

Marine Carbohydrates in Arctic Aerosol Particles and Fog – Diversity of Oceanic Sources and Atmospheric Transformations

Sebastian Zeppenfeld¹, Manuela van Pinxteren¹, Markus Hartmann², Moritz Zeising³, Astrid Bracher^{3,4}, and Hartmut Herrmann¹

¹ Atmospheric Chemistry Department (ACD), Leibniz-Institute for Tropospheric Research (TROPOS), Leipzig, Germany

² Atmospheric Microphysics (AMP), Leibniz-Institute for Tropospheric Research (TROPOS), Leipzig, Germany

³ Alfred-Wegener-Institute Helmholtz Centre for Polar and Marine Research, Bremerhaven, Germany

⁴ Institute of Environmental Physics, University of Bremen, Bremen, Germany

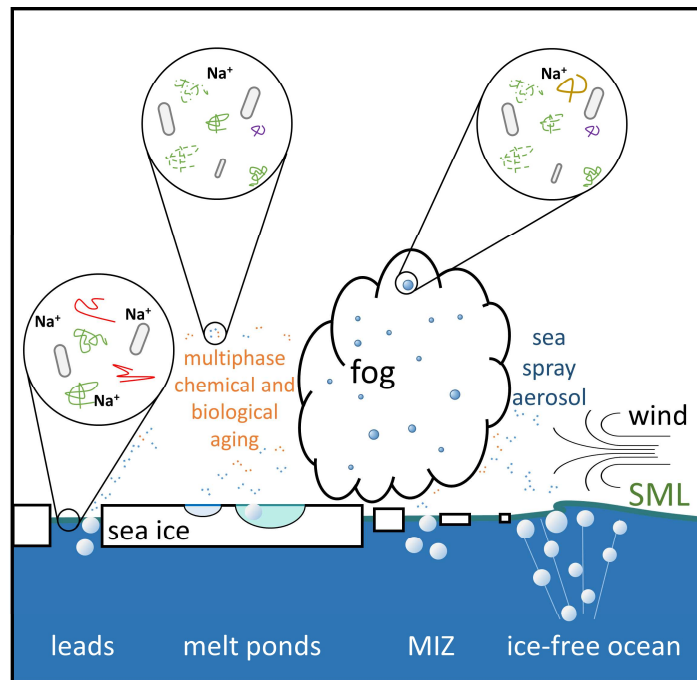
*Correspondence to: Hartmut Herrmann (herrmann@tropos.de)

Submitted to Atmospheric Chemistry and Physics 2023

Abstract

Carbohydrates, produced and released by marine microorganisms in the ocean, enter the atmosphere as part of sea spray aerosol (SSA) and can influence fog and cloud microphysics by acting as cloud condensation nuclei (CCN) or ice nucleating particles (INP). Particularly in the remote Arctic region, significant knowledge gaps persist about the sources, the sea-to-air transfer mechanisms, atmospheric concentrations, and processing of this substantial organic group. In this study, we present the results of a ship-based field study conducted from May to July 2017 in the Fram Strait, Barents Sea and central Arctic Ocean about the sea-air transfer of marine combined carbohydrates (CCHO) from concerted measurements of the bulk seawater, the sea surface microlayer (SML), aerosol particles and fog. In seawater, CCHO ranged between 22–1070 $\mu\text{g L}^{-1}$ with large differences among the different sea-ice-related sea surface compartments: ice-free ocean, marginal ice zone (MIZ), open leads/polynyas within the pack ice and melt ponds. Enrichment factors in the SML (EF_{SML}) relative to the bulk water were very variable in the dissolved (0.4–16) and particulate (0.4–49) phases with highest values in the MIZ and aged melt ponds. In the atmosphere, CCHO appeared in super- and submicron aerosol particles ($\text{CCHO}_{\text{aer,super}}$: 0.07–2.1 ng m^{-3} ; $\text{CCHO}_{\text{aer,sub}}$: 0.26–4.4 ng m^{-3}) and fog water ($\text{CCHO}_{\text{fog,liquid}}$: 18–22000 $\mu\text{g L}^{-1}$; $\text{CCHO}_{\text{fog,atmos}}$: 3–4300 ng m^{-3}). The enrichment factors for the sea-air transfer were calculated for super- and submicron aerosol particles and fog, however strongly varied depending on which of the sea-ice-related sea surface compartments were assumed as the oceanic emission source. Finally, we observed a quick atmospheric aging of CCHO after their emission with indications for both

37 biological/enzymatic processes (based on very selective changes within the monosaccharide
38 compositions of CCHO) and abiotic degradation (based on the depolymerization of long-chained CCHO
39 to short free monosaccharides). All in all, the present study highlights the diversity of marine emission
40 sources in the Arctic Ocean and atmospheric processes influencing the chemical composition of
41 aerosol particles and fog.



TOC Figure

42

43 1. Introduction

44 Sea spray aerosol (SSA) represents one of the major aerosol species in the lower troposphere over the
45 remote Arctic Ocean, particularly during the spring and summer months in the Northern Hemisphere
46 (Chi et al., 2015; Hara et al., 2003; Kirpes et al., 2018; May et al., 2016). Depending on the size
47 distribution and chemical composition, SSA particles strongly contribute to the populations of cloud
48 condensation nuclei (CCN) and ice nucleating particles (INP) affecting the polar radiative budget
49 through the formation of liquid droplets and ice crystals in fog and clouds (DeMott et al., 2016; Lawler
50 et al., 2021; McCluskey et al., 2018; Penner et al., 2001; Schiffer et al., 2018; Wilbourn et al., 2020).
51 Notably in the Arctic, one of the regions most affected by global warming, there is still a lack of
52 knowledge about the relationship between the formation and evolution of clouds and specific
53 chemical properties of SSA particles (Wendisch et al., 2023).

54 SSA is emitted directly from the ocean surface through wind-driven processes and, as a consequence,
55 contains the salts and the organic matter (OM) present in seawater, including carbohydrates (CHO) as
56 one of the largest identified organic fractions (Quinn et al., 2015 and references therein). In
57 microalgae, bacteria and also more complex marine organisms (e.g. kelp, krill), carbohydrates have
58 important metabolic, structural and protective functions or are released in response to environmental
59 stress, such as freezing or lack of nutrients (Krembs et al., 2002; Krembs and Deming, 2008; McCarthy
60 et al., 1996; Mühlenbruch et al., 2018; Suzuki and Suzuki, 2013; Wietz et al., 2015). In seawater, most
61 carbohydrates appear as linear or branched oligo- and polysaccharides, commonly referred to as
62 combined carbohydrates (CCHO). They can be found in both dissolved (*d*CCHO) and particulate
63 (*p*CCHO) phases, distinguished operationally by a 0.2 μm filtration. These macromolecules consist of
64 several monosaccharides, such as hexoses, pentoses, deoxy sugars, amino sugars, uronic acids and
65 amino sugar acids, which are connected via glycosidic bonds (Benner and Kaiser, 2003; Engel and
66 Händel, 2011; Panagiotopoulos and Sempéré, 2005). Most CCHO are quite stable within the marine
67 environment unless they are either hydrolyzed in the presence of specific enzymes or in a very acidic
68 setting (Arnosti, 2000; Panagiotopoulos and Sempéré, 2005). Heterotrophic bacteria use extracellular
69 enzymes to selectively degrade CCHO into absorbable shorter molecules leaving a certain part as
70 recalcitrant, more persistent OM (Alderkamp et al., 2007; Becker et al., 2020; Goldberg et al., 2011;
71 Wietz et al., 2015). While *p*CCHO is mostly attributed to recent productions by local phytoplankton
72 indicated by high positive correlations with total chlorophyll *a* (TChl-*a*), *d*CCHO appears to be the result
73 of more complex metabolic and transformation processes after its release (Becker et al., 2020; Fabiano
74 et al., 1993; Goldberg et al., 2011; Zeppenfeld et al., 2021a). In contrast, dissolved free carbohydrates
75 (*d*FCHO), short sugars in their monomer form, are quickly consumed by marine microorganisms

76 resulting in much lower concentrations of *d*FCHO compared to CCHO in ambient seawater (Engbrodt,
77 2001; Engel and Händel, 2011; Ittekkot et al., 1981; Zeppenfeld et al., 2020).

78 In the remote marine atmosphere, carbohydrates are suggested to significantly impact cloud
79 properties by contributing to both the CCN and INP populations (Alpert et al. 2022; Leck et al., 2013;
80 Orellana et al., 2011; van Pinxteren et al., 2022). Carbohydrates appear both in super- and submicron
81 SSA particles (Aller et al., 2017; Leck et al., 2013; Russell et al., 2010; Zeppenfeld et al., 2021a), most
82 likely resulting from their emission from the surface of the ocean after bubble bursting as part of jet
83 and film droplets (Veron, 2015; Wang et al., 2017). In addition to the bulk surface seawater, the sea
84 surface microlayer (SML) as the uppermost layer of the oceanic water column is an important source
85 of OM, and thus marine carbohydrates, in the SSA. The SML is described as a gelatinous film on top of
86 the ocean, which is often enriched in surface-active substances or buoyant gel particles compared to
87 the underlying bulk water (Engel et al., 2017; Wurl et al., 2009, 2011; Zäncker et al., 2017). Entrained
88 air bubbles rise within the upper part of the water column collecting surface-active organics on the
89 bubble surfaces from the bulk seawater (Burrows et al., 2014). Eventually they pass the thin SML and
90 burst there releasing film and jet droplets containing a mixture of substances found within the bulk
91 water and the SML (Burrows et al., 2014). At the same time, surfactants, exopolymers and microgels
92 in the SML increase the stability of the cap films of the bubbles, extend their lifetimes and enable the
93 drainage of water-soluble compounds (Bigg and Leck, 2008; Bikerman, 2013; Sellegri et al., 2006).
94 Consequently, the sea-air transfer occurs in a chemo-selective manner leading to a strong size-
95 dependent enrichment of surface-active organics relative to water-soluble sodium (Na⁺) and, hence, a
96 relative chemical composition of SSA different to the surface seawater (Facchini et al., 2008; O'Dowd
97 et al., 2004; van Pinxteren et al., 2017; Prather et al., 2013; Quinn et al., 2015; Triesch et al., 2021a, b).
98 These chemo-selective enrichments of organic substances in the SSA relative to bulk water, especially
99 in the submicron size range, usually exceed the enrichments in the SML by orders of magnitude (van
100 Pinxteren et al., 2017; Schmitt-Kopplin et al., 2012). The underlying mechanisms for the chemo-
101 selective sea-air transfer of carbohydrates, including co-adsorption, are complex and subject of several
102 recent and ongoing laboratory tank and modelling studies (Burrows et al., 2016; Hasenecz et al., 2020;
103 Schill et al., 2018; Xu et al., 2023). After their emission, fresh SSA particles, including the contained
104 carbohydrates, undergo atmospheric aging due to a not yet well-understood interplay of several
105 atmospheric processes, such as atmospheric acidification, abiotic radical chemistry and biological and
106 enzymatic modifications (Angle et al., 2021; Hasenecz et al., 2020; Malfatti et al., 2019; Trueblood et
107 al., 2019; Zeppenfeld et al., 2021a), potentially also altering their microphysical properties.

108 Besides SSA, high concentrations of marine carbohydrates in fog and low-level clouds in the marine
109 environment are plausible due to the high hygroscopicity of SSA serving as good CCN (Xu et al., 2022)

110 transferring OM from the particle into the liquid phase, the high water-solubility of carbohydrates, and
111 cloud-borne microorganisms potentially forming carbohydrates in-situ (Matulová et al., 2014). Only a
112 few studies conducted at field sites exposed to marine air masses measured certain subgroups of
113 carbohydrates, such as primary saccharides (Dominutti et al., 2022) or transparent exopolymer
114 particles (TEP) (Orellana et al., 2011; van Pinxteren et al., 2022) so far in fog/clouds. However, the
115 sources of marine carbohydrates in marine ambient fog/clouds, including $dFCHO_{\text{fog}}$ and $CCHO_{\text{fog}}$, and
116 their relationship to the bulk seawater, SML and aerosol particles still lack elucidation.

117 During the summer months, the chemical compounds of natural SSA and marine fog can be studied in
118 the Arctic Ocean due to the low influence of long-range transported anthropogenic pollution (Bozem
119 et al., 2019; Schmale et al., 2021). However, the presence and seasonal evolution of Arctic sea ice
120 divides this pristine region into a complex ensemble of several sea-ice-related sea surface
121 compartments. These encompass the open leads - sea ice fractures with variable widths ranging from
122 several to hundreds of meters - and polynyas, which are larger, more persistent areas of open water
123 within the pack ice. Furthermore, there is the ice-free ocean, the marginal ice zone (MIZ) defined by a
124 sea ice concentration threshold between 15 and 80% (Rolph et al., 2020), and melt ponds forming and
125 developing during the melting season on top of the ice floes. These environments are characterized by
126 different chemical, physical and biological characteristics potentially influencing the quantity and
127 properties of the SSA emitted. Recent studies observed, for instance, that the number and efficiency
128 of Arctic INP are strongly dominated by the type of sea-ice-related sea surface compartments that the
129 air masses had passed before sampling (Creamean et al., 2022; Hartmann et al., 2021;
130 Papakonstantinou-Presvelou et al., 2022; Porter et al., 2022). However, the individual conclusions still
131 appear controversial and might be biased by seasonal and interannual variabilities. Consequently,
132 more systematic studies in the Arctic, also with regard to the chemical properties of the aerosol
133 particles, are required to achieve more conclusive results.

134 To increase the knowledge about marine carbohydrates as important constituents of SSA and potential
135 CCN and INP, we present here the results of a comprehensive field study conducted onboard the
136 German icebreaker RV *Polarstern* from May to July 2017. We performed concerted measurements of
137 bulk seawater, SML, size-resolved aerosol particles and fog water at different locations dominated by
138 different sea-ice-related sea surface compartments (ice-free, leads/polynyas, MIZ, melt ponds) in the
139 Arctic Ocean. All marine and atmospheric compartments are discussed and compared on absolute
140 CCHO concentrations, calculated $CCHO/Na^+$ ratios, the relative monosaccharide contribution to CCHO
141 and the occurrence of $dFCHO$. The complex nature of these primary emission mechanisms and
142 subsequent atmospheric aging of marine CCHO in the Arctic Ocean are discussed in relation to our
143 findings. Our Arctic results are collated with those from the Southern Ocean at the Antarctic peninsula

144 during the austral summer, as presented in Zeppenfeld et al. (2021a) following a similar experimental
145 design. While both polar locations are remote marine regions with comparable meteorological
146 conditions during the sampling periods, the presence of Arctic sea ice adds another dimension of
147 complexity to data interpretation.

148 2. Experimental

149 2.1 Study area and field sampling

150 Field samples were gathered during the PS106 (PASCAL/SiPCA) campaign (Macke and Flores, 2018;
151 Wendisch et al., 2018) conducted from May to July 2017 on board the German icebreaker RV
152 *Polarstern* in the Fram Strait, Barents Sea and central Arctic Ocean, including a period operating from
153 a drifting ice station (03–16 June 2017).

154 Marine SML and corresponding bulk water samples were collected from various locations as shown in
155 [Figure SI 1](#). These include the ice-free ocean (four sampling events), open water areas within the pack
156 ice (20 sampling events, without distinguishing between open leads and polynyas), the MIZ (five
157 sampling events), and young and aged melt ponds (six sampling events). Using visual characteristics,
158 melt ponds were categorized as young (small, bluish, clear) or aged (larger, darker blue to greenish,
159 and turbid with particulates and microalgae). To minimize contamination from exhausts and
160 wastewater, water samples were taken at distances greater than 100 m from the ship. Seawater was
161 collected either using a rubber boat or directly from the ice edge. SML samples were obtained by
162 immersing a glass plate (length: 50 cm, width: 20 cm, thickness: 0.5 cm, sampling area: 2000 cm²)
163 vertically into the surface water and slowly withdrawing it at a speed of approximately 15 cm s⁻¹ (van
164 Pinxteren et al., 2012; Zeppenfeld et al., 2021a). The adhered SML film was drawn off the glass plate
165 surface into a prewashed wide-neck plastic bottle by a framed Teflon wiper. The average thickness of
166 the SML collected during this field study was 76±10 µm, which was calculated based on the volume of
167 the SML sample collected, the area of the immersed glass plate and the number of dips as described
168 by Cunliffe and Wurl (2014). Despite air temperatures during PS106 (median: -0.5°C; minimum: -7.6°C)
169 hovering around or slightly below the freezing point of seawater, the SML remained unfrozen on the
170 glass plate during sampling. The corresponding bulk water was taken from a defined depth of 1 m into
171 LDPE bottles attached to a telescopic rod, except in the bottom closed melt ponds where it was
172 scooped from the bottom at approximately 20–40 cm depth. Whenever melt pond sampling took
173 place, snow samples were collected from the ice floe surface roughly 10 m away from the melt pond.
174 Before each sampling, the sampling containers were first rinsed with a few milliliters of the
175 corresponding aqueous sample which was disposed immediately after. On board, small aliquots of the
176 water samples were analyzed immediately for salinity using a conductivity meter (pH/Cond 3320,
177 WTW), colored dissolved organic matter (CDOM) and particulate absorption (PAB), with more details
178 in section 2.6. For later chemical analyses (inorganic ions, pH, carbohydrates) 500–1000 mL of 0.2 µm
179 filtered water sample (dissolved fraction), 0.2 µm polycarbonate filters (particulate fraction) and field
180 blanks were stored at -20°C.

181 The sampling of ambient aerosol particles was conducted at the starboard side of RV *Polarstern* at the
182 top of the observation deck at a height of approx. 25 m above sea level as already described in Kecorius
183 et al. (2019). Size-segregated aerosol particles were sampled in five size ranges (stage 1: 0.05–0.14 μm ,
184 stage 2: 0.14–0.42 μm , stage 3: 0.42–1.2 μm , stage 4: 1.2–3.5 μm , stage 5: 3.5–10 μm aerodynamic
185 particle diameter with a 50% cut-off) on aluminum foils by using two synchronized low-pressure Berner
186 impactors (Hauke, Austria) with a flow rate of 75 L min^{-1} and a sampling time of three to six days. To
187 avoid the condensation of atmospheric water and subsequent microbial activities on the aluminum
188 foils, a 3 m long heated tubes between the isokinetic inlets and the impactors reduced the relative
189 humidity of the sampled air to 75–80%, when the ambient relative humidity was higher. During this
190 field study, the difference of the temperatures of the ambient air at the inlet and the sampled air after
191 the heating never exceeded 9 K. Consequently, losses of semi-volatile compounds or changes by heat-
192 induced chemical reactions are expected to be neglectable. Furthermore, the Berner impactors were
193 thermally insulated by a polystyrene shell. After sampling, the foils were stored in aluminum containers
194 at -20°C until analysis. In this study, the results from stages 1–3, 4–5 and 1–5 were summed up as
195 submicron (sub), supermicron (super) and PM_{10} , respectively. Details about the size-resolved aerosol
196 particle samples and corresponding meteorological information are given in (Table SI 1, in total 15
197 complete sets of Berner foils).

198 Close to the aerosol sampling, fog was collected using the Caltech Active Strand Cloud Collector Version
199 2 (CASCC2) as described by Demoz et al. (1996). Bulk fog droplets were impacted on Teflon strands
200 with a diameter of 508 μm and collected into a prewashed Nalgene polyethylene bottle. The flow rate
201 was 5.3 $\text{m}^3 \text{min}^{-1}$ and the 50% lower cut-off was determined to be approximately 3.5 μm . Further
202 information about the 22 fog samples collected during the PS106 campaign including meteorological
203 information can be found in Table SI 2 and in Hartmann et al. (2021).

204 **2.2 Total aerosol particle mass concentrations**

205 Before and after sampling, the aluminum foils were equilibrated (three days, 20°C , 50% relative
206 humidity) and weighed using a precise microbalance (Mettler Toledo XP2U, weighing error: $\pm 4.6 \mu\text{g}$).
207 Total particle mass concentrations ($\text{mass}_{\text{aer, stage } y}$) were calculated for each Berner stage as the ratio
208 between the difference of the absolute foil masses after and before sampling and the sampled air
209 volume. Afterwards, aluminum foils were divided for further chemical analyses.

210 **2.3 OC/EC in aerosol samples**

211 Organic carbon (OC_{aer}) and elemental carbon on Berner aerosol foils were determined as described by
212 Müller et al. (2010) using a two-step thermographic method (C/S MAX, Seifert Laborgeräte, Germany)
213 with a nondispersive infrared sensor.

214 **2.4 Carbohydrates in aerosol particles, fog, snow, seawater and melt ponds**

215 Marine carbohydrates in the particulate ($pCCHO$, $>0.2 \mu\text{m}$) and dissolved ($dCCHO/dFCHO$, $<0.2 \mu\text{m}$)
216 phases, including truly dissolved molecules and small colloids, were quantified from seawater and melt
217 pond samples following the protocol presented by Zeppenfeld et al. (2020, 2021a) using high-
218 performance anion-exchange chromatography with pulsed amperometric detection (HPAEC-PAD)
219 equipped with a Dionex CarboPac PA20 analytical column (3 mm \times 150 mm) and a Dionex CarboPac
220 PA20 guard column (3 mm \times 30 mm). The monosaccharides fucose (Fuc), rhamnose (Rha), arabinose
221 (Ara), galactose (Gal), glucose (Glc), xylose (Xyl), mannose (Man), fructose (Fru), galactosamine (GalN),
222 glucosamine (GlcN), muramic acid (MurAc), galacturonic acid (GalAc), and glucuronic acid (GlcAc) were
223 identified by their retention times. $dFCHO$ represent the sum of identifiable monosaccharides before,
224 and $dCCHO$ and $pCCHO$ additionally released after an acid hydrolysis (0.8 M HCl, 100°C, 20 h). $CCHO$ is
225 the sum of $dCCHO$ and $pCCHO$. CHO represents the sum of $CCHO$ and $dFCHO$, and consequently
226 encompasses all carbohydrates measured within this study. **Figure 1** gives an overview of the here
227 used carbohydrate-related abbreviations. Marine carbohydrates in fog water, snow and extracts from
228 size-resolved aerosol particles were measured with ($CCHO_{\text{fog}}$, $CCHO_{\text{aer}}$) or without ($dFCHO_{\text{fog}}$, $dFCHO_{\text{aer}}$)
229 prior acid hydrolysis.

230

231

232

233

234

235

236

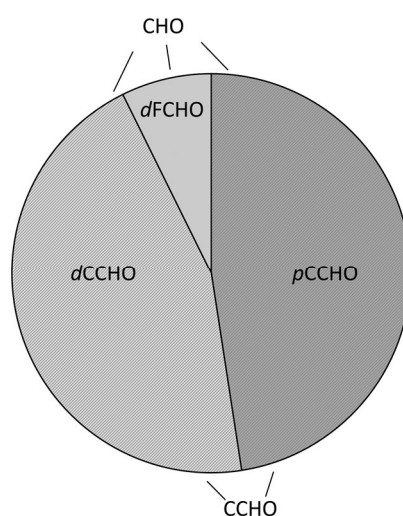


Figure 1. Overview of the abbreviations for carbohydrates (CHO) in seawater. CCHO: combined carbohydrates; $pCCHO$: particulate combined carbohydrates, $dCCHO$: dissolved combined carbohydrates; $dFCHO$: dissolved free carbohydrates.

