scans at different beam intensities were carried out to determine the pressure dependent background on all beams (Bernasconi et al., 2013; Meckler et al., 2014). Using the Easotope software (John and Bowen, 2016) a pressure-sensitive baseline (PBL) correction was applied, and raw carbon and oxygen values were converted to VPDB using the Brand parameters as suggested by Daëron et al. (2016). Further, the raw vs accepted Δ_{47} values for the carbonate standards and an empirical transfer function (ETF) were used to convert and normalize the sample Δ_{47} data to the Intercarb carbon dioxide equilibrium scale (I-CDES; Bernasconi et al., 2021).

Each individual reported clumped isotope value consists of at least 10 replicates., with an average of 13 replicates. Long-term accuracy and reproducibility of the Δ_{47} measurements were evaluated based on monitoring of the IAEA-C2 international carbonate standard, which was treated as an unknown sample (MAT253: $\Delta_{47} = 0.6385 \pm 0.034$, MAT253 Plus: $\Delta_{47} = 0.6411 \pm 0.026$‰; 1σ n=241) and fit within the accepted values (0.6409‰ ± 0.003‰; Bernasconi et al., 2021). Analytical errors are reported at the 95% confidence interval (Fernandez et al., 2017). 2017) without taking into account the standardisation error. However, as the measurements are carried out with a 50:50 standard to sample ratio, the additional uncertainty related to standardisation would be small (Bernasconi et al. 2021).

2.3.2 DIC and seawater carbon and oxygen isotopes

DIC and pCO_2 were measured on a Picarro G-2131-i Cavity Ringdown Spectrometer (CRDS; Picarro Inc, USA) coupled to a AS-D1 DIC-$\delta^{13}C$ Analyzer (Apollo Scitech, USA). For DIC measurements, 3 mL of centrifuged media were taken and reacted within the AS-D1 analyser with 0.9 mL of a 3% H_2PO_4 and 7% NaCl solution. The absolute DIC concentration was found through quantification of the extracted CO_2 concentration as described in Deng et al. (2022). A threshold of ≤ 20 ppm pCO_2 for background synthetic air (80% N_2, 20% O_2; PanGas) was first defined. After at least 10 measurements of background air below this threshold, the stable baseline was reached and the sample is injected into the measuring chamber. As the extracted

220 CO_2 flows into the measuring chamber, measurements of pCO_2 were taken every 3-5 seconds until all of the extracted CO_2 had passed through the measuring chamber, and the stable baseline was reached again. For the absolute DIC concentration, the integrated area of pCO_2 between the two baselines is plotted against the total DIC content. The latter is defined as the volume multiplied by the measured DIC. A Certified Reference Material ~~(CRM),~~. NOAA batch 186 with a known absolute DIC concentration of 2012 μM, is used for calibration. Through variation of the ~~CRM~~Certified Reference Material volume, a
225 range of integration areas and total DIC content was obtained and internally defined. Two secondary standards with calibrated DIC concentrations were also used. Each measurement was repeated twice with the same centrifuged media in quick succession to reduce CO_2 exchange with the atmosphere, which gave an average DIC and uncertainty for each measured ~~sample.~~replicate. At least three of these replicates were taken and measured for each experiment.

For $\delta^{13}C_{DIC}$ measurements, 1 mL of centrifuged media was acidified with 150μL of 104% H_2PO_4 in a He-flushed vial.
230 Subsequently the $\delta^{13}C_{DIC}$ was measured on a Gasbench II coupled to a Delta V Plus mass spectrometer (Thermo Fischer Scientific, USA). Two in-house $NaHCO_3$ standards dissolved in deionized water with $\delta^{13}C_{DIC}$ values of -4.66‰ and -7.94‰ were used.

The oxygen isotopic composition of the seawater ($\delta^{18}O_{sw}$) was measured on a Picarro L2130-*i* Isotope Wavelength-Scanned Cavity Ring-Down Spectrometer ~~(WS-CRDS)~~ following Gupta et al. (2009). The water sample is first passed through a salt
235 catchment trap, vaporized, and injected at a uniform concentration and flow rate into the ~~WS-CRDS.~~Picarro spectrometer. This delivers a pulse with a constant concentration profile, during which the ~~δ^2D~~δ^2H and $\delta^{18}O$ are measured. Three in-house standards calibrated to three IAEA and USGS standards (SLAP2, GRESP, and VSMOW2) were run every 20 samples. Another in-house standard is run as an unknown sample. All standards fell within 1 standard deviation of their accepted value. Measurement precision was affected by the residual salt from the seawater. At least two seawater measurements were taken
240 for each experiment. Subsequently an average $\delta^{18}O_{sw}$ and uncertainty (mean σ=0.32‰) for each condition was obtained, see Table S1. This uncertainty is used for the uncertainty of the oxygen isotope offset from seawater, $\delta^{18}O_c - \delta^{18}O_{sw}$ ($\Delta^{18}O_{c-sw}$) as reported in Table 1, and will be used in subsequent figures. ~~.~~ (mean σ=0.29‰).

pH measurements were made using a Mettler Toledo LE410 pH-probe (Mettler Toledo, Greifensee, Switzerland) and were calibrated with three NBS standards (pH=4.00, 7.01, 9.03 at 21°C), with a standard deviation of ±0.01. All pH measurements
245 are given in the NBS scale. Seawater carbon chemistry was calculated through CO2SYS (Lewis and Wallace, 1998), with input of measured pH and DIC, and using the K1, K2 constants of Leuker et al. (2000), $KHSO_4$ of Dickson (1990), KHF of Perez and Fraga (1987), and $[B]_I$ value of Lee et al. (2010).

DIC and pH measurements are compared and normalized to the initial measured values to detect potential drift and stability of the carbon chemistry of each culture. These are reported as initial DIC – measured DIC (Δ_{DIC}) and initial pH – measured
250 pH (Δ_{pH}) in Table S1. We report the uncertainty of the fractionation between coccolith calcite and DIC ($\Delta^{13}C_{c-DIC}$) ~~as~~from both the standard deviation of $\delta^{13}C_c$ and the range of all measured $\delta^{13}C_{DIC}$ over the course of the sampled culture, to take into account the potential effect of DIC evolution and drift, and will be used in subsequent figures.

The same media was used for both batch and continuous cultures and thus the same $\delta^{13}C_{DIC}$ would be expected for both. However, through the continuous bubbling and gas-exchange in the continuous cultures, there is a potential for different $\delta^{13}C_{DIC}$ values than the batch culture setup. Indeed, there is a non-systematic enrichment of 0-1‰ for the batch culture $\delta^{13}C_{DIC}$. However, the fractionation between coccolith calcite and DIC i.e. $\Delta^{13}C_{c\text{-}DIC}$, is not affected by these differences.

An often-used index in coccolithophore geochemistry studies (McClelland et al., 2017; Phelps et al., 2021) for the relative usage of carbon supply and demand is the dimensionless τ:

$$\tau = \frac{r*\rho*\mu}{3*CO_{2(aq)}*P_c} \tag{3}$$

It reflects the degree of carbon utilization by the coccolithophore as measured through growth rate (μ), defined in Eqs. 1 and 2 for batch and continuous culture setup respectively, the cell radius (r), and cellular carbon density (ρ) against the ~~DIC~~diffusive CO_2 supply into the cell set by ($CO_{2(aq)}$) and the permeability to CO_2 (P_c). We use a P_c of 1.4×10^{-3} µm day^{-1} (Blanco-Ameijeiras et al., 2020) and ρ of 2×10^{-3} µM (McClelland et al., 2017). It is assumed that there is only a diffusive supply of CO_2 into the cell. Our cell size measurements assume a perfect coccosphere for each counted cell, which results in a coccosphere radius that is slightly different than the cell radius. This is taken into account, together with uncertainties in μ and $CO_{2(aq)}$, into the uncertainty of the final τ value.

Pearson correlation coefficients (ρ) are used to determine whether two parameters are linearly correlated. If the ρ value is near 1 or -1, there is a strong correlation and one parameter has an effect on the other. Weak correlations are here defined as $\rho \leq \pm 0.40$ and significance is given by a p-value < 0.05.

3 Results

3.1 Carbon chemistry

3.1.1 DIC, pH, and $CO_{2(aq)}$

The stability of the carbon chemistry was monitored with at least three DIC and pH measurements for each culture. Drift from the initial conditions was less than ±20% and ±0.20 for Δ_{DIC} and Δ_{pH} respectively for 40 culture experiments (Fig. S2 and Table S1). To ensure only well-constrained culture conditions are considered, fifteen experiments with $\Delta_{DIC} \geq 20\%$ or $\Delta_{pH} \geq 0.20$ are excluded from our main analysis in Figs. 2 through 5, but are evaluated in a subsequent comparison (see Sect. 4.3).

3.1.2 Carbon isotopes

The carbon isotope composition of the DIC ($\delta^{13}C_{DIC}$) varies between -5.10‰ and 0.80‰, with a similar large range of -5.52‰ and -0.07‰ for all species' coccolith carbon isotopes ($\delta^{13}C_c$; Table S1). This is mostly due to the large variability in $\delta^{13}C_c$ for *G. oceanica*, which can be attributed to the range of different experimental conditions. The $\delta^{13}C_c$ in *G. muellerae* varies between -4.68‰ and -3.66‰, while the $\delta^{13}C_c$ in *C. leptoporus* varies between -5.15‰ and -5.04‰.

