

Carbon emission and export from Ket River, western Siberia

Artem G. Lim¹, Ivan V. Krickov¹, Sergey N. Vorobyev¹,

Mikhail A. Korets², Sergey Kopysov¹,

Liudmila S. Shirokova³, Jan Karlsson⁴, and Oleg S. Pokrovsky^{5*}

¹BIO-GEO-CLIM Laboratory, Tomsk State University, Tomsk, Russia

² V.N. Sukachev Institute of Forest of the Siberian Branch of Russian Academy of Sciences – separated department of the KSC SB RAS, Krasnoyarsk, 660036, Russia

³ N. Laverov Federal Center for Integrated Arctic Research, Russian Academy of Sciences, Arkhangelsk, Russia

⁴Climate Impacts Research Centre (CIRC), Department of Ecology and Environmental Science, Umeå University, Linnaeus väg 6, 901 87 Umeå, Sweden.

⁵ Geosciences and Environment Toulouse, UMR 5563 CNRS, 14 Avenue Edouard Belin 31400 Toulouse, France

Key words: CO₂, C, emission, boreal, river, export, landscape, Siberia

* email: oleg.pokrovsky@get.omp.eu

Abstract

Despite recent progress in the understanding of the carbon (C) cycle of Siberian permafrost-affected rivers, spatial and seasonal dynamics of C export and emission from medium-size rivers (50,000 - 300,000 km² watershed area) remain poorly known. Here we studied one of the largest tributaries of the Ob River, the Ket River (watershed = 94,000 km²) which drains through pristine taiga forest of the boreal zone in western Siberian Lowland (WSL). We combined continuous and discrete measurements of carbon dioxide (CO₂) concentration using submersible CO₂ sensor and floating chamber flux (FCO₂), with methane (CH₄), organic and inorganic C (DOC and DIC, respectively), particulate organic C and total bacterial concentrations over a 800-km transect of the Ket River main stem and its 26 tributaries during spring flood (May 2019) and 12 tributaries during summer baseflow (end of August – beginning of September 2019). The partial pressure of CO₂ (pCO₂) was lower and less variable in the main stem (2000 to 2500 μ atm) compared to that in tributaries (2000 to 5000 μ atm). In the tributaries, the pCO₂ was 40 % higher during baseflow compared to spring flood, whereas in the main stem, it did not vary significantly across the seasons. The methane concentration in the main stem and tributaries was a factor of 300 to 1900 (flood period) and 100 to 150 times lower than that of CO₂, and ranged from 0.05 to 2.0 μ mol L⁻¹. The FCO₂ ranged from 0.4 to 2.4 g C m⁻² d⁻¹ in the main channel and from 0.5 to 5.0 g C m⁻² d⁻¹ in the tributaries, being the highest during August in tributaries and weakly dependent on season in the main channel. During summer baseflow, the DOC aromaticity, bacterial number, and needleleaf forest coverage of the watershed positively affected CO₂ concentrations and fluxes. We hypothesize that relatively low spatial and seasonal variability in FCO₂ of the Ket River is due to flat homogeneous landscape (bogs and taiga forest) that results in long water residence times and stable input of allochthonous DOM, which dominate the FCO₂. The open water period (May to October) C emission from the fluvial network (main stem and tributaries) of the Ket River was estimated to 127 $\pm$ 11 Gg C y⁻¹ which is lower than the downstream dissolved and particulate C export during the same period. The estimated fluvial C emissions are highly conservative and contain uncertainties, linked to ignoring hot spots and hot moments of emissions, notably in the floodplain zone. This stresses the need of improving temporal resolution of FCO₂ and water coverage across seasons and emphasizes the important role of WSL rivers for release of CO₂ to the atmosphere.

60 **Introduction**

61 Assessment of greenhouse gas (GHG) emission from rivers is crucially important for understanding
62 the C cycle under various climate change scenarios (Campeau and del Giorgio, 2014; Chadburn et al., 2017;
63 Tranvik et al., 2018; Vonk et al., 2019; Vachon et al., 2020). Rivers receive terrestrial C and process and emit
64 a significant share of this C during transit to the sea (Liu et al., 2022). Quantifications of riverine C emissions
65 are sufficiently robust for relatively well studied regions of the world such as the European and N American
66 boreal zone (Dawson et al., 2004; Dinsmore et al., 2013; Wallin et al., 2013; Leith et al., 2015; Zolkos et al.,
67 2019; Hutchins et al., 2020), or Arctic and subarctic rivers of Alaska (Striegl et al., 2012; Crawford et al.,
68 2013; Stackpoole et al., 2017), although subjected to great uncertainty. Despite significant progress in
69 assessing riverine pCO₂ in previously under-represented or ignored regions such as lotic systems of Asia (Ran
70 et al., 2015, 2017; Varol and Li, 2017) or South America (Almeida et al., 2017), these studies generally use
71 a combination of pH and alkalinity (DIC) to calculate the pCO₂ instead of direct in-situ measurements, alike
72 the studies of global emissions (Raymond et al., 2013; Lauerwald et al., 2015). At the same time, there is a
73 growing number of studies reporting directly measured riverine pCO₂ – either discretely (Alin et al. 2011;
74 Borges et al., 2015; Amaral et al. 2018; 2022; Leng et al. 2022), continuously at fixed sites (Crawford et al.
75 2016a, Schneider et al. 2020; Gómez-Gener et al. 2021a), or along the river flow (Abril et al. 2014; Crawford
76 et al. 2016b; 2017; Borges et al. 2019). However, these studies are limited to tropical and temperate zones of
77 the world, and boreal regions of Western Europe and Northern America, and thus, further continuous and
78 discrete measurements of CO₂ concentration and fluxes in rivers from under-represented regions such as
79 Northern Eurasia, and in particular, Siberia, are needed. The on-going interest to Siberia comes from the fact
80 that this region hosts large C stocks in soils and wetlands intersected by extensive river networks that deliver
81 majority of water and C to the Arctic Ocean (Feng et al., 2013).

82 A few works on Siberian fluvial systems dealt with small (Castro-Morales et al., 2022) and large
83 (Denfeld et al., 2013; Vorobyev et al., 2021) rivers, but these were performed in Eastern Siberia, under
84 continuous permafrost zone. More progress has been achieved in quantification of downstream carbon export
85 by permafrost-affected Great Arctic Rivers of Siberia (Lobbies et al., 2000; Raymond et al., 2007; Cooper et
86 al., 2008; Semiletov et al., 2011; Feng et al., 2013; Griffin et al., 2018; Wild et al., 2019). However, spatial

and seasonal features of C emission from tributaries of large Siberian rivers are still remain poorly known. Existing data on western Siberia (Serikova et al., 2018; Karlsson et al., 2021) suggest that C (predominantly as CO₂) emissions from rivers can vary largely over space and time. Such high variations do not allow reliable quantitative assessment of C emission and integrating these values into regional and global C models.

In order to better understand and constrain the magnitude of C emission from Siberian rivers, we studied the Ket River (watershed 94,000 km²), a typical tributary of the Ob River in western Siberia. The Ob river is the largest (in terms of watershed area) Siberian river and drains large pristine territories of taiga forest and bogs. The catchment of Ob includes extensive regions of permafrost but a major part of it (> 80 %) is situated in the permafrost-free zone of which very few data exist on riverine C emissions (Karlsson et al., 2021). The Ket river drains through dense southern taiga forest and abundant wetlands with almost no human activity, thus serving a representative system for understanding C cycling in permafrost-free Siberian rivers. We followed, via a boat routing over the main stem and main tributaries of the river, the in-situ CO₂ concentrations combined with discrete sampling for dissolved CH₄, DOC, DIC, total bacterial number and particulate organic matter. These measurements were complemented with regular floating chamber measurements of CO₂ emission fluxes. We performed these observations during two main open water seasons of the year - the peak of the spring flood and the end of the summer baseflow. Our first objective was to quantify the difference in C concentration and emission during two seasons for the main stem and the tributaries and to relate these differences to main physico-chemical parameters of the water column and physio-geographical parameters (land cover) of the river watersheds. Our second objective was to obtain total C emission flux from the river watershed area and compare it to downstream export yield of dissolved and particulate carbon.

2. Study Site, Materials and Methods

2.1. Ket River and its tributaries

The Ket River main stem and its 26 tributaries sampled in this study include watersheds of distinct sizes (catchment area ranged from 94,000 km² at the Ket's mouth to 20 km² of smallest tributary), but rather similar lithology, climate and vegetation (**Fig. 1, Table S1**). The Strahler's order of sampled rivers and stream

114 ranges from 9 for Ket at its mouth to 2 for the smallest stream. The poorly accessible Ket River basin is fully
115 pristine (50 % forest, 40 % wetlands), and has almost no agricultural and forestry activity. The watershed of
116 Ket has very low population density ($0.27 \text{ person km}^{-2}$) and lacks road infrastructure due to absence of oil
117 and gas development and production. In this regard, this river can serve as a model for medium size bog-
118 forest rivers of the western Siberia Lowland and results obtained from this watershed can be extrapolated to
119 much larger territory, comprising about 1 million km^2 of permafrost-free taiga forest and bog regions of the
120 southern part of WSL.

121 The mean annual air temperatures (MAAT) is $-0.7 \pm 0.1 \text{ }^{\circ}\text{C}$ and the mean annual precipitation is 520
122 $\pm 20 \text{ mm y}^{-1}$ in the central part of the basin. The lithology of this part of western Siberian lowland is dominated
123 by Pleistocene silts and sands with carbonate concretions overlayed by quaternary deposits (loesses, fluvial,
124 glacial and lacustrine deposits). The dominant soils are podzols in forest areas and histosols in peat bog
125 regions. Further description of climate, lithology and landscape features of the territory is provided in former
126 studies (Frey and Smith, 2007; Pokrovsky et al., 2015).

127 The peak of annual discharge in 2019 occurred in the end of May; in August, the discharge was 3 to
128 5 times smaller (**Fig. 1**). Note that low runoff, lack of relief and highly homogenous landscape coverage of
129 the permafrost-free zone of western Siberia in general and of the Ket River basin in particular provide quite
130 smooth hydrographs of the rivers. In this regard, the spring flood period is extended over 2 months, from the
131 beginning of May to middle of July, whereas summer baseflow includes second half of July, August and
132 September. As a result, similar to previous study of rivers along a 2500 km transect of the WSL territory, the
133 timing of the two sampling campaigns covered approximately 80% of the annual water discharge in the basins
134 (Serikova et al., 2018). From May 18 to May 28, 2019, and from August 30 to September 2, 2019, we started
135 the boat trip in the middle course of the Ket River (Beliy Yar), and moved, first, 475 km upstream the Ket
136 river till its most headwaters, and then moved 834 km downstream till the river mouth, with an average speed
137 of 20 km h^{-1} . During summer baseflow, the 4-days trip was shortened by 200 km due to too low water level
138 in the upper reaches of the main stem and some small tributaries. We stopped each 30-50 km along the Ket
139 River and sampled for major hydrochemical parameters, GHG, river suspended matter and total bacterial
140 number of the main stem. We also moved several km upstream of selected tributaries to record CO_2

141 concentrations for at least 1 h and to sample for river hydrochemistry. At several occasions during spring
142 flood, we monitored CO₂ concentration and performed chamber measurements in the main stem and
143 tributaries during both day and night time period.

144

145 2.2. CO₂ and CH₄ concentrations and CO₂ fluxes by floating chambers

146 Surface water CO₂ concentration was measured continuously, *in-situ* by deploying a portable infrared
147 gas analyzer (IRGA, GMT222 CARBOCAP® probe, Vaisala®; accuracy ± 1.5%) of two ranges (2 000 and
148 10 000 ppm) as described in previous work of our group on the Lena River (Vorobyev et al., 2021). Sensor
149 preparation was conducted in the lab following the method described by Johnson et al. (2009). The
150 measurement unit (MI70, Vaisala®; accuracy ± 0.2%) was connected to the sensor allowing instantaneous
151 readings of pCO₂. The sensors were calibrated in the lab against standard gas mixtures (0, 800, 3 000, 8 000
152 ppm; linear regression with R² > 0.99) before and after the field campaign. The sensors' drift was 0.03-0.06%
153 per day and overall error was 4-8% (relative standard deviation, RSD). Following calibration, post-
154 measurement correction of the sensor output induced by changes in water temperature and barometric
155 pressure was done by applying empirically derived coefficients following Johnson et al. (2009). These
156 corrections never exceeded 5% of the measured values. During the cruise, we routinely measured atmospheric
157 CO₂ with the probe as a check for its good functioning. Furthermore, we tested two different sensors in several
158 sites of the river transect: a main probe used for continuous measurements and another probe used as a control
159 and never employed for continuous measurements. We did not find any sizable (>10%) difference in
160 measured CO₂ concentration between these two probes.

161 The probe was enclosed within a waterproof and gas-permeable membrane. For this, we used a
162 protective expanded polytetrafluoroethylene (PTFE) sleeve that is highly permeable to CO₂ but impermeable
163 to water (Johnson et al., 2009). The sensor was placed into a tube which was submerged 0.5 m below the
164 water surface. A Campbell logger was connected to the system allowing continuous recording of the CO₂
165 concentration, water temperature and pressure every minute. These readings were averaged over 10 minute
166 intervals yielding 732 individual pCO₂, water temperature and pressure values. The CO₂ concentrations in the
167 Ket River tributaries included between 10 and 20 averaged pCO₂ values for each tributary (250 measurements

168 in total) during spring flood period. In addition to continuous *in-situ* CO₂ measurements, we estimated pCO₂
169 via measured pH and DIC values, using the set of constants typically applied for riverine pCO₂ estimation in
170 organic-rich waters (Cai and Wang, 1998; DelDuco and Xu, 2017). The U-test (Mann-Whitney) demonstrated
171 a lack of significant difference in CO₂ concentrations measured by Vaissala and calculated from the pH and
172 DIC of the river water.

173 Discrete CO₂ fluxes were measured by using two floating CO₂ chambers equipped with non-
174 dispersive infrared SenseAir® CO₂ loggers (Bastviken et al., 2015), at each of the 7 (spring flood) and 6
175 (summer baseflow) sampling location of the main stem and 26 tributaries following the procedures described
176 elsewhere (Serikova et al., 2019; Krickov et al., 2021). The chambers were not anchored but slowly free-
177 drifted together with the boat, because it is known that anchored chambers can artificially enhance fluxes due
178 to turbulence thus providing erroneous estimates (Lorke et al., 2015). The CO₂ accumulation rate inside each
179 chamber was recorded continuously at 300 s interval. We used first 0.5–1 h of measurements for computing
180 CO₂ accumulation rate inside each chamber by linear regression.

