



1 **Wetting and drying cycles, organic matter and gypsum play a key role in structure formation and**
2 **stability of sodic Vertisols**

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13 **Abstract**

14 In the natural environment, soils undergo wetting and drying (WD) cycles due to precipitation
15 and evapotranspiration. The WD cycles have a profound impact on soil physical, chemical, and
16 biological properties and drive the development of structure in soils. Degraded soils are often
17 lacking structure and the effect of organic amendments and WD cycles on structure formation
18 of these soils is poorly understood. The aim of this study was to evaluate the role of biotic and
19 abiotic factors on aggregate formation and stabilisation of sodic soils after the addition of
20 gypsum and organic amendments (feedlot manure, chicken manure, lucerne pallets, and
21 anionic poly acrylamide). Amended soils were incubated at 25°C over four WD cycles, with
22 assessment of soil microbial respiration, electrical conductivity, pH, sodium adsorption ratio
23 (SAR), aggregate stability in water (ASWAT), aggregate size distribution, and mean weight
24 diameter. Our results demonstrate that WD cycles can improve aggregate stability after the
25 addition of amendments in sodic Vertisols, but this process depends on the type of organic
26 amendment. Lucerne pellets resulted in highest soil microbial respiration, proportions of large
27 macroaggregates, and mean weight diameter. In contrast, dispersion was significantly reduced



28 when soils were treated with chicken manure, whilst PAM only had a transient effect on
29 aggregate stability. When these organic amendments were applied together with gypsum, the
30 stability of aggregates was further enhanced, and dispersion became negligible after the second
31 WD cycle. The formation and stability of small macroaggregates was less dependent on the
32 type of organic amendments and more dependent on WD cycles as the proportion of small
33 macroaggregates also increased in control soils after four WD cycles, highlighting the role of
34 WD cycles as one of the key factors that improves aggregation and stability of sodic Vertisols.
35 **Key words:** Sodic soils, organic amendments, aggregate stability, mean weight diameter,
36 wetting and drying cycles

37 1 Introduction

38 Soils are subjected to seasonal and daily variations of water and temperature. These variations
39 effect the physical, biological, and chemical properties of soils. Natural variations in soil water
40 content can lead to wetting and drying (WD) cycles, which are affected by rainfall, solar
41 radiation, capillarity, wind and condensation (Utomo and Dexter 1982) and evapotranspiration.
42 In most terrestrial ecosystems, surface soils experience rapid changes in soil water content with
43 a longer dry period followed by a relatively rapid re-wetting (Borken and Matzner 2009).
44 Southern Queensland (Australia) usually has hot wet summers with cool dry winters, and soils
45 of this region and other semi-arid areas are particularly susceptible to drying and re-wetting
46 stresses due to the infrequency of rainfall. In the field, most soils experience more than one
47 WD cycle throughout the year. These WD cycles have a substantial influence on soil
48 aggregation and structure stabilisation because of its direct effect on hydration of minerals, and
49 indirect effect on soil microbial activity (Denef *et al.* 2001; Cosentino *et al.* 2006).
50 Soil aggregation is an important mechanism for stabilisation of soil organic matter (Six *et al.*
51 2000a). Furthermore, it also supports soil fertility as it reduces soil erosion and controls soil
52 aeration, water infiltration, hydraulic conductivity and nutrient cycling (Oades 1984; Six *et al.*



53 2000b). Soil aggregation is caused by various aggregate stabilising compounds, which work
54 together at different spatial scales (Tisdall and Oades 1982). Aggregate formation also depends
55 on soil microbial activity as it influences the production of binding materials such as microbial
56 exudates and hyphae (Rahman *et al.* 2017). Aggregate stability often exhibits seasonal and
57 inter-annual variability that is controlled mainly by rainfall, temperature, humidity, insolation,
58 and organic matter (Perfect *et al.* 1990).

59 Aggregation is important to rehabilitate sodic soils. Sodic soils are generally characterised as
60 soils with poor structure which makes those soils difficult to work when wet or dry (Rengasamy
61 and Olsson 1991). Poor structural stability of sodic soils restricts seedling emergence and root
62 growth which directly limits crop growth and development, and indirectly affects plant
63 nutrition by limiting water infiltration, nutrient uptake, and gaseous exchange (Curtin and
64 Naidu 1998). Rehabilitation of sodic soils requires an understanding of the complex
65 interactions between biological and physico-chemical factors that contributes to soil structure
66 formation and stability (Oades 1993; Nelson and Oades 1998).

67 In soils containing shrink-swell (smectitic) clays WD cycles can lead to more intensive changes
68 in soil structure as they can affect aggregation process directly through physical processes
69 (Utomo and Dexter 1982; Deneff *et al.* 2001). However, there is no consensus in the literature
70 regarding the role of WD cycles in improving soil aggregation and its stability. For example,
71 Rahman *et al.* (2018) reported that WD cycles improved (increased) the mean weight diameter
72 (MWD, a proxy of aggregate stability measurement) of Vertisols when treated with maize
73 straw after four WD cycles. In a similar manner, WD cycles also increased the proportion of
74 large macro aggregates of smectite Vertisols (Bravo-Garza *et al.* 2009). However, in contrast,
75 Peng *et al.* (2011), while comparing swelling and non-swelling soils, reported that WD cycles
76 decreased the MWD of swelling soils but not of non-swelling soils. Due to the physical
77 disturbance caused by WD cycles, Six *et al.* (2000b) found that repeated WD cycles decreased



78 the MWD, with this being related to the loss of soil organic matter. These apparently
79 contradictory results can potentially be explained on the basis of the initial conditions of the
80 soil, such as the physical conditions of aggregates, organic matter content and quality, and the
81 intensities and duration of the WD cycles (Cosentino *et al.* 2006).

82 It is well established that the addition of organic matter effects aggregate stability within a
83 period of days to weeks due to the stimulation of microbial activity (Six *et al.* 2004), depending
84 upon the quality and quantity of organic matter (Monnier, 1965 cited in Abiven *et al.* (2009)).
85 Studies investigating the effects of WD cycles on microbial activity were inconclusive, with
86 results differing due to varying experimental designs, incubation period and temperatures, soil
87 properties, and treatments applied. Xiang *et al.* (2008) found that multiple WD cycles increased
88 the microbial respiration of grassland soils up to 6-fold when compared to the soil that remained
89 wet. Drying and rewetting of these soils gave a new pulse of respiration with each WD cycle,
90 but when these soils were kept at constant water content, microbial respiration decreased to
91 almost zero. In contrast, however, Rahman *et al.* (2018) reported that repeated WD cycles in
92 Vertisols decreased soil respiration significantly but that the magnitude of this decrease became
93 smaller over the time.

