

Main Comments:

RW1: The one major nagging question that I had throughout the entire review was that the authors have not accounted for, or even discussed anywhere, the potential limitations of using absorbance alone to partition colored versus non-colored DOC concentrations. Specifically, can shifts in absorbance in response to salinization induce optical changes in the DOM pool that alter absorbance through concentration-independent processes? Salt additions that alter ion binding patterns in the soil matrix could liberate/precipitate stuff like iron that may impact absorbance, no? It could also alter the chelation of DOM and metals and other materials, altering optical conditions? I am not raising these potential mechanisms to say that they invalidate the results. It is likely that at these DOC concentrations, such effects would be less important. BUT, using knowledge from the literature, and any other information on hand, can the authors speak to what role, if any, these effects have on their interpretations and conclusions (even if only to caveat them)?

Author: The reviewer raises two good points, one that solution salinity affects the optical properties of CDOM and secondly that salinity affects the binding to the soil matrix of other components, like iron, that may also affect absorbance properties. The first effect can be seen from the results in Table 6 which show slight decreases in CDOM slopes and slope ratios within each sample category as salinity is increased. Salinity increases are also linked to increased specific- absorbance at long wavelengths (Gao et al. 2015), but this effect contrasts with the observed trend in our post-incubation data of decreased specific absorbance (increased DOC slope, Table 1). As far as possible metal:DOC interactions in the GDS solution, the DOM was sampled from a freshwater peat marsh with very low soil iron – which is why DOM is high to begin with. Moreover, all absorbance measurements were made under anaerobic conditions. Thus dissolved iron, to the extent it was present, would mainly be in non-absorbing Fe^{2+} form not the UV absorbing Fe^{3+} form. We now include new text describing the absorbance methods (this text was inadvertently left out of our first submission) which notes that scans were done under anaerobic conditions:

“Due to the highly colored nature of the Great Dismal Swamp DOC, we diluted subsamples of pre-incubation standards by a factor of 10 using dilutant from the same salinity treatment while we diluted the filtered post-incubation supernatant by the same factor using ultrapure water + NaCl (Sigma Aldrich, 99.5% purity) to match sample salinity. All solutions were degassed with N_2 , then dilutions were performed, cuvettes loaded and sealed in an anaerobic chamber. We performed absorbance scans at 2 nm intervals (270 – 750 nm) for all replicates using a Thermo Scientific Evolution 220 UV-Vis spectrophotometer.”

In addition, we will add comments on the effects of salinity and iron as well as pH change (a request from another reviewer) on our absorbance results to the discussion (following the paragraph discussing how isotherm parameters varied with salinity):

“We have considered possible physicochemical effects on DOM absorbance in our incubations, but these do not appear to substantially affect our interpretation of CDOC dynamics. Increasing

salinity, and thus ionic-strength, increases CDOM absorbance, with proportionally greater increases at longer wavelengths (Gao et al., 2015). This is attributed to deprotonation of DOM, and results in a lowering of the spectral slope of CDOM absorption spectra and increase of specific absorbance, e.g. at 355 nm (Gao et al 2015). While there was some increase in spectral slope and specific-absorbance with increasing treatment salinity for the pre-incubation standards (see Tables 1 and 6), we observed the opposite effect in the post-incubation solutions, i.e. adding salinity increased DOC vs absorbance slope (Table 1) which implies decreased specific absorbance. Increasing pH also increases DOM absorbance (Gao et al., 2015), and postincubation pH increased in all our incubations from 4.6 ± 0.09 in the standards to 6.74 ± 0.03 , 6.64 ± 0.04 , 5.15 ± 0.05 and 6.94 ± 0.10 for Kirkpatrick, Taskinas, Jug Bay and Wachapreague soils (respectively) (Pinsonneault et al., 2021). The increase in pH did not have an important effect on postincubation absorbance since specific absorbance decreased during the incubation, the opposite direction from what would be expected from the pH change alone.

Ferric (Fe^{3+}) iron also absorbs in the UV and its concentration in some surface waters is high enough to significantly bias estimates of CDOM absorbance (Poulin et al., 2014; Logozzo et al., 2022). However, Fe is not a significant factor for DOM from the Great Dismal Swamp since it is a peat wetland with very low iron (USFWS, 2006). Release of iron into solution during the incubation seems unlikely since it would have increased specific absorbance, again, the opposite of what we observed. Finally, we can exclude interference from dissolved iron on our absorbance measurements since all solutions were anaerobic and dissolved iron would be in the non-absorbing Fe^{2+} state.”

Specific Comments:

RW1: L23- GDS DOC – mention what this is at the start of abst. It’s introduced in a strange spot.

Author: The source of the test solution will be moved to the first sentence describing our experiments:

“To test this hypothesis, we generated initial mass sorption isotherms of CDOC and noncolored dissolved organic carbon (NCDOC) using anaerobic batch incubations of Great Dismal Swamp DOC with four tidal wetland soils, representing a range of organic carbon content (1.77 ± 0.12 % to 36.2 ± 2.2 %) and across four salinity treatments (0, 10, 20, and 35).”