The fractionation $\Delta^{13}C_{c\text{-DIC}}$, ranges between -1.41‰ and 1.22‰ for *G. oceanica*, -0.88‰ and 0.21‰ for *G. muellerae* and -2.86‰ and -2.22‰ for *C. leptoporus*. ~~Pearson correlation coefficients (ρ) are used to determine whether two parameters are linearly correlated. If the ρ value is near 1 or -1, there is a strong correlation and one parameter has an effect on the other.~~
285 ~~Weak correlations are here defined as ρ ≤ ±0.40 and significance is given by a p-value < 0.05.~~ No significant linear correlations were identified between *Gephyrocapsa* $\Delta^{13}C_{c\text{-DIC}}$ and carbonate system parameters (DIC, pH, $CO_{2(aq)}$) or culture parameters (cell density and Δ_{DIC}) (Table S2). *Gephyrocapsa* culture data is also not significantly correlated with τ, the index of carbon demand vs supply (Fig. 2; ρ = 0.06, p-value = 0.74). Including *Calcidiscus* does not improve the correlation or significance with any parameter apart from Δ_{DIC}.

3.2 Oxygen isotopes

The $\delta^{18}O_{sw}$ are given in Table S1 for all cultures and vary by ~1.6‰ across with no systematic differences between batch and continuous cultures. For a given temperature, *C. leptoporus* has systematically lower $\delta^{18}O_c$ values by ~3‰ while *G. oceanica* and *G. muellerae* have similar $\delta^{18}O_c$ values at the same temperature. The interspecies variations are independent of culture setup and experimental condition.

295 The oxygen isotope offset from seawater, $\delta^{18}O_c - \delta^{18}O_{sw}$ ($\Delta^{18}O_{c\text{-sw}}$), varies systematically for all experimental conditions as seen in Fig. 3 and Table 1. *Calcidiscus* has a systematic offset of ~3‰ relative to *Gephyrocapsa* at the cultured growth temperatures, which agrees with previous culturing studies (Ziveri et al., 2003; Candelier et al., 2013; Stevenson et al., 2014; Hermoso et al., 2016; Katz et al., 2017). For *G. oceanica*, there is a significant negative correlation between $\Delta^{18}O_{c\text{-sw}}$ and temperature (ρ = -0.94, p-value <0.05), which remains when including *G. muellerae* (ρ = -0.97, p-value <0.05). The $\Delta^{18}O_{c\text{-sw}}$-
300 temperature relationship is consistent between setups and experimental conditions and is not influenced by the variability in $\delta^{18}O_{sw}$ values. The available data indicate that the range of $\Delta^{18}O_{c\text{-sw}}$ is ~~thus~~ similar within the *Gephyrocapsa* genus ~~but not species specific.~~.

3.3 Clumped isotopes

The Δ_{47} values are shown in Table 1 and Fig. 4, and range from 0.589‰ – 0.640‰ for temperatures between 12-27°C for *G.*
305 *oceanica*, 0.618‰ – 0.659‰ between 6-18°C for *G. muellerae*, and 0.636‰ – 0.652‰ at 12°C for *C. leptoporus*. There is variation of ~25 ppm at given experimental conditions, in particular at 21°C and 24°C, although all datapoints fall within the long-term standard deviation of the standards used for correction of 0.020‰. There is no resolvable difference between species or genus at given temperatures~~.~~; differences are within ±0.016‰ of each other.

4 Discussion

310 ~~Disequilibrium in the carbon, oxygen, and clumped isotopic systems is driven by different mechanisms within the DIC-H₂O system and coccolithophore cells. Thus, in order to fully evaluate potential vital effects in the Δ_{47}-temperature relationship in~~

coccolith calcite, we first characterize and discuss the vital effects in carbon and oxygen isotopes and compare them to previous culture studies.

4.1 For our culture experiments, in order to evaluate whether processes promoting variable stable isotope effects would systematically affect Δ_{47}, we manipulated the media carbonate chemistry in ways that have produced variable stable isotope vital effects in coccoliths in previous cultures. We also contrasted two coccolith genera that exhibit different ranges of vital effects. Multiple physiological explanations for coccolith vital effects have been proposed (Rickaby et al., 2010; Moolna and Rickaby, 2012; Hermoso et al., 2014, 2016) and simulated previously in cellular models (Ziveri et al., 2012; Bolton and Stoll 2013; Holtz et al., 2015, 2017; McClelland et al., 2017). In our discussion, in Section 4.1 and 4.2 we briefly summarize these mechanisms, and quantify the vital effect in each experiment, so that in section 4.3 we may compare the vital effect to the Δ_{47} variations. The focus of this study is the calibration of the temperature dependence of Δ_{47} in coccolith calcite. Therefore, we will only briefly discuss the causes and mechanisms leading to isotopic disequilibrium in the cultures, focusing in particular on those that could also affect Δ_{47}.

4.1 Carbon isotope vital effects in coccolith calcite

In this study (Fig. 5), all continuous (and most batch) cultures for *G. oceanica* fall within the expected range of $0 \pm 1.5‰$ for $\Delta^{13}C_{c\text{-}DIC}$ as found in other studies (see Fig. S3; Rickaby et al., 2010; Moolna and Rickaby, 2012; Hermoso, 2014; Katz et al., 2017). There are no comparable culture studies for *G. muellerae*. The $\Delta^{13}C_{c\text{-}DIC}$ values fall within the range of *G. oceanica*. Both species will be combined in any inter-genera analyses for Sect. 4. *C. leptoporus* has a $\Delta^{13}C_{c\text{-}DIC}$ of $-2.0 \pm 1.0‰$ comparable to previous culturing studies (Fig. 5; Ziveri et al., 2003; Hermoso et al., 2014; Katz et al., 2017).

Models of the vital effect in coccolith carbon universally consider two fundamental controls on $\Delta^{13}C_{c\text{-}DIC}$ (Bolton and Stoll, 2013; Holtz et al., 2017; McClelland et al., 2017). For a given cell size and growth rate, all available models simulate a decrease in $\Delta^{13}C_{c\text{-}DIC}$ with increasing concentrations of $CO_{2(aq)}$. An increase in $CO_{2(aq)}$ relative to the cellular carbon demand will result in a decrease in $\Delta^{13}C_{c\text{-}DIC}$. The carbon supply versus demand index τ is often used to describe this behaviour (Eq. 3, Fig. 2). However, we cannot discern whether our *Gephyrocapsa* experiments show such a decrease in $\Delta^{13}C_{c\text{-}DIC}$. One contributing factor may be the positive correlation of $CO_{2(aq)}$ with DIC in our experiments, due to our strategy of manipulating the carbon system through bubbling CO_2 into the system. Previous models simulate an increase in $\Delta^{13}C_{c\text{-}DIC}$ when there is an increase in the proportion of HCO_3^- relative to CO_2 entering the cell as the greater proportion of HCO_3^- will cause an enrichment in the $\delta^{13}C$ of the intracellular reservoir. Although carbon limitation at low $CO_{2(aq)}$ levels is observed to increase HCO_3^- uptake into the cell (Rost et al., 2003; Bach et al., 2013), it is also possible that higher media DIC can also lead to an increased HCO_3^- uptake and compensate the expected effect of higher $CO_{2(aq)}$. Interspecific variations in $\Delta^{13}C_{c\text{-}DIC}$ are strongly conditioned by the PIC:POC ratio. The high PIC:POC ratio of *Calcidiscus* contributes to a lowering of the $\delta^{13}C_c$ and $\Delta^{13}C_{c\text{-}DIC}$ (Fig. 5; Bolton and Stoll, 2013; McClelland et al., 2017). While vital effects are present with clear differences between *Gephyrocapsa* and *Calcidiscus*, further physiological explanations are outside the scope of this study.

~~4.2~~ Oxygen isotope vital effects in coccolith calcite

345 At a given temperature, equilibrium oxygen isotope fractionation between calcite and water is inferred to be most closely represented by natural ~~precipitation featuring very~~carbonates precipitated at extremely slow ~~growth~~ rates and independent of pH (Coplen, 2007; Daëron et al., 2019~~) or laboratory experiments using carbonic anhydrase (CA) to maintain oxygen isotopic exchange between DIC and H₂O ()~~. The model of Watkins et al.~~, .~~ (2013~~:~~, 2014~~). This fractionation ~~) approximates this equilibrium, and is ~~illustrated~~shown as the 'equilibrium limit' in Fig. ~~3.~~3. This approximation is derived from their

350 experimental setup, which is not necessarily in equilibrium, and therefore potential small growth rate and pH effects are still present. Non-equilibrium fractionation effects between DIC and calcite that manifest at faster growth rates in experiments both with and without carbonic anhydrase (CA) include lower $\Delta^{18}O_{c-w}$, and a pH-dependence. This effect presumably but not necessarily occurs because calcite forms from both bicarbonate and carbonate ions in proportion to their abundance in solution. At a higher pH the proportion of calcite carbon derived from the carbonate ion increases (McConnaughey, 1989; Clark et al.,

355 1992; Dietzel et al., 1992; Zeebe and Wolf-Gladrow, 2001; Watkins et al., 2014; Devriendt et al., 2017). ~~This process is illustrated by the 'kinetic limit' in Fig. 3.~~Here, a pH of 8.3 at the crystallisation-site and the fastest growth rate is assumed in the model of Watkins et al. (2013, 2014) and Watkins and Devriendt (2022), with which the 'kinetic limit' is derived and illustrated in Fig. 3. This gives an approximate 2‰ offset and incorporates a large range of experimentally derived and modelled inorganic calcites precipitated in presence of CA. Additionally, numerous experiments and potentially many natural

360 biogenic and abiogenic systems may precipitate calcite from a solution in which equilibrium between DIC and H₂O is not maintained due to a lack of CA or fast calcification rates (Devriendt et al., 2017; Daëron et al. 2019). Rayleigh fractionation of oxygen isotopes in the internal DIC pool occurs as a result, which is transferred to the isotopic composition of the calcite and leads to lower $\Delta^{18}O_{c-w}$ values~~.~~, thus exacerbating the disequilibrium fractionation potentially present between the DIC and calcite as described above.

