



1 **Differential vulnerability of mineral-associated and particulate soil organic**  
2 **carbon to nitrogen addition in a subtropical forest**

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20



21 **Abstract**

22 Nitrogen (N) deposition rates of terrestrial ecosystems have gradually declined but are  
23 still high in some areas, and previous studies have reported that N addition elicits  
24 diverse impacts on soil organic carbon (SOC) pools. SOC can be divided into different  
25 functional fractions, namely particulate organic carbon (POC) and mineral-associated  
26 organic carbon (MAOC). The responses of these fractions to N addition should be  
27 elucidated to better understand the changes in SOC pools. We investigated the effects  
28 of N addition treatments (+0, +40, and +80 kg N ha<sup>-1</sup> yr<sup>-1</sup>) in a subtropical *Castanopsis*  
29 *fabri* forest to simulate N deposition. The surface (0-10 cm) SOC content in different  
30 fractions, aboveground litter biomass, fine root (diameter < 2 mm) biomass, soil  
31 exchangeable cation content, and soil enzyme activity under different N addition  
32 treatments were measured. The results showed that 1) N addition exerted a positive  
33 effect on POC content but did not significantly affect MAOC content. 2) POC content  
34 was negatively correlated with pH and soil enzyme activity and positively correlated  
35 with aboveground litter biomass, suggesting that POC accumulation was influenced by  
36 aboveground litter input and microbial decomposition. 3) Root biomass was unaffected  
37 significantly by the addition of N, which could be responsible for the limited response  
38 of MAOC to N addition. Furthermore, a close negative relationship was observed  
39 between exchangeable Al<sup>3+</sup> and Ca<sup>2+</sup> or K<sup>+</sup> contents, indicating the presence of a trade-  
40 off between negative effects of exchangeable cations on SOC bridging and their  
41 positive effects on SOC adsorption, thus resulting in an insignificant reaction of MAOC  
42 to N addition. Overall, N addition reduces the persistence and increases the nutrient  
43 density of SOC, and MAOC with more protection is less vulnerable to N addition than  
44 POC with less protection. By incorporating these nuances into ecosystem models, it is  
45 possible to more accurately predict SOC dynamics in response to global change.

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47

48 **Keywords:** carbon persistence; exchangeable cations; nitrogen addition; soil organic  
49 carbon fractions



## 50 **1 Introduction**

51 As the biggest carbon (C) sink in terrestrial ecosystems, the soil organic carbon  
52 (SOC) pool is essential to the C cycle-climate feedback (Bradford et al., 2016). C and  
53 nitrogen (N) cycles are considered tightly linked (Greaver et al., 2016; Cheng et al.,  
54 2019). Increased atmospheric N deposition resulting from excessive agricultural,  
55 industrial, and urban development has adversely affected SOC quality and ecological  
56 diversity in recent decades. This phenomenon has garnered notable attention from the  
57 scientific community concerning its impact on SOC pool (Clark and Tilman, 2008).  
58 Although the N deposition rate has gradually stabilized, it is higher in southeast China  
59 than in other regions within the country (Yu et al., 2019). Several researches have  
60 examined the effects of N addition on SOC and have reported that SOC pool may rise,  
61 fall, or remain unchanged with N addition (Li et al., 2022; Wang et al., 2023; Wu et al.,  
62 2023); overall, there is no unified conclusion on the effect of adding N on SOC.

63 Changes in plant C supply, SOC decomposition, microbial turnover, and  
64 interactions between SOC and soil minerals all may affect the response of SOC to N  
65 addition (Ye et al., 2018; Chen et al., 2021). Complex SOC composition may be the  
66 reason for the varied responses of SOC to N addition (Six et al., 2002; Lugato et al.,  
67 2021). Recently, the theory that the chemical complexity of SOC primarily governs its  
68 persistence has been criticized for lacking thermodynamic evidence and for neglecting  
69 microbial accessibility to substrate components (Schmidt et al., 2011). On the contrary,  
70 the mineral-aggregate protective mechanism is considered as an acceptable explanation  
71 for SOC persistence. Based on particle size widely used in SOC fractionation, SOC can  
72 be divided into particulate organic carbon (POC, >53  $\mu\text{m}$ ) and mineral-associated  
73 organic carbon (MAOC, <53  $\mu\text{m}$ ) (Lavallee et al., 2020). Dividing SOC into different  
74 functional fractions may better elucidate SOC dynamics under N addition (Chen et al.,  
75 2020; Lavallee et al., 2020). The majority of POC consists of plant components that are  
76 readily decomposable (i.e., carbohydrates and aliphatic compounds), and it has a faster  
77 turnover rate (Witzgall et al., 2021). MAOC, is primarily made up of low-molecular-



78 weight compounds (i.e., root exudates) and is shielded from breakdown by the chemical  
79 bonds between SOC and minerals, as well as microaggregates; thus, MAOC has a  
80 slower turnover rate (Poehlau et al., 2018) and is relatively stable (Lavellee et al., 2020).