94 Although the interaction of biotic and abiotic factors and its effect on aggregate stability and
95 formation is complex and inconsistent, little effort has been put into studying the underlying
96 mechanisms, particularly in sodic Vertisols. Furthermore, there is little information available
97 on the relationship between aggregate formation and stability following repeated WD cycles
98 after addition of gypsum and organic amendments. Thus, in the present study we used two
99 sodic Vertisols, with the aim to: i) investigate the effect of WD cycles on microbial respiration,
100 ii) assess the effects of WD cycles on aggregate formation and stability, and iii) determine how
101 many WD cycles are needed to improve aggregate stability. We hypothesised that i) organic
102 amendments enhance the formation of large aggregates and their stability increase with WD



103 cycles, and ii) the addition of gypsum reduces dispersion and aggregate stability is enhanced
104 with the introduction of WD cycles compared with a continuous wet regime.

105 **2 Materials and methods**

106 **2.1 Soils**

107 The two soils used in this experiment were collected from a farm located in southern
108 Queensland near Goondiwindi (28.54° S, 150.30° E), Australia. The soils were being cropped
109 using a maize (*Zea mays*) and wheat (*Triticum aestivum*) rotation system. Both soils were
110 classified as sodic Vertisols according to the FAO-World Reference Base (2015) (Vertosols in
111 the Australian Soil Classification). Soils were collected to a depth of 10 cm at two locations
112 from the cropped land. These sites were selected as they were in close proximity to each other
113 but were slightly different in regard to their physical behaviour (Table 1) with Soil 1 being
114 more dispersive than Soil 2. Soil cores for bulk density determinations were also collected
115 separately from each site and dried at 105°C for several days. After collection, soils were air
116 dried, and the larger clods gently broken into smaller aggregates by hand. The soils were then
117 passed through a 10 mm sieve to remove stones, visible roots, and plant litter. For chemical
118 analysis, a portion of each soil was sieved to <2 mm. Soil pH (ISO, 2005) and electrical
119 conductivity (EC) (ISO, 1994) were measured in 1:5 soil: water suspensions. Particle size
120 analysis was performed using the pipette method (Day, 1965), and field water capacity (-10
121 kPa) was measured using a pressure plate apparatus (Cassel and Nielsen, 1986). Exchangeable
122 cations (Ca^{2+} , Mg^{2+} , Na^+ , K^+), and effective cation exchange capacity were determined after
123 pre-washing with 60% ethanol, and then leaching with non-alcoholic 1 M NH_4Cl solution at
124 pH 7 (Tucker, 1985). The exchangeable sodium percentage was calculated as exchangeable Na
125 concentration divided by the effective cation exchange capacity. Total organic C, and total N
126 analyses were conducted using a LECO TruMac instrument using the Dumas method (Nelson
127 and Sommers, 1996). The electrochemical stability index was used as an indication of the



128 potential for soil dispersion. The threshold electrochemical stability index below which
129 structural breakdown occurs is 0.05 (McKenzie, 1998). The dispersion index (DI), another
130 measure of soil structural stability, is defined as the amount of dispersed silt+clay expressed as
131 a percentage of the total silt+clay of the soil (see section 2.3) (Mustafa and Letey, 1969). Soil
132 dispersion assessed by aggregate stability in water (ASWAT) is described in detail in Section
133 2.3.

134 *Table 1 Physico-chemical properties of soil samples*

	Soil 1	Soil 2
pH (1:5 water)	7.07	7.03
EC (1:5 dS m ⁻¹)	0.26	0.22
Sand (%)	34	37
Silt (%)	17	17
Clay (%)	49	46
Bulk density (g cm ⁻³)	1.29	1.29
Field capacity (%)	33	29
Total organic C (g kg ⁻¹)	8.8	9.8
Total N (g kg ⁻¹)	0.90	0.90
Effective cation exchange capacity (cmol ⁽⁺⁾ kg ⁻¹)	23	24
Exchangeable sodium percentage	15	16
Electrochemical stability index	0.02	0.01
ASWAT	15	11
DI (%)	48	28

151 2.2 Collection of different organic materials used as amendments

152 Four different organic amendments were used for this study, viz. feedlot manure (FLM),
153 chicken manure (CM), lucerne pellets (LP), and polyacrylamide (PAM). The FLM, and CM
154 were collected from cattle, and chicken sheds, respectively, located at The University of
155 Queensland (Gatton, Australia) before being air dried, whilst the LPs were a commercial



156 animal feed product. All organic amendments were ground and sieved through a 0.5 mm sieve
157 in order to minimise the effect of different size particles and to create a homogenised sample.
158 Anionic PAM Flobond L33 liquid (SNF Australia) had a charge density of 30% and the
159 molecular weight was 12-15 million Dalton (as per product specifications). Gypsum
160 ($\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$) was laboratory grade obtained from Sigma Aldrich.
161 A chemical analysis of the organic amendments was performed before mixing with the soils
162 (Table 2). Total C and N analysis were performed using the LECO TruMac instrument with
163 the Dumas method (Nelson and Sommers, 1996). The major cations, including Na, Ca, Mg,
164 and K, were quantified by inductively coupled plasma-optical emission spectrometry (ICP-
165 OES) after nitric acid microwave digestion (Kovács et al., 1996). The pH and EC of organic
166 amendments were measured in (1:5) water suspensions after shaking for 1 h.