365 Recent studies suggest that in certain systems such as synthetically precipitated calcite, oxygen isotope disequilibrium does not affect Δ_{47}, even for systems with intermediate oxygen isotope fractionations that fall between the kinetic and equilibrium limits, as seen in Fig. 3 (Kelson et al., 2017; Levitt et al., 2018; Jautzy et al., 2020; Fiebig et al., 2021). Models also suggest ~~this~~little to no Δ_{47} disequilibrium for biogenic carbonates such as foraminifera and bivalves, as the magnitude of potential Δ_{47} disequilibrium is below the current analytical resolution for Δ_{47} measurements although this is pH and growth rate dependent

370 (Defliese and Lohmann, 2015; Watkins and Hunt, 2015; Watkins and Devriendt ~~et al., 2017~~, 2022). However, this does not hold for corals, brachiopods, and speleothems, where systematic $\delta^{18}O$ and Δ_{47} disequilibria are present, and should be explored further in more biogenic carbonates (Watkins and Hunt, 2015; Guo and Zhou, 2019, Guo, 2020).

Offsets from the equilibrium limit ($\Delta\Delta^{18}O_{off}$) were calculated to quantify the vital effect in oxygen isotopes. Offsets may potentially arise from pH variability, growth rate, calcification rate, or disequilibrium within the DIC-H₂O system. *Calcidiscus*

375 is significantly different from equilibrium (t-test; t(4) = -58.68, p-value <0.05) and falls outside the equilibrium and kinetic limits as seen in Fig. 3. Although for *Gephyrocapsa* the mean $\Delta\Delta^{18}O_{off}$ value is around zero and there is no significant

difference from the abiogenically defined equilibrium (t-test; $t(72)$ = -0.47, p-value = 0.64), there is a range of ~1.5‰ in $\Delta\Delta^{18}O_{off}$ among different experiments. Previously published *Gephyrocapsa* data has a positive $\Delta\Delta^{18}O_{off}$ of 0-1‰ (Fig. 5; Ziveri et al., 2003; Hermoso et al., 2016).

380 CA has been suggested as a potential equilibration ~~mechanism~~catalyst for both oxygen isotopes and Δ_{47} within the DIC-H$_2$O system in calcifying organisms. Even at an elevated pH, the equilibration time is shortened considerably for abiogenic carbonates precipitated in the presence of CA (Uchikawa and Zeebe, 2012; Kelson et al., 2017). A lack or low activity of CA can potentially cause the variable $\Delta\Delta^{18}O_{off}$ seen in our data. While the pH of the intracellular calcification site for coccoliths is not well constrained, there is experimental evidence for CA in the biomineralization pathway of *Emiliania huxleyi* (Zhang

385 et al., 2021). Although CA has not been explicitly found in *Gephyrocapsa*, it is associated with the cytosol, chloroplast, or extracellularly in other coccolithophores (Nimer et al., 1994; Elzenga et al., 2000; Herfort et al., 2002; Rost et al., 2003). Furthermore, genes have been found associated with biomineralization and CA expression in *E. huxleyi* (Quinn et al., 2006; Soto et al., 2006; Richier et al., 2011). Models show a requirement of CA activity in the calcification pathway of coccolithophores, either in the cytosol or coccolith vesicle itself, as a purely uncatalyzed exchange would have too enriched

390 carbon isotope values in the coccolith calcite (Holtz et al., 2015; McClelland et al., 2017). Thus, it is probable that CA is present in coccolithophores. However, the activity of CA is not well constrained so it remains uncertain whether it is sufficient to ensure full equilibration of the DIC-H$_2$O system for both oxygen isotopes and Δ_{47}.

For *Gephyrocapsa*, the 1.5‰ range in $\Delta\Delta^{18}O_{off}$ also did not correlate with pH, growth rate, DIC, CO$_{2(aq)}$, or τ (Figs. S4, S5; Table S2). The 1.5‰ range in $\Delta\Delta^{18}O_{off}$ may be explained by varying degrees of isotopic equilibration between the DIC pool

395 and intracellular water in different experiments. Non-systematic disequilibria effects could be present for our continuous culture setup as the equilibration time for oxygen isotopic exchange between our bubbled CO$_2$ gas and the seawater media is in the order of hours (Zeebe and Wolf-Gladrow, 2001; Uchikawa and Zeebe, 2012). In support of this interpretation, there is a difference of ~0.5‰ between all continuous and batch culture $\Delta\Delta^{18}O_{off}$ (t-test; $t(38)$ = 1.76, p-value = 0.09, Fig. 5, Table S1). Further variability in $\Delta\Delta^{18}O_{off}$ could be a result of variable intracellular pH and/or varying calcification rates as well as the

400 presence and activity of CA. However, these physiological variations do not exhibit systematic relationships with the individual environmental parameters such as growth rate, external pH, or carbonate system. This suggests a potential for complex co-regulation of the physiological factors and/or multiple environmental controls, which are best evaluated by cellular models. and beyond the scope of this study.

4.2 Carbon isotope vital effects in coccolith calcite

405 In this study (Fig. 5), all continuous (and most batch) cultures for *G. oceanica* fall within the expected range of 0 ± 1.5‰ for $\Delta^{13}C_{c\text{-}DIC}$ as found in other studies (see Fig. S3; Rickaby et al., 2010; Moolna and Rickaby, 2012; Hermoso, 2014; Katz et al., 2017). There are no comparable culture studies for *G. muellerae*. The $\Delta^{13}C_{c\text{-}DIC}$ values fall within the range of *G. oceanica*. Both species will be combined in any inter-genera analyses for Sect. 4. *C. leptoporus* has a $\Delta^{13}C_{c\text{-}DIC}$ of -2.0 ± 1.0‰ comparable to previous culturing studies (Fig. 5; Ziveri et al., 2003; Hermoso et al., 2014; Katz et al., 2017).

410   Models of the vital effect in coccolith carbon simulate multiple processes affecting $\Delta^{13}C_{c\text{-}DIC}$ (Bolton and Stoll, 2013; Holtz et al., 2017; McClelland et al., 2017). The carbon utilization (or demand) as a function of the supply, termed τ, is an index used to describe changes in cellular carbon uptake (relative significance of diffusive CO_2 uptake vs active HCO_3^- uptake), and depletion of the intracellular dissolved carbon pool that impact the carbon isotopic fractionation (Eq. 3, Fig. 2). The cellular calcification (particulate inorganic carbon; PIC) relative to photosynthesis (particulate organic carbon; POC) is described as

415   the PIC:POC ratio. The PIC:POC intensity will modulate the $\Delta^{13}C_{c\text{-}DIC}$ response to τ. For moderate to heavily calcified cells with relatively high PIC:POC ratios, models simulate a decrease in $\Delta^{13}C_{c\text{-}DIC}$ with increasing τ, for example as might be triggered by decreasing concentrations of $CO_{2(aq)}$ (Eq. 3, Fig. 2). However, lightly calcified cells with PIC:POC of 0.5 exhibit a slight increase in $\Delta^{13}C_{c\text{-}DIC}$ with increasing τ, while cells with PIC:POC near 1 have $\Delta^{13}C_{c\text{-}DIC}$ nearly insensitive to τ. The *G.*

oceanica in our experiments feature a range of PIC:POC ratios from 0.4 to 1.3 (Torres-Romero et al., 2024A), which likely

420   contributes to the absence of a single clear trend between $\Delta^{13}C_{c\text{-}DIC}$ and τ. We have not determined the PIC:POC in our *Calcidiscus*, but it is generally characterized by a PIC:POC of 1.2-2.3 (Langer et al., 2006, 2012; Bolton and Stoll, 2013.). The lower $\Delta^{13}C_{c\text{-}DIC}$ of *Calcidiscus* correlate with lower $\Delta\Delta^{18}O_{off}$ (Figure 5), as observed in size fractions from sediments (e.g. Bolton and Stoll, 2013), but the physiological mechanism of this correlation has not yet been quantitatively explored in a coupled model of carbon and oxygen isotope fractionation in coccoliths.

425 **4.3 Calibration of coccolith clumped isotopes and temperature**

Despite the significant vital effects in carbon and oxygen isotopes, our coccolith Δ_{47} values show a consistent relationship with temperature (Fig. 6), ~~and~~which is similar to previous calibration studies (see Sect. 4.4, 4.5). In order to calculate a reliable coccolith Δ_{47}-temperature regression, we used a ~~bivariate~~simple least-squares fitting following Williamson (1968) and York et al. (2004), using the Excel spreadsheet by Cantrell (2008). This methodology considers the uncertainties from both the Δ_{47}

430   and temperature measurements. ~~However, as~~While the omni-variant generalised least squares regression would be better suited, as this incorporates the full error covariance (Daëron and Vermeesch, 2024), our data is standardised through reference materials in a moving time window and thus cannot be analysed through this method. As pointed out in a number of studies (Bonifacie et al., 2017; Fernandez et al., 2017; Katz et al., 2017; Kelson et al., 2017; de Winter et al., 2022), the bias from a low number of analytical replicates and the small temperature range can lead to significant differences in calibration slope and

435   intercepts. ~~Thus, as *G.*~~In order to determine potential species- or genus-dependent effects, an unorthodox approach was initially tested. *G. oceanica* contains the most diverse and largest range of temperature datapoints, while *C. leptoporus* only contains one temperature point and three Δ_{47} data points. Thus, the other two species' datasets will be successively included and evaluated for significance~~.~~ relative to the *G. oceanica* dataset. Further, each Δ_{47} ~~measurement~~biological and technical Δ_{47} sample and uncertainty ~~is~~are taken individually as to have an equal contribution of each datapoint to the final calibration. The

440   initial uncertainty of 0.1°C for temperature is 0.018 K^{-2} after conversion to $1/T^2$. The resulting slopes and intercepts of the Δ_{47}-temperature regressions are seen in Table 2. With the successive inclusion of the two other species, there is no significant change or offset in slope or intercept~~.~~ relative to the initial *G. oceanica* regression. All regression lines fall within 0.0012‰

error of each other, which shows that with the available data there is no species- or genus-specific vital effect on the Δ_{47}-temperature relationship.