81 Acidification is a common reaction of soil to N addition because of the 1) release  
82 of  $H^+$  during nitrification, 2) release of  $H^+$  after plant uptake of  $NH_4^+-N$ , and 3) leaching  
83 of exchangeable cations (Tian and Niu, 2015). The different functional fractions of SOC  
84 are susceptible to soil acidification. For instance, Rocci et al. (2021) elucidated that N  
85 addition resulted in increased POC and MAOC levels, and attributed the accumulation  
86 of POC to soil acidification-induced reduction in microbial biomass and SOC  
87 decomposition rate (Riggs and Hobbie, 2016). Moreover, N addition can enhance plant  
88 productivity (Feng et al., 2023), which may promote POC accumulation. According to  
89 Angst et al. (2021), about 60% of MAOC comes from plants, and plant-derived  
90 compounds are directly adsorbed onto soil minerals or aggregates (Craig et al., 2022),  
91 which promotes MAOC formation. Conversely, studies have shown adverse effects of  
92 N addition on POC and MAOC levels (Ye et al., 2018; Zhang et al., 2019; Chen et al.,  
93 2020). Guo et al. (2010) proposed that N addition reduced the production of microbial  
94 necromass and did not support the stable soil aggregate formation. Therefore, in the  
95 absence of physical barrier protection, POC is more easily decomposed by  
96 microorganisms (Lehmann and Kleber, 2015), resulting in a reduction in POC content.  
97 The size of MAOC pool is determined by the equilibrium between adsorption and  
98 desorption rates of charged components (Wu et al., 2023). The fast leaching of cations  
99 (e.g.,  $Ca^{2+}$ ) brought on by N addition changed the adsorption rate of C compounds on  
100 mineral surface, which was not conducive for the formation of MAOC, thus affecting  
101 the persistence of SOC (Ye et al., 2018). Therefore, the mechanism of how POC and  
102 MAOC react to N addition has not yet been determined.

103 We carried out a N addition experiment in a subtropical *Castanopsis fabri* forest  
104 to investigate the effects of N addition on POC and MAOC as well as their underlying  
105 mechanisms. The SOC fractions and various parameters, including aboveground litter



106 biomass, fine root biomass, soil enzyme activities, soil physicochemical properties, and  
107 exchangeable cations were examined. Based on previous findings, we proposed two  
108 hypotheses: 1) N addition promotes plant biomass and enhances C input, which may  
109 promote POC accumulation; 2) N addition causes the loss of exchangeable cations (e.g.,  
110  $\text{Ca}^{2+}$ ) through leaching, affecting the bridge between SOC and polyvalent cations,  
111 which may decrease MAOC content.

112

## 113 **2 Materials and methods**

### 114 **2.1 Study site**

115 Daiyun Mountain National Nature Reserve in southern China's has been selected  
116 as an experimental area (25.64°–25.73° N, 118.09°–118.34° E, 1800 m a.s.l.), which  
117 has a typical subtropical oceanic monsoon climate. The reserve's average annual  
118 temperature and rainfall are roughly 17.6 °C and 1700–2000 mm, respectively, and  
119 precipitation mainly occurs during March–September. The dominant tree species are  
120 *Castanopsis fabri*. The understory vegetation includes *Micheliamaudiae*, *Lindera*  
121 *aggregate*, *Schima superba*, and *Ilex pubescens*. The soil type is Ultisol according to  
122 US Department of Agriculture soil order, with 45% sand, 33% silt, and 22% clay.

123

### 124 **2.2 Experimental design**

125 Based on the rate of atmospheric N deposition in the area reported earlier, we  
126 selected the following three treatments: control (CT, +0 kg N ha<sup>-1</sup> yr<sup>-1</sup>), low-N addition  
127 (LN, +40 kg N ha<sup>-1</sup> yr<sup>-1</sup>), and high-N addition (HN, +80 kg N ha<sup>-1</sup> yr<sup>-1</sup>). Each treatment  
128 has four plots or replicates, with each plot measuring 10 m × 10 m and having a 10-m  
129 buffer zone between them. Beginning in early May 2020, urea (CO(NH<sub>2</sub>)<sub>2</sub>) was  
130 consistently sprayed over the low-N addition and high-N addition plots using a  
131 backpack sprayer after being dissolved in deionized water. On the control plots, the  
132 same volume of deionized water was sprayed. We did not observe any marked  
133 differences between basic physicochemical properties of the soils of different plots



134 before N addition. In December 2021, we used the five-point sampling method to  
135 sample soil from each plot. The surface litter was removed, and the topsoil (0–10 cm)  
136 was collected from each plot. Each plot's samples were thoroughly combined, placed  
137 in a low-temperature insulated box, and taken right away to the lab for further  
138 processing. In the lab, the soil was sieved through a 2-mm screen after being freed of  
139 fine gravel, trash, and plant roots using tweezers. The sieved soil was partially stored  
140 in a 4 °C refrigerator. Within one week, soil enzyme activity, microbial biomass,  
141 mineral N were evaluated. The other soil was air-dried and the SOC fractions and soil  
142 pH were examined. To quantify the soil physicochemical characteristics and  
143 exchangeable cations (Table S1), a piece of the air-dried soil was filtered through a  
144 mesh of 0.149 mm.