237 **2.5 Sodium and pH in aerosol particles, fog, seawater and melt ponds**

238 Major inorganic ions, including sodium (Na^+), were determined from 0.45 μm filtered aqueous extracts
239 of the size-resolved aerosol samples (50% of the Berner foil in 2 mL ultrapure water), fog water, diluted
240 (1:15 000) seawater and melt pond samples using ion chromatography (ICS-3000, Dionex) as described
241 by Müller et al. (2010). In this study, we discuss the results for Na^+ as a proxy for SSA emissions in
242 remote marine regions. Additionally, the pH was monitored by an additional autosampler sample

243 conductivity and pH accessory (Dionex) in all seawater, melt pond and, whenever enough sample
244 volume was available, in fog water.

245 **2.6 Absorption by phytoplankton, non-algal particles and colored dissolved** 246 **organic matter in seawater and melt pond samples**

247 For the investigation of bio-optical parameters in seawater and melt pond samples, the particulate
248 fraction was collected by filtering the water samples (5–500 mL) onto glass-fiber filters (GF/F,
249 Whatman), while the dissolved fraction was filtered through 0.2 μm Spartan syringe filters (Whatman,
250 Germany) immediately after sampling. The GF/F filters were analyzed to determine the absorption
251 spectra (i.e. 320–844 nm, 2 nm resolution) using the quantitative filtration technique with an
252 integrative-cavity absorption meter setup (QFT-ICAM) as developed by Röttgers et al. (2016). We
253 followed the protocol by Liu et al. (2018) for the instrument used here and the determination of the
254 absorption coefficients by total particles (a_{p440}), phytoplankton (a_{ph440}) and non-algal particles
255 (a_{NAP440}) at $\lambda=440$ nm.

256 The absorption for the dissolved fraction ($a_{CDOM}(\lambda)$) between 270 and 750 nm (1 nm resolution) were
257 measured as triplicates using a long path length liquid waveguide capillary cell (LWCC) system following
258 the procedure by Lefering et al. (2017) and including the correction for salinity effects by Röttgers et
259 al. (2014) as described for our instrumentation in Álvarez et al. (2022). The absorption coefficients in
260 the visible at 443 nm ($a_{CDOM443}$) and UV at 350 nm ($a_{CDOM350}$) bands were used as indicators of CDOM
261 magnitude.

262 **2.7 Supporting observations**

263 The German research vessel *Polarstern* performs continuous meteorological surface measurements
264 during times of ship operation. For this study, we used the data from the HMP155
265 thermometer/hygrometer probe (Vaisala), the ultrasonic anemometer (Thies Clima) and the FS11
266 visibility sensor (Vaisala) installed at a height of 29 m, 39 m and 20 m above sea level, respectively. The
267 quality-controlled data made available by the operators on the public repository PANGAEA
268 (Schmithüsen, 2018, 2019) supported the interpretation of the results of this study.

269 The 120 h back-trajectories were computed for the sampling periods of the size-resolved aerosol
270 particles and fog water events using the NOAA HYSPLIT model (Stein et al., 2015). The back-trajectories
271 were calculated on an hourly basis using the GDAS1 meteorological fields (Global Data Assimilation
272 System; 1° latitude/longitude; 3-hourly) and at arrival heights of 50, 250 and 1000 m. Sea ice
273 concentration data were retrieved from ERDDAP (Environmental Research Division's Data Access
274 Program), a data server maintained by NOAA (National Oceanic and Atmospheric Administration). The
275 MIZ was defined here as the oceanic region with a sea ice concentration between 15 and 80%. Data

276 on melt pond fractions were accessed from the sea ice remote sensing data achieve of the University
277 of Bremen (<https://data.seaice.uni-bremen.de>, Istomina (2020)).

278 **2.8 Statistics, calculations and visualization**

279 Statistical analyses, calculations and visualization were performed in OriginPro, Microsoft Excel and R
280 version 4.2.1 using the following packages: oce, ocedata, ncdf4, openair, ggplot2, reshape2, scales,
281 lubridate, cmocean, maps, mapdata, rgdal, raster, RColorBrewer, sp. Time-resolved back-trajectories
282 and sea ice maps were combined using R to compute and visualize the air mass history regarding the
283 sea-ice-related sea surface compartments that have been passed. As a result, relative residence times
284 of the air masses over certain surface features (ice-free, MIZ, pack ice, land) 12 hours before sampling
285 were calculated based on defined thresholds for sea ice concentration: less than 15% for ice-free
286 ocean, 15-80% for MIZ and over 80% for pack ice. Based on the remote sensing data used, we did not
287 distinguish between open leads and melt ponds as the air traversed the pack ice. Box-whisker plots
288 represent the interquartile range (box), median (horizontal line within the box), average (open square)
289 and the minimum and maximum values of the datasets (whiskers). Measured mean values are given
290 together with the calculated standard deviations (\pm). Correlations between two measured variables
291 were expressed via the Pearson correlation coefficient R . The thresholds of significance were set for
292 the p-values 0.1, 0.05, 0.01 and 0.001.

293 Enrichment factors for CCHO in the SML (EF_{SML}) relative to the corresponding bulk sample in different
294 sea-ice-related sea surface compartments (ice-free, leads/polynyas, MIZ, melt ponds) were calculated
295 based on **Formula 1** with $[x]_{SML}$ and $[x]_{bulk}$ representing the concentrations of either $pCCHO$ or $dCCHO$.
296 For the calculation of enrichment factors of CCHO in aerosol particles on Berner stage y ($EF_{aer,stage y}$;
297 **Formula 2**) and fog water (EF_{fog} ; **Formula 3**) relative to the bulk water samples, the ocean was assumed
298 as the most likely source of atmospheric Na^+ . For the calculations of EF_{aer} and EF_{fog} , we used the median
299 value of all $CCHO_{bulk} / Na^+_{bulk}$ ratios found in the samples of a certain sea-ice-related sea surface
300 compartment (ice-free, leads/polynyas, MIZ, melt ponds) over the whole campaign.

301

$$302 \quad EF_{SML} = \frac{[x]_{SML}}{[x]_{bulk}} \quad (1)$$

$$303 \quad EF_{aer,stage y} = \frac{[x]_{aer,stage y} / [Na^+]_{aer,stage y}}{[x]_{bulk} / [Na^+]_{bulk}} \quad (2)$$

$$304 \quad EF_{fog} = \frac{[x]_{fog} / [Na^+]_{fog}}{[x]_{bulk} / [Na^+]_{bulk}} \quad (3)$$

305 **3. Results and Discussion**

306 The sources of SSA particles, and hence of atmospheric marine carbohydrates, microbial cells and
307 fragments, in the Arctic are diverse and influenced by the prevailing sea ice conditions. Here, we
308 present the concentrations and relative compositions of CCHO in the SML and bulk water from the ice-
309 free ocean, open leads and polynyas within the pack ice, melt ponds and the MIZ. After this, the
310 different sea-ice-related sea surface compartments are linked with the atmospheric CCHO found in
311 ambient size-resolved aerosol particles and fog water. The influences of the air mass history,
312 enrichments of CCHO towards Na⁺ during the sea-air transfer and secondary atmospheric
313 transformations processes altering atmospheric CCHO are discussed.

314 **3.1 Sea ice influences the properties of the sea surface water**

315 **Variable CCHO concentrations in the Arctic surface water.** CCHO were found in the dissolved (*d*CCHO)
316 and particulate (*p*CCHO) phases of the SML and bulk water samples collected from the ocean and the
317 melt ponds during the PS106 campaign. Among all aqueous samples, regardless of the sampling
318 environment and depth (SML versus bulk), *d*CCHO (13–640 µg L⁻¹; mean_{*d*CCHO} = 82±110 µg L⁻¹; n=70)
319 and *p*CCHO (4–810 µg L⁻¹; mean_{*p*CCHO} = 84±160 µg L⁻¹; n=70) concentrations were highly variable.
320 However, the minimum, maximum and mean values of both *d*CCHO and *p*CCHO ranged within the
321 same orders of magnitude. CCHO as the sum of *d*CCHO and *p*CCHO ranged between 22–1070 µg L⁻¹
322 (mean_{CCHO} = 166±250 µg L⁻¹; n=70).

323 Large differences in the mean values and standard deviations of CCHO were observed among the four
324 sea-ice-related sea surface compartments in the Arctic (leads/polynyas within the pack ice, MIZ, ice-
325 free ocean, melt ponds) as shown in [Figure 2a+b](#). The highest mean values for *d*CCHO and *p*CCHO were
326 observed in the SML of the MIZ (mean_{*d*CCHO, SML, MIZ} = 190±160 µg L⁻¹; mean_{*p*CCHO, SML, MIZ} = 370±310 µg L⁻¹;
327 n=5) and melt ponds (mean_{*d*CCHO, SML, melt ponds} = 190±240 µg L⁻¹; mean_{*p*CCHO, SML, melt ponds} = 200±310 µg L⁻¹;
328 n=6), while the SML of the lead/polynya (mean_{*d*CCHO, SML, lead/polynya} = 70±75 µg L⁻¹;
329 mean_{*p*CCHO, SML, lead/polynya} = 70±120 µg L⁻¹; n=20) and ice-free open ocean (mean_{*d*CCHO, SML, ice-}
330 _{free} = 73±12 µg L⁻¹; mean_{*p*CCHO, SML, ice-free} = 36±5 µg L⁻¹; n=4) samples tended to contain less CCHO. The
331 lower SML concentrations from this study for the Arctic ice-free open ocean and lead/polynya samples
332 align closely with several other investigations. Specifically, our results are comparable to Gao et al.
333 (2012), who studied the SML of Arctic leads (mean_{*d*CCHO, SML, Arctic leads} = 163±104 µg L⁻¹; mean_{*p*CCHO, SML, Arctic}
334 _{leads} = 35±25 µg L⁻¹; n=4), and Zeppenfeld et al. (2021), focusing on the ice-free part of the Southern
335 Ocean west of the Antarctic peninsula during the austral summer
336 (mean_{*d*CCHO, SML, Southern Ocean} = 48±63 µg L⁻¹; mean_{*p*CCHO, SML, Southern Ocean} = 72±53 µg L⁻¹; n=18). Similarly, our
337 data mirror findings from the tropical Cape Verde (mean_{*d*CCHO, SML, Cape Verde} = 85±30 µg L⁻¹; van Pinxteren

338 et al., 2023) and the Peruvian upwelling region (mean_{dCCHO, SML, Peru} $\approx 92 \pm 32 \mu\text{g L}^{-1}$; Zäncker et al., 2017).
 339 Consequently, the Arctic MIZ and melt ponds, especially the aged ones with advanced microbiological
 340 activities, stood out with elevated CCHO within the Arctic and also compared to tropical and other
 341 polar regions.

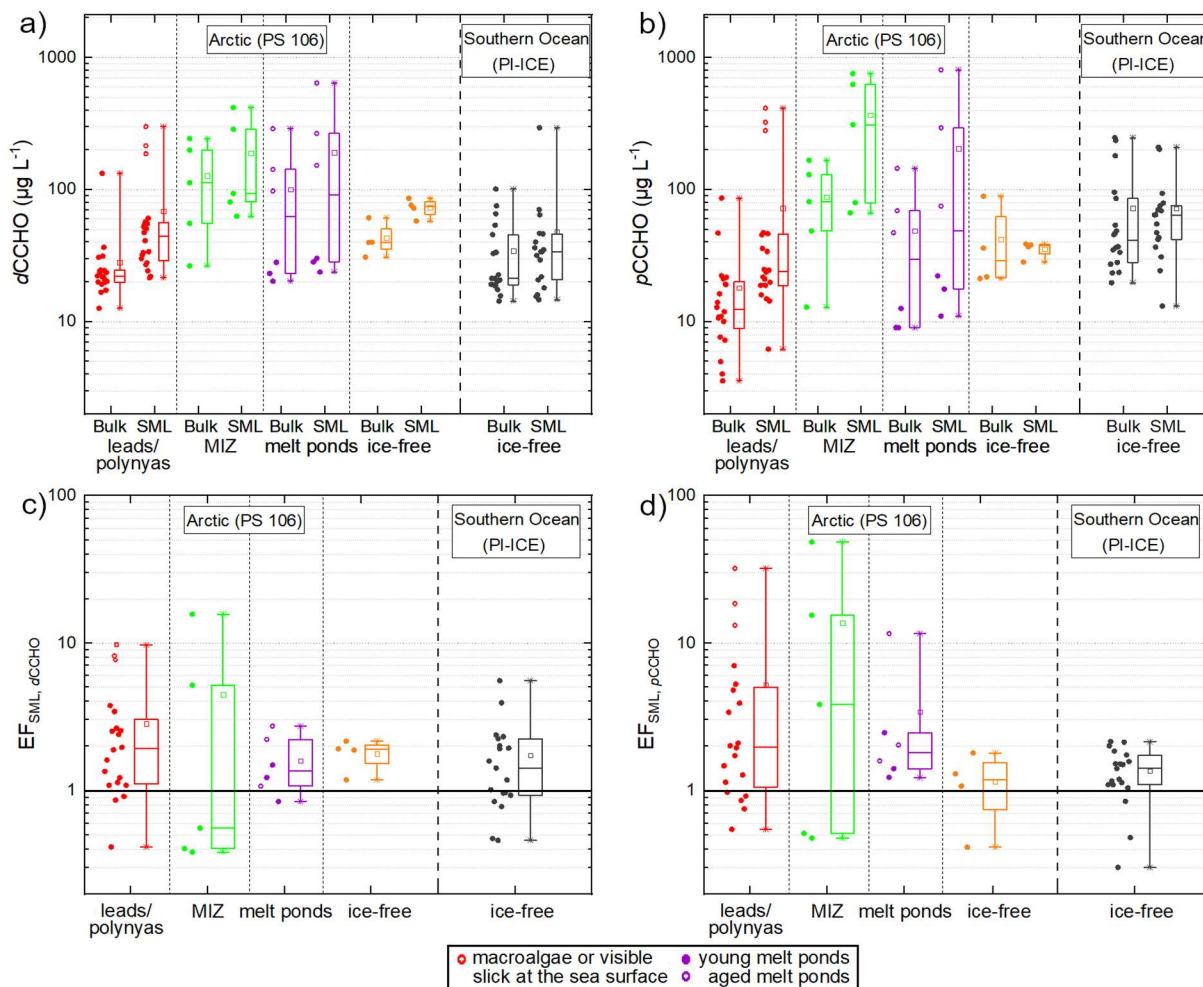


Figure 2. Scattered box-whisker plots showing the concentrations of a) *dCCHO* and b) *pCCHO* in the bulk and SML samples from the open leads and polynyas in the pack ice (red), the MIZ (green), ice-free open ocean (orange) and young and aged melt ponds (purple) collected during the PS106 campaign in the Arctic in comparison to the ice-free part of the Southern Ocean west of the Antarctic Peninsula investigated during the PI-ICE campaign in 2019 (black) as published in Zeppenfeld et al. (2021). EFs between SML and bulk water are shown in c) for *dCCHO* and d) for *pCCHO*. The black horizontal line represents an EF=1 meaning no enrichment or depletion.

342 **Variable enrichments of CCHO in the SML.** The enrichment factors (EF_{SML}) of the CCHO in the SML
 343 relative to the corresponding bulk water ranged between 0.4 and 16 for *dCCHO* (Figure 2c), while the
 344 EF_{SML} for *pCCHO* varied between 0.4 and 49 (Figure 2d). 80% of the SML samples were moderately or
 345 highly enriched in marine carbohydrates with only a few cases of depletion (7 for *dCCHO* and 8 for
 346 *pCCHO* out of 35 in total). With a median EF_{SML, pCCHO, MIZ} value of 3.8 and a mean of 13.8, the enrichment
 347 of *pCCHO* in the MIZ stood out compared to the *pCCHO* in other sea-ice-related sea surface
 348 compartments and to *dCCHO* overall. However, it should be noted that the number of MIZ samples
 349 was low and median and mean values were dominated by three sample pairs with very high EF_{SML}

350 values. Low to moderate enrichments for *d*CCHO and *p*CCHO were typically found in the lead/polynya
351 samples from the pack ice (median $EF_{SML,dCCHO,leads/polynyas}=1.9$; median $EF_{SML,pCCHO,leads/polynyas}=2.0$, $n=20$).
352 However, three lead samples showed quite high *d*CCHO & *p*CCHO concentrations in the SML compared
353 to the corresponding bulk samples resulting in high $EF_{SML,dCCHO,leads/polynyas}$ up to 10 and
354 $EF_{SML,pCCHO,leads/polynyas}$ up to 32. The exceptionally high EFs of these three samples can be explained by
355 the observation of slicks - visible films on the sea surface with altered reflectance and typically high
356 enrichments of organics (Cunliffe et al., 2013; Stolle et al., 2010; Williams et al., 1986; Wurl et al., 2009)
357 as well as the presence of macroalgae floating at the ocean's surface near the sampling site. Even
358 though the macroalgae were not collected themselves, their exudates or fragments might have been
359 released, accumulated and distributed in the SML close-by and thus sampled. Consequently, the few
360 samples with high EFs in open leads might rather represent exceptional events as spatially small-scale
361 phenomena.

362 The slight to high enrichments for *d*CCHO and *p*CCHO in this study are in good agreement with the
363 values reported by Gao et al. (2012), who determined $EF_{SML,dCCHO}$ between 3.5 and 12, and $EF_{SML,pCCHO}$
364 between 1.7 and 7.0 for open leads within the central Arctic Ocean. Furthermore, the $EF_{SML,dCCHO}$ of the
365 four Arctic sea-ice-related sea surface compartments reported here were not significantly different
366 compared to values found in the ice-free part of the Southern Ocean (ANOVA, one-way, 0.05
367 significance level). For the *p*CCHO, however, the average EF_{SML} in the Arctic MIZ was significantly higher
368 than the one of the Southern Ocean, whereas the $EF_{SML,pCCHO}$ of the ice-free ocean in the Arctic were
369 similar to the Southern Ocean.

370 For explaining the accumulation in the SML, previous studies proposed several mechanisms and
371 processes, which fundamentally differ for the dissolved and particulate carbohydrates. The enrichment
372 of *p*CCHO in the SML might be dominated by an interplay of density-related and wind-driven processes.
373 For instance, the positive buoyancy of TEP, a subgroup of *p*CCHO, leads to an upward flux serving as a
374 continual vehicle for marine organisms and attached chemical compounds (Azetsu-Scott and Passow,
375 2004; Mari et al., 2017). Furthermore, strong winds can cause a short-term mixing of the upper water
376 column reducing the EF_{SML} of particulates (Obernosterer et al., 2008) or TEP (Wurl et al., 2009; Zäncker
377 et al., 2021), while the wind-induced entrainment of air and the bubbling of seawater convert dissolved
378 negatively charged *d*CCHO and colloids into larger aggregates due to their sticky properties leading to
379 an enrichment of *p*CCHO in the SML (Passow, 2002; Robinson et al., 2019; Wurl et al., 2011). The
380 enrichment of *d*CCHO and also *d*FCHO in the SML is attributed to co-adsorption to other surface-active
381 compounds from the seawater matrix being scavenged at the surface of rising bubbles (Burrows et al.,
382 2016; Hasenecz et al., 2020; Schill et al., 2018; Xu et al., 2023). Additionally, microbial processes in the
383 SML could enhance the enrichment through in-situ *d*CCHO production by micro- or macroalgae, while

384 photolysis and enzymatic degradation of *d*CCHO into *d*FCHO by heterotrophic bacteria could decrease
385 the SML enrichment. Specific to the Arctic, the release of meltwater from the sea ice could be an
386 additional source for carbohydrates in the SML, considering the production of CCHO, exopolymeric
387 substances (EPS) and TEP by sea ice algae and bacteria as a protection strategy against freezing damage
388 and fluctuating salinity in sea ice (Aslam et al., 2016; Krembs et al., 2002; Krembs and Deming, 2008).
389 This aligns well with the finding by Galgani et al. (2016) who observed labile, fresh OM in the SML of
390 melt ponds compared to the rather old, refractory nature of the SML in the surrounding open leads.
391 Hence, melting of sea ice could explain the extraordinarily high EF_{SML} observed in some, but not all,
392 SML samples from the MIZ and melt ponds. In summary, several processes might be responsible for
393 enrichment processes in the SML, especially in the Arctic, where the melting of sea ice could strongly
394 bias the physiochemical processes usually observed in controlled tank experiments.

395 ***High and low salinities due to freezing and melting of sea ice.*** While the surface seawater of the Arctic
396 Ocean is very saline, the Arctic sea ice is much fresher due the separation into salt-free ice crystals and
397 a salty brine during its formation from seawater and a subsequent salt loss from gravity drainage in
398 winter and flushing during summer (Notz and Worster, 2009). During the late spring and summer
399 period of this study, when strong melting of sea ice occurs, a large amount of freshwater enters the
400 surface of the ocean creating inhomogeneities of salinity within the surface of the ocean. In both the
401 ice-free ocean and the pack ice, where sea ice exists, but the melting rate is low, salinities of the SML
402 and the bulk water ranged in this study between 30.9 and 34.5 (Zeppenfeld et al., 2019b), which is
403 typical for the SML and the surface bulk water of the Arctic Ocean (Vaqué et al., 2021). Within the MIZ,
404 where freshwater from melting sea ice quickly mixes with the salty ocean water, salinities were similar
405 with values in this study between 30.1 and 33.4, however, also with an exception in the SML of 25.7.
406 Melt ponds that were not yet joined at the bottom with the ocean below, were much fresher with
407 lower and more variable salinities ranging from 4.3 to 19.5 (Zeppenfeld et al., 2019b). With a few
408 exceptions, salinity discrepancies between the SML and the corresponding bulk water were small in
409 most cases.

410 Sea-air transfer studies usually refer to open ocean scenarios with high salinities in the seawater and
411 without the presence of melting sea ice. For the calculation of enrichment factors of organics in aerosol
412 particles (EF_{aer}) or fog (EF_{fog}), the concentrations of Na^+ – a major compound of sea salt – in the
413 seawater bulk is included by default (see equations 2 and 3). However, the Arctic is a more complex
414 marine environment where salinities, and hence Na^+ concentrations, can vary widely as melting
415 progresses. This may strongly influence the mechanisms behind the bubble bursting process, the
416 CCHO/ Na^+ ratios in the bulk seawater and the SML, and thus also the EF_{aer} and EF_{fog} as it will be

417 discussed in section 3.4. Consequently, the variability of salinity in Arctic seawater and melt ponds
418 should be considered for sea-air transfer studies that rely on Na^+ values.

