

1 **Wetting and drying cycles, organic amendments and gypsum play a key role in structure formation and**
2 **stability of sodic Vertisols**

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12 **Abstract**

13 In the natural environment, soils undergo wetting and drying (WD) cycles due to precipitation
14 and evapotranspiration. The WD cycles have a profound impact on soil physical, chemical, and
15 biological properties and drive the development of structure in soils. Degraded soils are often
16 lacking structure and the effect of organic amendments and WD cycles on structure formation
17 of these soils is poorly understood. The aim of this study was to evaluate the role of biotic and
18 abiotic factors on aggregate formation and stabilisation of sodic soils after the addition of
19 gypsum and organic amendments (feedlot manure, chicken manure, lucerne pellets, and
20 anionic poly acrylamide). Amended soils were incubated at 25°C over four WD cycles, with
21 assessment of soil microbial respiration, electrical conductivity, pH, sodium adsorption ratio
22 (SAR), aggregate stability in water (ASWAT), aggregate size distribution, and mean weight
23 diameter. Our results demonstrate that WD cycles can improve aggregate stability after the
24 addition of amendments in sodic Vertisols, but this process depends on the type of organic
25 amendment. Lucerne pellets resulted in highest soil microbial respiration, proportions of large
26 macroaggregates (>2000 µm), and mean weight diameter. In contrast, dispersion was
27 significantly reduced when soils were treated with chicken manure, whilst anionic

28 polyacrylamide only had a transient effect on aggregate stability. When these organic
29 amendments were applied together with gypsum, the stability of aggregates was further
30 enhanced, and dispersion became negligible after the second WD cycle. The formation and
31 stability of small macroaggregates (2000-250 μm) was less dependent on the type of organic
32 amendments and more dependent on WD cycles as the proportion of small macroaggregates
33 also increased in control soils after four WD cycles, highlighting the role of WD cycles as one
34 of the key factors that improves aggregation and stability of sodic Vertisols.

35 **Key words:** Sodic soils, organic amendments, aggregate stability, mean weight diameter,
36 wetting and drying cycles

37 **1 Introduction**

38 Soils are subjected to seasonal and daily variations of water and temperature. These variations
39 effect the physical, biological, and chemical properties of soils. Natural variations in soil water
40 content can lead to wetting and drying (WD) cycles, which are affected by rainfall, solar
41 radiation, capillarity, wind, condensation (Utomo and Dexter 1982) and evapotranspiration. In
42 most terrestrial ecosystems, surface soils experience rapid changes in soil water content with a
43 longer dry period followed by a relatively rapid re-wetting (Borken and Matzner 2009).
44 Southern Queensland (Australia) usually has hot wet summers with cool dry winters, and soils
45 of this region and other semi-arid areas are particularly susceptible to drying and re-wetting
46 stresses due to the infrequency of rainfall. In the field, most soils experience more than one
47 WD cycle throughout the year. These WD cycles have a substantial influence on soil
48 aggregation and structure stabilisation because of its direct effect on hydration of minerals, and
49 indirect effect on plant ecology and soil microbial activity (Denef *et al.* 2001; Cosentino *et al.*
50 2006).

51 Soil aggregation is an important mechanism for stabilisation of soil organic matter (Six *et al.*
52 2000a). Furthermore, it also supports soil fertility as it reduces soil erosion and controls soil

53 aeration, water infiltration, hydraulic conductivity and nutrient cycling (Oades 1984; Six *et al.*
54 2000b). Soil aggregation is caused by various aggregate stabilising compounds, which work
55 together at different spatial scales (Tisdall and Oades 1982). Aggregate formation also depends
56 on soil microbial activity, since the latter influences the production of binding materials such
57 as microbial exudates and hyphae (Rahman *et al.* 2017). Aggregate stability often exhibits
58 seasonal and inter-annual variability that is controlled mainly by rainfall, temperature,
59 humidity, insolation, and organic matter (Perfect *et al.* 1990).

60 Sodic soils are generally characterised as soils with poor structure which makes those soils
61 difficult to work when wet or dry (Rengasamy and Olsson 1991). Poor structural stability of
62 sodic soils restricts seedling emergence and root growth which directly limits crop growth and
63 development, and indirectly affects plant nutrition by limiting water infiltration, nutrient
64 uptake, and gaseous exchange (Curtin and Naidu 1998). Rehabilitation of sodic soils requires
65 an understanding of the complex interactions between biological and physico-chemical factors
66 that contributes to soil structure formation and stability (Oades 1993; Nelson and Oades 1998).

67 Traditionally the management practices used to improve the structure of sodic soils involves
68 the displacement of Na ions from the soil exchange complex with the help of divalent cations
69 such as Ca or increasing the ionic strength of soil solution (Ghosh *et al.* 2010), both of which
70 can be achieved by the application of gypsum to these soils. The effect of gypsum on increasing
71 ionic strength is immediate, but short lived. In contrast, the effect of gypsum for providing the
72 counter ion (Ca to replace Na) is permanent unless additional Na is added to system (e.g., using
73 poor quality irrigation water). Another frequently used management practice to ameliorate
74 sodic soils is the use of organic amendments. Addition of organic matter effect aggregate
75 stability within a period of days to weeks due to the stimulation of microbial activity (Six *et al.*
76 2004), depending upon the quality and quantity of organic matter (Monnier, 1965 cited in
77 Abiven *et al.* (2009)). While organic matter increases soil microbial respiration resulting in the

78 formation of extracellular polysaccharides which help in the formation of soil aggregates
79 (Bossuyt *et al.*, 2001), studies investigating the effect of organic amendments in improving the
80 soil structure are inconclusive. For instance, the extracellular polysaccharides and large
81 polyanions can bind clay particles into stable macroaggregates. On the other hand, organic
82 anions can enhance dispersion by increasing the negative charge on clay particles and by
83 complexing calcium and other polyvalent cations (such as those of aluminium), hence reducing
84 their activity in soil solution (Ghosh *et al.* 2010)

85 Apart from the changes in soil structure due to the addition of different ameliorants, WD cycles
86 can lead to more intensive changes in structure of soils dominated by smectitic clays (Vertisols)
87 through physical processes (Utomo and Dexter 1982; Deneff *et al.* 2001). These soils are
88 generally characterised as self-mulching soils as they exhibit shrink-swell properties imposed
89 by the WD cycles (Pal *et al.* 2012). Vertisols cover a total of an estimated 340 million ha in the
90 world (Australia, Asia, Africa, and America), out of which approximately 150 million ha is
91 potential crop land. However, the physical properties and moisture regimes of Vertisols
92 represent serious management constraints (Pal *et al.* 2012). Sodic Vertisols are common in
93 arid parts of the world. The effect of sodicity on the physical properties of Vertisols is still a
94 subject of debate. For example, Rahman *et al.* (2018) reported that WD cycles improved
95 (increased) the mean weight diameter (MWD, a proxy of aggregate stability measurement) of
96 Vertisols when treated with maize straw after four WD cycles. In a similar manner, WD cycles
97 also increased the proportion of large macro aggregates of smectite Vertisols (Bravo-Garza *et*
98 *al.* 2009). Peng *et al.* (2011), comparing swelling and non-swelling soils, reported that WD
99 cycles decreased the MWD of swelling soils but not of non-swelling soils. However, Six *et al.*
100 (2000b) found that repeated WD cycles decreased the MWD due to physical disturbance of
101 aggregates, with this being related to the loss of soil organic matter. These apparently
102 contradictory results can potentially be explained on the basis of the initial conditions of the

103 soil, such as the physical conditions of aggregates, organic matter content and quality, and the
104 intensities and duration of the WD cycles (Cosentino *et al.* 2006).