145

### 146 **2.3 Soil physicochemical characteristics**

147 Soil pH was measured using a glass electrode (soil: water; 1: 2.5) (Starter 300,  
148 USA). After inorganic C was eliminated from the soil samples using 0.5 mol L<sup>-1</sup> HCl,  
149 SOC and total N (TN) contents were measured using a CN analyzer (Elementar Vario  
150 EL III, Germany). According to Vance et al. (1987), the chloroform fumigation-  
151 extraction method was used to measure the amount of microbial biomass carbon (MBC).  
152 Using a continuous flow on auto-analyzer (SKALAR San++, Netherlands), soil mineral  
153 N was extracted in a 2 mol L<sup>-1</sup> KCl solution and quantified.

154

### 155 **2.4 SOC fractions**

156 SOC fractions were investigated using the physical fractionation (wet sieving)  
157 method described by Marriott and Wander (2006). Specifically, we weighed 20.0 g of  
158 dried soil samples into plastic bottles, and added 100 mL of sodium hexametaphosphate  
159 solution (5 g L<sup>-1</sup>) to each sample. The mixture was first shaken for 18 h. Subsequently,  
160 the dispersion was transferred to a 53-µm sieve of a reciprocating shaker (AS200  
161 control, Germany) with an amplitude of 3 cm, and the samples were subjected to 100



162 vibrations. A continuous flow of deionized water was used to flush the sieve to disperse  
163 the SOC within aggregates. Meanwhile, a clean beaker was used to collect the sample  
164 passing through the 53- $\mu\text{m}$  sieve. Soil samples collected on the sieve and in the beaker  
165 were dried at 60 °C until their weight remained stable. The SOC content in each fraction  
166 was evaluated using an elemental analyzer (Elementar Vario EL III, Germany). The  
167 following equations were used to obtain the POC (g kg<sup>-1</sup>) and MAOC (g kg<sup>-1</sup>) contents:

$$168 \quad \text{Mass recovery (\%)} = \frac{\text{Mass}_{\text{POC}} + \text{Mass}_{\text{MAOC}}}{\text{Mass}_{\text{Bulk soil}}} \times 100 \quad [1]$$

$$169 \quad \text{POC (g kg}^{-1}\text{)} = \frac{(\text{Mass}_{\text{POC}} \times \text{OC}_{\text{MAOC}})}{\text{Mass}_{\text{Bulk soil}} \times \text{Mass recovery}/100} \quad [2]$$

$$170 \quad \text{MAOC (g kg}^{-1}\text{)} = \frac{(\text{Mass}_{\text{MAOC}} \times \text{OC}_{\text{MAOC}})}{\text{Mass}_{\text{Bulk soil}} \times \text{Mass recovery}/100} \quad [3]$$

171 where  $\text{Mass}_{\text{Bulk}}$  is the soil mass used for sieving (g),  $\text{Mass}_{\text{POC}}$  and  $\text{Mass}_{\text{MAOC}}$  are the  
172 masses of the POC and MAOC fractions obtained after sieving (g), and  $\text{OC}_{\text{POC}}$  and  
173  $\text{OC}_{\text{MAOC}}$  are the C contents of POC and MAOC (g C kg<sup>-1</sup> fraction), respectively.

174

## 175 2.5 Exchangeable cations and soil enzyme activities

176 Soil exchangeable cations (K<sup>+</sup>, Na<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, and Al<sup>3+</sup>) were extracted using  
177 0.1 mol L<sup>-1</sup> BaCl<sub>2</sub>, and their contents were measured using inductively coupled plasma  
178 emission spectrometry (ICP-OES; USA). Activities of soil enzymes, including  $\beta$ -  
179 glucosidase (BG) and cellulolytic enzyme cellobiohydrolase (CBH), were measured  
180 using the method reported by Saiya-Cork et al. (2002). We added 1.0 g of soil sample  
181 and 125 mL of 50 mmol L<sup>-1</sup> acetate buffer solution to a triangular flask. The mixture  
182 was homogenized for 5 min using a magnetic stirrer, and the supernatant was collected  
183 as the test solution. In a 96-well microplate, the test solutions were applied to different  
184 substrates (4-methylumbelliferyl-D-glucopyranoside and 4-methylumbelliferyl-D-  
185 cellobioside) to measure BG and CBH activity, respectively. A multi-purpose  
186 microplate reader (SpectraMax M5, USA) equipped with 365 nm excitation and 450  
187 nm emission filters was used to measure fluorescence.

188



## 189 2.6 Aboveground litter biomass and fine root biomass

190 In each plot, three litter traps (1 m × 1 m) were installed approximately 1 m above  
191 the ground surface. Around the 20<sup>th</sup> of each month, litter was collected from the  
192 *Castanopsis fabri* forest. Litter from the same plot (with three traps) was collected and  
193 mixed in the laboratory. The collected litter was dried and weighed to determine the  
194 biomass. This process was continuously performed for one year. In December 2021, a  
195 five-point sampling technique was used to collect fresh soil cores from each plot at a  
196 depth of 0–10 cm, using a 3.5 cm-diameter soil auger. The diameter < 2 mm fine roots  
197 in the soil cores were carefully selected and thoroughly washed, then dried at a constant  
198 temperature of 60 °C, and finally weighed.