419 ***Four sea-ice-related sea surface compartments with different characteristics.*** The high Arctic differs
420 from other oceanic regions in the presence, formation and melting of sea ice creating sea-ice-related
421 sea surface compartments (ice-free, leads/polynyas, MIZ, melt ponds) with individual biological and
422 chemical characteristics, such as CCHO concentrations, enrichments in the SML and salinities. This
423 might potentially impact the transfer of substances from the ocean to the atmosphere, chemo-
424 selective enrichment processes of marine CCHO in the primary marine aerosol particles and thus their
425 microphysical properties. The next sections will elucidate if and how these differences within the
426 individual compartments relate to CCHO_{aer} and CCHO_{fog} .

427 **3.2 Sea spray aerosol and therein contained combined carbohydrates**

428 ***Breaking waves as the main mechanism for SSA emissions is not unambiguous in the Arctic.*** In the
429 open ocean, the emission flux of SSA and hence its inorganic and organic constituents mainly depend
430 on the wind speed as the driving force for breaking waves and bubble bursting, and furthermore on
431 the seawater temperature, salinity, wave properties and organic surface-active substances (Grythe et
432 al., 2014). In this study, atmospheric sodium ($\text{Na}^+_{\text{aer,PM10}}$), the best tracer for SSA (Barthel et al., 2019),
433 ranged between 12 and 765 ng m^{-3} (Table SI 1). $\text{Na}^+_{\text{aer,PM10}}$ showed a good correlation ($R=0.80$, $p<0.001$,
434 Figure 3a) with wind speed, measured at the sampling site and averaged over the sampling time, if all
435 aerosol samples are included. However, the strength of this correlation decreased sharply ($R=0.59$,
436 $p<0.1$), when only samples collected over the MIZ and the pack ice were included, while the few
437 samples from the open ocean characterized by high Na^+ values were excluded. This is due to the
438 presence of sea ice in the high Arctic, which likely alters and conceals the classical wind-driven
439 mechanisms of breaking waves and bubble bursting resulting in SSA emission. Firstly, sea ice covers a
440 significant part of the Arctic Ocean strongly reducing the area releasing SSA. Secondly, the presence of
441 sea ice causes an attenuation of the high-frequency wind-sea waves, while longer waves, such as
442 swells, can remain (Thomson, 2022). Consequently, the effect of wind on the SSA emission mechanisms
443 within the open leads and the MIZ might be different than in the ice-free ocean. For those sea-ice-
444 dominated compartments, alternative wind-independent sources of ascending bubbles were
445 suggested, such as melting sea ice nearby, respiration of phytoplankton or sea-air heat exchange below
446 the sea surface (Chen et al., 2022 and references therein). Thirdly, in contrast to other marine regions
447 with quite homogeneous ocean salinities, and hence sodium concentrations, the salinities among the
448 different Arctic sea-ice-related sea surface compartments are more variable due to the melting of sea
449 ice. Previously, the results of a sea-air transfer tank experiment with artificial seawater showed the
450 influence of salinity on the relative particle number concentrations of emitted SSA for salinities below
451 15 – values especially relevant for melt ponds in the Arctic – while changes at higher salinities did not
452 result in a measurable effect (Zábori et al., 2012). Additionally, organics with potential surface-active
453 properties are very variable in these disparate Arctic environments, as discussed for CCHO in section
454 3.1. Organic surfactants can alter the ocean surface's ability to form whitecaps and the lifetime of
455 bubbles (Bigg and Leck, 2008; Callaghan et al., 2012; Grythe et al., 2014) and therefore SSA properties.
456 Finally, blowing snow over the sea ice could serve as an additional source of atmospheric Na^+_{aer} when
457 a certain air-temperature-dependent wind speed threshold is exceeded. (Chen et al., 2022; Gong et
458 al., 2023; Huang and Jaeglé, 2017; Yang et al., 2008). Consequently, connections and correlations for
459 the release of SSA particles in the heterogeneous high Arctic are more difficult to explore than other

460 marine environments without sea ice. It can be assumed that this complex setting influences not only
 461 the release of inorganic constituents from seawater, but also its organic compounds, such as CCHO.

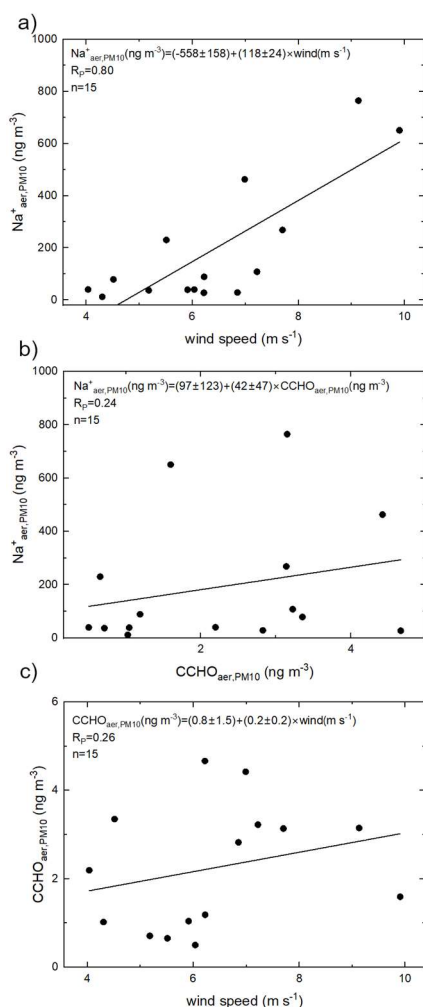
462

463

464

465

466



467

468

469

470

471

472

473

474

475

476 **Figure 3.** Correlations between a) $\text{Na}^+_{\text{aer,PM10}}$ and the averaged wind speed, b) $\text{Na}^+_{\text{aer,PM10}}$ and $\text{CCHO}_{\text{aer,PM10}}$, c) $\text{CCHO}_{\text{aer,PM10}}$ and averaged wind speed.

477 ***CCHO_{aer} distributed in all size modes.*** During the PS106 campaign, the overall atmospheric
 478 concentrations of $\text{CCHO}_{\text{aer,PM10}}$ ranged between 0.5 and 4.7 ng m⁻³ (Table SI 1). Combined
 479 carbohydrates were found on both supermicron ($\text{CCHO}_{\text{aer,super}} = 0.07\text{--}2.1$ ng m⁻³) and submicron
 480 particles ($\text{CCHO}_{\text{aer,sub}} = 0.26\text{--}4.4$ ng m⁻³). Thus, these CCHO_{aer} values ranged within the same orders of
 481 magnitude as in the Arctic studies by Karl et al. (2019) and Leck et al. (2013) or the study conducted at
 482 the western Antarctic peninsula by Zeppenfeld et al. (2021a). CCHO_{aer} appeared in all of the five size
 483 classes in variable concentrations (Figure 4a). Although the average concentrations were similar on all
 484 stages, local maxima were observed on stages 2 (0.14–0.42 μm) and 5 (1.2–10 μm). A similar size
 485 distribution of marine CCHO_{aer} in these specific size ranges, but more pronounced, has been already
 486 observed in the ice-free part of the Southern Ocean by Zeppenfeld et al. (2021a) explaining these
 487 findings with a likely release of marine polysaccharides from the ocean as part of film and jet droplets.
 488 Possibly, the aerosol size distribution of marine polysaccharides resulting from wind-driven bubble

489 bursting emissions are not as obvious in this Arctic study as it was in the ice-free Southern Ocean due
 490 to the presence of Arctic sea ice suppressing and altering the local SSA emission mechanisms as
 491 indicated in the previous section. The relative contribution of $CCHO_{aer}$ to $mass_{aer}$ varied between 0.01%
 492 and 4% (Figure 4b), while the carbon contained within the combined carbohydrates (C- $CCHO_{aer}$)
 493 contributed 0.06 to 4.9% to the OC_{aer} in the size-resolved aerosol particles (Figure 4c). These
 494 contributions agree well with the findings in marine aerosol particles from the Southern Ocean
 495 (Zeppenfeld et al., 2021a).

496

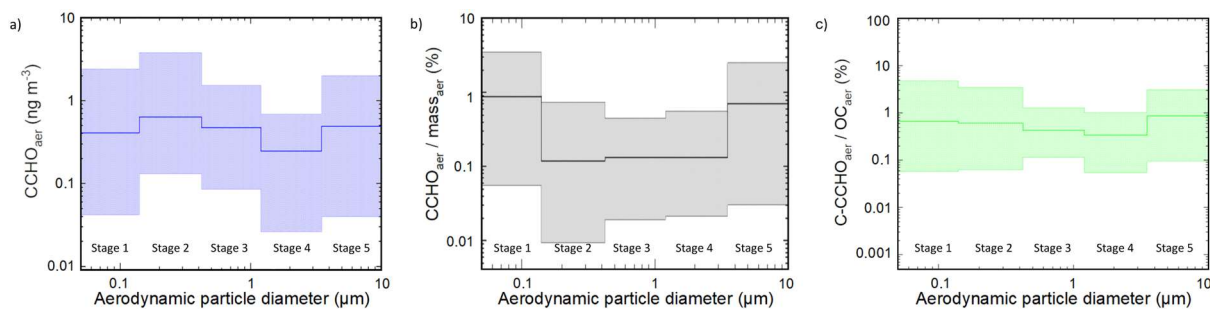


Figure 4. a) Concentration of combined carbohydrates in size-resolved aerosol particles ($CCHO_{aer}$), b) ratio of $CCHO_{aer}$ to the total particle mass concentration ($mass_{aer}$), c) ratios of carbon contained within the combined carbohydrates in aerosol particles (C- $CCHO_{aer}$) to organic carbon in aerosol particles (OC_{aer}). The bold lines represent the average concentrations during the PS106 campaign. The hatched areas show the range between the maximum and minimum values. The aerodynamic particle diameter refers to sampling conditions at relative humidity of max. 80%.

497 Unlike the study conducted in the Southern Ocean (Zeppenfeld et al., 2021a), $CCHO_{aer,PM10}$ in this study
 498 showed no significant correlations with $Na^+_{aer,PM10}$ ($R=0.24$, $p>0.1$, Figure 3b) or wind speed ($R=0.26$,
 499 $p>0.1$, Figure 3c). The presence of sea ice resulting in melt ponds and MIZ regions, and the interplay of
 500 multiple emission mechanisms in the Arctic, as discussed earlier, could account for this complexity.
 501 However, if the correlations are resolved for the different Berner impactor stages (i.e. size ranges), a
 502 large variability can be observed (Figure 5). A higher correlation was found especially on stage 4
 503 ($1.2\text{--}3.5\ \mu\text{m}$) between $CCHO_{aer,stage\ 4}$ and $Na^+_{aer,stage\ 4}$ ($R=0.76$, $p<0.01$), while the Pearson correlations
 504 coefficients for the other Berner stages were much lower. This could indicate the same marine source
 505 and wind-driven emission mechanism for both chemical constituents in this supermicron aerosol size
 506 mode, while other aerosol size modes might have been influenced by atmospheric aging and wind-
 507 independent emission mechanisms as already mentioned for Na^+_{aer} in the previous section. This
 508 observation agrees well with the findings by Bigg and Leck (2008) and Leck (2002) reporting submicron
 509 polymer gel particles, likely consisting of polysaccharides, in the atmosphere of the high Arctic
 510 containing almost no sea salt and showing large similarities to those particles found in open leads
 511 close-by. This is quite surprising considering that the mechanism of wind-driven wave breaking is quite
 512 limited due to the lack of long fetches of open water (Held et al., 2011; Norris et al., 2011).

513
514
515
516
517
518
519
520

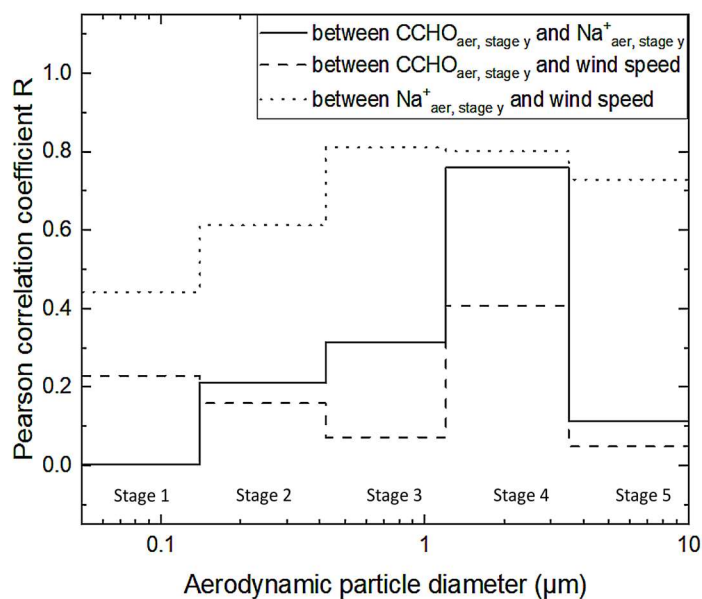


Figure 5. Pearson correlation coefficient R between CCHO_{aer,stage y} and Na⁺_{aer,stage y} (solid line), between CCHO_{aer,stage y} and the average wind speed (dashed line), and between Na⁺_{aer,stage y} and the average wind speed (dotted line) for each stage y of the Berner impactor.

521 Blowing snow has been discussed as a possible additional source for atmospheric Na⁺, raising the
522 question, if it could be a source for atmospheric carbohydrates, too. During this study, the
523 measurements of dFCHO and CCHO in five Arctic snow samples collected resulting in low values mostly
524 below the limits of detection. This finding supports the conclusion that blowing snow does not serve
525 as a competitive source for the emission of atmospheric marine carbohydrates.

526 3.3 Marine combined carbohydrates in fog

527 The concentrations of $\text{Na}^+_{\text{fog, liquid}}$ (1.7–903 mg L^{-1} ; mean = $130 \pm 220 \text{ mg L}^{-1}$; n=22) and $\text{CCHO}_{\text{fog, liquid}}$
528 (18–22000 $\mu\text{g L}^{-1}$; mean = $1380 \pm 4600 \mu\text{g L}^{-1}$; n=22) were very variable in fog water (Table SI 2).
529 Atmospheric concentrations of these chemical constituents in fog droplets (indicated by the index
530 'fog,atmos') can be calculated under consideration of the liquid water content (LWC) during the fog
531 events. Since LWC was not measured during PS106 directly, the LWC was approximated from the
532 measured CCN concentrations at the lowest quality assured supersaturation of 0.15% and an assumed
533 average droplet diameter of 17 μm . This approach resulted in LWCs of $0.62 \pm 0.39 \text{ g m}^{-3}$ for the fog
534 collected over the North Sea and Norwegian Sea, and $0.10 \pm 0.09 \text{ g m}^{-3}$ for the fog over the Arctic Ocean
535 (Hartmann et al., 2021). Following this approach, atmospheric concentrations in fog ranged between
536 0.12 and 150 $\mu\text{g m}^{-3}$ (mean = $25 \pm 43 \mu\text{g m}^{-3}$; n=16) for $\text{Na}^+_{\text{fog, atmos}}$, and between 3 and 4300 ng m^{-3}
537 (mean = $390 \pm 1100 \text{ ng m}^{-3}$; n=16) for $\text{CCHO}_{\text{fog, atmos}}$, respectively. These atmospheric concentrations in
538 fog are for both $\text{Na}^+_{\text{fog, atmos}}$ and $\text{CCHO}_{\text{fog, atmos}}$ by one to three orders of magnitude higher than the
539 atmospheric concentrations in aerosols discussed in section 3.2. This divergence may be explained by
540 the following:

- 541 - Fog scavenging is a transfer process of aerosol particles into the liquid phase of fog droplets
542 (Gilardoni et al., 2014). As fog forms and grows, it can capture aerosol particles in the air and
543 increase their concentration within the fog droplets. This could lead to higher atmospheric
544 concentrations of aerosol particle compounds, especially for the water-soluble and
545 hygroscopic ones, inside the fog compared to the surrounding air.
- 546 - The activation of aerosol particles to fog droplets is a process dominated by particle size with
547 larger particles tending to activate first. It is conceivable that SSA particles larger than 10 μm ,
548 usually few in number, but with a large mass contribution, were available near the sea surface,
549 where sampling occurred. These SSA particles were activated into fog droplets and contributed
550 significantly to the Na^+ and CCHO in the fog. In contrast, aerosol sampling was restricted by
551 the Berner impactor's 10 μm diameter cut-off neglecting the larger particles in the
552 consideration.
- 553 - The LWC values were not measured but estimated, which could be a source of errors. This
554 approach resulted in values representing rather the upper limit of LWC values typically
555 reported for Arctic summer fog (0.001-0.17 g m^{-3} (Kumai, 1973)) or sea fog (0.02-0.1 g m^{-3}
556 (Herrmann et al., 2015)), but appear within a realistic range. Consequently, they are likely not
557 responsible for the large difference between aerosol and fog concentrations of several orders
558 of magnitude.

559 Since both, organic and inorganic constituents, showed higher atmospheric concentrations in
 560 fog/clouds compared to ambient aerosol particles, we conclude that a physical phenomenon, such as
 561 fog scavenging, might explain this observation and not an in-situ formation within the cloud droplets.
 562 Similar to the findings of this study discussing marine CCHO and Na⁺ in Arctic fog, Triesch et al. (2021a)
 563 found strikingly high concentrations of free amino acids (FAA) and Na⁺ in marine clouds compared to
 564 aerosol particles both collected on top of the Mt. Verde on Cape Verde as shown in [Table 1](#).

565 While $dFCHO_{\text{fog}}$ and derivatives, such as anhydrosugars and sugar alcohols, have been readily reported
 566 for fog water with terrestrial and marine background (Dominutti et al., 2022), we here present for the
 567 first time ambient CCHO concentrations in marine fog.

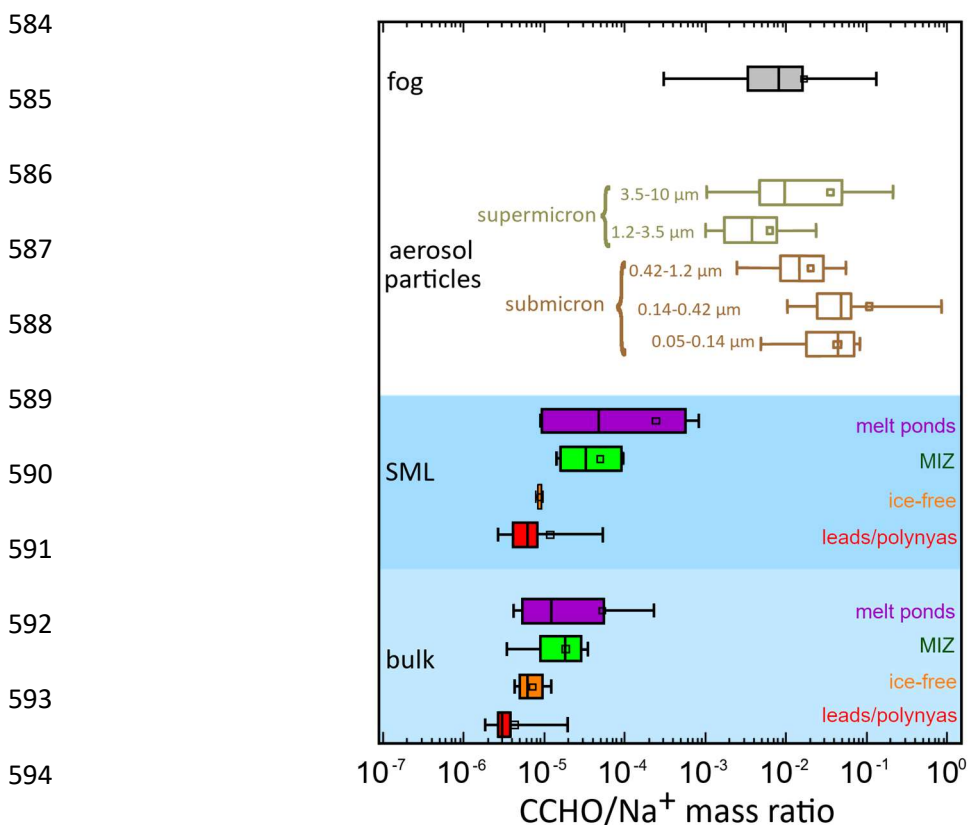
568 **Table 1.** Atmospheric concentrations of selected SSA constituents in fog/clouds compared to ambient aerosol particles during
 569 marine field studies.

Chemical constituent	Fog/cloud	PM ₁₀	Sampling location	Sampling height	Sampling period	Reference
	(ng m ⁻³)	(ng m ⁻³)		(m a.s.l.) ^h		
<i>d</i> FCHO	9.2–52 ^{a,b}	–	Reunion	1760 ^d	March–April 2019	Dominutti et al. (2022)
	1.5–1040 (mean:80±260)	<LOD–2.0	Arctic	25 ^e	May–July 2017	this study
CCHO	3–4300 (mean:390±1100)	0.5–4.7	Arctic	25 ^e	May–July 2017	this study
FAA	11–490	1.0–4.8	Cape Verde	744 ^f	Sept.–Oct. 2017	Triesch et al. (2021a)
	6–79 ^b	–	Reunion	1760 ^d	March–April 2019	Dominutti et al. (2022)
	(μg m ⁻³)	(μg m ⁻³)		(m a.s.l.) ^h		
Na ⁺	1.6–7.2	0.17–0.40	Cape Verde	744 ^f	Sept.–Oct. 2017	Triesch et al. (2021a)
	0.1–2.2 ^b	–	Reunion	1760 ^d	March–April 2019	Dominutti et al. (2022)
	0.014–0.063 ^c	–	Arctic	180–374 ^g	Aug.–Sept. 2018	Zinke et al. (2021)
	0.12–150 (mean:25±43)	0.012–0.77	Arctic	25 m ^e	May–July 2017	this study

570 ^aonly includes free glucose and rhamnose; sugar alcohols and anhydrosugars were not included for this table. ^bvalues were
 571 calculated from LWCs, molecular weights and concentrations in fog water given within the reference; terrestrial contributions
 572 are likely. ^ccalculated from concentration in fog water and an assumed LWC of 0.1 g m⁻³. ^dPiste Omega. ^eRV *Polarstern*. ^fMt.
 573 Verde. ^gtethered balloon, ^h'm a.s.l.' abbreviates 'meters above sea level'.