181 For CH₄ analyses, unfiltered water was sampled in 60-mL Serum bottles. For this, the bottles and caps
182 were manually submerged at approx. 30 cm depth from the water surface. The bottles were closed without
183 air bubbles using vinyl stoppers and aluminum caps and immediately poisoned by adding 0.2 mL of saturated
184 HgCl₂ via a two-way needle system. The samples were stored approximately one week in the refrigerator
185 before the analyses. In the laboratory, a headspace was created by displacing approximately 40% of water
186 with N₂ (99.999%). Two 0.5-mL replicates of the equilibrated headspace were analyzed for their
187 concentrations of CH₄, using a Bruker GC-456 gas chromatograph (GC) equipped with flame ionization and
188 thermal conductivity detectors (Serikova et al., 2019; Vorobyev et al., 2021). After every 10 samples, a
189 calibration of the detectors was performed using Air Liquid gas standards (i.e. 145 ppmv). Duplicate injection
190 of the samples showed that results were reproducible within ±5%. The specific gas solubility for CH₄
191 (Yamamoto et al., 1976) were used in calculation of the CH₄ content in the water. We calculated instantaneous
192 diffusive CH₄ fluxes for each of the chambers using chamber-specific gas transfer velocity (K_T) and the
193 concentrations of dissolved CH₄ in the water and in air-water equilibrium (atm = 1.8 ppm), following the
194 procedure outlined in Serikova et al. (2018), who used the same setup for measurements of GHG emissions

from small and medium-size rivers of the WSL. Note that this setup does not allow measuring the ebullitive CH_4 fluxes and thus it is possible that the evasion of CH_4 , especially in the stagnant zone of the river flow and floodplain in this study is sizably underestimated (i.e., Spawn et al., 2015; Stanley et al., 2016; Villa et al., 2021).

2.3. Chemical analyses of the river water

The dissolved oxygen (CellOx 325; accuracy of $\pm 5\%$), specific conductivity (TetraCon 325; $\pm 1.5\%$), and water temperature ($\pm 0.2^\circ\text{C}$) were measured in-situ at 20 cm depth using a WTW 3320 Multimeter. The pH was measured using portable Hanna instrument via combined Schott glass electrode calibrated with standard buffer solutions (4.01, 6.86 and 9.18 at 25°C), with an uncertainty of 0.01 pH units. The temperature of buffer solutions was within $\pm 2^\circ\text{C}$ of that of the river water. The water was sampled in pre-cleaned polypropylene bottle from 20-30 cm depth in the middle of the river and immediately filtered through disposable single-use sterile Sartorius filter units (0.45 μm pore size). The first 50 mL of filtrate was discarded. The DOC and Dissolved Inorganic Carbon (DIC) were determined by a Shimadzu TOC-VSCN Analyzer (Kyoto, Japan) with an uncertainty of 3% and a detection limit of 0.1 mg/L. Blanks of MilliQ water passed through the filters demonstrated negligible release of DOC from the filter material. The SUVA was measured via ultraviolet absorbance at 254 nm using a 10-mm quartz cuvette on a Bruker CARY-50 UV-VIS spectrophotometer.

The concentration of C and N in suspended material (Particulate Organic Carbon and Nitrogen (POC and PON, respectively)) was determined via filtration of 1 to 2 L of freshly collected river water (at the river bank or in the boat) with pre-weighted GFF filters (47 mm, 0.45 μm) and Nalgene 250-mL polystyrene filtration units using a Mityvac® manual vacuum pump. Particulate C and N were measured using catalytic combustion with Cu-O at 900°C with an uncertainty of $\leq 0.5\%$ using Thermo Flash 2000 CN Analyzer at EcoLab, Toulouse. The samples were analyzed before and after 1:1 HCl treatment to distinguish between total and inorganic C; however the ratio of $C_{\text{organic}} : C_{\text{carbonate}}$ in the river suspended matter (RSM) was always above 20 and the contribution of carbonate C to total C in the RSM was equal in average $0.3 \pm 0.3\%$ (2 s.d., $n = 30$).

Total microbial cell concentration was measured after sample fixation in glutaraldehyde, by a flow cytometry (Guava® EasyCyte™ systems, Merck). Cells were stained using 1 µL of a 10 times diluted SYBR GREEN solution (10000x, Merck), added to 250 µL of each sample before analysis. Particles were identified as cells based on green fluorescence and forward scatter (Marie et al., 2001).

2.4. Riverine carbon export flux by the Ket catchment

The C export flux over active (unfrozen) period (May to October) from the Ket basin was calculated based on monthly-averaged discharge at the river mouth in 2019 available from Russian Hydrological Survey and DOC, DIC and POC concentrations measured in the low reaches of the Ket River in this study (see hydrograph in **Fig. 1**). Riverine element fluxes should be usually estimated using a LOADEST method (Holmes et al., 2012) from calculated daily element loads. The latter typically obtained from a calibration regression, applied to daily discharge. This calibration regression can be constructed from time series of paired streamflow and measured element concentration data for sufficient period of the year. In our previous works in this and other similar boreal regions, we demonstrated that this method provides reasonable (within 10 to 30 %) agreement with monthly export fluxes calculated by multiplying mean monthly discharge by mean monthly concentration (Chupakov et al., 2020; Pokrovsky et al., 2022; Vorobyev et al., 2019). Given that the intrinsic uncertainties on mean monthly discharge are also between 10 and 20 % (see discussion for the WSL rivers in Pokrovsky et al., 2020), in this study, for open-water period export flux calculation, we used DOC, DIC and POC concentrations measured during spring flood (for May and June period) and baseflow (for August, September and October period). For the month of July, we used the mean concentrations of end of May and August-September which is in accord with seasonal discharge pattern of the Ket River. Note that the contribution of non-studied October month to total open water period water flux is < 10 % and thus cannot provide sizable uncertainties

2.5. Landscape parameters and water surface area of the Ket River basin

The physio-geographical characteristics of the 26 Ket tributaries and the 7 points of the Ket main stem (**Table S1, Fig. S1**) were determined by applying available digital elevation model (DEM GMTED2010), soil, vegetation and lithological maps. The landscape parameters were typified using TerraNorte Database of Land Cover of Russia (Bartalev et al., 2020; <http://terranorte.iki.rssi.ru>). This included various type of forest (evergreen, deciduous, needleleaf/broadleaf), grassland, tundra, wetlands, water bodies and riparian zones. Note that the land cover data correspond to the whole catchment area upstream of the sampling point. The climate parameters the watershed were obtained from CRU grids data (1950-2016) (Harris et al., 2014) and NCSCD data (Hugelius et al., 2013), respectively. The biomass was obtained from BIOMASAR2 dataset in raster format with spatial resolution of 1 x 1 km (Santoro et al., 2010). The soil OC content was taken from the Northern Circumpolar Soil Carbon Database (NCSCD). The original NCSCD dataset produced in GIS vector format corresponding to 1:1,000,000 scale of topographic map. It could be rasterized to 1 x 1 km pixel resolution. The lithology layer was taken from GIS version of Geological map of the Russian Federation (scale 1:5,000,000, <http://www.geolkarta.ru/>). We quantified river water surface area using the global SDG database with 30 m² resolution (Pekel et al., 2016) including both seasonal and permanent water for the open water period of 2019 and for the multiannual average (reference period 2000-2004). We also used a more recent GRWL Mask Database which incorporates first order temporary non-active streams (Allen and Pavelsky, 2018).

2.6. Data analysis

Carbon concentrations and fluxes for all dataset were tested for normality using a Shapiro-Wilk test. In case if the data were not normally distributed, we used non-parametric statistics. Comparisons of GHG parameters in the main stem and tributaries during two sampling seasons were conducted using a non-parametric Mann Whitney test at a significance level of 0.05. For comparison of unpaired data, a non-parametric H-criterion Kruskal-Wallis test was used to reveal the differences between different study sites. The Pearson rank order correlation coefficient ($p < 0.05$) was used to determine the relationship between CO₂ concentrations and emission fluxes and main landscape parameters of the Ket River tributaries, as well as

276 other potential drivers such as pH, O₂, water temperature, specific conductivity, DOC, DIC, particulate carbon
277 and nitrogen, and total bacterial number.

278 Further identification of C pattern drivers in river waters included a Principal Component Analysis
279 which allowed to test the effect of various hydrochemical and landscape parameters on CO₂ and CH₄
280 concentrations and CO₂ emissions. In addition to PCA, a Redundancy analysis (RDA) was used to extract
281 and summarize the variation in C pattern that can be explained by a set of explanatory variables
282 (environmental, climatic and hydrochemical factors). The RDA combines a PCA and multiple regression
283 analysis and it was run in XLSTAT is a statistical software that works as an add-on to Excel.

284

285

286 **3. Results**

287 *3.1. Greenhouse gases and dissolved and particulate C*

288 The main hydrochemical parameters and greenhouse gases concentration and exchange fluxes of the
289 Ket River and its tributaries are listed in **Table 1** and primary data are provided in **Table S2** of the
290 Supplement. Continuous pCO₂ measurements in the main stem during the spring (764 individual data points
291 over the full distance of the boat route (834 km) demonstrated a lack of systematic change in CO₂
292 concentration from headwaters to the mouth. The CO₂ concentration in tributaries was generally higher than
293 that in the main stem. As a result, the pCO₂ changed by a factor of 1.5 to 2 when tributaries with high CO₂
294 concentrations join the main stem (**Fig. 2 A**). There were strong but non-systematic variations in CO₂
295 concentration in the tributaries during the summer (**Fig. 2 C**). The CH₄ concentration (**Table 1 and Fig. S2**
296 **A, B**) was low in the Ket River (around 0.17 and 0.86 $\mu\text{mol L}^{-1}$ in May and August, respectively) and in the
297 tributaries (range 0.09 to 2.6 $\mu\text{mol L}^{-1}$, 2 to 3 times higher values during the baseflow). These values are
298 generally higher than the range of CH₄ concentration in large Siberian Rivers such as Lena (0.03 to 0.199
299 $\mu\text{mol L}^{-1}$, Bussman, 2013; Vorobyev et al., 2021) but consistent with concentrations in surface layers of East
300 Siberian ponds (0.6-2.4 $\mu\text{mol L}^{-1}$, Rehder et al., 2021). In the Ket River main stem and tributaries, the CH₄
301 concentrations are 300-2000 and 100-150 times lower than those of CO₂ during spring and summer,

respectively, and ranged from 0.05 to 2.0 $\mu\text{mol L}^{-1}$. Consequently, diffuse CH_4 emissions (**Table 1, Fig. S2 C, D**) constituted 0.1 to 0.5% of total C emissions and are not discussed in further detail.

During spring flood, CO_2 fluxes ranged from 0.26 to 3.2 $\text{g C m}^{-2} \text{d}^{-1}$ in the main stem and tributaries (**Table 1; Fig. 2 B**). During baseflow, the flux in the tributaries varied from 0.37 to 7.4 $\text{g C m}^{-2} \text{d}^{-1}$ and was a factor of 2 to 3 higher than that in the main stem (**Fig. 2 D; Table 1**). The CO_2 concentration in the river water and gas transfer velocity assessed from discrete measurements by floating chambers ($K_T = 0.08\text{-}1.83 \text{ m d}^{-1}$ in the main stem; $0.2\text{-}1.86 \text{ m d}^{-1}$ in the tributaries, **Table 1**) allowed for calculation of the continuous CO_2 fluxes (**Fig. 2 B**). For this, we used an average value of K_T measured between two chamber sites (separated by a distance of 50 to 100 km) to calculate the FCO_2 from in-situ measured pCO_2 in the river section between these two sites.

The DIC concentration increased 5 to 10 times between the spring (2.4 to 2.8 mg L^{-1}) and summer baseflow (18 to 20 mg L^{-1}) and the pH increased by 0.5-0.7 units between spring freshet and summer baseflow (**Fig. 3** and **Fig. S3 A, B** of the Supplement). The DOC concentration ranged from 18 to 25 mg L^{-1} during flood and from 15 to 18 mg L^{-1} during baseflow (**Fig. 3**). There was no systematic variations in DOC concentration over the 834 km of the main stem (20.7 ± 3.6 and $15.0 \pm 1.4 \text{ mg L}^{-1}$ in May and August, respectively); however, it was slightly higher and more variable in the tributaries (22.0 ± 4.0 and $16.5 \pm 7.4 \text{ mg L}^{-1}$, **Fig. S3 C, D**). The SUVA_{254} remained highly stable throughout the seasons for both the tributaries and the main stem (range from 4.2 to 4.9 $\text{L mg C}^{-1} \text{m}^{-1}$, **Table 1**). The POC was 3 times higher during baseflow compared to spring and ranged from 2 to 10 mg L^{-1} (**Fig. 3** and **Fig. S3 E, F**). The total bacterial number ranged from 5.0×10^5 to $8.7 \times 10^5 \text{ cells mL}^{-1}$ for the main stem and tributaries without significant ($p > 0.05$) seasonal variation (**Fig. 3** and **S3 G, H**).

3.2. Diurnal and spatial variation in CO_2 concentration and flux

The diel (day/night) measurements of CO_2 concentrations have been performed on six tributaries of the Ket River during the spring flood period (**Fig. 4**). In two of them (Sochur ad Lopatka), we measured both CO_2 concentration and CO_2 fluxes via floating chambers. Continuous CO_2 concentrations over 10-38 h exhibited a variation between 5 and 25% of the average value. Only in the case of a small tributary

Segondenka (**Fig. 4 E**), when we measured CO₂ over 38 h, there was a local maximum in concentration between 6 and 7 pm during the first and second day of monitoring, without any significant link to the water temperature. The deviation of FCO₂ from the average value over the period of observation in two tributaries (**Fig. 4 A, B**) did not exceed 20%, without any detectable difference between day and night period.

The spatial variation in pCO₂ and FCO₂ were tested during spring time in the flood zone of the Ket River middle course, where the flood zone was connected to the main channel. Regardless of the distance from the main stem and the size of the water body, the variation in pCO₂ and chamber-based fluxes were within 30% of the values measured in the main stem. This suggests that the main stem parameters can be used for upscaling the C emissions to the overall flood plain during May, provided that the water bodies are connected to the rivers. Further tests of spatial variation were performed on selected small tributaries, when we moved 8 to 16 km upstream towards the headwaters and monitored the CO₂ concentration in the river water. There was no sizable trend in CO₂ concentration over several km length of the tributary, consistent with small fluctuations over the hundred km-scale of the main stem (**Fig. S4 A**). Altogether, rather minor spatial and diel variations in both CO₂ concentration and emission fluxes support the chosen sampling strategy and allow reliable extrapolation of obtained results to full surface of lotic waters of the Ket River basin, during open water period.

3.3. Impact of water chemistry and catchment characteristics on CO₂ concentrations and emissions

There were generally no strong correlations between CO₂ and CH₄ and the main parameters of the water column (DOC, DIC, POC, TBC and SUVA (**Table 2**). The CO₂ concentration negatively correlated with O₂ concentration ($R_{\text{Pearson}} = -0.68$, $p < 0.05$) and FCO₂ positively correlated with SUVA₂₅₄ ($R = 0.34$, $p < 0.05$), **Fig. 5 A, B**. Other hydrochemical characteristics of the water column did not impact CO₂ and CH₄ concentration and CO₂ flux. During spring flood, there was no positive correlation between FCO₂ of the river water and various hydrochemical characteristics. During the summer baseflow, there were positive correlations between CO₂ concentration or flux and SUVA and total bacterial number (**Table 2**).