199

## 200 2.7 Statistical analyses

201 R 4.2.3 (R Core Team, 2023) and SPSS 27.0 (IBM, USA) were used to conduct  
202 statistical analyses. One-way analysis of variance was examined to test the effects of N  
203 addition on SOC fractions, soil physicochemical characteristics, exchangeable cations,  
204 soil enzyme activities, aboveground litter biomass, and fine root biomass. The least  
205 significant difference method was employed to compare treatment differences at a  
206 significance level of  $P < 0.05$ . Principal component analysis was examined using the  
207 “*factoextra*” package in R 4.2.3 to explore the influence of soil factors (exchangeable  
208 cations contents), plant factors (aboveground litter biomass and fine root biomass), and  
209 soil microbial factors (enzyme activities) on SOC fractions. Graphs were created using  
210 Origin 9.1 software (OriginLab, USA).

211

## 212 3 Results

### 213 3.1 Responses of SOC fractions to short-term N addition

214 The POC content of soil samples was significantly affected by short-term N  
215 addition in a content-dependent manner ( $P < 0.01$ ), but N addition did not exert any  
216 significant effect on the MAOC content ( $P > 0.05$ ; Fig. 1a and 1b). Moreover, the





217 MAOC: POC ratio was significantly affected by N addition ( $P < 0.01$ ), declining with  
218 N addition (Fig. 1c).

219

### 220 **3.2 Responses of soil exchangeable cations and enzyme activities to short-term N** 221 **addition**

222 N addition had a significant impact on exchangeable cation content, except for  $\text{Na}^+$   
223 content. Specifically, the  $\text{Al}^{3+}$  content of the low-N addition and high-N addition groups  
224 was significantly higher than that of control group, whereas the  $\text{K}^+$ ,  $\text{Ca}^{2+}$ , and  $\text{Mg}^{2+}$   
225 contents were significantly lower than those of the control group ( $P < 0.01$ ; Fig. 2).  
226 With rising N-addition levels, soil enzyme activities (BG and CBH) reduced ( $P < 0.01$ ;  
227 Fig. 3).

228

### 229 **3.3 Responses of aboveground litter biomass and fine root biomass to short-term** 230 **N addition**

231 N addition significantly affected the aboveground litter biomass ( $P < 0.01$ ). In  
232 comparison to that in the control plots, the aboveground litter biomass in the low-N  
233 addition and high-N addition plots was significantly risen by 17% and 28%,  
234 respectively. In contrast, the fine root biomass was unaffected by the addition of N ( $P$   
235 = 0.33; Fig. 4).

236

### 237 **3.4 Analysis of key factors influencing SOC fractions changes**

238 Principal component analysis revealed a significant positive correlation between  
239 POC and aboveground litter biomass ( $R^2 = 0.48$ ), and significant negative correlations  
240 between POC and soil exchangeable cations ( $\text{Mg}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{K}^+$ ), pH, and soil enzyme  
241 activities (BG, CBH) ( $R^2_{\text{POC-Mg}} = 0.73$ ,  $R^2_{\text{POC-Ca}} = 0.52$ ,  $R^2_{\text{POC-K}} = 0.57$ ,  $R^2_{\text{POC-pH}} = 0.82$ ,  
242  $R^2_{\text{POC-BG}} = 0.88$ ,  $R^2_{\text{POC-CBH}} = 0.91$ ). Moreover, we found that a close negative connection  
243 between  $\text{Al}^{3+}$  and  $\text{Ca}^{2+}$  or  $\text{K}^+$  ( $R^2_{\text{Al-Ca}} = 0.57$ ,  $R^2_{\text{Al-K}} = 0.55$ ; Fig. 5).

244



## 245 **4 Discussion**

### 246 **4.1 Positive response of POC to short-term N addition**

247 Our results demonstrated that short-term low and high N additions increased POC  
248 content ( $P < 0.01$ ; Fig. 1a). This is in line with previous findings (Riggs et al., 2015;  
249 Chen et al., 2018; Chen et al., 2020) and confirms our first hypothesis. The positive  
250 response of POC to N addition can be attributed to the following reasons (Fig. 6). First,  
251 POC is predominantly formed by plant-based compounds (Cotrufo and Lavelle, 2022),  
252 so the rise in POC content following the addition of N could be due to the increase in  
253 the inputs of plant residues to soils. In our study, N addition enhanced aboveground  
254 litter biomass considerably, but not fine root biomass. This result in combination with  
255 the significant correlation between POC and aboveground litter biomass ( $R^2 = 0.48$ , Fig.  
256 5) suggested that N-induced increase in POC was mainly derived from plant  
257 aboveground rather than belowground roots. Some previous studies suggested that POC  
258 pool is mainly affected by plant belowground roots (Zhang et al., 2019; Sokol et al.,  
259 2019; Sun et al., 2023). However, this phenomenon may be not applicable to our study,  
260 as N addition can enhance soil nutrient resources and decrease the proportion of  
261 biomass allocation to belowground and thus the amount of belowground C inputs (Feng  
262 et al., 2023). Second, the increase in POC may be also associated with the lower soil  
263 enzyme activities under N additions, as soil enzymes are important drivers of SOC  
264 decomposition (Liang et al., 2017). We found the significant decrease in soil enzyme  
265 activity in the low-N addition and high-N addition plots ( $P < 0.01$ ; Fig. 3), and a  
266 significant negative relationship between soil enzyme activity and POC ( $R^2_{\text{POC-BG}} =$   
267  $0.88$ ,  $R^2_{\text{POC-CBH}} = 0.91$ ; Fig. 4). This may be due to the leaching of soil exchangeable  
268 cations, which decrease soil pH, thereby inhibiting soil microbial biomass and soil  
269 enzyme activities (Averill and Waring, 2018), and reducing the decomposition and loss  
270 of POC. Overall, the increases in POC content under N addition can be not only related  
271 to N-induced increase in plant-derived C inputs but also related to N-induced decrease  
272 in microbial decomposition activities.