105 Similarly, studies investigating the effects of WD cycles on microbial activity were
106 inconclusive, with results differing due to varying experimental designs, incubation period and
107 temperatures, soil properties, and treatments applied. Xiang *et al.* (2008) found that multiple
108 WD cycles increased the microbial respiration of grassland soils up to 6-fold when compared
109 to the soil that remained wet. Drying and rewetting of these soils gave a new pulse of respiration
110 with each WD cycle, but when these soils were kept at constant water content, microbial
111 respiration decreased to almost zero. An increase in cumulative respiration from forest soils of
112 China was also observed when these soils experience WD cycle compared to the constant
113 moisture conditions (Zhang *et al.* 2022). In contrast, however, Rahman *et al.* (2018) reported
114 that repeated WD cycles in Vertisols decreased soil respiration significantly but that the
115 magnitude of this decrease became smaller over the time. Similarly, Yu *et al.* (2014) found that
116 repeated WD cycles decreased the cumulative soil respiration compared to constant moist
117 conditions in a loamy sand soil.

118 Although the interaction of biotic and abiotic factors and its effect on aggregate stability and
119 formation is complex and inconsistent, little effort has been put into studying the underlying
120 mechanisms, particularly in sodic Vertisols. Furthermore, there is little information available
121 on the relationship between aggregate formation and stability following repeated WD cycles
122 after addition of gypsum and organic amendments. Thus, in the present study we used two
123 sodic Vertisols, with the aim to: i) determine the role of gypsum and different organic
124 amendments on aggregate formation and stability, ii) explore the combined effect of gypsum
125 and organic amendments on soil physico-chemical and microbial properties, iii) investigate the
126 effect of WD cycles on microbial respiration, iv) assess the effects of WD cycles on aggregate
127 formation and stability, and v) determine how many WD cycles are needed to improve

128 aggregate stability. We hypothesised that i) organic amendments will increase the microbial
129 respiration and improves the formation of large macroaggregates and MWD, (ii) gypsum will
130 improve aggregate stability due to increases Ca concentration and ionic strength, iii) organic
131 amendments act synergistically with gypsum on aggregation, and iv) repeated WD cycles will
132 increase the process of aggregate formation and stability.

133 **2 Materials and methods**

134 **2.1 Soils**

135 The two soils used in this experiment were collected from a farm located in southern
136 Queensland near Goondiwindi (28.54° S, 150.30° E), Australia. The soils were being cropped
137 using a maize (*Zea mays*) and wheat (*Triticum aestivum*) rotation system. Both soils were
138 classified as sodic Vertisols according to the FAO-World Reference Base (2015) (Vertisols in
139 the Australian Soil Classification). Soils were collected with a shovel to a depth of 10 cm at
140 two locations from the cropped land. These sites were selected as they were in close proximity
141 to each other but were slightly different in regard to their physical properties (Table 1) with
142 Soil 1 being more dispersive than Soil 2. Three core samples were also collected for the
143 measurement of bulk density from each site (Soil 1 and Soil 2) and dried at 105°C for several
144 days. After collection, soils were air dried, and the larger clods gently broken into smaller
145 aggregates by hand. The soils were then passed through a 10 mm sieve to remove stones, visible
146 roots, and plant litter. For chemical analysis, a portion of each soil was sieved to <2 mm. Soil
147 pH (ISO, 2005) and electrical conductivity (EC) (ISO, 1994) were measured in 1:5 soil: water
148 suspension. Particle size analysis was performed using the pipette method (Day, 1965), and
149 field water capacity (-10 kPa) was measured using a pressure plate apparatus (Cassel and
150 Nielsen, 1986). Exchangeable cations (Ca^{2+} , Mg^{2+} , Na^+ , K^+), and effective cation exchange
151 capacity were determined after pre-washing with 60% ethanol, and then leaching with non-
152 alcoholic 1 M NH_4Cl solution at pH 7 (Tucker, 1985). The exchangeable sodium percentage

153 was calculated as exchangeable Na concentration divided by the effective cation exchange
 154 capacity. Total organic C, and total N analyses were conducted using a LECO TruMac
 155 instrument using the Dumas method (Nelson and Sommers, 1996). The electrochemical
 156 stability index was used as an indication of the potential for soil dispersion. The threshold
 157 electrochemical stability index below which structural breakdown occurs is 0.05 (McKenzie,
 158 1998). The dispersion index (DI), another measure of soil structural stability, is defined as the
 159 amount of dispersed silt+clay expressed as a percentage of the total silt+clay of the soil (see
 160 section 2.3) (Mustafa and Letey, 1969). Soil dispersion assessed by aggregate stability in water
 161 (ASWAT) is described in detail in Section 2.3.

Table 1 Physico-chemical properties of soil samples

	Soil 1	Soil 2
pH (1:5 water)	7.07	7.03
EC (1:5 dS m ⁻¹)	0.26	0.22
Sand (%)	34	37
Silt (%)	17	17
Clay (%)	49	46
Bulk density (g cm ⁻³)	1.29	1.29
Field capacity (%)	33	29
Total organic C (g kg ⁻¹)	8.8	9.8
Total N (g kg ⁻¹)	0.90	0.90
Effective cation exchange capacity (cmol ⁽⁺⁾ kg ⁻¹)	23	24
Exchangeable sodium percentage	15	16
Electrochemical stability index	0.02	0.01
ASWAT	15	11
DI (%)	48	28

162 2.2 Collection of different organic materials used as amendments

163 Four different organic amendments were used for this study, viz. feedlot manure (FLM),
164 chicken manure (CM), lucerne pellets (LP), and polyacrylamide (PAM). The FLM, and CM
165 were collected from a cattle feedlot and chicken sheds, respectively, located at The University
166 of Queensland (Gatton, Australia) before being air dried, whilst the LP was a commercial
167 animal feed product. The four organic amendments were chosen because of three reasons: 1)
168 they were easily available and are being used by farmers, 2) LP is used as green manure and
169 studies have shown it is effective in ameliorating sodic soils, and 3) PAM is used in mining
170 and construction to treat sodic dispersive soils. Furthermore, these amendments were different
171 in terms of their chemical properties (Table 2) and C functional groups (Niaz et al., 2022) and
172 may give a good contrast between the amendments. All organic amendments were ground and
173 sieved through a 0.5 mm sieve in order to minimise the effect of different size particles and to
174 create a homogenised sample. Anionic PAM Flobond L33 liquid (SNF Australia) had a charge
175 density of 30% and the molecular weight was 12-15 million Dalton (as per product
176 specifications). Gypsum ($\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$) was laboratory grade obtained from Sigma Aldrich.

177 A chemical analysis of the organic amendments was performed before mixing with the soils
178 (Table 2). Total C and N analysis were performed using the LECO TruMac instrument with
179 the Dumas method (Nelson and Sommers, 1996). The major cations, including Na^+ , Ca^{2+} ,
180 Mg^{2+} , and K^+ , were quantified by inductively coupled plasma-optical emission spectrometry
181 (ICP-OES) after nitric acid microwave digestion (Kovács et al., 1996). The pH and EC of
182 organic amendments were measured in (1:5) water suspensions after shaking for 1 h.