RW1: L33 - Intro – Polydisp. – define what this is in brackets.

Author: Definition will be added to the sentence (underlined):

“Studies have found that CDOM composition in flooding and ebbing tidal waters differs with more strongly colored and aromatic CDOM of higher average molecular weight and polydispersity [breadth of the molecular weight distribution] being exported from tidal marshes into estuarine waters during ebbing tide relative to that imported into marshes during flooding tide (Tzortziou et al., 2008; Tzortziou et al., 2011).”

RW1: L58 – Run on sentence.

Author: Not clear to which sentence the reviewer is referring. We have checked the text in this part of the mss to ensure that we don't have a run on sentence.

RW1: L96 – What is a “filter cale”? Can you reword with something more generic? I have never seen the word “cale” before like this.

Author: This was a typo, will be corrected to “filter capsule”

RW1: L106- Is the NaN₃ a preservative? Explain purpose briefly.

Author: Description will be added to sentence (underlined):

“The resulting filtered DOC concentrate was then treated with 1 mM sodium azide (NaN₃), a microbial inhibitor, and DOC concentration ([DOC]) was measured with a Shimadzu TOC-L using high-temperature combustion.”

RW1: L123- Any details about this regression R²/strength criteria used? Were relationships scrutinized in any way?

Author: Regression r² values and parameter standard errors are presented in Table 2, the r² values were all very near 1.

RW1: L163- Fig 1 caption. Explain blue solid/dashed arrows so readers don't need to sift through text to interpret figure.

Author: Arrows will be identified in the caption:

“Figure 1. Scatterplot of DOC content vs absorption coefficient at 355 nm in pre-incubation (triangles) and post-incubation (+ symbols) treatments in triplicate from the Kirkpatrick soil, S=10, isotherm experiment with fitted linear regressions. The dashed arrows indicate the difference in DOC between the regression lines for selected post-incubations absorbances, the solid arrows identify the DOC of the pre-incubation line at these absorbances.”

RW1: L204 – Fig 3. I spent a bunch of time trying to figure this out, then realized you are using negative values to mean desorption. Please state this up front in the caption. Even better would be to add a dashed zero line to signify the difference.

Author: The figures did have a zero gray line, this will be made into a dashed line that will be more visible. Also, we will note the meaning of negative values in the figure caption:

“Figure 3. Isotherms of adsorption-desorption of CDOC (filled shapes) and NCDOC (open shapes) for tidal marsh soils. Net adsorption of organic carbon based on the difference between initial and final concentrations of soil organic carbon (ΔC , mg C g-soil⁻¹) as a function of initial concentration of CDOC ($[CDOC]_i$, mg L⁻¹). Negative values indicate net desorption. Error bars show standard deviation of triplicate incubations, in most cases error bars are smaller than the symbol. Incubations used soils from (a) Kirkpatrick, (b) Taskinas, (c) Jug Bay, and (d) Wachapreague marshes. Red circles, green triangles, blue squares, and purple diamonds denote salinities of 0, 10, 20, and 35, respectively.”

RW1: L215- All of this isotherm theory and mathematical model interpretation would be far easier to follow as a section fully developed in the methods, perhaps with a conceptual diagram of the isotherm graph.

Author: The text describing the linear desorption isotherm will be moved to the methods. Figure 4 was included to assist the reader in visualizing the context for the different isotherm models (see comment below).

RW1: L240- Throughout results, the ability for readers to wade through this info would be improved with the use of sub-headings to group info by themes.

Author: We will add additional sub-headings in the results, 3.2 CDOC Sorption Isotherms, 3.3 NCDOC Desorption Isotherms and 3.4 Relationship of Isotherm Parameters to Soil Characteristics

RW1: L251- Figure 4- Confusing. Is the y axis still initial minus final? If so, make that one clear.

Author: This is stated clearly in the figure caption.

“Net adsorption of organic carbon, based on the difference between initial and final concentrations of soil carbon (ΔC , mg C g-soil⁻¹), is plotted vs solution TDOC (for Total) or CDOC (Colored and Noncolored).”

To further emphasize the occurrence of net desorption for some initial concentrations of CDOC a dashed line will be added at zero.

RW1: L273- Again, a bunch of this background definition content would be nice to see in a dedicated methods section up front.

Author: The definition of null point and description of its meaning will be moved to methods:

“The null point (NP) is defined as the concentration at which there is no net removal of CDOC from solution (adsorption) or release from soil (desorption), the biogeochemical significance of which is a sorptive equilibrium between the soil mineral surfaces and the aqueous phase. It is derived by setting Eq 2 to zero and solving for NP concentration of $[CDOC]_i$:

$$NP = \frac{C_{CO}}{(Q_{Cmax} - C_{CO}) * K_c} \quad (3)$$

RW1: L281- Initial sentence not needed.