273

#### 274 **4.2 Non-significant response of MAOC to short-term N addition**

275 N addition had a neutral effect on the MAOC content ( $P = 0.27$ ; Fig. 1b), such  
276 result is contrary to our second hypothesis and previous studies (Lu et al., 2021; Rocci  
277 et al., 2021). Chen et al. (2021) showed that microbial turnover to produce microbial  
278 necromass is thought to be a major driver of MAOC, and the undetectable change in  
279 MAOC content might be regulated by maintaining an equilibrium between microbial  
280 necromass input and microbial decomposition output. Here, we attributed the non-  
281 response of MAOC to N addition to the following reasons. First, previous studies have  
282 shown that compared to aboveground litter, belowground roots make a greater  
283 contribution to the persistence of SOC, as they can interact with soil minerals,  
284 microorganisms, and aggregates right away after death (Wu et al., 2018; Whalen et al.,  
285 2021). However, fine root biomass was not influenced by N short-term addition, which  
286 may be a reason for the lack of a marked response of MAOC to N addition in this study.  
287 Second, the effect of N-induced soil acidification on adsorption and desorption of  
288 exchangeable cations, such as  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Fe}^{3+}$ , and  $\text{Al}^{3+}$ , is also likely to be important  
289 factors regulating MAOC changes (Sun et al., 2023; Tang et al., 2023). However, we  
290 identified that few studies have reported the relationship between SOC pools and  
291 exchangeable cations following N addition (Yu et al., 2017; Chen et al., 2020; Sun et  
292 al., 2023). Specifically, Yu et al. (2017) demonstrated that fertilization promoted the  
293 enrichment of aluminium and iron ions on the surface of soil particles, leading to  
294 enhanced soil aggregation. This increased soil aggregation reduced the rate of soil  
295 mineralization, potentially increasing MAOC storage and preservation (Waychunas et  
296 al., 2005). In contrast, Chen et al. (2020) have indicated that N addition had no  
297 discernible effect on soil  $\text{Al}^{3+}$  content but decreased soil  $\text{Ca}^{2+}$  content (Chen et al., 2020;  
298 Sun et al., 2023). The reduction in  $\text{Ca}^{2+}$  content could potentially lead to decreased  
299 mineral sorption from microbial necromass and products, resulting in the depletion of  
300 MAOC content. In this study, a significant rise in soil  $\text{Al}^{3+}$  content and a simultaneous



301 decline in  $Mg^{2+}$ ,  $Ca^{2+}$ , and  $K^+$  contents were observed following the addition of N ( $P <$   
302 0.01; Fig. 2). Principal component analysis revealed a strong negative correlation  
303 between  $Al^{3+}$  and  $Ca^{2+}$  (or  $K^+$ ) ( $R^2_{Al-Ca} = 0.57$ ,  $R^2_{Al-K} = 0.55$ ; Fig. 5). In light of these  
304 results, we suggest a new framework that the negative effects of leaching of  
305 exchangeable cations on SOC bridging and the positive effects of exchangeable cation  
306 dissolution on SOC adsorption may offset each other, leading to an insignificant  
307 response of MAOC to the addition of N in this subtropical forest. Moreover, we  
308 observed a decrease in MBC content under N addition (Table S1). Long-term changes  
309 in microbial biomass may markedly affect MAOC by altering the adsorption of soil  
310 minerals on microbial necromass or litters (Feng et al., 2022).

311

#### 312 **4.3 Effects of short-term N addition on SOC persistence**

313 MAOC with a long turnover rate is critical for maintaining the persistence of SOC  
314 (Kristensen et al., 2022). Although POC constitutes a smaller proportion of soil, its  
315 faster turnover rate is essential for maintaining or improving soil fertility (Benbi et al.,  
316 2011). Compared with MAOC, POC has a lower density, contains more chemically  
317 complicated compounds on average, and has a higher C: N ratio (Cotrufo and Lavelle,  
318 2022). Here, the MAOC: POC ratio was significantly decreased with the addition of N  
319 ( $P < 0.01$ ; Fig. 1c), indicating that N addition reduces the persistence and increases the  
320 nutrient density of SOC (Sokol et al., 2022). Similar findings have been published by  
321 Tang et al. (2023). There was a strong positive relationship between POC and SOC by  
322 principal component analysis ( $R^2 = 0.89$ ; Fig 5), suggesting that the change in SOC with  
323 the addition of N was driven by a change in POC rather than MAOC. However, Lugato  
324 et al. (2021) reported that POC in forest soils is susceptible to depletion under global  
325 change. They recommended the implementation of coniferous forest management  
326 strategies that are aimed at increasing plant inputs into the soil to counteract POC losses.  
327 Overall, it is likely that the accumulation in POC under N addition will offset the loss  
328 of POC with global change. A positive relationship between SOC loss and SOC



329 persistence was shown in Liang et al. (2023), suggesting that efforts should be directed  
330 towards improving SOC persistence. Thus, we propose enhancing SOC persistence by  
331 regulating soil exchangeable cations to facilitate mineral sorption of microbial  
332 necromass and its products.