183 **Table 2** Chemical properties of organic amendments

pH	EC (1:5)	TC	TN	C:N	Ca^{2+}	Na^+	Mg^{2+}	K^+
(1:5)	(dS m^{-1})	(%)	(%)		(g kg^{-1})			

FLM	8.4	3.0	14.9	1.4	10:1	18.1	4.2	12.8	12.5
CM	8.2	10.7	26.0	2.6	10:1	179.0	4.7	9.7	29.7
LP	5.6	5.7	43.8	3.1	14:1	13.8	2.9	3.7	12.8
PAM			49.7	0.9	56:1				

184 2.3 Incubation experiment

185 An incubation experiment was conducted using a complete randomised design. The treatments
 186 consisted of the five amendments (gypsum (G), PAM, FLM, CM, and LP) plus an unamended
 187 control. In addition, the FLM, CM, LP, and PAM were also applied in combination with G in
 188 order to check for synergistic effects between gypsum and organic amendments. This yielded
 189 a total of 10 treatments, each with three replicates, being a total of 60 experimental units. Soil
 190 samples (300 g) were mixed with the respective organic amendments (Table 3) added at rates
 191 that were commercially feasible (10 Mg ha⁻¹) except PAM which was added at 1 kg ha⁻¹. The
 192 gypsum requirement of both soil samples was calculated based on the formula given by Oster
 193 and Jayawardane (1998) as follows;

$$194 \text{ Gypsum requirement (GR)} = 0.00086 \times F \times D \times \partial b \times (CEC) \times (ESP_i - ESP_f) \quad (1)$$

195 Where F is exchanged efficiency of Ca-Na and for this case considered equal to 1, D is the
 196 depth of soil to be reclaimed (cm), ∂b is soil bulk density (g/cm³), CEC is cation exchange
 197 capacity (cmol⁺/kg), ESP_i is initial soil exchangeable sodium percentage, ESP_f is final or
 198 desired exchangeable sodium percentage. For simplicity, a single gypsum rate of 2.5 Mg/ha
 199 was selected for both soils

200 The experiment was performed jars with 12 cm diameter and 15 cm height, and the soil was
 201 packed to a height of 3-5 cm. The soil water content at field capacity (-10 kPa) was ~ 0.30 g
 202 water g⁻¹ soil. The WD cycles were imposed as follows. First deionised water was added to the
 203 soils (0.30 g g⁻¹ for Soil 1 and 0.31 g g⁻¹ for Soil 2) at 25°C, and after 1 d, the lids were removed,
 204 and soils were allowed to dry at 25°C during which the soil water content decreased to air-dry

205 conditions (~ 0.1 g water g^{-1} soil). The dry-down typically completed in 14 d. The WD regime
 206 was applied four times with the total duration of the experiment being 60 d. Although the WD
 207 regime might not entirely represent the field conditions due to lack of plant growth, it is
 208 representative of the mean temperature and the field water content.

209 **Table 3** *Treatments and their application rates*

	Treatments	Application rates
1	Control	No amendment
2	G	2.5 Mg ha ⁻¹
3	FLM	10 Mg ha ⁻¹
4	CM	10 Mg ha ⁻¹
5	LP	10 Mg ha ⁻¹
6	PAM	1 kg ha ⁻¹
7	G + FLM	2.5 Mg ha ⁻¹ + 10 Mg ha ⁻¹
8	G + CM	2.5 Mg ha ⁻¹ + 10 Mg ha ⁻¹
9	G + LP	2.5 Mg ha ⁻¹ + 10 Mg ha ⁻¹
10	G + PAM	2.5 Mg ha ⁻¹ + 1 kg ha ⁻¹

210 Soil solutions (~ 3 -5 ml) were extracted using polyacrylonitrile hollow fibre samplers (Menzies
 211 and Guppy, 2000) embedded in the jars containing the treated soils. Soil solution was collected
 212 four times at the start of each WD cycle after the water was added to soils. Soil solution pH,
 213 EC, NH₄⁺-N, NO₃⁻-N, dissolved organic carbon (DOC), Na, Ca, Mg, and K were measured as
 214 follows: NH₄⁺-N by the phenate method (Rice et al., 2017), NO₃⁻-N by cadmium reduction
 215 (Rice et al., 2017) using a segmented flow analyser (SEAL AA3), DOC using a total organic
 216 carbon liquid analyser (Shimadzu, Japan), and the cations using ICP-OES. The
 217 sodium adsorption ratio (SAR) was calculated from the soil solution concentrations of Na⁺,
 218 Ca²⁺, and Mg²⁺.

219 Soil dispersion was determined using the ASWAT test (Field *et al.* 1997) and DI (Mustafa and
 220 Letey 1969). Air dried aggregates from each sample after completion of each cycle were placed

221 in a Petri dish filled with deionised water. A visual assessment of aggregate dispersion was
222 made after 10 min and 2 h, assessing dispersion on a scale of 0-4. The scores were as follows:
223 0: no dispersion, 1: slight dispersion (slight milkiness adjacent to aggregates in water), 2:
224 moderate dispersion (obvious milkiness), 3: strong dispersion (considerable milkiness with
225 about half of the material dispersed in water), and 4: complete dispersion (leaving sand particles
226 in a cloud of dispersed clay). Dispersion scores that were determined after 10 min and 2 h were
227 added together and then added to 8, thus giving a range of values from 9-16. Aggregates which
228 were not dispersed after 2 h were remolded, submerged in water and dispersion reassessed
229 again after 10 min and 2 h. Scores were given from 0-4 for both times and added (giving values
230 from 0-8). Thus, aggregates were considered stable if they had a low ASWAT score, and the
231 higher the ASWAT score, the more unstable the aggregate.

232 For the measurement of DI, 15 g of air-dry soil (< 2 mm) was placed in a sedimentation cylinder
233 with an approximate volume of 1.2 L. Deionised water was added to the cylinder to bring the
234 volume to a 1 L. The suspension was then shaken for 30 min on an end-over-end shaker. The
235 suspension was then stirred with 10 strokes of a plunger. The temperature was noted, the
236 suspension was allowed to settle, and after an appropriate time (8 h) the percentage of clay was
237 measured with hydrometer. The measurements were done after the first, second, and fourth
238 WD cycles. Results were expressed as DI (Mustafa and Letey 1969):

$$239 \quad DI(\%) = 100 \times \frac{\text{silt+clay (easily dispersed,\%)}}{\text{silt+clay (particle size analysis,\%)}} \quad (2)$$

240 Aggregate size distribution was also determined to quantify changes in large and small
241 aggregates with WD cycles. Air dried soil samples (25 g) were broken gently to pass through
242 a 10 mm sieve and then slowly wetted up in water for 5 min (Hernandez *et al.* 2017). They
243 were then wet sieved for 5 min at 33 oscillations per minute (Cook *et al.* 1992) using a set of
244 sieves: 2000 μm , 250 μm and 53 μm (Kemper and Rosenau 1986). Three replications were
245 made of each sample. The water level was adjusted so that the aggregates on the upper sieve

246 were submerged in water at the highest point of the oscillation. The material retained on each
247 sieve and the fraction passing through after wet sieving was collected, dried at 40 °C, and
248 weighed. (Kemper and Rosenau 1986). Measurements were made after the first, second, and
249 fourth WD cycles. The >2000 µm aggregates hereafter are referred to as large
250 macroaggregates, 2000-250 µm as small macroaggregates, 250-53 µm as micro-aggregates
251 (MIC), and <53 µm as silt+clay fraction. The MWD was calculated from the aggregate
252 fractions obtained after wet sieving on each sieve size, as follows:

$$253 \quad MWD = \sum_{i=1}^n x^i w^i \quad (3)$$

254 Where x^i is the mean diameter of each fraction and w^i is the proportion of the total sample
255 weight in the corresponding size fraction (Kemper and Rosenau 1986).