574 3.4 Chemo-selective sea-air transfer of marine carbohydrates

575 The chemo-selective sea-air transfer of organics towards inorganic sea salt constituents has been
576 described both in tank and ambient field studies for organic carbon in general (Gantt et al., 2011;
577 Hoffman and Duce, 1976; van Pinxteren et al., 2017) or several chemical constituents, such as
578 carbohydrates (Hasenecz et al., 2020; Schill et al., 2018; Zeppenfeld et al., 2021a), lipids (Triesch et al.,
579 2021b) and free and combined amino acids (Triesch et al., 2021a, c). The calculation of dimensionless
580 ratios between the concentrations of the examined organic parameter and Na^+ allows a comparison
581 of aquatic and atmospheric samples within the marine environment. Figure 6 shows the CCHO/Na^+
582 ratios for the bulk and SML in the four sea-ice-related sea surface compartments, size resolved aerosol
583 particles and fog water collected during the PS106 cruise.



594
595 **Figure 6.** CCHO/Na^+ ratios for CCHO in Arctic fog, size-resolved aerosol particles and the surface seawater (SML and bulk)
596 from melt ponds, the marginal ice zone (MIZ), the ice-free ocean and leads/polynyas from the pack ice.

595 **Wide range of CCHO/Na^+ ratios in Arctic surface seawater.** In the surface seawater samples of this
596 study, the CCHO/Na^+ ratios spanned from 2×10^{-6} to 8×10^{-4} , representing a wider range than those
597 found in the Southern Ocean (9×10^{-7} and 3×10^{-5} ; Zeppenfeld et al., 2021). While the ratios in the SML
598 and bulk water in general ranged in the same orders of magnitude, large differences were observed in
599 the individual Arctic sea-ice-related sea surface compartments. In the SML, lowest median values were
600 found in the leads/polynyas and ice-free ocean samples with 6×10^{-6} and 9×10^{-6} , respectively, while
601 higher median values appeared in the SML of the MIZ (3×10^{-5}) and melt ponds (4×10^{-5}), or even

602 6×10^{-4} , when only aged melt ponds were considered. This large variability of CCHO/Na⁺ ratios can be
603 explained by the variable content of CCHO (high CCHO content in aged melt ponds & MIZ versus lower
604 CCHO content in ice-free ocean & leads/polynyas) and Na⁺ (low salinity in the SML of melt ponds versus
605 higher salinities in ice-free ocean & leads/polynyas & MIZ) in the different sea-ice-related sea surface
606 compartments. It can be expected, that the different CCHO/Na⁺ ratios in the individual seawater
607 compartments impacted the corresponding CCHO/Na⁺ ratios in fog and aerosol particles during the
608 sea-air transfer, and consequently the enrichment factors for the sea-air transfer (EF_{aer} , EF_{fog}), which
609 are calculated from those ratios.

610 ***Air mass history influences CCHO_{aer}/Na⁺_{aer} ratios in Arctic aerosol particles.*** In contrast to the
611 seawater samples, CCHO_{aer}/Na⁺_{aer} ratios were much higher for aerosol particles considering the size
612 resolution (1×10^{-3} – 9×10^{-1}) supporting the concept of the chemo-selective enrichment of
613 carbohydrates towards Na⁺ during the transfer from the ocean into the atmosphere. In this context,
614 submicron particles showed much higher median ratios of 4×10^{-2} (0.05–0.14 μm) and 4×10^{-2}
615 (0.14–0.42 μm) than supermicron particles with 4×10^{-3} (1.2–3.5 μm) and 1×10^{-2} (3.5–10 μm).
616 Regarding PM₁₀ (sum of all five Berner stages), the CCHO_{aer,PM10}/Na⁺_{aer,PM10} ratios varied much more in
617 the Arctic study presented here (2×10^{-3} – 2×10^{-1} , see [Table SI 1](#)) than in the ice-free part of the Southern
618 Ocean (8×10^{-4} – 7×10^{-3} ; Zeppenfeld et al. (2021b)).

619 During four aerosol sampling periods (24/05/17–26/05/17; 26/05/17–29/05/17; 29/05/17–01/06/17;
620 19/06/17–25/06/17), air masses had predominantly passed over the ice-free ocean (45–100% of the
621 12 hours prior to sampling, as shown in [Table SI 1](#) & [Figure SI 2](#)). Interestingly, these periods exhibited
622 the lowest CCHO_{aer,PM10}/Na⁺_{aer,PM10} ratios (2×10^{-3} – 9×10^{-3} , detailed in [Table SI 1](#)), values that are
623 strikingly similar to those observed in the ice-free Southern Ocean. In contrast, higher ratios were
624 found, when the air masses had rested a significant time over the pack ice or the MIZ. This could be an
625 indication that the chemical composition of the sea-ice-related sea surface compartments, here the
626 ice-free ocean with low CCHO/Na⁺ ratios, strongly influences the relative composition of aerosol
627 particles. In contrast, the influence of the MIZ, pack ice and melt ponds exhibiting quite different
628 chemical, physical and biological properties on CCHO_{aer,PM10}/Na⁺_{aer,PM10} could not be resolved in further
629 details following this approach using back-trajectory calculations and satellite data. This is certainly
630 due to the proximity of these sea-ice-related sea surface compartments on a small spatial scale
631 (especially melt ponds in direct vicinity to open leads), the long sampling periods of aerosol particles,
632 the lacking knowledge of deposition rates, the effect of wind on wave propagation and bubble bursting
633 processes within the individual sea-ice-related sea surface compartments and missing data on the
634 biological activities in individual melt ponds.

635 **Similar CCHO/Na⁺ ratios in aerosol particles and fog.** For fog, CCHO_{fog}/Na⁺_{fog} ratios ranged from
636 3×10^{-4} to 1×10^{-1} , which covers the same orders of magnitude of aerosol particles. Even though absolute
637 atmospheric concentrations of CCHO are much higher in fog than in aerosol particles possibly due to
638 fog scavenging (as discussed in 3.3), the CCHO/Na⁺ ratios were similar. This strongly implies that
639 CCHO_{fog} actually originated from the ambient marine aerosol particles. The attempt to find matches or
640 common trends between aerosol particles and the fog in individual samples was not successful,
641 certainly due to the very different resolutions of sampling times and in addition due to the probability
642 of fog droplets containing aerosol particles bigger than 10 μm .

643 **Calculated EF_{aer} and EF_{fog} depend on the sea-ice-related marine source under consideration.** EF_{aer} and
644 EF_{fog} are calculated as a quotient between the CCHO/Na⁺ ratios in the size-resolved aerosol
645 particles/fog and the corresponding bulk water. The CCHO/Na⁺ ratios in the Arctic seawater of this
646 study were very variable depending on the regarded sea-ice-related sea surface compartment
647 environment, as well in the aerosol particles and in fog water. This fact strongly impacted the resulting
648 hypothetical EF_{aer} and EF_{fog}, enabling calculated values ranging between 10^1 and 10^4 for supermicron
649 aerosol particles, between 10^2 and 10^5 for submicron particles and between 10^0 and 10^4 for fog
650 depending on which sea-ice-related sea surface compartment was assumed as the marine source of
651 SSA as shown in [Figure 7](#). Due to missing information, including SSA emission fluxes from the four sea-
652 ice-related compartments, aerosol deposition rates, biological activities in melt ponds, wind effects on
653 wave propagation and bubble bursting, and the comparative importance of melt ponds versus open
654 leads (which are in close proximity, making it difficult to resolve them in back-trajectory analyses) as
655 SSA sources – we didn't perform calculations based on the back-trajectory history of each atmospheric
656 sample. Instead, subsequent calculations for EF_{aer} and EF_{fog} employed a hypothetical approach,
657 assessing the range of enrichment factors by considering only one of the four sea-ice-related
658 compartments—represented by the corresponding median CCHO_{bulk}/Na⁺_{bulk} ratios—as the only source,
659 while excluding the others.

660 Lower atmospheric EFs were calculated when aged melt ponds (EF_{aer,super}=19–750; EF_{aer,sub}=127–5100;
661 EF_{fog}=5–2400) or the MIZ (EF_{aer,super}=60–2310; EF_{aer,sub}=390–16000; EF_{fog}=17–7400) were assumed as
662 the only (theoretical) source of CCHO and Na⁺, while higher values were found with the ice-free ocean
663 (EF_{aer,super}=175–6800; EF_{aer,sub}=1100–46000; EF_{fog}=50–22000) or open leads/polynyas
664 (EF_{aer,super}=360–14000; EF_{aer,sub}=2360–95000; EF_{fog}=103–44600). It is important to note that EFs were
665 most consistent with results from other CCHO sea-air transfer studies in the tank (Hasenecz et al.,
666 2020) and the field (Zeppenfeld et al., 2021a), when aged melt ponds or the MIZ were considered as
667 the only emission source. If leads/polynyas and the ice-free ocean were regarded as the only emission
668 source, higher EF_{aer} and EF_{fog} values were obtained, and hence a possible overestimation of the

669 mechanistic process of enrichment. As the results on back-trajectory calculations and sea ice maps
 670 demonstrated (Table SI 1 & Figure SI 2), most air masses were exposed to several of the sea-ice-related
 671 sea surface compartments before sampling. Consequently, none of the Arctic sea-ice-related sea
 672 surface compartments discussed above should be overlooked when discussing of sea-air transfer of
 673 organic substances.

674 During the same Arctic field campaign, Hartmann et al. (2021) investigated INP in ambient aerosol
 675 particles and compared it to bulk and SML in seawater from all the different sea-ice-related sea surface
 676 compartments using similar EF_{aer} calculations as reported here. They concluded that an enrichment of
 677 3 to 5 orders of magnitude was necessary during the sea-air transfer to fully attribute atmospheric INP
 678 to oceanic sources. Here, we show that such high EF_{aer} and EF_{fog} for organics, and hence marine
 679 biogenic INP, can be calculated, e.g. when open leads/polynyas were referred to as the only oceanic
 680 source. In summary, Arctic air masses have been impacted by different types of sea-ice-related sea
 681 surface compartments before sampling, whereas it is still unclear which one has the biggest effect on
 682 the chemical composition of the marine aerosol particles. This aspect should be considered when the
 683 marine SSA constituents are modelled for the Arctic from remote sensing data.

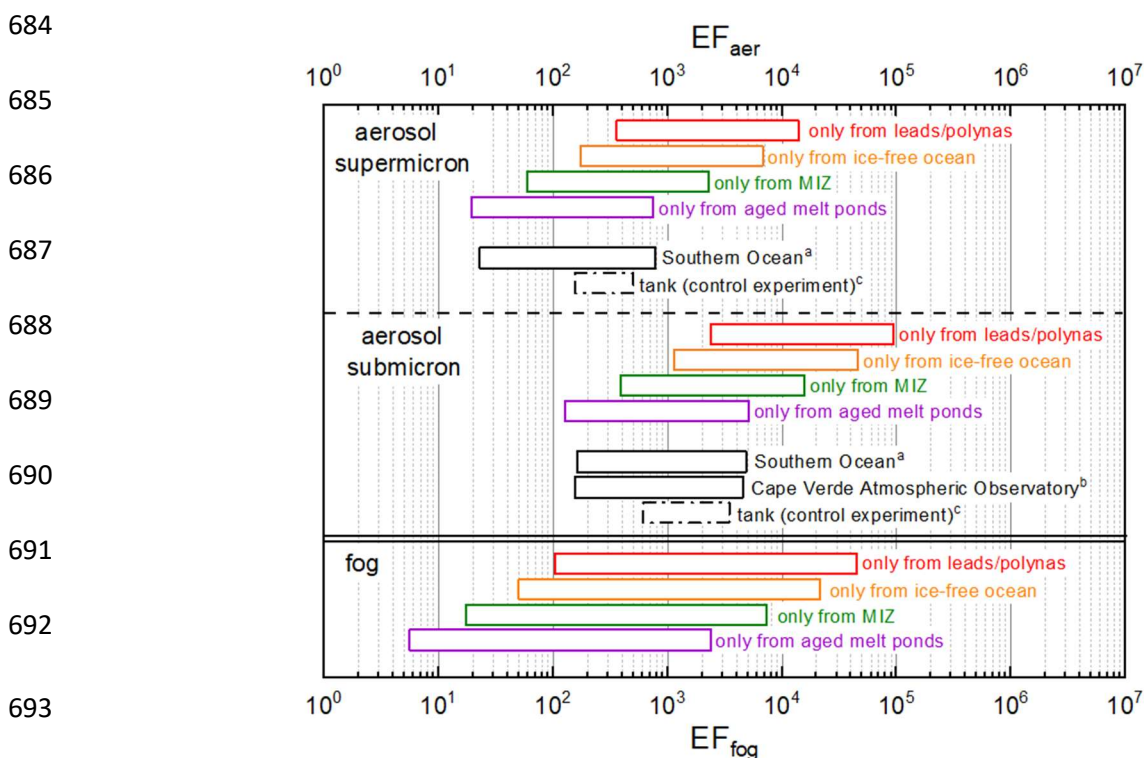


Figure 7. Range of calculated hypothetical enrichment factors EF_{aer} and EF_{fog} assuming either leads/polynyas, the ice-free ocean, the MIZ or aged melt ponds as the only marine source for the sea-air transfer of CCHO in the Arctic. For the calculation of EF_{aer} and EF_{fog} , the minimum and maximum values of the $CCHO_{aer/fog}/Na^+_{aer/fog}$ ratios and the median values of $CCHO_{bulk}/Na^+_{bulk}$ were used. The EF_{aer} values of this study were compared with the results of a) the field study conducted in the Southern Ocean by Zeppenfeld et al. (2021), b) the field study conducted at Cape Verde Atmospheric Observatory (CVAO) by van Pinxteren et al. (2023) and c) the results of the CCHO tank study by Hasenecz et al. (2020) without any addition of heterotrophic bacteria (control experiment). Here, EF_{aer} values were calculated from the experimental data published by Hasenecz et al. (2020b).

694 **3.5 Atmospheric aging of marine carbohydrates**

695 To resolve the fate of marine carbohydrates in the atmosphere after their ejection from the ocean, the
696 relative molar contributions of monosaccharides to CCHO were compared between the bulk and SML
697 from the leads/polynyas, MIZ, ice-free ocean and melt pond samples, as well as the sub-and
698 supermicron aerosol particles and fog water (Figure 8). The composition of marine carbohydrates in
699 seawater strongly depends on the dominating microbial species, season, diagenetic state, availability
700 of nutrients and environmental stress factors (Engbrodt, 2001; Goldberg et al., 2011) leading to a
701 natural variability among individual samples even within small spatial scales. Consequently, to enable
702 the direct comparison of seawater with atmospheric samples of this field study with an elevated level
703 of statistical certainty, here we compare the mean values of the entire data set, instead of individual
704 samples. Finally, in addition to the changes of the monosaccharide patterns of CCHO, the systematic
705 degradation of CCHO to *d*FCHO was observed in the atmosphere and will be discussed within this
706 section.

707 ***CCHO composition in different sea-ice-related sea surface compartments and depths is similar.*** In
708 seawater (bulk and SML), glucose (means= 35–48 mol%), galactose (means= 13–18 mol%) and xylose
709 (means= 7–16 mol%) dominated the CCHO composition followed by smaller contributions of other
710 neutral sugars, amino sugars, uronic acids and muramic acid (Figure 8). Considering the natural
711 variability among individual samples, there were no significant differences in means between the bulk
712 and SML, nor between the lead/polynya, MIZ, ice-free ocean and melt pond samples. Variations were
713 observed between the dissolved and particulate fractions (Figure SI 3), nevertheless the combined
714 carbohydrates within all sea-ice-related sea surface compartments followed the same pattern of the
715 predominance of glucose, galactose and xylose. Overall, the relative monosaccharide compositions of
716 glucose > (galactose ≈ xylose) > other (neutral or charged) monosaccharides of the seawater samples
717 from this Arctic study appear similar to the monosaccharide compositions investigated in the SML and
718 bulk water from the Central Arctic Ocean (Gao et al., 2012) and at the western Antarctic peninsula
719 (Zeppenfeld et al., 2021a), the meltwater of Arctic multiyear sea ice (Amon et al., 2001) and the
720 epipelagic water from the Ross Sea (Kirchman et al., 2001).

721 ***Less galactose, but more muramic acid in atmospheric CCHO_{aer} and CCHO_{fog}.*** Atmospheric samples
722 showed a different monosaccharide pattern within the hydrolyzed CCHO in comparison to the
723 seawater and melt pond samples. While glucose (means= 41 mol% for fog; 50 mol% for submicron and
724 60 mol% for supermicron aerosol particles) and xylose (means= 16; 15 and 15 mol%) still prevailed
725 over the relative monosaccharide pattern, the contribution of galactose (means= 6; 3 and 3 mol%) was
726 strongly reduced, both in fog and aerosol particles. On the other hand, the ratio of muramic acid was

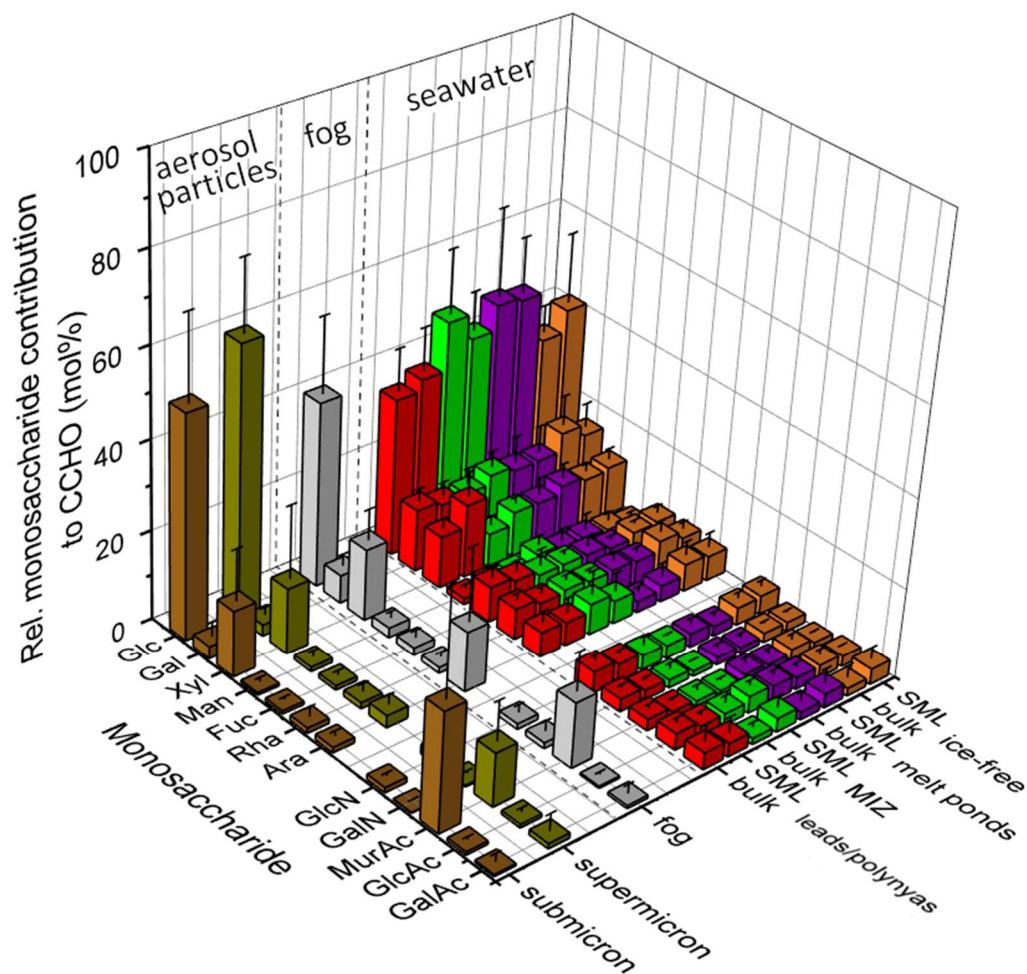


Figure 8. Relative monosaccharide composition of combined carbohydrates (CCHO) after acid hydrolysis in sub-/ supermicron aerosol particles, fog water, bulk and SML samples from the leads and polynyas within the pack ice, the MIZ, the ice-free ocean and young and aged melt ponds. The 3D bar chart shows the averages and standard deviations of the relative contributions. Glc: glucose, Gal: galactose, Xyl: xylose, Man: mannose, Fuc: fucose, Ara: arabinose, GlcN: glucosamine, GalN: galactosamine, MurAc: muramic acid, GlcAc: glucuronic acid, GalAc: galacturonic acid.

727 strongly elevated in aerosol particles (means= 12 and 26 mol%) and fog water (mean= 14 mol%) in
 728 comparison to the oceanic samples (means= 0.9–2.6 mol%). These differences of the relative
 729 monosaccharide contributions to CCHO among the seawater and the atmospheric samples described
 730 within this study are in good agreement with the sea-air transfer investigations conducted in the
 731 Southern Ocean at the western Antarctic peninsula (Zeppenfeld et al., 2021a). Consequently, the
 732 occurring phenomenon might be independent from the sampling location and could be explained by
 733 three possible atmospheric processes, such as (1) a chemo-selective sea-air transfer of certain oligo-
 734 or polysaccharides over others, (2) an atmospheric transformation due to abiotic chemical reactions
 735 or (3) an atmospheric transformation due to microbiological activities. Among these possible
 736 pathways, Zeppenfeld et al. (2021) presumed the secondary atmospheric transformation caused by
 737 microbiological metabolism as the most probable or at least most dominant one supported by the
 738 prevalence of muramic acid, an amino sugar acid naturally occurring in bacterial cell walls (Mimura

739 and Romano, 1985; Sud and Tyler, 1964), and the very selective absence of certain monosaccharides
740 in the CCHO_{aer} in aerosol particles as it was observed in this Arctic study as well.

741 **Formation of combined arabinose in fog.** A comparison of the monosaccharide composition of aerosol
742 particles and fog water showed great similarity regarding dominant contributions from glucose, xylose
743 and muramic acid. It seems plausible that the fog water droplets contained the same inorganic and
744 organic compounds found in the SSA particles assuming that SSA particles activated the formation of
745 fog droplets as CCN due to their rather large diameters and high hygroscopicity. Apart from that,
746 however, a significant difference was observed in the increased relative contribution of arabinose in
747 fog (mean= 13 mol%) compared to aerosol particles (means= 1.2 and 2.7 mol%) indicating a formation
748 of arabinose in the liquid phase. During a marine microcosm experiment performed by Hasenecz et al.
749 (2020), a strong link was observed between the release of arabinose-containing polysaccharides in
750 form of EPS and the presence of heterotrophic bacteria and stressed phytoplankton. Furthermore, a
751 strain of the psychrotolerant marine bacterium *Pseudoalteromonas* sp. has been shown to produce
752 EPS mainly composed from glucose, arabinose and xylose (Casillo et al., 2018; Qin et al., 2007).
753 Consequently, the release of arabinose-containing EPS in fog could be a plausible protection
754 mechanism of microorganisms contained within a droplet against freezing damage under low Arctic
755 temperatures.