333

### 334 **5 Conclusion**

335 In this study, we elucidated that MAOC was less vulnerable to short-term N  
336 addition than POC in a subtropical forest. The reason for the accumulation of POC  
337 caused by N addition could be the combination of increased plant litter input and  
338 decreased microbial decomposition. By contrast, short-term N addition did not exhibit  
339 marked effect on MAOC. Such results could be related to cancellation of effects by the  
340 lack of marked changes in fine root biomass and the adsorption and desorption of  
341 organic molecules on the mineral surface. The marked decrease in MAOC: POC ratio  
342 under N addition indicated that SOC persistence and nutrient density decreased. Overall,  
343 our study links the responses of POC and MAOC to the plant–microbial–soil response,  
344 providing insights into the mechanisms of SOC dynamics under ongoing N deposition  
345 in subtropical forest ecosystems.

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347

### 348 **Data availability statement**

349 The data that supports the findings of this study are available in the supplementary  
350 material of this article.

351

352

### 353 **Authors' contributions**

354 Qiufang Zhang, Yuehmin Chen and Jingqi Chen conceived and designed the study; Hui  
355 Dai, Quanxin Zeng, Xueqi Sun, and Wenwei Chen collected the samples; Jingqi Chen  
356 and Yuanzhen Peng analyzed the samples/data and wrote the manuscript with help from  
357 Qiufang Zhang; Qiufang Zhang, Jiguang Feng, Biao Zhu, and Yuehmin Chen revised



358 the manuscript; all authors approved the final manuscript.

359

360

361 **Competing interests**

362 The authors declare that they have no conflict of interest.

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553 **Figure captions**

554

555 **Fig. 1** Effects of N addition on the contents of particulate organic carbon (POC, **a**) and  
556 mineral-associated organic carbon (MAOC, **b**) and MAOC: POC ratio (**c**). Boxes and  
557 horizontal lines represent the quartile range and median, respectively, with lines  
558 extending 1.5 times above and below the quartile range ( $n = 4$ ).  $P$  represents the N  
559 addition effect. Significant differences between various N addition treatments are  
560 shown by different letters above the boxes ( $P < 0.05$ ). CT, LN and HN represent control,  
561 low-nitrogen and high-nitrogen, respectively, and the corresponding N addition rates  
562 are 0, 40 and 80 kg N ha<sup>-1</sup> yr<sup>-1</sup>.

563

564

565 **Fig. 2** Effects of N addition on the content of exchangeable cations (K<sup>+</sup>, **a**; Na<sup>+</sup>, **b**;  
566 Ca<sup>2+</sup>, **c**; Mg<sup>2+</sup>, **d**; Al<sup>3+</sup>, **e**) in soil samples. Boxes and horizontal lines represent the  
567 quartile range and median, respectively, with lines extending 1.5 times above and below  
568 the quartile range ( $n = 4$ ).  $P$  represents the N addition effect. Significant differences  
569 between various N addition treatments are shown by different letters above the boxes  
570 ( $P < 0.05$ ). CT, LN and HN represent control, low-nitrogen and high-nitrogen,  
571 respectively, and the corresponding N addition rates are 0, 40 and 80 kg N ha<sup>-1</sup> yr<sup>-1</sup>.

572

573

574 **Fig. 3** Effects of N addition on the activities of β-glucosidase (BG, **a**) and 1, 4-β-  
575 cellobiohydrolase (CBH, **b**). Boxes and center lines represent the quartile range and  
576 median, respectively, with lines extending 1.5 times above and below the quartile range  
577 ( $n = 4$ ).  $P$  represents the N addition effect. Significant differences between various N  
578 addition treatments are shown by different letters above the boxes ( $P < 0.05$ ). CT, LN  
579 and HN represent control, low-nitrogen and high-nitrogen, respectively, and the  
580 corresponding N addition rates are 0, 40 and 80 kg N ha<sup>-1</sup> yr<sup>-1</sup>.

581

582

583 **Fig. 4** Effects of N addition on aboveground litter biomass (**a**) and fine root  
584 biomass(**b**). Boxes and center lines represent the quartile range and median,  
585 respectively, with lines extending 1.5 times above and below the quartile range ( $n = 4$ ).



586 *P* represents the N addition effect. Significant differences between various N addition  
587 treatments are shown by different letters above the boxes ( $P < 0.05$ ). CT, LN and HN  
588 represent control, low-nitrogen and high-nitrogen, respectively, and the corresponding  
589 N addition rates are 0, 40 and 80 kg N ha<sup>-1</sup> yr<sup>-1</sup>.

590

591

592 **Fig. 5** Principal component analysis of soil organic carbon fractions and other soil  
593 parameters. POC: particulate organic carbon; MAOC: mineral-associated organic  
594 carbon; SOC: soil organic carbon; MN: mineral nitrogen; Litter: aboveground litter  
595 biomass; Root: fine root biomass; BG: β-glucosidase; CBH: 1, 4-β-cellobiohydrolase.