256 **2.4 Soil microbial respiration**

257 The biolability (susceptibility to microbial decomposition) of the organic amendments was
258 determined by measuring CO₂ release over a 60 d incubation period comprising four WD
259 cycles. Briefly, 50 g of each soil sample was added to 250 mL jars with the relevant
260 amendments (Table 3) and covered with lids having two 5 mm holes. All the treatments were
261 replicated three times and randomised. The soils were wetted to field capacity on the first day
262 of incubation using deionised water and incubated at 25 °C. After the first day of soil
263 incubation, the jars were left open to allow the soil to dry at 25 °C. For the microbial respiration
264 measurements, rubber tubes with Luer locks were inserted into the holes and connected to a
265 CO₂ analyser (WMA-4 CO₂ analyzer, John Morris USA). The lids were kept closed for 1 h
266 prior to the measurement being taken. The readings were then converted in g C-CO₂ kg⁻¹ of
267 soil d⁻¹. Cumulative CO₂ produced was calculated as the sum of the daily rate and the interval
268 days between the two measurements (by linear extrapolation) for the incubation period. The
269 measurements were taken daily for the first 7 d. Thereafter, there was no significant change in

270 microbial respiration, so the measurements were performed after 10 d and 15 d of each WD
271 cycle.

272 **2.5 Statistical analysis**

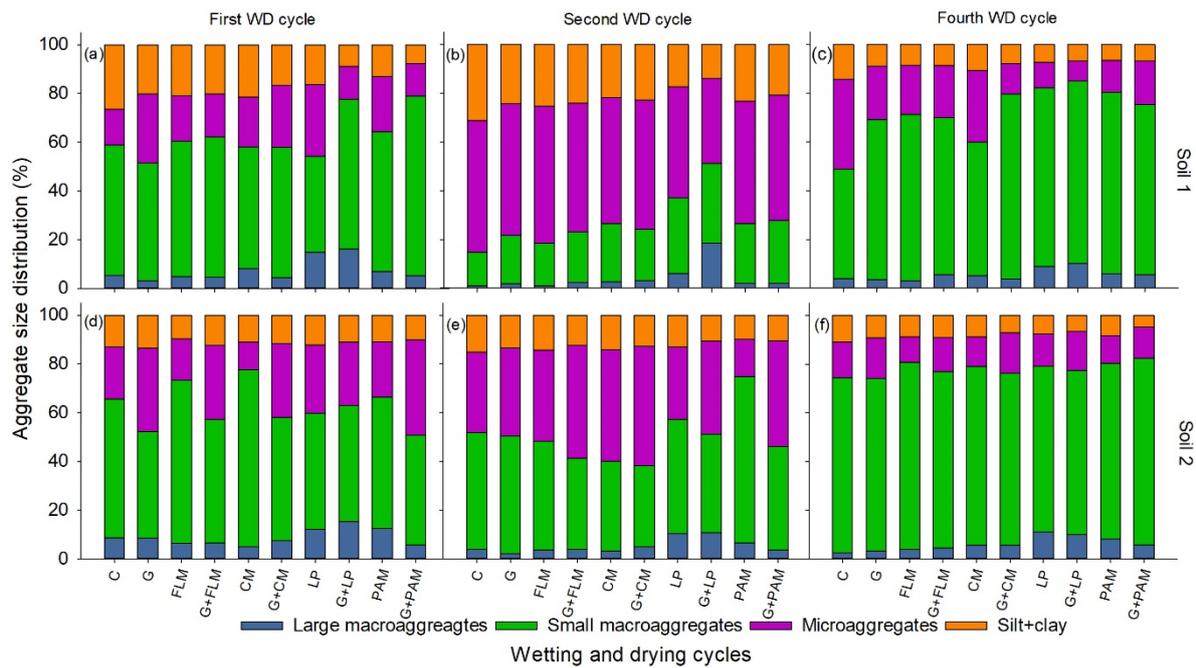
273 Statistical analyses were performed using R x 64 3.3.3 statistical software (R Core Team,
274 2020). Histograms were plotted to check the normality of each parameter separately and found
275 that transformation of data was not necessary. Soil microbial respiration was recorded on a
276 daily basis hence respiration data were subjected to analysis of variance with days as the
277 repeated measures (mixed effect model). All the remaining parameters including EC, pH,
278 $\text{NH}_4\text{-N}$ and $\text{NO}_3\text{-N}$, SAR, ASWAT, DI, aggregate size distribution, and MWD were measured
279 after each WD cycle. Hence, these data were analysed using a two-way analysis of variance
280 (ANOVA) taking treatments and WD cycles as two factors. The ANOVA for each soil was
281 conducted separately using general linear model. Tukey's honest significant difference was
282 used for pairwise comparisons between treatment means. After checking the significance of
283 data, mean data were graphed using Sigma Plot v14.0 (Systat Software Inc., 2017). Principal
284 component analyses (PCA) were performed to identify the roles of microbial, chemical and
285 physical properties after the addition of different treatments in each WD cycle.

286 **3 Results**

287 **3.1 Soil aggregation dynamics measured by aggregate size distribution and mean** 288 **weight diameter**

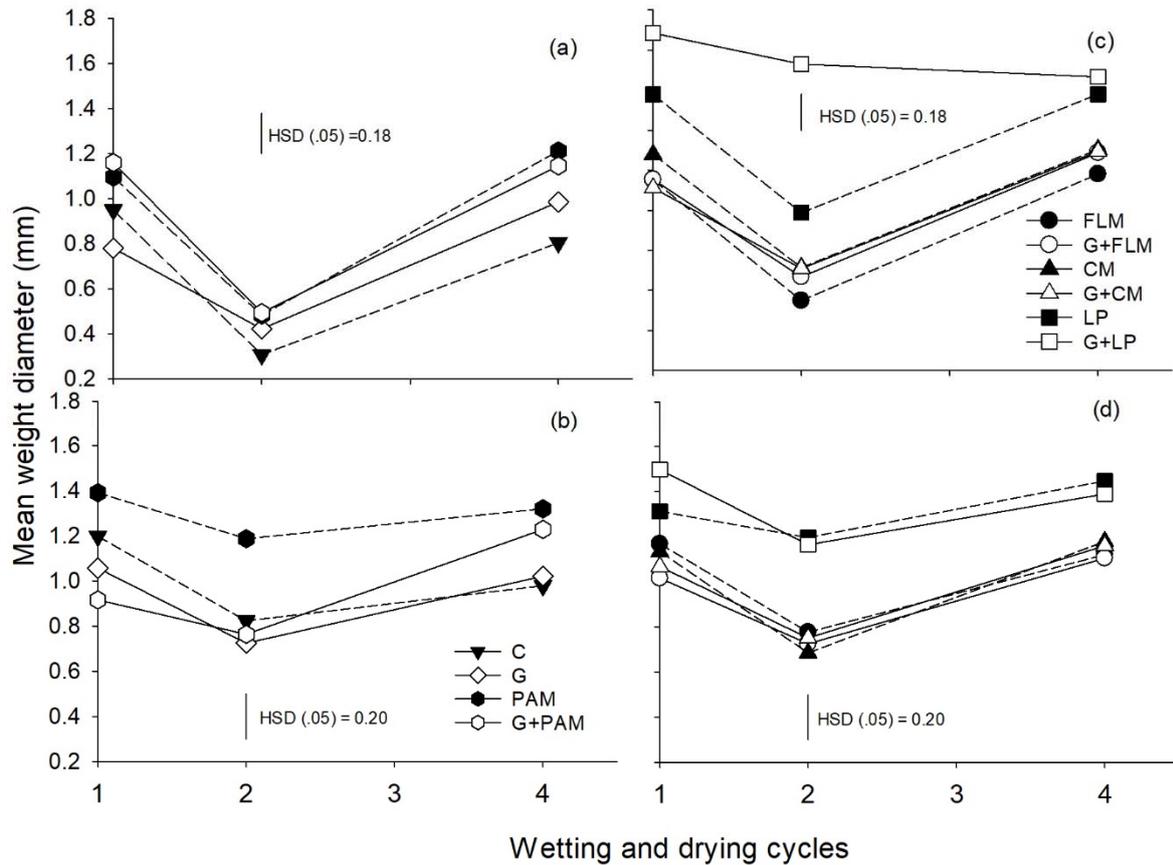
289 A significant ($p < 0.001$) interaction between WD cycles and amendments was found for all the
290 aggregate sizes in both soils except for the proportion of large macroaggregates and silt+clay
291 fraction in Soil 2 (Fig. 1), although the main effects of treatments and WD cycles were highly
292 significant ($p < 0.001$) in Soil 2. The finding of a significant interaction between amendments
293 and WD cycles indicates that although aggregate size distribution changed with the WD cycles,
294 the pattern of change depended on the amendment. Overall, the proportion of large