There was a decrease in pCO₂ with an increase in the stream order (**Fig. S5 A**), consistent with negative correlation between pCO₂ and S_{watershed} during the spring (**Table 2**). However, neither FCO₂ nor gas

transfer coefficient exhibited significant link to the stream order (**Fig. S5 B, C**) or the watershed surface area (**Table 2**). Among different landscape factors, only deciduous light needleleaf forest (larch trees) exhibited significant ($p < 0.01$) positive correlations ($0.6 \leq R_{\text{Pearson}} \leq 0.7$) with CO₂ concentration and flux of the Ket River main stem and tributaries, detectable only during the summer baseflow period (**Fig. 5 C**). The peatland and bogs at the watershed exhibited only weak, although positive ($0.2 < R_{\text{Pearson}} < 0.4$), correlation with pCO₂ and FCO₂ (**Table 2**). The other potentially important landscape factors of the river watershed (type of forest, riparian and total aboveground vegetation, recent burns, water bodies) did not significantly impact the CO₂ and CH₄ concentration and measured CO₂ fluxes in the Ket River basin (**Table 2**). The mean annual precipitation (MAP) at the watershed positively correlated with pCO₂ and FCO₂ during the baseflow (**Fig. 5 D**).

Principal Component Analysis (PCA) demonstrated a general lack of control of physico-chemical parameters of the water column and watershed land cover on C emission pattern in the river waters. The PCA identified two factors that had generally low ability to describe the variance (19 and 7%, respectively; **Table S3** of the Supplement). None of the factors acted significantly on dissolved CO₂, CH₄ or CO₂ flux in the river water. The RDA treatment did not provide additional insights into environmental control of C pattern across the rivers and seasons. After normalization, the main result was that the analyses are not statistically significant ($p > 0.05$).

373

374 *3.4. Areal C emissions and export fluxes*

The C emissions ($> 99.5\%$ CO₂, $< 0.5\%$ CH₄) from the lotic waters of the Ket River basin were assessed based on total river water coverage of the Ket watershed in 2019 (856 km², of which 691 km² is seasonal water, according to the Global SDG database). Given that the measurements were performed at the peak of spring flood in 2019, we used the maximal water coverage of the Ket River basin to calculate the emissions during May and June, and baseflow coverage for measurements during July-October period.

For C emission calculation, we used the mean values of FCO₂ of the main stem and the tributaries (1.31 ± 0.81 g C m⁻² d⁻¹ for spring flood; 2.11 ± 1.86 g C m⁻² d⁻¹ for summer-autumn baseflow) which covers full variability of both tributaries and the Ket River main channel (**Table 1, Figure 3**). For the month of July

383 which was not sampled in this work and which represents a transition period between the flood and the
384 baseflow, we used the mean value of May and August ($1.55 \text{ g C m}^{-2} \text{ d}^{-1}$). For the two months of maximal
385 water flow (May - June), the C emission from the whole Ket basin amounts to $68 \pm 42 \text{ Gg}$. When summed up
386 with July ($25 \pm 20 \text{ Gg}$) and summer-autumn baseflow period (August to October) emission ($32 \pm 28 \text{ Gg}$), the
387 total open water season emission flux is 125 Gg . The uncertainty on the total emission over 6 months of the
388 open water period is difficult to quantify but it can be estimated as between 30 and 50 %. This range covers
389 both the uncertainty of the water coverage of the territory (i.e., Krickov et al., 2021) and the seasonal and
390 spatial variations of CO_2 emission in the Ket basin assessed in the present study.

391 Based on yield calculations described in section 2.4, the total annual (excluding ice-covered period)
392 riverine C export from the Ket River basin ($S_{\text{watershed}} = 94,000 \text{ km}^2$) is 0.35 Tg ($3.7 \text{ t C km}^{-2}_{\text{land}} \text{ y}^{-1}$), of which
393 DOC, DIC and POC accounts for 56, 24 and 20%, respectively. Therefore, over the 6 month of open water
394 period, the C emissions from lotic waters of Ket watershed constituted less than 30% of the dissolved and
395 particulate downstream export of carbon.

396

397

398 4. DISCUSSION

399 4.1. Temporal and spatial pattern of CO_2 emissions from the river waters

400 The first important result of the present study is quite low spatial and seasonal variability in both CO_2
401 concentration and emissions, as well as in DOC concentration and aromaticity (reflected by SUVA_{254}) in the
402 main channel (**Fig. 3, S3, Table 1**). The variability in the tributaries was much larger, with differences in
403 dissolved and gaseous C parameters between spring flood and summer-autumn baseflow (**Table S4 A**). While
404 CO_2 concentrations were different between tributaries and the main stem during both flood and baseflow, the
405 CO_2 flux was not different between the main stem and tributaries regardless of season (**Table S4 B**). This,
406 together with lack of diel variations in CO_2 concentrations and emissions during spring period of maximal
407 water coverage (**Fig. 4**), suggest rather stable pattern of CO_2 in the river water, not linked to short-scale
408 processes (primary productivity, photolysis, daily temperature variation). Indeed, negligible primary
409 productivity in the water column may stem from low water temperatures ($9.3 \text{ }^\circ\text{C}$), shallow photic layer of

410 organic-rich waters (DOC of 22 mg L⁻¹) and lack of periphyton activity during high flow of the spring flood.
411 Note that this finding contrasts the recent results of high frequency pCO₂ measurements in temperate rivers
412 that show a 30 % higher nocturnal emission compared to daytime observations due to
413 photosynthesis/respiration cycle (Gómez-Gener et al., 2021b). In the Ambolikha River of Eastern Siberia, a
414 small (S_{watershed} = 121 km²) Arctic stream of continuous permafrost zone, the diel CO₂ cycle exhibited a
415 moderate increase during the day, which was attributed to external lateral sources and photochemical
416 oxidation of terrestrial DOC, rather than in-stream metabolism (Castro-Morales et al., 2022). At the same
417 time, several studies in tropical DOM-rich rivers such as Congo (Borges et al. 2019) have not detected diel
418 variations of CO₂ because aquatic pelagic primary production was low (Descy et al., 2018) due to strong light
419 attenuation in the water column by DOM.

420 Concerning spatial variability of C concentrations and emissions during the spring flood, the pCO₂
421 did not demonstrate sizable variation along the main stem of the Ket River and some of its tributaries, when
422 moving from the mouth to the headwaters. The SUVA also remained highly stable along the river flow. This,
423 together with a lack of FCO₂ correlation with river watershed area during this period (**Table 2**) and the
424 absence of link between the stream's Strahler order and measured FCO₂ and K_T (**Fig. S5 B, C**), suggest
425 relatively modest control of headwater C cycling by 'fresh' unprocessed organic matter from upland mire
426 waters on CO₂ emissions from the Ket River basin. Much stronger control of mire waters is reported in boreal
427 zone of the Northern Europe (Wallin et al., 2013, 2018). Furthermore, our results on the Ket River main stem
428 and tributaries are in contrast to the general view of disproportional importance of headwater streams in
429 overall CO₂ emission from river basins (Li et al., 2021). Thus, across the United States fluvial system, the
430 stream's Strahler order was shown to be important driver of CO₂ evasion from river water surfaces, with
431 lower order streams exhibiting the highest pCO₂ and gas transfer velocity (Butman and Raymond, 2011). A
432 likely explanation is relative low values of gas transfer velocity measured in the small streams of the Ket
433 basin in this study (0.2 - 2.0 m d⁻¹, **Table 1**). Based on a hydraulic model of stream velocity and mean channel
434 slope (Eqn. 4 in Raymond et al., 2012), we calculated the gas transfer velocities in studied rivers as median
435 of 1.02, IQR from 0.27 to 1.52 m d⁻¹, in very good agreement with chamber-measured values for the Ket
436 River main stem and tributaries. Although these calculated values are also consistent with transfer coefficients

for western Siberia calculated by Liu et al. (2022) based on reach-slope and flow velocity (i.e., $K_T \leq 2 \text{ m d}^{-1}$), they are typical of lakes rather than rivers (i.e., Kokic et al., 2015). We believe that low K_T values for the Ket River basin stem from low channel slope (0.2 to 0.7 m km^{-1}) and flow rate (1-2 km h^{-1}), strongly forested and wind-protected river bed without distinct valley due to generally flat orographic context of this part of the WSL (Serikova et al., 2018). Furthermore, due to small size and short fetch of the Ket River and its tributaries (see pictures of typical environments in **Fig. S4 B-D** of the Supplement), extended floodplain zone also contributes to low values of K_T measured in the studied river basin. This is consistent with observations in other flooded regions, where a canopy of vegetation protects the water-air interface from wind stress thus rendering the gas transfer velocity lower compared to open water such as large river (i.e., Foster-Martinez and Variano, 2016; Ho et al., 2018; Abril and Borges, 2019). We therefore warn against the use of high value of transfer velocity, suitable for large Siberian rivers (i.e., Karlsson et al., 2021; Vorobyev et al., 2021), for assessing the emissions in medium and small size, sheltered streams with extensive riparian vegetation. Another important aspect linked to C emissions from flooded forest (notably birch trees, see **Fig. S4 B**) of the floodplain (e.g. Pangala et al., 2017), not investigated in this study.

451

4.2. *Environmental factors possibly controlling CO₂ concentration and emission pattern in the Ket River main stem and tributaries*

Despite sizable variability of pCO_2 in the tributaries, especially during the baseflow, there were no correlations between either pCO_2 or FCO_2 and main hydrochemical parameters of the water column (**Table 2**). The only exception is O_2 concentration, which negatively correlated with pCO_2 during spring flood and both pCO_2 and FCO_2 during summer baseflow (**Fig. 5 A**). This finding suggests potential importance of shallow suboxic riparian flooded zone, meadows and forest, as well as floodplain lakes, in controlling CO_2 build up in the water column due to diffusion from sediments or decaying macrophytes, as it was shown for the floodplain of the Ob River middle course (Krickov et al., 2021). We believe that main reasons of remarkable stability in CO_2 concentrations and emissions and weak environmental control on dissolved and gaseous pattern in the Ket River basin are (1) essentially homogeneous landscapes, lithology and quaternary deposits of the whole river basin (20-25 % bogs, 60-70% forest, 3-5 % riparian zone), and (2) strong

dominance of allochthonous sources in both dissolved and particulate organic matter. Indeed, the SUVA and bacterial number (TBC) positively correlated with both $p\text{CO}_2$ and FCO_2 during summer (**Fig. 5 B; Table 2**), which may indicate non-negligible role of bacterial processing of allochthonous (aromatic) DOC delivered to the water column from wetlands and mires. As such, homogeneous land cover and essentially allochthonous DOC can still lead to variations of CO_2 per stream size, with small systems showing higher values than large systems as predicted conceptually (Hotchkiss et al., 2015) and verified at basin-scale (e.g. Borges et al., 2019). Consistent with this, we observed systematically higher CO_2 concentration and flux in small tributaries [which were fed by mire waters with ‘non-processed’ OM] compared to the main stem (**Table 2**). Furthermore, the positive correlation between mean annual precipitation (MAP) and $p\text{CO}_2$ and FCO_2 during the baseflow (**Table 2, Fig. 5 D**) could reflect the importance of water storage in the mires and wetlands (which also showed positive but less significant correlations, **Table 2**) during the summer time, and progressive release of CO_2 and DOC-rich waters from the wetlands to the streams. Another indirect evidence of the mire water control on CO_2 emission from the river comes from daily CO_2 pattern in a tributary of the Ket River (**Fig. 4 E**). For this relatively small river ($S_{\text{watershed}} = 472 \text{ km}^2$), we noted that there was quite heavy rainfall, between 7 am and 3 pm, prior to the CO_2 peak which was observed at 7 pm. Given that water residence time is very short during spring flood, when the soils are partially frozen, the delivery of allochthonous DOM and elevated CO_2 from adjacent mires could be the cause of observed CO_2 peak. Generally, the terrestrial source controlling CO_2 pattern in the Ket River could be either soil litter leachates (in spring) or bog water (during baseflow, when the river water is substantially derived from wetlands, Alahó et al., 2018a, b). Therefore, the patterns in CO_2 emissions observed in the present study during summer baseflow thus suggest the importance of allochthonous organic matter from the peatland for CO_2 production in the water column and in soils where the degradation of DOC is enhanced by the presence of bacteria. This is consistent with observations in other regions that, during summer-time, numerous processes contribute to increase CO_2 in rivers such as higher temperature stimulating microbial metabolism, longer residence time and enhanced flow paths of soil water (Borges et al. 2018).

A correlation between CO_2 flux during baseflow and the proportion of deciduous needleleaf forest at the watershed (**Fig. 5 C**) may suggest the importance of C cycling by larch trees and their possible control on

491 the delivery of degradable organic matter to the river. Similar control of larch vegetation on riverine CO₂ has
492 been suggested for the Lena River, Eastern Siberia (Vorobyev et al., 2021) although we acknowledge that
493 further observations on contrasted Siberian watersheds are necessary to confirm the observation that larch
494 trees litterfall led to export of degradable OM to the river.

495 In the Ket River basin, the local soil/groundwater effects are expected to be more pronounced during
496 baseflow, due to lower impact of dilution, compared to the spring flood period. The hypothesis of deeper flow
497 path in summer compared to spring is confirmed for the WSL (Frey and McClelland, 2009; Pokrovsky et al.,
498 2015; Serikova et al., 2018) and is supported in this study by a strong increase in DIC concentration between
499 spring and summer (**Fig. 3**). Thus, although the pairwise correlations between parameters do not support any
500 particular mechanism, it is not excluded that OM bio- and photo degradation and local mire water feeding
501 drive FCO₂ in spring, and that deeper flowpaths and DIC export drive the elevated FCO₂ in summer. The
502 latter is consistent with results of analysis of streams and rivers across the contiguous United States, which
503 demonstrated that ~60% of CO₂ evasion is from external sources rather than internal production (Hotchkiss
504 et al., 2015). In view of lack of correlation of CO₂ emissions in the Ket River and tributaries with
505 hydrochemical parameters of the water column, we believe that external source of CO₂ in studied river system
506 represents sizable contribution to total riverine CO₂ evasion across the seasons and sampling sites. In
507 particular, in small peatland streams, the CO₂-rich deep peat/groundwater is known to be the major source of
508 aquatic CO₂ under low flow conditions (Dinsmore and Billett, 2008), whereas in boreal headwater streams
509 of N Sweden the main source of stream CO₂ was inflowing CO₂-rich soil waters (Winterdahl et al., 2016).