596

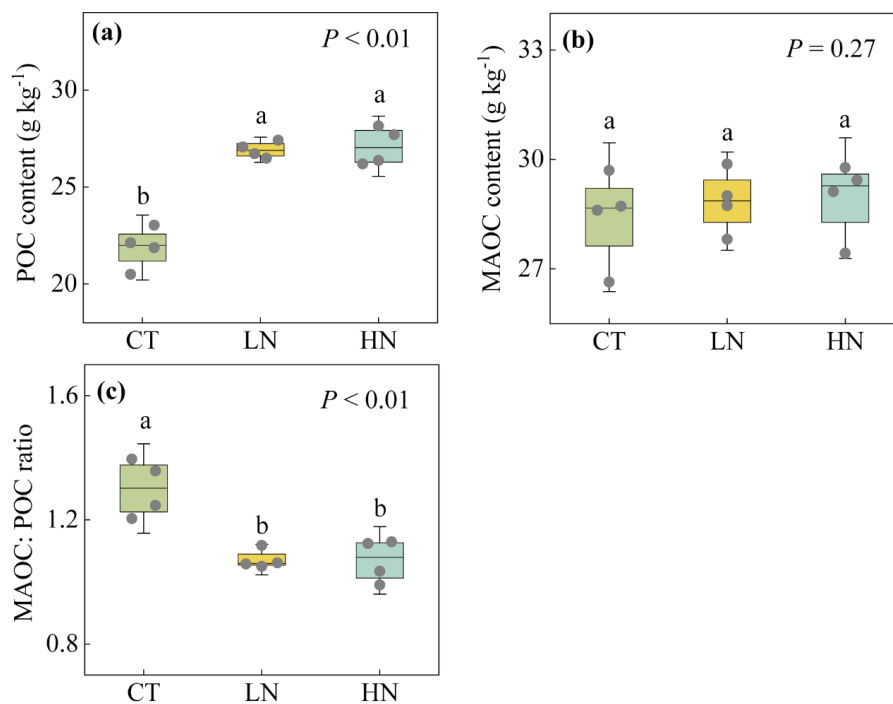
597

598 **Fig. 6** Conceptual diagram representing how the soil organic carbon (SOC) fractions  
599 in the subtropical *Castanopsis fabri* forest react to the addition of N. Notably, N  
600 addition increases particulate organic carbon (POC) content but exerts no marked  
601 influence on mineral-associated organic carbon (MAOC) content; however, it reduces  
602 SOC persistence. The white up, down, and horizontal arrows represent the positive,  
603 negative, and neutral effects of N addition, respectively.

604 Note: ① N addition increases aboveground litter input, which enhances POC content.

605 ② N addition leads to soil acidification and inhibits soil enzyme activities. This  
606 function potentially reduces POC decomposition, ultimately increasing POC content.

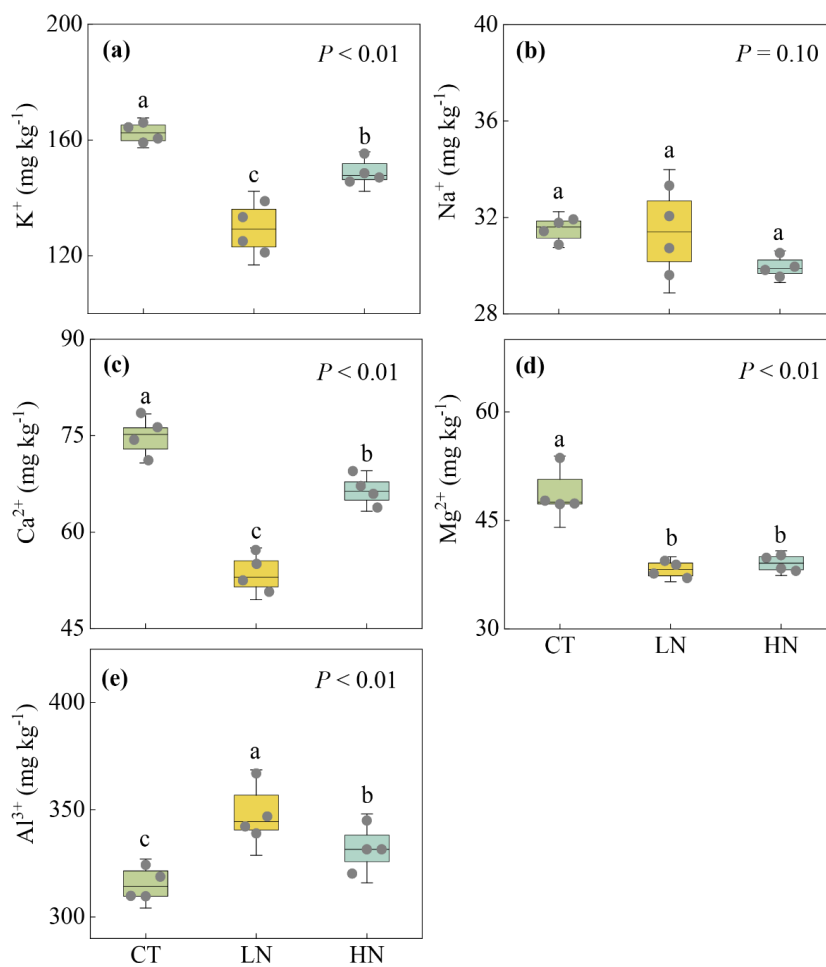
607 ③ Ca<sup>2+</sup> and K<sup>+</sup> leaching negatively affect SOC bridging, whereas Al<sup>3+</sup> dissolution  
608 positively influences SOC adsorption. These opposing effects may offset each other,  
609 resulting in a scenario in which MAOC content experiences no marked changes upon  
610 N addition. ④ Fine root biomass remains largely unaffected by N addition,  
611 contributing to the protection of soil aggregates and, therefore, to the preservation of  
612 MAOC content.