295 macroaggregates decreased after the first WD cycle and increased by the end of fourth WD
296 cycle. Among the various amendments, the LP and G+LP treatments had the highest proportion
297 of large macroaggregates, whereas PAM, FLM, and CM had little or no effect on large
298 macroaggregates for either soil. We also observed changes in the small macroaggregate
299 fraction, with this markedly decreasing by the second WD cycle but greatly increasing by the
300 fourth WD cycle. In contrast to the changes in small macroaggregates, the proportion of
301 microaggregates increased considerably (ca. two-fold) by the second WD cycle and greatly
302 decreased by the fourth WD cycle in both soils. The silt+clay fraction slightly increased by the
303 end of second WD cycle and again decreased slightly by the fourth WD cycle in both soils.
304 The MWD of both soils was calculated from the aggregate size distribution. The main effects
305 of amendments and WD cycles were found highly significant ($p < 0.001$) in both soils, however,
306 the interaction between amendments and WD cycles was significant only for Soil 1. It was
307 observed that MWD decreased after the second WD cycle (Fig. 2) and increased after the fourth
308 WD cycle, irrespective of amendments or soil, reflecting the changes in actual aggregate sizes
309 (see Fig.1). Expressing changes in MWD relative to the control or gypsum treated soil showed
310 that LP increased the MWD in both soils over the four WD cycles (Fig. S1), with PAM being
311 less effective than LP, whereas the other amendments had no significant effect.



312

313 *Figure 1. Distribution of the aggregate sizes in the two soils after addition of amendments after first,*
 314 *second, and fourth WD cycles: First WD cycle (a; Soil 1, and d; Soil 2), Second WD cycle (b; Soil 1,*
 315 *and e; Soil 2), and fourth WD cycle (c; Soil 1, and f; Soil 2) (C: control, G: gypsum, PAM: anionic*
 316 *polyacrylamide, FLM: feedlot manure, CM: chicken manure, LP: lucerne pellets). Large*
 317 *macroaggregates (>2000 μm); small macroaggregates (2000-250 μm); microaggregates (250-53 μm),*
 318 *and silt+clay fraction (<53 μm). Error bars have been omitted to improve readability.*



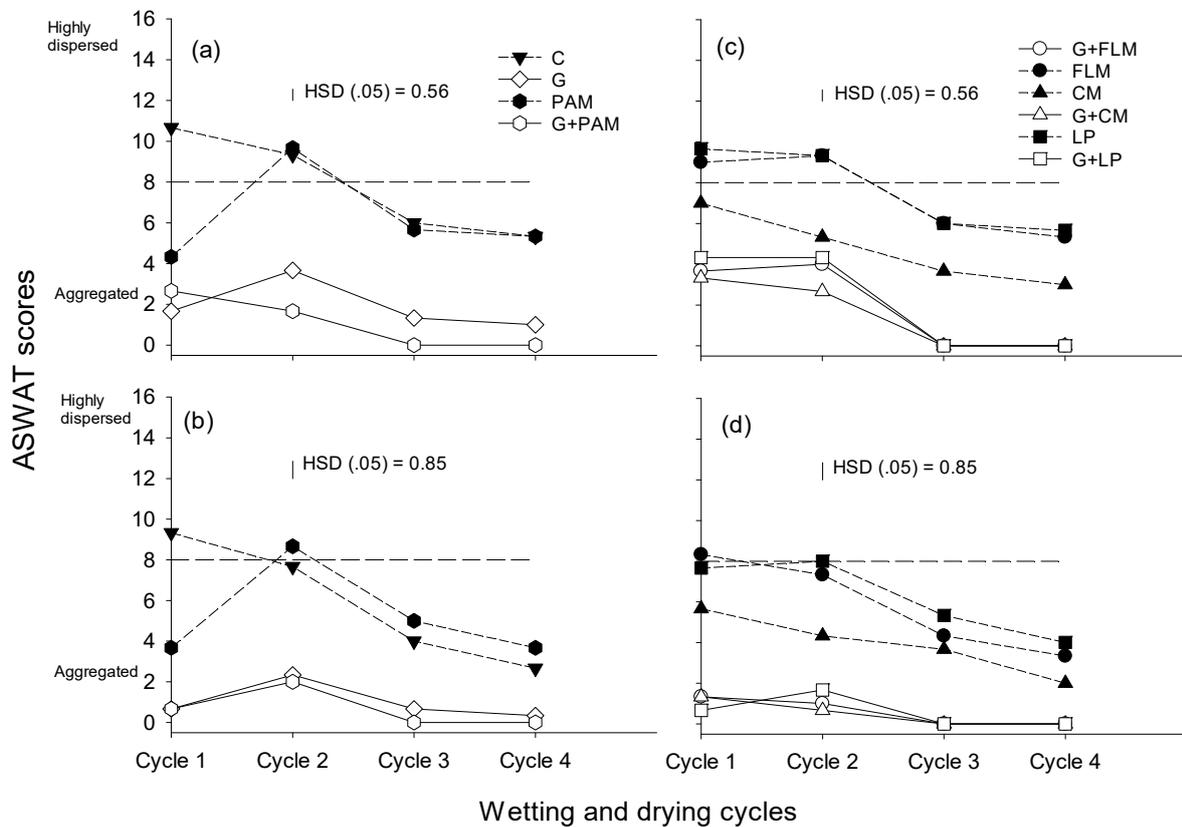
319

320 *Figure 2. The mean weight diameter (MWD) of the soils after addition of amendments during four*
 321 *wetting and drying (WD) cycles. Soil 1 (a, c), and Soil 2 (b, d). The treatments are C: control, G:*
 322 *gypsum, PAM: anionic polyacrylamide, FLM: feedlot manure, CM: chicken manure, LP: lucerne*
 323 *pellets. Vertical bars represent Tukey's honest significant difference (HSD) values at P=0.05 for*
 324 *pairwise comparisons among cycles.*