445 Secondly, we tested the potential influence of variable carbonate chemistry and the carbon and oxygen "vital effects" on the Δ_{47}-temperature relationship. To this end, we normalized our Δ_{47} data against the Meinicke et al. (2020) Δ_{47}-temperature calibration at each temperature to generate residual values ($\Delta\Delta_{47,off}$). Pearson correlation tests were then performed for the normalized $\Delta\Delta_{47,off}$ residual values against the different carbonate chemistry parameters. The resulting Pearson correlation coefficients are shown in Table 3. Apart from a weak non-significant, positive and negative correlation for the batch and

450 continuous culture setup $\Delta\Delta_{47,off}$ and pH respectively and a moderate but significant positive correlation between the continuous culture setup $\Delta\Delta_{47,off}$ and $\Delta\Delta^{18}O_{off}$, there are no conclusive or significant correlations for all measurements and experimental setups. Any difference in Δ_{47} as a result of the two culture setups; continuous and batch, must also be established. There is no significant difference in the normalized $\Delta\Delta_{47,off}$ values between batch and continuous culture setups (t-test; t(38) = 0.35, p-value = 0.73), further indicating that carbonate chemistry does not have an effect on the measured Δ_{47} value. The non-

455 significant correlation in $\Delta^{13}C_{c\text{-}DIC}$ and $\Delta\Delta_{47,off}$ for all setups, shows that there is no carbon isotope vital effect affecting the coccolith Δ_{47}-values (Fig. S6). While there is a moderate, significant positive correlation between $\Delta\Delta^{18}O_{off}$-$\Delta\Delta_{47,off}$, it is only present for the continuous culture setup and has an r^2 of 0.20. Thus, there is no evidence of an important impact of the oxygen isotope vital effect on coccolith Δ_{47}-values.

Thirdly, average Δ_{47} values were calculated for each species at every growth temperature. These temperature-weighted

460 averages can highlight bias from a low number of measurement replicates at certain growth temperatures, such as at 6°C and 27°C. The resulting Δ_{47}-temperature regression is indistinguishable from regressions using individual Δ_{47} sample datapoints ((±6.1 ppm; Table 2).

Lastly, if there are no species-specific nor carbonate chemistry related vital effects in the Δ_{47}-temperature relationship, the data that were initially excluded due to the poorly characterized carbonate system (see Sect. 3.1.1) and poor constraining of the

465 stable carbon and oxygen isotope fractionations (i.e. datapoints with $\Delta_{DIC}\geq20\%$ or $\Delta_{pH}\geq0.20$) can also be included. If the Δ_{47}-temperature relationship in coccoliths is purely related to temperature, this will not interfere with the resulting calibration. Indeed, the slope and intercept fall within error of the other regressions, albeit with a slightly higher slope and lower intercept, seen in Table 2 and Fig. 6.

Based on the above analysis we conclude that all 55 datapoints can be considered for a coccolith Δ_{47}-temperature calibration

470 (T in K, ±1σ):

$$\Delta_{47}(I - CDES) = 0.0375 \pm 0.004 * \frac{10^6}{T^2} + 0.181 \pm 0.048 \tag{4}$$

4.4 A unified coccolith Δ_{47}-temperature calibration?

The similar culturing study of three coccolithophore species by Katz et al. (2017) also found no species-specific vital effects affecting the Δ_{47} values and a consistent Δ_{47}-temperature correlation. Furthermore, two common coccolithophore species (E.

475 *huxleyi* and *C. braarudii*) were cultured that were not included in our study. Comparable and overlapping Δ_{47} values were obtained for all temperatures. If we combine the clumped isotope data from this study and Katz et al. (2017) (Fig. 7) we obtain a coccolith Δ_{47}-temperature calibration that falls within the 95% confidence interval of Eq. 4:

$$\Delta_{47}(I-CDES) = 0.0381 \pm 0.003 * \frac{10^6}{T^2} + 0.176 \pm 0.037 \hspace{3cm} (5)$$

However, the study was conducted before the introduction of the I-CDES standardisation methodology using carbonates and
480 used gas-based standardization, consequently the data could have a systematic difference that cannot be resolved with certainty. Thus, when comparing to other calibration studies we will not include Katz et al. (2017) in the dataset and use Eq. 4 as a coccolith Δ_{47}-temperature calibration, which is only based on our culture data in the I-CDES frame.

4.5 Comparison with other calibrations

4.5 A coccolith or biogenic specific calibration?

485 In Fig. 7 and 8 we compare our data with the individual data of previous biogenic Δ_{47}-temperature calibration studies. The slopes and the intercepts of the equations with a limited number of replicates and a limited range of temperatures have larger uncertainties (Bonifacie et al., 2017; Fernandez et al., 2017; Kelson et al., 2017) and all datasets have a significant scatter due to their small variability. We focus on three five biogenic and one inorganic carbonate calibration studies (Peral et al., 2018; Meinicke et al., 2020; Anderson Caldarescu et al., 2021; de Winter et al., 2022). We also include the ‘; Huyghe et al., 2022)
490 and one inorganic (“MIT calibration’ equation of calibration”; Anderson et al., 2021; Daëron and Gray (, 2023), however as no new data was generated in that study only the recalculated calibration curve is shown.) carbonate studies. All datasets are converted to I-CDES using the methodology described in Bernasconi et al. (2021 (2021). The biogenic data sets are combined into a general ‘biogenic’ calibration, excluding this study. The dataset of Caldarescu et al. (2021) is included in the de Winter et al. (2022) calibration, using their equation 3. As suggested by Huyghe et al. (2022), the juvenile specimen that shows Δ_{47}
495 disequilibrium effects related to growth rate is not included. The planktic foraminifera data recalculated to I-CDES of Meinicke et al. (2020) and seen in Fig. 8 Peral et al. (2018) are combined. Both the original studies' and recalculated calcification temperatures by Daëron and Gray (2023) are used, and named ‘Daëron & Gray 2023 orig’ and ‘Daëron & Gray 2023 recal’ respectively.

While overall the individual Δ_{47} values for coccolith calcite fall within the scatter of the Peral et al. (2018) and Meinicke et al.
500 (2020) previous calibrations, our coccolith Δ_{47}-temperature calibration regression plots slightly systematically above previous calibrations, but generally below Katz et al. (2017). To identify and evaluate how this would impact temperature reconstructions, a similar exercise to Sect. 4.3 was performed. At all growth temperatures, a Δ_{47} value was calculated from each calibration. $\Delta\Delta_{47,off}$ residuals were then generated through comparison with our study's calibration at each temperature. Katz et al. (2017) yield positive residuals whereas the All non-coccolith calibrations yield negative residuals ($\Delta\Delta_{47,off}$) relative
505 to our coccolith Δ_{47}-temperature calibration (Fig. 9). This results in lower calculated Δ_{47} temperatures compared to our calibration. Similar results (Fig. 8). Positive offsets were found for aragonite and brachiopods by (de Winter et al. (., 2022)

and brachiopods (Bajnai et al., 2018; Davies et al., 2023; Letulle et al. ~~(2023) respectively,~~), when compared to the Anderson

et al. (2021) inorganic carbonate calibration. However, brachiopods show a complex system of kinetic disequilibrium effects

in both Δ_{47} and Δ_{48} associated with early-stage CO_2 absorption in the DIC, and do not have the CA enzyme. de Winter et al.

510 (2022~~)~~), in addition, argued that the Δ_{47}-temperature relationship is non-linear at temperatures >100°C and calibrations are

substantially biased by data with a large temperature range such as Anderson et al. (2021~~). Indeed, applying)~~ when used at

temperatures <30°C. The 'MIT' calibration uses only the inorganic carbonates from the Anderson et al. (2021) ~~calibration~~

~~dataset and indeed~~ yields lower and non-overlapping $\Delta\Delta_{47,off}$ residuals relative to our calibration~~, see Table 4 and~~ (Fig. ~~9.~~8).

Using only the data <30°C from Anderson et al. (2021) to calculate a temperature regression does yield overlapping $\Delta\Delta_{47,off}$

515 residuals relative to our calibration, however ~~it is~~ due to the steeper slope of the calibration, the $\Delta\Delta_{47,off}$ residuals are strongly

temperature dependent ~~(see Table S1). Yet, Letulle et al. (2023) argues against this bias from warmer temperatures and shows~~

~~that their offsets are similar for calibrations derived from the marine~~. The non-linearity of the Δ_{47}-temperature ~~realm (<30°C).~~

~~Offsets are instead~~relationship is thus most likely not a major cause for the consistent offset of our coccolith calibration (see

Table S3).