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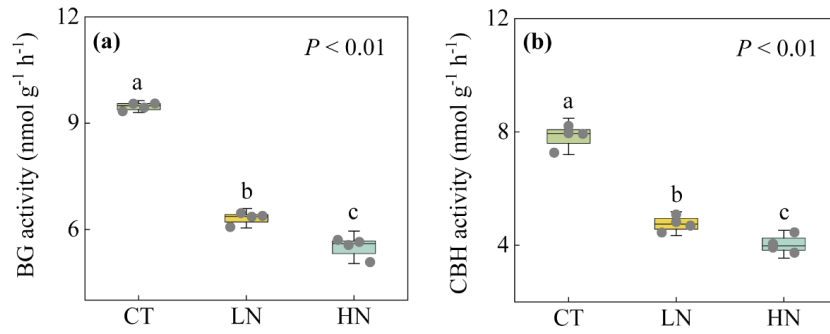
614 **Fig. 1**

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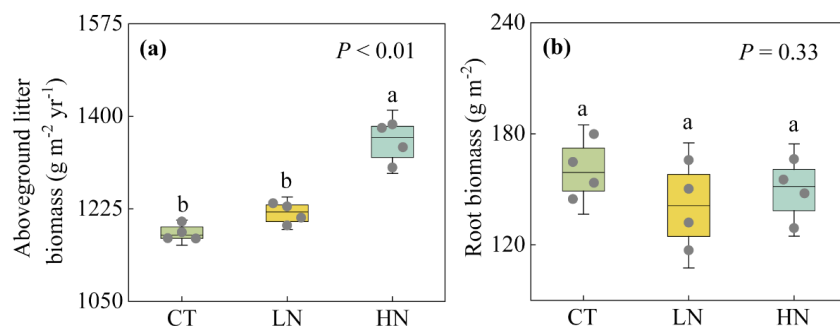
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617 **Fig. 2**  
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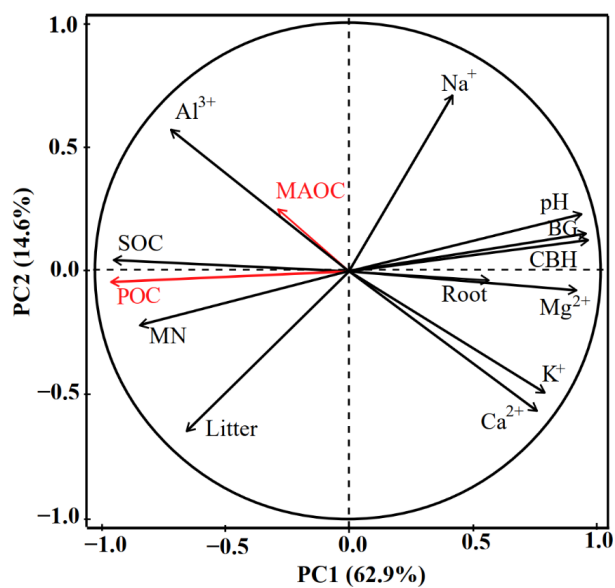
619  
620 **Fig. 3**

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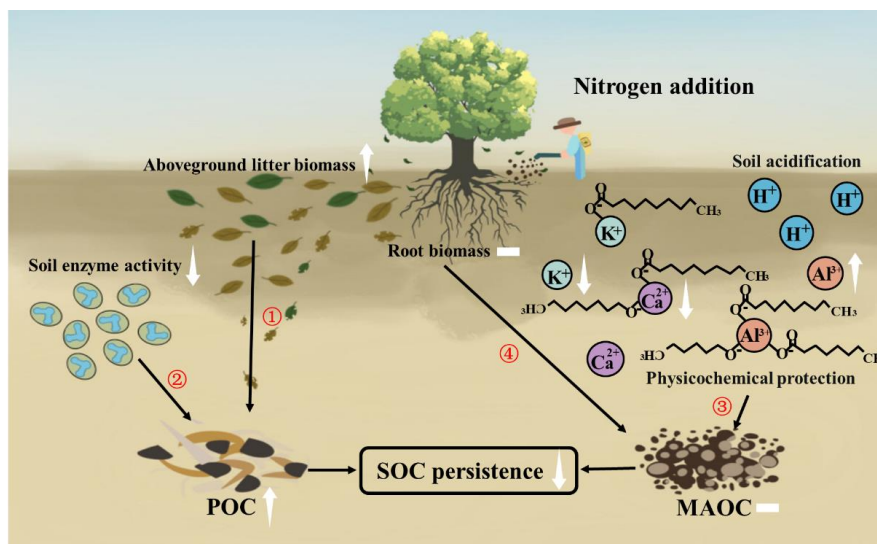


622  
623 **Fig. 4**

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625  
626 Fig. 5  
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628

629 **Fig. 6**

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