756 **Indication for microbial activities in the atmosphere.** Intact bacterial cells at atmospheric
757 concentrations between 5×10^2 and 8×10^4 cells m^{-3} for remote marine and ice-covered regions (Šantl-
758 Temkiv et al., 2018; Mayol et al., 2017), cell-bound and free enzymes have been detected in ambient
759 and nascent marine super- and submicron aerosol particles during several field and tank studies (Aller
760 et al., 2005; Hasenecz et al., 2020; Malfatti et al., 2019; Marks et al., 2001; Rastelli et al., 2017; Šantl-
761 Temkiv et al., 2020; Uetake et al., 2020). For surviving in this hostile environment, some of these
762 microbes have developed a remarkable resilience towards extreme environmental stressors, such as
763 high UV radiation, radical exposure, changing osmolarity, freezing temperatures and desiccation. As
764 survival strategies could serve the selective enzymatic consumption of airborne labile carbohydrates
765 explaining the here observed loss of galactose and the persistence of xylose, the release of protecting
766 biofilms from EPS, carotenoid pigmentation or the formation of own precipitating hydrometeors by
767 enabling condensation on a surface as a CCN or freezing by IN active surfaces to reduce their
768 atmospheric residence time (Delort et al., 2010; Matulová et al., 2014; Šantl-Temkiv et al., 2020).
769 Consequently, an enzymatic transformation might serve as a plausible explanation for the selective
770 removal of certain monosaccharides within CCHO_{aer} and CCHO_{fog} observed here. However, the survival
771 and the metabolic activity of microorganisms is restricted by the presence of water (Ervens and Amato,
772 2020; Haddrell and Thomas, 2017) identifying liquid hydrometeors or fresh SSA as the most biologically

773 active atmospheric hotspots. In contrast to most of the ambient aerosol particles, fog droplets provide
774 enough water essential for bacterial activities. However, they might freeze under Arctic sub-zero
775 temperatures possibly causing damage to the microbial cells, which might explain an in-situ formation
776 of a protecting biofilm from arabinose-containing EPS. In a previous Arctic study, Orellana et al. (2011)
777 readily detected microgels in aerosol particles, cloud and fog water most likely emitted from the
778 surface water and the SML via bubble bursting. Indications for an in-situ generation of marine
779 microgels in fog water as an additional source to the primary release from the ocean by bubble bursting
780 have been observed by van Pinxteren et al. (2022) in the tropical Atlantic Ocean.

781 The selective sea-air transfer of certain carbohydrates over others and the abiotic degradation as
782 further possible pathways to the biotic transformation of marine CCHO_{aer} have been discussed in detail
783 in Zeppenfeld et al. (2021), but do not appear, based on the current state of knowledge, as likely
784 explanations of the very selective CCHO degradation and formation of other CCHO observed here.
785 More future lab and mesocosm experiments are required to elucidate the contribution of each of these
786 processes. Finally, the similarity between the carbohydrate compositions of fog water and aerosol
787 particles, both two atmospheric compartments collected with different instrumentation, allows to rule
788 out artefacts of the different sampling and extraction techniques as a reason for the observed
789 differences to the seawater.

790 ***Depolymerization of CCHO to dFCHO, seawater versus atmosphere.*** Free glucose, by far the most
791 prevailing monosaccharide among dFCHO in seawater, ranged between 0.6 and 51 $\mu\text{g L}^{-1}$ during the
792 PS106 cruise in the bulk and the SML (Zeppenfeld et al., 2019a). Thus, dFCHO/CHO ratios, meaning the
793 contribution of sugar monomers to all marine carbohydrates measured in this study, varied between
794 1–14% with an average of $5\pm 3\%$. Conversely, 86–99% (mean: 95 ± 3) of carbohydrates in the bulk and
795 SML of ocean seawater and melt ponds were incorporated into an oligo- or polysaccharidic structure.
796 CCHO can be hydrolyzed to dFCHO either in an acidic environment or enzymatically by heterotrophic
797 bacteria (Arnosti, 2000; Panagiotopoulos and Sempéré, 2005). Seawater from the Arctic Ocean is
798 slightly alkaline with reported pH values between 7.98 and 8.49 (Rérolle et al., 2016; Tynan et al.,
799 2016), while the pH of melt pond water has been observed to be more variable from mildly acidic (6.1)
800 to more alkaline (10.8) (Bates et al., 2014). In agreement with previous findings, the oceanic surface
801 seawater (pH: 7.98–8.66), including the samples from the MIZ, ice-free ocean and open
802 leads/polynyas, and the melt pond samples (pH: 7.26–8.62) were slightly alkaline in this study.
803 Consequently, it is more plausible that the depolymerization of CCHO in seawater can be ascribed to
804 bacterial activities rather than acid hydrolysis. Since dFCHO are readily resorbed by heterotrophic
805 bacteria with high turnover rates (Ittekkot et al., 1981; Kirchman et al., 2001), concentrations of these
806 monosaccharides are rather low in seawater.

807 In contrast, in aerosol particles, higher $d\text{FCHO}/\text{CHO}$ ratios up to 35% occurred in some selected
808 samples, which is much higher than in seawater, suggesting that CCHO might be depolymerized in the
809 atmosphere. SSA particles are known to significantly acidify within minutes after their release due to
810 the uptake of acidic gases, atmospheric aging reactions with sulfuric dioxide and water loss (Angle et
811 al., 2021). In this context, the surface-to-volume ratio determines the efficiency of the acidification
812 effect, which means that it is most pronounced for submicron SSA particles with reported pH values
813 of 1.5–2.6 within a few minutes in a tank study (Angle et al., 2021), and less pronounced for
814 supermicron SSA particles or cloud droplets (Angle et al., 2022). Consequently, it is conceivable that
815 an acid hydrolysis of CCHO_{aer} to monomeric $d\text{FCHO}_{\text{aer}}$ occurs at the surface or within the bulk of SSA
816 particles leading to quick atmospheric aging. However, due to analytical constraints, such as the limits
817 of detections (LODs) of the methodology, the $d\text{FCHO}$ in size-resolved aerosol particles could not be
818 detected in all samples and the data availability is not strong enough to draw more conclusions for
819 aerosol particles.

820 In fog, where LODs did not represent an issue due to the high concentrations, $d\text{FCHO}/\text{CHO}$ ratios
821 exceeding those in seawater were also observed, ranging from 1-60% (mean: $27\pm 16\%$). The
822 monosaccharide composition of $d\text{FCHO}_{\text{fog}}$ was primarily made up of glucose, arabinose, fructose, and
823 xylose, with minor contributions from glucosamine, galactose, mannose, rhamnose, and fucose. While
824 $d\text{FCHO}_{\text{fog}}$ and CCHO_{fog} shared similar dominant monosaccharides, fructose was only present in
825 $d\text{FCHO}_{\text{fog}}$. This absence in CCHO_{fog} is attributed to fructose's low stability during the analytical
826 preparation for CCHO analysis (Panagiotopoulos and Sempéré, 2005). As a result, fructose won't be
827 further discussed. In this study, pH values of fog water ranged between 5.7 and 6.8, which is 1–2
828 magnitudes more acidic than in seawater. Polysaccharides are known to depolymerize due to acid
829 hydrolysis, especially at elevated temperatures. The pH-stability can be largely variable among the
830 different polysaccharides; however, we are not aware of studies that have shown such fast
831 depolymerizations, in the sense of time scales relevant for atmospheric lifetime of aerosol particles, at
832 such mildly acid conditions and low temperatures as those of the Arctic atmosphere. Furthermore,
833 there was no significant correlation between the pH and the $d\text{FCHO}/\text{CHO}$ of these cloud samples.
834 Consequently, there are no indications that the majority of CCHO was hydrolyzed inside the cloud
835 droplets, however it might be conceivable that hydrolysis had readily occurred within the non-
836 activated SSA particle where pH values were much lower.

837 Besides an acid hydrolysis induced by quick atmospheric acidification of SSA particles, atmospheric
838 radicals, such as OH (Trueblood et al., 2019), or photolytic cleavages of glycosidic bonds (Kubota et al.,
839 1976) could have contributed to the degradation of atmospheric CCHO to monomeric $d\text{FCHO}$ in SSA
840 and marine fog. For these processes, however, still hardly any systematic lab studies have been

841 conducted for the plurality of marine polysaccharides, which makes a classification of the meaning of
842 these processes difficult. A preferred sea-air transfer of *d*FCHO over CCHO to explain this observation
843 seems unlikely based on the missing enrichment of neutral *d*FCHO in contrast to the high EF_{aer} of CCHO
844 shown in tank studies (Hasenecz et al., 2019, 2020). Finally, a microbial depolymerization of CCHO by
845 extracellular enzymes in fog cannot be entirely ruled out considering that the activity of some
846 polysaccharide-degrading enzymes, such as α - and β -glucosidase, have been found to accelerate in
847 seawater with increasing acidity (Piontek et al., 2010). However, this finding was conducted for a pH
848 range only 0.3 pH units lower than the typical pH of seawater and it is not sure, if this finding can be
849 transferred to the more acid conditions in aerosol particles and fog water.

850 ***Several aging processes in the atmosphere.*** We observed significant changes between the chemical
851 composition of marine carbohydrates in the surface seawater, including the bulk and SML, and
852 atmospheric carbohydrates, including aerosol particles and fog. Based on the changing
853 monosaccharide composition pattern of CCHO with selective degradation and formation of specific
854 monosaccharides within CCHO, we conclude microbial or enzymatic activities within the aerosol
855 particles of fog droplets. Furthermore, the increasing contribution of *d*FCHO to the total carbohydrate
856 pool in fog and aerosol particles might be attributed to a hydrolytic cleavage of the glycosidic linkages
857 between monosaccharide units within the oligo- and polysaccharides after a quick atmospheric
858 acidification of SSA particles. Consequently, atmospheric carbohydrates experience quick atmospheric
859 aging, potentially due to both biological and abiotic processes, after their release from the ocean.
860 Possibly, this could affect the CCN and INP properties of marine carbohydrates and hence the
861 formation and properties of clouds.

862 **3.6 Perspective assessment of CCHO via bio-optical parameters**

863 The absorption of phytoplankton (a_{ph}) and CDOM (a_{CDOM}) are bio-optical parameters providing
864 additional information about the chemical and microbiological history of the water masses within the
865 particulate and dissolved phase, respectively. They can be measured on discrete water samples and
866 can also be assessed as products from satellites (Lefering et al., 2017; Matsuoka et al., 2012, 2013;
867 Röttgers et al., 2016). Here we tested, if a_{ph} or CDOM parameters correlate with CCHO in seawater to
868 potentially enable the remote-sensing approximation of marine CCHO in seawater and potentially in
869 the atmosphere.

870 **Good assessment of CCHO in seawater via a_{ph440} .** a_{ph440} derived from the phytoplankton absorption
871 spectrum is directly related to the biomarker TChl-*a* indicating phytoplankton biomass (Bricaud et al.,
872 2004; Phongphattarawat, 2016). The advantage of using a_{ph440} over pigment data, including TChl-*a*
873 from full high-performance liquid chromatography (HPLC) analysis (e.g. Barlow et al., 1997; Taylor et
874 al., 2011), is the lower need of sample volume for the analysis. This allows the determination of values
875 in the SML samples as well (Zäncker et al., 2017), which are laborious to collect and therefore limited
876 in availability. In this study, a_{ph440} strongly correlated with $pCCHO$ ($R=0.90$, $p<0.001$) in bulk and SML
877 samples (Figure 9a) showing a direct link with fresh phytoplankton biomass production. A similar link
878 has been described before for TChl-*a* and $pCCHO$ in the photic layer of the Ross Sea (Fabiano et al.,
879 1993), in the ocean west of the Antarctic peninsula (Zeppenfeld et al., 2021a) and between TChl-*a* and
880 the particulate form of laminarin, an algal polysaccharide, in Arctic and Atlantic water samples (Becker
881 et al., 2020). $dCCHO$ showed a good, but weaker correlation with a_{ph440} ($R=0.66$, $p<0.001$) than
882 $pCCHO$. This finding supports the assumption that $pCCHO$ are rather freshly produced by local
883 autotrophs, while the link between $dCCHO$ with their primary production was already obscured by
884 subsequent transformation processes resulting in a more recalcitrant, long-lived mix of
885 macromolecules (Goldberg et al., 2011; Hansell, 2013; Keene et al., 2017). Nevertheless, CCHO, the
886 sum from $dCCHO$ and $pCCHO$, showed a high correlation with a_{ph440} ($R=0.84$, $p<0.001$, Figure 9b)
887 leading to the conclusion that this bio-optical parameter derived from the $a_{ph}(\lambda)$ spectrum is suitable
888 to assess the total amount of CCHO in the surface seawater of the different sea-ice-related sea surface
889 compartments of the Arctic.

890 **Good assessment of CCHO in seawater via $a_{CDOM350}$.** In this study, high correlations were observed
891 between $dCCHO$ and $a_{CDOM350}$ ($R=0.66$, $p<0.001$, Figure 10a), and weaker correlations between $dCCHO$
892 and $a_{CDOM443}$ ($R=0.53$, $p<0.001$, Figure 10b). The better correlation at $\lambda=350$ nm compared to 443 nm
893 can be explained by the fact that a_{CDOM} exponentially decreases with wavelength. While absorption by
894 CDOM is higher at $\lambda=350$ nm, it is much closer to the method detection limit at $\lambda=443$ nm and is

895 therefore more error-prone. However, with current satellite products only a_{CDOM} at 440 nm can be
896 retrieved.

897 Previous studies reported strong correlations between $a_{CDOM350}$ and dissolved organic carbon (DOC)
898 in Arctic seawater (Gonçalves-Araujo et al., 2015; Spencer et al., 2009; Stedmon et al., 2011; Walker
899 et al., 2013). Consequently, it is conceivable that $dCCHO$, an important constituent of DOC, shows good
900 correlations as well. Surprisingly, the correlation between CCHO (sum of $dCCHO$ and $pCCHO$) and
901 $a_{CDOM350}$ was strongest ($R=0.85$, $p<0.001$, **Figure 10c**), indicating that CDOM retrieval from high-
902 resolution satellite data could allow a good approximation of CCHO in Arctic seawater.

903

904

905

906

907

908

909

910

911

912

913

914

915

916

917

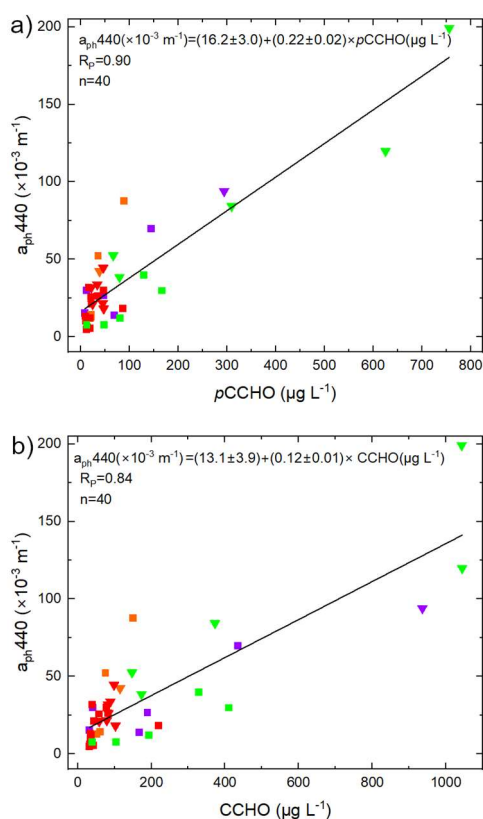


Figure 9. Correlation plots of a_{ph440} derived from PAB spectra against a) $pCCHO$ and b) CCHO. Triangles: SML, squares: bulk. Green: marginal ice zone (MIZ), purple: melt ponds, orange: ice-free ocean, red: open leads/polynyas in the pack ice.

918

919

920

921

922

923

924
 925
 926
 927
 928
 929
 930
 931
 932
 933
 934
 935
 936
 937
 938
 939
 940
 941

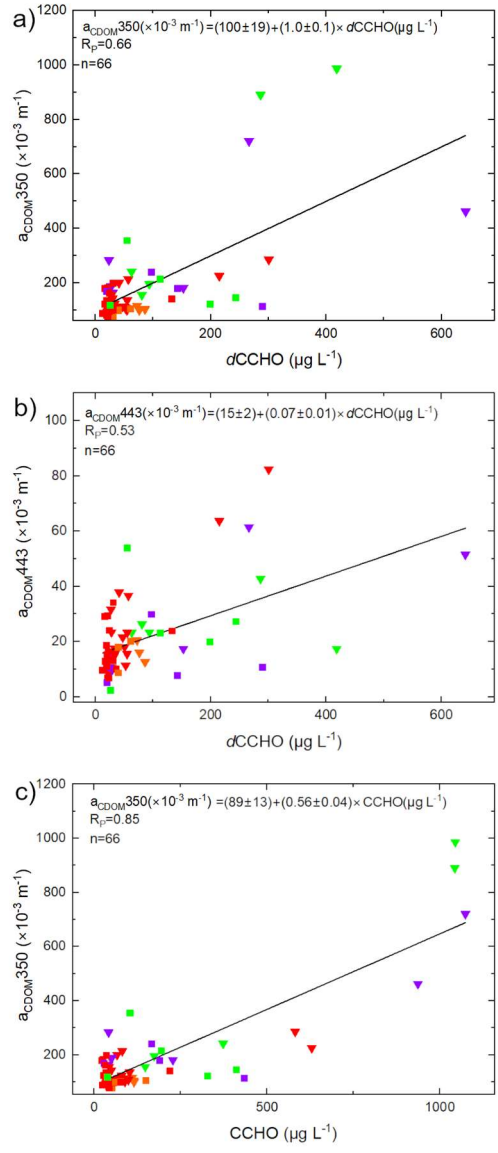


Figure 10. Correlation plots of a) $a_{CDOM350}$ against $dCCHO$, b) $a_{CDOM443}$ against $dCCHO$ and c) $a_{CDOM350}$ against $CCHO$. Triangles: SML, squares: bulk. Green: marginal ice zone (MIZ), purple: melt ponds, orange: ice-free ocean, red: open leads/polynyas in the pack ice.

942 **4. Summary and Atmospheric Implications**

943 We studied the sea-air transfer of marine carbohydrates from field samples collected in the Arctic
944 during the PS106 campaign from May to July 2017. Large differences of absolute CCHO concentrations
945 and SML enrichments were observed among the different sea-ice-related sea surface compartments
946 (leads/polynyas within the pack ice, ice-free ocean, MIZ, melt ponds). CCHO_{aer} were detected in the
947 sub- and supermicron aerosol particles with indications for primary emissions from the sea through
948 bubble bursting, though the correlations with the SSA tracer Na⁺ and wind speed were possibly
949 reduced due to the presence of sea ice influencing the wind-induced SSA emission mechanisms.
950 Atmospheric CCHO and Na⁺ concentrations in fog strongly exceeded those of the aerosol particles likely
951 due to a physical phenomenon. Large enrichments of CCHO in aerosol and fog compared to bulk
952 seawater were observed. The extent of these enrichments varied based on the type of sea-ice-related
953 sea surface compartment assumed as the oceanic source for atmospheric CCHO. We observed a
954 subsequent atmospheric aging of CCHO in the atmosphere, both in aerosol particles and fog, noticed
955 by the selective loss and formation of certain monosaccharide units within CCHO suggesting selective
956 enzymatic/microbial activities, and a depolymerization of CCHO to *d*FCHO, most measurable in fog
957 water and likely due to abiotic degradation, e.g. acid hydrolysis. CCHO correlated well with bio-optical
958 parameters, such as a_{ph440} from phytoplankton absorption and $a_{CDOM350}$. These parameters can be
959 measured via remote sensing and may allow the retrieval of CCHO from satellite data, which
960 potentially will enable an accurate modelling of atmospheric CCHO concentrations as soon as all
961 emission and atmospheric aging processes are sufficiently understood. This study shows that the Arctic
962 is a complex environment, where the diversity of sea-ice-related sea surface compartments needs to
963 be considered as primary sources of marine CCHO or other organic compounds, and where these
964 molecules can be transformed after their primary sea-air transfer by biological and abiotic processes
965 in the atmosphere.

966 Marine carbohydrates are assumed to impact cloud properties by acting as CCN and INP (Alpert et al.
967 2022; Leck et al., 2013; Orellana et al., 2011; van Pinxteren et al., 2022). Studying the chemical identity
968 of those atmospheric nucleation particles, their emission mechanisms and their transformation due to
969 atmospheric aging can strongly improve the understanding of the cloud formation in the Arctic, cloud
970 microphysical properties, the radiation budget, cryosphere-ocean-atmosphere interactions and
971 eventually feedback mechanisms in the frame of Arctic amplification. It can be assumed that within
972 the warming Arctic, where sea ice extent is continuously shrinking, the MIZ area will expand (Strong
973 and Rigor, 2013) and the number of biologically-active melt ponds will increase during the summer
974 season in the next years. These new MIZ regions and melt ponds could potentially produce more
975 marine carbohydrates than the ice-free ocean or open leads within the pack ice leading to enhanced

976 CCN and INP populations in the Arctic atmosphere serving as a still not well-explored feedback
977 mechanism within Arctic amplification.

978 **Data availability.** All data are available on the public repository PANGAEA:
979 <https://doi.org/10.1594/PANGAEA.962208> and <https://doi.org/10.1594/PANGAEA.932573> (for fog
980 samples); <https://doi.org/10.1594/PANGAEA.962210> and <https://doi.org/10.1594/PANGAEA.932569>
981 (for aerosol particles); <https://doi.org/10.1594/PANGAEA.961004> (for seawater samples).

982 **Author contribution.** SZ wrote the manuscript with contributions from MvP, MH, MZ, AB and HH. SZ,
983 MvP and MH collected the field samples during the PS106 campaign. SZ performed the laboratory
984 carbohydrate analysis and statistical evaluation. MZ and AB assessed the bio-optical parameters. All
985 co-authors proofread and commented the manuscript.

986 **Competing interests.** The authors declare that they have no conflict of interest.

987 **Acknowledgements.** We gratefully acknowledge the funding by the Deutsche Forschungsgemeinschaft
988 (DFG, German Research Foundation, Projektnummer 268020496–TRR 172) within the Transregional
989 Collaborative Research Center “Arctic Amplification: Climate Relevant Atmospheric and Surface
990 Processes, and Feedback Mechanisms (AC)³”. This research has been supported by the DFG SPP 1158,
991 grant number 424326801 by enabling the access to melt pond data. We thank Andreas Macke and
992 Hauke Flores, chief scientists for the RV *Polarstern* cruises PS106.1 and PS106.2 (expedition grant
993 number AWI_PS106_00), and the captain and the crew of RV *Polarstern* for their support during the
994 expedition from May to July 2017. We thank Andrea Haudek and Hartmut Haudek for the development
995 and construction of the conditioning tube and the wind control system connected to the Berner
996 impactor. We thank Anett Dietze, Susanne Fuchs and Anke Rödger for the mass, inorganic ion and
997 OC/EC measurements. We acknowledge René Rabe and Sonja Wiegmann for supporting the
998 preparation of PS106 chemical equipment and optical instrumentation, respectively, and Yangyang Liu
999 for introducing the optical measurement procedure before PS106.