325 3.2 Soil dispersion dynamics as measured by ASWAT and DI

326 We assessed soil dispersion using both the ASWAT test (Fig. 3) and DI (see Supplementary
 327 Data Fig. S2), with good agreement between the DI results and ASWAT scores (Fig. S3).
 328 Overall, we observed that soil dispersion decreased with WD cycles in both soils. Although,
 329 Soil 1 was more dispersive than Soil 2, both soils showed a similar decrease in dispersion with
 330 WD cycles. Addition of gypsum significantly decreased dispersion in all WD cycles, and this
 331 effect of gypsum was greater than the effect of organic amendments in decreasing soil

332 dispersion. PAM had only a short-term effect, with PAM decreasing dispersion for the first
 333 WD cycle but not thereafter. Of the organic amendments, only CM decreased the dispersion in
 334 both soils, but FLM and LP did not decrease dispersion in both soils in the first and second
 335 WD cycles. Given that both EC (Fig. S4) and SAR (Fig. S5) are known to affect soil dispersion,
 336 it was not surprising that the ASWAT scores were negatively correlated with EC (Fig. S6), and
 337 positively correlated with SAR (Fig. S7). However, the ASWAT scores became less affected
 338 by SAR after second WD cycle, with this highlighting the importance of WD cycles on
 339 structure improvement.

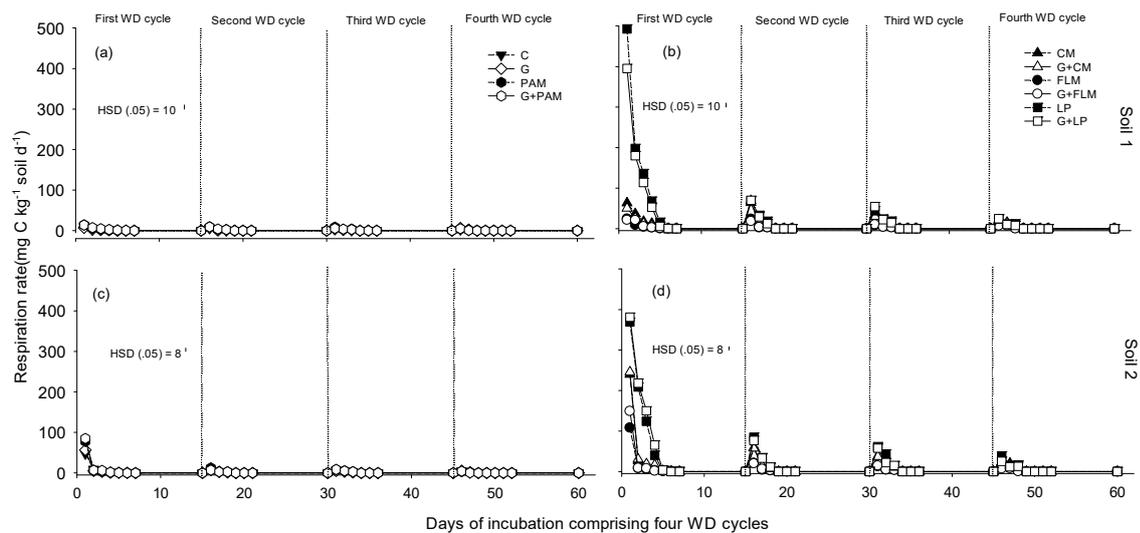


340
 341 *Figure 3. The ASWAT scores of soils after addition of amendments during four wetting and drying*
 342 *(WD) cycles; Soil 1 (a, c), and Soil 2 (b, d). The amendments are C: control, G: gypsum, PAM: anionic*
 343 *polyacrylamide, G+PAM: gypsum+anionic polyacrylamide, FLM: feedlot manure, G+FLM:*
 344 *gypsum+feedlot manure, CM: chicken manure, G+CM: gypsum+chicken manure, LP: lucerne pellets,*

345 *G+LP: gypsum+lucerne pellets*). Vertical bars represent Tukey's honest significant difference (HSD)
346 values at $P=0.05$ for pairwise comparisons among cycles.

347 **3.3 Soil microbial respiration**

348 Soil WD cycles impose a significant stress on the soil microbial community. The addition of
349 treatments resulted in a significant increase ($p<0.001$) in soil microbial respiration for both the
350 soils (Fig. 4). Rewetting of the dried soils caused a transient increase in microbial respiration
351 (Fig. 4), with respiration being highest during the first WD cycle and decreasing during
352 subsequent WD cycles. The lowest microbial respiration rates were observed in the fourth WD
353 cycle in both soils (Fig. 4). The highest microbial respiration rates were observed in soils
354 amended with LP or CM in the first WD cycle, whereas FLM had little effect on microbial
355 respiration, whilst PAM and gypsum had no effect on microbial respiration.



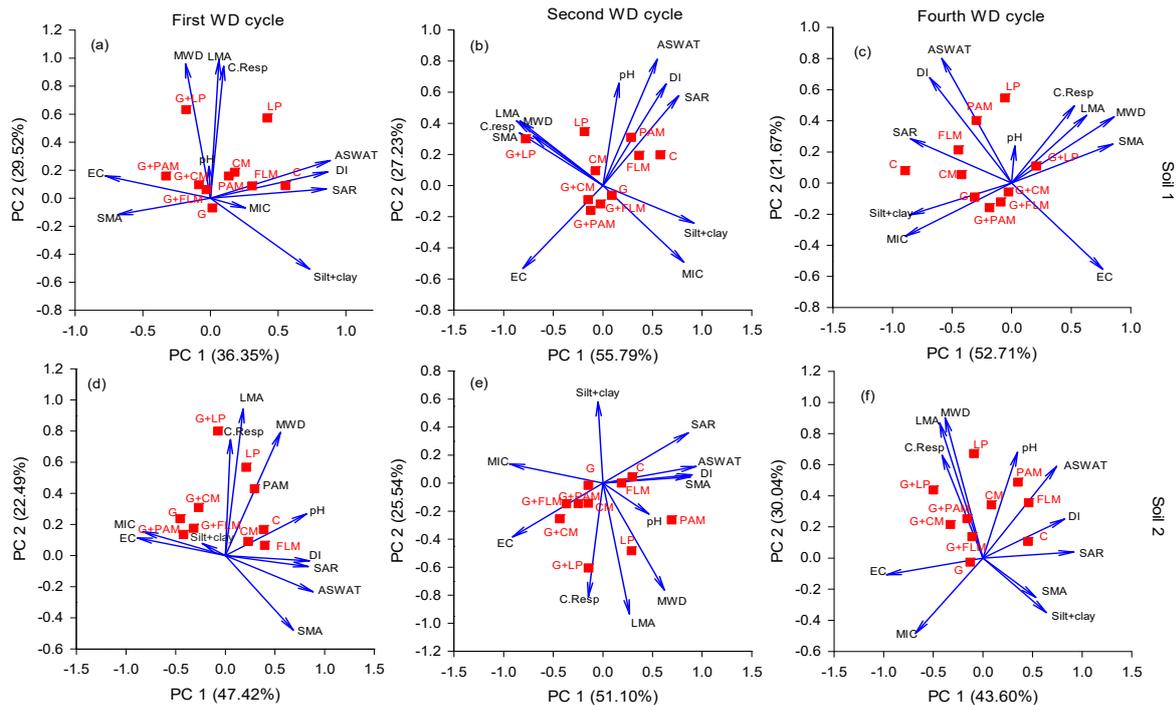
356

357 *Figure 4. Changes in soil respiration rate after addition of amendments during four alternate WD*
 358 *cycles; Soil 1 (a, b) Soil 2 (c, d) The treatments were C: control, G: gypsum, PAM: anionic*
 359 *polyacrylamide, G+PAM: gypsum+anionic polyacrylamide, FLM: feedlot manure, G+FLM:*
 360 *gypsum+feedlot manure, CM: chicken manure, G+CM: gypsum+chicken manure, LP: lucerne pallets,*
 361 *G+LP: gypsum+lucerne pallets. Vertical bars represent Tukey's honest significant difference (HSD)*
 362 *values at P=0.05 for pairwise comparisons among the four WD cycles. The vertical dotted lines are*
 363 *showing the days when deionised water was added to start the next WD cycle.*