520 To a certain degree, offsets are also likely due to the uncertainty related to ~~imprecise~~the determination of calcification

temperatures in previous calibrations based on foraminifera collected from core tops. Often indirectly inferred or estimated

from other proxies, these uncertainties in the calcification temperatures can result in large variabilities, obscure potential effects

related to temperature, and can result in ~~small absolute differences between calibrations (see Daëron and Gray, 2023 for~~

~~discussion).~~ differences between calibrations. Meinicke et al. (2020) compared and discussed three different methods of

525 determining the calcification temperature of the foraminifera used in their study, and concluded that using their oxygen isotope

composition and the temperature calibration of Shackleton et al. (1974) provides the most robust estimate of the true

calcification temperatures. In an extensive study, Daëron and Gray (2023) re-determined the oxygen isotope fractionation in

the foraminifera species that were used by Meinicke et al. (2020) and Peral et al. (2018) for their clumped isotope calibrations

using samples from laboratory cultures and plankton tows. They concluded that foraminifera calcification temperatures are

530 best approximated by using the Kim and O'Neil (1997) calibration with species-specific offsets. They tested this concept by

comparing the calcification temperatures determined for the Meinicke et al. (2020) and Peral et al. (2018) datasets, with

reconstructed water column temperatures at the sites of the core tops (Fig. 7 in Daeron and Gray, 2023), and concluded that

this is a better estimate of calcification temperatures than previously published. The applications of these revised species-

specific oxygen isotope fractionation curves result in a non-systematic 1-2°C offset to colder temperatures from the original

535 studies of Peral et al. (2018) and Meinicke et al. (2020; Fig. 8), especially at temperatures below 15°C and leads to a match

between these two foraminifera-based calibrations and the inorganic calibration of Anderson et al. (2021). A close examination

of Fig 7 in Daeron and Grey (2023) however, shows that the majority of their discordant samples (i.e. those that record

temperatures outside the measured water column temperatures at the living depth of the specific species) are on the cold part

of the calibration, 12 are below 13°C. Moreover, at least 7 of the ones that are considered concordant as the error bars overlap

540 with the measured temperatures, have an average that is 1-2°C below the lowest measured temperature. About 27 of the 37

datapoints below 15 °C are either colder than the lowest temperature or only overlap with the coldest temperatures of the observed water column temperature range. We suggest that this would impart a cold bias on those samples. Further, the proposed extension of the calcification depths to 500 m would also give a cold bias and is not supported by foraminifera living depths (Schiebel and Hembleben, 2017). Based on these considerations, we suggest that the use of calcification temperatures

545 from oxygen isotopes need further testing, ideally on laboratory cultured specimens and warrants caution. Both the variability and absolute $\Delta\Delta_{47,off}$ residuals differ for all compared calibrations. Katz et al. (2017), Anderson et al. (2021), and Daëron and Gray (2023) yield $\Delta\Delta_{47,off}$ residuals that do not overlap with our dataset. Peral et al. (2018), Meinicke et al. (2020), and de Winter et al. (2022) yield $\Delta\Delta_{47,off}$ residuals within the uncertainty of our dataset, see Table 4 and Fig. 9. Temperature offsets at the average growth temperature of our dataset, 18.7°C, are between -3°C and 3°C for the different calibrations, although

550 vary in magnitude at different temperatures, see Table 4 and Fig. 9.

Both the variability and absolute $\Delta\Delta_{47,off}$ residuals differ for all compared calibrations. Our tightly-controlled growth temperatures of the culture data eliminate the problem of uncertain calcification temperatures as in the case of foraminifera calibrations (Meinicke et al., 2020; Daëron and Gray, 2023). It also allows the exclusion of effects from environmental variabilities, such as pCO₂, pH, growth rate, and vital effects on measured Δ_{47} values as discussed in Sect. 4.3. It is important

555 to note that the vital effects in the coccolith carbon and oxygen isotopes have no impact on their Δ_{47}. Furthermore, modelling studies for inorganic calcite, grown in the presence of CA and at growth rates and pH ranges relevant for coccolithophores, also show clear vital effects for carbon and oxygen but find no distinction between equilibrium and modelled Δ_{47} (Watkins and Hunt, 2015; Hill et al., 2020; Uchikawa et al., 2021; Watkins and Devriendt, 2022). While all compared biogenic calibrations fall within ±1°C of this studies' calibration, the inorganic carbonate calibrations yield systematically colder

560 temperatures by at least 2°C (Fig. 9a, c). As our study was performed with strict temperature regulation, any related uncertainty can be eliminated as a source of inter-calibration variability. As all compared biogenic carbonates are within uncertainty of our study, we suggest that a constant biological mechanism is causing the offset of biogenic relative to inorganic carbonates. A combination of all data from Peral et al. (2018), Meinicke et al. (2020), de Winter et al. (2022), and this study yields a general biogenic Δ_{47} temperature calibration (Eq. 6; Fig. 9b).

565 $$\Delta_{47}(I - CDES) = 0.0371 \pm 0.001 * \frac{10^6}{T^2} + 0.184 \pm 0.016 \tag{6}$$

All previous biogenic calibrations fall within ±1°C and the 95% confidence interval of the general biogenic calibration (Fig. 9b). Thus, when reconstructing calcification temperatures from biogenic carbonates, we suggest the use of a well-constrained and biogenic specific calibration such as Eq. 4.

The Daëron and Gray (2023) and MIT calibrations yield $\Delta\Delta_{47,off}$ residuals that do not overlap with our dataset between 2-28°C.

570 The original Peral et al. (2018), Meinicke et al. (2020), de Winter et al. (2022), and Huyghe et al. (2022) calibrations do yield $\Delta\Delta_{47,off}$ residuals within the uncertainty of our dataset. Temperature offsets at the average growth temperature of our dataset, 18.7°C, are between 0°C and 4°C for the different calibrations, but vary in magnitude at different temperatures, see Table 4.

The high variability of the temperature offsets over the given temperature range for certain biogenic carbonate studies such as de Winter et al. (2022), suggest non-uniform disequilibrium effects are present. Thus, a pooled calibration including all biogenic studies mentioned, except this study, and using either the original and recalculated calcification temperatures were performed to identify whether there is potential for a general biogenic-specific calibration (Fig. 8). Our dataset still has a positive $\Delta\Delta_{47,off}$ relative to the biogenic calibrations, for the original calcification temperatures from 2°C to 1.5°C for 27°C to 6°C respectively and from 3°C to 2°C for the recalculated dataset. From this, two potential conclusions can be argued; there is a coccolith specific-calibration or the calcification temperatures for previous studies are underestimated. Our well-constrained coccolith cultures allow for the exclusion of effects from environmental variabilities, such as pCO_2, pH, coccolithophore growth rate, and vital effects on Δ_{47} values as discussed in Sect. 4.3. The vital effects observed in the coccolith carbon and oxygen isotopes do not have an impact on the Δ_{47}, where $\Delta\Delta_{47,off}$ are within 0.0012‰ despite up to ~3‰ variability for carbon and oxygen isotopes for our experimental conditions. This is below our analytical uncertainty for Δ_{47}, thus while we can't fully rule out that vital effects are present on Δ_{47}, the offset from the inorganic equilibrium calibration must be similar at all temperatures, systematic and unrelated to vital effects. Indeed, for the available dataset no species- or genus-related effects are present, although this must be further examined in future studies with in situ temperatures such as sediment traps or cultures. Foraminifera also show variable oxygen and carbon isotope vital effects, although less extreme than coccolithophores, but follow the same Δ_{47} relationship.

The tightly-controlled growth temperatures of our cultures eliminate the problem of uncertain calcification temperatures as in the case of foraminifera calibrations (Meinicke et al., 2020; Daëron and Gray, 2023). Indeed, the offsets of our coccolith calibration relative to the inorganic MIT calibration are consistent and suggest a uniform disequilibrium mechanism ($\Delta\Delta_{47,off}$ = -0.0079±0.0005‰). The purely foraminifera-based calibrations, both the original and recalculated calcification temperatures of Peral et al. (2018) and Meinicke et al. (2020), show more variability relative to the MIT calibration ($\Delta\Delta_{47,off}$ = 0.0023±0.0012‰ and $\Delta\Delta_{47,off}$ = -0.0013±0.0015‰ respectively), and suggests variable disequilibrium effects. However, at this stage it is difficult to state whether this is due to general foraminifera, species-specific disequilibria, or due to uncertainties related to the calcification temperature. This highlights the importance and uncertainties related to the calcification temperature and their implications. Modelling studies for inorganic calcite, grown in the presence of CA and at growth rates and pH ranges relevant for coccolithophores, show offsets for carbon and oxygen and find kinetic disequilibrium effects that are pH and crystal-growth rate dependent (Hill et al., 2014; Watkins and Hunt, 2015; Uchikawa et al., 2021; Watkins and Devriendt, 2022). Between our cultured temperatures of 6-27°C and in saline conditions, a shift from pH 8 to 8.5 can give an offset of ~3‰ for $\delta^{18}O$ and ~90 ppm for Δ_{47} at very fast crystal growth rates (10^{-5} mol$m^{-2}s^{-1}$) and offsets of ~1‰ for $\delta^{18}O$ and ~1 ppm for Δ_{47} for 1000x slower crystal growth rates (10^{-7} mol$m^{-2}s^{-1}$; Fig. 6 in Watkins and Devriendt, 2022). The latter is comparable to estimated coccolith calcification rates of 10^{-7}-10^{-8} mol$m^{-2}s^{-1}$ (Langer et al., 2006, 2012; Sett et al., 2014). Also, the modelled difference of Δ_{47} between HCO_3^- relative to CO_3^{2-} at 25°C, with a difference of ~5 pH units, and saline conditions is ~0.034‰, and ~7‰ for $\delta^{18}O$ (Hill et al., 2014; Watkins and Devriendt, 2022). Thus, the difference due to calcification mechanisms by regulating internal pH and thus the HCO_3^-/CO_3^{2-} ratio that has been postulated to drive the carbon and oxygen vital effect

differences between different coccolithophore and foraminifera species cannot be applied on its own to explain the Δ_{47} offset from equilibrium (Spero et al., 1997; Rickaby et al., 2010; Ziveri et al., 2012; Devriendt et al., 2017; Holtz et al., 2017). Rather, a crystal growth rate variability between species is likely the cause of the 3‰ offset in $\Delta\Delta^{18}O_{off}$ between *C. leptoporus* and *G. oceanica* and only ±1.2 ppm difference in Δ_{47}. Perhaps, a difference due to species-specific coccolith-associated-proteins that can influence different calcite saturation states within the coccolith vesicle and lead to more crystal-growth rate variability, can also have an effect (Lee et al., 2016). However, as the offset from the equilibrium inorganic calcite is systematic across the three coccolithophore species cultured here, there is more work needed to identify the disequilibrium processes for other coccolithophore species as well as for foraminifera. Future studies with constrained and in situ temperature measurements such as in sediment traps (Clark et al., 2024) or cultures are recommended to disentangle and validate our findings of this biogenic disequilibrium.