167 *Table 2 Chemical properties of organic amendments*

	pH	EC (1:5)	TC	TN	C:N	Ca	Na	Mg	K
	(1:5)	(dS m^{-1})	(%)	(%)		(g kg^{-1})	(g kg^{-1})	(g kg^{-1})	(g kg^{-1})
FLM	8.4	3.0	14.9	1.4	10:1	18.1	4.2	12.8	12.5
CM	8.2	10.7	26.0	2.6	10:1	179.0	4.7	9.7	29.7
LP	5.6	5.7	43.8	3.1	14:1	13.8	2.9	3.7	12.8
PAM			49.7	0.9	56:1				

168 2.3 Incubation experiment

169 An incubation experiment was conducted using a complete randomised design. The treatments
170 consisted of the five amendments (gypsum (G), PAM, FLM, CM, and LP) plus an unamended
171 control. In addition, the FLM, CM, LP, and PAM were also applied in combination with G in
172 order to check for synergistic effects between gypsum and organic amendments. This yielded
173 a total of 10 treatments, each with three replicates, being a total of 60 experimental units. Soil
174 samples (300 g) were mixed with the respective amendments (Table 3) added at rates that were
175 commercially feasible (10 Mg ha^{-1}). The gypsum requirement of both soil samples was



176 calculated based on the formula given by Oster and Jayawardane (1998). The soil water content
177 at field capacity (-10 kPa) was ~ 0.30 g water g^{-1} soil. The WD cycles were imposed as follows.
178 First deionised water was added to the soils (0.30 g g^{-1} for Soil 1 and 0.31 g g^{-1} for Soil 2) at
179 $25^{\circ}C$, and after 1 d, the lids were removed and soils were allowed to dry at $25^{\circ}C$ during which
180 the soil water content decreased to air-dry conditions (~ 0.1 g water g^{-1} soil). The dry-down
181 typically completed in 14 d. The WD regime was applied four times with the total duration of
182 the experiment being 60 d. Although the WD regime might not entirely represent the field
183 conditions due to lack of plant growth, it is representative of the mean temperature and the field
184 water content.

185 **Table 3** *Treatments and their application rates*

	Treatments	Application rates
1	Control	No amendment
2	G	2.5 Mg ha^{-1}
3	FLM	10 Mg ha^{-1}
4	CM	10 Mg ha^{-1}
5	LP	10 Mg ha^{-1}
6	PAM	1 kg ha^{-1}
7	G + FLM	2.5 Mg ha^{-1} + 10 Mg ha^{-1}
8	G + CM	2.5 Mg ha^{-1} + 10 Mg ha^{-1}
9	G + LP	2.5 Mg ha^{-1} + 10 Mg ha^{-1}
10	G + PAM	2.5 Mg ha^{-1} + 1 Mg ha^{-1}

186 Soil solutions (~ 3 - 5 ml) were extracted using polyacrylonitrile hollow fibre samplers (Menzies
187 and Guppy, 2000) embedded in the jars containing the treated soils. Soil solution was collected
188 four times at the start of each WD cycle after the water was added to soils. Soil solution pH,
189 EC, NH_4^+ -N, NO_3^- -N, dissolved organic carbon (DOC), Na, Ca, Mg, and K were measured as
190 follows: NH_4^+ -N by the phenate method (Rice et al., 2017), NO_3^- -N by cadmium reduction
191 (Rice et al., 2017) using a segmented flow analyser (SEAL AA3), DOC using a total organic



192 carbon liquid analyser (Shimadzu, Japan), and the cations using ICP-OES. The
193 sodium adsorption ratio (SAR) was calculated from the soil solution concentrations of Na, Ca,
194 and Mg.

195 Soil dispersion was determined using the ASWAT test (Field *et al.* 1997) and DI (Mustafa and
196 Letey 1969). Air dried aggregates from each sample after completion of each cycle were placed
197 in a Petri dish filled with deionised water. A visual assessment of aggregate dispersion was
198 made after 10 min and 2 h, assessing dispersion on a scale of 0-4. The scores were as follows:
199 0: no dispersion, 1: slight dispersion (slight milkiness adjacent to aggregates in water), 2:
200 moderate dispersion (obvious milkiness), 3: strong dispersion (considerable milkiness with
201 about half of the material dispersed in water), and 4: complete dispersion (leaving sand particles
202 in a cloud of dispersed clay). Dispersion scores that were determined after 10 min and 2 h were
203 added together and then added to 8, thus giving a range of values from 9-16. Aggregates which
204 were not dispersed after 2 h were remolded, submerged in water and dispersion reassessed
205 again after 10 min and 2 h. Scores were given from 0-4 for both times and added (giving values
206 from 0-8). Thus, aggregates were considered stable if they had a low ASWAT score, and the
207 higher the ASWAT score, the more unstable the aggregate.

208 For the measurement of DI, 15 g of air-dry soil (< 2 mm) was placed in a sedimentation cylinder
209 with an approximate volume of 1.2 L. Deionised water was added to the cylinder to bring the
210 volume to a 1 L. The suspension was then shaken for 30 min on an end-over-end shaker. The
211 suspension was then stirred with 10 strokes of a plunger. The temperature was noted, the
212 suspension was allowed to settle, and after an appropriate time (8 h) the percentage of clay was
213 measured with hydrometer. The measurements were done after the first, second, and fourth
214 WD cycles. Results were expressed as DI (Mustafa and Letey 1969):

$$215 \quad DI(\%) = 100 \times \frac{\text{silt+clay (easily dispersed, \%)}}{\text{silt+clay (particle size analysis, \%)}} \quad (1)$$

216 Aggregate size distribution was also determined to quantify changes in large and small



217 aggregates with WD cycles. Air dried soil samples (25 g) were broken gently to pass through
218 a 10 mm sieve and then slowly wetted up in water for 5 min (Hernandez *et al.* 2017). They
219 were then wet sieved for 5 min at 33 oscillations per minute (Cook *et al.* 1992) using a set of
220 sieves: 2000 μm , 250 μm and 53 μm (Kemper and Rosenau 1986). Three replications were
221 made of each sample. The water level was adjusted so that the aggregates on the upper sieve
222 were submerged in water at the highest point of the oscillation. The material retained on each
223 sieve and the fraction passing through after wet sieving was collected, dried at 40 °C, and
224 weighed. (Kemper and Rosenau 1986). Measurements were made after the first, second, and
225 fourth WD cycles. The >2000 μm aggregates hereafter are referred to as large
226 macroaggregates, 2000-250 μm as small macroaggregates, 250-53 μm as micro-aggregates
227 (MIC), and <53 μm as silt+clay fraction. The MWD was calculated from the aggregate
228 fractions obtained after wet sieving on each sieve size, as follows:

$$229 \quad MWD = \sum_{i=1}^n x^i w^i \quad (2)$$

230 Where x^i is the mean diameter of each fraction and w^i is the proportion of the total sample
231 weight in the corresponding size fraction (Kemper and Rosenau 1986).