1000 **Financial support.** This research has been supported by the Deutsche Forschungsgemeinschaft (DFG,
1001 German Research Foundation, Projektnummer 268020496–TRR 172) within the Transregional
1002 Collaborative Research Center “Arctic Amplification: Climate Relevant Atmospheric and Surface
1003 Processes, and Feedback Mechanisms (AC)³” in subprojects B04 and C03.

1004 **List of abbreviations**

1005	a_{CDOM}	absorption coefficient by colored dissolved organic carbon
1006	aer	aerosol particles
1007	a_{NAP}	absorption coefficient by non-algal particles
1008	ANOVA	Analysis of Variance
1009	a_{p}	absorption coefficient by total particles
1010	a_{ph}	absorption coefficient by phytoplankton

1011	Ara	arabinose
1012	atmos	atmospheric concentrations
1013	C-CCHO	carbon contained within the combined carbohydrate
1014	CCHO	combined carbohydrates
1015	CCN	cloud condensation nuclei
1016	CDOM	colored dissolved organic matter
1017	CHO	carbohydrates
1018	<i>d</i> CCHO	dissolved combined carbohydrates
1019	<i>d</i> FCHO	dissolved free carbohydrates
1020	EF	enrichment factor
1021	EPS	exopolymeric substances
1022	ERDDAP	Environmental Research Division's Data Access Program
1023	FAA	free amino acids
1024	Fru	fructose
1025	Fuc	fucoese
1026	Gal	galactose
1027	GalN	galactosamine
1028	GalAc	galacturonic acid
1029	Glc	glucose
1030	GlcAc	glucuronic acid
1031	GlcN	glucosamine
1032	HPAEC-PAD	high-performance anion-exchange chromatography with pulsed amperometric detection
1033	HPLC	high-performance liquid chromatography
1034	INP	ice nucleating particles
1035	LWCC	liquid waveguide capillary cell
1036	Man	mannose
1037	MIZ	marginal ice zone
1038	MurAc	muramic acid
1039	Na ⁺	sodium ion
1040	NOAA	National Oceanic and Atmospheric Administration
1041	OC	organic carbon
1042	OM	organic matter
1043	PAB	particulate absorption
1044	<i>p</i> CCHO	particulate combined carbohydrates
1045	PM	particulate matter
1046	Rha	rhamnose
1047	SML	sea surface microlayer
1048	SSA	sea spray aerosol
1049	sub	submicron
1050	super	supermicron
1051	TChl- <i>a</i>	total chlorophyll <i>a</i>
1052	TEP	transparent exopolymer particles
1053	QFT-ICAM	quantitative filtration technique with an integrative-cavity absorption meter setup
1054	Xyl	xylose

1055 References

- 1056 Alderkamp, A.-C., Buma, A. G. J., and van Rijssel, M.: The carbohydrates of *Phaeocystis* and their degradation in the microbial
1057 food web, *Biogeochemistry*, 83, 99–118, <https://doi.org/10.1007/s10533-007-9078-2>, 2007.
- 1058 Aller, J. Y., Kuznetsova, M. R., Jahns, C. J., and Kemp, P. F.: The sea surface microlayer as a source of viral and bacterial
1059 enrichment in marine aerosols, *Journal of Aerosol Science*, 36, 801–812, <https://doi.org/10.1016/j.jaerosci.2004.10.012>,
1060 2005.
- 1061 Aller, J. Y., Radway, J. C., Kilhau, W. P., Bothe, D. W., Wilson, T. W., Vaillancourt, R. D., Quinn, P. K., Coffman, D. J., Murray,
1062 B. J., and Knopf, D. A.: Size-resolved characterization of the polysaccharidic and proteinaceous components of sea spray
1063 aerosol, *Atmospheric Environment*, 154, 331–347, <https://doi.org/10.1016/j.atmosenv.2017.01.053>, 2017.
- 1064 Alpert, P. A., Kilhau, W. P., O'Brien, R. E., Moffet, R. C., Gilles, M. K., Wang, B., Laskin, A., Aller, J. Y., and Knopf, D. A.: Ice-
1065 nucleating agents in sea spray aerosol identified and quantified with a holistic multimodal freezing model, *Science Advances*,
1066 8, eabq6842, <https://doi.org/10.1126/sciadv.abq6842>, 2022.
- 1067 Álvarez, E., Losa, S. N., Bracher, A., Thoms, S., and Völker, C.: Phytoplankton Light Absorption Impacted by Photoprotective
1068 Carotenoids in a Global Ocean Spectrally-Resolved Biogeochemistry Model, *Journal of Advances in Modeling Earth Systems*,
1069 14, e2022MS003126, <https://doi.org/10.1029/2022MS003126>, 2022.
- 1070 Amon, R. M. W., Fitznar, H.-P., and Benner, R.: Linkages among the bioreactivity, chemical composition, and diagenetic state
1071 of marine dissolved organic matter, *Limnology and Oceanography*, 46, 287–297, 2001.
- 1072 Angle, K., Grassian, V. H., and Ault, A. P.: The rapid acidification of sea spray aerosols, *Physics today*, 75,
1073 <https://doi.org/10.1063/PT.3.4926>, 2022.
- 1074 Angle, K. J., Crocker, D. R., Simpson, R. M. C., Mayer, K. J., Garofalo, L. A., Moore, A. N., Garcia, S. L. M., Or, V. W., Srinivasan,
1075 S., Farhan, M., Sauer, J. S., Lee, C., Pothier, M. A., Farmer, D. K., Martz, T. R., Bertram, T. H., Cappa, C. D., Prather, K. A., and
1076 Grassian, V. H.: Acidity across the interface from the ocean surface to sea spray aerosol, *PNAS*, 118,
1077 <https://doi.org/10.1073/pnas.2018397118>, 2021.
- 1078 Arnosti, C.: Substrate specificity in polysaccharide hydrolysis: Contrasts between bottom water and sediments, *Limnology
1079 and Oceanography*, 45, 1112–1119, <https://doi.org/10.4319/lo.2000.45.5.1112>, 2000.
- 1080 Aslam, S. N., Michel, C., Niemi, A., and Underwood, G. J. C.: Patterns and drivers of carbohydrate budgets in ice algal
1081 assemblages from first year Arctic sea ice, *Limnology and Oceanography*, 61, 919–937, <https://doi.org/10.1002/lno.10260>,
1082 2016.
- 1083 Azetsu-Scott, K. and Passow, U.: Ascending marine particles: Significance of transparent exopolymer particles (TEP) in the
1084 upper ocean, *Limnology and Oceanography*, 49, 741–748, <https://doi.org/10.4319/lo.2004.49.3.0741>, 2004.
- 1085 Barlow, R., Cummings, D., and Gibb, S.: Improved resolution of mono- and divinyl chlorophylls a and b and zeaxanthin and
1086 lutein in phytoplankton extracts using reverse phase C-8 HPLC, *Marine Ecology Progress Series*, 161, 303–307,
1087 <https://doi.org/10.3354/meps161303>, 1997.
- 1088 Barthel, S., Tegen, I., and Wolke, R.: Do new sea spray aerosol source functions improve the results of a regional aerosol
1089 model?, *Atmospheric Environment*, 198, 265–278, <https://doi.org/10.1016/j.atmosenv.2018.10.016>, 2019.
- 1090 Bates, N. R., Garley, R., Frey, K. E., Shake, K. L., and Mathis, J. T.: Sea-ice melt CO₂-carbonate chemistry in the western Arctic
1091 Ocean: meltwater contributions to air–sea CO₂ gas exchange, mixed-layer properties and rates of net community production
1092 under sea ice, *Biogeosciences*, 11, 6769–6789, <https://doi.org/10.5194/bg-11-6769-2014>, 2014.
- 1093 Becker, S., Tebben, J., Coffinet, S., Wiltshire, K., Iversen, M. H., Harder, T., Hinrichs, K.-U., and Hehemann, J.-H.: Laminarin is
1094 a major molecule in the marine carbon cycle, *PNAS*, 117, 6599–6607, <https://doi.org/10.1073/pnas.1917001117>, 2020.
- 1095 Benner, R. and Kaiser, K.: Abundance of amino sugars and peptidoglycan in marine particulate and dissolved organic matter,
1096 *Limnology and Oceanography*, 48, 118–128, <https://doi.org/10.4319/lo.2003.48.1.0118>, 2003.
- 1097 Bigg, E. K. and Leck, C.: The composition of fragments of bubbles bursting at the ocean surface, *Journal of Geophysical
1098 Research: Atmospheres*, 113, <https://doi.org/10.1029/2007JD009078>, 2008.
- 1099 Bikerman, J. J.: *Foams*, Springer Science & Business Media, 344 pp., 2013.
- 1100 Bozem, H., Hoor, P., Kunkel, D., Köllner, F., Schneider, J., Herber, A., Schulz, H., Leaitch, W. R., Aliabadi, A. A., Willis, M. D.,
1101 Burkart, J., and Abbatt, J. P. D.: Characterization of transport regimes and the polar dome during Arctic spring and summer
1102 using in situ aircraft measurements, *Atmospheric Chemistry and Physics*, 19, 15049–15071, <https://doi.org/10.5194/acp-19-15049-2019>, 2019.
- 1104 Bricaud, A., Claustre, H., Ras, J., and Oubelkheir, K.: Natural variability of phytoplanktonic absorption in oceanic waters:
1105 Influence of the size structure of algal populations, *Journal of Geophysical Research: Oceans*, 109,
1106 <https://doi.org/10.1029/2004JC002419>, 2004.

- 1107 Burrows, S. M., Ogunro, O., Frossard, A., Russell, L. M., Rasch, P. J., and Elliott, S.: A Physically Based Framework for Modelling
1108 the Organic Fractionation of Sea Spray Aerosol from Bubble Film Langmuir Equilibria, *Atmospheric Chemistry and Physics*,
1109 14(24):13601–13629, <https://doi.org/10.5194/acp-14-13601-2014>, 2014.
- 1110 Burrows, S. M., Gobrogge, E., Fu, L., Link, K., Elliott, S. M., Wang, H., and Walker, R.: OCEANFILMS-2: Representing
1111 coadsorption of saccharides in marine films and potential impacts on modeled marine aerosol chemistry, *Geophysical*
1112 *Research Letters*, 43, 8306–8313, <https://doi.org/10.1002/2016GL069070>, 2016.
- 1113 Callaghan, A. H., Deane, G. B., Stokes, M. D., and Ward, B.: Observed variation in the decay time of oceanic whitecap foam,
1114 *Journal of Geophysical Research: Oceans*, 117, <https://doi.org/10.1029/2012JC008147>, 2012.
- 1115 Casillo, A., Lanzetta, R., Parrilli, M., and Corsaro, M. M.: Exopolysaccharides from Marine and Marine Extremophilic Bacteria:
1116 Structures, Properties, Ecological Roles and Applications, *Marine Drugs*, 16, 69, <https://doi.org/10.3390/md16020069>, 2018.
- 1117 Chen, Q., Mirrieles, J. A., Thanekar, S., Loeb, N. A., Kirpes, R. M., Upchurch, L. M., Barget, A. J., Lata, N. N., Raso, A. R. W.,
1118 McNamara, S. M., China, S., Quinn, P. K., Ault, A. P., Kennedy, A., Shepson, P. B., Fuentes, J. D., and Pratt, K. A.: Atmospheric
1119 particle abundance and sea salt aerosol observations in the springtime Arctic: a focus on blowing snow and leads,
1120 *Atmospheric Chemistry and Physics*, 22, 15263–15285, <https://doi.org/10.5194/acp-22-15263-2022>, 2022.
- 1121 Chi, J. W., Li, W. J., Zhang, D. Z., Zhang, J. C., Lin, Y. T., Shen, X. J., Sun, J. Y., Chen, J. M., Zhang, X. Y., Zhang, Y. M., and Wang,
1122 W. X.: Sea salt aerosols as a reactive surface for inorganic and organic acidic gases in the Arctic troposphere, *Atmospheric*
1123 *Chemistry and Physics*, 15, 11341–11353, <https://doi.org/10.5194/acp-15-11341-2015>, 2015.
- 1124 Creamean, J. M., Barry, K., Hill, T. C. J., Hume, C., DeMott, P. J., Shupe, M. D., Dahlke, S., Willmes, S., Schmale, J., Beck, I.,
1125 Hoppe, C. J. M., Fong, A., Chamberlain, E., Bowman, J., Scharien, R., and Persson, O.: Annual cycle observations of aerosols
1126 capable of ice formation in central Arctic clouds, *Nat Commun*, 13, 3537, <https://doi.org/10.1038/s41467-022-31182-x>, 2022.
- 1127 Cunliffe, M., Engel, A., Frka, S., Gašparović, B., Guitart, C., Murrell, J. C., Salter, M., Stolle, C., Upstill-Goddard, R., and Wurl,
1128 O.: Sea surface microlayers: A unified physicochemical and biological perspective of the air–ocean interface, *Progress in*
1129 *Oceanography*, 109, 104–116, <https://doi.org/10.1016/j.pocean.2012.08.004>, 2013.
- 1130 Cunliffe, M. and Wurl, O.: Guide to best practices to study the ocean’s surface., *Marine Biological Association of the United*
1131 *Kingdom for SCOR*, 2014.
- 1132 Delort, A.-M., Vaïtilingom, M., Amato, P., Sancelme, M., Parazols, M., Mailhot, G., Laj, P., and Deguillaume, L.: A short
1133 overview of the microbial population in clouds: Potential roles in atmospheric chemistry and nucleation processes,
1134 *Atmospheric Research*, 98, 249–260, <https://doi.org/10.1016/j.atmosres.2010.07.004>, 2010.
- 1135 DeMott, P. J., Hill, T. C. J., McCluskey, C. S., Prather, K. A., Collins, D. B., Sullivan, R. C., Ruppel, M. J., Mason, R. H., Irish, V. E.,
1136 Lee, T., Hwang, C. Y., Rhee, T. S., Snider, J. R., McMeeking, G. R., Dhaniyala, S., Lewis, E. R., Wentzell, J. J. B., Abbatt, J., Lee,
1137 C., Sultana, C. M., Ault, A. P., Axson, J. L., Martinez, M. D., Venero, I., Santos-Figueroa, G., Stokes, M. D., Deane, G. B., Mayol-
1138 Bracero, O. L., Grassian, V. H., Bertram, T. H., Bertram, A. K., Moffett, B. F., and Franc, G. D.: Sea spray aerosol as a unique
1139 source of ice nucleating particles, *PNAS*, 113, 5797–5803, <https://doi.org/10.1073/pnas.1514034112>, 2016.
- 1140 Demoz, B. B., Collett, J. L., and Daube, B. C.: On the Caltech Active Strand Cloudwater Collectors, *Atmospheric Research*, 41,
1141 47–62, [https://doi.org/10.1016/0169-8095\(95\)00044-5](https://doi.org/10.1016/0169-8095(95)00044-5), 1996.
- 1142 Dominutti, P. A., Renard, P., Vaïtilingom, M., Bianco, A., Baray, J.-L., Borbon, A., Bourianne, T., Burnet, F., Colomb, A., Delort,
1143 A.-M., Dufлот, V., Houdier, S., Jaffrezo, J.-L., Joly, M., Leremboure, M., Metzger, J.-M., Pichon, J.-M., Ribeiro, M., Rocco, M.,
1144 Tulet, P., Vella, A., Leriche, M., and Deguillaume, L.: Insights into tropical cloud chemistry in Réunion (Indian Ocean): results
1145 from the BIO-MAÏDO campaign, *Atmospheric Chemistry and Physics*, 22, 505–533, <https://doi.org/10.5194/acp-22-505-2022>,
1146 2022.
- 1147 Engbrodt, R.: Biogeochemistry of dissolved carbohydrates in the Arctic, *Berichte zur Polar-und Meeresforschung (Reports on*
1148 *Polar and Marine Research)*, 396, 106pp, 2001.
- 1149 Engel, A. and Händel, N.: A novel protocol for determining the concentration and composition of sugars in particulate and in
1150 high molecular weight dissolved organic matter (HMW-DOM) in seawater, *Marine Chemistry*, 127, 180–191,
1151 <https://doi.org/10.1016/j.marchem.2011.09.004>, 2011.
- 1152 Engel, A., Bange, H. W., Cunliffe, M., Burrows, S. M., Friedrichs, G., Galgani, L., Herrmann, H., Hertkorn, N., Johnson, M., Liss,
1153 P. S., Quinn, P. K., Schartau, M., Soloviev, A., Stolle, C., Upstill-Goddard, R. C., van Pinxteren, M., and Zäncker, B.: The Ocean’s
1154 Vital Skin: Toward an Integrated Understanding of the Sea Surface Microlayer, *Front. Mar. Sci.*, 4, 165,
1155 <https://doi.org/10.3389/fmars.2017.00165>, 2017.
- 1156 Ervens, B. and Amato, P.: The global impact of bacterial processes on carbon mass, *Atmospheric Chemistry & Physics*, 20,
1157 1777–1794, <https://doi.org/10.5194/acp-20-1777-2020>, 2020.
- 1158 Fabiano, M., Povero, P., and Danovaro, R.: Distribution and composition of particulate organic matter in the Ross Sea
1159 (Antarctica), *Polar Biol*, 13, 525–533, <https://doi.org/10.1007/BF00236394>, 1993.

- 1160 Facchini, M. C., Rinaldi, M., Decesari, S., Carbone, C., Finessi, E., Mircea, M., Fuzzi, S., Ceburnis, D., Flanagan, R., Nilsson, E. D.,
 1161 Leeuw, G. de, Martino, M., Woeltjen, J., and O'Dowd, C. D.: Primary submicron marine aerosol dominated by insoluble organic
 1162 colloids and aggregates, *Geophysical Research Letters*, 35, <https://doi.org/10.1029/2008GL034210>, 2008.
- 1163 Galgani, L., Piontek, J., and Engel, A.: Biopolymers form a gelatinous microlayer at the air-sea interface when Arctic sea ice
 1164 melts, *Scientific Reports*, 6, 29465, <https://doi.org/10.1038/srep29465>, 2016.
- 1165 Gantt, B., Meskhidze, N., Facchini, M. C., Rinaldi, M., Ceburnis, D., and O'Dowd, C. D.: Wind speed dependent size-resolved
 1166 parameterization for the organic mass fraction of sea spray aerosol, *Atmospheric Chemistry and Physics*, 11, 8777–8790,
 1167 <https://doi.org/10.5194/acp-11-8777-2011>, 2011.
- 1168 Gao, Q., Leck, C., Rauschenberg, C., and Matrai, P. A.: On the chemical dynamics of extracellular polysaccharides in the high
 1169 Arctic surface microlayer, *Ocean Science*, 8, 401–418, 2012.
- 1170 Gilardoni, S., Massoli, P., Giulianelli, L., Rinaldi, M., Paglione, M., Pollini, F., Lanconelli, C., Poluzzi, V., Carbone, S., Hillamo, R.,
 1171 Russell, L. M., Facchini, M. C., and Fuzzi, S.: Fog scavenging of organic and inorganic aerosol in the Po Valley, *Atmospheric
 1172 Chemistry and Physics*, 14, 6967–6981, <https://doi.org/10.5194/acp-14-6967-2014>, 2014.
- 1173 Goldberg, S. J., Carlson, C. A., Brzezinski, M., Nelson, N. B., and Siegel, D. A.: Systematic removal of neutral sugars within
 1174 dissolved organic matter across ocean basins, *Geophysical Research Letters*, 38, <https://doi.org/10.1029/2011GL048620>,
 1175 2011.
- 1176 Gonçalves-Araujo, R., Stedmon, C. A., Heim, B., Dubinenkov, I., Kraberg, A., Moiseev, D., and Bracher, A.: From Fresh to Marine
 1177 Waters: Characterization and Fate of Dissolved Organic Matter in the Lena River Delta Region, Siberia, *Frontiers in Marine
 1178 Science*, 2, 2015.
- 1179 Gong, X., Zhang, J., Croft, B., Yang, X., Frey, M. M., Bergner, N., Chang, R. Y.-W., Creamean, J. M., Kuang, C., Martin, R. V.,
 1180 Ranjithkumar, A., Sedlacek, A. J., Uin, J., Willmes, S., Zawadowicz, M. A., Pierce, J. R., Shupe, M. D., Schmale, J., and Wang, J.:
 1181 Arctic warming by abundant fine sea salt aerosols from blowing snow, *Nat. Geosci.*, 16, 768–774,
 1182 <https://doi.org/10.1038/s41561-023-01254-8>, 2023.
- 1183 Grythe, H., Ström, J., Krejci, R., Quinn, P., and Stohl, A.: A review of sea-spray aerosol source functions using a large global set
 1184 of sea salt aerosol concentration measurements, *Atmospheric Chemistry and Physics*, 14, 1277–1297,
 1185 <https://doi.org/10.5194/acp-14-1277-2014>, 2014.
- 1186 Haddrell, A. E. and Thomas, R. J.: Aerobiology: Experimental Considerations, Observations, and Future Tools, *Appl. Environ.
 1187 Microbiol.*, 83, <https://doi.org/10.1128/AEM.00809-17>, 2017.
- 1188 Hansell, D. A.: Recalcitrant Dissolved Organic Carbon Fractions, *Annual Review of Marine Science*, 5, 421–445,
 1189 <https://doi.org/10.1146/annurev-marine-120710-100757>, 2013.
- 1190 Hara, K., Yamagata, S., Yamanouchi, T., Sato, K., Herber, A., Iwasaka, Y., Nagatani, M., and Nakata, H.: Mixing states of
 1191 individual aerosol particles in spring Arctic troposphere during ASTAR 2000 campaign, *Journal of Geophysical Research:
 1192 Atmospheres*, 108, <https://doi.org/10.1029/2002JD002513>, 2003.
- 1193 Hartmann, M., Gong, X., Kecorius, S., van Pinxteren, M., Vogl, T., Welti, A., Wex, H., Zeppenfeld, S., Herrmann, H.,
 1194 Wiedensohler, A., and Stratmann, F.: Terrestrial or marine – indications towards the origin of ice-nucleating particles during
 1195 melt season in the European Arctic up to 83.7°N, *Atmospheric Chemistry and Physics*, 21, 11613–11636,
 1196 <https://doi.org/10.5194/acp-21-11613-2021>, 2021.
- 1197 Hasencz, E., Jayarathne, T., Pendergraft, M. A., Santander, M. V., Mayer, K. J., Sauer, J., Lee, C., Gibson, W. S., Kruse, S. M.,
 1198 Malfatti, F., Prather, K. A., and Stone, E. A.: Marine bacteria affect saccharide enrichment in sea spray aerosol during a
 1199 phytoplankton bloom, *ACS Earth Space Chem.*, 4, 1638–1649, <https://doi.org/10.1021/acsearthspacechem.0c00167>, 2020.
- 1200 Hasencz, E. S., Kaluarachchi, C. P., Lee, H. D., Tivanski, A. V., and Stone, E. A.: Saccharide Transfer to Sea Spray Aerosol
 1201 Enhanced by Surface Activity, Calcium, and Protein Interactions, *ACS Earth Space Chem.*, 3, 2539–2548,
 1202 <https://doi.org/10.1021/acsearthspacechem.9b00197>, 2019.
- 1203 Held, A., Brooks, I. M., Leck, C., and Tjernström, M.: On the potential contribution of open lead particle emissions to the
 1204 central Arctic aerosol concentration, *Atmospheric Chemistry and Physics*, 11, 3093–3105, <https://doi.org/10.5194/acp-11-3093-2011>, 2011.
- 1206 Herrmann, H., Schaefer, T., Tilgner, A., Styler, S. A., Weller, C., Teich, M., and Otto, T.: Tropospheric Aqueous-Phase Chemistry:
 1207 Kinetics, Mechanisms, and Its Coupling to a Changing Gas Phase, *Chem. Rev.*, 115, 4259–4334,
 1208 <https://doi.org/10.1021/cr500447k>, 2015.
- 1209 Hoffman, E. J. and Duce, R. A.: Factors influencing the organic carbon content of marine aerosols: A laboratory study, *Journal
 1210 of Geophysical Research (1896-1977)*, 81, 3667–3670, <https://doi.org/10.1029/JC081i021p03667>, 1976.
- 1211 Huang, J. and Jaeglé, L.: Wintertime enhancements of sea salt aerosol in polar regions consistent with a sea ice source from
 1212 blowing snow, *Atmos. Chem. Phys.*, 17, 3699–3712, <https://doi.org/10.5194/acp-17-3699-2017>, 2017.