510 Another important factor responsible for higher CO₂ production in the water column in summer
511 compared to spring could be POC degradation. The riverine POC is known to be much more biodegradable
512 than DOC (Attermeyer et al., 2018), and the POC concentration in the Ket River basin increased 4-fold
513 between spring and summer (**Table 1**). The origin of summer-time POC and its lability remain elusive, but
514 could be a combination of plankton bloom and mire- or forest-derived DOC coagulation products in the water
515 column (Krickov et al., 2018). Furthermore, pronounced heterogeneity in CO₂ emission during baseflow
516 among tributaries may also reflect the heterogeneity of riverine organic matter which is known to be the
517 maximal during low flow conditions and minimal during high flow (Lynch et al., 2019).

518 The main unexpected result of this study is that none of the physiochemical parameters of the water
 519 column and the land cover factor is sufficiently strong to drive the CO₂ and CH₄ patterns, although they show
 520 pronounced spatial and seasonal variations. Although correlations do not necessary imply causation and some
 521 correlations could be spurious or indirect, this analysis, together with PCA treatment, allow first order
 522 assessment of possible governing factors or dismissing the environmental parameters that do not contribute
 523 in GHG pattern control. A likely explanation is that simultaneous operation of multiple aquatic processes that
 524 include carbon, oxygen, nutrient, and plankton and peryphyton dynamics as well as sediment respiration
 525 control the CO₂ and CH₄ exchanges with the atmosphere, as it is known for boreal lakes and floodplain zones
 526 of the boreal rivers (i.e., Bayer et al., 2019; Zabelina et al., 2021; Krickov et al., 2019). Given that even a
 527 multiparametric statistical treatment (PCA) did no demonstrate sizable explanation capacity of the data set,
 528 we cannot exclude that these potential physico-chemical, microbiological and landscape drivers are working
 529 in different (opposing) directions and have counteracted each other. However, further in-depth analysis of
 530 these interactions requires much better seasonal resolution, ideally over full period of the year, which was
 531 beyond the scope of the present study.

532

533 4.3. Emissions from the Ket River basin compared to downstream export of riverine carbon

534 The estimated C emissions ($> 99.5\%$ C; $< 0.5\%$ CH₄) from the Ket River main channel over 830 km
 535 distance (0.5 to $2.5\text{ g C m}^{-2}\text{ d}^{-1}$) are comparable to those of the Ob River main channel ($1.32 \pm 0.14\text{ g C m}^{-2}\text{ d}^{-1}$
 536 ¹ in the permafrost-free zone; Karlsson et al., 2021). The CO₂ emission in Ket's tributaries (1 to $2\text{ g C m}^{-2}\text{ d}^{-1}$
 537 ¹ in spring; 1 to $5\text{ g C m}^{-2}\text{ d}^{-1}$ in summer) are within the range reported for small rivers and streams of the
 538 permafrost-free zone of western Siberia (0 to $3.6\text{ g C m}^{-2}\text{ d}^{-1}$ in spring; 4 to $9\text{ g C m}^{-2}\text{ d}^{-1}$ in summer; Serikova
 539 et al., 2018), forest and wetland headwater streams of northern Sweden (0.5 to $5\text{ g C m}^{-2}\text{ d}^{-1}$; Gómez-Gener
 540 et al., 2021a), and boreal streams in Canada and Alaska (0.8 to $5.2\text{ g C m}^{-2}\text{ d}^{-1}$, Koprivnjak et al., 2010;
 541 Teodoru et al., 2009; Crawford et al., 2013; Campeau et al., 2014). Total C emissions from the water surfaces
 542 of the Ket River basin assessed in this study ($148\text{ g C-CO}_2\text{ m}^{-2}_{\text{water}}\text{ y}^{-1}$, assuming no emission under ice), when
 543 normalized to the Ket river basin area ($S_{\text{watershed}} = 94,000\text{ km}^2$), amounts to $1.35\text{ g C m}^{-2}_{\text{land}}\text{ y}^{-1}$. Generally
 544 higher land area - specific emissions, comparable or exceeding those of the Ket River, were reported in

545 Québec (1.0 to 4.6 g C m⁻² y⁻¹; Campeau and del Giorgio, 2014; Hutchins et al., 2019; Teodoru et al., 2009),
546 Sweden (1.6 to 8.6 g C m⁻² y⁻¹; Humborg et al., 2010; Jonsson et al., 2007; Lundin et al., 2013; Wallin et al.,
547 2011, 2018) and boreal portions of the Yukon River (7 to 9 g C m⁻² y⁻¹; Striegl et al., 2012; Stackpoole et al.,
548 2017). Possible reasons for these differences could be different areal coverage of the territory by river
549 network, the calculated rather than measured CO₂ fluxes, or the higher gas transfer velocity in the rivers from
550 mountainous regions.

551 The regional assessment of the Ket River basin performed in this study are based on direct chamber
552 measurements of emissions and as such provide rigorous basis for upscaling the CO₂ emissions from currently
553 understudied lotic waters of permafrost-free zone of Western Siberia. The C evasion from the fluvial network
554 of the Ket River assessed in the present work (127 ± 11 Gg y⁻¹, ignoring the emission during the ice breakup
555 in early spring) is 3 times lower than the total (DOC+DIC+POC) downstream export by this river from the
556 same territory (0.35 Tg C y⁻¹). The riverine C yield for the Ket River (3.7 t C km⁻²_{land} y⁻¹) is in agreement
557 with regional C (DOC+DIC) yield by permafrost-free small and medium size rivers of the WSL (3 to 4 t C
558 km⁻²_{land} y⁻¹, Pokrovsky et al., 2020) and with the Ob River in the permafrost-free zone (3.6 t C km⁻²_{land} y⁻¹,
559 Vorobyev et al., 2019). Note that the latter study of the Ob River, which is very similar in the environmental
560 context to the Ket River, included high frequency weekly sampling over several years of monitoring. Thus,
561 the similarity of downstream export fluxes of the Ket and Ob Rivers support the validity of approaches for
562 sampling and C yield calculation employed in the present study. Such high C yields in the southern,
563 permafrost-free part of the WSL stem from essentially inorganic carbon originated from groundwater
564 discharge of carbonate mineral rich reservoirs, abundant in this region (Pokrovsky et al., 2015). At the same
565 time, the organic C yield in rivers of this region is quite low and represents less than 20 % of total C yield
566 (Pokrovsky et al., 2020; Vorobyev et al., 2019). This can explain anomalously low value of C evasion : C
567 export of the Ket River (1 : 3) measured in this work as compared to the average values for permafrost-free
568 zone of Western Siberia (1 : 1, Serikova et al., 2019). Another factor potentially leading to underestimation
569 of C evasion in this study is GIS-based minimal water coverage which does not include seasonal oxbow lakes,
570 flooded forest and temporary water bodies of the floodplain which provide sizable emissions (see Krickov et
571 al., 2021). We also do not exclude that some important hot moments / hot spots of C emission were missed

in our sampling campaign, such as summer baseflow/autumn peaks (Serikova et al., 2019) or stagnant zones of the floodplain in summer (Krickov et al., 2021; Castro-Morales et al., 2021). This calls a need for higher spatial and temporal resolution monitoring of C emission, with special focus on important events across full hydrological continuum.

5. Concluding remarks

Via combination of discrete floating chamber and hydrochemistry and continuous CO₂ concentration measurements over 830 km of large pristine boreal river of western Siberia main channel and its 26 tributaries during the peak of spring flood and the summer-autumn baseflow, we quantified spatial and temporal variations, overall emissions of C (CO₂, CH₄) and export of (DOC, DIC and POC) during the 6 months of open water period. The range of CO₂ and CH₄ concentrations in the main channel and tributaries as well as CO₂ emissions were consistent with other boreal and subarctic regions but demonstrated rather low seasonal and spatial variability. The diel CO₂ flux by floating chambers and continuous pCO₂ measurements in the tributaries of the Ket River during spring flood demonstrated negligible impact of day/night period on the CO₂ concentrations and emission fluxes.

We hypothesize that homogeneous landscape coverage (bog and taiga forest) provide stable allochthonous input of DOM as confirmed by very weak spatial and seasonal variations of DOM aromaticity. Among possible driving factors of CO₂ production in the water column (bio- and photo-degradation of DOC and POC, plankton metabolism), none seems to be sizably important for persistent CO₂ supersaturation and relevant emissions. The landscape factors of the watershed (bog and forest coverage, soil organic carbon stock) of the tributaries and along the main stem did not sizably affected the C concentration and emission pattern across two seasons. We hypothesize that stable terrestrial input of strongly aromatic DOM, shallow photic layer and humic waters of the Ket River basin preclude sizable daily and seasonal variations of C parameters. Punctual discharge of groundwater, resuspension of sediments or shallow subsurface influx from mires and riparian zone may be responsible for small-scale heterogeneities in C emissions and concentrations along the main stem and among the tributaries. These effects are much stronger pronounced during summer

baseflow compared to spring flood. Overall, deeper flow paths in summer compared to spring enhance the DIC discharge within the river bed and the tributaries, thus leading to elevated CO₂ flux in summer. Additional factor responsible for higher CO₂ emission during this season could be mire-originated particulate organic matter (POM) processing in the water column.

The six month open-water period C emissions from the lotic waters of the Ket River basin were sizably lower than the downstream total C export by this river during the same period. We conclude that regional estimations of C balance in lotic systems should be based on a combination of direct chamber measurements, discrete hydrochemical sampling and continuous in-situ monitoring with submersible sensors, at least during two most important hydrological periods of the year which are, for boreal regions, the spring flood and the summer-autumn baseflow. We believe that this is the best trade-off between scientific rigor and logistical feasibility in poorly accessible, pristine and strongly understudied regions.

Acknowledgements.

We acknowledge support from RSF grant 22-17-00253, the TSU Development Program “Priority-2030”, and the Swedish Research Council (grant no. 2016-05275).

Authors contribution.

AL and OP designed the study and wrote the paper; AL, SV, IK and OP performed sampling, analysis and their interpretation; LS performed bacterial assessment and DOC/DIC analysis and interpretation; MK performed landscape characterization of the Ket River basin and calculated water surface area; SK performed hydrological analysis; JK provided analyses of literature data, transfer coefficients for FCO₂ calculations and global estimations of areal emission vs export.

Competing interests.

The authors declare that they have no conflict of interest.

Data availability.

Pokrovsky, O., Lim, A., Krickov, I., Korets, M., Vorobyev, S.: “Ket River hydrochemistry, CO₂ concentration and landscape parameters”, Mendeley Data, V1, doi: 10.17632/snwbkvg6tc.1, 2022.

References

630 Abril, G., Martinez, J.-M., Artigas, L. F., Moreira-Turcq, P., Benedetti, M. F., Vidal, L., Meziane, T., Kim,
631 J.-H., Bernardes, M. C., Savoye, N., Deborde, J., Albéric, P., Souza, M. F. L., Souza, E. L., and Roland,
632 F.: Amazon river carbon dioxide outgassing fueled by wetlands, *Nature*, 505, 395-398,
633 <https://doi.org/10.1038/nature12797>, 2014.

634 Abril, G. and Borges, A. V.: Ideas and perspectives: Carbon leaks from flooded land: do we need to replumb
635 the inland water active pipe? *Biogeosciences*, 16, 769–784, <https://doi.org/10.5194/bg-16-769-2019>,
636 2019.

637 Ala-Aho, P., Soulsby, C., Pokrovsky, O. S., Kirpotin, S. N., Karlsson, J., Serikova, S., Manasypov, R., Lim,
638 A., Krickov, I., Kolesnichenko, L. G., Laudon, H., and Tetzlaff, D.: Permafrost and lakes control river
639 isotope composition across a boreal Arctic transect in the Western Siberian lowlands, *Environ. Res.*
640 *Lett.*, 13, <https://doi.org/10.1088/1748-9326/aaa4fe>, 2018a.

641 Ala-aho, P., Soulsby, C., Pokrovsky, O. S., Kirpotin, S. N., Karlsson, J., Serikova, S., Vorobyev, S. N.,
642 Manasypov, R. M., Loiko, S., and Tetzlaff, D.: Using stable isotopes to assess surface water source
643 dynamics and hydrological connectivity in a high-latitude wetland and permafrost influenced
644 landscape, *J. Hydrol.*, 556, 279–293, <https://doi.org/10.1016/j.jhydrol.2017.11.024>, 2018b.

645 Alin, S. R., Rasera, M. D. F. F. L., Salimon, C. I., Richey, J. E., Holtgrieve, G. W., Krusche, A. V., and
646 Snidvongs, A.: Physical controls on carbon dioxide transfer velocity and flux in low-gradient river
647 systems and implications for regional carbon budgets, *J. Geophys. Res. Biogeosciences*, 116,
648 <https://doi.org/10.1029/2010JG001398>, 2011.

649 Allen, G. H. and Pavelsky, T. M.: Global extent of rivers and streams, *Science*, 361, 585–588,
650 <https://doi.org/10.1126/science.aat0636>, 2018.

651 Almeida, R. M., Pacheco, F. S., Barros, N., Rosi, E., and Roland, F.: Extreme floods increase CO₂ outgassing
652 from a large Amazonian river, *Limnol. Oceanogr.*, 62, 989–999, <https://doi.org/10.1002/lno.10480>,
653 2017.

654 Amaral, J. H. F., Borges, A. V., Melack, J. M., Sarmiento, H., Barbosa, P. M., Kasper, D., Melo, M. L., de
655 Fex Wolf, D., da Silva, J. S., and Forsberg, B. R.: Influence of plankton metabolism and mixing
656 depth on CO₂ dynamics in an Amazon floodplain lake, *Sci. Total. Env.* 630, 1381-1393,
657 <https://doi.org/10.1016/j.scitotenv.2018.02.331>, 2018.

658 Amaral, J. H. F., Melack, J. M., Barbosa, P. M., Borges, A. V., Kasper, D., Cortes, A. C., Zhou, W.,
659 MacIntyre, S., and Forsberg, B. R.: Inundation, hydrodynamics and vegetation influences carbon
660 dioxide concentrations in Amazon floodplain lakes, *Ecosystem*, 25(4), 911–930,
661 <https://doi.org/10.1007/s10021-021-00692-y>, 2022.

662 Attermeyer, K., Catalán, N., Einarsdottir, K., Freixa, A., Groeneveld, M., Hawkes, J. A., Bergquist, J., and
663 Tranvik, L. J.: Organic carbon processing during transport through boreal inland waters: particles as
664 important sites, *J. Geophys. Res. Biogeosciences*, 123, 2412–2428,
665 <https://doi.org/10.1029/2018JG004500>, 2018.

666 Bartalev, S. A., Egorov, V. A., Ershov, D. V., Isaev, A. S., Lupyan, E. A., Plotnikov, D. E., and Uvarov, I.
667 A.: Remote mapping of vegetation land cover of Russia based on data of MODIS spectroradiometer,
668 *Mod. Probl. Earth Remote Sens. Space*, 8, 285–302, 2018.

669 Bastviken, D., Sundgren, I., Natchimuthu, S., Reyier, H., and Gålfalk, M.: Technical Note: Cost-efficient
670 approaches to measure carbon dioxide (CO₂) fluxes and concentrations in terrestrial and aquatic
671 environments using mini loggers, *Biogeosciences*, 12, 3849–3859, [https://doi.org/10.5194/bg-12-3849-](https://doi.org/10.5194/bg-12-3849-2015)
672 2015, 2015.