Author: We agree that it was not needed for this paragraph. The purpose was to provide a segue to the spectral results, which is better accomplished by having a rephrased version as the topic sentence of the following paragraph:

“Given that spectral characteristics of CDOM absorbance and fluorescence are used to infer sources, sinks, reactivity, and other biogeochemical processes (Chin et al., 2002; Helms et al., 2008; Tzortziou et al., 2008), we examined how spectral properties of CDOC were affected by sorption processes.”

RW1: L293- “than less” – grammar.

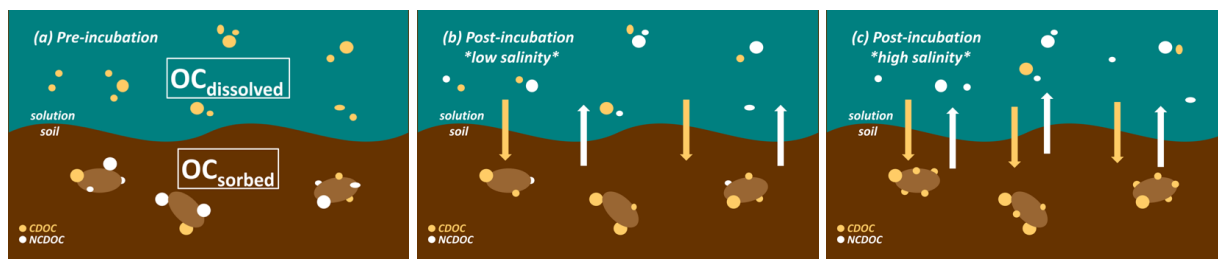
Author: Changed to “and less”

RW1: L330- Do the authors find this odd that these components decrease? NCDOC increases, so should these components not also go up? I am not saying that they must, I am likely missing something here, but just want the authors to explain this to me/readers.

Author: It is not clear to which component decrease the reviewer is referring. Line 333 speaks of the decrease in the phenylalanine component, C4. As something prevalent in the GDS, it is regarded as a component of the CDOC, not the NCDOC, so there is no inconsistency with the increase in NCDOC.

RW1: L345- This figure feels a bit unnecessary/of limited use in current format. I’d swap out the text “dismalswamp” with something generic so that someone outside the region will understand is the stock solution without knowing the location it came from, and not having to consult the methods again. The grey/brown DOC could be defined in a legend so it is visually obvious what the color coding means. The figure could be redone to reflect what increasing salinity does to DOC sorption, which is a key question as I take it in the paper.

Author: Based on the reviewer’s helpful comments the figure will be revised:



The different phases are now described in general terms and the color code for CDOC and NCDOC is explicit in the diagram. Separate panels will be shown to depict the increase in net adsorption of CDOC and desorption of NCDOC at high salinity.

RW1: L362- Ref format in brackets, run on sentence as written.

Author: The text will be broken up into two sentences:

“However, this was a minor process under our experimental conditions because incubations had minimal effects on the slopes and slope ratios of absorbance spectra. Moreover, such differential adsorption would be expected to cause differential changes in the specific absorbance of CDOC (an indicator of average molecular weight) as a function of CDOC (cf. adsorption of fulvic acid on goethite Zhou et al., 2001) whereas we observed uniform specific absorbance over all incubation conditions (Fig. 1).”

RW1: L364- Why redefining GDS here?

Author: Abbreviation redefinition deleted.

RW1: L393- Photodegradation reference needed here.

Author: Citation added to Dainard et al (2015).

RW1: L420- If the conceptual figure could link to this sentence it would be more useful. See my comment above about incorporating salinity.

Author: The figure is now modified to illustrate our interpretation of the salinity effect

RW1: L437- You come back to flood/ebb differences in DOM here. A few more sentences integrating the mechanisms exposed by the experiment, and how they potentially help to explain the empirical observations in the literature, would strengthen the discussion.

Author: We will add new text placing sorption processes in the context of flood/ebb differences:

“Porewater draining from tidal creek banks at low-tide is a primary source of CDOM in the ebbing tide (Menendez et al., 2022). Our results are consistent with this CDOM being derived by exchange with marsh sediment, as low DOC flood waters infiltrate into the sediment and desorb CDOC from the sediment. Our results also suggest that sediments are a significant source of NCDOC which, for example, comprises ~50% of the DOC in Kirkpatrick Marsh tidal creeks at low tide”

References:

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Menendez, A., Tzortziou, M., Neale, P., Megonigal, P., Powers, L., Schmitt-Kopplin, P., and Gonsior, M.: Strong Dynamics in Tidal Marsh DOC Export in Response to Natural Cycles and Episodic Events From Continuous Monitoring, *J. Geophys. Res.-Biogeo.*, 127, 10.1029/2022jg006863, 2022.

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