- 1213 Istomina, L.: Retrieval of Sea Ice Surface Melt Using OLCI Data Onboard Sentinel-3, 2020, C017-07, 2020.
- 1214 Ittekkot, V., Brockmann, U., Michaelis, W., and Degens, E. T.: Dissolved free and combined carbohydrates during a
1215 phytoplankton bloom in the northern North Sea, *Marine Ecology Progress Series*, 4, 299–305, 1981.
- 1216 Karl, M., Leck, C., Rad, F. M., Bäcklund, A., Lopez-Aparicio, S., and Heintzenberg, J.: New insights in sources of the sub-
1217 micrometre aerosol at Mt. Zeppelin observatory (Spitsbergen) in the year 2015, *Tellus B: Chemical and Physical Meteorology*,
1218 71, 1613143, <https://doi.org/10.1080/16000889.2019.1613143>, 2019.
- 1219 Kecorius, S., Vogl, T., Paasonen, P., Lampilahti, J., Rothenberg, D., Wex, H., Zeppenfeld, S., van Pinxteren, M., Hartmann, M.,
1220 Henning, S., Gong, X., Welti, A., Kulmala, M., Stratmann, F., Herrmann, H., and Wiedensohler, A.: New particle formation and
1221 its effect on cloud condensation nuclei abundance in the summer Arctic: a case study in the Fram Strait and Barents Sea,
1222 *Atmospheric Chemistry and Physics*, 19, 14339–14364, <https://doi.org/10.5194/acp-19-14339-2019>, 2019.
- 1223 Keene, W. C., Long, M. S., Reid, J. S., Frossard, A. A., Kieber, D. J., Maben, J. R., Russell, L. M., Kinsey, J. D., Quinn, P. K., and
1224 Bates, T. S.: Factors That Modulate Properties of Primary Marine Aerosol Generated From Ambient Seawater on Ships at Sea,
1225 *Journal of Geophysical Research: Atmospheres*, 122, 11,961–11,990, <https://doi.org/10.1002/2017JD026872>, 2017.
- 1226 Kirchman, D. L., Meon, B., Ducklow, H. W., Carlson, C. A., Hansell, D. A., and Steward, G. F.: Glucose fluxes and concentrations
1227 of dissolved combined neutral sugars (polysaccharides) in the Ross Sea and Polar Front Zone, Antarctica, *Deep Sea Research*
1228 *Part II: Topical Studies in Oceanography*, 48, 4179–4197, [https://doi.org/10.1016/S0967-0645\(01\)00085-6](https://doi.org/10.1016/S0967-0645(01)00085-6), 2001.
- 1229 Kirpes, R. M., Bondy, A. L., Bonanno, D., Moffet, R. C., Wang, B., Laskin, A., Ault, A. P., and Pratt, K. A.: Secondary sulfate is
1230 internally mixed with sea spray aerosol and organic aerosol in the winter Arctic, *Atmos. Chem. Phys.*, 18, 3937–3949,
1231 <https://doi.org/10.5194/acp-18-3937-2018>, 2018.
- 1232 Krembs, C. and Deming, J. W.: The role of exopolymers in microbial adaptation to sea ice, in: *Psychrophiles: from biodiversity*
1233 *to biotechnology*, Springer, 247–264, 2008.
- 1234 Krembs, C., Eicken, H., Junge, K., and Deming, J. W.: High concentrations of exopolymeric substances in Arctic winter sea ice:
1235 implications for the polar ocean carbon cycle and cryoprotection of diatoms, *Deep Sea Research Part I: Oceanographic*
1236 *Research Papers*, 49, 2163–2181, [https://doi.org/10.1016/S0967-0637\(02\)00122-X](https://doi.org/10.1016/S0967-0637(02)00122-X), 2002.
- 1237 Kubota, H., Ogiwara, Y., and Matsuzaki, K.: Photo-Induced Formation of Peroxide in Saccharides and Related Compounds,
1238 *Polymer Journal*, 8, 557–563, <https://doi.org/10.1295/polymj.8.557>, 1976.
- 1239 Kumai, M.: Arctic Fog Droplet Size Distribution and Its Effect on Light Attenuation, *Journal of the Atmospheric Sciences*, 30,
1240 635–643, [https://doi.org/10.1175/1520-0469\(1973\)030<0635:AFDSDA>2.0.CO;2](https://doi.org/10.1175/1520-0469(1973)030<0635:AFDSDA>2.0.CO;2), 1973.
- 1241 Lawler, M. J., Saltzman, E. S., Karlsson, L., Zieger, P., Salter, M., Baccharini, A., Schmale, J., and Leck, C.: New Insights Into the
1242 Composition and Origins of Ultrafine Aerosol in the Summertime High Arctic, *Geophysical Research Letters*, 48,
1243 e2021GL094395, <https://doi.org/10.1029/2021GL094395>, 2021.
- 1244 Leck, C.: Chemical composition and sources of the high Arctic aerosol relevant for cloud formation, *Journal of Geophysical*
1245 *Research*, 107, <https://doi.org/10.1029/2001JD001463>, 2002.
- 1246 Leck, C., Gao, Q., Mashayekhy Rad, F., and Nilsson, U.: Size-resolved atmospheric particulate polysaccharides in the high
1247 summer Arctic, *Atmospheric Chemistry and Physics*, 13, 12573–12588, <https://doi.org/10.5194/acp-13-12573-2013>, 2013.
- 1248 Lefering, I., Röttgers, R., Utschig, C., and McKee, D.: Uncertainty budgets for liquid waveguide CDOM absorption
1249 measurements, *Appl. Opt.*, AO, 56, 6357–6366, <https://doi.org/10.1364/AO.56.006357>, 2017.
- 1250 Liu, Y., Röttgers, R., Ramírez-Pérez, M., Dinter, T., Steinmetz, F., Nöthig, E.-M., Hellmann, S., Wiegmann, S., and Bracher, A.:
1251 Underway spectrophotometry in the Fram Strait (European Arctic Ocean): a highly resolved chlorophyll a data source for
1252 complementing satellite ocean color, *Opt. Express*, OE, 26, A678–A696, <https://doi.org/10.1364/OE.26.00A678>, 2018.
- 1253 Macke, A. and Flores, H.: The Expeditions PS106/1 and 2 of the Research Vessel POLARSTERN to the Arctic Ocean in 2017,
1254 Bremerhaven, Germany, 171 pp., https://doi.org/10.2312/BzPM_0719_2018, 2018.
- 1255 Malfatti, F., Lee, C., Tinta, T., Pendergraft, M. A., Celussi, M., Zhou, Y., Sultana, C. M., Rotter, A., Axson, J. L., Collins, D. B.,
1256 Santander, M. V., Anides Morales, A. L., Aluwihare, L. I., Riemer, N., Grassian, V. H., Azam, F., and Prather, K. A.: Detection of
1257 Active Microbial Enzymes in Nascent Sea Spray Aerosol: Implications for Atmospheric Chemistry and Climate, *Environ. Sci.*
1258 *Technol. Lett.*, 6, 171–177, <https://doi.org/10.1021/acs.estlett.8b00699>, 2019.
- 1259 Mari, X., Passow, U., Migon, C., Burd, A. B., and Legendre, L.: Transparent exopolymer particles: Effects on carbon cycling in
1260 the ocean, *Progress in Oceanography*, 151, 13–37, <https://doi.org/10.1016/j.pocean.2016.11.002>, 2017.
- 1261 Marks, R., Kruczalac, K., Jankowska, K., and Michalska, M.: Bacteria and fungi in air over the Gulf of Gdańsk and Baltic sea,
1262 *Journal of Aerosol Science*, 32, 237–250, [https://doi.org/10.1016/S0021-8502\(00\)00064-1](https://doi.org/10.1016/S0021-8502(00)00064-1), 2001.
- 1263 Matsuoka, A., Bricaud, A., Benner, R., Para, J., Sempéré, R., Prieur, L., Bélanger, S., and Babin, M.: Tracing the transport of
1264 colored dissolved organic matter in water masses of the Southern Beaufort Sea: relationship with hydrographic
1265 characteristics, *Biogeosciences*, 9, 925–940, <https://doi.org/10.5194/bg-9-925-2012>, 2012.

- 1266 Matsuoka, A., Hooker, S. B., Bricaud, A., Gentili, B., and Babin, M.: Estimating absorption coefficients of colored dissolved
1267 organic matter (CDOM) using a semi-analytical algorithm for southern Beaufort Sea waters: application to deriving
1268 concentrations of dissolved organic carbon from space, *Biogeosciences*, 10, 917–927, [https://doi.org/10.5194/bg-10-917-](https://doi.org/10.5194/bg-10-917-2013)
1269 2013, 2013.
- 1270 Matulová, M., Husárová, S., Capek, P., Sancelme, M., and Delort, A.-M.: Biotransformation of Various Saccharides and
1271 Production of Exopolymeric Substances by Cloud-Borne *Bacillus* sp. 3B6, *Environ. Sci. Technol.*, 48, 14238–14247,
1272 <https://doi.org/10.1021/es501350s>, 2014.
- 1273 May, N. W., Quinn, P. K., McNamara, S. M., and Pratt, K. A.: Multiyear study of the dependence of sea salt aerosol on wind
1274 speed and sea ice conditions in the coastal Arctic: ARCTIC SEA SALT AEROSOL, *J. Geophys. Res. Atmos.*, 121, 9208–9219,
1275 <https://doi.org/10.1002/2016JD025273>, 2016.
- 1276 Mayol, E., Arrieta, J. M., Jiménez, M. A., Martínez-Asensio, A., Garcias-Bonet, N., Dachs, J., González-Gaya, B., Royer, S.-J.,
1277 Benítez-Barrios, V. M., Fraile-Nuez, E., and Duarte, C. M.: Long-range transport of airborne microbes over the global tropical
1278 and subtropical ocean, *Nature Communications*, 8, 1–9, <https://doi.org/10.1038/s41467-017-00110-9>, 2017.
- 1279 McCarthy, M., Hedges, J., and Benner, R.: Major biochemical composition of dissolved high molecular weight organic matter
1280 in seawater, *Marine Chemistry*, 55, 281–297, [https://doi.org/10.1016/S0304-4203\(96\)00041-2](https://doi.org/10.1016/S0304-4203(96)00041-2), 1996.
- 1281 McCluskey, C. S., Hill, T. C. J., Humphries, R. S., Rauker, A. M., Moreau, S., Strutton, P. G., Chambers, S. D., Williams, A. G.,
1282 McRobert, I., Ward, J., Keywood, M. D., Harnwell, J., Ponsonby, W., Loh, Z. M., Krummel, P. B., Protat, A., Kreidenweis, S. M.,
1283 and DeMott, P. J.: Observations of Ice Nucleating Particles Over Southern Ocean Waters, *Geophysical Research Letters*, 45,
1284 11,989–11,997, <https://doi.org/10.1029/2018GL079981>, 2018.
- 1285 Mimura, T. and Romano, J. C.: Muramic Acid measurements for bacterial investigations in marine environments by high-
1286 pressure liquid chromatography, *Appl. Environ. Microbiol.*, 50, 229–237, <https://doi.org/10.1128/AEM.50.2.229-237.1985>,
1287 1985.
- 1288 Mühlenbruch, M., Grossart, H.-P., Eigemann, F., and Voss, M.: Mini-review: Phytoplankton-derived polysaccharides in the
1289 marine environment and their interactions with heterotrophic bacteria, *Environmental Microbiology*, 20, 2671–2685,
1290 <https://doi.org/10.1111/1462-2920.14302>, 2018.
- 1291 Müller, K., Lehmann, S., Pinxteren, D. van, Gnauk, T., Niedermeier, N., Wiedensohler, A., and Herrmann, H.: Particle
1292 characterization at the Cape Verde atmospheric observatory during the 2007 RHaMBLe intensive, *Atmospheric Chemistry
1293 and Physics*, 10, 2709–2721, <https://doi.org/10.5194/acp-10-2709-2010>, 2010.
- 1294 Norris, S. J., Brooks, I. M., de Leeuw, G., Sirevaag, A., Leck, C., Brooks, B. J., Birch, C. E., and Tjernström, M.: Measurements of
1295 bubble size spectra within leads in the Arctic summer pack ice, *Ocean Science*, 7, 129–139, [https://doi.org/10.5194/os-7-129-](https://doi.org/10.5194/os-7-129-2011)
1296 2011, 2011.
- 1297 Notz, D. and Worster, M. G.: Desalination processes of sea ice revisited, *Journal of Geophysical Research: Oceans*, 114,
1298 <https://doi.org/10.1029/2008JC004885>, 2009.
- 1299 Obernosterer, I., Catala, P., Lami, R., Caparros, J., Ras, J., Bricaud, A., Christine, D., Van Wambeke, F., and Lebaron, P.:
1300 Biochemical characteristics and bacterial community structure of the sea surface microlayer in the South Pacific Ocean,
1301 *Biogeosciences*, 5, 693–705, 2008.
- 1302 O’Dowd, C. D., Facchini, M. C., Cavalli, F., Ceburnis, D., Mircea, M., Decesari, S., Fuzzi, S., Yoon, Y. J., and Putaud, J.-P.:
1303 Biogenically driven organic contribution to marine aerosol, *Nature*, 431, 676–680, <https://doi.org/10.1038/nature02959>,
1304 2004.
- 1305 Orellana, M. V., Matrai, P. A., Leck, C., Rauschenberg, C. D., Lee, A. M., and Coz, E.: Marine microgels as a source of cloud
1306 condensation nuclei in the high Arctic, *PNAS*, 108, 13612–13617, <https://doi.org/10.1073/pnas.1102457108>, 2011.
- 1307 Panagiotopoulos, C. and Sempéré, R.: Analytical methods for the determination of sugars in marine samples: A historical
1308 perspective and future directions, *Limnology and Oceanography: Methods*, 3, 419–454,
1309 <https://doi.org/10.4319/lom.2005.3.419>, 2005.
- 1310 Papakonstantinou-Presvelou, I., Sourdeval, O., and Quaas, J.: Strong Ocean/Sea-Ice Contrasts Observed in Satellite-Derived
1311 Ice Crystal Number Concentrations in Arctic Ice Boundary-Layer Clouds, *Geophysical Research Letters*, 49, e2022GL098207,
1312 <https://doi.org/10.1029/2022GL098207>, 2022.
- 1313 Passow, U.: Transparent exopolymer particles (TEP) in aquatic environments, *Progress in Oceanography*, 55, 287–333,
1314 [https://doi.org/10.1016/S0079-6611\(02\)00138-6](https://doi.org/10.1016/S0079-6611(02)00138-6), 2002.
- 1315 Penner, J. E., Andreae, M. O., Annegarn, H., Barrie, L., Feichter, J., Hegg, D., Jayaraman, A., Leitch, R., Murphy, D., Nganga,
1316 J., and Pitari, G.: Aerosols, their Direct and Indirect Effects, *Climate Change 2001: The Scientific Basis. Contribution of Working
1317 Group I to the Third Assessment Report of the Intergovernmental Panel on Climate Change*, 289–348, 2001.
- 1318 Phongphattarawat, S.: Variability in pigment composition and bio-optical characteristics of phytoplankton populations in the
1319 Atlantic basin, <http://purl.org/dc/dcmitype/Text>, University of Oxford, 2016.

- 1320 van Pinxteren, M., Müller, C., Iinuma, Y., Stolle, C., and Herrmann, H.: Chemical Characterization of Dissolved Organic
 1321 Compounds from Coastal Sea Surface Microlayers (Baltic Sea, Germany), *Environmental Science & Technology*, 46, 10455–
 1322 10462, <https://doi.org/10.1021/es204492b>, 2012.
- 1323 van Pinxteren, M., Barthel, S., Fomba, K. W., Müller, K., Von Tümpling, W., and Herrmann, H.: The influence of environmental
 1324 drivers on the enrichment of organic carbon in the sea surface microlayer and in submicron aerosol particles – measurements
 1325 from the Atlantic Ocean, *Elem Sci Anth*, 5, <https://doi.org/10.1525/elementa.225>, 2017.
- 1326 van Pinxteren, M., Robinson, T.-B., Zeppenfeld, S., Gong, X., Bahlmann, E., Fomba, K. W., Triesch, N., Stratmann, F., Wurl, O.,
 1327 Engel, A., Wex, H., and Herrmann, H.: High number concentrations of transparent exopolymer particles in ambient aerosol
 1328 particles and cloud water – a case study at the tropical Atlantic Ocean, *Atmospheric Chemistry and Physics*, 22, 5725–5742,
 1329 <https://doi.org/10.5194/acp-22-5725-2022>, 2022.
- 1330 van Pinxteren, M., Zeppenfeld, S., Fomba, K. W., Triesch, N., Frka, S., and Herrmann, H.: Amino acids, carbohydrates, and
 1331 lipids in the tropical oligotrophic Atlantic Ocean: sea-to-air transfer and atmospheric in situ formation, *Atmospheric Chemistry
 1332 and Physics*, 23, 6571–6590, <https://doi.org/10.5194/acp-23-6571-2023>, 2023.
- 1333 Piontek, J., Lunau, M., Händel, N., Borchard, C., Wurst, M., and Engel, A.: Acidification increases microbial polysaccharide
 1334 degradation in the ocean, *Biogeosciences*, 7, 1615–1624, <https://doi.org/10.5194/bg-7-1615-2010>, 2010.
- 1335 Porter, G. C. E., Adams, M. P., Brooks, I. M., Ickes, L., Karlsson, L., Leck, C., Salter, M. E., Schmale, J., Siegel, K., Sikora, S. N. F.,
 1336 Tarn, M. D., Vüllers, J., Wernli, H., Zieger, P., Zinke, J., and Murray, B. J.: Highly Active Ice-Nucleating Particles at the Summer
 1337 North Pole, *Journal of Geophysical Research: Atmospheres*, 127, e2021JD036059, <https://doi.org/10.1029/2021JD036059>,
 1338 2022.
- 1339 Prather, K. A., Bertram, T. H., Grassian, V. H., Deane, G. B., Stokes, M. D., DeMott, P. J., Aluwihare, L. I., Palenik, B. P., Azam,
 1340 F., Seinfeld, J. H., Moffet, R. C., Molina, M. J., Cappa, C. D., Geiger, F. M., Roberts, G. C., Russell, L. M., Ault, A. P., Baltrusaitis,
 1341 J., Collins, D. B., Corrigan, C. E., Cuadra-Rodriguez, L. A., Ebben, C. J., Forestieri, S. D., Guasco, T. L., Hersey, S. P., Kim, M. J.,
 1342 Lambert, W. F., Modini, R. L., Mui, W., Pedler, B. E., Ruppel, M. J., Ryder, O. S., Schoepp, N. G., Sullivan, R. C., and Zhao, D.:
 1343 Bringing the ocean into the laboratory to probe the chemical complexity of sea spray aerosol, *PNAS*, 110, 7550–7555,
 1344 <https://doi.org/10.1073/pnas.1300262110>, 2013.
- 1345 Qin, G., Zhu, L., Chen, X., Wang, P. G., and Zhang, Y.: Structural characterization and ecological roles of a novel
 1346 exopolysaccharide from the deep-sea psychrotolerant bacterium *Pseudoalteromonas* sp. SM9913, *Microbiology*, 153, 1566–
 1347 1572, <https://doi.org/10.1099/mic.0.2006/003327-0>, 2007.
- 1348 Quinn, P. K., Collins, D. B., Grassian, V. H., Prather, K. A., and Bates, T. S.: Chemistry and Related Properties of Freshly Emitted
 1349 Sea Spray Aerosol, *Chemical Reviews*, 115, 4383–4399, <https://doi.org/10.1021/cr500713g>, 2015.
- 1350 Rastelli, E., Corinaldesi, C., Dell’Anno, A., Lo Martire, M., Greco, S., Cristina Facchini, M., Rinaldi, M., O’Dowd, C., Ceburnis, D.,
 1351 and Danovaro, R.: Transfer of labile organic matter and microbes from the ocean surface to the marine aerosol: an
 1352 experimental approach, *Scientific Reports*, 7, 1–10, <https://doi.org/10.1038/s41598-017-10563-z>, 2017.
- 1353 Rérolle, V., Ruiz-Pino, D., Rafizadeh, M., Loucaides, S., Papadimitriou, S., Mowlem, M., and Chen, J.: Measuring pH in the
 1354 Arctic Ocean: Colorimetric method or SeaFET?, *Methods in Oceanography*, 17, 32–49,
 1355 <https://doi.org/10.1016/j.mio.2016.05.006>, 2016.
- 1356 Robinson, T.-B., Wurl, O., Bahlmann, E., Jürgens, K., and Stolle, C.: Rising bubbles enhance the gelatinous nature of the air–
 1357 sea interface, *Limnology and Oceanography*, 64, 2358–2372, <https://doi.org/10.1002/lno.11188>, 2019.
- 1358 Rolph, R. J., Feltham, D. L., and Schröder, D.: Changes of the Arctic marginal ice zone during the satellite era, *The Cryosphere*,
 1359 14, 1971–1984, <https://doi.org/10.5194/tc-14-1971-2020>, 2020.
- 1360 Röttgers, R., McKee, D., and Utschig, C.: Temperature and salinity correction coefficients for light absorption by water in the
 1361 visible to infrared spectral region, *Opt. Express*, OE, 22, 25093–25108, <https://doi.org/10.1364/OE.22.025093>, 2014.
- 1362 Röttgers, R., Doxaran, D., and Dupouy, C.: Quantitative filter technique measurements of spectral light absorption by aquatic
 1363 particles using a portable integrating cavity absorption meter (QFT-ICAM), *Opt. Express*, OE, 24, A1–A20,
 1364 <https://doi.org/10.1364/OE.24.0000A1>, 2016.
- 1365 Russell, L. M., Hawkins, L. N., Frossard, A. A., Quinn, P. K., and Bates, T. S.: Carbohydrate-like composition of submicron
 1366 atmospheric particles and their production from ocean bubble bursting, *Proc. Natl. Acad. Sci. U.S.A.*, 107, 6652–6657,
 1367 <https://doi.org/10.1073/pnas.0908905107>, 2010.
- 1368 Šantl-Temkiv, T., Gosewinkel, U., Starnawski, P., Lever, M., and Finster, K.: Aeolian dispersal of bacteria in southwest
 1369 Greenland: their sources, abundance, diversity and physiological states, *FEMS Microbiol Ecol*, 94,
 1370 <https://doi.org/10.1093/femsec/fiy031>, 2018.
- 1371 Šantl-Temkiv, T., Sikoparija, B., Maki, T., Carotenuto, F., Amato, P., Yao, M., Morris, C. E., Schnell, R., Jaenicke, R., Pöhlker, C.,
 1372 DeMott, P. J., Hill, T. C. J., and Huffman, J. A.: Bioaerosol field measurements: Challenges and perspectives in outdoor studies,
 1373 *Aerosol Science and Technology*, 54, 520–546, <https://doi.org/10.1080/02786826.2019.1676395>, 2020.