673 Bayer, T. K., Gustafsson, E., Brakebusch, M., and Beer, C.: Future carbon emission from boreal and
674 permafrost lakes are sensitive to catchment organic carbon loads, *J. Geophys. Res. Biogeosciences*,
675 124, 1827-1848, <https://doi.org/10.1029/2018JG004978>, 2019.

676 Borges, A. V., Darchambeau, F., Lambert, T., Bouillon, S., Morana, C., Brouyère, S., Hakoun, V., Jurado,
677 A., Tseng, H.-C., Descy, J.-P., and Roland, F. A. E.: Effects of agricultural land use on fluvial carbon
678 dioxide, methane and nitrous oxide concentrations in a large European river, the Meuse (Belgium),
679 *Sci. Total Environ.*, 610-611, 342-355, <https://doi.org/10.1016/j.scitotenv.2017.08.047>, 2018.

680 Borges, A. V., Darchambeau, F., Teodoru, C. R., Marwick, T. R., Tammooh, F., Geeraert, N., Omengo, F. O.,
681 Guérin, F., Lambert, T., Morana, C., Okuku, E., and Bouillon, S.: Globally significant greenhouse gas

emissions from African inland waters, *Nature Geoscience*, 8, 637–642, <https://doi.org/10.1038/NNGEO2486>, 2015.

Busmann, I.: Distribution of methane in the Lena Delta and Buor-Khaya Bay, Russia, *Biogeosciences*, 10, 4641–4652, <https://doi.org/10.5194/bg-10-4641-2013>, 2013.

Butman, D. and Raymond, P.: Significant efflux of carbon dioxide from streams and rivers in the United States, *Nature Geoscience*, 4, 839–842. <https://doi.org/10.1038/ngeo1294>, 2011.

Cai, W.-J. and Wang, Y.: The chemistry, fluxes, and sources of carbon dioxide in the estuarine waters of the Satilla and Altamaha Rivers, Georgia, *Limnol. Oceanogr.*, 43, 657–668, <https://doi.org/10.4319/lo.1998.43.4.0657>, 1998.

Campeau, A. and del Giorgio, P. A.: Patterns in CH₄ and CO₂ concentrations across boreal rivers: Major drivers and implications for fluvial greenhouse emissions under climate change scenarios, *Glob. Change Biol.*, 20, 1075–1088, <https://doi.org/10.1111/gcb.12479>, 2014.

Campeau, A., Lapierre, J.-F., Vachon, D., and del Giorgio, P. A.: Regional contribution of CO₂ and CH₄ fluxes from the fluvial network in a lowland boreal landscape of Québec, *Glob. Biogeochem. Cycles*, 28, 57–69, <https://doi.org/10.1002/2013GB004685>, 2014.

Castro-Morales, K., Canning, A., Körtzinger, A., Göckede, M., Küsel, K., Overholt, W. A., Wichard, T., Redlich, S., Arzberger, S., Kolle, O., and Zimov, N.: Effects of reversal of water flow in an Arctic floodplain river on fluvial emissions of CO₂ and CH₄, *J. Geophys. Res. Biogeosciences*, 127, e2021JG006485, <https://doi.org/10.1029/2021JG006485>, 2022.

Chadburn, S. E., Krinner, G., Porada, P., Bartsch, A., Beer, C., Beletti Marchesini, L., Boike, J., Ekici, A., Elberling, B., Friberg, T., Hugelius, G., Johansson, M., Kuhry, P., Kutzbach, L., Langer, M., Lund, M., Parmentier, F.-J. W., Peng, S., Van Huissteden, K., Wang, T., Westermann, S., Zhu, D., and Burke, E. J.: Carbon stocks and fluxes in the high latitudes: using site-level data to evaluate Earth system models, *Biogeosciences*, 14, 5143–5169, <https://doi.org/10.5194/bg-14-5143-2017>, 2017.

Cooper, L. W., McClelland, J. W., Holmes, R. M., Raymond, P. A., Gibson, J. J., Guay, C. K., and Peterson, B. J.: Flow-weighted values of runoff tracers ($\delta^{18}\text{O}$, DOC, Ba, alkalinity) from the six largest Arctic rivers, *Geophys. Res. Lett.*, 35, L18606, <https://doi.org/10.1029/2008GL035007>, 2008.

Crawford, J. T., Striegl, R. G., Wickland, K. P., Dornblaser, M. M., and Stanley, E. H.: Emissions of carbon dioxide and methane from a headwater stream network of interior Alaska, *J. Geophys. Res. Biogeosciences*, 118, 482–494, <https://doi.org/10.1002/jgrg.20034>, 2013.

Crawford, J. T., Loken, L. C., Casson, N. J., Smith, C., Stone, A. G., and Winslow, L. A.: High-speed limnology: using advanced sensors to investigate spatial variability in biogeochemistry and hydrology, *Environ. Sci. Technol.*, 49, 442–450, <https://doi.org/10.1021/es504773x>, 2015.

Crawford, J. T., Stanley, E. H., Dornblaser, M. M., and Striegl, R. G.: CO₂ time series patterns in contrasting headwater streams of North America, *Aquatic Sciences*, 79, 473–486, <https://doi.org/10.1007/s00027-016-0511-2>, 2016a.

Crawford, J. T., Loken, L. C., Stanley, E. H., Stets, E. G., Dornblaser, M. M., and Striegl, R. G.: Basin scale controls on CO₂ and CH₄ emissions from the Upper Mississippi River, *Geophys. Res. Lett.*, 43, 1973–1979, <https://doi.org/10.1002/2015GL067599>, 2016b.

Crawford, J. T., Butman, D. E., Loken, L. C., Stadler, P., Kuhn, C., and Striegl, R. G.: Spatial variability of CO₂ concentrations and biogeochemistry in the Lower Columbia River, *Inland Waters*, 7, 417–427, <https://doi.org/10.1080/20442041.2017.1366487>, 2017.

Dawson, J. J., Billett, M. F., Hope, D., Palmer, S. M., and Deacon, C.: Sources and sinks of aquatic carbon in a peatland stream continuum, *Biogeochemistry*, 70, 71–92, 2004.

Descy, J. P., Darchambeau, F., Lambert, T., Stoyneva, M. P., Bouillon, S., and Borges, A. V.: Phytoplankton dynamics in the Congo River, *Freshwater Biology*, 62, 87–101, <https://doi.org/10.1111/fwb.12851>, 2017.

DelDuco, E. M. and Xu, Y. J.: Dissolved carbon transport and processing in North America’s largest swamp river entering the Northern Gulf of Mexico, *Water*, 11, 1395, <https://doi.org/10.3390/w11071395>, 2019.

Denfeld, B. A., Frey, K. E., Sobczak, W. V., Mann, P. J., and Holmes, R. M.: Summer CO₂ evasion from streams and rivers in the Kolyma River basin, north-east Siberia, *Polar Res.*, 32, 19704, <https://doi.org/10.3402/polar.v32i0.19704>, 2013.

734 Dinsmore, K. J. and Billett, M. F.: Continuous measurement and modeling of CO₂ losses from a peatland
735 stream during stormflow events, *Water Resour. Res.*, 44, W12417,
736 <https://doi.org/10.1029/2008WR007284>, 2008.

737 Dinsmore, K. J., Billett, M. F., and Dyson, K. E.: Temperature and precipitation drive temporal variability in
738 aquatic carbon and GHG concentrations and fluxes in a peatland catchment, *Glob. Change Biol.*, 19,
739 2133–2148, <https://doi.org/10.1111/gcb.12209>, 2013.

740 Feng, X., Vonk, J. E., Dongen, B. E. V., Gustafsson, Ö., Semiletov, I. P., Dudarev, O. V., Wang, Z.,
741 Montluçon, D. B., Wacker, L., and Eglinton, T. I.: Differential mobilization of terrestrial carbon pools
742 in Eurasian Arctic river basins, *Proc. Natl. Acad. Sci. U. S. A.*, 110, 14168–14173,
743 <https://doi.org/10.1073/pnas.1307031110>, 2013.

744 Foster-Martinez, M. R. and Variano, E. A.: Air-water gas exchange by waving vegetation stems, *J. Geophys.*
745 *Res. Biogeosciences*, 121, 1916–1923, <https://doi.org/10.1002/2016JG003366>, 2016.

746 Frey, K. E. and Smith, L. C.: How well do we know northern land cover? Comparison of four global
747 vegetation and wetland products with a new ground-truth database for West Siberia. *Glob. Biogeochem.*
748 *Cycles*, 21, GB1016. <https://doi.org/10.1029/2006GB002706>, 2007.

749 Frey, K. E. and McClelland, J. W.: Impacts of permafrost degradation on arctic river biogeochemistry,
750 *Hydrol. Process.*, 23, 169–182, <https://doi.org/10.1002/hyp.7196>, 2009.

751 Gómez-Gener, L., Rocher-Ros, G., Battin, T., Cohen, M. J., Dalmagro, H. J., Dinsmore, K. J., Drake, T. W.,
752 Duvert, C., Enrich-Prast, A., Horgby, Å., Johnson, M. S., Kirk, L., Machado-Silva, F., Marzolf, N. S.,
753 McDowell, M. J., McDowell, W. H., Miettinen, H., Ojala, A. K., Peter, H., Pumpanen, J., Ran, L.,
754 Riveros-Iregui, D. A., Santos, I. R., Six, J., Stanley, E. H., Wallin, M. B., White, S. A., and Sponseller,
755 R. A.: Global carbon dioxide efflux from rivers enhanced by high nocturnal emissions, *Nat. Geosci.*,
756 1–6, <https://doi.org/10.1038/s41561-021-00722-3>, 2021a.

757 Gómez-Gener, L., Hotchkiss, E. R., Laudon, H., and Sponseller, R. A.: Integrating discharge-concentration
758 dynamics across carbon forms in a boreal landscape, *Water Resour. Res.*, 57, e2020WR028806,
759 <https://doi.org/10.1029/2020WR028806>, 2021b.

760 Griffin, C. G., McClelland, J. W., Frey, K. E., Fiske, G., and Holmes, R. M.: Quantifying CDOM and DOC
761 in major Arctic rivers during ice-free conditions using Landsat TM and ETM+ data, *Remote Sens.*
762 *Environ.*, 209, 395–409, <https://doi.org/10.1016/j.rse.2018.02.060>, 2018.

763 Harris, I., Jones, P. d., Osborn, T. j., and Lister, D. h.: Updated high-resolution grids of monthly climatic
764 observations – the CRU TS3.10 Dataset, *Int. J. Climatol.*, 34, 623–642,
765 <https://doi.org/10.1002/joc.3711>, 2014.

766 Ho, D. T., Engel, V. C., Ferrón, S., Hickman, B., Choi, J., and Harvey, J. W.: On factors influencing air-water
767 gas exchange in emergent wetlands, *J. Geophys. Res. Biogeosciences*, 123, 178–192,
768 <https://doi.org/10.1002/2017JG004299>, 2018.

769 Holmes, R. M., McClelland, J. W., Peterson, B. J., & Tank S. E. et al. Seasonal and annual fluxes of nutrients
770 and organic matter from large rivers to the Arctic Ocean and surrounding seas, *Estuaries and Coasts*,
771 35, 369–382. doi: 10.1007/s12237-011-9386-6, 2012.

772 Holmes, R. M., Coe, M. T., Fiske, G. J., Gurtovaya, T., McClelland, J. W., Shiklomanov, A. I., Spencer, R.
773 G. M., Tank, S. E., and Zhulidov, A. V.: Climate change impacts on the hydrology and biogeochemistry
774 of Arctic Rivers, in: *Climatic Changes and Global warming of Inland Waters: Impacts and Mitigation*
775 *for Ecosystems and Societies*, edited by: Goldman, C. R., Kumagi, M., and Robarts, R. D., John Wiley
776 and Sons, 1–26, 2013.

777 Hotchkiss, E. R., Hall Jr, R. O., Sponseller, R. A., Butman, D., Klaminder, J., Laudon, H., Rosvall, M., and
778 Karlsson, J.: Sources of and processes controlling CO₂ emissions change with the size of streams and
779 rivers, *Nat. Geosci.*, 8, 696–699, <https://doi.org/10.1038/ngeo2507>, 2015.

780 Hugelius, G., Tarnocai, C., Broll, G., Canadell, J. G., Kuhry, P., and Swanson, D. K.: The northern
781 circumpolar soil carbon database: Spatially distributed datasets of soil coverage and soil carbon storage
782 in the northern permafrost regions, *Earth Syst. Sci. Data*, 5, 3–13, [https://doi.org/10.5194/essd-5-3-](https://doi.org/10.5194/essd-5-3-2013)
783 2013, 2013.

784 Humborg, C., Mörrth, C.-M., Sundbom, M., Borg, H., Blenckner, T., Giesler, R., and Ittekkot, V.: CO₂
785 supersaturation along the aquatic conduit in Swedish watersheds as constrained by terrestrial

786 respiration, aquatic respiration and weathering, *Glob. Change Biol.*, 16, 1966–1978,
787 <https://doi.org/10.1111/j.1365-2486.2009.02092.x>, 2010.

788 Hutchins, R. H. S., Prairie, Y. T., and del Giorgio, P. A.: Large-Scale Landscape Drivers of CO₂, CH₄, DOC,
789 and DIC in Boreal River Networks, *Glob. Biogeochem. Cycles*, 33, 125–142,
790 <https://doi.org/10.1029/2018GB006106>, 2019.

791 Hutchins, R. H. S., Tank, S. E., Olefeldt, D., Quinton, W. L., Spence, C., Dion, N., Estop-Aragonés, C., and
792 Mengistu, S. G.: Fluvial CO₂ and CH₄ patterns across wildfire-disturbed ecozones of subarctic Canada:
793 Current status and implications for future change, *Glob. Change Biol.*, 26, 2304–2319,
794 <https://doi.org/10.1111/gcb.14960>, 2020.

795 Johnson, M. S., Billett, M. F., Dinsmore, K. J., Wallin, M., Dyson, K. E., and Jassal, R. S.: Direct and
796 continuous measurement of dissolved carbon dioxide in freshwater aquatic systems—method and
797 applications, *Ecohydrology*, 3, 68–78, <https://doi.org/10.1002/eco.95>, 2009.

798 Jonsson, A., Algesten, G., Bergström, A.-K., Bishop, K., Sobek, S., Tranvik, L. J., and Jansson, M.:
799 Integrating aquatic carbon fluxes in a boreal catchment carbon budget, *J. Hydrol.*, 334, 141–150,
800 <https://doi.org/10.1016/j.jhydrol.2006.10.003>, 2007.

801 Karlsson, J., Serikova, S., Vorobyev, S. N., Rocher-Ros, G., Denfeld, B., and Pokrovsky, O. S.: Carbon
802 emission from Western Siberian inland waters, *Nat. Commun.*, 12, 825, [https://doi.org/10.1038/s41467-](https://doi.org/10.1038/s41467-021-21054-1)
803 021-21054-1, 2021.