364 3.4 Relationship between soil physical, chemical, and microbial properties

365 Soil properties determined for both soils after the first, second, and fourth WD cycles (Fig. 5)
 366 were used to examine inter-relationships by PCA. Overall, the PCA biplots indicated that soil
 367 microbial respiration, MWD, and the proportion of large macroaggregates were positively
 368 correlated with each other. A positive correlation between ASWAT, DI with SAR was
 369 observed in both soils, whereas EC had a negative correlation with ASWAT and DI.
 370 Surprisingly, there was not a strong correlation between silt+clay and DI or ASWAT scores.
 371 The effect of SAR and EC on soil dispersion decreased after the second WD cycle (Fig. 5),
 372 underlining the importance of WD cycles on structure formation and stability, which offsets
 373 the detrimental effects of SAR on stability. Obvious treatment effects on different soil
 374 properties could also be seen in each WD cycle (Fig. 5). The G+LP and LP treatments were

375 positively correlated with microbial respiration, the proportion of large macroaggregates and
 376 MWD in all the cycles. The gypsum added with and without organic amendments produced
 377 the highest EC in all the cycles. The control soils had highest SAR, ASWAT scores and DI
 378 followed by FLM and LP. It was also observed that the proportion of large macroaggregates
 379 and the MWD were negatively correlated with the silt+clay fraction in each WD cycle.



380
 381 *Figure 5. Principal component analysis (PCA) biplot of the physicochemical and microbial properties*
 382 *effected by different treatments in two soils after first, second, and fourth WD cycle: Soil 1(a, b, and c)*
 383 *and Soil 2 (d, e, and f). The treatments are C: control, G: gypsum, PAM: anionic polyacrylamide, FLM:*
 384 *feedlot manure, CM: chicken manure, LP: lucerne pellets. The data presented in these PCA biplots are*
 385 *the mean data of all the days for each parameter (blue arrows) and factorial scores of all the treatments*
 386 *(red squares). The parameters are ASWAT scores (aggregate stability in water test), DI (Dispersion*
 387 *index), C. Resp (cumulative microbial respiration), SAR (sodium adsorption ratio), EC (electrical*
 388 *conductivity 1:5), LMA (large macroaggregates), SMA (small macroaggregates), MIC*
 389 *(microaggregates), Silt+Clay, and MWD (mean weight diameter).*

390 4 Discussion

391 The results of this experiment showed clear differences in the role of organic amendments,
392 gypsum (Ca), and WD cycles on the formation and stabilisation of soil aggregates. We showed
393 that the formation and stability of large macroaggregates was controlled by the type of organic
394 amendments, whereas the WD cycles increased the formation and stability of small
395 macroaggregates. Formation of microaggregates is beneficial in that it decreases dispersion,
396 but microaggregates may not be sufficient to improve soil water infiltration (Collis-George and
397 Greene 1979; Nemati *et al.* 2002). Regardless, even a small increase either in large
398 macroaggregates and small macroaggregates may increase infiltration, with this facilitating the
399 leaching of excess Na. Here, we discuss the importance of each factor (WD cycles, organic
400 amendments, and Ca) individually.

401 4.1 The relative role of WD cycles on aggregation

402 The WD cycles result in rearrangement of pores and soil particles and may lead to increased
403 rigidity and stability of soil aggregates (Horn *et al.* 2014). We observed a marked change in
404 aggregate size distribution with repeated WD cycles (Fig. 1), from macroaggregates (large
405 macroaggregates and small macroaggregates) at the completion of the first WD cycle, to
406 microaggregates at the completion of the second WD cycle, and back to macroaggregates (large
407 macroaggregates and small macroaggregates) at the completion of the fourth WD cycle. We
408 suggest that extracellular polysaccharides formed by microbial activity (indicated by soil
409 microbial respiration, Fig. 3) are responsible for the formation of large macroaggregates at the
410 completion of the first WD cycle. After the second WD cycle, the microbial activity greatly
411 decreased (Fig. 3) and macroaggregates (large macroaggregates and small macroaggregates)
412 were broken down into microaggregates and silt+clay. This allowed the soil particles to settle
413 into tightly packed configurations, resulting in stronger interconnections upon WD cycles
414 (Kemper and Rosenau 1984). Macroaggregates were more susceptible to disintegration during

415 wet sieving compared to microaggregates at the completion of second WD cycle. By the fourth
416 WD cycle, some rearrangements of soil particles likely occurred, facilitated by soil drying,
417 thereby rebuilding macroaggregates. The decrease in large macroaggregates (Fig.1) at the
418 completion of the first WD cycle is also consistent with the findings of Deneff *et al.* (2001),
419 Rahman *et al.* (2018) and Zhang *et al.* (2022). Macroaggregates are expected to be more
420 susceptible to disintegration due to water content changes, because of the large number of
421 planes of weakness and greater angular momentum (Kay, 1990; Deneff *et al.* 2001; Zhang *et al.*
422 2022).

423 The changes observed in the proportion of small macroaggregates followed the same pattern
424 as large macroaggregates, with an initial increase in the proportion of small macroaggregates
425 when the soils were treated with organic amendments. In contrast to these results, no significant
426 differences were observed when the same soils were treated with organic amendments under
427 continuous wet conditions (Niaz *et al.*, 2022). Also, the increase in proportion of larger
428 aggregates (large macroaggregates and small macroaggregates) is 2x greater when the same
429 soils were exposed to WD cycles as compared to constant wet regime (Niaz *et al.*, 2022). In
430 this study, the proportion of small macroaggregates did not increase after the second WD cycle,
431 but the proportion of microaggregates and silt+clay fraction increased, suggesting that upon
432 repeated WD cycles, large macroaggregates break down into microaggregates and silt+clay
433 fractions. These observations support the finding that macroaggregates are composed of
434 microaggregates (Six *et al.*, 2000a). In addition, we observed that the control soils (where no
435 amendments were added) also showed an increase in the proportion of small macroaggregates
436 with WD cycles. This highlights the importance of WD cycles and suggests that the formation
437 of small macroaggregates is less dependent on the organic matter content, in agreement with
438 the PCA biplots (Fig. 5). The significant increase in the proportion of small macroaggregates
439 in control soils after the fourth WD cycle also indicates that there could be an increase in

440 interparticle bond strength with ageing or time (Utomo and Dexter 1982; Dexter 1988; Kong
441 *et al.* 2005; Bravo-Garza *et al.* 2009).