232 **2.4 Soil microbial respiration**

233 The biolability (susceptibility to microbial decomposition) of the organic amendments was
234 determined by measuring CO₂ release over a 60 d incubation period comprising four WD
235 cycles. Briefly, 50 g of each soil sample was added to 250 mL jars with the relevant
236 amendments (Table 3) and covered with lids having two 5 mm holes. All the treatments were
237 replicated three times and randomised. The soils were wetted to field capacity on the first day
238 of incubation using deionised water and incubated at 25 °C. After the first day of soil
239 incubation, the jars were left open to allow the soil to dry at 25 °C. For the microbial respiration
240 measurements, rubber tubes with Luer locks were inserted into the holes and connected to a
241 CO₂ analyser (WMA-4 CO₂ analyzer, John Morris USA). The lids were kept closed for 1 h



242 prior to the measurement being taken. The readings were then converted in $\text{g C-CO}_2 \text{ kg}^{-1}$ of
243 soil d^{-1} . Cumulative CO_2 produced was calculated as the sum of the daily rate and the interval
244 days between the two measurements (by linear extrapolation) for the incubation period. The
245 measurements were taken daily for the first 7 d. Thereafter, there was no significant change in
246 microbial respiration, so the measurements were performed after 10 d and 15 d of each WD
247 cycle.

248 **2.5 Statistical analysis**

249 Statistical analyses were performed using R x 64 3.3.3 statistical software (R Core Team,
250 2020). Histograms were plotted to check the normality of each parameter separately and found
251 that transformation of data was not necessary. Soil microbial respiration was recorded on a
252 daily basis hence respiration data were subjected to analysis of variance with days as the
253 repeated measures (mixed effect model). All the remaining parameters including EC, pH,
254 $\text{NH}_4^+\text{-N}$ and $\text{NO}_3^-\text{-N}$, SAR, ASWAT, DI, aggregate size distribution, and MWD were measured
255 after each WD cycles. Hence, these data were analysed using a two-way analysis of variance
256 (ANOVA) taking treatments and WD cycles as two factors. The ANOVA for each soil was
257 conducted separately using general linear model. Tukey's honest significant difference was
258 used for pairwise comparisons between treatment means. After checking the significance of
259 data, mean data were graphed using Sigma Plot v14.0 (Systat Software Inc., 2017). Principal
260 component analyses (PCA) were performed to identify the roles of microbial, chemical and
261 physical properties after the addition of different treatments in each WD cycle.

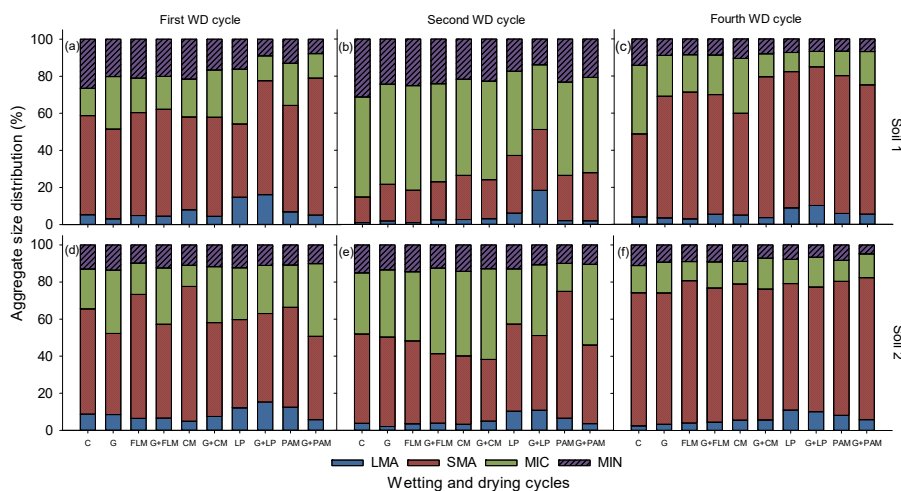
262 **3 Results**

263 **3.1 Soil aggregation dynamics measured by aggregate size distribution and mean** 264 **weight diameter**

265 Aggregate size distribution was significantly ($p < 0.001$) affected by amendments and WD
266 cycles in both soils except for the proportion of large macroaggregates and silt+clay fraction