804 Kokic, J., Wallin, M. B., Chmiel, H. E., Denfeld, B. A., and Sobek, S.: Carbon dioxide evasion from
805 headwater systems strongly contributes to the total export of carbon from a small boreal lake catchment,
806 *J. Geophys. Res. Biogeosciences*, 120, 13–28, <https://doi.org/10.1002/2014JG002706>, 2015.

807 Koprivnjak, J.-F., Dillon, P. J., and Molot, L. A.: Importance of CO₂ evasion from small boreal streams, *Glob.*
808 *Biogeochem. Cycles*, 24, GB4003, <https://doi.org/10.1029/2009GB003723>, 2010.

809 Krickov, I. V., Lim, A. G., Manasypov, R. M., Loiko, S. V., Shirokova, L. S., Kirpotin, S. N., Karlsson, J.,
810 and Pokrovsky, O. S.: Riverine particulate C and N generated at the permafrost thaw front: Case study
811 of western Siberian rivers across a 1700 km latitudinal transect, *Biogeosciences*, 6867–6884,
812 <https://doi.org/10.5194/bg-15-6867-2018>, 2018.

813 Krickov, I. V., Serikova, S., Pokrovsky, O. S., Vorobyev, S. N., Lim, A. G., Siewert, M. B., and Karlsson, J.:
814 Sizable carbon emission from the floodplain of Ob River, *Ecol. Indic.*, 131, 108164,
815 <https://doi.org/10.1016/j.ecolind.2021.108164>, 2021.

816 Lauerwald, R., Laruelle, G. G., Hartmann, J., Ciais, P., and Regnier, P. A. G.: Spatial patterns in CO₂ evasion
817 from the global river network, *Glob. Biogeochem. Cycles*, 29, 534–554,
818 <https://doi.org/10.1002/2014GB004941>, 2015.

819 Leith, F. I., Dinsmore, K. J., Wallin, M. B., Billett, M. F., Heal, K. V., Laudon, H., Öquist, M. G., and Bishop,
820 K.: Carbon dioxide transport across the hillslope–riparian–stream continuum in a boreal headwater
821 catchment, *Biogeosciences*, 12, 1881–1892, <https://doi.org/10.5194/bg-12-1881-2015>, 2015.

822 Leng, P., Li, Z., Zhang, Q., Li, F., and Koschorreck, M.: Fluvial CO₂ and CH₄ in a lowland agriculturally
823 impacted river network: Importance of local and longitudinal controls, *Environ. Pollution* 303, Art No
824 119125, <https://doi.org/10.1016/j.envpol.2022.119125>, 2022.

825 Li, M., Peng, C., Zhang, K., Xu, L., Wang, J., Yang, Y., Li, P., Liu, Z., and He, N.: Headwater stream
826 ecosystem: an important source of greenhouse gases to the atmosphere, *Water Res.*, 190, 116738,
827 <https://doi.org/10.1016/j.watres.2020.116738>, 2021.

828 Liu S., Kuhn, C., Amatulli, G., Aho, K., Butman, D.E., Allen, G.H., Lin, P., Pan, M., Yamazaki, D.,
829 Brinkerhoff, C., Gleason, C., Xia, X., and Raymond, P.A.: The importance of hydrology in routing
830 terrestrial carbon to the atmosphere via global streams and rivers, *PNAS* 119 No. 11, e2106322119;
831 <https://doi.org/10.1073/pnas.2106322119>, 2022.

832 Lobbes, J. M., Fitznar, H. P., and Kattner, G.: Biogeochemical characteristics of dissolved and particulate
833 organic matter in Russian rivers entering the Arctic Ocean, *Geochim. Cosmochim. Acta*, 64, 2973–
834 2983, [https://doi.org/10.1016/S0016-7037\(00\)00409-9](https://doi.org/10.1016/S0016-7037(00)00409-9), 2000.

835 Lorke A., Bodmer, P., Noss, C., Alshboul, Z., Koschorreck, M., Somlai-Haase, C., Bastviken, D., Flury, S.,
836 McGinnis, D.F., Maeck, A., Müller, D., and Premke, K.: Technical note: drifting versus anchored
837 flux chambers for measuring greenhouse gas emissions from running waters, *Biogeosciences*, 12,
838 7013–7024, <https://doi.org/10.5194/bg-12-7013-2015>, 2015.

839 Lundin, E. J., Giesler, R., Persson, A., Thompson, M. S., and Karlsson, J.: Integrating carbon emissions from
 840 lakes and streams in a subarctic catchment, *J. Geophys. Res. Biogeosciences*, 118, 1200–1207,
 841 <https://doi.org/10.1002/jgrg.20092>, 2013.
 842 Lynch, L. M., Sutfin, N. A., Fegcl, T. S., Boot, C. M., Covino, T. P., and Wallenstein, M. D.: River channel
 843 connectivity shifts metabolite composition and dissolved organic matter chemistry, *Nat. Commun.*, 10,
 844 459, <https://doi.org/10.1038/s41467-019-08406-8>, 2019.
 845 Marie, D., Partensky, F., Vaulot, D., and Brussaard, C.: Enumeration of Phytoplankton, Bacteria, and Viruses
 846 in Marine Samples, *Curr. Protoc. Cytom.*, 10, 11111–11115,
 847 <https://doi.org/10.1002/0471142956.cy1111s10>, 1999.
 848 Pangala, S., Enrich-Prast, A., Basso, L., Peixoto, R. B., Bastviken, D., Hornibrook, E. R. C., Gatti, L. V.,
 849 Ribeiro, H., et al.: Large emissions from floodplain trees close the Amazon methane budget, *Nature*
 850 552, 230–234, <https://doi.org/10.1038/nature24639>, 2017.
 851 Pekel, J.-F., Cottam, A., Gorelick, N., and Belward, A. S.: High-resolution mapping of global surface water
 852 and its long-term changes, *Nature*, 540, 418–422, <https://doi.org/10.1038/nature20584>, 2016.
 853 Pokrovsky, O. S., Manasypov, R. M., Loiko, S., Shirokova, L. S., Krickov, I. A., Pokrovsky, B. G.,
 854 Kolesnichenko, L. G., Kopysov, S. G., Zemtsov, V. A., Kulizhsky, S. P., Vorobyev, S. N., and Kirpotin,
 855 S. N.: Permafrost coverage, watershed area and season control of dissolved carbon and major elements
 856 in western Siberian rivers, *Biogeosciences*, 12, 6301–6320, <https://doi.org/10.5194/bg-12-6301-2015>,
 857 2015.
 858 Pokrovsky, O. S., Manasypov, R. M., Kopysov, S. G., Krickov, I. V., Shirokova, L. S., Loiko, S. V., Lim, A.
 859 G., Kolesnichenko, L. G., Vorobyev, S. N., and Kirpotin, S. N.: Impact of permafrost thaw and climate
 860 warming on riverine export fluxes of carbon, nutrients and metals in Western Siberia, *Water (MDPI)*,
 861 12, 1817, <https://doi.org/10.3390/w12061817>, 2020.
 862 Pokrovsky, O. S., Manasypov, R. M., Chupakov, A. V., and Kopysov, S.: Element transport in the Taz River,
 863 western Siberia, *Chemical Geology*, 614, Art No 121180
 864 <https://doi.org/10.1016/j.chemgeo.2022.121180>, 2022.
 865 Ran, L., Lu, X. X., Richey, J. E., Sun, H., Han, J., Yu, R., Liao, S., and Yi, Q.: Long-term spatial and temporal
 866 variation of CO₂ partial pressure in the Yellow River, China, *Biogeosciences*, 12, 921–932,
 867 <https://doi.org/10.5194/bg-12-921-2015>, 2015.
 868 Ran, L., Lu, X. X., and Liu, S.: Dynamics of riverine CO₂ in the Yangtze River fluvial network and their
 869 implications for carbon evasion, *Biogeosciences*, 14, 2183–2198, [https://doi.org/10.5194/bg-14-2183-](https://doi.org/10.5194/bg-14-2183-2017)
 870 2017, 2017.
 871 Raymond, P. A., McClelland, J. W., Holmes, R. M., Zhulidov, A. V., Mull, K., Peterson, B. J., Striegl, R. G.,
 872 Aiken, G. R., and Gurtovaya, T. Y.: Flux and age of dissolved organic carbon exported to the Arctic
 873 Ocean: A carbon isotopic study of the five largest arctic rivers, *Glob. Biogeochem. Cycles*, 21, GB4011,
 874 <https://doi.org/10.1029/2007GB002934>, 2007.
 875 Raymond, P. A., Zappa, C.J., Butman, D., Bott, T.L., Potter, J., Mulholland, P., Laursen, A. E., McDowell,
 876 W. H., and Newbold, D.: Scaling the gas transfer velocity and hydraulic geometry in streams and small
 877 rivers. *Limnology and Oceanography: Fluids and Environments*, 2(1), 41–53.
 878 <https://doi.org/10.1215/21573689-1597669>, 2012.
 879 Raymond, P. A., Hartmann, J., Lauerwald, R., Sobek, S., McDonald, C., Hoover, M., Butman, D., Striegl,
 880 R., Mayorga, E., Humborg, C., Kortelainen, P., Dürr, H., Meybeck, M., Ciais, P., and Guth, P.: Global
 881 carbon dioxide emissions from inland waters, *Nature*, 503, 355–359,
 882 <https://doi.org/10.1038/nature12760>, 2013.
 883 Rehder, Z., Zaplavnova, A., and Kutzbach, L.: Identifying drivers behind spatial variability of methane
 884 concentrations in East Siberian ponds. *Front. Earth Sci.* 9, Art No 617662, doi:
 885 10.3389/feart.2021.617662, 2021.
 886 Rocher-Ros, G., Sponseller, R. A., Lidberg, W., Mörtz, C.-M., and Giesler, R.: Landscape process domains
 887 drive patterns of CO₂ evasion from river networks, *Limnol. Oceanogr. Lett.*, 4, 87–95,
 888 <https://doi.org/10.1002/lol2.10108>, 2019.
 889 Santoro, M., Beer, C., Cartus, O., Schmullius, C., Shvidenko, A., McCallum, I., Wegmüller, U., and
 890 Wiesmann, A.: The BIOMASAR algorithm: An approach for retrieval of forest growing stock volume

891 using stacks of multi-temporal SAR data, in: Proceedings of ESA Living Planet Symposium, Bergen,
892 Norway, 28 June – 2 July, 2010.

893 Schneider, C. L., Herrera, M., Raisle, M. L., Suarez, E., and Riveros-Iregui, D. A.: Carbon Dioxide (CO₂)
894 Fluxes from terrestrial and aquatic environments in a high altitude tropical catchment. *J Geophys.*
895 *Res. Biogeosciences* 125, e2020JG005844. <https://doi.org/10.1029/2020JG005844>, 2020.

896 Semiletov, I. P., Pipko, I. I., Shakhova, N. E., Dudarev, O. V., Pugach, S. P., Charkin, A. N., McRoy, C. P.,
897 Kosmach, D., and Gustafsson, Ö.: Carbon transport by the Lena River from its headwaters to the Arctic
898 Ocean, with emphasis on fluvial input of terrestrial particulate organic carbon vs. carbon transport by
899 coastal erosion, *Biogeosciences*, 8, 2407–2426, <https://doi.org/10.5194/bg-8-2407-2011>, 2011.

900 Serikova, S., Pokrovsky, O. S., Ala-Aho, P., Kazantsev, V., Kirpotin, S. N., Kopysov, S. G., Krickov, I. V.,
901 Laudon, H., Manasypov, R. M., Shirokova, L. S., Soulsby, C., Tetzlaff, D., and Karlsson, J.: High
902 riverine CO₂ emissions at the permafrost boundary of Western Siberia, *Nat. Geosci.*, 11, 825–829,
903 <https://doi.org/10.1038/s41561-018-0218-1>, 2018.

904 Serikova, S., Pokrovsky, O. S., Laudon, H., Krickov, I. V., Lim, A. G., Manasypov, R. M., and Karlsson, J.:
905 High carbon emissions from thermokarst lakes of Western Siberia, *Nat. Commun.*, 10,
906 <https://doi.org/10.1038/s41467-019-09592-1>, 2019.

907 Spawn, S. A., Dunn, S. T., Fiske, G. E., Natali, S. M., Schade, J. D., and Zimov, N. S.: Summer methane
908 ebullition from a headwater catchment in Northeastern Siberia, *Inland Waters*, 5(3), 224–230, DOI:
909 10.5268/IW-5.3.845, 2015.

910 Stackpoole, S. M., Butman, D. E., Clow, D. W., Verdin, K. L., Gaglioti, B. V., Genet, H., and Striegl, R. G.:
911 Inland waters and their role in the carbon cycle of Alaska, *Ecol. Appl.*, 27, 1403–1420,
912 <https://doi.org/10.1002/eap.1552>, 2017.

913 Stanley, E. H., Casson, N. J., Christel, S. T., Crawford, J. T., Loken, L. C., and Oliver, S.K.: The ecology of
914 methane in streams and rivers: patterns, controls, and global significance, *Ecol. Monogr.* 86, 146–171,
915 2016.

916 Striegl, R. G., Dornblaser, M. M., McDonald, C. P., Rover, J. A., and Stets, E. G.: Carbon dioxide and
917 methane emissions from the Yukon River system, *Glob. Biogeochem. Cycles*, 26, GB0E05,
918 <https://doi.org/10.1029/2012GB004306>, 2012.

919 Teodoru, C. R., del Giorgio, P. A., Prairie, Y. T., and Camire, M.: Patterns in pCO₂ in boreal streams and
920 rivers of northern Quebec, Canada, *Glob. Biogeochem. Cycles*, 23, GB2012,
921 <https://doi.org/10.1029/2008GB003404>, 2009.

922 Tranvik, L., Cole, J. J., and Prairie, Y. T.: The study of carbon in inland waters-from isolated ecosystems to
923 players in the global carbon cycle, *Limnol. Oceanogr. Lett.*, 3, 41–48,
924 <https://doi.org/10.1002/lol2.10068>, 2018.

925 Vachon, D., Sponseller, R. A., and Karlsson, J.: Integrating carbon emission, accumulation and transport in
926 inland waters to understand their role in the global carbon cycle, *Glob. Change Biol.*, 27, 719–727,
927 <https://doi.org/10.1111/gcb.15448>, 2021.

928 Villa, J. A., Ju, Y., Yazbeck, T., Waldo, S., Wrighton, K. C., and Bohrer, G.: Ebullition dominates methane
929 fluxes from the water surface across different ecohydrological patches in a temperate freshwater marsh
930 at the end of the growing season, *Sci. Total Environ.* 767, 144498, 2021.