5 Conclusions

In this study we cultured three species of coccolithophores (*Gephyrocapsa oceanica*, *G. muellerae*, and *Calcidiscus leptoporus*) in continuous and batch culture setups for temperatures between 6 and 27°C and $CO_{2(aq)}$ between 5 and 45 µM. Vital effects in carbon and oxygen isotopes in coccoliths were observed and their magnitude is consistent with previous culturing studies. We show that for our well-constrained continuous culture setup there are no systematic external environmental influences from pH, DIC, or $CO_{2(aq)}$ on the carbon and oxygen isotopic values for both *G. oceanica* and *G. muellerae*. Both species precipitate coccolith calcite close to isotopic equilibrium with the water. ($\Delta^{13}C_{c-DIC}$ = ±1.5‰; $\Delta\Delta^{18}O_{off}$ = ±1‰). *C. leptoporus* shows pronounced vital effects in both carbon and oxygen isotopes, although no clear physiological conclusions can be drawn on the source of these vital effects. A calcification model would aid in describing the sources of the vital effects.

We establish a coccolith-specific Δ_{47}-temperature calibration and ~~find no distinguishable vital~~observe a consistent offset from previous inorganic calibrations ($\Delta\Delta_{47,off}$ = -0.0079±0.0005‰), which suggests disequilibrium effects in coccolith Δ_{47} for all three coccolithophore species. There are ~~also~~however, no effects on the calibration are due to differences in environmental parameters or species. Our coccolith Δ_{47} data is largely consistent with a previous coccolith culture study (Katz et al., 2017), and indicates that coccolithophores precipitate coccolith calcite in clumped isotope ~~equilibrium~~disequilibrium with their environment. Our well-constrained coccolith Δ_{47}-temperature calibration (Eq. 4) also shows ~~no consistent offset~~ offsets from previous biogenic calibration studies ~~(≤2°C). Combining,~~ dependent on the ~~previous biogenic calibrations into a general Δ_{47}-~~ choice of calcification temperature~~-~~. The discrepancies derived from the differences in calcification temperature render it difficult to conclusively state whether a general biogenic calibration ~~(Eq. 6) yields better constraints and no consistent offsets between calibrations (±2°C). However, there is a systematic offset of ≥2°C from previous inorganic calibrations. Thus, while all previous biogenic calibrations can~~should be used ~~interchangeably~~. Thus, we suggest the use of our well-constrained Δ_{47}-temperature calibration to reconstruct temperatures from well screened/preserved coccolith calcite~~.~~, regardless of species.

Author contributions.

640 AJC: cell culture, analyses, and writing, ITR: cell culture and analyses, MJ: analyses, SMB: supervision, analyses, funding acquisition, and writing, HMS: supervision, funding acquisition, and writing.

Competing interests

The authors declare that they have no conflict of interest.

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Tables

Species	Temperature (°C)	Final $CO_{2(aq)}$ (μM)	Culture setup type	$\Delta^{13}C_{c\text{-DIC}}$ (‰ V-PDB ± 1σ)	$\Delta^{18}O_{c\text{-sw}}$ (‰ V-SMOW ± 1σ)	Δ_{47} (‰ I-CDES ± 1SE)
G. oceanica	12	14.66	Batch	-0.40 ± 0.47	32.95 ± 0.34	0.640 ± 0.009
	15	21.56	Continuous	-0.87 ± 1.12	31.29 ± 0.26	0.621 ± 0.009
	15	33.01	Continuous	0.03 ± 0.18	31.57 ± 0.25	0.634 ± 0.008
	15	40.18	Continuous	-0.24 ± 0.13	31.68 ± 0.26	0.637 ± 0.013
	18	6.94	Continuous	-1.15 ± 0.69	31.37 ± 0.13	0.628 ± 0.009
	18	7.95	Continuous	-1.41 ± 0.70	31.03 ± 0.26	0.629 ± 0.005
	18	18.41	Continuous	0.54 ± 1.00	30.65 ± 0.15	0.620 ± 0.004
	18	24.12	Continuous	-0.95 ± 1.00	30.74 ±0.13	0.620 ± 0.010
	18	27.73	Continuous	0.51 ± 1.00	30.63 ± 0.12	0.629 ± 0.008
	18	42.58	Continuous	-0.78 ± 0.35	30.45 ± 0.11	0.633 ± 0.005
	21	5.62	Continuous	-0.24 ± 0.25	30.45 ± 0.14	0.624 ± 0.007
	21	6.18	Continuous	0.75 ± 0.25	30.47 ± 0.17	0.611 ± 0.008
	21	6.18	Continuous	0.69 ± 0.27	30.37 ± 0.22	0.626 ± 0.004
	21	7.63	Continuous	-0.61 ± 0.25	30.44 ± 0.35	0.621 ± 0.005
	21	11.68	Continuous	-0.38 ± 0.15	30.15 ± 0.11	0.618 ± 0.012
	21	12.83	Continuous	-0.43 ± 0.13	30.26 ± 0.07	0.608 ± 0.012
	21	21.23	Continuous	1.22 ± 1.05	30.77 ± 0.33	0.620 ± 0.005
	21	21.23	Continuous	1.22 ± 1.05	30.76 ± 0.33	0.604 ± 0.008
	21	29.48	Continuous	-0.49 ± 0.32	30.74 ± 0.86	0.630 ± 0.007
	21	30.76	Continuous	-0.66 ± 0.32	30.92 ± 0.86	0.625 ± 0.011
	21	43.22	Continuous	-1.21 ± 1.05	30.57 ± 0.34	0.603 ± 0.004

	21	43.22	Continuous	-1.25 ± 1.05	30.56 ± 0.33	0.607 ± 0.006
	21	43.76	Continuous	-0.98 ± 1.05	30.45 ± 0.33	0.611 ± 0.010
	24	13.05	Continuous	0.78 ± 0.34	29.39 ± 0.09	0.599 ± 0.005
	24	13.05	Continuous	0.85 ± 0.34	29.44 ± 0.10	0.598 ± 0.008
	24	13.73	Batch	-0.80 ± 0.28	29.09 ± 0.41	0.605 ± 0.007
	24	14.07	Batch	-0.21 ± 0.28	29.70 ± 0.37	0.611 ± 0.008
	24	16.95	Continuous	0.65 ± 0.34	29.72 ± 0.10	0.607 ± 0.010
	24	19.18	Continuous	-0.87 ± 0.39	29.38 ± 0.14	0.589 ± 0.006
	27	15.11	Continuous	0.61 ± 0.82	29.10 ± 0.13	0.603 ± 0.007
	27	15.11	Continuous	0.61 ± 0.82	29.03 ± 0.11	0.597 ± 0.007
G. muellerae	6	14.40	Batch	-0.88 ± 0.03	33.90 ± 0.28	0.659 ± 0.006
	6	14.57	Batch	-0.57 ± 0.03	34.00 ± 0.28	0.655 ± 0.005
	12	16.34	Batch	0.03 ± 0.21	32.48 ± 0.71	0.648 ± 0.005
	12	21.22	Batch	0.21 ± 0.21	32.98 ± 0.70	0.645 ± 0.008
	18	7.94	Batch	-0.88 ± 0.35	31.28 ± 0.24	0.618 ± 0.010
	18	9.80	Batch	-0.26 ± 0.35	31.31 ± 0.24	0.629 ± 0.009
C. leptoporus	12	17.94	Batch	-2.34 ± 0.42	29.78 ± 0.36	0.645 ± 0.006
	12	17.94	Batch	-2.22 ± 0.42	29.93 ± 0.35	0.652 ± 0.005
	12	19.12	Batch	-2.86 ± 0.49	29.89 ± 0.57	0.636 ± 0.008

945 **Table 1. Overview table for all included data points for Sect. 3, 4.1, 4.2. including temperature (°C), culture setup type, $CO_{2(aq)}$ (µM), $\Delta^{13}C_{c\text{-DIC}}$ (‰; VPDB), $\Delta^{18}O_{c\text{-sw}}$ (‰; VSMOW), and Δ_{47} values (‰; I-CDES). 1 standard deviation (σ) is included for the $\Delta^{13}C_{c\text{-DIC}}$ and $\Delta^{18}O_{c\text{-sw}}$, which take the evolution of the media over time into account, and 1 standard error (SE) for Δ_{47}.**

Data type	Slope	Intercept
G. oceanica	0.0377 ± 0.009	0.179 ± 0.104
G. oceanica + *G. muellerae*	0.0360 ± 0.005	0.198 ± 0.063
G. oceanica + *C. leptoporus*	0.0387 ± 0.007	0.168 ± 0.083
Temperature-weighted averages	0.0358 ± 0.006	0.202 ± 0.073
All species	0.0367 ± 0.005	0.190 ± 0.059
Including excluded data	0.0375 ± 0.004	0.181 ± 0.048

Table 2. Slope and intercepts with 1σ for the Δ_{47}-temperature regression using the Williamson-York bivariate least-squares method.

950

	pH	DIC	$CO_{2(aq)}$	$\Delta^{13}C_{C\text{-DIC}}$	$\Delta\Delta^{18}O_{off}$
All measured $\Delta\Delta_{47,off}$	0.17	-0.01	-0.09	0.04	0.08
Continuous setup $\Delta\Delta_{47,off}$	0.27	-0.04	-0.13	0.09	0.45*
Batch setup $\Delta\Delta_{47,off}$	-0.32	0.20	0.11	0.13	-0.19

Inserted Cells

Inserted Cells

Table 3. Pearson correlation coefficients (ρ) between ΔΔ$_{47,off}$ and pH, DIC, and CO$_{2(aq)}$ for all measurements, continuous culture, and batch culture setup. ~~For all p,~~Asterisk indicates where p-~~values >~~value is < 0.05.