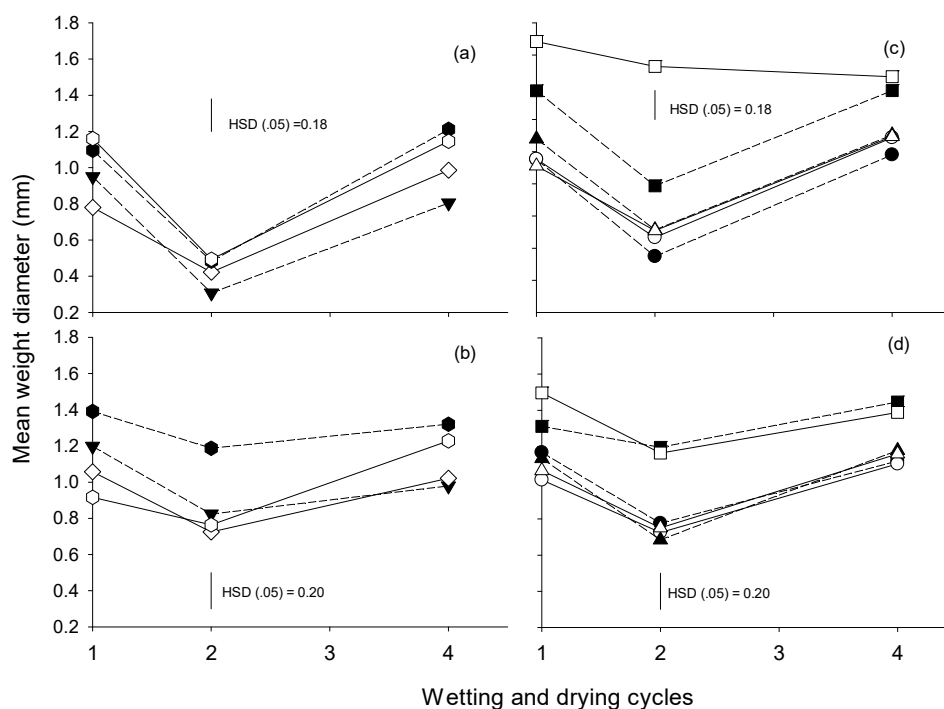


267 in Soil 2 (Fig. 1), although the main effects of treatments and WD cycles were highly significant
268 ($p < 0.001$) in Soil 2. The finding of a significant interaction between amendments and WD
269 cycles indicates that although aggregate size distribution changed with the WD cycles, the
270 pattern of change depended on the amendment. For example, we observed significant changes
271 in the proportion of large macroaggregates in both soils. Overall, the proportion of large
272 macroaggregates decreased after the first WD cycle and increased by the end of fourth WD
273 cycle. Among the various amendments, the LP and G+LP treatments had the highest proportion
274 of large macroaggregates, whereas PAM, FLM, and CM had little or no effect on large
275 macroaggregates for either soil. We also observed changes in the small macroaggregate
276 fraction, with this markedly decreasing by the second WD cycle but greatly increasing by the
277 fourth WD cycle. In contrast to the changes in small macroaggregates, the proportion of
278 microaggregates increased considerably (ca. two-fold) by the second WD cycle and greatly
279 decreased by the fourth WD cycle in both soils. The silt+clay fraction slightly increased by the
280 end of second WD cycle and again decreased slightly by the fourth WD cycle in both soils.
281 The MWD of both soils was calculated from the aggregate size distribution. The main effects
282 of amendments and WD cycles were found highly significant ($p < 0.001$) in both soils, however,
283 the interaction between amendments and WD cycles was significant only for Soil 1. It was
284 observed that MWD decreased after the second WD cycle (Fig. 2) and increased after the fourth
285 WD cycle, irrespective of amendments or soil, reflecting the changes in actual aggregate sizes
286 (see Fig.1). Expressing changes in MWD relative to the control or gypsum treated soil showed
287 that LP increased the MWD in both soils over the four WD cycles (Fig. S1), with PAM being
288 less effective than LP, whereas the other amendments had no significant effect.



289

290 *Figure 1. Distribution of the aggregate sizes in the two soils after addition of amendments after first,*
 291 *second, and fourth WD cycles: First WD cycle (a; Soil 1, and d; Soil 2), Second WD cycle (b; Soil 1,*
 292 *and e; Soil 2), and fourth WD cycle (c; Soil 1, and f; Soil 2) (C: control, G: gypsum, PAM: anionic*
 293 *polyacrylamide, FLM: feedlot manure, CM: chicken manure, LP: lucerne pellets). Large*
 294 *macroaggregates (>2000 μm); small macroaggregates (2000-250 μm); microaggregates (250-53 μm),*
 295 *and silt+clay fraction (<53 μm). Error bars have been omitted to improve readability.*



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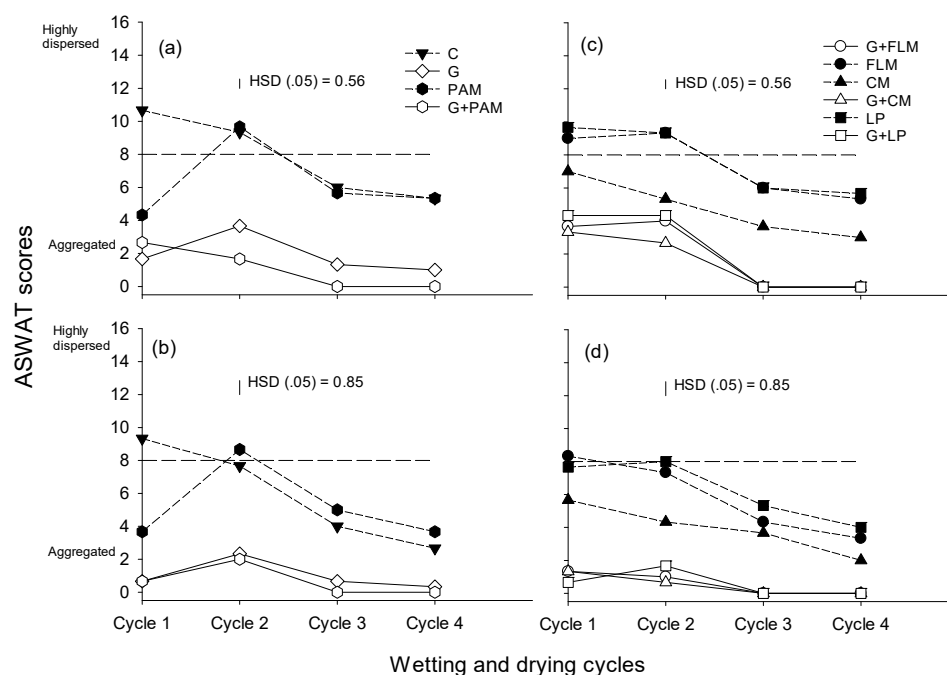
297 *Figure 2. The mean weight diameter (MWD) of the soils after addition of amendments during four*
298 *wetting and drying (WD) cycles. Soil 1 (a, c), and Soil 2 (b, d). The treatments are C: control, G:*
299 *gypsum, PAM: anionic polyacrylamide, FLM: feedlot manure, CM: chicken manure, LP: lucerne*
300 *pellets. Vertical bars represent Tukey's honest significant difference (HSD) values at P=0.05 for*
301 *pairwise comparisons among cycles.*

302 3.2 Soil dispersion dynamics as measured by ASWAT and DI

303 We assessed soil dispersion using both the ASWAT test (Fig. 3) and DI (see Supplementary
304 Data Fig. S2), with good agreement between the DI results and ASWAT scores (Fig. S3).
305 Overall, we observed that soil dispersion decreased with WD cycles in both soils. Although,
306 Soil 1 was more dispersive than Soil 2, both soils showed a similar decrease in dispersion with
307 WD cycles. Addition of gypsum significantly decreased dispersion in all WD cycles, and this
308 effect of gypsum was greater than the effect of organic amendments in decreasing soil



309 dispersion. PAM had only a short-term effect, with PAM decreasing dispersion for the first
 310 WD cycle but not thereafter. Of the organic amendments, only CM decreased the dispersion in
 311 both soils, but FLM and LP did not decrease dispersion in both soils in the first and second
 312 WD cycles. Given that both EC (Fig. S4) and SAR (Fig. S5) are known to affect soil dispersion,
 313 it was not surprising that the ASWAT scores were negatively correlated with EC (Fig. S6), and
 314 positively correlated with SAR (Fig. S7). However, the ASWAT scores became less affected
 315 by SAR after second WD cycle, with this highlighting the importance of WD cycles on
 316 structure improvement.