931 Vorobyev, S. N., Pokrovsky, O. S., Kolesnichenko, L. G., Manasypov, R. M., Shirokova, L. S., Karlsson, J.,
932 and Kirpotin, S. N.: Biogeochemistry of dissolved carbon, major, and trace elements during spring flood
933 periods on the Ob River, *Hydrol. Process.*, 33, 1579–1594, <https://doi.org/10.1002/hyp.13424>, 2019.

934 Vorobyev, S. N., Karlsson, J., Kolesnichenko, Y. Y., Korets, M. A., and Pokrovsky, O. S.: Fluvial carbon
935 dioxide emission from the Lena River basin during the spring flood, *Biogeosciences*, 18, 4919–4936,
936 <https://doi.org/10.5194/bg-18-4919-2021>, 2021.

937 Wallin, M. B., Öquist, M. G., Buffam, I., Billett, M. F., Nisell, J., and Bishop, K. H.: Spatiotemporal
938 variability of the gas transfer coefficient (KCO₂) in boreal streams: Implications for large scale
939 estimates of CO₂ evasion, *Glob. Biogeochem. Cycles*, 25, GB3025,
940 <https://doi.org/10.1029/2010GB003975>, 2011.

941 Wallin, M. B., Grabs, T., Buffam, I., Laudon, H., Ågren, A., Öquist, M. G., and Bishop, K.: Evasion of CO₂
942 from streams – The dominant component of the carbon export through the aquatic conduit in a boreal
943 landscape, *Glob. Change Biol.*, 19, 785–797, <https://doi.org/10.1111/gcb.12083>, 2013.

- Wallin, M. B., Campeau, A., Audet, J., Bastviken, D., Bishop, K., Kokic, J., Laudon, H., Lundin, E., Löfgren, S., Natchimuthu, S., Sobek, S., Teutschbein, C., Weyhenmeyer, G. A., and Grabs, T.: Carbon dioxide and methane emissions of Swedish low-order streams—a national estimate and lessons learnt from more than a decade of observations, *Limnol. Oceanogr. Lett.*, 3, 156–167, <https://doi.org/10.1002/lol2.10061>, 2018.
- Wild, B., Andersson, A., Bröder, L., Vonk, J., Hugelius, G., McClelland, J. W., Song, W., Raymond, P. A., and Gustafsson, Ö.: Rivers across the Siberian Arctic unearth the patterns of carbon release from thawing permafrost, *PNAS*, 116, 10280–10285, <https://doi.org/10.1073/pnas.1811797116>, 2019.
- Winterdahl, M., Wallin, M. B., Karlsen, R. H., Laudon, H., Öquist, M., and Lyon, S. W.: Decoupling of carbon dioxide and dissolved organic carbon in boreal headwater streams, *J. Geophys. Res. Biogeosciences*, 121, 2630–2651, <https://doi.org/10.1002/2016JG003420>, 2016.
- Yamamoto, S., Alcauskas, J. B., and Crozier, T. E.: Solubility of methane in distilled water and seawater. *J. Chem. Eng. Data*, 21, 78–80, 1976.
- Zabelina, S. V., Shirokova, L. S., Klimov, S. I., Chupakov, A. V., Lim, A. G., Polishchuk, Y. M., Polishchuk, V. M., Bogdanov, A. N., Muratov, I. N., Gueren, F., Karlsson, J., and Pokrovsky, O. S.: Seasonal and spatial variations of CO₂ and CH₄ concentrations and fluxes in surface waters of frozen peatlands (NE Europe): morphological and hydrochemical control, *Limnology Oceanography*, 66 (S1), S216–S230, doi: 10.1002/lno.11560, 2021.
- Zolkos, S., Tank, S. E., Striegl, R. G., and Kokelj, S. V.: Thermokarst effects on carbon dioxide and methane fluxes in streams on the Peel Plateau (NWT, Canada), *J. Geophys. Res. Biogeosciences*, 124, 1781–1798, <https://doi.org/10.1029/2019JG005038>, 2019.

986
987
988
989
990
991
992
993

994
995
996
997
998
999
1000
1001
1002
1003
1004
1005
1006
1007
1008
1009
1010
1011
1012
1013
1014
1015
1016

Table 1. Measured hydrochemical and GHG exchange parameters in the Ket River main stem and tributaries (average $\pm$ s.d.; (*n*) is number of measurements). The FCO₂ and K_T are chamber-measured CO₂ flux and gas transfer velocity, respectively, whereas diffusive CH₄ flux (FCH₄) was calculated using chamber-specific transfer coefficient.

Parameter	unit	Tributaries		Main stem	
		Flood (<i>n</i> =26)	Base flow (<i>n</i> =12)	Flood (<i>n</i> =7)	Base flow (<i>n</i> =6)
Water temperature	°C	9.48±2.25	14.9±1.24	9.06±1.59	16.5±0.54
pH		6.31±0.45	6.71±0.57	6.2±0.43	7.29±0.26
Dissolved O ₂	mg L ⁻¹	8.53±1.26	8.02±1.13	8.85±0.83	8.78±0.18
Specific Conductivity	µS cm ⁻¹	40.7±22.7	126.9±62.1	39±14.9	181±36.8
DIC	mg L ⁻¹	2.83±2.58	17.8±10.4	2.43±1.49	20.5±5.22
DOC	mg L ⁻¹	21.7±3.94	15.7±7.04	21.9±4.28	16.6±3.57
SUVA ₂₅₄	L mg C ⁻¹ m ⁻¹	4.34±0.33	4.9±0.66	4.29±0.18	4.26±0.52
PON	mg L ⁻¹	0.08±0.06	0.64±0.27	0.1±0.07	0.96±0.22
POC	mg L ⁻¹	2.41±1.17	8±2.36	2.55±1.2	9.49±1.98
TBC	*10 ⁵ cells ml ⁻¹	5.89±3.26	8.69±3.21	5.95±2.83	4.94±2.15
K _T	m d ⁻¹	0.53±0.38	1.21±0.52	0.77±0.55	1.22±0.37
FCO ₂	g C m ⁻² d ⁻¹	1.3±0.76	2.63±2.15	1.35±1.08	1.16±0.5
pCO ₂	µatm	2880±680	4000±1500	2400±330	2520±980
FCH ₄	mmol C m ⁻² d ⁻¹	0.39±0.95	1.38±1.21	0.06±0.05	0.95±0.88
CH ₄	µmol L ⁻¹	0.65±0.66	1.17±0.81	0.17±0.01	0.86±0.91

Table 2. Pearson correlation coefficients of measured FCO₂, CO₂, and CH₄ concentration with hydrochemical parameters of the water column (DOC, SUVA, particulate organic carbon and nitrogen, total bacterial cells) and landscape parameters of the tributaries and the main stem of the Ket River. Significant (p < 0.05) values are labeled by asterisk.

	all seasons			spring flood			summer baseflow		
	CH ₄	CO ₂	FCO ₂	CH ₄	CO ₂	FCO ₂	CH ₄	CO ₂	FCO ₂
Hydrochemical parameters									
pH	0.2	-0.1	-0.2	-0.1	0.1	-0.2	0.0	-0.6*	-0.6*
Dissolved O ₂	-0.1	-0.7*	-0.1	0.0	-0.8*	0.1	-0.2	-0.8*	-0.7*
Specific conductivity	0.3	0.0	0.1	-0.2	0.0	0.1	0.2	-0.3	-0.6*
DIC	0.3	0.0	0.0	-0.1	0.0	0.1	0.2	-0.4	-0.7*
DOC	-0.1	0.0	0.1	0.3	0.0	-0.1	-0.2	-0.1	0.2
SUVA ₂₅₄	0.1	0.2	0.3	0.4	-0.3	0.1	-0.2	0.5*	0.6*
PON	0.1	-0.1	0.2	-0.2	-0.4*	0.2	-0.4	-0.5*	-0.5
POC	0.1	-0.1	0.2	-0.2	-0.4*	0.1	-0.3	-0.3	0.1
TBC	0.2	0.2	0.1	0.3	-0.2	-0.1	0.0	0.5*	0.5*
Climatic characteristics									
MAAT	0.2	0.0	-0.5*	0.1	0.0	-0.4*	0.2	0.1	-0.5
MAP	0.0	0.3*	0.5*	0.1	0.0	0.3	0.1	0.6*	0.7*
Land-cover characteristics									
Watershed area	-0.3	-0.3*	0.2	-0.4	-0.5*	0.0	-0.2	-0.1	0.5
Dark Needleleaf Forest	0.1	0.0	-0.3	0.1	0.0	-0.3	0.2	-0.1	-0.2
Light Needleleaf Forest	0.3*	0.4*	0.2	0.4	0.2	0.0	0.4	0.7*	0.6*
Broadleaf Forest	-0.3	-0.4*	0.1	-0.5*	-0.4	0.1	-0.3	-0.6*	-0.2
Mixed Forest	0.0	-0.2	-0.3	0.1	-0.1	-0.3	-0.1	-0.4	-0.4
Peatlands and bogs	0.0	0.2	0.3	-0.1	0.0	0.2	0.1	0.2	0.4
Riparian Vegetation	-0.1	0.0	-0.1	-0.2	0.1	0.0	-0.2	-0.2	-0.5
Grassland	0.1	-0.1	0.0	-0.1	-0.2	0.1	0.3	0.0	-0.5
Recent Burns	-0.1	-0.1	0.2	-0.1	-0.2	0.1	-0.3	0.1	0.4
Water Bodies	-0.2	-0.1	0.3	-0.3	-0.3	0.2	-0.2	-0.1	0.3

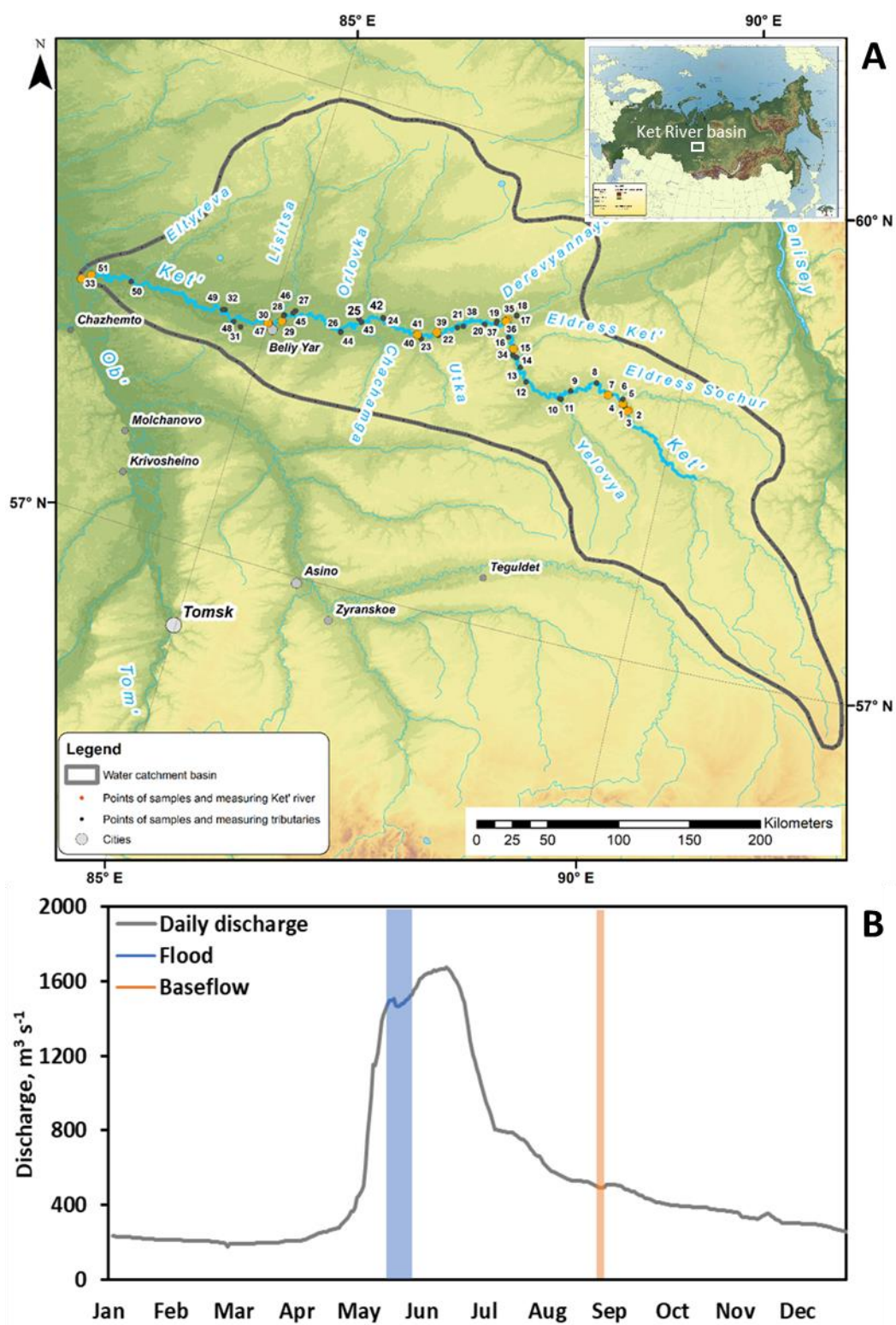
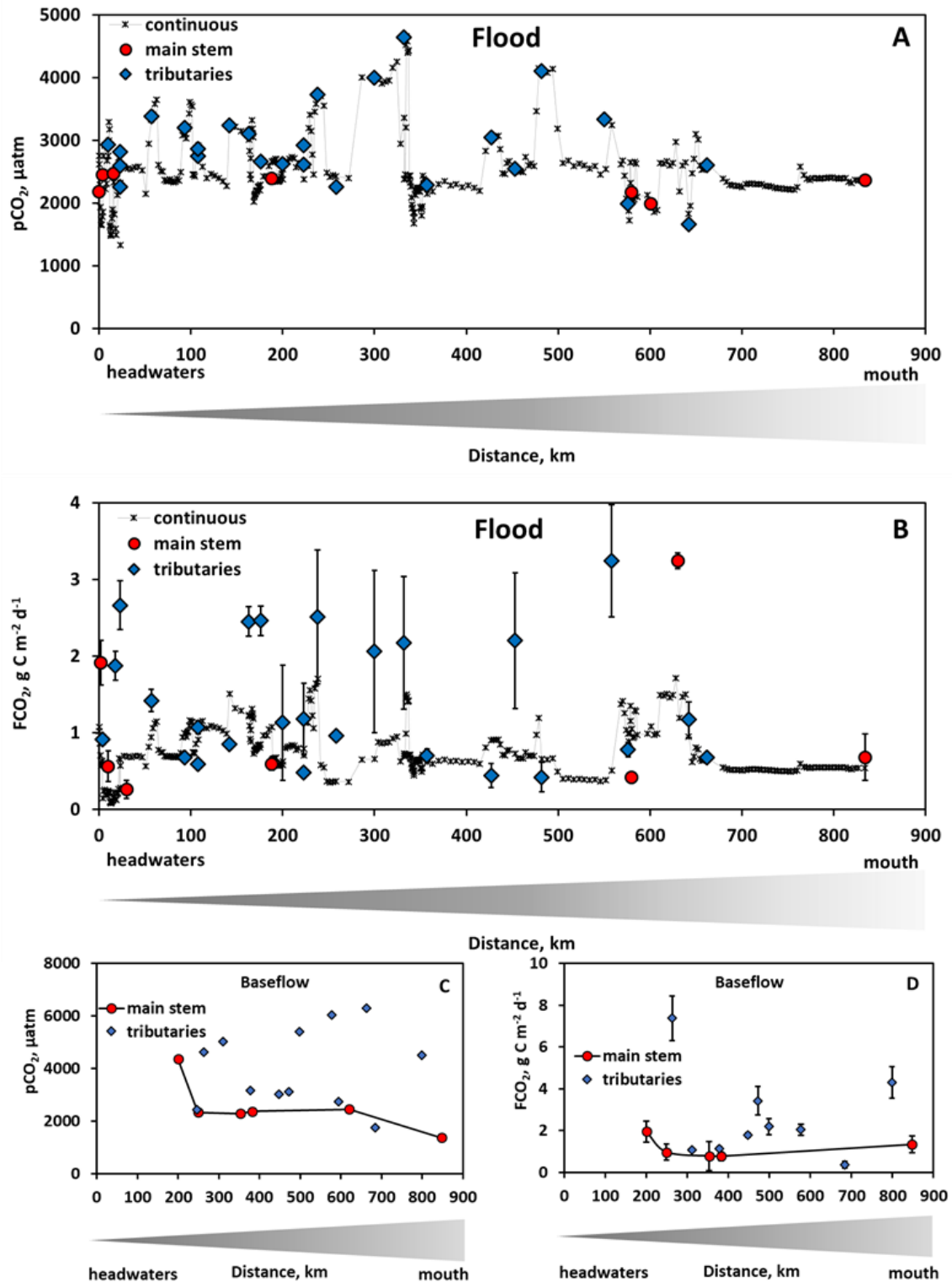


Fig. 1. A: Map of the studied Ket River watershed with continuous pCO_2 measurements in the main stem. **B:** Daily discharge (Q) at the gauging station of the Ket mouth, Rodionovka, in 2019. Highlighted in blue and orange are two sampling campaigns of this study, spring flood and summer-autumn baseflow.