	Measured Δ$_{47}$	~~Katz et al. (2017)~~	Peral et al. (2018)		Meinicke et al. (2020)		~~Anderson et al. (2021)~~Daëron and Vermeesch (2024) MIT	~~de Winter et al. (2022)~~Daëron and Gray (2023) orig	Daëron and Gray (2023) recal
ΔΔ$_{47,off}$ residuals (ppm ± 1σ)	-0.6 ± 3.4		~~8~~3.2 ± 0.~~2~~7	~~0.9 ± 2.4~~	-3.1 ± 1.3		-~~8~~7.9 ± ~~0 ±~~ ~~1.0~~.5	-4.3 ± 1~~5~~.7 ± 0.7	-~~7.6~~8.5 ± 0.6
Temperature offset (°C ± 1σ)	0.19 ± 1.12		-~~2.67~~1.03 ± 0.~~88~~02		-~~0.30~~99 ± 0.~~81~~36	~~1.04 ± 0.43~~	-2.7~~05~~6 ± 0.3~~20~~5	-1.4~~48~~6 ± 0.5~~53~~5	-2.5~~57~~7 ± 0.1~~93~~9

955 **Table 4. Average ΔΔ$_{47,off}$ residuals relative to our dataset comparing our coccolith calibration equations and those from ~~Katz et al. (2017), the~~ Peral et al. (2018), Meinicke et al. (2020), ~~Anderson et al. (2021), de Winter et al. (2022), and~~Daëron and Vermeesch (2024) MIT, Daëron and Gray (2023~~).~~) original temperatures, and Daëron and Gray (2023) recalculated temperatures. Equivalent temperature offset is based on the equivalent ~~Meinicke et al. (2020)~~ Δ$_{47}$ value at the average growth temperature (18.7°C~~).~~) using Equation 4.**

Figures

(a)

(b)
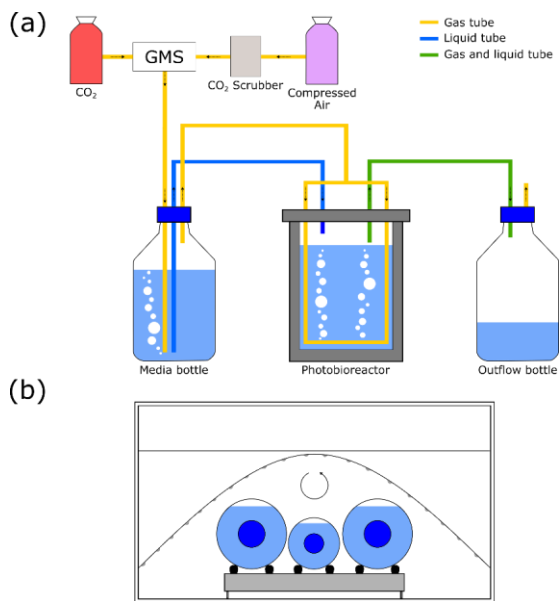
Figure 1. Photobioreactor and incubator systems. (a) Photobioreactor with controlled CO₂ gas input, adjusted from Zhang et al. (2022), (b) Incubator with roller containing 2 2L and 1 1L bottle and LED strip for batch cultures.

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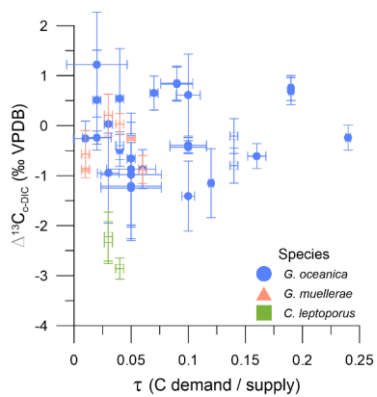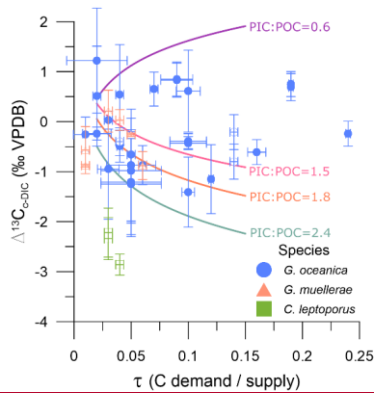
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Figure 2. Coccolith carbon isotope fractionation relative to external DIC ($\Delta^{13}C_{c\text{-DIC}}$) against the carbon demand vs supply (τ). Blue circles are *G. oceanica*, orange triangles are *G. muellerae*, and green squares are *C. leptoporus*. Open and filled symbols are batch and continuous cultures respectively. Error bars are as described in methods. Lines represent the model simulations of McClelland et al. (2017) at different PIC:POC ratios; purple = 0.6, pink = 1.5, orange = 1.8, grey = 2.4.

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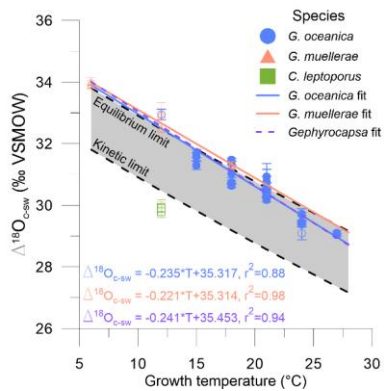
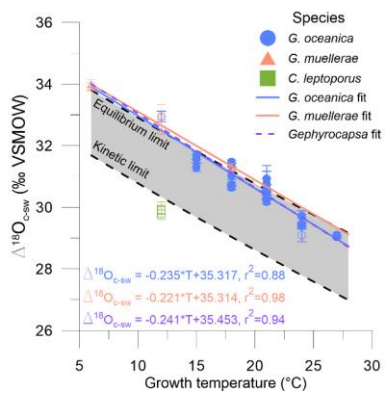


Figure 3. The fractionation of oxygen isotopes in coccolith calcite from seawater ($\Delta^{18}O_{c\text{-}sw}$), both in VSMOW, in relation to the growth temperature in °C. Blue circles are *G. oceanica*, orange triangles are *G. muellerae*, and green squares are *C. leptoporus*. Open and filled symbols are batch and continuous cultures respectively. Linear regression fits are calculated for *G. oceanica* and *G. muellerae* separately, as well as for the whole *Gephyrocapsa* genus, with equivalent formulae. Error bars are as described in methods, temperature error bars are smaller than symbols. The kinetic and equilibrium limits are from Watkins et al. (2014) and defined as $\left(\frac{17747}{T+273.15}\right) - 29.77$ and $\left(\frac{\cancel{17747}}{\cancel{T+273.15}}\right)\left(\frac{17965}{T+273.15}\right) - \cancel{31.77}32.67$ respectively.

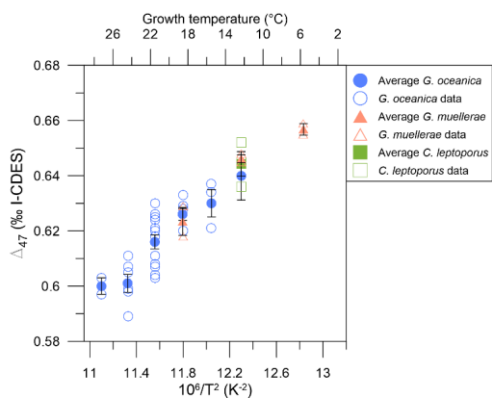



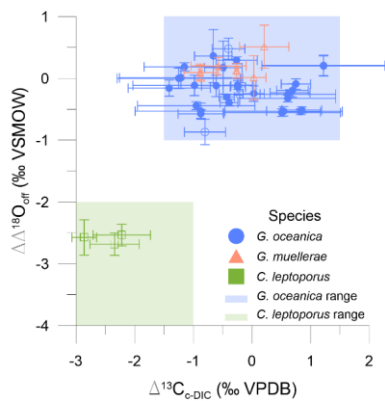
Figure 4. Δ_{47} values from coccolith calcite versus $10^6/T^2$ for all data points and averages. Averages are full symbols, while individual datapoints are empty symbols. Blue circles are *G. oceanica*, orange triangles are *G. muellerae*, and green squares are *C. leptoporus* data. Error bars are as described in methods, temperature error bars are smaller than symbols.

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Figure 5. The oxygen isotope offset from the equilibrium limit in Watkins et al., (2014) ($\Delta\Delta^{18}O_{off}$) against the coccolith carbon isotope fractionation relative to external DIC ($\Delta^{13}C_{c-DIC}$). Blue circles are *G. oceanica*, orange triangles are *G. muellerae*, and green squares are *C. leptoporus* data. Open and filled symbols are batch and continuous cultures respectively. The $\Delta^{13}C_{c-DIC}$ and $\Delta\Delta^{18}O_{off}$ range of previous *G. oceanica* and *C. leptoporus* studies are given as a blue and green background respectively. Error bars are as described in methods.
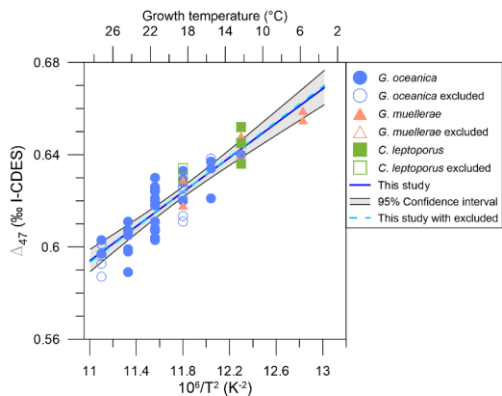
Figure 6. Δ_{47} values compared to temperature (in K) for cultured coccoliths. York regression and its 95% confidence interval are shown as a blue line and light grey envelope. The species are colour and symbol coded following Fig. 2. Blue circles are *G. oceanica*, orange triangles are *G. muellerae*, and green triangles are *C. leptoporus*. Empty symbols are excluded data but still included for illustration and the coccolith Δ_{47}-temperature calibration.