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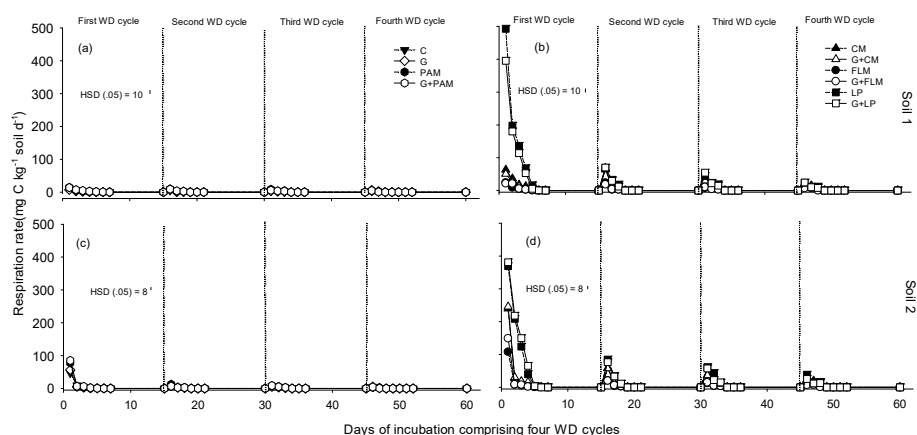
318 *Figure 3. The ASWAT scores of soils after addition of amendments during four wetting and drying*
 319 *(WD) cycles; Soil 1 (a, c), and Soil 2 (b, d). The amendments are C: control, G: gypsum, PAM: anionic*
 320 *polyacrylamide, G+PAM: gypsum+anionic polyacrylamide, FLM: feedlot manure, G+FLM:*
 321 *gypsum+feedlot manure, CM: chicken manure, G+CM: gypsum+chicken manure, LP: lucerne pellets,*



322 *G+LP: gypsum+lucerne pellets*). Vertical bars represent Tukey's honest significant difference (HSD)
323 values at $P=0.05$ for pairwise comparisons among cycles.

324 **3.3 Soil microbial respiration**

325 Soil WD cycles impose a significant stress on the soil microbial community. The addition of
326 treatments resulted in a significant increase ($p<0.001$) in soil microbial respiration for both the
327 soils (Fig. 4). Rewetting of the dried soils caused a transient increase in microbial respiration
328 (Fig. 4), with respiration being highest during the first WD cycle and decreasing during
329 subsequent WD cycles. The lowest microbial respiration rates were observed in the fourth WD
330 cycle in both soils (Fig. 4). The highest microbial respiration rates were observed in soils
331 amended with LP or CM in the first WD cycle, whereas FLM had little effect on microbial
332 respiration, whilst PAM and gypsum had no effect on microbial respiration.



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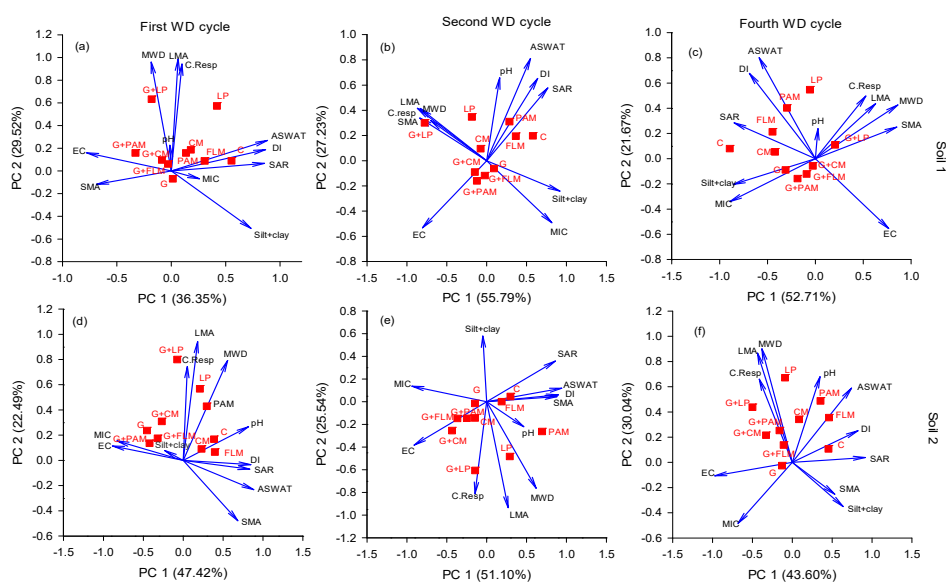
334 *Figure 4. Changes in soil respiration rate after addition of amendments during four alternate WD*
335 *cycles; Soil 1 (a, b) Soil 2 (c, d) The treatments were C: control, G: gypsum, PAM: anionic*
336 *polyacrylamide, G+PAM: gypsum+anionic polyacrylamide, FLM: feedlot manure, G+FLM:*
337 *gypsum+feedlot manure, CM: chicken manure, G+CM: gypsum+chicken manure, LP: lucerne pallets,*
338 *G+LP: gypsum+lucerne pallets. Vertical bars represent Tukey's honest significant difference (HSD)*
339 *values at P=0.05 for pairwise comparisons among the four WD cycles. The vertical dotted lines are*
340 *showing the days when deionised water was added to start the next WD cycle.*

341 **3.4 Relationship between soil physical, chemical, and microbial properties**