1042

1043 **Figure 2.** The measured pCO₂ (A, C) and CO₂ fluxes (B, D) during spring flood (A, B) and summer
 1044 baseflow (C, D) of the Ket River main stem and tributaries (over the 830 km distance, from the headwaters to
 1045 the mouth (left to right)). The symbols represent discrete in situ pCO₂ (Vaissala) and FCO₂ (floating
 1046 chambers) measurements of the main stem (red circles) and tributaries (blue diamonds). Continuous in-situ
 1047 pCO₂ measurements and calculated FCO₂ are available only for the main stem in spring (black crosses). For
 1048 the latter, we used an average value of measured gas transfer velocity (K_T) between two chamber sites
 1049 (separated by a distance of 50 to 100 km) to calculate the FCO₂ from in-situ measured pCO₂ in the river
 1050 section between these two sites. Note that during summer baseflow, the water level did not allow reaching
 1051 the headwaters of the Ket River (first 0-200 km on the river course).

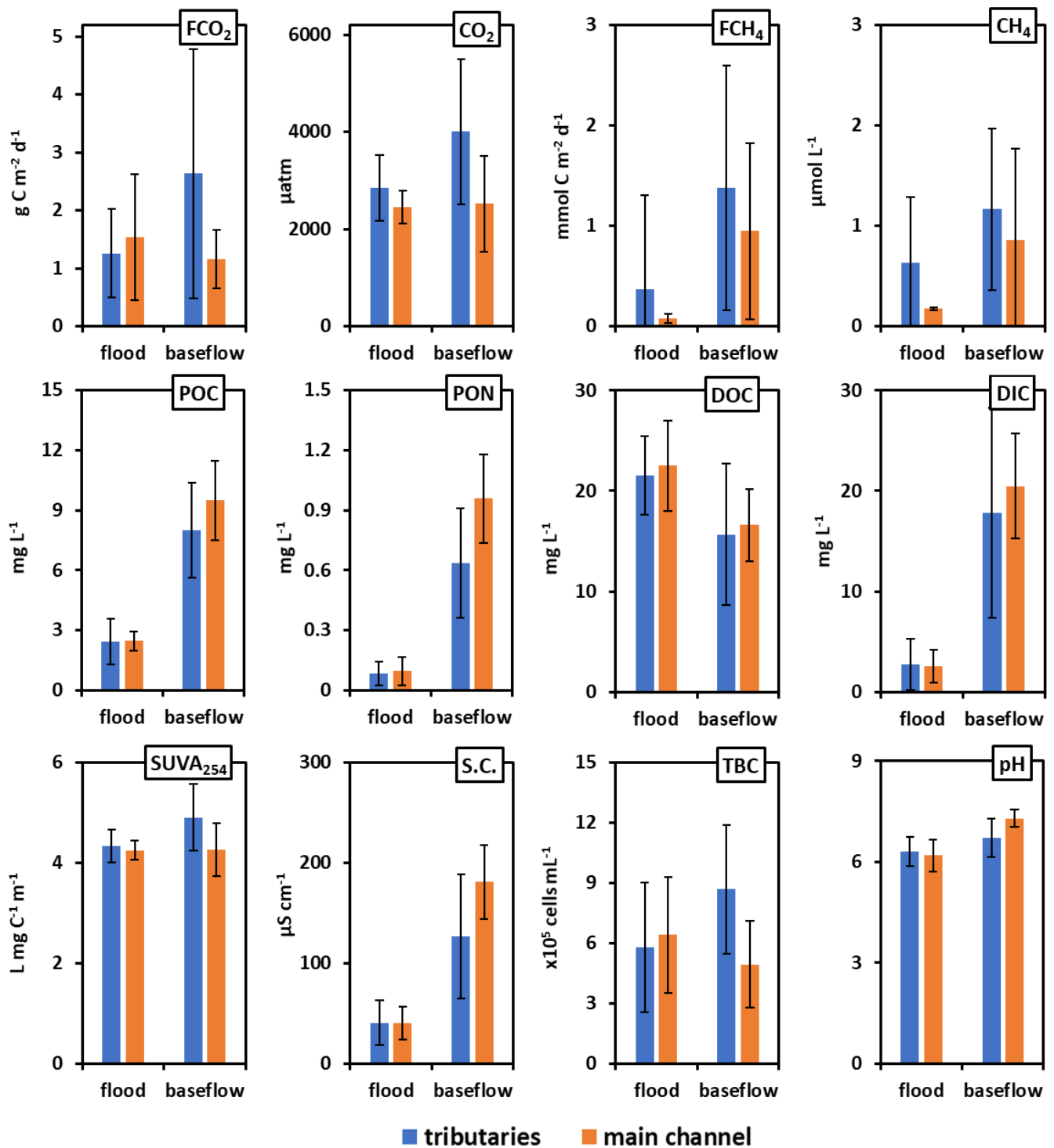
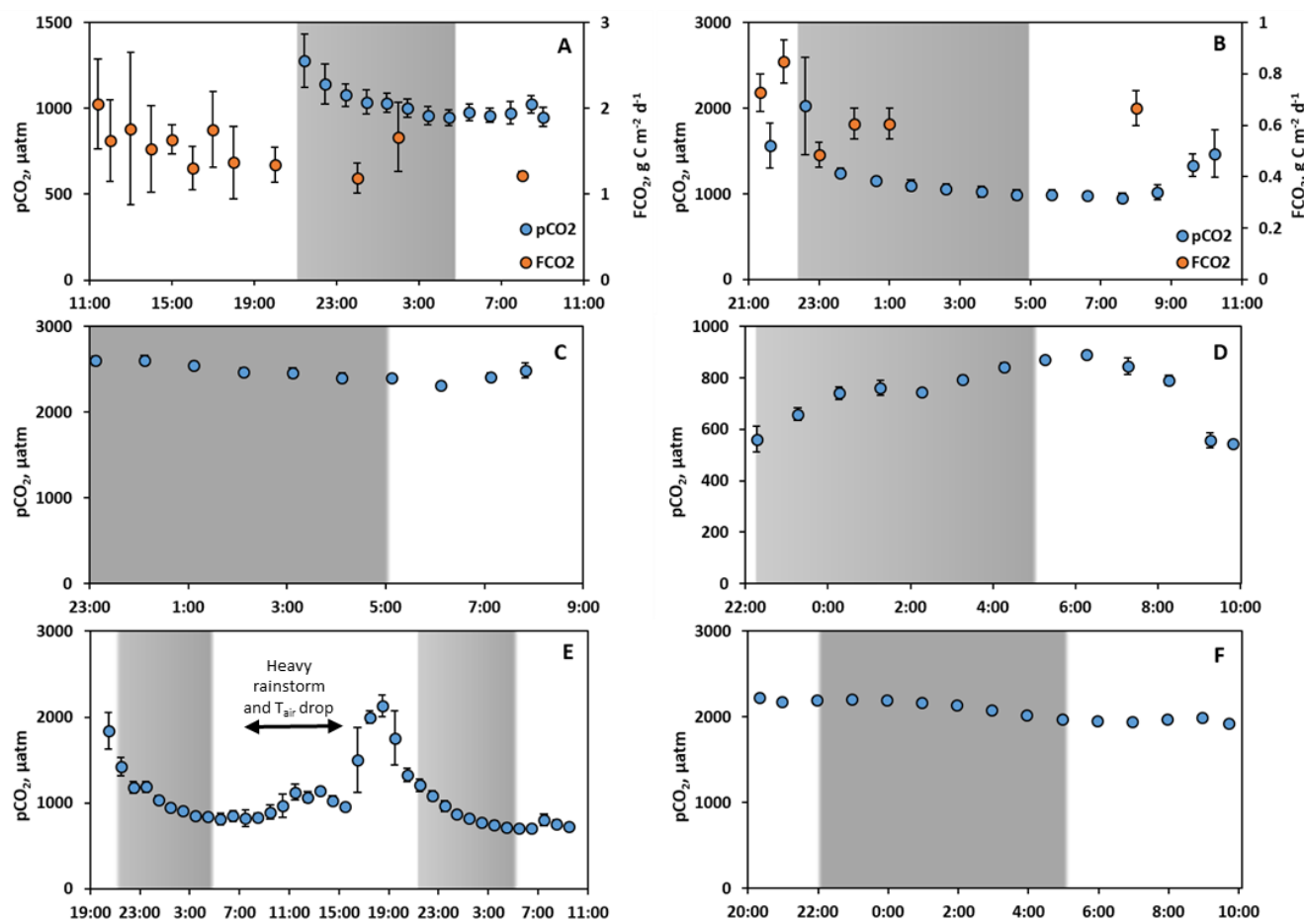


Figure 3. Mean ($\pm$ s.d.) GHG concentration and chamber-measured fluxes (FCO₂), hydrochemical parameters, particulate organic carbon and nitrogen (POC and PON, respectively) and total bacteria count (TBC), in the main channel (orange column) and the tributaries (blue column) of the Ket River in spring flood and summer (early fall) baseflow.



1063

1064

1065

1066

1067

Figure 4. Continuous pCO₂ concentration (A-F, blue circles) and chamber-based fluxes (A, B) measured during spring flood period in tributaries (A Sochur No 3, B Lopatka No 8, C Derevyannaya No 12, D Ob river entrance, E Segondenka No 26) and in the Ket River main stem (middle course) near Stepanovka village (F) including night time measurements (shaded area). The measurement frequency was one per hour. Variations of water temperature were within the range of 0.3 to 0.6 °C and did not exhibit significant correlations with pCO₂ and FCO₂. Note that, for the small river Segondenka (S_{watershed} = 472 km²), where the CO₂ peak was observed at 7 pm (E), there was quite heavy rainfall between 7 am and 3 pm.

1075

1076

1077

1078

1079

1084
1085
1086
1087
1088
1089
1090
1091
1092
1093
1094
1095
1096
1097

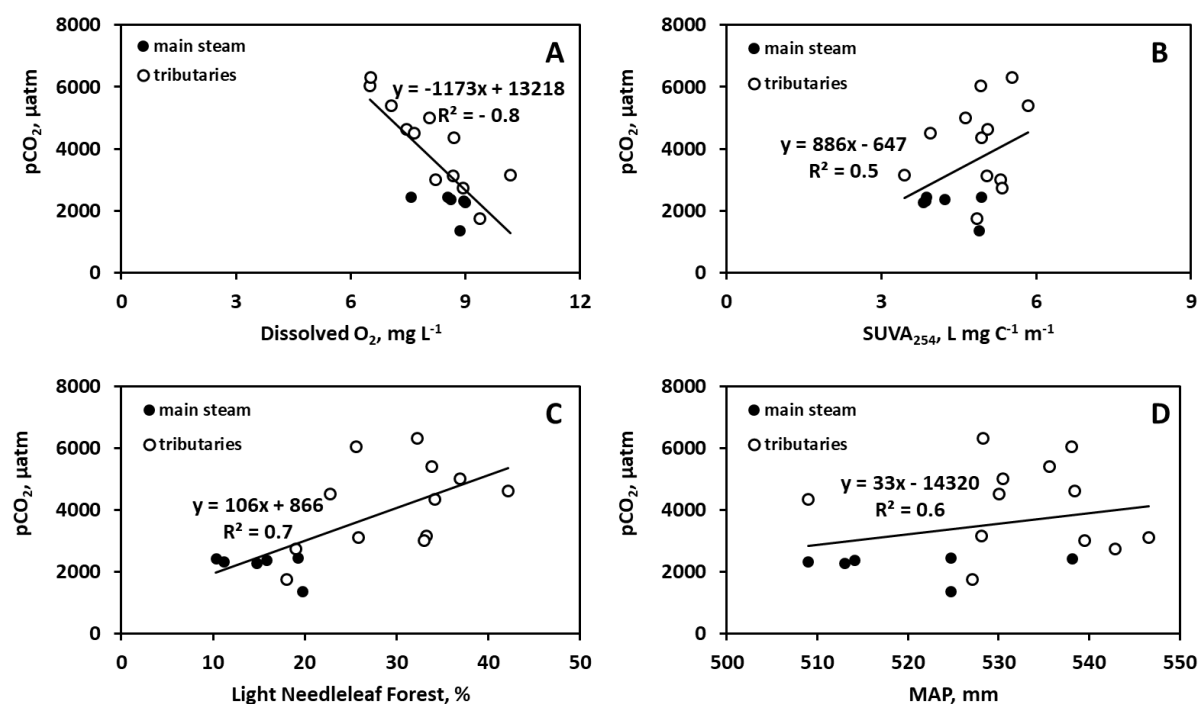


Figure 5. Significant ($p < 0.05$) control of dissolved oxygen (**A**), SUVA₂₅₄ (**B**), light needleleaf forest (**C**), and mean annual precipitation (**D**) on CO₂ concentration in the Ket River and tributaries during summer baseflow.