442 **4.2 The role of organic amendments on soil respiration and aggregation**

443 The addition of organic amendments provided an energy and nutrient source for
444 microorganisms and resulted in increased respiration rates, although we observed that
445 respiration rate decreased with repeated WD cycles (Fig. 4), presumably due to depletion of
446 easily metabolisable organic compounds (Harrison-Kirk *et al.* 2013; Yu *et al.* 2014; Rahman
447 *et al.* 2018; Fraser *et al.* 2016; Brangari *et al.* 2022; Zhang *et al.* 2022). In contrast to LP, CM
448 had a modest effect on microbial respiration, whereas FLM had no significant effect on
449 microbial respiration as it was already decomposed. Rewetting of dry soil led to immediate
450 flush of microbial respiration (Fig. 4) similar to the results of Brangari *et al.* (2022). This could
451 be attributed to the slaking of larger aggregates (Fig. 1), thereby exposing some occluded
452 organic matter, or due to the utilization of substrates that become available upon rewetting (Wu
453 and Brookes 2005; Borken and Matzner 2009; Yu *et al.* 2014; Zhang *et al.* 2022). These
454 available (labile) substrates after rewetting include remnants of the added organic matter and
455 microbial biomass (Wu and Brookes 2005; Zhang *et al.* 2022). Over time, with successive WD
456 cycles, it is likely that the labile organic C pool was either exhausted or became physically
457 protected within aggregates and hence became less accessible to microbial degradation (Zhang
458 *et al.* 2022).

459 For the LP and G+LP treatments, where the increase in respiration rate was ~4-fold greater in
460 Soil 1 and ~2 fold greater in Soil 2 than for any other treatment (Fig. 4), we observed that the
461 soils also had a significant increase in the proportion of large macroaggregates (Fig. 1) and
462 MWD (Fig. 2), suggesting that labile organic compounds are important for structure formation
463 (also confirmed by PCA biplots Fig. 5). The formation of large macroaggregates after
464 incorporation of organic amendments, as observed here for LP, has also been reported by Deneff

465 *et al.* (2001), Bravo-Garza *et al.* (2009) and Rahman *et al.* (2018). Although the maximum
466 respiration rate was observed during the first WD cycle for LP and G+LP, the proportion of
467 large macroaggregates and MWD increased at the end of fourth WD cycle. During later WD
468 cycles, it is likely conversion of readily metabolisable organic compounds (e.g., microbial
469 polymers) to more resistant forms resulted in the formation of macroaggregates (large
470 macroaggregates and small macroaggregates), but in this case, aggregation may have been
471 caused by more hydrophobic compounds. However, the proportion of large macroaggregates
472 did not increase much as compared to the first WD cycle, likely because microbial activity was
473 lower. One of the possible reasons of increased MWD could be the accumulation of microbial
474 binding agents over time that are released continuously from microbial activity due to organic
475 matter decomposition (Rahman *et al.* 2018).

476 When the soils were treated with organic amendments, a decrease in dispersion was observed,
477 but was not significantly different from control except in CM treated soils. The possible reason
478 for increased stability in CM treated soils was its higher Ca content (Table 2), increased EC
479 (Fig. S4) and decreased SAR (Fig. S5). For the remaining organic amendments viz; FLM, LP,
480 and PAM, an improvement in aggregate stability was observed after the second WD cycle.
481 Although LP had the highest MWD, it still did not reduce dispersion in the first WD cycle. We
482 suggest that high MWD in itself is a misleading measure of soil stability since only a few large
483 aggregates can result in a high MWD (Niaz *et al.*, 2022). Some dispersion can occur in parallel
484 which is sensitively detected by the ASWAT test, but not by the MWD. Both assays detect
485 different physical processes - hence a high MWD (Fig. 1) and dispersion (Fig. 2) are not
486 mutually exclusive (Niaz *et al.*, 2022). Organic amendments rich in aliphatic compounds or
487 waxes can lead to hydrophobicity of aggregate surfaces during the drying step which slows
488 down the wetting of aggregate surfaces, reducing the disruption caused during rewetting
489 (Piccolo and Mbagwu 1999; Borken and Matzner 2009). Monnier (1965) proposed that fresh

490 organic amendments can increase aggregate stability in the time frame of weeks and months as
491 compared to already decomposed stable organic amendments. But in our study, we found no
492 significant differences in dispersion after the addition of LP (fresh) and FLM (partially
493 decomposed). These findings suggest that there might be some other mechanisms which are
494 responsible for the binding of organic matter with clay particles to control dispersion, such as
495 the size of the organic matter molecule and charge density. This requires further investigation.

496 **4.3 The relative role of Ca (gypsum) in improving aggregate stability**

497 The results of this experiment showed that addition of gypsum (Ca) significantly reduced soil
498 dispersion and increased aggregate stability. This improvement in aggregate stability is because
499 of the increased EC (ionic strength, Fig. S4) and decreased SAR (Fig. S5) after the addition of
500 gypsum. The increased EC likely resulted in the flocculation of soil particles by reducing the
501 diffuse double layer (van Olphen 1977, Ghosh et al., 2010, Bennett et al. 2015). Improved
502 stability was also observed when organic amendments were applied with gypsum, especially
503 in G+PAM, G+LP, and G+FLM treated soils. The PAM had an initial positive effect but led to
504 decreased stability at completion of the second WD cycle. Although the addition of gypsum
505 increased aggregate stability, it was observed that addition of gypsum did not affect the
506 proportion of large macroaggregates and MWD. However, when organic amendments were
507 added with gypsum an improvement in proportion of large macroaggregates and MWD was
508 observed in G+LP treated soils. This can be explained as Ca-bridging effect through which
509 clay particles are attached to organic matter and polyvalent cations, resulting in the formation
510 of macro and micro aggregates (Wuddivira and Camps-Roach 2007).

511 **Conclusion**

512 The stability of dispersive sodic Vertisols was improved by the application of organic
513 amendments and gypsum, which was further enhanced by WD cycles. Gypsum reduced soil
514 dispersion but did not affect the proportion of large macroaggregates and MWD. We observed

515 that not all organic amendments were equally beneficial in improving soil aggregation and
516 aggregate stability. LP significantly increased the proportion of large macroaggregates
517 compared to FLM and PAM. In contrast, CM significantly reduced soil dispersion as it had
518 higher calcium content. It was also found that PAM only had a transient effect in controlling
519 dispersion. In the absence of organic amendments, repeated WD cycles reduced the dispersion
520 of sodic soils, but when organic amendments were added (with or without gypsum) soil
521 aggregation and soil stability was improved even more. It is likely that soil microbial activity
522 contributed to the aggregate formation. Implementation of these findings in the field would
523 favour the use of organic amendments with gypsum to improve the physicochemical properties
524 of sodic soils, which is further enhanced by WD cycles. The aim should be initially to prevent
525 soil dispersion which can be achieved by the application of Ca (through application of gypsum)
526 and then to build larger aggregates which can be achieved by the application of organic
527 amendments.

528 **Author contribution**

529 Conceptualization: Niaz. S., Wehr, J. B. ., Kopittke, P.M., Dalal, R.C., and Menzies, N.W.;;
530 Formal analysis, investigation, and writing original draft: Niaz. S; Review and editing: Wehr,
531 J. B., Kopittke, P.M., Dalal, R.C., and Menzies, N.W.; Supervision: Wehr, J. B., and Menzies,
532 N.W.

533 **Competing interests**

534 The authors declare that they have no conflict of interest.

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