342 Soil properties determined for both soils after the first, second, and fourth WD cycles (Fig. 5)
343 were used to examine inter-relationships by PCA. Overall, the PCA biplots indicated that soil
344 microbial respiration, MWD, and the proportion of large macroaggregates were positively
345 correlated with each other. A positive correlation between ASWAT, DI with SAR was
346 observed in both soils, whereas EC had a negative correlation with ASWAT and DI.
347 Surprisingly, there was not a strong correlation between silt+clay and DI or ASWAT scores.
348 The effect of SAR and EC on soil dispersion decreased after the second WD cycle (Fig. 5),
349 underlining the importance of WD cycles on structure formation and stability, which offsets
350 the detrimental effects of SAR on stability. Obvious treatment effects on different soil
351 properties could also be seen in each WD cycle (Fig. 5). The G+LP and LP treatments increased



352 microbial respiration, the proportion of large macroaggregates and MWD in all the cycles. The
 353 gypsum added with and without organic amendments produced the highest EC in all the cycles.
 354 The control soils had highest SAR, ASWAT scores and DI followed by FLM and LP. It was
 355 also observed that the proportion of large macroaggregates and the MWD were negatively
 356 correlated with the silt+clay fraction in each WD cycle.



357
 358 *Figure 5. Principal component analysis (PCA) biplot of the physicochemical and microbial properties*
 359 *effected by different treatments in two soils after first, second, and fourth WD cycle: Soil 1(a, b, and c)*
 360 *and Soil 2 (d, e, and f). The treatments are C: control, G: gypsum, PAM: anionic polyacrylamide, FLM:*
 361 *feedlot manure, CM: chicken manure, LP: lucerne pellets. The data presented in these PCA biplots are*
 362 *the mean data of all the days for each parameter (blue arrows) and factorial scores of all the treatments*
 363 *(red squares). The parameters are ASWAT scores (aggregate stability in water test), DI (Dispersion*
 364 *index), C. Resp (cumulative microbial respiration), SAR (sodium adsorption ratio), EC (electrical*
 365 *conductivity 1:5), LMA (large macroaggregates), SMA (small macroaggregates), MIC*
 366 *(microaggregates), Silt+Clay, and MWD (mean weight diameter).*



367 4 Discussion

368 The results of this experiment showed clear differences in the role of organic amendments,
369 gypsum (Ca), and WD cycles on the formation and stabilisation of soil aggregates. We showed
370 that the formation and stability of large macroaggregates was controlled by the type of organic
371 amendments, whereas the WD cycles increased the formation and stability of small
372 macroaggregates. Formation of microaggregates is beneficial in that it decreases dispersion,
373 but microaggregates may not be sufficient to improve soil water infiltration (Collis-George and
374 Greene 1979; Nemati *et al.* 2002). Regardless, even a small increase either in large
375 macroaggregates and small macroaggregates may increase infiltration, with this facilitating the
376 leaching of excess Na. Here, we discuss the importance of each factor (WD cycles, organic
377 amendments, and Ca) individually.

378 4.1 The relative role of WD cycles on aggregation

379 The WD cycles result in rearrangement of pores and soil particles and may lead to increased
380 rigidity and stability of soil aggregates (Horn *et al.* 2014). We observed a marked change in
381 aggregate size distribution with repeated WD cycles (Fig. 1), from macroaggregates (large
382 macroaggregates and small macroaggregates) after first WD cycle, to microaggregates after
383 second WD cycle, and back to macroaggregates (large macroaggregates and small
384 macroaggregates) after the fourth WD cycle. We suggest that extracellular polysaccharides
385 formed by microbial activity are responsible for the formation of large macroaggregates after
386 the first WD cycle. After the second WD cycle, the microbial activity greatly decreased (Fig.
387 3) and macroaggregates (large macroaggregates and small macroaggregates) were broken
388 down into microaggregates and silt+clay. This allowed the soil particles to settle into tightly
389 packed configurations, resulting in stronger interconnections upon WD cycles (Kemper and
390 Rosenau 1984). The decrease in large macroaggregates (Fig.1) after the first WD cycle is also
391 consistent with the findings of Rahman *et al.* (2018). Macroaggregates are expected to be more



392 susceptible to disintegration due to water content changes, because of the large number of
393 planes of weakness and greater angular momentum (Kay 1990). As a result, macroaggregates
394 break more easily during WD cycles compared to microaggregates.

395 The changes observed in the proportion of small macroaggregates followed the same pattern
396 as large macroaggregates, with an initial increase in the proportion of small macroaggregates
397 when the soils were treated with organic amendments. In contrast to these results, no significant
398 differences were observed when the same soils were treated with organic amendments under
399 continuous wet conditions (Niaz *et al.*, unpublished). Also, the increase in proportion of larger
400 aggregates (large macroaggregates and small macroaggregates) is 2x greater when the same
401 soils were exposed to WD cycles as compared to constant wet regime (Niaz *et al.*, unpublished).
402 In this study, the proportion of small macroaggregates did not increase after the second WD
403 cycle, but the proportion of microaggregates and silt+clay fraction increased, suggesting that
404 upon repeated WD cycles, large macroaggregates break down into microaggregates and
405 silt+clay fractions. These observations support the finding that macroaggregates are composed
406 of microaggregates (Six *et al.*, 2000a). In addition, we observed that the control soils (where
407 no amendments were added) also showed an increase in the proportion of small
408 macroaggregates with WD cycles. This highlights the importance of WD cycles and suggests
409 that the formation of small macroaggregates is less dependent on the organic matter content, in
410 agreement with the PCA biplots (Fig. 5). The significant increase in the proportion of small
411 macroaggregates in control soils after the fourth WD cycle also indicates that there could be an
412 increase in interparticle bond strength with ageing or time (Utomo and Dexter 1982; Dexter
413 1988; Kong *et al.* 2005; Bravo-Garza *et al.* 2009).