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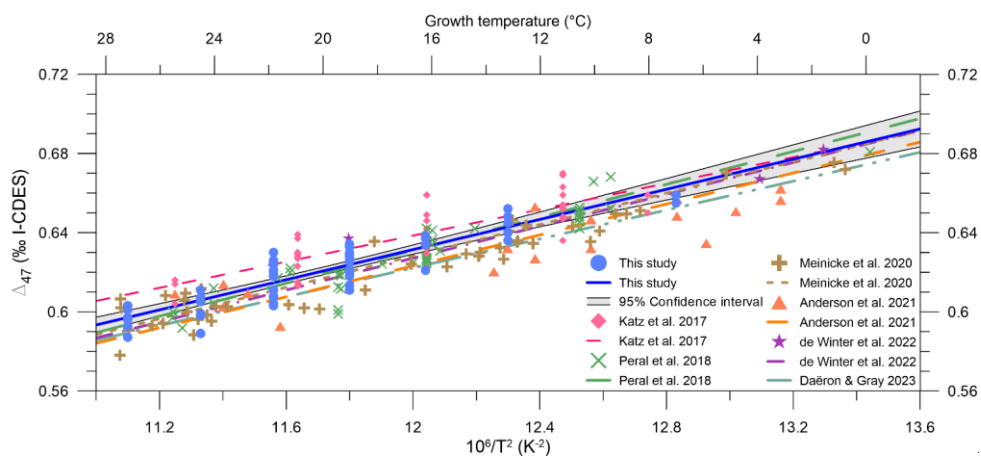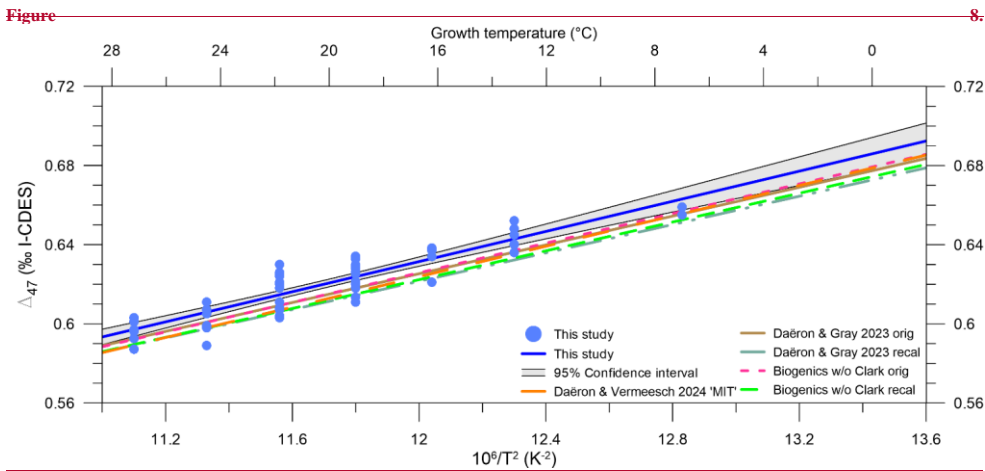
Figure 7. Δ_{47} versus temperature for coccolith calcite and previous biogenic and inorganic calibrations. The York linear fit and 95% confidence interval for the unified coccolith calcite Δ_{47}-temperature is shown as a blue line with grey shading. ~~Katz et al. (2017; pink diamonds), Peral et al. (2018; green X's), Meinicke et al. (2020; brown crosses), Anderson et al. (2021; orange triangles), and de Winter et al. (2022 purple stars). The calibration curves are also shown as dashed lines, with the same colour scheme, Daëron and Gray (2023) is shown as a grey dashed-dot line.~~Daëron and Gray (2023 orig; brown dash-dot dot-dash line), Daëron and Gray (2023 recal; grey dash-dot dot-dash line), Daëron and Vermeesch (2024 'MIT'; orange dashed line), biogenic calibration without this study using original published temperatures (pink dashed line), and biogenic calibration without this study using recalculated temperatures (green dashed line).
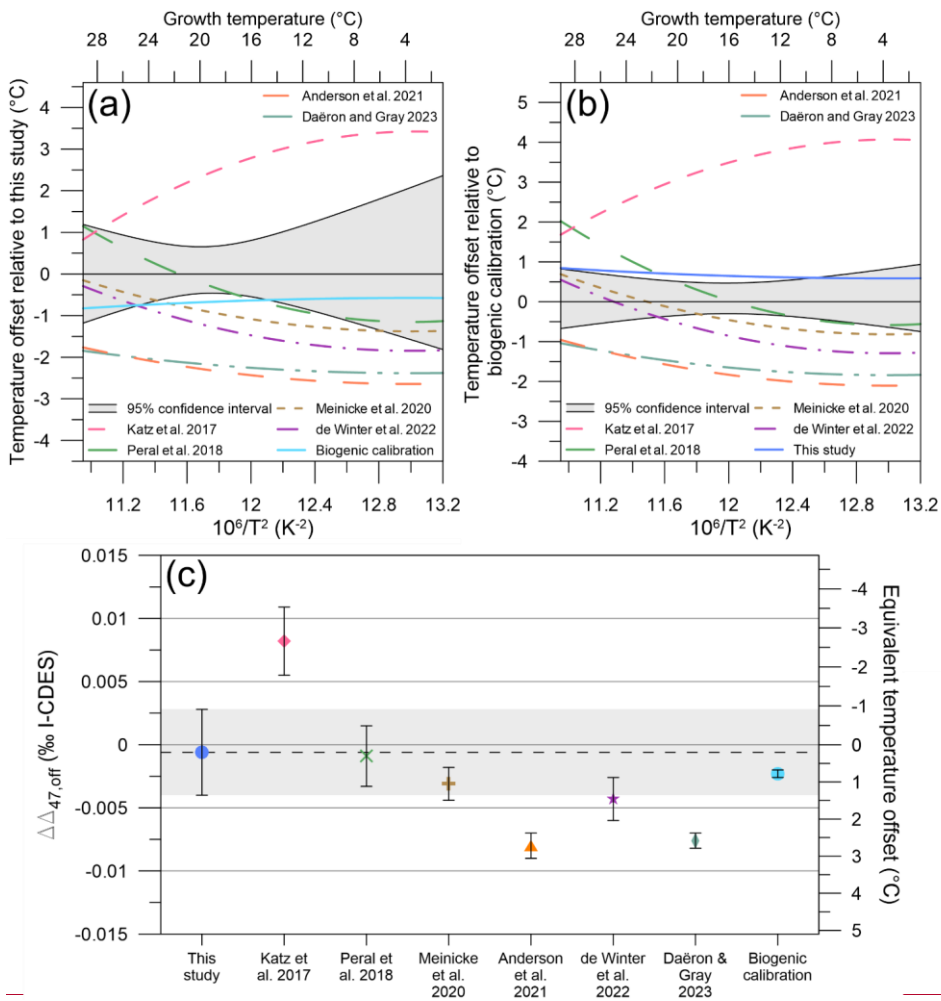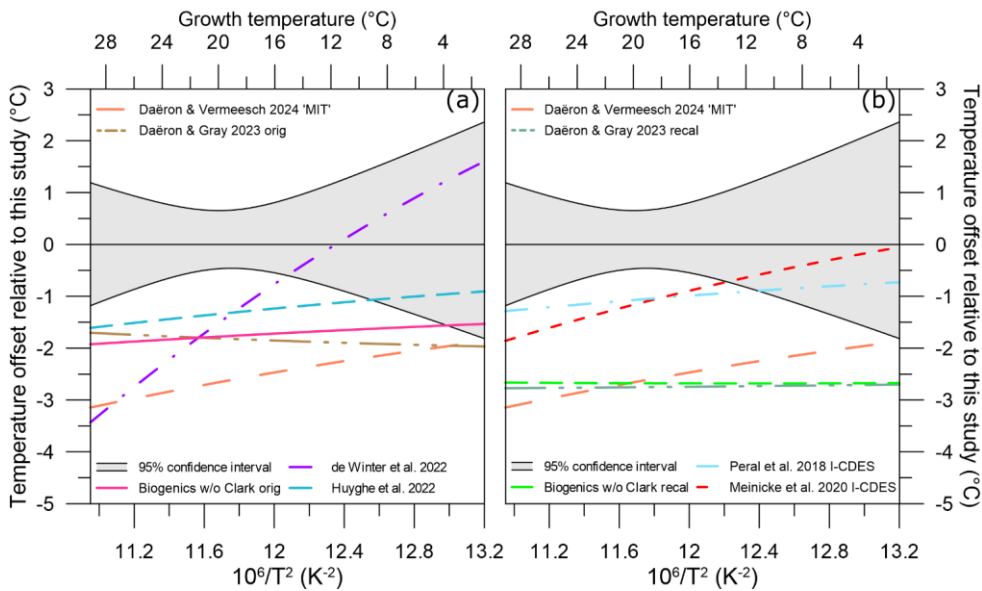
Figure ~~9~~8. Temperature offset of different Δ_{47}-temperature calibrations relative to ~~(a)~~ our study's equivalent temperature~~, (b) the general biogenic Δ_{47}-temperature calibration, (c) our study's equivalent at the average growth temperature (18.7°C), with average $\Delta\Delta_{47,off}$ residuals of different Δ_{47}-temperature calibrations relative to this study's dataset.~~. The other studies' calibration data are colour coded following Fig. ~~8. The general biogenic Δ_{47}-temperature calibration is given as~~7, with additional studies of Peral et al. (2018; light blue~~.~~ dash-dot-dash line) and Meinicke et al. (2020; red dashed line), de Winter et al. (2022; purple dash-dot-dash line), and Huyghe et al. (2022; blue dashed line) using original published temperatures. The 95% confidence interval for our study is given as light grey shading ~~in (a), (b). This study's average $\Delta\Delta_{47,off}$ and range is given as a dotted line and light grey shading in (c).~~.