- 1374 Schiffer, J. M., Mael, L. E., Prather, K. A., Amaro, R. E., and Grassian, V. H.: Sea Spray Aerosol: Where Marine Biology Meets
1375 Atmospheric Chemistry, *ACS Cent. Sci.*, 4, 1617–1623, <https://doi.org/10.1021/acscentsci.8b00674>, 2018.
- 1376 Schill, S. R., Burrows, S. M., Hasenecz, E. S., Stone, E. A., and Bertram, T. H.: The Impact of Divalent Cations on the Enrichment
1377 of Soluble Saccharides in Primary Sea Spray Aerosol, *Atmosphere*, 9, 476, <https://doi.org/10.3390/atmos9120476>, 2018.
- 1378 Schmale, J., Zieger, P., and Ekman, A. M. L.: Aerosols in current and future Arctic climate, *Nature Climate Change*, 11, 95–105,
1379 <https://doi.org/10.1038/s41558-020-00969-5>, 2021.
- 1380 Schmithüsen, H.: Continuous meteorological surface measurement during POLARSTERN cruise PS106/1 (ARK-XXXI/1.1),
1381 <https://doi.org/10.1594/PANGAEA.886302>, 2018.
- 1382 Schmithüsen, H.: Continuous meteorological surface measurement during POLARSTERN cruise PS106/2 (ARK-XXXI/1.2),
1383 <https://doi.org/10.1594/PANGAEA.901179>, 2019.
- 1384 Schmitt-Kopplin, P., Liger-Belair, G., Koch, B. P., Flerus, R., Kattner, G., Harir, M., Kanawati, B., Lucio, M., Tziotis, D., Hertkorn,
1385 N., and Gebefügi, I.: Dissolved organic matter in sea spray: a transfer study from marine surface water to aerosols,
1386 *Biogeosciences*, 9, 1571–1582, 2012.
- 1387 Sellegri, K., O'Dowd, C. D., Yoon, Y. J., Jennings, S. G., and Leeuw, G. de: Surfactants and submicron sea spray generation,
1388 *Journal of Geophysical Research: Atmospheres*, 111, <https://doi.org/10.1029/2005JD006658>, 2006.
- 1389 Spencer, R. G. M., Aiken, G. R., Butler, K. D., Dornblaser, M. M., Striegl, R. G., and Hernes, P. J.: Utilizing chromophoric dissolved
1390 organic matter measurements to derive export and reactivity of dissolved organic carbon exported to the Arctic Ocean: A
1391 case study of the Yukon River, Alaska, *Geophysical Research Letters*, 36, <https://doi.org/10.1029/2008GL036831>, 2009.
- 1392 Stedmon, C. A., Amon, R. M. W., Rinehart, A. J., and Walker, S. A.: The supply and characteristics of colored dissolved organic
1393 matter (CDOM) in the Arctic Ocean: Pan Arctic trends and differences, *Marine Chemistry*, 124, 108–118,
1394 <https://doi.org/10.1016/j.marchem.2010.12.007>, 2011.
- 1395 Stein, A. F., Draxler, R. R., Rolph, G. D., Stunder, B. J. B., Cohen, M. D., and Ngan, F.: NOAA's HYSPLIT Atmospheric Transport
1396 and Dispersion Modeling System, *Bull. Amer. Meteor. Soc.*, 96, 2059–2077, <https://doi.org/10.1175/BAMS-D-14-00110.1>,
1397 2015.
- 1398 Stolle, C., Nagel, K., Labrenz, M., and Jürgens, K.: Succession of the sea-surface microlayer in the coastal Baltic Sea under
1399 natural and experimentally induced low-wind conditions, *Biogeosciences*, 7, 2975–2988, [https://doi.org/10.5194/bg-7-2975-
1400 2010](https://doi.org/10.5194/bg-7-2975-2010), 2010.
- 1401 Strong, C. and Rigor, I. G.: Arctic marginal ice zone trending wider in summer and narrower in winter, *Geophysical Research
1402 Letters*, 40, 4864–4868, <https://doi.org/10.1002/grl.50928>, 2013.
- 1403 Sud, I. J. and Tyler, M. E.: Cell-Wall Composition and Osmotic Fragility of Selected Marine Bacteria, *Journal of Bacteriology*,
1404 87, 696–700, 1964.
- 1405 Suzuki, E. and Suzuki, R.: Variation of Storage Polysaccharides in Phototrophic Microorganisms, *Journal of Applied
1406 Glycoscience*, 60, 21–27, https://doi.org/10.5458/jag.jag.JAG-2012_016, 2013.
- 1407 Taylor, B. B., Torrecilla, E., Bernhardt, A., Taylor, M. H., Peeken, I., Röttgers, R., Piera, J., and Bracher, A.: Bio-optical provinces
1408 in the eastern Atlantic Ocean and their biogeographical relevance, 8, 7165–7219, <https://doi.org/10.5194/bgd-8-7165-2011>,
1409 2011.
- 1410 Thomson, J.: Wave propagation in the marginal ice zone: connections and feedback mechanisms within the air–ice–ocean
1411 system, *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences*, 380, 20210251,
1412 <https://doi.org/10.1098/rsta.2021.0251>, 2022.
- 1413 Triesch, N., van Pinxteren, M., Engel, A., and Herrmann, H.: Concerted measurements of free amino acids at the Cabo Verde
1414 islands: high enrichments in submicron sea spray aerosol particles and cloud droplets, *Atmospheric Chemistry and Physics*,
1415 21, 163–181, <https://doi.org/10.5194/acp-21-163-2021>, 2021a.
- 1416 Triesch, N., van Pinxteren, M., Frka, S., Stolle, C., Spranger, T., Hoffmann, E. H., Gong, X., Wex, H., Schulz-Bull, D., Gašparović,
1417 B., and Herrmann, H.: Concerted measurements of lipids in seawater and on submicrometer aerosol particles at the Cabo
1418 Verde islands: biogenic sources, selective transfer and high enrichments, *Atmospheric Chemistry and Physics*, 21, 4267–4283,
1419 <https://doi.org/10.5194/acp-21-4267-2021>, 2021b.
- 1420 Triesch, N., van Pinxteren, M., Salter, M., Stolle, C., Pereira, R., Zieger, P., and Herrmann, H.: Sea Spray Aerosol Chamber Study
1421 on Selective Transfer and Enrichment of Free and Combined Amino Acids, *ACS Earth Space Chem.*, 5, 1564–1574,
1422 <https://doi.org/10.1021/acsearthspacechem.1c00080>, 2021c.
- 1423 Trueblood, J. V., Wang, X., Or, V. W., Alves, M. R., Santander, M. V., Prather, K. A., and Grassian, V. H.: The Old and the New:
1424 Aging of Sea Spray Aerosol and Formation of Secondary Marine Aerosol through OH Oxidation Reactions, *ACS Earth Space
1425 Chem.*, 3, 2307–2314, <https://doi.org/10.1021/acsearthspacechem.9b00087>, 2019.

- 1426 Tynan, E., Clarke, J. S., Humphreys, M. P., Ribas-Ribas, M., Esposito, M., Rérolle, V. M. C., Schlosser, C., Thorpe, S. E., Tyrrell,
1427 T., and Achterberg, E. P.: Physical and biogeochemical controls on the variability in surface pH and calcium carbonate
1428 saturation states in the Atlantic sectors of the Arctic and Southern Oceans, *Deep Sea Research Part II: Topical Studies in*
1429 *Oceanography*, 127, 7–27, <https://doi.org/10.1016/j.dsr2.2016.01.001>, 2016.
- 1430 Uetake, J., Hill, T. C. J., Moore, K. A., DeMott, P. J., Protat, A., and Kreidenweis, S. M.: Airborne bacteria confirm the pristine
1431 nature of the Southern Ocean boundary layer, *PNAS*, 117, 13275–13282, <https://doi.org/10.1073/pnas.2000134117>, 2020.
- 1432 Vaqué, D., Boras, J. A., Arrieta, J. M., Agustí, S., Duarte, C. M., and Sala, M. M.: Enhanced Viral Activity in the Surface Microlayer
1433 of the Arctic and Antarctic Oceans, *Microorganisms*, 9, 317, <https://doi.org/10.3390/microorganisms9020317>, 2021.
- 1434 Veron, F.: Ocean Spray, *Annual Review of Fluid Mechanics*, 47, 507–538, <https://doi.org/10.1146/annurev-fluid-010814-014651>, 2015.
- 1436 Walker, S. A., Amon, R. M. W., and Stedmon, C. A.: Variations in high-latitude riverine fluorescent dissolved organic matter:
1437 A comparison of large Arctic rivers, *Journal of Geophysical Research: Biogeosciences*, 118, 1689–1702,
1438 <https://doi.org/10.1002/2013JG002320>, 2013.
- 1439 Wang, X., Deane, G. B., Moore, K. A., Ryder, O. S., Stokes, M. D., Beall, C. M., Collins, D. B., Santander, M. V., Burrows, S. M.,
1440 Sultana, C. M., and Prather, K. A.: The role of jet and film drops in controlling the mixing state of submicron sea spray aerosol
1441 particles, *PNAS*, 114, 6978–6983, <https://doi.org/10.1073/pnas.1702420114>, 2017.
- 1442 Wendisch, M., Macke, A., Ehrlich, A., Lüpkes, C., Mech, M., Chechin, D., Dethloff, K., Barientos, C., Bozem, H., Brückner, M.,
1443 Clemen, H.-C., Crewell, S., Donth, T., Dupuy, R., Ebell, K., Egerer, U., Engelmann, R., Engler, C., Eppers, O., Gehrman, M.,
1444 Gong, X., Gottschalk, M., Goubeyre, C., Griesche, H., Hartmann, J., Hartmann, M., Heinold, B., Herber, A., Herrmann, H.,
1445 Heygster, G., Hoor, P., Jafariserajehlou, S., Jäkel, E., Järvinen, E., Jourdan, O., Kästner, U., Kecorius, S., Knudsen, E. M., Köllner,
1446 F., Kretzschmar, J., Lelli, L., Leroy, D., Maturilli, M., Mei, L., Mertes, S., Mioche, G., Neuber, R., Nicolaus, M., Nomokonova, T.,
1447 Notholt, J., Palm, M., van Pinxteren, M., Quaas, J., Richter, P., Ruiz-Donoso, E., Schäfer, M., Schmieder, K., Schnaiter, M.,
1448 Schneider, J., Schwarzenböck, A., Seifert, P., Shupe, M. D., Siebert, H., Spreen, G., Stapf, J., Stratmann, F., Vogl, T., Welti, A.,
1449 Wex, H., Wiedensohler, A., Zanatta, M., and Zeppenfeld, S.: The Arctic Cloud Puzzle: Using ALOUD/PASCAL Multi-Platform
1450 Observations to Unravel the Role of Clouds and Aerosol Particles in Arctic Amplification, *Bull. Amer. Meteor. Soc.*,
1451 <https://doi.org/10.1175/BAMS-D-18-0072.1>, 2018.
- 1452 Wendisch, M., Brückner, M., Crewell, S., Ehrlich, A., Notholt, J., Lüpkes, C., Macke, A., Burrows, J. P., Rinke, A., Quaas, J.,
1453 Maturilli, M., Schemann, V., Shupe, M. D., Akansu, E. F., Barrientos-Velasco, C., Bärfuss, K., Blechschmidt, A.-M., Block, K.,
1454 Bougoudis, I., Bozem, H., Böckmann, C., Bracher, A., Bresson, H., Bretschneider, L., Buschmann, M., Chechin, D. G., Chylik, J.,
1455 Dahlke, S., Deneke, H., Dethloff, K., Donth, T., Dorn, W., Dupuy, R., Ebell, K., Egerer, U., Engelmann, R., Eppers, O., Gerdes, R.,
1456 Gierens, R., Gorodetskaya, I. V., Gottschalk, M., Griesche, H., Gryanik, V. M., Handorf, D., Harm-Altstädter, B., Hartmann, J.,
1457 Hartmann, M., Heinold, B., Herber, A., Herrmann, H., Heygster, G., Höschel, I., Hofmann, Z., Hölemann, J., Hünnerbein, A.,
1458 Jafariserajehlou, S., Jäkel, E., Jacobi, C., Janout, M., Jansen, F., Jourdan, O., Jurányi, Z., Kalesse-Los, H., Kanzow, T., Käthner,
1459 R., Kliesch, L. L., Klingebiel, M., Knudsen, E. M., Kovács, T., Körtke, W., Krampe, D., Kretzschmar, J., Kreyling, D., Kulla, B.,
1460 Kunkel, D., Lampert, A., Lauer, M., Lelli, L., Lerber, A. von Linke, O., Löhnert, U., Lonardi, M., Losa, S. N., Losch, M., Maahn,
1461 M., Mech, M., Mei, L., Mertes, S., Metzner, E., Mewes, D., Michaelis, J., Mioche, G., Moser, M., Nakoudi, K., Neggers, R.,
1462 Neuber, R., Nomokonova, T., Oelker, J., Papakonstantinou-Presvelou, I., et al.: Atmospheric and Surface Processes, and
1463 Feedback Mechanisms Determining Arctic Amplification: A Review of First Results and Prospects of the (AC)3 Project, *Bulletin*
1464 *of the American Meteorological Society*, 104, E208–E242, <https://doi.org/10.1175/BAMS-D-21-0218.1>, 2023.
- 1465 Wietz, M., Wemheuer, B., Simon, H., Giebel, H.-A., Seibt, M. A., Daniel, R., Brinkhoff, T., and Simon, M.: Bacterial community
1466 dynamics during polysaccharide degradation at contrasting sites in the Southern and Atlantic Oceans, *Environ. Microbiol.*, 17,
1467 3822–3831, <https://doi.org/10.1111/1462-2920.12842>, 2015.
- 1468 Wilbourn, E. K., Thornton, D. C. O., Ott, C., Graff, J., Quinn, P. K., Bates, T. S., Betha, R., Russell, L. M., Behrenfeld, M. J., and
1469 Brooks, S. D.: Ice Nucleation by Marine Aerosols Over the North Atlantic Ocean in Late Spring, *Journal of Geophysical*
1470 *Research: Atmospheres*, 125, e2019JD030913, <https://doi.org/10.1029/2019JD030913>, 2020.
- 1471 Williams, P. M., Carlucci, A. F., Henrichs, S. M., Van Vleet, E. S., Horrigan, S. G., Reid, F. M. H., and Robertson, K. J.: Chemical
1472 and microbiological studies of sea-surface films in the Southern Gulf of California and off the West Coast of Baja California,
1473 *Marine Chemistry*, 19, 17–98, [https://doi.org/10.1016/0304-4203\(86\)90033-2](https://doi.org/10.1016/0304-4203(86)90033-2), 1986.
- 1474 Wurl, O., Miller, L., Röttgers, R., and Vagle, S.: The distribution and fate of surface-active substances in the sea-surface
1475 microlayer and water column, *Marine Chemistry*, 115, 1–9, <https://doi.org/10.1016/j.marchem.2009.04.007>, 2009.
- 1476 Wurl, O., Miller, L., and Vagle, S.: Production and fate of transparent exopolymer particles in the ocean, *Journal of Geophysical*
1477 *Research: Oceans*, 116, <https://doi.org/10.1029/2011JC007342>, 2011.
- 1478 Xu, M., Tsona Tchinda, N., Li, J., and Du, L.: Insoluble lipid film mediates transfer of soluble saccharides from the sea to the
1479 atmosphere: the role of hydrogen bonding, *Atmospheric Chemistry and Physics*, 23, 2235–2249, <https://doi.org/10.5194/acp-23-2235-2023>, 2023.

- 1481 Xu, W., Ovadnevaite, J., Fossum, K. N., Lin, C., Huang, R.-J., Ceburnis, D., and O'Dowd, C.: Sea spray as an obscured source for
1482 marine cloud nuclei, *Nat. Geosci.*, 15, 282–286, <https://doi.org/10.1038/s41561-022-00917-2>, 2022.
- 1483 Yang, X., Pyle, J. A., and Cox, R. A.: Sea salt aerosol production and bromine release: Role of snow on sea ice, *Geophysical
1484 Research Letters*, 35, <https://doi.org/10.1029/2008GL034536>, 2008.
- 1485 Zábory, J., Matisāns, M., Krejci, R., Nilsson, E. D., and Ström, J.: Artificial primary marine aerosol production: a laboratory study
1486 with varying water temperature, salinity, and succinic acid concentration, *Atmospheric Chemistry and Physics*, 12, 10709–
1487 10724, <https://doi.org/10.5194/acp-12-10709-2012>, 2012.
- 1488 Zäncker, B., Bracher, A., Röttgers, R., and Engel, A.: Variations of the Organic Matter Composition in the Sea Surface
1489 Microlayer: A Comparison between Open Ocean, Coastal, and Upwelling Sites Off the Peruvian Coast, *Frontiers in
1490 Microbiology*, 8, 2369, <https://doi.org/10.3389/fmicb.2017.02369>, 2017.
- 1491 Zäncker, B., Cunliffe, M., and Engel, A.: Eukaryotic community composition in the sea surface microlayer across an east–west
1492 transect in the Mediterranean Sea, *Biogeosciences*, 18, 2107–2118, <https://doi.org/10.5194/bg-18-2107-2021>, 2021.
- 1493 Zeppenfeld, S., van Pinxteren, M., Hartmann, M., Bracher, A., Stratmann, F., and Herrmann, H.: Glucose as a Potential
1494 Chemical Marker for Ice Nucleating Activity in Arctic Seawater and Melt Pond Samples, *Environ. Sci. Technol.*, 53, 8747–8756,
1495 <https://doi.org/10.1021/acs.est.9b01469>, 2019a.
- 1496 Zeppenfeld, S., van Pinxteren, M., Hartmann, M., Bracher, A., Wiegmann, S., Stratmann, F., and Herrmann, H.: Glucose, T50
1497 and salinity in the surface microlayer and bulk water samples from the Arctic during POLARSTERN cruise PS106 (2017),
1498 <https://doi.org/10.1594/PANGAEA.899258>, 2019b.
- 1499 Zeppenfeld, S., van Pinxteren, M., Engel, A., and Herrmann, H.: A protocol for quantifying mono- and polysaccharides in
1500 seawater and related saline matrices by electro-dialysis (ED) – combined with HPAEC-PAD, *Ocean Science*, 16, 817–830,
1501 <https://doi.org/10.5194/os-16-817-2020>, 2020.
- 1502 Zeppenfeld, S., van Pinxteren, M., van Pinxteren, D., Wex, H., Berdalet, E., Vaqué, D., Dall'Osto, M., and Herrmann, H.: Aerosol
1503 Marine Primary Carbohydrates and Atmospheric Transformation in the Western Antarctic Peninsula, *ACS Earth Space Chem.*,
1504 5, 1032–1047, <https://doi.org/10.1021/acsearthspacechem.0c00351>, 2021a.
- 1505 Zeppenfeld, S., Fuchs, S., Rödger, S., Dietze, A., van Pinxteren, M., and Herrmann, H.: Marine carbohydrates and inorganic
1506 ions in size-resolved atmospheric particles collected over the Southern Ocean, <https://doi.org/10.1594/PANGAEA.927565>,
1507 2021b.
- 1508 Zeppenfeld, S., van Pinxteren, M., Fuchs, S., Hartmann, M., Gong, X., and Herrmann, H.: Inorganic ions in fog water sampled
1509 from the Arctic in 2017, <https://doi.org/10.1594/PANGAEA.932573>, 2021c.
- 1510 Zeppenfeld, S., van Pinxteren, M., Fuchs, S., Hartmann, M., Gong, X., and Herrmann, H.: Inorganic ions in size-resolved aerosol
1511 particles sampled from the Arctic in 2017, <https://doi.org/10.1594/PANGAEA.932569>, 2021d.
- 1512 Zeppenfeld, S., van Pinxteren, M., Fuchs, S., Hartmann, M., and Herrmann, H.: Combined carbohydrates, dissolved free
1513 carbohydrates and pH in Arctic fog water sampled during PS106, <https://doi.org/10.1594/PANGAEA.962208>, 2023a.
- 1514 Zeppenfeld, S., van Pinxteren, M., Fuchs, S., Hartmann, M., and Herrmann, H.: Combined carbohydrates, organic carbon and
1515 total aerosol mass concentrations in size-resolved aerosol particles sampled from the Arctic in 2017,
1516 <https://doi.org/10.1594/PANGAEA.962210>, 2023b.
- 1517 Zeppenfeld, S., Bracher, A., Wiegmann, S., Zeising, M., Fuchs, S., van Pinxteren, M., and Herrmann, H.: Dissolved and
1518 particulate combined carbohydrates, pH, inorganic ions, CDOM and particulate absorption of SML and bulk water in Arctic
1519 surface seawater and melt ponds, <https://doi.org/10.1594/PANGAEA.961004>, 2023c.
- 1520 Zinke, J., Salter, M. E., Leck, C., Lawler, M. J., Porter, G. C. E., Adams, M. P., Brooks, I. M., Murray, B. J., and Zieger, P.: The
1521 development of a miniaturised balloon-borne cloud water sampler and its first deployment in the high Arctic, *Tellus B:
1522 Chemical and Physical Meteorology*, 73, 1–12, <https://doi.org/10.1080/16000889.2021.1915614>, 2021.
- 1524