414 **4.2 The role of organic amendments on soil respiration and aggregation**

415 The addition of organic amendments provided an energy and nutrient source for
416 microorganisms and resulted in increased respiration rates, although we observed that



417 respiration rate decreased with repeated WD cycles (Fig. 4), presumably due to depletion of
418 easily metabolisable organic compounds (Harrison-Kirk *et al.* 2013; Rahman *et al.* 2018). In
419 contrast to LP, CM had a modest effect on microbial respiration. Whereas, FLM had no
420 significant effect on microbial respiration as it was already decomposed. Rewetting of dry soil
421 led to immediate flush of microbial respiration (Fig. 4). This could be attributed to the slaking
422 of larger aggregates (Fig. 1), thereby exposing some occluded organic matter, or due to the
423 utilization of substrates that become available upon rewetting (Wu and Brookes 2005; Borke
424 and Matzner 2009). These available (labile) substrates after rewetting include remnants of the
425 added organic matter and microbial biomass (Wu and Brookes 2005). Over time, with
426 successive WD cycles, it is likely that the labile organic C pool was either exhausted or became
427 physically protected within aggregates and hence became less accessible to microbial
428 degradation.

429 For the LP and G+LP treatments, where the increase in respiration rate was ~4-fold greater in
430 Soil 1 and ~2 fold greater in Soil 2 than for any other treatment (Fig. 4), we observed that the
431 soils also had a significant increase in the proportion of large macroaggregates (Fig. 1) and
432 MWD (Fig. 2), suggesting that labile organic compounds are important for structure formation
433 (also confirmed by PCA biplots Fig. 5). The formation of large macroaggregates after
434 incorporation of organic amendments, as observed here for LP, has also been reported by Deneff
435 *et al.* 2001; Bravo-Garza *et al.* 2009; Rahman *et al.* 2018. Although the maximum respiration
436 rate was observed during the first WD cycle for LP and G+LP, the proportion of large
437 macroaggregates and MWD increased at the end of fourth WD cycle. During later WD cycles,
438 it is likely conversion of readily-metabolisable organic compounds (e.g., microbial polymers)
439 to more resistant forms resulted in the formation of macroaggregates (large macroaggregates
440 and small macroaggregates), but in this case, aggregation may have been caused by more
441 hydrophobic compounds. However, the proportion of large macroaggregates did not increase



442 much as compared to the first WD cycle, likely because microbial activity was lower. One of
443 the possible reasons of increased MWD could be the accumulation of microbial binding agents
444 over time that are released continuously from microbial activity due to organic matter
445 decomposition (Rahman *et al.* 2018).

446 When the soils were treated with organic amendments, a decrease in dispersion was observed
447 but not significantly different from control except in CM treated soils. The possible reason for
448 increased stability in CM treated soils was its higher Ca content (Table 2), increased EC (Fig.
449 S4) and decreased SAR (Fig. S5). For the remaining organic amendments viz; FLM, LP, and
450 PAM, an improvement in aggregate stability was observed after the second WD cycle.
451 Although, LP had the highest MWD, it still did not reduce dispersion in the first WD cycle.
452 We suggest that high MWD in itself is a misleading measure of soil stability since only a few
453 large aggregates can result in a high MWD (Niaz *et al.*, unpublished). Some dispersion can
454 occur in parallel which is sensitively detected by the ASWAT test, but not by the MWD. Both
455 assays detect different physical processes - hence a high MWD (Fig. 1) and dispersion (Fig. 2)
456 are not mutually exclusive (Niaz *et al.*, unpublished). Organic amendments rich in aliphatic
457 compounds or waxes can lead to hydrophobicity of aggregate surfaces during the drying step
458 which slows down the wetting of aggregate surfaces reducing the disruption caused during
459 rewetting (Piccolo and Mbagwu 1999; Borken and Matzner 2009). Monnier (1965) proposed
460 that fresh organic amendments can increase aggregate stability in the time frame of weeks and
461 months as compared to already decomposed stable organic amendments. But in our study, we
462 found no significant differences in dispersion after the addition of LP (fresh) and FLM
463 (partially decomposed). These findings suggest that there might be some other mechanisms
464 which are responsible for the binding of organic matter with clay particles to control dispersion,
465 such as the size of the organic matter molecule and charge density. This requires further
466 investigation.



467 **4.3 The relative role of Ca (gypsum) in improving aggregate stability**

468 In this experiment we found that gypsum (Ca) flocculated soil particles and reduced soil
469 dispersion (Fig. 3, Fig. S2), because the addition of Ca led to a significant increase in the EC
470 (Fig. S4) and a decrease in SAR (Fig. S5), with this presumably resulting in the flocculation of
471 soil particles (van Olphen 1977; Muneer and Oades 1989; Chorom and Rengasamy 1997;
472 Bennett *et al.* 2015). Improved aggregation and stability were also observed when the soils
473 were treated with gypsum and organic amendments. Gypsum reduces dispersion by increasing
474 flocculation, which is not same as improving aggregation. For the latter, organic amendments
475 are required. It is noteworthy, however, that there are also other mechanisms by which Ca can
476 interact with organic substances to form Ca-bridging through which clay particles are attached
477 to organic matter, with polyvalent cations forming micro-and macroaggregates (Edwards and
478 Bremner 1967; Muneer and Oades 1989; Tisdall 1996; Chenu *et al.* 1998).

479 **Conclusion**

480 Aggregate formation in two sodic Vertisols was significantly affected by WD cycles, organic
481 matter amendments and gypsum. However, we observed that not all organic amendments were
482 beneficial for increasing the formation of macroaggregates. Organic amendments LP and CM
483 increased the proportion of larger aggregates, whereas FLM was ineffective which can be
484 attributed to the effects of amendments on microbial activity. While G helped in reducing soil
485 dispersion, it had no significant role in the formation of larger aggregates. Implementation of
486 these findings in the field would favour the use of organic amendments with gypsum to improve
487 the physicochemical properties of sodic soils, which is further enhanced by WD cycles. The
488 aim should be initially to prevent soil dispersion which can be achieved by the application of
489 Ca (through application of gypsum) and then to build larger aggregates which can be achieved
490 by the application of organic amendments, and imposing wetting and drying cycles. The large
491 aggregates then improve water and air entry in soil.



492 **Author contribution**

493 Conceptualization: Niaz. S., J. Wehr, B. J., Kopittke, P.M., Dalal, R.C., and Menzies, N.W.;

494 Formal analysis, investigation, and writing original draft: Niaz. S; Review and editing: J.

495 Wehr, B. J., Kopittke, P.M., Dalal, R.C., and Menzies, N.W.; Supervision: Wehr, B. J., and

496 Menzies, N.W.

497 **Competing interests**

498 The authors declare that they have no conflict of interest.

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